Electronic Supplementary Information For

An Acid Catalyzed Reversible Ring-closure Reaction of Cyano-Rhodamine Spirolactam

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Materials and general methods:

Chemicals used in the syntheses were purchased from Aldrich without further purification. ¹H-NMR and ¹³C-NMR spectra were recorded on a Varian Mercury/VX 300 spectrometer and chemical shifts are reported as ppm (in CDCl₃ or with CD₃OD and DCl mixture, TMS as internal standard). Mass spectra were recorded using Micromass Q-TOF I mass spectrometer. IR data were recorded with a Shimadzu FTIR Spectrophotometer. Fluorescence emission and excitation spectra were recorded using a Varian Cary Eclipse Fluorescence Spectrophotometer. All pH measurements were made on an Accumet Basic AB15 pH meter. X-ray intensity data were collected using a Bruker SMART APEX diffractometer. Cell images were obtained by Olympus X81 fluorescence microscopy.

pH titration experiment

A 5 mM stock solution of **RBCN** in CH₃CN was prepared and then diluted in H₂O to give the 10 μ M of **RBCN** sample. The **RBCN** sample was stirred at room temperature and pH was adjusted by 0.1 M, 2.0 M and 10.0 M of HCl_(aq) and NaOH_(aq), then absorption and fluorescence signals were recorded. For the reversible pH-dependent fluorescence changes experiment, the pH of **RBCN** sample was first adjusted to ~10 and then to ~2 and then adjusted between ~10 and ~2 for several cycles. Fluorescence signals were recorded in different cycle, respectively. 2.0 M and 10.0 M of HCl_(aq) and NaOH_(aq) were adopted in this assay to avoid changing the volume so much (totally ~2 % for 10 cycles).

Cell imaging

SW620 cells were cultured in RPMI1640(VWR) supplemented with 10% FCS (Thermoscientific). One day before imaging, cells were seeded into 12-well flat bottomed plates. After 24 h, the cells were co-stained with 5 μ M **RBCN** and trackers (15 nM for LysoTracker Green or 500 nM for Mitotracker Green) for 30 min at 37 °C under 5% CO₂ and washed with phosphate-buffered saline (PBS) three times. After replacement of medium, cells were imaged using Olympus X81 fluorescence microscopy.

Synthesis of RBCN

In a 50 mL flask, rhodamine B (100.0 mg, 0.21 mmol) was dissolved in 25 mL ethanol. The red solution was heated to reflux under stirring, then cyanamide (50.0 mg, 1.2 mmol) was added. The resultant reaction mixture was refluxed for 72 h. TLC showed around 50% of the starting material was converted to product (**RBCN**) with much weaker polarity. After removal of ethanol under vacuum, the residue was purified by flash chromatography with CH₂Cl₂/EtOAc = 4/1 as eluent to give the light pink powder **RBCN** (53.5 mg, yield: 54.7%). ¹H-NMR (400 MHz, CDCl₃), δ_H (ppm): 7.98 (d, 1H, J = 7.4 Hz, C₆H₄), 7.65 (t, 1H, J = 7.4 Hz, C₆H₄), 7.55 (t, 1H, J = 7.4 Hz, C₆H₄), 7.16 (d, 1H, J = 7.4 Hz, C₆H₄), 6.53 (d, 2H, J = 8.8 Hz, Xanthene-H), 6.41 (s, 2H, Xanthene-H), 6.33 (d, 2H, J = 8.8 Hz, Xanthene-H), 3.33 (q, 8H, J = 6.8 Hz, CH₂), 1.17 (t, 12H, CH₃); ¹³C-NMR (100 MHz, CDCl₃), δ_C (ppm): 166.6489, 153.3394, 152.5031, 149.4970, 135.5330, 129.3200, 128.4062, 126.6485, 124.8635, 124.2857, 108.4320, 106.9820, 103.1700, 97.7815, 69.2468, 44.4259, 12.5743; IR (KBr): 2235 (s), -CN; HRMS *m/e* calculated for (M+H)⁺ C₂₉H₃₀N₄O₂ 467.2447; found 467.2449.

Crystallography

X-ray intensity data from a colorless plate like crystal were collected at 100(2) K using a Bruker SMART APEX diffractometer (Mo Ka radiation, 1 = 0.71073 Å).¹ The raw area detector data frames were reduced with the SAINT+ program.¹ Final unit cell parameters were determined by least-squares refinement of 4087 reflections from the data set. Direct methods structure solution, difference Fourier calculations and full-matrix least-squares refinement against F^2 were performed with SHELXS/L² as implemented in OLEX2.³

The compound crystallizes in the triclinic crystal system. The space group P-1 (No. 2) was assumed and confirmed by the successful solution and refinement of the structure. The asymmetric unit consists of two crystallographically independent molecules and a methanol molecule of crystallization with a population of 50%. The methanol is disordered about an inversion center, and thus only half is present per asymmetric unit. It was further modeled as occupying two disordered sites with the asymmetric unit with occupancies O1S-C1S/O2S-C2S = 0.263(7)/0.237(7). These occupancies were constrained to sum to 0.5. The disordered methanol atoms were refined with C-O distances restrained to 1.45(2) Å, and were assigned a common isotropic displacement parameter. All other non-hydrogen atoms were refined with anisotropic displacement parameters. Hydrogen atoms were placed in geometrically idealized positions and included as riding atoms. Hydrogen atoms were not located or calculated for the disordered methanol atoms.

Identification code	RBCN
Empirical formula	C29.25H31N4O2.25
Formula weight	474.58
Temperature/K	100(2)
Crystal system	triclinic
Space group	P-1
a/Å	11.2094(16)
b/Å	14.479(2)
c/Å	16.455(2)
$\alpha ^{\prime \circ}$	80.674(3)
β/°	86.095(3)
$\gamma/^{\circ}$	75.836(3)
Volume/Å3	2554.1(6)
Z	4
pcalcmg/mm3	1.234
m/mm-1	0.079
F(000)	1010.0
Crystal size/mm3	0.44 imes 0.32 imes 0.1
2Θ range for data collection	2.5 to 50.06°
Index ranges	$-13 \le h \le 13, -17 \le k \le 17, -19 \le l \le 19$
Reflections collected	28263
Independent reflections	9012[R(int) = 0.0653]
Data/restraints/parameters	9012/2/653
Goodness-of-fit on F2	0.855
Final R indexes [I>= 2σ (I)]	R1 = 0.0459, wR2 = 0.0744
Final R indexes [all data]	R1 = 0.0847, wR2 = 0.0843
Largest diff. peak/hole / e Å-3	0.26/-0.19



Table S1. X-ray crystallographic data for RBCN.

Figure S1. (a) Components of the crystal structure. The asymmetric unit contains two crystallographically independent, chemically identical molecules and a disordered methanol molecule of crystallization. (b) The two independent molecules overlaid. Green = molecule with label suffix "A"; red = "B".





Figure S3. Photos of **RBCN** in other protic solvents under sunlight (top) and UV light (bottom). From left to right: 1 = isopropanol alcohol, 2 = isobutyl alcohol, 3 = 1-butabol, 4 = ethylene glycol, 5 = diethylene glycol, 6 = glycerin.



Figure S4. (a) Fluorescence intensity changes of **RBCN** (at 570 nm) in acetone, acetonitrile, THF, DMF and DMSO with different portions (v/v) of water. (b) Fluorescence spectrum changes of **RBCN** in acetonitrile with different portions of water. Inset shows the photo of **RBCN** before and after addition of



5.2 % water under UV-light. 5 µM RBCN for each sample and the excitation was performed at 530 nm.

Figure S5 ¹³C-NMR spectrums of **RBCN** in (a) CDCl₃, (b) CDCl₃ (40%) + CD₃OD (60%) and (c) CDCl₃ (33.3%) + CD₃OD (50%) + DCl (16.7%).



Figure S6. ¹H-NMR spectrums of **RBCN** in (a) $CDCl_3$, (b) $CDCl_3$ (40%) + CD_3OD (60%) and (c) $CDCl_3$

$(33.3\%) + CD_3OD(50\%) + DCl(16.7\%).$



Figure S7. ¹H-NMR spectrum of **RBCN** after treatment of DCl (3days later).



Figure S8. MS spectrum of RBCN after treatment of DCl (3days later).

After treatment by DCl for 3 days, 3'-H was deuterated (Figure S7) that was also supported by MS result (Figure S8).

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Figure S9. ¹H-NMR of **RBCN** (in CDCl₃).



PPNLEDUEDDIDOGO/GCO4BO010 Henning/15554 Harristaning/Constant Marting/15344 Harristaning/Data Barristaning/Constant Marting/15344 Harristaning/Data Barristaning/Data Barristaning/Data

Figure S10. ¹³C-NMR of **RBCN** (in CDCl₃).

References:

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(3) Dolomanov, O. V.; Bourhis, L. J.; Gildea, R. J.; Howard J. A. K.; Puschmann, H. J. Appl. Cryst. 2009, 42, 339-341.