## **Supplementary Information**

# Comparative analysis of cross strand aromatic-Phe interactions in designed peptide β-hairpins

Kamlesh Madhusudan Makwana and Radhakrishnan Mahalakshmi\*

Molecular Biophysics Laboratory, Department of Biological Sciences, Indian Institute of Science Education and Research, Bhopal 462023. India.



#### Supplementary Experimental Section

**Mass spectra and HPLC profiles:** Mass spectra were recorded on a micrOTOF-Q II mass spectrometer (Bruker Daltonik GmbH). Calculated molecular weights and observed peptide adducts are annotated. Analytical HPLC profiles of purified peptides were obtained on a Shimadzu UFLC system on a C<sub>18</sub> reverse phase column (5µm particle size) using methanol-water gradients. The HPLC profile is provided as an inset and peak retention times (RT) of the desired peptide (in minutes) are also indicated. A water-methanol gradient of 70-100% methanol in 30 minutes and 100-100% methanol in next 10 minutes was used in each case.

			<u>Sup</u>	plementary	<u>Figures</u>		
		<u>4β</u> 2,3, 2, 2, 2, 2	8β 3β/4γ 1. 2.0. 1.9. 1	6β 6γ 1β 	6δ 3γ 1.4 1.3 1.2 11 10 09 08	8γ	
	F2	5.2 5.0	V3	P4 V8 G5 P4 ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	7β Μ β ppm	
	V3     F7     F2 v8     L6     L1       9.0     8.5     8.0     7.5     7.0     6.5     ppm						
		9 8	lulu		4 3 2	))( 1 ppm	
Residue	NH	CαH	$C^{\beta}H$	СүН	Others	${}^{3}J_{\rm NH-C}{}^{\alpha}{}_{\rm H}$	dδ/dT
	ppm	ppm	ppm	ppm	ppm	Hz	ppb/K
Leu1	7.93	4.48	1.48	1.47	C <sup>8</sup> H: 0.89	8.34	-3.7
Phe2	8.12	5.25	3.15 2.79	-	C <sup>8</sup> H: 7.17 C⁵H: 7.23 C³H: 7.11	9.18	-8.0
Val3	8.94	4.50	2.04	0.925	-	9.45	-4.8
<sup>D</sup> Pro4	-	4.35	2.22	1.99	С <sup>8</sup> Н: 3.75	-	-
Gly5	8.31	4.04 3.73	-	-	-	-	-10.2
Leu6	8.15	4.67	1.78	1.63	C <sup>8</sup> H: 0.92	8.84	-2.2
Phe7	8.66	4.54	2.85 2.74	-	C <sup>8</sup> H: 6.67 C⁵H: 7.03 C <sup>ζ</sup> H: 7.04	7.79	-9.6
Val8	8.20	4.10	1.98	0.82 0.73	-	9.59	-3.6

**Figure S1a:** <sup>1</sup>H 1D spectrum (top) and NMR parameters (table) of peptide **FF** in CD<sub>3</sub>OH at 303K.

## Electronic Supplementary Material (ESI) for Organic & Biomolecular Chemistry This journal is The Royal Society of Chemistry 2014



Residue	NH	CαH	$C^{\beta}H$	СүН	Others	${}^{3}J_{\rm NH-C}{}^{\alpha}{}_{\rm H}$	dδ/dT
	ppm	ppm	ppm	ppm	ppm	Hz	ppb/K
Leu1	7.99	4.49	1.58	1.48	C <sup>8</sup> H: 0.87	8.17	-3.2
Tyr2	8.25	5.14	3.03 2.70	-	C <sup>8</sup> H: 6.93 C <sup>€</sup> H: 6.65	9.19	-7.3
Val3	8.92	4.48	2.03	0.90	-	9.58	-4.3
<sup>D</sup> Pro4	-	4.32	2.24 2.14	1.95	C <sup>8</sup> H: 3.72	-	-
Gly5	8.00	4.00 3.81	-	-	-	-	-11.7
Leu6	8.06	4.67	1.76	1.61	C <sup>8</sup> H: 0.92	8.90	-3.8
Phe7	8.69	4.60	2.85	-	C <sup>8</sup> H: 6.79 Cध्H: 7.07 टरॅंH: 7.08	7.88	-9.4
Val8	8.29	4.14	2.01	0.84 0.74	-	9.63	-1.6

**Figure S1b:** <sup>1</sup>H 1D spectrum (top) and NMR parameters (table) of peptide **YF** in CD<sub>3</sub>OH at 303K.



Residue	NH	CαH	$C^{\beta}H$	СүН	Others	${}^{3}J_{\rm NH-C}{}^{\alpha}{}_{\rm H}$	dδ/dT
	ppm	Ppm	ppm	ppm	ppm	Hz	ppb/K
Leu1	7.78	4.51	1.55	1.44	С <sup><b>6</b>1</sup> Н: 0.89	8.51	-3.4
Trp2	8.28	5.18	3.43 2.95	-	C <sup>δ1</sup> H: 7.18 N <sup>€1</sup> H: 10.24 C <sup>€3</sup> H: 7.57 C <sup>72</sup> H: 7.17 C <sup>73</sup> H: 7.00 C <sup>η2</sup> H: 7.07	9.21	-7.9
Val3	9.25	4.55	2.09	0.92	-	9.71	-6.3
<sup>D</sup> Pro4	-	4.38	2.21	1.97	C <sup>δ1</sup> H: 3.75	-	-
Gly5	8.45	4.24 3.72	-	-	-	-	-11.0
Leu6	8.26	4.75	1.79	1.63	C <sup>δ1</sup> H: 0.91	9.50	-3.0
Phe7	8.48	4.28	2.45 1.70	-	C <sup>8</sup> H: 6.15 C <sup>€</sup> H: 6.89 C <sup>₹</sup> H: 6.95	8.24	-8.5
Val8	8.15	4.02	1.94	0.79 0.71	-	9.75	-1.7

**Figure S1c:** <sup>1</sup>H 1D spectrum (top) and NMR parameters (table) of peptide **WF** in CD<sub>3</sub>OH at 303K.



**Figure S2:** Comparison of the amide region of the <sup>1</sup>H 1D spectra of the three peptides in CD<sub>3</sub>OH at 303K to check for peptide aggregation. The top spectrum (i) was recorded with 16 scans and a peptide concentration of ~2-3mM. Spectra (ii) and (iii) were recorded after 10-fold dilution of (i), using 16 scans (ii) or 96 scans (iii). Resonance assignments are shown only for the top spectrum in each peptide. Complete assignment for (i) is available in Figure S1.



**Figure S3:** Partial expansions of homonuclear <sup>1</sup>H-<sup>1</sup>H NMR ROESY spectra of **FF**, **YF** and **WF**, obtained in CD<sub>3</sub>OH, highlighting diagnostic  $\beta$ -hairpin NH-NH (top) and NH-C<sup> $\alpha$ </sup>H (bottom) NOEs. Note that the NH-NH region is also shown in Fig. 2 of the main text, but is included here for completion.



**Figure S4:** Temperature dependence of exposed versus hydrogen bonded amides, probed by monitoring <sup>1</sup>H chemical shifts with temperature, from 223K to 323K at intervals of 10K, for (A) **FF**, (B) **YF** and (C) **WF**. Residues involved in internal hydrogen bonds in the folded hairpin conformation are indicated as filled symbols and those in the non-hydrogen bonding position are represented as open symbols. The change in chemical shift for each residue across the three peptides is compared in (D).







**Figure S6:** Partial expansions of the ROESY spectra in CD<sub>3</sub>OH, highlighting key NOEs that establish ring orientation in the three peptides. NOEs observed from the aromatic ring protons to the  $C^{\alpha/\beta/\gamma}H$  region (top), aromatic resonances (middle) and amide resonances (bottom) are indicated



**Figure S7:** Schematic representation of key cross-strand ring proton NOEs observed in the ROESY spectrum of the three peptides.

Electronic Supplementary Material (ESI) for Organic & Biomolecular Chemistry This journal is © The Royal Society of Chemistry 2014



**Figure S8:** Partial expansion of the ROESY spectrum of peptide **YF**, highlighting the corresponding diagnostic NOEs for conformer II,<sup>1</sup> which confirms the presence of two alternate ring orientations in this peptide with sufficient occupancy.



**Figure S9:** Data from the observed Phe7 C<sup> $\delta$ </sup>H proton chemical shifts in CD<sub>3</sub>OH at the respective temperatures (shown as symbols) were fit to an exponential function to predict the C<sup> $\delta$ </sup>H shift at higher (to ~375K) and lower (200K) temperatures. The rate of change in chemical shift for this resonance is dramatic in **WF**, while it is nearly linear for **FF** and **YF**, over the temperature range examined. This anomalous upfield shift in **WF** is likely to arise from strong indole-benzene interactions in the peptide upon adopting a  $\beta$ -hairpin conformation.



**Figure S10:** Influence of ring currents on <sup>1</sup>H NMR chemical shift of geminal  $C^{\beta}H$  protons of F/Y/W2 and F7. An ~0.4ppm upfield shift is seen for the geminal  $7C^{\beta}H$  in **FF** and **YF**, when compared with  $2C^{\beta}H$  in **FF**, despite variation in the chemical nature of residues at position 2, while in **WF**, an additional 0.4-1.2ppm change is obtained.



**Figure S11:** Temperature dependence of Phe7 C<sup> $\alpha$ </sup>H proton resonance under the influence of aromatic interactions. Anomalous upfield chemical shift can be observed in all three peptides, which is a direct evidence for the resonance being under the shielding zone of the N-terminal ring. The d $\delta$ /dT values for each peptide are: -5.1ppb/K (**FF**); -5.5ppb/K (**YF**); -5.3ppb/K (**WF**). The corresponding upfield shifted Phe7 C<sup> $\beta$ </sup>H resonance is in line with this observation.



**Figure S12:** Comparison of the far-UV electronic circular dichroism (ECD) spectra of the three peptides in methanol. The far-UV CD arising from the backbone  $n \rightarrow \pi^*$  and  $\pi \rightarrow \pi^*$  transitions of the peptide unit are often masked by contributions from aromatic interactions. We observe anomalous spectra in all three cases, due to significant contribution of aromatic residues to the far-UV CD and the characteristic exciton couplet at 195nm and 215nm for a  $\beta$ -sheet structure is not obtained. In **FF**, we observe two negative bands at ~195nm and ~210nm arising from Phe-Phe interactions. Similarly, the spectrum for **YF** displays a shallow trough centred at ~220nm and a second negative maximum at ~198nm, and resembles the spectrum observed for Tyr-Tyr interactions, suggesting that the contributions of Phe to far-UV CD is surpassed by the influence of Tyr. The anomalous contribution becomes even stronger in the case of **WF**, with a minor positive band at 228nm and negative maxima at ~200nm and ~210nm.

## **Supplementary Tables**

**Table S1:** Summary of experimental constraints used in the NMR structure calculation and observed violations in the calculated structures.

Experimental Constraints	No. of Constraints			
Experimental Constraints	FF	YF	WF	
Intraresidue NOEs	38	37	40	
Sequential NOEs	13	13	15	
Long range NOEs	14	20	25	
Hydrogen bonds	4	4	4	
Angle constraints	12	12	12	
Violations observed	0	0	0	

**Table S2**: Average backbone torsion angles calculated for 35 best structures for the three peptides.

<b>FF</b> : Ac-LFV- <sup>D</sup> PG-LFV-NH₂					
Residue	Phi <sup>a</sup>	Psi <sup>a</sup>			
Leu1	-	75.5 +/- 5.0			
Phe 2	-126.1 +/- 6.2	137.3 +/- 7.5			
Val 3	-136.0 + 4.5	73.4 +/- 2.6			
<sup>D</sup> Pro 4	69.7 +/- 0.03	-113.3 +/- 1.6			
Gly 5	-90.1 +/- 0.1	15.7 +/- 0.4			
Leu 6	-133.1 +/- 2.9	156.7 +/- 7.8			
Phe 7	-130.2 +/- 5.0	141.5 +/- 6.0			
Val 8	-129.5 +/- 6.0	-			

<b>YF</b> : Ac-LYV- <sup>D</sup> PG-LFV-NH <sub>2</sub>					
Residue	Phi <sup>a</sup>	Psi <sup>a</sup>			
Leu1	-	81.3 +/- 2.4			
Tyr 2	-130.3 +/- 8.6	143.1 +/- 7.9			
Val 3	-136.0 + 5.7	73.2 +/- 2.5			
<sup>D</sup> Pro 4	69.7 +/- 0.03	-113.0 +/- 2.2			
Gly 5	-90.5 +/- 0.5	16.4 +/- 4.9			
Leu 6	-128.5 +/- 6.1	168.5 +/- 11.9			
Phe 7	-129.6 +/- 8.0	147.3 +/- 2.7			
Val 8	-129.1 +/- 7.6	-			

WF: Ac-LWV- <sup>D</sup> PG-LFV-NH <sub>2</sub>					
Residue	Phi <sup>a</sup>	Psi <sup>a</sup>			
Leu1	-	84.8 +/- 0.1			
Trp 2	-99.1 +/- 0.1	125.8 +/- 0.9			
Val 3	-126.1 + 0.6	75.3 +/- 0.9			
<sup>D</sup> Pro 4	69.7 +/- 0.04	-111.8 +/- 0.9			
Gly 5	-91.4 +/- 0.1	15.7 +/- 2.0			
Leu 6	-128.0 +/- 2.1	142.3 +/- 0.0			
Phe 7	-119 +/- 0.2	144.2 +/- 0.3			
Val 8	-140.2 +/- 0.2	-			

<sup>a</sup>Values in degree

### **Supplementary References**

1. R. Sonti, R. Rai, S. Ragothama and P. Balaram, *J. Phys. Chem. B*, 2012, **116**, 14207-14215.