

## Supplementary Information

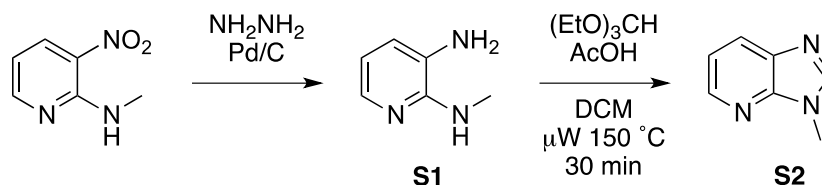
### Regioselective C2-Arylation of Imidazo[4,5-*b*]pyridines.

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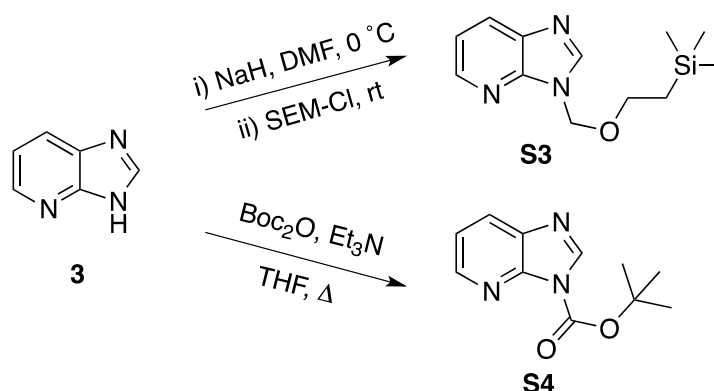
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<b>1. Synthetic Schemes to Intermediates .....</b>	<b>S2</b>
<b>2. Deprotection of C-H Arylation Products.....</b>	<b>S4</b>
<b>3. Kinetic Isotope Experiment .....</b>	<b>S5</b>
<b>4. Experimental data for synthesis of intermediates .....</b>	<b>S6</b>
<b>5. NMR Spectra .....</b>	<b>S12</b>
<b>5.1 Spectra of intermediates .....</b>	<b>S12</b>
<b>5.2 Spectra of C-H arylation products .....</b>	<b>S31</b>
<b>5.3 Spectra of deprotected C2-arylated products.....</b>	<b>S61</b>

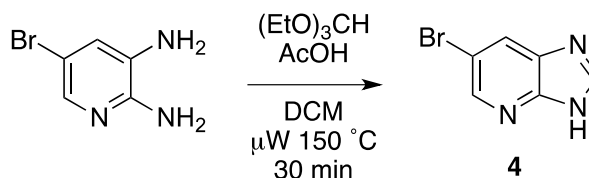
## 1. Synthetic Schemes to Intermediates



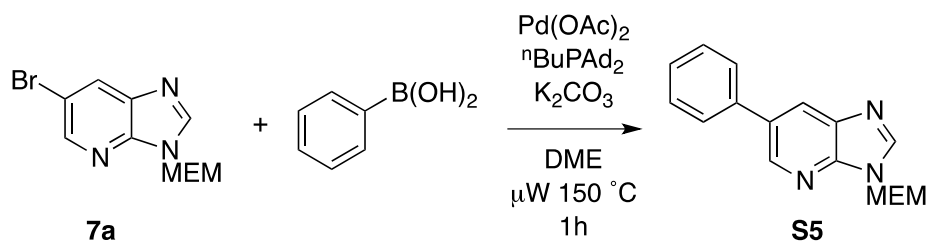
**Scheme S1.** Synthesis of *N*3-methylimidazo[4,5-*b*]pyridine, **S2**.



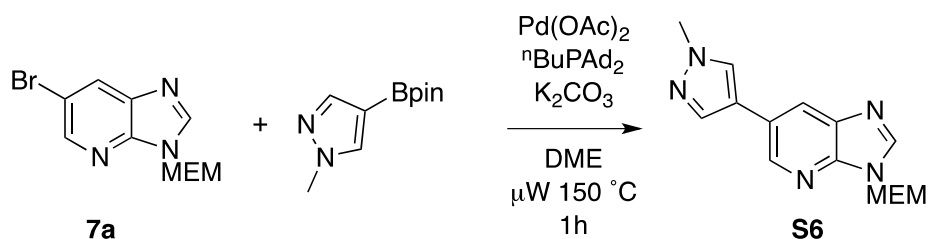
**Scheme S2.** Synthesis of *N*3-SEM and Boc protected imidazo[4,5-*b*]pyridines.



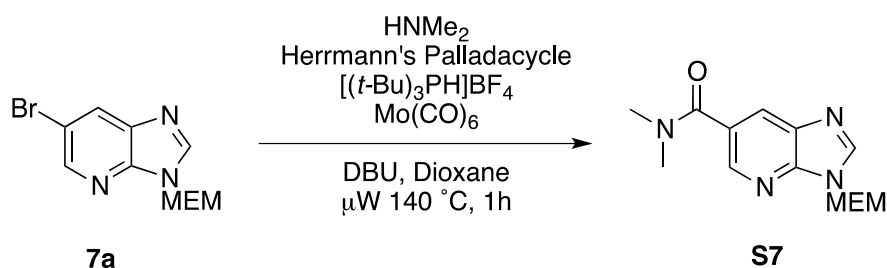
**Scheme S3.** Synthesis of 6-bromoimidazo[4,5-*b*]pyridine **4**.



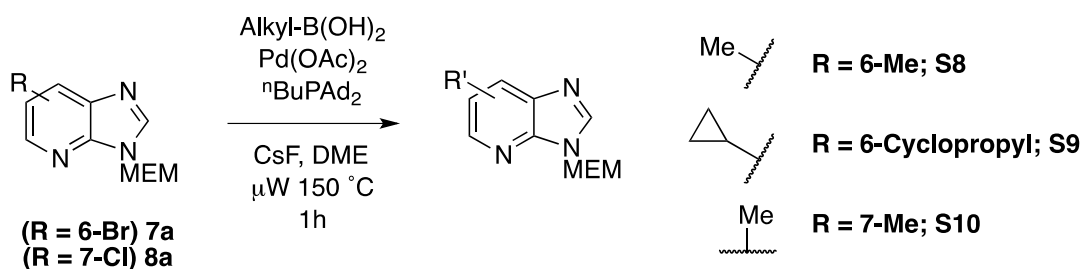
**Scheme S4.** Suzuki cross-coupling to 3-((2-methoxyethoxy)methyl)-6-phenyl-3H-imidazo[4,5-*b*]pyridine, **S5**.



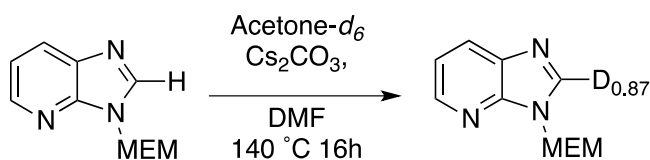
**Scheme S5.** Suzuki cross-coupling to 3-((2-methoxyethoxy)methyl)-6-(1-methyl-1H-pyrazol-4-yl)-3H-imidazo[4,5-b]pyridine, **S6**.



**Scheme S6.** Aminocarbonylation protocol affording 3-((2-methoxyethoxy)methyl)-N,N-dimethyl-3H-imidazo[4,5-b]pyridine-6-carboxamide, **S7**.<sup>1</sup>



**Scheme S7.** Preparation of alkyl-substituted *N*3-MEM-imidazo[4,5-*b*]pyridines, **S8-S10**.

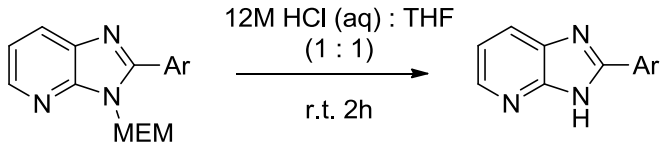
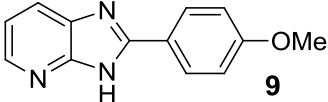
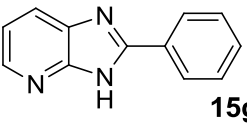
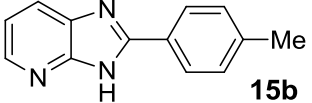
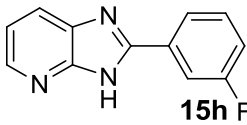
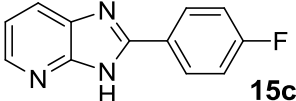
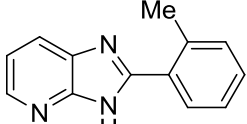
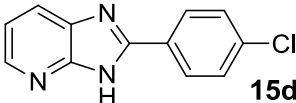
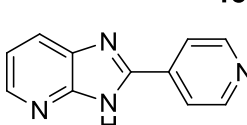
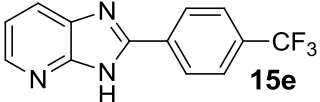
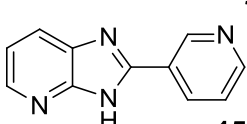
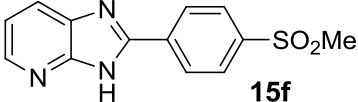
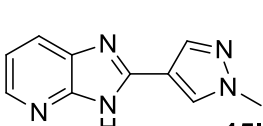


**Scheme S8:** Investigating the reaction mechanism; C2-deprotonation under C-H arylation conditions.

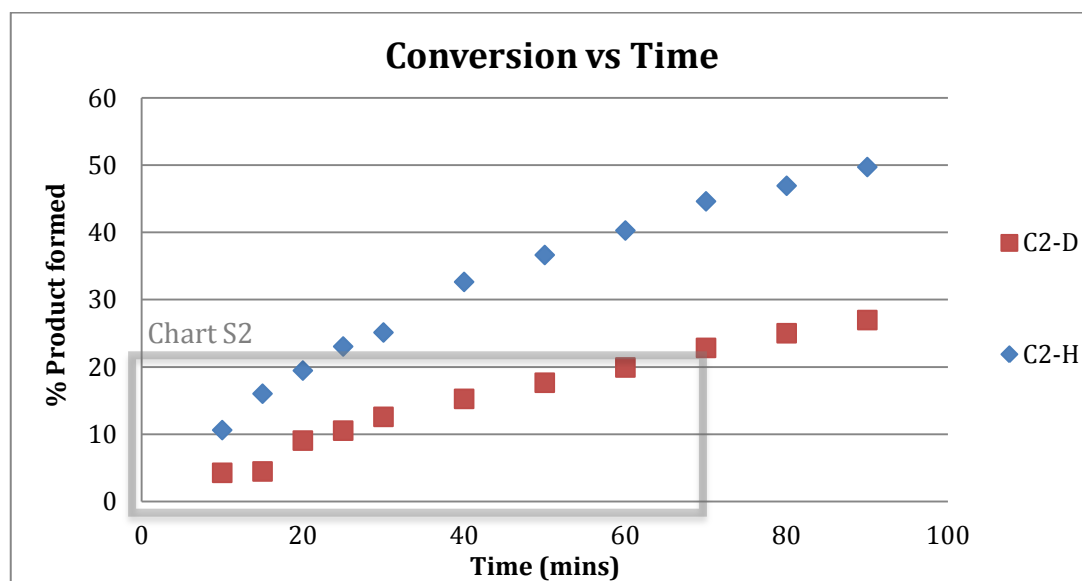
<sup>1</sup> (a) Appukkuttan, P.; Axelsson, L.; der Eycken, E. V.; Larhed, M. *Tet. Lett.* **2008**, 49, 5625. (b) Lagerlund, O.; Larhed, M. *J. Comb. Chem.* **2006**, 8,4.

## 2. Deprotection of C-H Arylation Products

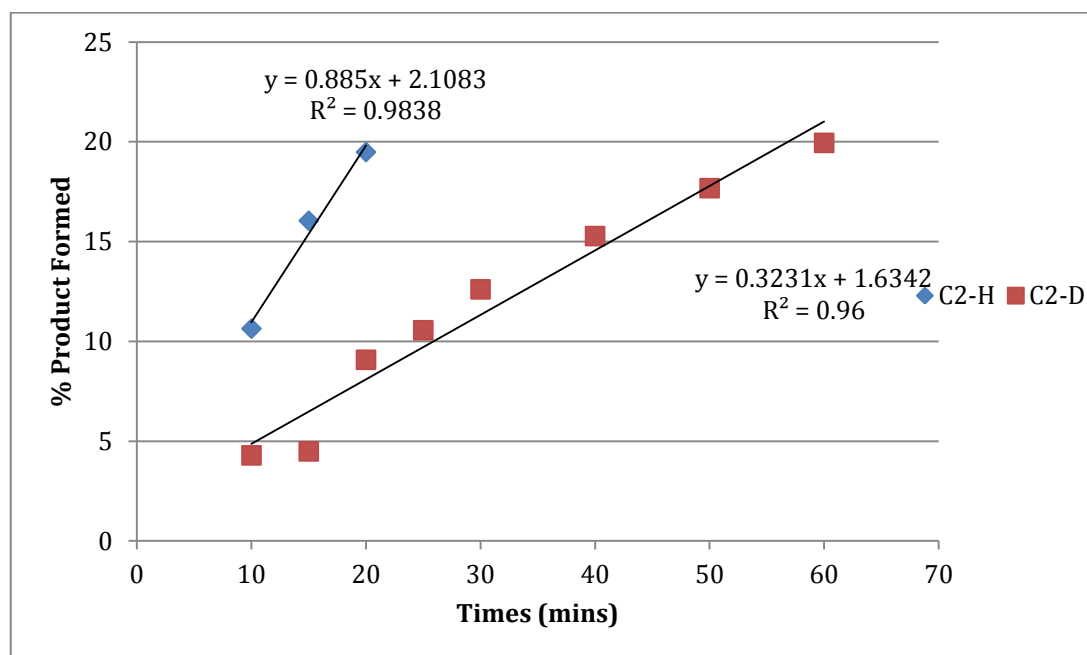
**Table S1.** Acid-mediated deprotection of C-H arylation products.

 <p style="text-align: center;"> <chem>Cc1ccc(cc1)C2=NC3=CC=CC=C3N2NMEM</chem> <math>\xrightarrow[\text{r.t. 2h}]{12\text{M HCl (aq) : THF (1 : 1)}}</math> <chem>Cc1ccc(cc1)C2=NC3=CC=CC=C3N2</chem> </p> <p style="text-align: center;"><b>13a-l</b> <span style="margin-left: 150px;"><b>9 and 15b-l</b></span></p>					
Entry	Product	Isolated Yield	Entry	Product	Isolated Yield
1	 <b>9</b>	98%	7	 <b>15g</b>	81%
2	 <b>15b</b>	86%	8	 <b>15h</b>	90%
3	 <b>15c</b>	98%	9	 <b>15i</b>	76%
4	 <b>15d</b>	50%	10	 <b>15j</b>	46%
5	 <b>15e</b>	53%	11	 <b>15k</b>	38%
6	 <b>15f</b>	67%	13	 <b>15l</b>	48%

### 3. Kinetic Isotope Experiment



**Chart S1.** Chart showing conversion versus time for KIE experiments. The ‘% product formed’ is defined as the ratio of product formed to remaining starting material, as measured by LCMS UV-Absorbance at  $\lambda_{254}$  nm.



**Chart S2.** Initial rate corresponding to 20% conversion.

Kinetic Isotope Effect calculation: 
$$KIE = \frac{k_H}{k_D} = \frac{0.885}{0.323} = 2.74$$

#### 4. Experimental for synthesis of intermediates

See main manuscript for general experimental.

**N<sup>2</sup>-Methylpyridine-2,3-diamine, S1:** A flask containing *N*-methyl-3-nitropyridin-2-amine (2 g, 13.1 mmol) and Pd/C (10 %) (60 mg) was flushed with N<sub>2</sub> and DME (15 mL) added. The mixture was stirred at 40 °C and hydrazine monohydrate (1.8 mL, 40 mmol) was added drop-wise. Upon complete addition the mixture was heated to reflux for 90 minutes. Upon cooling, the mixture was filtered through a bed of celite (upper) and silica (lower). The filter bed was washed with EtOAc:Et<sub>2</sub>O (1:1; 60 mL), the solvent mixture was removed under reduced pressure to yield the product as a purple solid (1.6 g, quant.) which was used without further purification. <sup>1</sup>H NMR: (500 MHz, CDCl<sub>3</sub>) δ<sub>H</sub> 7.81 (dd, *J* = 5.1, 1.5 Hz, 1H), 6.85 (dd, *J* = 7.4, 1.5 Hz, 1H), 6.54 (dd, *J* = 7.4, 5.1 Hz, 1H), 4.17 (s, 1H), 3.17 (s, 2H), 3.04 (d, *J* = 2.8 Hz, 3H); <sup>13</sup>C NMR: (126 MHz, CDCl<sub>3</sub>) δ<sub>C</sub> 151.2, 139.3, 128.4, 121.8, 113.2, 28.7; LCMS *t*<sub>R</sub> 0.41 min, *m/z* 124 (M+H)<sup>+</sup>; Purity (AUC) ≥ 95%; HRMS (M+H)<sup>+</sup> *calculated* for C<sub>6</sub>H<sub>10</sub>N<sub>3</sub> = 124.0869, *found* = 124.0959.

**3-Methyl-3H-imidazo[4,5-*b*]pyridine, S2:** To a suspension of *N*<sup>2</sup>-methylpyridine-2,3-diamine, **S1** (1.16 g, 9.4 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (5 mL) was added triethylorthoformate (10 mL) and acetic acid (1.5 mL). The mixture was reacted under microwave irradiation at 150 °C for 30 min. The residue was purified by flash chromatography; conditions (CH<sub>2</sub>Cl<sub>2</sub>:EtOH:NH<sub>3</sub> (95:4:1) yielding the product as a brown solid (1.17g, 8.79 mmol, 93 %). Lit. m.p. 76-78;<sup>2</sup> Observed m.p. 74-77°C; <sup>1</sup>H NMR: (500 MHz, CDCl<sub>3</sub>) δ<sub>H</sub> 8.45 (dd, *J* = 4.8, 1.4 Hz, 1H), 8.10 (dd, *J* = 8.0, 1.4 Hz, 1H), 8.06 (s, 1H), 7.28-7.23 (m, 1H), 3.96 (s, 3H); <sup>13</sup>C NMR: (126 MHz, CDCl<sub>3</sub>) δ<sub>C</sub> 147.0, 144.0, 143.8, 134.9, 127.4, 117.6, 29.2; LCMS *t*<sub>R</sub> = 0.81 mins, *m/z* = 134 (M+H)<sup>+</sup>; Purity (AUC) ≥ 95%; HRMS (M+H)<sup>+</sup> *calculated* for C<sub>7</sub>H<sub>8</sub>N<sub>3</sub> = 134.0713, *found* = 134.0718, (M+Na)<sup>+</sup> *calculated* for C<sub>7</sub>H<sub>7</sub>N<sub>3</sub>Na = 156.0532, *found* = 156.0535.

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<sup>2</sup> Y. Mizuno, M. Ikehara, T. Itoh and K. Saito, *J. Org. Chem.*, 1963, **28**, 1837-1841.

**3-((2-(Trimethylsilyl)ethoxy)methyl)-3H-imidazo[4,5-b]pyridine, S3:**

Imidazo[4,5-*b*]pyridine, **3** (1191 mg, 10 mmol) in DMF (100 mL) was cooled to 0 °C in an ice/water bath. To this was added portionwise NaH (480 mg, 12 mmol) and upon complete addition the mixture was stirred for 15 minutes. SEMCl (2.12 mL, 12 mmol) was added dropwise before stirring the mixture at room temperature for 24 h. The mixture was concentrated *in vacuo*, the residue was taken in 1:1 EtOAc:Cyclohexane (200 mL), washed with water (50 mL), sat. aq. NaCl (3 x 50 mL), dried (MgSO<sub>4</sub>) and concentrated to give crude product which was purified by column chromatography (10 – 50 % EtOAc in cyclohexane) to afford a yellow oil (1.52g, 61%). <sup>1</sup>H NMR: (500 MHz, CDCl<sub>3</sub>) δ<sub>H</sub> 8.44 (dd, *J* = 1.4, 8.8 Hz, 1H), 8.21 (s, 1H), 8.10 (dd, *J* = 1.5, 8.0, 1H), 7.28 (dd, *J* = 4.8, 8.0 Hz, 1H), 5.70 (s, 2H), 3.65 – 3.61 (m, 2H), 0.95 – 0.92 (m, 2H), -0.05 (s, 9H); <sup>13</sup>C NMR: (126 MHz, CDCl<sub>3</sub>) δ<sub>C</sub> 147.1, 144.7, 144.2, 135.2, 128.1, 118.7, 72.0, 67.1, 17.7, -1.48; LCMS *t<sub>R</sub>* = 2.95 mins, *m/z* = 250 (M+H)<sup>+</sup>; Purity (AUC) ≥ 95%; HRMS (M+H)<sup>+</sup> *calculated* for C<sub>12</sub>H<sub>20</sub>N<sub>3</sub>OSi = 250.1376, *found* = 250.1367.

**tert-Butyl 3H-imidazo[4,5-*b*]pyridine-3-carboxylate, S4:** To a stirred suspension of imidazo[4,5-*b*]pyridine, **3** (1191 mg, 10 mmol), Et<sub>3</sub>N (334 μL, 2.4 mmol) and DMAP (24 mg, 0.2 mmol) in THF (10 mL) at r.t. under an N<sub>2</sub> atmosphere, was added Boc<sub>2</sub>O (2M in THF) (1.1 mL, 2.2 mmol). The mixture was heated to 65 °C for 1 hour. Upon cooling the mixture was concentrated *in vacuo* and purified by flash column chromatography (20-40% EtOAc in cyclohexane) to yield a white solid (951 mg, 43%). Observed m.p. = 90 – 92 °C; <sup>1</sup>H NMR: (500 MHz, CDCl<sub>3</sub>) δ<sub>H</sub> 8.67 (s, 1H), 8.62 (dd, *J* = 4.8, 1.6 Hz, 1H), 8.29 (dd, *J* = 8.1, 1.6 Hz, 1H), 7.35 (dd, *J* = 8.1, 4.8 Hz, 1H), 1.72 (s, 9H); <sup>13</sup>C NMR: (126 MHz, CDCl<sub>3</sub>) δ<sub>C</sub> 156.5, 147.4, 146.5, 144.1, 124.2, 122.6, 120.2, 85.6, 28.0; LCMS *t<sub>R</sub>* = 2.54 mins, *m/z* = 164 (loss of *t*-Bu, M+H<sup>+</sup>); Purity (AUC) ≥ 95%; HRMS (M+H)<sup>+</sup> *calculated* for C<sub>11</sub>H<sub>14</sub>N<sub>3</sub>O<sub>2</sub> = 220.1081, *found* = 220.1077, also 164.0626 (loss of *tert*-butyl).

**6-Bromo-3*H*-imidazo[4,5-*b*]pyridine, 4:** A solution of 5-bromopyridine-2,3-diamine (500 mg, 2.66 mmol), triethylorthoformate (5 mL), acetic acid (0.75 mL) and CH<sub>2</sub>Cl<sub>2</sub> (2.5 mL) was irradiated in a  $\mu$ W at 150 °C for 30 mins. The solvents were removed under reduced pressure and the residue was purified by Biotage SP1 (50 g SNAP column, 40 mL/min, 3 CV Cyclohexane, 9 CV 0-10 % MeOH in CH<sub>2</sub>Cl<sub>2</sub>). The product was obtained as a cream solid (358 mg, 69 %). Lit. m.p.<sup>3</sup> 227-229; Observed m.p. 229-231°C; <sup>1</sup>H NMR (CD<sub>3</sub>OD, 500MHz)  $\delta_{\text{H}}$  8.48 (d, *J* = 2.1 Hz, 1H), 8.42 (s, 1H), 8.23 (d, *J* = 2.1 Hz, 1H); <sup>13</sup>C NMR (CD<sub>3</sub>OD, 126 MHz)  $\delta_{\text{C}}$  149.6, 145.0, 144.9, 126.0, 113.5, one q C does not appear; LCMS *t<sub>R</sub>* 1.74 min, *m/z* 197, 199 (M+H)<sup>+</sup> bromine isotopic pattern; Purity (AUC)  $\geq$  95%; HRMS (M+H)<sup>+</sup> *calculated* for C<sub>6</sub>H<sub>5</sub>N<sub>3</sub>Br = 197.9661, *found* = 197.9670; CHN Microanalysis *calculated* for C<sub>6</sub>H<sub>4</sub>BrN<sub>3</sub> = C, 36.39; H, 2.04; N, 21.22%, *observed* = C, 36.33; H, 2.06; N, 21.16%.

**3-((2-Methoxyethoxy)methyl)-6-phenyl-3*H*-imidazo[4,5-*b*]pyridine, S5:** 6-Bromo-3-((2-methoxyethoxy)methyl)-3*H*-imidazo[4,5-*b*]pyridine, **7a** (589 mg, 2.0 mmol), phenyl boronic acid (488mg, 4.0 mmol), K<sub>2</sub>CO<sub>3</sub> (4.0 ml, 1 M, 4.0 mmol), Pd(OAc)<sub>2</sub> (18 mg, 4 mol%) and nBuPAd<sub>2</sub> (57 mg, 8 mol%) were dissolved in DME (5.0 ml) and heated in a microwave reactor at 150 °C for 1 h. The mixture was diluted with ethyl acetate (100 mL) and washed with H<sub>2</sub>O. The crude mixture was concentrated *in vacuo* and purified on Biotage SP1 (12g SINGLE StEP column, 15 mL/min, 3 CV CH<sub>2</sub>Cl<sub>2</sub> then 12 CV 0-100% (5% MeOH in CH<sub>2</sub>Cl<sub>2</sub>)) to afford product as an orange oil (521 mg, 92%). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta_{\text{H}}$  8.66 (d, *J* = 1.9 Hz, 1H), 8.28 (s, 1H), 8.27 (d, *J* = 1.9 Hz, 1H), 7.65 - 7.61 (m, 2H), 7.53 - 7.47 (m, 2H), 7.44 - 7.38 (m, 1H), 5.80 (s, 2H), 3.78 - 3.73 (m, 2H), 3.56 - 3.51 (m, 2H), 3.36 (s, 3H); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta_{\text{C}}$  146.6, 144.9, 144.2, 138.6, 135.3, 133.0, 129.1, 127.6, 127.5 126.5, 73.0, 71.6, 68.9, 59.1; LCMS *t<sub>R</sub>* = 2.74 min, *m/z* 284 (M+H)<sup>+</sup>; Purity (AUC)  $\geq$  95%; HRMS (M+H)<sup>+</sup> *calculated* for C<sub>16</sub>H<sub>18</sub>N<sub>3</sub>O<sub>2</sub> = 284.1394, *found* = 284.1388.

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<sup>3</sup> H. Graboyes and A. R. Day, *J. Am. Chem. Soc.*, 1957, **79**, 6421-6426.



**3-((2-Methoxyethoxy)methyl)-6-(1-methyl-1H-pyrazol-4-yl)-3H-**

**imidazo[4,5-*b*]pyridine, S6:** 6-Bromo-3-((2-methoxyethoxy)methyl)-3H-imidazo[4,5-*b*]pyridine, **7a** (1.0M in DMF, 0.25 mL, 0.25 mmol), 1-methylpyrazole-4-boronic acid pinacol ester (78 mg, 0.375 mmol), Pd(OAc)<sub>2</sub> (2.2 mg, 0.01 mmol) and nBuPAD<sub>2</sub> (7 mg, 0.02 mmol) were taken in 1.0 M aq. K<sub>2</sub>CO<sub>3</sub> (0.5 mL, 0.5 mmol) and DME (0.75 mL), the combined reagents were irradiated in  $\mu$ W at 150 °C for 1h. The mixture was filtered through a bed of celite (top layer) and silica (bottom layer), eluting with CHCl<sub>3</sub>/MeOH affording crude product which was purified on Biotage SP1 (12g SINGLE StEP column, 15 mL/min, 0 – 5% MeOH in CH<sub>2</sub>Cl<sub>2</sub>) to afford the product as a brown oil (62 mg, 86%). <sup>1</sup>H NMR: (500 MHz, CDCl<sub>3</sub>)  $\delta_{\text{H}}$  8.60 (d, *J* = 2.0 Hz, 1H), 8.30 (s, 1H), 8.16 (d, *J* = 2.0 Hz, 1H), 7.83 (d, *J* = 0.9 Hz, 1H), 7.70 (d, *J* = 0.9 Hz, 1H), 5.80 (s, 2H), 4.01 (s, 3H), 3.79 – 3.73 (m, 2H), 3.57 – 3.52 (m, 2H), 3.37 (s, 3H); <sup>13</sup>C NMR: (126 MHz, CDCl<sub>3</sub>)  $\delta_{\text{C}}$  145.7, 144.5, 143.1, 136.9, 127.1, 124.9, 124.5, 120.3, 73.1, 71.6, 69.0, 59.1, 39.2; LCMS *t*<sub>R</sub> = 2.06 mins, *m/z* = 288 (M+H)<sup>+</sup>; Purity (AUC)  $\geq$  95%; HRMS (M+H)<sup>+</sup> *calculated* for C<sub>14</sub>H<sub>18</sub>N<sub>5</sub>O<sub>2</sub> = 288.1455, *found* = 288.1459.

**3-((2-Methoxyethoxy)methyl)-*N,N*-dimethyl-3H-imidazo[4,5-*b*]pyridine-**

**6-carboxamide, S7:** 6-Bromo-3-((2-methoxyethoxy)methyl)-3H-imidazo[4,5-*b*]pyridine, **7a** (1.0M in DMF, 0.25 mL, 0.25 mmol), dimethylamine (2.0M in THF, 625  $\mu$ L, 1.25 mmol), Herrmann's Palladacycle (6 mg, 0.00625 mmol), [(*t*-Bu)<sub>3</sub>PH]BF<sub>4</sub> (5 mg, 0.0175 mmol), Mo(CO)<sub>6</sub> (66mg, 0.25 mmol) and DBU (1.0M in THF, 0.75 mL, 0.75 mmol) were combined in 1,4-dioxane (2.5 mL) and irradiated in  $\mu$ W at 140 °C for 1 h. The reaction mixture was filtered through a bed of celite (top layer) and silica (bottom layer) with CHCl<sub>3</sub>/MeOH to afford crude product. Purification using Biotage SP1 (10g SNAP, 15 mL/min, 0 – 5% MeOH in CH<sub>2</sub>Cl<sub>2</sub>) afforded the product as a brown oil (60mg, 86%). <sup>1</sup>H NMR: (500 MHz, CDCl<sub>3</sub>)  $\delta_{\text{H}}$  8.57 (d, *J* = 1.9 Hz, 1H), 8.32 (s, 1H), 8.18 (d, *J* = 1.9 Hz, 1H), 5.80 (s, 2H), 3.78 – 3.70 (m, 2H), 3.56 – 3.51 (m, 2H), 3.36 (s, 3H), 3.19 (bs, 3H), 3.09 (bs, 3H); <sup>13</sup>C NMR: (126 MHz, CDCl<sub>3</sub>)  $\delta_{\text{C}}$  169.6, 147.5, 145.6, 144.2, 134.3, 127.6, 127.2, 73.0, 71.5, 69.0, 59.1, 39.9, 35.7; LCMS *t*<sub>R</sub> = 1.62 mins, *m/z* = 279

(M+H)<sup>+</sup>; Purity (AUC) ≥ 95%; HRMS (M+H)<sup>+</sup> *calculated* for C<sub>13</sub>H<sub>19</sub>N<sub>4</sub>O<sub>3</sub> = 279.1452, *found* = 279.1455.

**3-((2-Methoxyethoxy)methyl)-6-methyl-3H-imidazo[4,5-*b*]pyridine, S8:**

6-Bromo-3-((2-methoxyethoxy)methyl)-3H-imidazo[4,5-*b*]pyridine, **7a** (589 mg, 2.0 mmol), methylboronic acid (239 mg, 4.0 mmol), CsF (636 mg, 4.0 mmol), Pd(OAc)<sub>2</sub> (18 mg, 4 mol%) and nBuPAd<sub>2</sub> (57 mg, 8 mol%) were dissolved in DME (5.0 ml) and heated in a microwave reactor at 150 °C for 1 h. The crude mixture was concentrated *in vacuo* and purified by column chromatography (CH<sub>2</sub>Cl<sub>2</sub> to EtOH:CH<sub>2</sub>Cl<sub>2</sub> 10:90) to afford the product as a yellow oil (354 mg, 80%); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ<sub>H</sub> 8.27 (d, *J* = 1.8 Hz, 1H), 8.19 (s, 1H), 7.90 (d, *J* = 1.8 Hz, 1H), 5.75 (s, 2H), 3.72 – 3.70 (m, 2H), 3.52 – 3.50 (m, 2H), 3.36 (s, 3H), 2.50 (s, 3H); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ<sub>C</sub> 145.6, 145.5, 144.2, 135.1, 128.4, 128.1, 72.9, 71.5, 68.8, 59.0, 18.6; LCMS *t<sub>R</sub>* = 1.62 min, *m/z* = 222 (M+H)<sup>+</sup>; Purity (AUC) ≥ 95%; HRMS (M+H)<sup>+</sup> *calculated* for C<sub>11</sub>H<sub>16</sub>N<sub>3</sub>O<sub>2</sub> = 222.1237, *found* = 222.1232.

**6-Cyclopropyl-3-((2-methoxyethoxy)methyl)-3H-imidazo[4,5-*b*]pyridine, S9:**

**S9:** 6-Bromo-3-((2-methoxyethoxy)methyl)-3H-imidazo[4,5-*b*]pyridine, **7a** (589 mg, 2.0 mmol), cyclopropylboronic acid (344 mg, 4.0 mmol), CsF (636 mg, 4.0 mmol), Pd(OAc)<sub>2</sub> (18 mg, 4 mol%) and nBuPAd<sub>2</sub> (57 mg, 8 mol%) were dissolved in DME (5.0 ml) and heated in a microwave reactor at 150 °C for 1 h. The crude mixture was concentrated *in vacuo* and purified by column chromatography (CH<sub>2</sub>Cl<sub>2</sub> to EtOH: CH<sub>2</sub>Cl<sub>2</sub> 10:90) to afford the product as a yellow oil (440 mg, 89%); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ<sub>H</sub> 8.36 (d, *J* = 2.0 Hz, 1H), 8.08 (s, 1H), 7.45 (d, *J* = 2.0 Hz, 1H), 5.57 (s, 2H), 3.54 – 3.48 (m, 2H), 3.47 – 3.42 (m, 2H), 3.28 (s, 3H), 2.00 (ddd, *J* = 8.5, 5.2, 3.4 Hz, 1H), 1.04 – 0.96 (m, 2H), 0.70 (dt, *J* = 6.4, 4.8 Hz, 2H); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ<sub>C</sub> 155.1, 145.0, 144.6, 134.9, 125.9, 115.1, 75.4, 71.7, 67.7, 59.0, 13.4, 9.1; LCMS *t<sub>R</sub>* = 1.95 min, *m/z* = 248 (M+H)<sup>+</sup>; Purity (AUC) ≥ 95%; HRMS (M+H)<sup>+</sup> *calculated* for C<sub>13</sub>H<sub>18</sub>N<sub>3</sub>O<sub>2</sub> = 248.1394, *found* = 248.1394.

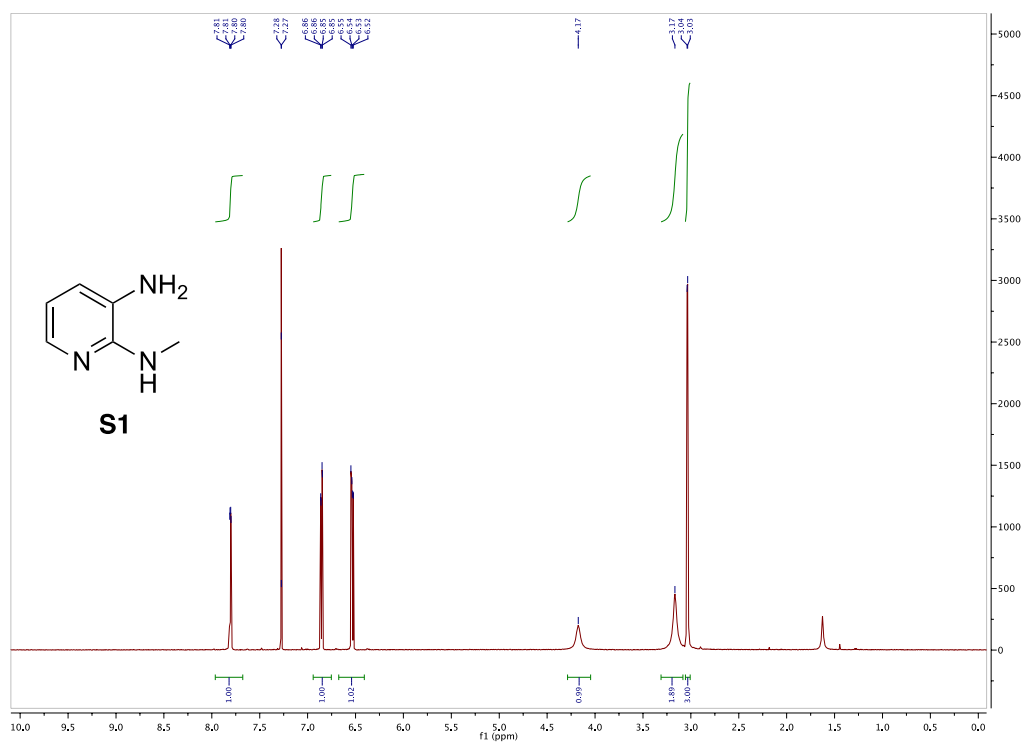
**3-((2-Methoxyethoxy)methyl)-7-methyl-3H-imidazo[4,5-*b*]pyridine, S10:**

7-Chloro-3-((2-methoxyethoxy)methyl)-3H-imidazo[4,5-*b*]pyridine, **8a** (1.0M in DMF, 0.2 mL, 0.2 mmol), methylboronic acid (18 mg, 0.3 mmol), CsF (60 mg, 0.4 mmol), Pd(OAc)<sub>2</sub> (2.2 mg, 5 mol%) and nBuPAd<sub>2</sub> (7.2 mg, 10 mol%) were dissolved in DME (1.0 ml) and heated in a microwave reactor at 150 °C for 2.5 h. The reaction mixture was filtered through a plug of silica (bottom layer) and celite (top layer), eluting with CHCl<sub>3</sub> & MeOH. The filtrate was concentrated *in vacuo* and purified by column chromatography (0 – 5% MeOH in CH<sub>2</sub>Cl<sub>2</sub>) to afford the product as a yellow oil (35 mg, 80%). <sup>1</sup>H NMR: (500 MHz, CDCl<sub>3</sub>) δ<sub>H</sub> 8.32 (d, *J* = 4.9 Hz, 1H), 8.29 (s, 1H), 7.12 (dd, *J* = 4.9 0.8 Hz, 1H), 5.78 (s, 2H), 3.75 – 3.71 (m, 2H), 3.54 – 3.50 (m, 2H), 3.35 (s, 3H), 2.73 (d, *J* = 0.8 Hz, 3H); <sup>13</sup>C NMR: (126 MHz, CDCl<sub>3</sub>) δ<sub>C</sub> 146.2, 144.8, 142.9, 139.9, 134.2, 120.1, 73.2, 71.5, 68.9, 59.1, 16.3; LCMS *t*<sub>R</sub> = 1.96 mins, *m/z* = 222 (M+H)<sup>+</sup>; Purity (AUC) ≥ 95%; HRMS (M+H)<sup>+</sup> *calculated* for C<sub>11</sub>H<sub>16</sub>N<sub>3</sub>O<sub>2</sub> = 222.1237, *found* = 222.1240.

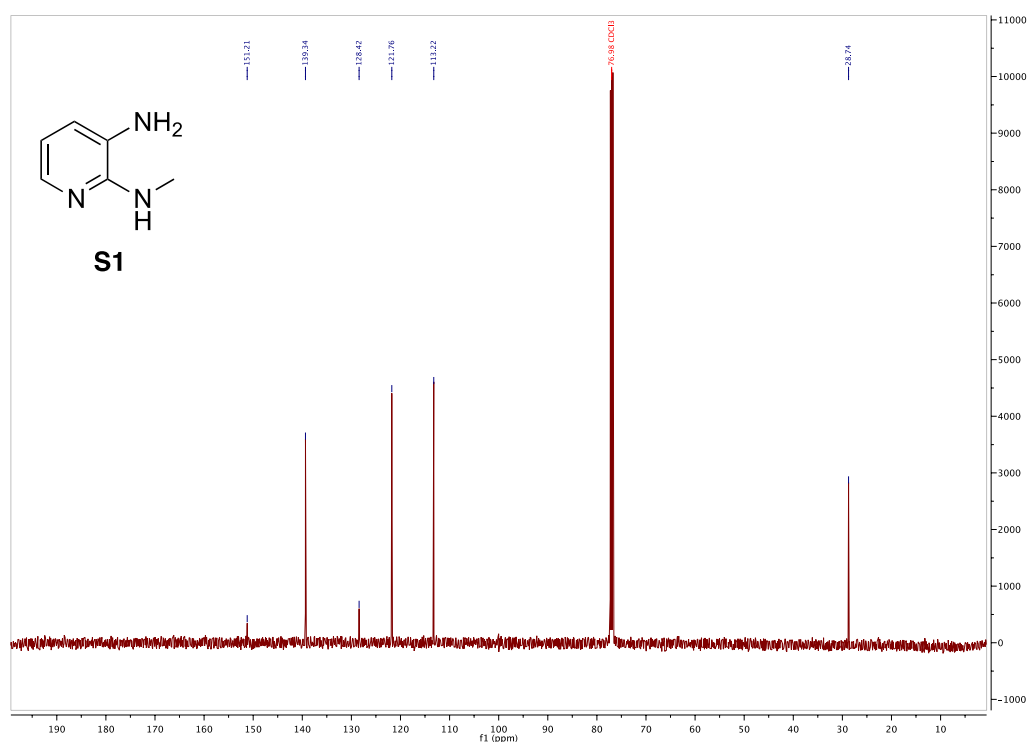
## 5. NMR Spectra

### 5.1 Spectra of intermediates

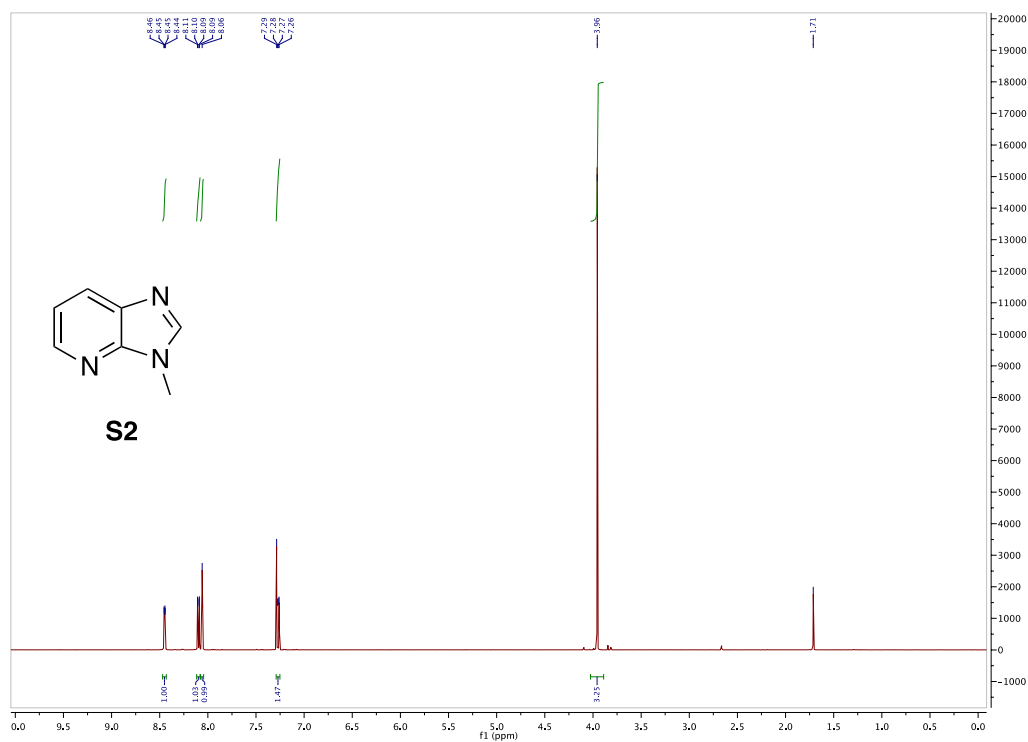
$^1\text{H}$  NMR:



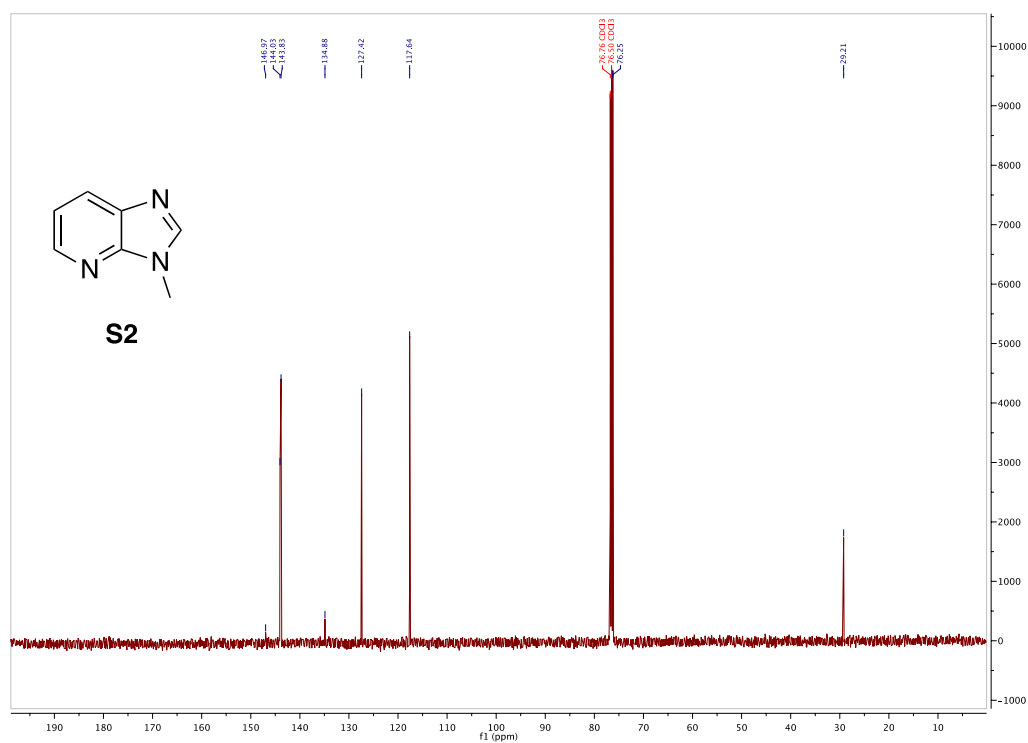
$^{13}\text{C}$  NMR:



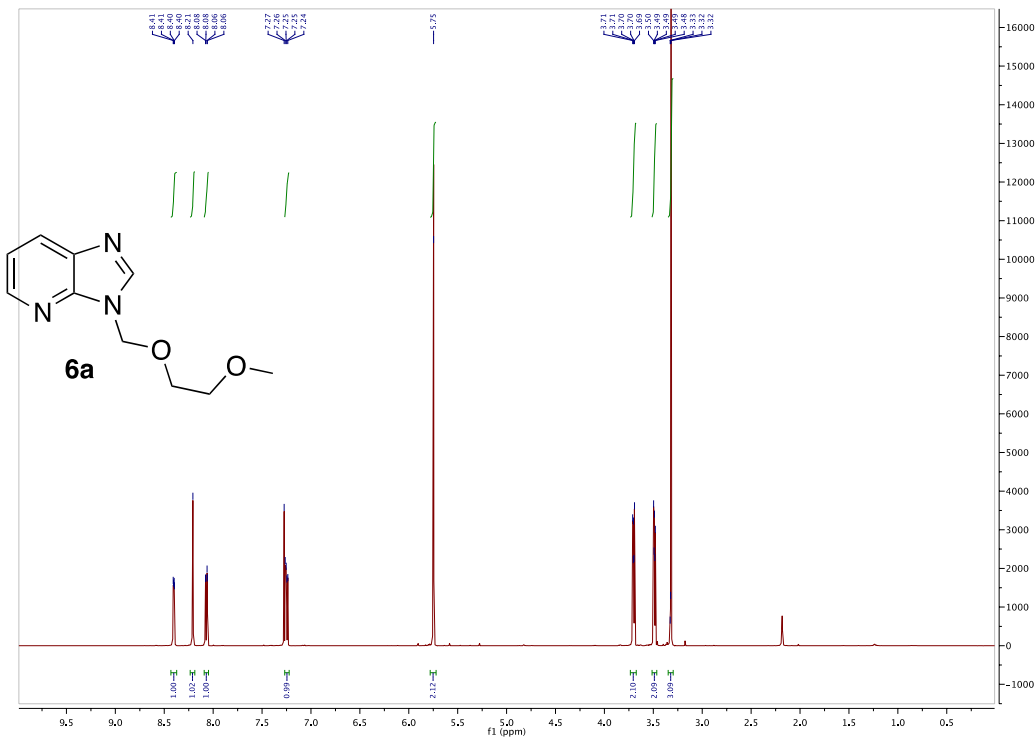
<sup>1</sup>H NMR:



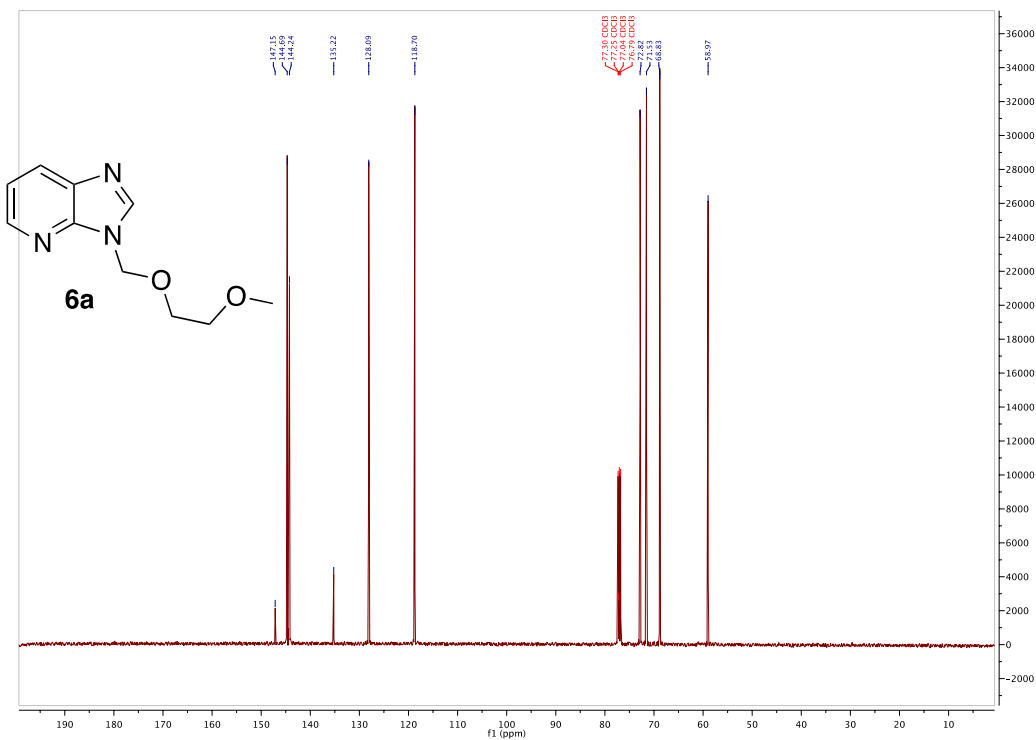
<sup>13</sup>C NMR:



<sup>1</sup>H NMR:



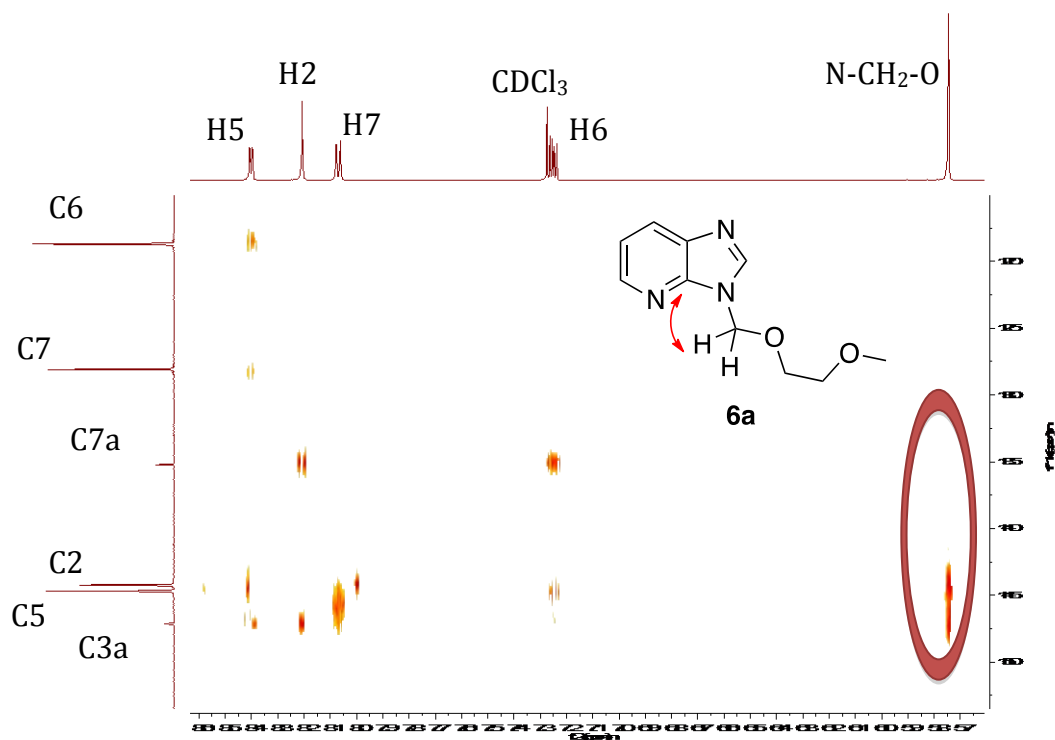
<sup>13</sup>C NMR:



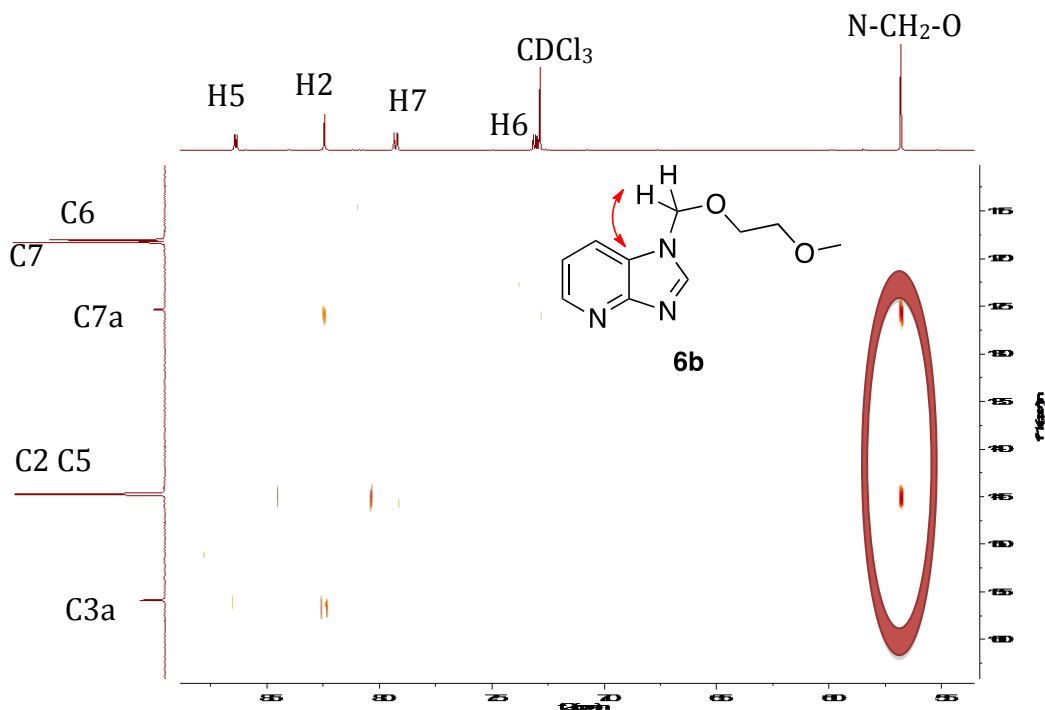


## 2D NMR – HMBC Experiment

**N3 MEM, 4a** – N-CH<sub>2</sub>-O correlates to C3a and C2 and not C7a.

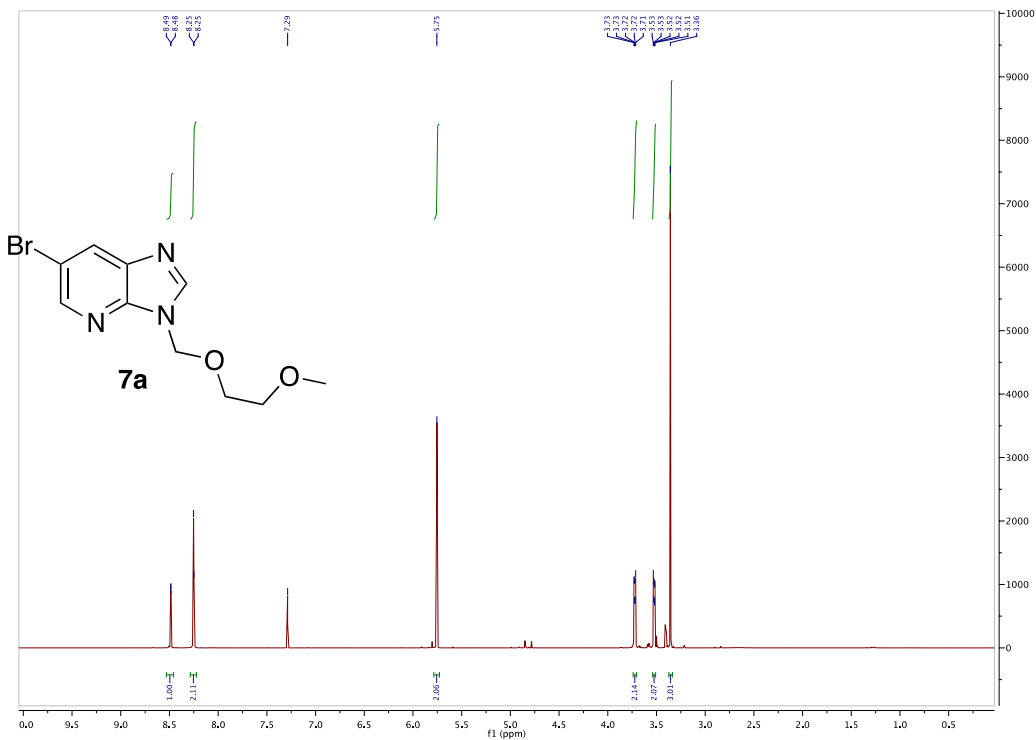


**N1 MEM, 4b** – N-CH<sub>2</sub>-O correlates to C7a and C2 and not C3a.

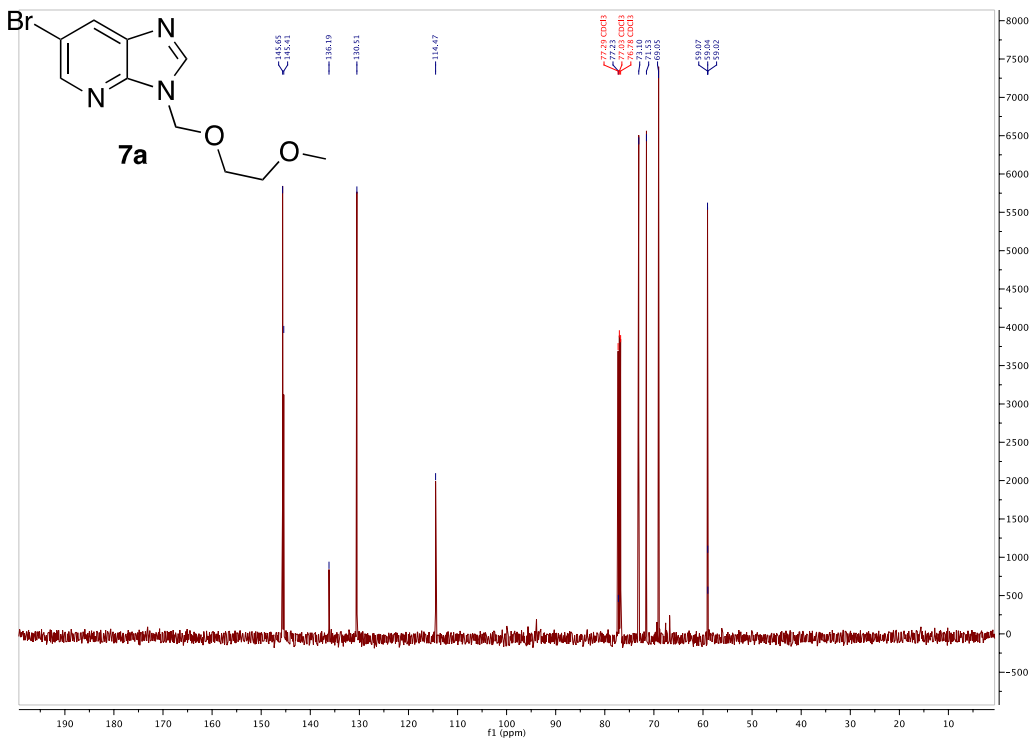




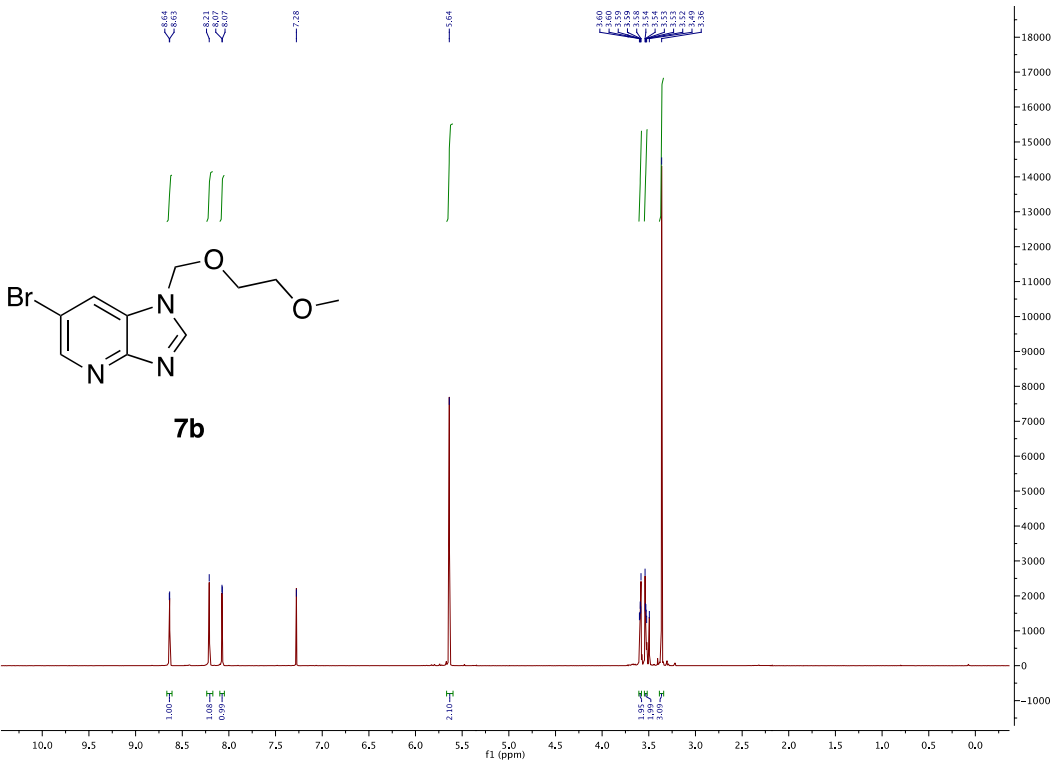
<sup>1</sup>H NMR:



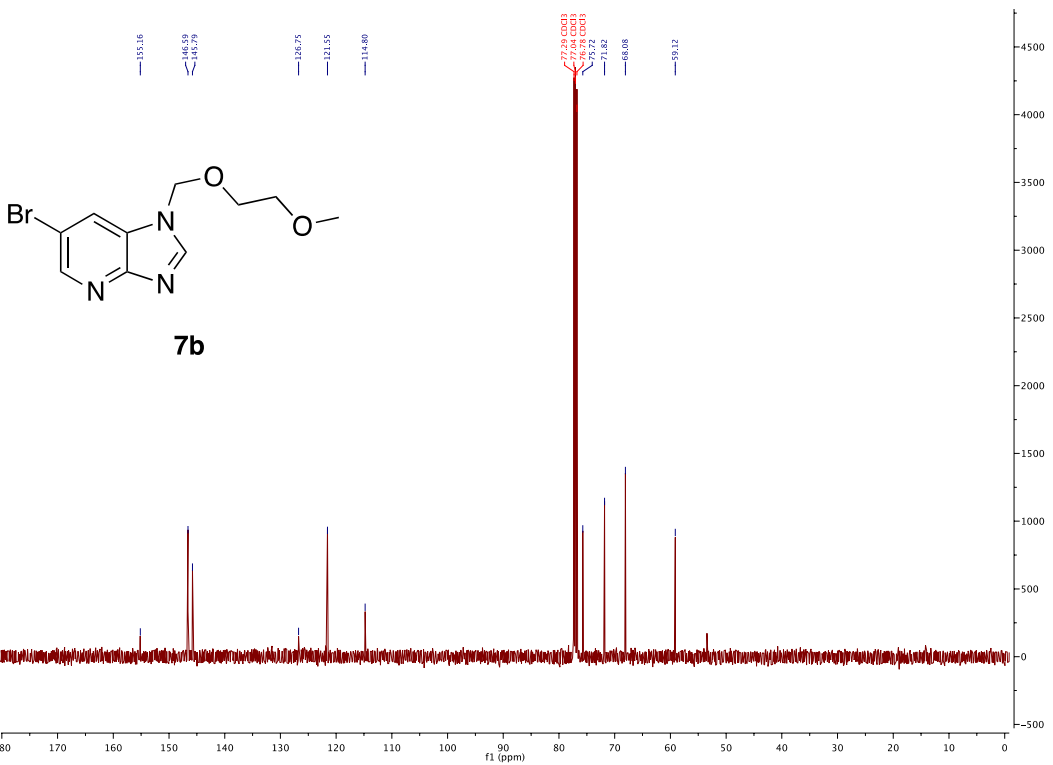
<sup>13</sup>C NMR:



<sup>1</sup>H NMR:

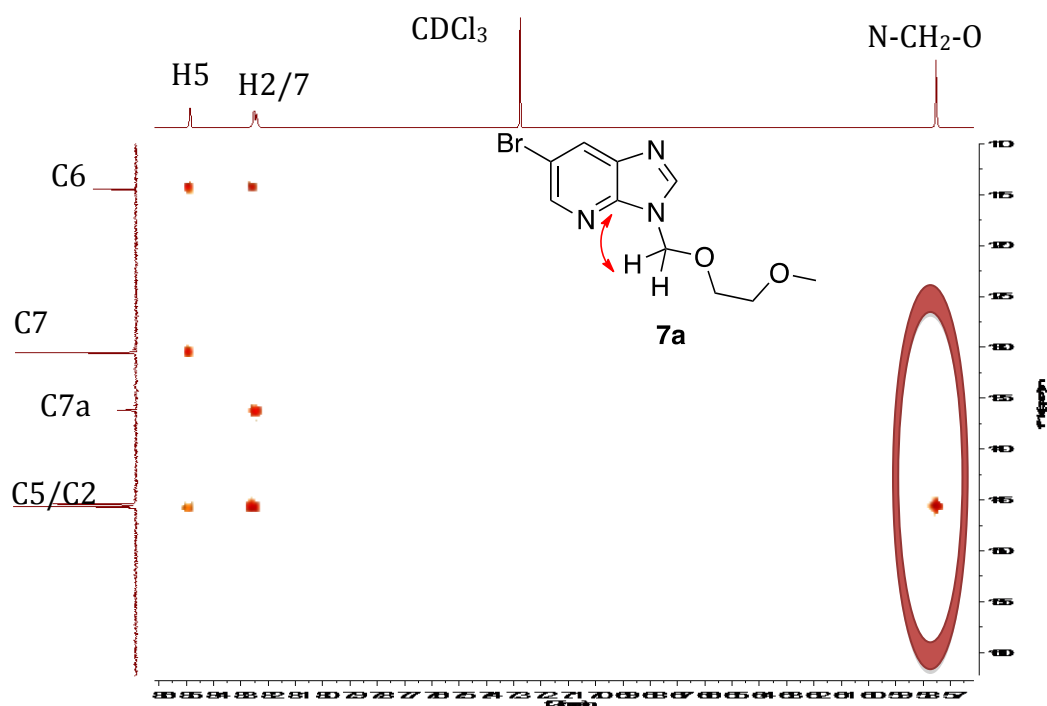


<sup>13</sup>C NMR:

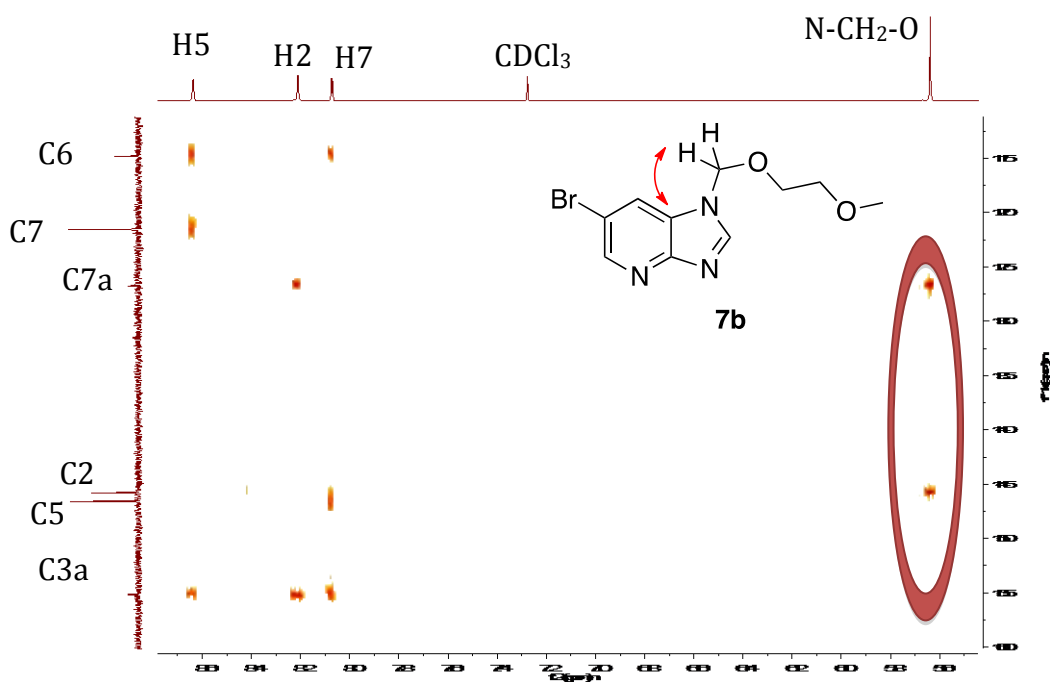


## 2D NMR – HMBC Experiment

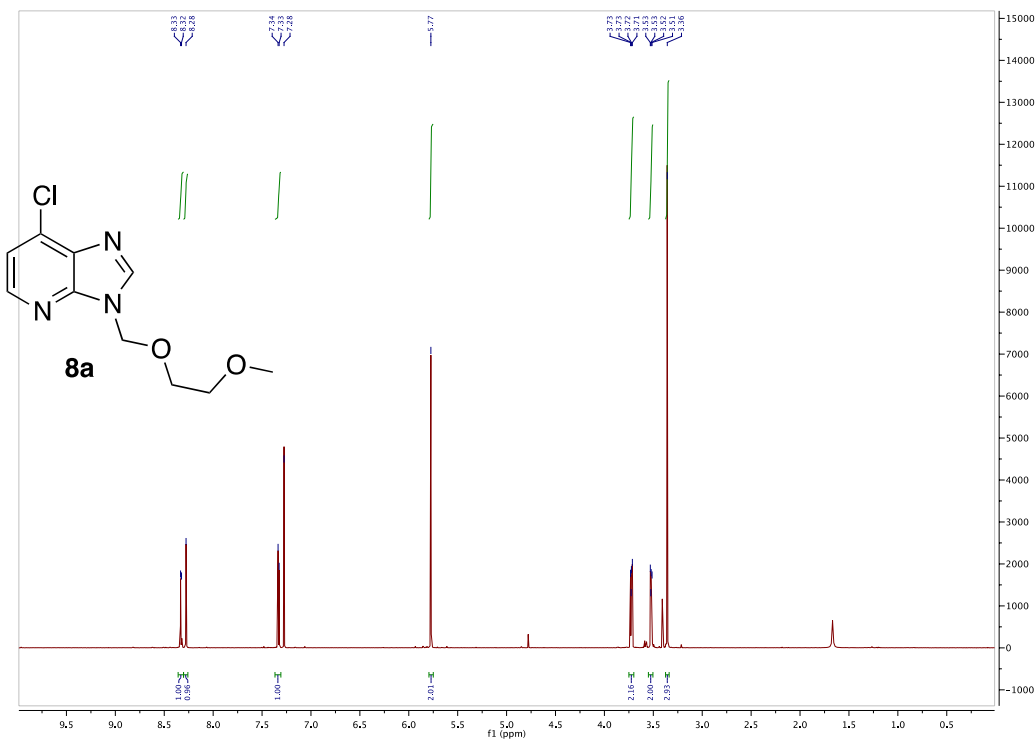
**6-Br N3 MEM, 5a** – N-CH<sub>2</sub>-O does not show correlation to either quaternary C, but it does to C2 - inconclusive.



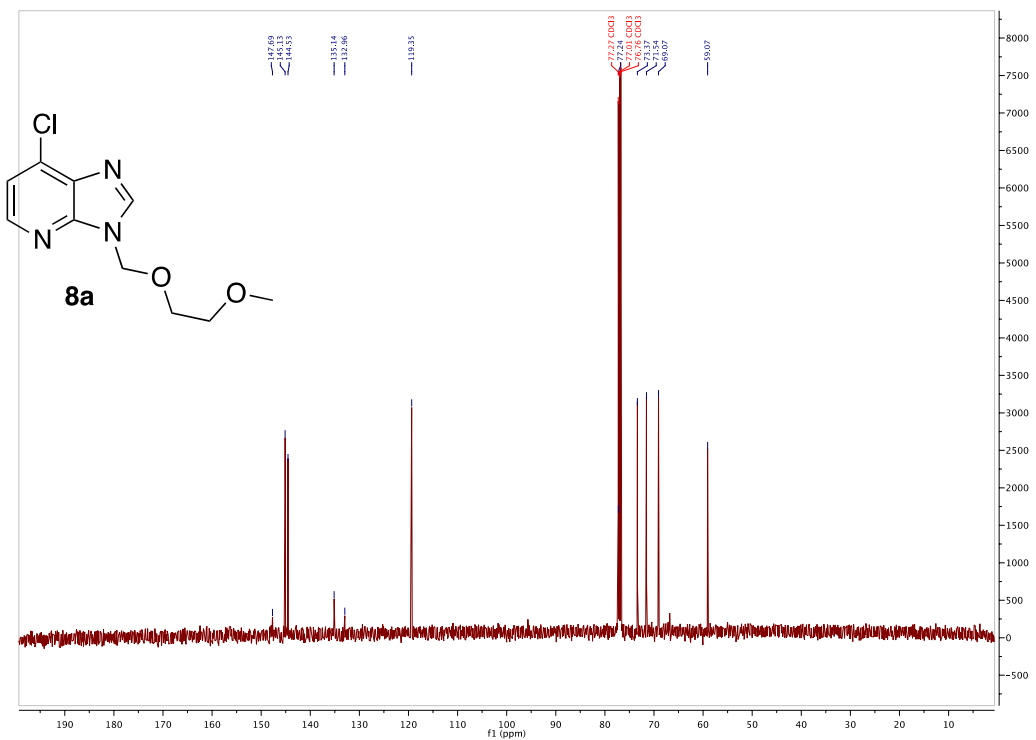
**6-Br N1 MEM, 5b** – However, the N1 regioisomer is easily distinguished as the N-CH<sub>2</sub>-O correlates to C7a and C2 and not C3a.



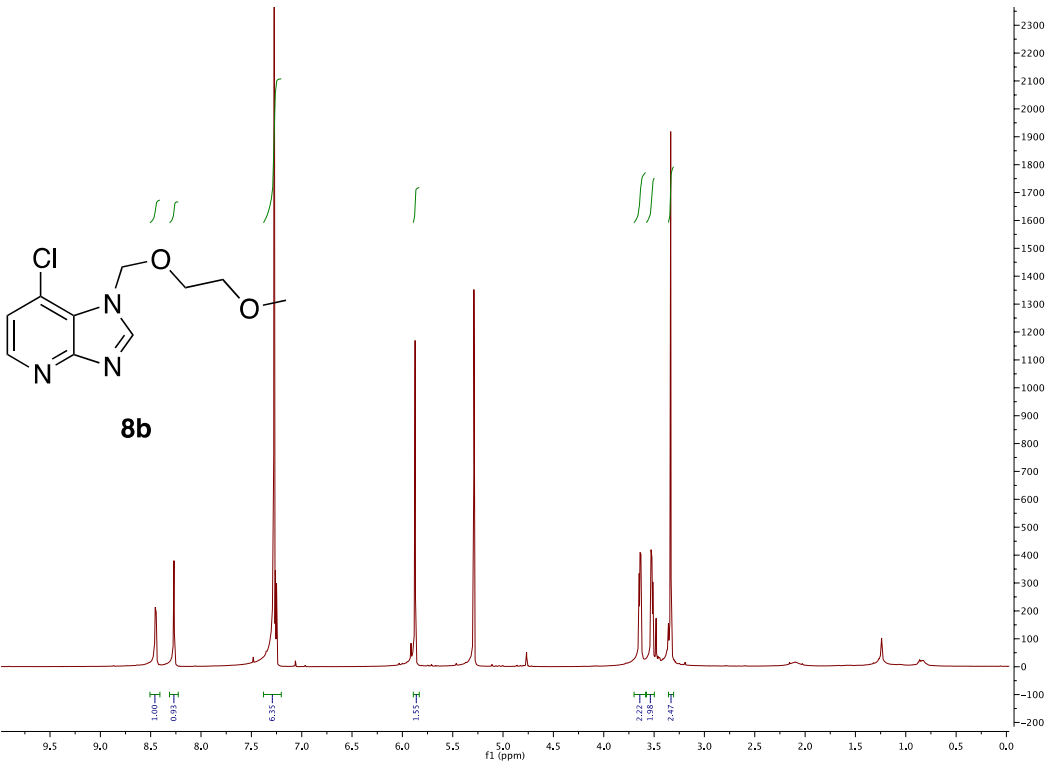
<sup>1</sup>H NMR:



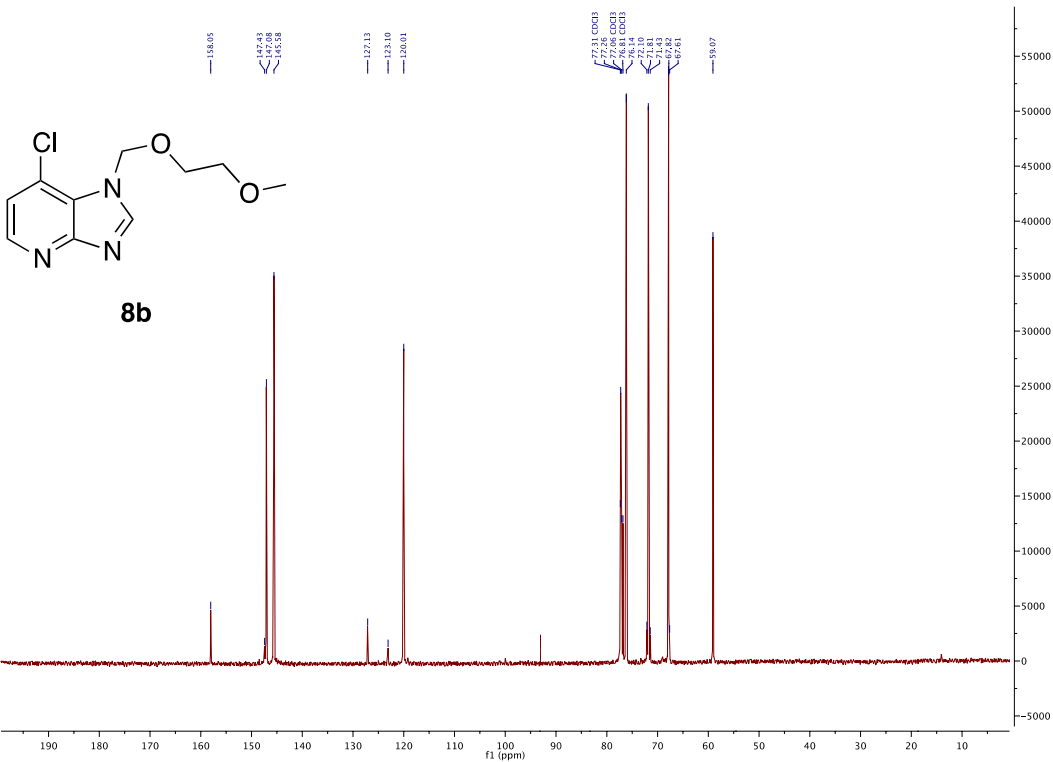
<sup>13</sup>C NMR:



<sup>1</sup>H NMR:

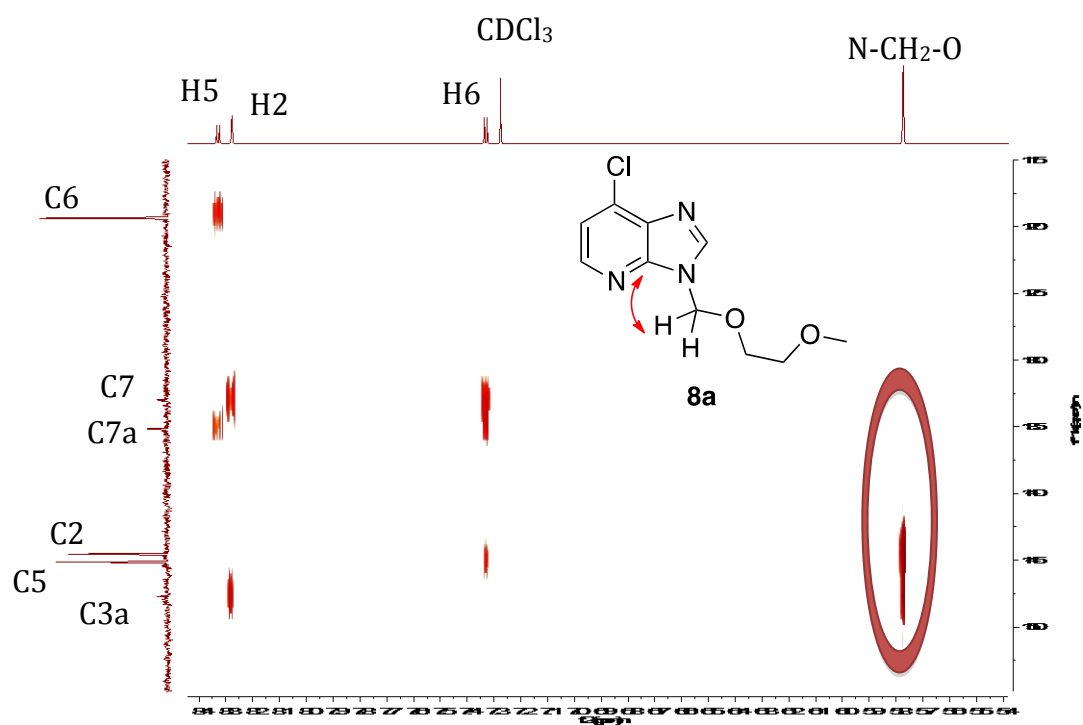


<sup>13</sup>C NMR:

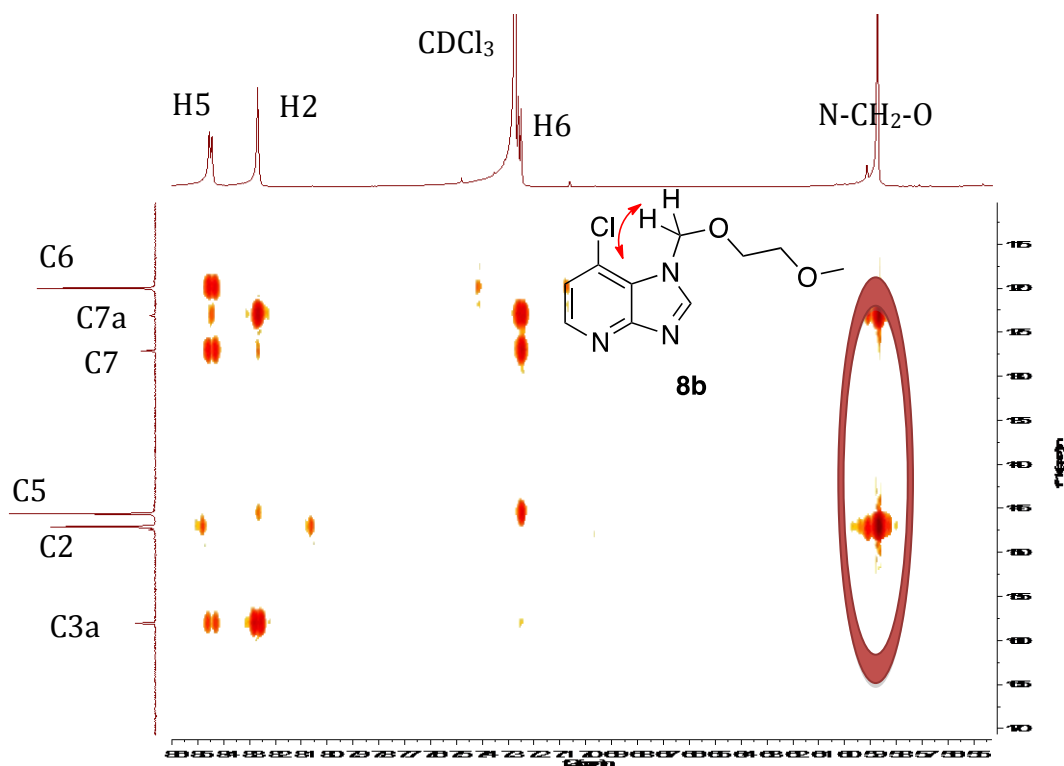


## 2D NMR – HMBC Experiment

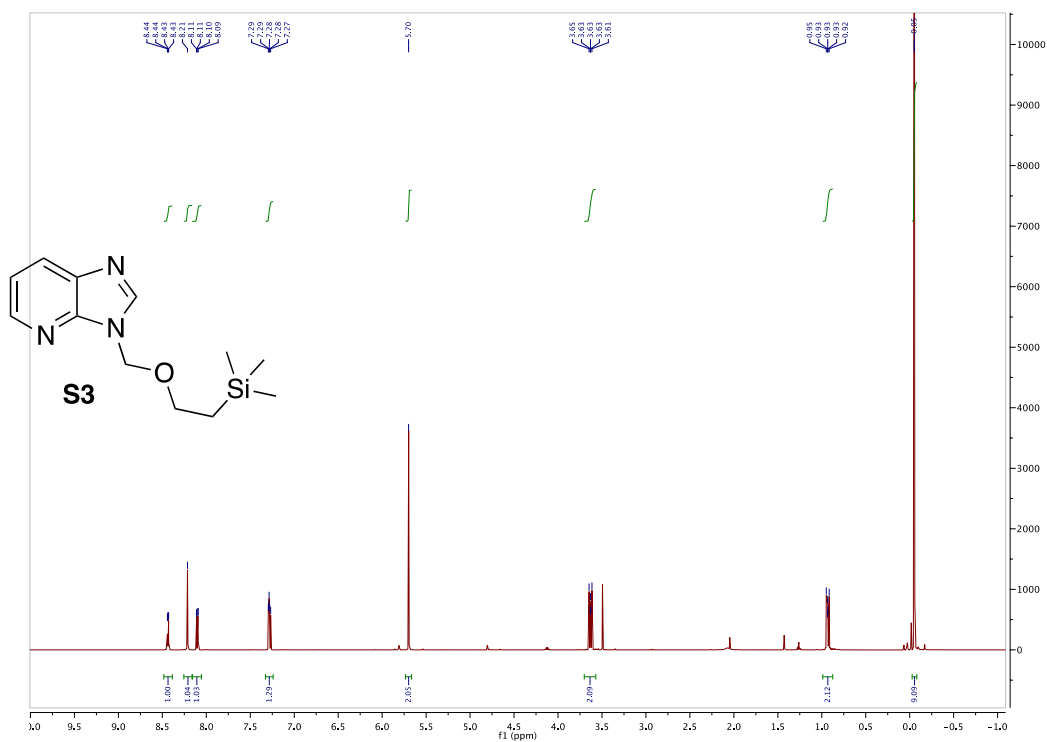
**7-Cl N3 MEM, 6a** – N-CH<sub>2</sub>-O correlates to C3a and C2 and not C7a.



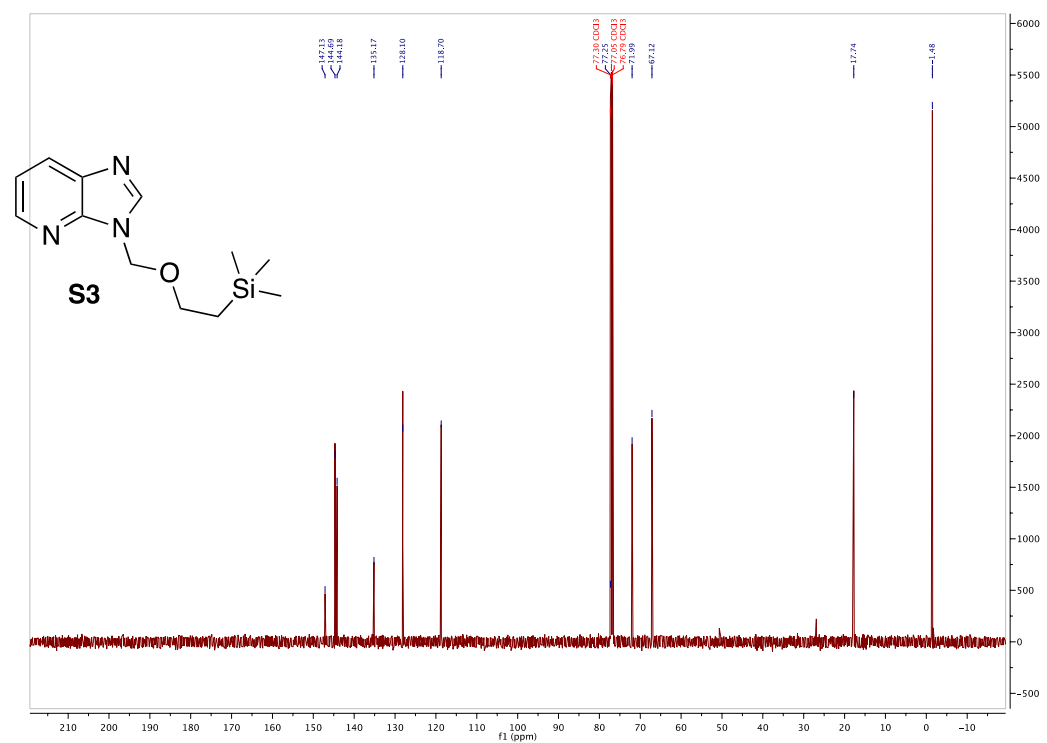
**7-Cl N1 MEM, 6b** – N-CH<sub>2</sub>-O correlates to C7a and C2 and not C3a.



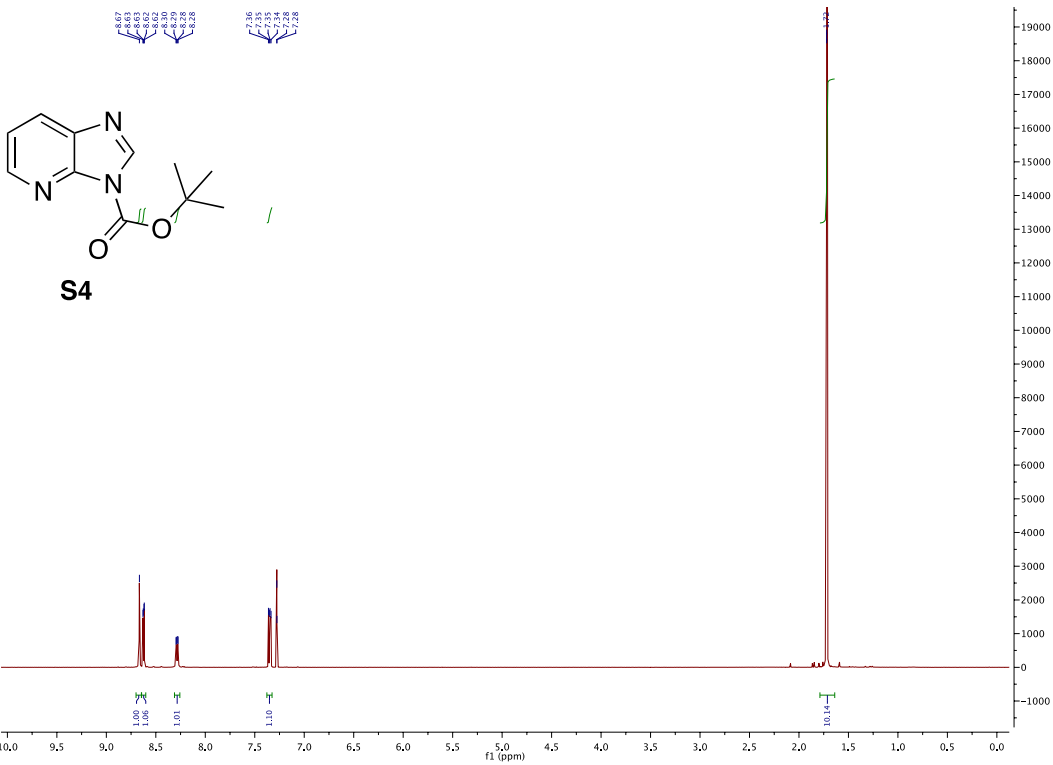
<sup>1</sup>H NMR:



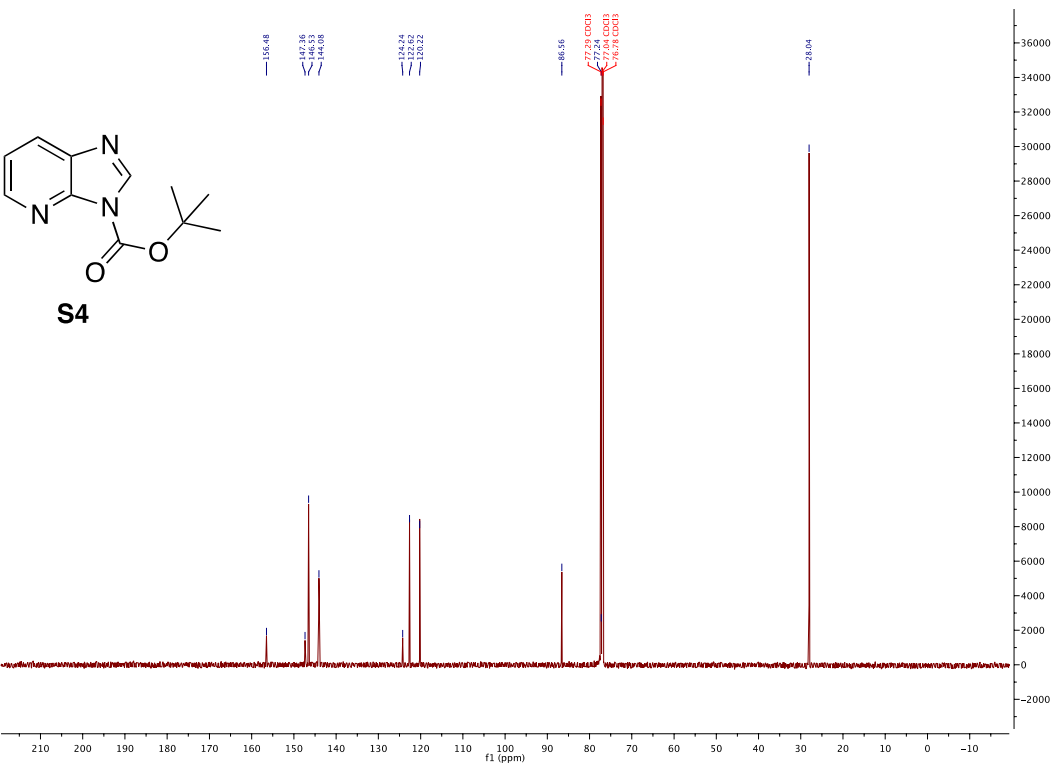
<sup>13</sup>C NMR:



<sup>1</sup>H NMR:

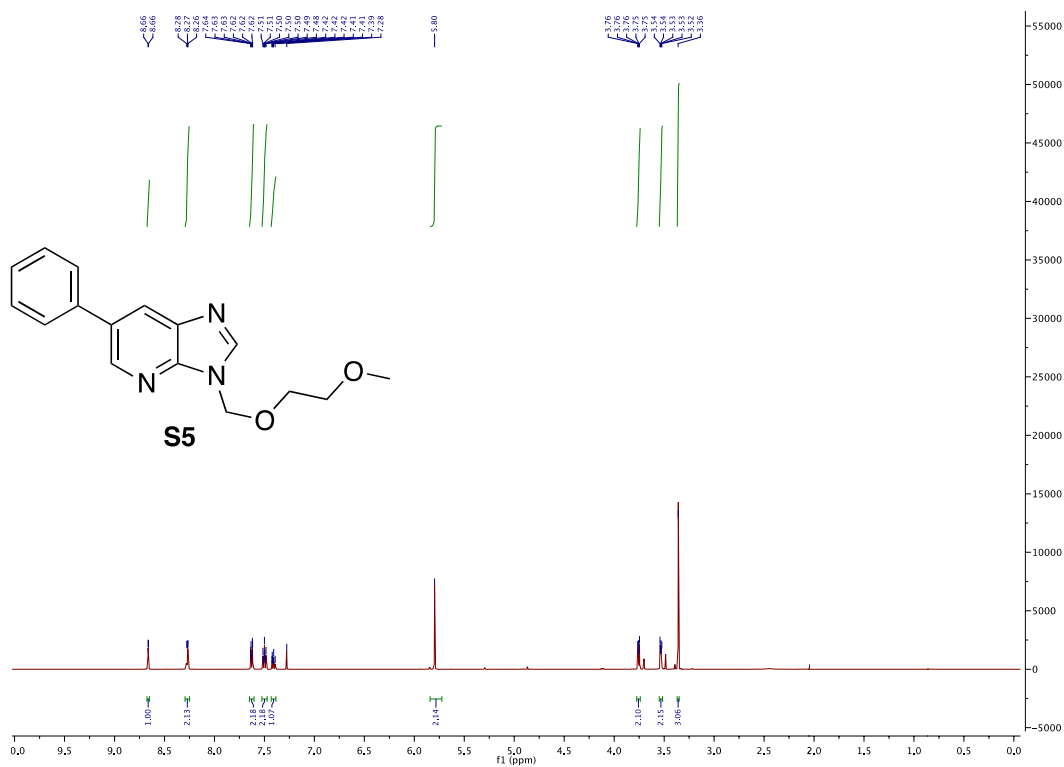


<sup>13</sup>C NMR:

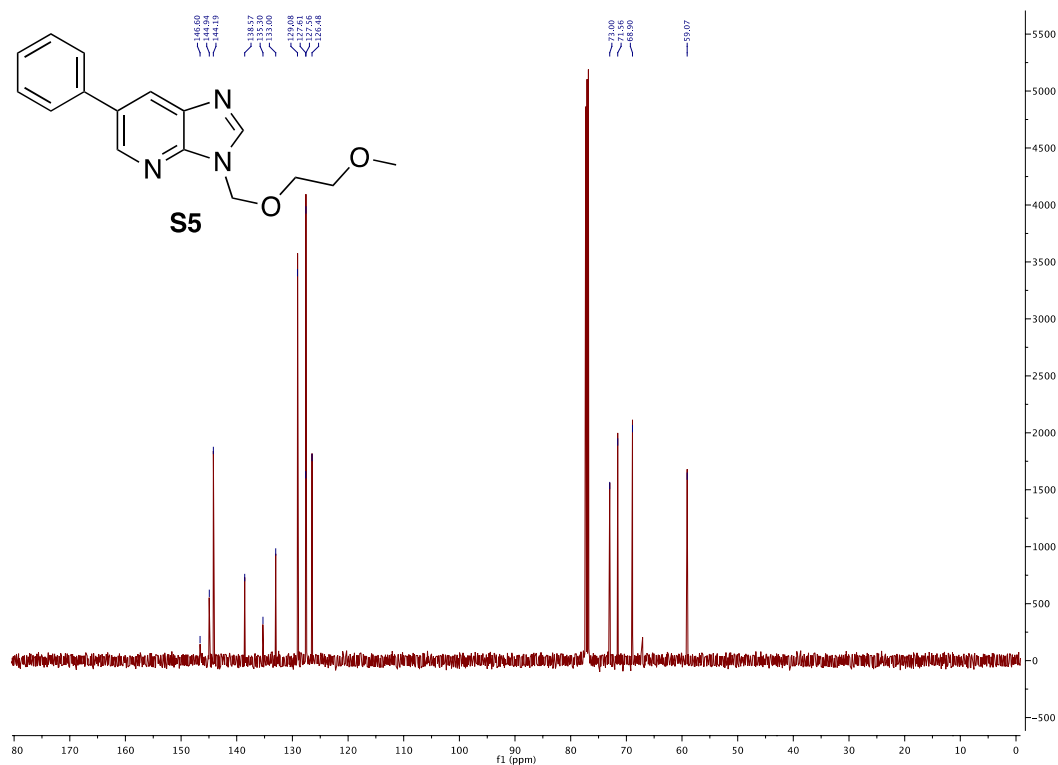




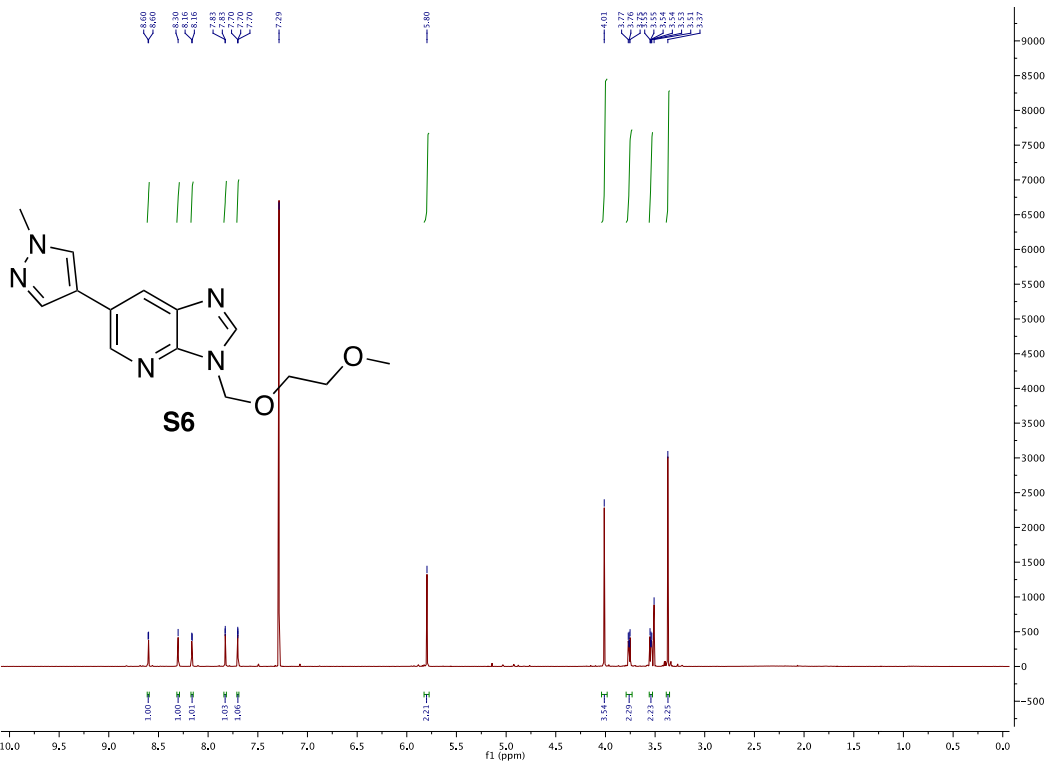
<sup>1</sup>H NMR:



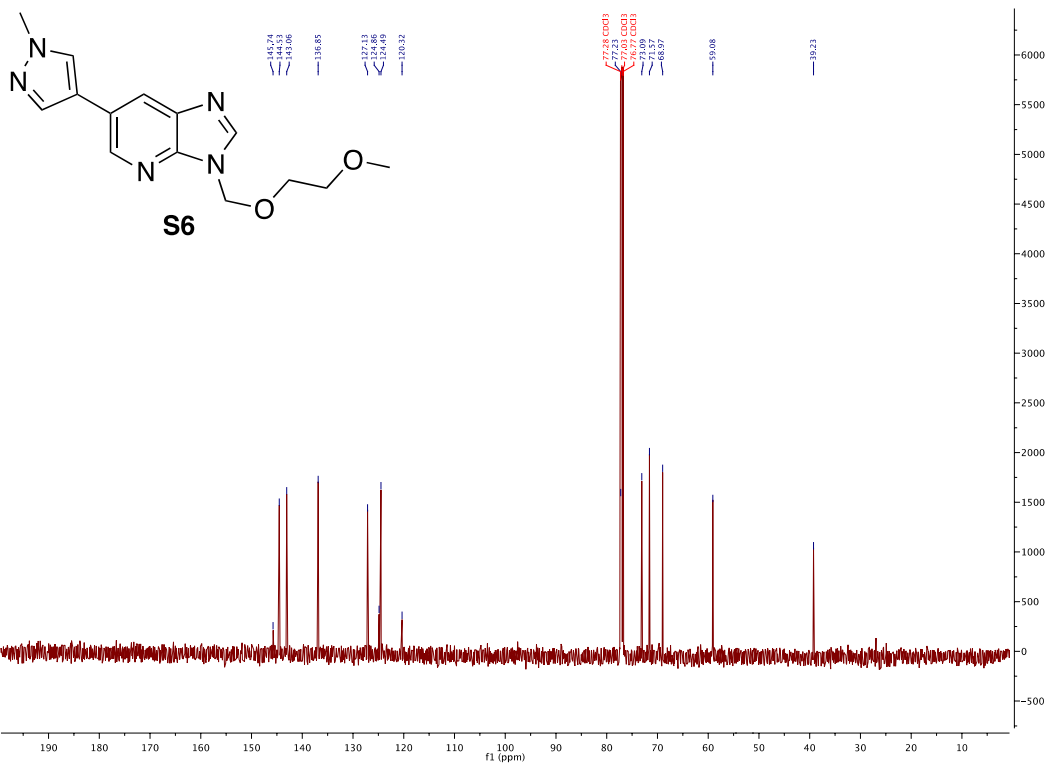
<sup>13</sup>C NMR:



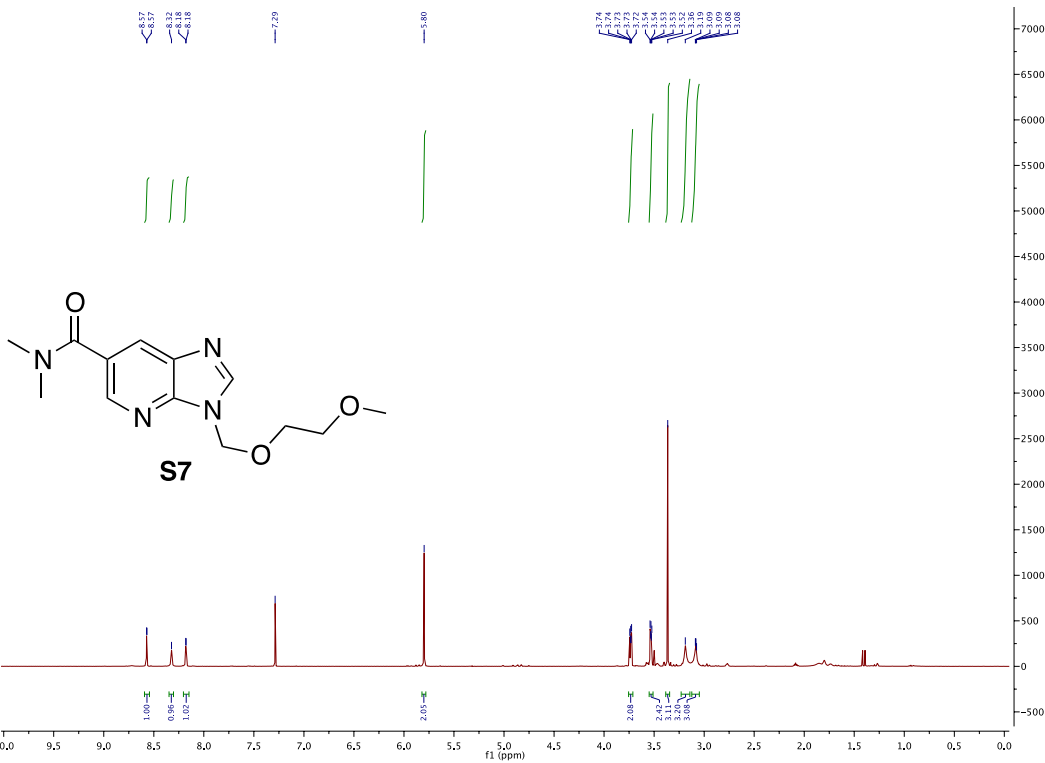
<sup>1</sup>H NMR:



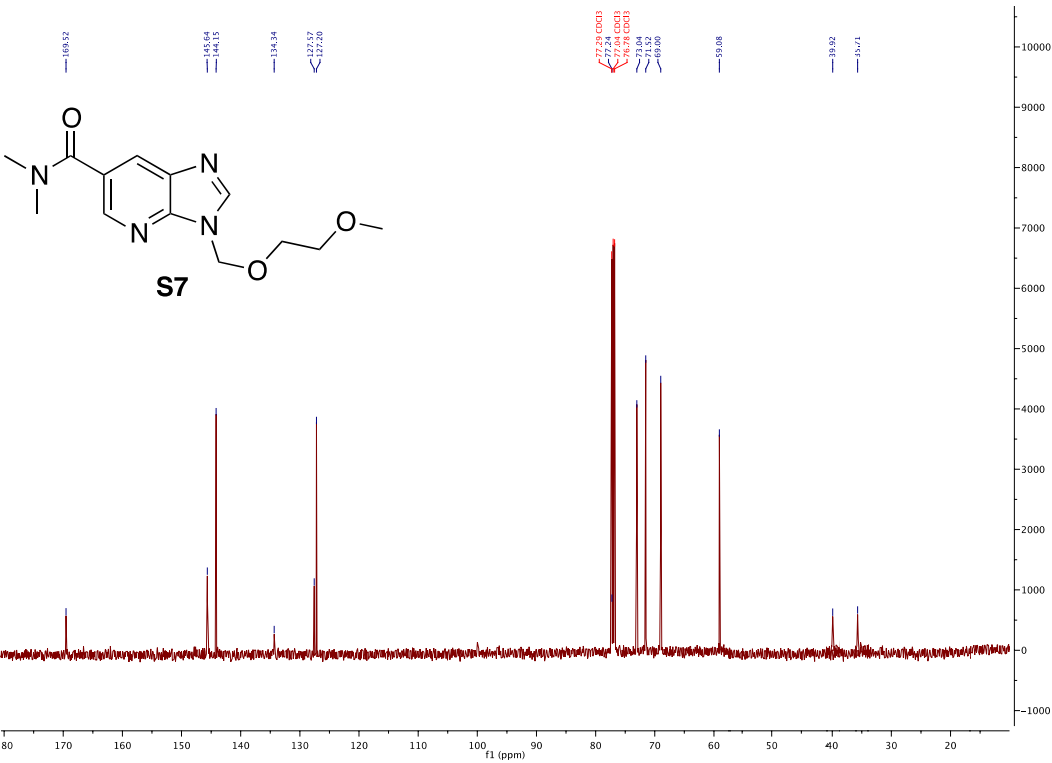
<sup>13</sup>C NMR:



<sup>1</sup>H NMR:



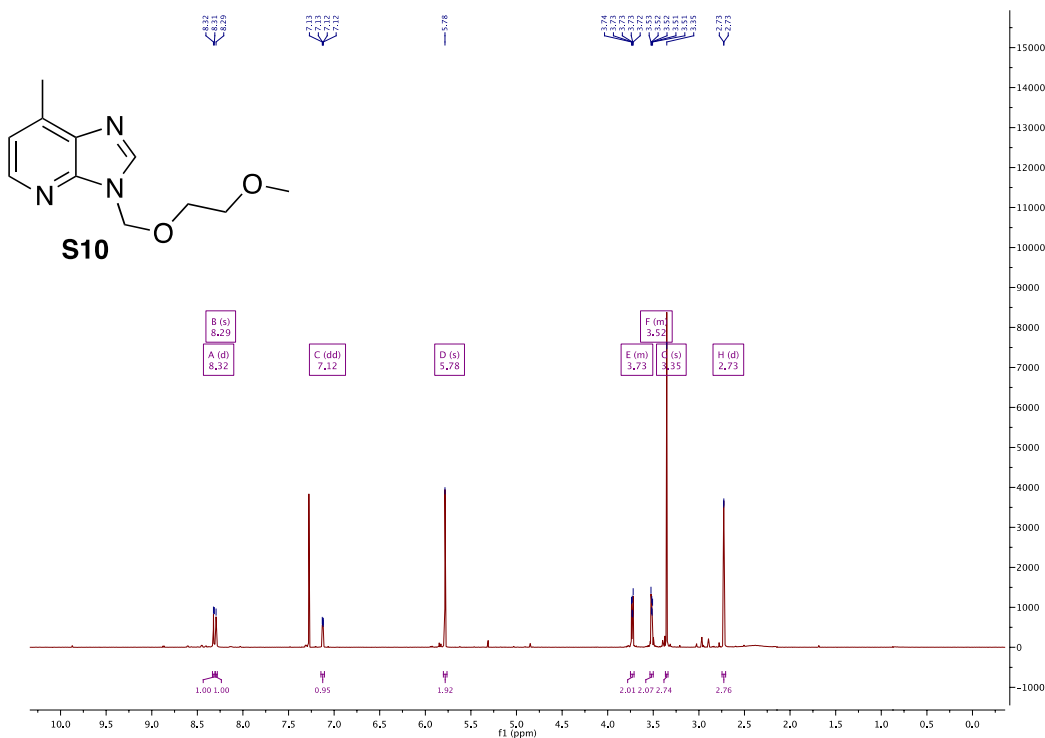
<sup>13</sup>C NMR:



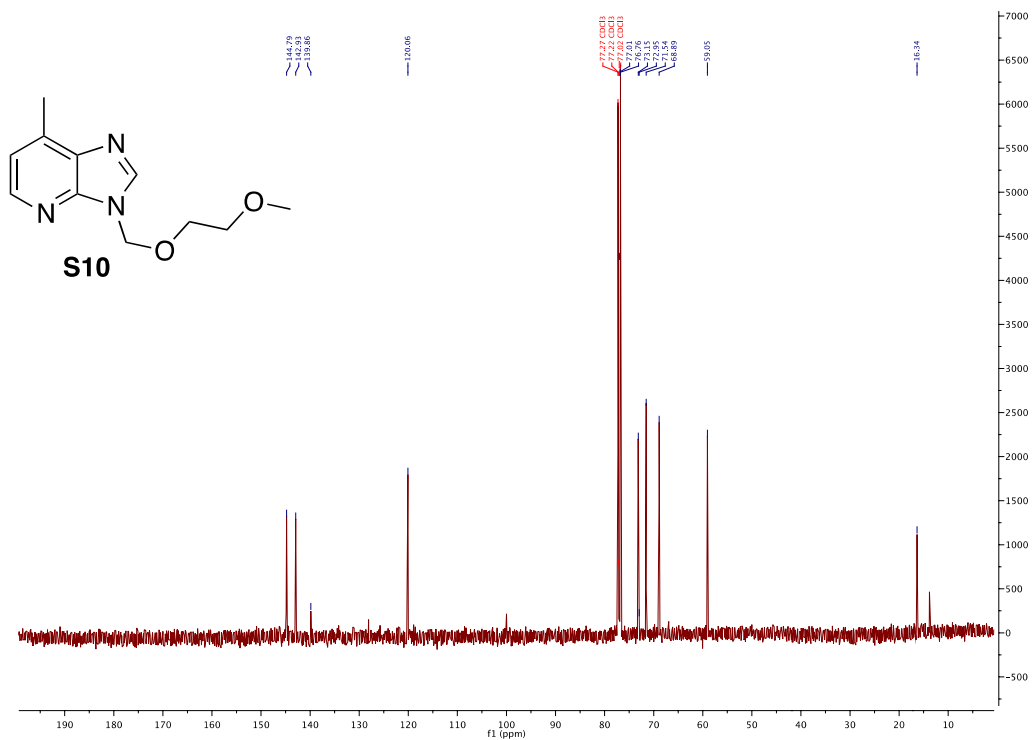




<sup>1</sup>H NMR:

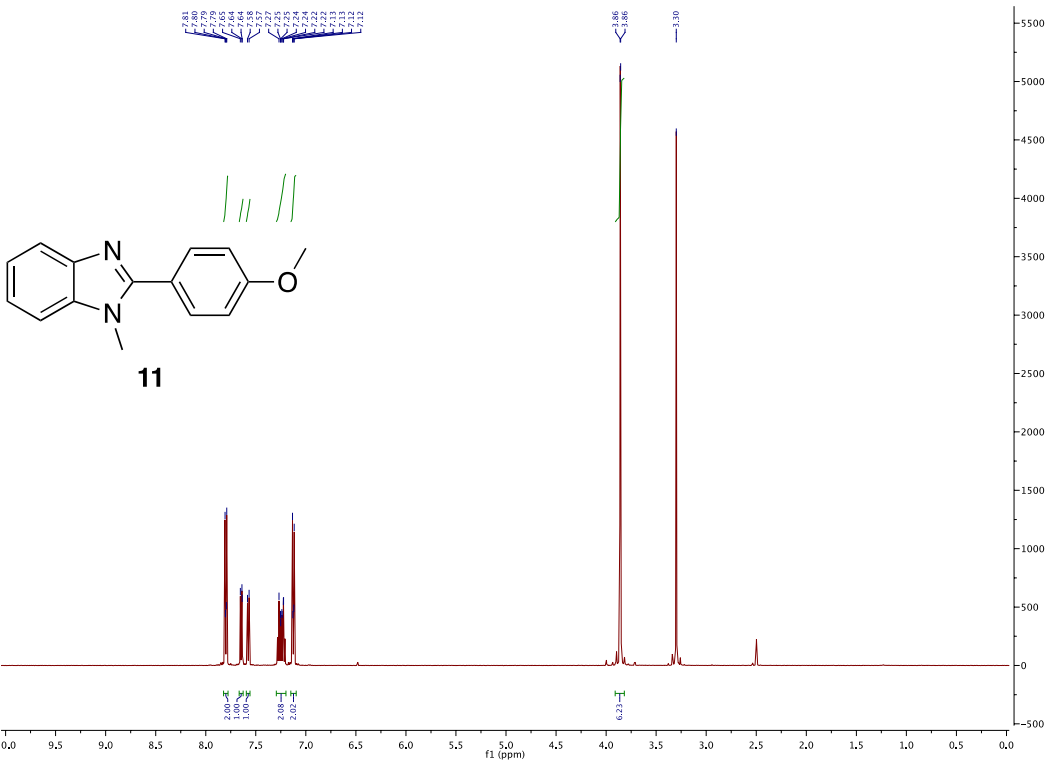


<sup>13</sup>C NMR:

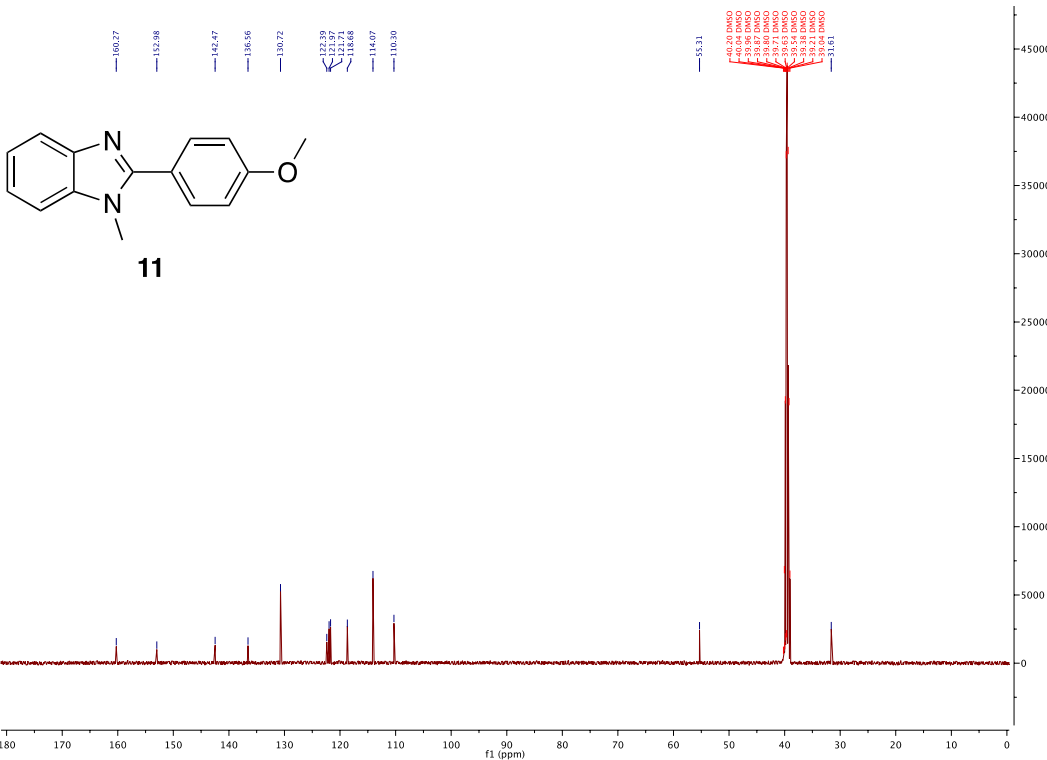


5.2 Spectra of C-H arylation products

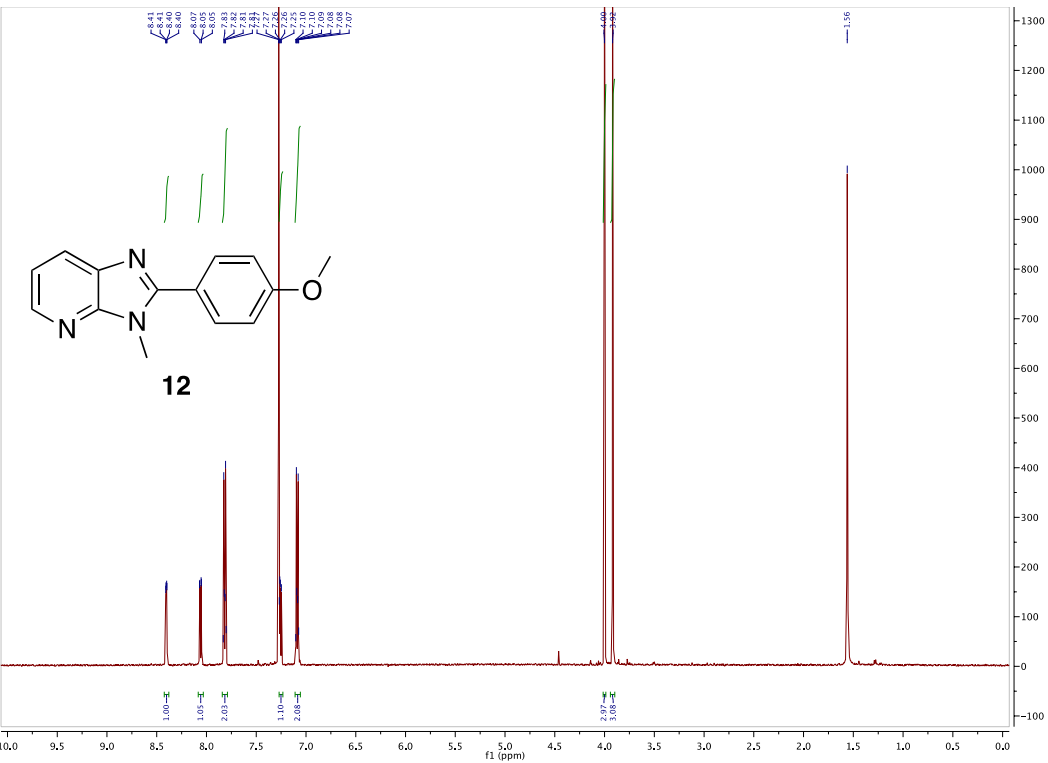
<sup>1</sup>H NMR:



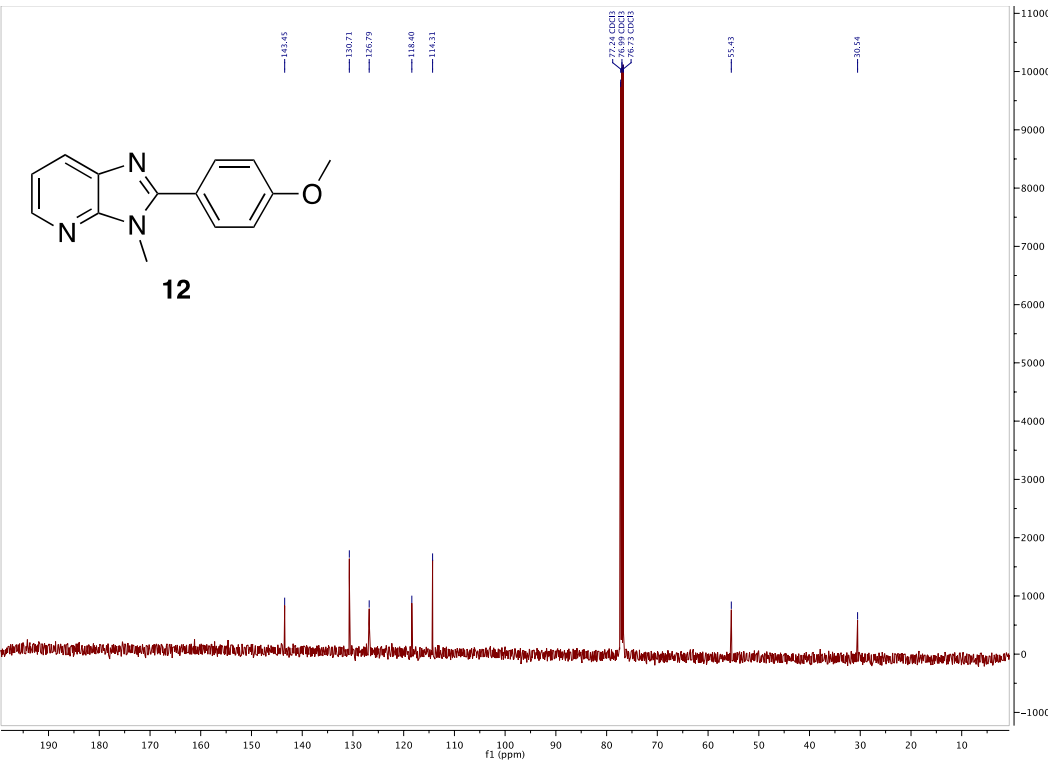
<sup>13</sup>C NMR:



<sup>1</sup>H NMR:

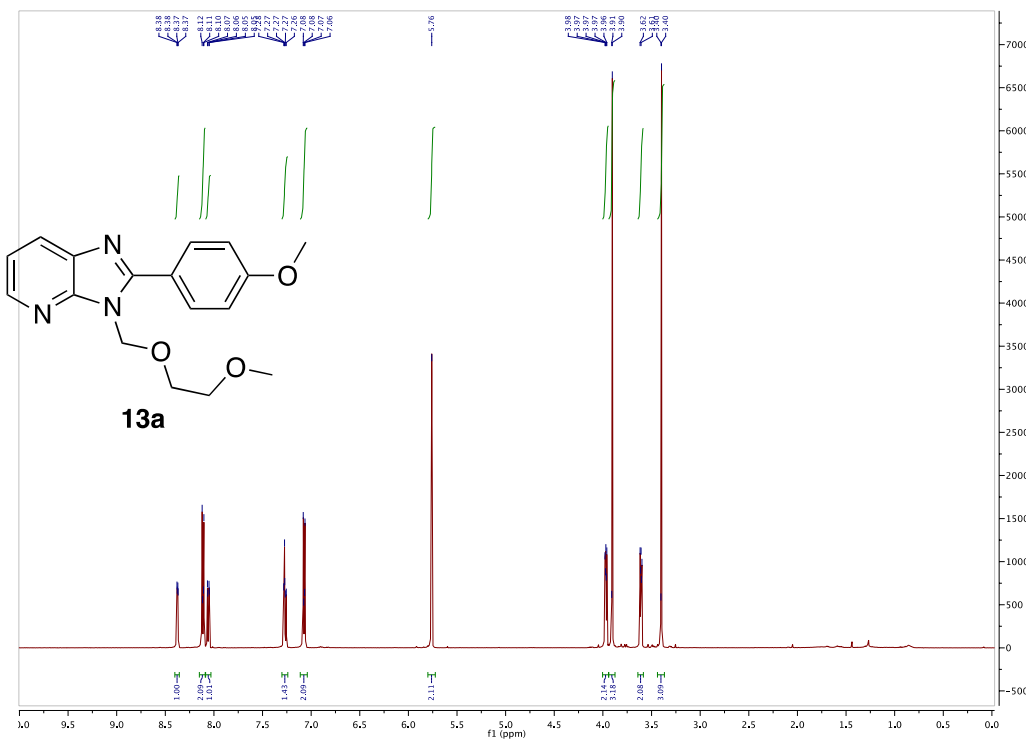


<sup>13</sup>C NMR:

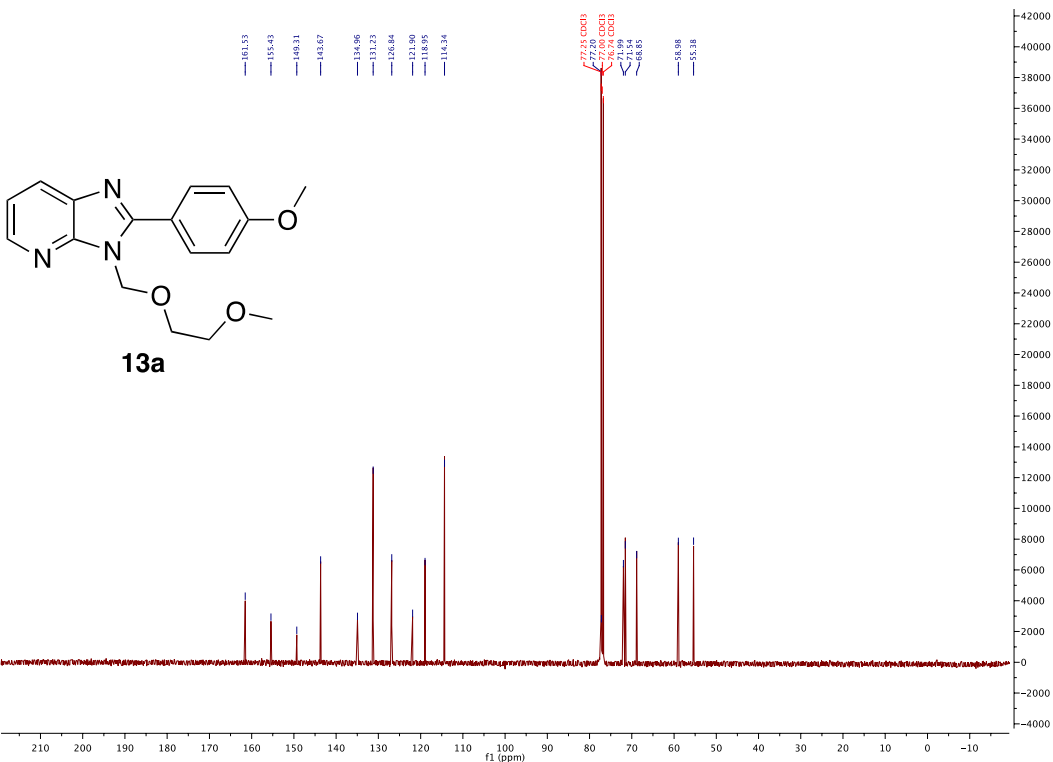




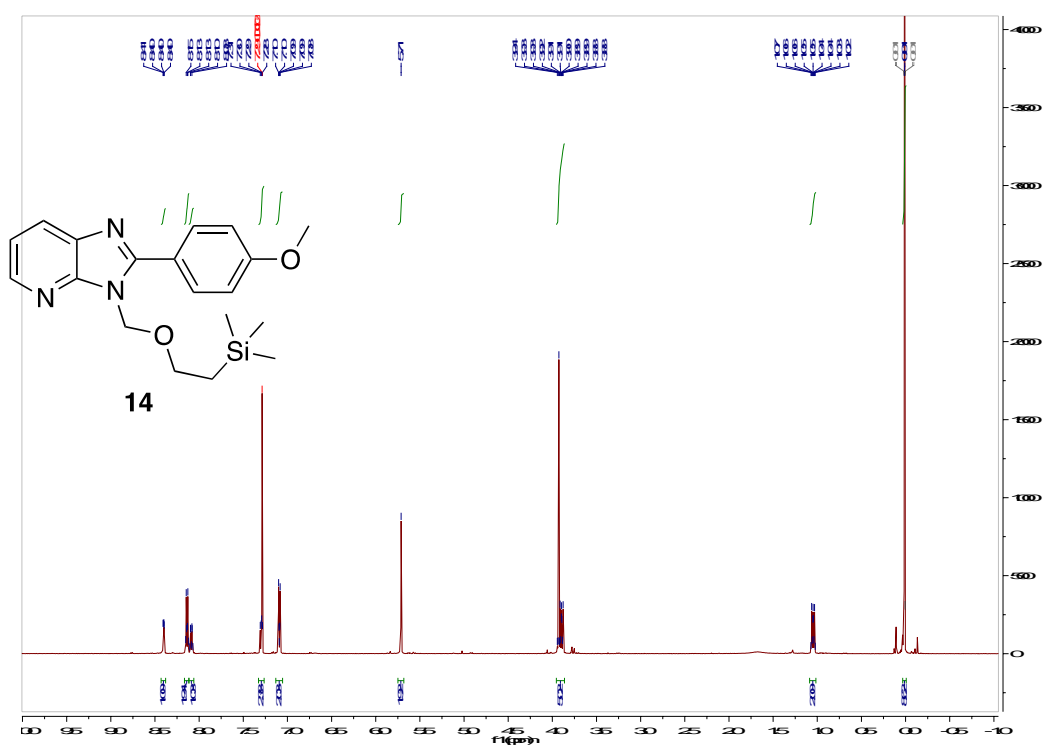
<sup>1</sup>H NMR:



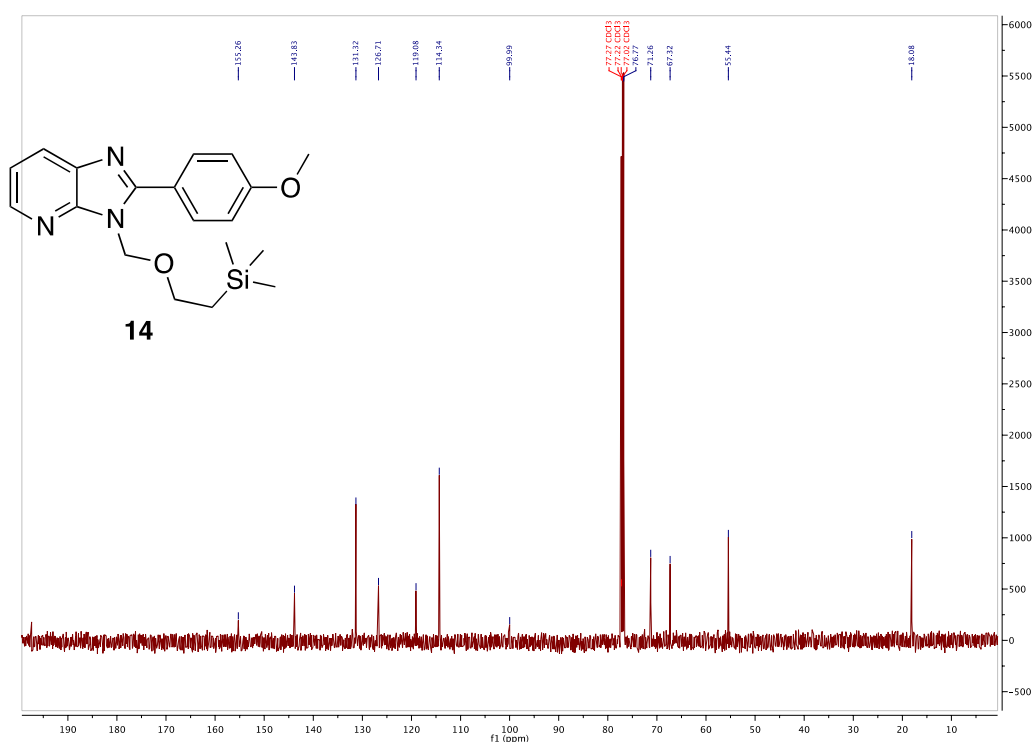
<sup>13</sup>C NMR:



$^1\text{H}$  NMR:



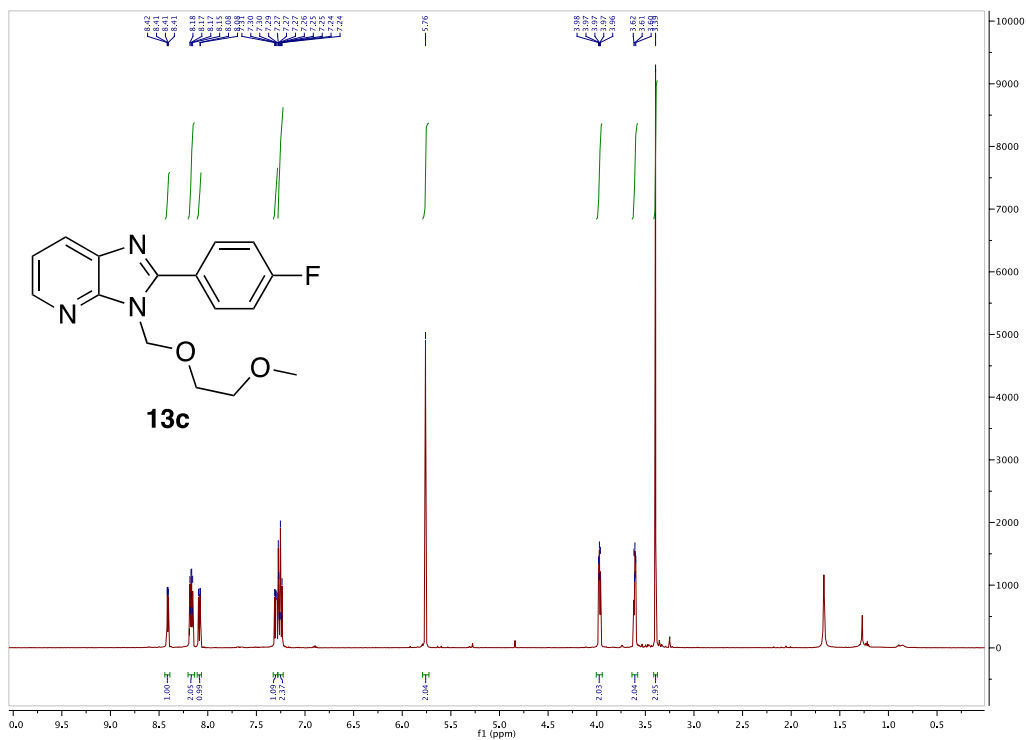
$^{13}\text{C}$  NMR:



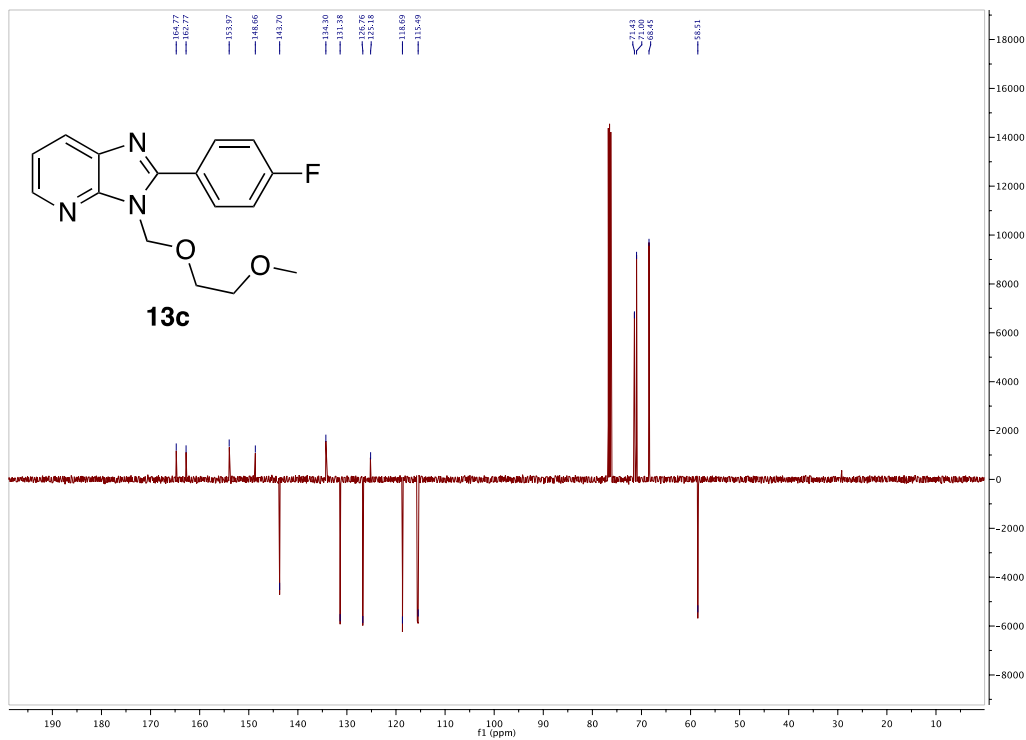
$^{13}\text{C}$  peak for Si(CH<sub>3</sub>)<sub>3</sub> assigned from HSQC experiment,  $\delta_{\text{C}} = -1.2$  ppm.



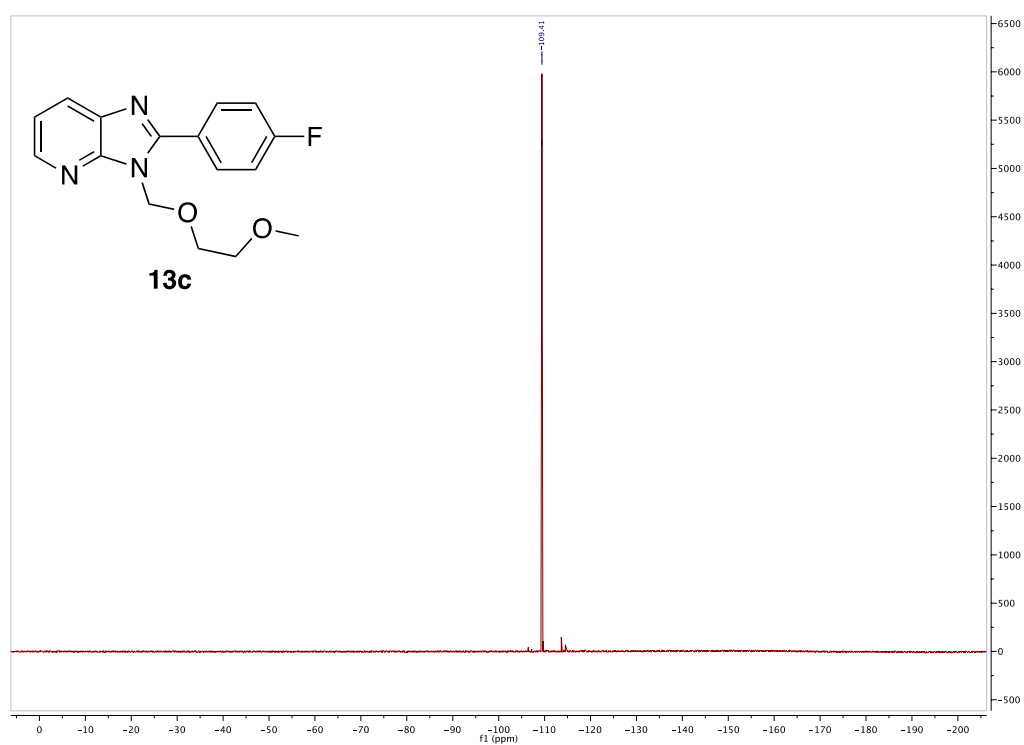
<sup>1</sup>H NMR:



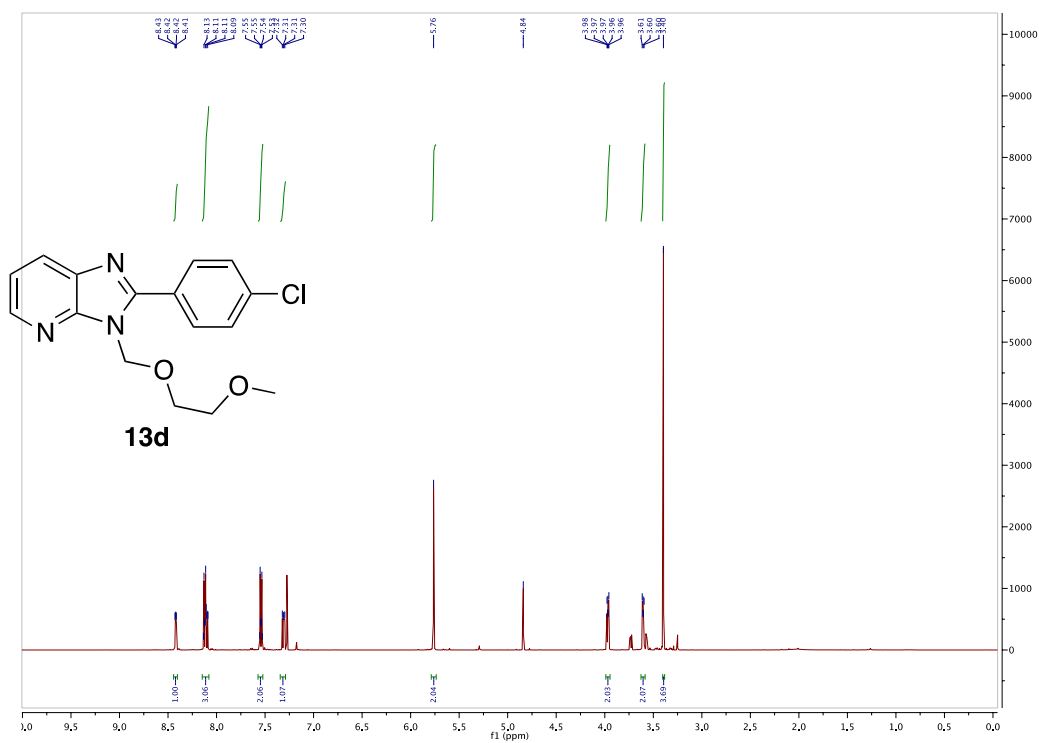
<sup>13</sup>C NMR, DEPT135:



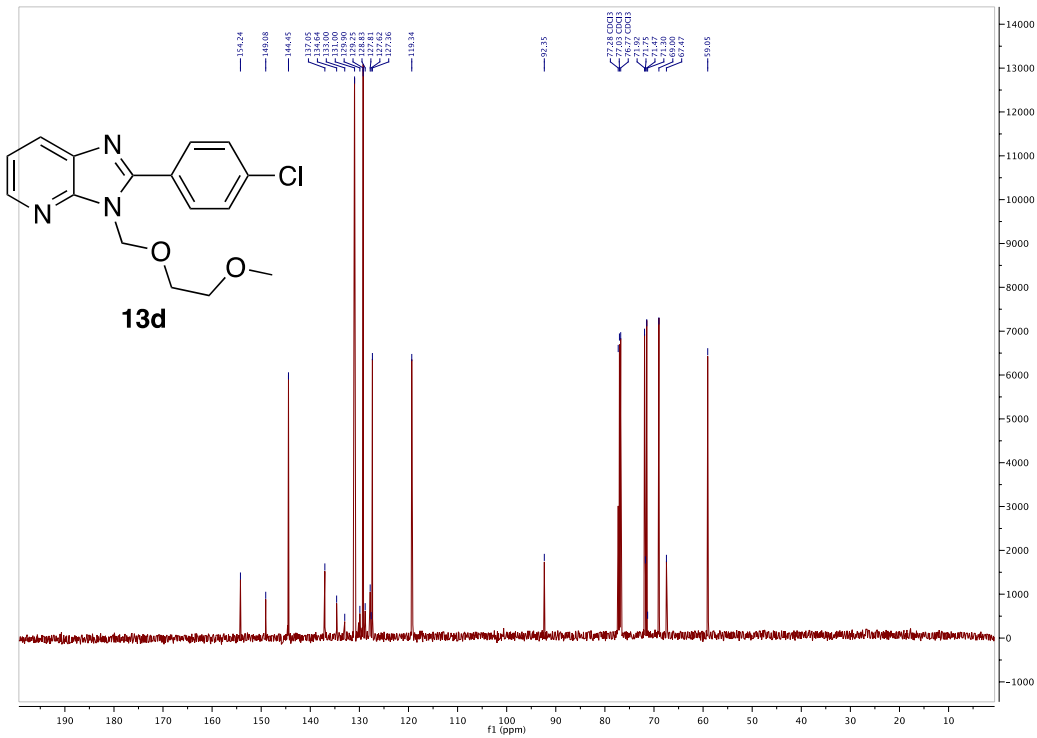
<sup>19</sup>F NMR:



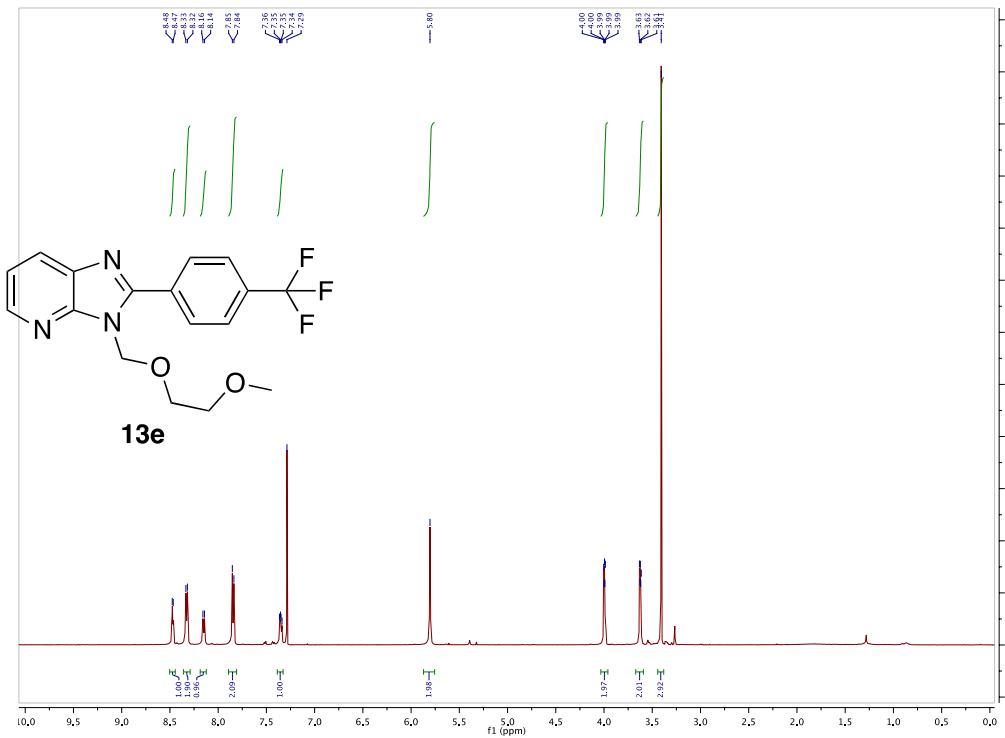
<sup>1</sup>H NMR:



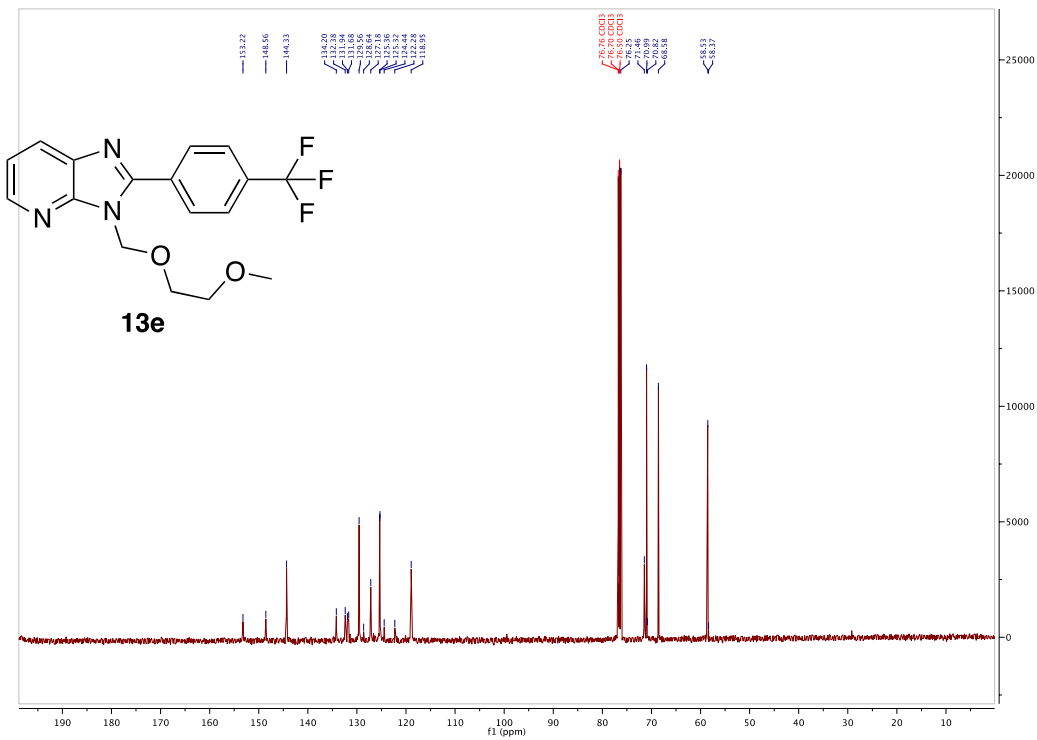
<sup>13</sup>C NMR:



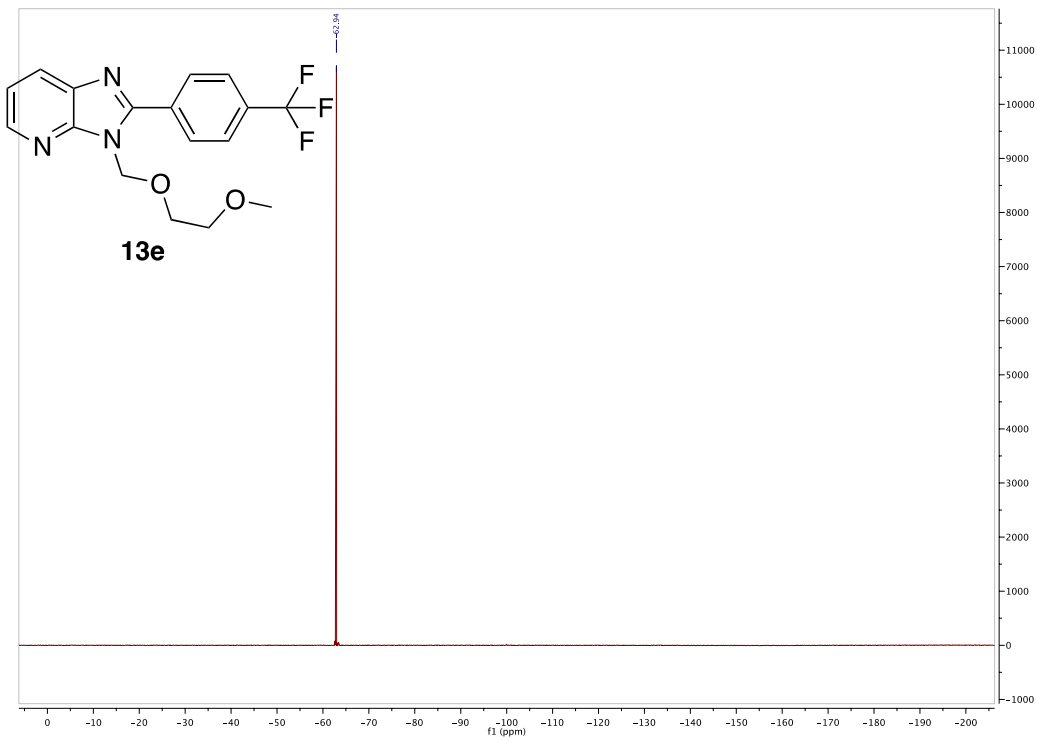
<sup>1</sup>H NMR:



<sup>13</sup>C NMR:



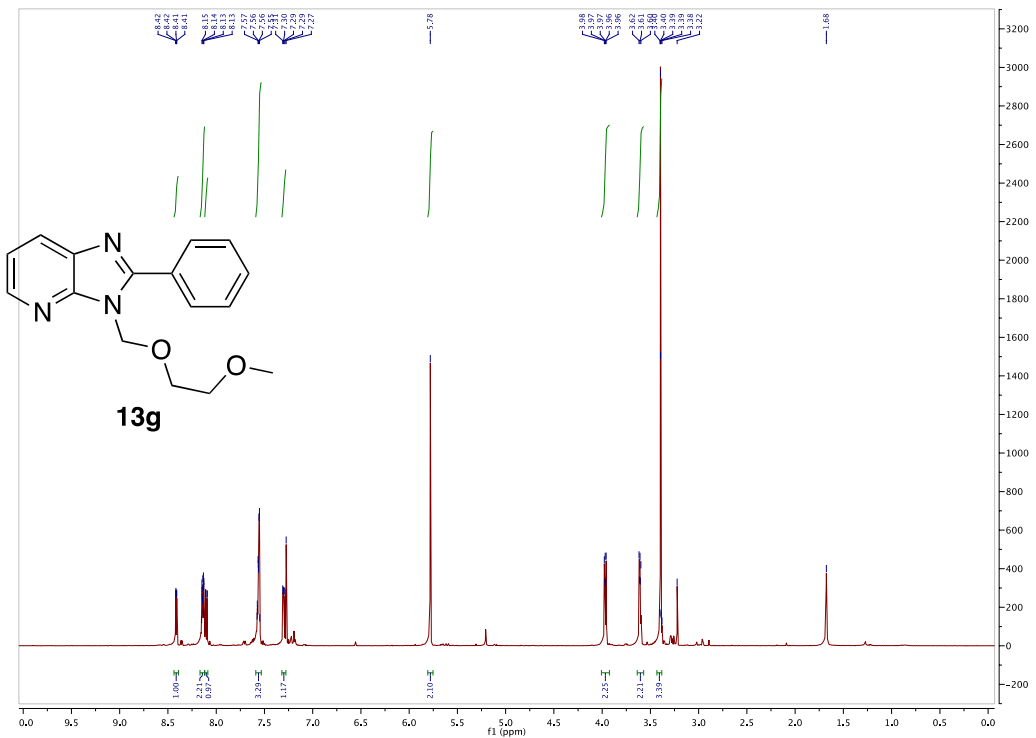
<sup>19</sup>F NMR:



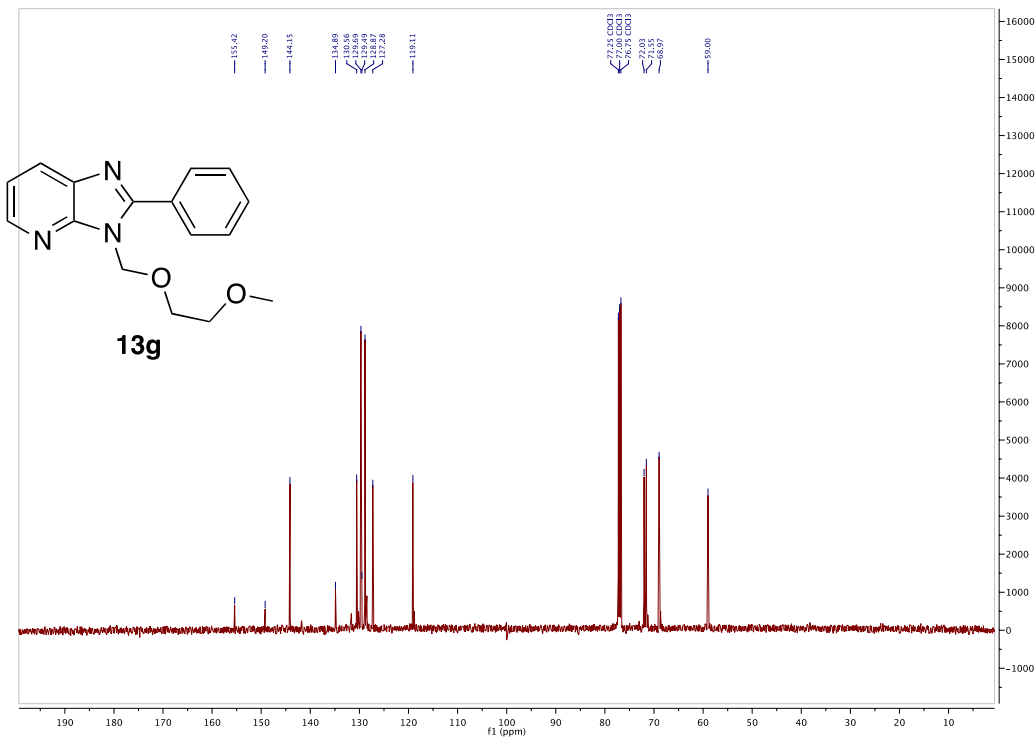




<sup>1</sup>H NMR:

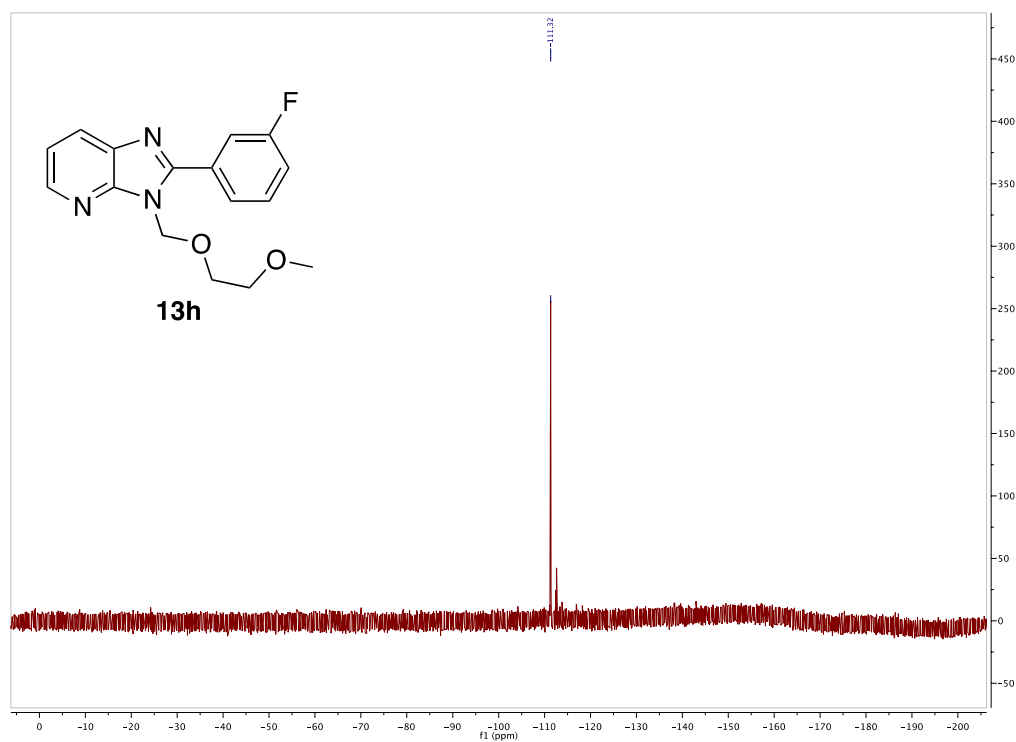


<sup>13</sup>C NMR:

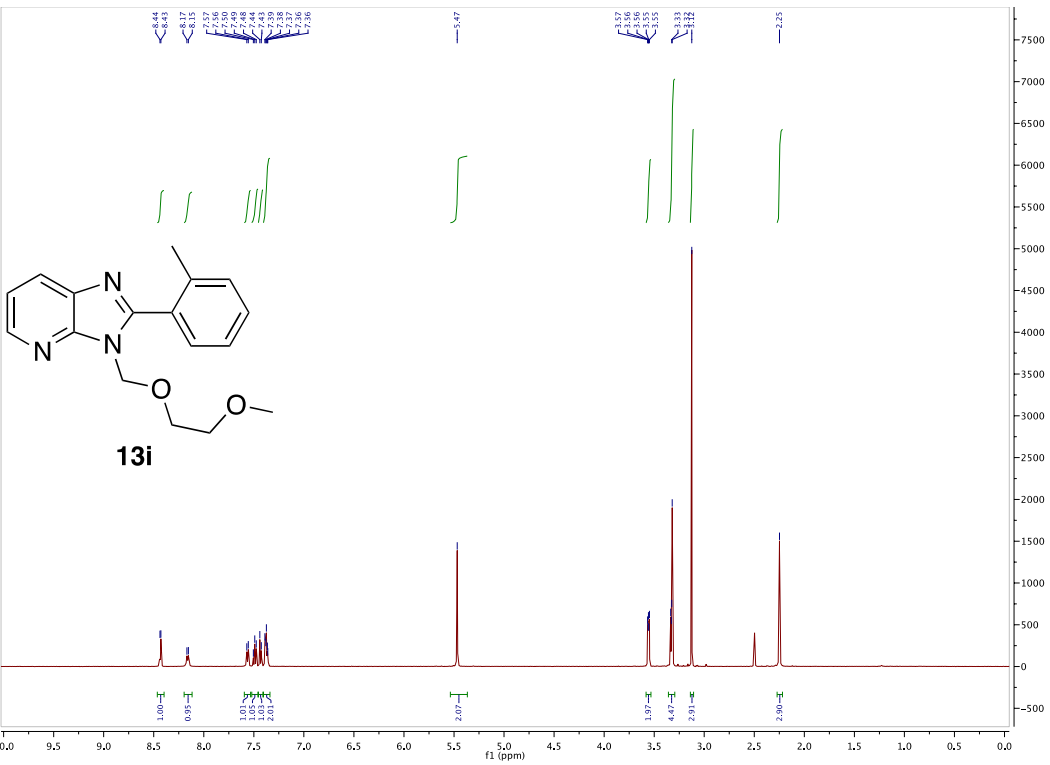




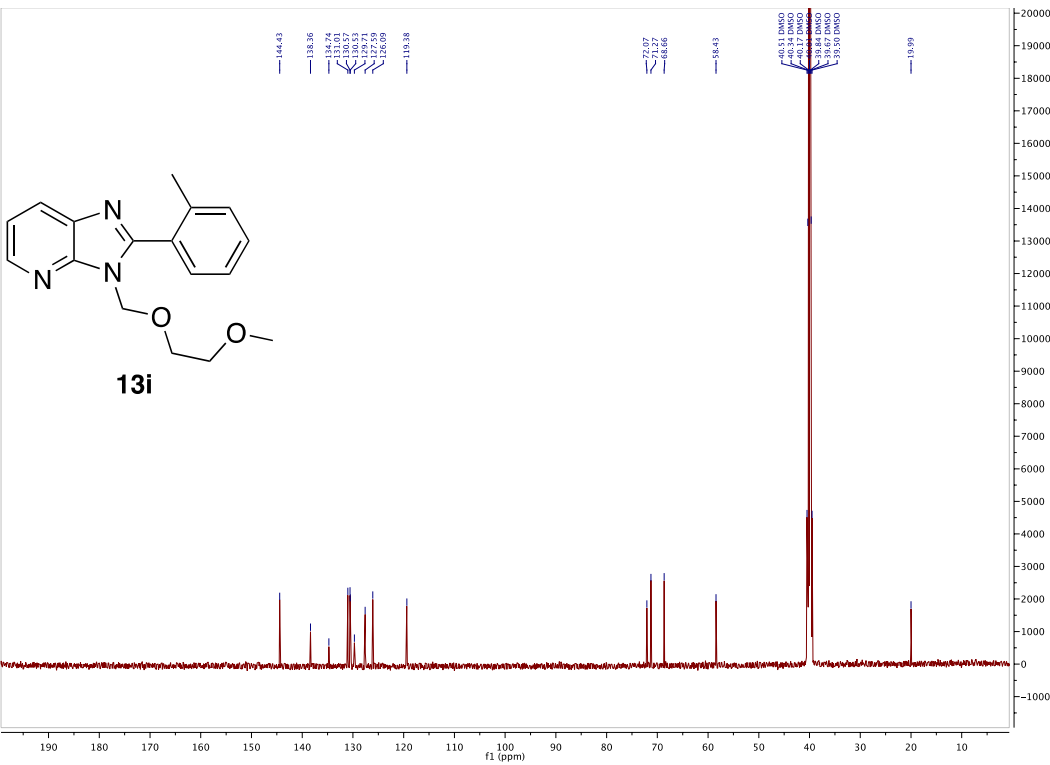
<sup>19</sup>F NMR:



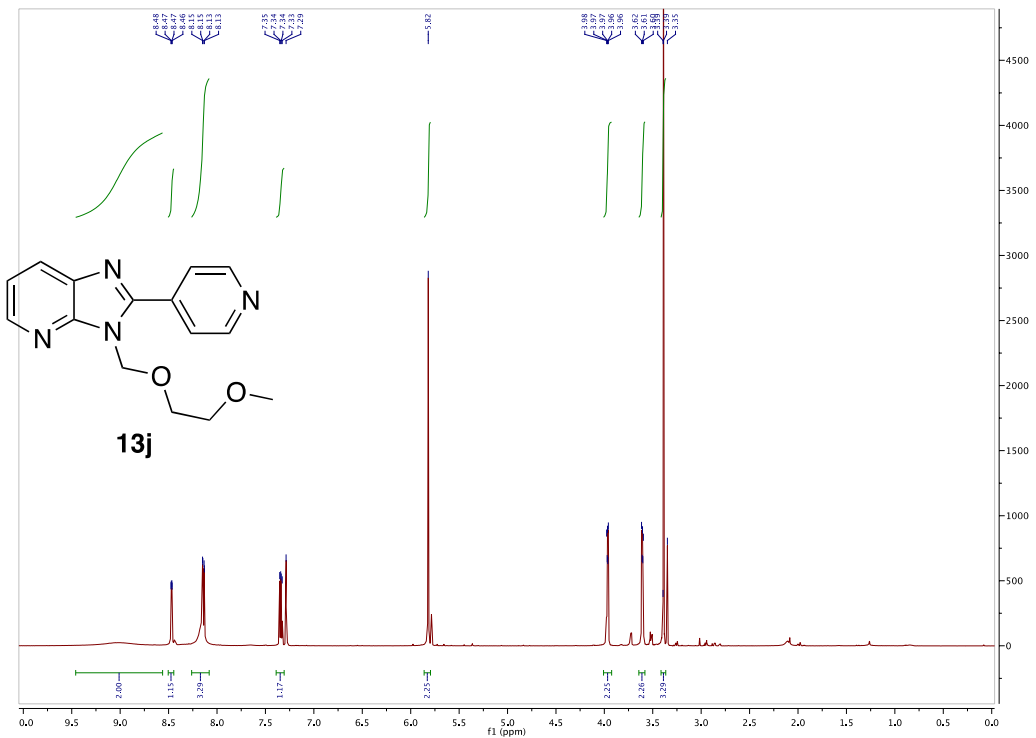
<sup>1</sup>H NMR:



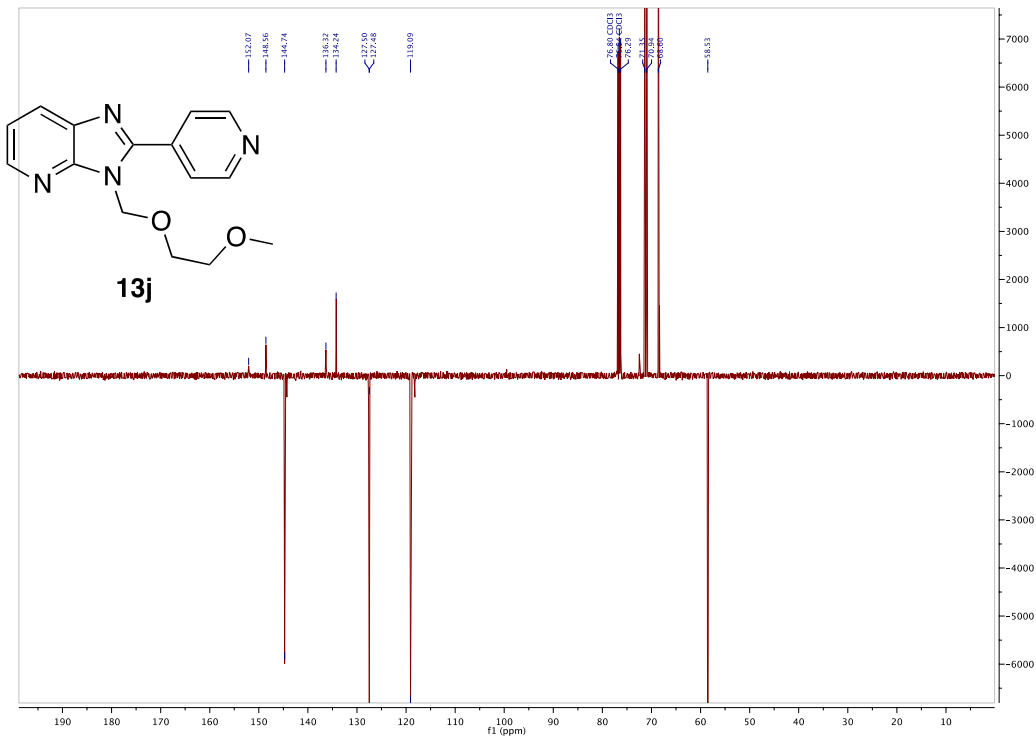
<sup>13</sup>C NMR:



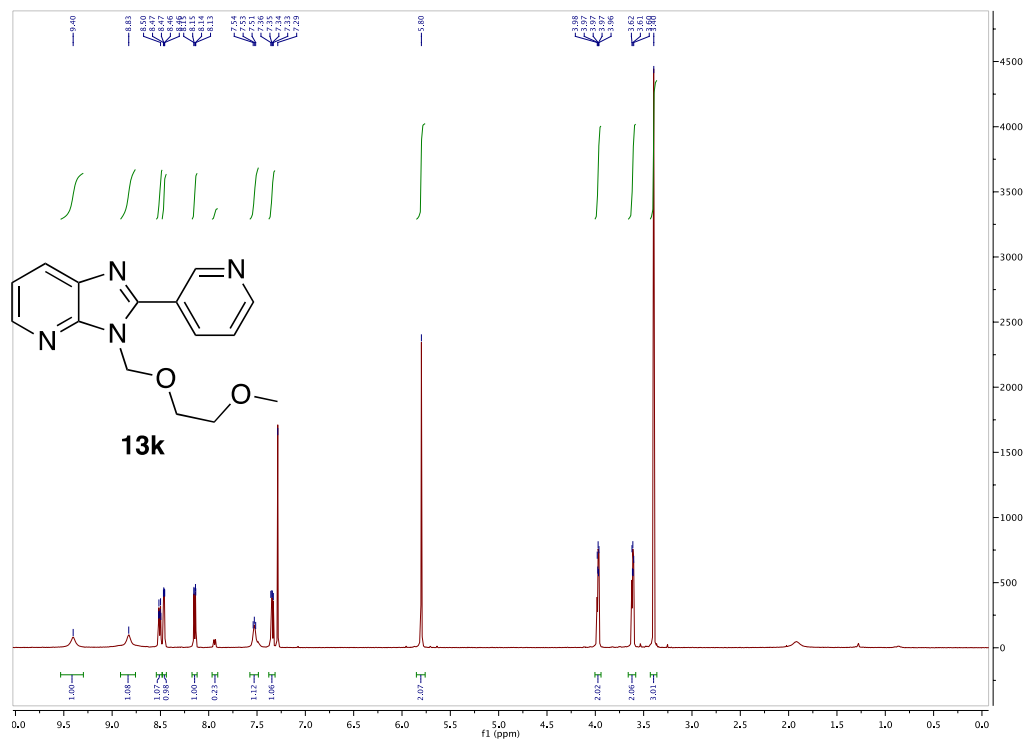
<sup>1</sup>H NMR:



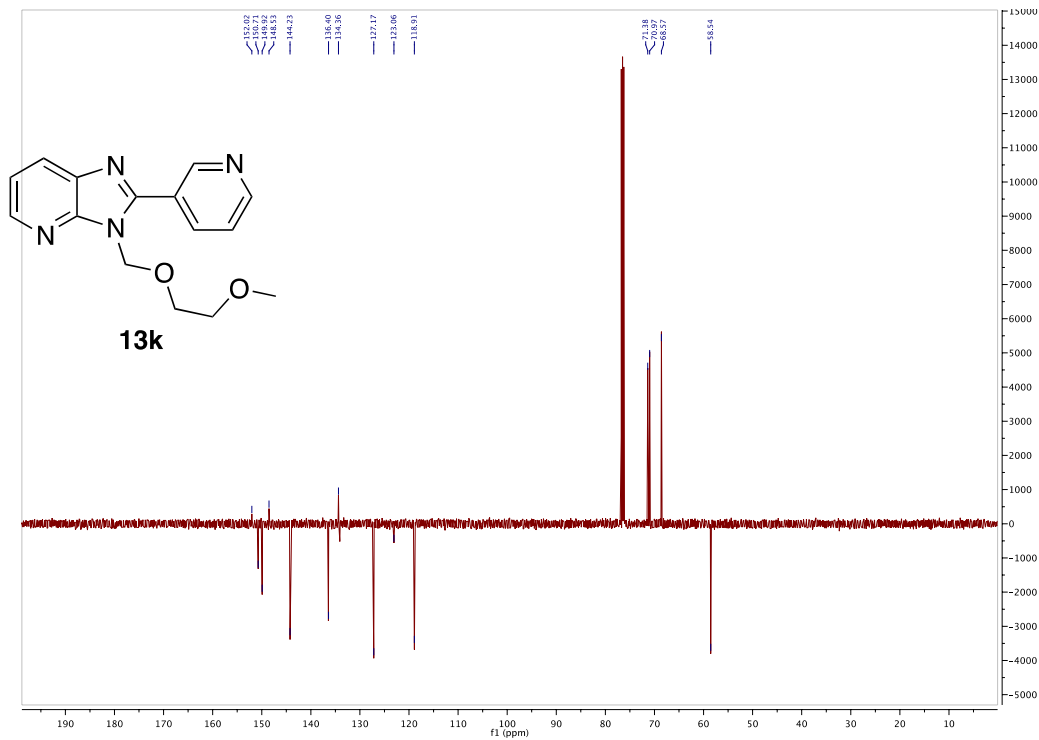
<sup>13</sup>C NMR, DEPT135:



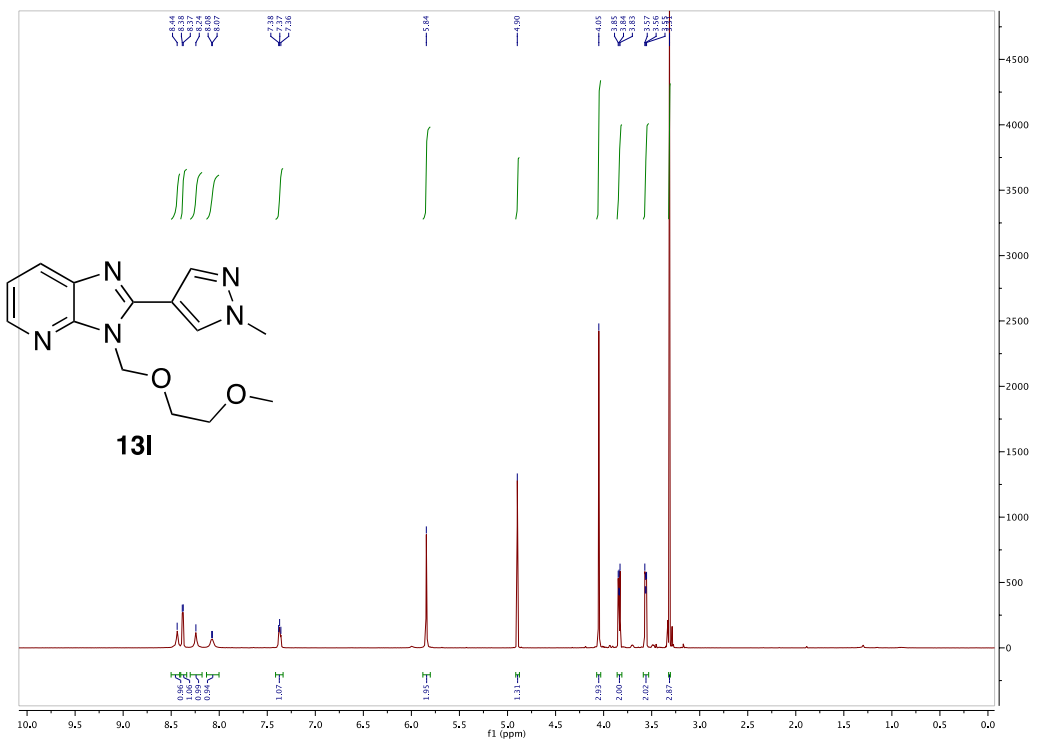
<sup>1</sup>H NMR:



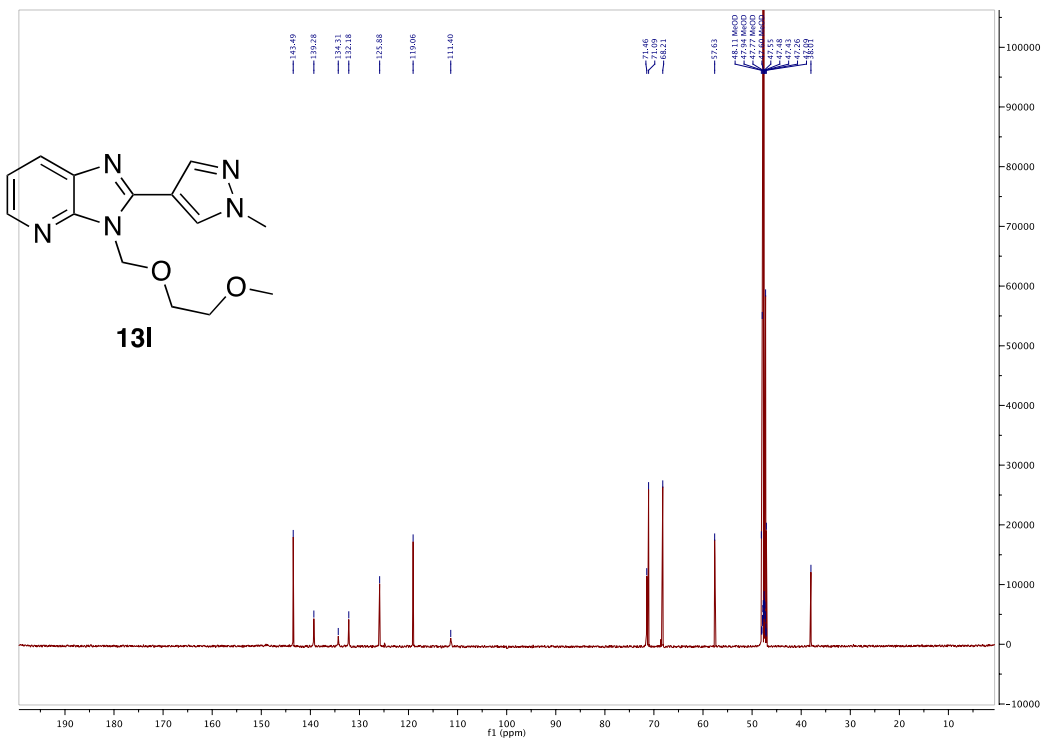
<sup>13</sup>C NMR, DEPT135:



<sup>1</sup>H NMR:



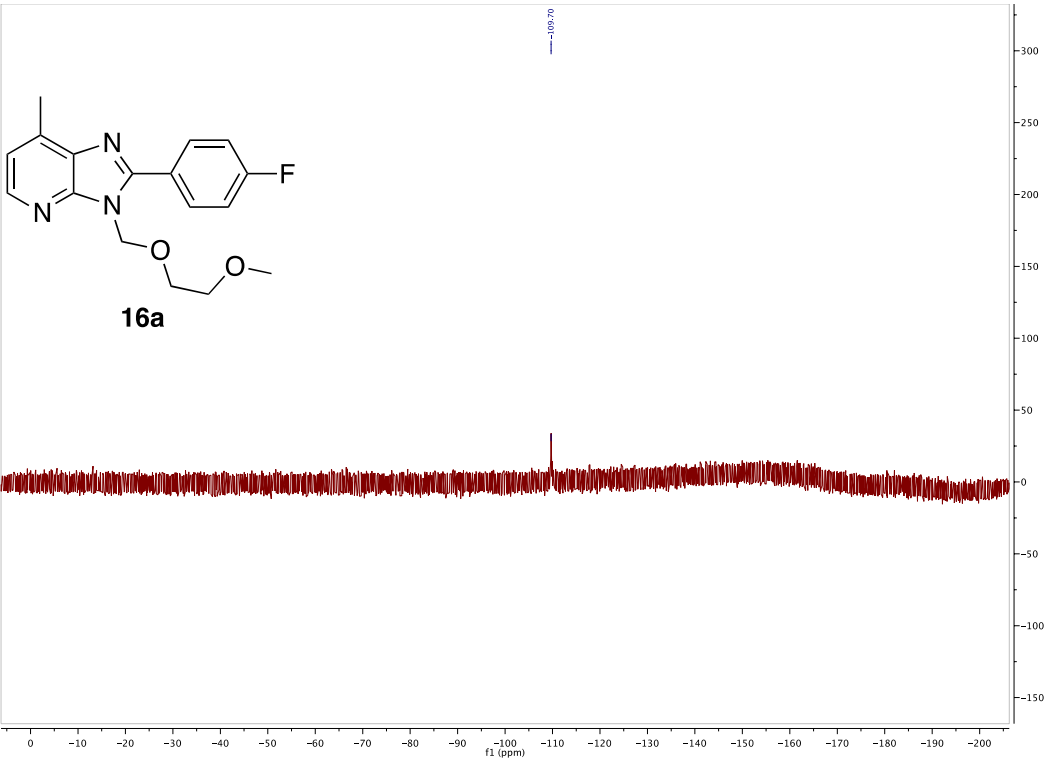
<sup>13</sup>C NMR:



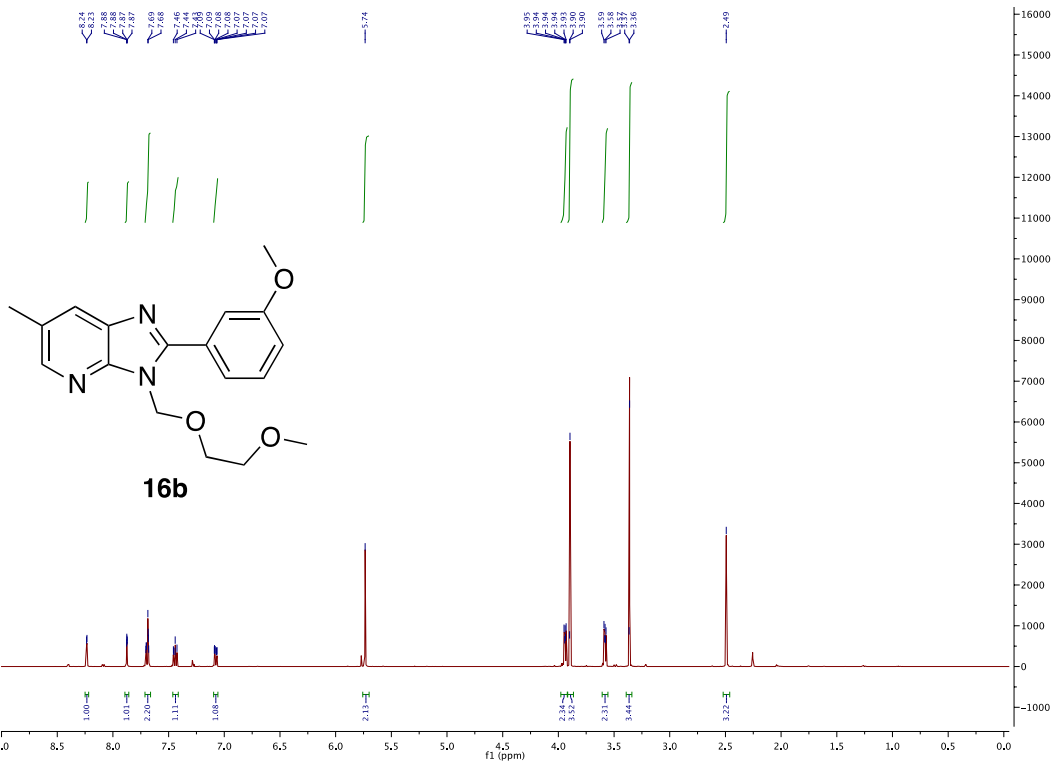




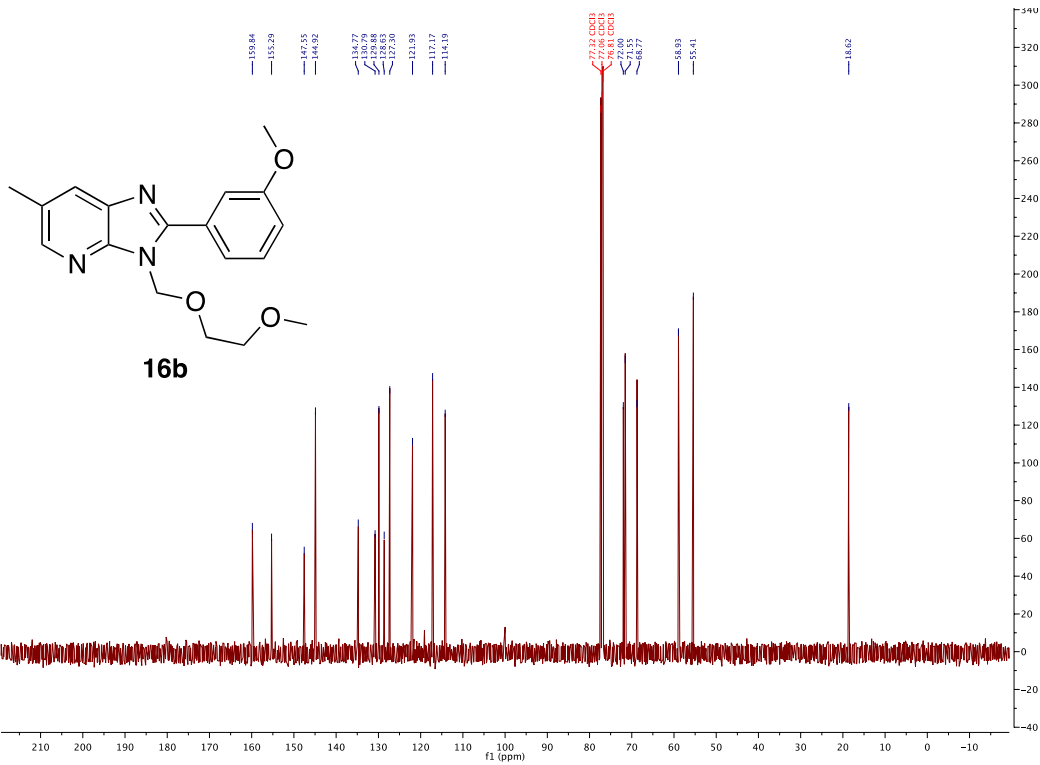
<sup>19</sup>F NMR:



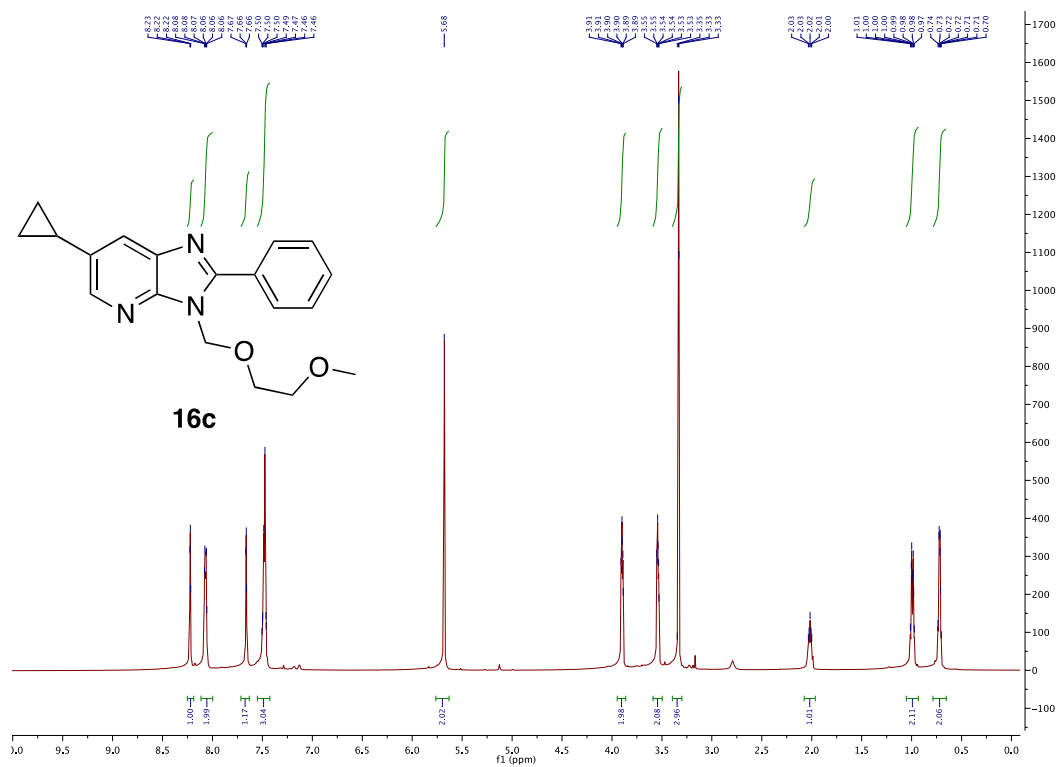
<sup>1</sup>H NMR:



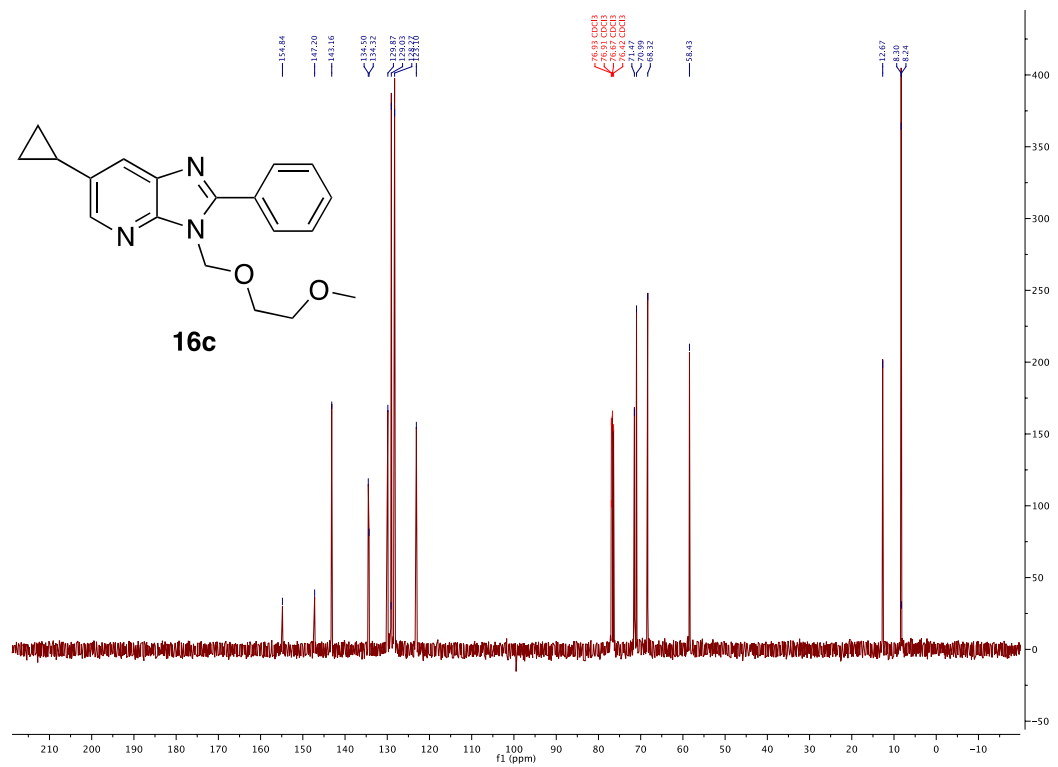
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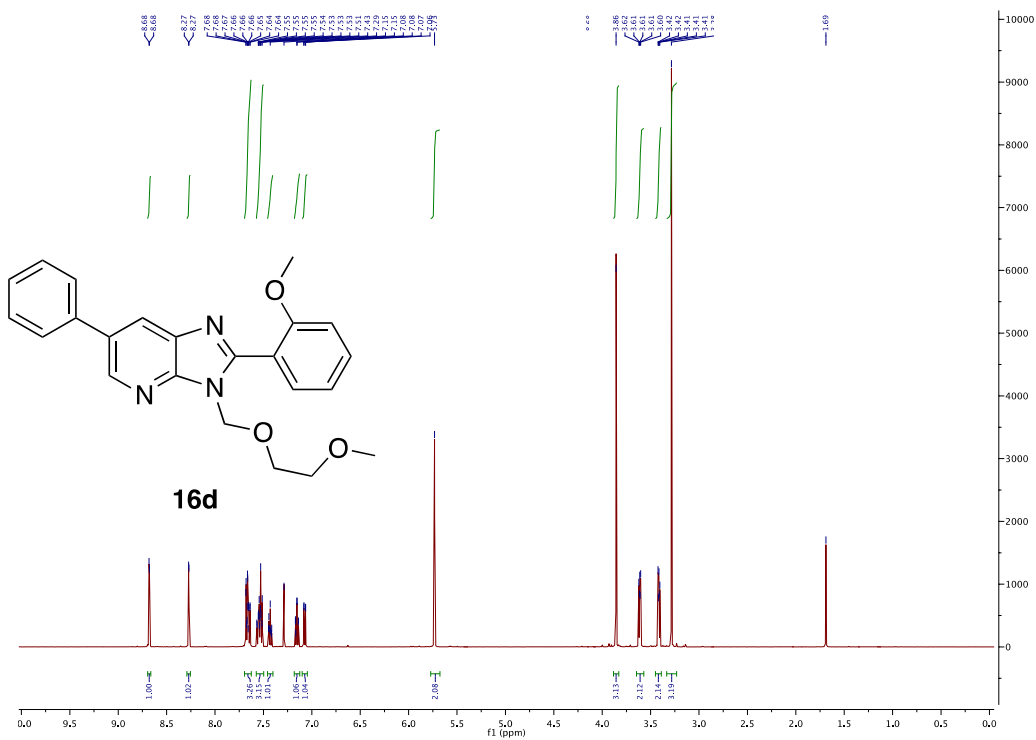
<sup>1</sup>H NMR:



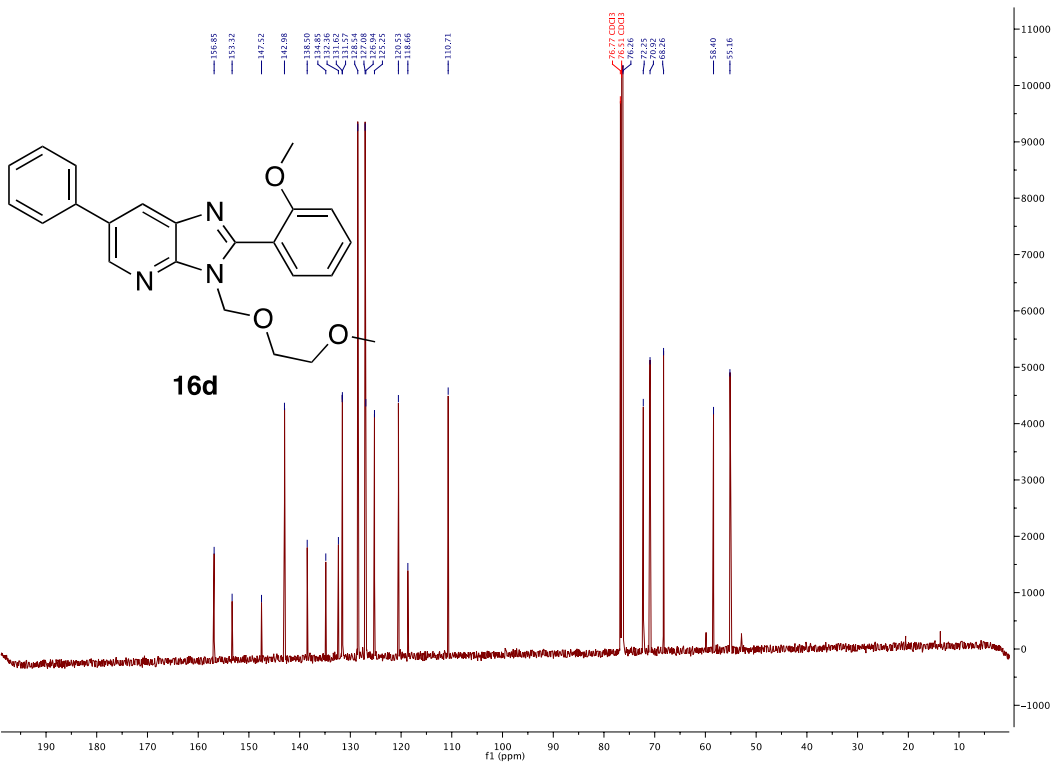
<sup>13</sup>C NMR:



<sup>1</sup>H NMR:

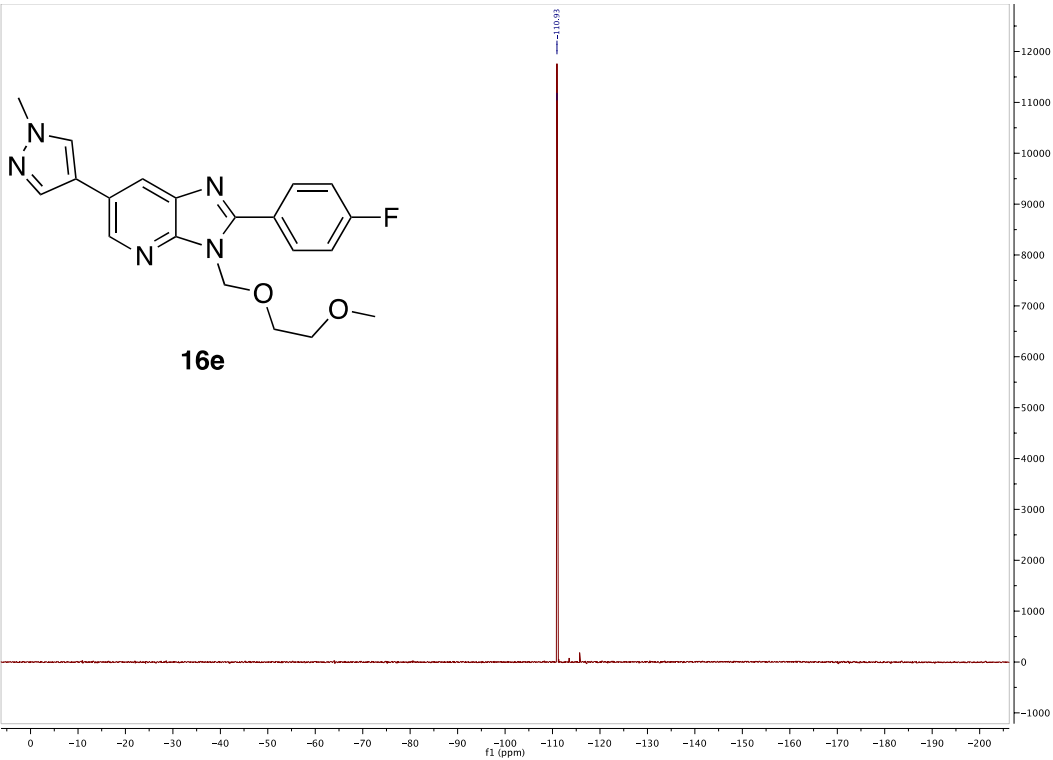


<sup>13</sup>C NMR:

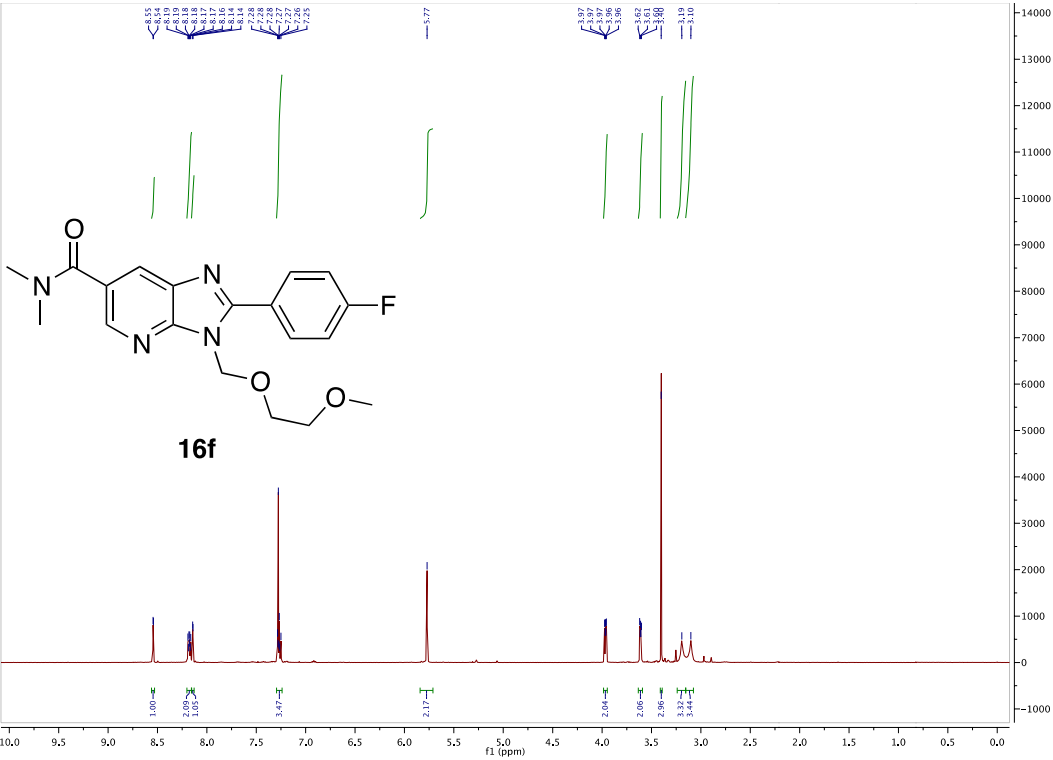




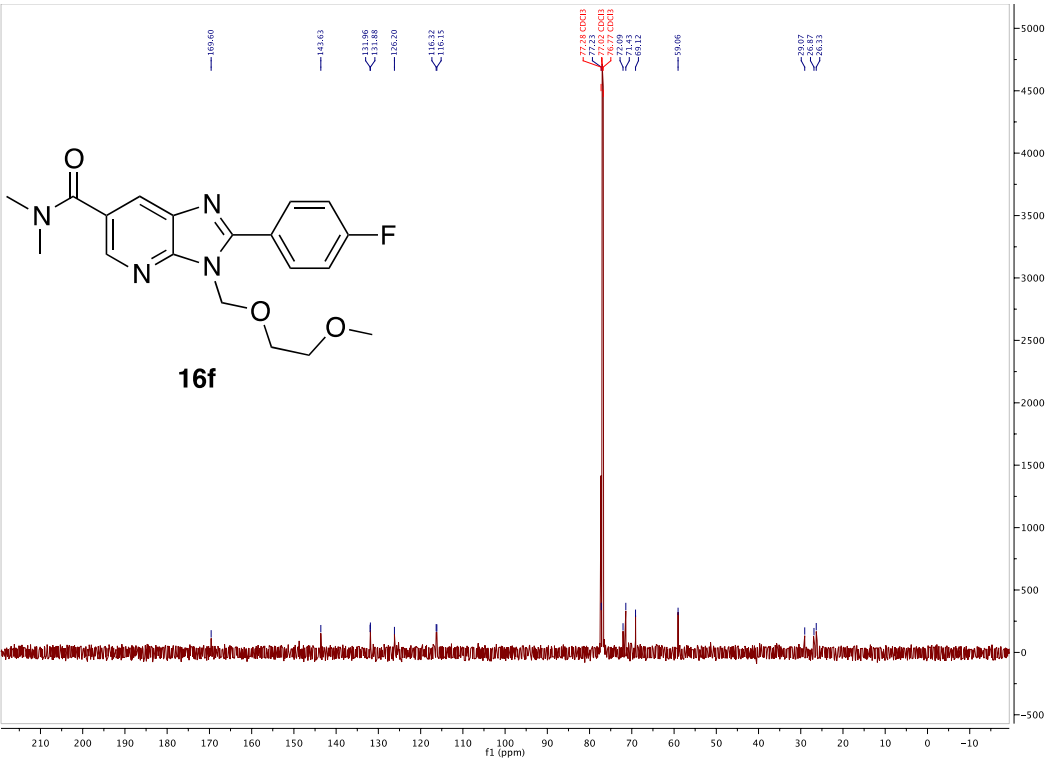
<sup>19</sup>F NMR:



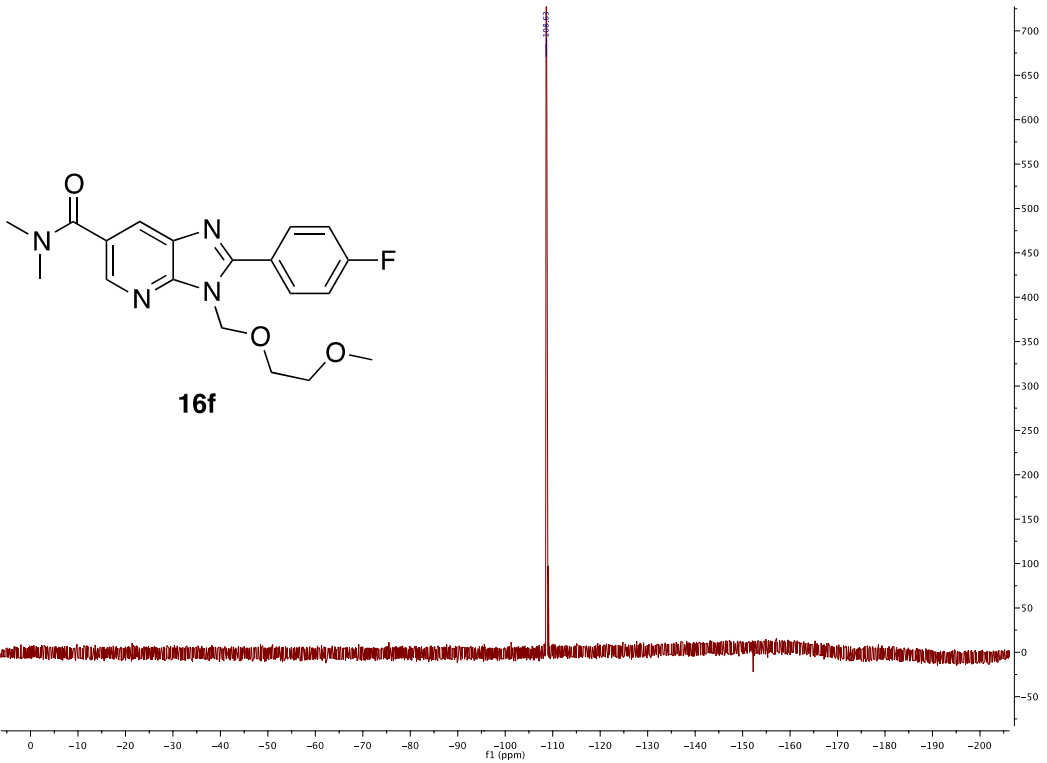
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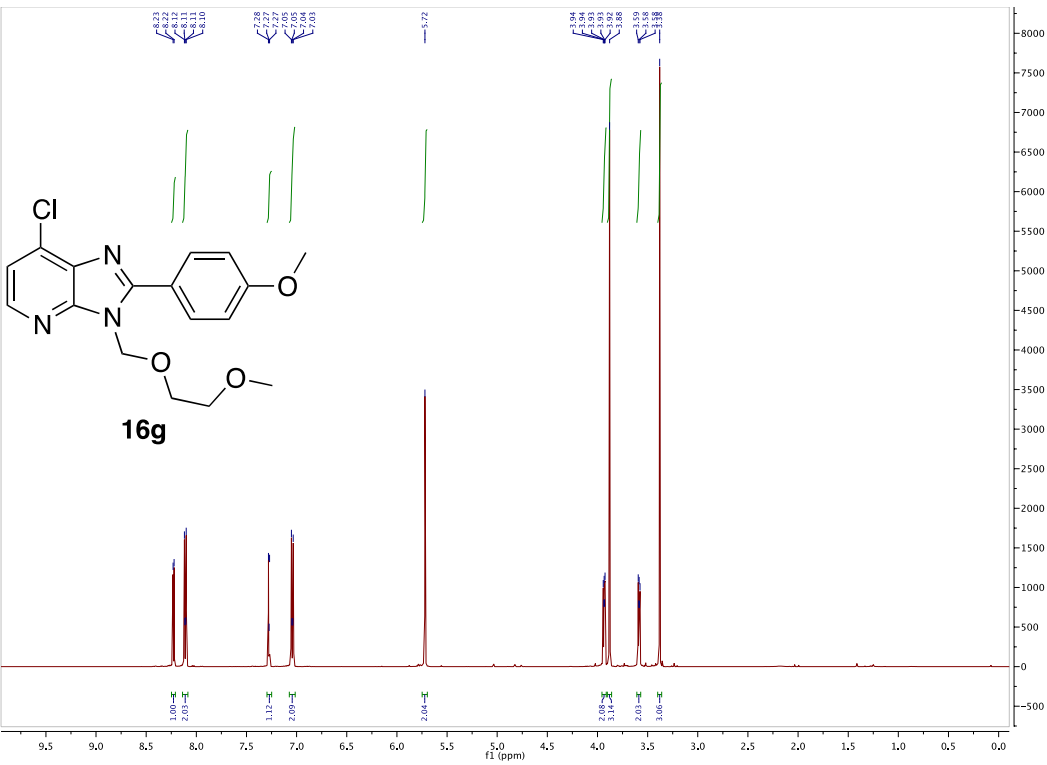
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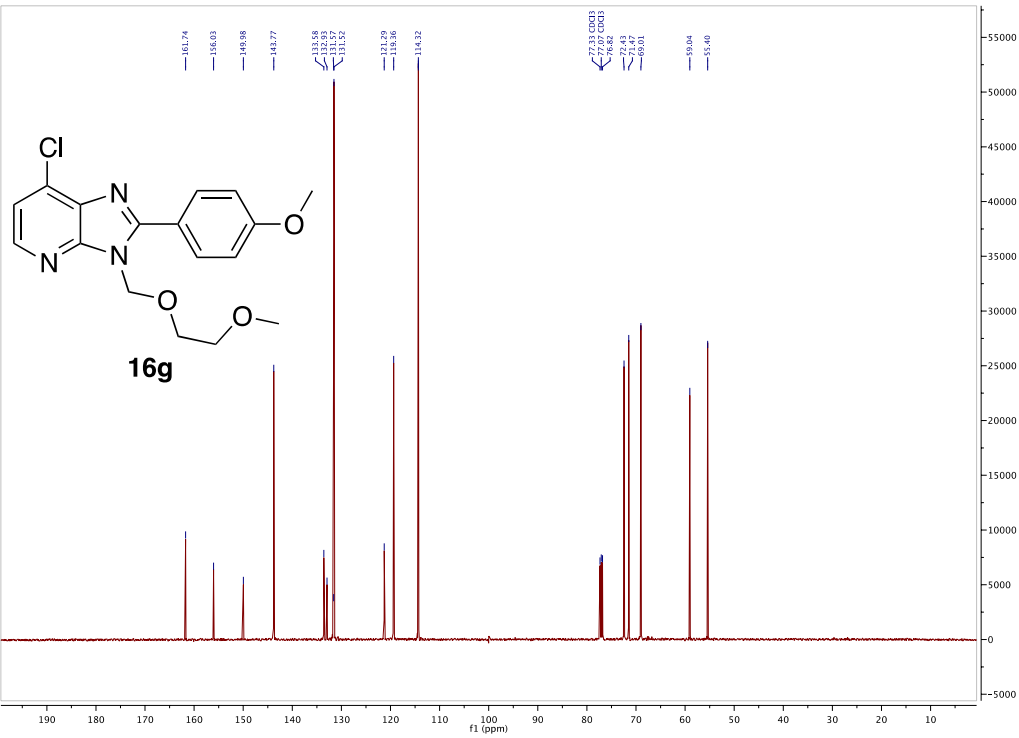
<sup>19</sup>F NMR:



<sup>1</sup>H NMR:

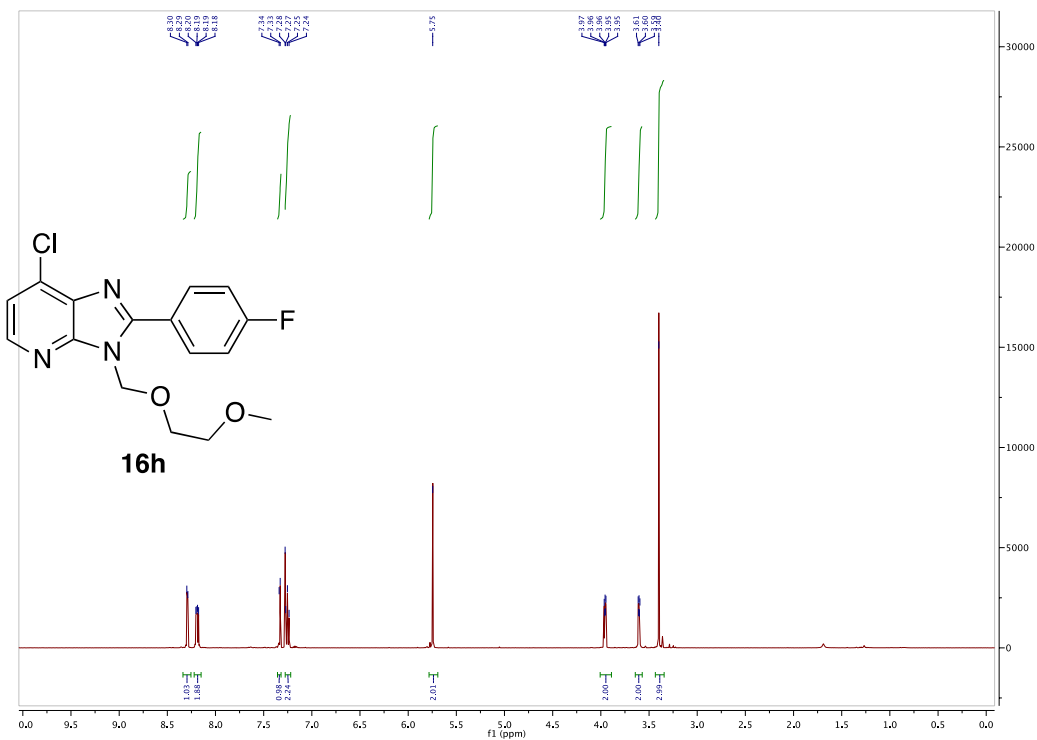


<sup>13</sup>C NMR:

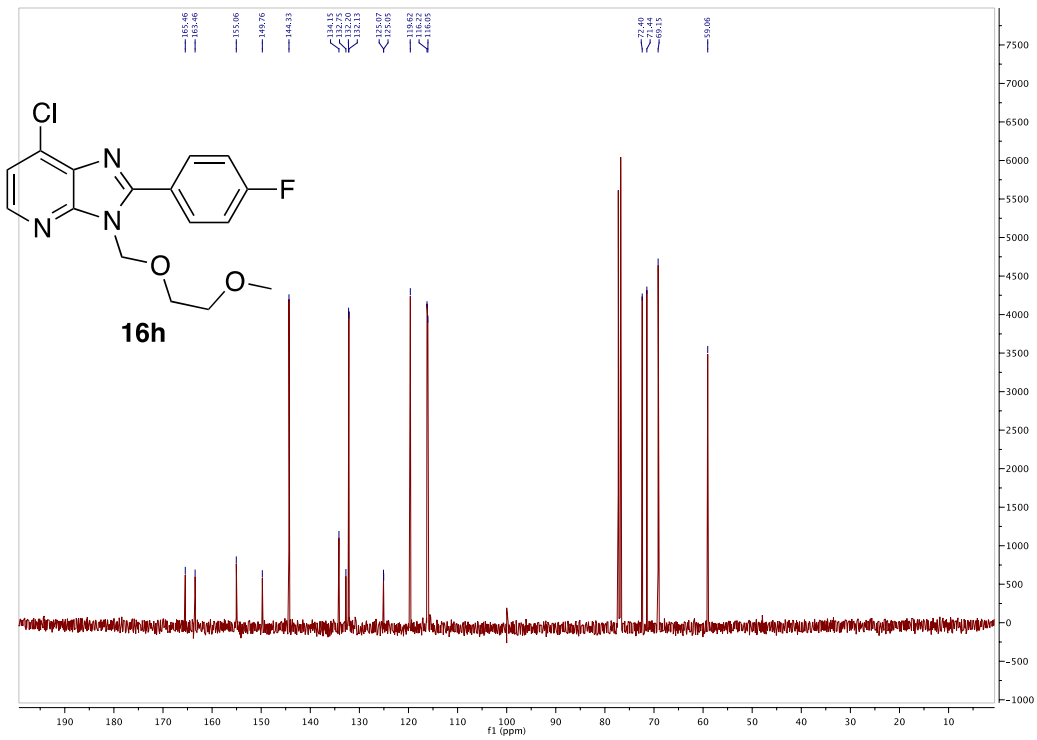




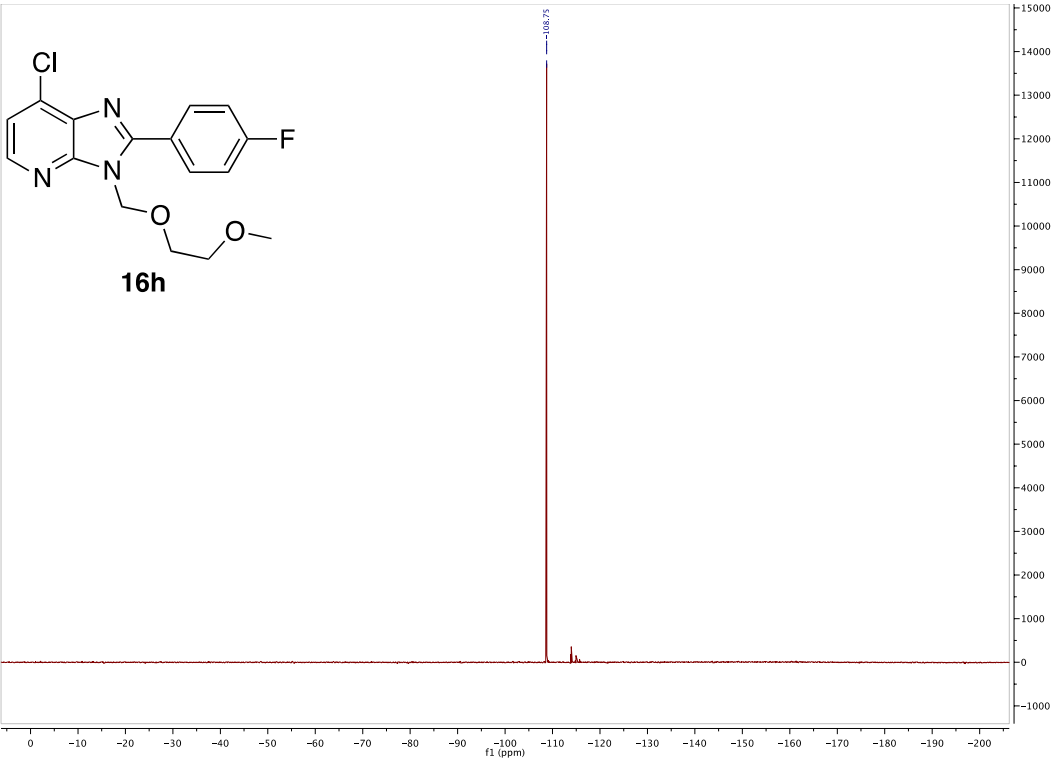
<sup>1</sup>H NMR:



<sup>13</sup>C NMR:



<sup>19</sup>F NMR:

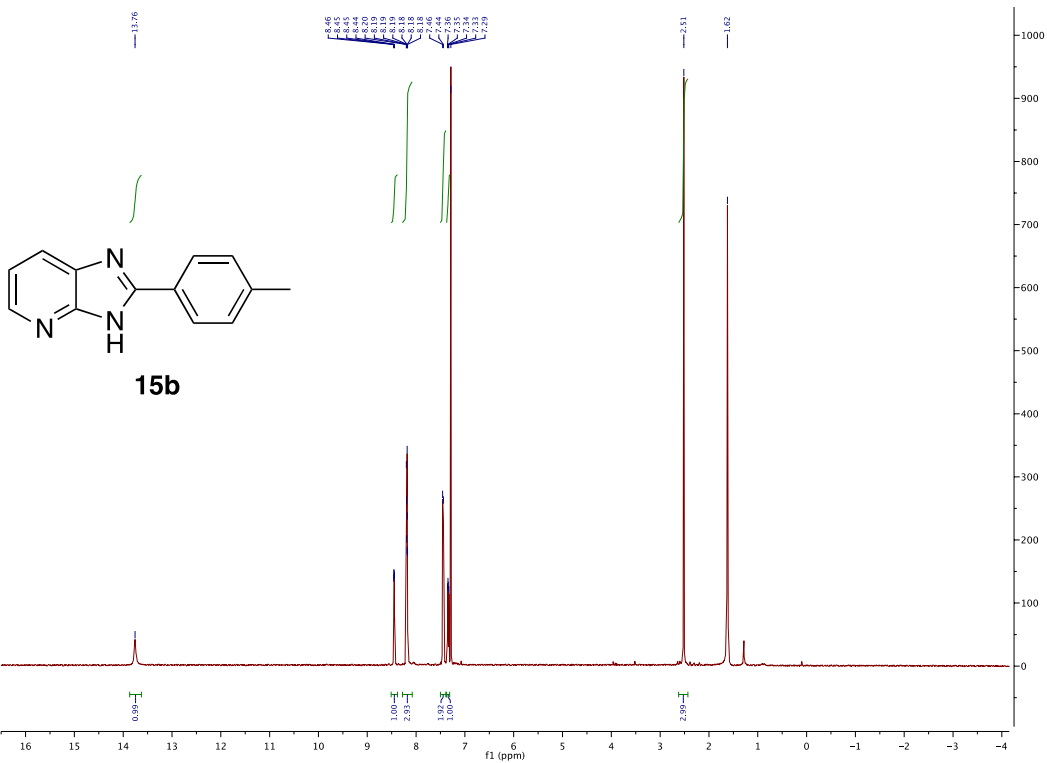






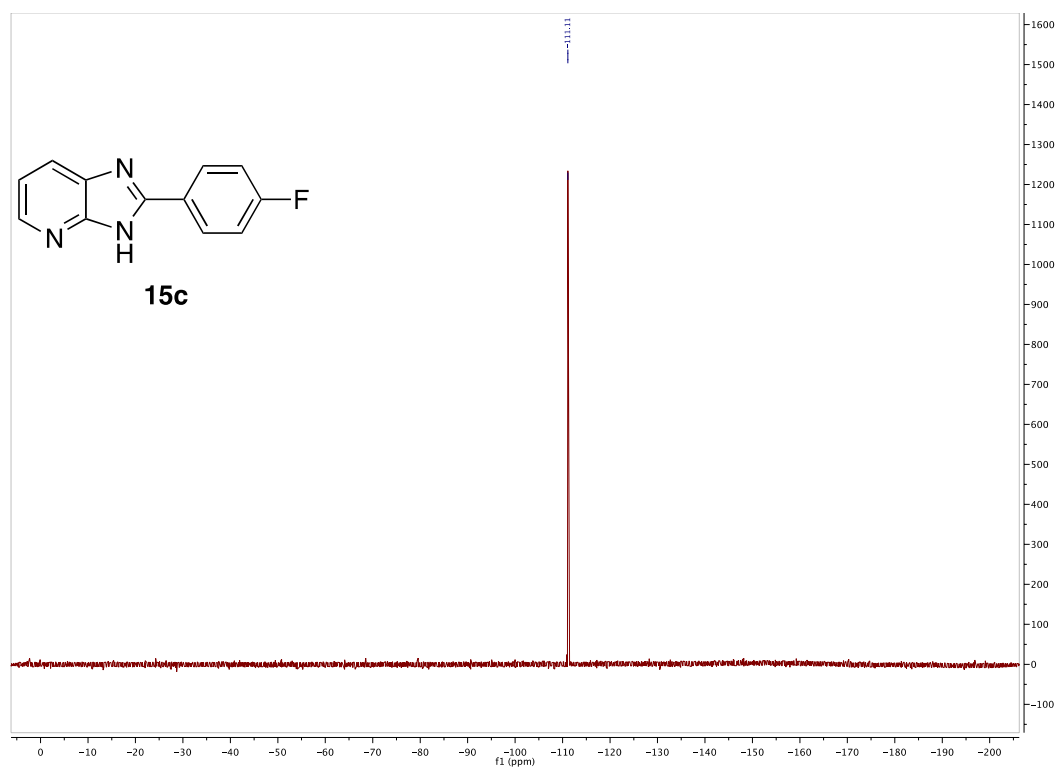


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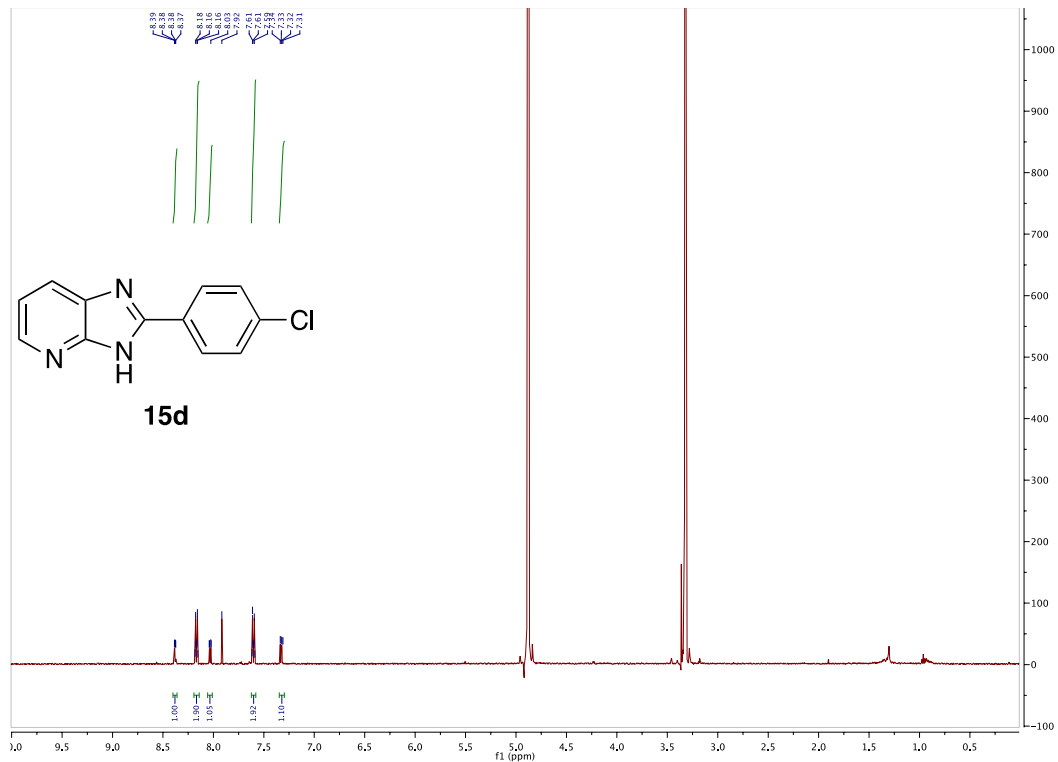




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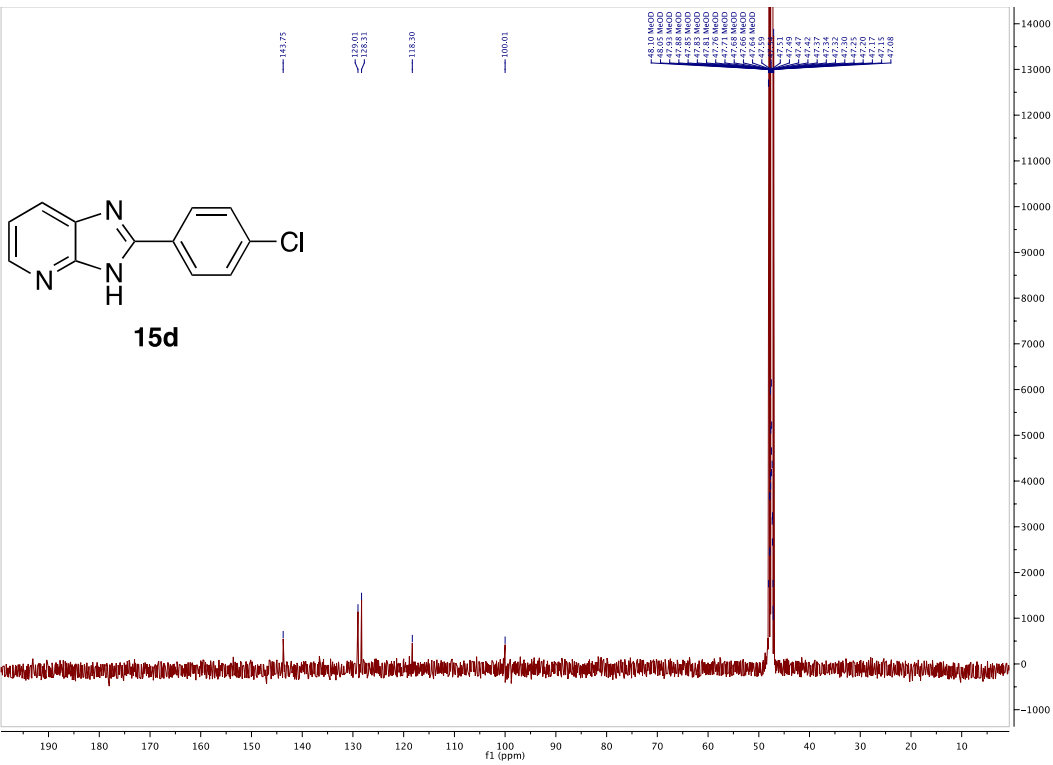


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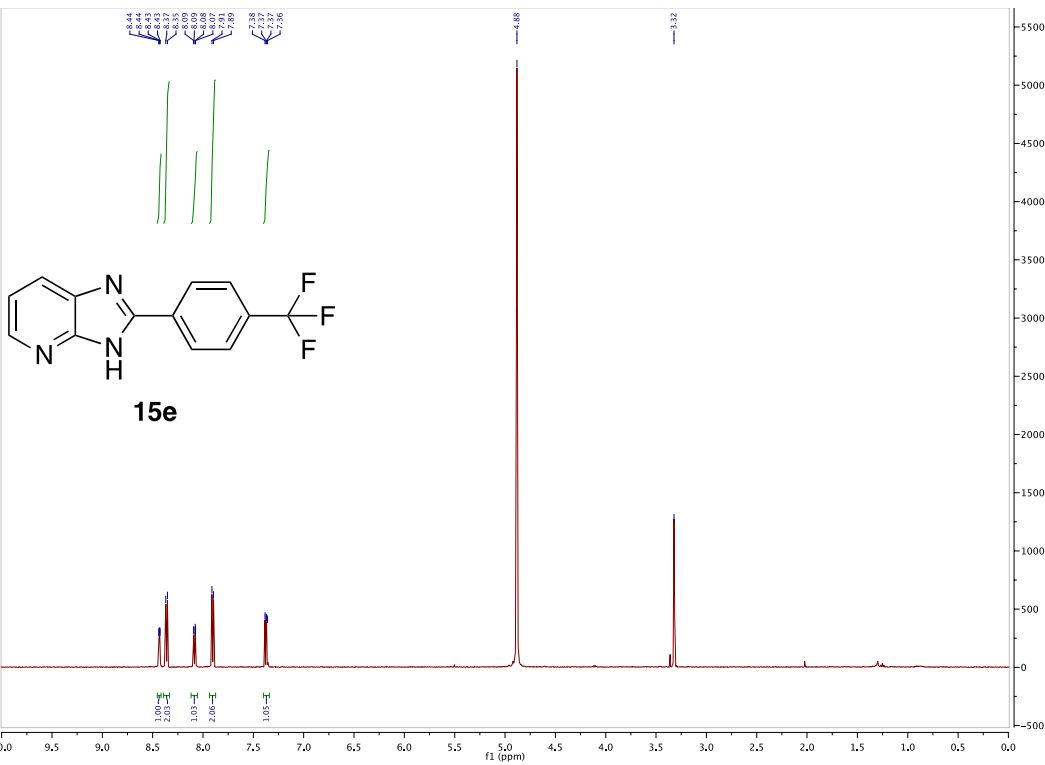




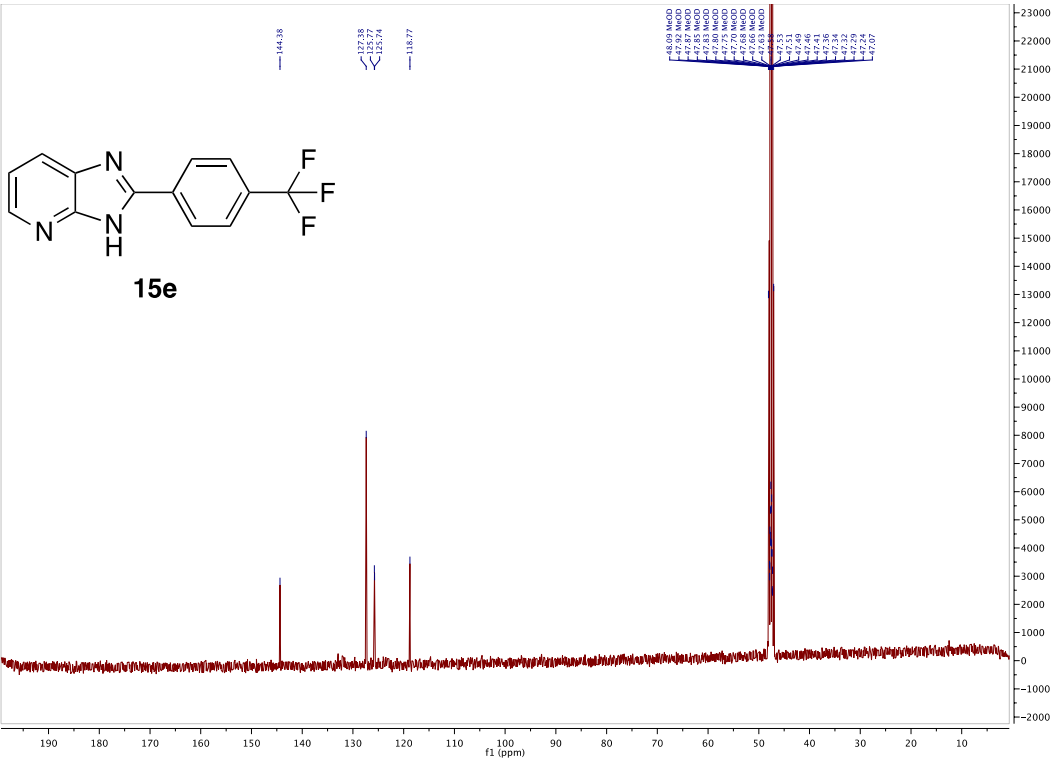
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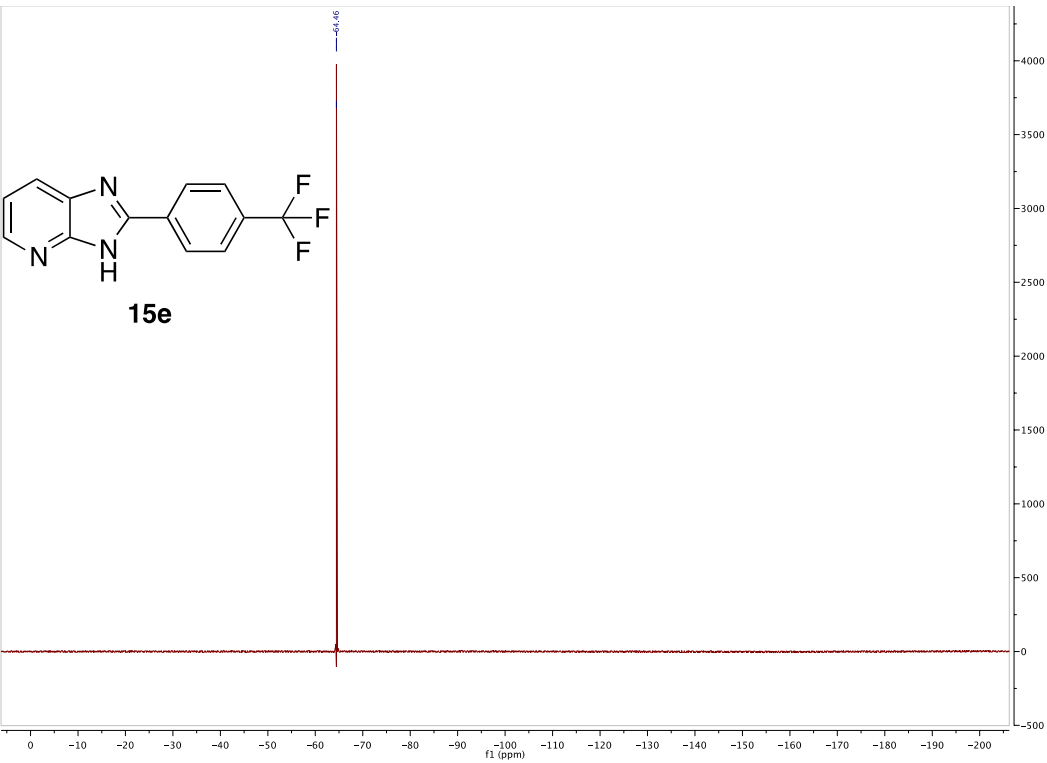
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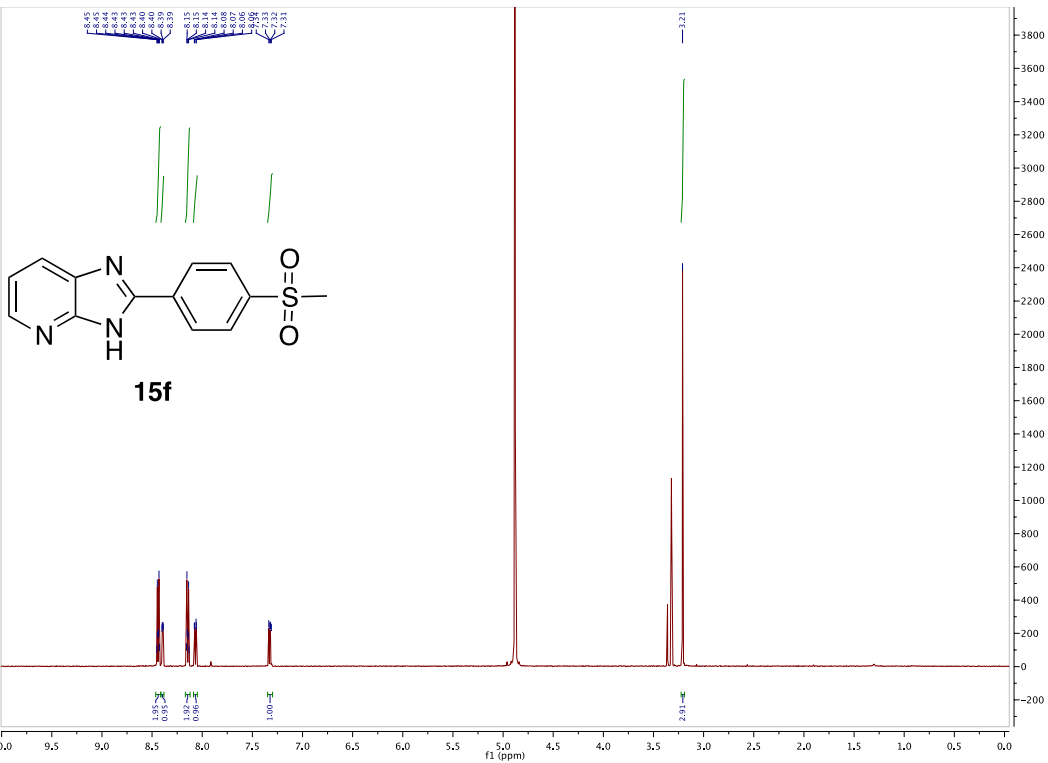
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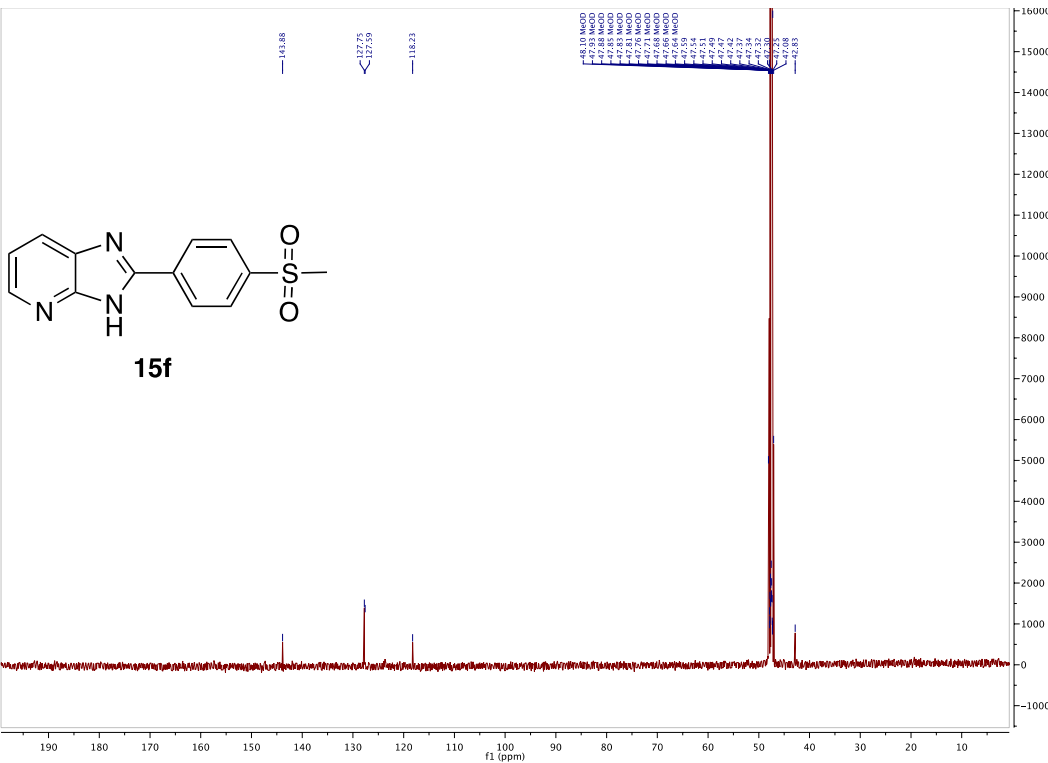
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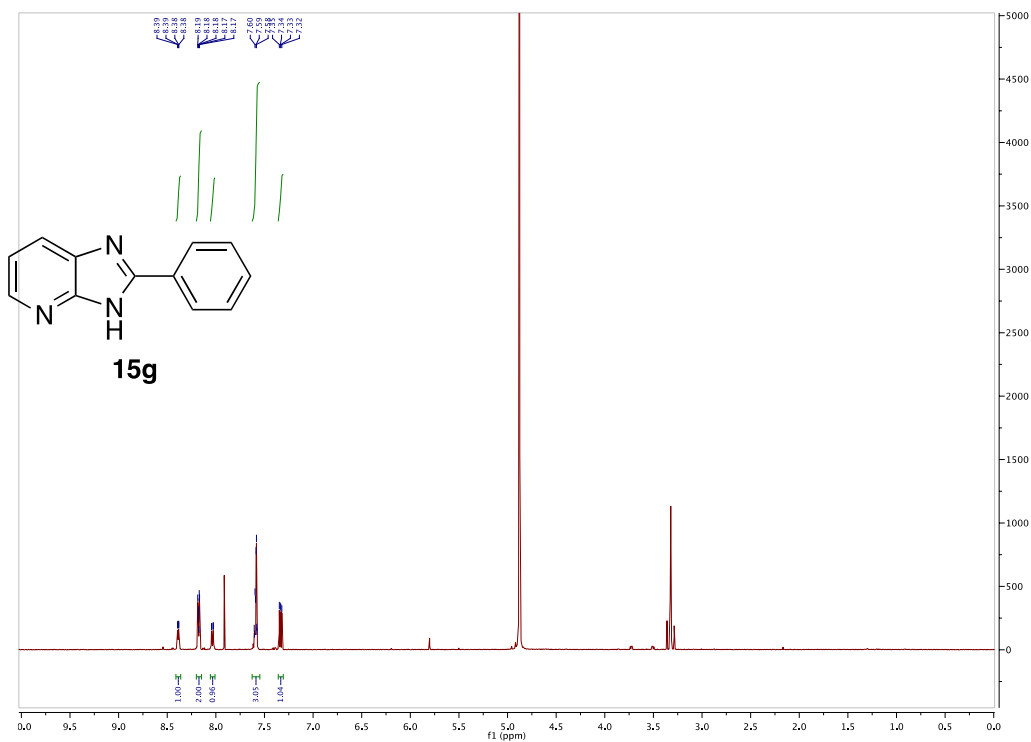
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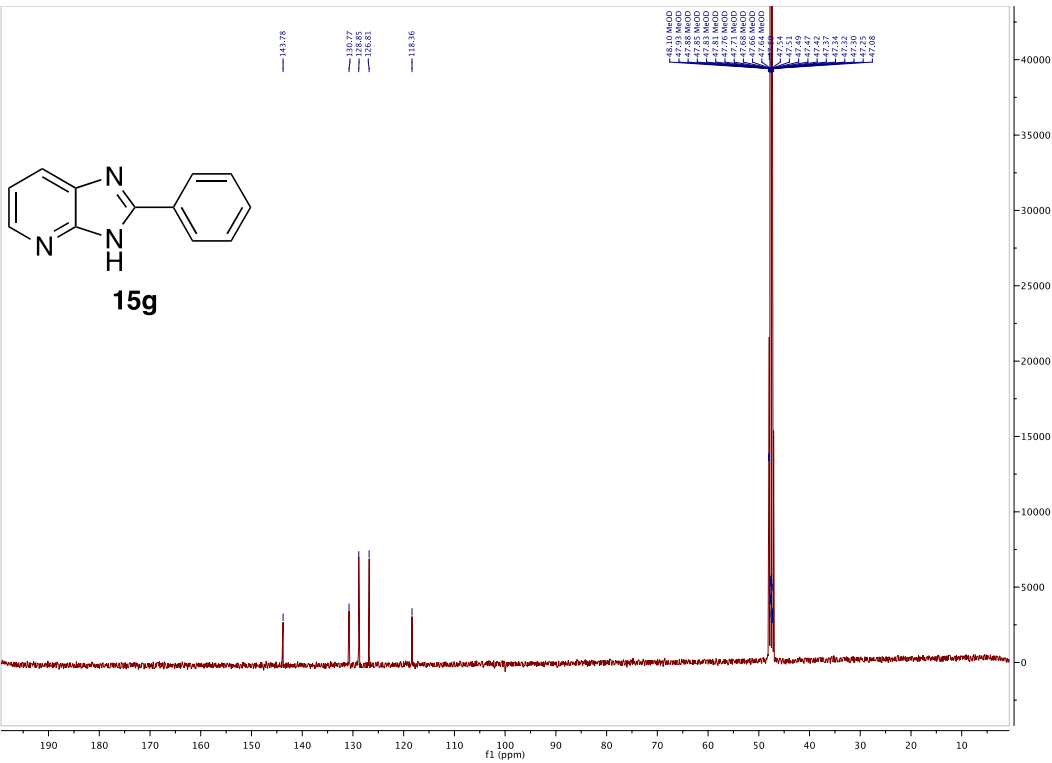
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<sup>1</sup>H NMR:

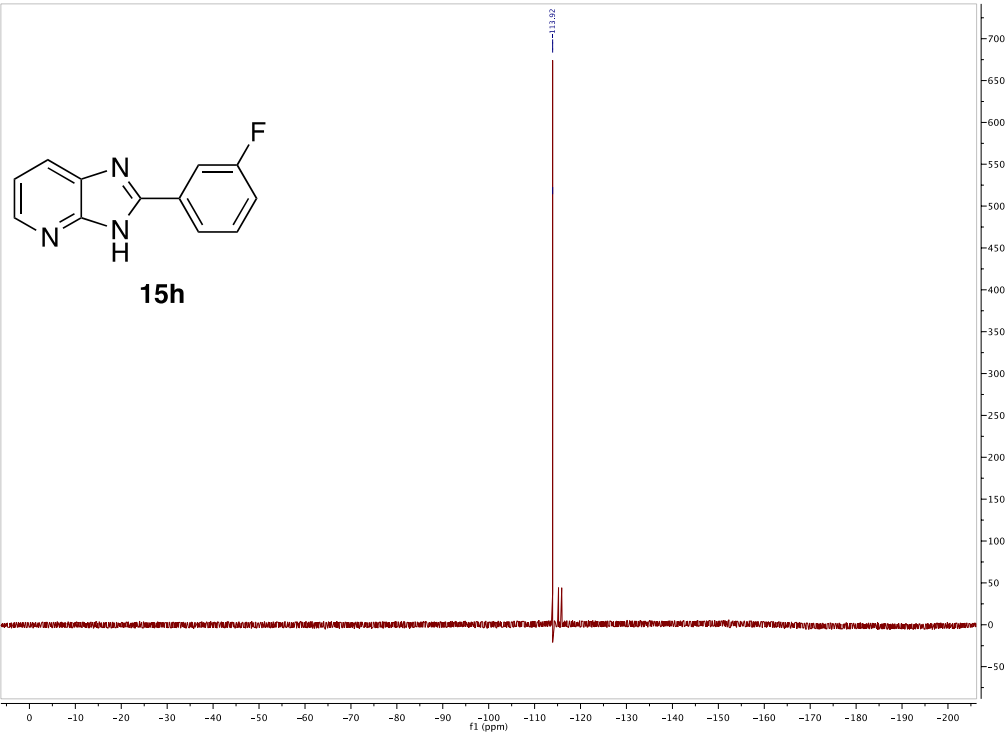


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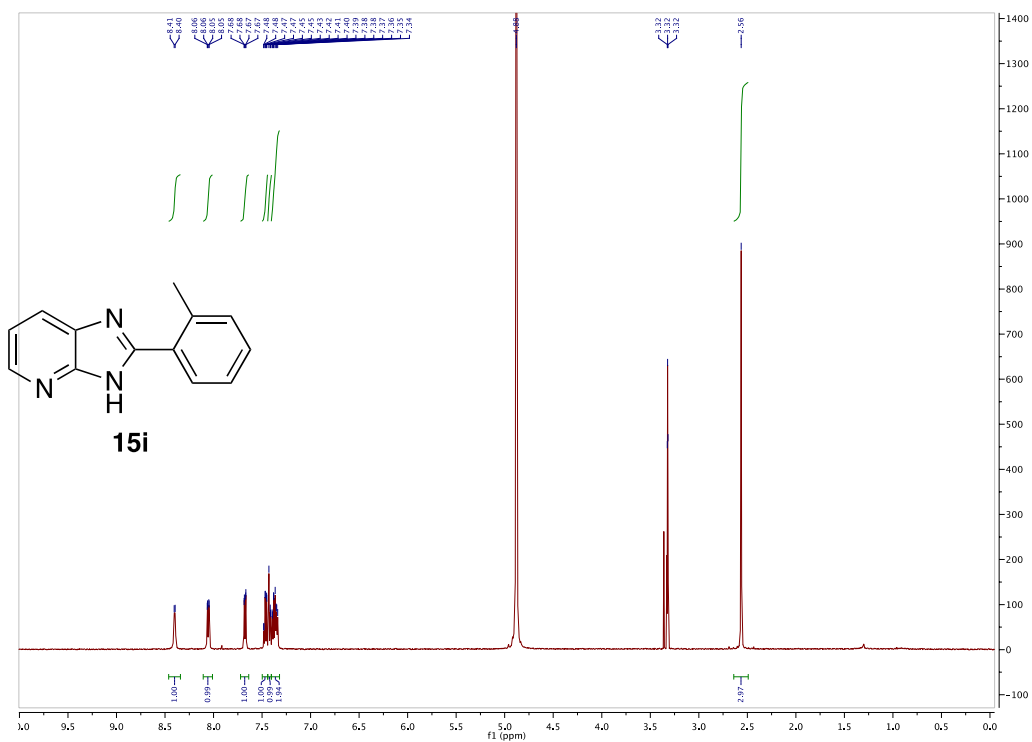




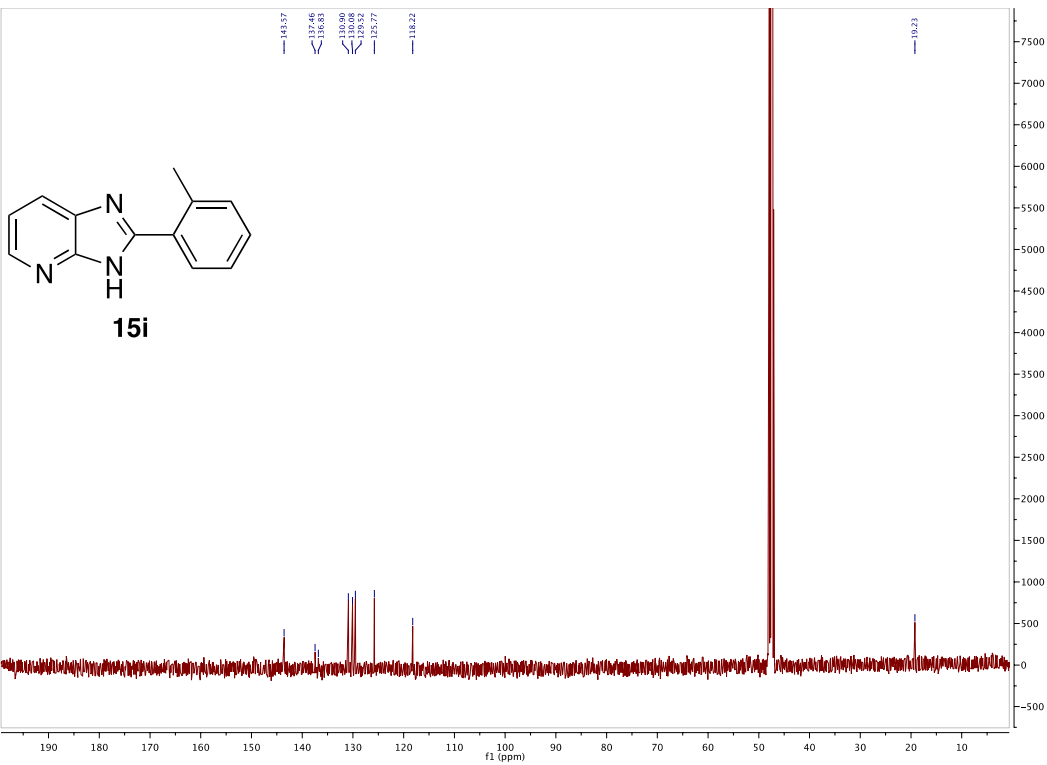
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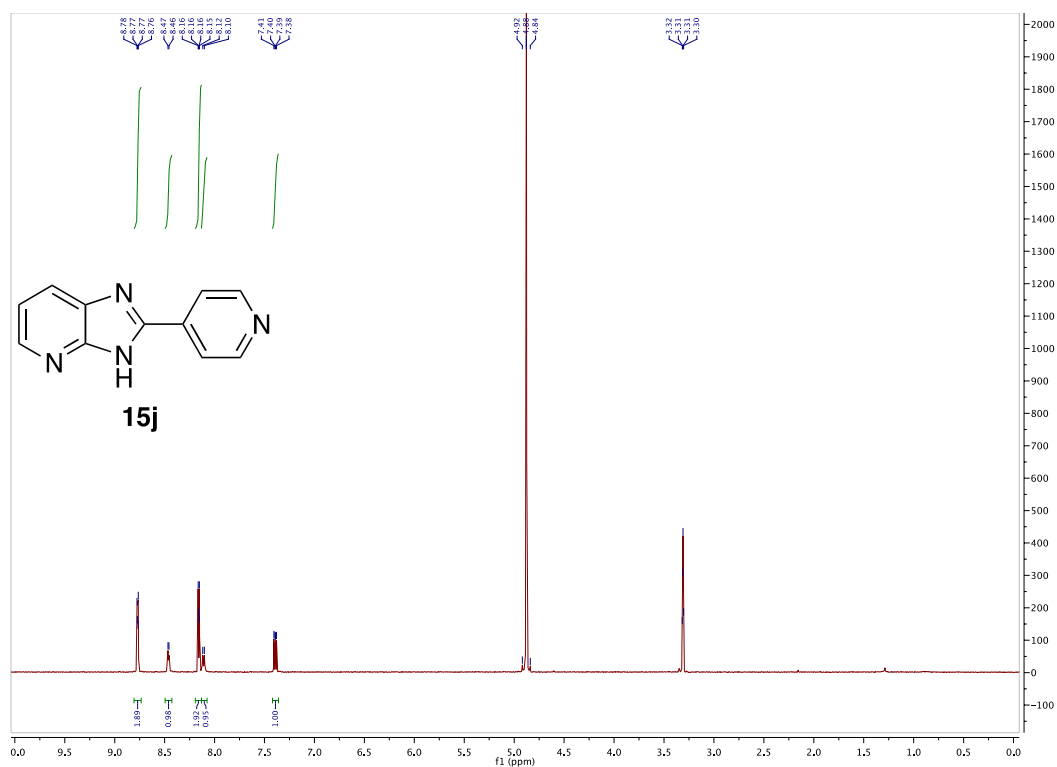
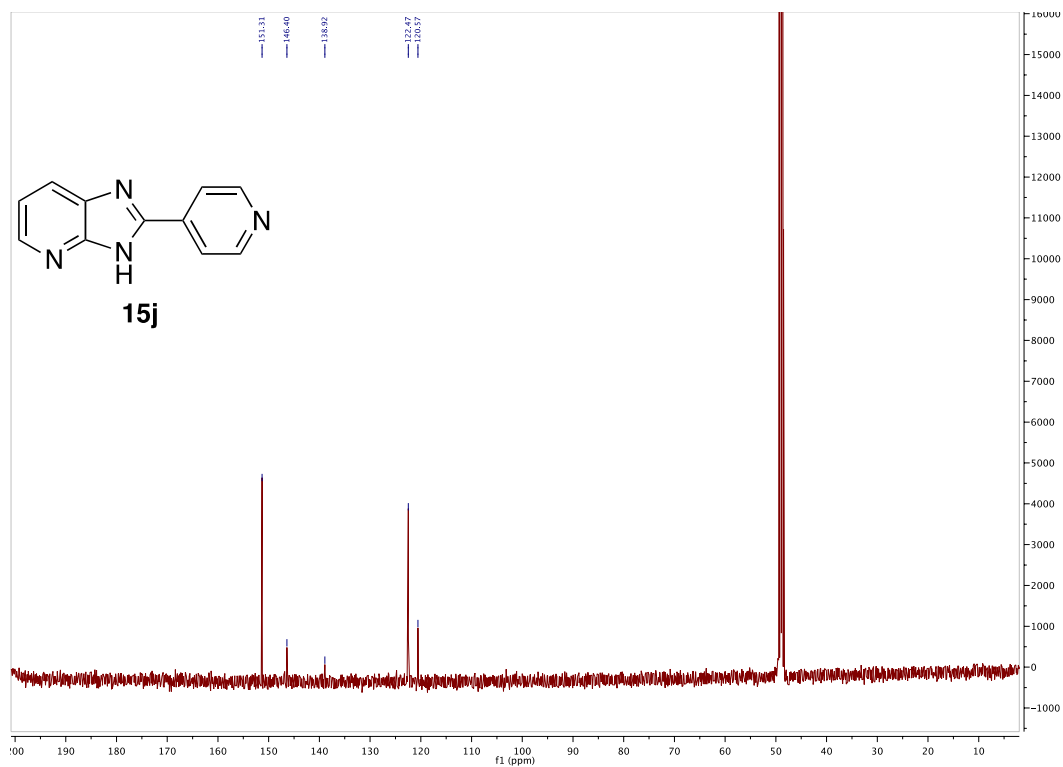


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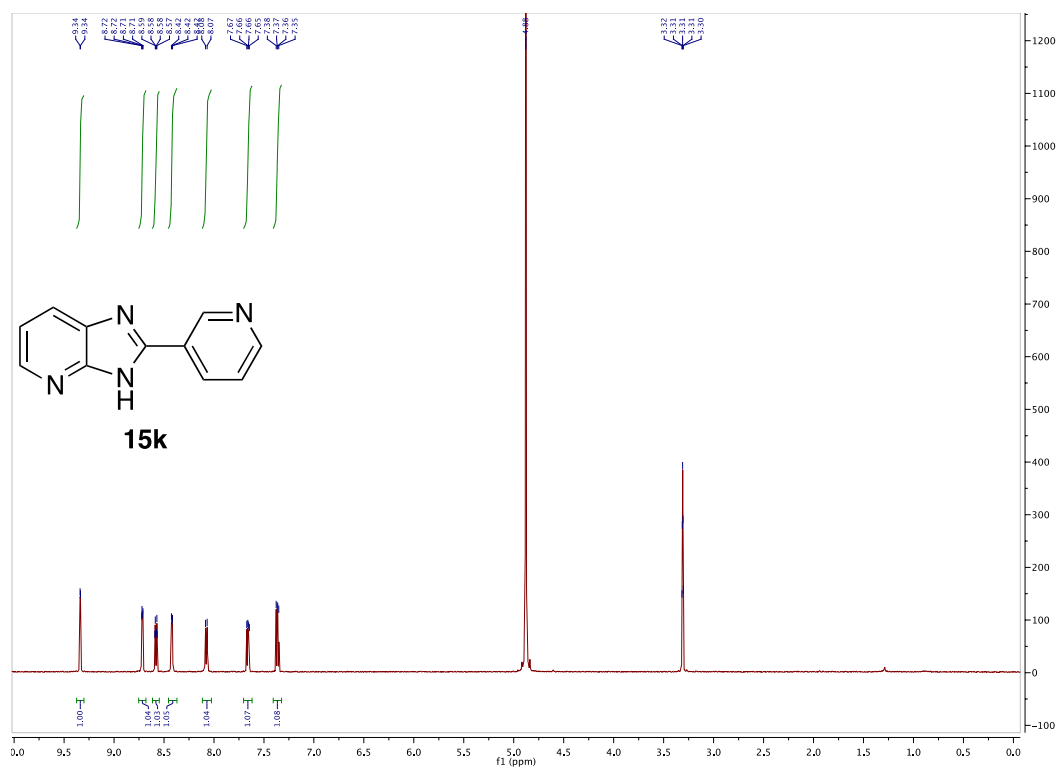
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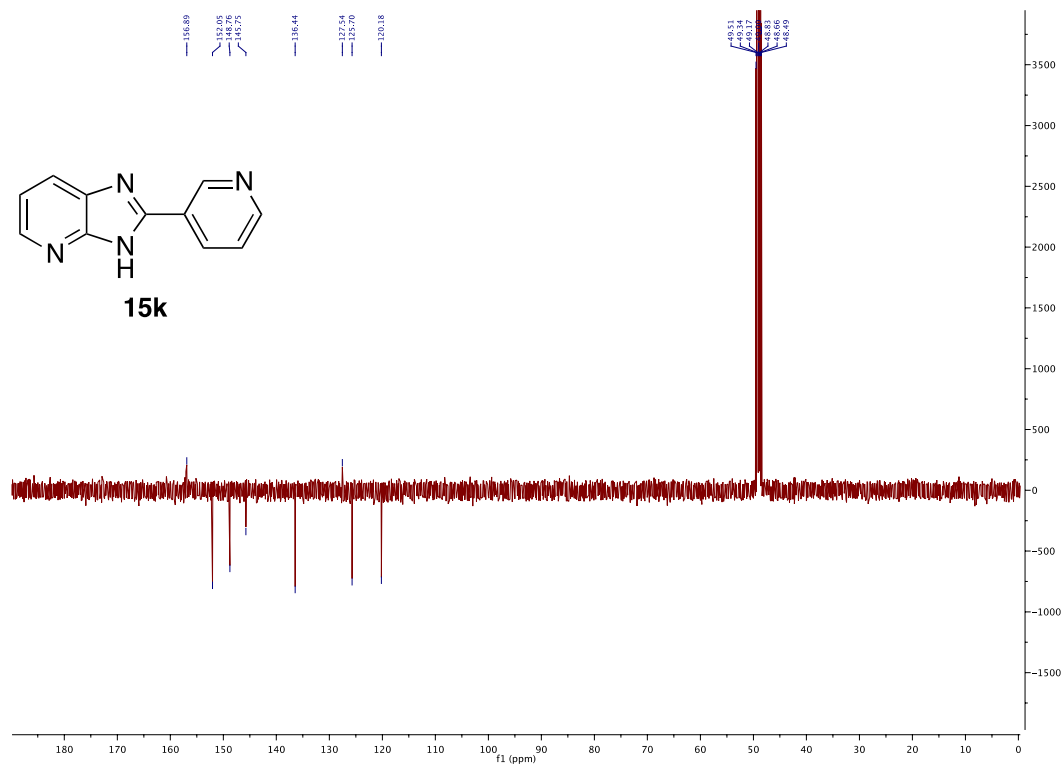
<sup>1</sup>H NMR:<sup>13</sup>C NMR:



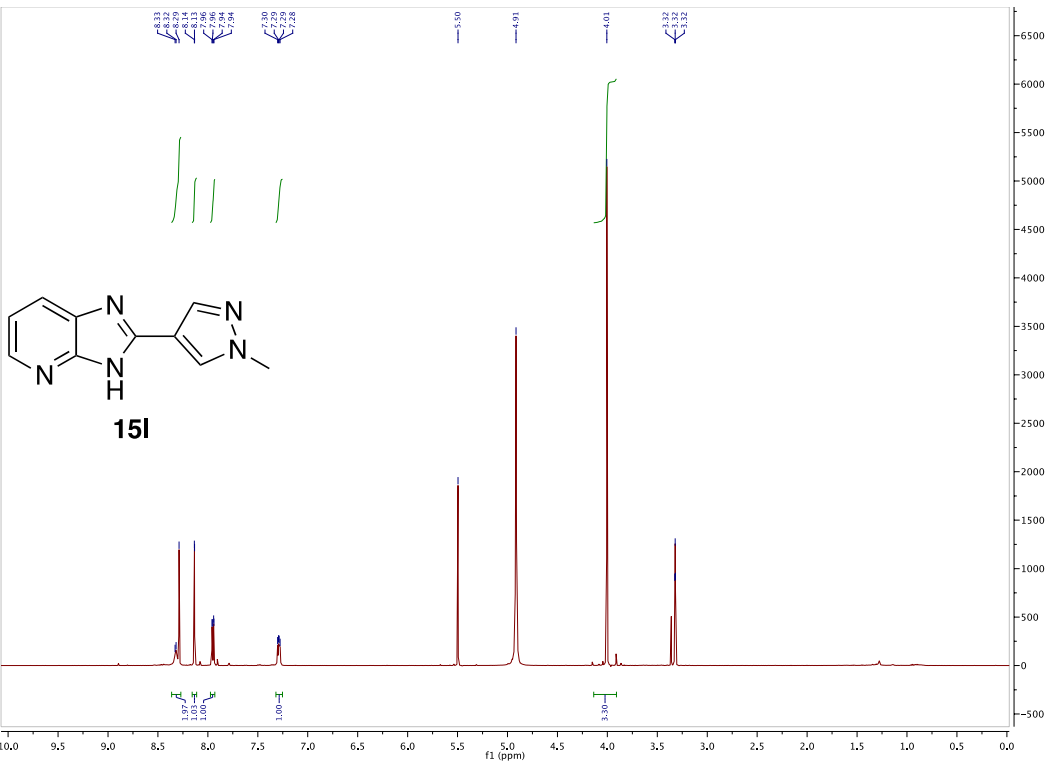
<sup>1</sup>H NMR:



<sup>13</sup>C NMR, DEPT:



<sup>1</sup>H NMR:



<sup>13</sup>C NMR:

