

Supporting Information

The first ratiometric fluorescent probes for aminopeptidase N cell imaging

Laizhong Chen, Wei Sun, Jing Li, Zhenzhen Liu, Zhao Ma, Wei Zhang, Lupei Du, Wenfang Xu, Hao

Fang and Minyong Li*

Department of Medicinal Chemistry, Key Laboratory of Chemical Biology of Natural Products (MOE), School of Pharmacy, Shandong University, Jinan, Shandong 250012, China

*Tel./fax: +86-531-8838-2076, E-mail: mli@sdu.edu.cn

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1. Synthesis of 6c and 6d.

Methyl 6-bromo-2-ethyl-1*H*-benzo[*de*]isoquinoline-1,3(2*H*)-dione (1a). The solution of 4-bromo-1,8-naphthalic anhydride (13.8 g, 50 mmol) and ethanamine (3.91 g, 60 mmol) in 500 mL 1,4-dioxane was refluxed at 120°C for 4 h. Then the mixture was cooled and poured to 500 mL water. The yielded yellowish sediment were collected by filtration and dried under 110°C air atmosphere to yield a yellowish solid, 14 g, yield = 91.0%, mp: 159.2-160.1 °C.

6-amino-2-ethyl-1*H*-benzo[*de*]isoquinoline-1,3(2*H*)-dione (2a). The mixture of **1a** (1.5 g, 5 mmol), conc. ammonin (10 mL), copper power (0.032 g, 0.5 mmol) and a stirring bar were sealed in a 30 mL screwed tube and stirred in a oil bath at 100°C for 10 h. After cooling to r.t., the suspension was extraced with 50 mL AcOEt, to the residue was added 50 mL water and extraced by another 50 mL AcOEt. The combination was washed with brine (100 mL*3) and dried over anhydrous MgSO₄ for 24 h. The solution was concentrated under reduced pressure and then purified by combiflash to yield a yellow power, 0.32 g, yield = 34.0%, mp: 280.0-282.3 °C.

2-((((9*H*-fluoren-9-yl)methoxy)carbonyl)amino)propanoic acid (3c). To the solution of Ala (3.5 g, 40 mmol) in 120 mL 10% Na₂CO₃ was added the solution of FmocOSu (13.5 g, 40 mmol) in acetone (100 mL) at 0°C. The mixture was stirred at r.t. over night. Acetone was evaporated under reduced pressure and the residue was wased by ether (100 mL*3). The water phase was adjusted pH to 2 with Conc. HCl and extracted with AcOEt (100 mL*3). The organic phase was dried over anhydrous MgSO₄ for 24 h. The solution was concentrated under reduced pressure and then recrystallized with AcOEt-*n*-hexane to yield a white solid, 10 g, yield = 80.4%, mp: 138.2-139.7 °C.

2-((((9*H*-fluoren-9-yl)methoxy)carbonyl)amino)pentanoic acid (3d). The synthesis is analogous to **3c** with Nva as the starting material. The product was obtained as a white solid. Yield = 79.4%, mp:134.1-135.4 °C.

(9*H*-fluoren-9-yl)methyl (1-chloro-1-oxopropan-2-yl)carbamate (4c). To the solution of **3c** (0.5 g, 1.6 mmol) in CH₂Cl₂ (8 mL) was added SOCl₂ (4 mL), the mixture was stirred at r.t. for 2 h. The solution was concentrated under reduced pressure and recrystallized with CH₂Cl₂-*n*-hexane to yield a white solid, 0.35 g, yield = 66.5%, mp: 94.5-96 °C.

(9*H*-fluoren-9-yl)methyl (1-chloro-1-oxopentan-2-yl)carbamate (4d). The synthesis is analogous to **4c** with **3d** as the starting material. The product was obtained as a white solid. Yield = 71.4%, mp: 95.3-97.1 °C.

(9*H*-fluoren-9-yl)methyl

(1-((2-ethyl-1,3-dioxo-2,3-dihydro-1*H*-benzo[*de*]isoquinolin-6-yl)amino)-1-oxopropa

n-2-yl)carbamate (5c). The suspension of **4c** (0.33 g, 1.0 mmol) and **2a** (0.12 g, 0.5 mmol) in AcOH (10 mL) was refluxed for 2 h. The mixture was cooled and poured into 50 mL water, the yield precipitation was filtered to yield a yellow power. The yellow power was purified by silica gel column to yield a pale-yellow power, 0.11 g, Yield = 41.3%, mp:112.7-113.4 °C.

(9H-fluoren-9-yl)methyl

(1-((2-ethyl-1,3-dioxo-2,3-dihydro-1H-benzo[de]isoquinolin-6-yl)amino)-1-oxopentan-2-yl)carbamate (5d). The synthesis is analogous to **5a** with **4d** and **2a** as the starting material. The product was obtained as a white solid. Yield = 73.5%, mp:95.3-97.1 °C.

2-amino-N-(2-ethyl-1,3-dioxo-2,3-dihydro-1H-benzo[de]isoquinolin-6-yl)propanamide hydrochloride (6c). The mixture of **5c** (0.06 g, 0.11 mmol) and 5 mL 20% piperidine-DMF was stirred at r.t. for 30 min then concentrated under reduced pressure at 90°C to remove excess piperidine and DMF. The residue was purified first by silica gel column and then recrystallization with AcOEt-*n*-hexane to yield a yellow solid. 5 mL EtOAc solution saturated with HCl gas was added and stirred at r.t. for 1 h. The yield precipitation was filtered and washed with AcOEt to yield a light yellow power, 20 mg, yield = 51.1%, mp: 255.3-257.8°C, $[\alpha]_D^{20} = 31.51$ ($c = 0.073$, MeOH). ¹H-NMR (D₂O, 300 MHz): δ 8.07 (d, 1 H, $J = 7.2$ Hz), 7.99 (t, 2 H, $J = 8.1$ Hz), 7.71 (d, 1 H, $J = 8.1$ Hz), 7.51 (t, 1 H, $J = 7.5$ Hz), 4.38 (q, 1 H, $J = 6.9$ Hz), 3.81 (q, 2 H, $J = 7.2$ Hz), 1.73 (d, 3H, $J = 6.9$ Hz), 1.12 (t, 3 H, $J = 7.2$ Hz); ¹³C-NMR (DMSO, 150 MHz): δ 170.3, 163.7, 163.1, 139.5, 131.8, 131.5, 129.9, 128.7, 127.2, 125.2, 122.9, 121.1, 119.2, 49.5, 39.2, 17.8, 13.6; HRMS (ESI) m/z calcd. for C₁₇H₁₈N₃O₃ ($[M + H]^+$) 312.1343; found 312.1338.

2-amino-N-(2-ethyl-1,3-dioxo-2,3-dihydro-1H-benzo[de]isoquinolin-6-yl)pentanamide hydrochloride (6d). The synthesis is analogous to **6c** with **5d** as the starting material. The product was obtained as a light yellow power. Yield =49.5%, mp:277.4-280.0°C. $[\alpha]_D^{20} = 40.00$ ($c = 0.080$, MeOH). ¹H-NMR (D₂O, 300 MHz): δ 8.04 (d, 1 H, $J = 7.5$ Hz), 7.97 (t, 2 H, $J = 8.1$ Hz), 7.69 (d, 1 H, $J = 8.1$ Hz), 7.49 (t, 1 H, $J = 8.1$ Hz), 4.32 (t, 1 H, $J = 6.6$ Hz), 3.79 (q, 2 H, $J = 6.9$ Hz), 2.00-2.09 (m, 2 H), 1.49-1.59 (m, 2 H), 1.02-1.14 (m, 6 H); ¹³C-NMR (DMSO, 150 MHz): δ 169.5, 163.6, 163.1, 139.2, 131.8, 131.5, 129.7, 128.7, 127.3, 125.2, 122.9, 121.3, 119.4, 54.4, 34.5, 32.3, 18.2, 14.1, 12.8; HRMS (ESI) m/z calcd. for C₁₉H₂₂N₃O₃ ($[M + H]^+$) 340.1656; found 340.1658.

2. Time course scan of 6c, 6d to APN

To the solution of **6c** or **6d** (80 μL , 250 μM) in 0.1 M tris-HCl buffer at pH 7.5 was added the solution of APN (20 μL , 10 IU/mL). The mixture was scanned for 30 min with excitation at 400 nm every two minutes by Varioskan microplate reader.

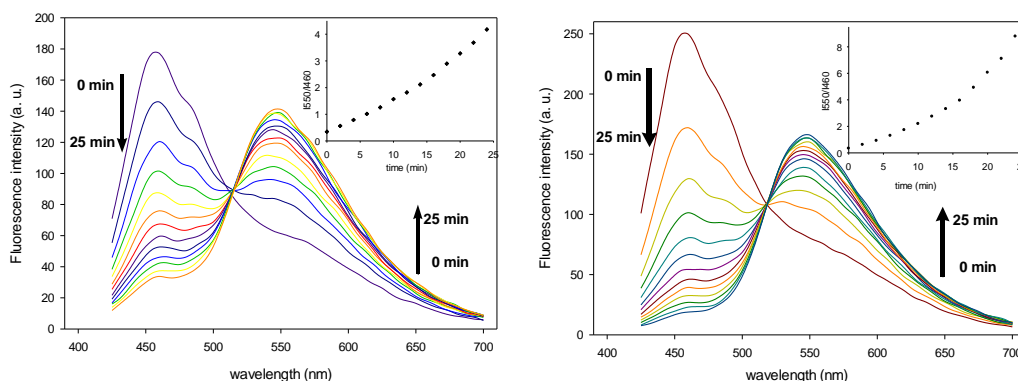
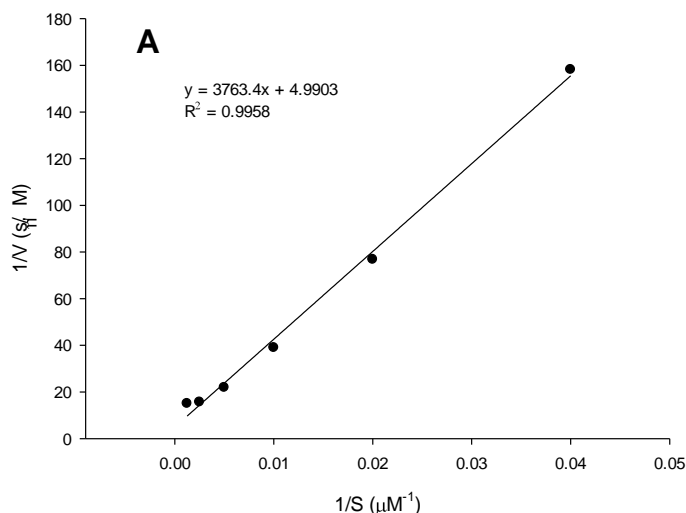


Figure S1 Emission spectra of **6c** (left) and **6d** (right) (200 μM) in the presence of APN (0.2 IU) in a tris-HCl buffer (100 mM, pH 7.5). Insets: ratios of fluorescent intensities at 550 and 460 nm as a function of time after adding APN ($\lambda_{\text{ex}} = 400 \text{ nm}$).

3. Lineweaver-Burk plot for V_{max} and K_{m} measurement



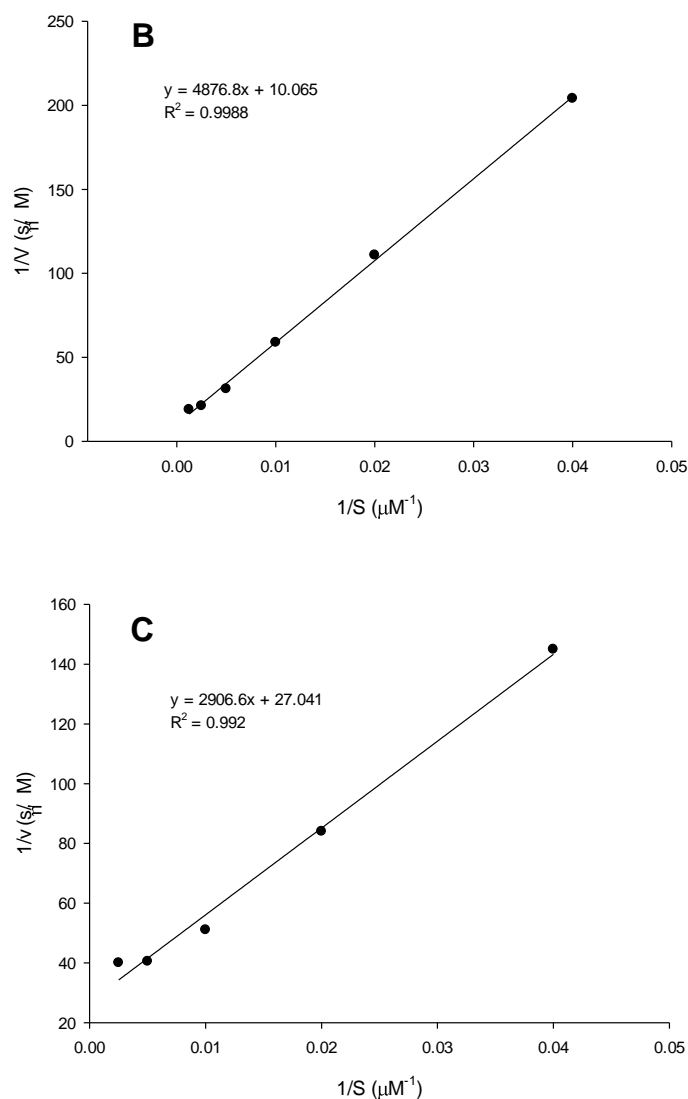


Figure S2 Lineweaver-Burk plot of **6c** (A), **6d** (B) and **6e** (C).

4. Stability of **6c**, **6d** and **6e**

Stability of **6c**, **6d** and **6e** in tris-HCl buffer, artificial gastric juice and artificial intestinal juice was tested by measurement of the time course for the changing of ratios of fluorescent intensities at 550 and 460 nm (excitation at 400 nm) in a solution of 100 μM .

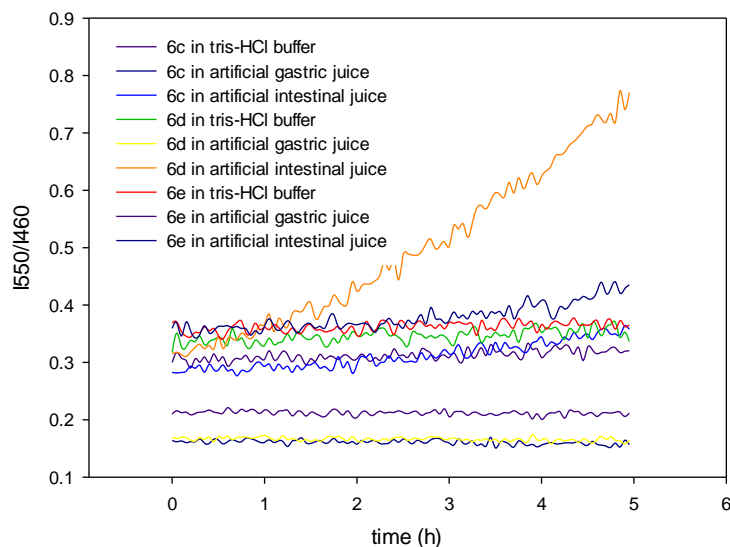


Figure S3 Time course for the generation of ratios of fluorescent intensities at 550 and 460 nm (excitation at 400 nm) of **6c**, **6d** and **6e** in the solution of tris-HCl buffer (100 mM, pH=7.5), artificial gastric juice and artificial intestinal juice (100 μ M).

5. ^1H NMR, ^{13}C NMR and HR-MS spectra

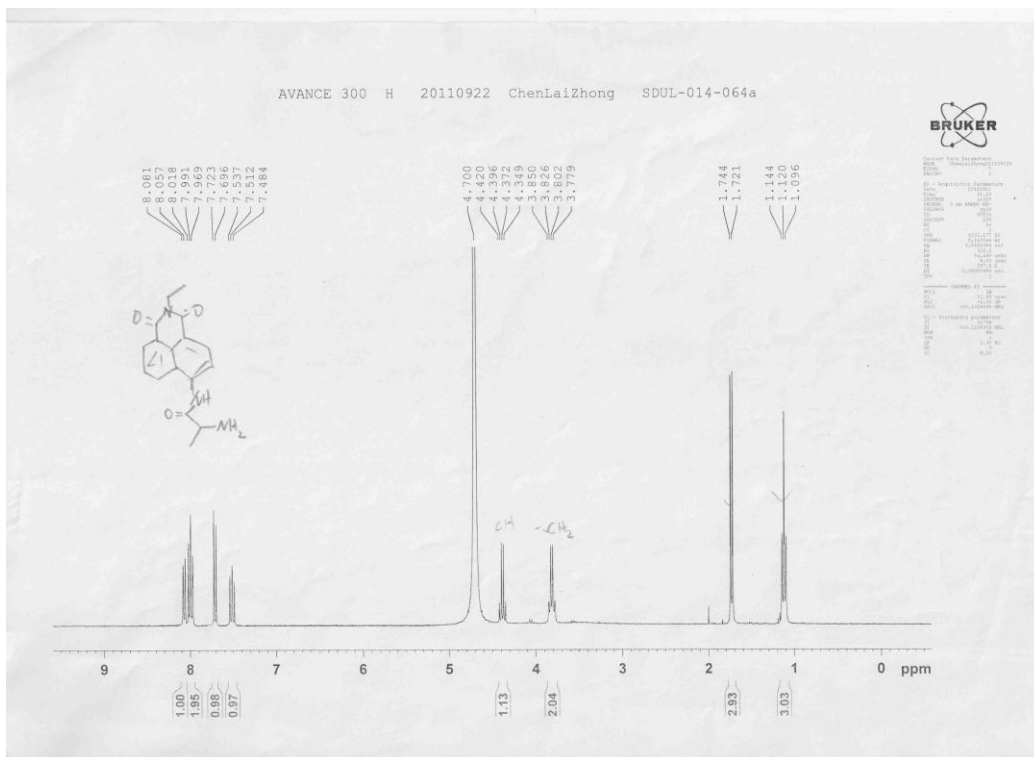


Figure S4 ^1H NMR (300 MHz) spectrum of Probe **6c**.

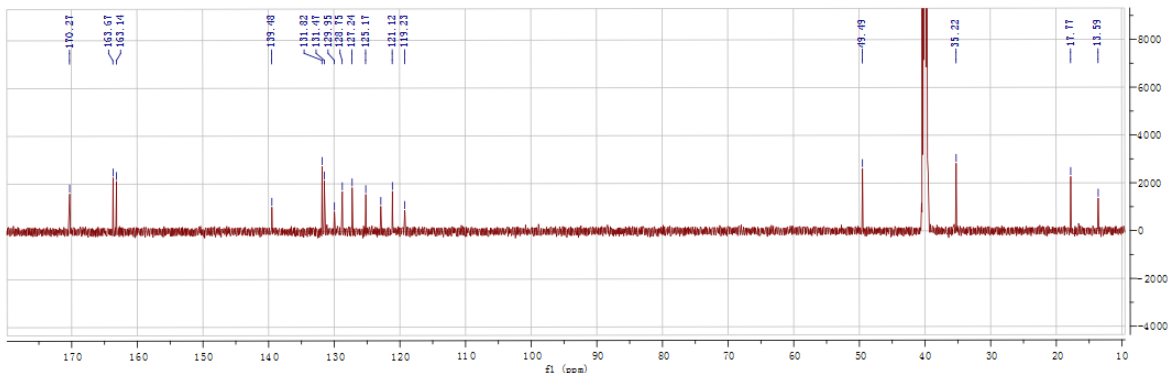


Figure S5 ¹³C NMR (150 MHz) spectrum of Probe 6c.

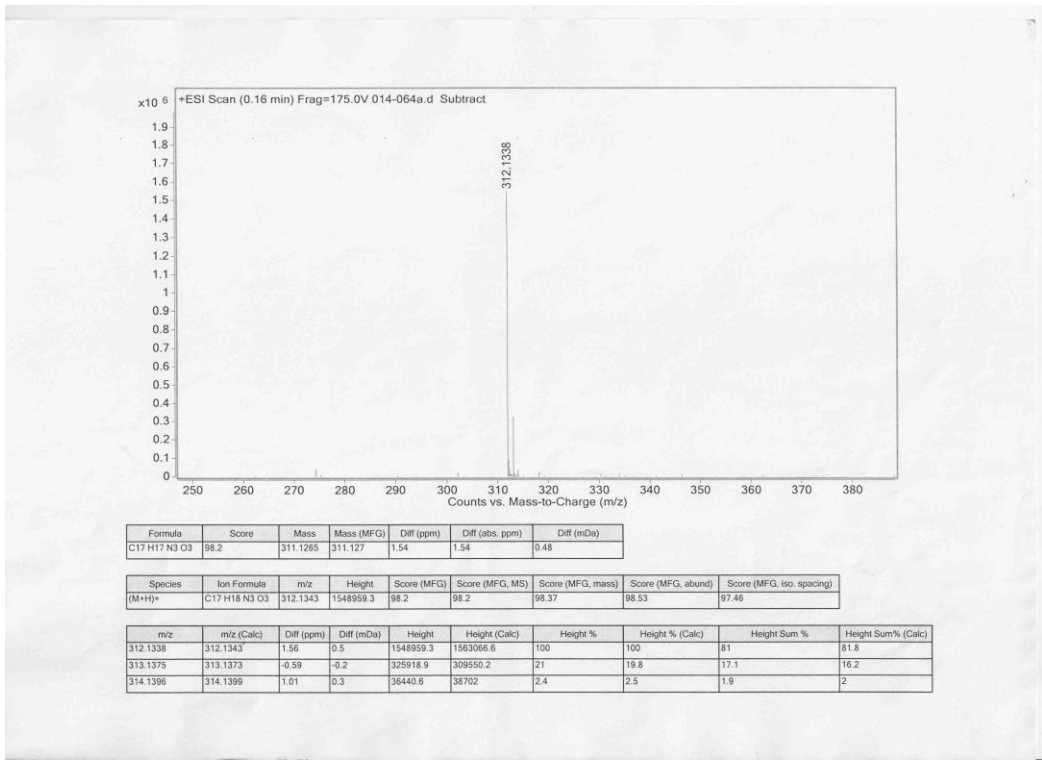


Figure S6 High resolution mass spectrum of Probe 6c.

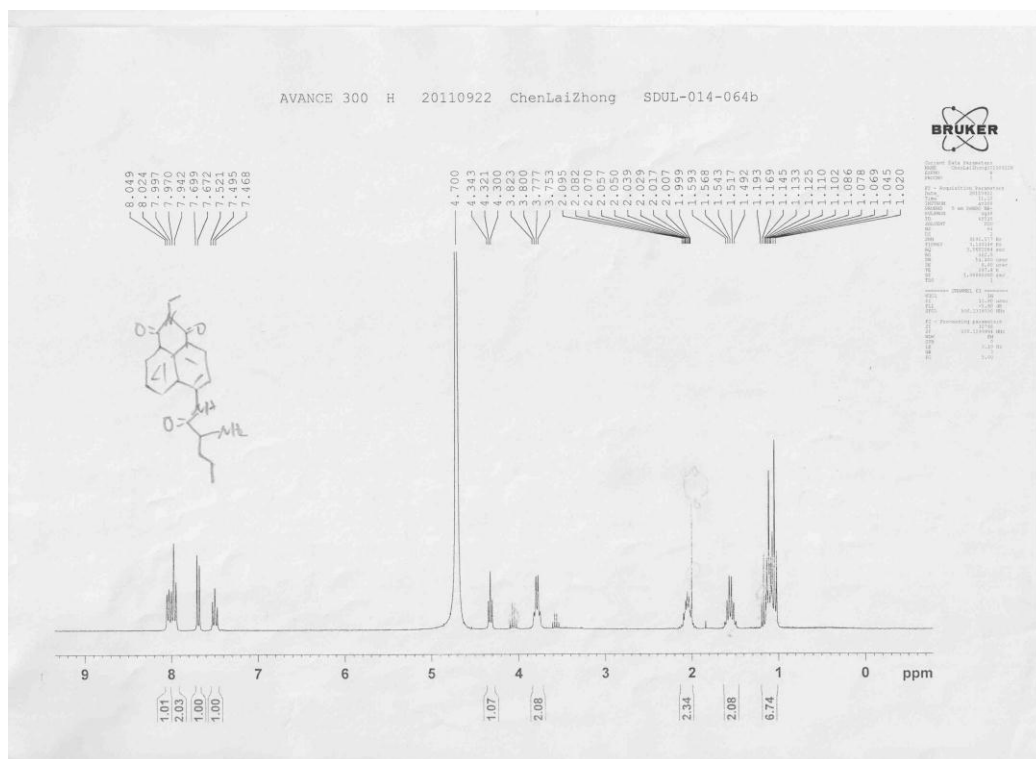


Figure S7 ¹H NMR (300 MHz) spectrum of Probe 6d.

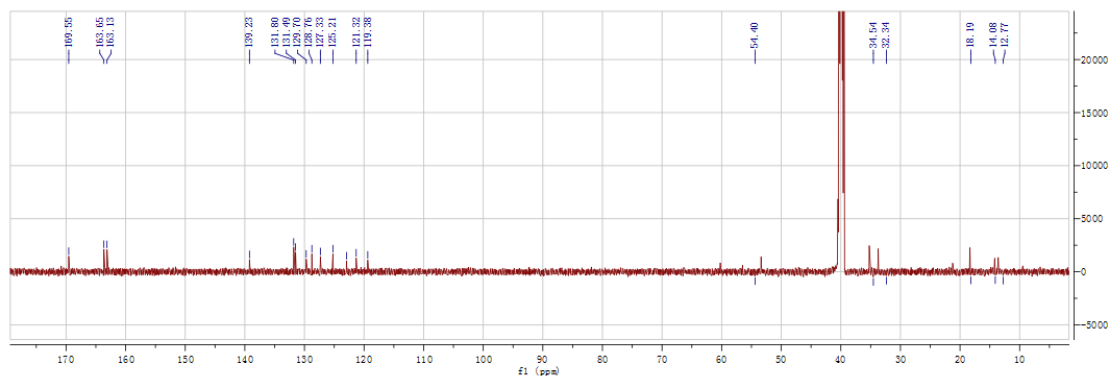


Figure S8 ¹³C NMR (150 MHz) spectrum of Probe 6d.

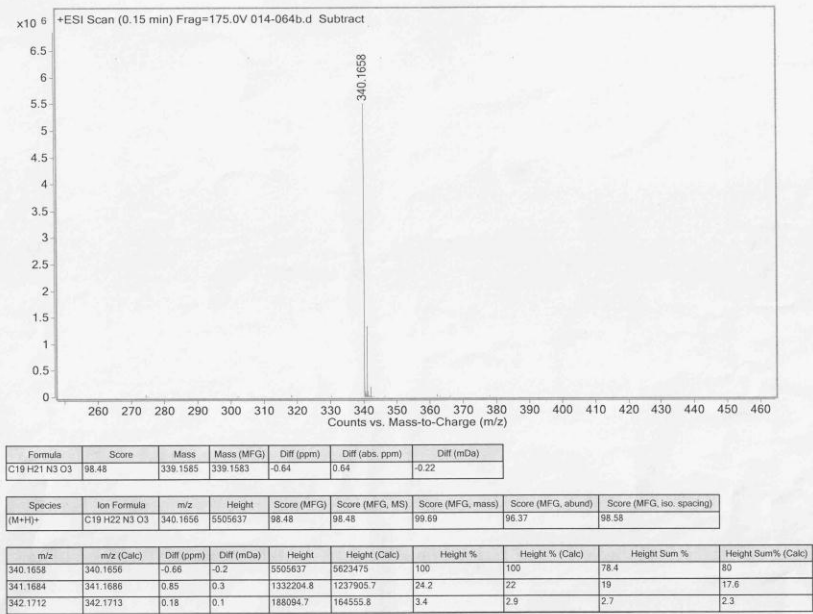


Figure S9 High resolution mass spectrum of Probe **6d**.

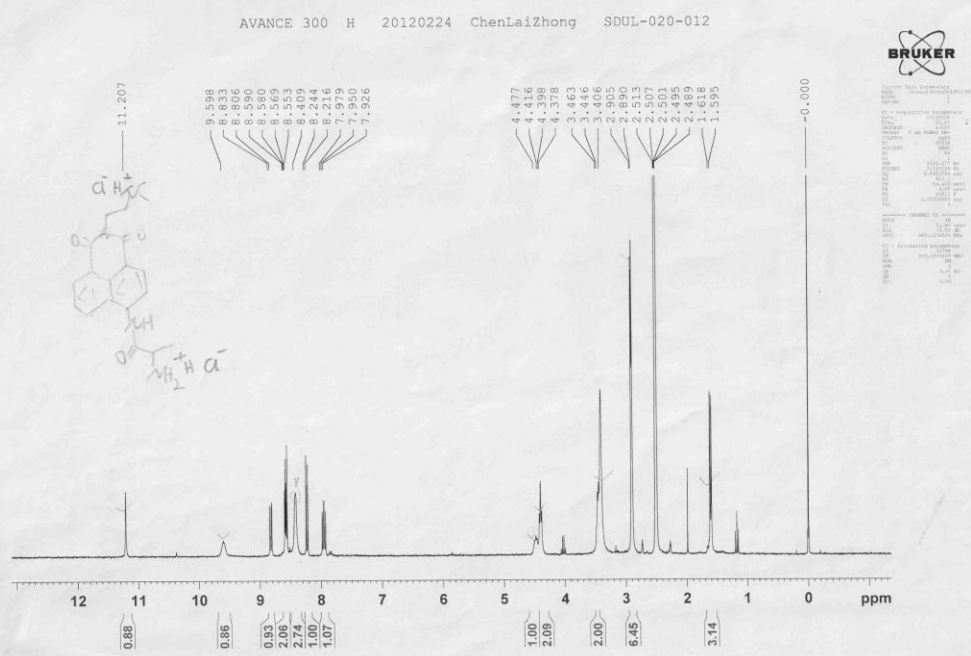


Figure S10 ^1H NMR (300 MHz) spectrum of Probe **6e**.

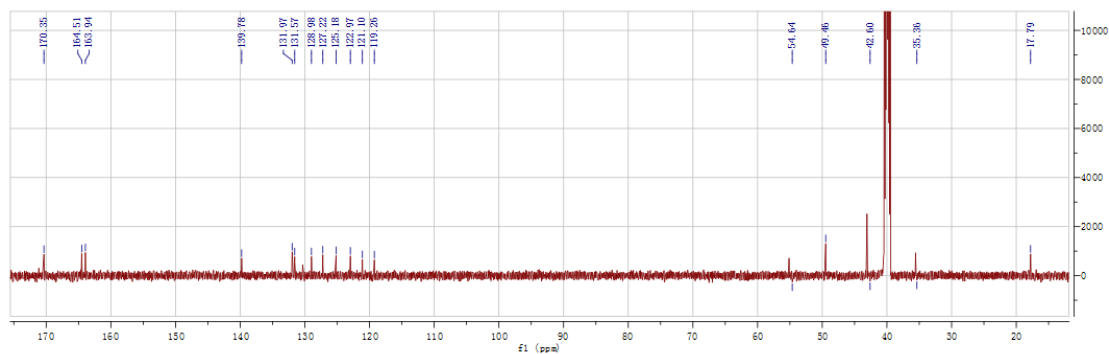


Figure S11 ¹³C NMR (150 MHz) spectrum of Probe **6e**.

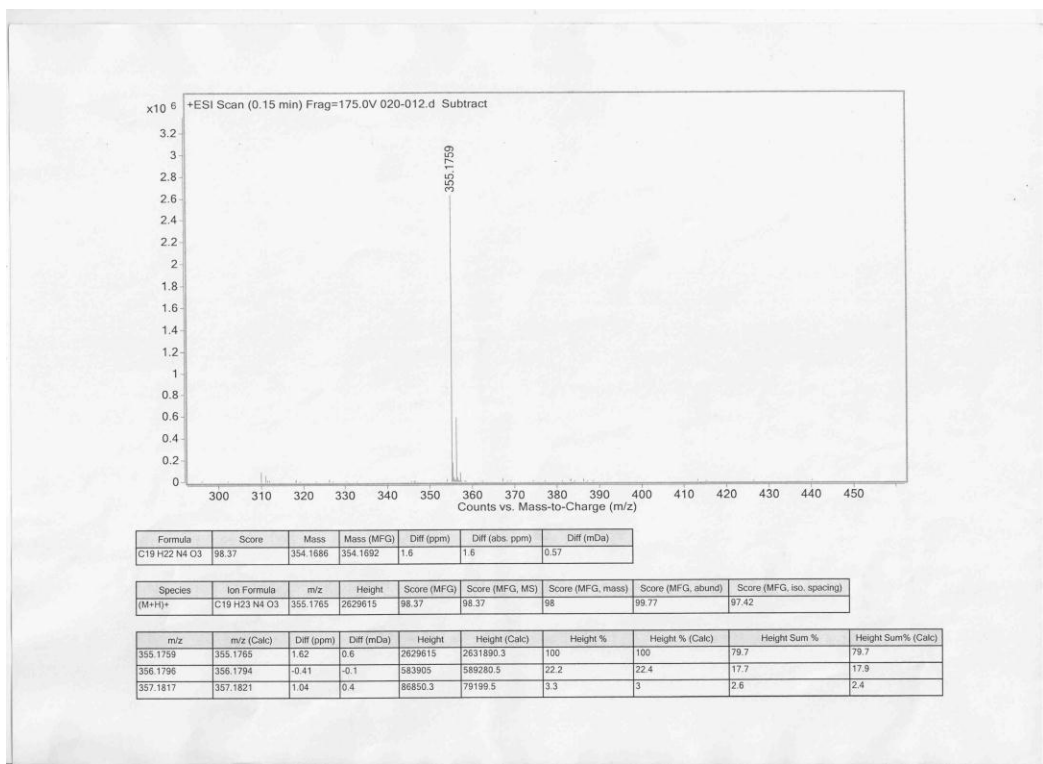


Figure S12 High resolution mass spectrum of Probe **6e**.