

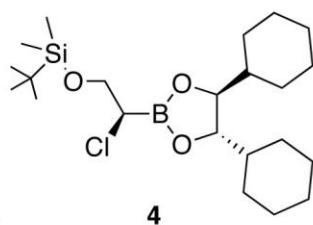
Synthesis and Properties of 1-(3-Dihydroxyboryl-2,3-dideoxyribosyl)pyrimidines”

Byung Ju Kim, Jinhua Zhang, Shenglan Tan, Donald S. Matteson\*, William H. Prusoff, and Yung-chi Cheng\*

Supporting Information

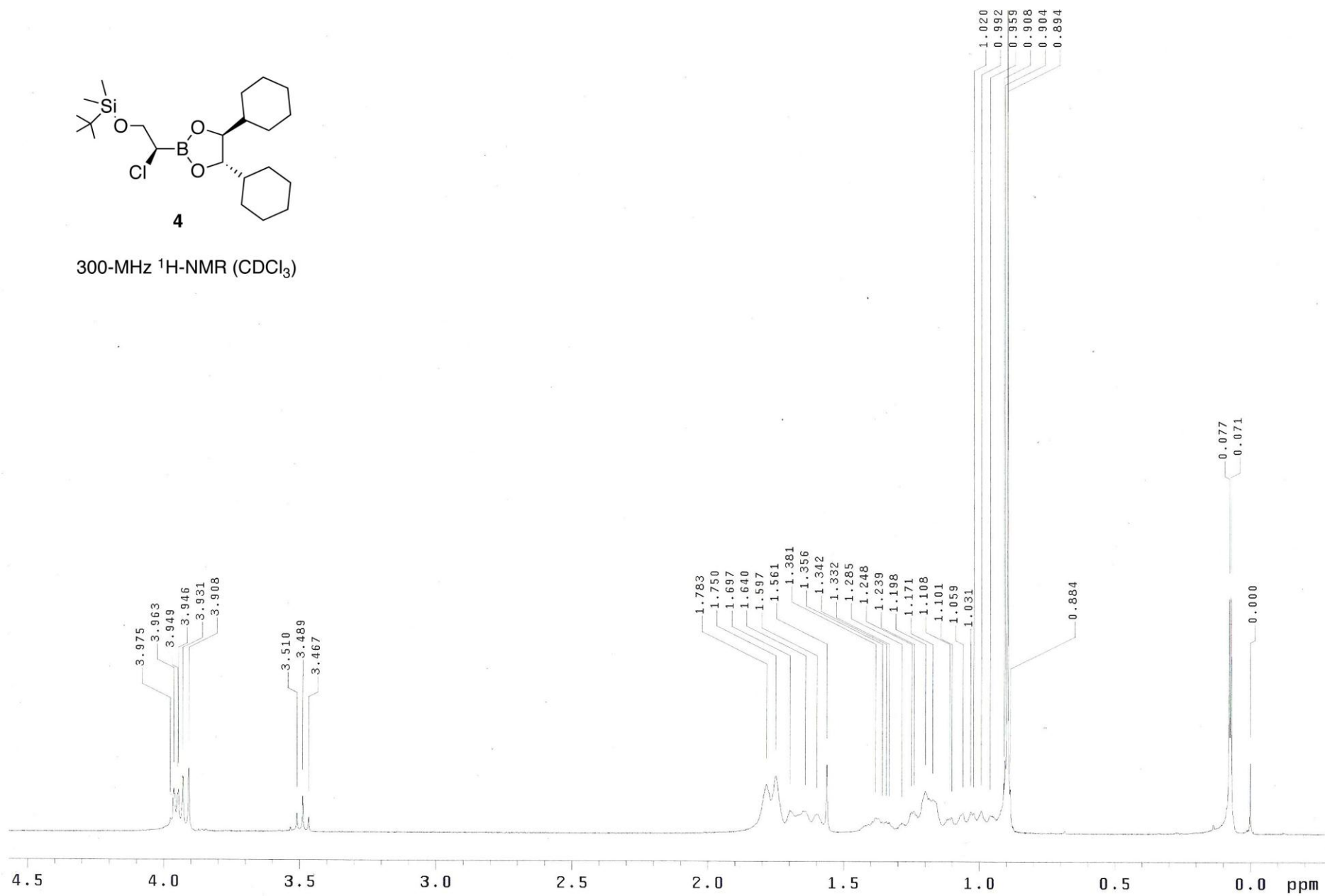
NMR spectra (in CDCl<sub>3</sub> if not otherwise specified); plots of cytotoxicity data and degradation of β-14b and β-14c.

Compound number	First NMR Page	Compound number	First NMR Page	Decomposition Studies	Page
4	2	13b	32	β-14b (BFUdR)	62
5	3	13c	36	β-14b (BFUdR) cytotoxicity data	63
6	5	13d	40	14a (BdThd)	64
7	7	14a	41	14c (BIdU)	65
8	9	14b	45	14d (BdCyd)	66
9	11	14c	49	Summary of all stabilities	66
10	13	14d	53		
11	15	16	54		
12a	17	17	55		
12d	21	18	56		
13a	23	19	59		
Thymidine from 13a	29				



4

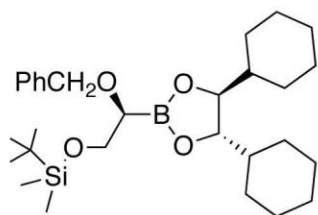
300-MHz  $^1\text{H}$ -NMR ( $\text{CDCl}_3$ )



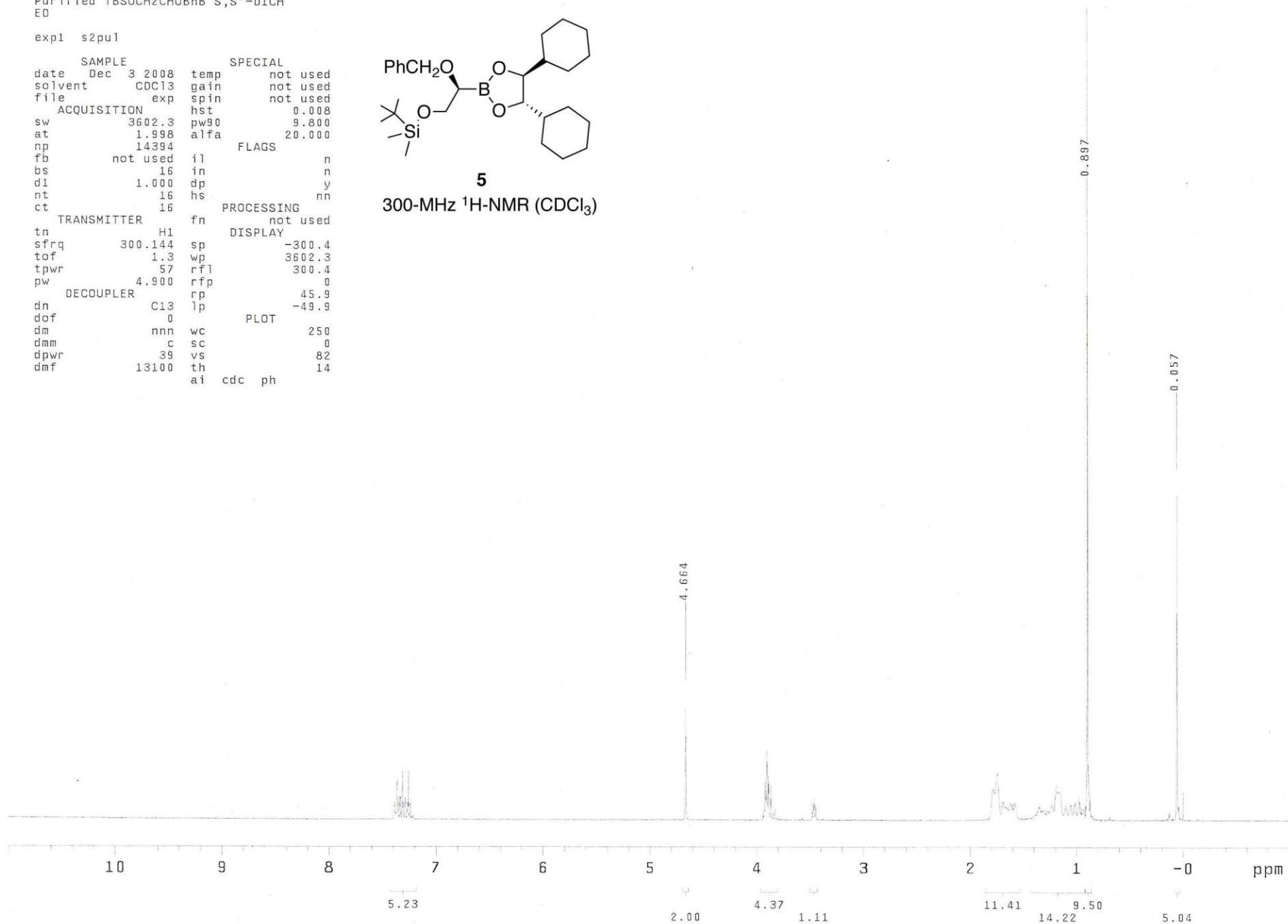
Purified TBSOCH<sub>2</sub>CHOBnB S,S'-DICH  
ED

exp1 s2pu1

SAMPLE		SPECIAL	
date	Dec 3 2008	temp	not used
solvent	CDC13	gain	not used
file	exp	spin	not used
ACQUISITION		hst	0.008
sw	3602.3	pw90	9.800
at	1.998	alfa	20.000
np	14394	FLAGS	
fb	not used	il	n
bs	16	in	n
dl	1.000	dp	y
nt	16	hs	nn
ct	16	PROCESSING	
TRANSMITTER		fn	not used
tn	H1	DISPLAY	
sfrq	300.144	sp	-300.4
tof	1.3	wp	3602.3
tpwr	57	rfl	300.4
pw	4.900	rfp	0
DECOUPLER		rp	45.9
dn	C13	lp	-49.9
dof	0	PLOT	
dm	nnn	wc	250
dmm	c	sc	0
dpwr	39	vs	82
dmf	13100	th	14
	ai	cdc	ph



**5**  
300-MHz <sup>1</sup>H-NMR (CDCl<sub>3</sub>)

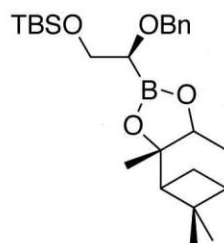


4

Transesterified  
TBSOCH<sub>2</sub>CHOBn pinanediol boronate

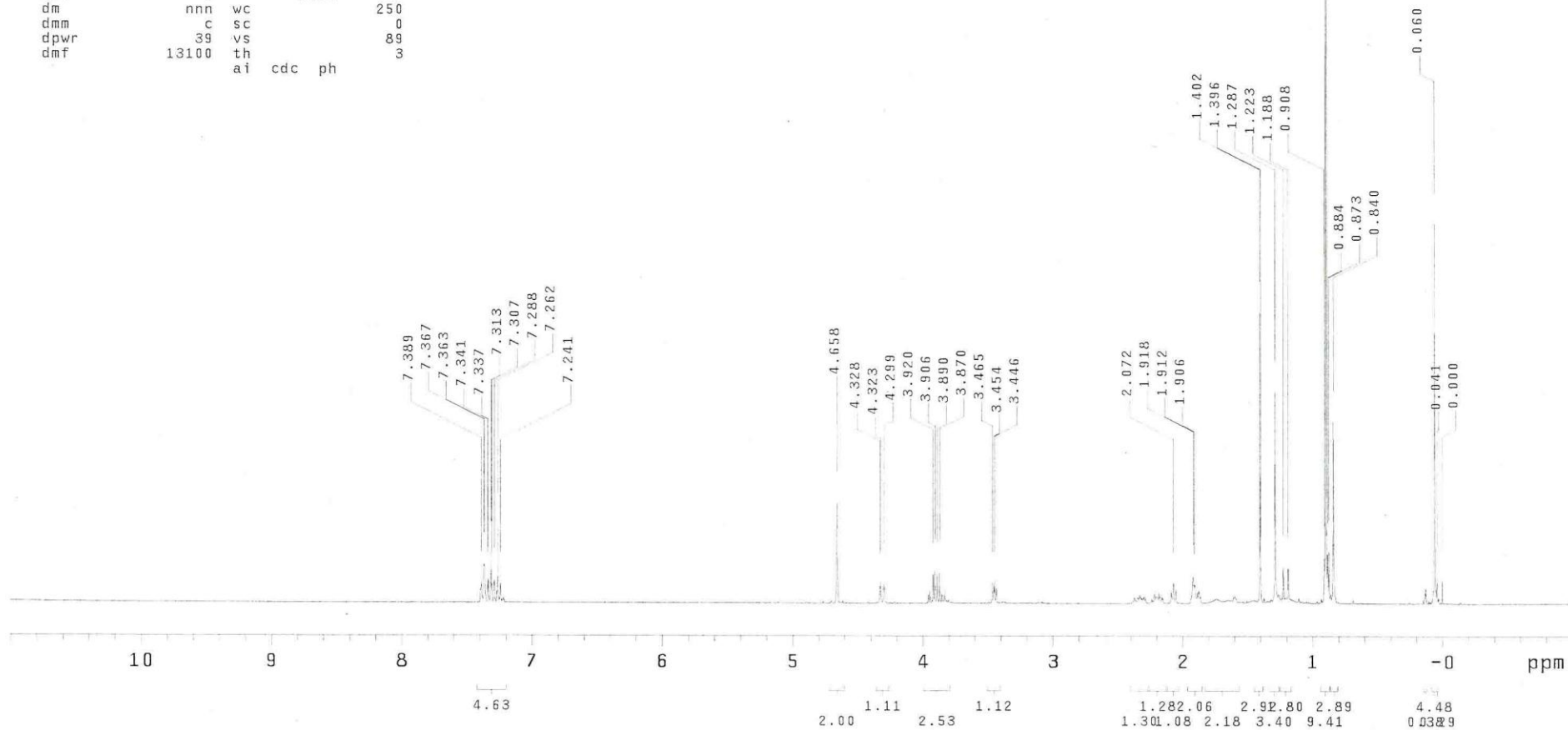
exp1 s2pul

SAMPLE		SPECIAL	
date	Dec 3 2008	temp	not used
solvent	CDCl <sub>3</sub>	gain	not used
file	exp	spin	not used
ACQUISITION			
sw	3602.3	hst	0.008
at	1.998	pw90	9.800
np	14394	alfa	20.000
TRANSMITTER		FLAGS	
fb	not used	il	n
bs	16	in	n
d1	1.000	dp	y
nt	16	hs	nn
ct	16	PROCESSING	
tn	H1	fn	not used
sfrq	300.144	DISPLAY	
tof	1.3	sp	-299.5
tpwr	57	wp	3602.3
pw	4.900	rfl	299.5
DECOUPLER		rfp	0
dn	C13	rp	44.9
dof	0	lp	-50.5
dmm	nnn	PLOT	
dpwr	39	wc	250
dmf	13100	sc	0
		vs	89
		th	3
		af	cdc ph



6

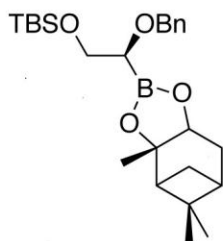
300-MHz <sup>1</sup>H-NMR  
(CDCl<sub>3</sub>)



TBSOCH<sub>2</sub>CHOBnBP boronate, 13C

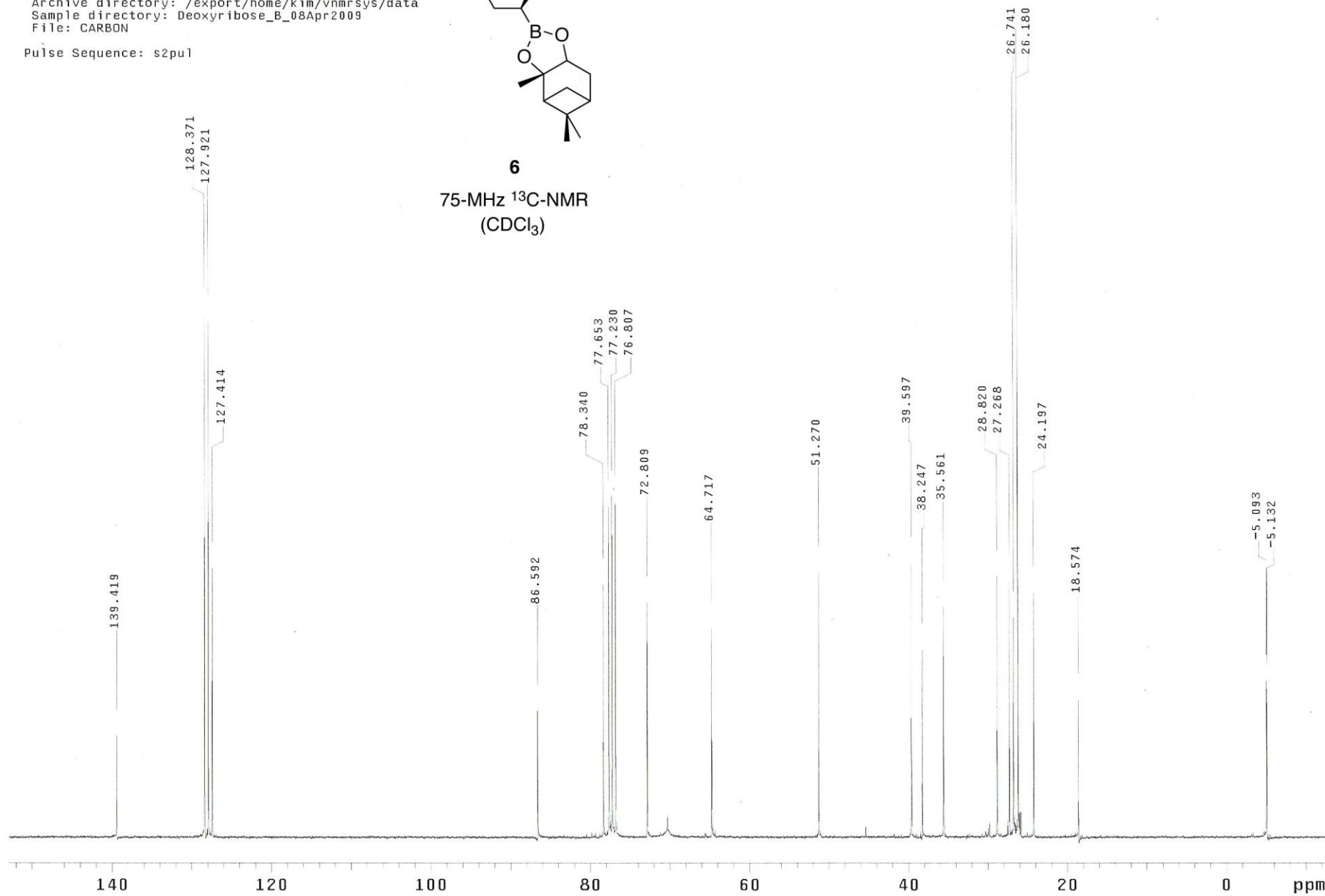
Archive directory: /export/home/kim/vnmrsys/data  
Sample directory: Deoxyribose\_B\_08Apr2009  
File: CARBON

Pulse Sequence: s2pul



6

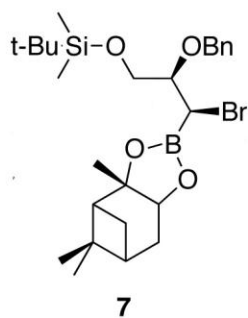
75-MHz <sup>13</sup>C-NMR  
(CDCl<sub>3</sub>)



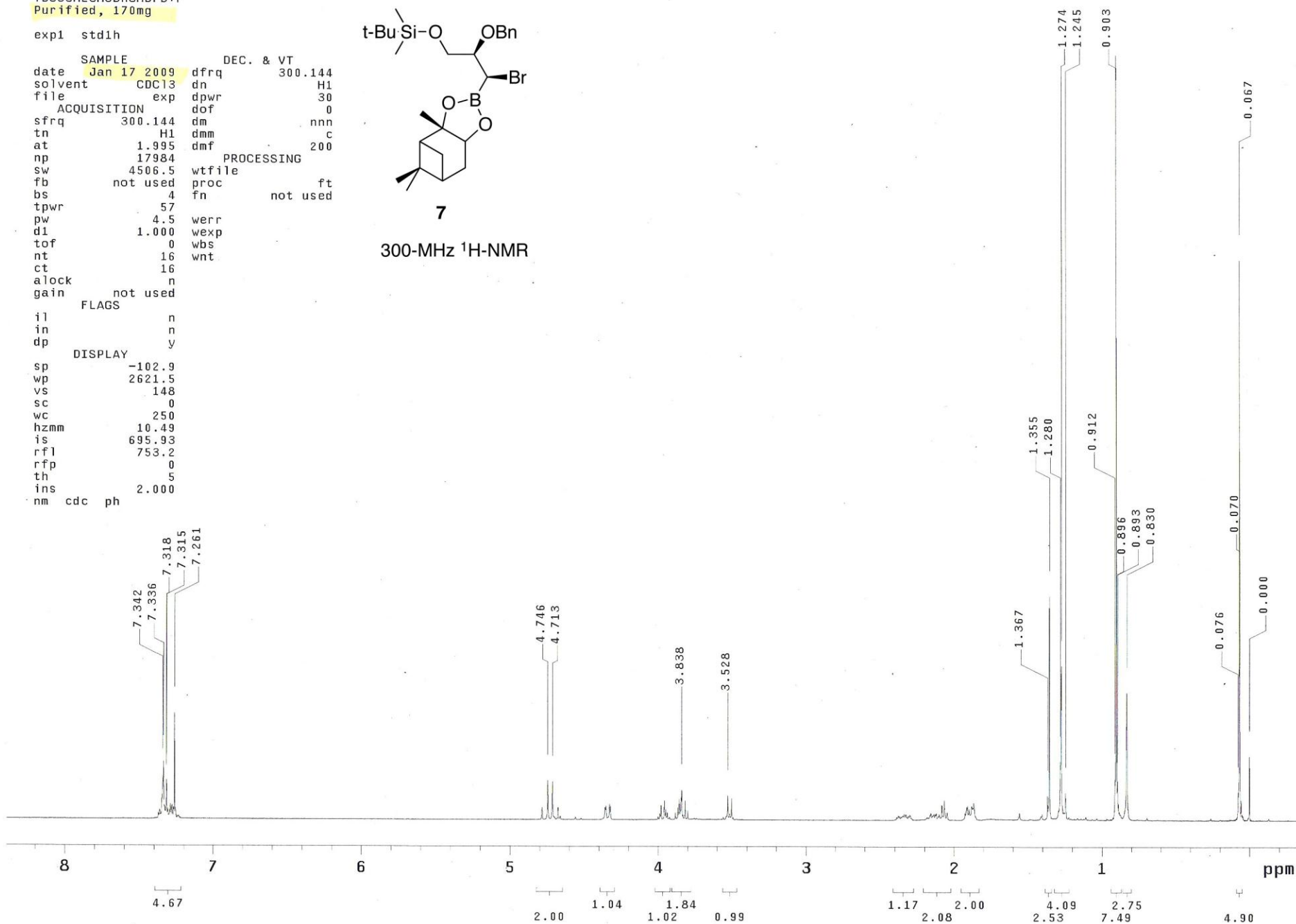
TBSOCH<sub>2</sub>CHOBNCHBrB+P  
Purified, 170mg

exp1 std1h

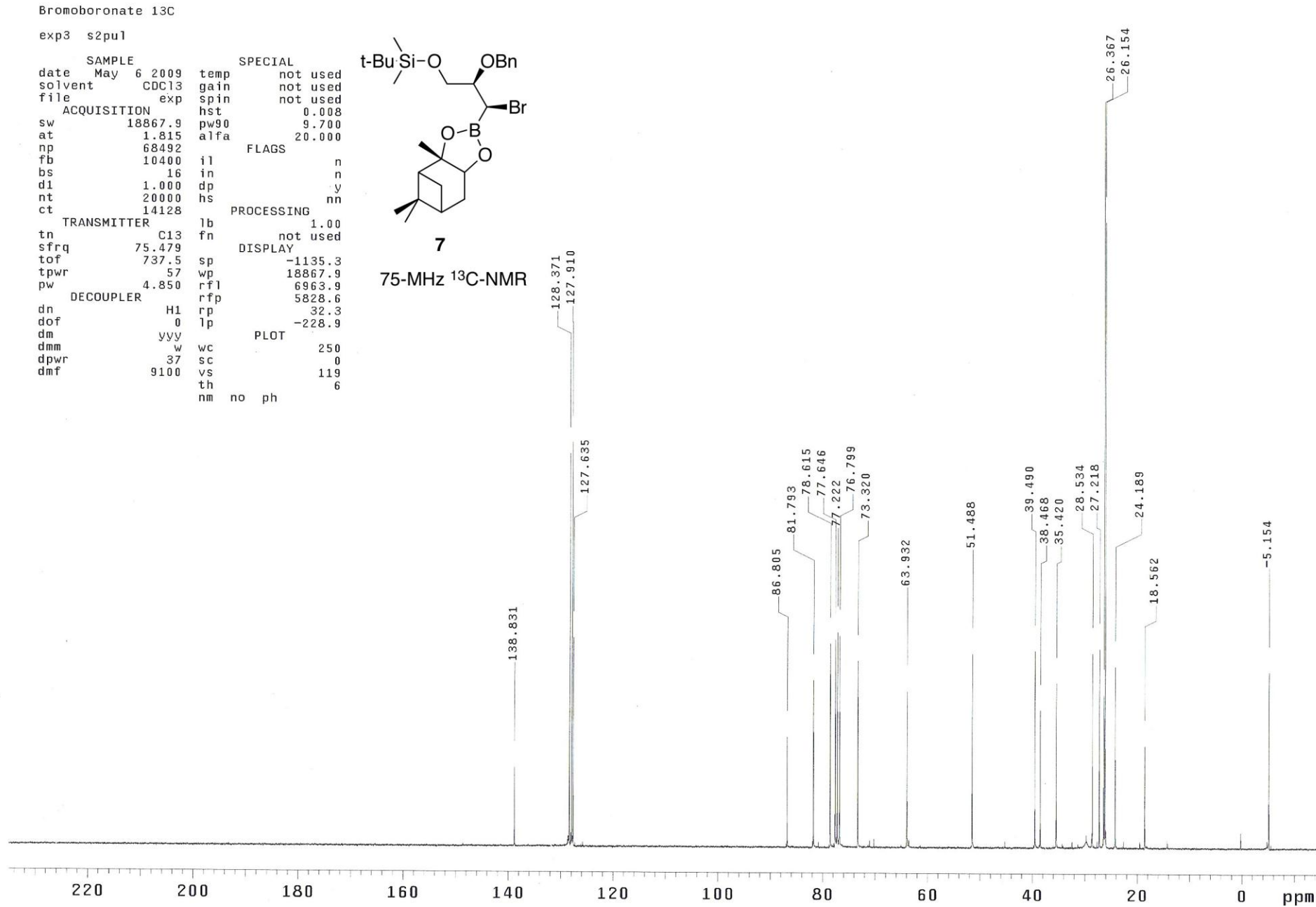
SAMPLE		DEC. & VT	
date	Jan 17 2009	dfrq	300.144
solvent	CDC13	dn	H1
file	exp	dpwr	30
ACQUISITION		dof	0
sfrq	300.144	dm	nnn
tn	H1	dmm	c
at	1.995	dmf	200
np	17984	PROCESSING	
sw	4506.5	wtfile	
fb	not used	proc	ft
bs	4	fn	not used
tpwr	57		
pw	4.5	werr	
d1	1.000	wexp	
tof	0	wbs	
nt	16	wnt	
ct	16		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	y		
DISPLAY			
sp	-102.9		
wp	2621.5		
vs	148		
sc	0		
wc	250		
hzmm	10.49		
is	695.93		
rfl	753.2		
rfp	0		
th	5		
ins	2.000		
nm	cdc	ph	



300-MHz <sup>1</sup>H-NMR



Solvent is CDCl<sub>3</sub> if not otherwise specified.



Pulse Sequence: s2pul

Solvent: CDCl<sub>3</sub>

Temp. 22.0 C / 295.1 K

INOVA-500 "nmrc500b"

Relax. delay 1.000 sec

Pulse 45.0 degrees

Acq. time 1.892 sec

Width 8000.0 Hz

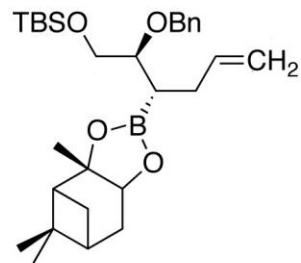
8 repetitions

OBSERVE H1, 499.8560475 MHz

DATA PROCESSING

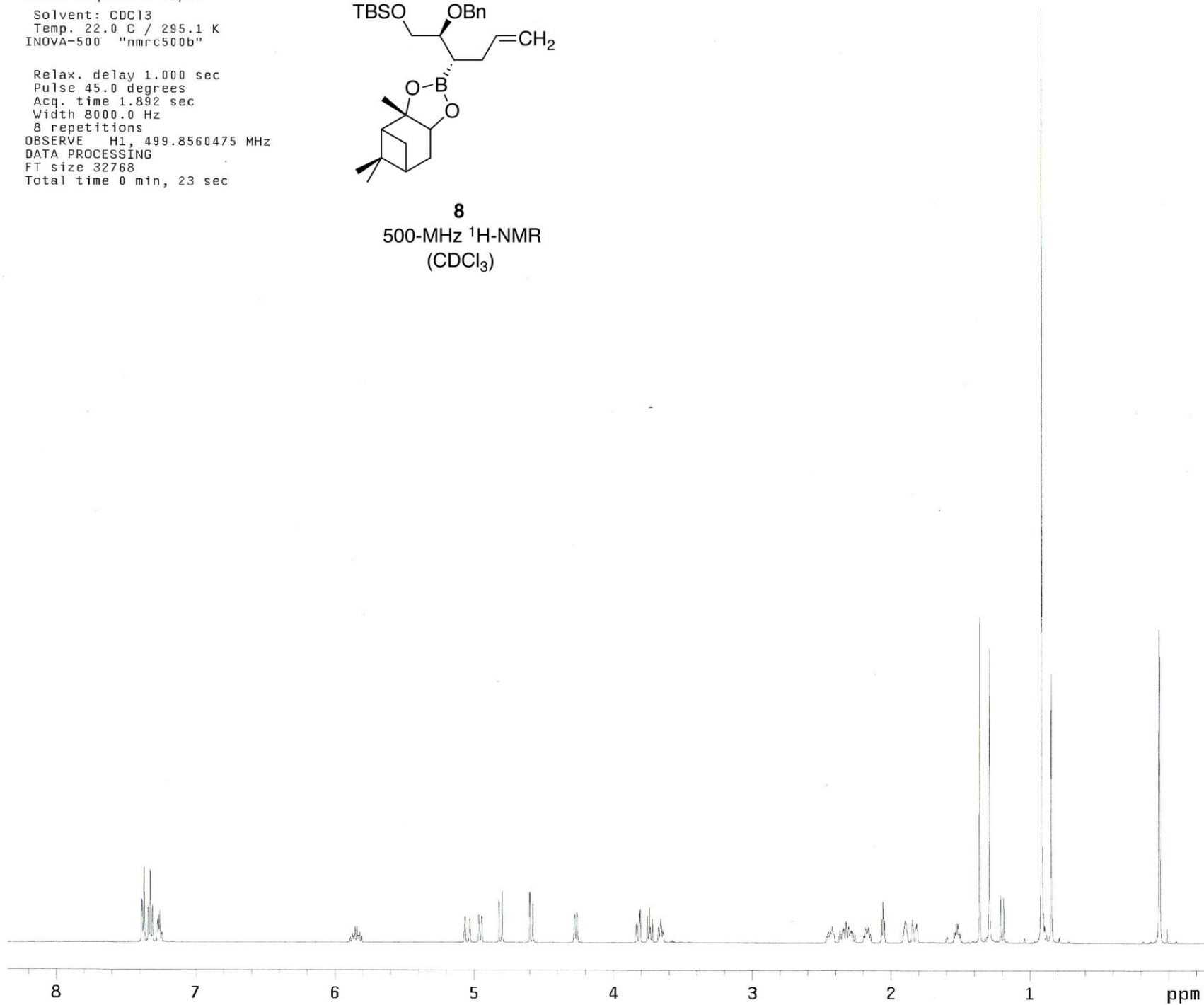
FT size 32768

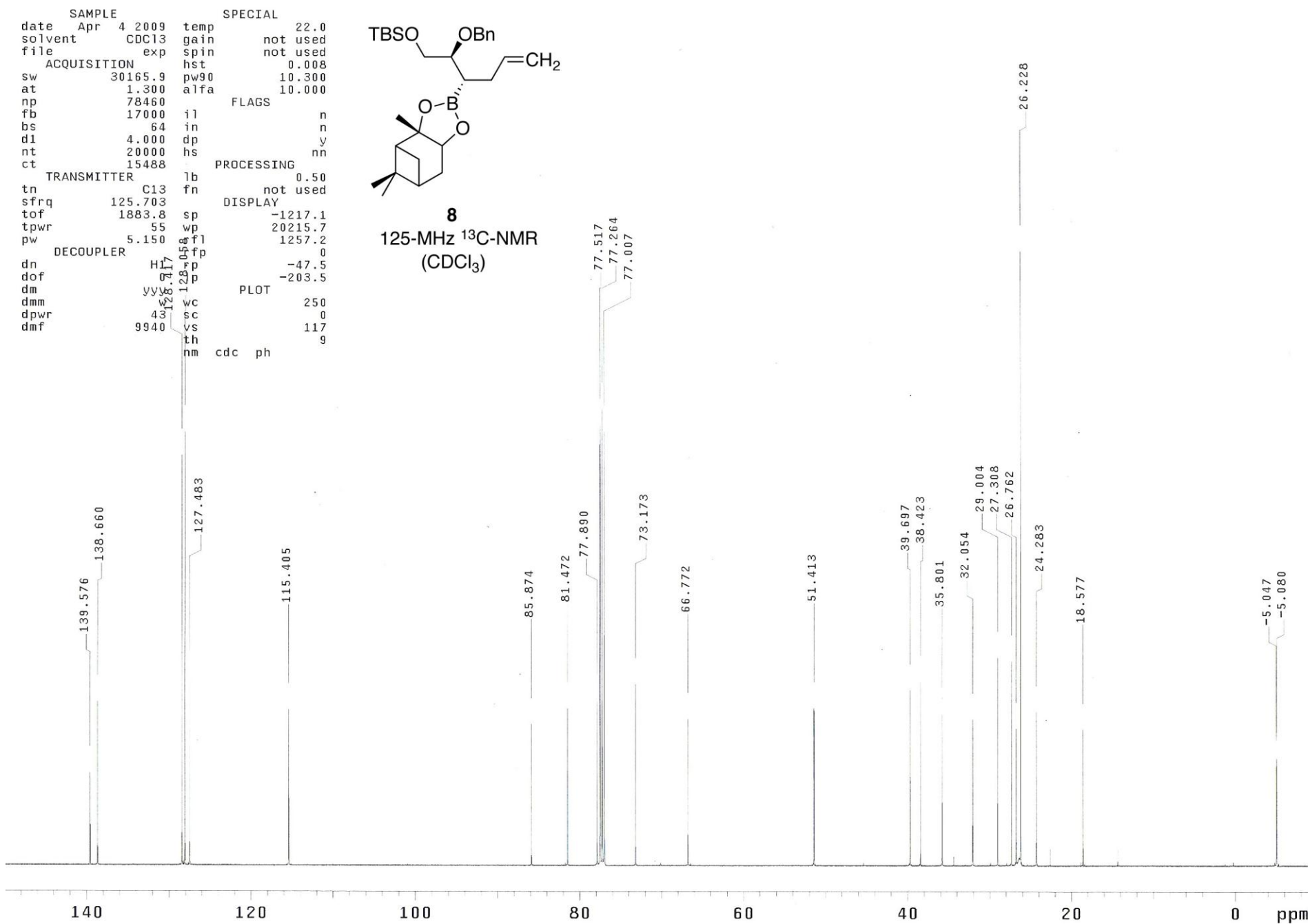
Total time 0 min, 23 sec

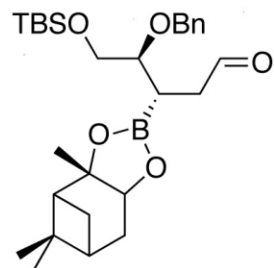


**8**

500-MHz <sup>1</sup>H-NMR  
(CDCl<sub>3</sub>)

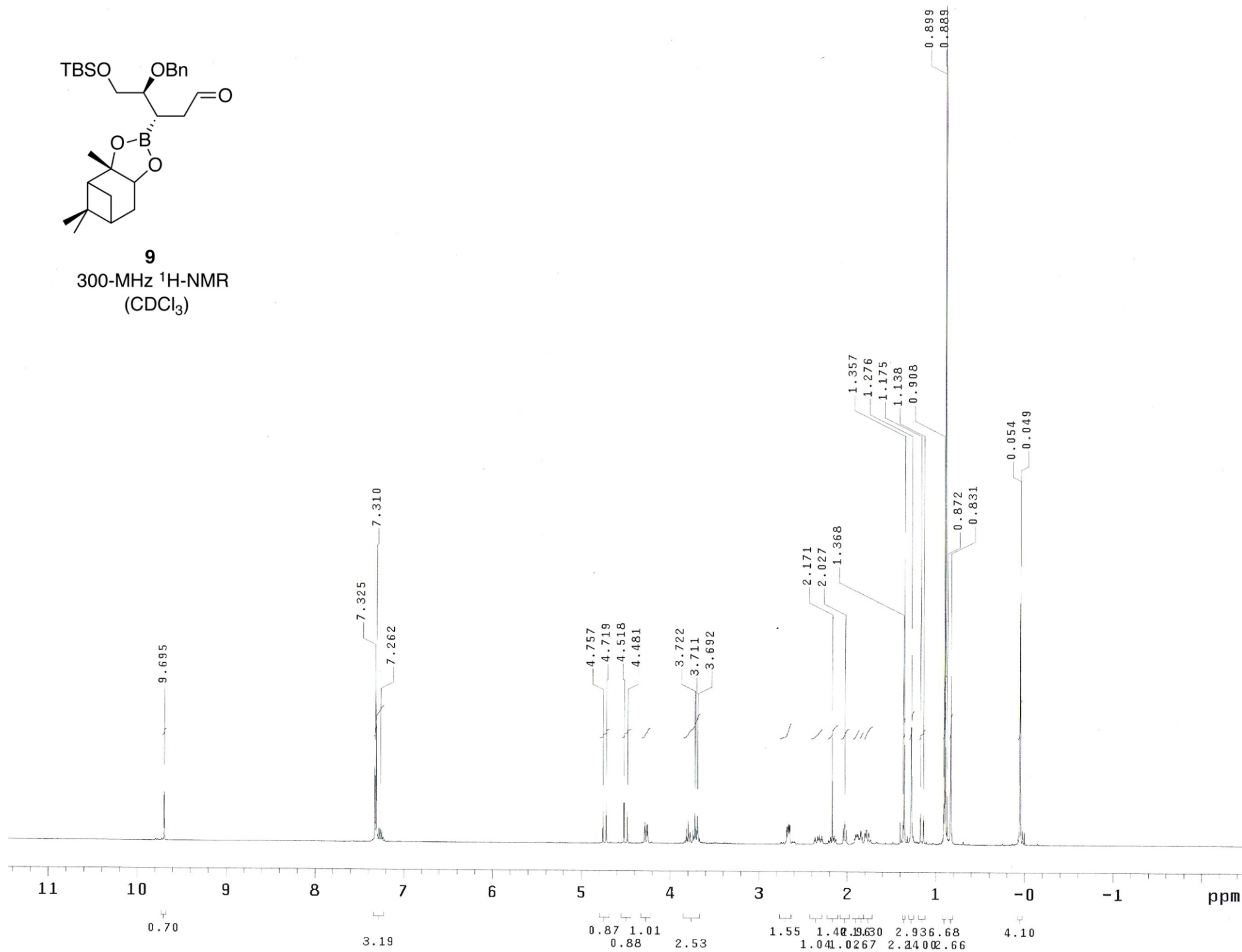






**9**

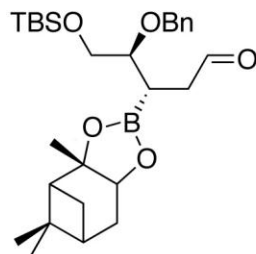
300-MHz  $^1\text{H}$ -NMR  
( $\text{CDCl}_3$ )



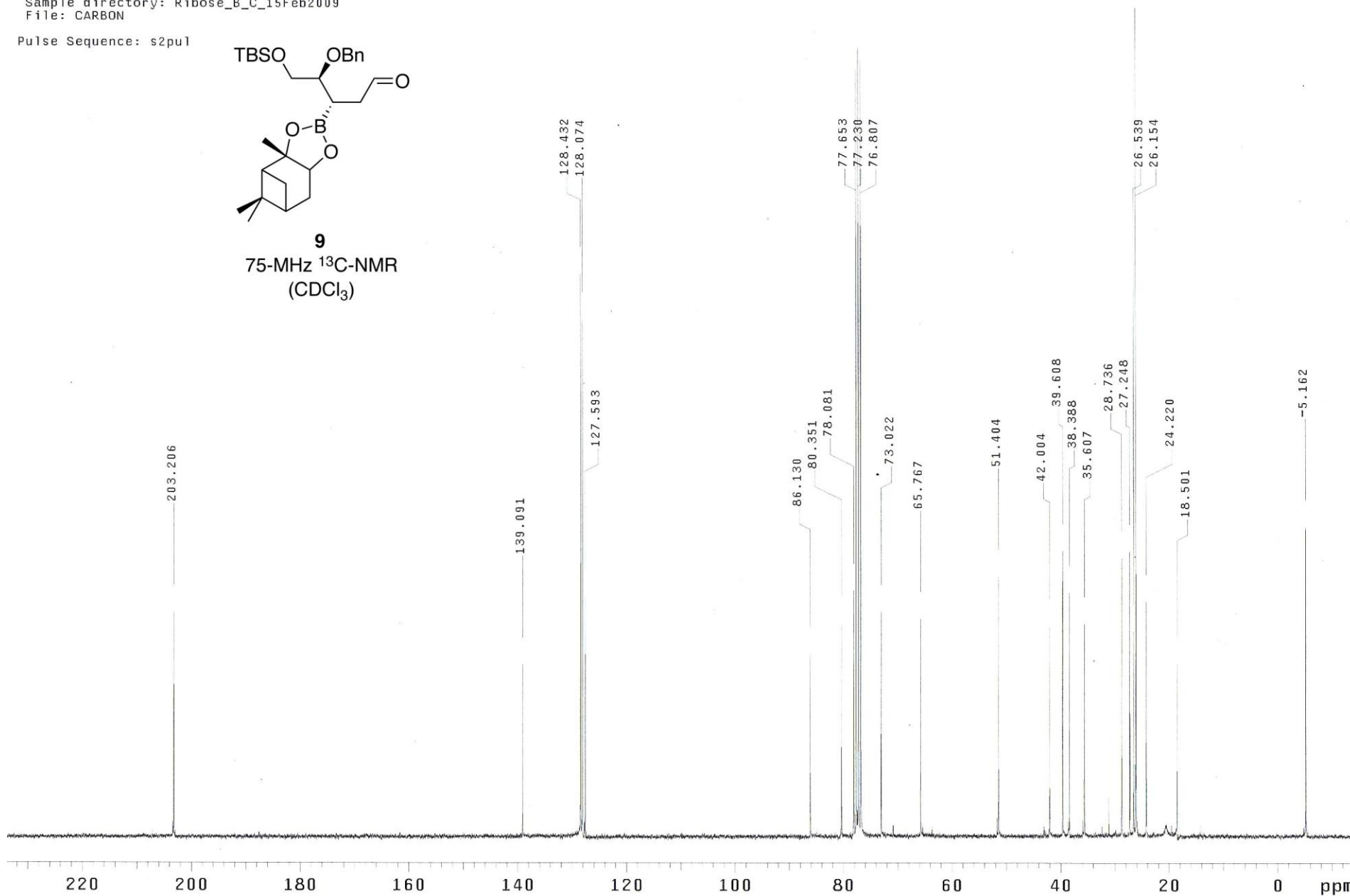
aldehyde boronate, <sup>13</sup>C

Archive directory: /export/home/kim/vnmrsys/data  
Sample directory: Ribose\_B\_C\_15Feb2009  
File: CARBON

Pulse Sequence: s2pu1



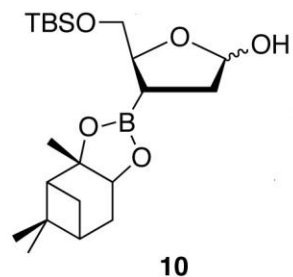
**9**  
75-MHz <sup>13</sup>C-NMR  
(CDCl<sub>3</sub>)



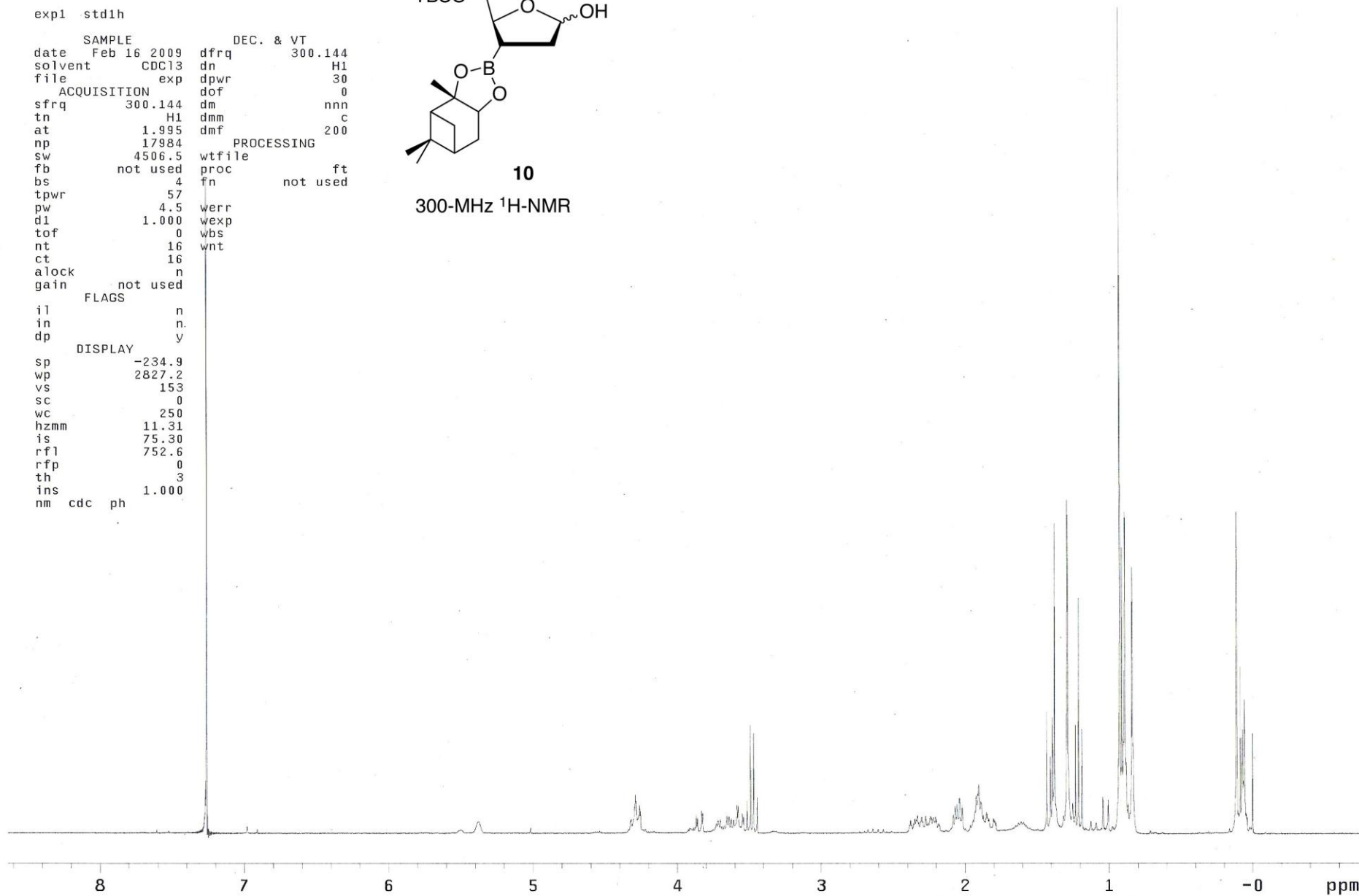
Ribose boronate  
latter fraction

exp1 std1h

SAMPLE		DEC. & VT	
date	Feb 16 2009	dfrq	300.144
solvent	CDC13	dn	H1
file	exp	dpwr	30
ACQUISITION		dof	0
sfrq	300.144	dm	nnn
tn	H1	dmm	c
at	1.995	dmf	200
np	17984	PROCESSING	
sw	4506.5	wtfile	
fb	not used	proc	ft
bs	4	fn	not used
tpwr	57		
pw	4.5	werr	
d1	1.000	wexp	
tof	0	wbs	
nt	16	wnt	
ct	16		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	y		
DISPLAY			
sp	-234.9		
wp	2827.2		
vs	153		
sc	0		
wc	250		
hzmm	11.31		
is	75.30		
rfl	752.6		
rfp	0		
th	3		
ins	1.000		
nm	cdc ph		



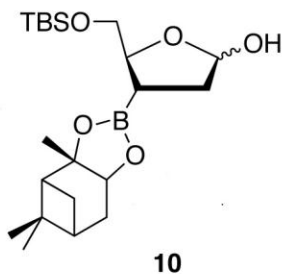
300-MHz <sup>1</sup>H-NMR



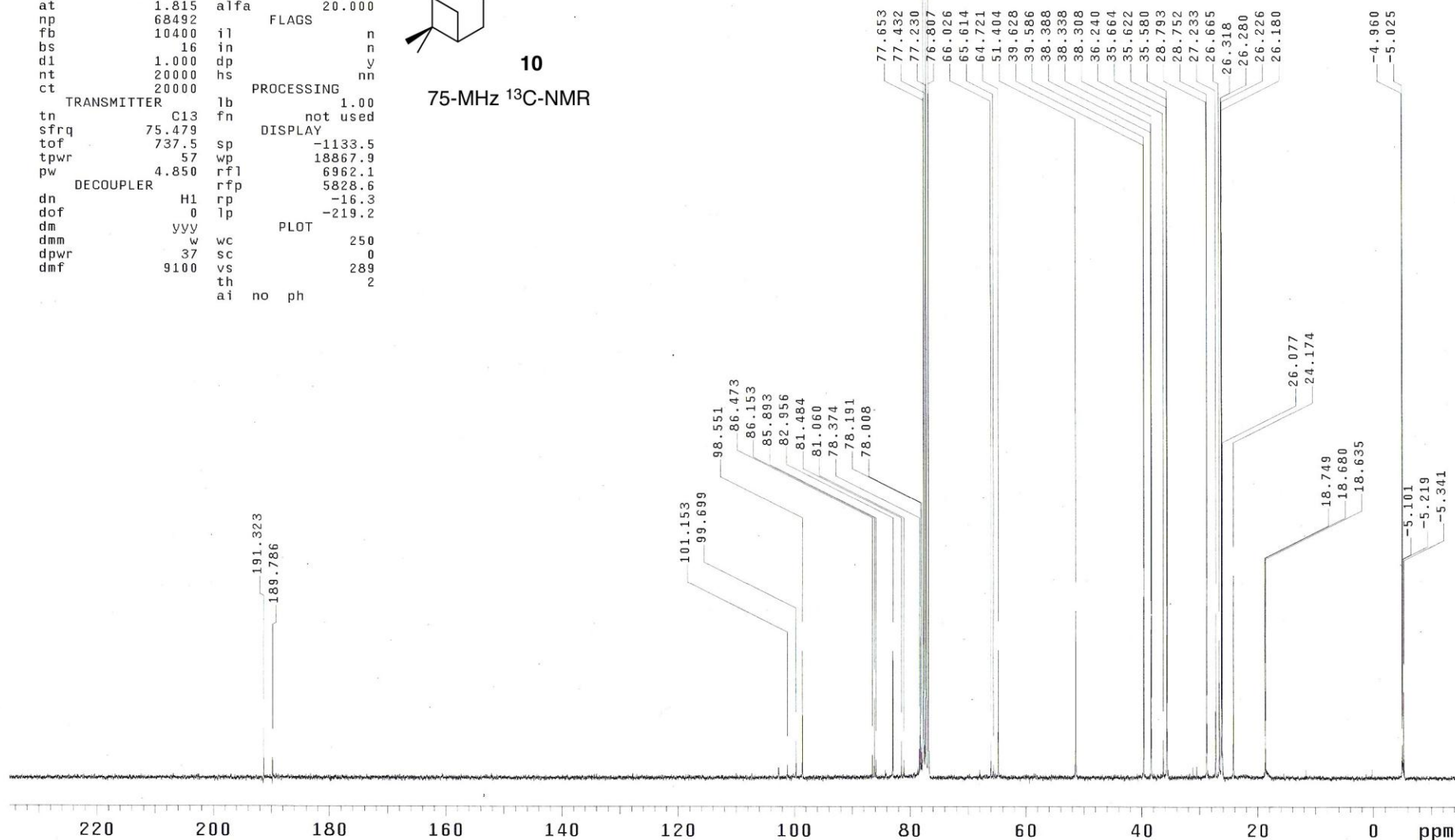
Ribose boronate  
Made on 2/15/09

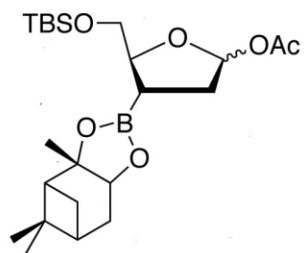
exp3 s2pu1

SAMPLE		SPECIAL	
date	Feb 15 2009	temp	not used
solvent	CDCl3	gain	not used
file	exp	spin	not used
ACQUISITION		hst	0.008
sw	18867.9	pw90	9.700
at	1.815	alfa	20.000
np	68492	FLAGS	
fb	10400	il	n
bs	16	in	n
dl	1.000	dp	y
nt	20000	hs	nn
ct	20000	PROCESSING	
TRANSMITTER		lb	1.00
tn	C13	fn	not used
sfrq	75.479	DISPLAY	
tof	737.5	sp	-1133.5
tpwr	57	wp	18867.9
pw	4.850	rfl	6962.1
DECOUPLER		rfp	5828.6
dn	H1	rp	-16.3
dof	0	lp	-219.2
dm	yyv	PLOT	
dmm	w	wc	250
dpwr	37	sc	0
dmf	9100	vs	289
		th	2
		ai	no ph



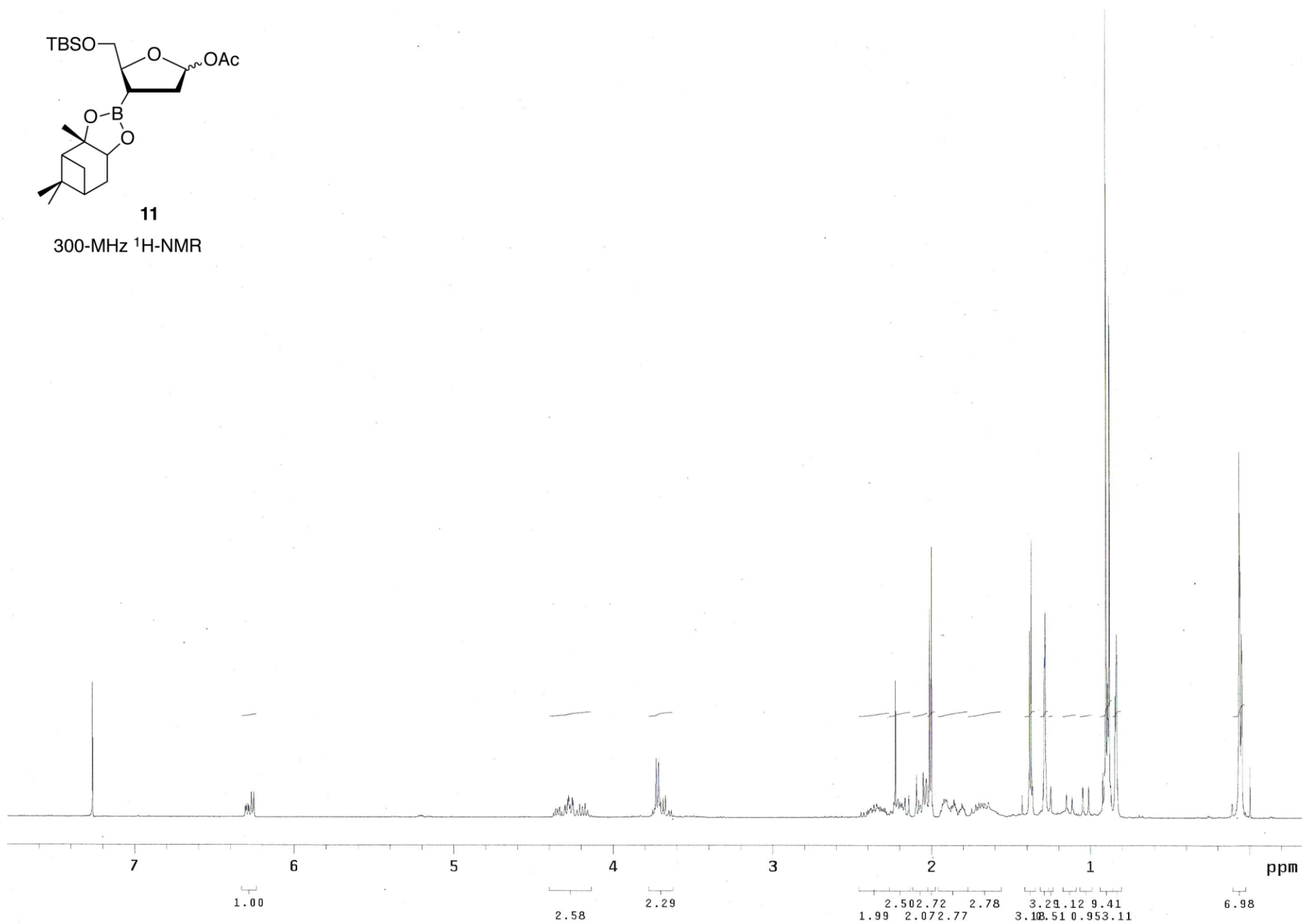
75-MHz <sup>13</sup>C-NMR





**11**

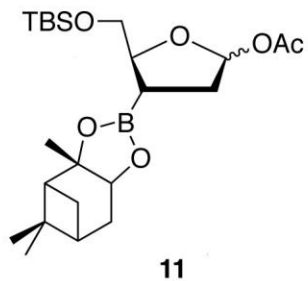
300-MHz  $^1\text{H}$ -NMR



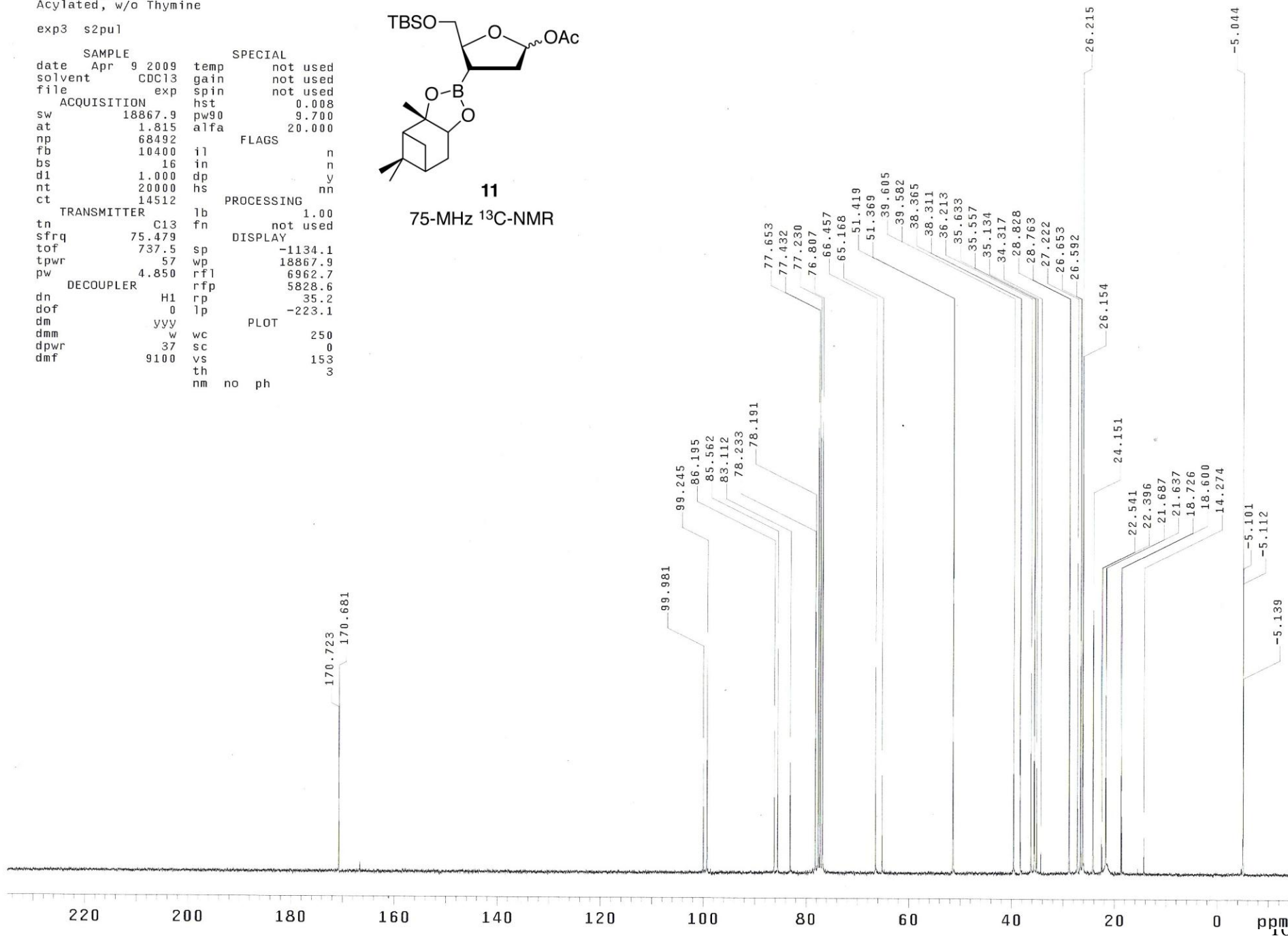
Deoxyribose boronate  
Acylated, w/o Thymine

exp3 s2pul

SAMPLE		SPECIAL	
date	Apr 9 2009	temp	not used
solvent	CDC13	gain	not used
file	exp	spin	not used
ACQUISITION		hst	0.008
sw	18867.9	pw90	9.700
at	1.815	alfa	20.000
np	68492	FLAGS	
fb	10400	il	n
bs	16	in	n
d1	1.000	dp	y
nt	20000	hs	nn
ct	14512	PROCESSING	
TRANSMITTER		lb	1.00
tn	C13	fn	not used
sfrq	75.479	DISPLAY	
tof	737.5	sp	-1134.1
tpwr	57	wp	18867.9
pw	4.850	rfl	6962.7
DECOUPLER		rfp	5828.6
dn	H1	rp	35.2
dof	0	lp	-223.1
dm	yyy	PLOT	
dmm	w	wc	250
dpwr	37	sc	0
dmf	9100	vs	153
		th	3
		nm	no ph



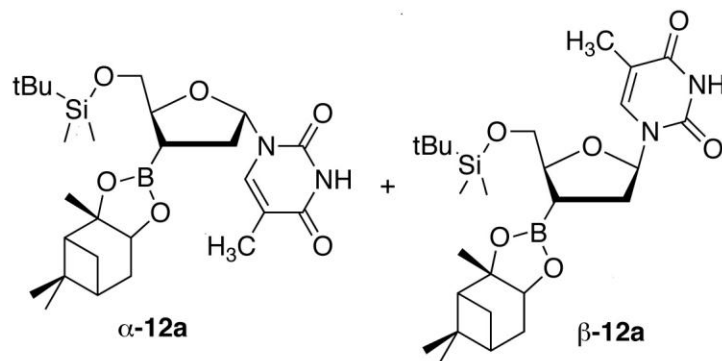
75-MHz  $^{13}\text{C}$ -NMR



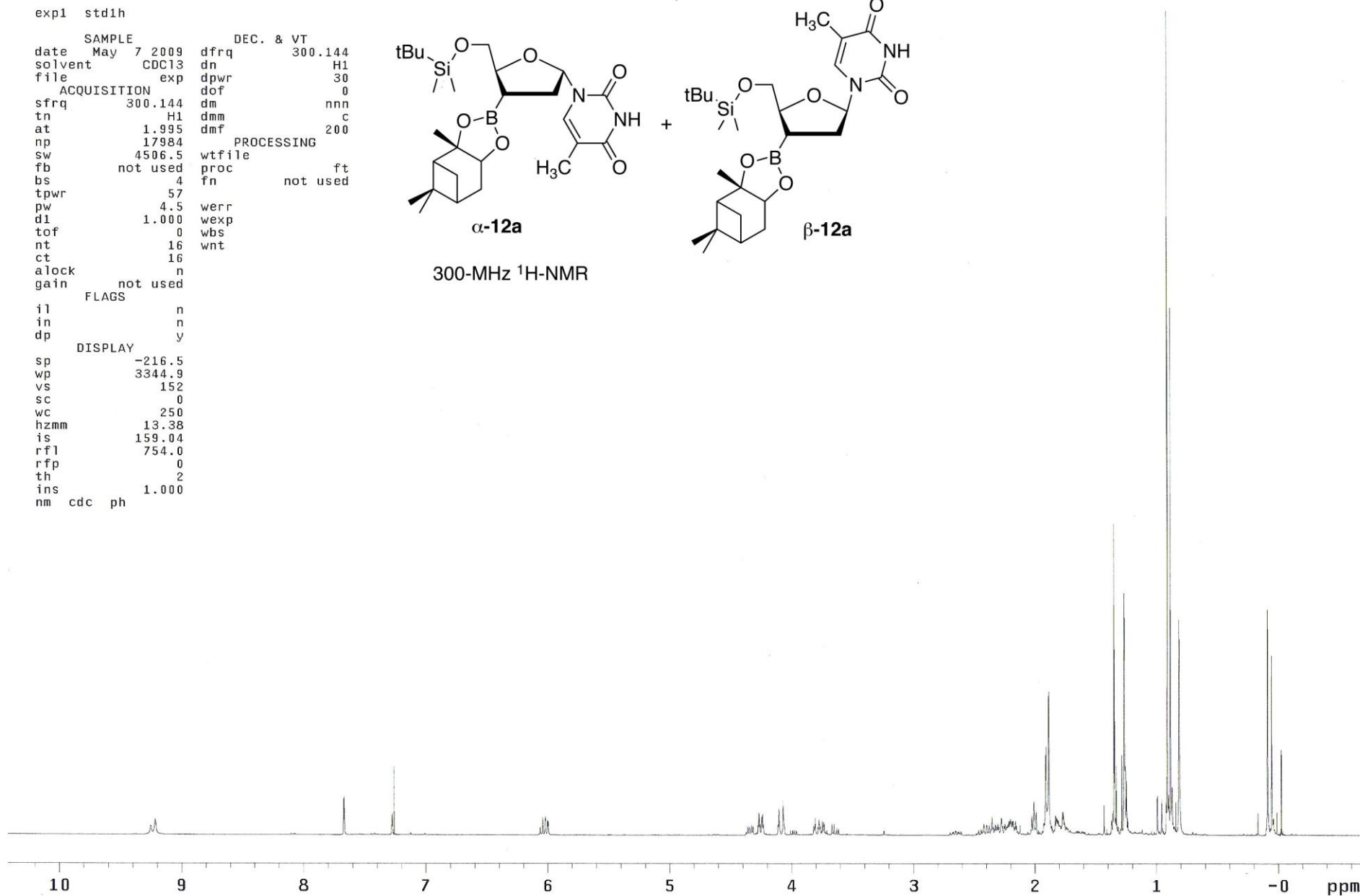
Silylated Thymidine boronate  
ab mixture

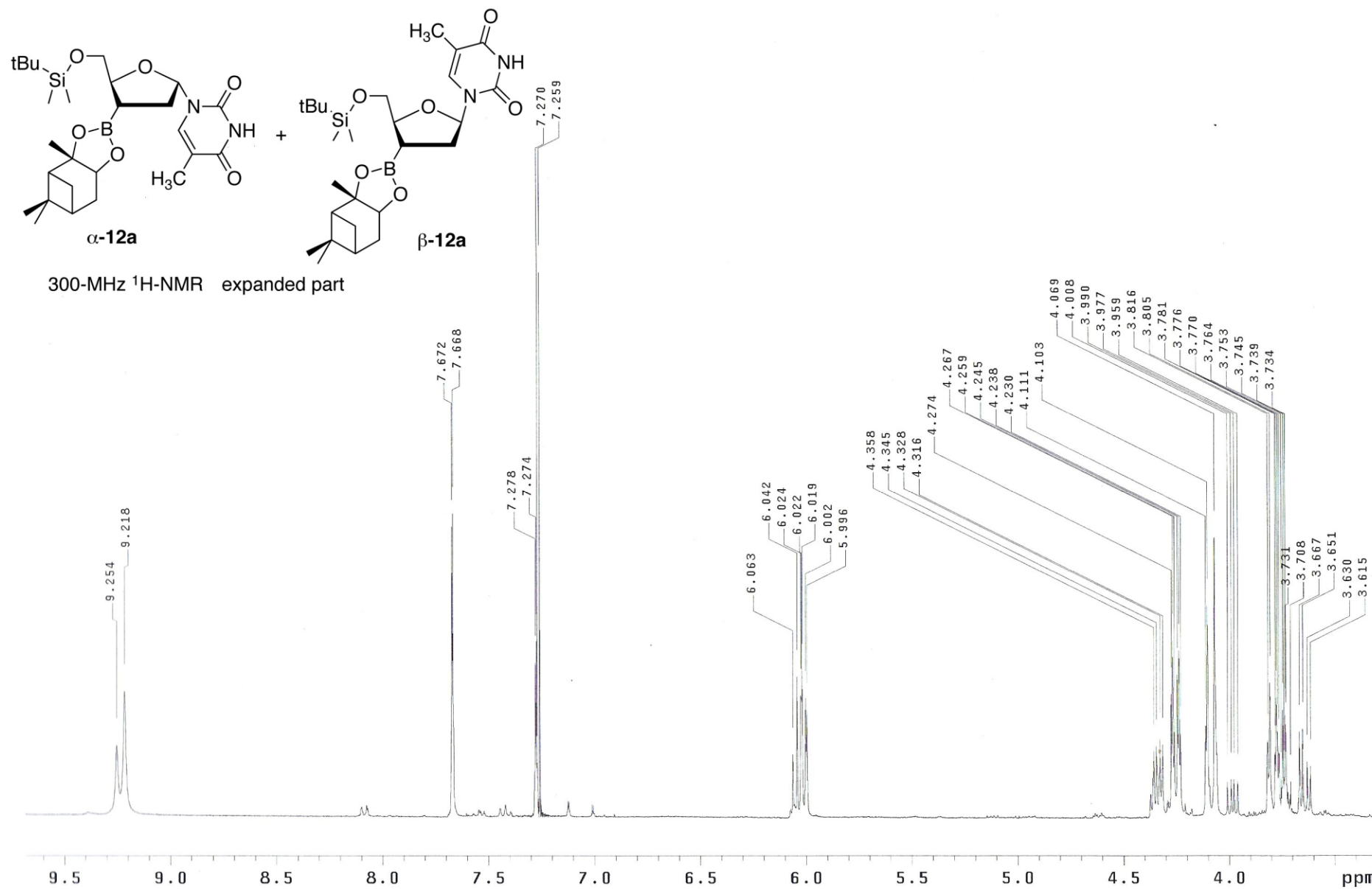
exp1 std1h

SAMPLE		DEC. & VT	
date	May 7 2009	dfrq	300.144
solvent	CDCl3	dn	H1
file	exp	dpwr	30
ACQUISITION		dof	0
sfrq	300.144	dm	nnn
tn	H1	dmm	c
at	1.995	dmf	200
np	17984	PROCESSING	
sw	4506.5	wtfile	ft
fb	not used	proc	not used
bs	4	fn	
tpwr	57		
pw	4.5	werr	
dl	1.000	wexp	
tof	0	wbs	
nt	16	wnt	
ct	16		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	y		
DISPLAY			
sp	-216.5		
wp	3344.9		
vs	152		
sc	0		
wc	250		
hzmm	13.38		
is	159.04		
rfl	754.0		
rfp	0		
th	2		
ins	1.000		
nm	cdc ph		



300-MHz <sup>1</sup>H-NMR

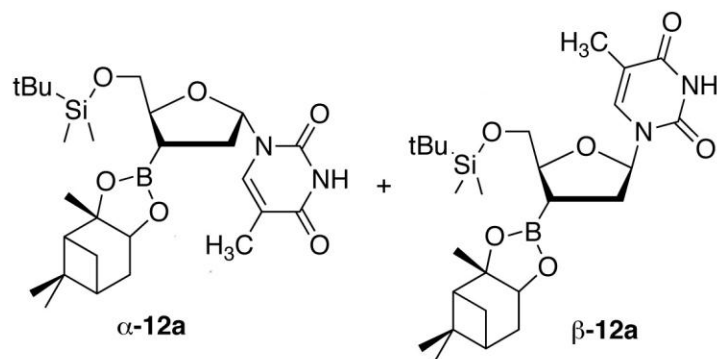




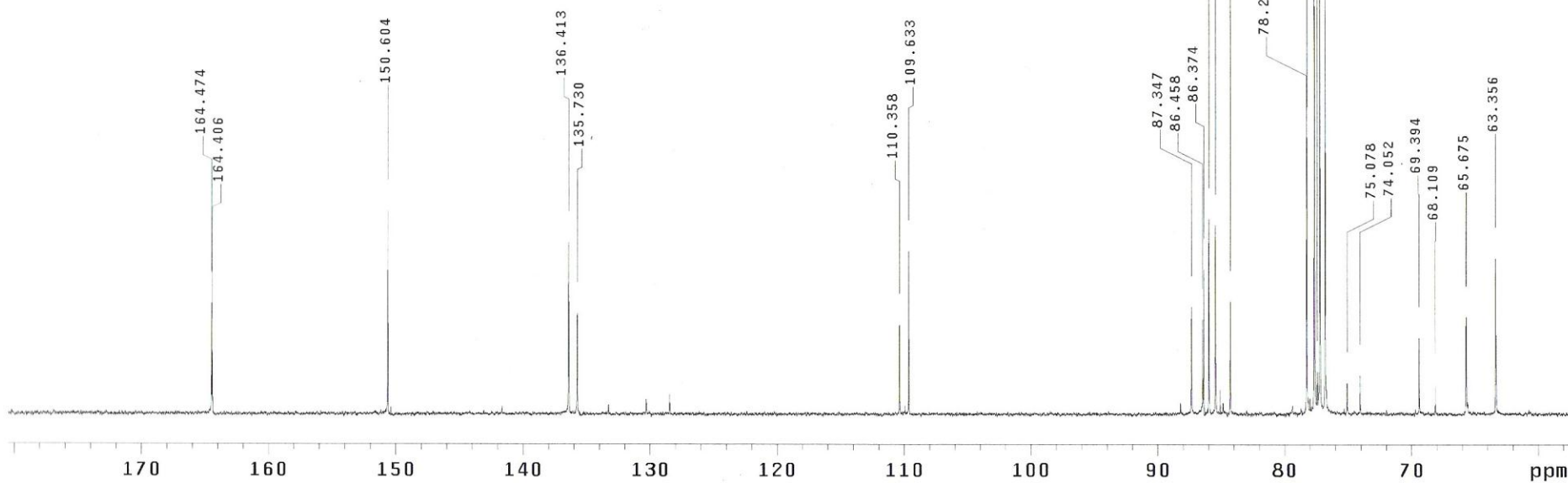
Silylated Thymidine BPab 13C

Archive directory: /export/home/kim/vnmrsys/data  
Sample directory: Deoxyribose\_B\_08Apr2009  
File: CARBON

Pulse Sequence: s2pu1



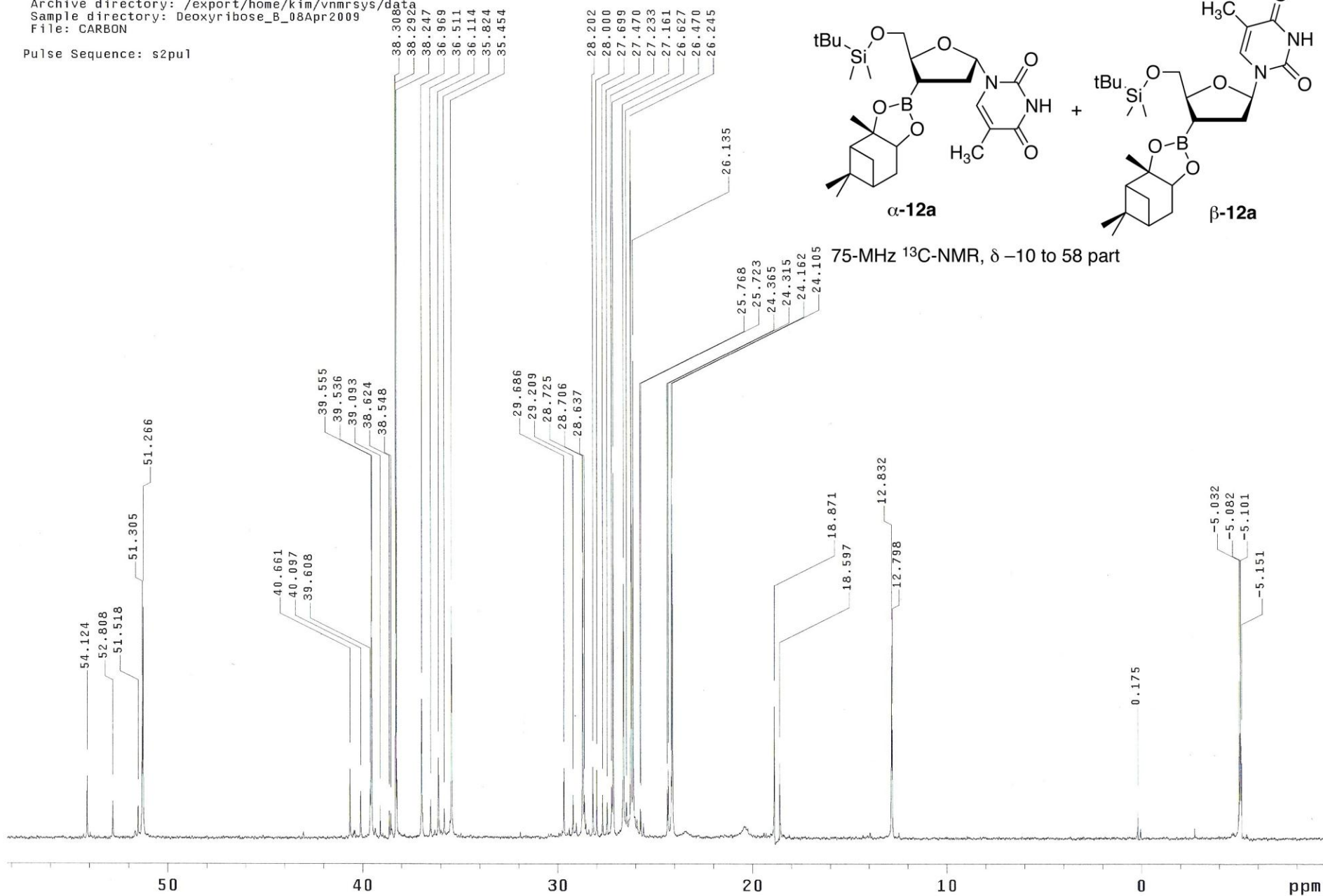
75-MHz  $^{13}\text{C}$ -NMR,  $\delta$  58 to 180 part



Silylated Thymidine BPab 13C

Archive directory: /export/home/kim/vnmrsys/data  
Sample directory: Deoxyribose\_B\_08Apr2009  
File: CARBON

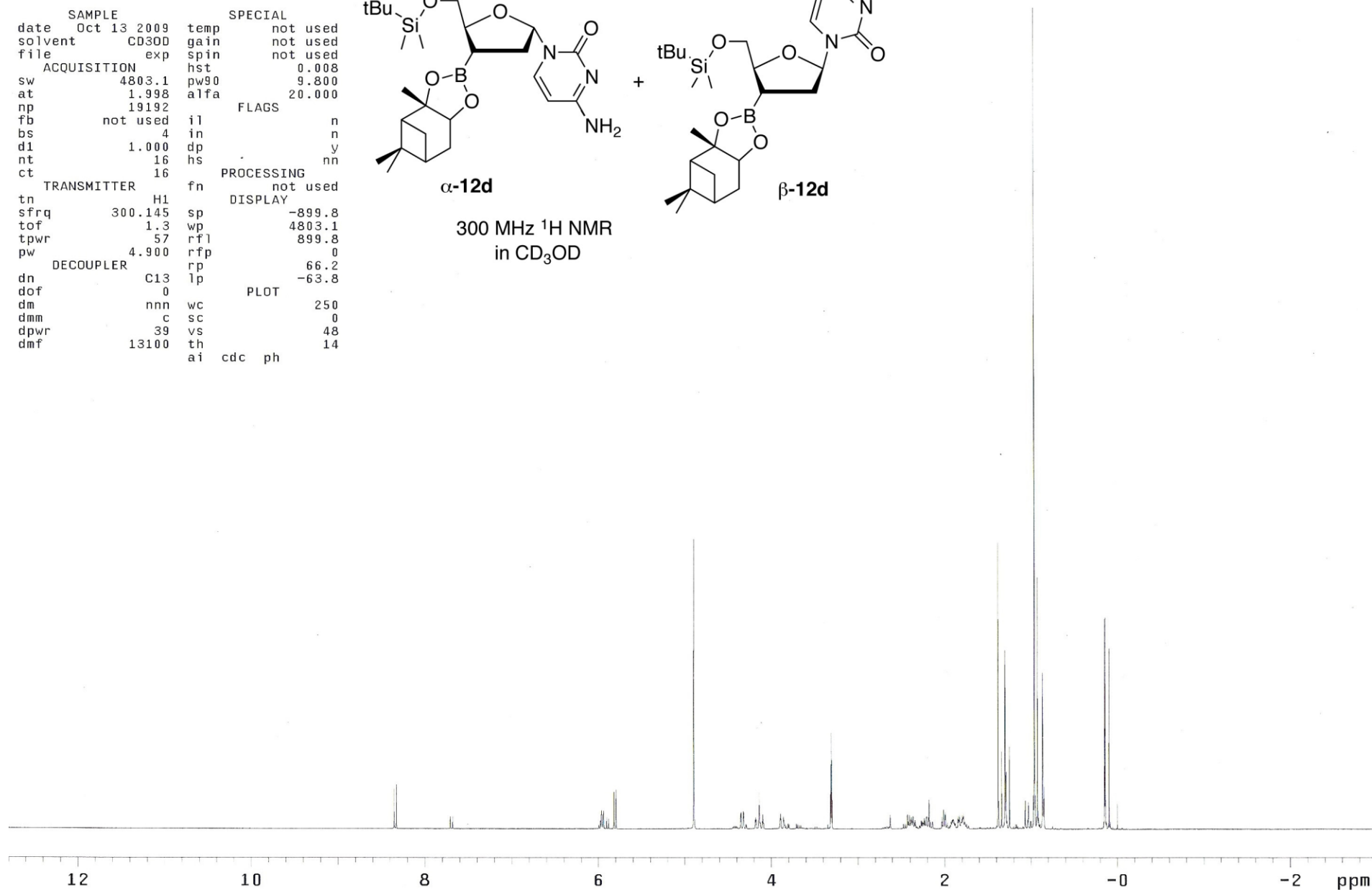
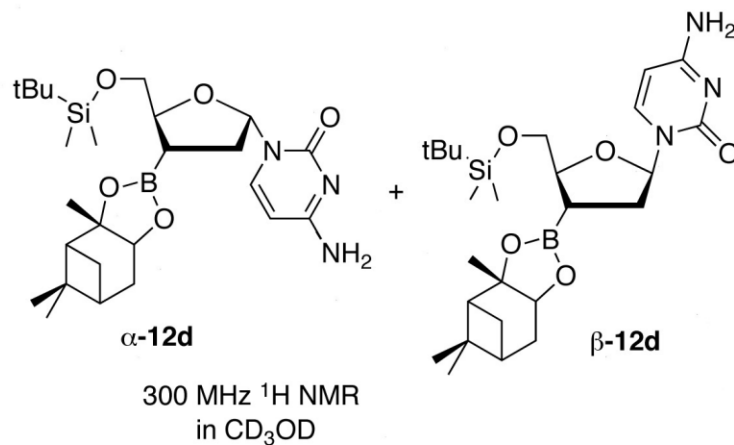
Pulse Sequence: s2pu1



Sililated cytidine boronate

exp5 s2pu1

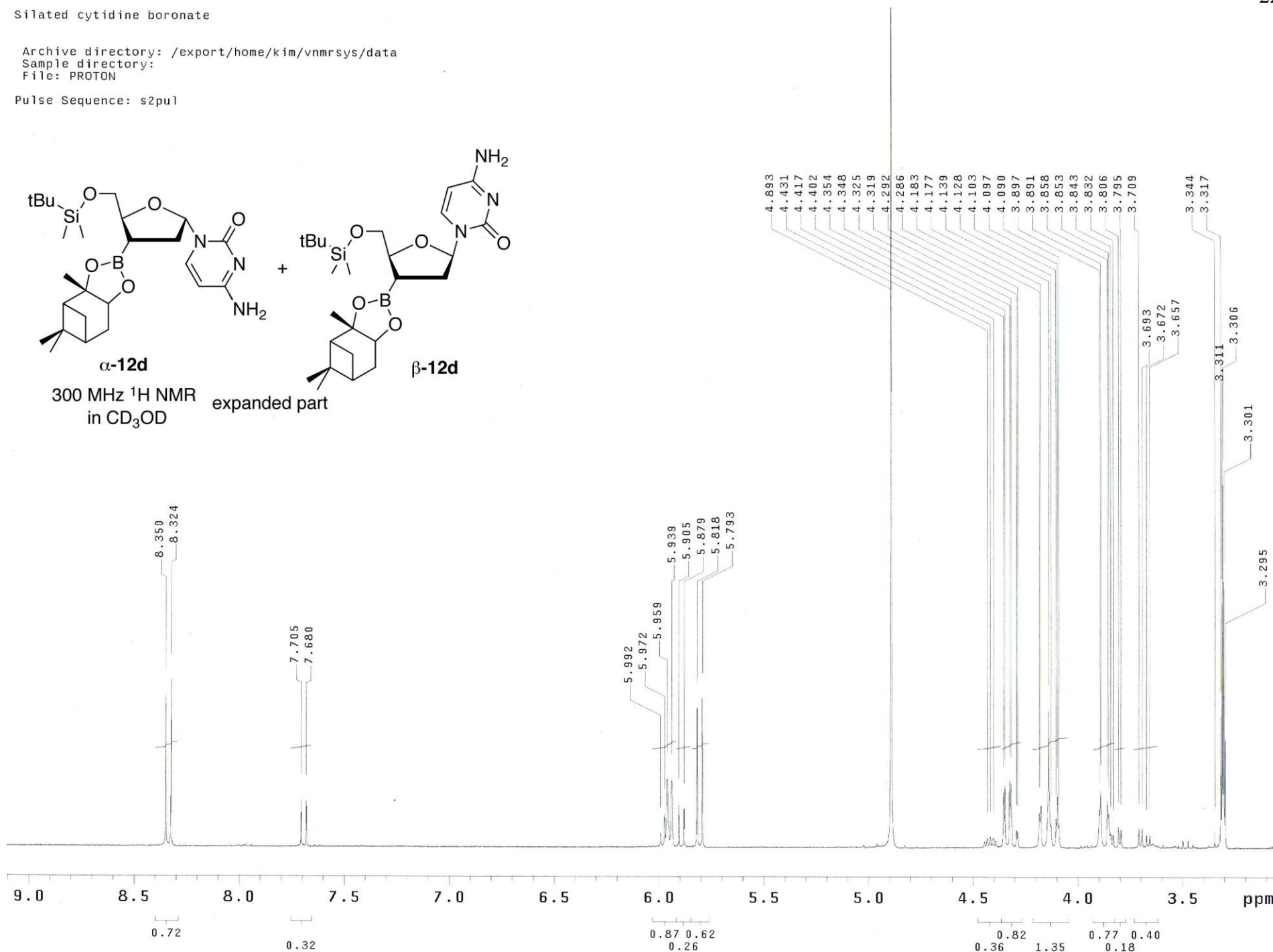
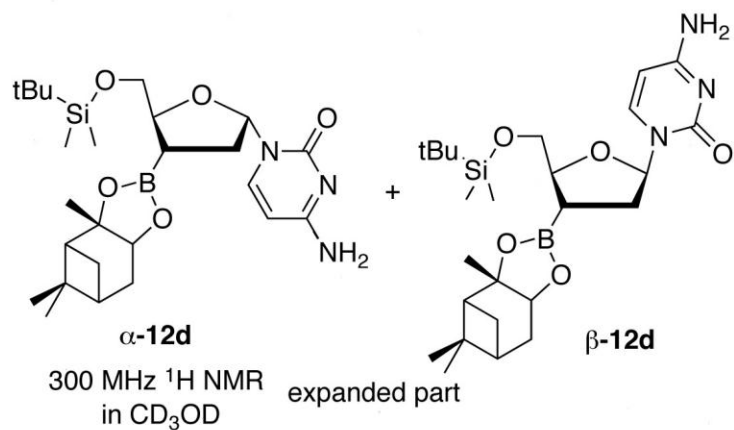
SAMPLE		SPECIAL	
date	Oct 13 2009	temp	not used
solvent	CD3OD	gain	not used
file	exp	spin	not used
ACQUISITION		hst	0.008
sw	4803.1	pw90	9.800
at	1.998	alfa	20.000
np	19192	FLAGS	
fb	not used	il	n
bs	4	in	n
d1	1.000	dp	y
nt	16	hs	nn
ct	16	PROCESSING	
TRANSMITTER		fn	not used
tn	H1	DISPLAY	
sfrq	300.145	sp	-899.8
tof	1.3	wp	4803.1
tpwr	57	rfl	899.8
pw	4.900	rfp	0
DECOUPLER		rp	66.2
dn	C13	lp	-63.8
dof	0	PLOT	
dm	nnn	wc	250
dmm	c	sc	0
dpwr	39	vs	48
dmf	13100	th	14
	ai	cdc	ph



Silated cytidine boronate

Archive directory: /export/home/kim/vnmrsys/data  
Sample directory:  
File: PROTON

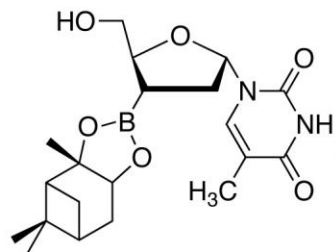
Pulse Sequence: s2pul



Deoxythymidine boronate  
b fraction, TMSBr treated

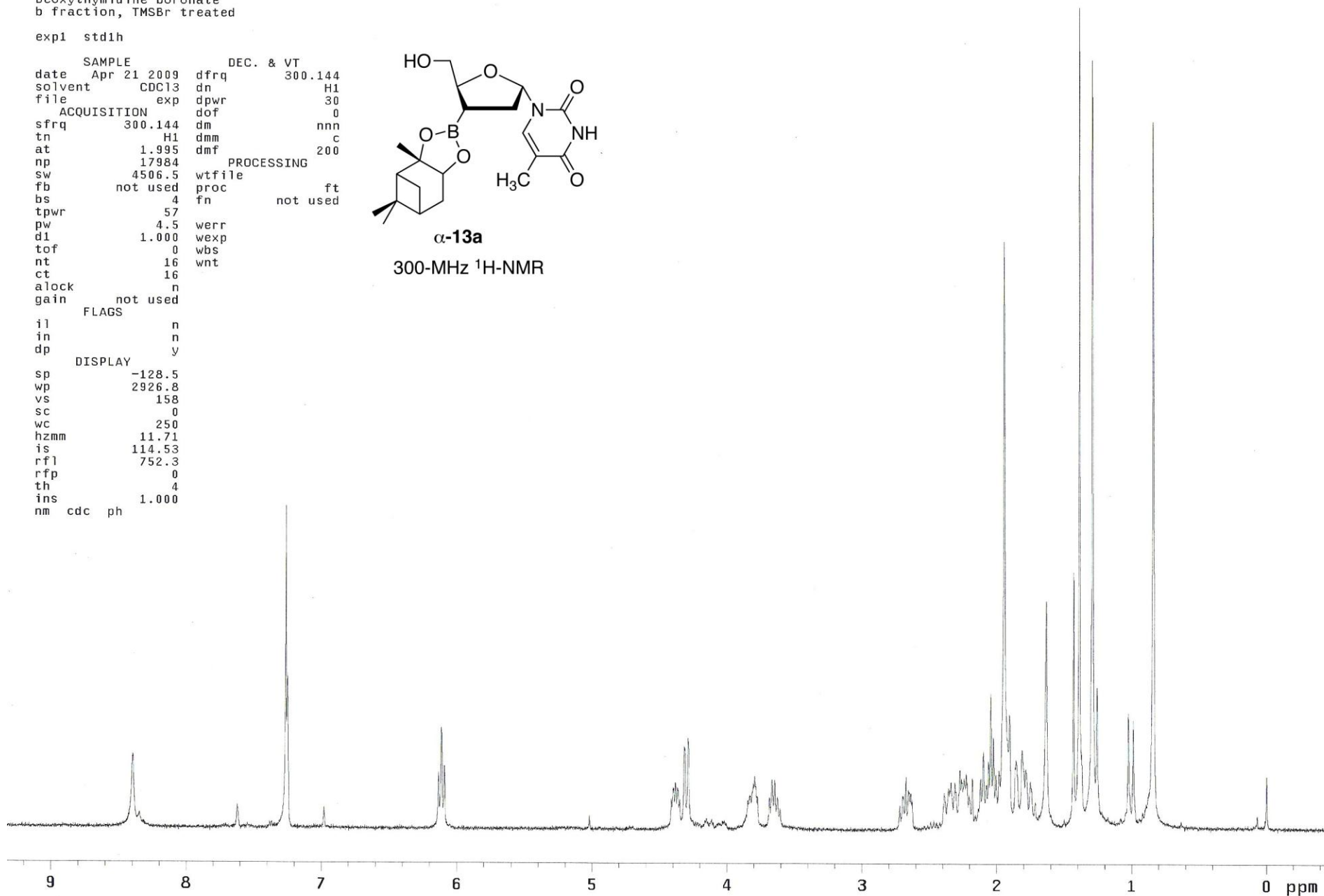
exptl std1h

SAMPLE		DEC. & VT	
date	Apr 21 2009	dfrq	300.144
solvent	CDC13	dn	H1
file	exp	dpwr	30
ACQUISITION		dof	0
sfrq	300.144	dm	nnn
tn	H1	dmm	c
at	1.995	dmf	200
np	17984	PROCESSING	
sw	4506.5	wtfile	
fb	not used	proc	ft
bs	4	fn	not used
tpwr	57		
pw	4.5	werr	
d1	1.000	wexp	
tof	0	wbs	
nt	16	wnt	
ct	16		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	y		
DISPLAY			
sp	-128.5		
wp	2926.8		
vs	158		
sc	0		
wc	250		
hzmm	11.71		
is	114.53		
rfl	752.3		
rfp	0		
th	4		
ins	1.000		
nm	cdc ph		



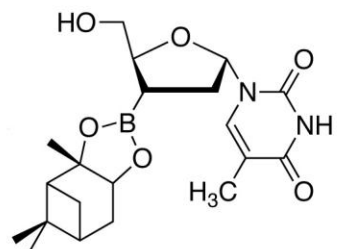
**$\alpha$ -13a**

300-MHz  $^1\text{H}$ -NMR



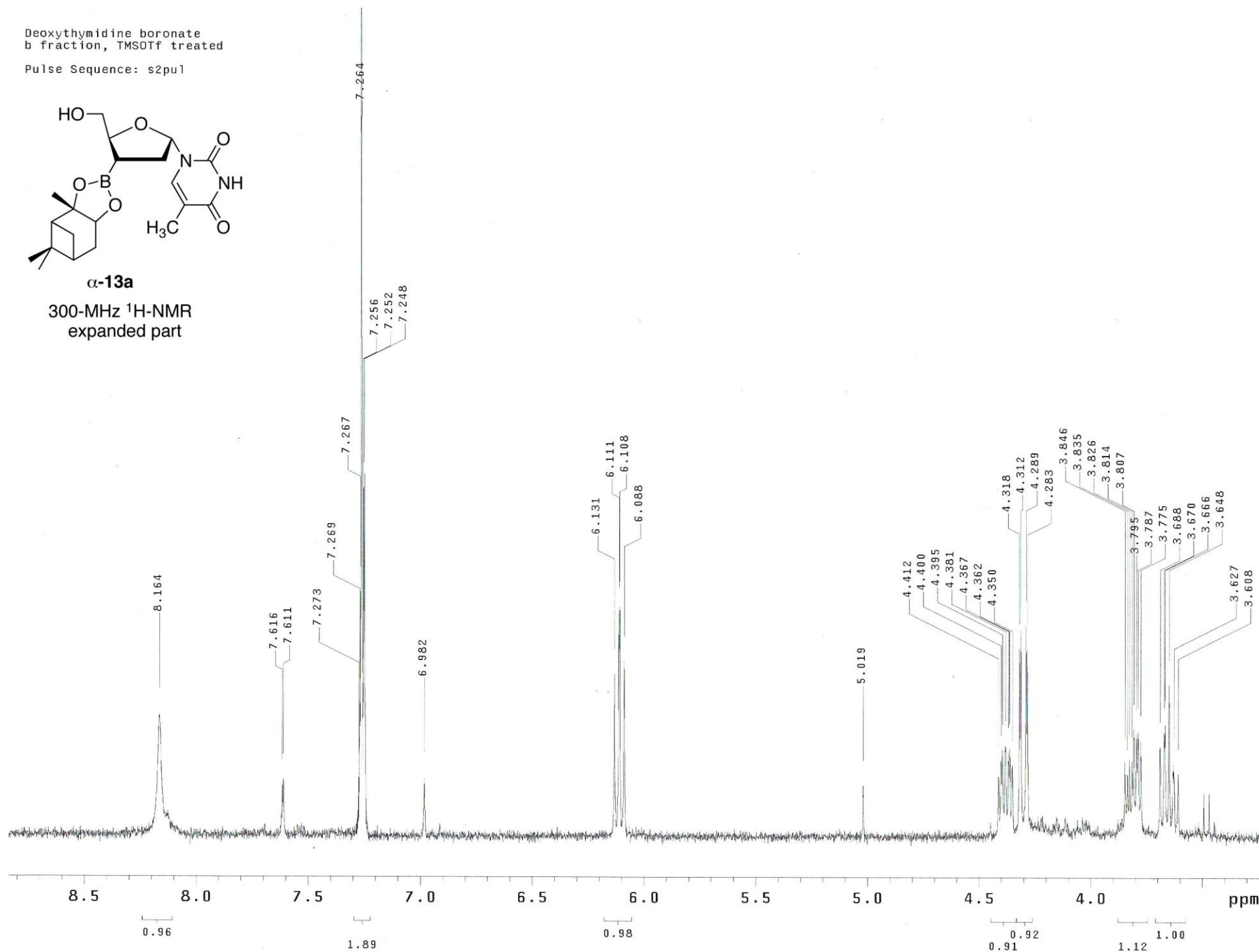
Deoxythymidine boronate  
b fraction, TMSOTf treated

Pulse Sequence: s2pu1



**$\alpha$ -13a**

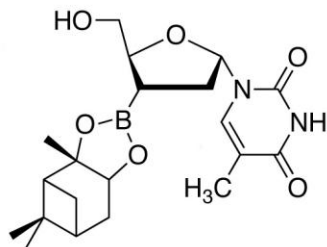
300-MHz  $^1\text{H}$ -NMR  
expanded part



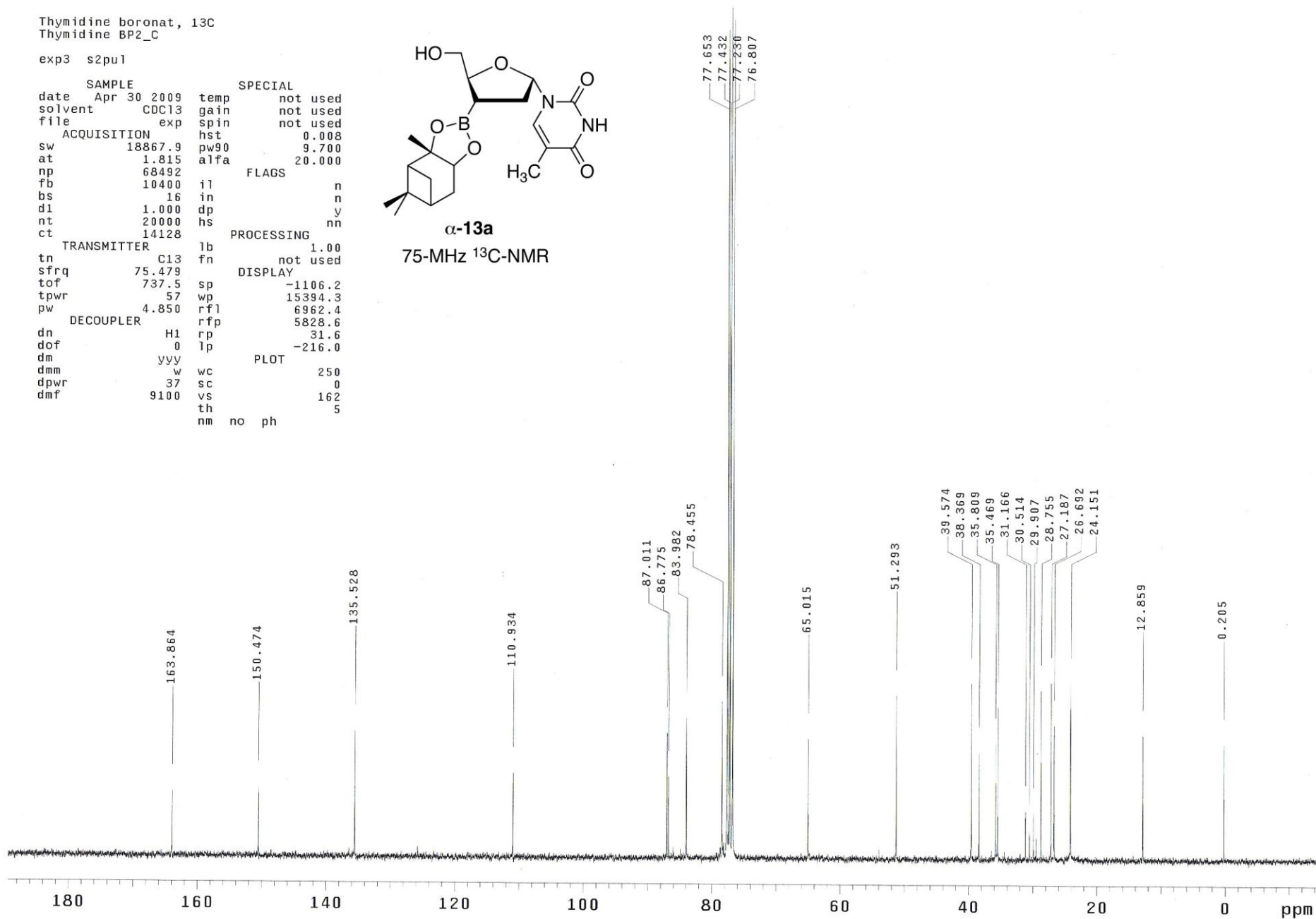
Thymidine boronat, <sup>13</sup>C  
Thymidine BP2\_C

exp3 s2pu1

SAMPLE		SPECIAL	
date	Apr 30 2009	temp	not used
solvent	CDC13	gain	not used
file	exp	spin	not used
ACQUISITION		hst	0.008
sw	18867.9	pw90	9.700
at	1.815	alfa	20.000
np	68492	FLAGS	
fb	10400	il	n
bs	16	in	n
d1	1.000	dp	y
nt	20000	hs	nn
ct	14128	PROCESSING	
TRANSMITTER		lb	1.00
tn	C13	fn	not used
sfrq	75.479	DISPLAY	
tof	737.5	sp	-1106.2
tpwr	57	wp	15394.3
pw	4.850	rfl	6962.4
DECOUPLER		rfp	5828.6
dn	H1	rp	31.6
dof	0	lp	-216.0
		PLOT	
dmm	yyy	wc	250
dpwr	37	sc	0
dmf	9100	vs	162
		th	5
		nm	no ph



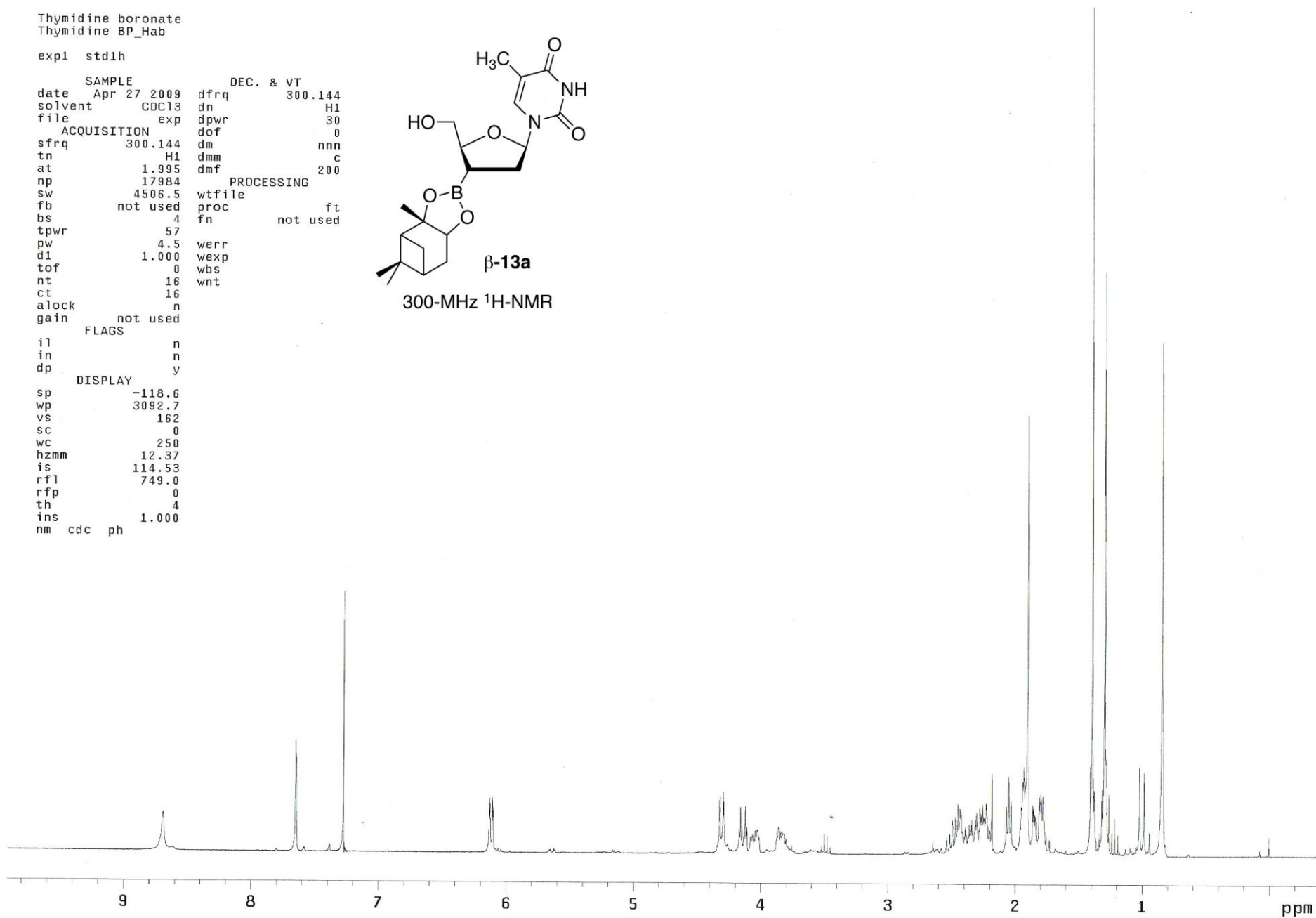
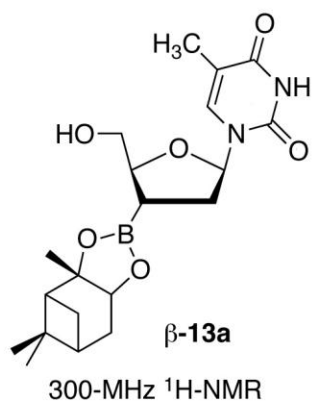
$\alpha$ -13a  
75-MHz <sup>13</sup>C-NMR



Thymidine boronate  
Thymidine BP\_Hab

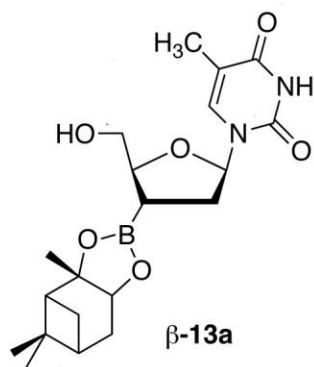
exp1 std1h

SAMPLE		DEC. & VT	
date	Apr 27 2009	dfrq	300.144
solvent	CDC13	dn	H1
file	exp	dpwr	30
ACQUISITION		dof	0
sfrq	300.144	dm	nnn
tn	H1	dmm	c
at	1.995	dmf	200
np	17984	PROCESSING	
sw	4506.5	wtfile	
fb	not used	proc	ft
bs	4	fn	not used
tpwr	57		
pw	4.5	werr	
dl	1.000	wexp	
tof	0	wbs	
nt	16	wnt	
ct	16		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	y		
DISPLAY			
sp	-118.6		
wp	3092.7		
vs	162		
sc	0		
wc	250		
hzmm	12.37		
is	114.53		
rfl	749.0		
rfp	0		
th	4		
ins	1.000		
nm	cdc ph		

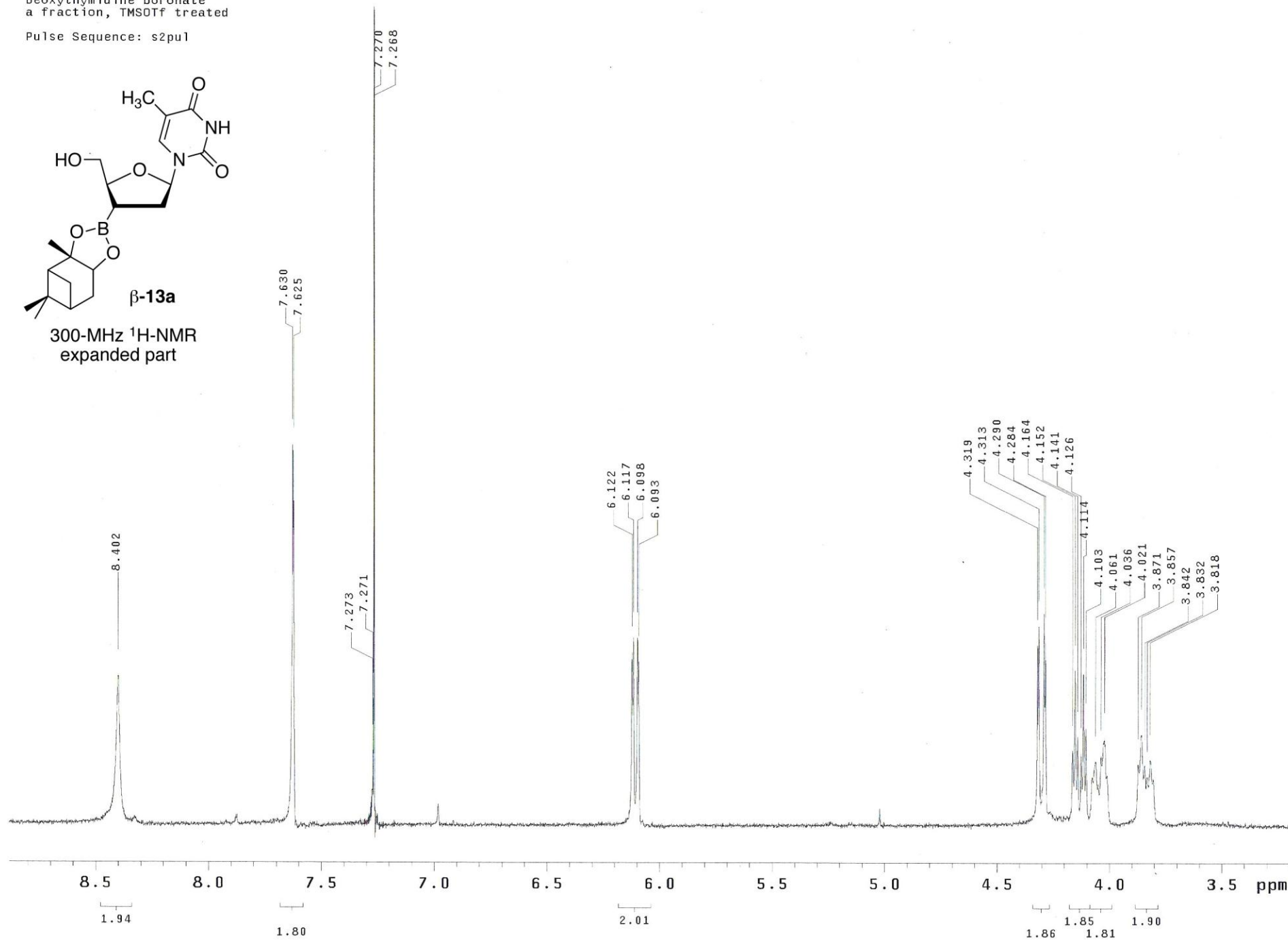


Deoxythymidine boronate  
a fraction, TMSOTf treated

Pulse Sequence: s2pu1



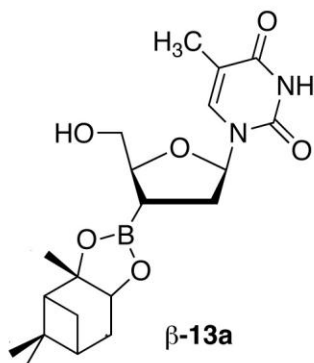
300-MHz  $^1\text{H}$ -NMR  
expanded part



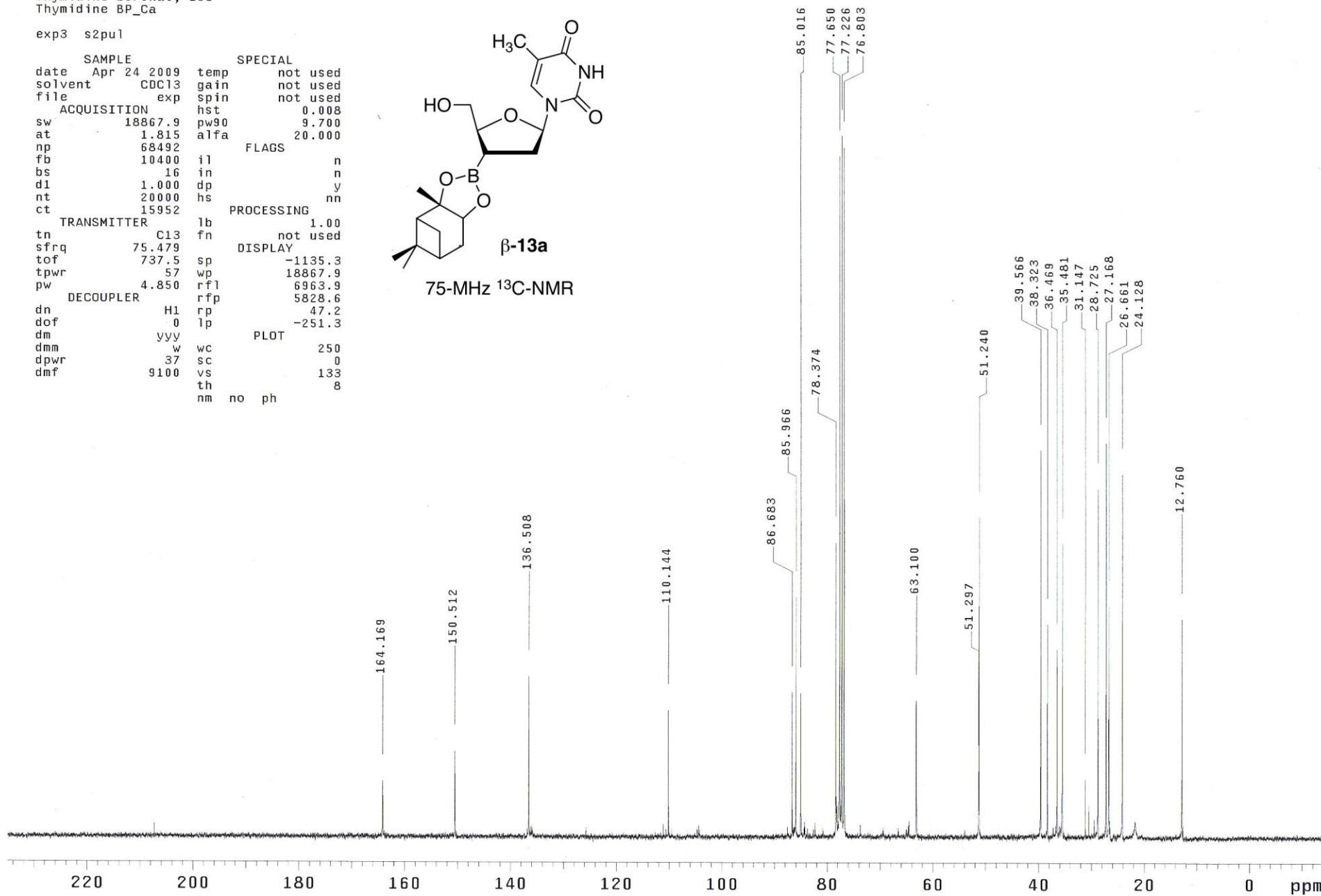
Thymidine boronat, <sup>13</sup>C  
Thymidine BP\_Ca

exp3 s2pu1

SAMPLE		SPECIAL	
date	Apr 24 2009	temp	not used
solvent	CDCl <sub>3</sub>	gain	not used
file	exp	spin	not used
ACQUISITION		hst	0.008
sw	18867.9	pw90	9.700
at	1.815	alfa	20.000
np	68492	FLAGS	
fb	10400	il	n
bs	16	in	n
d1	1.000	dp	y
nt	20000	hs	nn
ct	15952	PROCESSING	
TRANSMITTER		lb	1.00
tn	C13	fn	not used
sfrq	75.479	DISPLAY	
tof	737.5	sp	-1135.3
tpwr	57	wp	18867.9
pw	4.850	rfl	6963.9
DECOUPLER		rfp	5828.6
dn	H1	rp	47.2
dof	0	lp	-251.3
dm	yyy	PLOT	
dmm	w	wc	250
dpwr	37	sc	0
dmf	9100	vs	133
		th	8
		nm	no ph



75-MHz <sup>13</sup>C-NMR

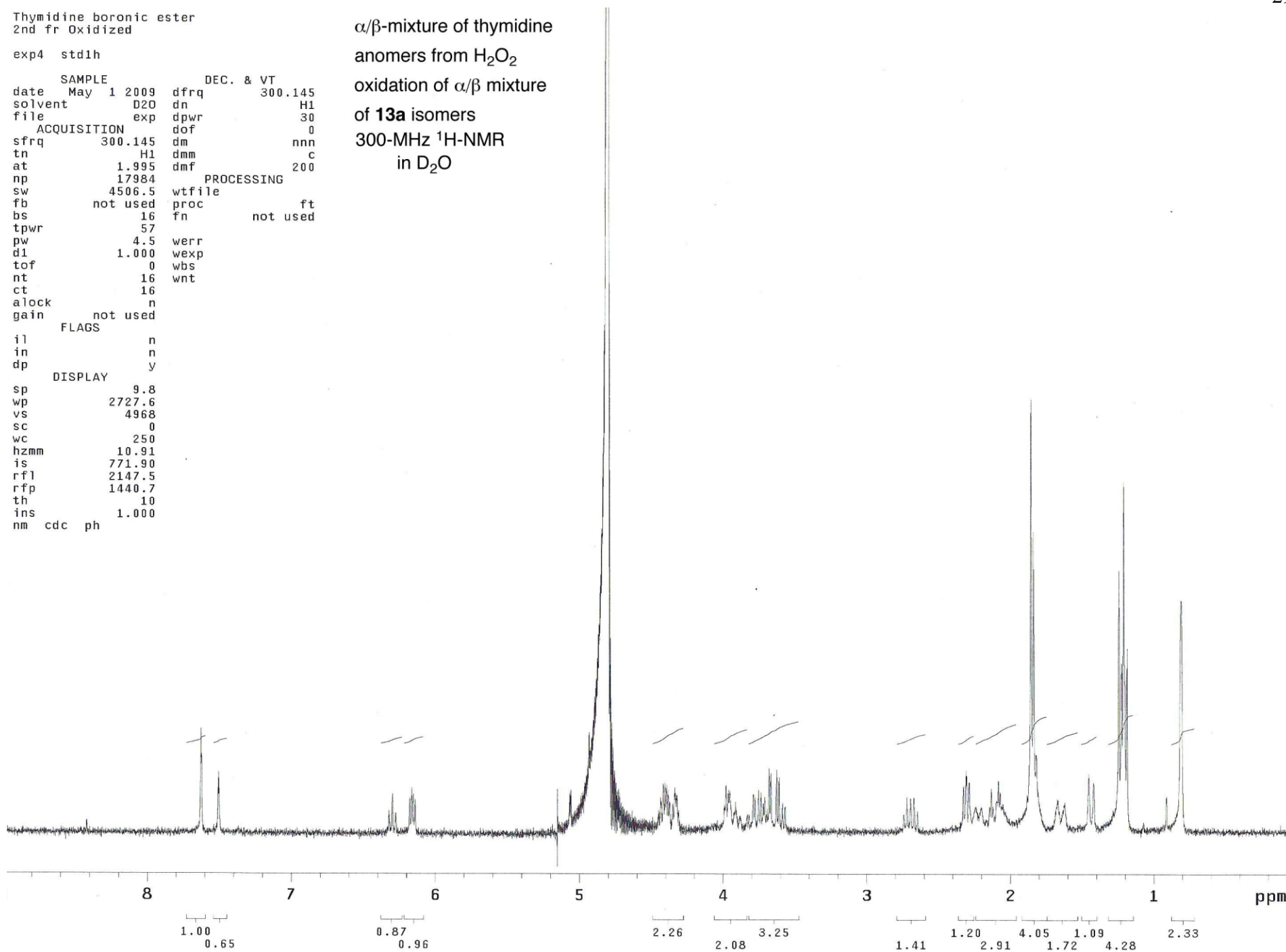


Thymidine boronic ester  
2nd fr Oxidized

exp4 std1h

SAMPLE		DEC. & VT	
date	May 1 2009	dfrq	300.145
solvent	D2O	dn	H1
file	exp	dpwr	30
ACQUISITION		dof	0
sfrq	300.145	dm	nnn
tn	H1	dmm	C
at	1.995	dmf	200
np	17984	PROCESSING	
sw	4506.5	wtfile	ft
fb	not used	proc	fn
bs	16	not used	
tpwr	57		
pw	4.5	werr	
d1	1.000	wexp	
tof	0	wbs	
nt	16	wnt	
ct	16		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	y		
DISPLAY			
sp	9.8		
wp	2727.6		
vs	4968		
sc	0		
wc	250		
hzmm	10.91		
is	771.90		
rfl	2147.5		
rfp	1440.7		
th	10		
ins	1.000		
nm	cdc ph		

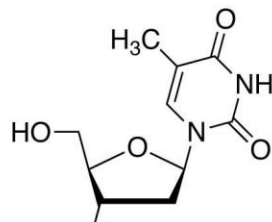
$\alpha/\beta$ -mixture of thymidine  
anomers from H<sub>2</sub>O<sub>2</sub>  
oxidation of  $\alpha/\beta$  mixture  
of **13a** isomers  
300-MHz <sup>1</sup>H-NMR  
in D<sub>2</sub>O



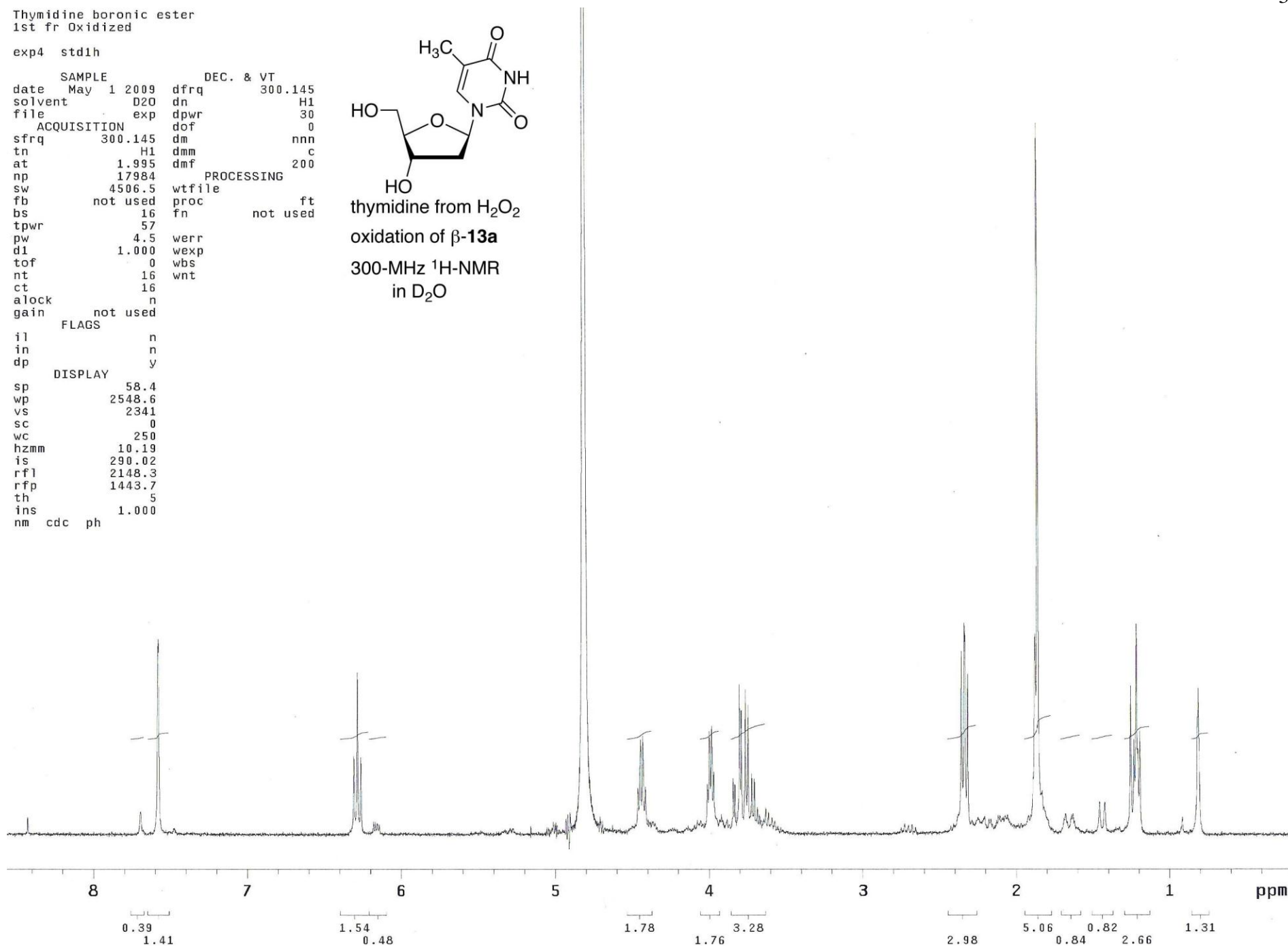
Thymidine boronic ester  
1st fr Oxidized

exp4 std1h

SAMPLE		DEC. & VT	
date	May 1 2009	dfrq	300.145
solvent	D2O	dn	H1
file	exp	dpwr	30
ACQUISITION		dof	0
sfrq	300.145	dm	nnn
tn	H1	dmm	c
at	1.995	dmf	200
np	17984	PROCESSING	
sw	4506.5	wtfile	ft
fb	not used	proc	fn
bs	16	not used	
tpwr	57	werr	
pw	4.5	wexp	
d1	1.000	wbs	
tof	0	wnt	
nt	16		
ct	16		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	y		
DISPLAY			
sp	58.4		
wp	2548.6		
vs	2341		
sc	0		
wc	250		
hzmm	10.19		
is	290.02		
rfl	2148.3		
rfp	1443.7		
th	5		
ins	1.000		
nm	cdc ph		



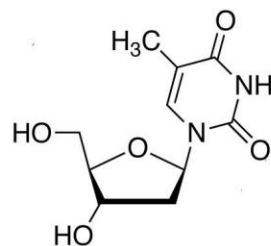
thymidine from H<sub>2</sub>O<sub>2</sub>  
oxidation of β-13a  
300-MHz <sup>1</sup>H-NMR  
in D<sub>2</sub>O



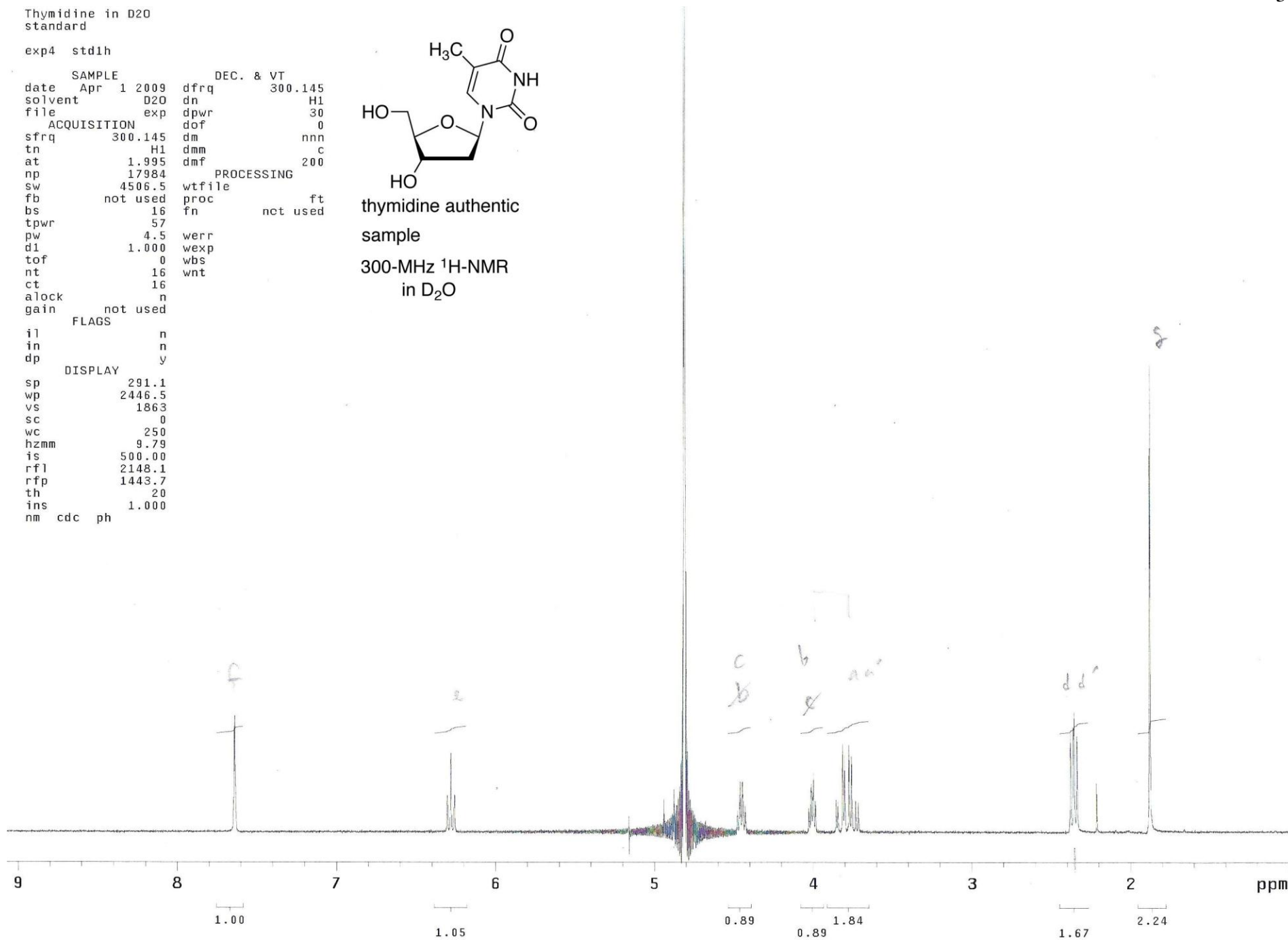
Thymidine in D2O  
standard

exp4 std1h

SAMPLE		DEC. & VT	
date	Apr 1 2009	dfrq	300.145
solvent	D2O	dn	H1
file	exp	dpwr	30
ACQUISITION		do	0
sfrq	300.145	dm	nnn
tn	H1	dmm	c
at	1.995	dmf	200
np	17984	PROCESSING	
sw	4506.5	wtfile	
fb	not used	proc	ft
bs	16	fn	not used
tpwr	57		
pw	4.5	werr	
d1	1.000	wexp	
tof	0	wbs	
nt	16	wnt	
ct	16		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	y		
DISPLAY			
sp	291.1		
wp	2446.5		
vs	1863		
sc	0		
wc	250		
hzmm	9.79		
is	500.00		
rfl	2148.1		
rfp	1443.7		
th	20		
ins	1.000		
nm	cdc ph		



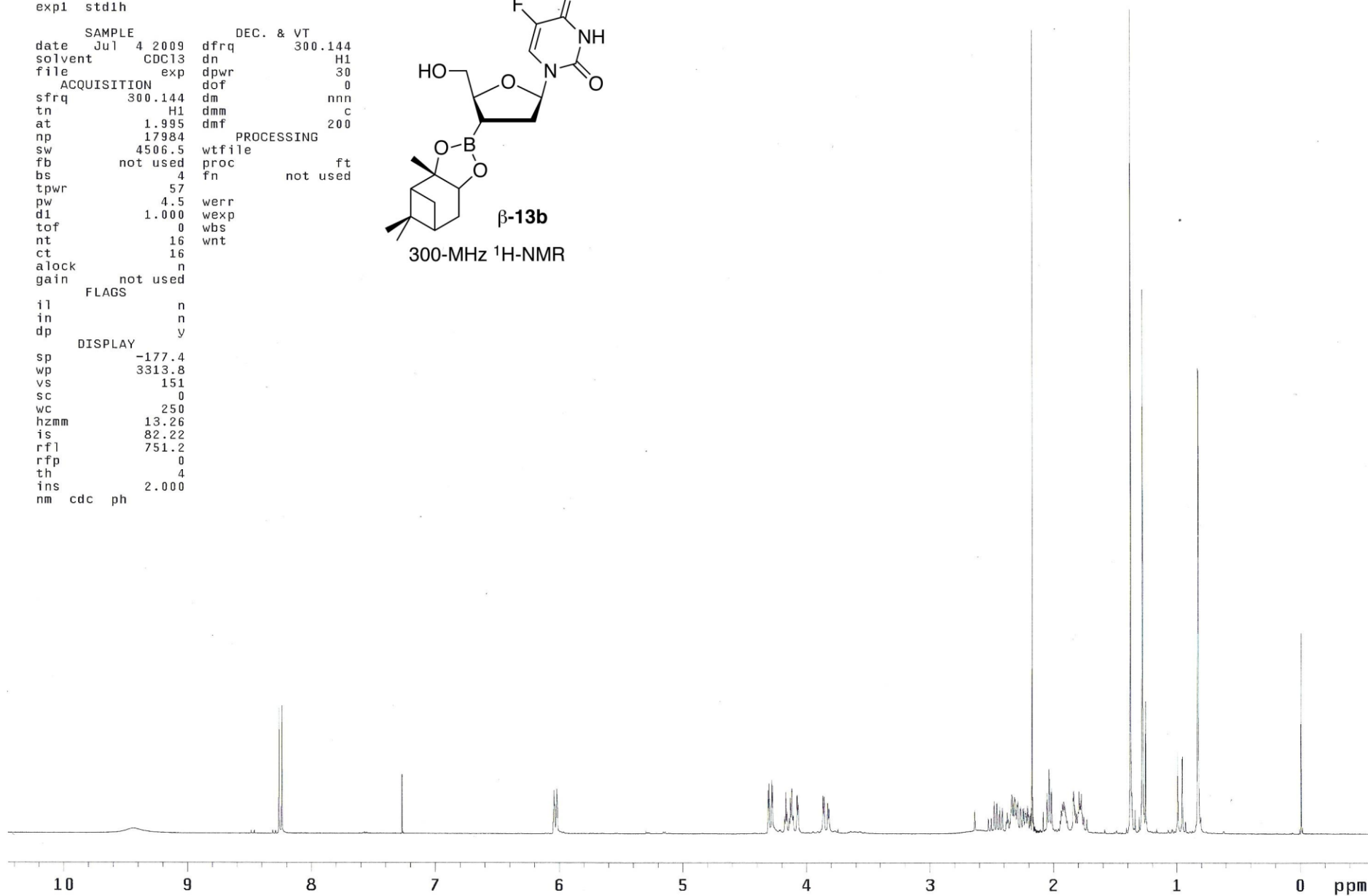
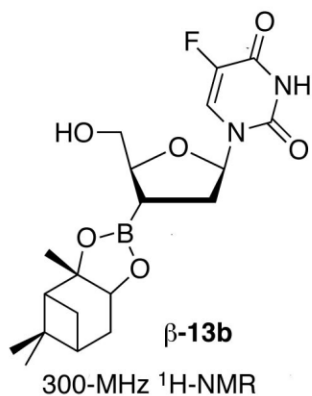
thymidine authentic  
sample  
300-MHz <sup>1</sup>H-NMR  
in D<sub>2</sub>O



Fluorouridine boronate  
PLC purified, 30mg

exp1 std1h

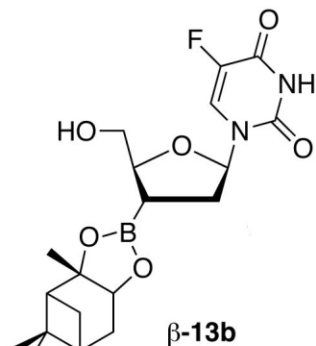
SAMPLE		DEC. & VT	
date	Jul 4 2009	dfrq	300.144
solvent	CDC13	dn	H1
file	exp	dpwr	30
ACQUISITION		dof	0
sfrq	300.144	dm	nnn
tn	H1	dmm	c
at	1.995	dmf	200
np	17984	PROCESSING	
sw	4506.5	wtfile	
fb	not used	proc	ft
bs	4	fn	not used
tpwr	57		
pw	4.5	werr	
d1	1.000	wexp	
tof	0	wbs	
nt	16	wnt	
ct	16		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	y		
DISPLAY			
sp	-177.4		
wp	3313.8		
vs	151		
sc	0		
wc	250		
hzmm	13.26		
is	82.22		
rfl	751.2		
rfp	0		
th	4		
ins	2.000		
nm	cdc ph		



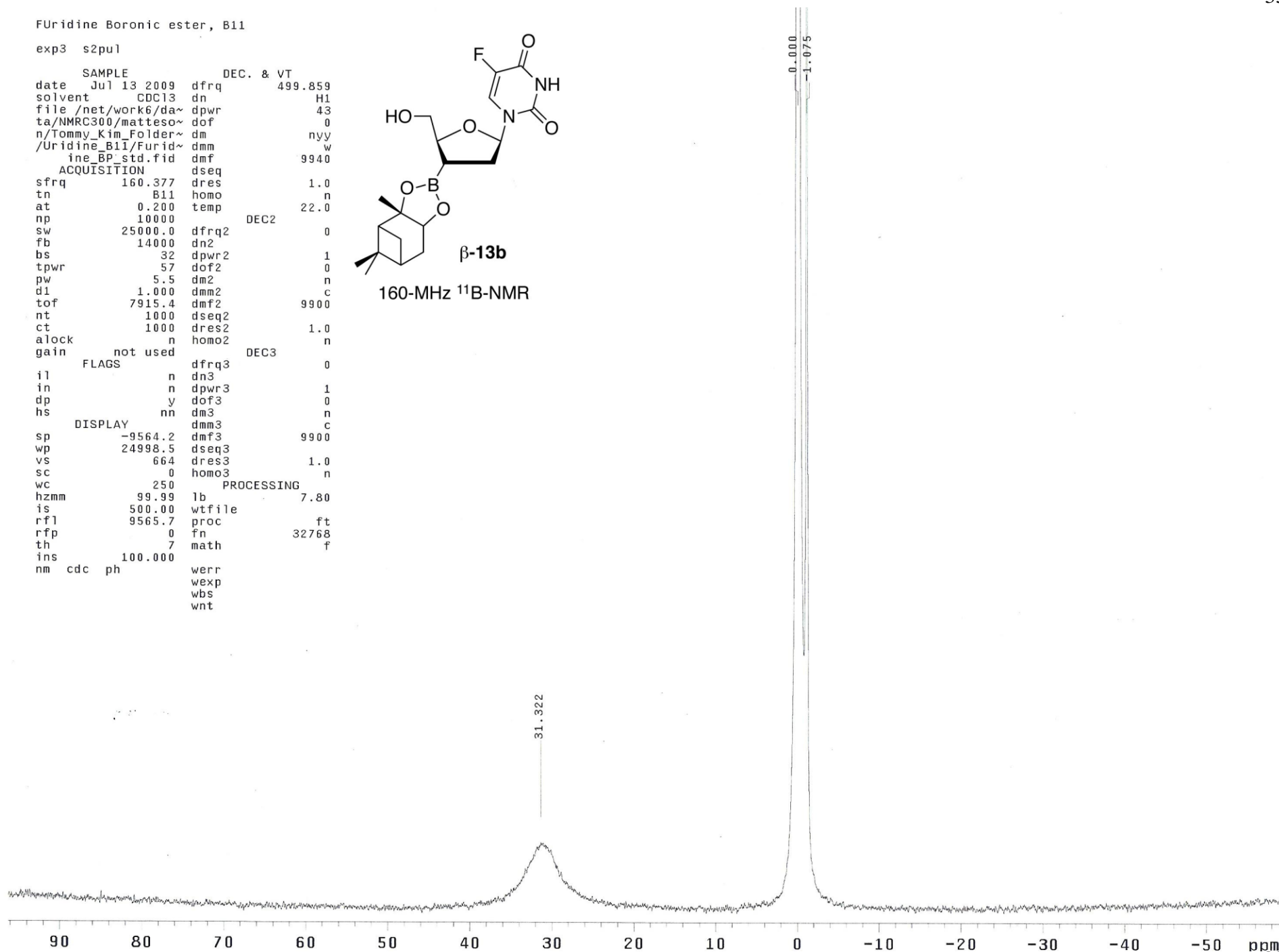
FUridine Boronic ester, B11

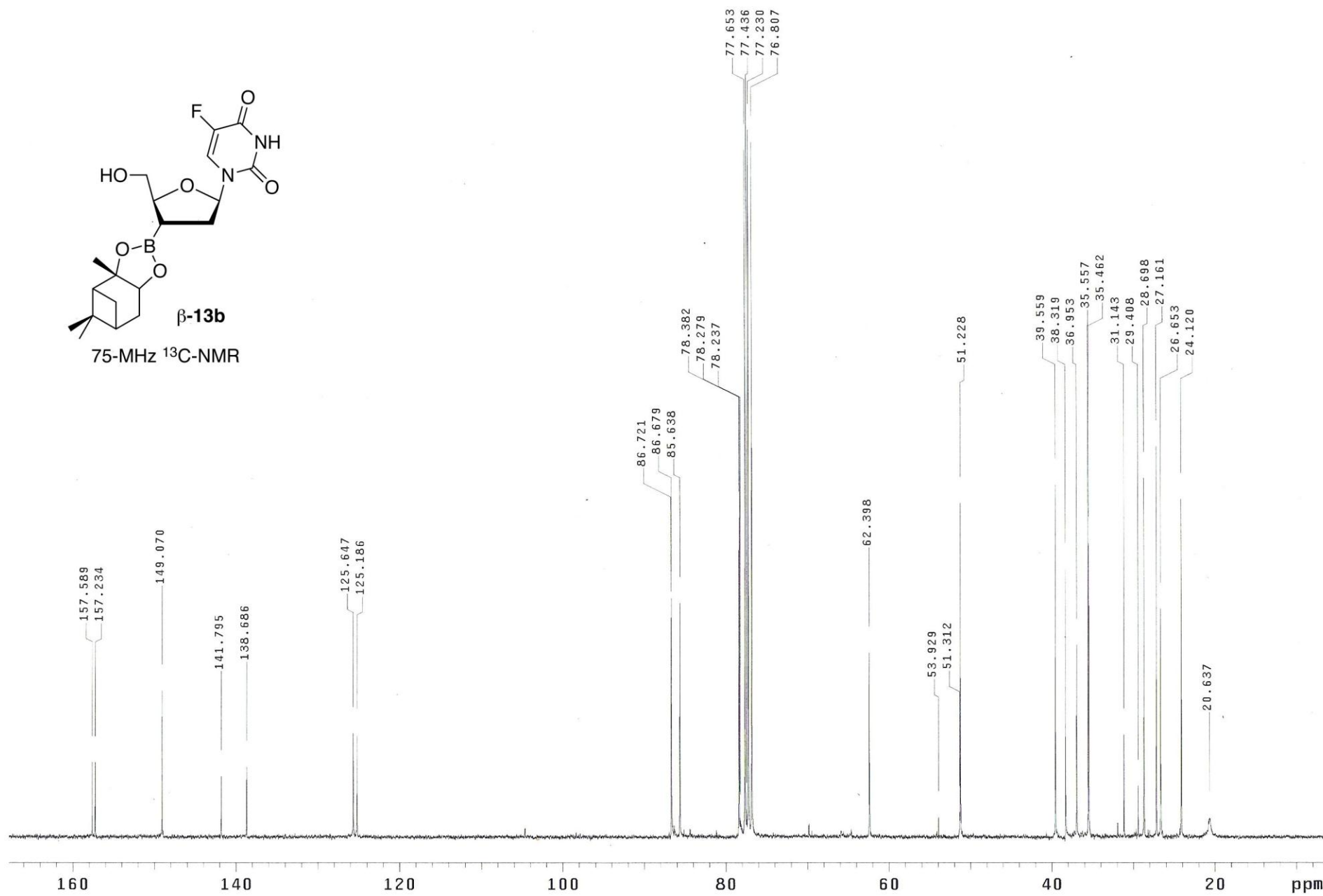
exp3 s2pu1

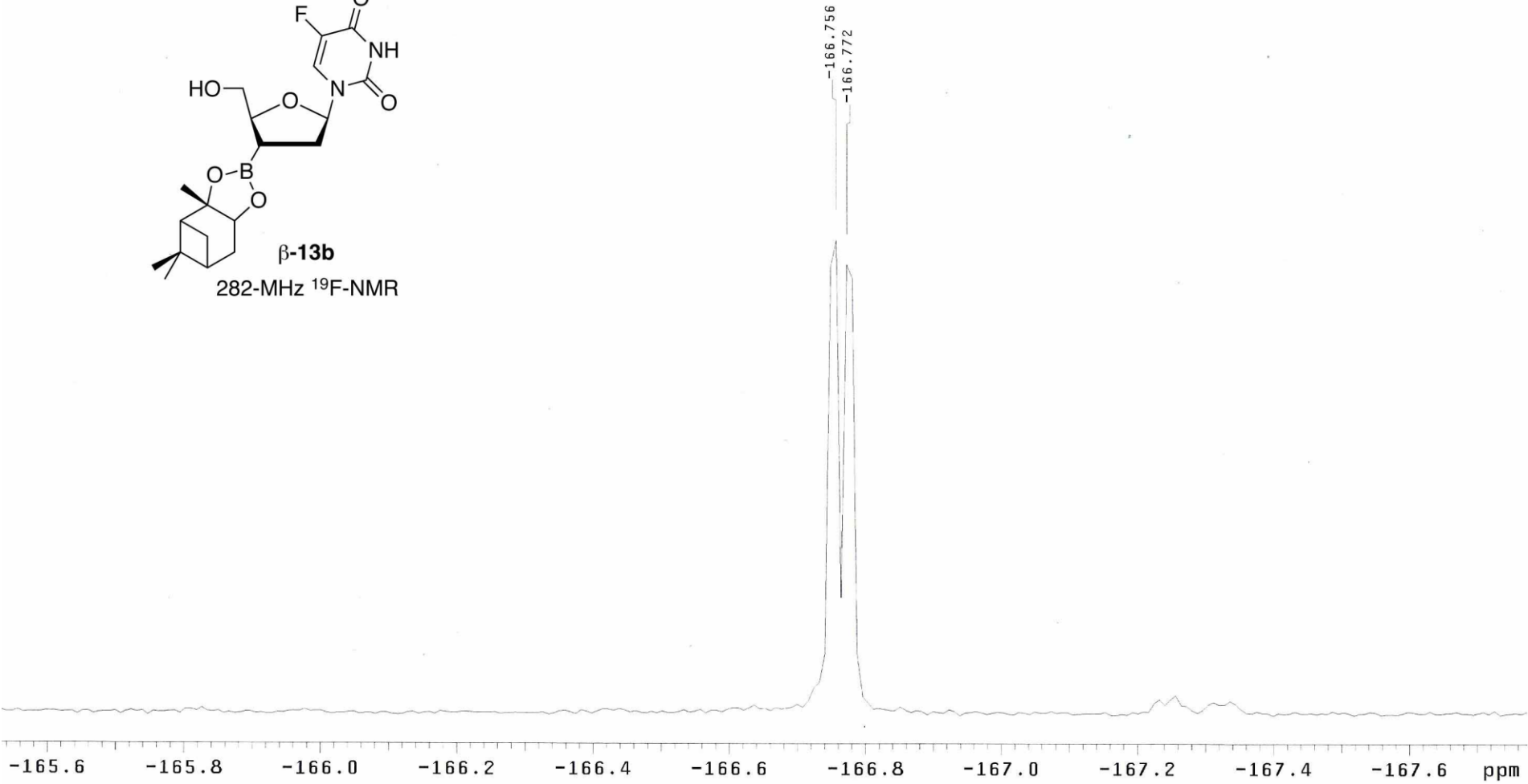
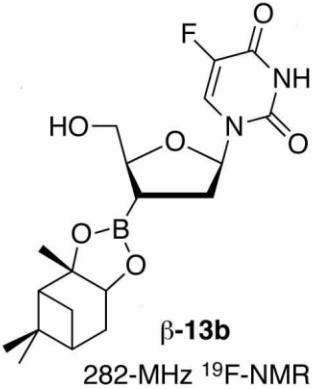
SAMPLE		DEC. & VT	
date	Jul 13 2009	dfrq	499.859
solvent	CDCl3	dn	H1
file	/net/work6/da~	dpwr	43
ta/NMRC300/matteso~		dof	0
n/Tommy_Kim_Folder~		dm	nyy
/Uridine_B11/Furid~		dmm	w
ine_BP_std.fid		dmf	9940
ACQUISITION			
sfrq	160.377	dres	1.0
tn	B11	homo	n
at	0.200	temp	22.0
np	10000	DEC2	
sw	25000.0	dfrq2	0
fb	14000	dn2	
bs	32	dpwr2	1
tpwr	57	dof2	0
pw	5.5	dm2	n
d1	1.000	dmm2	c
tof	7915.4	dmf2	9900
nt	1000	dseq2	
ct	1000	dres2	1.0
alock	n	homo2	n
gain	not used	DEC3	
FLAGS			
il	n	dfrq3	0
in	n	dn3	
dp	y	dpwr3	1
hs	nn	dof3	0
		dm3	n
		dmm3	c
DISPLAY			
sp	-9564.2	dmf3	9900
wp	24998.5	dseq3	
vs	664	dres3	1.0
sc	0	homo3	n
wc	250	PROCESSING	
hzmm	99.99	lb	7.80
is	500.00	wtfile	
rfl	9565.7	proc	ft
rfp	0	fn	32768
th	7	math	f
ins	100.000		
nm	cdc ph	werr	
		wexp	
		wbs	
		wnt	



160-MHz <sup>11</sup>B-NMR



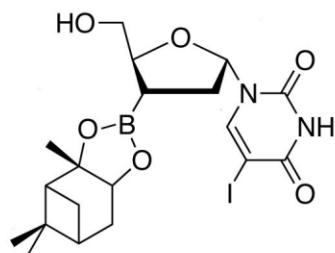




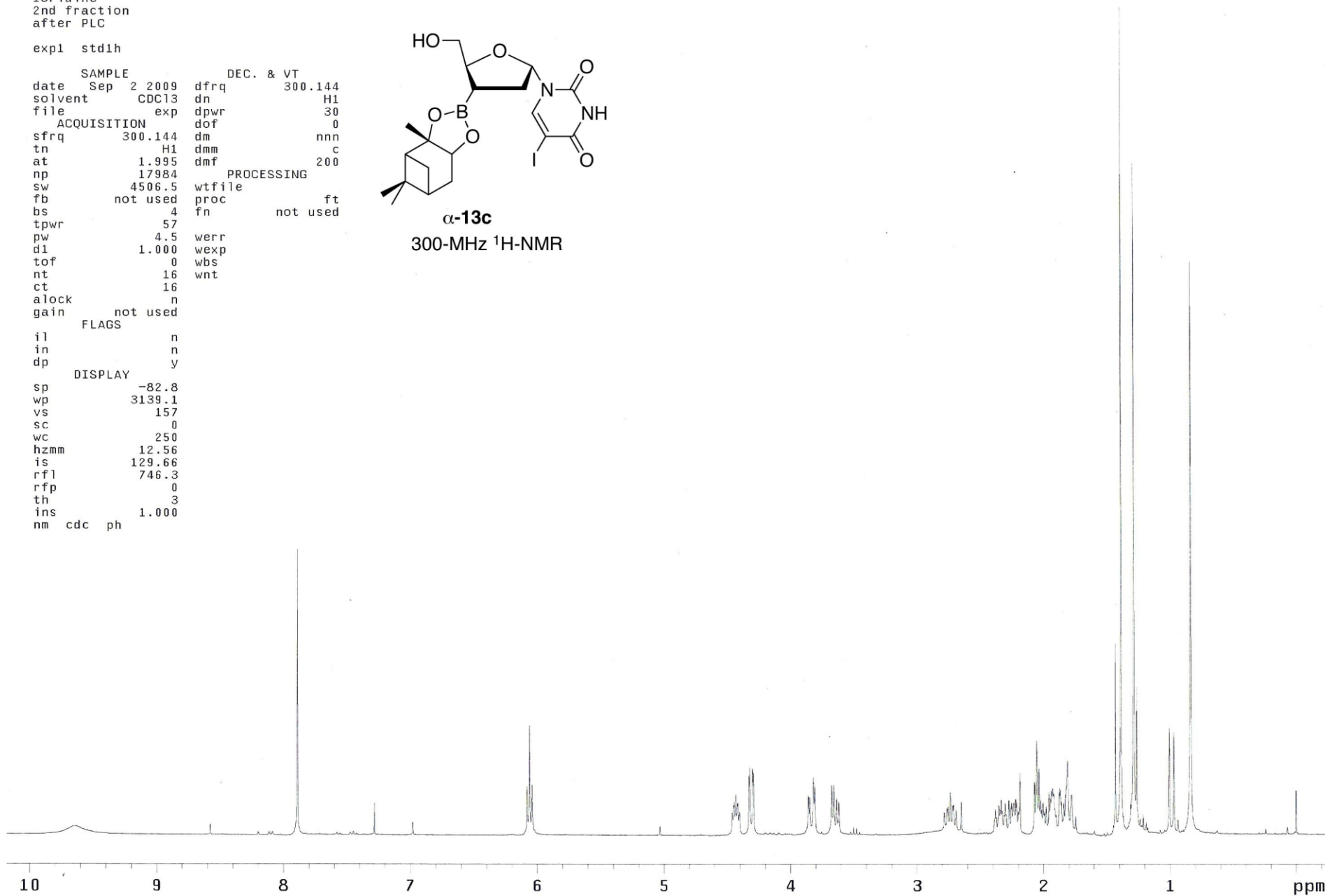
IUridine  
2nd fraction  
after PLC

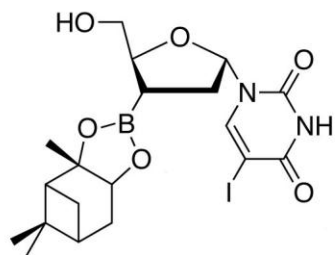
expl std1h

SAMPLE		DEC. & VT	
date	Sep 2 2009	dfrq	300.144
solvent	CDC13	dn	H1
file	exp	dpwr	30
ACQUISITION		dof	0
sfrq	300.144	dm	nnn
tn	H1	dmm	c
at	1.995	dmf	200
np	17984	PROCESSING	
sw	4506.5	wtfile	
fb	not used	proc	ft
bs	4	fn	not used
tpwr	57		
pw	4.5	werr	
d1	1.000	wexp	
tof	0	wbs	
nt	16	wnt	
ct	16		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	y		
DISPLAY			
sp	-82.8		
wp	3139.1		
vs	157		
sc	0		
wc	250		
hzmm	12.56		
is	129.66		
rfl	746.3		
rfp	0		
th	3		
ins	1.000		
nm	cdc ph		



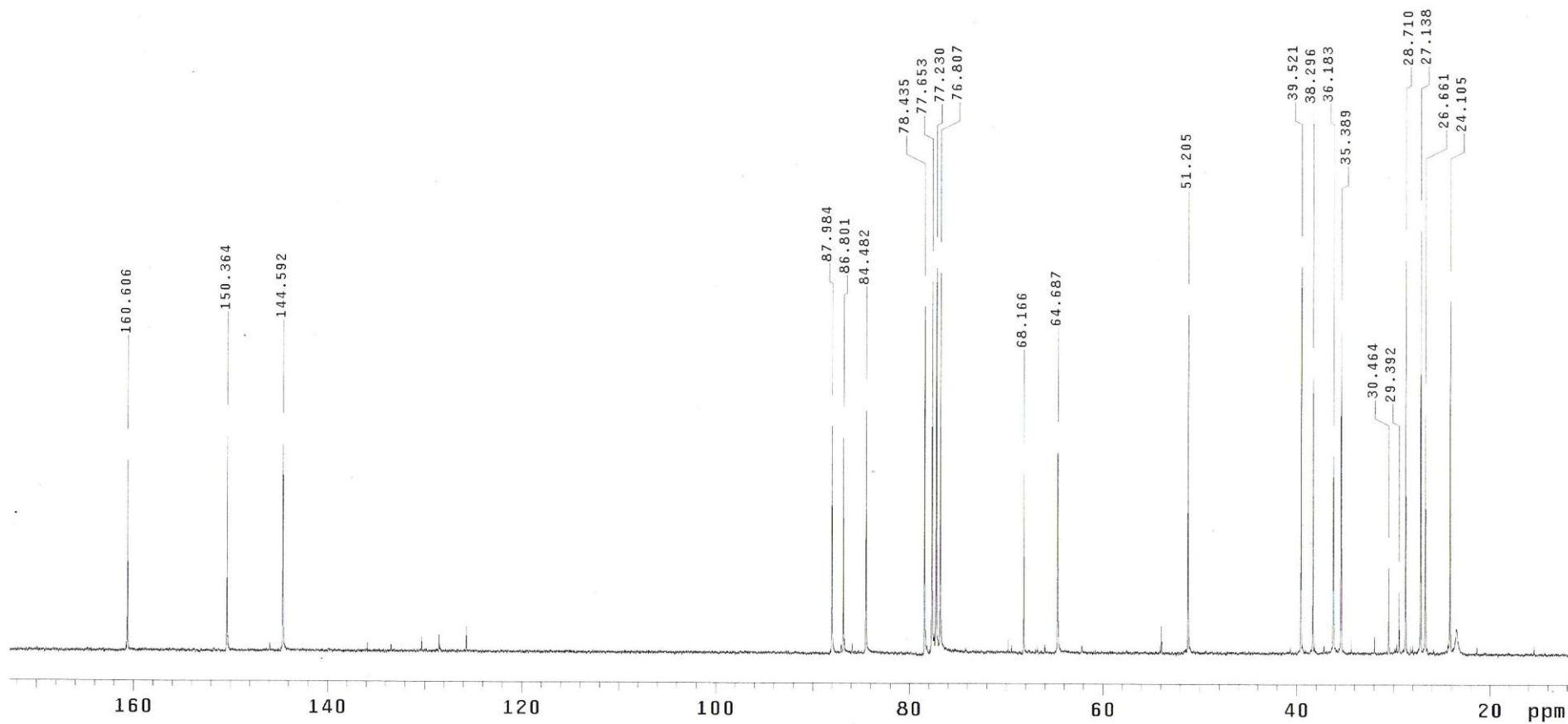
**α-13c**  
300-MHz <sup>1</sup>H-NMR





**α-13c**

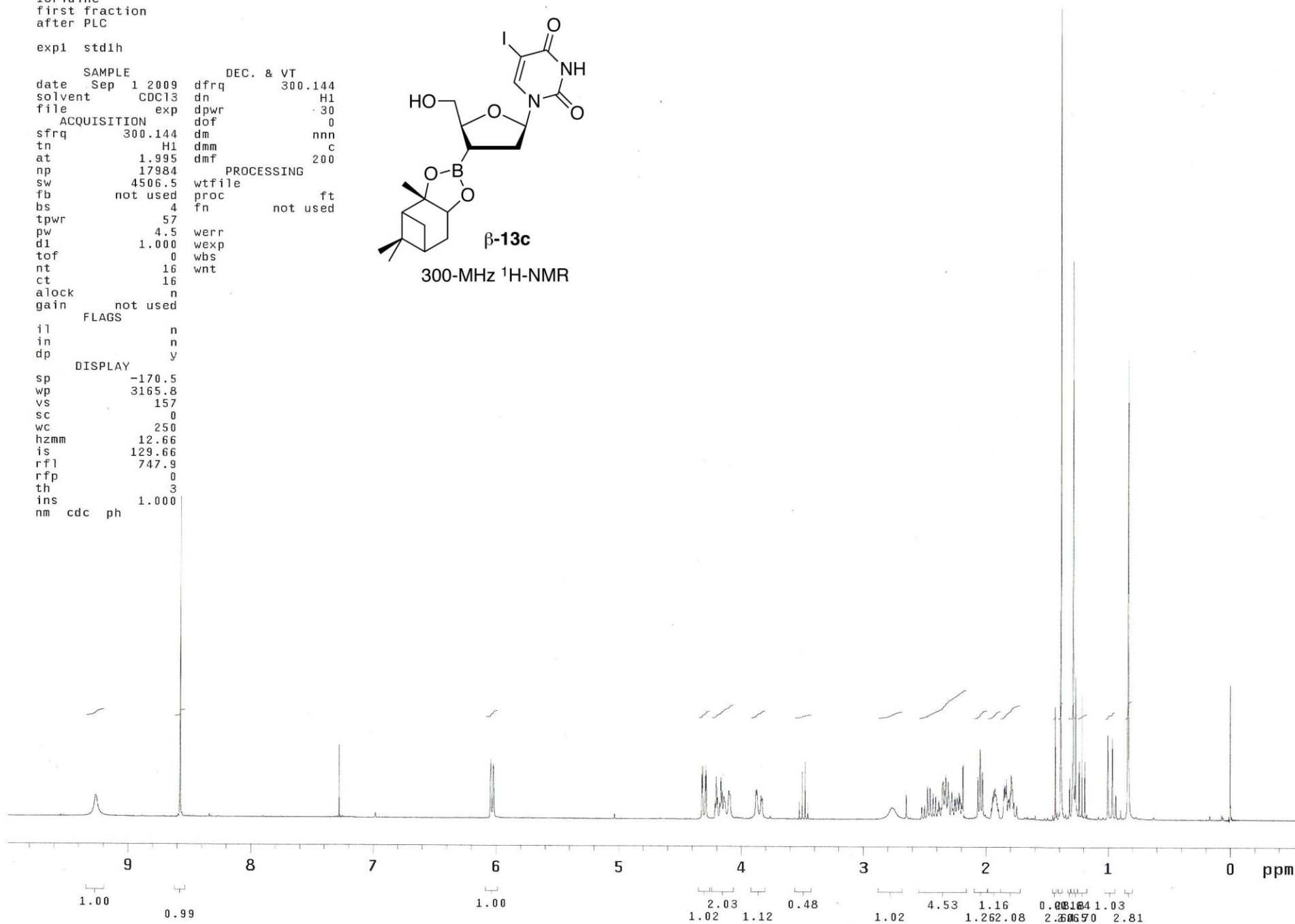
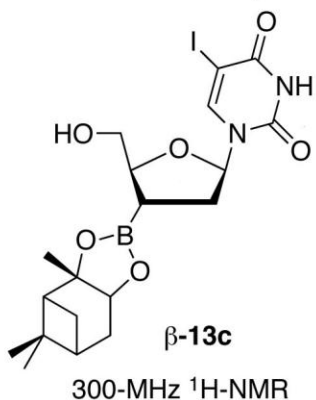
75-MHz <sup>13</sup>C-NMR

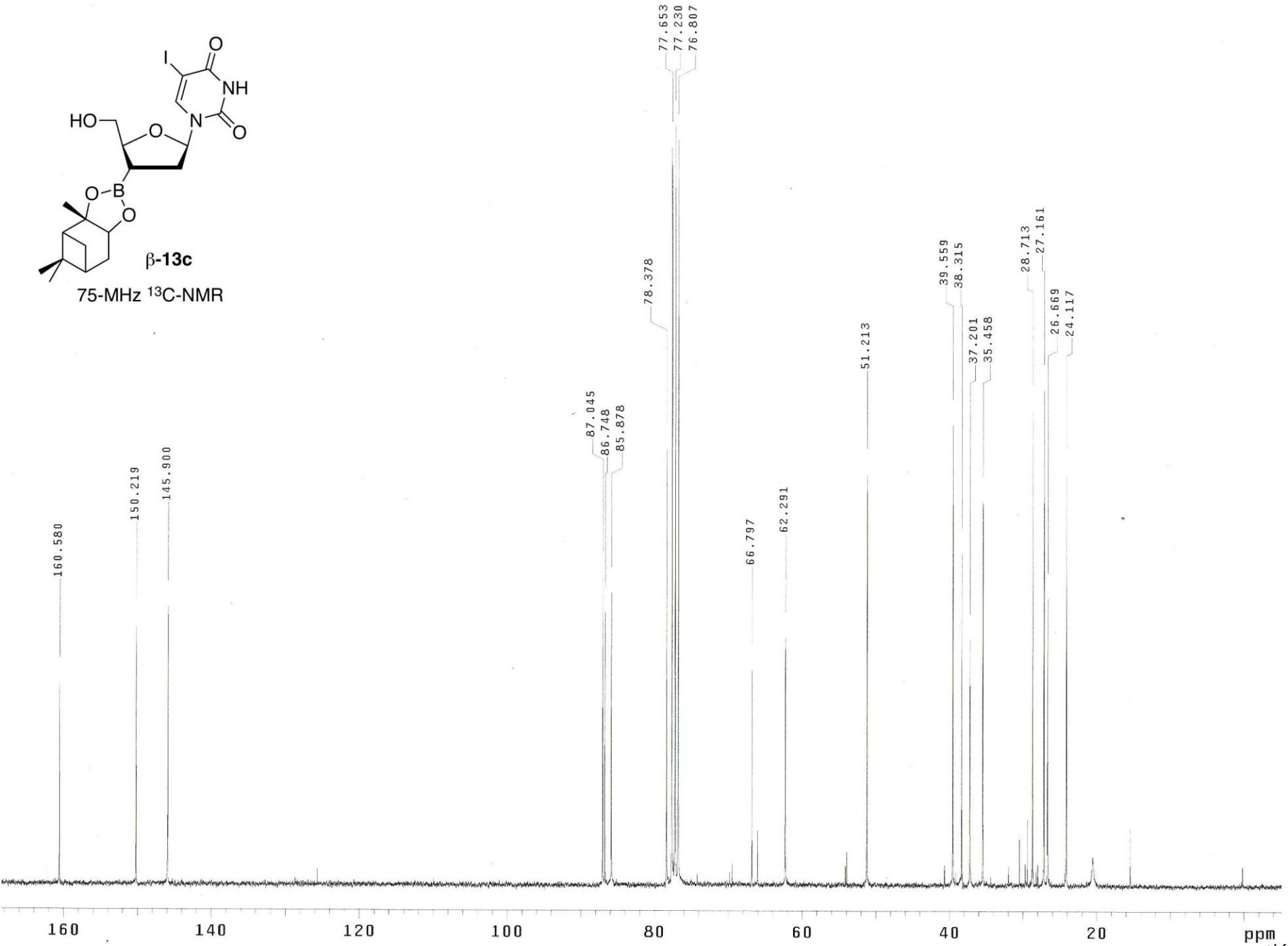


IUridine  
first fraction  
after PLC

exp1 std1h

SAMPLE		DEC. & VT	
date	Sep 1 2009	dfrq	300.144
solvent	CDC13	dn	H1
file	exp	dpwr	30
ACQUISITION		dof	0
sfrq	300.144	dm	nnn
tn	H1	dmm	c
at	1.995	dmf	200
np	17984	PROCESSING	
sw	4506.5	wtfile	
fb	not used	proc	ft
bs	4	fn	not used
tpwr	57		
pw	4.5	werr	
d1	1.000	wexp	
tof	0	wbs	
nt	16	wnt	
ct	16		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	y		
DISPLAY			
sp	-170.5		
wp	3165.8		
vs	157		
sc	0		
wc	250		
hzmm	12.66		
is	129.66		
rfl	747.9		
rfp	0		
th	3		
ins	1.000		
nm	cdc ph		

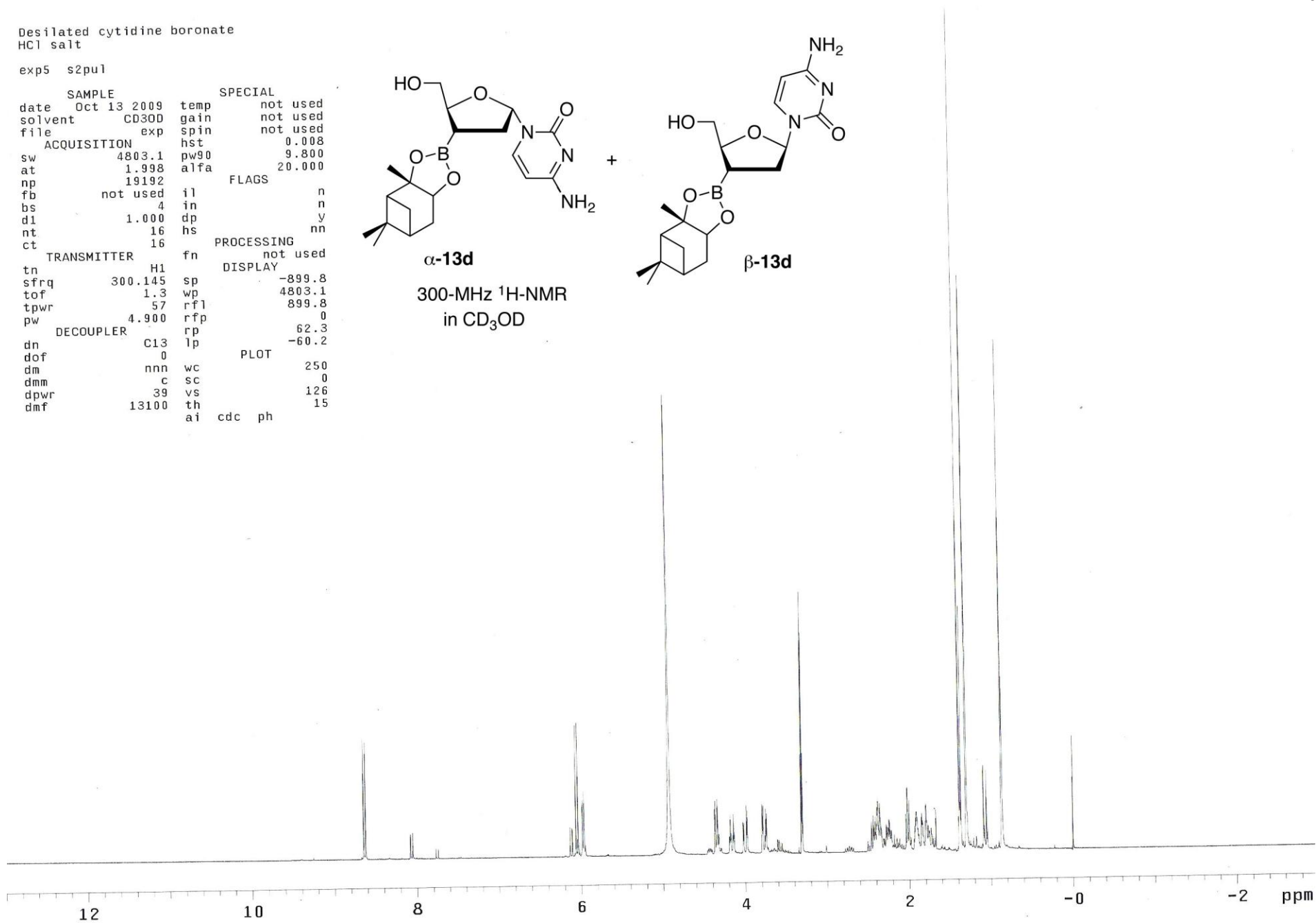
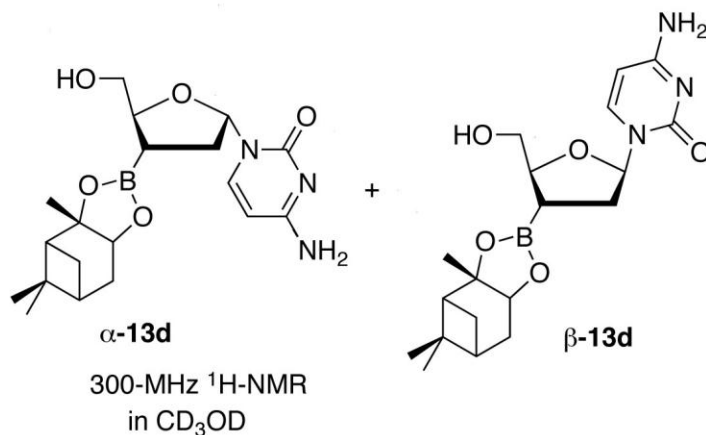




Desilated cytidine boronate  
HCl salt

exp5 s2pul

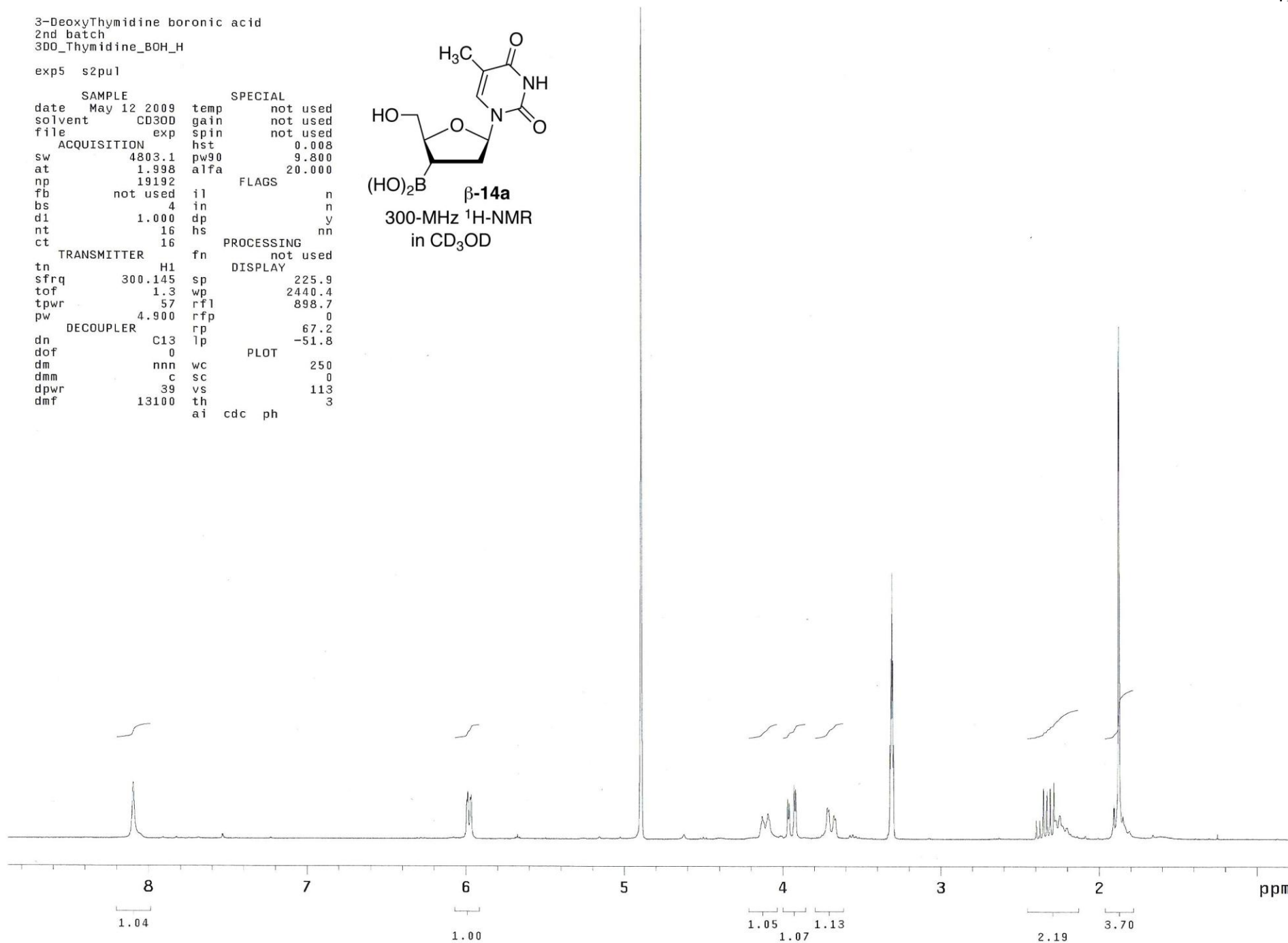
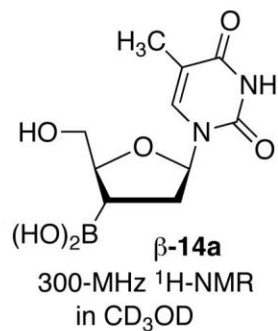
SAMPLE		SPECIAL	
date	Oct 13 2009	temp	not used
solvent	CD300	gain	not used
file	exp	spin	not used
ACQUISITION		hst	0.008
sw	4803.1	pw90	9.800
at	1.998	alfa	20.000
np	19192	FLAGS	
fb	not used	il	n
bs	4	in	n
d1	1.000	dp	y
nt	16	hs	nn
ct	16	PROCESSING	
TRANSMITTER		fn	not used
tn	H1	DISPLAY	
sfrq	300.145	sp	-899.8
tof	1.3	wp	4803.1
tpwr	57	rfl	899.8
pw	4.900	rfl	0
DECOUPLER		rp	62.3
dn	C13	lp	-60.2
dof	0	PLOT	
dm	nnn	wc	250
dmm	c	sc	0
dpwr	39	vs	126
dmf	13100	th	15
		ai	cdc ph



3-DeoxyThymidine boronic acid  
2nd batch  
300\_Thymidine\_BOH\_H

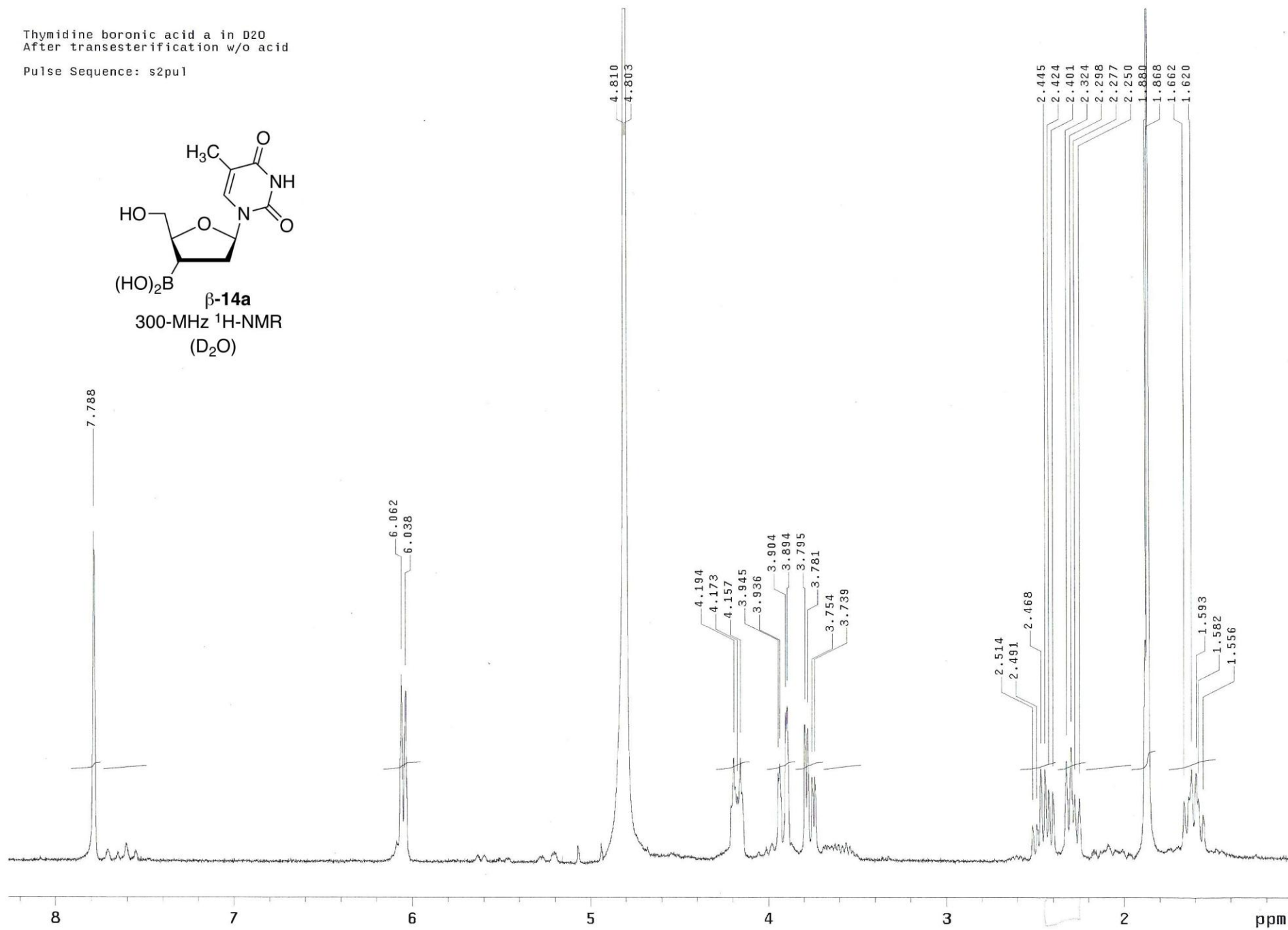
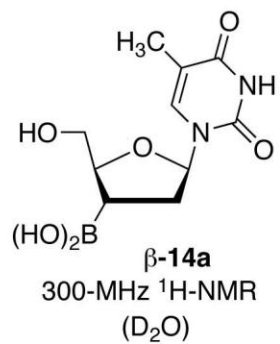
exp5 s2pu1

SAMPLE		SPECIAL	
date	May 12 2009	temp	not used
solvent	CD3OD	gain	not used
file	exp	spin	not used
ACQUISITION			
sw	4803.1	hst	0.008
at	1.998	pw90	9.800
np	19192	alfa	20.000
TRANSMITTER		PROCESSING	
tn	H1	fn	not used
sfrq	300.145	DISPLAY	
tof	1.3	sp	225.9
tpwr	57	wp	2440.4
pw	4.900	rfl	898.7
DECOUPLER		rpf	0
dn	C13	rp	67.2
dof	0	lp	-51.8
dm	nnn	PLOT	
dmm	c	wc	250
dpwr	39	sc	0
dmf	13100	vs	113
		th	3
		ai	cdc ph



Thymidine boronic acid **a** in D<sub>2</sub>O  
After transesterification w/o acid

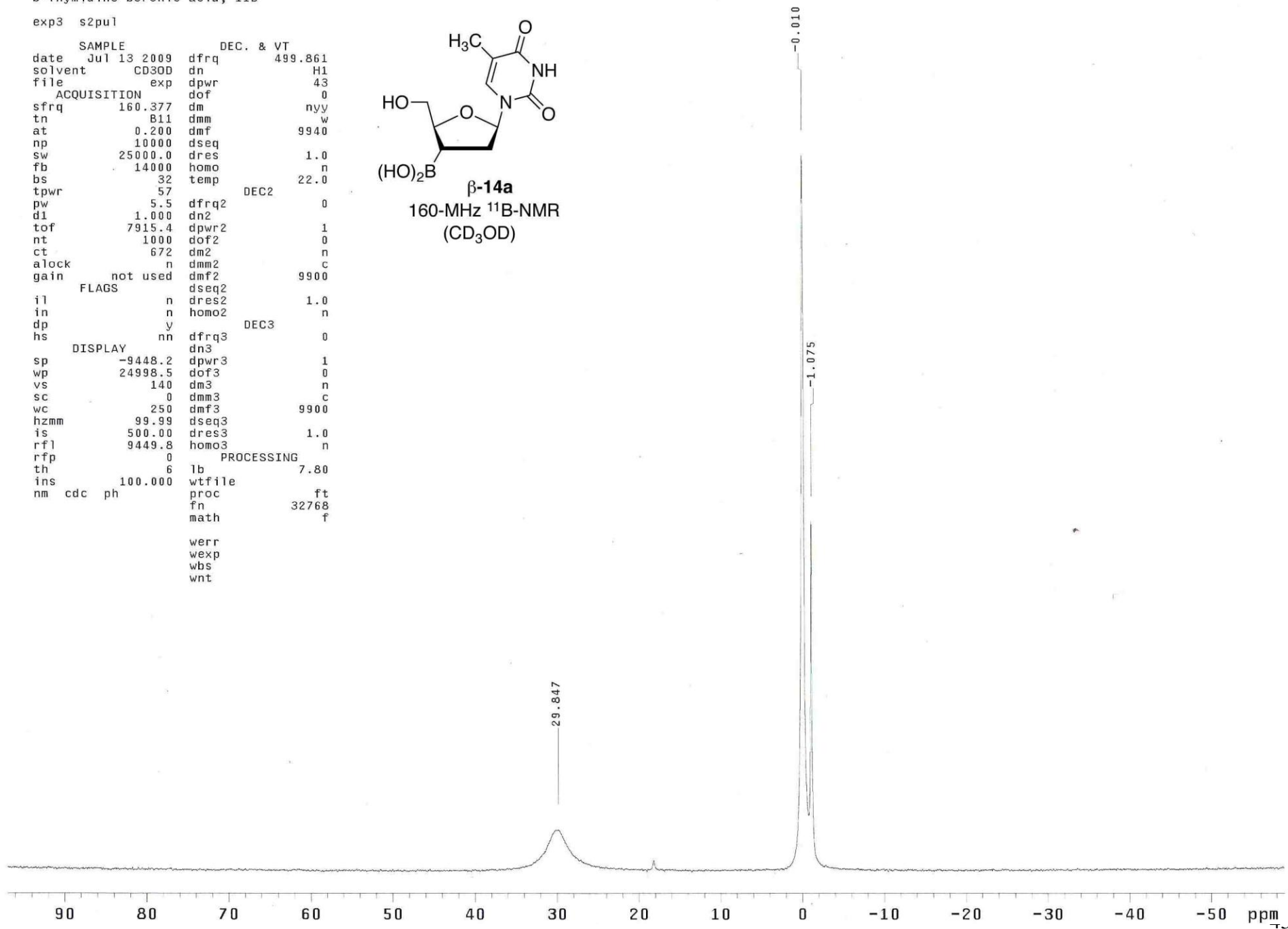
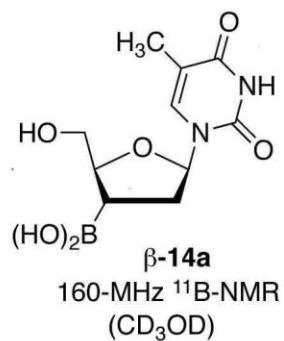
Pulse Sequence: s2pu1



b-Thymidine boronic acid, 11B

exp3 s2pul

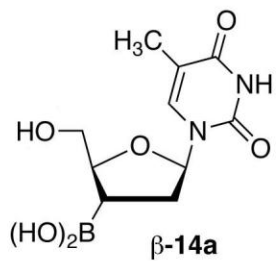
SAMPLE		DEC. & VT	
date	Jul 13 2009	dfrq	499.861
solvent	CD3OD	dn	H1
file	exp	dpwr	43
ACQUISITION		dof	0
sfrq	160.377	dm	nyy
tn	811	dmm	w
at	0.200	dmf	9940
np	10000	dseq	
sw	25000.0	dres	1.0
fb	14000	homo	n
bs	32	temp	22.0
tpwr	57		DEC2
pw	5.5	dfrq2	0
d1	1.000	dn2	
tof	7915.4	dpwr2	1
nt	1000	dof2	0
ct	672	dm2	n
alock	n	dmm2	c
gain	not used	dmf2	9900
FLAGS		dseq2	
il	n	dres2	1.0
in	n	homo2	n
dp	y		DEC3
hs	nn	dfrq3	0
DISPLAY		dn3	
sp	-9448.2	dpwr3	1
wp	24998.5	dof3	0
vs	140	dm3	n
sc	0	dmm3	c
wc	250	dmf3	9900
hzmm	99.99	dseq3	
is	500.00	dres3	1.0
rfl	9449.8	homo3	n
rtp	0		PROCESSING
th	6	lb	7.80
ins	100.000	wtfile	
nm	cdc ph	proc	ft
		fn	32768
		math	f
		werr	
		wexp	
		wbs	
		wnt	



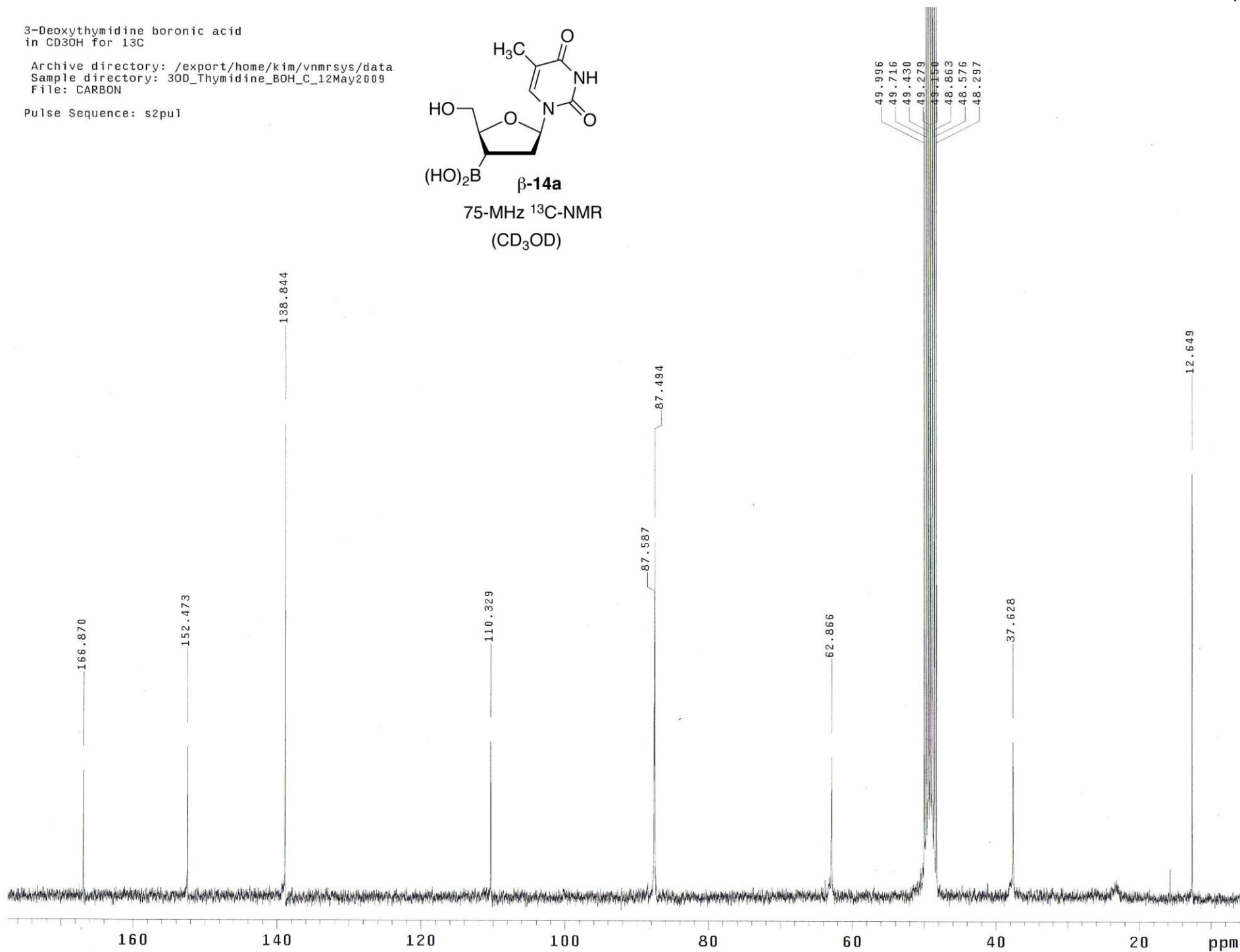
3-Deoxythymidine boronic acid  
in CD<sub>3</sub>OH for <sup>13</sup>C

Archive directory: /export/home/kim/vnmrsys/data  
Sample directory: 300\_Thymidine\_BOH\_C\_12May2009  
File: CARBON

Pulse Sequence: s2pu1



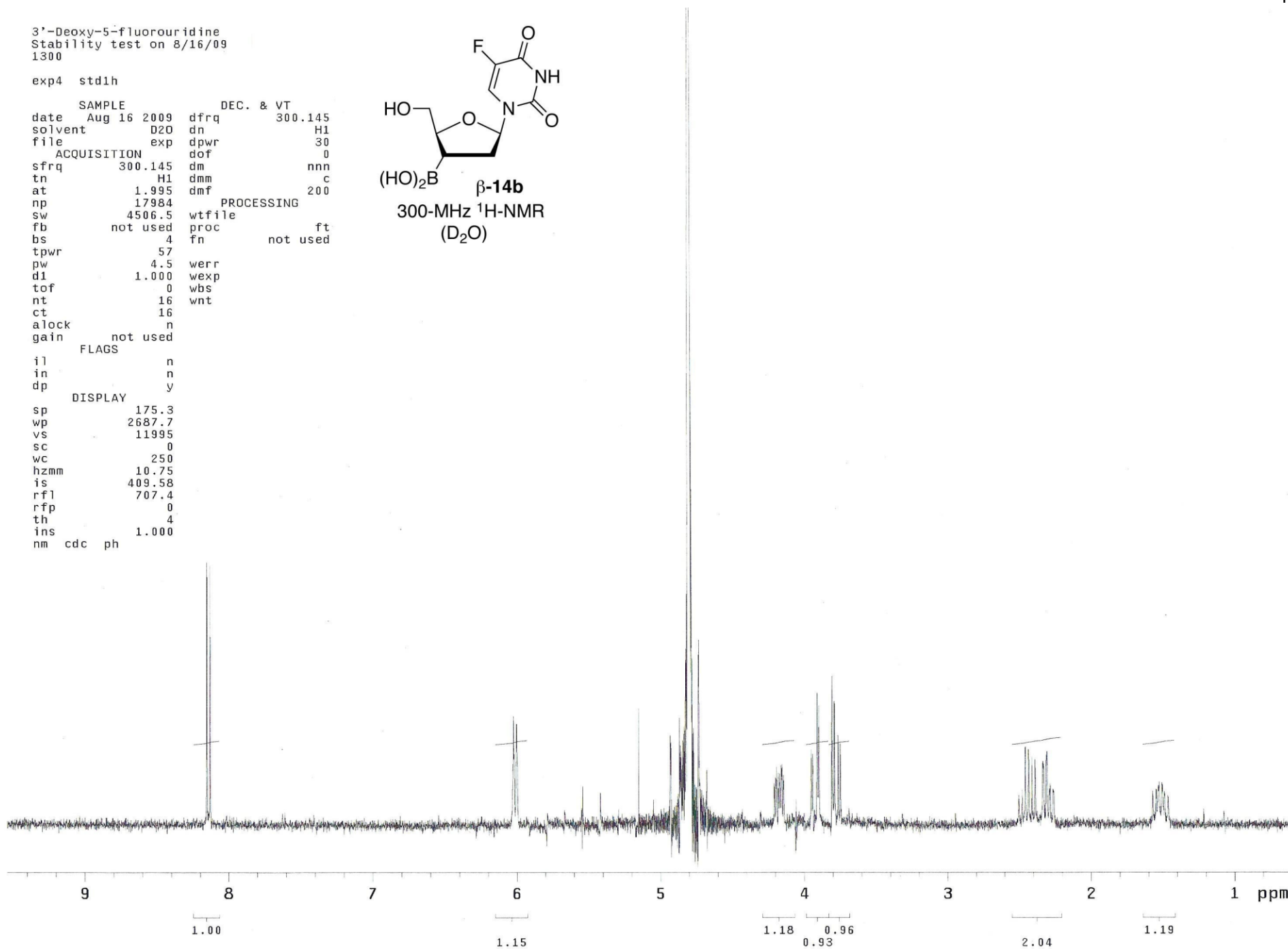
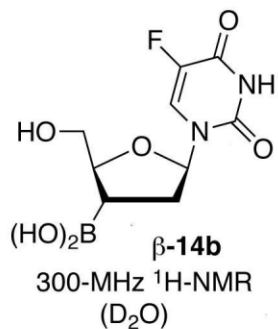
75-MHz <sup>13</sup>C-NMR  
(CD<sub>3</sub>OD)



3'-Deoxy-5-fluorouridine  
Stability test on 8/16/09  
1300

exp4 std1h

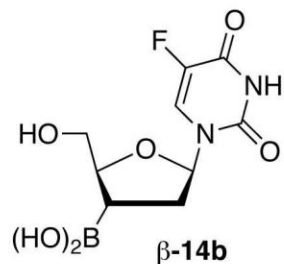
SAMPLE		DEC. & VT	
date	Aug 16 2009	dfrq	300.145
solvent	D2O	dn	H1
file	exp	dpwr	30
ACQUISITION		dof	0
sfrq	300.145	dm	nnn
tn	H1	dmm	c
at	1.995	dmf	200
np	17984	PROCESSING	
sw	4506.5	wtfile	
fb	not used	proc	ft
bs	4	fn	not used
tpwr	57		
pw	4.5	werr	
d1	1.000	wexp	
tof	0	wbs	
nt	16	wnt	
ct	16		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	y		
DISPLAY			
sp	175.3		
wp	2687.7		
vs	11995		
sc	0		
wc	250		
hzmm	10.75		
is	409.58		
rfl	707.4		
rfp	0		
th	4		
ins	1.000		
nm	cdc ph		



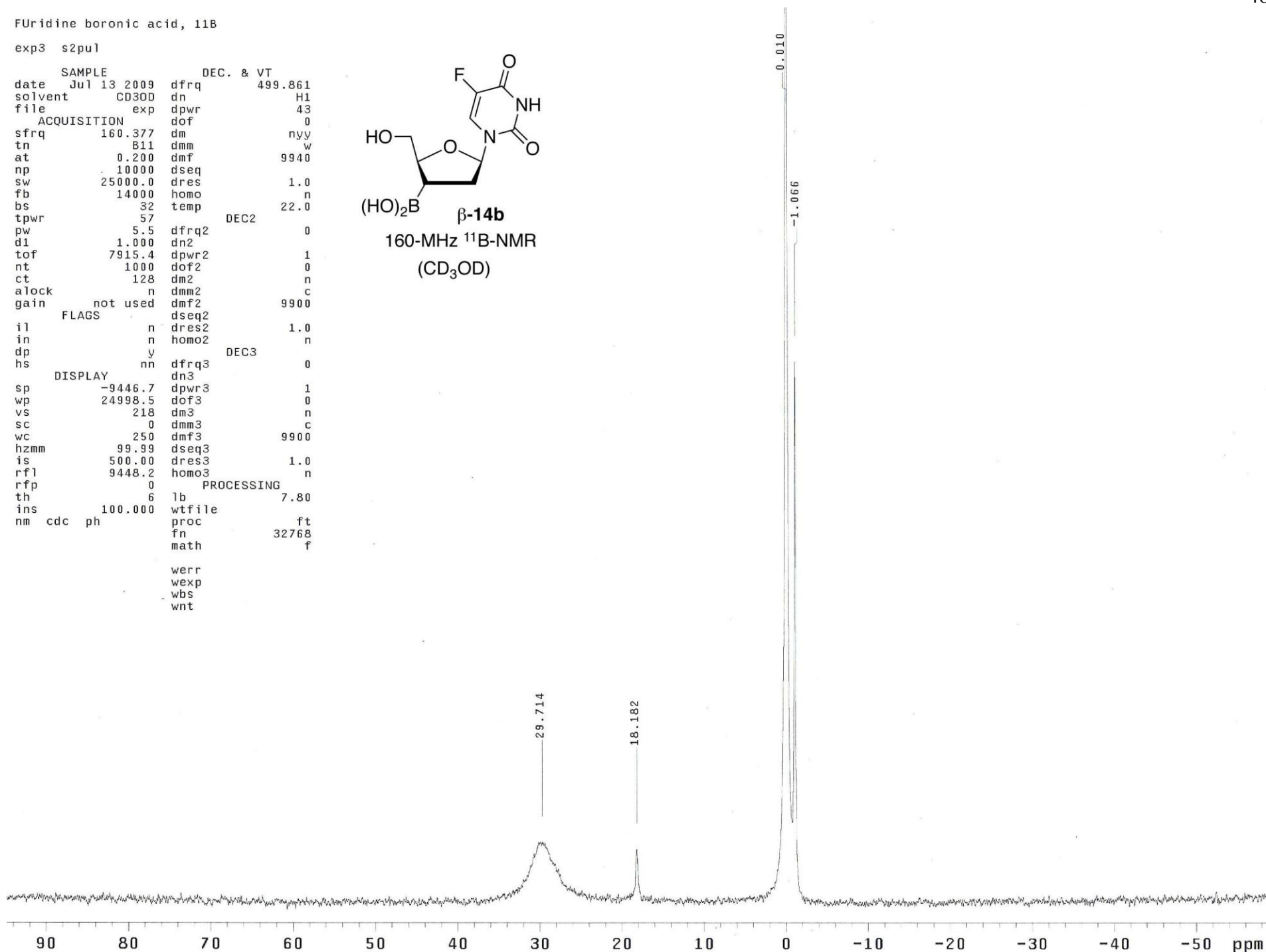
Furidine boronic acid, 11B

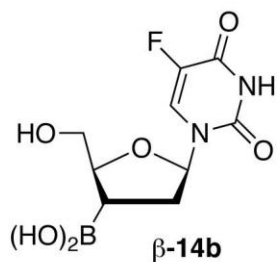
exp3 s2pu1

SAMPLE		DEC. & VT	
date	Jul 13 2009	dfrq	499.861
solvent	CD3OD	dn	H1
file	exp	dpwr	43
ACQUISITION			
sfrq	160.377	dm	nyy
tn	811	dmm	w
at	0.200	dmf	9940
np	10000	dseq	
sw	25000.0	dres	1.0
fb	14000	homo	n
bs	32	temp	22.0
tpwr	57		
pw	5.5	dfrq2	0
d1	1.000	dn2	
tof	7915.4	dpwr2	1
nt	1000	dof2	0
ct	128	dm2	n
alock	n	dmm2	c
gain	not used	dmf2	9900
FLAGS			
il	n	dseq2	
in	n	dres2	1.0
dp	y	homo2	n
hs	nn		
DISPLAY		DEC3	
sp	-9446.7	dfrq3	0
wp	24998.5	dn3	
vs	218	dpwr3	1
sc	0	dof3	0
wc	250	dm3	n
hzmm	99.99	dmm3	c
is	500.00	dmf3	9900
rfl	9448.2	dseq3	
rfl	0	dres3	1.0
th	6	homo3	n
ins	100.000	PROCESSING	
nm	cdc ph	lb	7.80
		wfile	
		proc	ft
		fn	32768
		math	f
		werr	
		wexp	
		wbs	
		wnt	

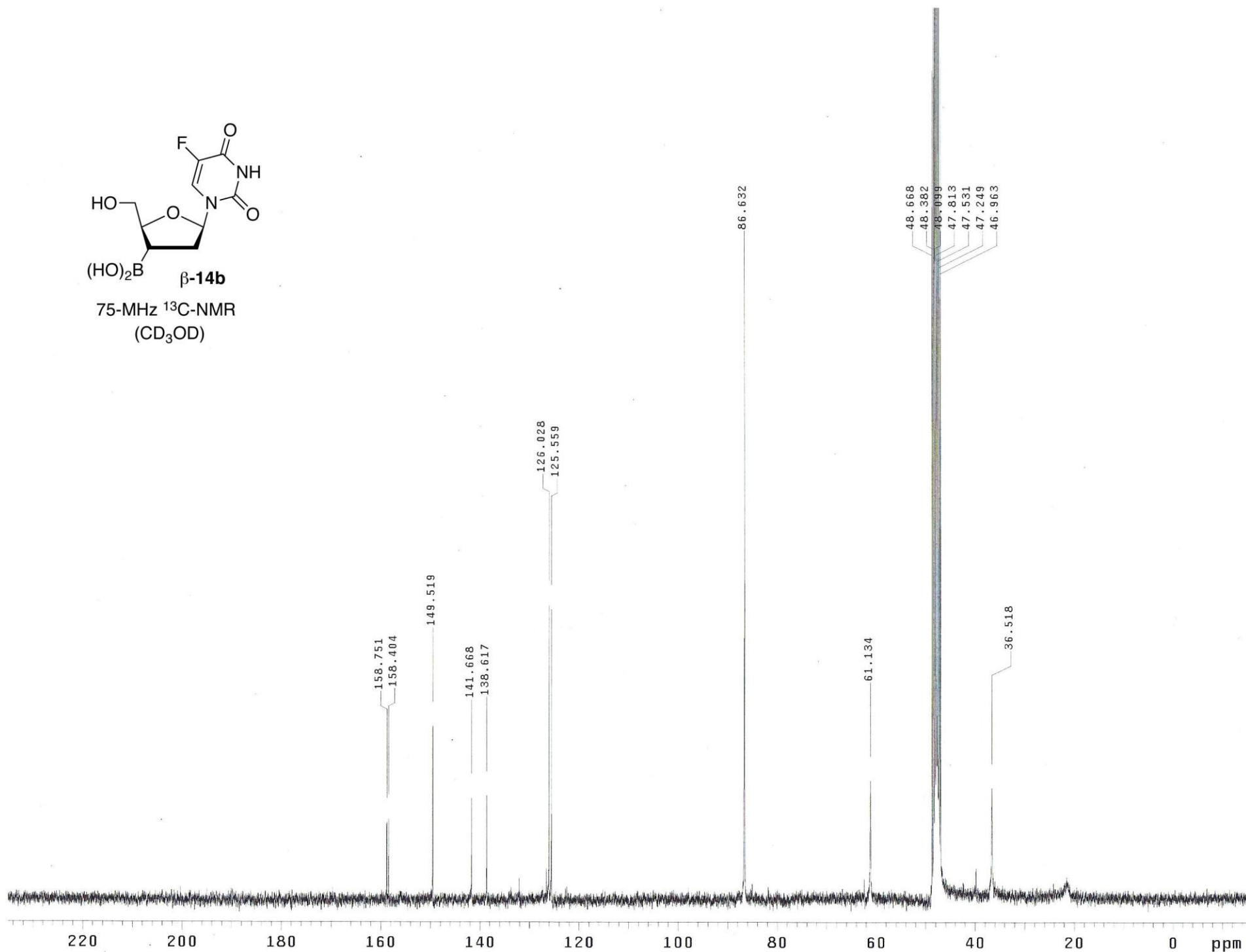


160-MHz <sup>11</sup>B-NMR  
(CD<sub>3</sub>OD)





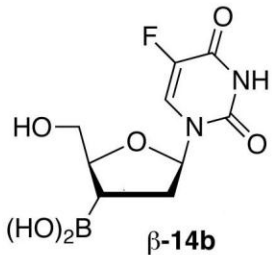
75-MHz <sup>13</sup>C-NMR  
(CD<sub>3</sub>OD)



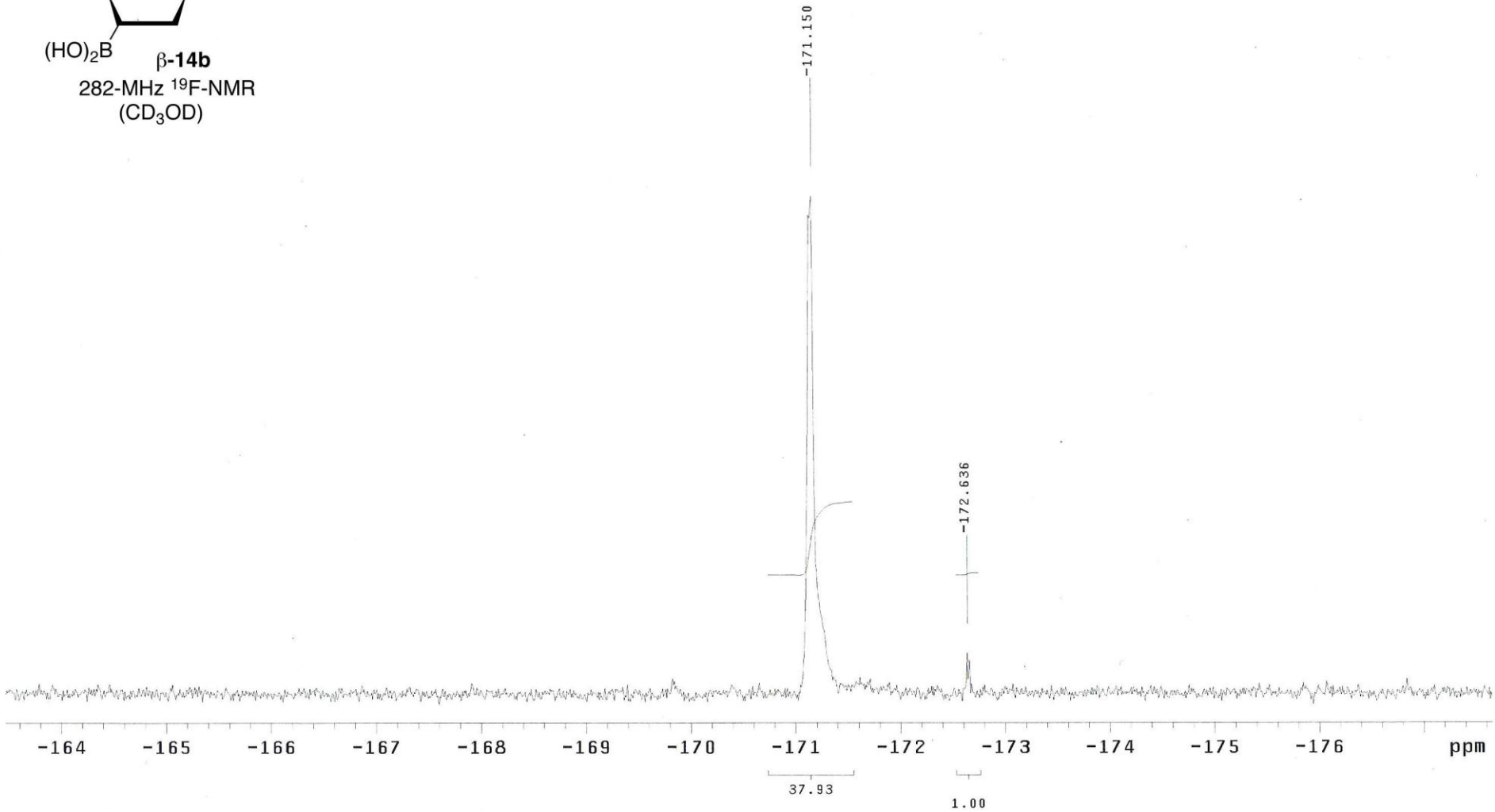
FUridine\_BOH\_F

Archive directory: /export/home/kim/vnmrsys/data  
Sample directory: FUridine\_BOHF\_07Jul2009

Pulse Sequence: s2pu1



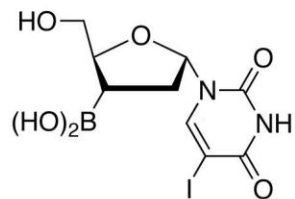
282-MHz <sup>19</sup>F-NMR  
(CD<sub>3</sub>OD)



3'-deoxyiodouridine boronic acid  
a anomer

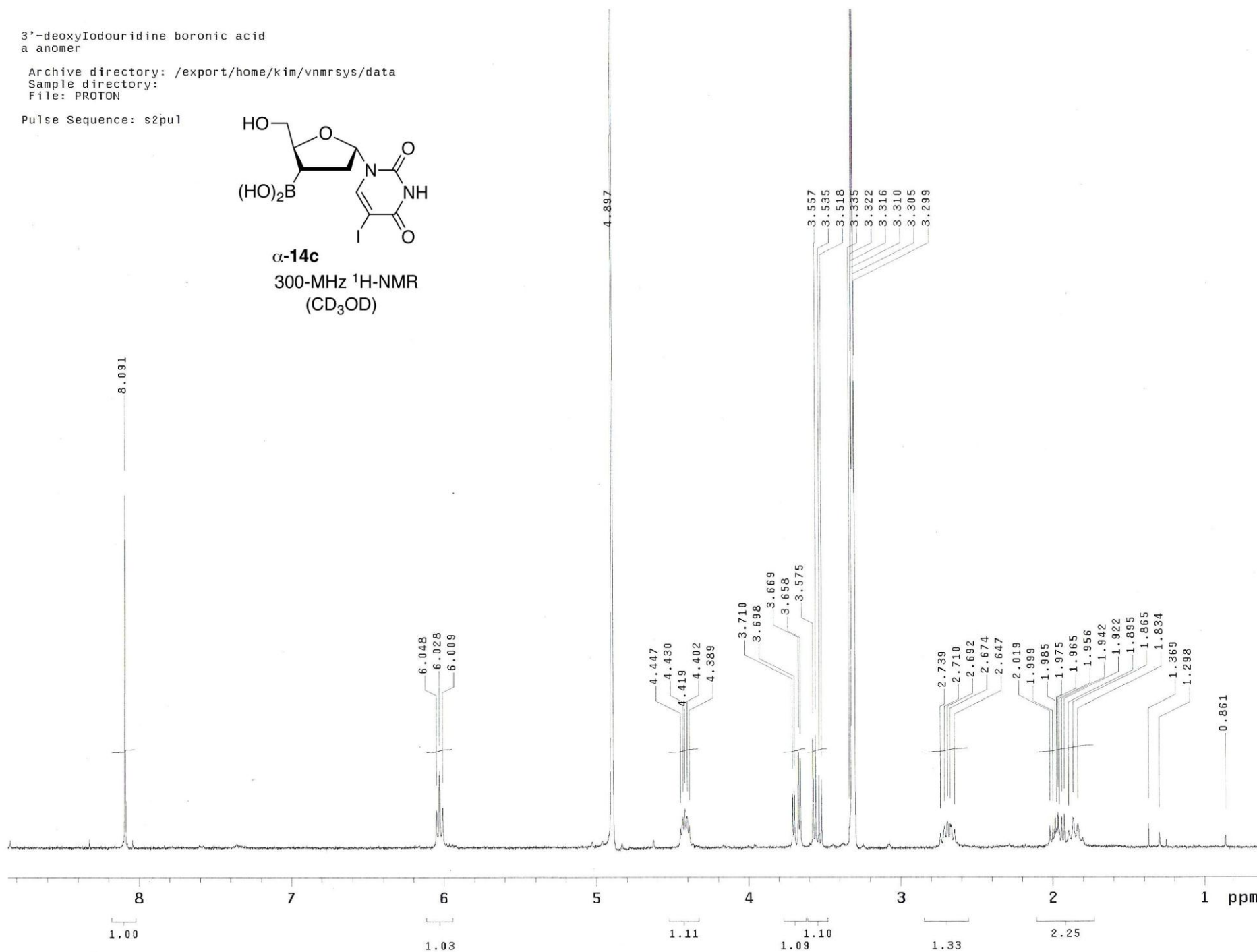
Archive directory: /export/home/kim/vnmr/sys/data  
Sample directory:  
File: PROTON

Pulse Sequence: s2pu1



**$\alpha$ -14c**

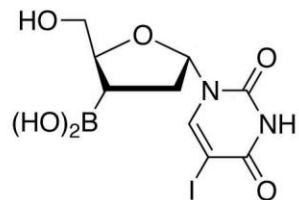
300-MHz  $^1\text{H}$ -NMR  
( $\text{CD}_3\text{OD}$ )



IUridine\_aBOH\_C

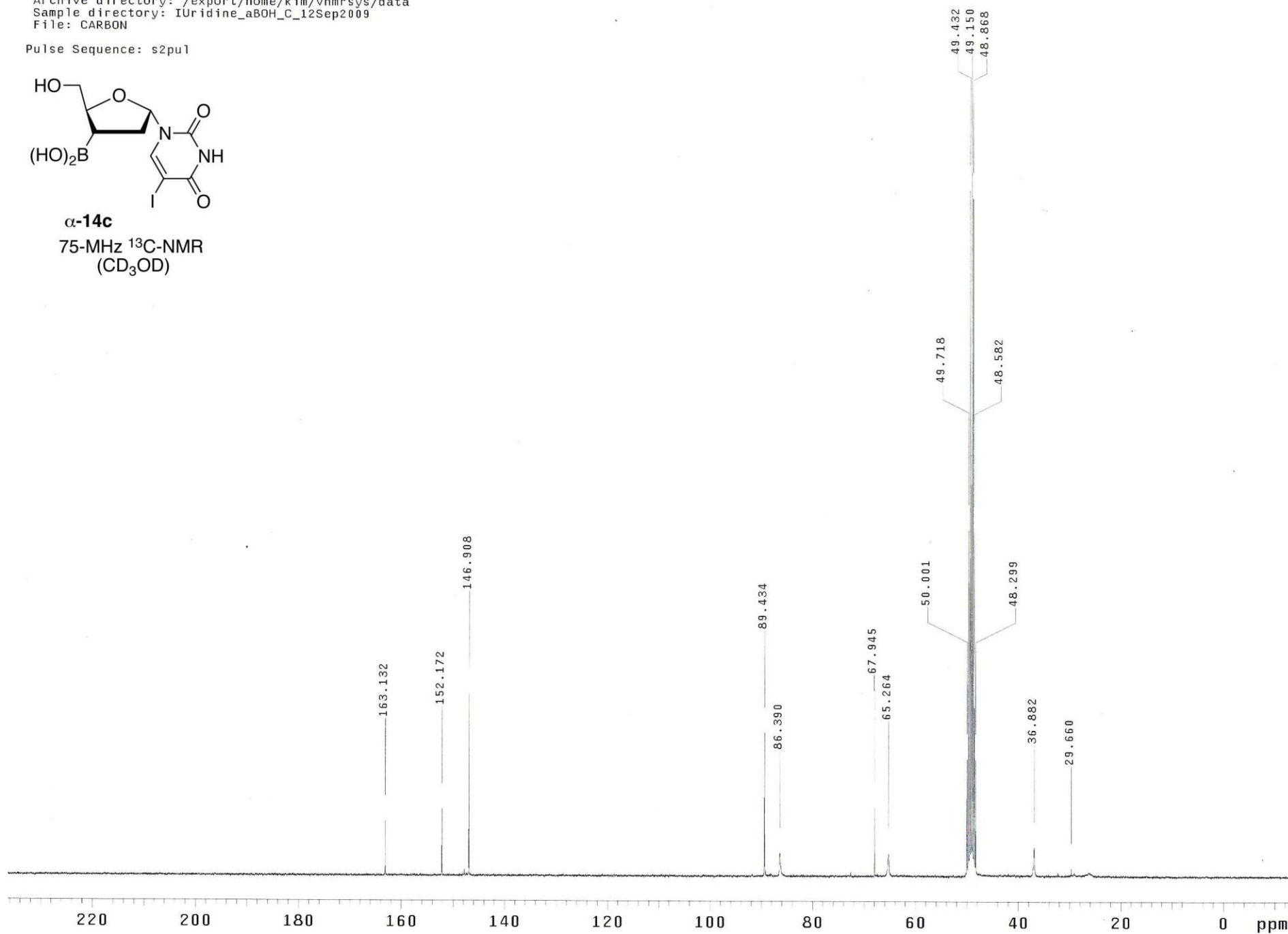
Archive directory: /export/home/kim/vnmrsys/data  
Sample directory: IUridine\_aBOH\_C\_12Sep2009  
File: CARBON

Pulse Sequence: s2pu1



**α-14c**

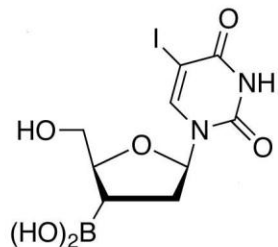
75-MHz <sup>13</sup>C-NMR  
(CD<sub>3</sub>OD)



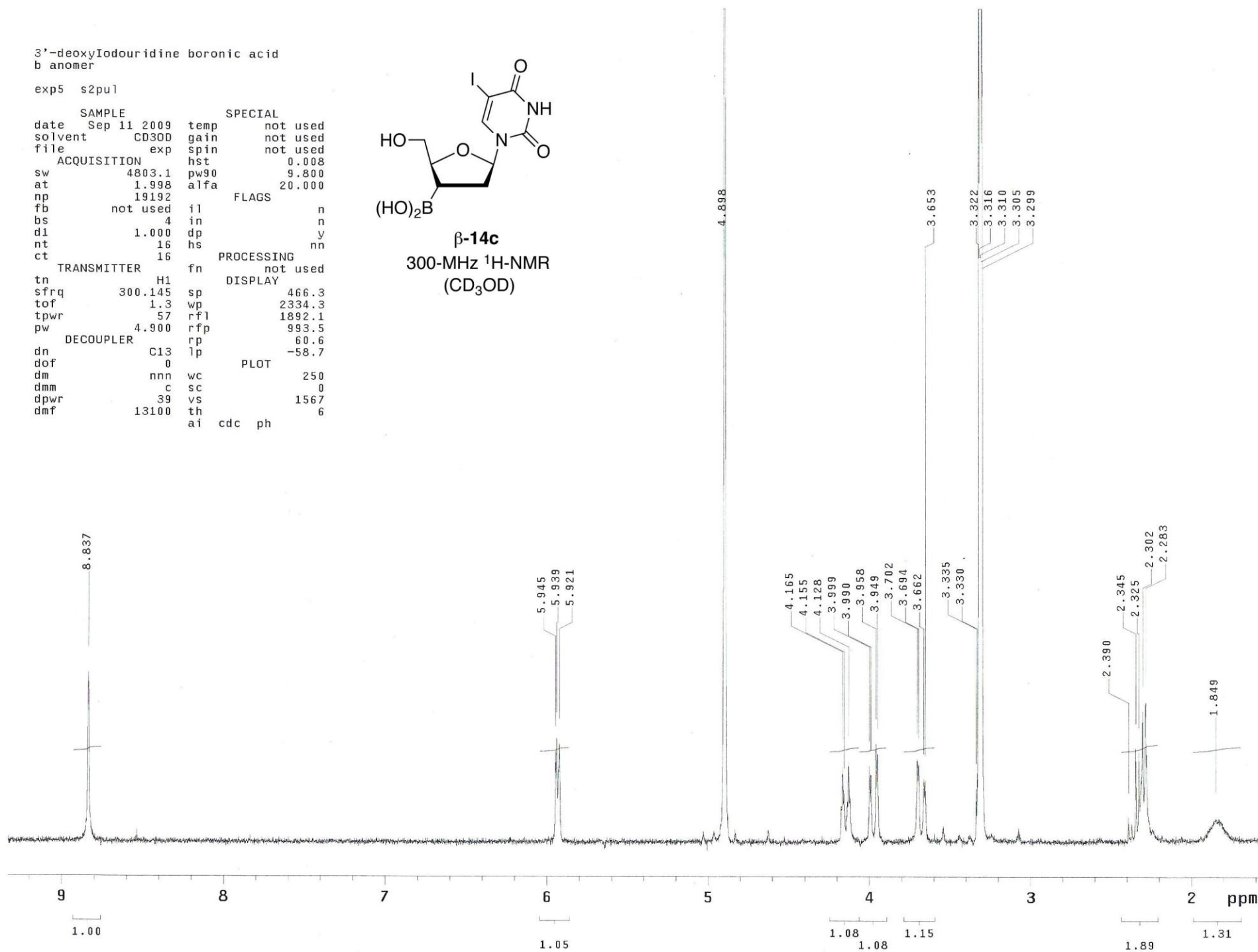
3'-deoxyIodouridine boronic acid  
b anomer

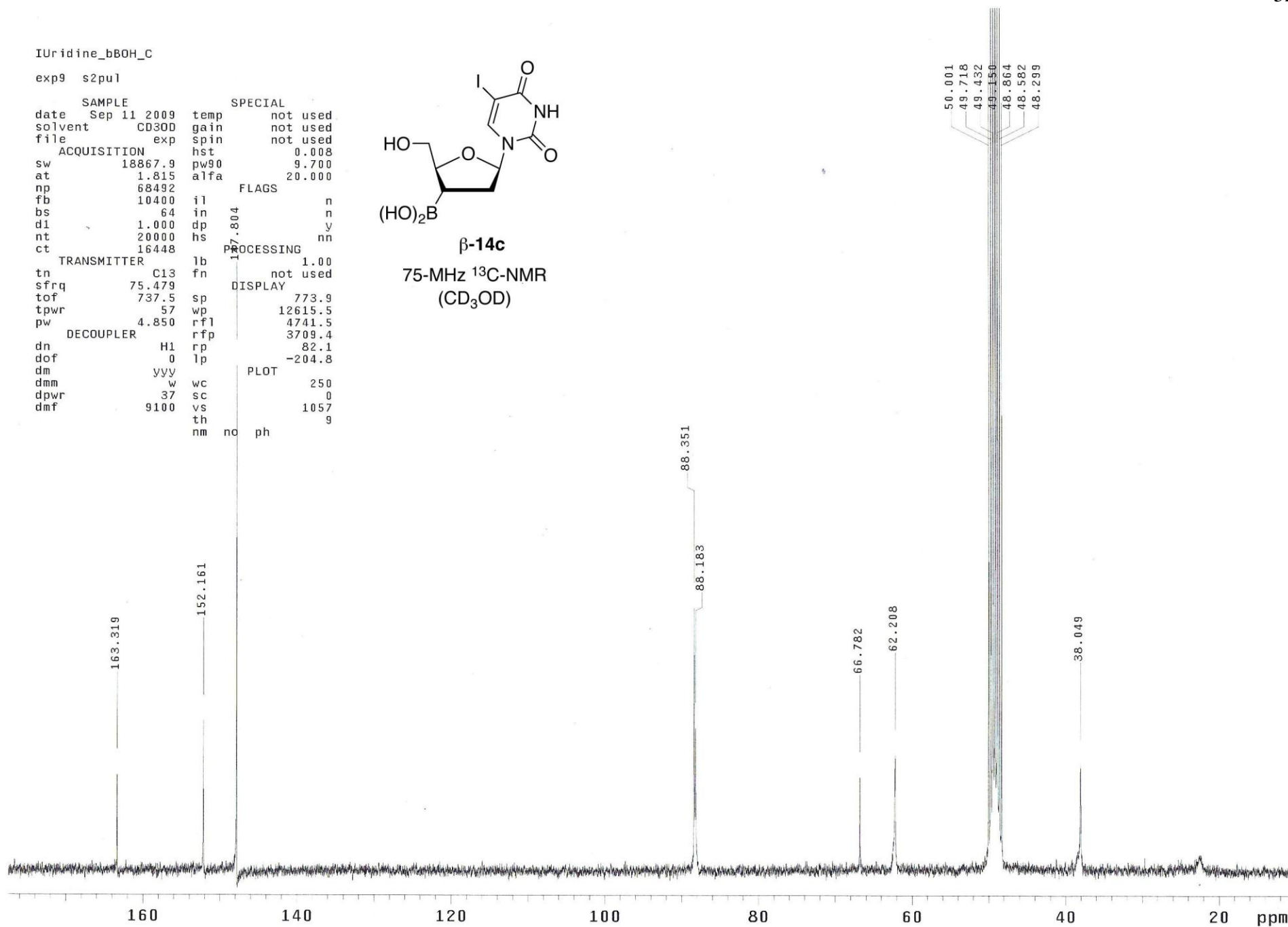
exp5 s2pu1

SAMPLE		SPECIAL	
date	Sep 11 2009	temp	not used
solvent	CD3OD	gain	not used
file	exp	spin	not used
ACQUISITION		hst	0.008
sw	4803.1	pw90	9.800
at	1.998	alfa	20.000
np	19192	FLAGS	
fb	not used	il	n
bs	4	in	n
d1	1.000	dp	y
nt	16	hs	nn
ct	16	PROCESSING	
TRANSMITTER		fn	not used
tn	H1	DISPLAY	
sfrq	300.145	sp	466.3
tof	1.3	wp	2334.3
tpwr	57	rfl	1892.1
pw	4.900	rfp	993.5
DECOUPLER		rp	60.6
dn	C13	lp	-58.7
dof	0	PLOT	
dm	nnn	wc	250
dmm	c	sc	0
dpwr	39	vs	1567
dmf	13100	th	6
	ai	cdc	ph



**β-14c**  
300-MHz <sup>1</sup>H-NMR  
(CD<sub>3</sub>OD)

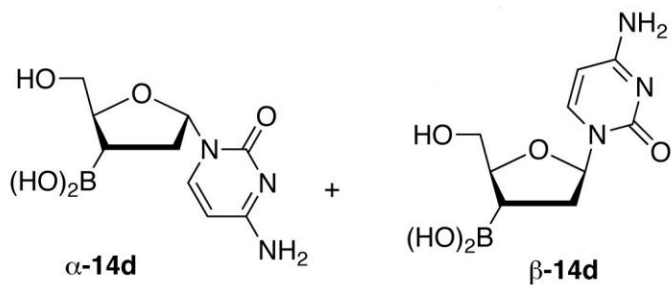




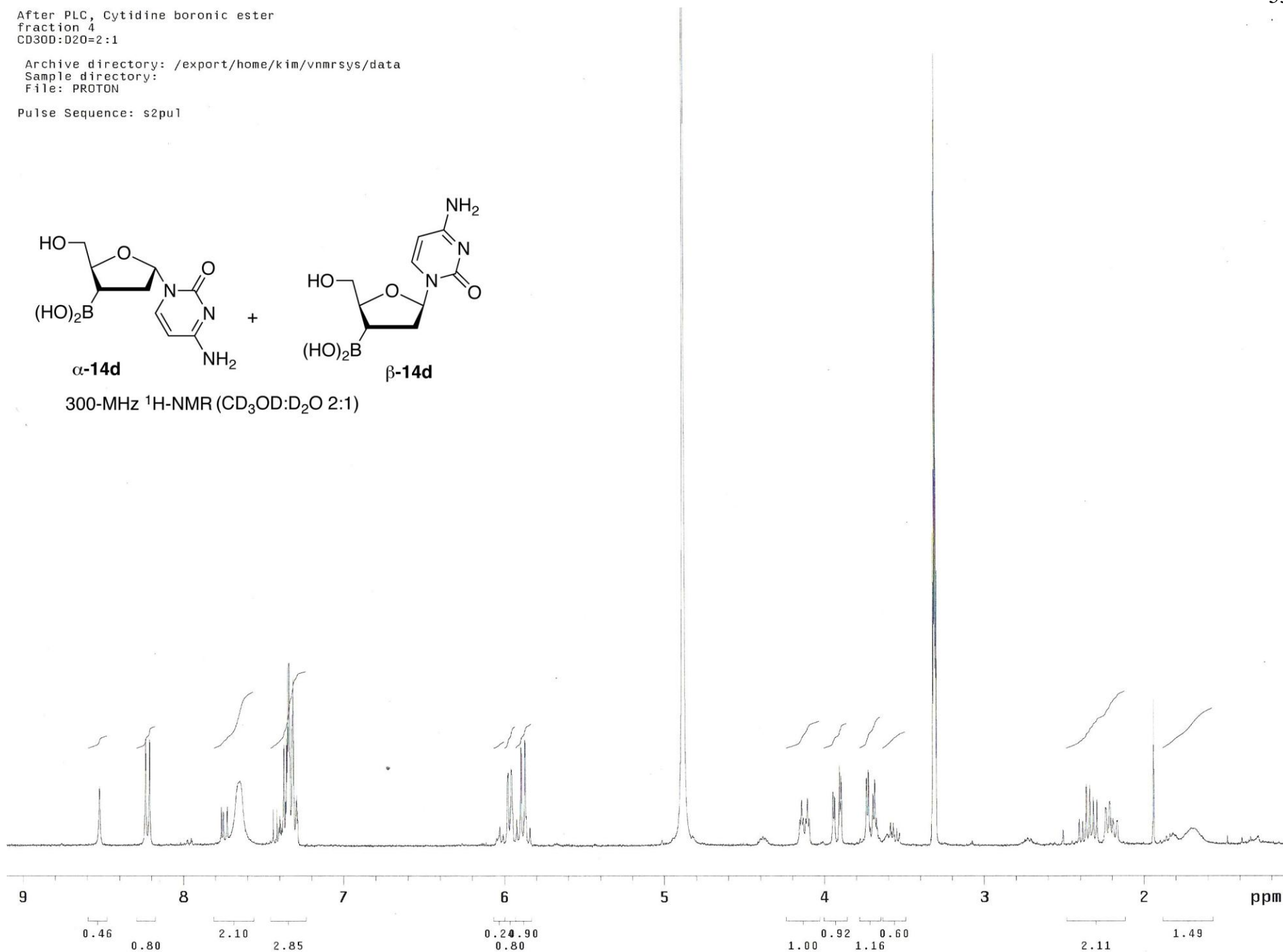
After PLC, Cytidine boronic ester  
fraction 4  
CD3OD:D2O=2:1

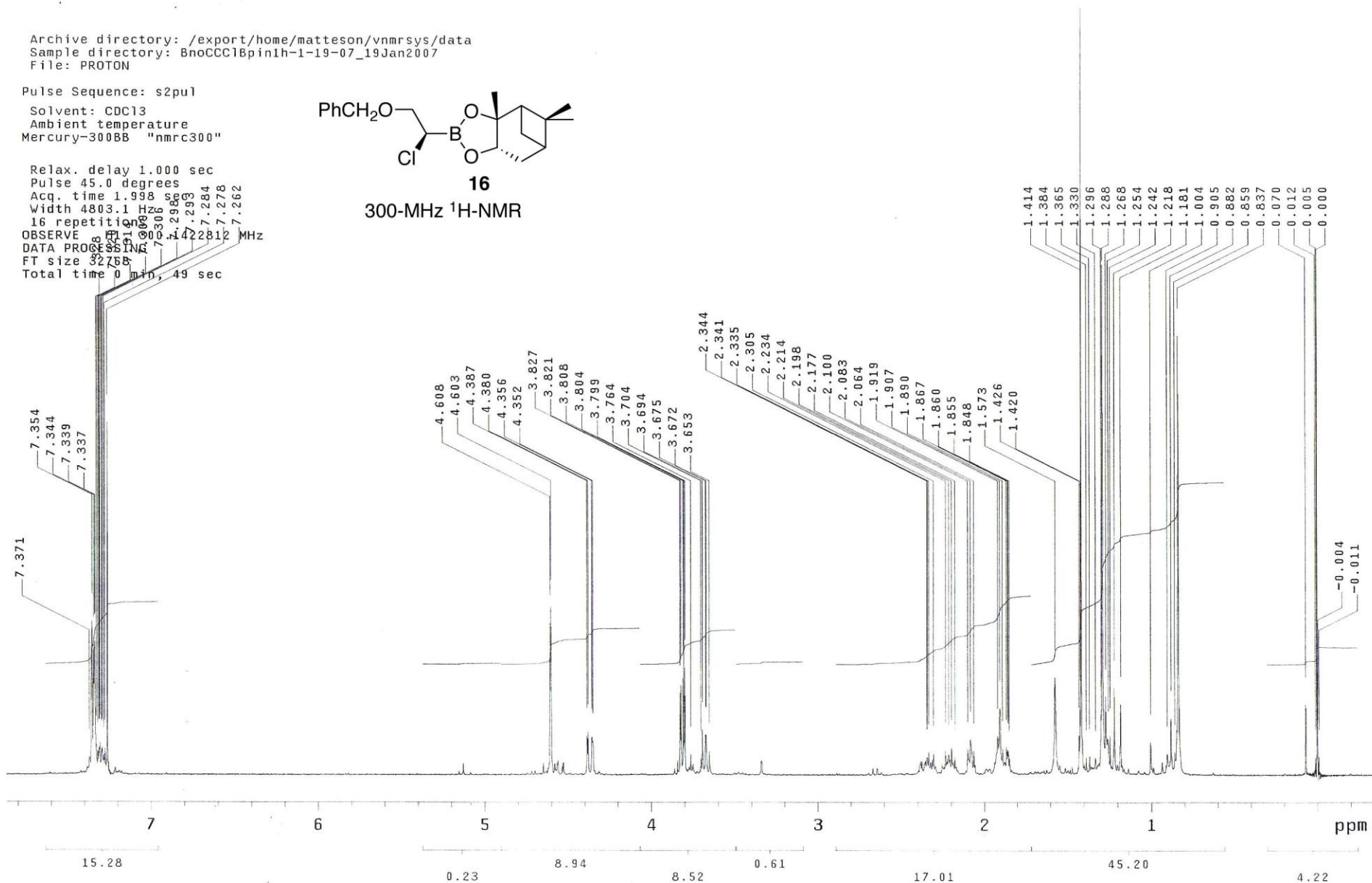
Archive directory: /export/home/kim/vnmrsys/data  
Sample directory:  
File: PROTON

Pulse Sequence: s2pu1



300-MHz  $^1\text{H}$ -NMR ( $\text{CD}_3\text{OD}:\text{D}_2\text{O}$  2:1)



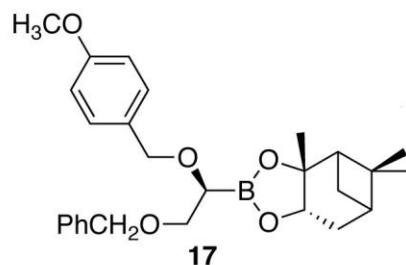


Archive directory: /export/home/matteson/vnmrsys/data  
Sample directory: BnOCH2CHOBnBpin1H\_26Jan2007  
File: PROTON

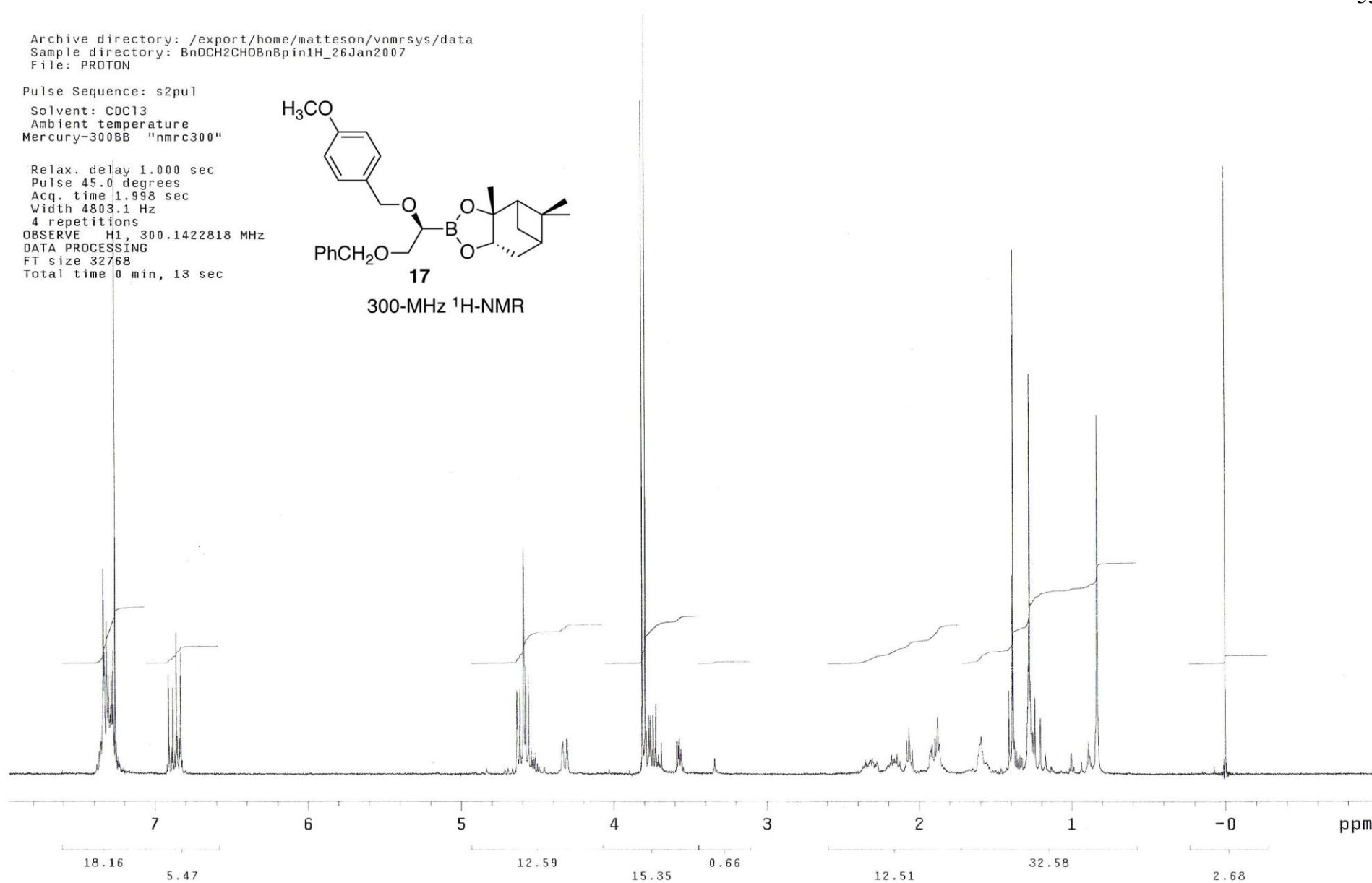
Pulse Sequence: s2pu1

Solvent: CDCl3  
Ambient temperature  
Mercury-300BB "nmrc300"

Relax. delay 1.000 sec  
Pulse 45.0 degrees  
Acq. time 1.998 sec  
Width 4803.1 Hz  
4 repetitions  
OBSERVE H1, 300.1422818 MHz  
DATA PROCESSING  
FT size 32768  
Total time 0 min, 13 sec



300-MHz <sup>1</sup>H-NMR

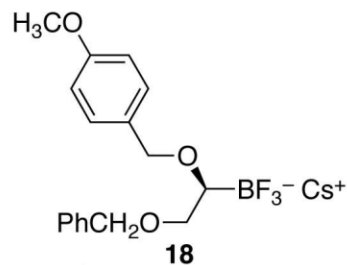


Archive directory: /export/home/matteson/vnmrsys/data  
Sample directory: BnOCCOPMBBF3Cs-1H\_27Feb2007-14:43:09  
File: PROTON

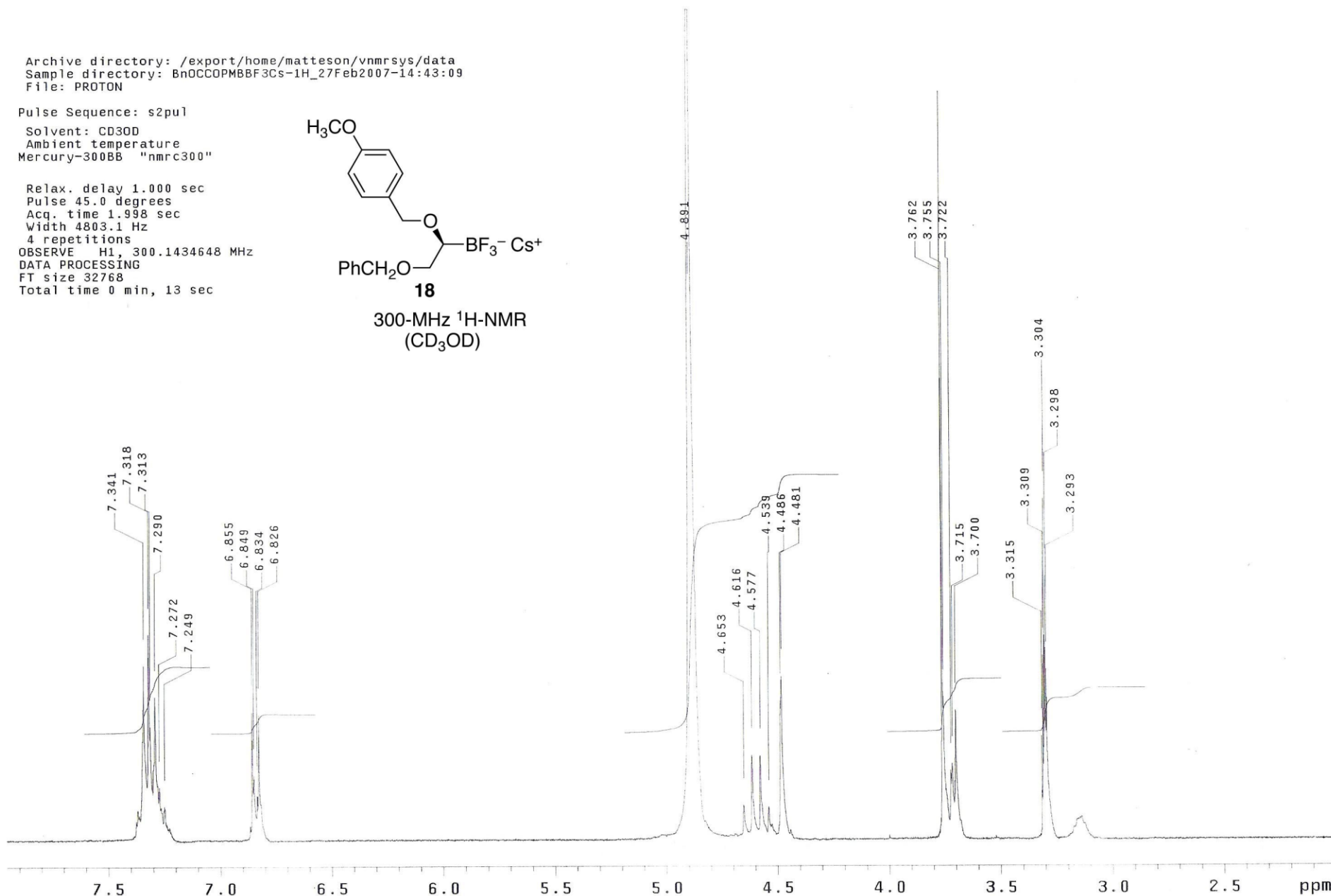
Pulse Sequence: s2pu1

Solvent: CD3OD  
Ambient temperature  
Mercury-300BB "nmrc300"

Relax. delay 1.000 sec  
Pulse 45.0 degrees  
Acq. time 1.998 sec  
Width 4803.1 Hz  
4 repetitions  
OBSERVE H1, 300.1434648 MHz  
DATA PROCESSING  
FT size 32768  
Total time 0 min, 13 sec



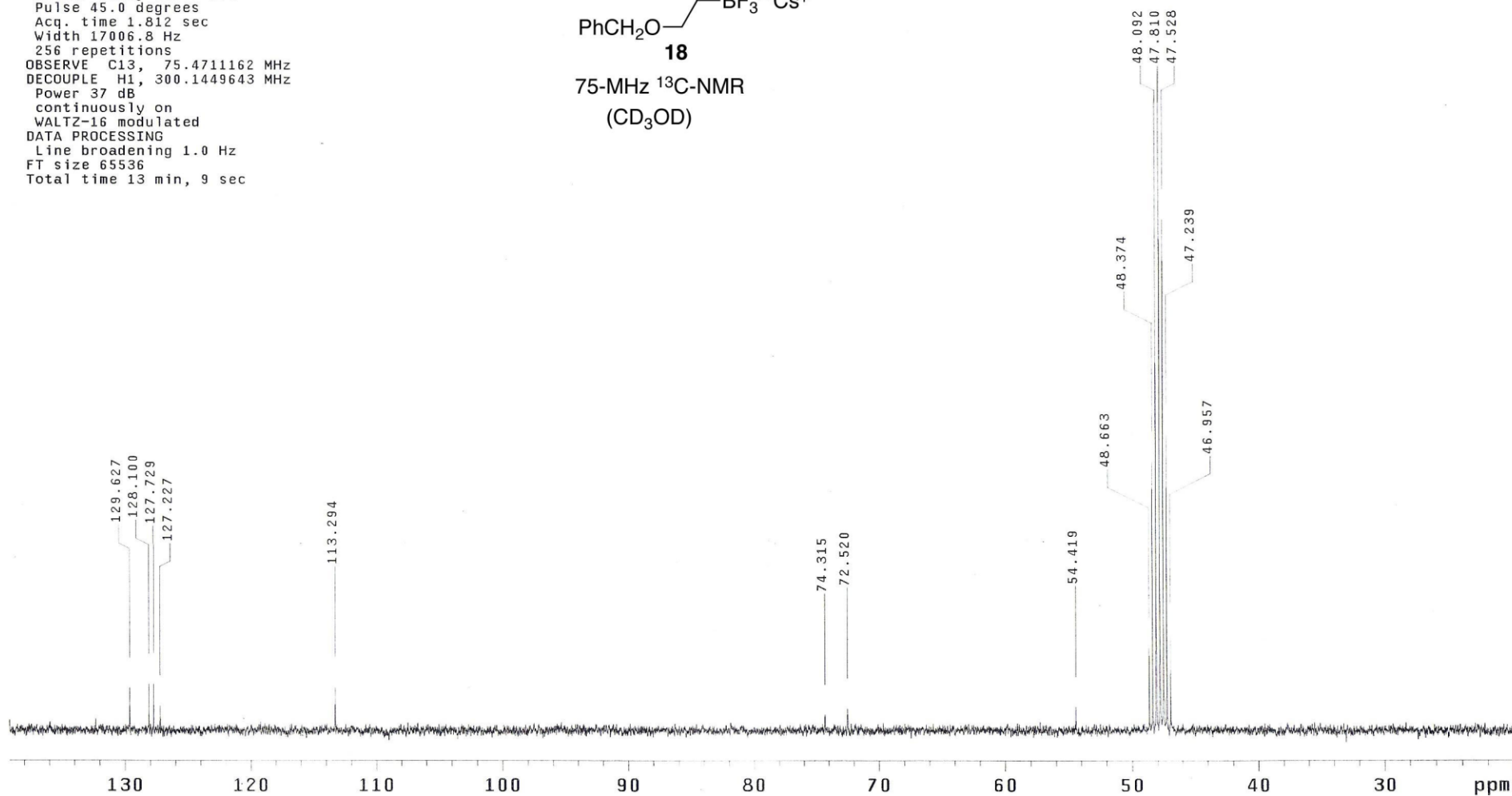
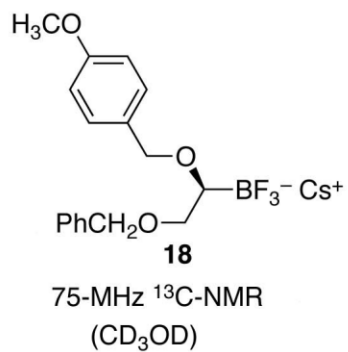
300-MHz <sup>1</sup>H-NMR  
(CD<sub>3</sub>OD)



Archive directory: /export/home/matteson/vnmrsys/data  
Sample directory: DSM3-21-07CD30D13C\_21Mar2007  
File: CARBON

Pulse Sequence: s2pul  
Solvent: CD3OD  
Ambient temperature  
Mercury-300BB "nmrc300"

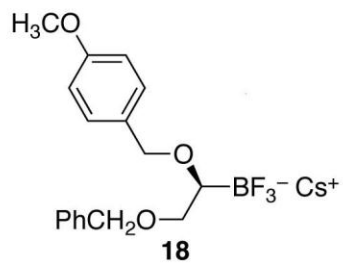
Relax. delay 1.000 sec  
Pulse 45.0 degrees  
Acq. time 1.812 sec  
Width 17006.8 Hz  
256 repetitions  
OBSERVE C13, 75.4711162 MHz  
DECOUPLE H1, 300.1449643 MHz  
Power 37 dB  
continuously on  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
FT size 65536  
Total time 13 min, 9 sec



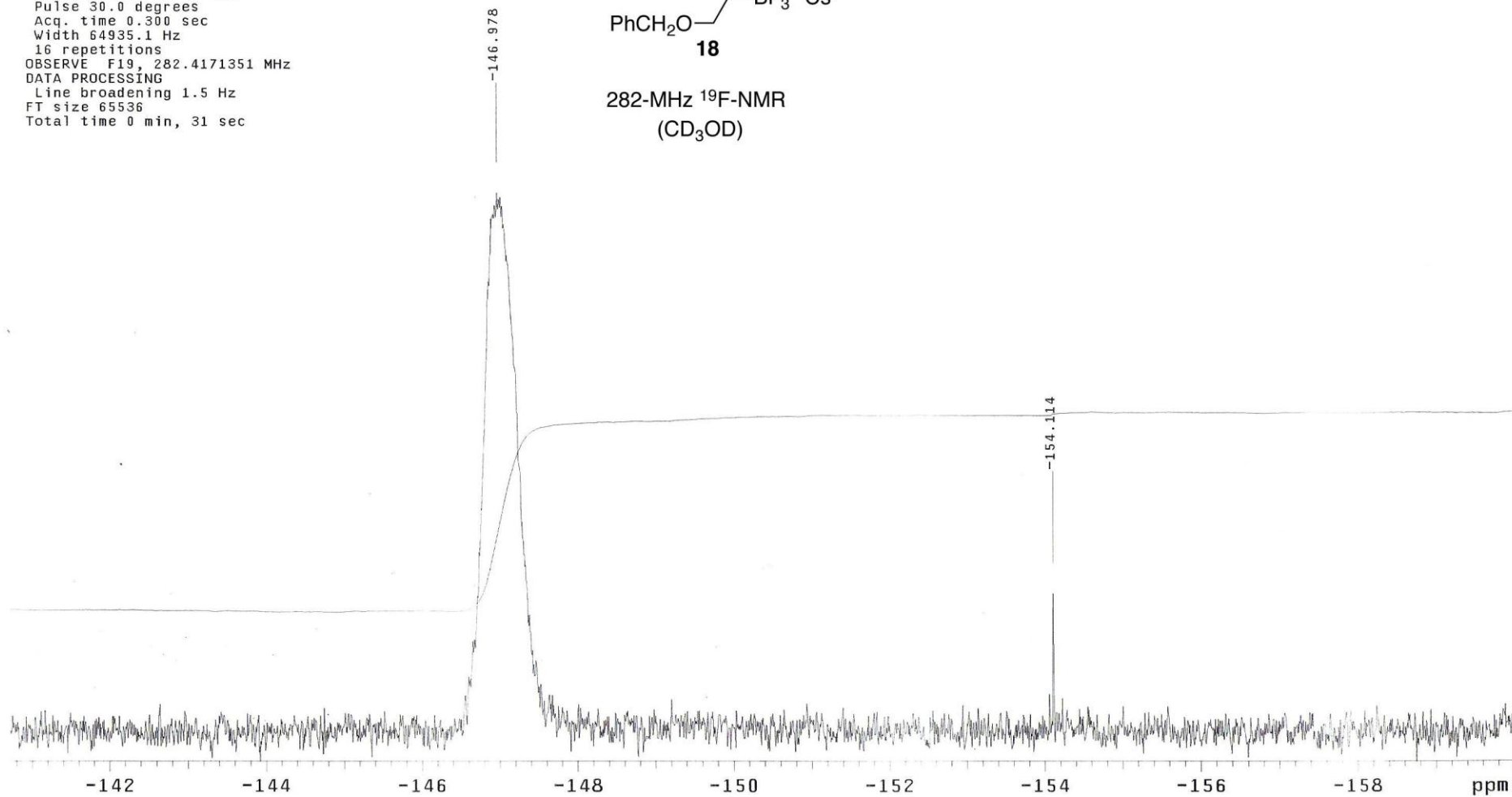
Archive directory: /export/home/matteson/vnmrsys/data  
Sample directory: DSM3-21-07CD30D19F\_21Mar2007  
File: FLUORINE

Pulse Sequence: s2pu1  
Solvent: CD3OD  
Ambient temperature  
Mercury-300BB "nmrc300"

Relax. delay 1.500 sec  
Pulse 30.0 degrees  
Acq. time 0.300 sec  
Width 64935.1 Hz  
16 repetitions  
OBSERVE F19, 282.4171351 MHz  
DATA PROCESSING  
Line broadening 1.5 Hz  
FT size 65536  
Total time 0 min, 31 sec



282-MHz  $^{19}\text{F}$ -NMR  
( $\text{CD}_3\text{OD}$ )



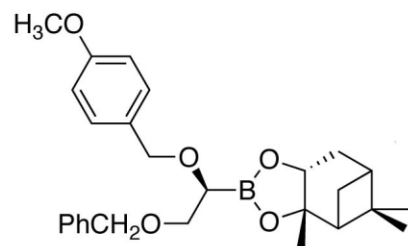
STANDARD 1H OBSERVE

Archive directory: /export/home/matteson/vnmrsys/data  
Sample directory: BnOCC(OPMB)B+PD1H\_03Apr2007  
File: PROTON

Pulse Sequence: s2pu1

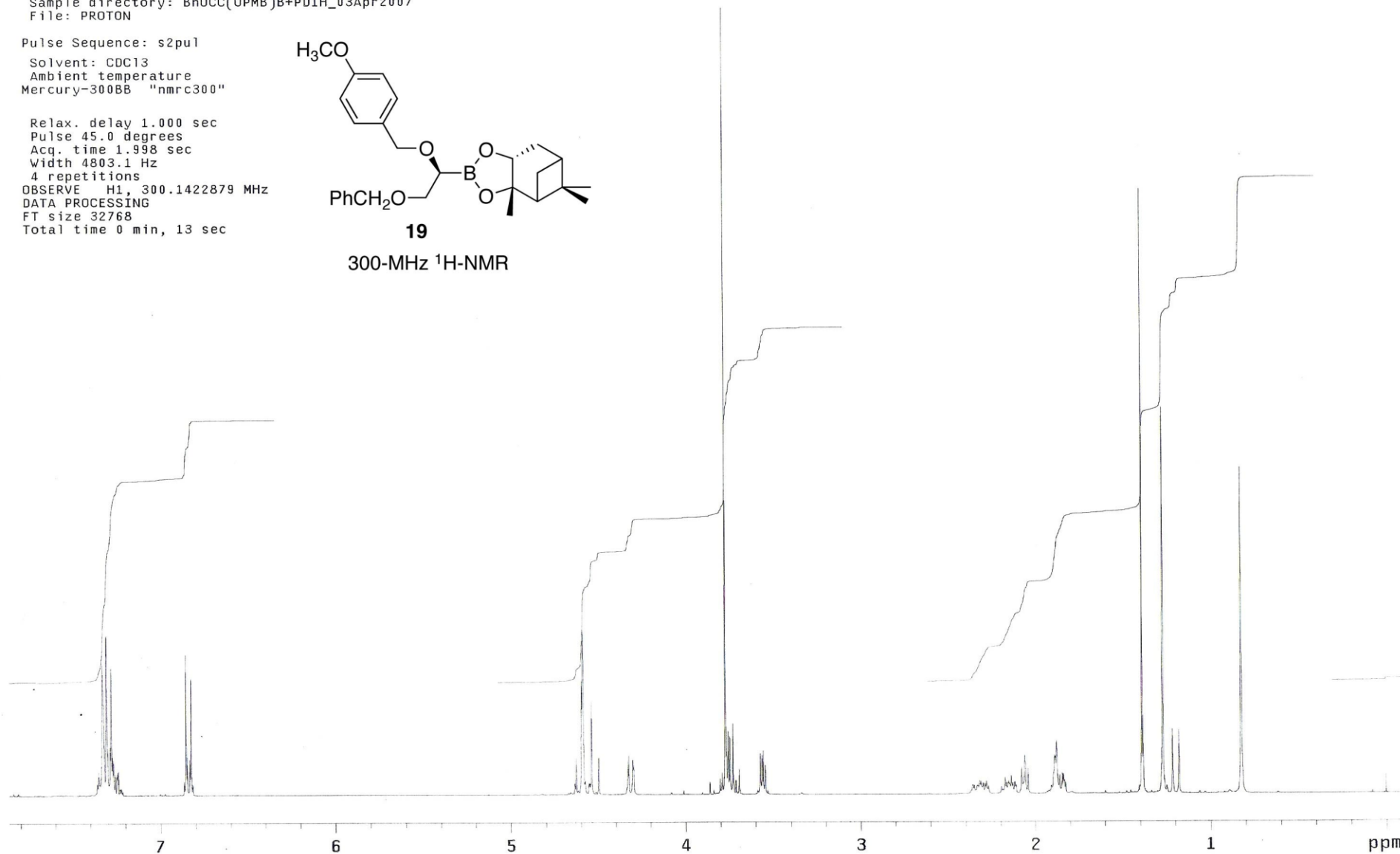
Solvent: CDCl3  
Ambient temperature  
Mercury-300BB "nmrc300"

Relax. delay 1.000 sec  
Pulse 45.0 degrees  
Acq. time 1.998 sec  
Width 4803.1 Hz  
4 repetitions  
OBSERVE H1, 300.1422879 MHz  
DATA PROCESSING  
FT size 32768  
Total time 0 min, 13 sec



**19**

300-MHz <sup>1</sup>H-NMR



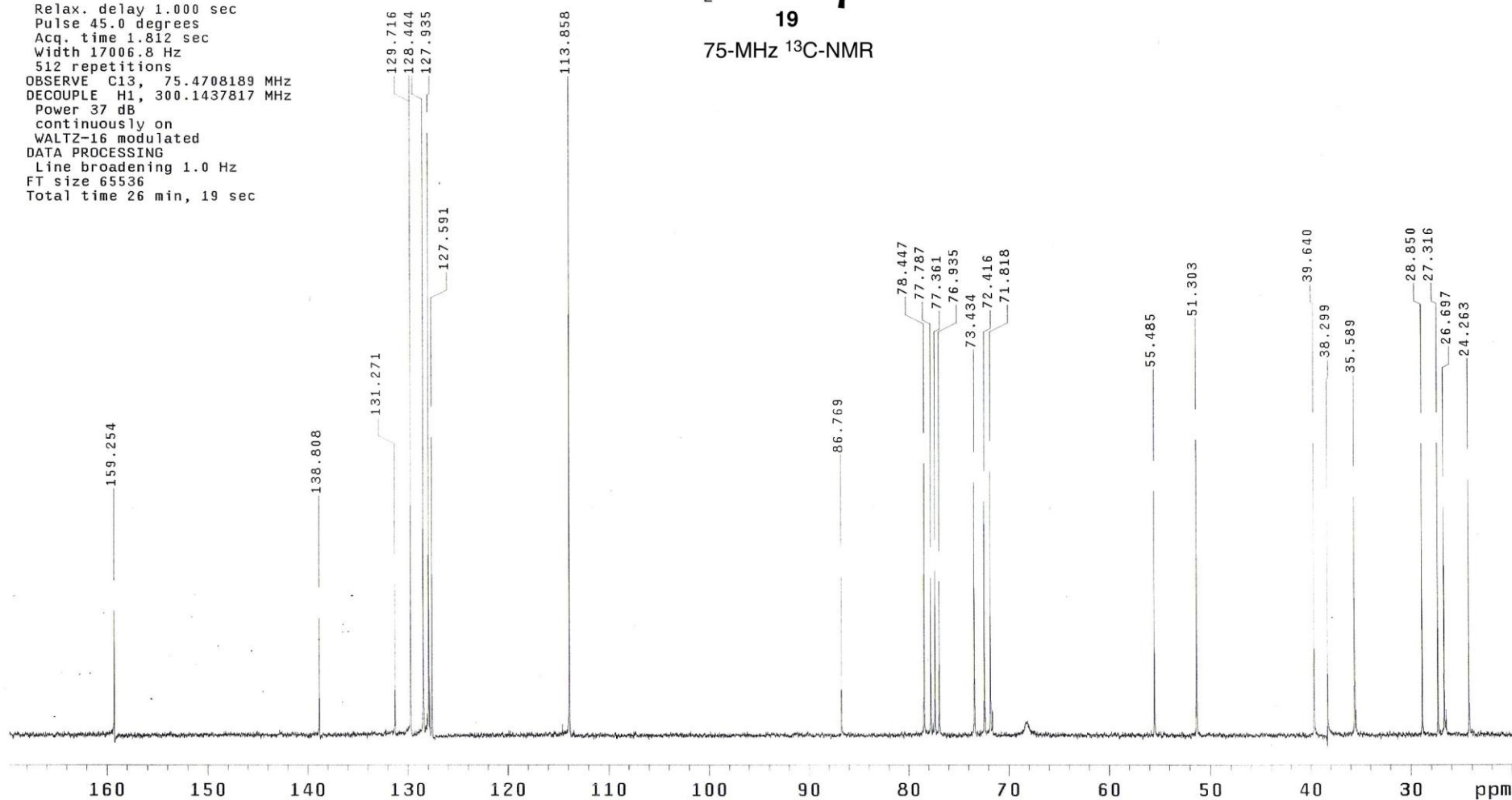
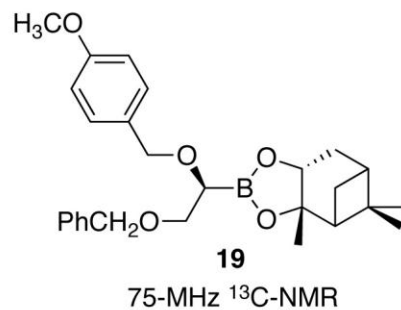
<sup>13</sup>C OBSERVE

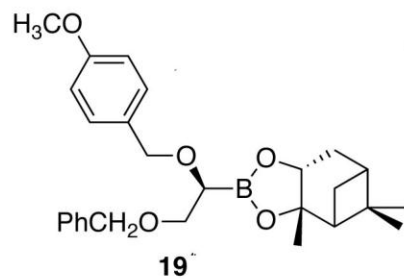
Archive directory: /export/home/matteson/vnmrsys/data  
Sample directory: BnOCC(OPMB)B+PD13C\_03Apr2007  
File: CARBON

Pulse Sequence: s2pu1

Solvent: CDCl<sub>3</sub>  
Ambient temperature  
Mercury-300BB "nmrc300"

Relax. delay 1.000 sec  
Pulse 45.0 degrees  
Acq. time 1.812 sec  
Width 17006.8 Hz  
512 repetitions  
OBSERVE C13, 75.4708189 MHz  
DECOUPLE H1, 300.1437817 MHz  
Power 37 dB  
continuously on  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
FT size 65536  
Total time 26 min, 19 sec





DEPT NMR

CH3 carbons



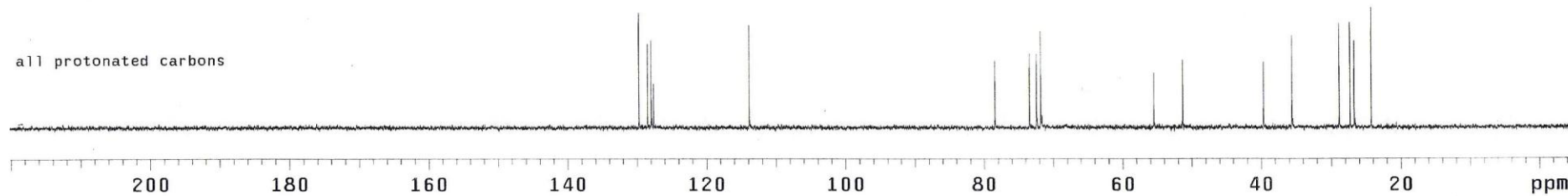
CH2 carbons



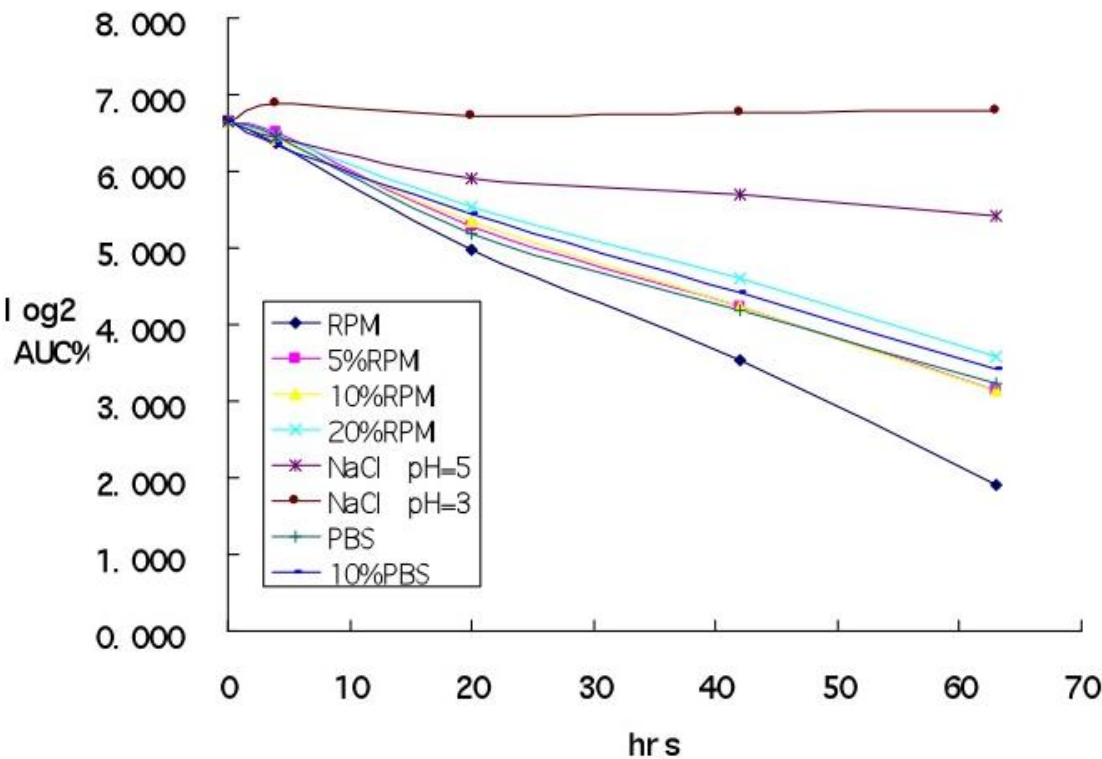
CH carbons



all protonated carbons



$\beta$  -BFUdR stability at 37 °C

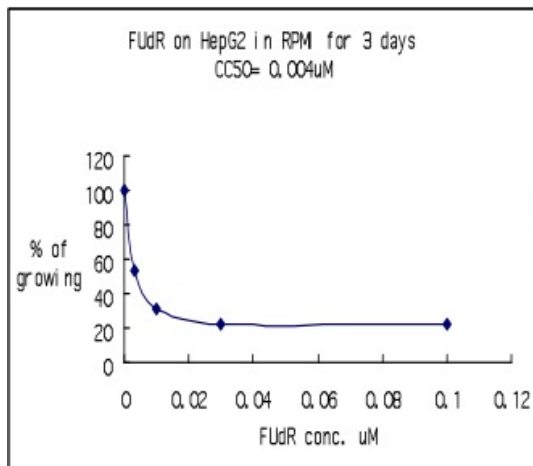


AUC = area under curve, integrated HPLC signal. RPM is RPMI-1640, 10%RPM is 10% bovine fetal serum/90% RPMI-1640, PBS is phosphate, BFUdR is  $\beta$ -14b.

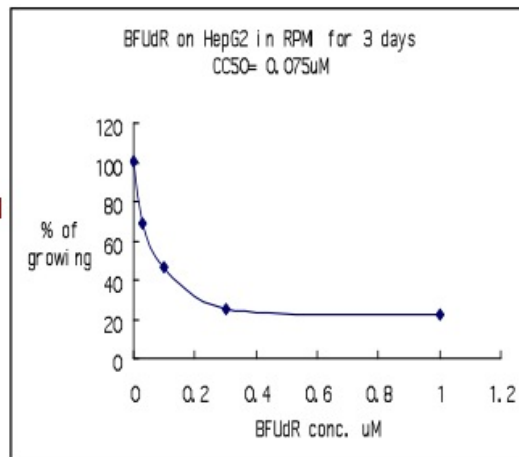
Half life of  $\beta$ -BFUdR in different conditions at 37 °C

condition	T1/2 (hrs)
RPMI pH=7.4	13.4
5 % FBS-RPMI pH=7.4	17.7
10 % FBS-RPMI pH=7.4	18.0
20 % FBS-RPMI pH=7.4	20.6
150 mM Nacl pH=5	54.1
150 mM Nacl pH=3	stable
PBS pH=7.4	18.3
10 % FBS-PBS pH=7.4	19.8

RPMI	$y = -0.0747x + 6.612$
	$R^2 = 0.9983$
5%FBS-RPMI	$y = -0.0564x + 6.6142$
	$R^2 = 0.9933$
10%FBS-RPMI	$y = -0.0556x + 6.5915$
	$R^2 = 0.9973$
20%FBS-RPMI	$y = -0.0485x + 6.6214$
	$R^2 = 0.9976$
150mM Nacl pH=5	$y = -0.0185x + 6.4973$
	$R^2 = 0.9268$
150Mm Nacl pH=3	stable
PBS	$y = -0.0545x + 6.5508$
	$R^2 = 0.9861$
10%FBS-PBS	$y = -0.0506x + 6.5558$
	$R^2 = 0.997$



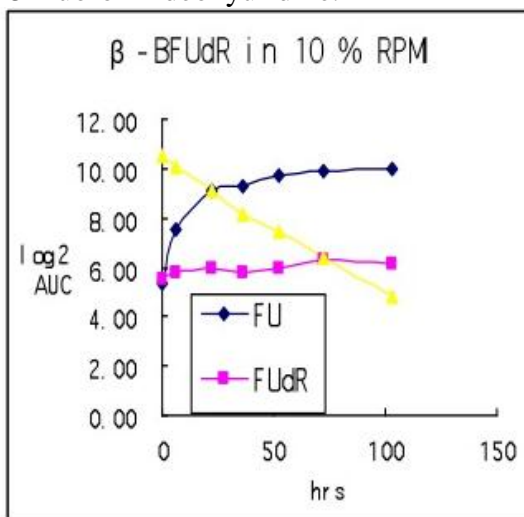
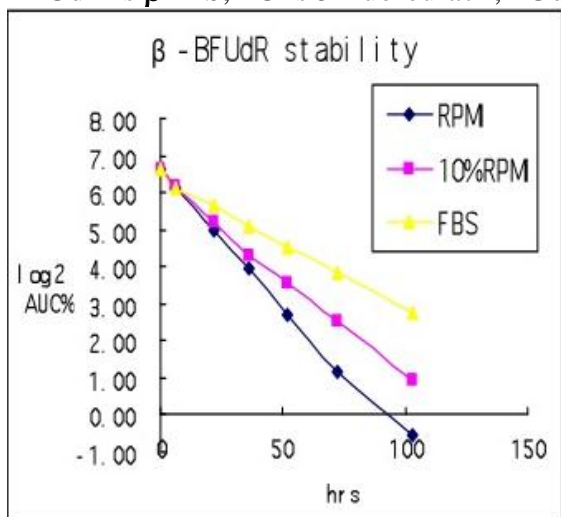
19 fold



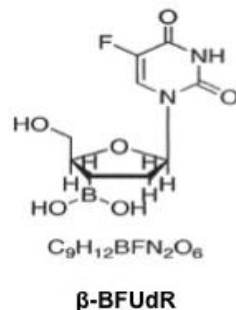
$\beta$ -BFDuR in 10% RPMI			
hrs	FU%	FUdR%	$\beta$ -BFDuR%
0	4.6	3.0	92.4
4	10.9	5.5	83.6
20	45.3	6.9	47.8
42	69.3	7.6	23.0
63	81.7	7.7	10.5

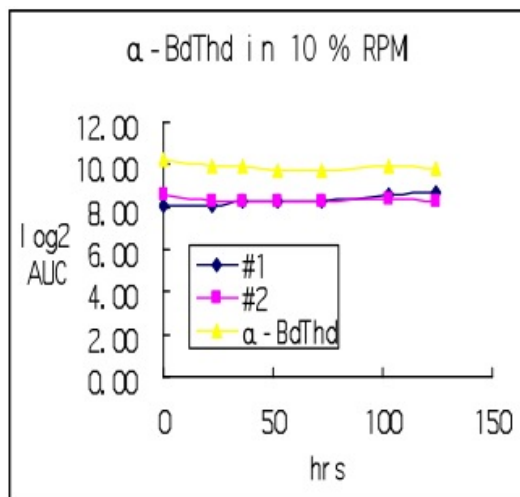
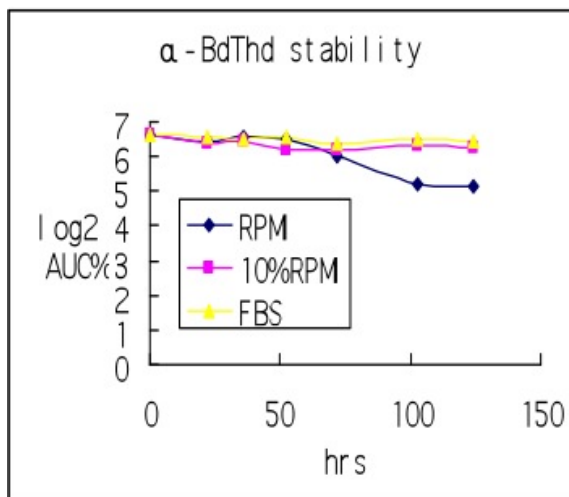
In 63 hrs, about 5%  $\beta$ -BFDuR was converted into FUdR, which might explain the phenomenon that the cytotoxicity of  $\beta$ -BFDuR is about 5% of FUdR.

BFDuR is  $\beta$ -14b, FU is 5-fluorouracil, FUdR is 5-fluoro-2'-deoxyuridine.

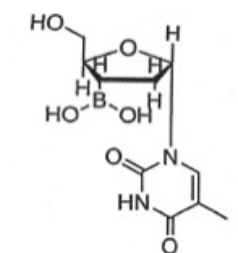


$\beta$ -BFDuR	RPMI	10 % RPMI	FBS
T1/2 (hrs)	14.1	18.3	27.3



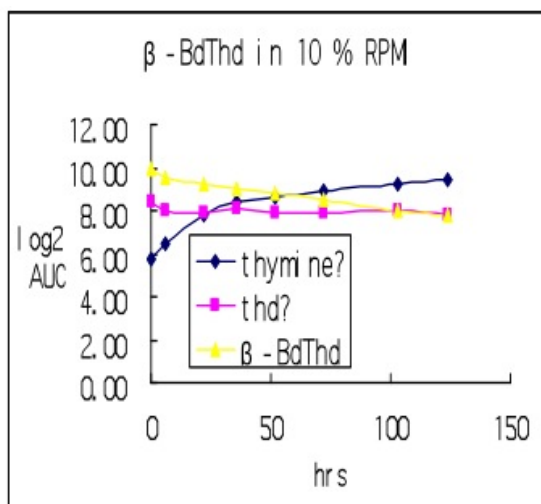
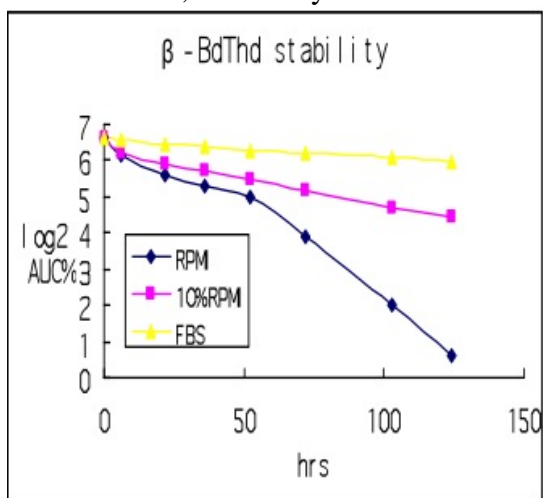


$\alpha$ -BdThd	RPMI	10 % RPMI	FBS
T1/2 (hrs)	72.5	454.5	769.2

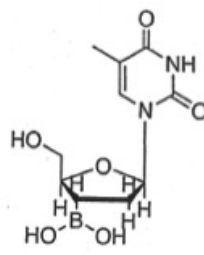


$\alpha$ -BdThd

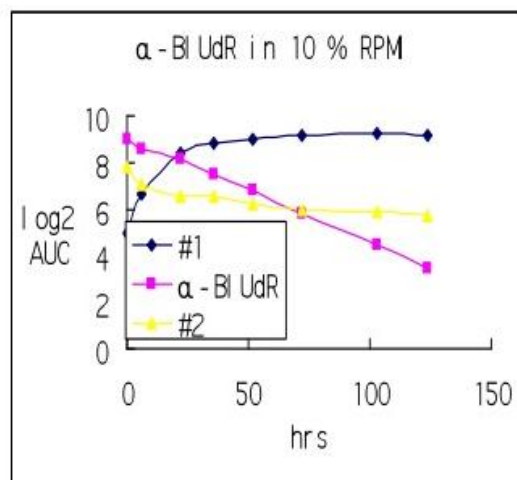
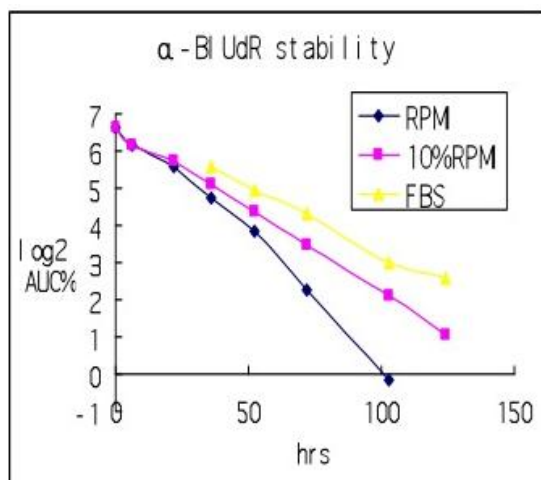
BdThd is **14a**, Thd is thymidine.



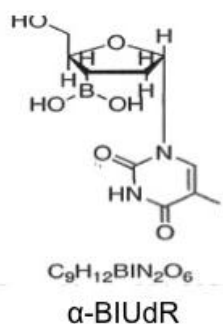
$\beta$ -BdThd	RPMI	10 % RPMI	FBS
T1/2 (hrs)	21.8	61.3	192.3



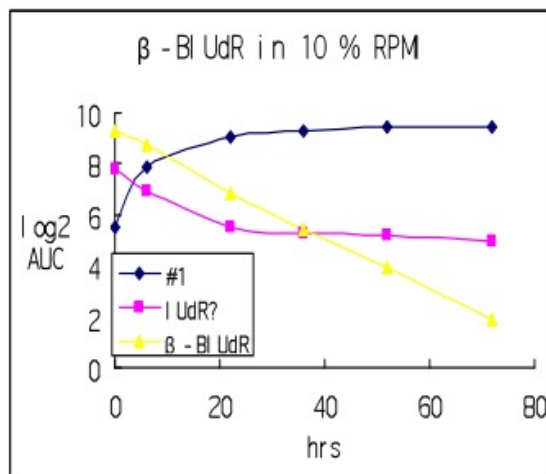
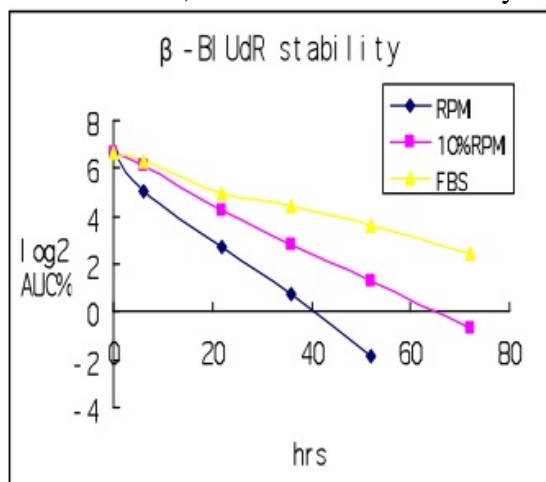
$\beta$ -BdThd



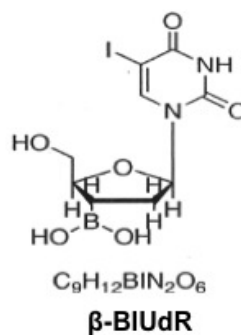
$\alpha$ -BIUdR	RPMI	10 % RPMI	FBS
T1/2 (hrs)	15.5	22.6	28.3

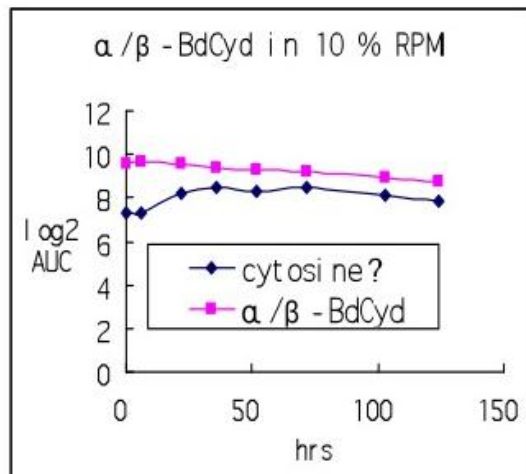
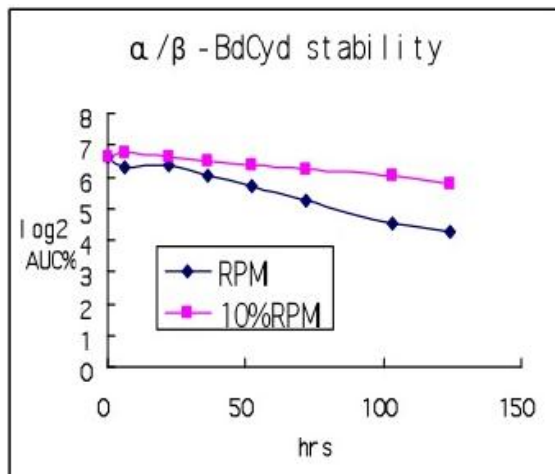


BIUdR is **14c**, IUdR is 5-iodo-2'-deoxyuridine.

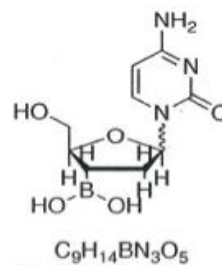


$\beta$ -BIUdR	RPMI	10 % RPMI	FBS
T1/2 (hrs)	6.4	9.7	17.5





$\alpha/\beta$ -BdCyd	RPMI	10 % RPMI	FBS
T1/2 (hrs)	51.8	138.9	400.0



**$\alpha/\beta$ -BdCyd**

BdCyd is **14d**.

Stability of the compounds at 37 °C			
T1/2 (hrs)	RPMI	10 % RPMI	FBS
$\beta$ -BFUdR	14.1	18.3	27.3
$\alpha$ -BdThd	72.5	454.5	769.2
$\beta$ -BdThd	21.8	61.3	192.3
$\alpha$ -BIUdR	15.5	22.6	28.3
$\beta$ -BIUdR	6.4	9.7	17.5
$\alpha/\beta$ -BdCyd	51.8	138.9	400.0
IUdR	769.2	625.0	166.7
FUdR	stable	stable	