

Electronic Supplementary Information (ESI)

Regioselective di- and tetra- functionalisation of γ -cyclodextrin using capping methodology

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Content

- NMR and mass spectra of **6-11**
- Crystal structure analyses of **9·2H₂O·0.5C₅H₁₂**
- General procedure for assigning the glucose units linked by a given capping unit
- References

NMR and mass spectra

NMR spectra of all compounds were recorded in CDCl₃ at 25 °C.

6^A,6^B-Bis-*O*-bis{benzene-1,3-bis[bis(4-*tert*-butylphenyl)methyl]}-2^A,2^B,2^C,2^D,2^E,2^F,2^G,2^H,3^A,3^B,3^C,3^D,3^E,3^F,3^G,3^H,6^C,6^D,6^E,6^F,6^G,6^H-docosa-*O*-methyl-γ-cyclodextrin (6):

¹ H NMR spectrum	5
¹ H/ ¹ H COSY spectrum	6
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¹ H/ ¹ H ROESY spectrum.....	8
¹³ C{ ¹ H} NMR spectrum.....	9
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Full assignment (Table 1, Table 2).....	13

6^A,6^B:6^D,6^E-Tetra-*O*-bis{benzene-1,3-bis[bis(4-*tert*-butylphenyl)methyl]}2^A,2^B,2^C,2^D,2^E,2^F,2^G,2^H,3^A,3^B,3^C,3^D,3^E,3^F,3^G,3^H,6^C,6^F,6^G,6^H-icosa-*O*-methyl-γ-cyclodextrin (7):

¹ H NMR spectrum	14
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¹ H/ ¹ H ROESY spectrum.....	17
¹³ C{ ¹ H} NMR spectrum.....	18
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Simulated mass spectrum	21
Full assignment (Table 3, Table 4).....	22

6^A,6^B:6^E,6^F-Tetra-*O*-bis{benzene-1,3-bis[bis(4-*tert*-butylphenyl)methyl]}2^A,2^B,2^C,2^D,2^E,2^F,2^G,2^H,3^A,3^B,3^C,3^D,3^E,3^F,3^G,3^H,6^C,6^D,6^G,6^H-icosa-*O*-methyl-γ-cyclodextrin (8):

¹ H NMR spectrum	23
¹ H/ ¹ H COSY spectrum	24
¹ H/ ¹ H TOCSY spectrum.....	25
¹ H/ ¹ H ROESY spectrum.....	26
¹³ C{ ¹ H} NMR spectrum.....	27
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Full assignment (Table 5, Table 6).....	31

2^A,2^B,2^C,2^D,2^E,2^F,2^G,2^H,3^A,3^B,3^C,3^D,3^E,3^F,3^G,3^H,6^C,6^D,6^G,6^H-icosa-*O*-methyl- γ -cyclodextrin (9):

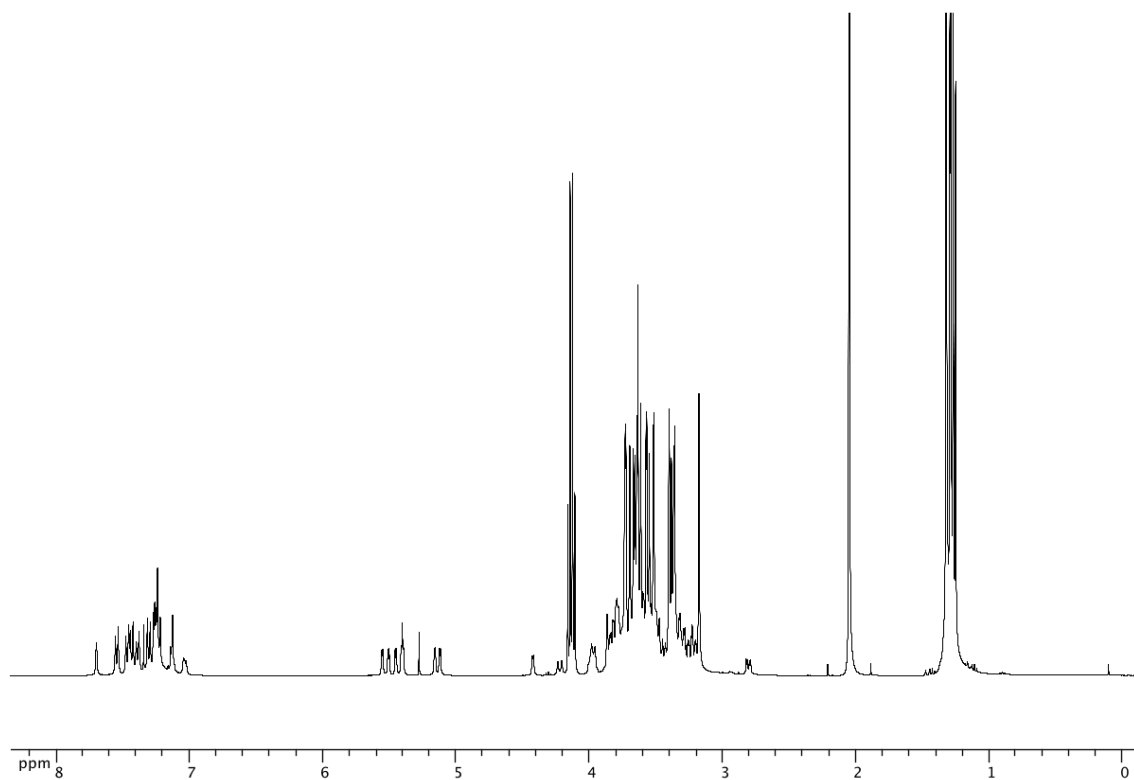
¹ H NMR spectrum	32
¹ H/ ¹ H COSY spectrum	33
¹³ C{ ¹ H} NMR spectrum	34
DEPT 135 spectrum	35
¹ H/ ¹³ C HMQC spectrum	36
Mass spectrum	37
Simulated mass spectrum	37

2^A,2^B,2^C,2^D,2^E,2^F,2^G,2^H,3^A,3^B,3^C,3^D,3^E,3^F,3^G,3^H,6^C,6^D,6^E,6^F,6^G,6^H-docosa-*O*-methyl- γ -cyclodextrin (10).

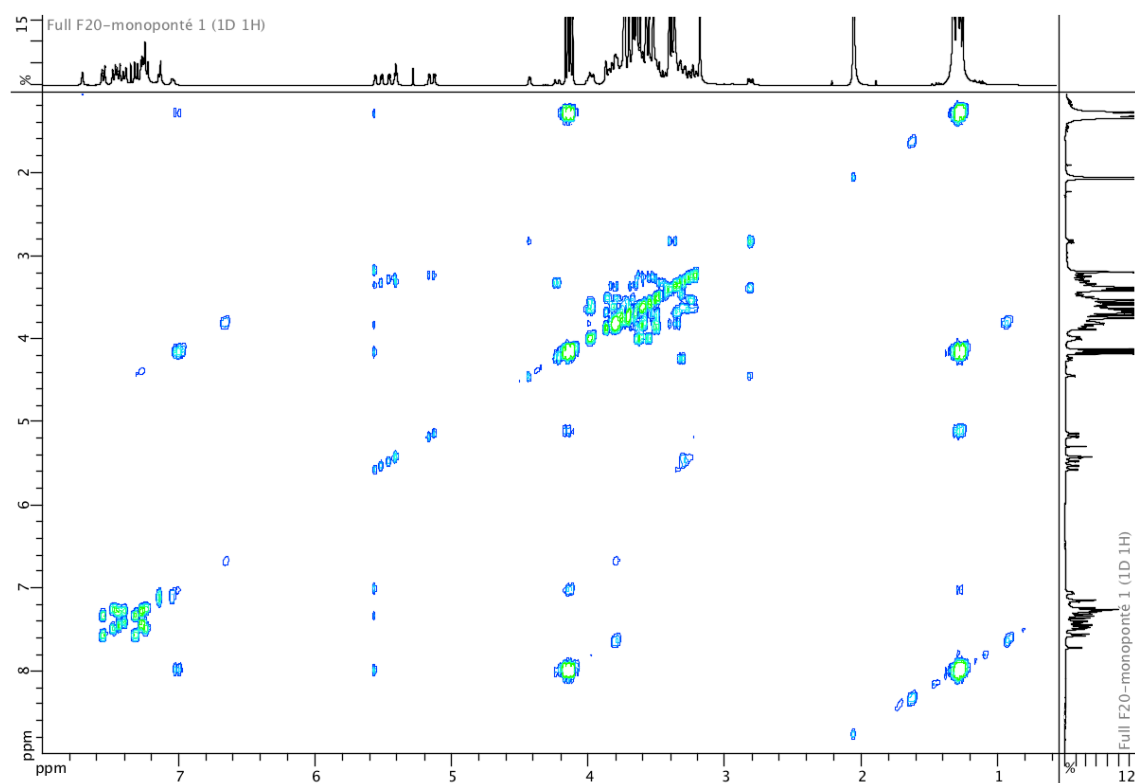
¹ H NMR spectrum	38
¹ H/ ¹ H COSY spectrum	39
¹³ C{ ¹ H} NMR spectrum	40
DEPT 135 spectrum	41
¹ H/ ¹³ C HMQC spectrum	42
Mass spectrum	43
Simulated mass spectrum	43

2^A,2^B,2^C,2^D,2^E,2^F,2^G,2^H,3^A,3^B,3^C,3^D,3^E,3^F,3^G,3^H,6^C,6^F,6^G,6^H-icosa-*O*-methyl- γ -cyclodextrin (11):

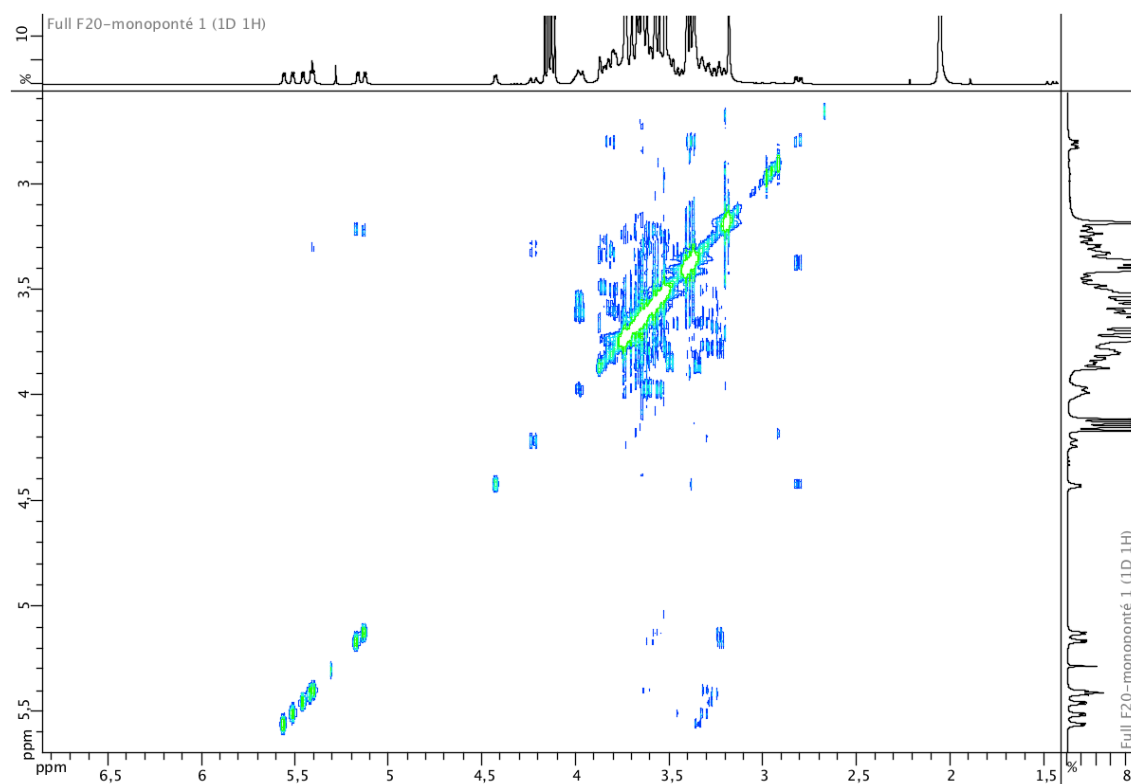
¹ H NMR spectrum	44
¹ H/ ¹ H COSY spectrum	45
¹³ C{ ¹ H} NMR spectrum	46
DEPT 135 spectrum	47
¹ H/ ¹³ C HMQC spectrum	48
Mass spectrum	49
Simulated mass spectrum	49



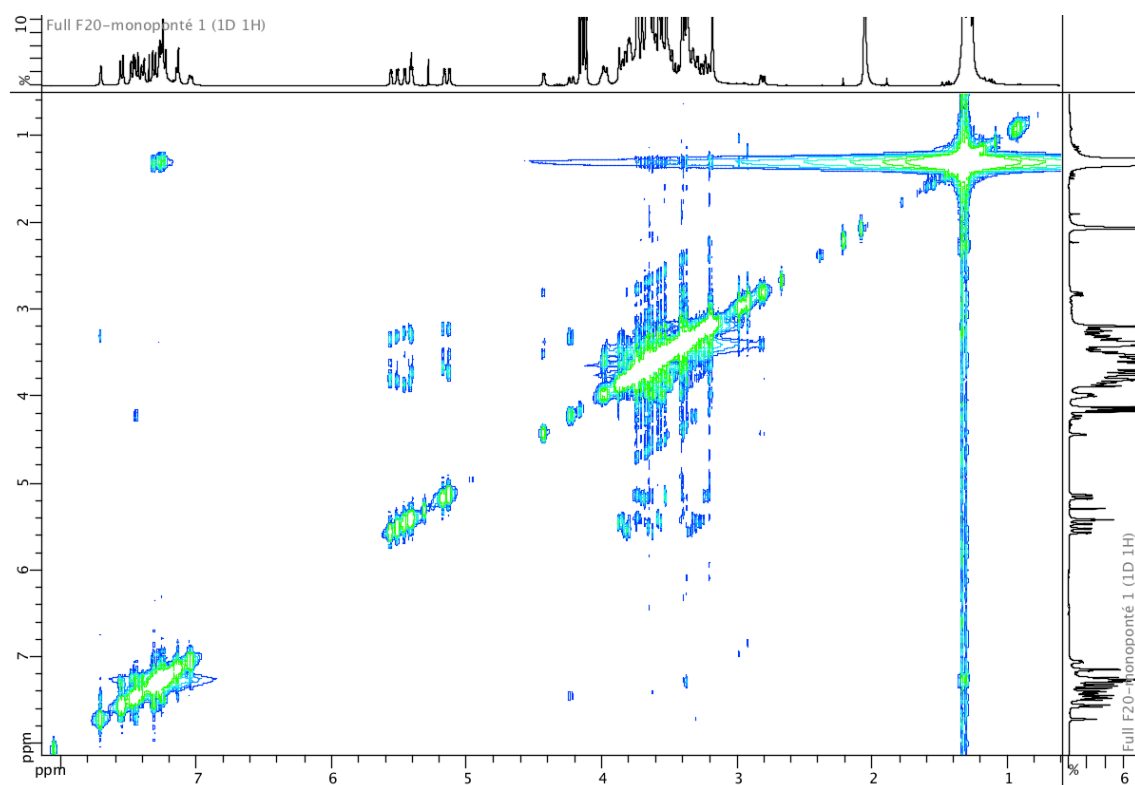
¹H NMR spectrum of **6**.



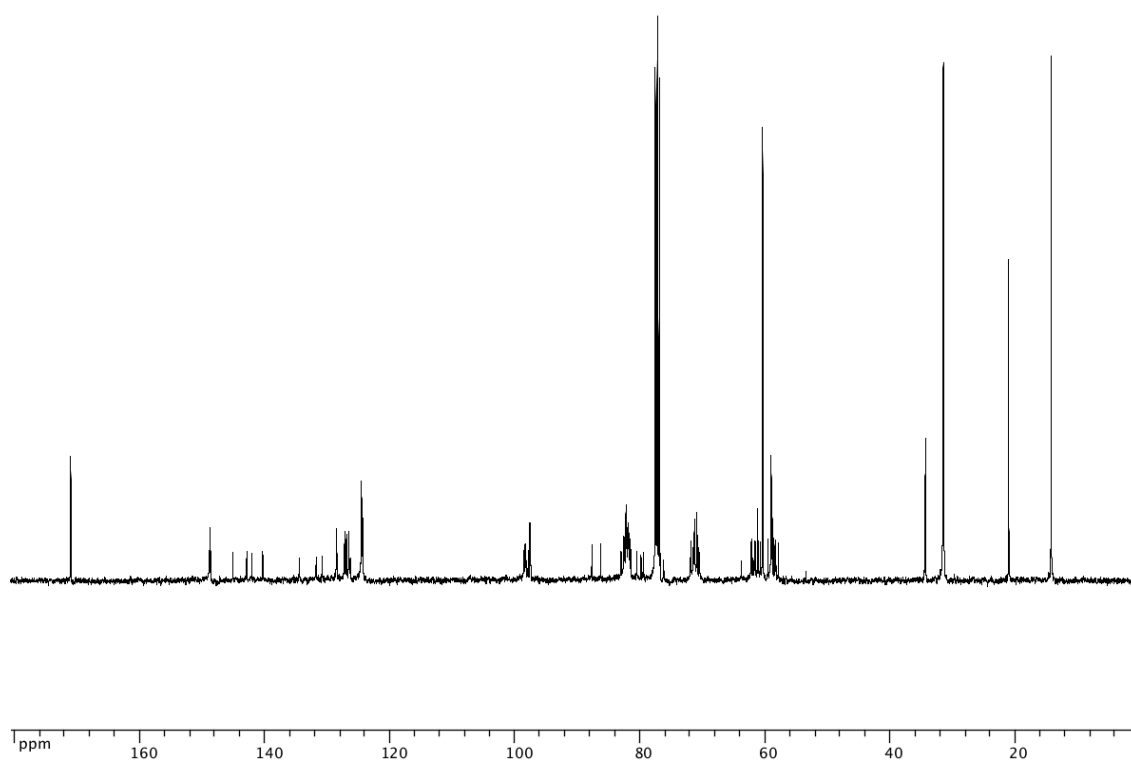
$^1\text{H}/^1\text{H}$ COSY spectrum of **6**.



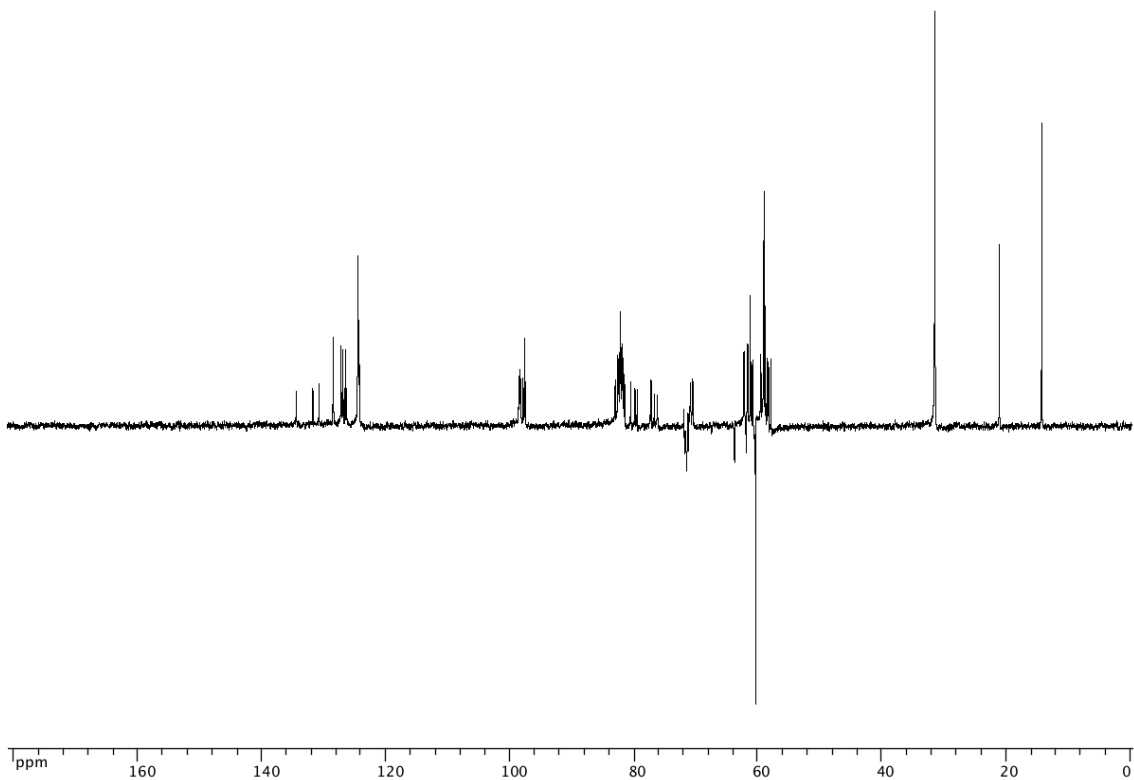
$^1\text{H}/^1\text{H}$ TOCSY spectrum of **6**.



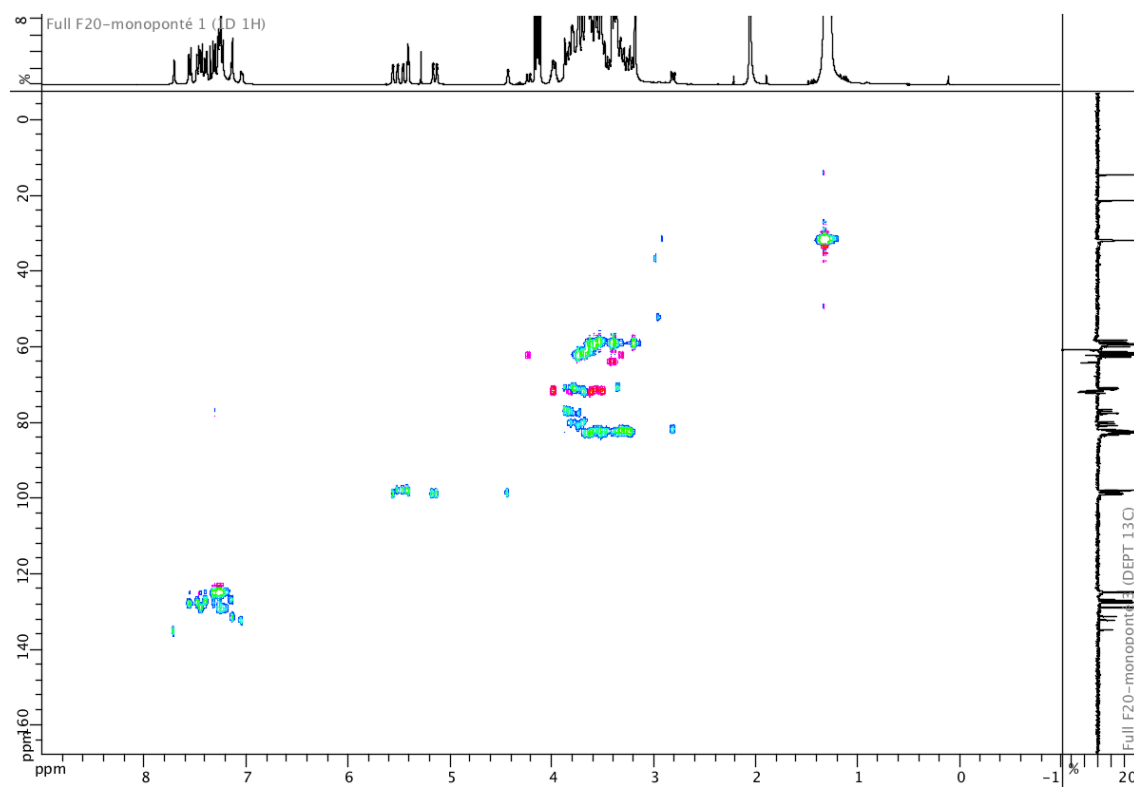
$^1\text{H}/^1\text{H}$ ROESY spectrum of **6**.



$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **6**.



DEPT 135 spectrum of **6**.



$^1\text{H}/^{13}\text{C}$ HSQC edited spectrum of **6**.

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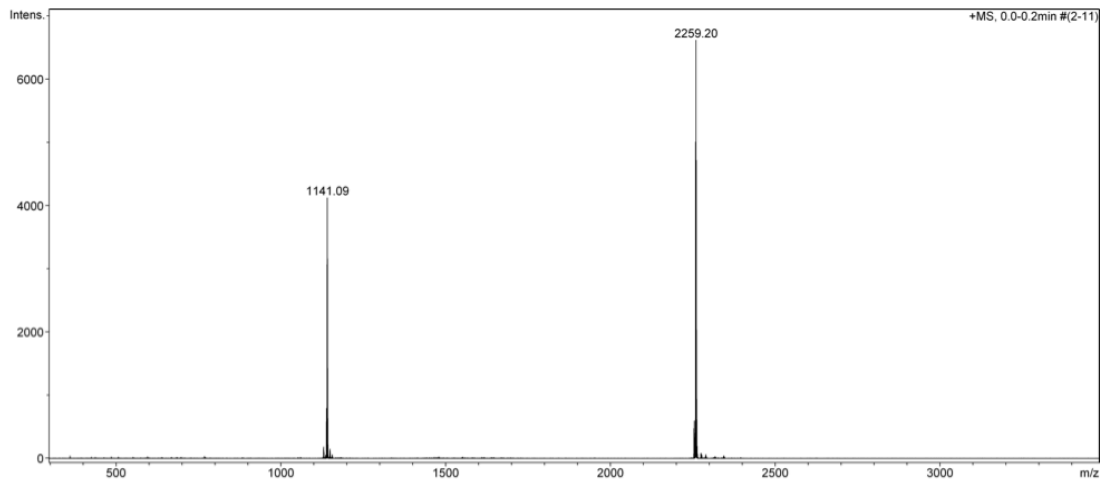
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Sample Name MJ83F20-AB monicap
Comment

Acquisition Date 10/14/2011 2:11:43 PM
Operator Administrator
Instrument micrOTOF

Acquisition Parameter

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Scan Range	n/a	Set Skimmer 1	50.0 V	Dry Heater	180 °C	APCI Heater	517 °C



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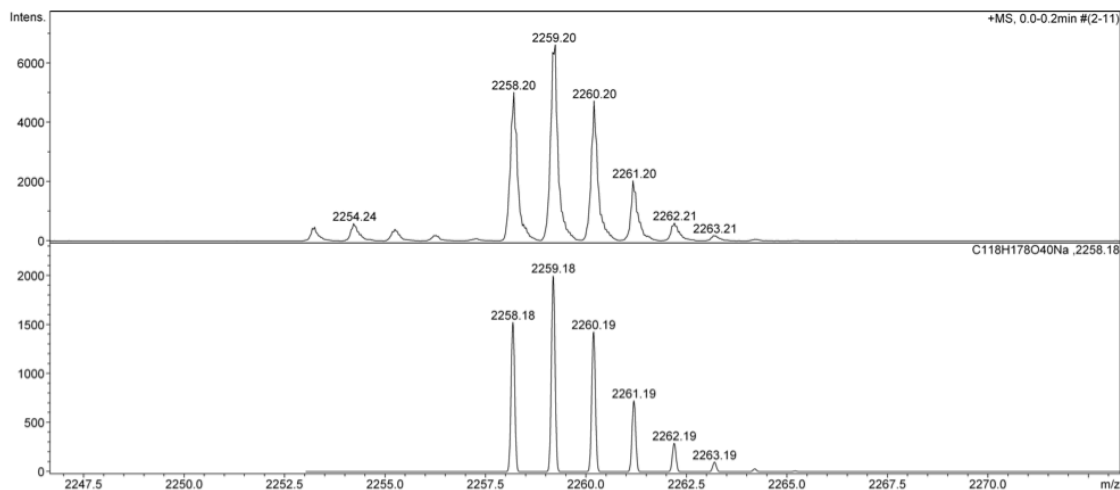
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Instrument micrOTOF

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Source Type	ESI	Capillary	4500 V	Nebulizer	0.4 Bar	Corona	219 nA
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Scan Range	n/a	Set Skimmer 1	50.0 V	Dry Heater	180 °C	APCI Heater	517 °C



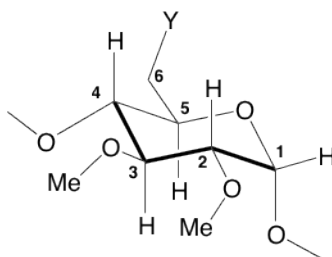
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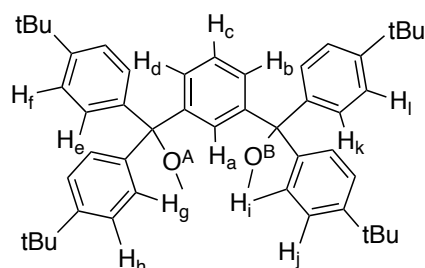
Mass spectrum of **6**, and simulation.

Table 1. Assignment of the sugar protons of **6** (based on TOCSY, COSY, ROESY and HMQC).

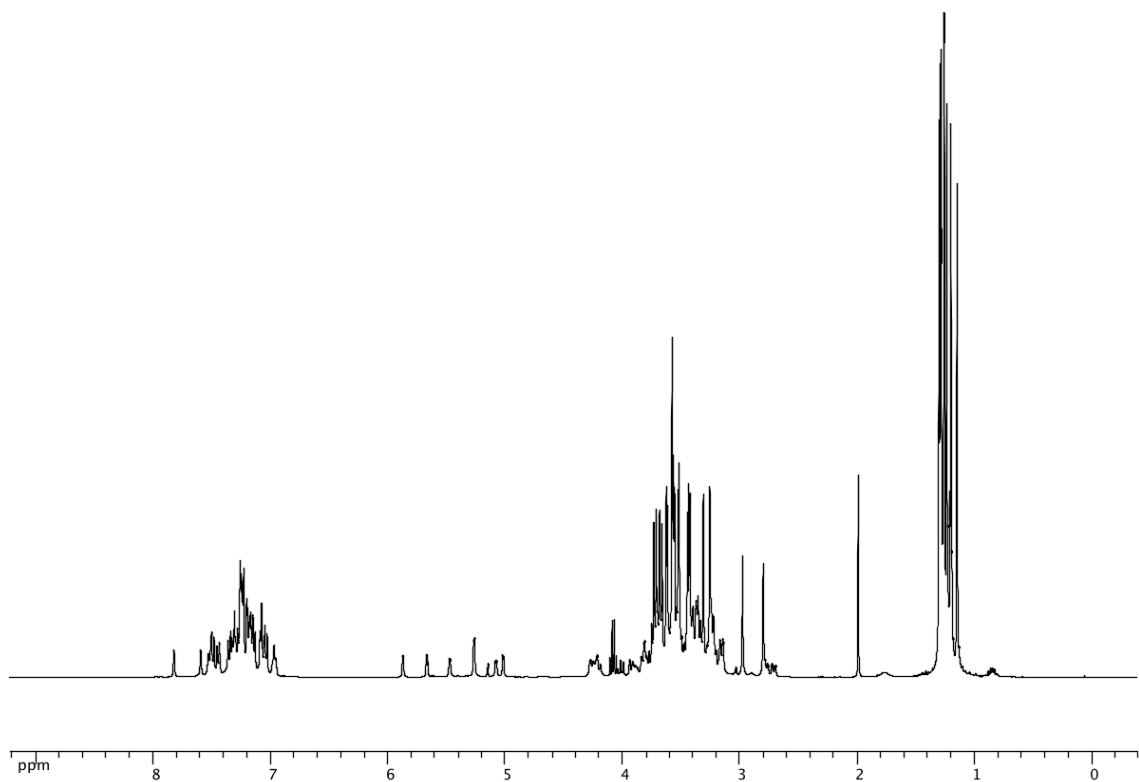


	A	B	C	D	E	F	G	H
H-1	5.51	4.42	5.56	5.45	5.41	5.12	5.16	5.40
H-2	3.31	2.81	3.33	3.27	3.25	3.22	3.21	3.30
H-3	3.45	3.37	3.65	3.61	3.51	3.55	3.59	3.60
H-4	3.49	3.81	3.87	3.82	3.71	3.67	3.73	3.81
H-5	3.66	3.33	3.85	3.78	3.68	3.76	3.79	3.76
H-6a	3.37	3.30	3.59	3.62	3.47	3.49	3.55	3.60
H-6b	3.42	4.22	3.61	3.97	3.61	3.63	3.97	3.98
OMe-6	—	—	3.40	3.36	3.18	3.18	3.37	3.41

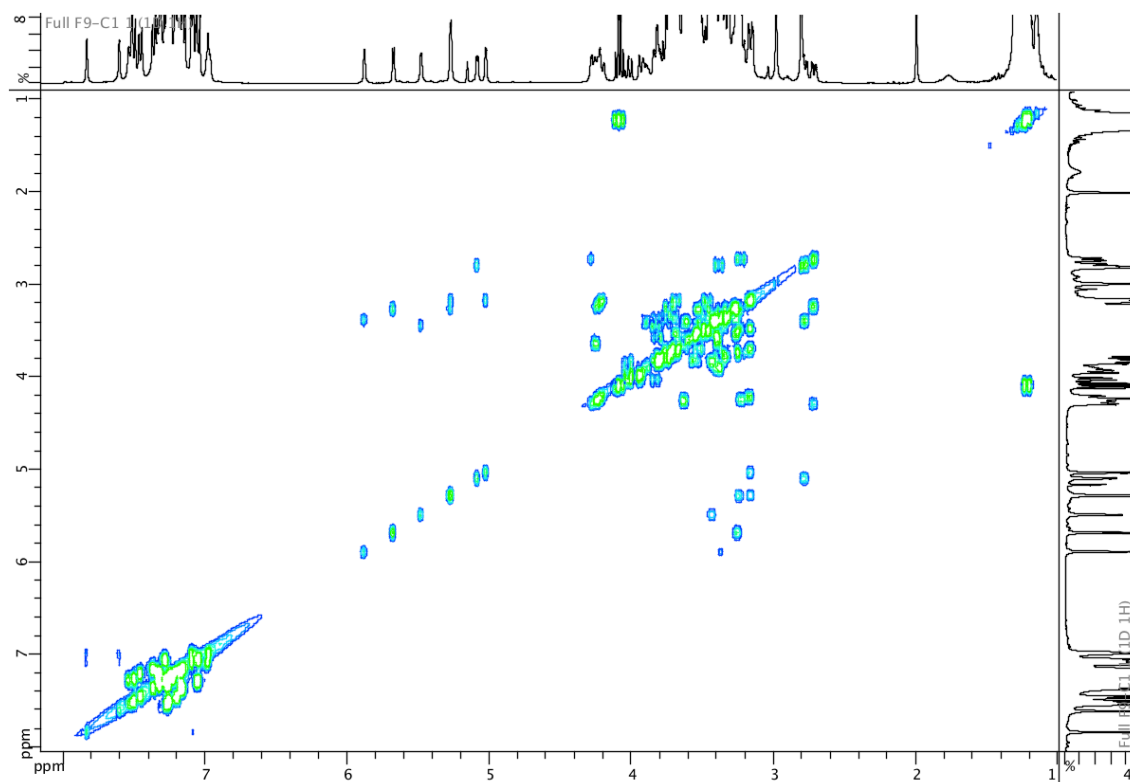
Table 2. Assignment of the aromatic protons of **6** (based on TOCSY, COSY, ROESY and HMQC).



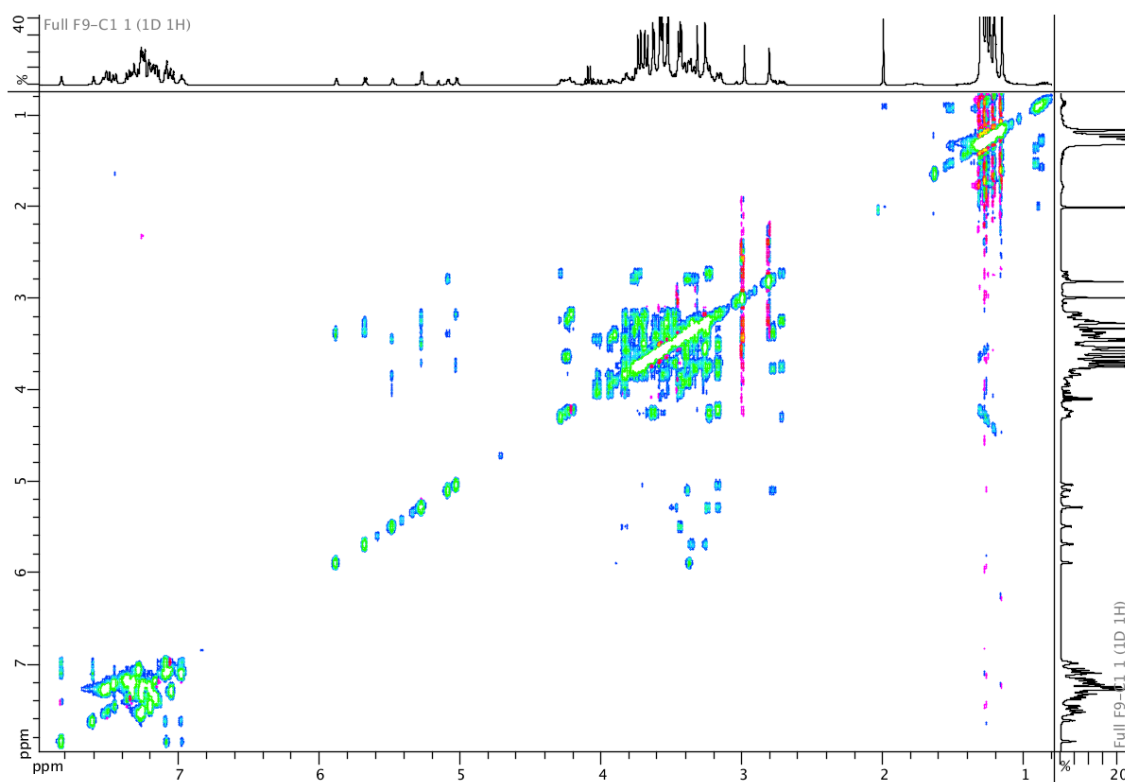
	A,B bridge
H _a	7.70
H _b	7.12
H _c	7.04
H _d	7.14
H _e	7.47
H _f	7.23
H _g	7.55
H _h	7.31
H _i	7.44
H _j	7.27
H _k	7.39
H _l	7.25



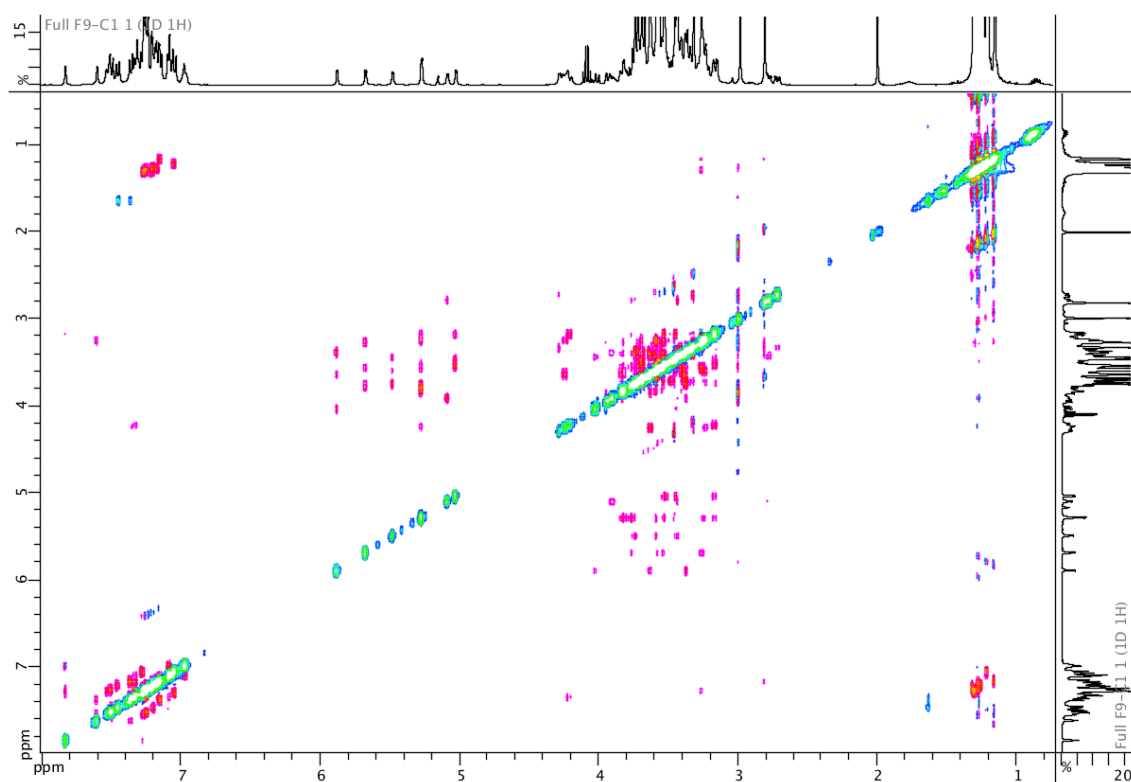
^1H NMR spectrum of **7**.



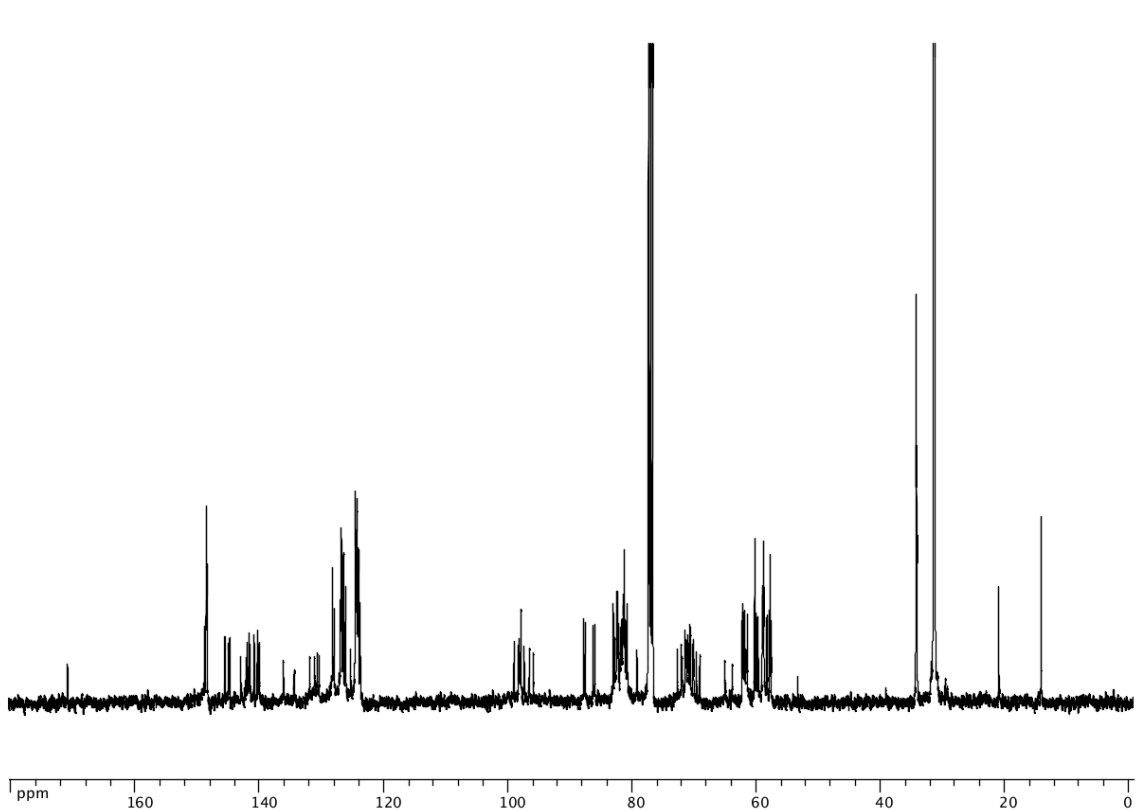
$^1\text{H}/^1\text{H}$ COSY spectrum of **7**.



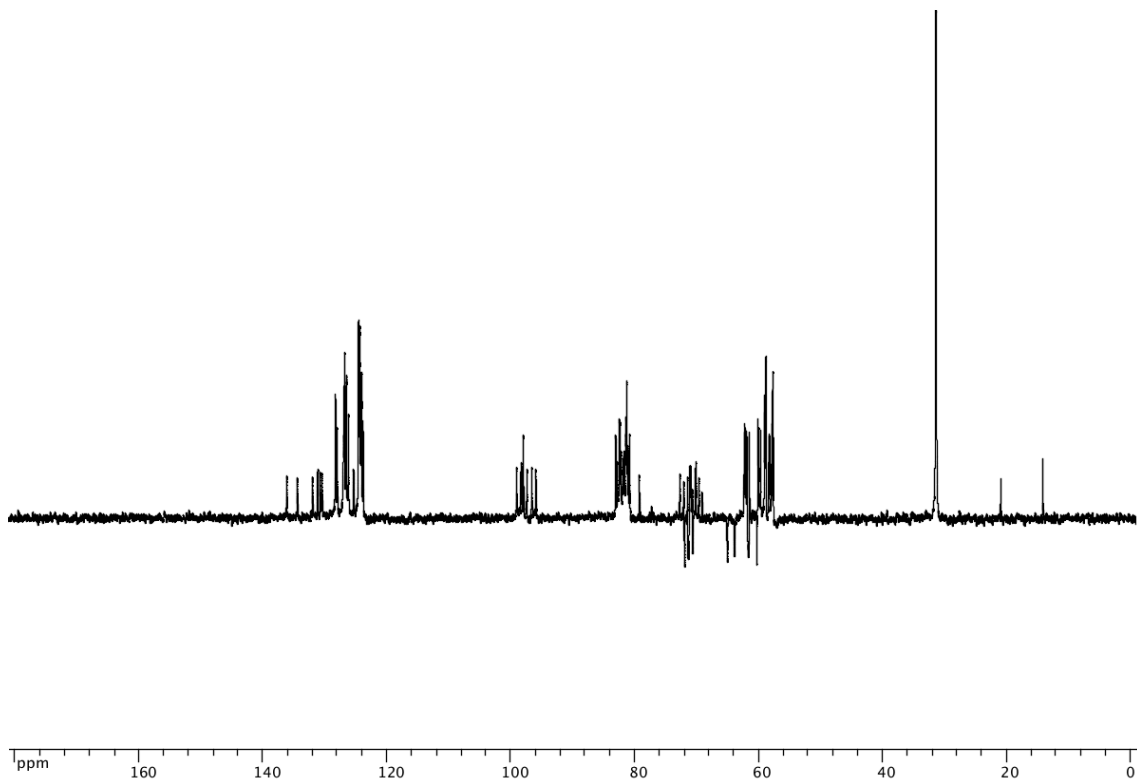
$^1\text{H}/^1\text{H}$ TOCSY spectrum of **7**.



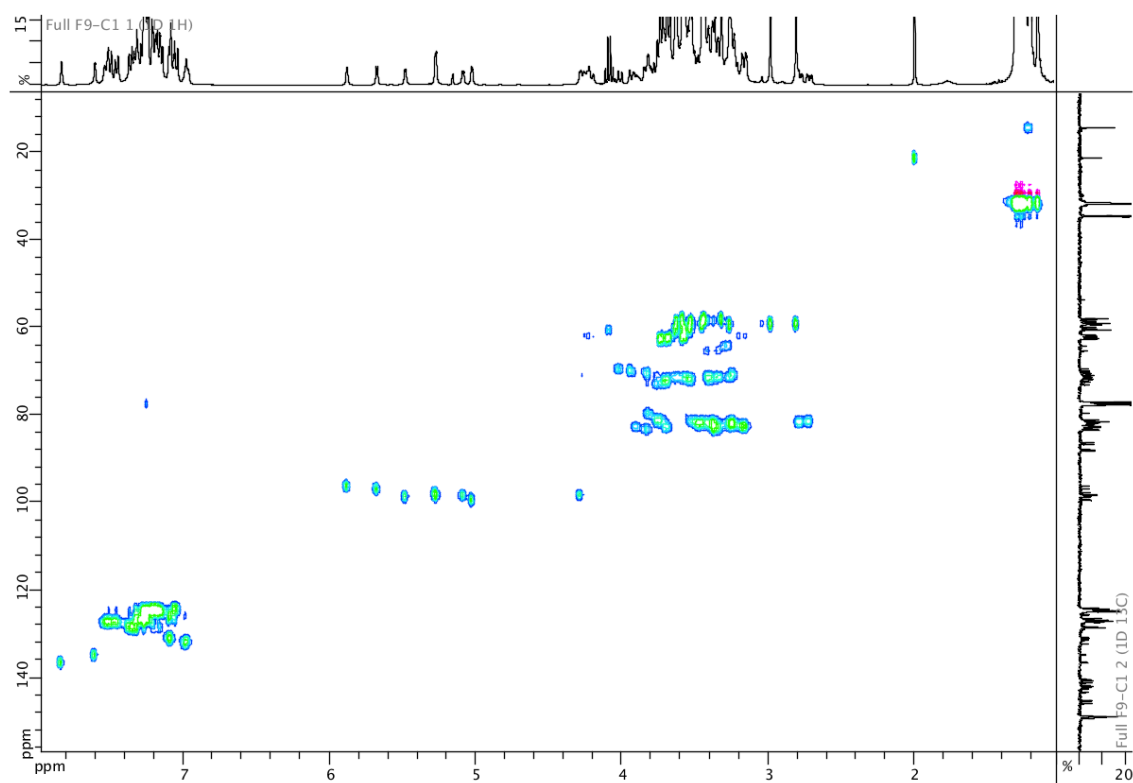
$^1\text{H}/^1\text{H}$ ROESY spectrum of **7**.



$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **7**.



DEPT 135 spectrum of **7**.



$^1\text{H}/^{13}\text{C}$ HSQC edited spectrum of **7**.

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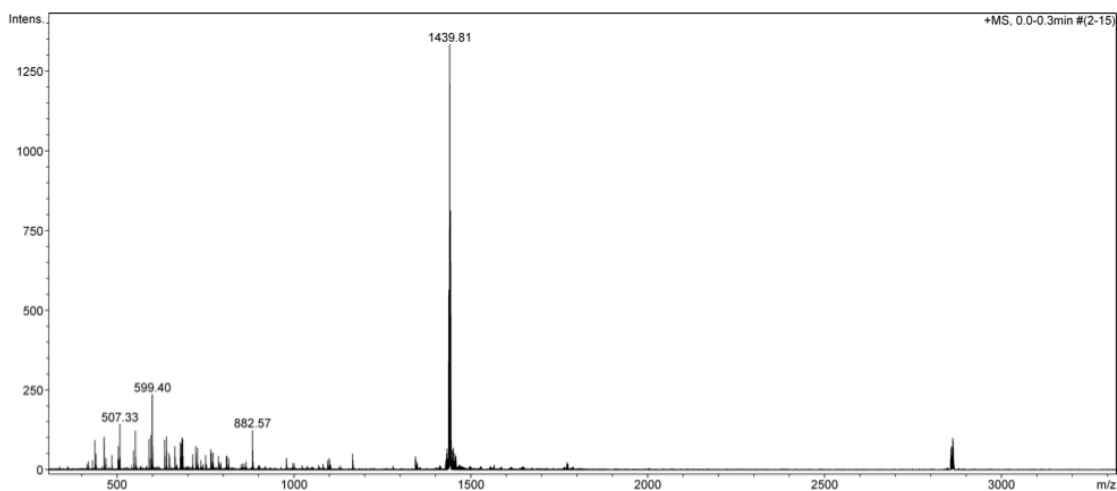
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Comment

Acquisition Date 10/14/2011 2:06:47 PM
Operator Administrator
Instrument micrOTOF

Acquisition Parameter

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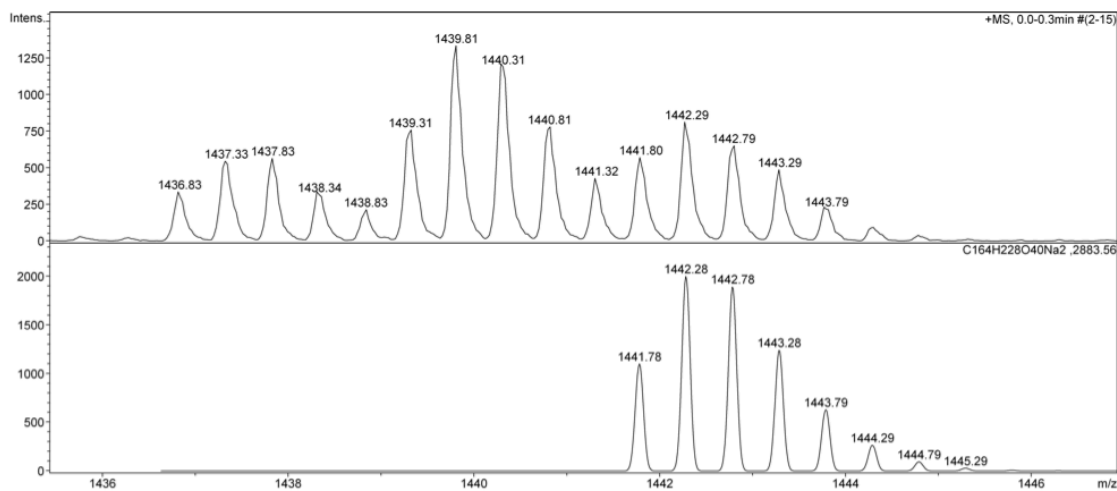
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Instrument micrOTOF

Acquisition Parameter

Source Type	ESI	Capillary	4500 V	Nebulizer	0.4 Bar	Corona	219 nA
Ion Polarity	Positive	Set Capillary Exit	150.0 V	Dry Gas	4.0 l/min	Set Hexapole RF	300.0 V
Scan Range	n/a	Set Skimmer 1	50.0 V	Dry Heater	180 °C	APCI Heater	517 °C



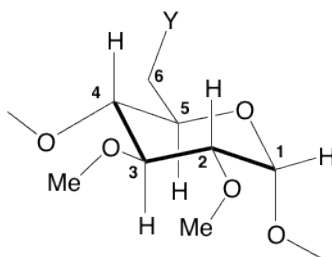
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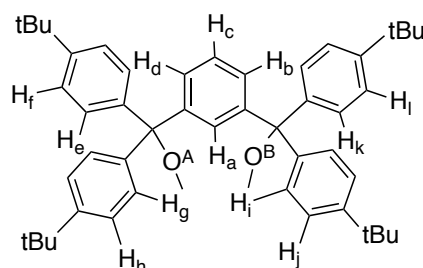
Mass spectrum of **7** and simulation.

Table 3. Assignment of the sugar protons of **7** (based on TOCSY, COSY, ROESY and HMQC).

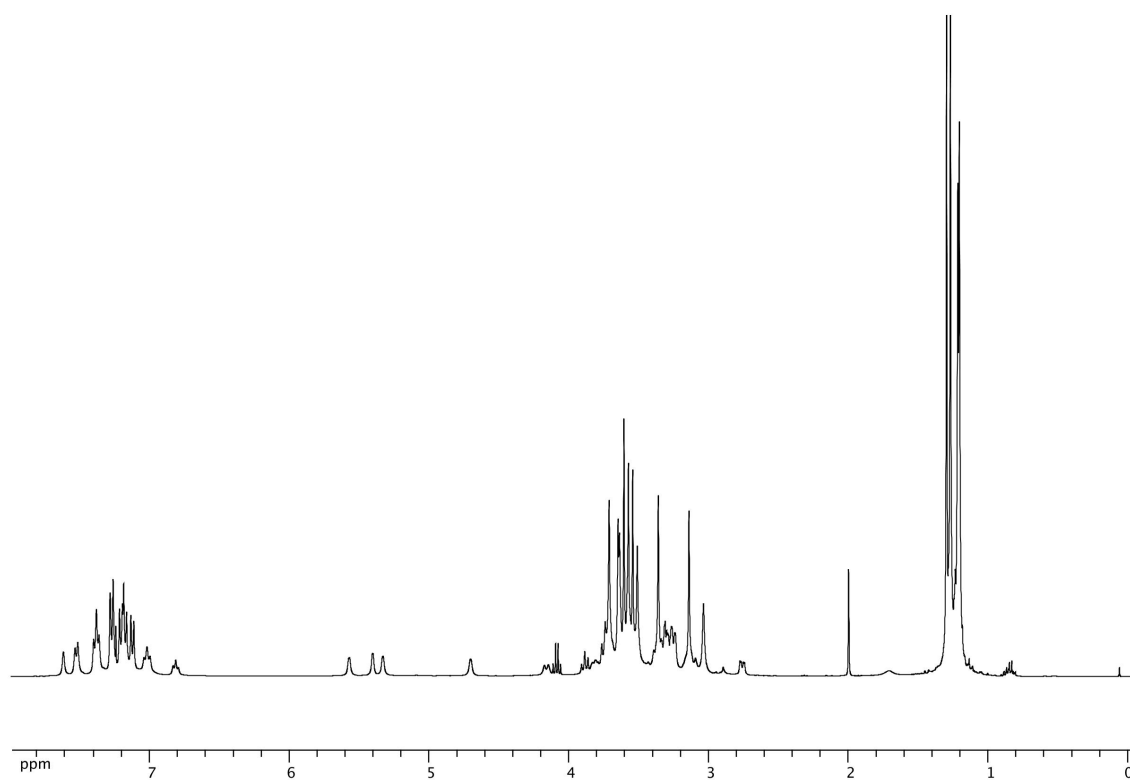


	A	B	C	D	E	F	G	H
H-1	5.26	4.27	5.48	5.87	5.08	5.67	5.02	5.27
H-2	3.15	2.71	3.42	3.36	2.78	3.24	3.16	3.23
H-3	3.46	3.22	3.82	3.41	3.38	3.35	3.76	3.81
H-4	3.34	3.73	4.02	3.89	3.76	3.52	3.83	3.68
H-5	3.40	3.23	3.94	3.68	3.32	3.82	3.69	3.55
H-6a	3.26	3.24	3.25	3.33	3.17	3.62	3.39	3.38
H-6b	3.30	4.23	3.52	3.41	4.20	4.23	3.53	3.60
OMe-6	—	—	3.26	—	—	2.80	2.98	3.31

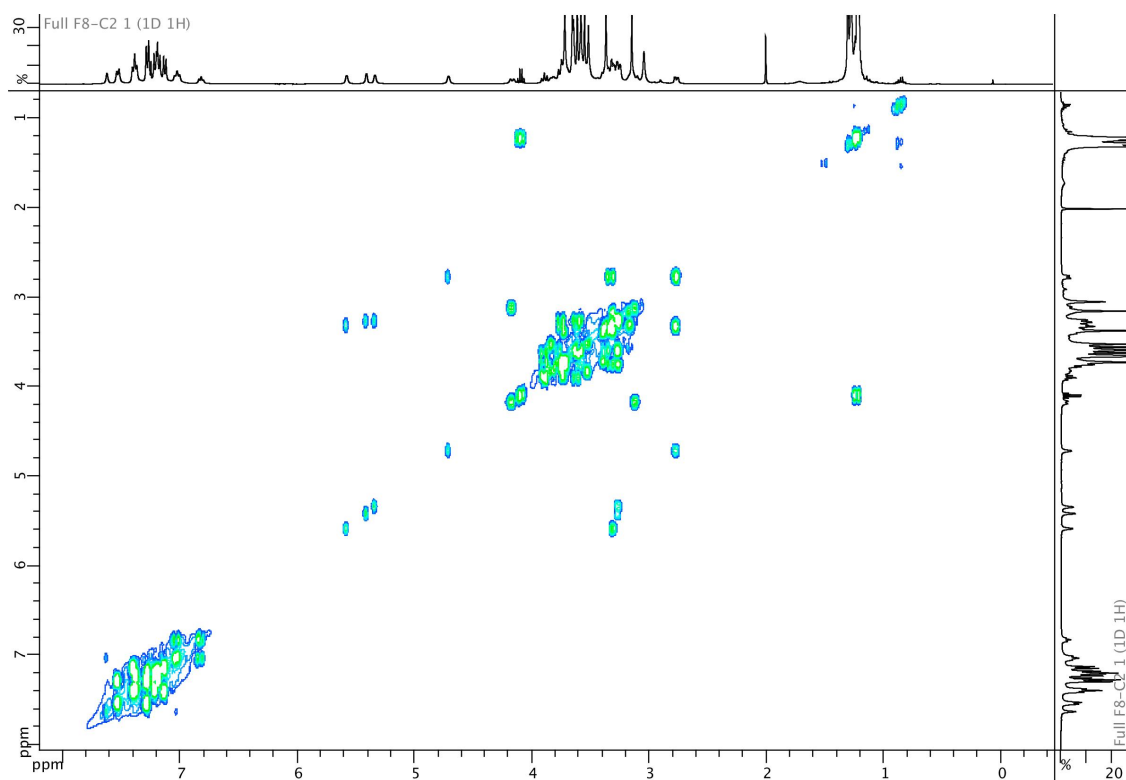
Table 4. Assignment of the aromatic protons of **7** (based on TOCSY, COSY, ROESY and HMQC).



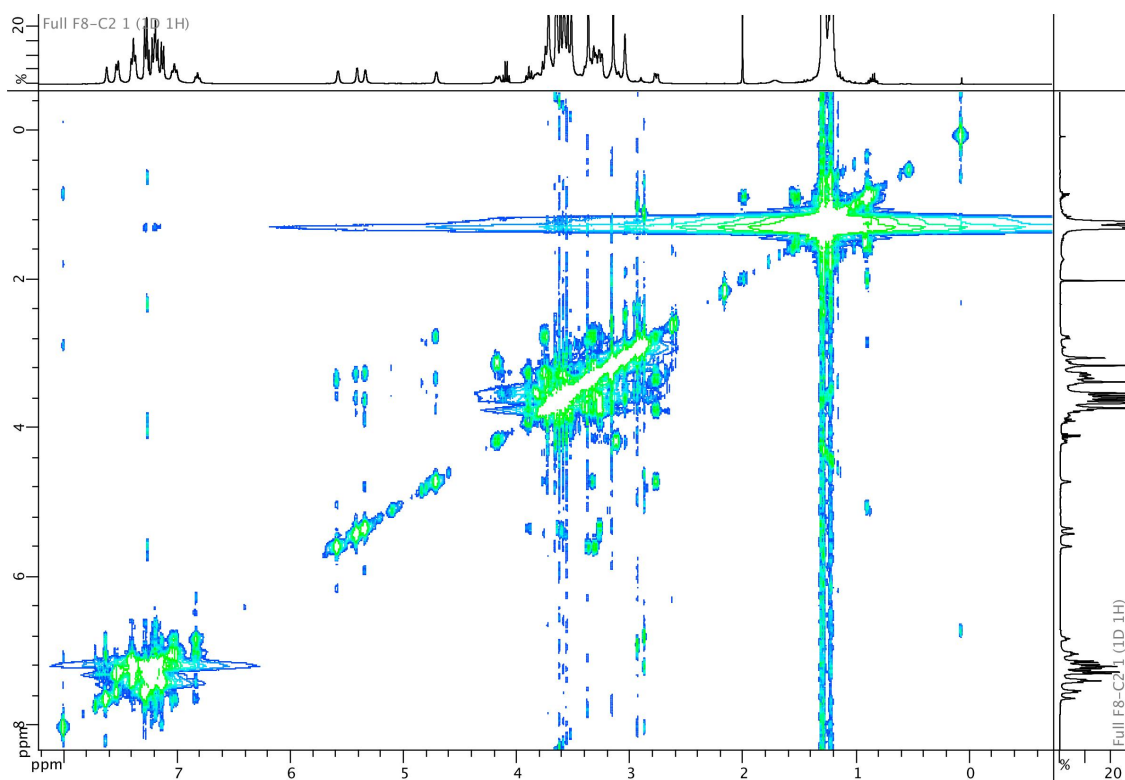
	A,B bridge		D,E bridge
H _a	7.60	H _{a'}	7.83
H _b	7.08	H _{b'}	7.09
H _c	7.04	H _{c'}	7.05
H _d	6.99	H _{d'}	6.97
H _e	7.27	H _{e'}	7.32
H _f	7.26	H _{f'}	7.16
H _g	7.36	H _{g'}	7.27
H _h	7.16	H _{h'}	7.05
H _i	7.52	H _{i'}	7.32
H _j	7.27	H _{j'}	7.23
H _k	7.45	H _{k'}	7.50
H _l	7.19	H _{l'}	7.26



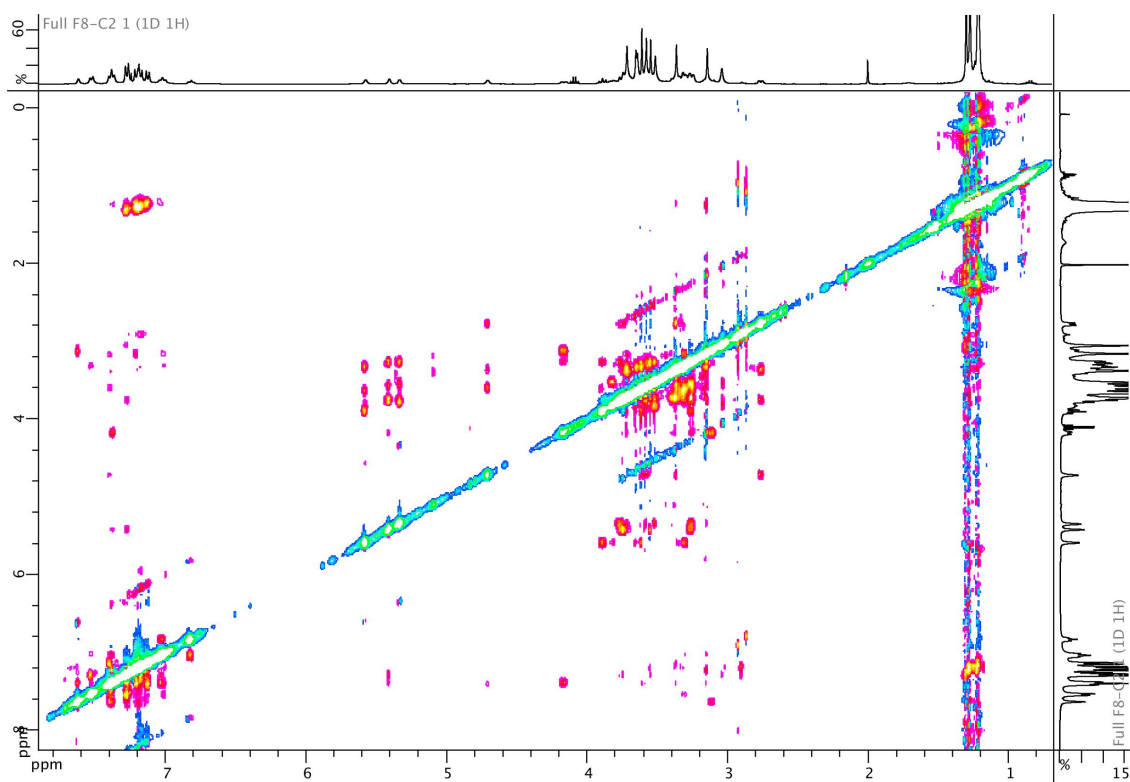
^1H NMR spectrum of **8**.



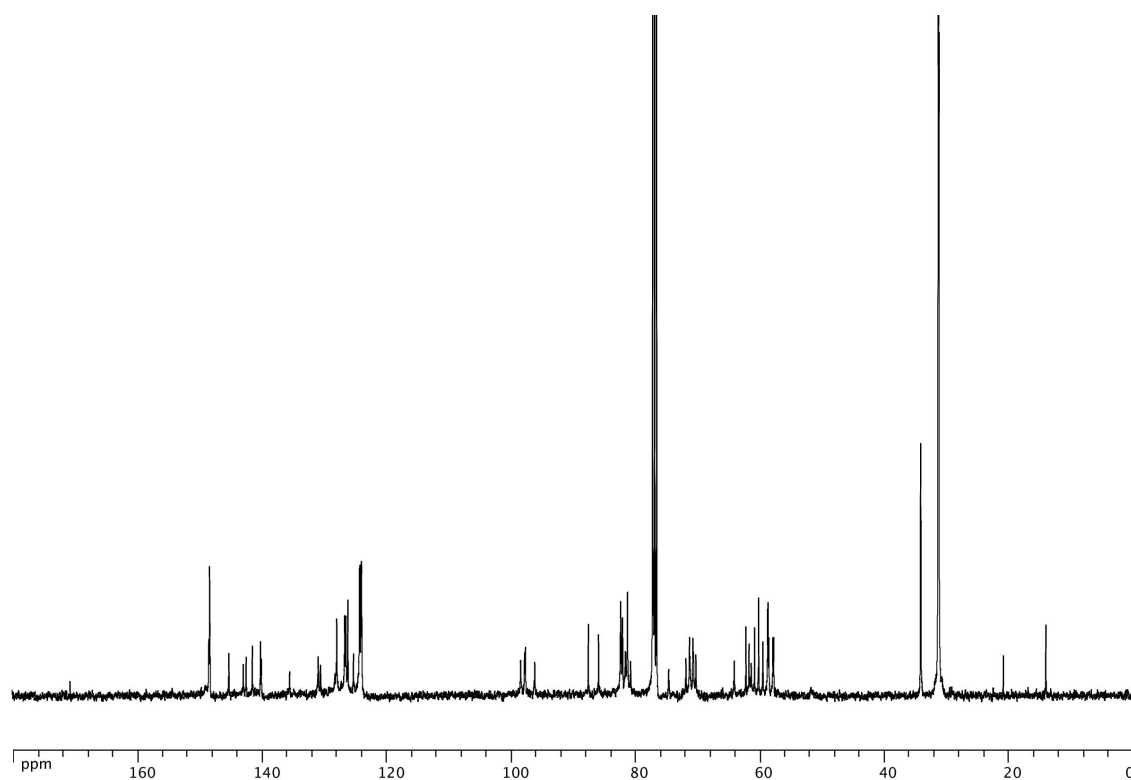
$^1\text{H}/^1\text{H}$ COSY spectrum of **8**.



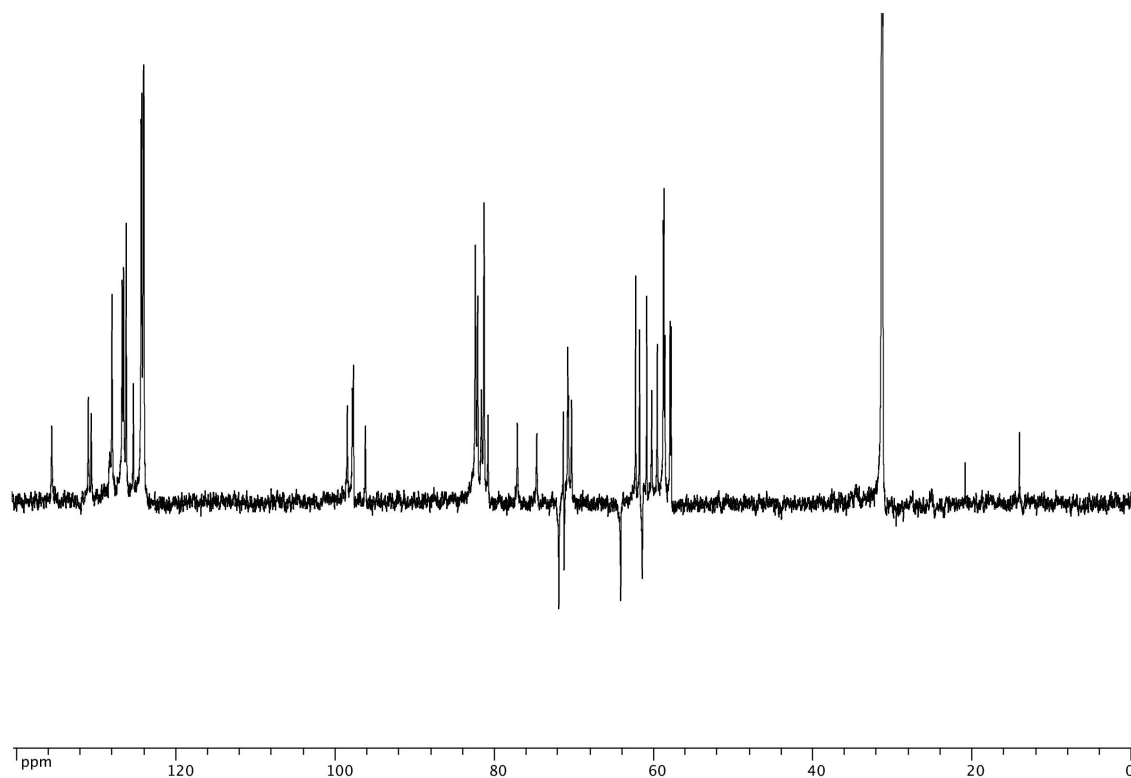
$^1\text{H}/^1\text{H}$ TOCSY spectrum of **8**.



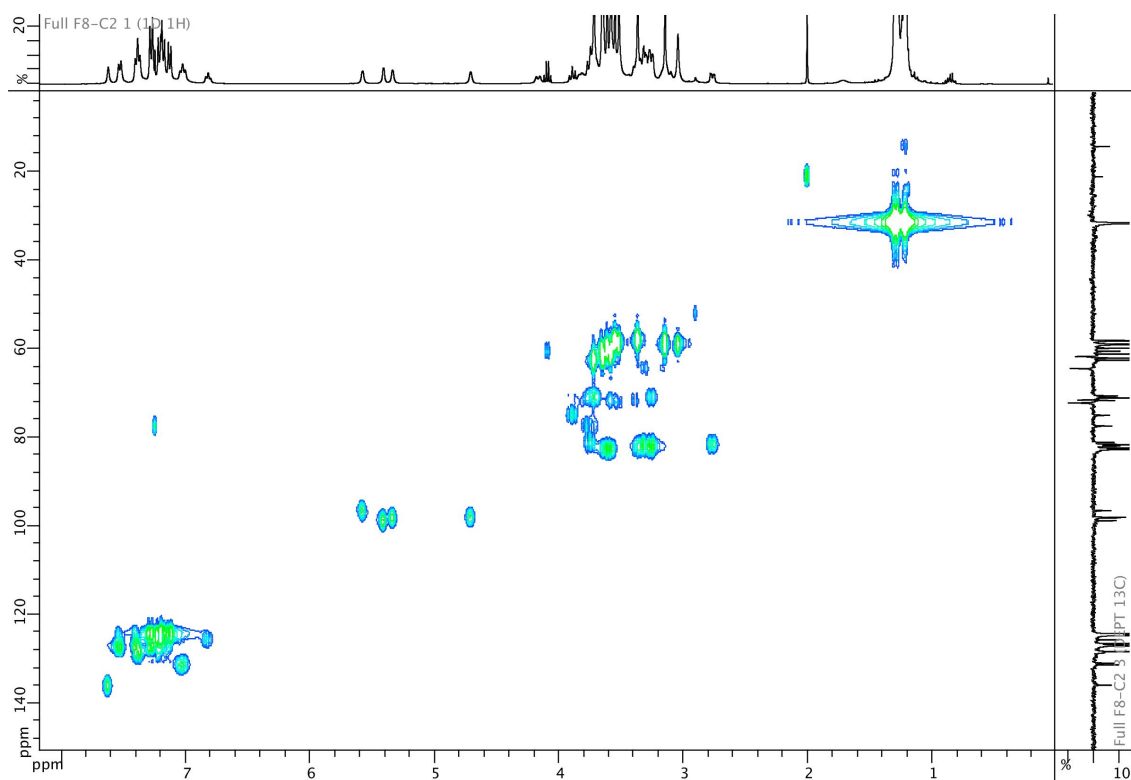
$^1\text{H}/^1\text{H}$ ROESY spectrum of **8**.



$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **8**.



DEPT 135 spectrum of **8**.



$^1\text{H}/^{13}\text{C}$ HMQC spectrum of **8**.

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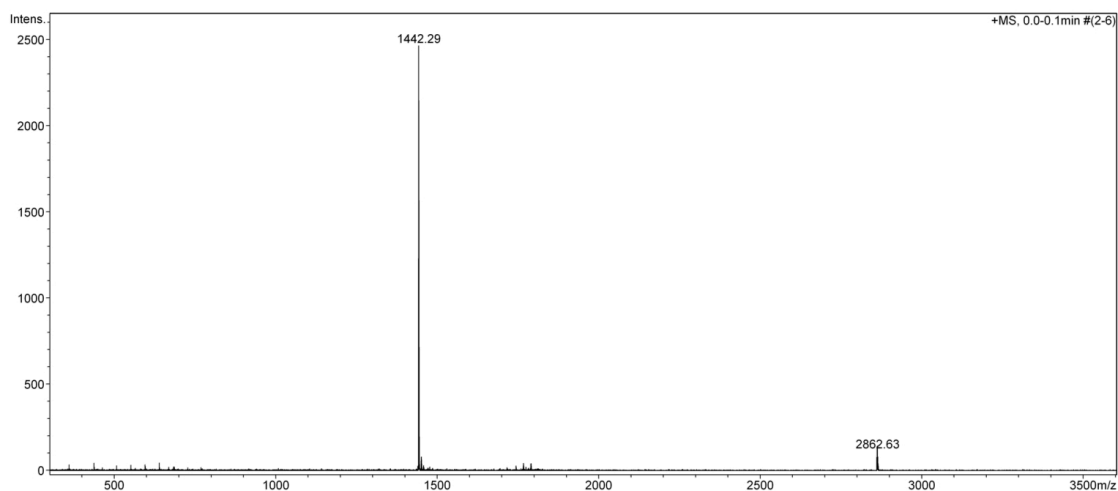
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Sample Name MJ83-F8-C2
Comment

Acquisition Date 10/14/2011 2:14:20 PM
Operator Administrator
Instrument micrOTOF

Acquisition Parameter

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Scan Range	n/a	Set Skimmer 1	50.0 V	Dry Heater	180 °C	APCI Heater	517 °C



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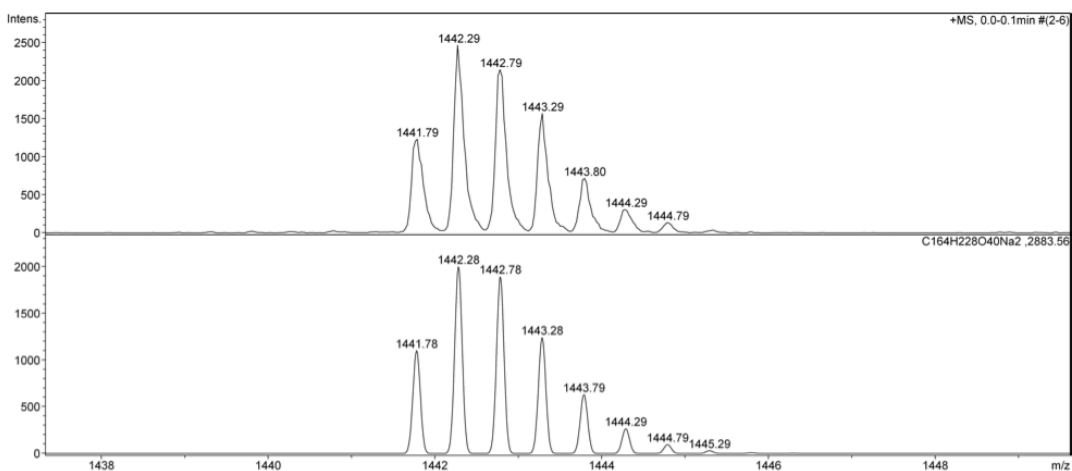
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Sample Name MJ83-F8-C2
Comment

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Operator Administrator
Instrument micrOTOF

Acquisition Parameter

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Scan Range	n/a	Set Skimmer 1	50.0 V	Dry Heater	180 °C	APCI Heater	517 °C



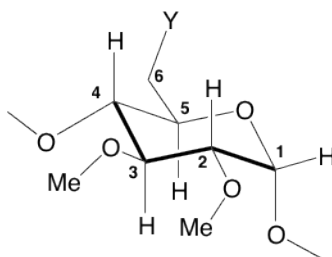
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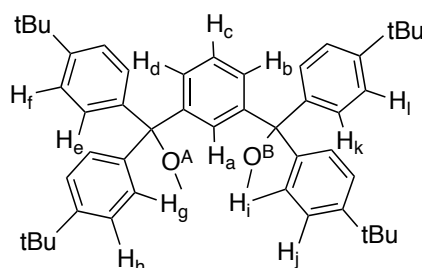
Mass spectrum of **8** and simulation.

Table 5. Assignment of the sugar protons of **8** (based on TOCSY, COSY, ROESY and HMQC).

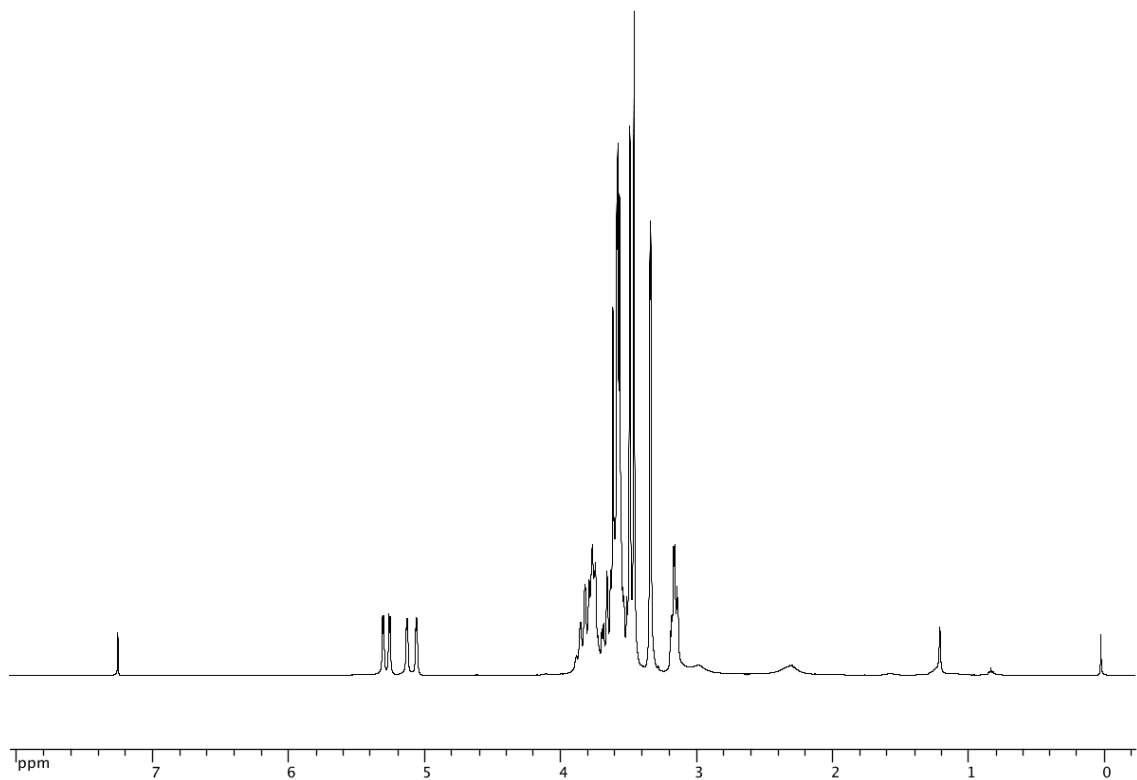


	A	B	C	D
H-1	5.57	4.70	5.40	5.33
H-2	3.29	2.76	3.25	3.26
H-3	3.35	3.32	3.59	3.63
H-4	3.57	3.75	3.77	3.89
H-5	3.56	3.23	3.70	3.73
H-6a	3.15	3.10	3.24	3.59
H-6b	3.30	4.16	3.33	3.61
OMe-6	—	—	3.14	3.03

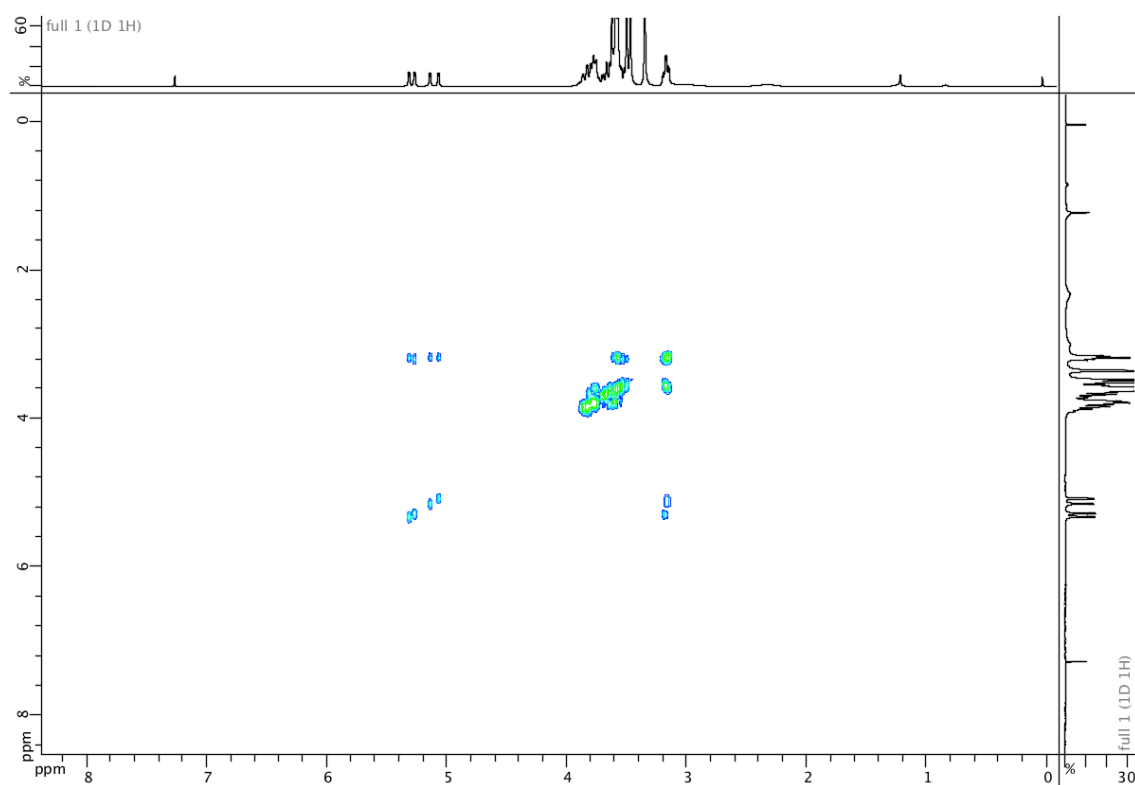
Table 6. Assignment of the aromatic protons of **8** (based on TOCSY, COSY, ROESY and HMQC).



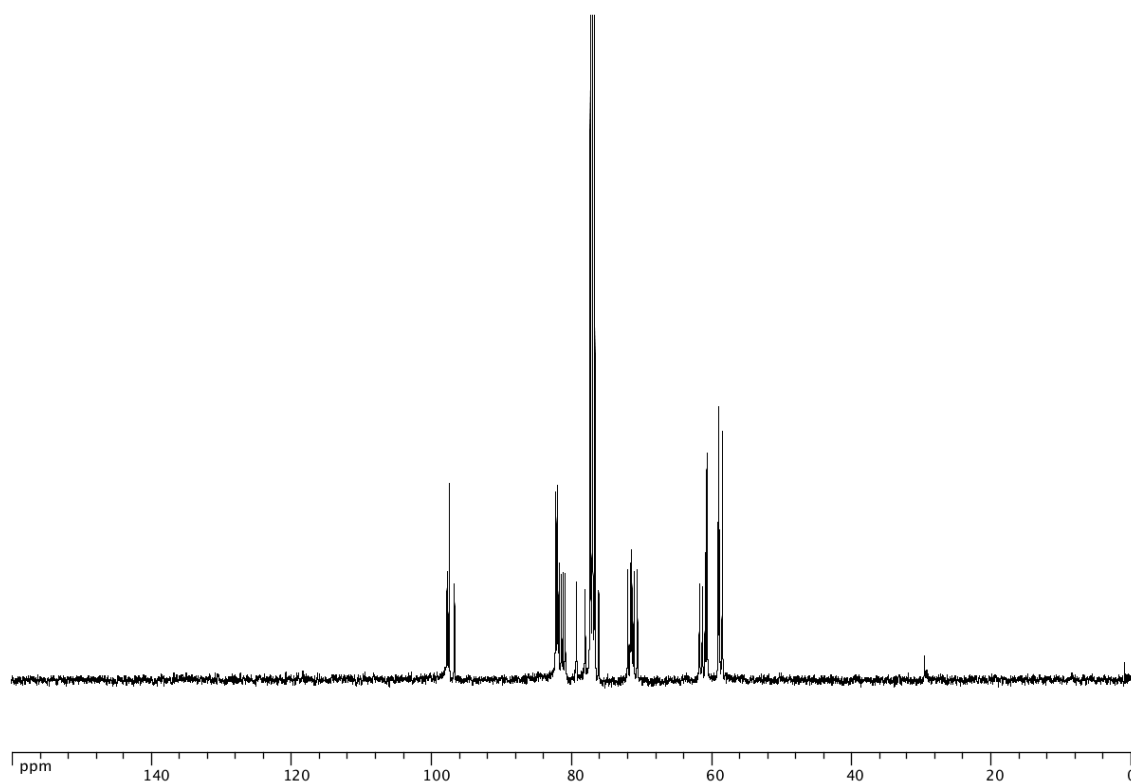
	A,B (or E,F) bridge
H _a	7.62
H _b	7.03
H _c	7.02
H _d	7.00
H _e	7.27
H _f	7.17
H _g	7.52
H _h	7.27
H _i	7.37
H _j	7.20
H _k	7.39
H _l	7.12



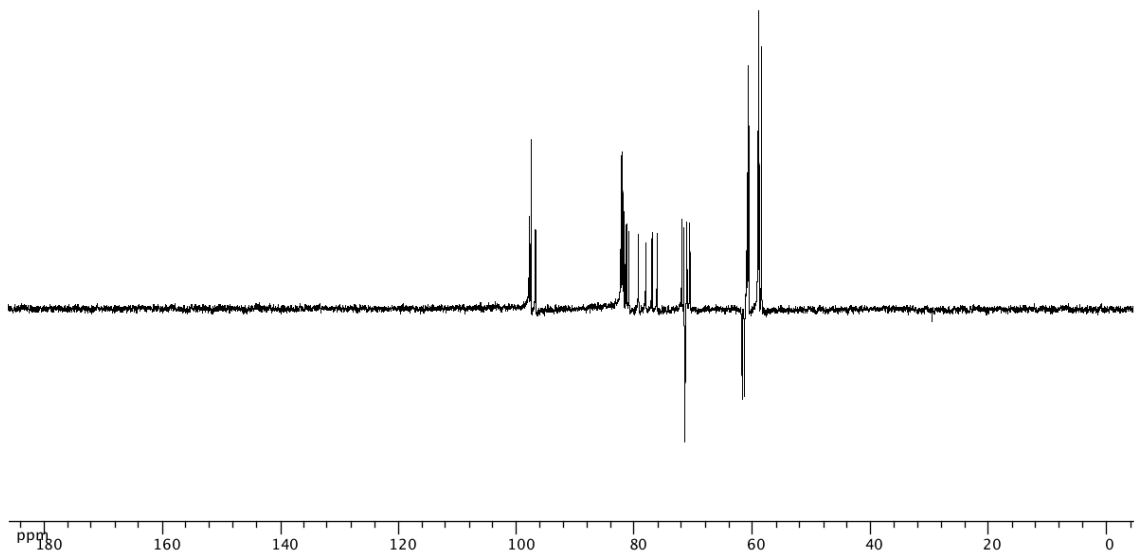
^1H NMR spectrum of **9**.



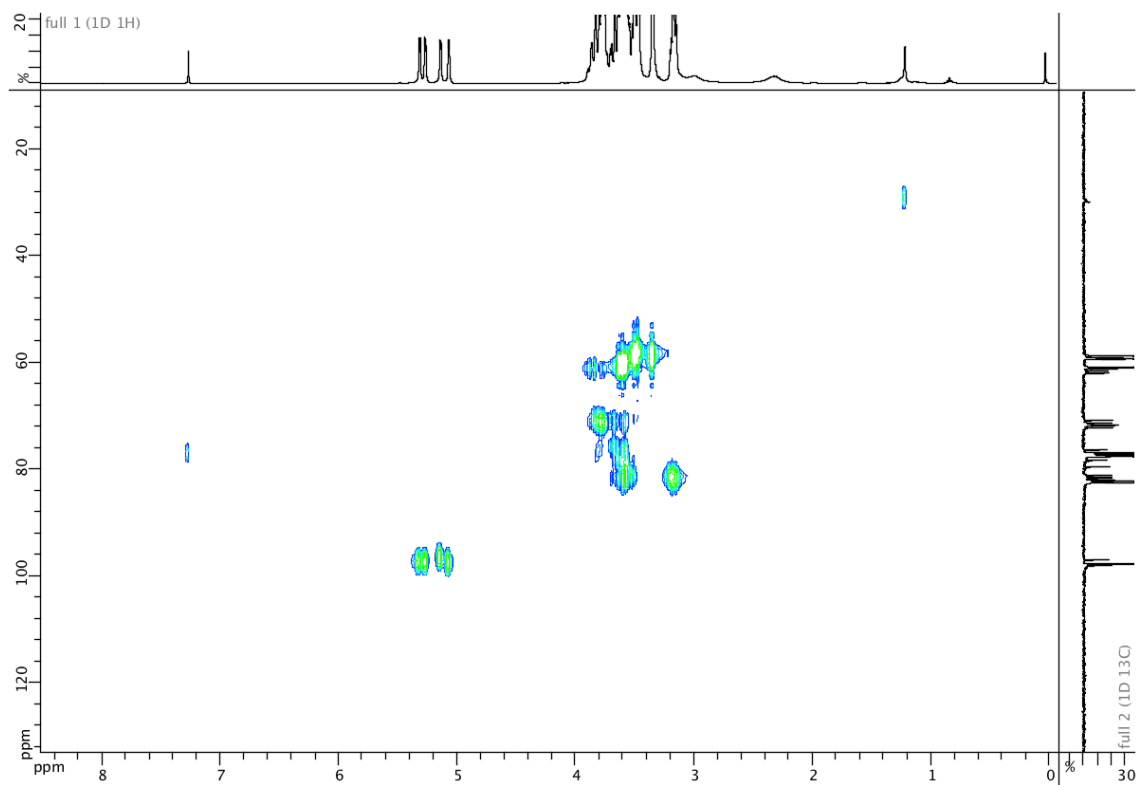
$^1\text{H}/^1\text{H}$ COSY spectrum of **9**.



$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **9**.



DEPT 135 spectrum of **9**.



$^1\text{H}/^{13}\text{C}$ HMQC spectrum of **9**.

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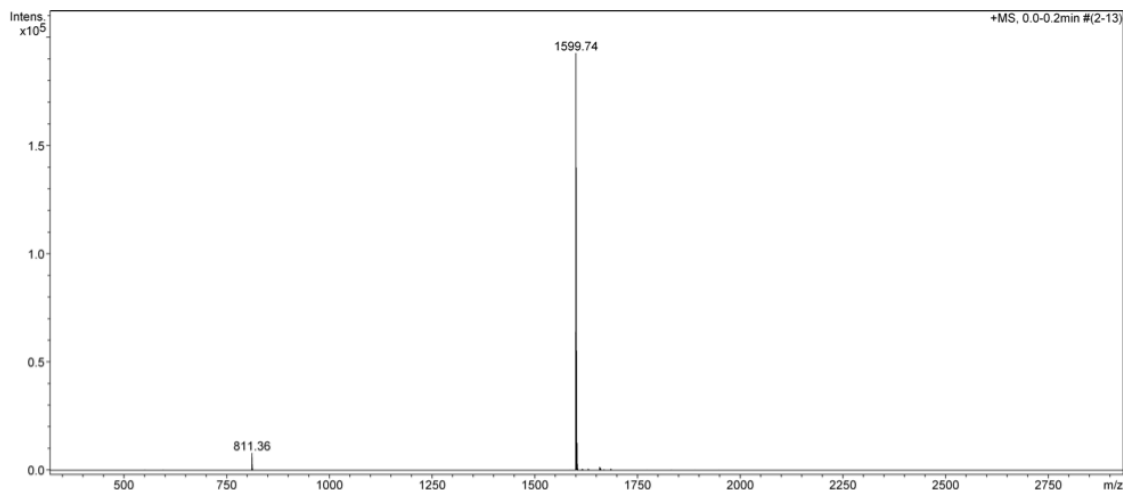
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Method esi wide pos.m
Sample Name MJ85 C2
Comment

Acquisition Date 11/10/2011 3:46:33 PM
Operator Administrator
Instrument micrOTOF

Acquisition Parameter

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Scan Range	n/a	Set Skimmer 1	50.0 V	Dry Heater	180 °C	APCI Heater	517 °C



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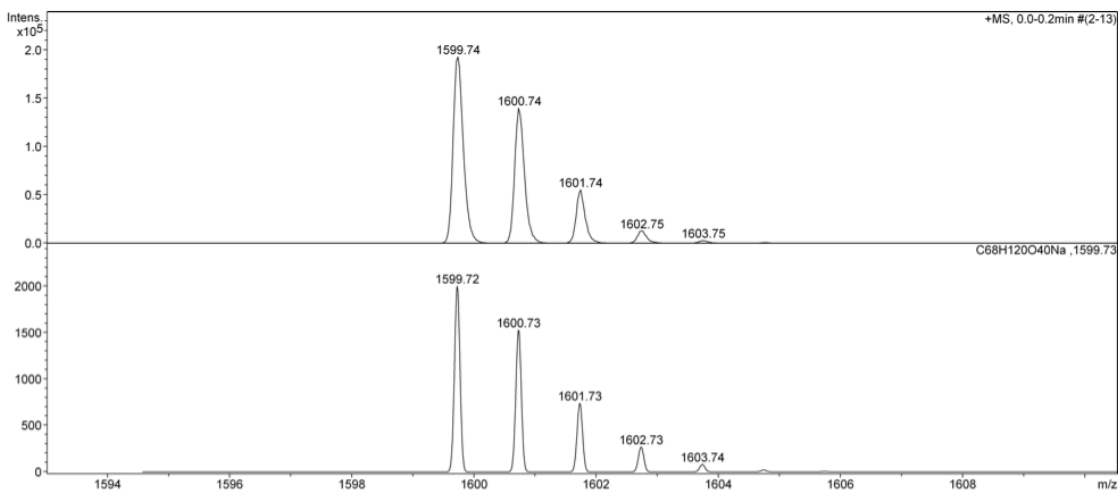
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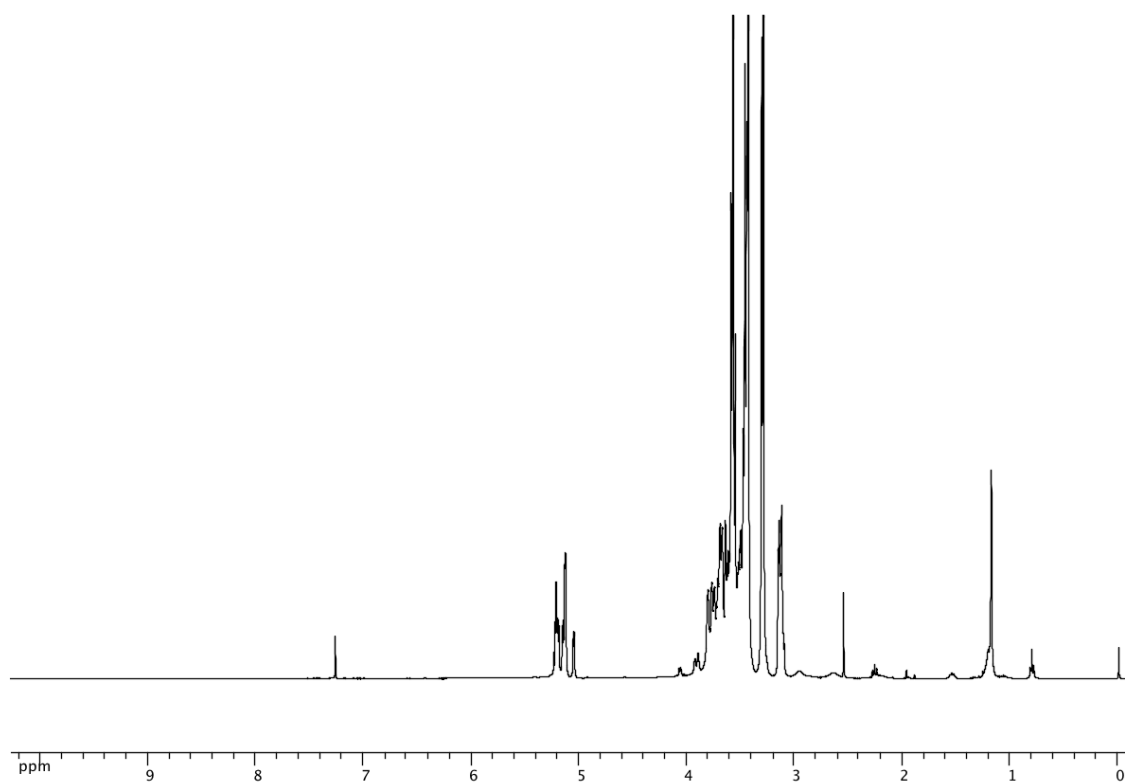


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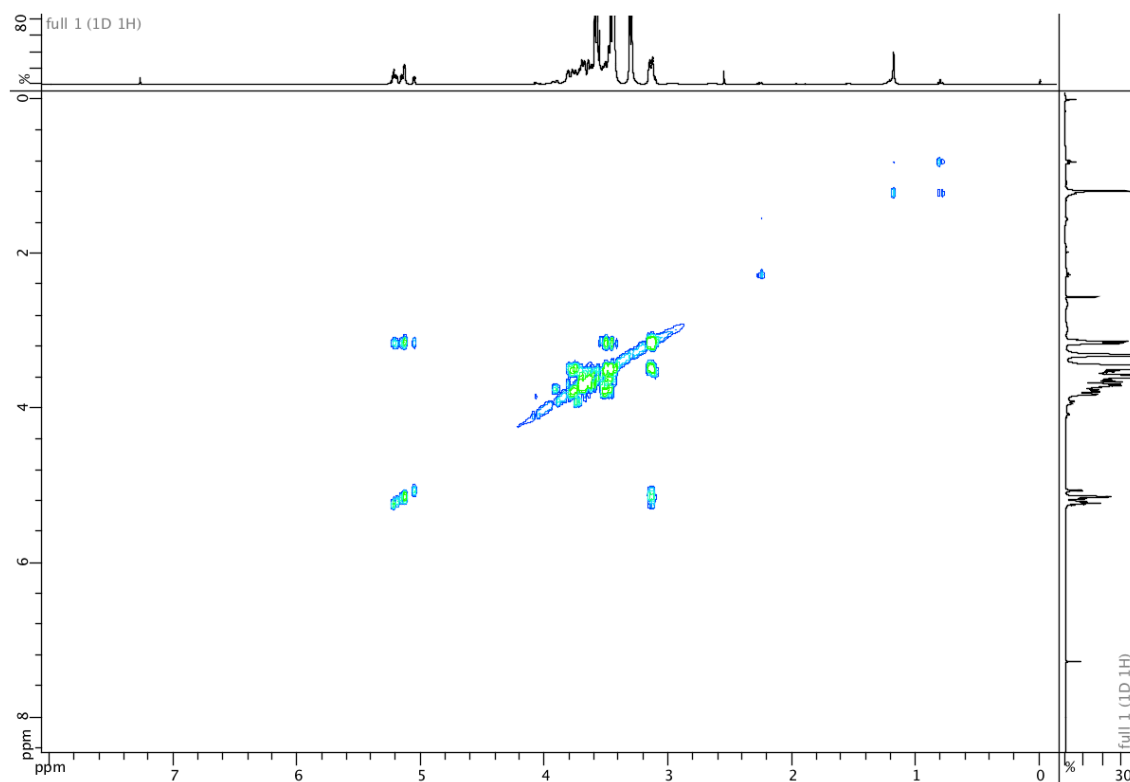
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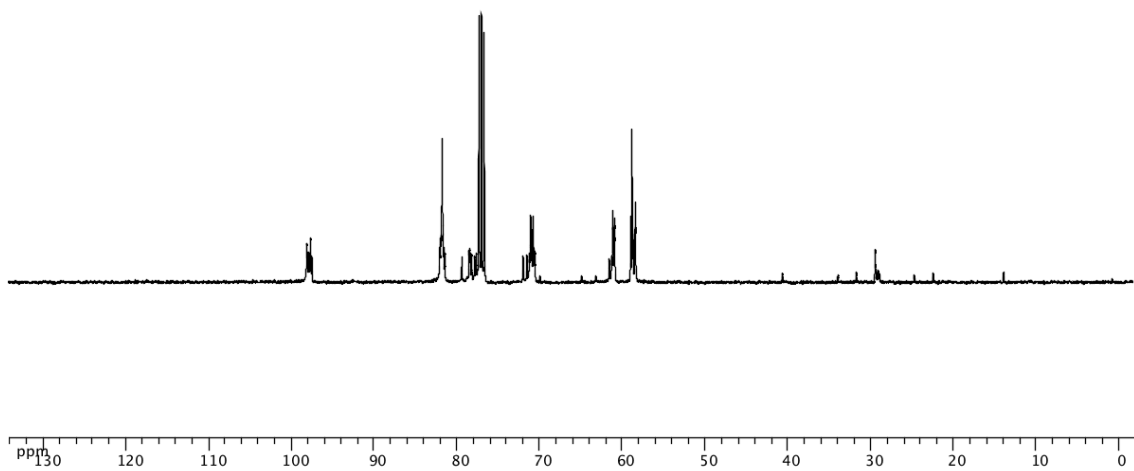
Mass spectrum of **9** and simulation.



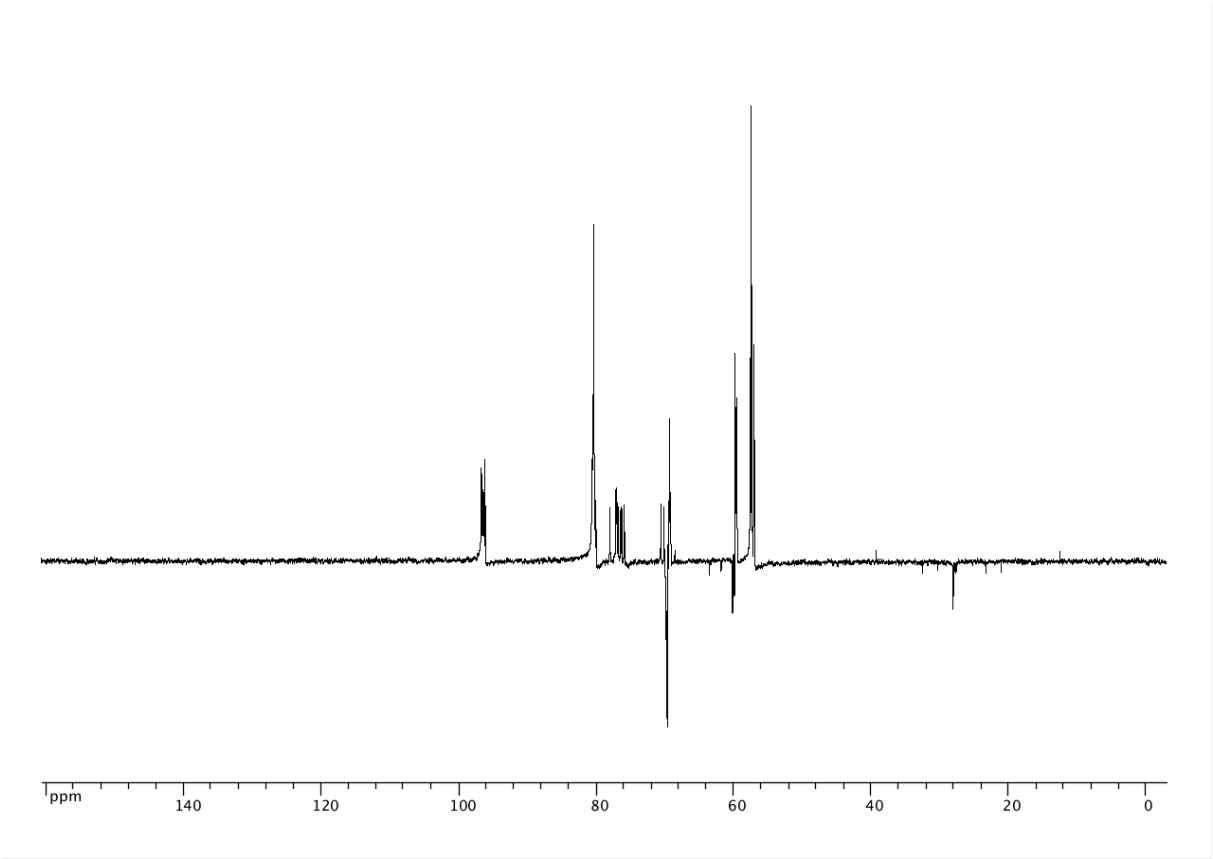
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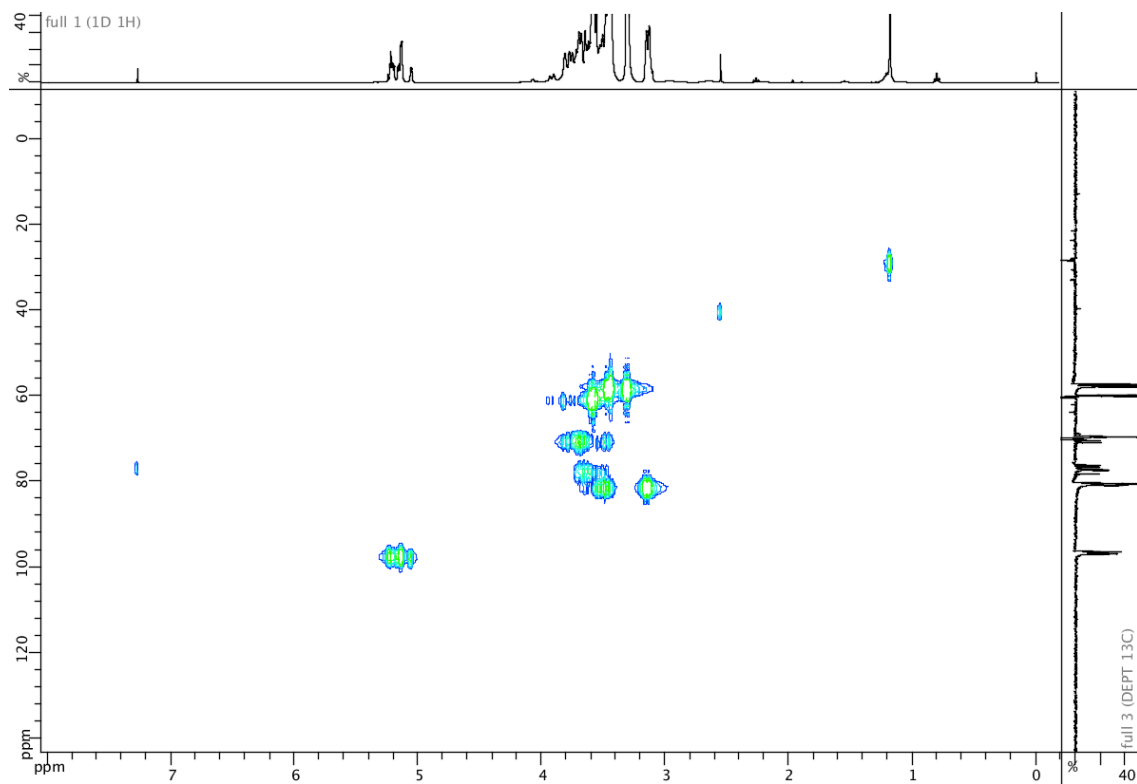
$^1\text{H}/^1\text{H}$ COSY spectrum of **10**.



$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **10**.



DEPT 135 spectrum of **10**.



$^1\text{H}/^{13}\text{C}$ HMQC spectrum of **10**.

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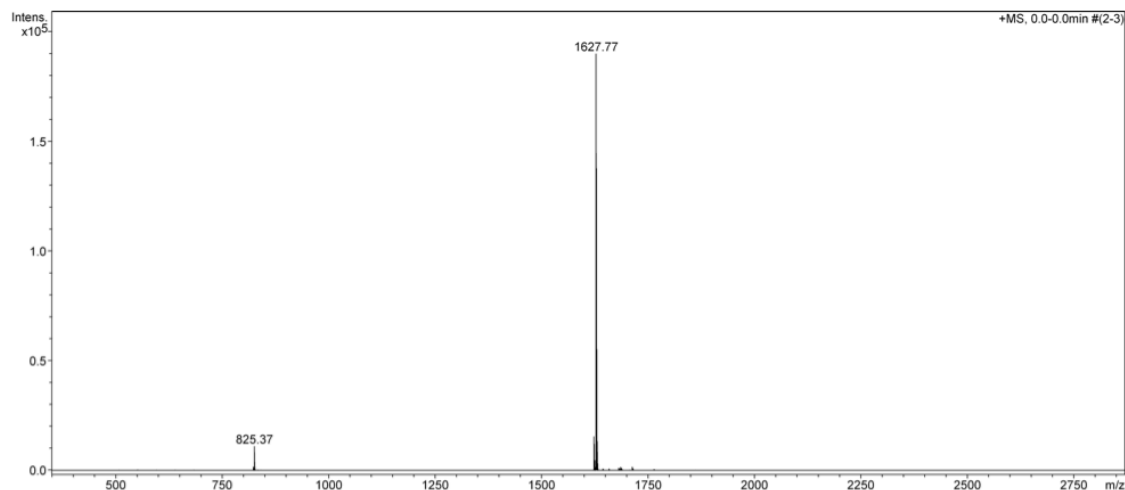
Analysis Info

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Method esi wide pos.m
Sample Name MJ85 diol
Comment

Acquisition Date 11/10/2011 3:39:03 PM
Operator Administrator
Instrument micrOTOF

Acquisition Parameter

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Bruker Daltonics DataAnalysis 3.1

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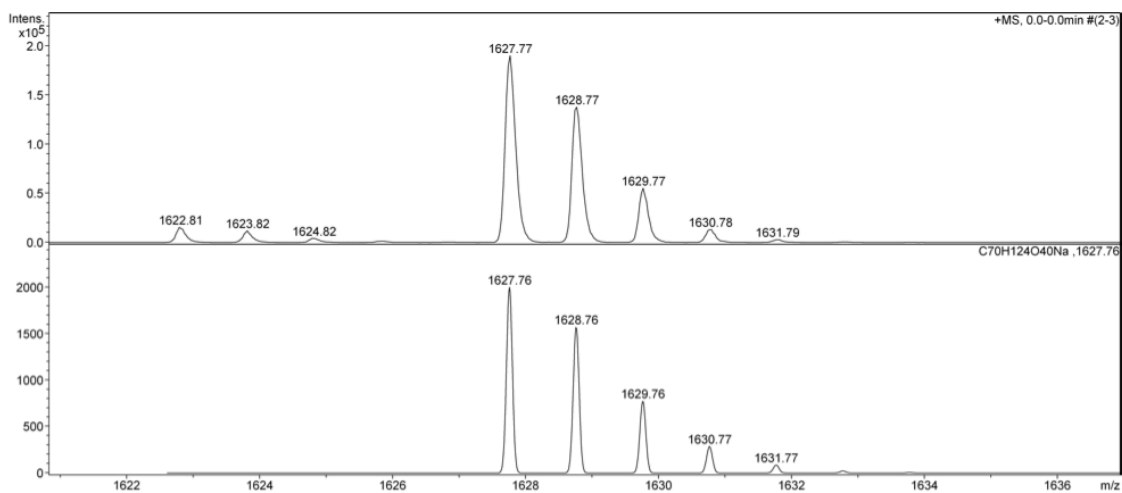
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Operator Administrator
Instrument micrOTOF

Acquisition Parameter

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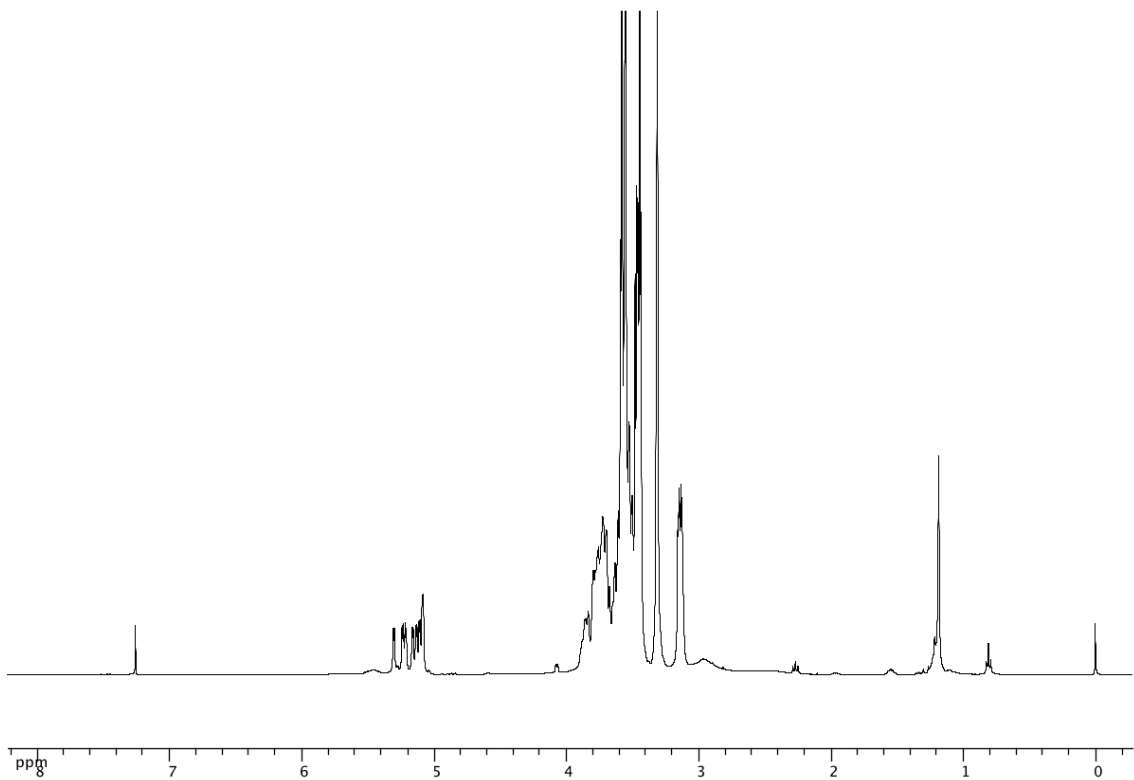


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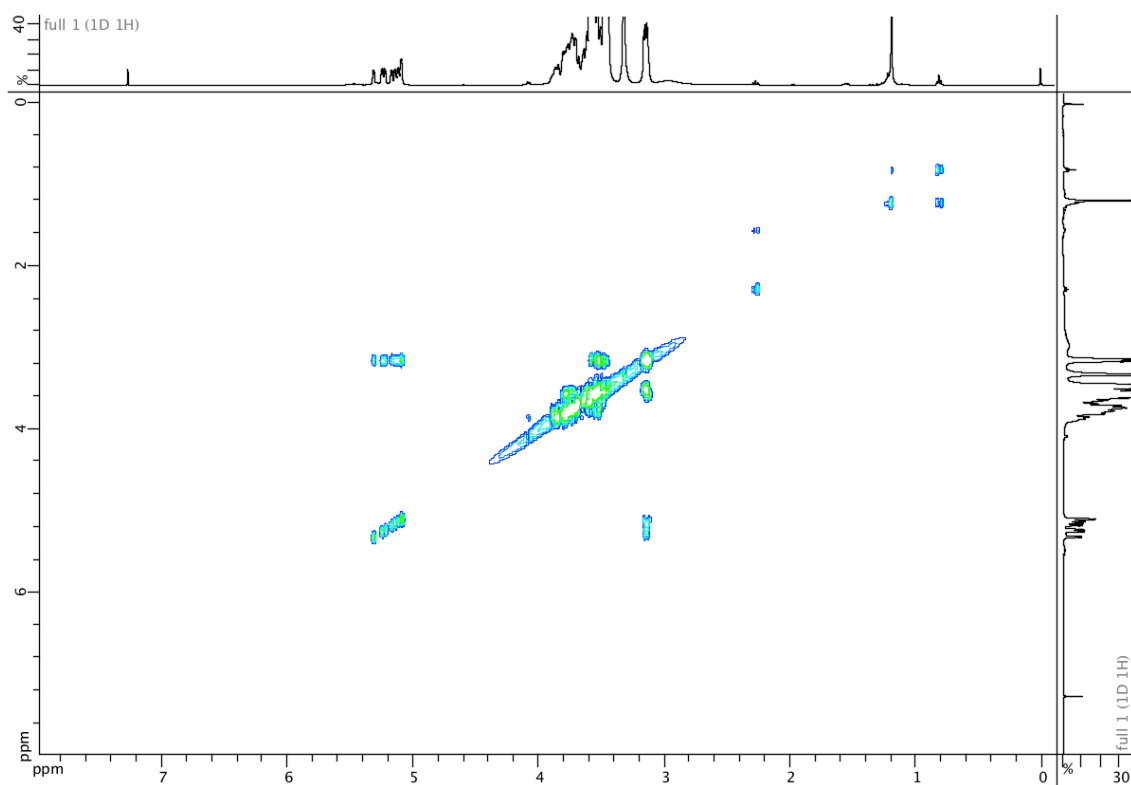
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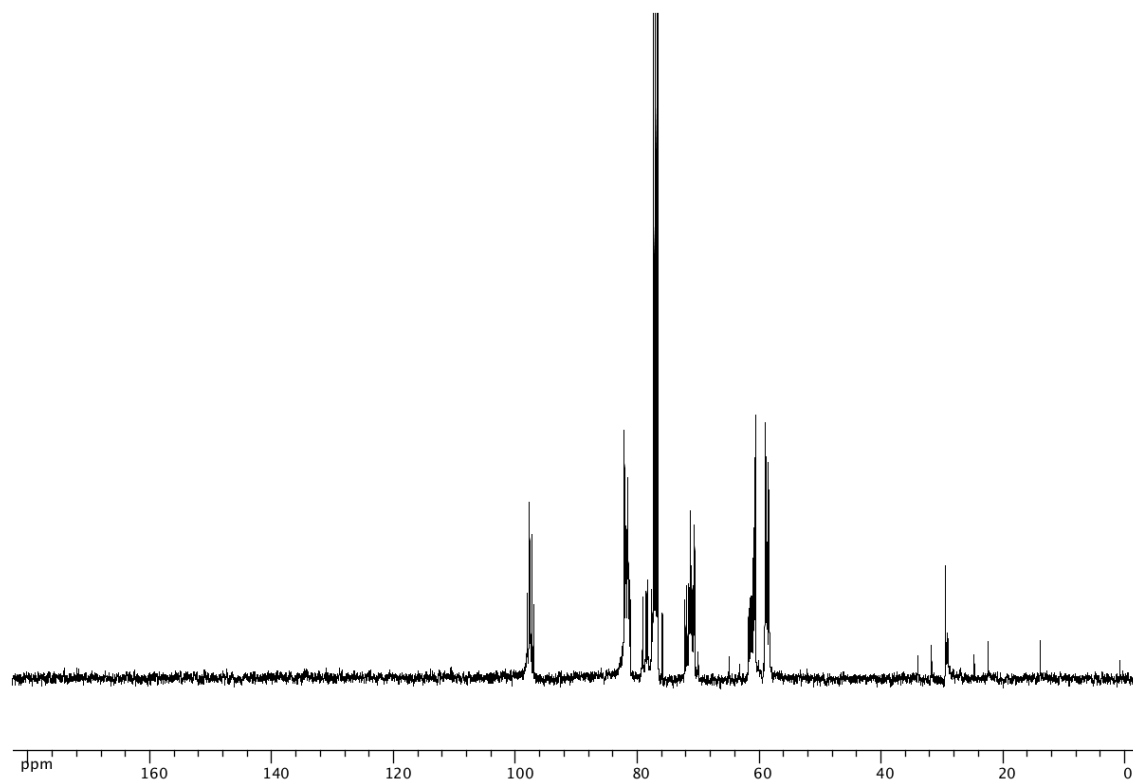
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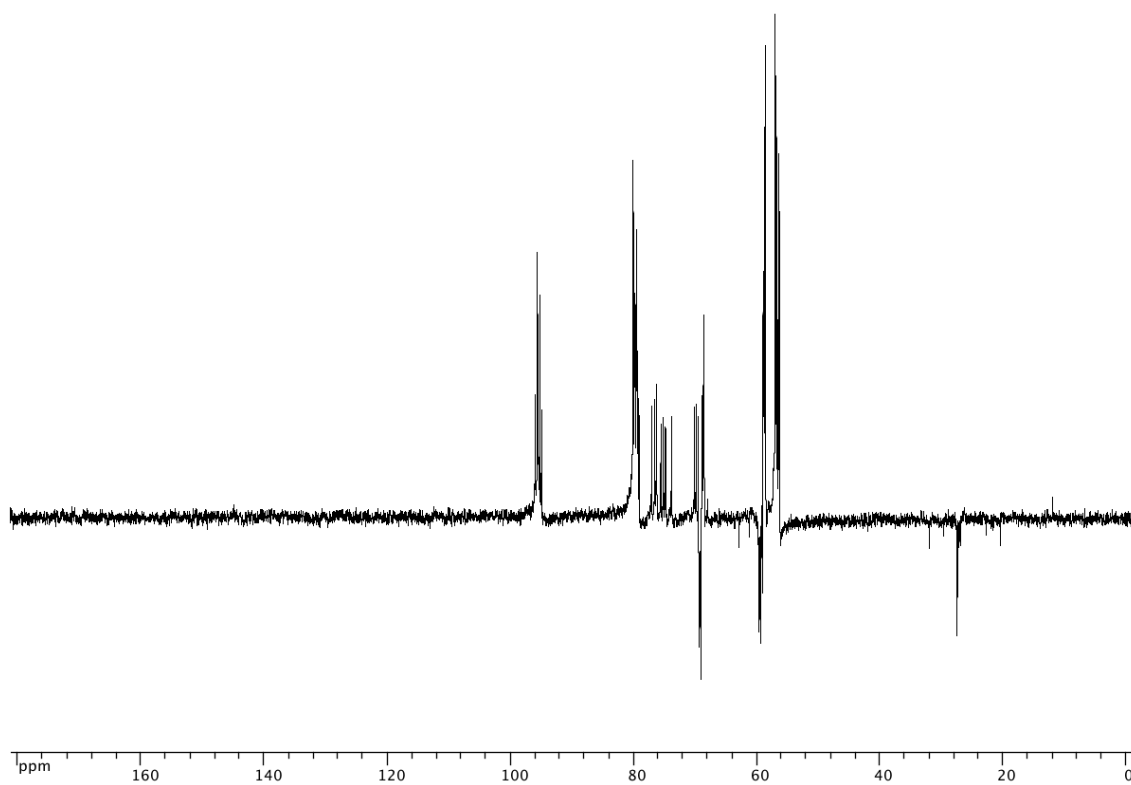
^1H NMR spectrum of **11**.



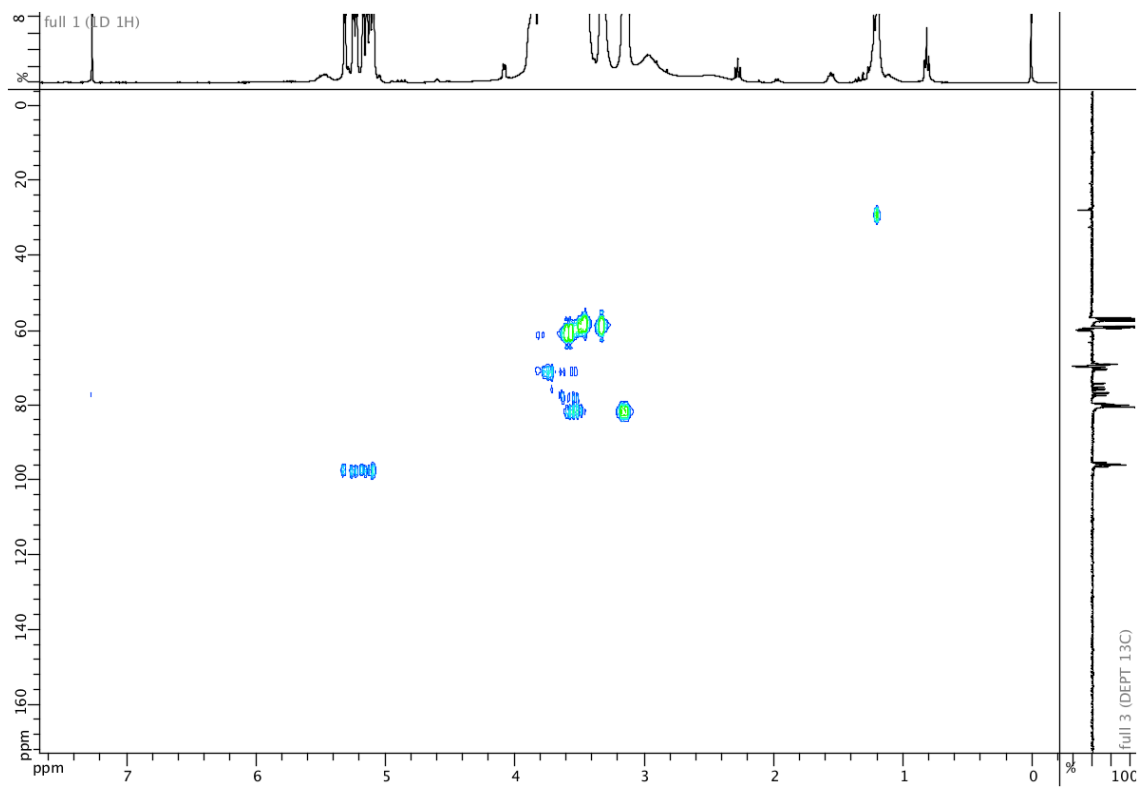
$^1\text{H}/^1\text{H}$ COSY spectrum of **11**.



$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **11**.



DEPT 135 spectrum of **11**.



$^1\text{H}/^{13}\text{C}$ HMQC spectrum of **11**.

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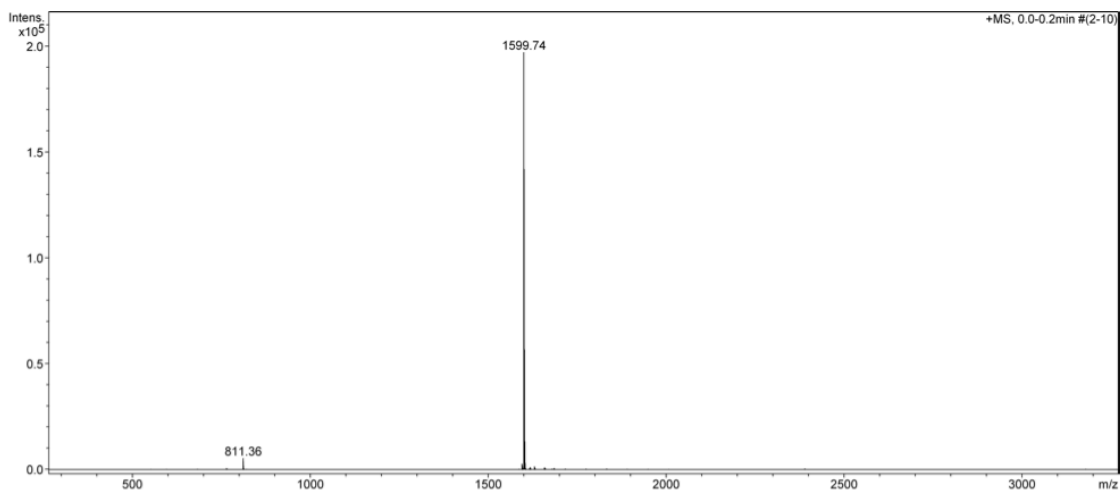
Analysis Info

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Method esi wide pos.m
Sample Name MJ85 C1
Comment

Acquisition Date 11/10/2011 3:42:37 PM
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Bruker Daltonics DataAnalysis 3.1

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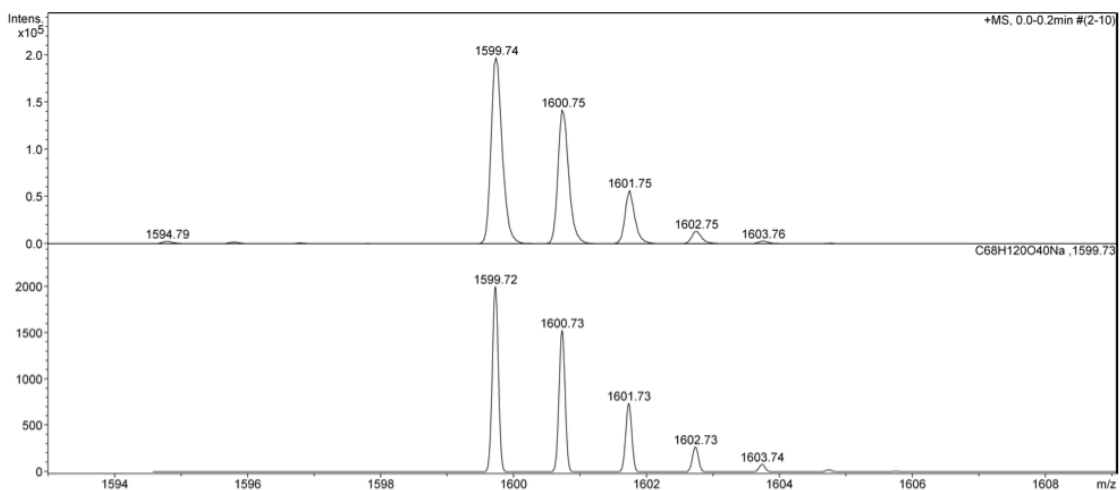
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Instrument micrOTOF

Acquisition Parameter

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Scan Range	n/a	Set Skimmer 1	50.0 V	Dry Heater	180 °C	APCI Heater	517 °C



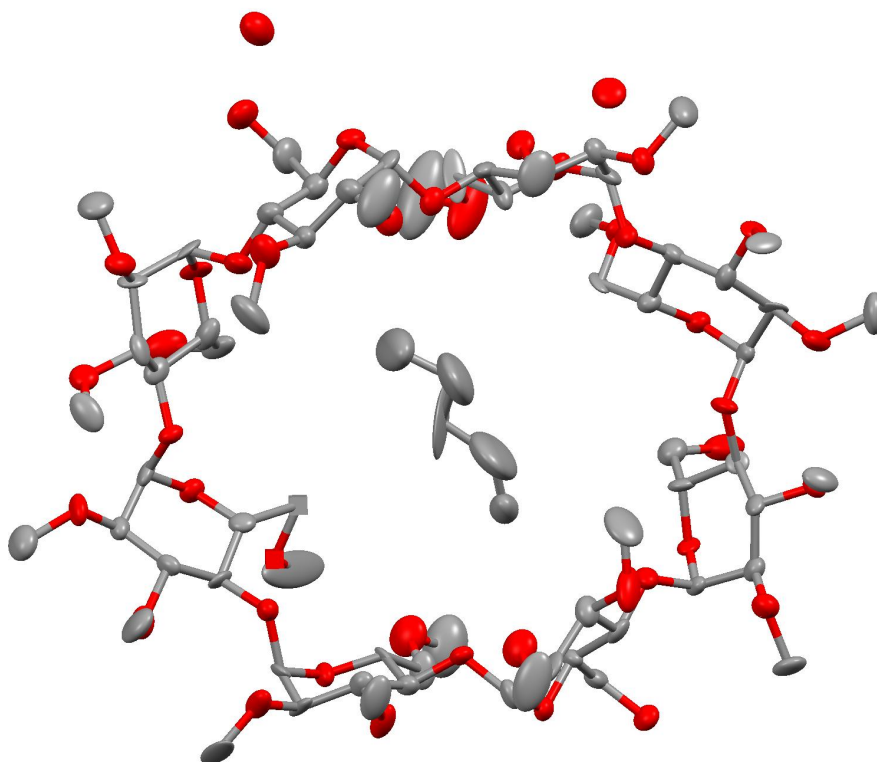
Bruker Daltonics DataAnalysis 3.1

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Mass spectrum of **11** and simulation.

Crystal structure analysis



X-ray crystallographic data of 9: Single crystals of **9** were obtained by slow diffusion of pentane into a dichloromethane solution of the compound. Crystal data for $C_{70.5}H_{130}O_{42}$ (**9**·2H₂O·0.5C₅H₁₂), $M_r = 1649.74$, orthorhombic, space group $P2_12_12_1$, $a = 15.485(1)$, $b = 23.254(1)$, $c = 27.718(1)$ Å, $V = 9981.2(8)$ Å³, $Z = 4$, $\rho_{\text{calcd}} = 1.098$ g cm⁻³, $\lambda(\text{MoK}\alpha) = 0.71073$ Å, $\mu = 0.090$ mm⁻¹, $F(000) = 3556$, $T = 100(2)$ K. The sample ($0.32 \times 0.25 \times 0.07$ mm) was studied with an Oxford Diffraction Xcalibur Saphir 3 CCD with graphite monochromatised MoK α radiation. The structure was solved with SIR-97,¹⁶ which revealed the non-hydrogen atoms of the molecule. After anisotropic refinement, many hydrogen atoms were found with a Fourier difference analysis. The whole structure was refined with SHELX-97¹⁷ and full-matrix least-square techniques. Use of F^2 magnitude; x, y, z, β_{ij} for C, O atoms, x, y, z , in riding mode for H atoms, 982 variables and 5983 observations with $[I > 2.0\sigma(I)]$, calcd $w = 1/[\sigma^2(F_o^2) + (0.1186 P)^2]$ where $P = (F_o^2 + 2 F_c^2)/3$ with the resulting $R = 0.0700$, $R_w = 0.2129$, and $S_w = 0.691$, $\Delta\rho < 0.409$ e Å⁻³. CCDC 862358 (**9**) contain the supplementary crystallographic data for this report. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

General procedure for assignment of the glucose units linked by a given capping unit.³

Our strategy for full structural assignment began with the differentiation between capped and non-capped C-6 carbon atoms by DEPT 135. These appear as two distinct sets of signals. The H-6 protons were then identified using ^1H - ^{13}C HMQC (Heteronuclear Multiple Quantum Coherence spectroscopy). By using TOCSY (TOtal Correlation SpectroscopY) and COSY (COrelated SpectroscopY), each H-6 proton was correlated to the set of protons belonging to the same glucose residue. The connectivity between individual glucose units was then established via a ROESY (Rotating frame Overhauser Effect SpectroscopY) experiment showing the proximity between H-4_N and H-1_{N+1} protons (N and N+1 standing for neighbouring glucose moieties labelled in the alphabetical order).

References

1. A. Altomare, J. Foadi, C. Giacovazzo, A. G. G. Moliterni, M. C. Burla and G. Polidori, *J. Appl. Crystallogr.*, 1998, **31**, 74-77.
2. G. M. Sheldrick, SHELXL-97, *Program for Crystal Structure Refinement*, 1997, University of Göttingen, Göttingen (Germany).
3. H.-J. Schneider, F. Hacket and V. Rüdiger, *Chem. Rev.*, 1998, **98**, 1755-1785.