Supporting Information for

# Fluoride as An Inducible DNA Cross-linking Agent for New Antitumor Prodrug

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#### Apparatus

<sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on Varian Mercury 300 spectrometers, respectively. HRMS were recorded on a Brucker APEX IV (7.0 T).

#### Materials

All solvents and reagents were commercially available and used without further purification unless for special needs: MEM (HyClone, Thermo Scientific), fetal bovine serum (FBS, HyClone), penicillin and streptomycin (Invitrogen), MTT (Sigma), and propidium iodide (Sigma-Aldrich). HeLa cells were purchased from China Center for Type Culture Collection.

#### Synthesis experiments

**General information:** Unless otherwise specified, chemicals were purchased from Alfa Aesar or Sigma-Aldrich and were used as received without further purification.



**Scheme S1**. Synthesis of compound **1**.a) *tert*-Butyldimethylsilyl chloride, chloroform, RT; b) sodium borohydride, ethanol, RT; c) TFA, THF; d) lithium bromide, THF; e) *N*-methyldiethanolamine, chloroform; f) thionyl chloride, RT.

#### Synthesis of compound 1

**4-(***tert***-Butyldimethylsilyloxy)benzaldehyde (1b)**<sup>1</sup>: To a mixture of **1a** (1.2 g, 10 mmol) in 50mL chloroforms was added *tert*-butyldimethylsilyl chloride (1.8 g, 12 mmol), TEA (1 ml), the reaction mixture was stirred at room temperature for 12 h. The mixture was diluted by DCM, washed by H<sub>2</sub>O ( $3\times50$  mL) and dried with anhydrous Na<sub>2</sub>SO<sub>4</sub>. The mixture was evaporated and the residue was subjected to column chromatography on silica gel with 0–5% DCM in hexane to give the desired

product **1b** as a white solid (1.9 g, 86%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 1.38$  (s, 4 H), 3.08 (s, 9 H), 7.26 (s, 2 H), 11.97 ppm (s, 1 H).

**4-**(*tert*-**Butyldimethylsilyloxy)phenylmethanol** (**1c**)<sup>1</sup> : NaBH<sub>4</sub> (0.28 g, 7.5 mmol) was added to a mixture of **1b** (1.2 g, 5 mmol) in 50mL ethanol. The mixture was stirred at room temperature for 2h. Then the reaction mixture was evaporated and diluted with chloroform, washed by H<sub>2</sub>O (3 × 50 mL) and dried with anhydrous Na<sub>2</sub>SO<sub>4</sub> respectively. The solution was evaporated and the residue was subjected to column chromatography on silica gel with ethyl acetate/cyclohexane (1: 5, v/v) as eluent to get the desired product as a white solid **1c** (0.9 g, 90%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  = 0.02 (s, 4 H), 0.89 (s, 9H), 4.87(s, 2 H), 5.31(s, 1 H), 6.70 (s, 2 H), 7.06 ppm (s, 2 H).

**4-**(*tert*-**Butyldimethylsilyloxy)benzyl 2,2,2-trifluoroacetate** (**1d**) <sup>1</sup>: TFA (1 ml, 1.2 mmol) was added to a mixture of **1c** (0.24 g, 1 mmol) in 30 mL THF. The reaction mixture was refluxed for 0.5h and then mixture was diluted by chloroform (30 mL), washed by H<sub>2</sub>O ( $3 \times 50$  mL). After the isolated solution was dried with anhydrous Na<sub>2</sub>SO<sub>4</sub>, it was evaporated and the residue was subjected to column chromatography on silica gel with ethyl acetate/cyclohexane (1:10, v/v) as eluent to get the desired product **1d** as a yellow oil (0.23 g, 70%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 0.06$  (s, 4 H), 1.23(s, 9H), 4.98(s, 2 H), 7.06(s, 2 H), 7.36 ppm (s, 2 H).

**4-(tert-Butyldimethylsilyloxy)benzylbromide** (1e) <sup>1</sup>: To a mixture of 1d (0.34 g, 1 mmol) in 30mL THF was added LiBr (0.1 g, 1.2 mmol), the reaction mixture was refluxed for 20h. After the mixture was diluted by chloroform (30 ml), washed by H<sub>2</sub>O ( $3 \times 50$  mL) and dried with anhydrous Na<sub>2</sub>SO<sub>4</sub>, the solution was evaporated and the residue was subjected to column chromatography on silica gel with ethyl acetate/cyclohexane (1:10, v/v) as eluent to get the desired product 1e as a yellow solid (0.23 g, 78%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 0.21$  (s, 4 H), 0.98 (s, 9 H), 4.56 (s, 2 H), 6.86 (s, 2 H), 7.16 ppm (s, 2 H).

**4-Ditert-butylmethylsilyloxy-***N*, *N*'-**bis**(**2-hydroxyethyl**)-*N*, *N*'-**dimethylphenyl** (**1f**) : A solution of 4-(*tert*-butyldimethylsilyloxy)benzylbromide (**1e**) (0.15 g, 0.5 mmol) and *N*-methyldiethanolamine (0.12 g, 1 mmol) in chloroform (40 mL) was refluxed for overnight. After filtration, the rude product was recrystallized by ethanol and ethylether for several times. The product **1f** was obtained as a white solid (0.21 g, 90%). <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>):  $\delta = 0.82$  (s, 4 H), 0.81(s, 9 H), 2.83(s, 3 H), 3.17-3.37 (m, 4 H), 3.76 (s, 4 H), 4.49 (s, 2 H), 5.20 (s, 2 H), 6.80 (s, 2 H), 7.35 ppm (s, 2 H). <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 600 MHz):  $\delta$  153.5, 129.6, 125.5, 120.3, 66.3, 64.5, 48.1, 25.5, 17.3, -4.03 ppm. ESI-MS found m/z = 340.9 [M-Br]<sup>-</sup>.

**4-Di***tert*-butylmethylsilyloxy-*N*, *N*'-bis(2-chloroethyl)-*N*, *N*'-dimethylphenyl (1) : A solution of 4-di*tert*-butylmethylsilyloxy-*N*, *N*'-bis(2-hydroxyethyl)-*N*,

*N'*-dimethylphenyl (**1f**) (0.1 g, 0.20 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (20 mL) was slowly added dropwise to thinoyl chloride (2 mL) in ice-water bath, then stirred at r. t. for 2 days. After evaporated, the residue was recrystallized with CH<sub>3</sub>OH/Et<sub>2</sub>O several times, then the deaired product **1** was obtained (0.086 g, 85%). <sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>):  $\delta = 0.23$  (s, 4 H), 0.96 (s, 9H), 3.07 (s, 3 H), 3.69-3.96 (m, 4H), 4.18 (s, 4 H), 4.72 (s, 2 H), 7.16 ppm (s, 2 H); <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 600 MHz):  $\delta 151.5$ , 130.6, 126.5, 119.3, 65.3, 63.5, 46.1, 23.5, 16.3, -4.09 ppm. HRMS-ES (*m*/*z*) [M-Br]<sup>+</sup> calcd. For C<sub>18</sub>H<sub>32</sub>Cl<sub>2</sub>NOSi, 376.1625; found, 376.1624

#### **DNA experiments:**

**General Information:** Plasmid DNA (pBR322) was purchased from Fermentas Co., Ltd. The DNA oligonucleotide was purchased from Takara BIO Co.Ltd., with the 5'-terminus fluorolabeled with TAMRA.

#### General protocol for alkaline agarose gel electrophoresis:

See reference 2.

#### Cross-linking of DNA oligonucleotide by compounds:

Cross-linking reaction was carried out in a volume of 10  $\mu$ L containing 5  $\mu$ M selective 5'-terminus fluoro-labeled oligonucleotide, 10 mM KF, 10 mM phosphate buffer (pH = 7.4) and 1 mM compound. The mixture was incubated at 37 °C for 3 h. Then the solution was mixed with 10  $\mu$ L formamide deionized to the final volume of 20  $\mu$ L. The residue was took out and then analyzed by a 20% denaturing polyacrylamide electrophoresis. The cross-linking products of compounds **1** and **2** with oligonucleotide were produced follow the same protocol mentioned as above.

#### **Piperidine treatment**

The cross-linked products were isolated by following procedures: first, ethanol and sodium acetate/acetic acid buffer (pH = 5) at -20 °C for one night, then the mixture was centrifugated 12000 rpm at 4 °C for 20 min, dried and treated with 100  $\mu$ L 1 M piperidine for 30 min at 90 °C. After stored and precipitated by ethanol at -20 °C for one night, the DNA product fragments were then obtained and further dissolved it in formamide deionized. The cross-linking results were analyzed by polyacrylamide gel electrophoresis under denaturing conditions.



Figure S1. Cross-linking results of compound 2 with different fluoride concentration. lane 1: 0.5µg pBR322 control ; lane 2: 0.5 µg pBR322 + 50 µM compound  $2 + 50 \mu$ M KF; lane 3 0.5µg pBR322 + 50 µM compound  $2 + 100 \mu$ M KF; lane 3 : 0.5µg pBR322 + 50 µM compound  $2 + 500 \mu$ M KF; lane 4: 0.5µg pBR322 + 50 µM compound  $2 + 1 \mu$ M KF.



Figure S2. Cross-linking results of compound 2 with different anion selectivity. lane 1:  $0.5\mu g \ pBR322$  control ; lane 2:  $0.5\mu g \ pBR322 + 50 \ \mu M$  compound 2 + 10 mM IO<sub>4</sub> ; lane 3  $0.5\mu g \ pBR322 + 50 \ \mu M$  compound 2 + 10 mM OCl ; lane 4 :  $0.5\mu g \ pBR322 + 50 \ \mu M$  compound 2 + 10 mM Br ; lane 5:  $0.5\mu g \ pBR322 + 50 \ \mu M$  compound 2 + 10 mM KF.



**Figure S3.** Cell viability in the presence of **2** at different concentrations (50 nM-100  $\mu$ M) with 100  $\mu$ M fluoride. The data were obtained through MTT assay and presented as mean  $\pm$  SD (n= 3).



**Figure S4.** Cell viability in the presence of fluoride at different concentrations (10  $\mu$ M-50mM). The data were obtained through MTT assay and presented as mean  $\pm$  SD (n= 3).

#### Reference

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