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An Unusual Glycosylation Product from a Partially Protected Fucosyl Donor

under Silver Triflate activation conditions

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Supporting information

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Experimental data - General Information

Unless otherwise stated; specific rotation was recorded in a Rudolph research autopol IV polarimeter, infrared spectra (IR) were recorded on a Perkin Elmer spectrometer. A Brüker Avance 400, and an Agilent 400 spectrometer were employed for ¹H (400.13 MHz) and ¹³C (100.61 MHz) NMR spectra, a ³ Brüker Ultrashield 600 spectrometer was employed for ¹H (600.13 MHz) and ¹³C (150.90 MHz) NMR spectra. Resonances δ , are in ppm units downfield from an internal reference in CDCl₃ (δ_{H} = 7.26 ppm, δ_{C} = 77.0 ppm), MeOH (δ_{H} = 3.31 ppm, δ_{C} = 49.0 ppm). For oligosaccharides the notation a, b, c.... refers to the monosaccharide from the reducing end. Mass spectrometry analysis was performed with a Q-Tof Premier Waters Maldi-quadrupole time-of-flight (Q-Tof) mass spectrometer equipped ¹⁰ with Z-spray electrospray ionization (ESI). X-ray crystallography was performed with a Brüker SMART APEX diffractometer. Silica gel Florisil (200 mesh; Aldrich) was used for column chromatography. Analytical thin-layer chromatography was performed using Merck 60 F₂₅₄ silica gel (pre-coated sheets, 0.2 mm thick, 20 cm x 20 cm) and visualised by UV irradiation or molybdenum staining. DCM, MeOH, THF and toluene were dried over flame dried 3 Å or 4 Å sieves. Triethylamine (Et₃N) and ¹⁵ trifluoroacetic acid (TFA) were used dry from sure/seal bottles. Other reagents were purchased from an industrial supplier; Sigma Aldrich, VWR, Carbosynth.

For experimental procedures and characterisation data for compounds **1**,**2**, **6** and **8** see references 19 and 35.

²⁰ 3-(2,4-Di-O-*tert*-butyldimethylsilyl-α-L-fucopyranosyl)-2,4-di-O-*tert*-butyl-dimethylsilyl-1-O-benzylα-L-fucopyranose, 3a

and,

3-(2,4-di-O-*tert*-butyldimethylsilyl- α -L-fucopyranosyl)-2,4-di-O-*tert*-butyl-dimethylsilyl-1-O-benzyl- β -L-fucopyranose, 3b

²⁵ Thioglycoside donor **1** (120 mg, 0.28 mmol), was dried under vacuum for 12 h, before dissolving in DCM (2 mL) containing pre-activated 3 Å ms under N₂. The mixture was cooled to 0 °C and Br₂ (104 μ L, 2.80 mmol) added. The mixture was stirred under N₂ for 5 min at 0 °C after which time the bromine was removed *in vacuo*, co-evaporating with DCM (1 mL) and toluene (1 mL). The mixture was re-dissolved in anhydrous DCM (2 mL), and BnOH (29 μ L, 0.28 mmol) was added. The ³⁰ temperature was lowered to -20 °C. AgOTf (141 mg, 0.55 mmol) in dry THF (1 mL) was cooled to -20 °C, and added. The mixture was stirred in the dark for 25 min before quenching with Et₃N (0.5 mL), and stirring for 20 min. The mixture was diluted with DCM (10 mL) and filtered through a plug of celite. The organic layer was washed with saturated aqueous NaHCO₃ solution (5 mL), deionised H₂O

(3 mL) then dried over MgSO₄. The mixture was filtered and the solvent removed *in vacuo*. The mixture was purified by column chromatography (Et₂O:Hex, 1:19 v:v) to yield the disaccharide products **3a** (28 mg, 24 % yield), and **3b** (23 mg, 20 % yield) as clear oils;

s $\alpha\alpha$ anomer characterised, **3a**

 $[\alpha]_{D}^{20} = 64^{\circ} (\text{deg cm}^{3} \text{g}^{-1} \text{dm}^{-1}) (\text{c} = 0.1 \text{ in CH}_{3}\text{Cl});$

v_{max} (thin film) 3590 cm⁻¹ (OH), 2928 cm⁻¹ (CH);

¹H NMR (600 MHz, CDCl₃) δ 7.38 (2H, d, *J* = 7.2 Hz, o-Ph), 7.33 (2H, t, *J* = 7.2 Hz, m-Ph), 7.28 (1H, t, *J* = 7.2Hz, p-Ph), 5.16 (1H, d, *J*_{1,2} = 3.0 Hz, H-1B), 4.85 (1H, d, *J*_{1,2} = 3.4 Hz, H-1A), 4.66 (1H, d, *J* = 11.8 Hz, OCH<u>H</u>), 4.51 (1H, d, *J* = 11.8 Hz, OC<u>H</u>H), 4.15 (1H, dd, *J*_{2,3} = 10.0 Hz, *J*_{2,1} = 3.3 Hz, H-2A), 4.10 (1H, q, *J*_{5,6} = 6.5 Hz, H-5B), 4.05 (1H, dd, *J*_{3,2} = 10.0 Hz, *J*_{3,4} = 2.0 Hz, H-3A), 3.99 (1H, br s, H-4A), 3.94 (1H, ddd, *J*_{3,2} = 9.3 Hz, *J*_{3,0H} = 5.2 Hz, *J*_{3,4} = 1.7 Hz, H-3B), 3.88 (2H, m, H-5A, H-2B), 3.81 (1H, d, *J*_{4,3} = 1.6 Hz, H-4B), 1.76 (1H, *J* = 5.2 Hz, 3B-OH), 1.15 (3H, d, *J*_{6,5} = 6.4 Hz, H-6A), 1.14 (3H, d, *J*_{6,5} = 6.4 Hz, H-6B), 0.96 (18H, s, SiC(CH₃)₃), 0.91, 0.86 (9H, SiC(CH₃)₃), 0.16, 0.16, 0.13, 0.11, 0.11, 0.11, 0.04, 0.04 ((3H, s, Si(CH₃)₂);

¹³C NMR (150 MHz, CDCl₃) δ 138.0 (Ar C), 128.2 (Ar CH), 128.1 (Ar CH), 127.1 (Ar CH), 100.4 (C-1B), 99.1 (C-1A), 77.7 (C-3A), 74.3 (C-4A), 74.0 (C-4B), 72.0 (C-2B), 70.5 (C-3 B), 70.4 (C-2A), 69.6 (PhCH₂O), 68.1 (C-5B), 68.0 (C-5A), 26.1, 26.1, 26.0, 25.9 (SiC(<u>C</u>H₃)₃), 18.6, 18.5, 18.1, 18.1 (Si<u>C</u>(CH₃)₃), 17.2 (C-6B), 17.0 (C-6A), -3.8, -3.9, -4.0, -4.1, -4.4, -4.7, -4.7, -4.8, -4.8 Si(<u>C</u>H₃);

M/z HRMS (ESI-TOF) calcd. for $C_{43}H_{84}O_9NaSi_4 = 879.5090$, (M+Na)⁺. Found = 879.5068.

 $_{20} \alpha \beta$ anomer characterised, **3b**

 $[\alpha]_{D}^{20} = 92^{\circ} (\text{deg cm}^{3} \text{g}^{-1} \text{dm}^{-1}) (\text{c} = 0.1 \text{ in CH}_{3}\text{Cl});$

v_{max} (thin film) 3599 cm⁻¹ (OH), 2929 cm⁻¹ (CH);

¹H NMR (600 MHz, CDCl₃) δ 7.39 (2H, d, *J* = 7.2 Hz, o-Ph), 7.34 (2H, t, *J* = 7.2 Hz, m-Ph), 7.30 (1H, t, *J* = 7.2 Hz, 5.07 (1H, d, *J*_{1,2} = 2.0 Hz, H-1B), 4.92 (1H, d, *J* = 11.6 Hz, PhC<u>H(H)</u>), 4.52 (1H, d, *J* = 11.6 Hz, PhCH(<u>H</u>)), 4.29 (1H, br s, H-1A), 4.11 (1H, q, *J*_{5,6} = 6.8 Hz, H-5B), 4.08 (1H, br s, H-4A), 3.92 (3H, m, H-3B, 2B, 5A), 3.76 (1H, br s, H-4B), 3.62 (2H, m, H-2A,3A), 1.73 (1H, br s, OH), 1.32 (3H, br s, H-6A), 1.15 (3H, d, *J*_{6,5} = 6.7 Hz, H-6B), 0.97, (18H, s, SiC(CH₃)₃), 0.94, 0.89 (9H, s, Si C(CH₃)₃), 0.15, 0.15, 0.14, 0.13, 0.12, 0.11, 0.11, 0.05 (3H, s, Si(CH₃)₂;

¹³C NMR (150 MHz, CDCl₃) δ 137.9 (Ar C), 128.2 (Ar CH), 128.0 (Ar CH), 127.3 (Ar CH), 102.6 (C-1A),
³⁰ 101.0 (C-1B), 81.5 (C-3A), 74.0-72.5 (br, C-4A), 72.3 (C-2B), 74.0 (C-4B), 72.0 (C-2A), 71.7 (C-5A), 70.4 (C-3B), 70.2 (OCH₂Ph), 68.0 (C-5B), 26.1, 26.1, 26.1, 26.0 (SiC(CH₃)₃), 18.6, 18.5, 18.3, 18.1 (SiC(CH₃)), 17.3 (C-6A), 17.2 (C-6B), - 3.6, - 3.9, - 4.0, - 4.0, - 4.2, - 4.4, - 4.6 (Si(CH₃));

M/z HRMS (ESI-TOF) calcd. for $C_{43}H_{84}O_9NaSi_4 = 879.5090$, (M+Na)⁺. Found = 879.5063.

Reverse Glycosylation conditions for preparation of 3a and 3b (Table 1; entry 12)

Thioglycoside donor **1** (120 mg, 0.28 mmol), was dried under vacuum for 12 h, before dissolving in ⁵ DCM (2 mL) containing pre-activated 3 Å ms under N₂. The mixture was cooled to 0 °C and Br₂ (104 μL, 2.80 mmol) added. The mixture was stirred under N₂ for 5 min at 0 °C before the bromine was removed *in vacuo*, co-evaporating with DCM and toluene. The bromide donor mixture was redissolved in anhydrous DCM (2 mL). BnOH (29 μL, 0.28 mmol) was added to AgOTf (141 mg, 0.55 mmol) in dry THF (1 mL) at -20 °C. The bromide donor mixture was added to the acceptor/activator ¹⁰ mixture dropwise, and stirred in the dark for 25 min. The mixture was quenching with Et₃N (0.5 mL), and stirred for 20 min before being worked up as general procedure to yield the disaccharide products **3a** (16 mg, 14 % yield), and **3b** (14 mg, 12 % yield) as clear oils;

$O-(2,4-Di-O-tert-butyl dimethyl silyl-\alpha-L-fuc opyranosyl)-2,4-di-O-tert-butyl-dimethyl silyl-1-O-tert-butyl dimethyl silyl-1$

15 propargyl-α-L-fucopyranose, 5a

and

$O-(2,4-Di-O-tert-butyl dimethylsilyl-\alpha-L-fucopyranosyl)-2,4-di-O-tert-butyl-dimethylsilyl-1-O-tert-butyl-1-O-tert-butyl-dimethylsilyl-1-O-tert-buty$

propargyl-β-L-fucopyranose 5b

Thioglycoside donor **1** (120 mg, 0.28 mmol), Br₂ (104 μL, 2.80 mmol) and propargyl alcohol **63** (15 μL, 0.28 mmol) were reacted according to procedure described for synthesis of **3a/3b**. The crude material was purified by column chromatography (EtOAc:Hexane, 1:49 (v/v)) to yield products **5a**, as a clear oil, (50 mg, 45 %) and the product **5b**, as a clear oil (9 mg, 8 %);

 $\alpha\alpha$ anomer characterised, 5a

 $[\alpha]_{D}^{20} = -76^{\circ} (\text{deg cm}^{3} \text{g}^{-1} \text{dm}^{-1}) (\text{c} = 0.1 \text{ in CH}_{3}\text{Cl});$

²⁵ ν_{max} (thin film) 3599 cm⁻¹ (OH), 2929 cm⁻¹ (CH);

¹H NMR (600 MHz, CDCl₃) δ 5.13 (1H, d, J_{1,2} = 3.1 Hz, H-1B), 4.97 (1H, d, J_{1,2} = 3.7 Hz, H-1A), 4.24 (2H OCH₂), 4.16 (1H, dd, J_{2,3} = 10.1 Hz, J_{2,1} = 3.5 Hz, H-2A), 4.10 (1H, q, J_{5,6} = 6.6 Hz, H-5B), 4.01 (1H, br s, H-4A), 4.00 (1H, dd, J_{3,2} = 9.9 Hz, J_{3,4} = 2.2 Hz, H-3A), 3.94 (1H, ddd, J_{3,2} = 9.6 Hz, J_{3,0H} = 6.0 Hz, J_{3,4} = 2.9 Hz, H-3B), 3.90 (1H, dd, J_{3,2} = 10.0 Hz, J_{3,4} = 3.1 Hz, H-2B) 3.87 (1H, q, J_{5,6} = 6.6 Hz, H-5A), 3.81 (1H, d, J_{3,4} = 1.9 Hz, H-4B), 2.36 (1H, t, J = 2.4 Hz, C≡<u>CH</u>), 1.76 (1H, d, J = 6.0 Hz, OH), 1.17 (3H, d, J_{6,5} = 6.8 Hz, H-6A), 1.14 (3H, d, J_{6,5} = 6.8 Hz, H-6B), 0.96, 0.95, 0.94, 0.93 (9H, s, SiC(CH₃)₃), 0.16, 0.15, 0.15, 0.15, 0.15, 0.11, 0.11 (3H, s, Si(CH₃)₂);

¹³C NMR (150 MHz, CDCl₃) δ 100.8 (C-1B), 98.0 (C-1A), 79.6 (<u>C</u>=CH), 77.9 (C-3A), 74.3 (C-4A), 74.1 (C=<u>C</u>H), 74.0 (C-4B), 72.1 (C-2B), 70.5 (C-3B), 70.0 (C-2A), 68.3 (C-5A), 68.2 (C-5B), 54.2 (OCH₂), 26.1, 26.1, 26.1, 26.0 (SiC(<u>C</u>H₃)₃), 18.6, 18.6, 18.5, 18.5, 18.3, 18.3, 18.2, 18.2 (Si<u>C</u>(CH₃)₃, 17.3 (C-6B), 16.9 (C-6A), -3.8, -3.9, -3.9, -4.2, -4.4, -4.7, -4.9 (Si(CH₃)₂;

 $_{3}$ **M/z HRMS (ESI-TOF)** calcd. for C₃₉H₈₀O₉NaSi₄ = 827.4777 (M+Na)⁺. Found = 827.4770.

 $\alpha\beta$ anomer characterised, **5b**

 $[\alpha]_{D}^{20} = -57^{\circ} (\text{deg cm}^{3} \text{g}^{-1} \text{dm}^{-1}) (\text{c} = 0.1 \text{ in CH}_{3}\text{Cl});$

v_{max} (thin film) 3587 cm⁻¹ (OH), 2931 cm⁻¹ (CH);

¹H NMR (400 MHz, CDCl₃) δ 5.07 (1H, d, $J_{1,2} = 2.8$ Hz, H-1b), 4.37 (1H, dd, J = 15.5 Hz, J = 2.4 Hz, ¹⁰ OCH(H)) 4.34 (1H, d, $J_{1,2} = 6.5$ Hz, H-1a), 4.31 (1H, dd, J = 15.6 Hz, J = 2.2 Hz, OC<u>H</u>(H)), 4.10 (1H, q, $J_{5,6} = 6.6$ Hz, H-5b), 4.03 (1H, br s, H-4a), 3.97 (1H, ddd, $J_{3,2} = 10.0$ Hz, $J_{3,OH} = 6.5$ Hz, $J_{3,4} = 2.6$ Hz, H-3b), 3.92 (1H, dd, $J_{2,3} = 10.1$ Hz, $J_{2,1} = 3.1$ Hz, H-2b), 3.85 (1H, dd, $J_{2,3} = 8.9$ Hz, $J_{2,1} = 6.4$ Hz, H-2a), 3.81 (1H, br s, H-4b), 3.60 (1H, dd, $J_{3,2} = 8.6$ Hz, $J_{3,4} = 1.2$ Hz, H-3a), 3.57 (1H, m, H-5a), 2.39 (-C=<u>CH</u>), 1.24 (3H, d, $J_{6,5} = C-6a$), 1.15 (3H, d, $J_{6,5} = C-6b$), 1.00, 0.95, 0.95, 0.94 (9H, s, SiC(<u>CH</u>₃)₃), 0.15, 0.15, 0.14, 0.14, ¹⁵ 0.14, 0.14, 0.12, 0.12 (3H, s, Si(CH₃)₂;

¹³C NMR (100 MHz, CDCl₃) δ 101.4 (C-1a), 101.0 (C-1b), 82.1 (C-3a), 78.8 (-C=CH), 74.5 (-C=CH), 74.1 (C-4b), 72.5 (C-4a), 72.1 (C-2b), 71.9 (C-2a), 70.1 (C-3b), 68.0 (C-5b), 55.0 (OCH₂), 26.0, 25.9, 25.9, 25.7 (SiC(CH₃)₃), 18.5, 18.4, 18.4, 18.53, 18.1, 18.1, 18.0, 18.0 (SiC(CH₃)₃), 17.8, 17.8 (C-6b, C-6a) -3.7, -3.7, -3.9, -4.1, -4.2, -4.5, -4.6, -4.7 (Si(CH₃)₂;

 $_{20}$ **M/z HRMS (ESI-TOF)** calcd. for C₃₉H₈₀O₉NaSi₄ = 827.4777 (M+Na)⁺. Found = 827.4736.

$\label{eq:2-Acetamido-2-deoxy-3,4-di-O-acetyl-6-O-(3-O-(2,4-di-O-tert-butyldimethylsilyl-α-L-fucopyranosyl)-2,4-di-O-tert-butyldimethylsilyl-α-L-fucopyranosyl)-1-O-propargyl-β-D-glucopyranose, 7$

²⁵ Thioglycoside donor **1** (128 mg, 0.30 mmol), Br₂ (104 μL, 2.80 mmol) and acceptor **6** (100 mg, 0.30 mmol) reacted according to procedure described for synthesis of **3a/3b**. The crude material was purified by column chromatography (EtOAc:Hex, 1:1 (v/v)) to yield the product as a clear oil (13 mg, 8 %);

 $[\alpha]_{D}^{20} = -83^{\circ} (\text{deg cm}^{3} \text{g}^{-1} \text{dm}^{-1}) (\text{c} = 0.1 \text{ in CH}_{3}\text{Cl});$

³⁰ ν_{max} (thin film) 3530 cm⁻¹ (OH), 3276 cm⁻¹ (C=CH), 3035 cm⁻¹ (NH), 2929 cm⁻¹ (CH), 1753 cm⁻¹ C=O, 1663 cm⁻¹ (NHC=O); 15

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¹H NMR (600 MHz, CDCl₃) δ 5.39 (1H, d, *J* = 9.1 Hz, NH), 5.24 (1H, app t, *J*_{3,4} = *J*_{3,2} = 9.8 Hz, H-3A), 5.10 (1H, d, *J*_{1,2} = 2.3 Hz, H-1C), 5.00 (1H, app t *J*_{4,3} = *J*_{4,5} = 9.6 Hz, H-4A), 4.76 (1H, d, *J*_{1,2} = 8.4 Hz, H-1A), 4.74 (1H, d, *J*_{1,2} = 3.0 Hz, H-1B), 4.37 (2H, m, <u>CH</u>₂C≡CH), 4.08 (1H, dd, *J*_{2,3} = 9.6 Hz, *J*_{2,1} = 2.7 Hz, H-2B), 4.06 (2H, m, H-5C, H-4B), 4.00 (1H, app q, *J*_{2,NH} = *J*_{2,1} = *J*_{2,3} =8.9 Hz, H-2A), 3.93 (4H, m, H-3B, H-3C, Hs 2C, H-5B), 3.83 (1H, br s, H-4C), 3.75 (1H, br d, *J*_{6',6} = 11.6 Hz, H-6'A), 3.73 (1H, m, H-5A), 3.60 (1H, dd, *J*_{6,6'} = 11.5 Hz, *J*_{6,5} = 5.8 Hz, H-6A), 2.46 (1H, s, C≡<u>CH</u>), 2.05, 2.03, 1.98 (3H, s, COCH₃), 1.79 (1H, d, *J* = 4.5Hz, 3C-OH), 1.19 (3H, d, *J*_{6,5} = 6.4 Hz, H-6B), 1.15 (3H, d, *J*_{6,5} = 6.4 Hz, H-6C), 0.97, 0.97, 0.95, 0.92 (9H, s, SiC(CH₃)₃), 0.20, 0.17, 0.16, 0.15, 0.13, 0.11, 0.11, 0.10 (3H, s, SiCH₃);

¹³C NMR (150 MHz, CDCl₃) δ 171.0, 169.9, 168.9 (C=O), 100.6 (C-1C), 99.0 (C-1B), 98.2 (C-1A), 78.7
¹⁰ (HC=<u>C</u>), 77.6 (C-3B), 74.9 (C=<u>CH</u>), 73.8 (C-4C), 73.4 (C-5A), 73.3 (C-4B), 72.7 (C-3A), 71.6 (C-3C), 70.3 (C-2C), 70.1 (C-2B), 68.9 (C-4A), 68.5 (C-5B), 68.0 (C-5C), 66.8 (C-6A), 55.4 (CH₂C=CH), 54.0 (C-2A), 25.9, 25.9, 25.9, 25.8 (SiC(CH₃)₃), 23.2 (NHCO<u>C</u>H₃), 20.5, 20.5 (COCH₃), 18.4, 18.3, 18.2, 18.0 (Si<u>C</u>(CH₃)₃), 17.1 (C-6C), 16.6 (C-6B), -3.6, -4.0, -4.0, -4.4, -4.6, -4.9, -4.9, -4.9 (Si<u>C</u>H₃);

M/z HRMS (ESI-TOF) calcd. for $C_{51}H_{96}NO_{16}Si_4 = 1090.5806$, (M-H)⁻. Found = 1090.5806.

Supplemental – S3 NMR data

$3-(2,4-Di-O-tert-butyldimethylsilyl-\alpha-L-fucopyranosyl)-2,4-di-O-tert-butyl-dimethylsilyl-1-O-benzyl-\alpha-L-fucopyranose, 3a$











3-(2,4-di-O-*tert*-butyldimethylsilyl-α-L-fucopyranosyl)-2,4-di-O-*tert*-butyl-dimethylsilyl-1-O-benzylβ-L-fucopyranose, 3b

¹H-NMR



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¹H-¹³C HSQC



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 $O-(2,4-Di-O-tert-butyldimethylsilyl-\alpha-L-fucopyranosyl)-2,4-di-O-tert-butyl-dimethylsilyl-1-O-propargyl-\alpha-L-fucopyranose, 5a$

⁵¹H-NMR





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s O-(2,4-Di-O-*tert*-butyldimethylsilyl-α-L-fucopyranosyl)-2,4-di-O-*tert*-butyl-dimethylsilyl-1-Opropargyl-β-L-fucopyranose 5b

¹H-NMR



¹H-¹³C HSQC



¹⁰ 2-Acetamido-2-deoxy-3,4-di-O-acetyl-6-O-(3-O-(2,4-di-O-*tert*-butyldimethylsilyl-α-L-fucopyranosyl)-2,4-di-O-*tert*-butyldimethylsilyl-α-L-fucopyranosyl)-1-O-propargyl-β-D-glucopyranose, 7







