#### **Electronic Supporting Information**

#### Rhodium(III)-Catalyzed Oxidative Olefination of N-Allyl Sulfonamides

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#### **1.** General Considerations

All reactions were carried out under an atmosphere of argon unless otherwise noted. Reactions temperatures are reported as those of the oil bath. The solvents used were purified by distillation and were transferred under argon.

Commercially available chemicals were obtained from Sigma-Aldrich, Alfa Aesar, TCI and Aladdin and were used as received unless otherwise stated. [RhCp\*Cl<sub>2</sub>]<sub>2</sub> were purchased from Strem.

Reactions were monitored with analytical thin-layer chromatography (TLC) on silica. GC analyses were performed on Shimadzu GC-2010 (Column: GL Science, TCWAX, 0.25 mm ID, 0.25 mm, 60.0 m; Gas pressure: 85 kPa; Total flow: 8.6 mL/min; Column flow: 1.14 mL/min; Velocity: 39.8 cm/sec; Column program: starting from 100°C, 10 min hold, 100°C/min to 270°C, 25 min hold. NMR spectra were recorded on Bruker-DRX (400 MHz or 500 MHz) spectrometers. Chemical shifts ( $\delta$ ) are given in ppm relative to SiMe<sub>4</sub>. The residual solvent signals were used as references and the chemical shifts were converted to the TMS scale (CDCl<sub>3</sub>:  $\delta_{\rm H} = 7.26$  ppm,  $\delta_{\rm C} = 77.00$  ppm; MeOD:  $\delta_{\rm H} = 3.31$  ppm,  $\delta_{\rm C} = 49.15$  ppm).

#### 2. Preparation of starting materials

#### Synthesis of N-allyl Sulfonamides



In a round-bottom flask, allylamine and triethylamine were combined in  $CH_2Cl_2$ . The flask was cooled in an ice bath, to which was slowly added the sulfonyl chloride. The ice bath was allowed to warm to room temperature and the mixture was stirred for 12 h. The reaction mixture was then diluted with  $CH_2Cl_2$  and washed with HCl (1M) and water. The organic layer was dried over  $Na_2SO_4$  and taken to dryness under reduced pressure. The product obtained was pure enough for further reaction.

### 3. Optimization of the Reaction Conditions

	) − S−NH + ≉ O 1a	O [RhC oxidant acetone, 10 2a	p*Cl <sub>2</sub> ]2 (2.1 equiv) 00 °C, 24 h	S-NH 3aa O-OMe
Entry	Catalyst	Loading (%)	Oxidant	Yield(%) <sup>[b]</sup>
1	[RhCp*Cl <sub>2</sub> ] <sub>2</sub>	5	Cu(OAc) <sub>2</sub>	42
2	[RhCp*Cl <sub>2</sub> ] <sub>2</sub>	5	$Ag_2CO_3$	53
3	[RhCp*Cl <sub>2</sub> ] <sub>2</sub>	5	Ag <sub>2</sub> O	nd
4	[RhCp*Cl <sub>2</sub> ] <sub>2</sub>	5	AgOAc	76
5	[RhCp*Cl <sub>2</sub> ] <sub>2</sub>	4	AgOAc	75
6	[RhCp*Cl <sub>2</sub> ] <sub>2</sub>	3	AgOAc	60
7 <sup>[c]</sup>	[RhCp*Cl <sub>2</sub> ] <sub>2</sub>	4	AgOAc	72
8 <sup>[d]</sup>	[RhCp*Cl <sub>2</sub> ] <sub>2</sub>	4	AgOAc	78

#### Table 1. Optimization Studies

<sup>[a]</sup> Conditions: *N*-Ts allylamine (0.3 mmol), methyl acrylate (0.45 mmol), [RhCp\*Cl<sub>2</sub>]<sub>2</sub> (3-5 mol %), oxidant (2.1 equiv), acetone (3 mL), 100 °C, 24 h, sealed tube under argon. <sup>[b]</sup> isolated yield. <sup>[c]</sup> 4.5 equiv of AgOAc was used. <sup>[d]</sup> 36 h.

#### 4. Preparation and Characterization of the Products

#### General procedure for the synthesis of 3 and 4

Sulfonamide (1a) (0.3 mmol),  $[RhCp*Cl_2]_2$  (4 mol%), acrylate (2) (0.45 mmol), and AgOAc (0.63 mmol for 3; 1.35 mmol for 4) were weighed into a 25 mL pressure tube, to which was added acetone (4 mL) in a glove box. The reaction vessel was heated in a preheated oil bath at 100 °C for 24 h (1 h for KIE studies) with efficient stirring. After cooled to room temperature, the mixture was concentrated under vacuum and the residue was purified by column chromatography on silica gel with a gradient eluent of petroleum ether and ethyl acetate to afford the product. Products 3 decompose in CDCl<sub>3</sub> so acetone- $d_6$  was used as an NMR solvent in most cases.

#### General procedure for the synthesis of 5a, 5b and 6

To a mixture of a butadiene product (0.3 mmol) and  $Pd(OAc)_2$  (0.015 mmol) in toluene (3.0 mL) was added pyridine (0.03 mmol). The solution was stirred under an oxygen atmosphere at 80 °C for 12 h. After cooled to room temperature, the mixture was filtered and was purified by column chromatography on silica gel with a gradient eluent of petroleum ether and ethyl acetate to give the product.

5. Characterization Data



<sup>1</sup>**H** NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.76 (d, J = 8.2 Hz, 2H), 7.44 – 7.30 (m, 3H), 6.15 (t, J = 11.3 Hz, 1H), 5.91 (d, J = 15.2 Hz, 1H), 5.72 (dt, J = 10.8, 7.2 Hz, 1H), 4.72 (t, J = 5.9 Hz, 1H), 3.83 (t, J = 6.6 Hz, 2H), 3.75 (s, 3H), 2.45 (s, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  167.1, 143.8, 137.6, 136.6, 134.0, 129.9, 129.0, 127.2, 123.5, 51.7, 40.4, 21.5. HRMS (ESI): Calcd for [C<sub>14</sub>H<sub>17</sub>NO<sub>4</sub>S+Na]<sup>+</sup> 318.0776; Found: 318.0774.



<sup>1</sup>**H** NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.75 (d, J = 8.3 Hz, 2H), 7.33 (d, J = 8.0 Hz, 2H), 6.14 (t, J = 11.2 Hz, 1H), 5.84 (d, J = 15.2 Hz, 1H), 5.67 (dt, J = 10.8, 7.0 Hz, 1H), 4.41 (t, J = 6.1 Hz, 1H), 3.82 (t, J = 5.9 Hz, 1H), 2.44 (s, 3H), 1.48 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  166.0, 143.5, 136.5, 133.3, 129.7, 128.3, 127.1, 125.6, 80.6, 40.3, 29.6, 27.9, 21.4. HRMS (ESI): Calcd for [C<sub>17</sub>H<sub>23</sub>NO<sub>4</sub>S+Na]<sup>+</sup> 360.1245; Found: 360.1249.



<sup>1</sup>**H** NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.75 (d, J = 8.3 Hz, 2H), 7.37 (m, 5H), 7.31 (d, J = 8.2 Hz, 2H), 6.16 (t, J = 11.4 Hz, 1H), 5.96 (d, J = 15.2 Hz, 1H), 5.79 – 5.62 (m, 1H), 5.19 (s, 2H), 4.47 (t, J = 6.0 Hz, 1H), 3.83 (t, J = 6.6 Hz, 2H), 3.72 (t, J = 6.0 Hz, 1H), 2.44 (s, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  166.4, 143.7, 137.9, 135.8, 134.3, 129.8, 128.5, 128.2, 128.2, 127.1, 123.4, 66.3, 40.4, 29.6, 21.5. HRMS (ESI): Calcd for  $[C_{20}H_{21}NO_4S+Na]^+$  394.1089; Found: 394.1092.



<sup>1</sup>**H** NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.76 (d, J = 8.0 Hz, 2H), 7.35 (m, 3H), 6.15 (t, J = 11.3 Hz, 1H), 5.90 (d, J = 15.2 Hz, 1H), 5.71 (dt, J = 10.7, 7.2 Hz, 1H), 4.74 (t, J = 6.0 Hz, 1H), 4.20 (q, J = 7.1 Hz, 2H), 3.83 (t, J = 6.7 Hz, 2H), 2.44 (s, 3H), 1.29 (t, J = 7.1 Hz, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  166.7, 143.8, 137.3, 136.5, 133.8, 129.9, 129.0, 127.1, 123.9, 60.6, 40.4, 21.5, 14.2. HRMS (ESI): Calcd for [C<sub>15</sub>H<sub>19</sub>NO<sub>4</sub>S+Na]<sup>+</sup> 332.0932; Found: 332.0939.



<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 7.75 (d, J = 8.3 Hz, 2H), 7.40 – 7.29 (m, 3H), 6.03 (d, J = 11.8 Hz, 1H), 5.80 (d, J = 15.1 Hz, 1H), 4.71 (t, J = 6.2 Hz, 1H), 3.75 (d, J = 6.3 Hz, 2H), 3.73 (s, 3H), 2.44 (s, 3H), 1.86 (s, 3H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 167.4, 143.7, 142.6, 138.5, 136.7, 129.8, 127.1, 127.0, 121.2, 51.6, 43.5, 22.3, 21.5. HRMS (ESI): Calcd for  $[C_{15}H_{19}NO_4S+Na]^+$  332.0932; Found: 332.0940.



<sup>1</sup>**H** NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.75 (d, J = 8.3 Hz, 2H), 7.33 (d, J = 8.1 Hz, 2H), 6.02 (d, J = 11.8 Hz, 1H), 5.74 (d, J = 15.0 Hz, 1H), 4.40 (t, J = 6.3 Hz, 1H), 3.74 (d, J = 6.4 Hz, 2H), 2.44 (s, 3H), 1.87 (s, 3H), 1.47 (s, 9H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  166.4, 143.7, 141.6, 137.2, 136.7, 129.9, 127.2, 127.1, 123.7, 80.5, 43.6, 28.1, 22.3, 21.5. HRMS (ESI): Calcd for [C<sub>18</sub>H<sub>25</sub>NO<sub>4</sub>S+Na]<sup>+</sup> 374.1402; Found: 374.1406.



<sup>1</sup>**H NMR** (**400 MHz**, **CDCl**<sub>3</sub>) δ 7.72 (d, J = 7.4 Hz, 2H), 7.49 – 7.21 (m, 8H), 6.00 (d, J = 11.7 Hz, 1H), 5.82 (d, J = 15.1 Hz, 1H), 5.31 (s, 1H), 5.15 (s, 2H), 3.71 (d, J = 3.9 Hz, 2H), 2.39 (s, 3H), 1.85 (s, 3H). <sup>13</sup>**C NMR** (**101 MHz**, **CDCl**<sub>3</sub>) δ 166.9, 143.4, 143.2, 139.1, 136.6, 135.9, 129.7, 128.4, 128.1, 128.0, 127.0, 126.7, 120.8, 66.1, 43.3, 22.2, 21.4. **HRMS** (**ESI**): Calcd for  $[C_{21}H_{23}NO_4S+Na]^+$  408.1245; Found: 408.1241.



<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 7.75 (d, J = 8.3 Hz, 2H), 7.45 – 7.29 (m, 3H), 6.03 (d, J = 11.7 Hz, 1H), 5.80 (d, J = 15.1 Hz, 1H), 4.71 (t, J = 6.3 Hz, 1H), 4.18 (q, J = 7.1 Hz, 2H), 3.75 (d, J = 6.3 Hz, 2H), 2.44 (s, 3H), 1.87 (s, 3H), 1.28 (t, J = 7.1 Hz, 3H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 167.0, 143.7, 142.4, 138.3, 136.7, 129.8, 127.13, 127.05, 121.7, 60.4, 43.5, 22.3, 21.5, 14.2. HRMS (ESI): Calcd for  $[C_{16}H_{21}NO_4S+Na]^+$  346.1089; Found: 346.1084.



3ae yield:70%

<sup>1</sup>**H NMR** (**500 MHz, CDCl**<sub>3</sub>) **major:** δ 7.73 (d, J = 8.3 Hz, 2H), 7.31 (d, J = 8.0 Hz, 2H), 7.03 (ddd, J = 11.9, 10.9, 1.1 Hz, 1H), 6.52 – 6.42 (m, 1H), 5.80 – 5.71 (m, 1H), 5.30 (d, J = 10.9 Hz, 1H), 5.17 (t, J = 6.0 Hz, 1H), 3.85 – 3.77 (m, 2H), 2.44 (s, 3H). **minor:** δ 7.80 (d, J = 8.3 Hz, 2H), 7.33 (d, J = 8.0 Hz, 2H), 7.12 – 7.04 (m, 1H), 6.14 – 6.05 (m, 1H), 5.84 – 5.76 (m, 1H), 5.36 (d, J = 15.9 Hz, 1H), 5.14 (t, J = 6.0 Hz, 1H), 3.77 – 3.72 (m, 2H), 2.43 (s, 3H). <sup>13</sup>C **NMR** (**126 MHz, CDCl**<sub>3</sub>) **major:** δ 143.87, 142.39, 136.69, 134.92, 129.80, 127.08, 126.97, 115.66, 99.80, 40.23, 21.47; **minor:** δ 143.66, 134.98, 129.87, 129.64, 128.33, 127.11, 126.32, 109.92, 101.28, 40.23, 21.47. **HRMS** (**ESI**): Calcd for  $[C_{13}H_{14}N_2O_2S+Na]^+$  285.0674; Found: 285.0683.



3be yield:67%

<sup>1</sup>**H** NMR (500 MHz, CDCl<sub>3</sub>) major: δ 7.72 (d, J = 8.3 Hz, 2H), 7.31 (d, J = 7.9 Hz, 2H), 7.07 – 6.96 (m, 1H), 6.34 (d, J = 11.6 Hz, 1H), 5.31 (t, J = 6.2 Hz, 1H), 5.15 (d, J = 10.9 Hz, 1H), 3.72 (d, J = 6.2 Hz, 2H), 2.44 (s, 3H), 1.82 (s, 3H); minor: δ 7.74 (d, J = 8.5 Hz, 2H), 7.33 (d, J = 7.9 Hz, 2H), 7.07 – 6.97 (m, 1H), 5.97 (d, J = 11.7 Hz, 1H), δ 5.24 (t, J = 6.2 Hz, 1H), 5.23 (d, J = 15.7 Hz, 1H), 3.67 (d, J = 6.1 Hz, 2H), 2.45 (s, 3H), 1.86 (s, 3H).<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) major: δ 144.0, 143.8, 143.3, 136.7, 129.7, 127.0, 125.2, 116.2, 96.9, 43.4, 22.4, 21.5; minor: δ 144.5, 143.97, 143.92, 136.4, 129.8, 127.1, 126.4, 118.1, 98.6, 43.3, 22.4, 21.5. HRMS (ESI): Calcd for [C<sub>14</sub>H<sub>16</sub>N<sub>2</sub>O<sub>2</sub>S+Na]<sup>+</sup> 299.0830; Found: 299.0838.



<sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.89 – 7.85 (m, 2H), 7.59 (ddd, J = 6.4, 3.7, 1.2 Hz, 1H), 7.53 (ddd, J = 8.3, 2.2, 0.9 Hz, 2H), 7.32 – 7.23 (m, 1H), 6.12 (t, J = 11.3 Hz, 1H), 5.83 (d, J = 15.2 Hz, 1H), 5.67 (dt, J = 10.8, 7.2 Hz, 1H), 4.89 (s, 1H), 3.84 (t, J = 6.6 Hz, 2H), 1.47 (s, 9H). <sup>13</sup>**C NMR** (126 MHz, CDCl<sub>3</sub>)  $\delta$  165.9, 139.8, 136.3, 133.0, 132.8, 129.22, 129.17, 127.1, 126.0, 80.7, 40. 5, 28.1. HRMS (ESI): Calcd for [C<sub>16</sub>H<sub>21</sub>NO<sub>4</sub>S+Na]<sup>+</sup> 360.1245; Found: 360.1240.



tBu3db yield:52%

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.38 (d, J = 7.9 Hz, 2H), 8.06 (d, J = 8.1 Hz, 2H), 7.32 – 7.21 (m, 1H), 6.17 (t, J = 11.3 Hz, 1H), 5.86 (d, J = 15.2 Hz, 1H), 5.65 (dd, J = 16.2, 7.7 Hz, 1H), 4.90 (s, 1H), 3.91 (s, 2H), 1.47 (s, 9H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 165.9, 150.3, 145.8, 135.8, 131.8, 129.9, 128.4, 126.6, 124.5, 81.0, 40.4, 28.1. HRMS (ESI): Calcd for [C<sub>16</sub>H<sub>20</sub>N<sub>2</sub>O<sub>6</sub>S+Na]<sup>+</sup> 405.1096; Found: 405.1102.



t<sup>Bu</sup> 3eb yield:66%

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.81 (d, J = 8.5 Hz, 2H), 7.50 (d, J = 8.5 Hz, 2H), 7.32 – 7.23 (m, 1H), 6.15 (t, J = 11.4 Hz, 1H), 5.85 (d, J = 15.2 Hz, 1H), 5.65 (dt, J = 10.9, 7.1 Hz, 1H), 4.73 (t, J = 5.4 Hz, 1H), 3.85 (t, J = 6.6 Hz, 2H), 1.48 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 136.1, 132.5, 129.54, 129.47, 128.6, 126.2, 124.4, 122.8, 80.8, 40.4, 28.1, 27.4. HRMS (ESI): Calcd for  $[C_{16}H_{20}CINO_4S+Na]^+$  394.0856; Found: 394.0861.



<sup>1</sup>**H** NMR (400 MHz, CDCl<sub>3</sub>) δ 8.01 (d, J = 7.9 Hz, 2H), 7.80 (d, J = 7.9 Hz, 2H), 7.28 (m, 1H), 6.16 (t, J = 11.3 Hz, 1H), 5.86 (d, J = 15.3 Hz, 1H), 5.66 (dd, J = 17.1, 7.6 Hz, 1H), 4.94 (s, 1H), 3.88 (t, J = 6.5 Hz, 2H), 1.47 (s, 9H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 165.9, 143.6, 136.0, 134.6 (q,  $J_{C-F} = 33.1$  Hz), 132.3, 129.6, 127.7, 126.4 (q,  $J_{C-F} = 3.6$  Hz), 126.3, 123.2 (q,  $J_{C-F} = 272.8$  Hz, CF<sub>3</sub>), 80.9, 40.4, 28.1. HRMS (ESI): Calcd for [C<sub>17</sub>H<sub>20</sub>F<sub>3</sub>NO<sub>4</sub>S+Na]<sup>+</sup> 428.1119; Found: 428.1121.



tBu yield:73%

<sup>1</sup>**H NMR** (400 **MHz**, **CDCl**<sub>3</sub>) δ 7.67 (m, 2H), 7.45 – 7.35 (m, 2H), 7.27 (m, 1H), 6.13 (t, J = 11.3 Hz, 1H), 5.84 (d, J = 15.2 Hz, 1H), 5.68 (dt, J = 11.1, 7.2 Hz, 1H), 4.71 (s, 1H), 3.83 (t, J = 6.7 Hz, 2H), 2.43 (s, 3H), 1.48 (s, 9H). <sup>13</sup>**C NMR** (101 MHz, CDCl<sub>3</sub>) δ 165.9, 139.5, 136.3, 133. 7, 133.1, 129.12, 129.09, 127.5, 125.9, 124.2, 80.7, 40.5, 29.3, 28.1, 21.3. **HRMS** (ESI): Calcd for  $[C_{17}H_{23}NO_4S+Na]^+$  374.1402; Found: 374.1405.



<sup>1</sup>**H NMR** (**400 MHz**, **MeOD**) δ 7.72 (d, J = 8.3 Hz, 2H), 7.38 (d, J = 8.1 Hz, 2H), 7.32 (dd, J = 3.1, 1.5 Hz, 1H), 6.25 (t, J = 3.3 Hz, 1H), 6.06 (s, 1H), 3.84 (s, 2H), 2.96 (s, 3H), 2.91 (s, 3H), 2.40 (s, 3H). <sup>13</sup>**C NMR** (**101 MHz**, **MeOD**) δ 171.9, 146.9, 137.6, 131.3, 129.7, 128.2, 124.1, 116.0, 112.9, 38.1, 36.1, 34.0, 21.7. **HRMS** (**ESI**): Calcd for  $[C_{15}H_{18}N_2O_3S+Na]^+$  329.0936; Found: 329.0942.

Ph-N 
$$H$$
  
H  $Me$   
O Me 4c vield:55%

<sup>1</sup>**H NMR** (400 MHz, MeOD) δ 7.85 (d, J = 7.6 Hz, 2H), 7.67 (t, J = 7.4 Hz, 1H), 7.57 (t, J = 7.7 Hz, 2H), 7.34 (dd, J = 3.2, 1.6 Hz, 1H), 6.27 (t, J = 3.4 Hz, 1H), 6.08 (s, 1H), 3.85 (s, 2H), 2.97 (s, 3H), 2.91 (s, 3H). <sup>13</sup>**C NMR** (101 MHz, MeOD) δ 171.9, 140.6, 135.4, 130.8, 129.9, 128.1, 124.2, 116.2, 113.1, 49.2, 38.1, 36.1, 33.9. **HRMS** (ESI): Calcd for [C<sub>14</sub>H<sub>16</sub>N<sub>2</sub>O+Na]<sup>+</sup> 251.1160; Found: 251.1167.



4e vield:58%

<sup>1</sup>**H NMR** (**400 MHz**, **CDCl**<sub>3</sub>)  $\delta$  7.77 – 7.71 (m, 2H), 7.50 – 7.43 (m, 2H), 7.22 (dd, *J* = 3.3, 1.7 Hz, 1H), 6.25 (t, *J* = 3.4 Hz, 1H), 6.10 (s, 1H), 3.82 (s, 2H), 2.97 (s, 3H), 2.93 (s, 3H). <sup>13</sup>**C NMR** (**101 MHz**, **CDCl**<sub>3</sub>)  $\delta$  168.9, 140.4, 137.3, 129.6, 128.6, 128.3, 122.5, 114.7, 112.2, 37.5, 35.5, 32.9. **HRMS** (**ESI**): Calcd for [C<sub>14</sub>H<sub>15</sub>ClN<sub>2</sub>O<sub>3</sub>S+H]<sup>+</sup> 349.0390; Found: 349.0384.

vield:69%

<sup>1</sup>**H NMR** (**400 MHz**, **CDCl**<sub>3</sub>) δ 7.75 (d, J = 6.0 Hz, 2H), 7.46 (s, 1H), 7.33 (d, J = 7.0 Hz, 2H), 6.45 (s, 1H), 5.99 (s, 1H), 4.53 (s, 2H), 4.14 (q, J = 6.9 Hz, 2H), 2.43 (s, 3H), 1.28 (t, J = 6.9 Hz, 3H). <sup>13</sup>**C NMR** (**101 MHz**, **CDCl**<sub>3</sub>) δ 167.4, 155.4, 144.9, 134.9, 134.4, 130.0, 127.6, 127.3, 93.3, 59.7, 57.5, 21.6, 14.4. **HRMS** (**ESI**): Calcd for  $[C_{15}H_{17}NO_4S+Na]^+$  330.0776; Found: 330.0779.

<sup>1</sup>**H NMR** (**400 MHz**, **CDCl**<sub>3</sub>) δ 7.75 (d, J = 8.3 Hz, 2H), 7.32 (d, J = 8.1 Hz, 2H), 7.15 (s, 1H), 5.91 (s, 1H), 4.38 (s, 2H), 4.13 (q, J = 7.1 Hz, 2H), 2.42 (s, 3H), 1.95 (s, 3H), 1.27 (t, J = 7.1 Hz, 3H). <sup>13</sup>**C NMR** (**101 MHz**, **CDCl**<sub>3</sub>) δ 167.6, 156.5, 147.5, 144.7, 134.4, 129.9, 127.2, 122.9, 91.4, 59.8, 59.4, 21.5, 14.5, 14.3. **HRMS** (**ESI**): Calcd for  $[C_{16}H_{19}NO_4S+Na]^+$  344.0932; Found: 344.0930.

CN 6 yield:58%

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.68 (d, J = 8.3 Hz, 2H), 7.32 (d, J = 8.1 Hz, 2H), 7.02 (s, 1H), 6.22 (s, 1H), 3.88 (s, 2H), 2.42 (s, 3H), 2.02 (s, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  145.4, 135.7, 130.2, 126.7, 122.7, 122.4, 120.7, 117.6, 116.2, 21.6, 17.3, 11.6. HRMS (ESI): Calcd for [C<sub>14</sub>H<sub>14</sub>N<sub>2</sub>O<sub>2</sub>S+Na]<sup>+</sup> 297.0674; Found: 297.0665.

#### 6. Deuterium-Labeling Experiment

*N*-Propargyl-*p*-tolylsulfonamide (105 mg, 0.5 mmol) was added in one portion to a stirred suspension of LiAlD<sub>4</sub> (25 mg, 0.65 mmol) in THF (2 mL) at -15 °C. The reaction mixture was stirred at -15 °C and was allowed to warm to room temperature (14 h). The reaction mixture was cooled to -10 °C, and was carefully quenched by slow addition of D<sub>2</sub>O (50  $\mu$ L). The mixture was stirred for another hour, filtered, and dried over MgSO<sub>4</sub>. The solvent was evaporated under reduced pressure, and the resulting residue was purified by column chromatography on silica gel to provide the desired product **1a**- $d_2$  (yield: 45%). The degree of deuteration is > 98% at both two positions.

#### 6.2 Kinetic Isotope Effect



A mixture of **1a**- $d_2$  (> 98% deuteration, 0.3 mmol), 1a(0.3mmol), ethyl acrylate (0.24 mmol), [RhCp\*Cl<sub>2</sub>]<sub>2</sub> (4 mol%), and AgOAc (0.63 mmol) were weighed into a pressure tube. Acetone (4 mL) was added. The mixture was then stirred at 100°C for 1 h. After cooled to room temperature, the mixture was filtered and the crude product was purified by silica gel column chromatography. <sup>1</sup>H NMR analysis (400 MHz, CDCl<sub>3</sub>) showed that a value of  $k_{\rm H}/k_{\rm D} = 4.2$  was obtained.





### 7. NMR Spectra of Products















2.00H

8.5 8.0

9.0

3.08H

7.0

7.5

1.00H 1.00H 1.01H

6.0

5.5

6.5



2.00H

4.0 f1 (ppm)

2.01H

3.5

0.98H

5.0

4.5



3.0

3. 05≖

2.5

2.0

3. 08⊣

1.0

0.5

0.0 -0.5 -1.(

1.5













































































