

Development of an *in situ* Culture-Free Screening Test for the Rapid Detection of *Staphylococcus aureus* within Healthcare Environments

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Bacterial Culture

Bacterial isolates were obtained from a clinical setting as a result of both MRSA (MRSA numbers 1-11,13,14,16,17 and 19) and MSSA (MSSA numbers 58,59,61,62, 63,64, 68, 69,71,72) infection, and were part of a cohort of bacteria taken from UK hospitals. *E.coli* samples were a combination of clinical isolates and wild type strains (*E.coli* 1-10), and coagulase negative Staphylococci include strains of *S.epidermidis*, *S. warneri*, *S.hominis* and *M. luteus*, and were obtained from clinical isolates from Sutton hospital. All bacteria were cultured initially on Brain Heart infusion agar, after which they were subcultured twice onto nutrient agar overnight at 37 °C, under aerobic conditions. Each bacterial isolate possesses individual characteristics, and thus displays a variance in the amount of coagulase produced, and therefore there is a degree of variance in results, lending to a greater amount of error in the results depicted.

Preparation of the LGX solution

A 100 µM and 50 µM solution of compound 3 was prepared by dissolving it in 2.5 % methanol followed by dilution in 1 x PBS. Solutions of Tris Base (0.05 M) and NaCl (0.1 M) in deionised H₂O were added to the solution until an optimum pH of 8.5 was achieved. The appropriate amount of human prothrombin in 1 x PBS was then added to produce final prothrombin concentrations of 83.6 nM and 41.8 nM, respectively. The resulting solution was termed “LGX”. The concentrations of LGX, subsequently discussed, refer to the concentration of the active constituent compound 3.

Testing procedure the efficacy of LGX as a selective and sensitive means of detecting *S. aureus*

In order to detect the presence of staphylocoagulase and thus the efficacy of LGX, varying cell concentrations (10^6 , 10^5 , 10^3 , 10^2 CFU/mL (50 µM LGX) and 10^4 , 10^3 , 10^2 , 10^0 CFU/mL (100 µM LGX) were added to a microtitre plate (Nunclon 96 well plates) in a 1:1 ratio with either 50 µM or 100 µM of prepared LGX solution. This provided a final LGX concentration of 25 µM or 50 µM, respectively. The relative fluorescence was then recorded every fifteen minutes over a six hour time period (λ_{ex} = 488 nm and λ_{em} = 525 nm). Positive controls used in each experiment were a control strain of MRSA (NCTC 12493), and the negative control was a clinical isolate of *E. coli*. A 1:1 ratio of both the 100 µM and 50 µM of LGX solution with 1 x PBS was used, and termed “LGX alone”. Fluorescence intensity was determined at 15 minute intervals over a 6 hour time period and n=3 samples were assayed in each case. We assayed 15 Strains of MRSA at varying cell concentration (10^6 , 10^5 , 10^3 , 10^2 CFU/mL (50 µM LGX) and 10^4 , 10^3 , 10^2 , 10^0 CFU/mL (100 µM LGX)) with equal volume of both 50 µM and 100 µM LGX, respectively, to give a final LGX concentration of 25 µM or 50 µM, respectively, in addition to a strain of *E.coli*, which served as a negative control as described above.

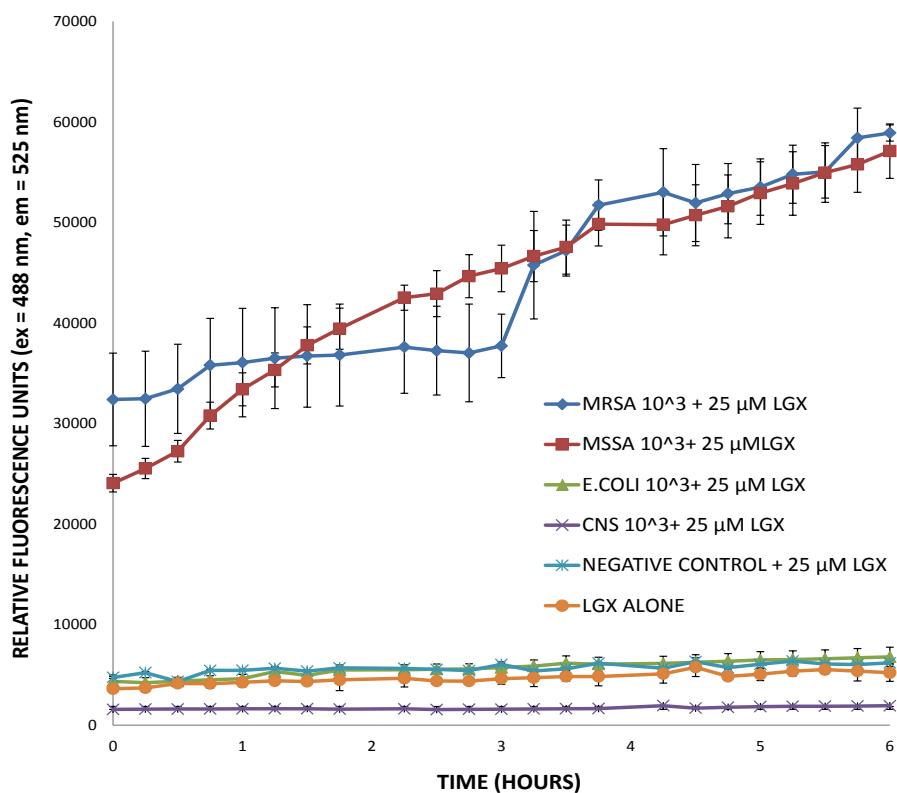


Figure 1 – The efficacy of 25 μ M LGX in detecting MRSA concentrations as low as 10^2 CFU/mL. Average of ten to fifteen bacterial strains (n=3). Error bars represent standard deviation.

As can be observed in figure 1, the results of all concentrations of MRSA (average of all 15 samples per data set illustrated) are plotted against a negative control and LGX alone, there is a significantly greater increase in fluorescence intensity in MRSA samples across the 6 hour time period. Taking a lower threshold limit of 20,000 relative fluorescence units (RFU) as a positive result, it can be seen that at a 25 μ M concentration of LGX, and at all 10^3 CFU/mL MRSA/MSSA, there is a clear positive result yet, however as expected, this is lower than the effect observed with the 50 μ M LGX. As with the 50 μ M data an instantaneous positive reaction can also be observed. This data shows that the compound harbours the ability to rapidly detect SA at low concentrations.

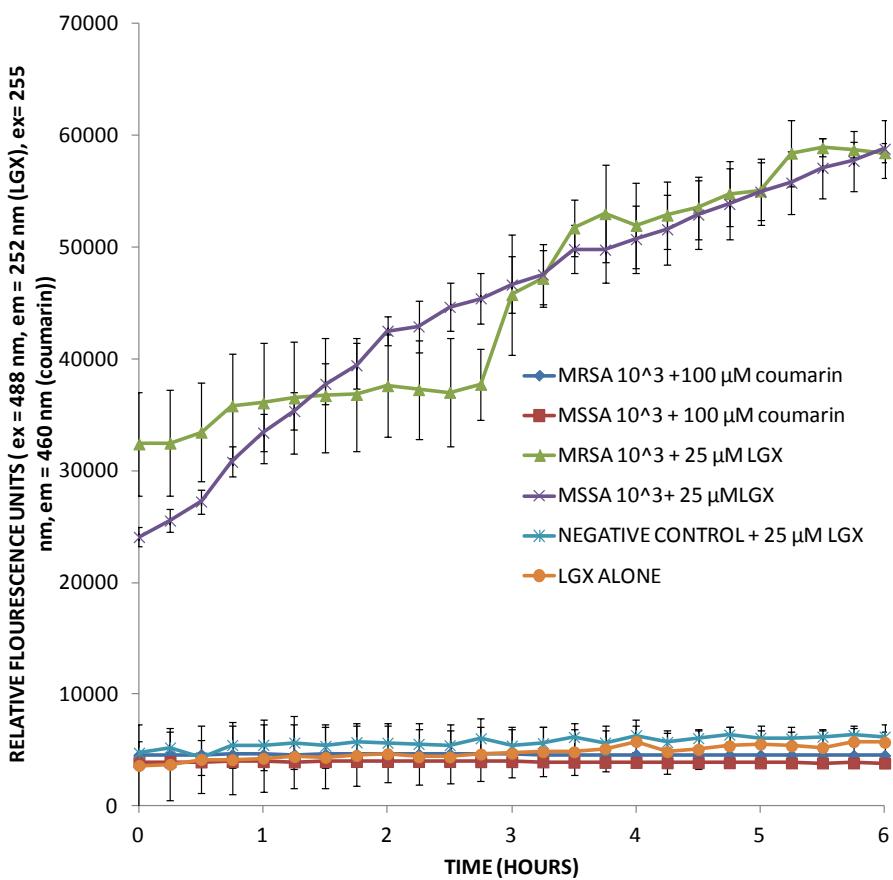
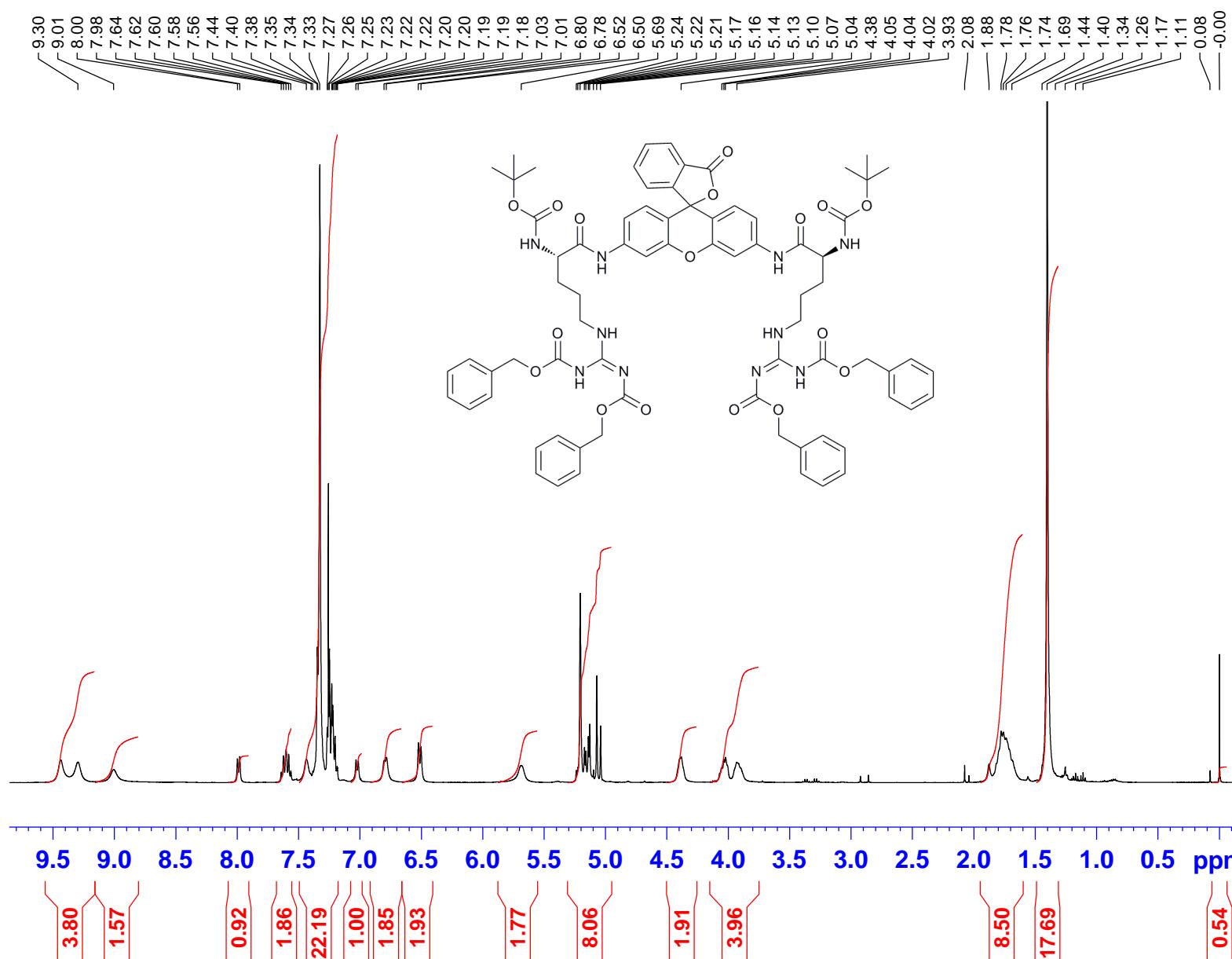


Figure 2 - The efficacy of 25 μM LGX at demonstrating both selectivity and sensitivity in detecting coagulase positive MRSA and MSSA, compared to the efficacy 100 μM coumarin, in bacterial concentrations of 10^3 CFU/mL. Average of ten to fifteen bacterial strains (n=3). Error bars represent standard deviation.

The data demonstrated in Figure 2 illustrates that, similarly to 50 μM LGX and 500 μM coumarin, there is significantly more fluorescence produced by LGX, when compared to the coumarin system. These results show that at low concentrations, the LGX system demonstrates considerable efficacy in terms of speed of detection, as well sensitivity, and is a vast improvement on the existing coumarin system.

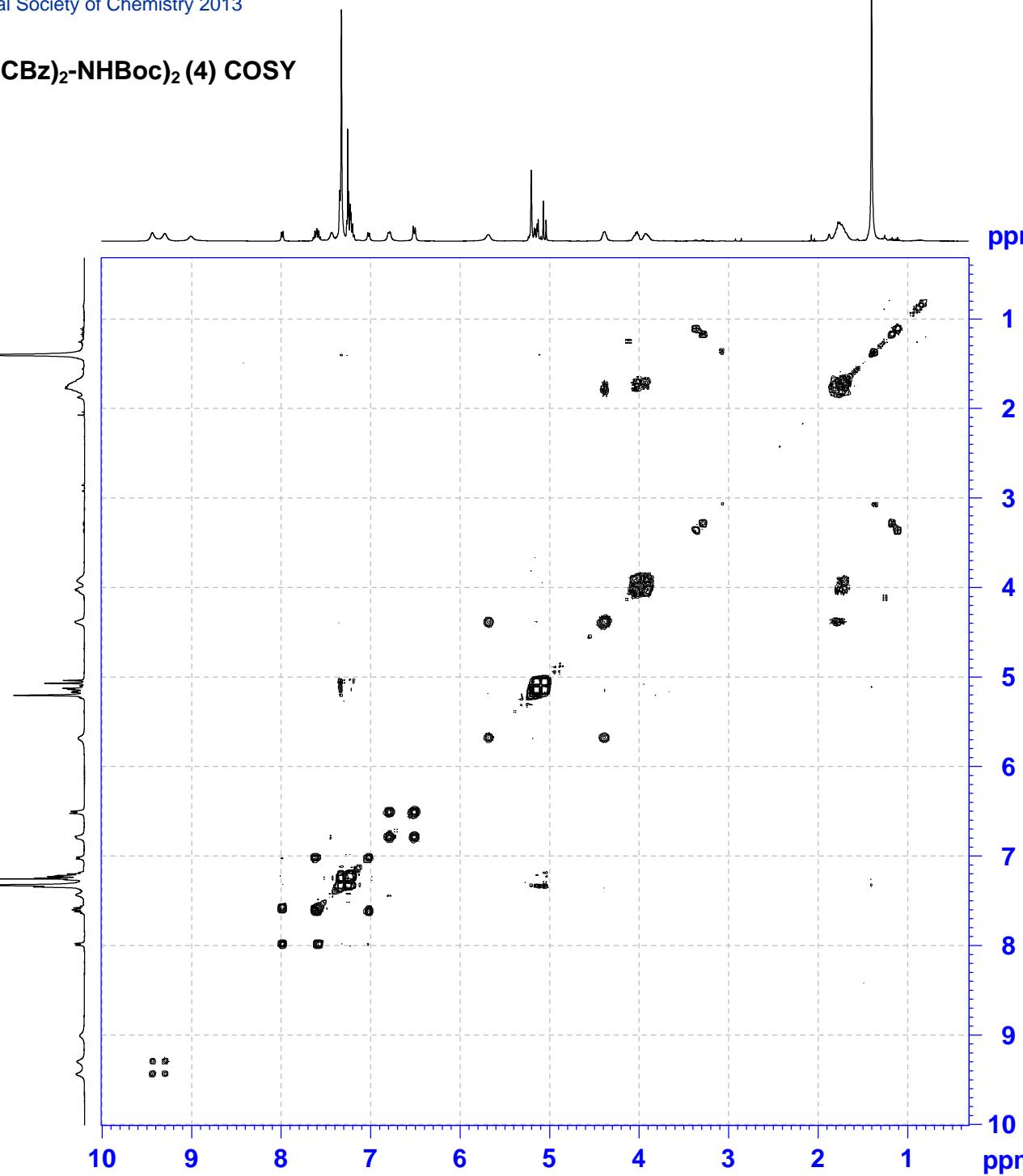
Rhod-(Arg-(CBz)₂-NHBoc)₂ (4)



NAME LG1-51 CHCl₃ C
EXPNO 30
PROCNO 1
Date_ 20101111
Time 20.08
INSTRUM av400
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl₃
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 90.5
DW 60.800 usec
DE 15.82 usec
TE 297.1 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 15.00 usec
PL1 -3.50 dB
PL1W 16.47660065 W
SFO1 399.7324685 MHz
SI 65536
SF 399.7300133 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

Rhod-(Arg-(CBz)₂-NH_{Boc})₂ (4) COSY

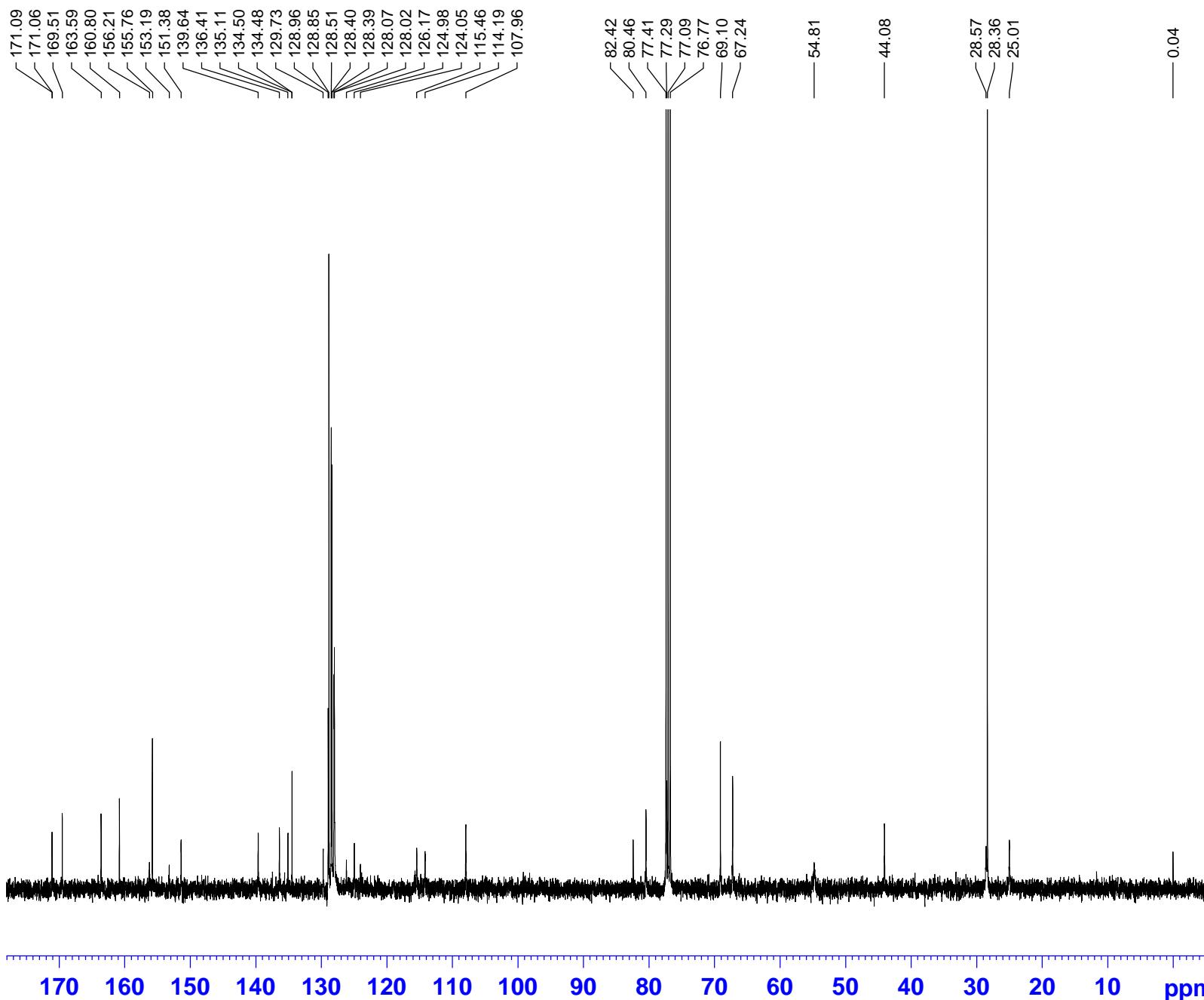


NAME LG1-51 CHCl₃ C
EXPNO 34
PROCNO 1
Date_ 20101111
Time 20.58
INSTRUM av400
PROBHD 5 mm PABBO BB-
PULPROG cosygppmfcf
TD 2048
SOLVENT CDCl₃
NS 2
DS 8
SWH 3875.969 Hz
FIDRES 1.892563 Hz
AQ 0.2642420 sec
RG 2050
DW 129.000 usec
DE 6.50 usec
TE 297.2 K
D0 0.00000300 sec
D1 1.92750096 sec
D13 0.00000400 sec
D16 0.00020000 sec
IN0 0.00025800 sec

===== CHANNEL f1 =====
NUC1 1H
P1 15.00 usec
PL1 -3.50 dB
PL1W 16.47660065 W
SFO1 399.7320771 MHz

===== GRADIENT CHANNEL =====
GPNAM1 SINE.100
GPNAM2 SINE.100
GPNAM3 SINE.100
GPZ1 16.00 %
GPZ2 12.00 %
GPZ3 40.00 %
P16 1000.00 usec
ND0 1
TD 128
SFO1 399.7321 MHz
FIDRES 30.281008 Hz
SW 9.696 ppm
FnMODE QF
SI 1024
SF 399.7300133 MHz
WDW SINE
SSB 0
LB 0.00 Hz
GB 0
PC 1.40
SI 1024
MC2 QF
SF 399.7300133 MHz
WDW SINE
SSB 0
LB 0.00 Hz
GB 0

Rhod-(Arg-(CBz)₂-NH_{Boc})₂ (4)



BRUKER

NAME LG1-51 CHCl₃ C
EXPNO 32
PROCNO 1
Date_ 20101111
Time 20.40
INSTRUM av400
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl₃
NS 256
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631988 sec
RG 2050
DW 20.800 usec
DE 9.66 usec
TE 297.1 K
D1 2.0000000 sec
D11 0.0300000 sec
TD0 1

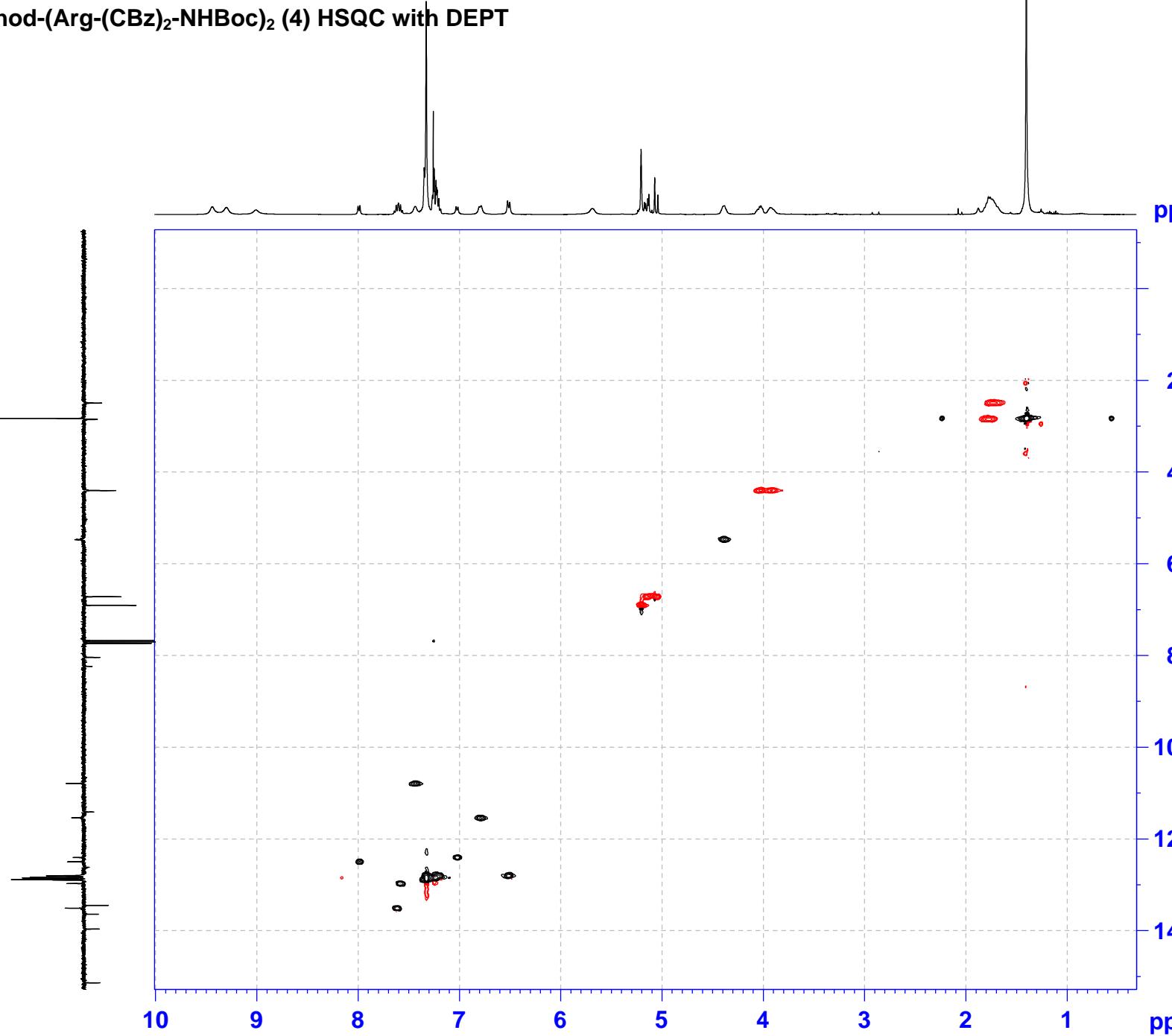
===== CHANNEL f1 =====
NUC1 13C
P1 9.60 usec
PL1 -3.70 dB
PL1W 74.19429779 W
SFO1 100.5222397 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -3.50 dB
PL12 11.04 dB
PL13 14.00 dB
PL2W 16.47660065 W
PL12W 0.57925212 W
PL13W 0.29300001 W
SFO2 399.7315989 MHz
SI 65536
SF 100.5121880 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

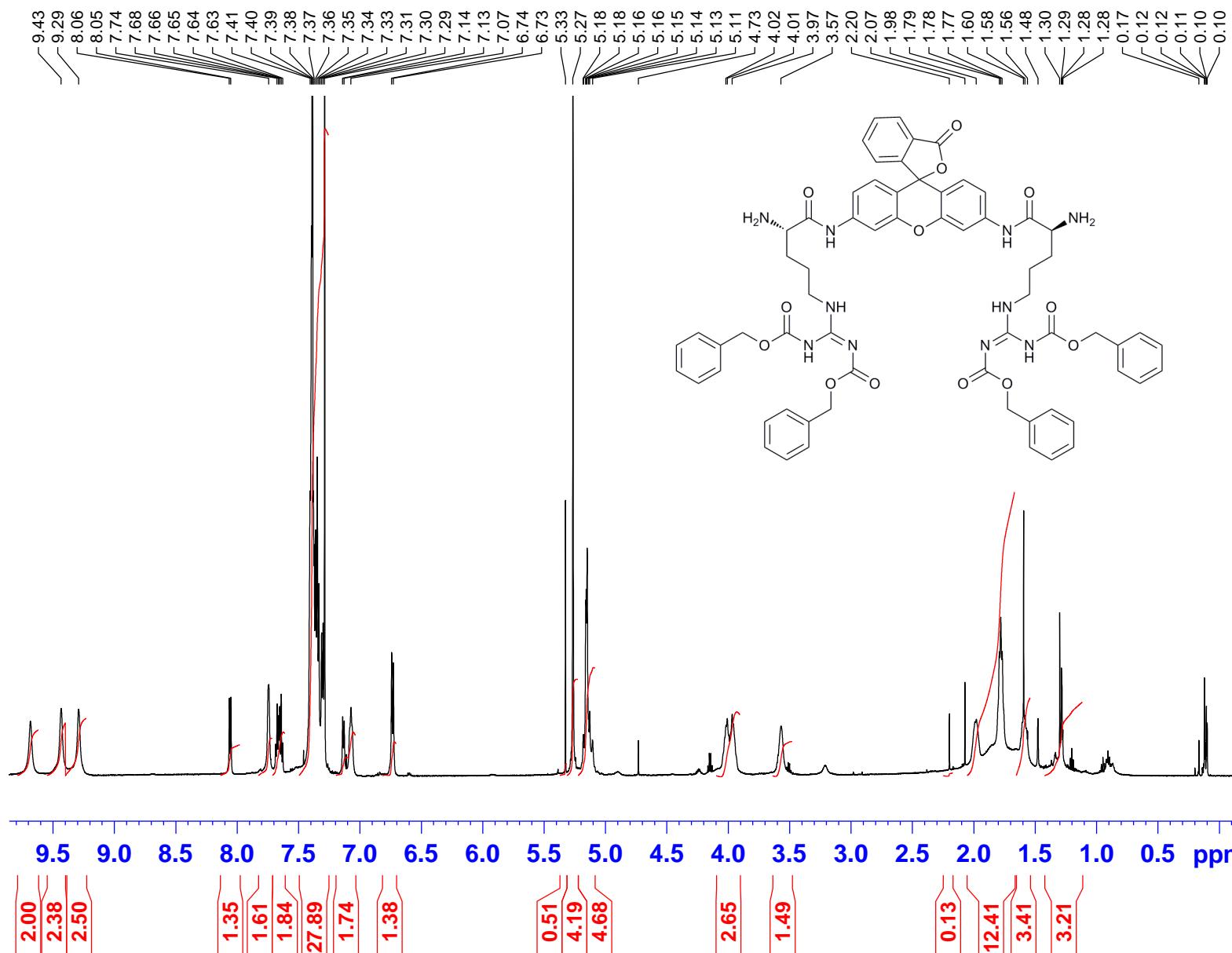
Rhod-(Arg-(CBz)₂-NH_{Boc})₂ (4) HSQC with DEPT



NAME LG1-51 CHCl₃ C
EXPNO 31
PROCNO 1
Date 20101111
Time 20.10
INSTRUM av400
PROBHD 5 mm PABBO BB-
PULPROG hsqcetgpsp.3
TD 1024
SOLVENT CDCl₃
NS 2
DS 16
SWH 3875.969 Hz
FIDRES 3.785126 Hz
AQ 0.1321460 sec
RG 2050
DW 129.000 usec
DE 6.50 usec
TE 297.2 K
CNUST2 145.000000
D0 0.000003 sec
D1 1.44 0.592 sec
D4 0.00172414 sec
D11 0.03000000 sec
D16 0.00020000 sec
D21 0.00360000 sec
IN0 0.00003005 sec
L31 1
===== CHANNEL f1 =====
NUC1 1H
P1 15.00 usec
P2 30.00 usec
P28 1.00 usec
PL1 -3.50 dB
PL1W 16.47660065 W
SF01 399.7320771 MHz
===== CHANNEL f2 =====
CPDPRG2 bi_p5m4sp_4sp.2
NUC2 ¹³C
P3 9.60 usec
P14 500.00 usec
P31 1900.00 usec
P63 1500.00 usec
PL0 120.00 dB
PL2 -3.00 dB
PL12 1.50 dB
PL0W 0.0000000 W
PL2W 74.19429779 W
PL12W 1.39434588 W
SF02 100.5192243 MHz
SP3 4.81 dB
SP14 6.29 dB
SP18 11.02 dB
SP31 12.31 dB
SPNAM3 Crp60_0.5_20.1
SPNAM14 Crp32_1.5_20.2
SPNAM8 Crp60_xflit.2
SPNAM31 Crp32_1.5_20.2
SPDAL3 0.500
SPDAL14 0.500
SPDAL18 0.500
SPDAL31 0.500
SPOFFS3 0.00 Hz
SPOFFS14 0.00 Hz
SPOFFS18 0.00 Hz
SPOFFS31 0.00 Hz
===== GRADIENT CHANNEL =====
GPNAME1 SINE_100
GPNAME2 SINE_100
GPZ1 20.00 %
GPZ2 20.10 %
P16 1000.00 usec
ND0 2
TD 256
SF01 100.5192 MHz
FIDRES 65.046150 Hz
SW 165.658 ppm
FnMODE Echo-Antiecho
SI 1024
SF 399.7300133 MHz
WDW QSINE
SSB 2
LB 0.00 Hz
GB 0
PC 1.40
SI 1024
MC2 echo-antiecho
SF 100.5121880 MHz
WDW QSINE
SSB 2
LB 0.00 Hz
GB 0



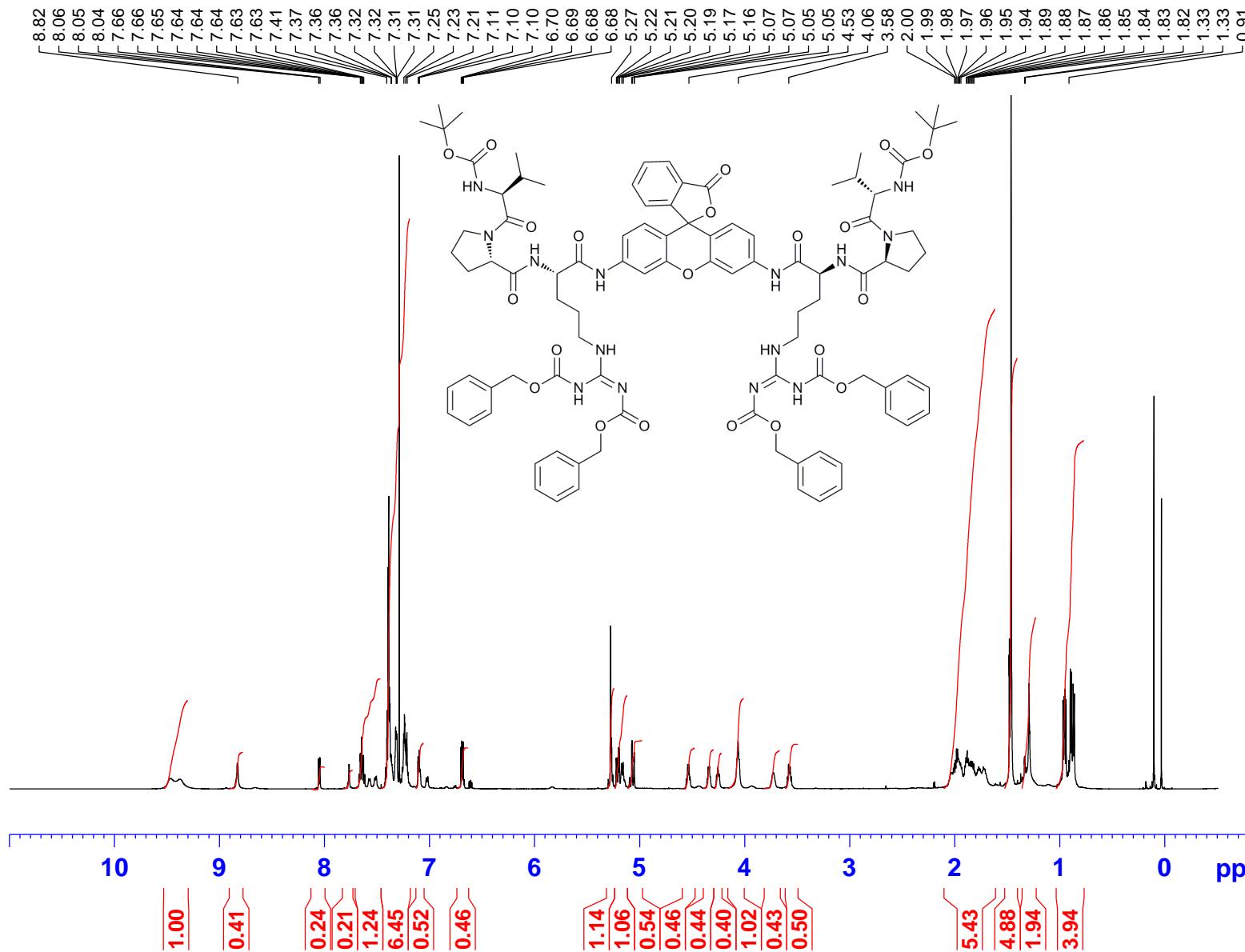
Rhodamine-(Arg-CBz)₂-NH₂)₂ (6)



NAME LG1-41
EXPNO 10
PROCNO 1
Date_ 20101004
Time 12.48
INSTRUM av600
PROBHD 5 mm PATXI 1H/
PULPROG zg30
TD 65536
SOLVENT CDCl₃
NS 16
DS 2
SWH 12345.679 Hz
FIDRES 0.188380 Hz
AQ 2.6542580 sec
RG 228
DW 40.500 usec
DE 11.34 usec
TE 298.1 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 7.00 usec
PL1 0.25 dB
PL1W 16.59961510 W
SFO1 600.1337060 MHz
SI 65536
SF 600.1300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

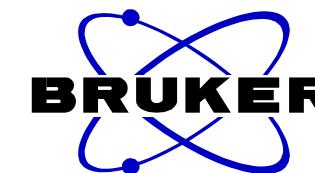
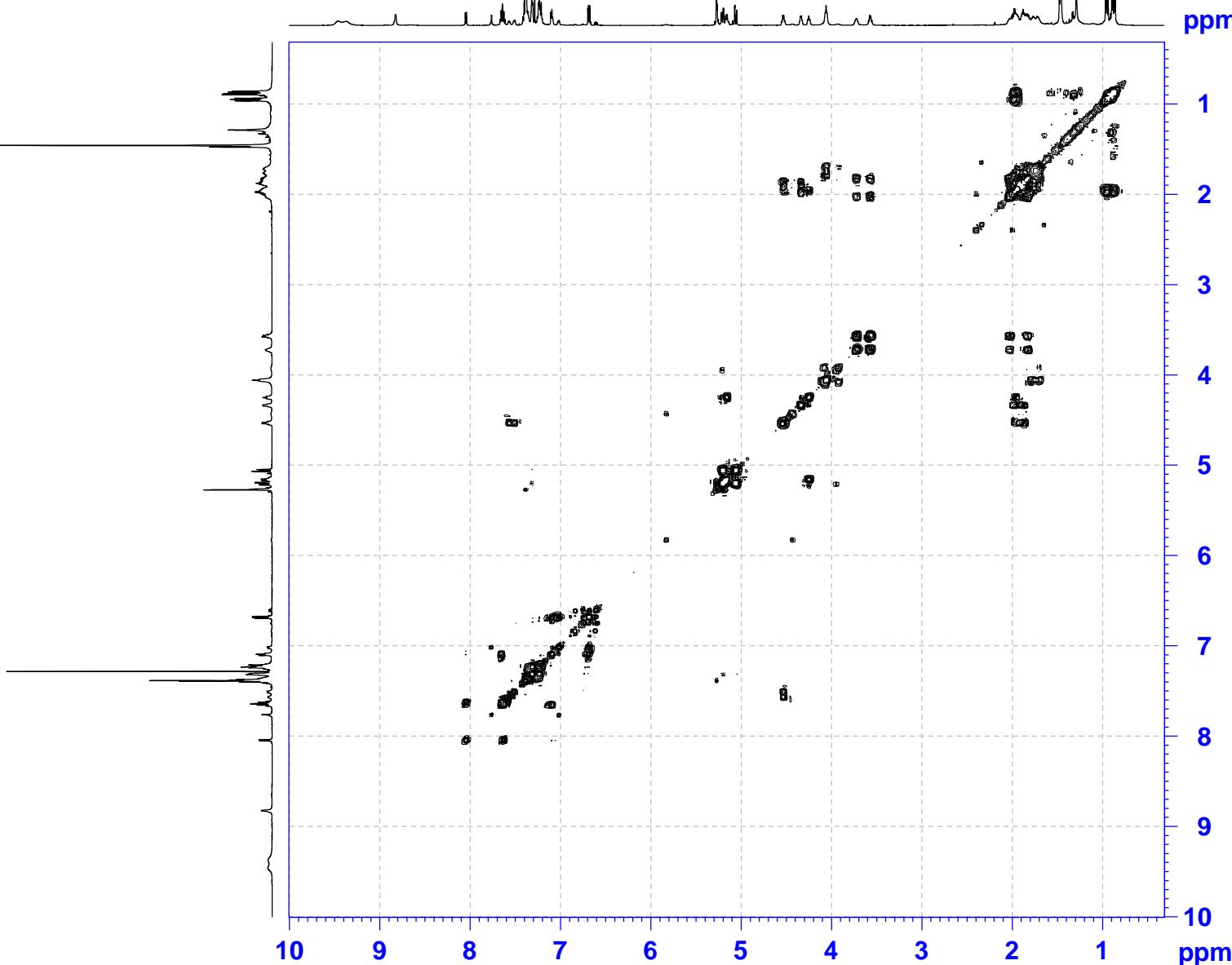
Rhodamine-(Arg-(Cbz)₂-Pro-Val-NHBoc)₂ (5)



NAME LG1-57
EXPNO 10
PROCNO 1
Date_ 20101124
Time 14.54
INSTRUM av600
PROBHD 5 mm PATXI 1H/
PULPROG zg30
TD 65536
SOLVENT CDCl₃
NS 16
DS 2
SWH 8417.509 Hz
FIDRES 0.128441 Hz
AQ 3.8928883 sec
RG 57
DW 59.400 usec
DE 6.50 usec
TE 308.0 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 ======
NUC1 1H
P1 7.00 usec
PL1 0.25 dB
PL1W 16.59961510 W
SFO1 600.1339008 MHz
SI 32768
SF 600.1300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

Rhodamine-(Arg-(Cbz)₂-Pro-Val-NHBoc)₂ (5) gradient COSY

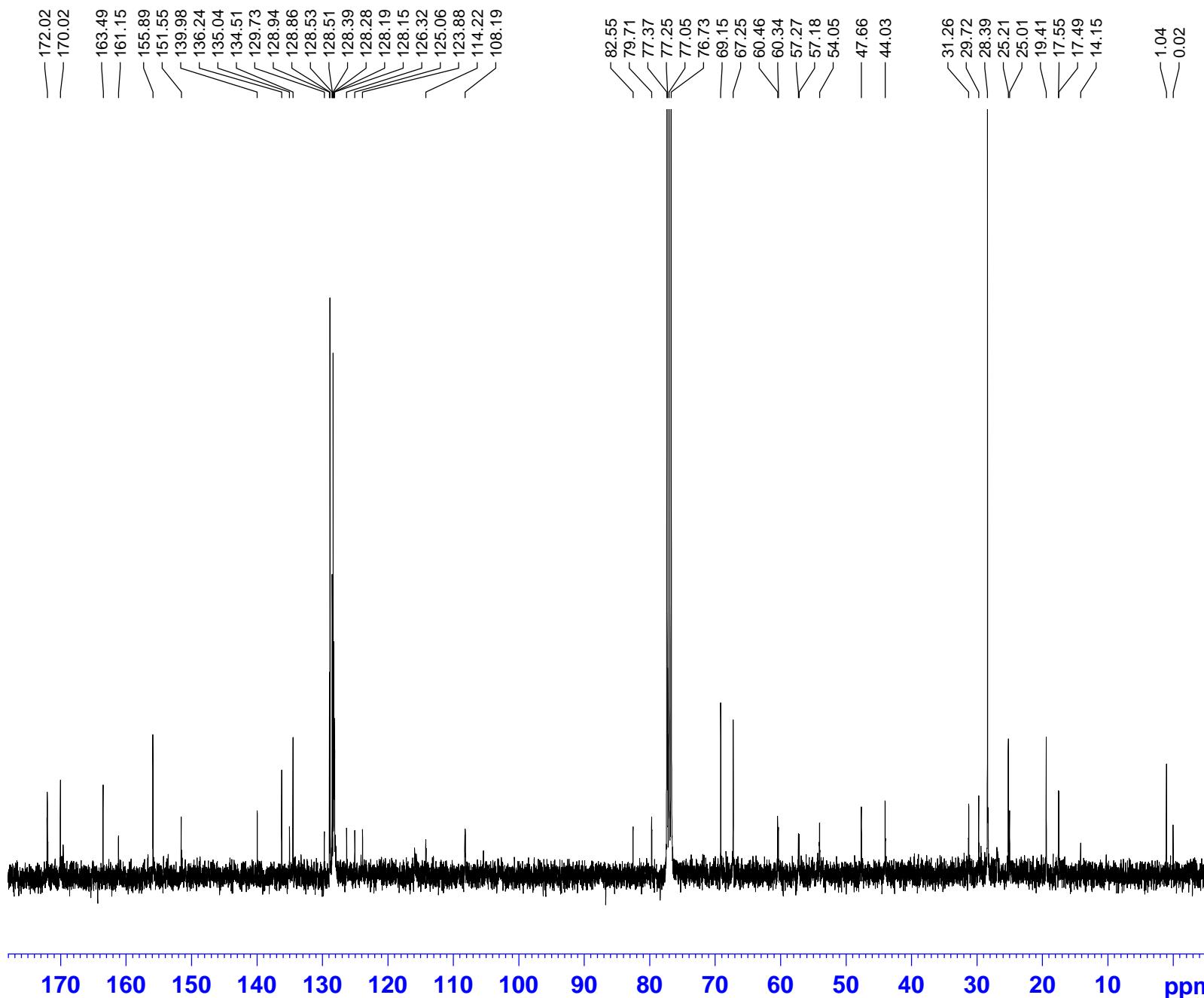


NAME LG1-57
EXPNO 13
PROCNO 1
Date_ 20101124
Time 15.31
INSTRUM av600
PROBHD 5 mm PATXI 1H/
PULPROG cosygppmfcf
TD 2048
SOLVENT CDCl₃
NS 2
DS 8
SWH 6009.615 Hz
FIDRES 2.934382 Hz
AQ 0.1704436 sec
RG 2050
DW 83.200 usec
DE 6.50 usec
TE 308.0 K
D0 0.00000300 sec
D1 2.00000000 sec
D13 0.00000400 sec
D16 0.00020000 sec
INO 0.00016640 sec

===== CHANNEL f1 =====
NUC1 1H
P1 7.00 usec
PL1 0.25 dB
PL1W 16.59961510 W
SFO1 600.1330006 MHz

===== GRADIENT CHANNEL =====
GPNAM1 SINE.100
GPNAM2 SINE.100
GPNAM3 SINE.100
GPZ1 16.00 %
GPZ2 12.00 %
GPZ3 40.00 %
P16 1000.00 usec
ND0 1
TD 128
SFO1 600.133 MHz
FIDRES 46.950092 Hz
SW 10.014 ppm
FnMODE QF
SI 1024
SF 600.1300000 MHz
WDW SINE
SSB 0
LB 0.00 Hz
GB 0
PC 1.00
SI 1024
MC2 QF
SF 600.1300000 MHz
WDW SINE
SSB 0
LB 0.00 Hz
GB 0

Rhodamine-(Arg-(Cbz)₂-Pro-Val-NHBoc)₂ (5)



BRUKER

NAME LG1-57carb
EXPNO 10
PROCNO 1
Date_ 20101125
Time 9.42
INSTRUM av400
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl₃
NS 1024
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631988 sec
RG 2050
DW 20.800 usec
DE 9.66 usec
TE 296.9 K
D1 2.0000000 sec
D11 0.0300000 sec
TD0 1

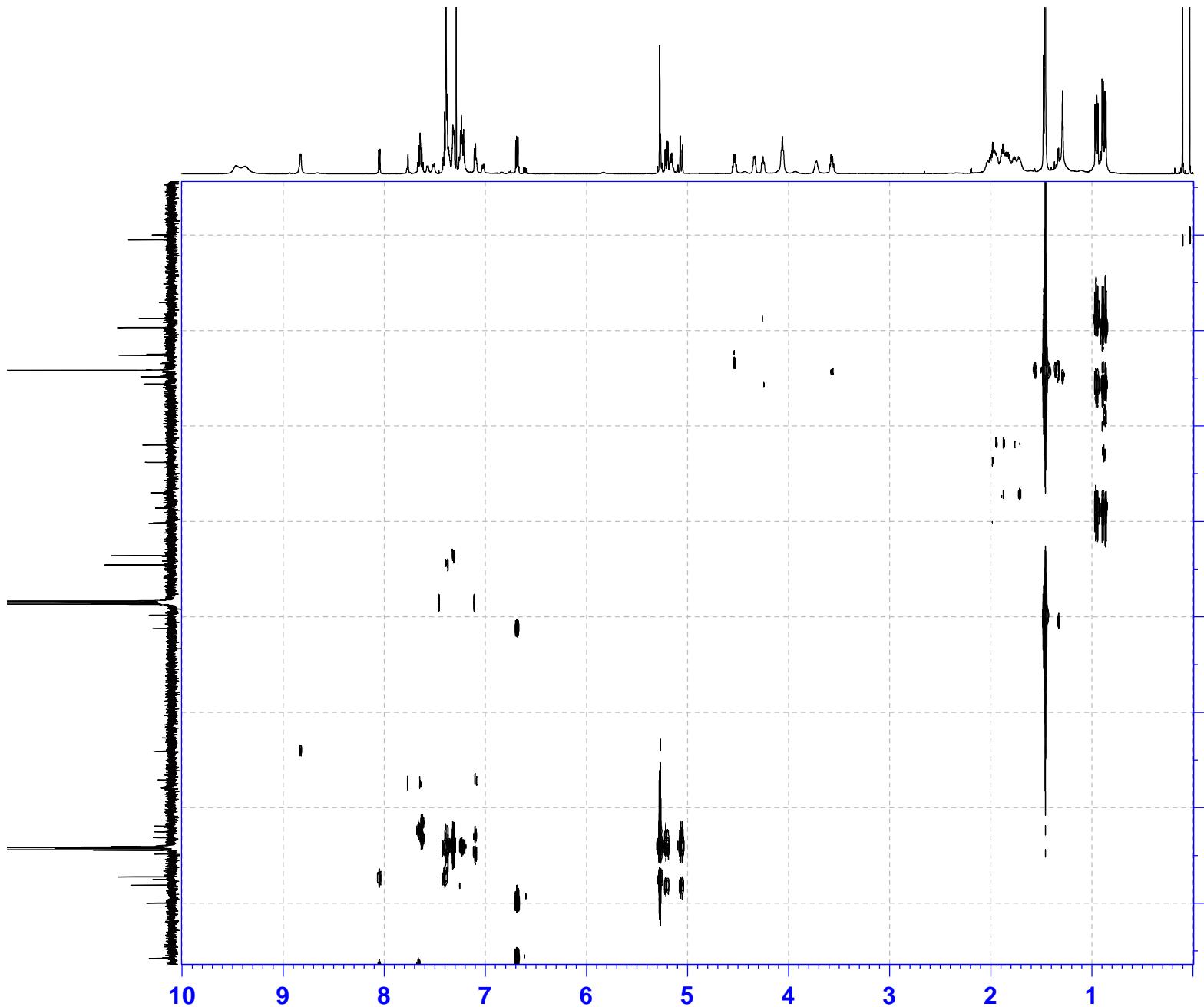
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NUC1 13C
P1 9.60 usec
PL1 -3.70 dB
PL1W 74.19429779 W
SFO1 100.5222397 MHz

===== CHANNEL f2 ======
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -3.50 dB
PL12 11.04 dB
PL13 14.00 dB
PL2W 16.47660065 W
PL12W 0.57925212 W
PL13W 0.29300001 W
SFO2 399.7315989 MHz
SI 65536
SF 100.5121880 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Rhodamine-(Arg-(Cbz)₂-Pro-Val-NHBoc)₂ (5) HMBC



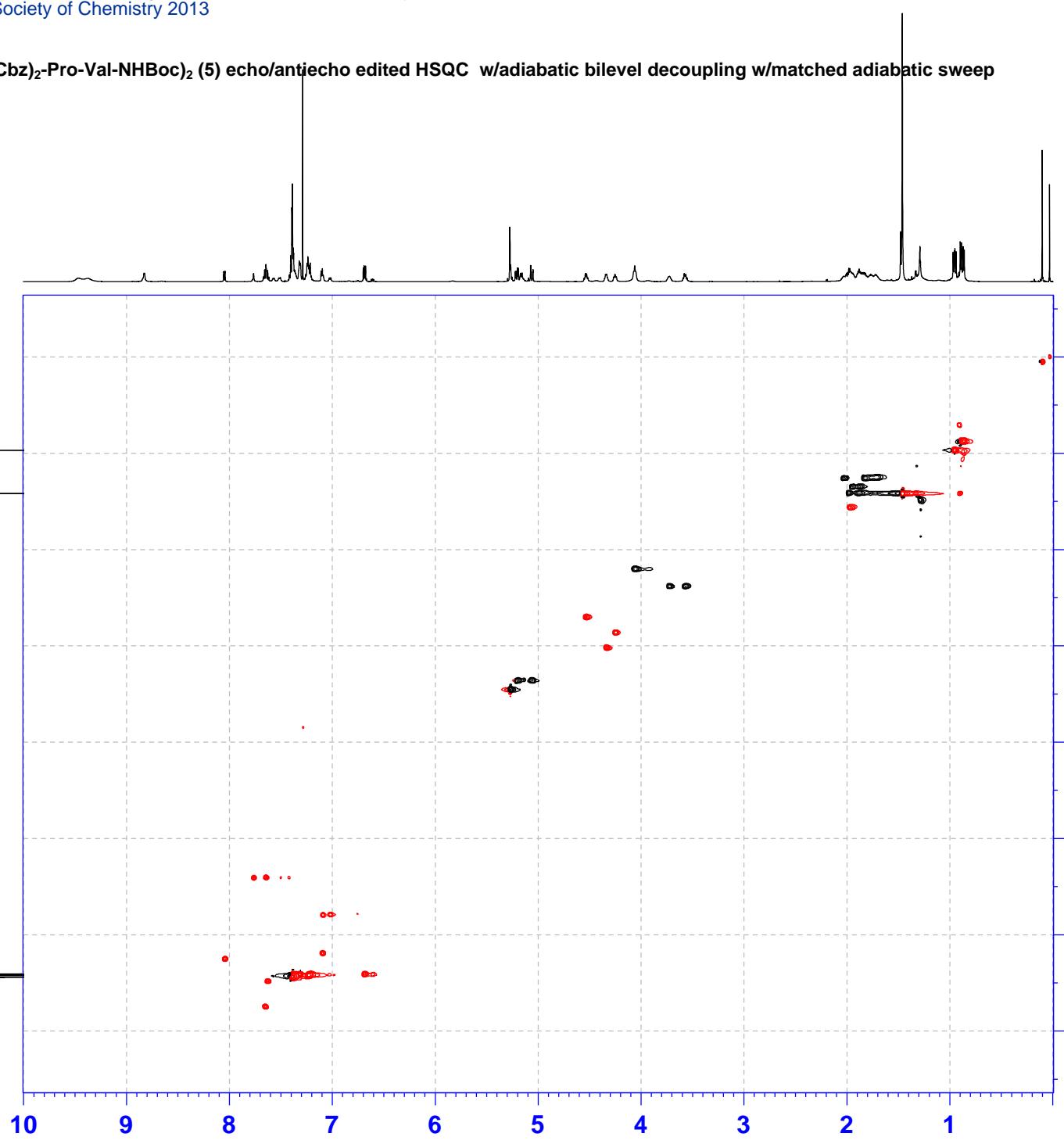
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PROCNO 1
Date_ 20101124
Time 15.14
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PULPROG hmbcetgpl3nd
TD 4096
SOLVENT CDCl₃
NS 4
DS 16
SWH 6009.615 Hz
FIDRES 1.467191 Hz
AQ 0.3408372 sec
RG 2050
DW 83.200 usec
DE 6.50 usec
TE 307.8 K
CNUST6 120.000000
CNUST7 165.000000
CNUST13 10.000000
CNUST30 0.5981154
D0 0.0000300 sec
D1 1.5000000 sec
D6 0.0500000 sec
D16 0.0002000 sec
INO 0.00001490 sec
===== CHANNEL f1 ======
NUC1 1H
P1 7.00 usec
P2 14.00 usec
PL1 0.25 dB
PL1W 16.59961510 W
SFO1 600.1330006 MHz
===== CHANNEL f2 ======
NUC2 13C
P3 12.00 usec
P24 2000.00 usec
PL2 -3.30 dB
PL2W 191.18331909 W
SFO2 150.9178750 MHz
SP7 3.28 dB
SPNAM7 Crp60comp.4
SPQAL7 0.500
SPOFFS7 0.00 Hz
===== GRADIENT CHANNEL =====
GPNAME1 SINE.100
GPNAME3 SINE.100
GPNAME4 SINE.100
GPNAME5 SINE.100
GPNAME6 SINE.100
GPZ1 80.00 %
GPZ3 14.00 %
GPZ4 -8.00 %
GPZ5 -4.00 %
GPZ6 -2.00 %
P16 1000.00 usec
ND0 2
TD 128
SFO1 150.9179 MHz
FIDRES 261.860260 Hz
SW 222.095 ppm
PmMode Echo-Antiecho
SI 2048
SF 600.1300000 MHz
WDW SINE
SSB 2
LB 0.00 Hz
GB 0
PC 1.00
SI 1024
MC2 echo-antiecho
SF 150.9028090 MHz
WDW SINE
SSB 2
LB 0.00 Hz
GB 0



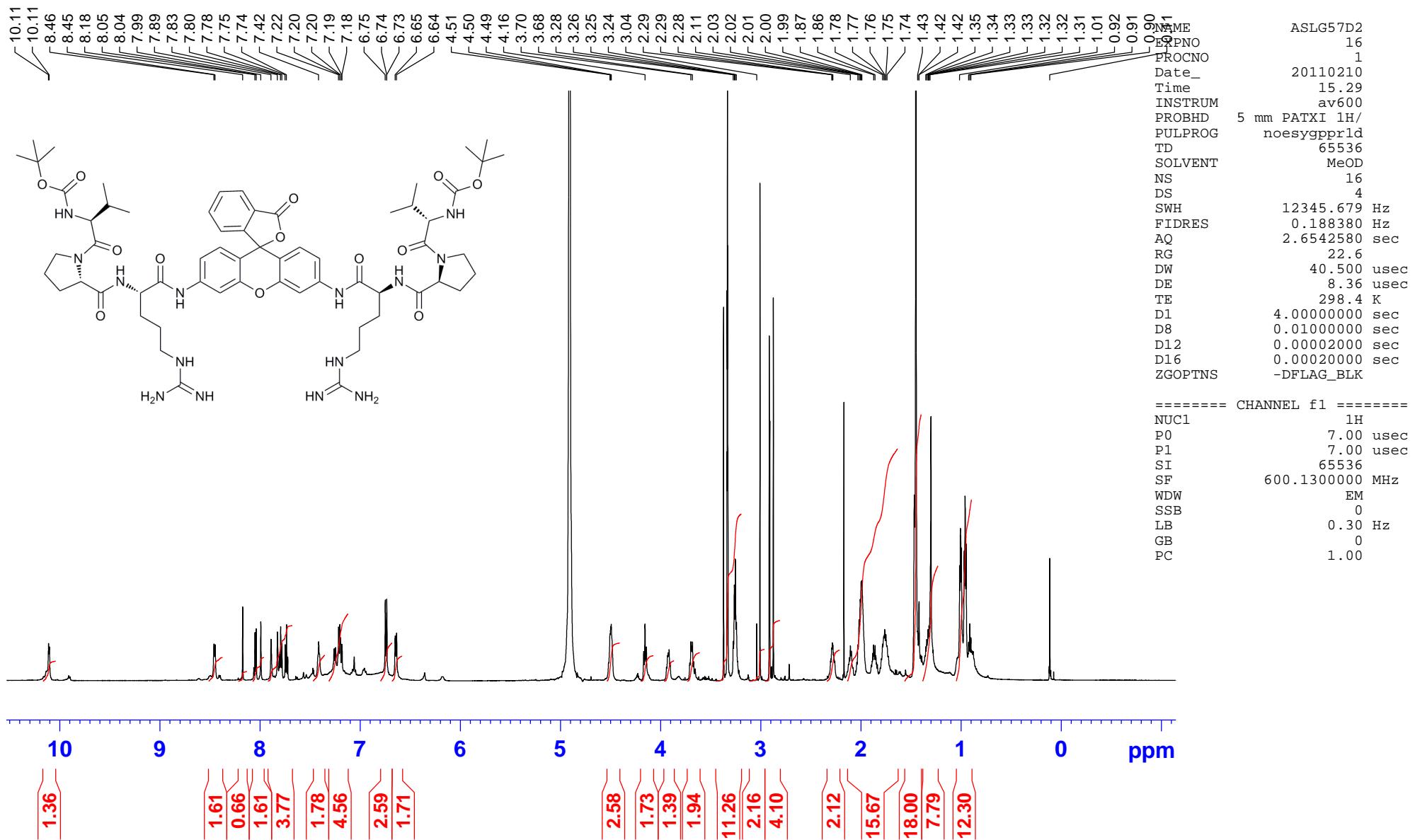
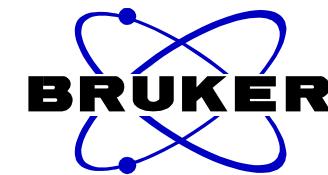
Rhodamine-(Arg-Cbz)₂-Pro-Val-NHBoc₂ (5) echo/antiecho edited HSQC w/adiabatic bilevel decoupling w/matched adiabatic sweep



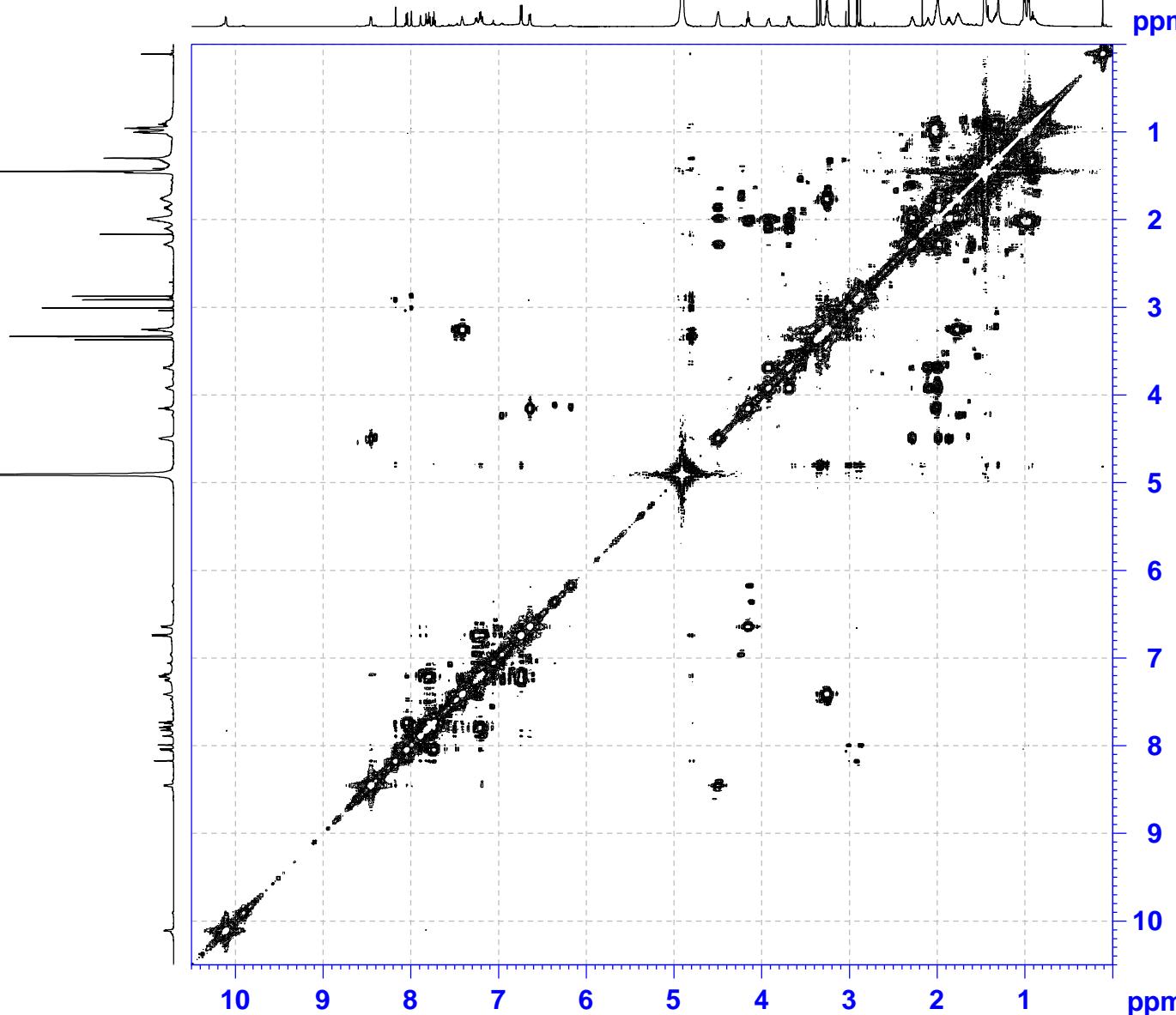
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EXPNO 11
PROCNO 1
Date 20101124
Time 15.00
INSTRUM av600
PROBHD 5 mm PATXI 1H/
PULPROG hsqcetgpsp.3
TD 1024
SOLVENT CDCl₃
NS 2
DS 16
SWH 6009.615 Hz
FIDRES 5.868765 Hz
AQ 0.0852468 sec
RG 2050
DW 83.200 usec
DE 6.50 usec
TE 308.3 K
CNS1 145.000000
D0 0.0000300 sec
D1 1.500000 sec
D4 0.00172414 sec
D11 0.03000000 sec
D16 0.00020000 sec
D21 0.00360000 sec
IN0 0.00002000 sec
L31 1
===== CHANNEL f1 ======
NUC1 1H
P1 7.00 usec
P2 14.00 usec
P28 0.00 usec
PL1 0.25 dB
PL1W 16.59961510 W
SF01 600.1330006 MHz
===== CHANNEL f2 ======
CPDPRG2 bi_p5m4sp_4sp.2
NUC2 ¹³C
P3 12.00 usec
P14 500.00 usec
P31 1730.00 usec
P63 1500.00 usec
PL0 120.00 dB
PL2 3.30 dB
PL12 1.00 dB
PL2W 0.0000000 W
PL1W 191.18331909 W
PL12W 5.61629534 W
SF02 150.99133719 MHz
SP3 3.28 dB
SP14 4.76 dB
SP18 8.67 dB
SP31 10.78 dB
SPNAM3 Crp60,0,5,20.1
SPNAM14 Crp32,1,5,20.2
SPNAM18 Crp60_xfilt.2
SPNAM31 Crp32,1,5,20.2
SPDAL3 0.500
SPDAL14 0.500
SPDAL18 0.500
SPDAL31 0.500
SPOFFS3 0.00 Hz
SPOFFS14 0.00 Hz
SPOFFS18 0.00 Hz
SPOFFS31 0.00 Hz
===== GRADIENT CHANNEL =====
GPNAME1 SINE,100
GPNAME2 SINE,100
GPZ1 20.00 %
GPZ2 20.10 %
P16 1000.00 usec
ND0 2
TD 256
SF01 150.9134 MHz
FIDRES 97.656288 Hz
SW 165.658 ppm
FnMODE Echo-Antiecho
SI 2048
SF 600.1300000 MHz
WDW QSINE
SSB 2
LB 0.00 Hz
GB 0
PC 1.00
SI 1024
MC2 echo-antiecho
SF 150.9028090 MHz
WDW QSINE
SSB 2
LB 0.00 Hz
GB 0



Rhodamine-(Arg-Pro-Val-NHBoc)₂ (3)



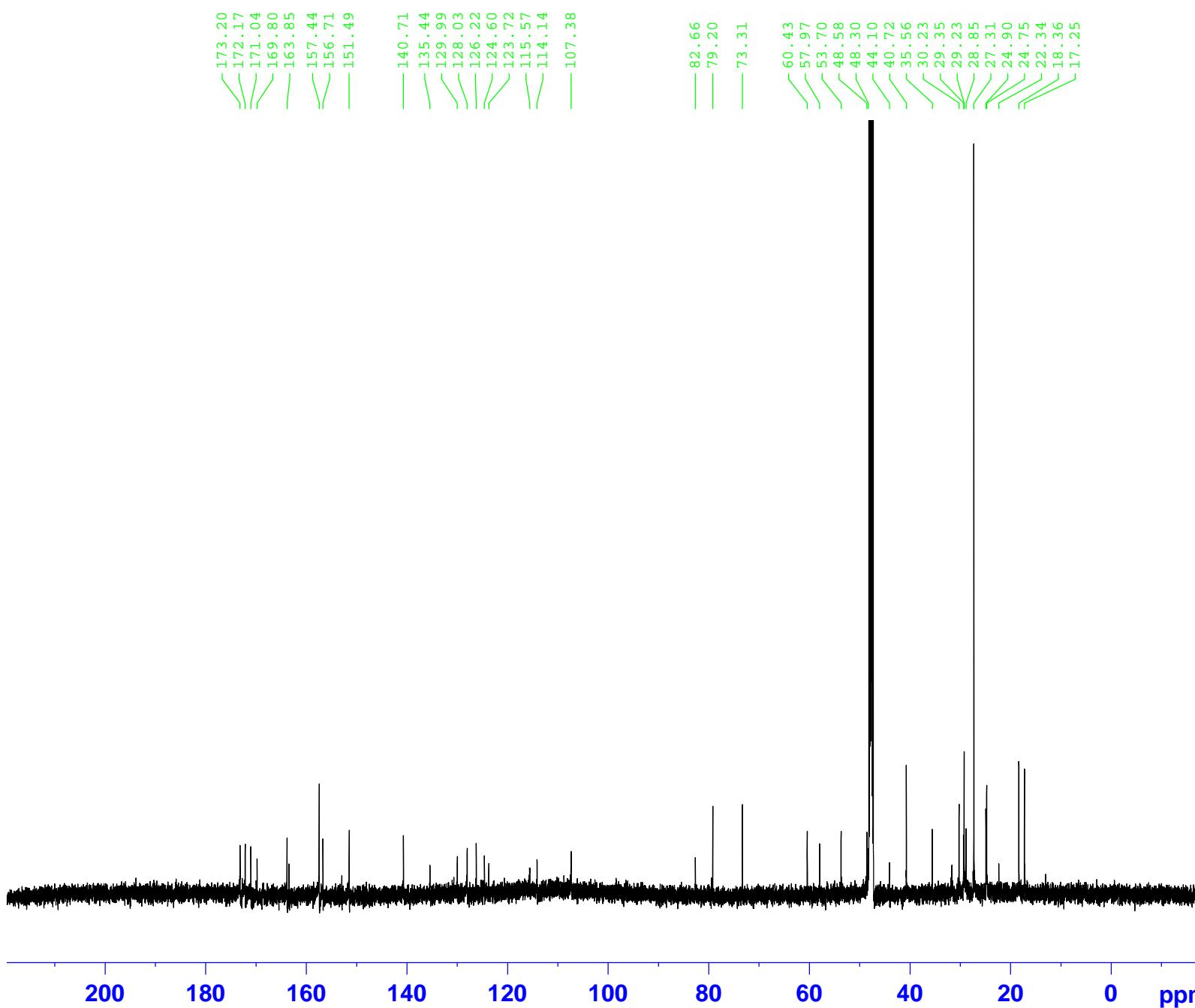
Rhodamine-(Arg-Pro-Val-NHBoc)₂ (3) COSY



NAME ASLG57D2
EXPNO 24
PROCNO 1
Date_ 20110211
Time 10.16
INSTRUM av600
PROBHD 5 mm PATXI 1H/
PULPROG cosygpprgrf
TD 4096
SOLVENT MeOD
NS 16
DS 8
SWH 9615.385 Hz
FIDRES 2.347506 Hz
AQ 0.2130420 sec
RG 128
DW 52.000 usec
DE 6.50 usec
TE 298.4 K
D0 0.00000300 sec
D1 2.00000000 sec
D11 0.03000000 sec
D12 0.00002000 sec
D16 0.00020000 sec
IN0 0.00010395 sec

===== CHANNEL f1 =====
NUC1 1H
P0 7.00 usec
P1 7.00 usec
ND0 1
TD 256
SFO1 600.1329 MHz
FIDRES 37.576378 Hz
SW 16.029 ppm
FnMODE QF
SI 2048
SF 600.1300000 MHz
WDW SINE
SSB 0
LB 0.00 Hz
GB 0
PC 1.00
SI 2048
MC2 QF
SF 600.1300000 MHz
WDW SINE
SSB 0
LB 0.00 Hz
GB 0

Rhodamine-(Arg-Pro-Val-NHBoc)₂ (3)

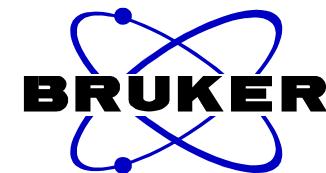


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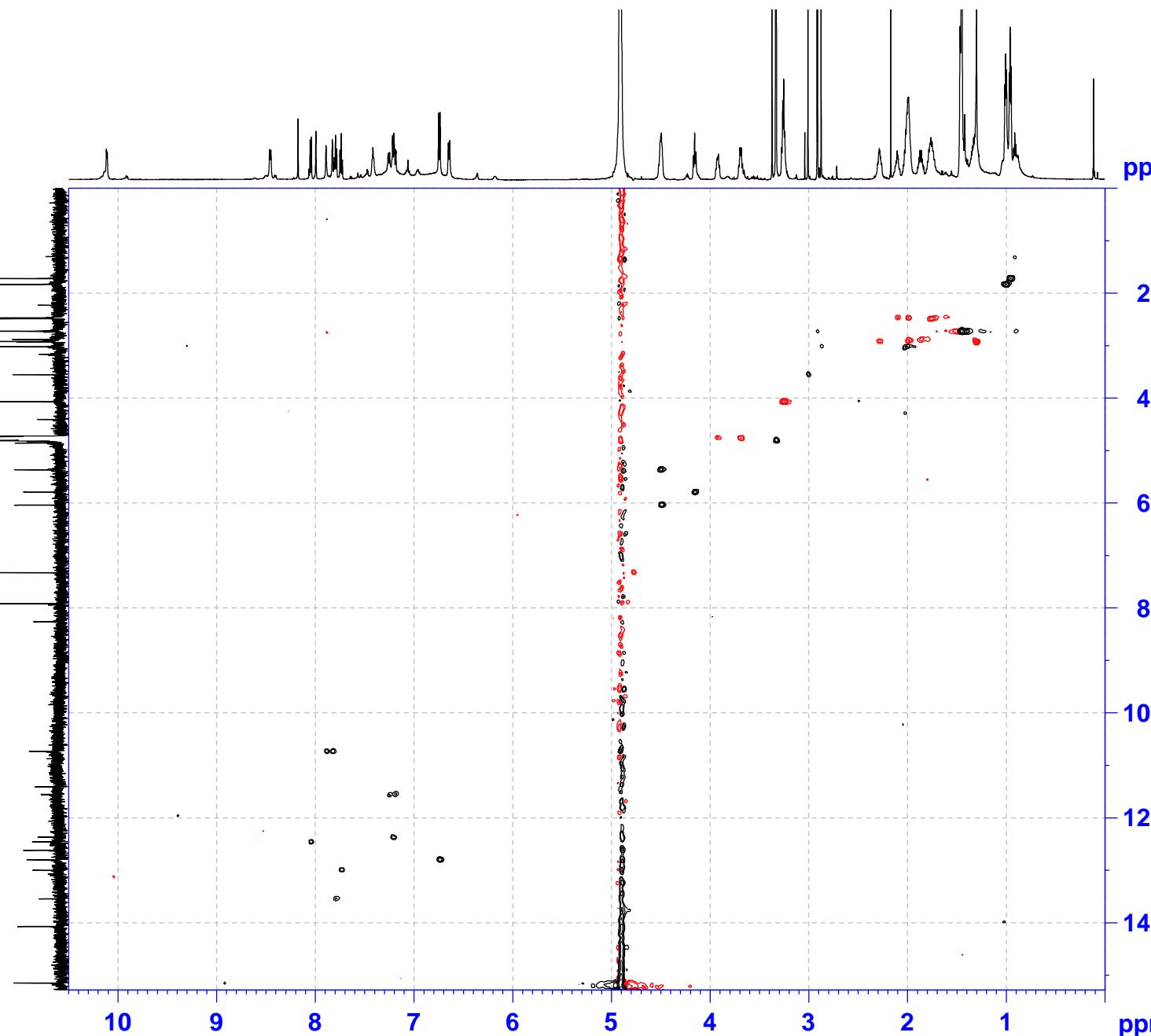
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EXPNO 23
PROCNO 1
Date_ 20110210
Time 20.51
INSTRUM av600
PROBHD 5 mm PATXI 1H/
PULPROG zgpg30
TD 65536
SOLVENT MeOD
NS 5120
DS 4
SWH 36057.691 Hz
FIDRES 0.550197 Hz
AQ 0.9088159 sec
RG 322
DW 13.867 usec
DE 6.50 usec
TE 298.4 K
D1 2.0000000 sec
D11 0.0300000 sec

===== CHANNEL f1 =====
NUC1 ¹³C
P1 12.00 usec
SI 32768
SF 150.9028090 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Rhodamine-(Arg-Pro-Val-NHBoc)₂ (3) HSQC



NAME ASLG57D2
EXPNO 13
PROCNO 2
Date_ 20110210
Time 13.21
INSTRUM av600
PROBHD 5 mm PATXI 1H/
PULPROG hsqcetgpsisp2.4
TD 1024
SOLVENT MeOD
NS 4
DS 32
SWH 9615.385 Hz
FIDRES 9.390024 Hz
AQ 0.0532980 sec
RG 1
DW 52.000 usec
DE 6.50 usec
TE 299.4 K
CNST2 145.0000000
CNST17 -0.5000000
D0 0.00000300 sec
D1 1.50000000 sec
D2 0.00344828 sec
D4 0.00172414 sec
D11 0.03000000 sec
D16 0.00020000 sec
D21 0.00360000 sec
D24 0.00089000 sec
IN0 0.00002000 sec
L0 0
TD0 2
===== CHANNEL f1 =====
NUC1 1H
P1 7.00 usec
P2 14.00 usec
P28 0.00 usec
ND0 2
TD 256
SFO1 150.9134 MHz
FIDRES 97.656288 Hz
SW 165.658 ppm
FnMODE Echo-Antiecho
SI 2048
SF 600.1300000 MHz
WDW QSINE
SSB 2
LB 0.00 Hz
GB 0
PC 1.40
SI 1024
MC2 echo-antiecho
SF 150.9028090 MHz
WDW QSINE
SSB 2
LB 0.00 Hz
GB 0

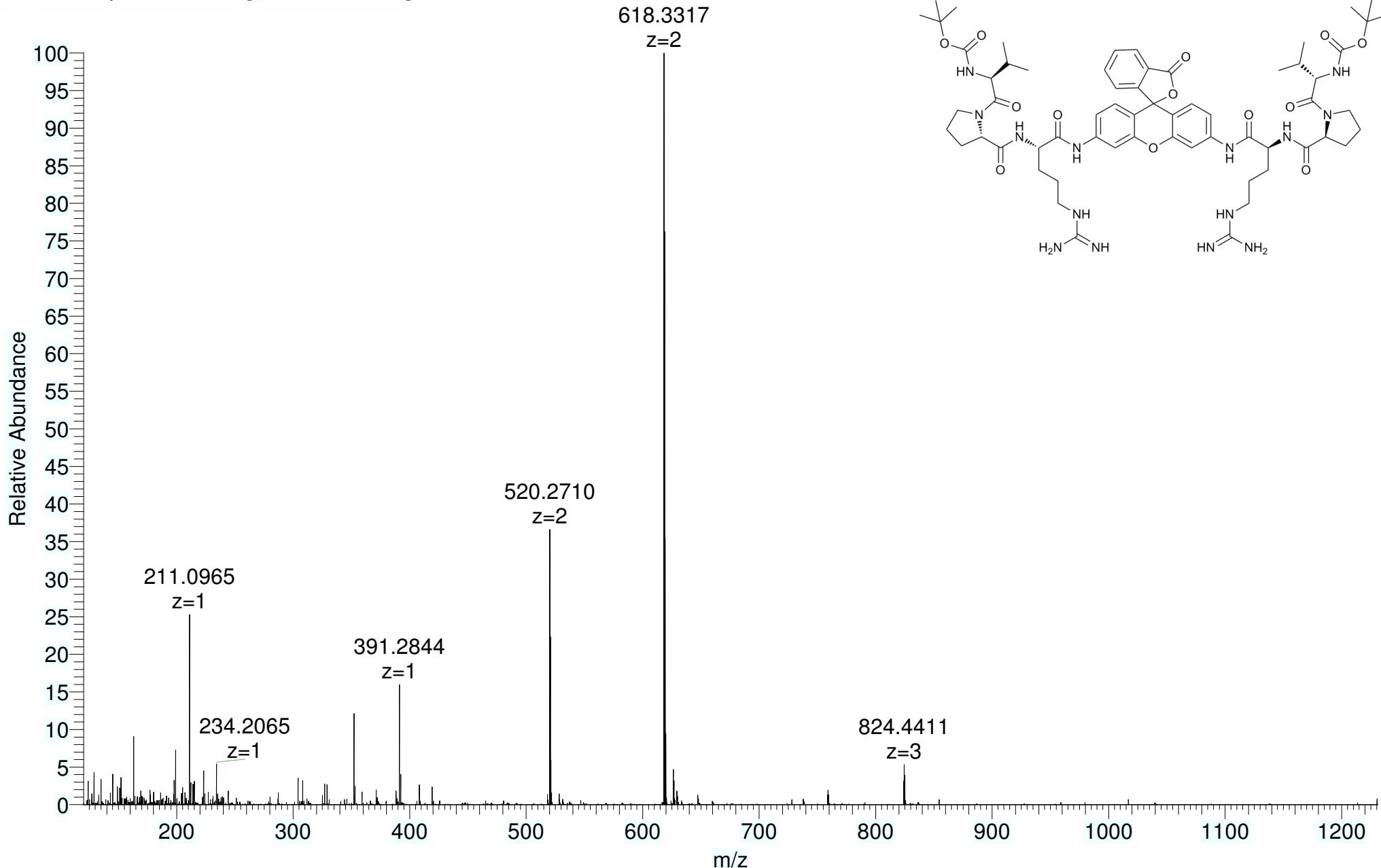


ASLG57D MW=1235?
(MeOH)/MeOH + NH₄OAc

EPSRC National Centre Swansea
LTQ Orbitrap XL

Alex Sinclair
02/02/2011 12:47:25

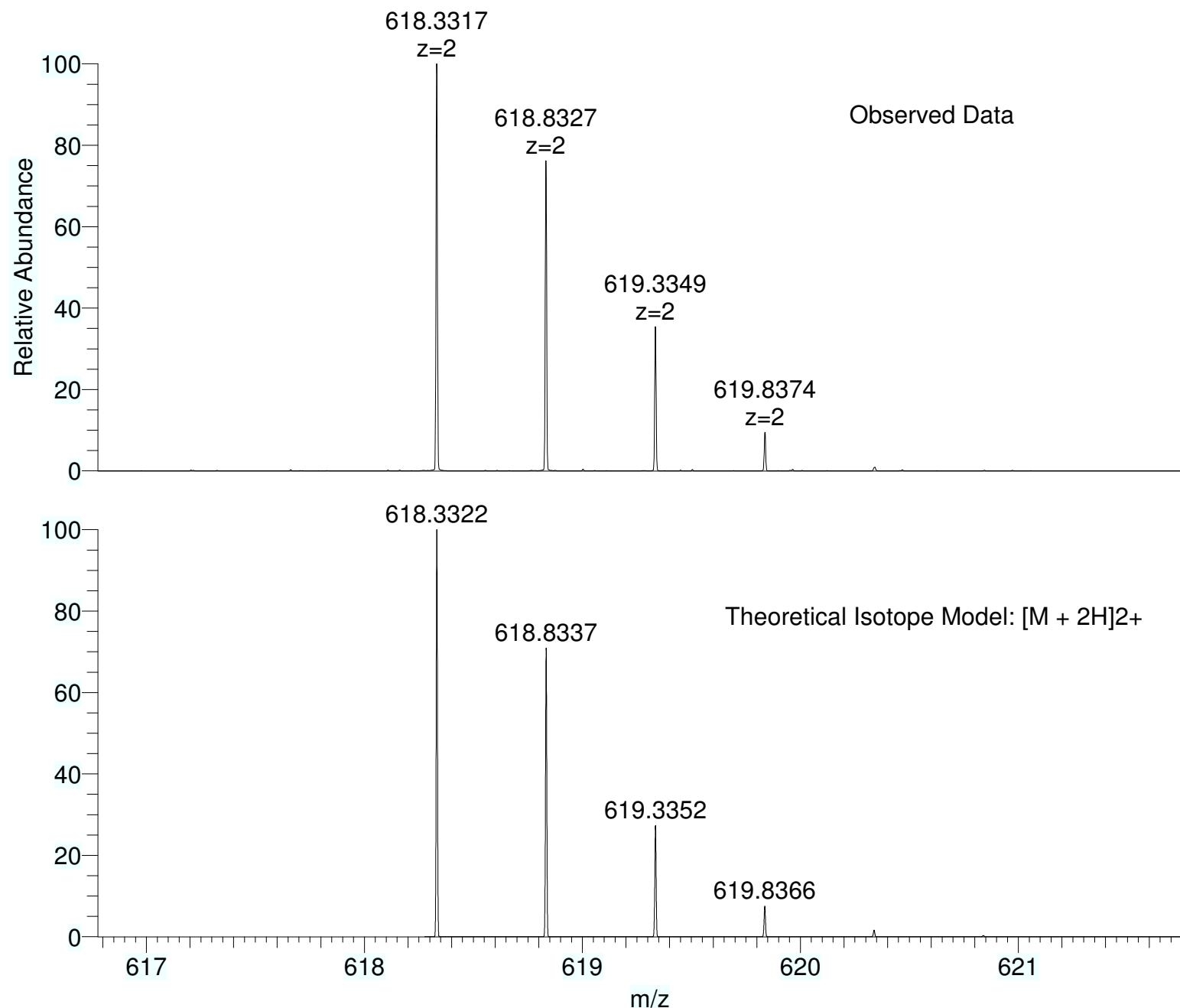
KINSIN001-OE-HNESP #18-22 RT: 0.82-0.91 AV: 5 SM: 7G NL: 6.42E6
T: FTMS + p NSI Full ms [120.00-2000.00]



ASLG57D MW=1235?
(MeOH)/MeOH + NH₄OAc

EPSRC National Centre Swansea
LTQ Orbitrap XL

Alex Sinclair
02/02/2011 12:47:25



NL:
6.42E6
KINSIN001-OE-HNESP#18-22
RT: 0.82-0.91 AV: 5 T: FTMS
+ p NSI Full ms
[120.00-2000.00]

NL:
1.10E4
C₆₂H₈₆N₁₄O₁₃ HH:
C₆₂H₈₈N₁₄O₁₃
p (gss, s /p:40) Chrg 2
R: 100000 Res .Pwr . @FWHM