

Supporting information

Design, synthesis and properties of artificial nucleic acids from (R)-4-amino-butane-1, 3-diol

Pengfei Li,^{a#} Jingjing Sun,^{a#} Meng Su^a, Xiaogai Yang^c and Xinjing Tang^{a,b*}

^aState Key Laboratory of Natural and Biomimetic Drugs, School of Pharmaceutical Sciences, Peking University, Beijing, 100191, China

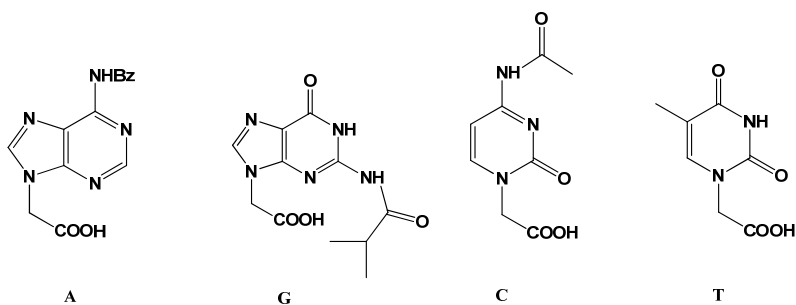
^bState key Laboratory of Drug Research, Shanghai Institute of Materia Medica, Chinese Academy of Sciences, Shanghai 201203, China

E-mail: xinjingt@bjmu.edu.cn; Tel: +86-010-82800535

^cDepartment of Chemical Biology, School of Pharmaceutical Sciences, Peking University, Beijing 100191, People's Republic of China

- S1.** Synthesis of acetate-nucleobases and acyclic nucleotide analogues.
- S2.** (S)- amino-propylene-glycol nucleic acid modified duplexes.
- S3.** Solid synthesis and evaluation of (R)-Am-BuNA modified oligonucleotides.
- S4.** Van't Hoff plot of (R)-AM-BuNA modified oligonucleotides.
- S5.** NMR-spectrum and ESI-MS of compounds in our work.
- S6.** ESI-MS of oligonucleotides in our work.

S1. Synthesis of acetate-nucleobases and acyclic nucleotide analogues.



Scheme S1 Acetate-nucleobases in our work. (A, G, C and T)

Acetate-nucleobases (A, G and C, T is commercially available) are synthesized according to previous reported methods with modifications.¹ NMR data were consistent to those in literature reports.

(N⁶-benzyladenin-9-yl) acetic acid (A)

¹H-NMR (400 MHz, DMSO-d₆) δ = 11.20 (s, 1 H), 8.74 (s, 1 H), 8.46 (s, 1 H), 8.06 (d, 2 H, J = 7.3 Hz), 7.64 (m, 1 H), 7.54-7.65 (m, 2 H), 5.11 (s, 2 H).

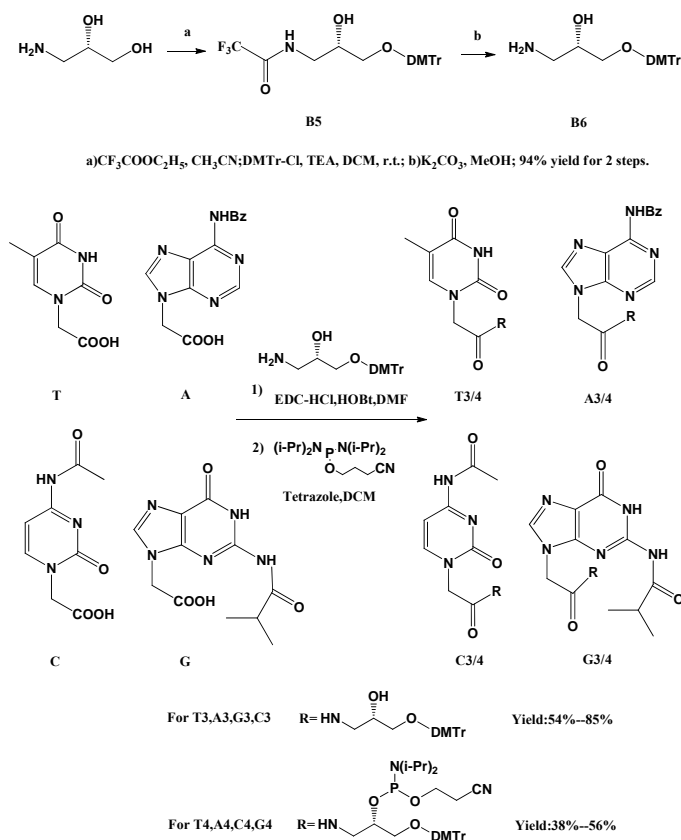
2-(4-acetamido-2-oxopyrimidin-1(2H)-yl) acetic acid (C)

¹H-NMR (400 MHz, DMSO-d₆) δ = 10.87 (s, 1 H), 8.04 (d, 1 H, J = 7.2 Hz), 7.17 (d, 1 H, J = 7.2 Hz), 4.54 (s, 2 H), 2.11 (s, 3 H). ¹³C-NMR (100 MHz, DMSO-d₆) δ = 170.9, 169.3, 162.8, 155.2, 150.7, 95.0, 50.54, 24.3.

2-(2-isobutyramido-6-oxo-1H-purin-9(6H)-yl) acetic acid (G)

¹H-NMR (400 MHz, DMSO-d₆) δ = 12.10 (s, 1 H), 11.69 (s, 1 H), 7.99 (s, 1 H), 4.90 (s, 2 H), 2.76 (m, 1 H), 1.10 (d, 6 H, J = 7Hz); ¹³C-NMR (100 MHz, DMSO-d₆) δ = 180.2, 169.0, 154.8, 148.1, 140.3, 119.5, 44.5 34.7, 27.7, 18.9.

S2. (S)-3-Amino-1, 2-propanediol nucleic acid modified duplexes[#]



Scheme S2. Synthesis of backbone and monomers of (S)-3-amino-1, 2-propanediol nucleic acid

Table S1. Sequences of (S)-3-Amino-1, 2-propanediol nucleic acid ((S)-AmpNA) modified duplex.[#]

Entry	Sequence ^S	Calculated Mw	Measured Mw
AmpNA-mmp 1	3'- <u>c</u> ACA <u>c</u> CTT <u>g</u> CC <u>a</u> TC <u>g</u> -2'	4545.9	4547.2
AmpNA-mmp 2	3'- <u>c</u> ac <u>a</u> ctt <u>g</u> ccat <u>g</u> -2'	4696.0	4697.6
AmpNA-mmp 3	<u>c</u> ACACCTTGCCATCG	4485.8	4487.0
AmpNA-mmp 4	CAC <u>a</u> CCCTTGCCATCG	4485.8	4487.0
AmpNA-mmp 5	CACACCT <u>t</u> GCCATCG	4485.8	4487.0
AmpNA-mmp 6	CACACCTTGCC <u>a</u> TCG	4485.8	4487.0
AmpNA-mmp 7	CACACCTTGCCATC <u>g</u>	4485.8	4487.0

^S underlined lower case letters represent (S)-3-Amino-1,2-propanediol nucleic acid modified nucleotides and upper case letters represent DNA nucleotides.

[#]Part of data came from published thesis of master student from our laboratory: Meng Su, "Functionalization and Reconstruction of Oligonucleotides", Master Thesis, 2012, Peking University.

S3. Solid synthesis and evaluation of (R)-Am-BuNA modified oligonucleotides.

DNA synthesis, cleavage and purification

DNA oligonucleotides were synthesized on an Applied Biosystems Incorporated 394 synthesizer. The synthesis was carried out on 1 μmol scale. The coupling time and concentration of phosphoramidite monomers for the acyclic nucleotides were the same as those for the natural bases. With a refurbished ABI 394, we set 90s coupling time and 0.1M concentration for all monomers. Trityl detection showed the same level of incorporation efficiency for acyclic nucleosides as that of natural DNA oligonucleotides with the average step yield over 97%.

After synthesis, fresh concentrated ammonium hydroxide was added to CPGs. The CPGs were shaken for 24 hours at room temperature and then concentrated. The residue was dissolved by small amount of water and was then purified with Waters 320 HPLC on an Eclipse XDB-C18 column (5 μM , 9.4 \times 250 mm). Conditions: solvent A, 0.05M TEAA buffer; solvent B, acetonitrile. Started at 0% B; liner gradient 2% B/min, flow rate: 2 mL/min. 80% aqueous acetic acid (500 μL) was added to dry oligonucleotides to remove trityl group. The solution was shaken for 30 min at room temperature, and was then concentrated and purified by HPLC. HPLC conditions: started at 0% B; liner gradient 2% B/min, flow rate: 2 mL/min. PAD detector wavelength: 260 nm.

Thermal denaturation studies for acyclic-modified oligonucleotides.

The oligonucleotides were mixed in corresponding buffers and ion concentrations. Oligonucleotide solutions were hybridized by first heating at 90 $^{\circ}\text{C}$ for 5 min and then slowly cooled down to room temperature. The melting profiles started with a denaturing run from 25 $^{\circ}\text{C}$ to 85 $^{\circ}\text{C}$ at a rate of 1 $^{\circ}\text{C}$ /min. The absorbance at 260 nm was monitored at 0.5 $^{\circ}\text{C}$ intervals on a Beckman Series 800 UV spectrometer (at 295 nm for denaturation of quadruplex). The melting temperatures of the oligonucleotides were determined as the peak of the first derivations of the corresponding melting curves.

Enzymatic stability of acyclic-modified oligonucleotides with FBS and SVPDE

Enzymatic stability of the acyclic-modified single-stranded DNA and thrombin-binding DNA aptamer was tested in the presence of fetal bovine serum (FBS) and snake venom phosphodiesterase (SVPDE). The reactions were performed with the oligonucleotide concentrations of 5 μM (SVPDE) and 10 μM (FBS) oligonucleotide at 37 $^{\circ}\text{C}$. To the above oligonucleotide solution, proper amount of enzymes was added for oligonucleotide degradation. Aliquots (10 μL) were taken at intervals, quenched by the addition of equal volume of 10 mM EDTA aqua on ice bath and kept at -80 $^{\circ}\text{C}$ for further PAGE analysis. Every sample was added 4 μL loading buffer (6-fold) before PAGE assay. The mixture was subjected to the gel electrophoresis using 20% native polyacrylamide gel with 1 \times TBE buffer at 150 V for 40min.

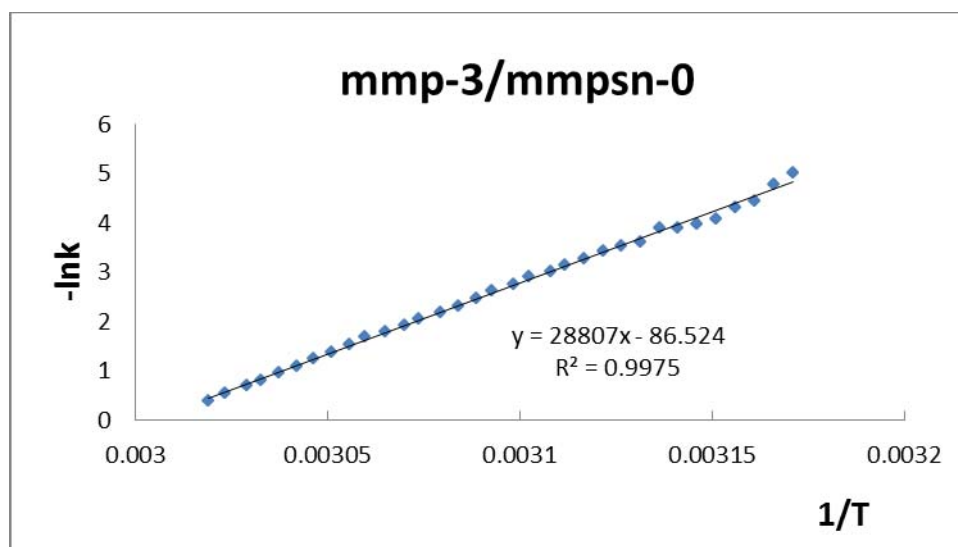
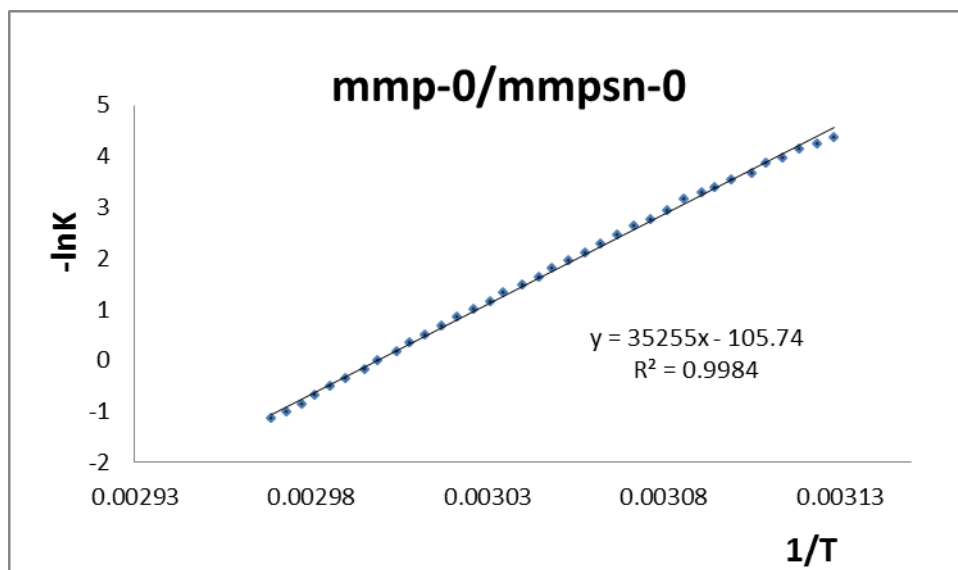
CD spectra measurement

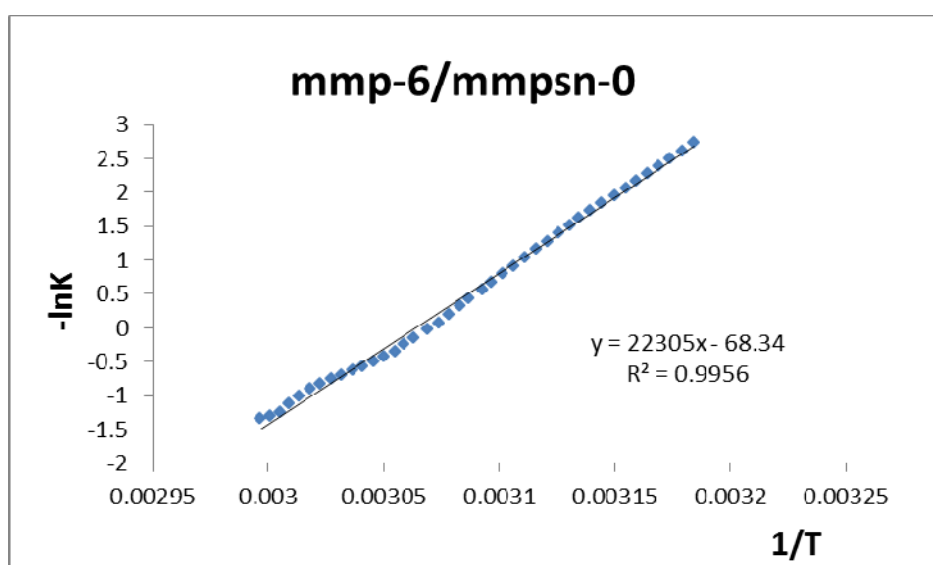
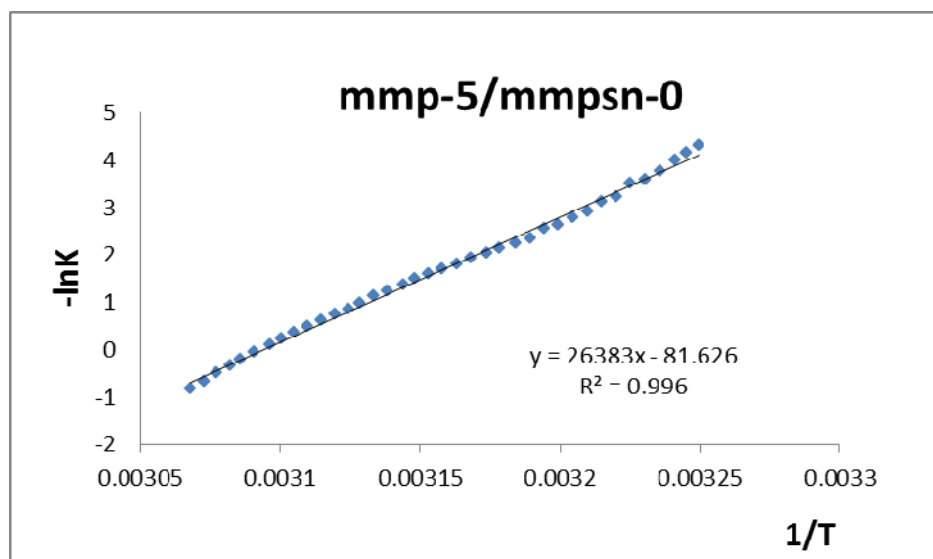
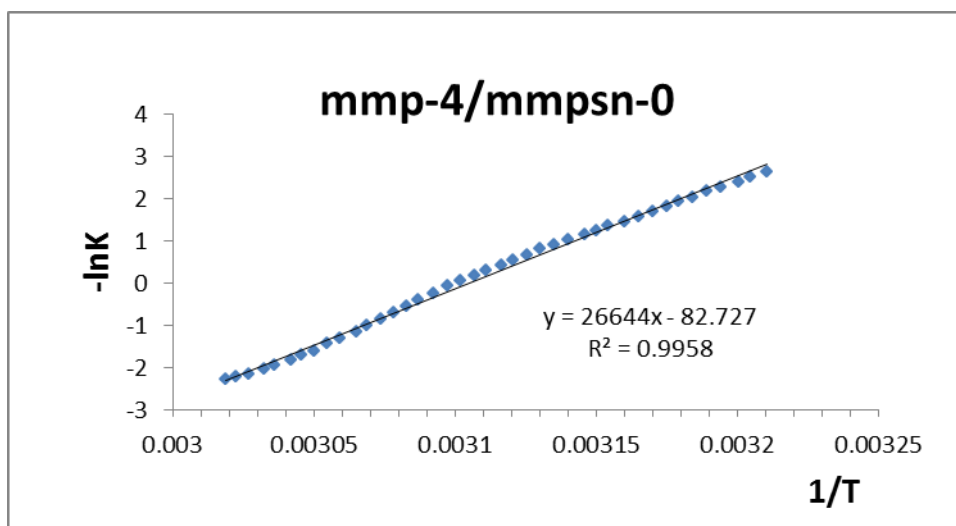
The samples of oligonucleotides in corresponding buffers and ion concentrations were first annealed. Then CD spectra of the samples were measured at room temperature (298 K) in 0.1cm cuvettes using a Jasco J-810 spectropolarimeter. The wavelength was scanned from 220 to 320 nm with a scanning speed of 200 nm/min for 3 times. The obtained curves were smoothed by the built-in software.

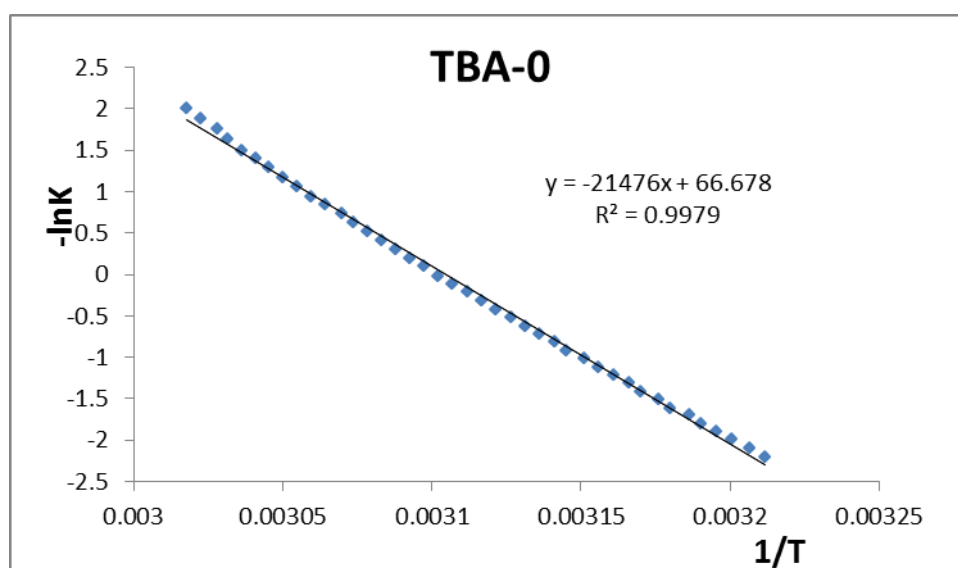
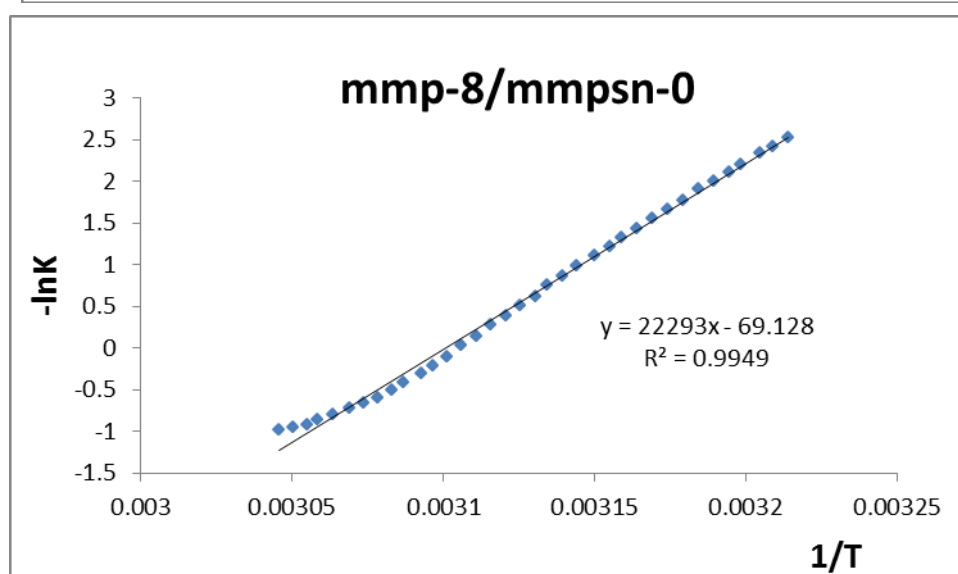
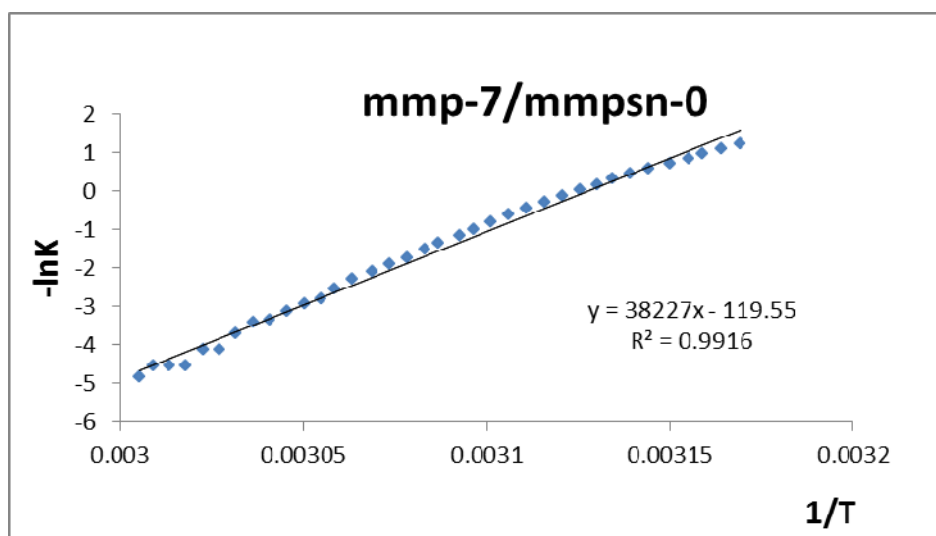
Methods for the calculation of thermodynamic parameters of duplex or quadruplex formation

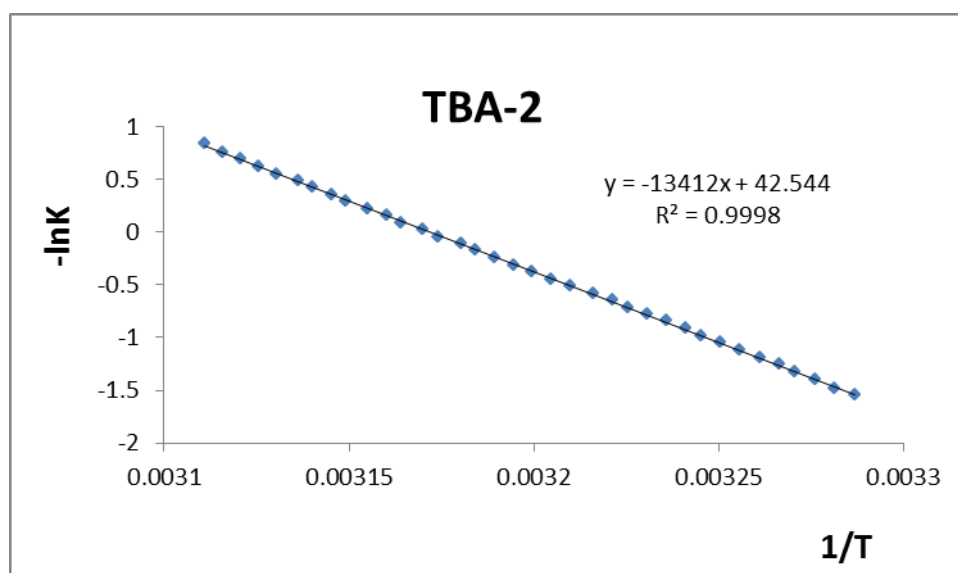
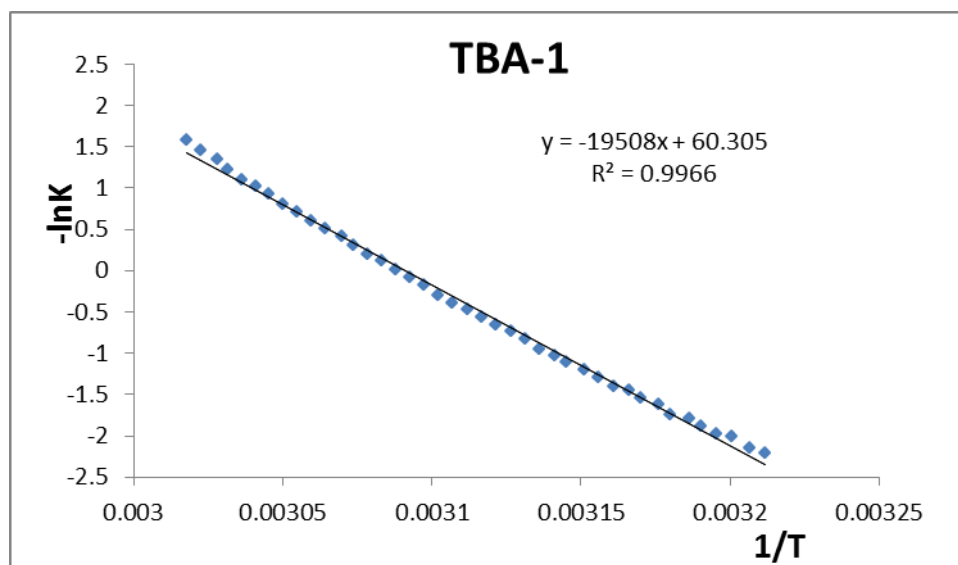
In melting processes of duplexes and quadruplexes, we supposed the existence of folded and unfolded states. The equilibrium constant, K , can be determined by the ratio of two folded and unfolded states ($K=\theta/(1-\theta)$). Supposed that θ referred to the percentage of duplex in all oligonucleotides, then $K=(A_{\text{max}}-A_n)/(A_n-A_{\text{min}})$. According to Gibbs fundamental equation: $\Delta G=-RT\ln(K)=\Delta H-T\Delta S$, a linear regression with $1/T$ as x and $\ln(K)$ as y was conducted to obtain the slope as $-(\Delta H/R)$ and vertical intercept as $(\Delta S/R)$. Accordingly, ΔH and ΔS can be obtained through the melting profiles. All the parameters were constructed by fitting melting curves in a temperature range of $T_m \pm 10^{\circ}\text{C}$

S4. Van't Hoff plot of (R)-AM-BuNA modified oligonucleotides.

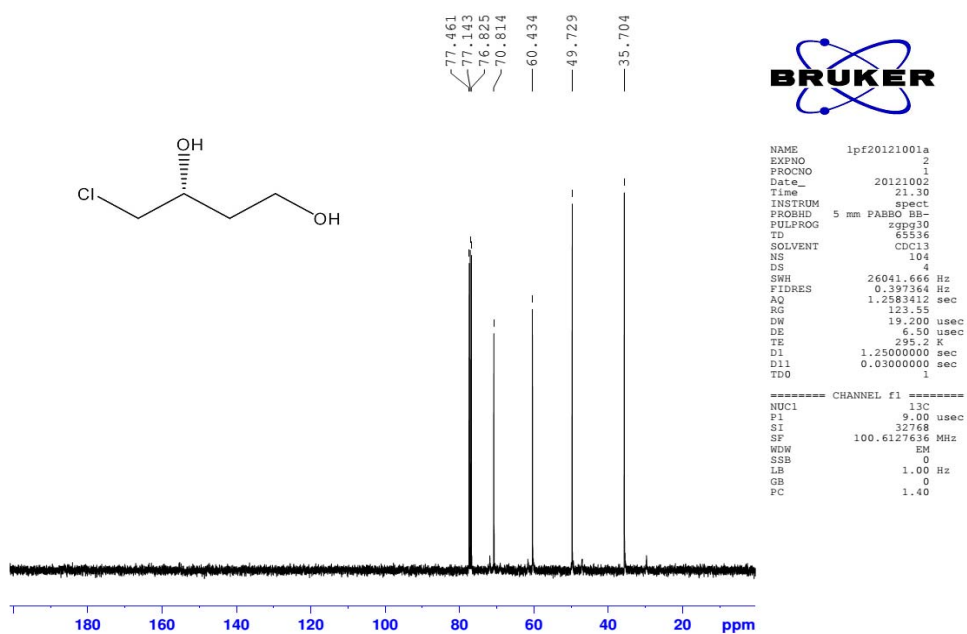
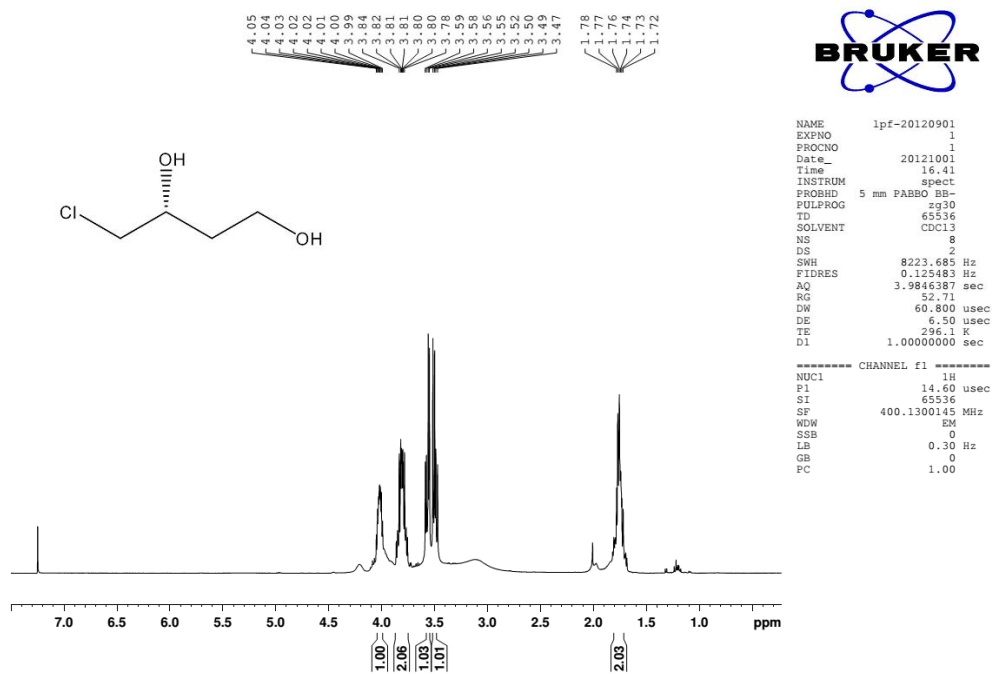


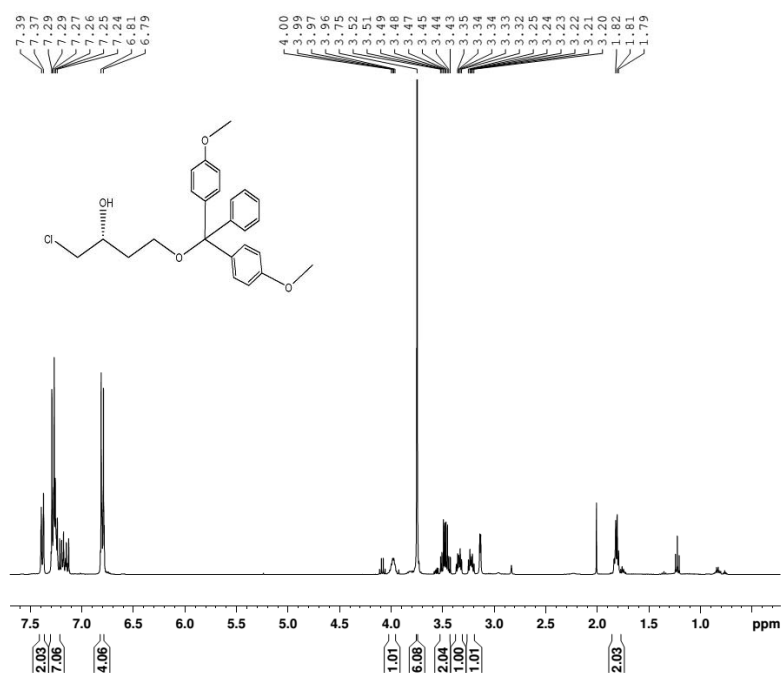






S5. NMR spectra and ESI-MS of compounds in our work.



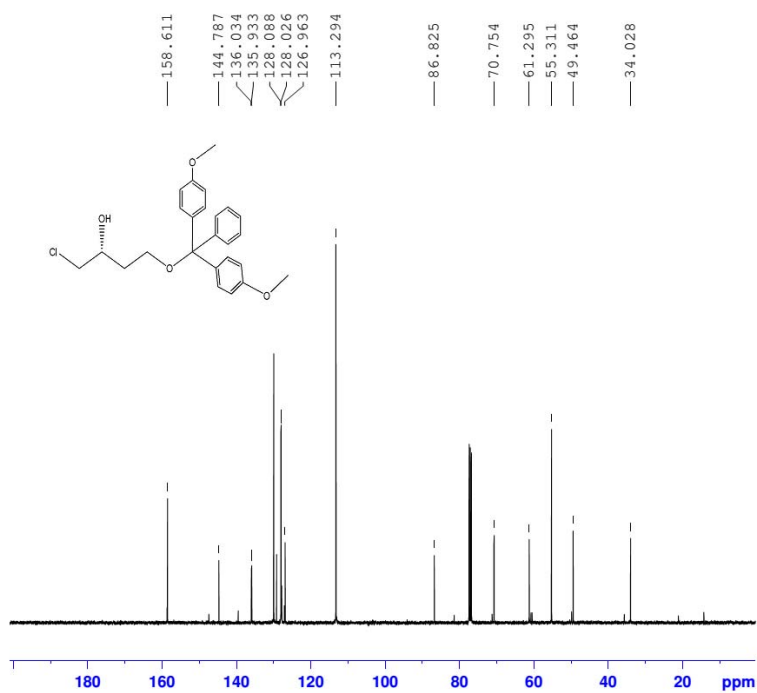


```

NAME      LPF-20121004
EXPNO     1
PROCNO    1
Date_     20121004
Time      18.15
INSTRUM   spect
PROBHD    5 mm PABBO BB-
PULPROG   zg30
ID        65536
SOLVENT   CDCl3
NS        16
DS        2
SWH       8223.685 Hz
FIDRES    0.125483 Hz
AQ        3.9846387 sec
RG        48.73
DW        60.800 usec
DE        6.50 usec
TE        294.0 K
D1        1.0000000 sec
TDO       1
    
```

```

===== CHANNEL f1 =====
NUC1      1H
P1        14.00 usec
SI        65536
SF        400.1300281 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        1.00
    
```

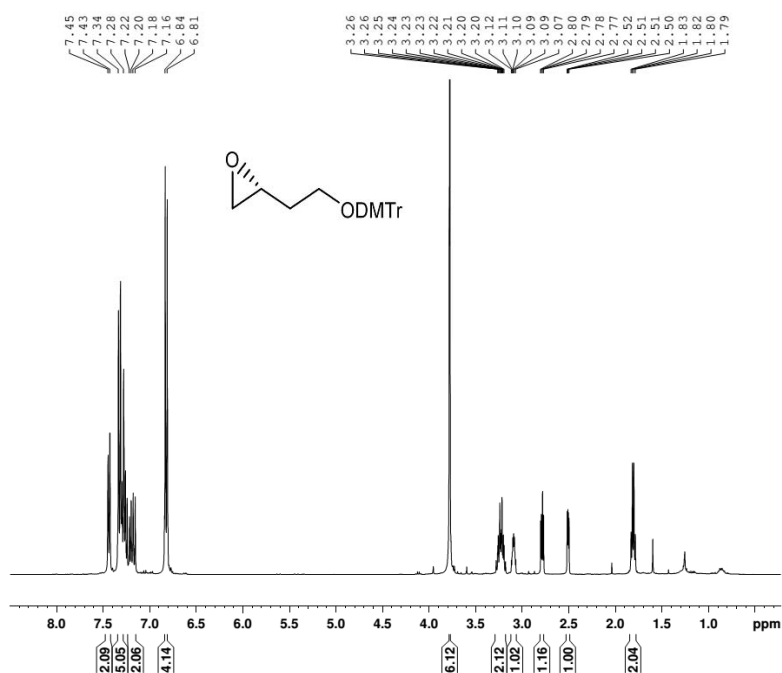


```

NAME      LPF-20121004
EXPNO     2
PROCNO    1
Date_     20121004
Time      18.27
INSTRUM   spect
PROBHD    5 mm PABBO BB-
PULPROG   zgpg30
TD        65536
SOLVENT   CDCl3
NS        256
DS        4
SWH       26041.666 Hz
FIDRES    0.397364 Hz
AQ        1.2583412 sec
RG        205.82
DW        19.200 usec
DE        6.50 usec
TE        295.5 K
D1        1.2500000 sec
D11       0.0300000 sec
TDO       1
    
```

```

===== CHANNEL f1 =====
NUC1      13C
P1        9.00 usec
SI        32768
SF        100.6127622 MHz
WDW       EM
SSB       0
LB        1.00 Hz
GB        0
PC        1.40
    
```

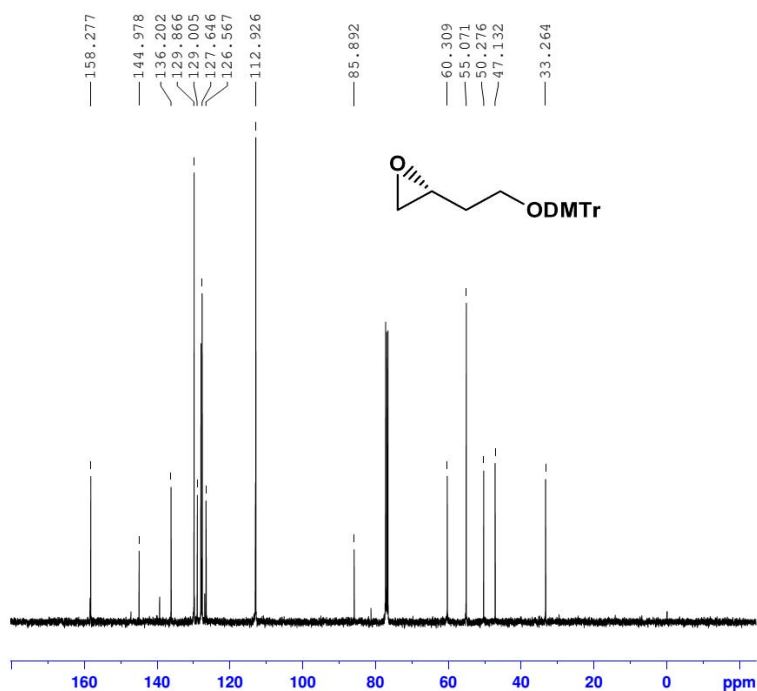


```

NAME      lpf1030b3
EXPNO    1
PROCNO   1
Date_    20121030
Time     22.47
INSTRUM  spect
PROBHD   5 mm PABBO BB-
PULPROG  zg30
ID       65536
SOLVENT  CDCl3
NS       16
DS       2
SWH      8223.685 Hz
FIDRES   0.125483 Hz
AQ       3.9846387 sec
RG       72.2
DW       60.800 usec
DE       6.50 usec
TE       294.2 K
D1       1.00000000 sec
TD0      1
    
```

```

===== CHANNEL f1 =====
NUC1     1H
P1       14.00 usec
SI       65536
SF       400.1300169 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.00
    
```

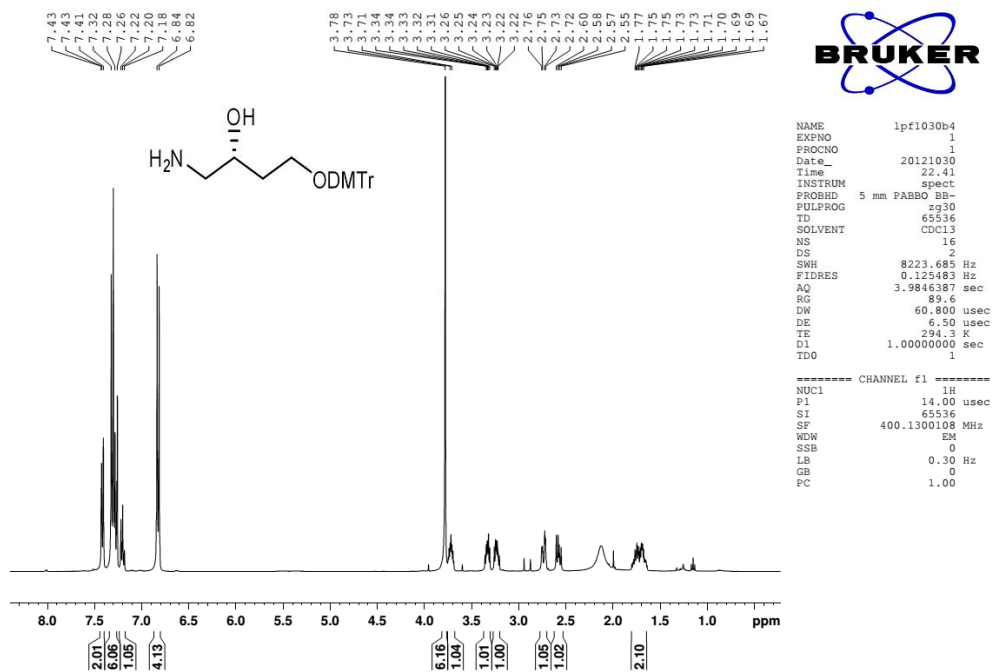
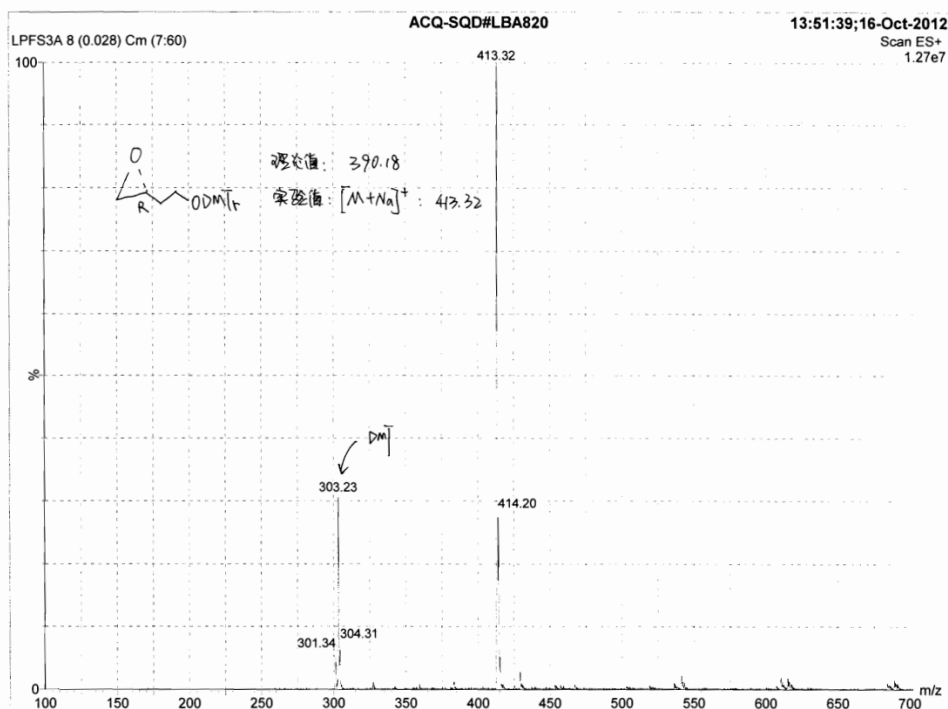


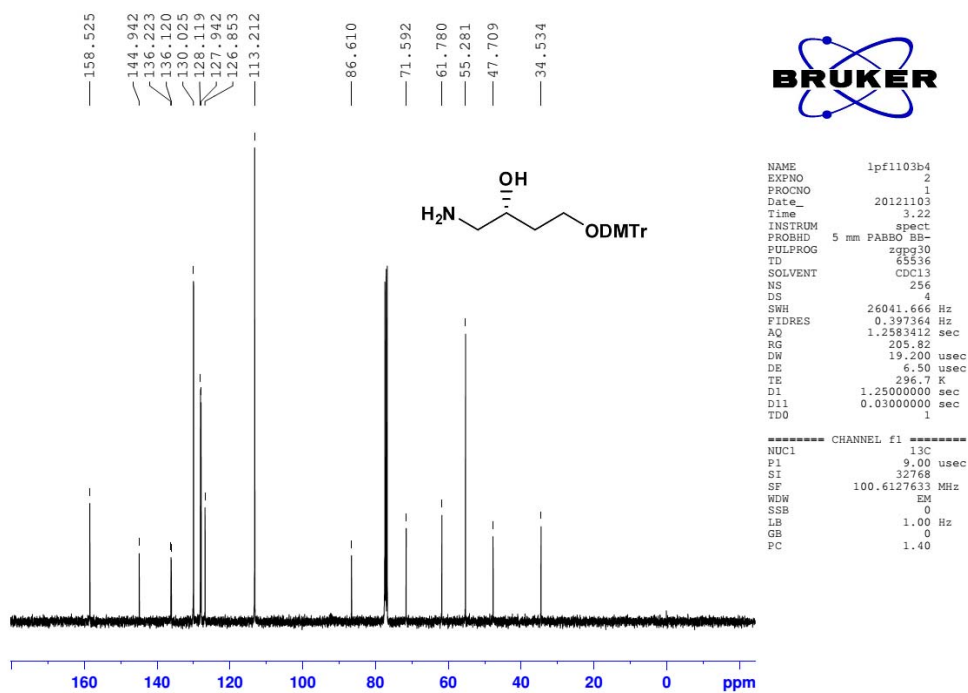
```

NAME      lpf1103b3
EXPNO    2
PROCNO   1
Date_    20121103
Time     3.06
INSTRUM  spect
PROBHD   5 mm PABBO BB-
PULPROG  zgpg30
TD       65536
SOLVENT  CDCl3
NS       256
DS       4
SWH      26041.666 Hz
FIDRES   0.197364 Hz
AQ       1.2583412 sec
RG       183
DW       19.200 usec
DE       6.50 usec
TE       296.6 K
D1       1.25000000 sec
D11      0.03000000 sec
TD0      1
    
```

```

===== CHANNEL f1 =====
NUC1     13C
P1       9.00 usec
SI       32768
SF       100.6127845 MHz
WDW      EM
SSB      0
LB       1.00 Hz
GB       0
PC       1.40
    
```



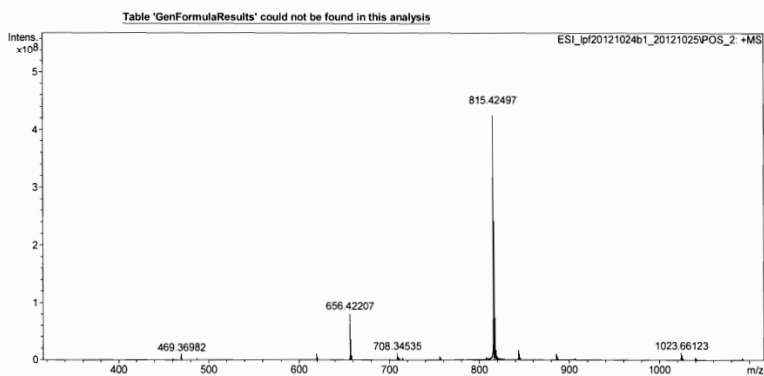
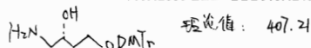


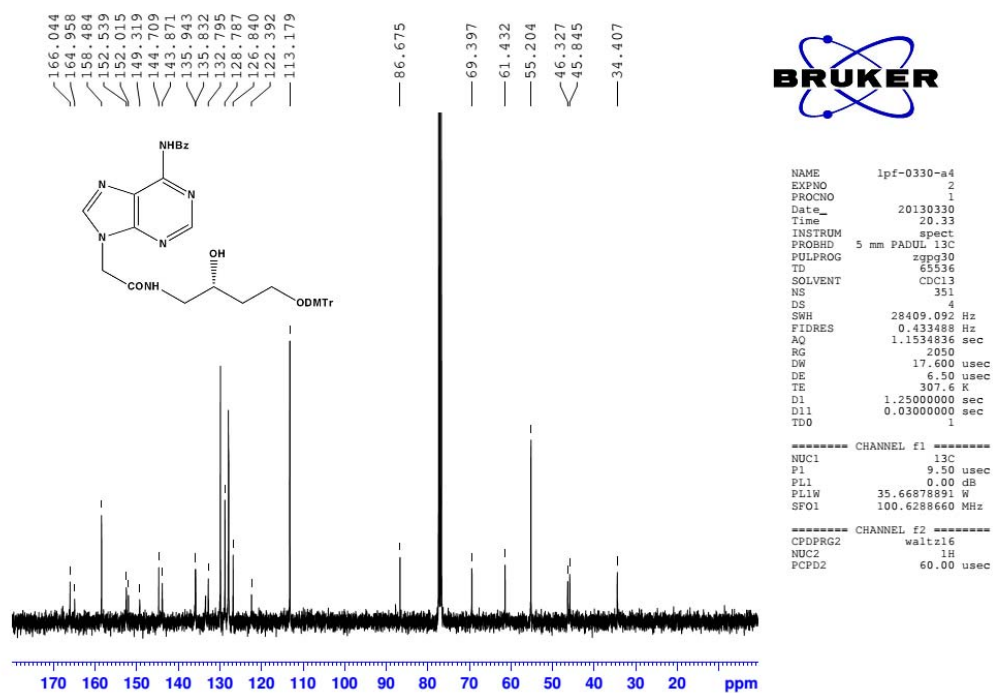
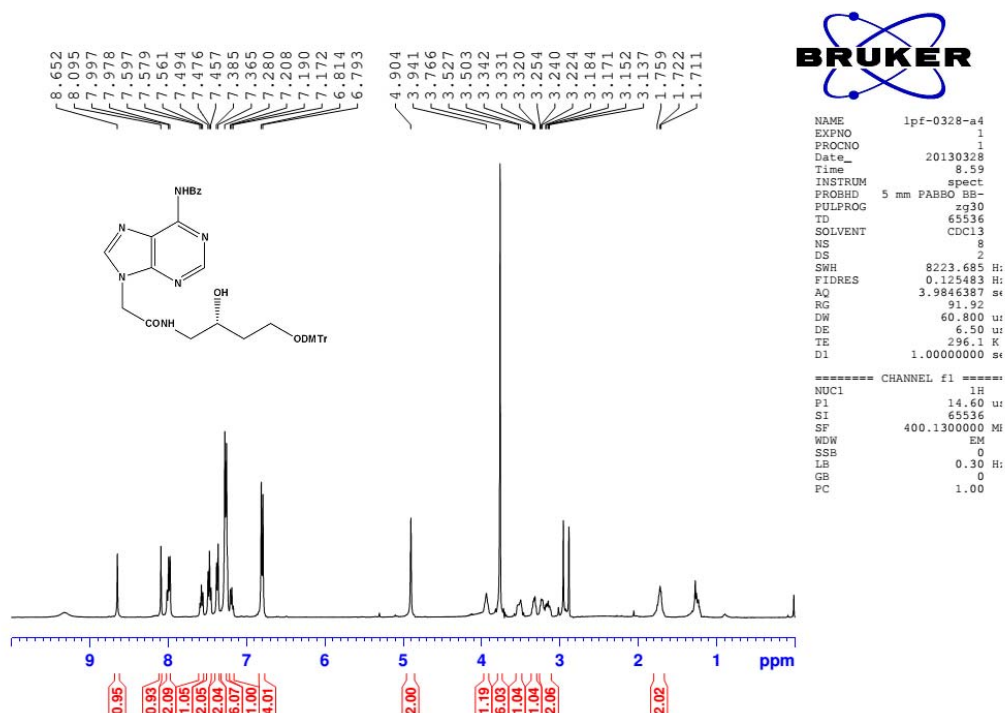
天然药物及仿生药物国家重点实验室

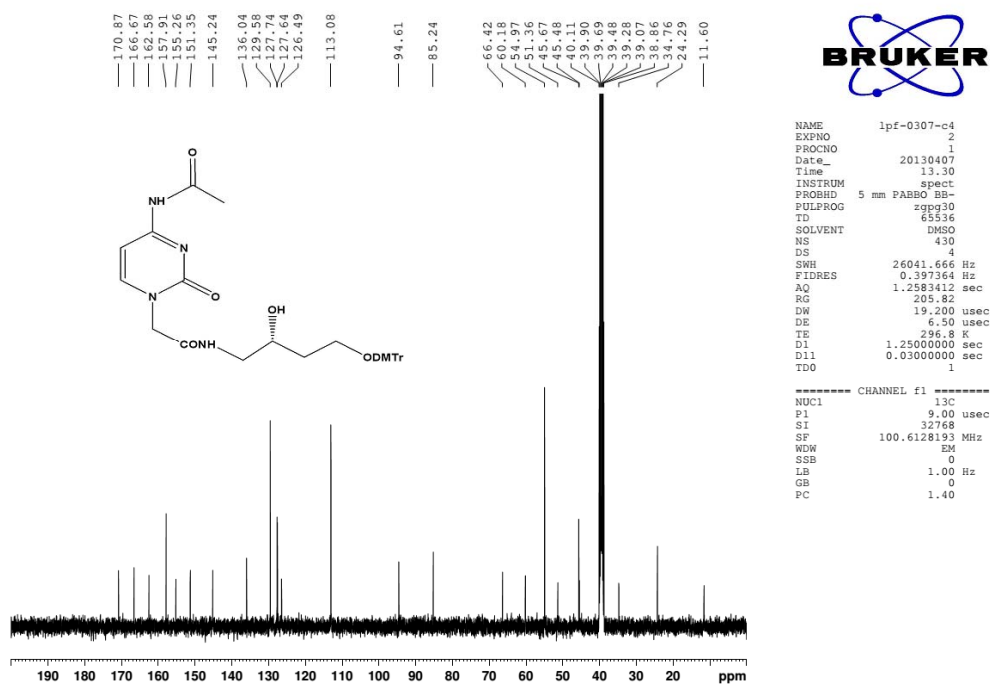
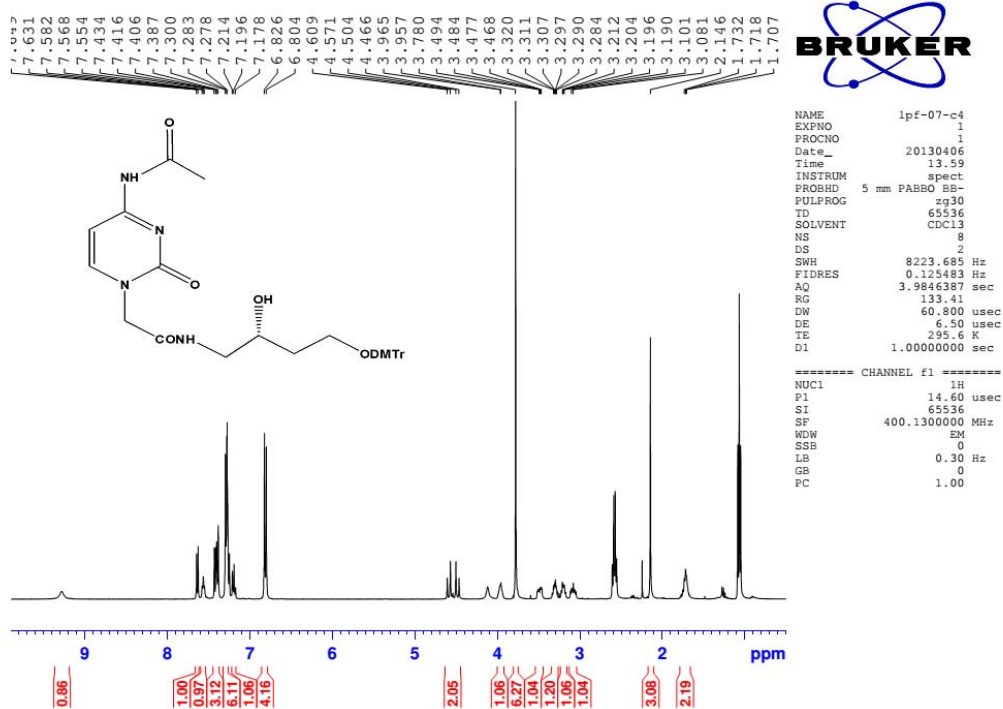
Analysis Name C:\data_sample\ESI_121020_30\ESI_lp20121024b1_201210 Acquisition Date 10/25/2012 4:19:27 PM
 25VPOS_2 Operator bpfsh@bjmu.edu.cn; Tel:010-82801437
 Instrument FT_MS_Bruker APEX IV (7.0 T)

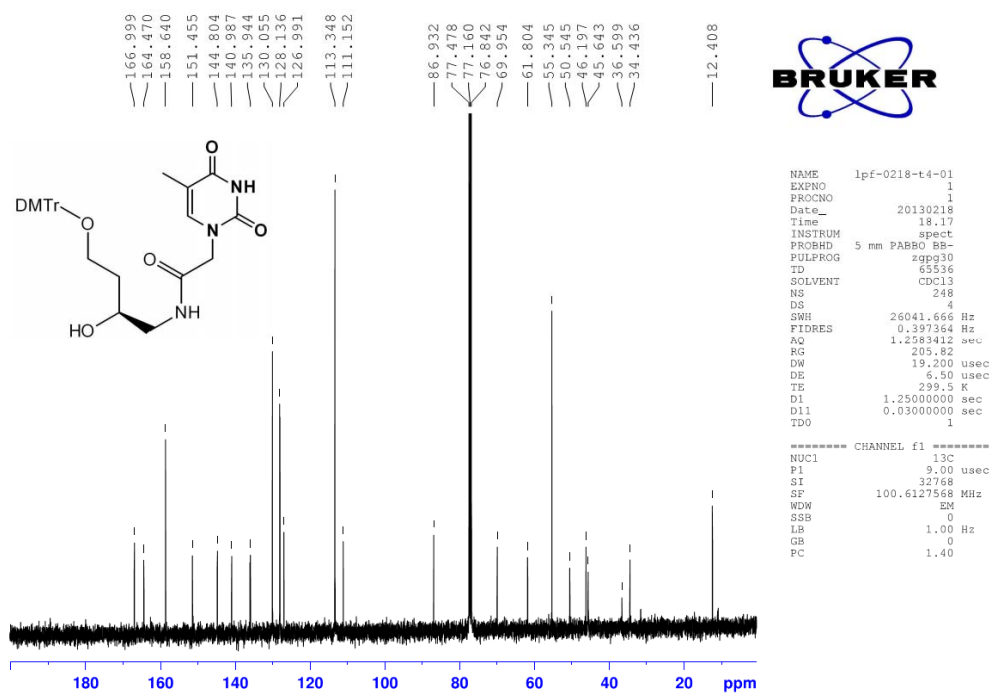
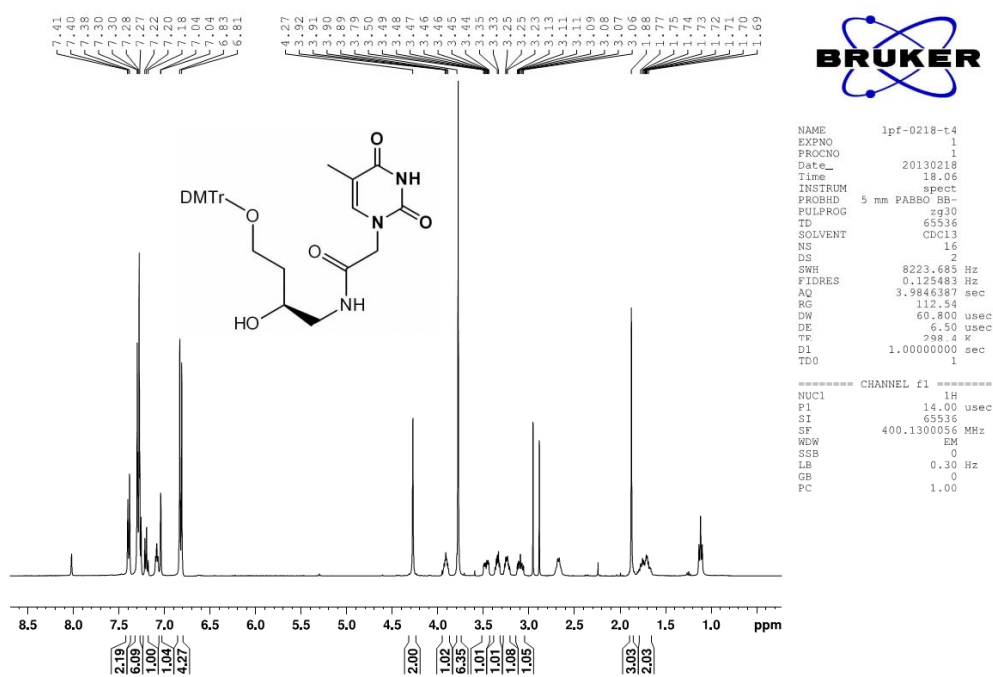
Comment ESI POS C25H29N1O4 MW 407.2097
 CAL588: 226.16718;249.15695;340.25887;301.14158;
 362.24081;391.28483;413.26647;453.34353;
 475.32548;509.25407;509.25407;568.42780;
 588.40954;679.51166;701.49361;826.47121;

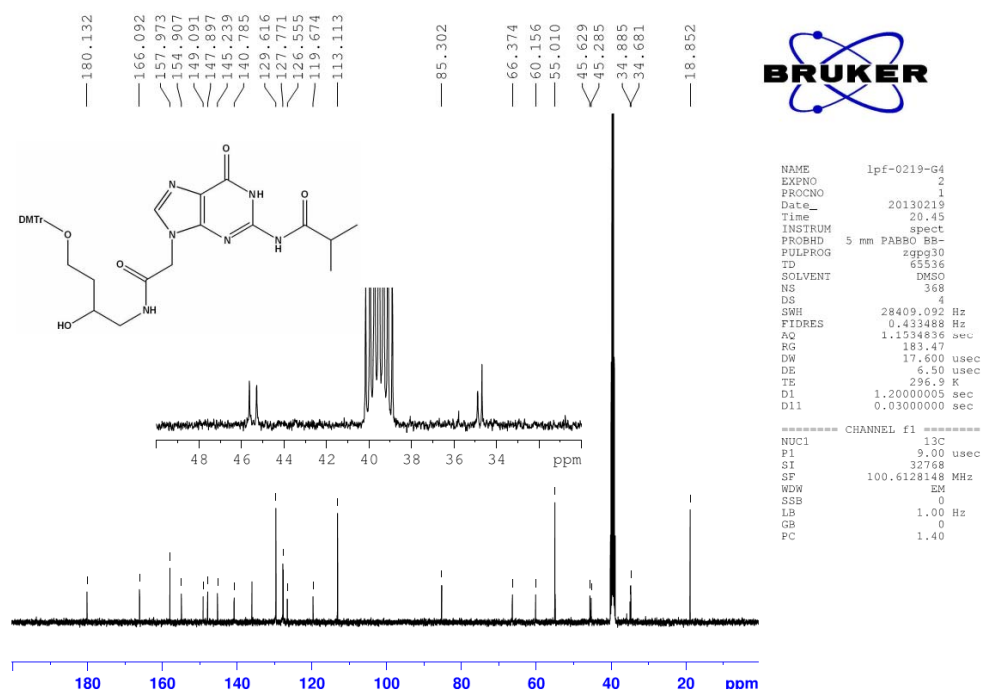
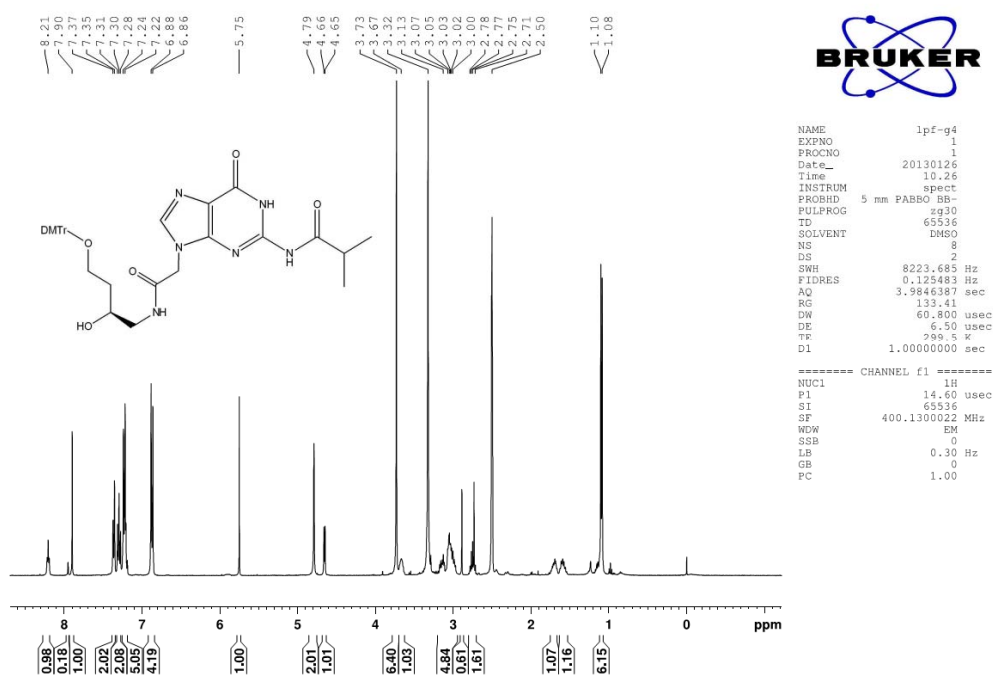
Acquisition Result: Exact Mass Measured Mass Error (mDa) Error (ppm) Description
 815.42659 815.42497 1.62 1.99 2M+H⁺;e

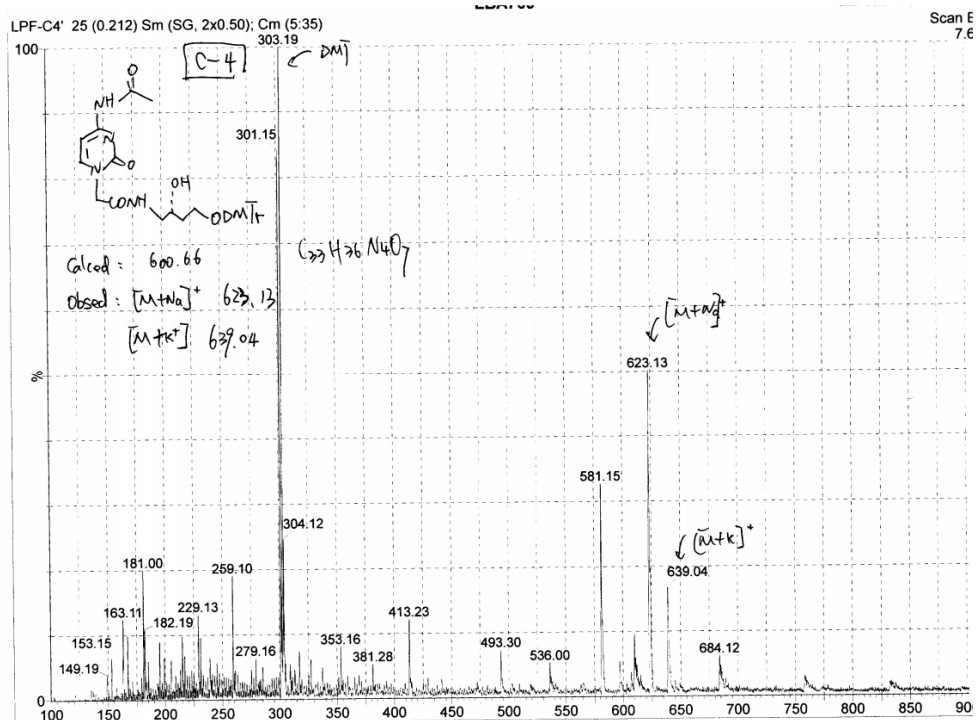
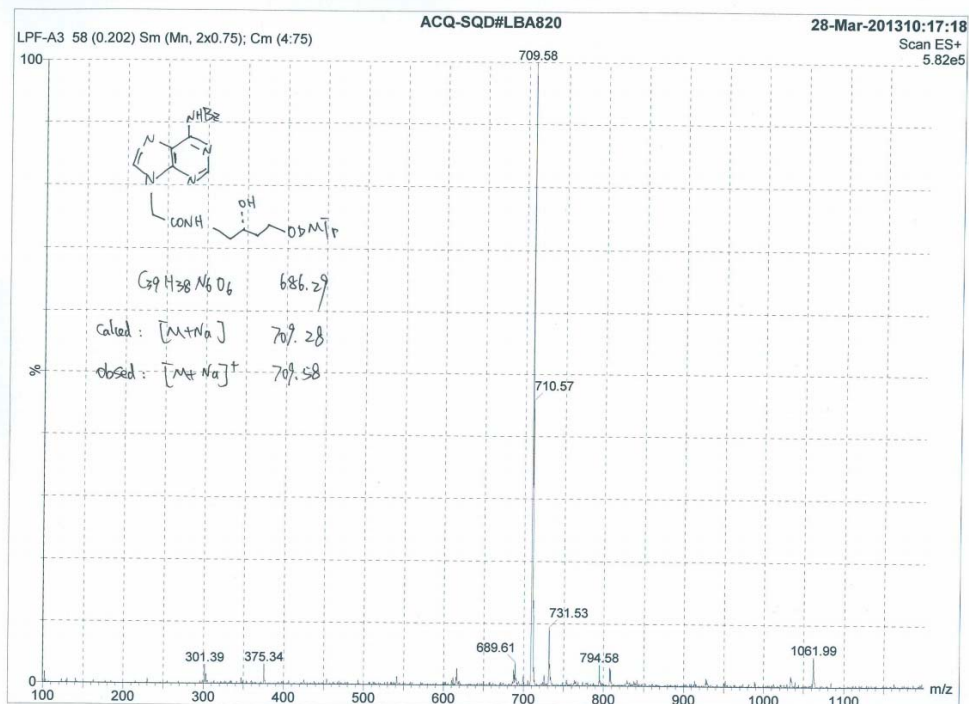


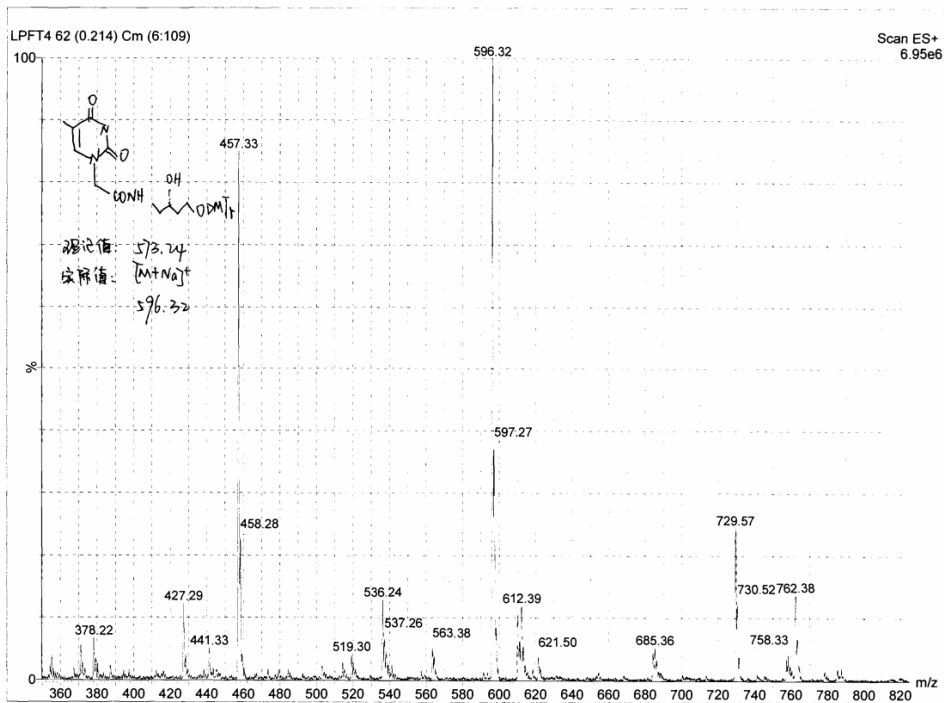
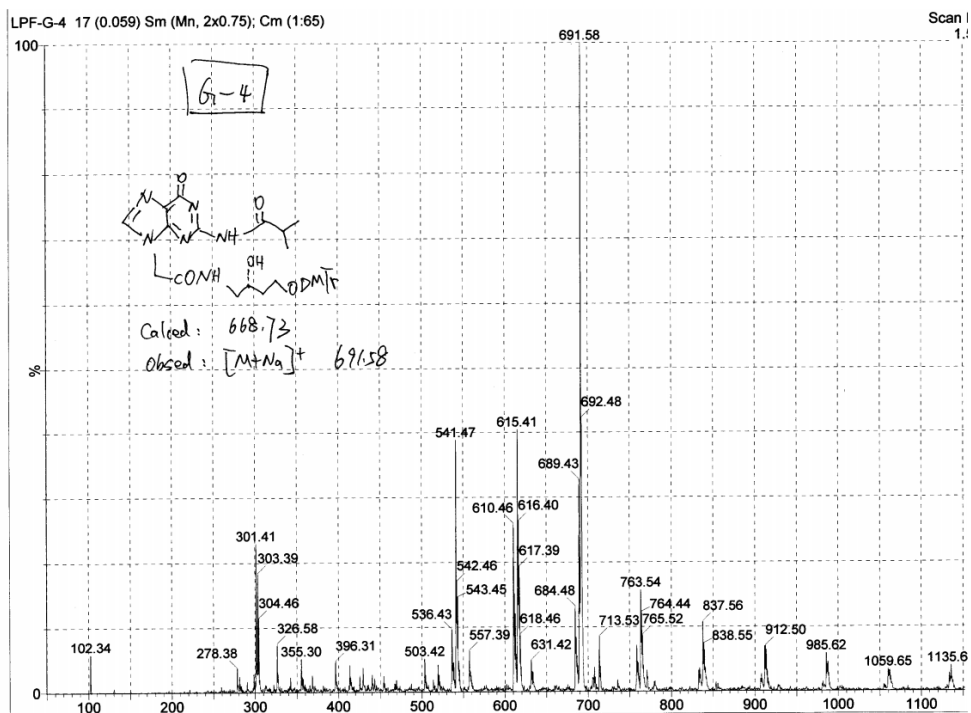


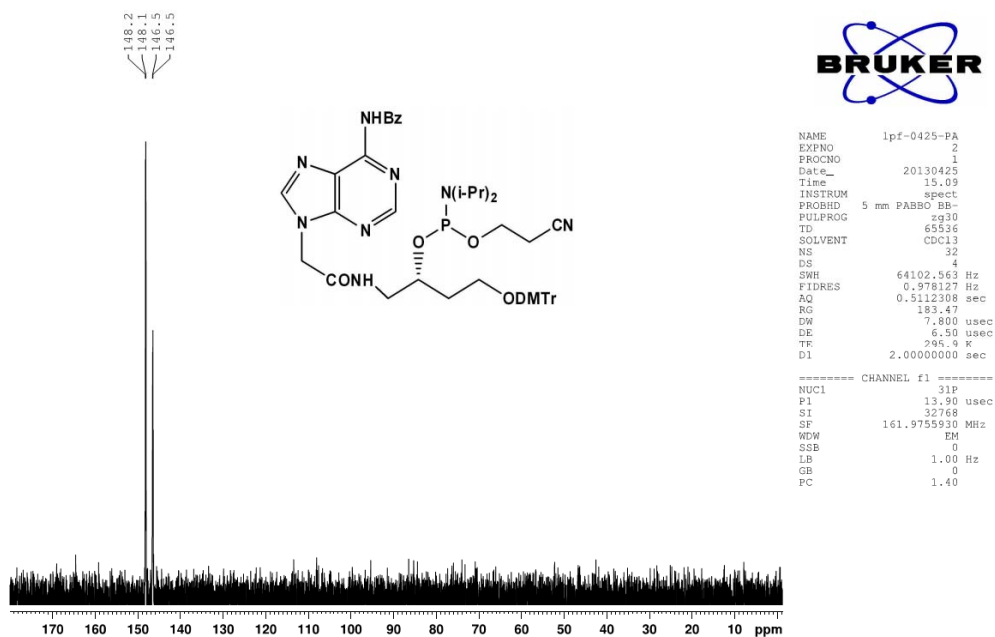
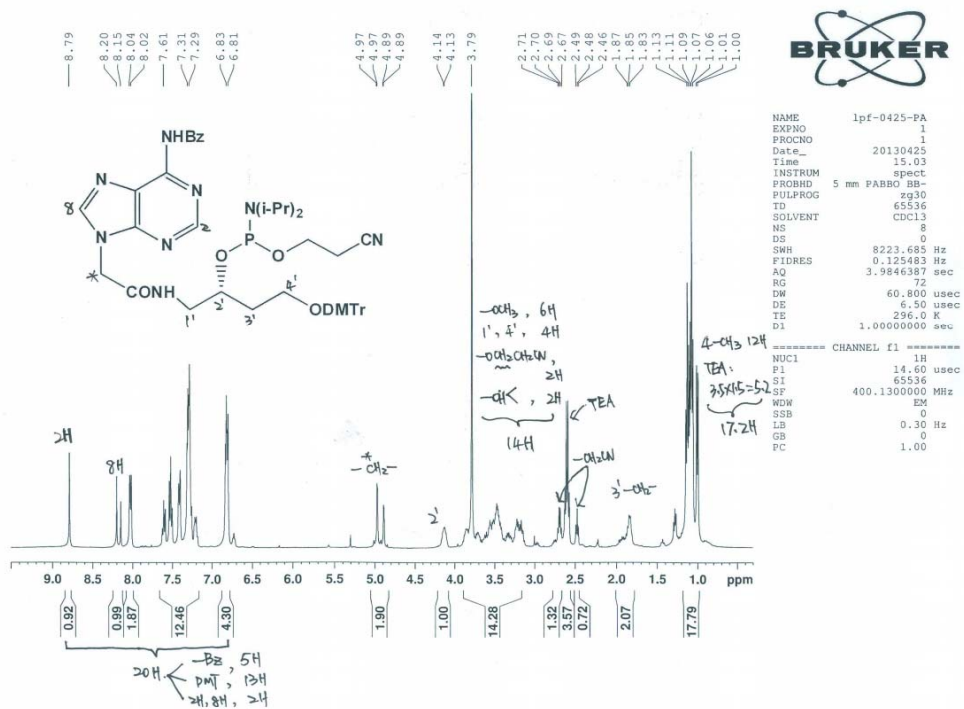


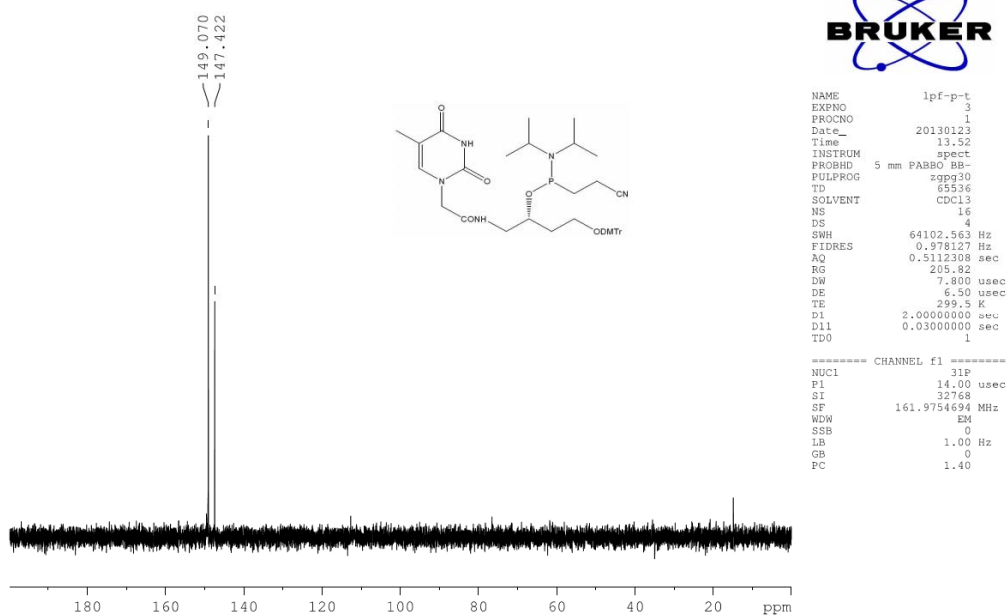
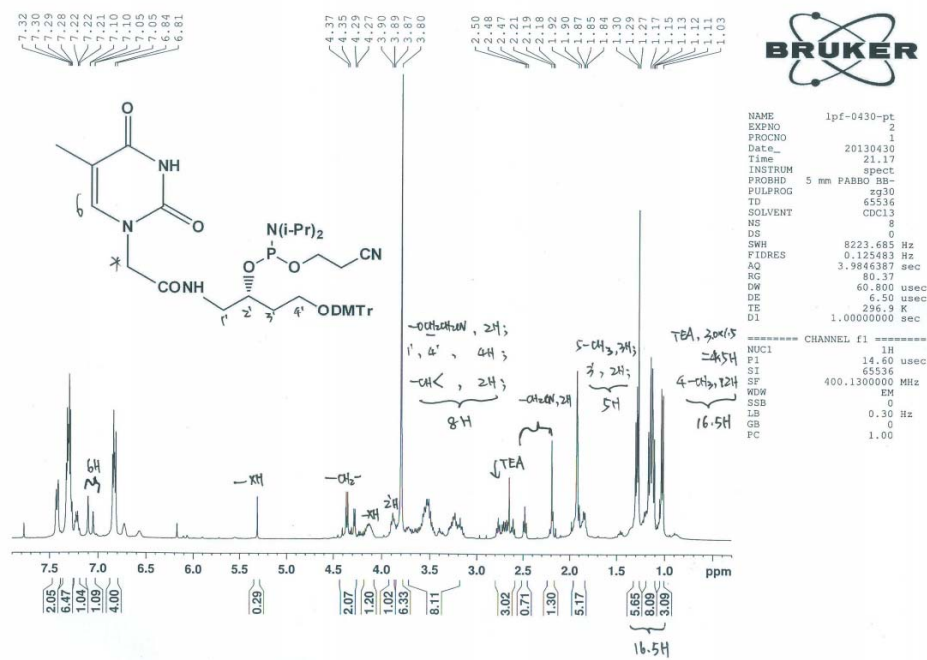


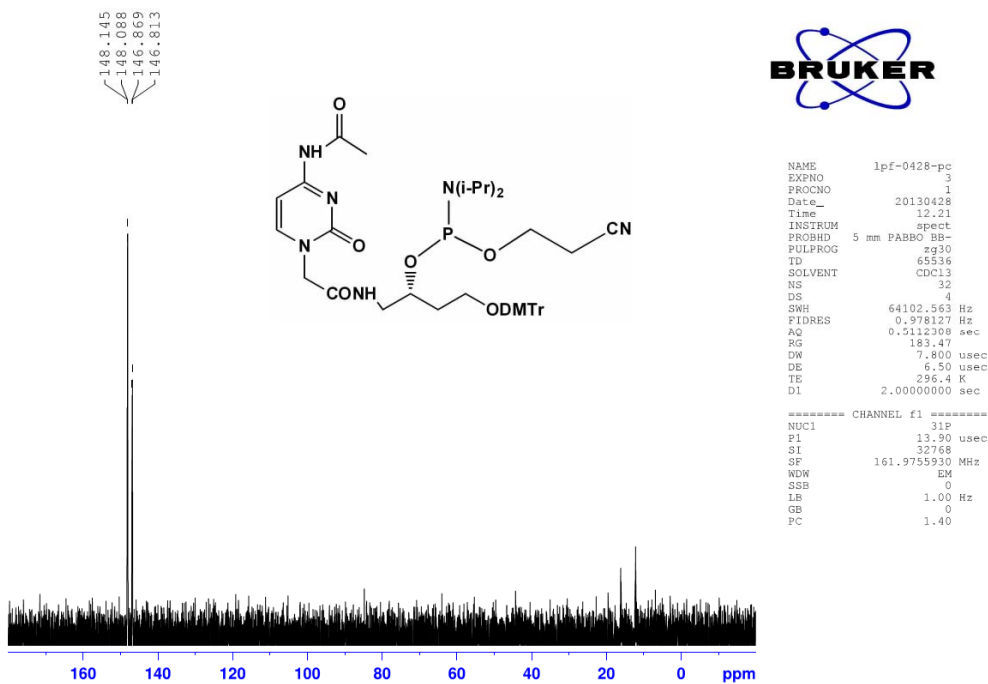
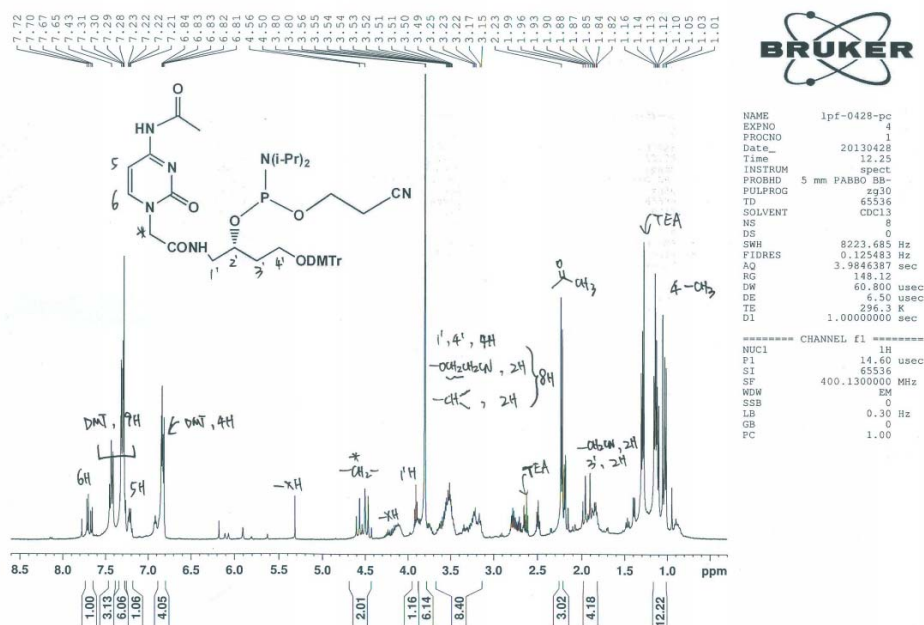


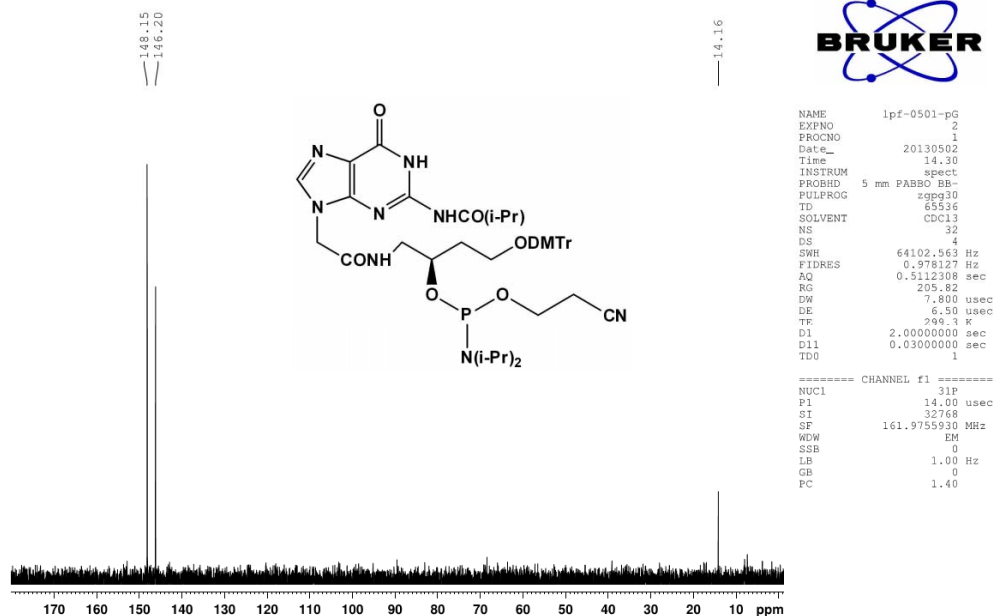
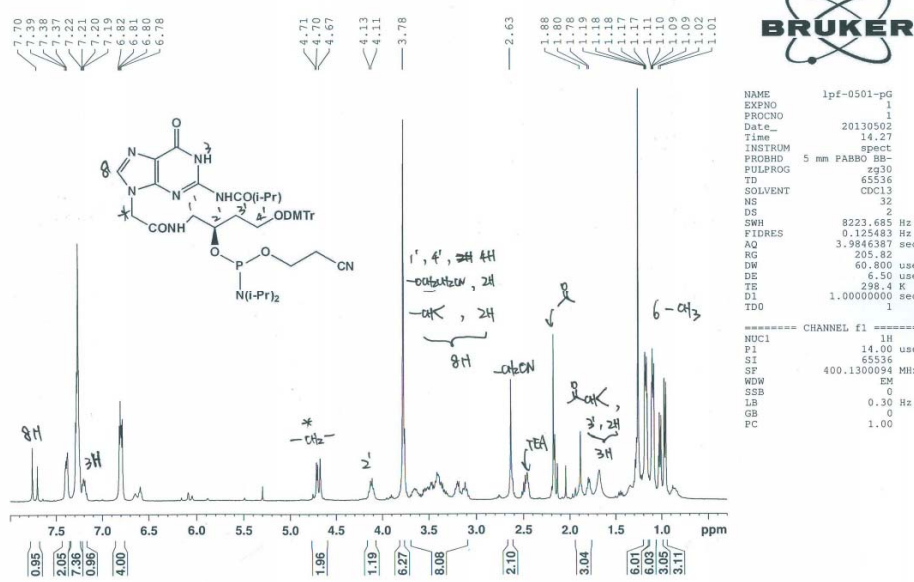


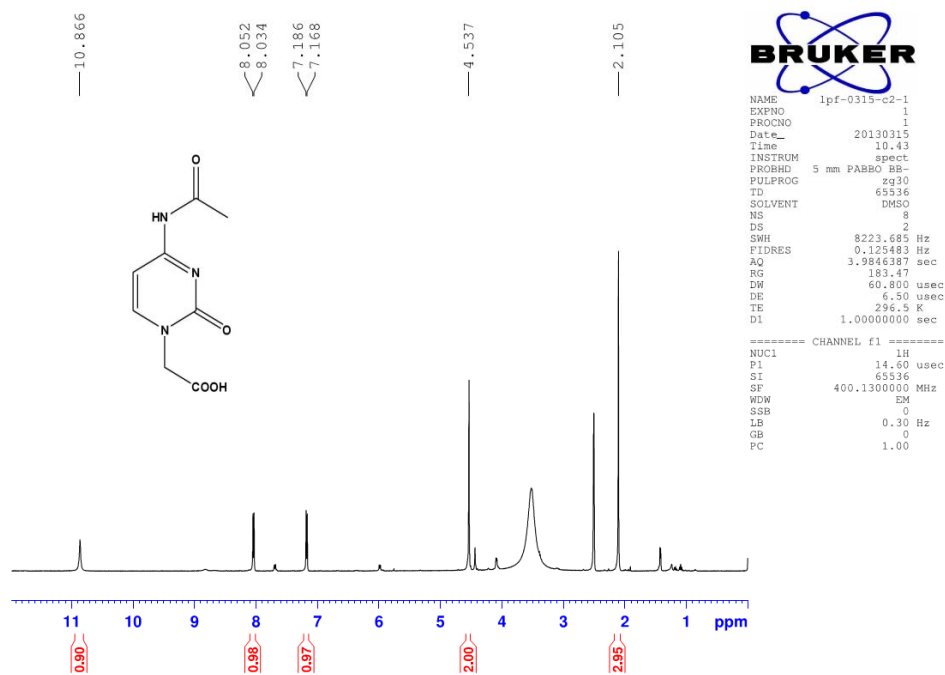
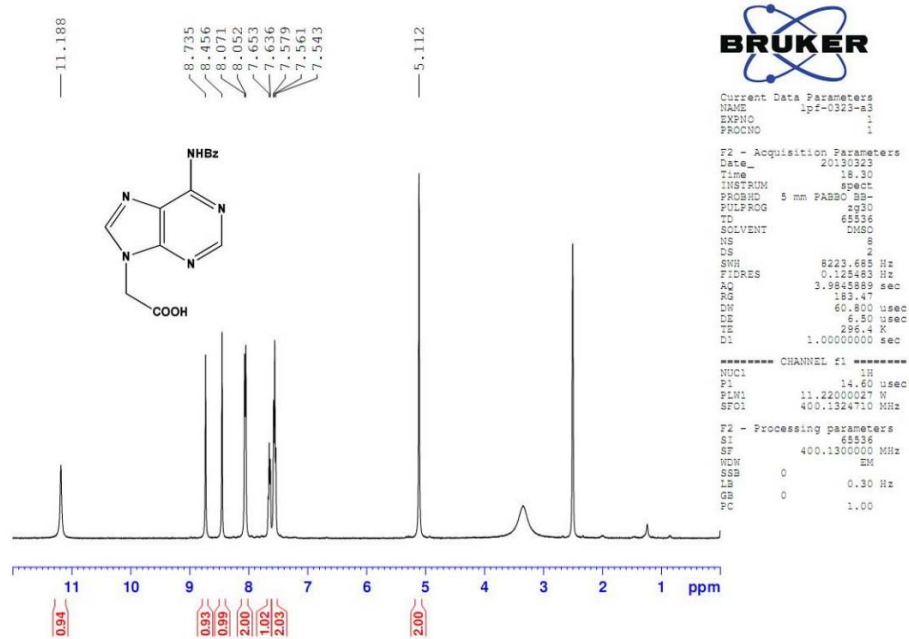


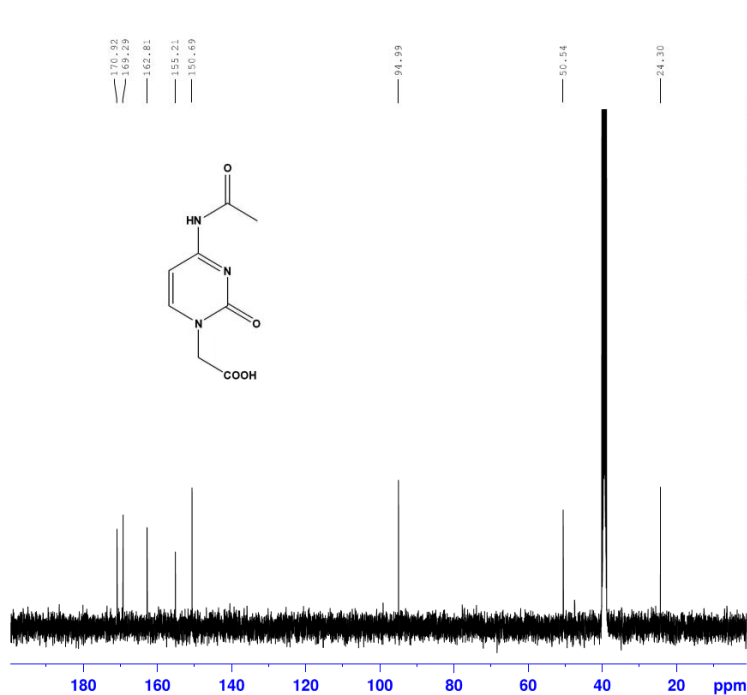












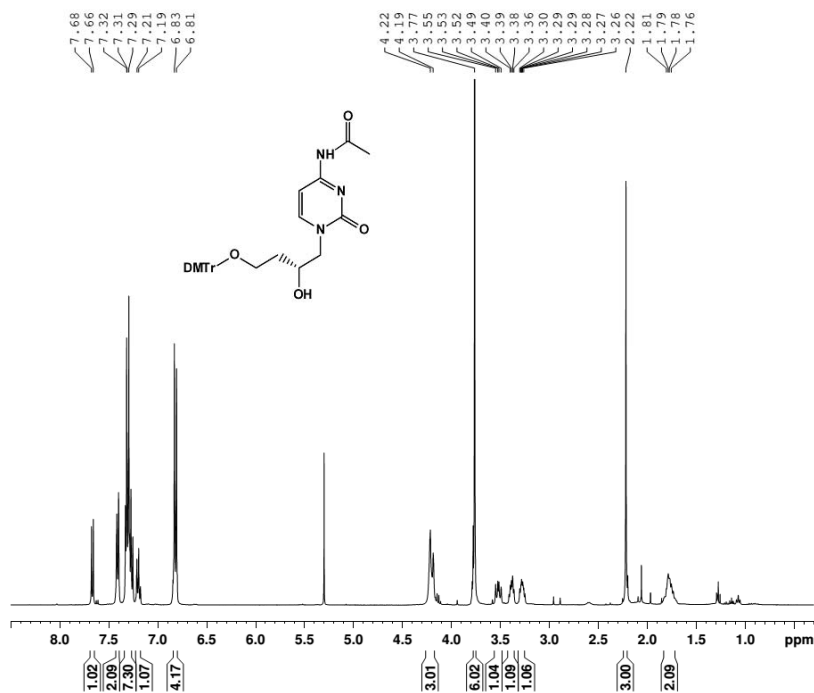
```

NAME      1pf-0315-c2-2
EXPNO    2
PROCNO   1
Date_    20130315
Time     22.23
INSTRUM  spect
PROBHD   5 mm FAPBO BB-
PULPROG  zgpg30
TD       65536
SOLVENT  DMSO
NS       502
DS       4
SWH      26041.666 Hz
FIDRES   0.397364 Hz
AQ       1.2583412 sec
RG       205.82
DW       19.203 usec
DE       6.50 usec
TE       298.7 K
D1       1.2500000 sec
D11      0.0300000 sec
TDO      1
    
```

==== CHANNEL f1 =====

```

NUC1      13C
P1        9.00 usec
SI        32768
SF        100.6128193 MHz
WDW       EM
SSB       0
LB        1.00 Hz
GB        0
PC        1.40
    
```



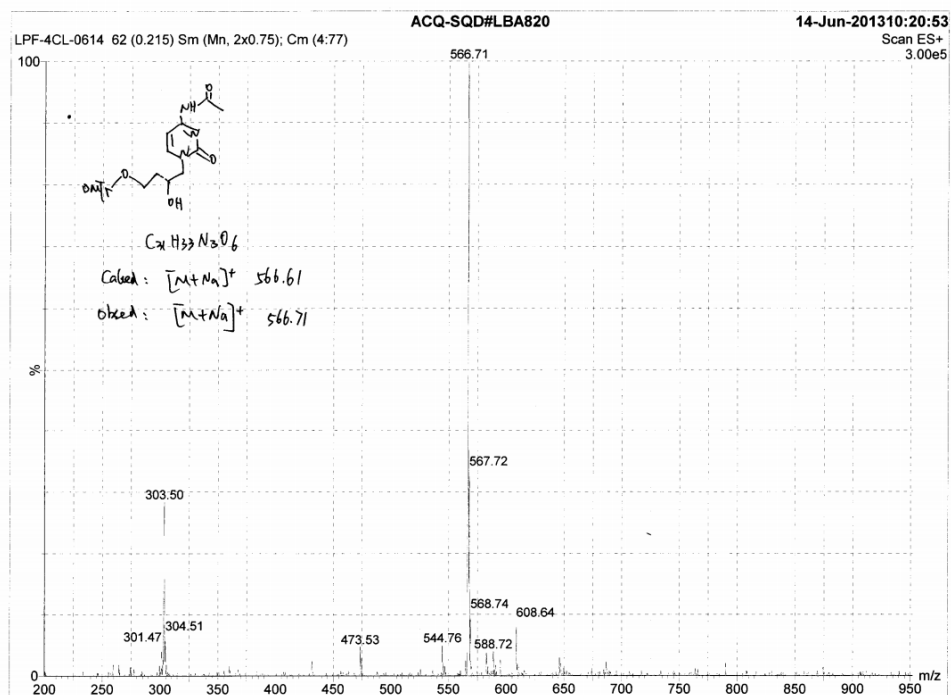
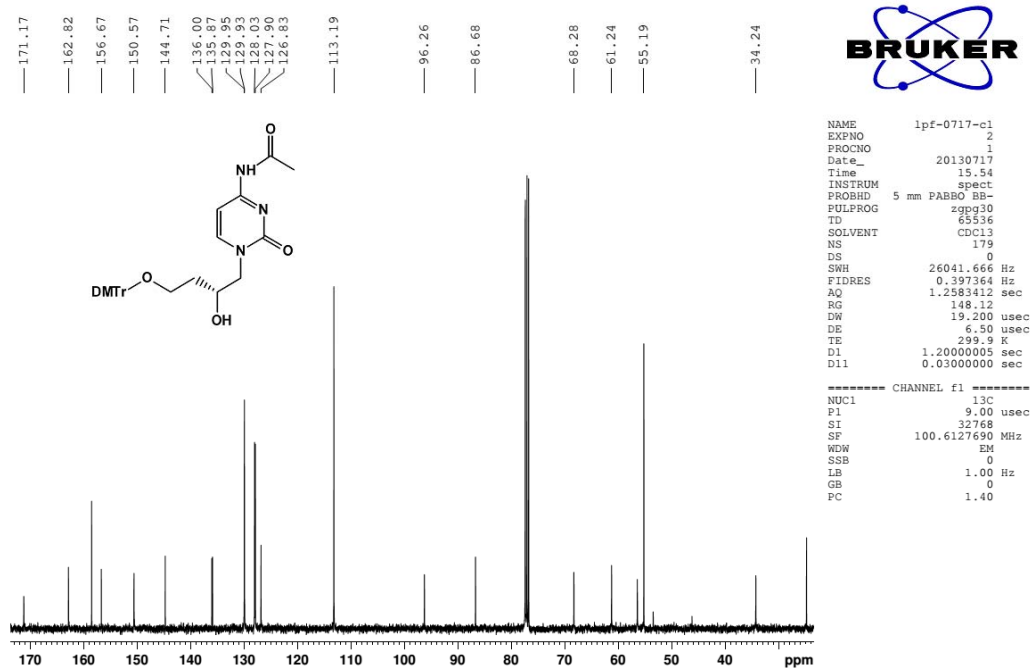
```

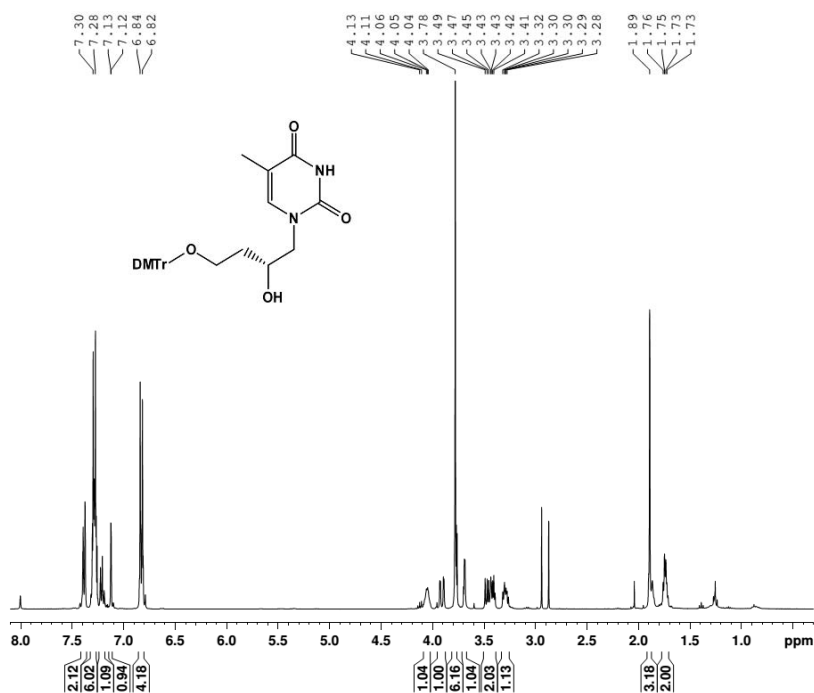
NAME      1pf-0717-c1
EXPNO    1
PROCNO   1
Date_    20130717
Time     15.51
INSTRUM  spect
PROBHD   5 mm FAPBO BB-
PULPROG  zg30
TD       65536
SOLVENT  CDCl3
NS       8
DS       0
SWH      8223.685 Hz
FIDRES   0.125483 Hz
AQ       3.9846387 sec
RG       64.88
DW       60.800 usec
DE       6.50 usec
TE       299.1 K
D1       1.0000000 sec
    
```

==== CHANNEL f1 =====

```

NUC1      1H
P1        14.60 usec
SI        65536
SF        400.1300000 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        1.00
    
```

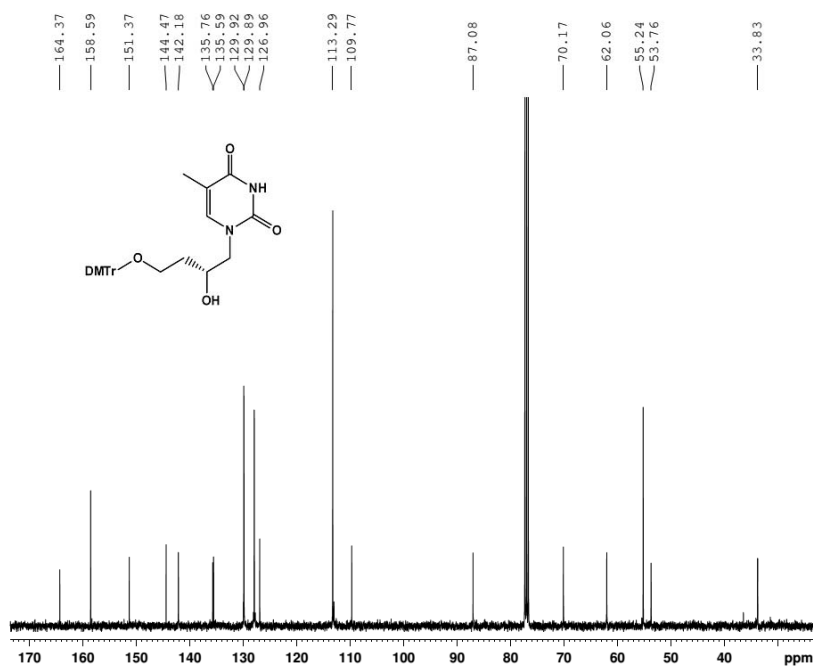




```

NAME      LPF-0912-T1
EXPNO    1
PROCNO   1
Date_    20130913
Time     9.03
INSTRUM  spect
PROBHD   5 mm PABBO BB-
PULPROG  zg30
TD       65536
SOLVENT  CDCl3
NS       16
DS       2
SWH      8223.685 Hz
FIDRES   0.125483 Hz
AQ       3.9846387 sec
RG       89.6
DW       60.800 usec
DE       6.50 usec
TE       297.4 K
D1       1.00000000 sec
TD0      1

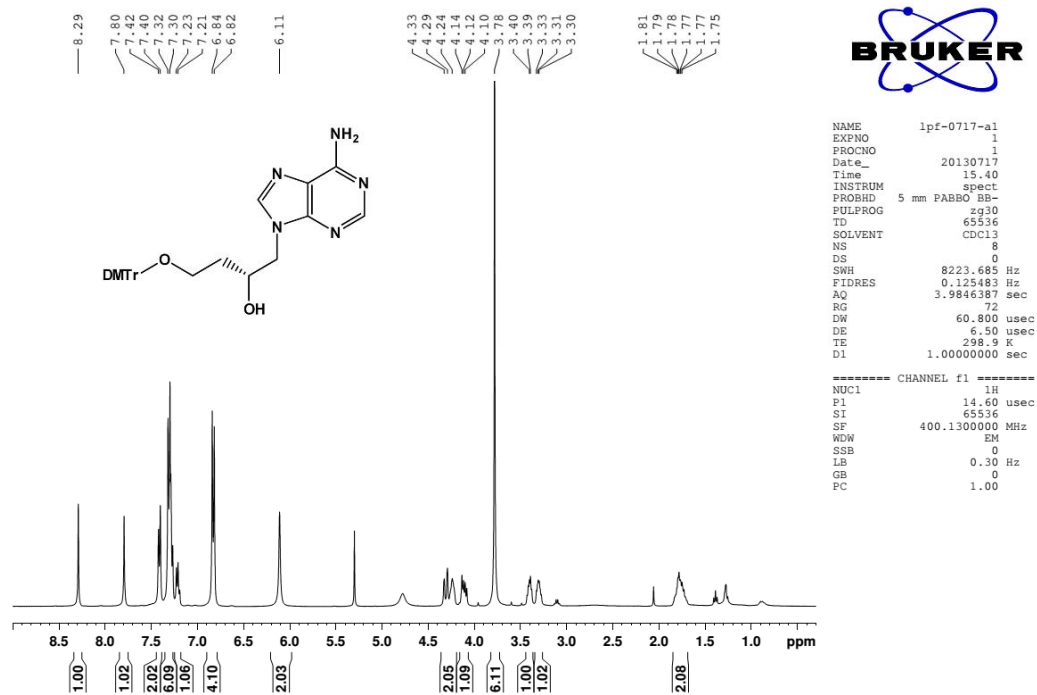
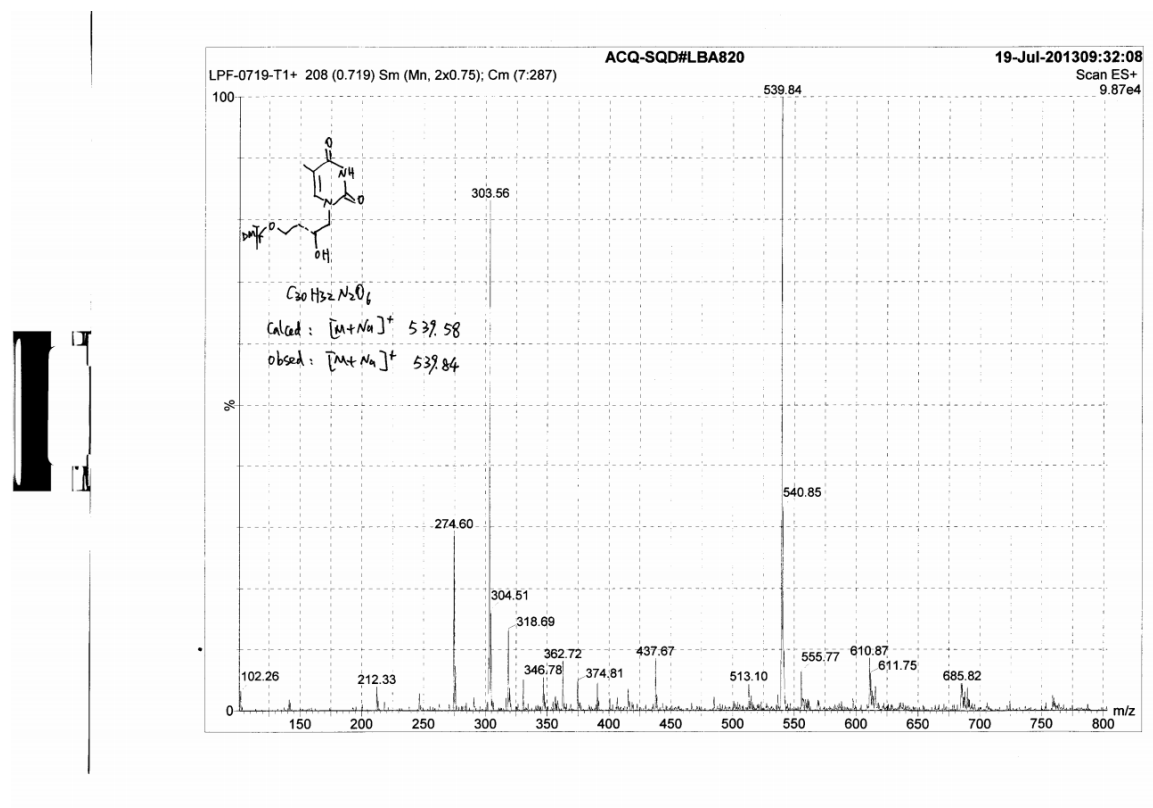
----- CHANNEL f1 -----
NUC1     1H
P1       14.00 usec
SI       65536
SF       400.1300104 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.00
    
```

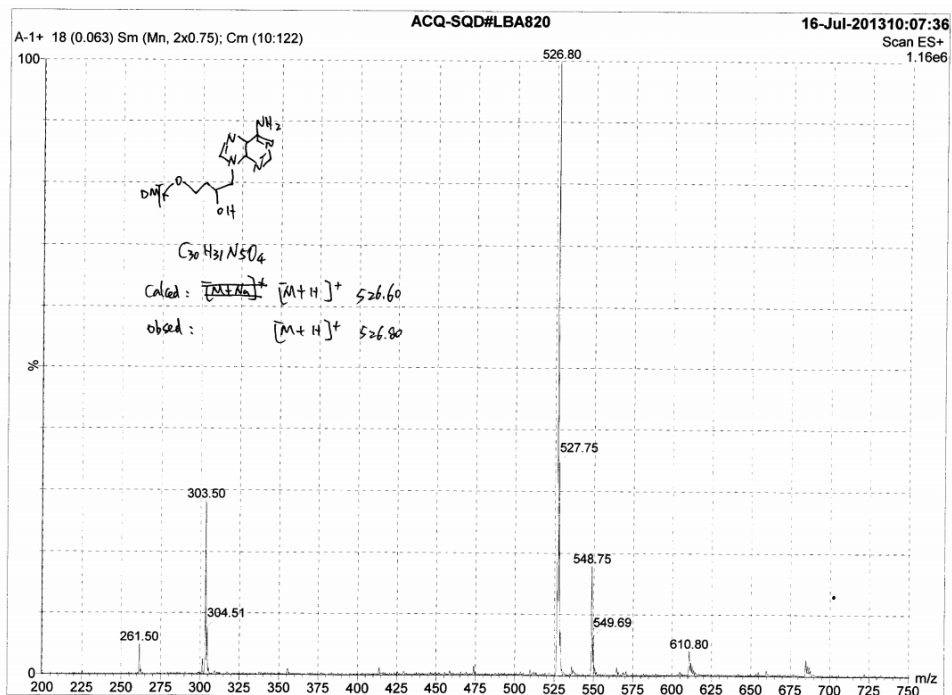
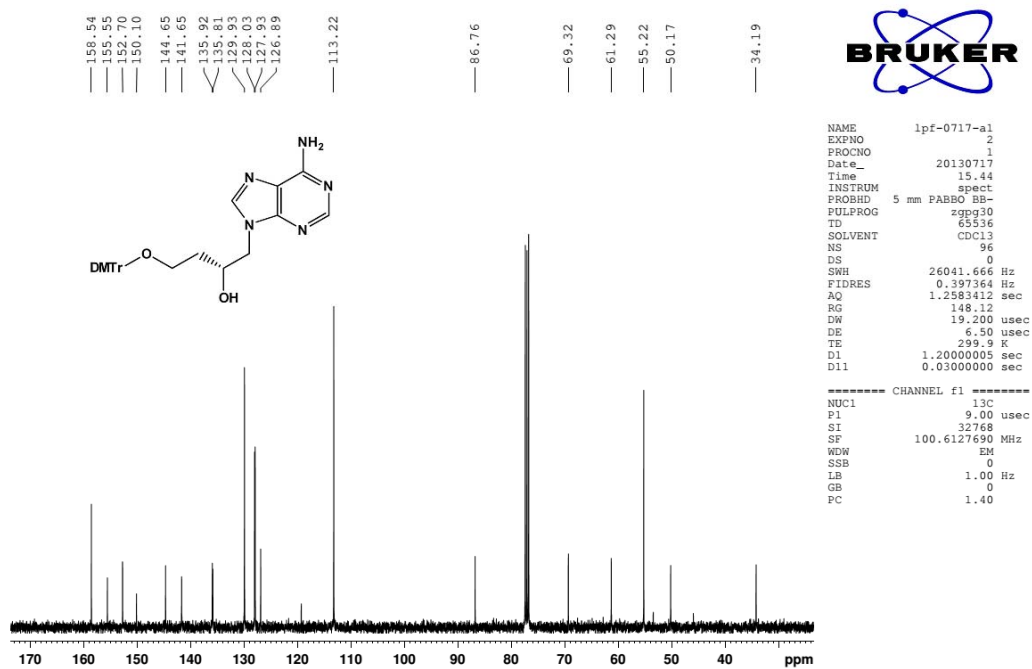


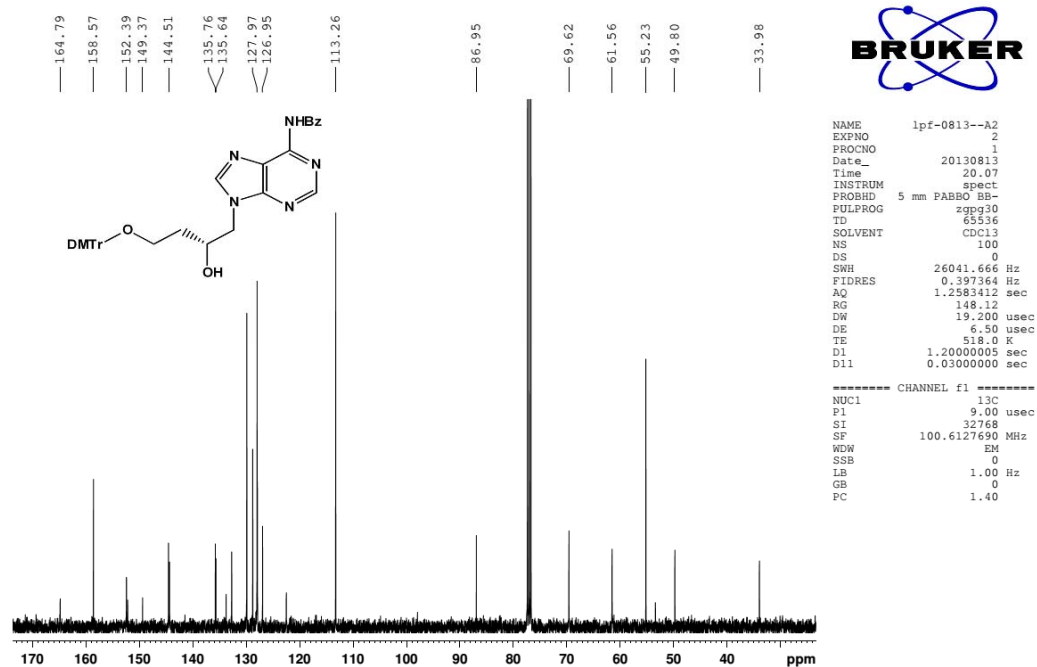
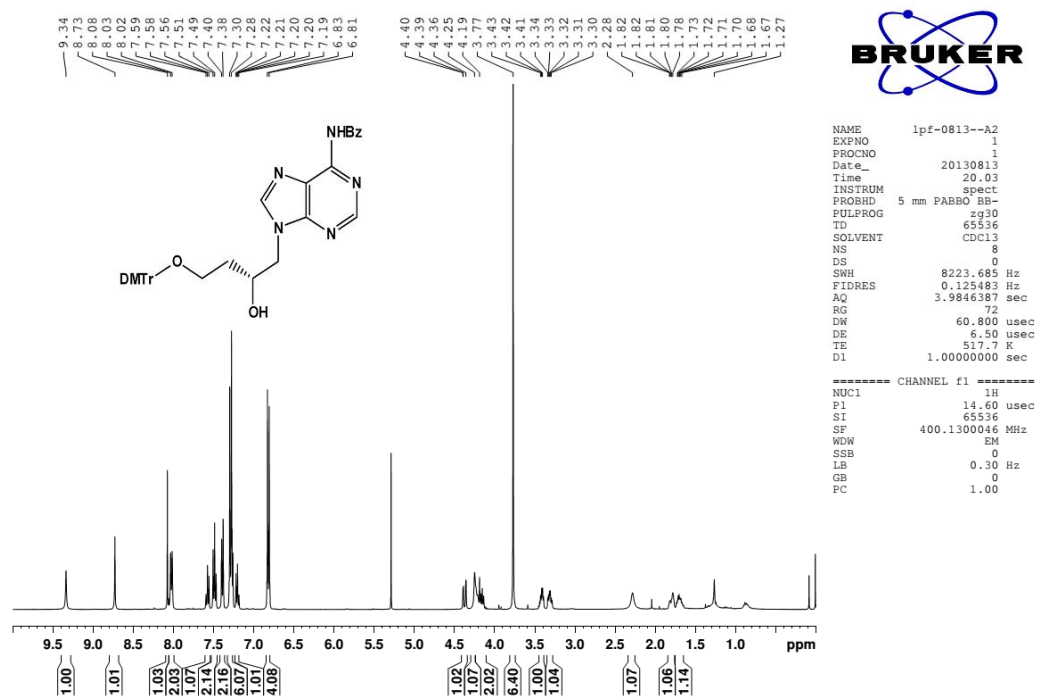
```

NAME      LPF-0912-T1
EXPNO    2
PROCNO   1
Date_    20130913
Time     9.04
INSTRUM  spect
PROBHD   5 mm PABBO BB-
PULPROG  zgpg30
TD       65536
SOLVENT  CDCl3
NS       302
DS       4
SWH      26041.666 Hz
FIDRES   0.397364 Hz
AQ       1.2583412 sec
RG       205.82
DW       19.200 usec
DE       6.50 usec
TE       298.0 K
D1       1.25000000 sec
D11      0.03000000 sec
TD0      1

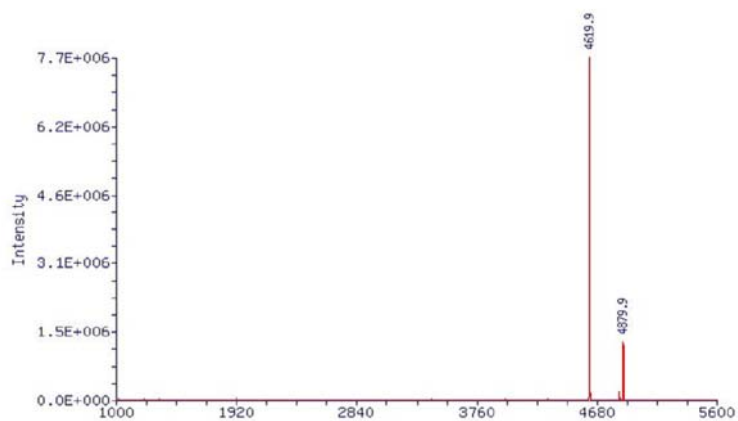
----- CHANNEL f1 -----
NUC1     13C
P1       9.00 usec
SI       32768
SF       100.6127690 MHz
WDW      EM
SSB      0
LB       1.00 Hz
GB       0
PC       1.40
    
```



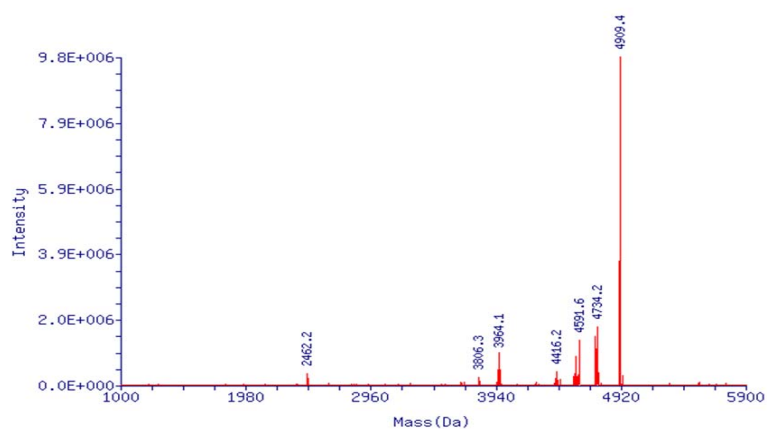




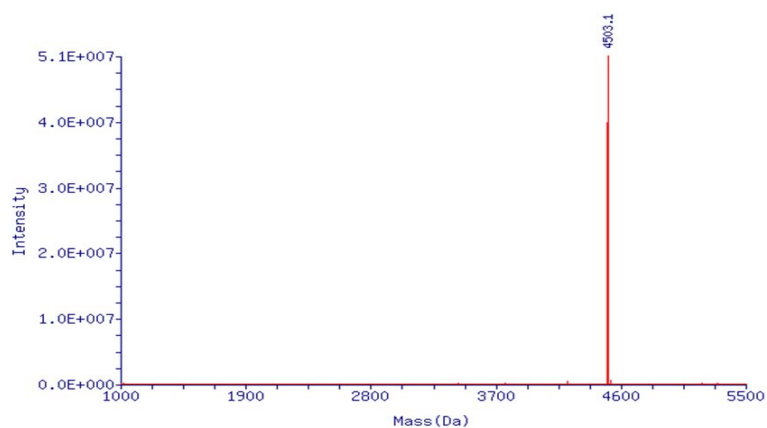
S6. SI-MS of oligonucleotides in our work.



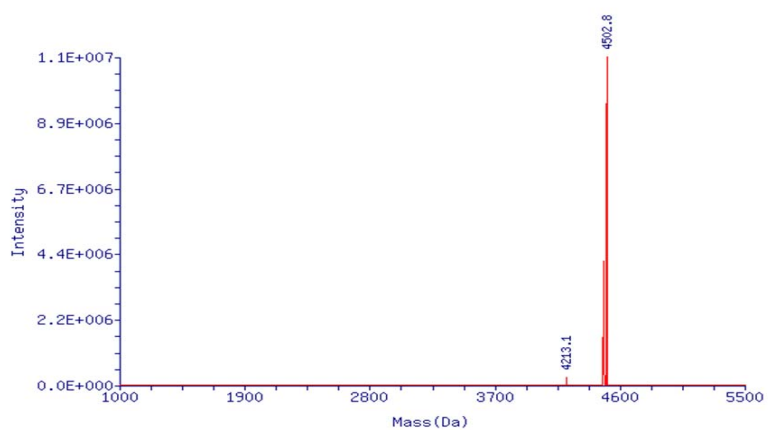
mmp-1



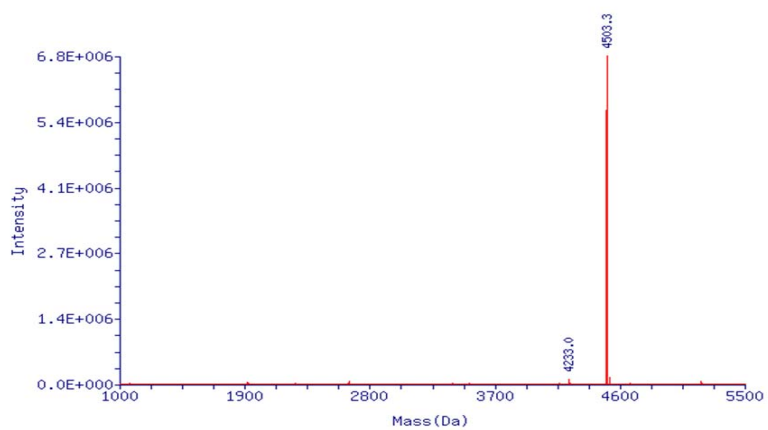
mmp-2



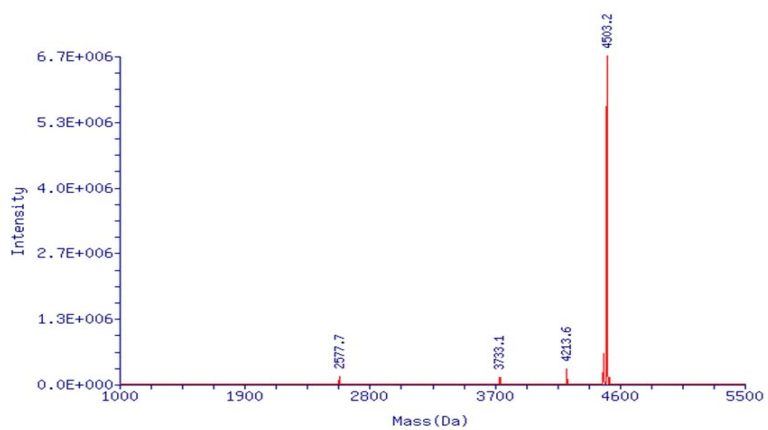
mmp-3



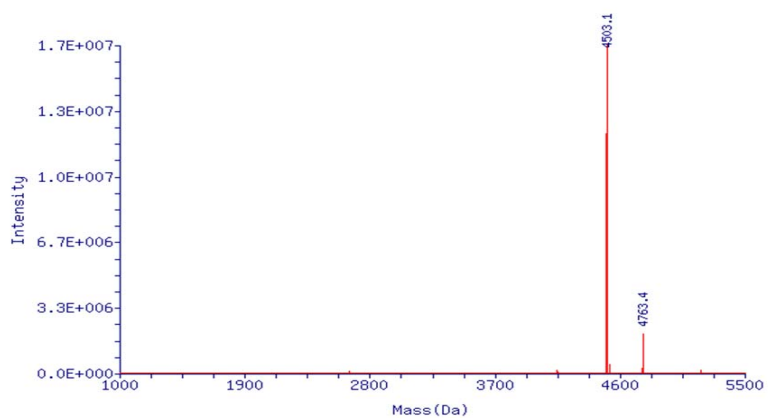
mmp-4



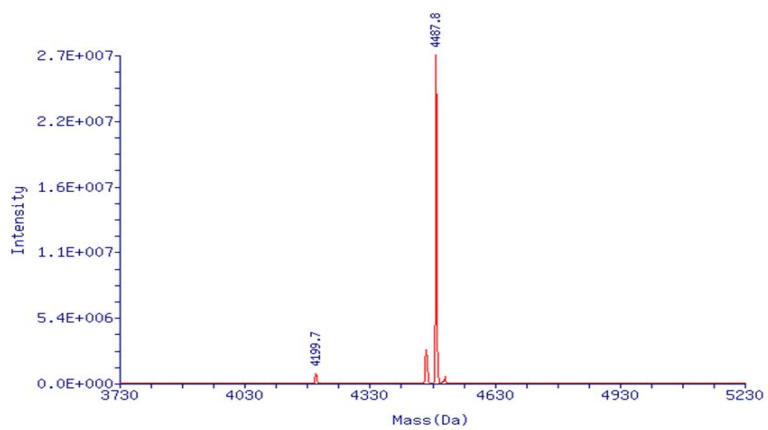
mmp-5



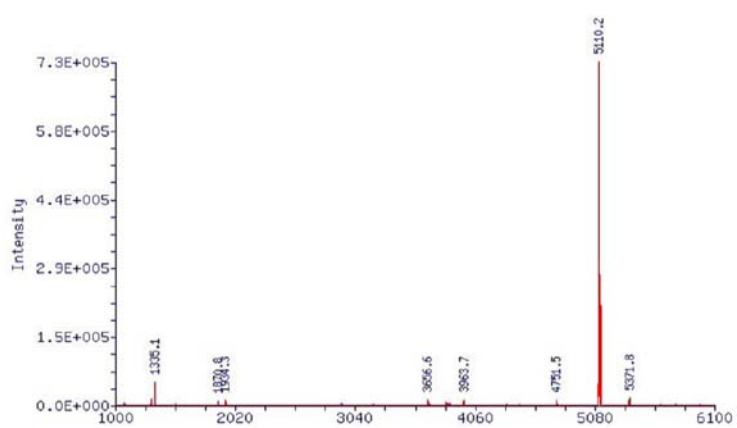
mmp--6



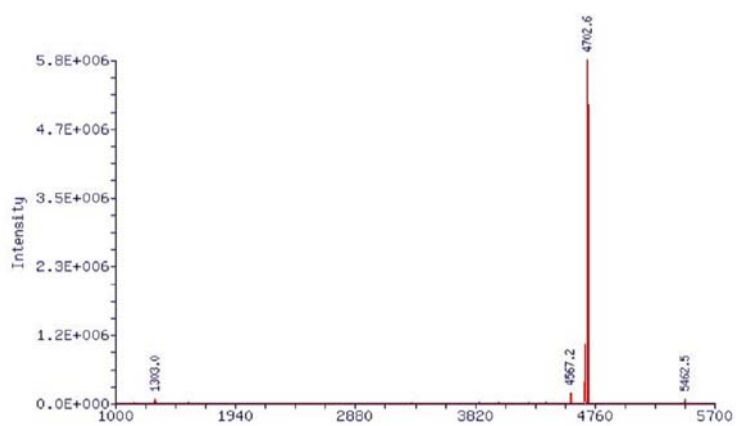
mmp-7



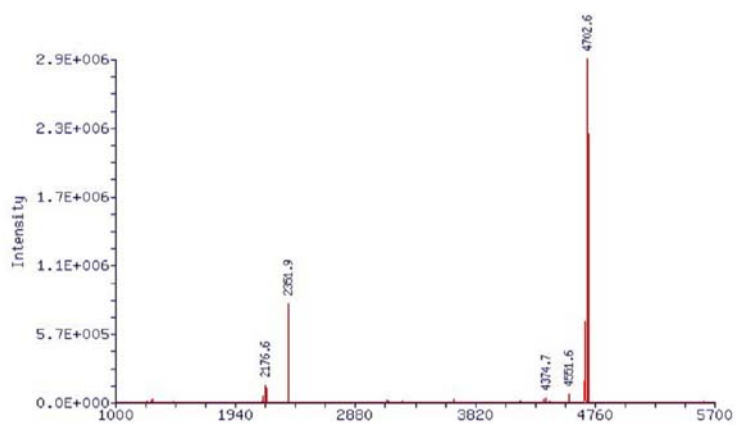
mmp-8



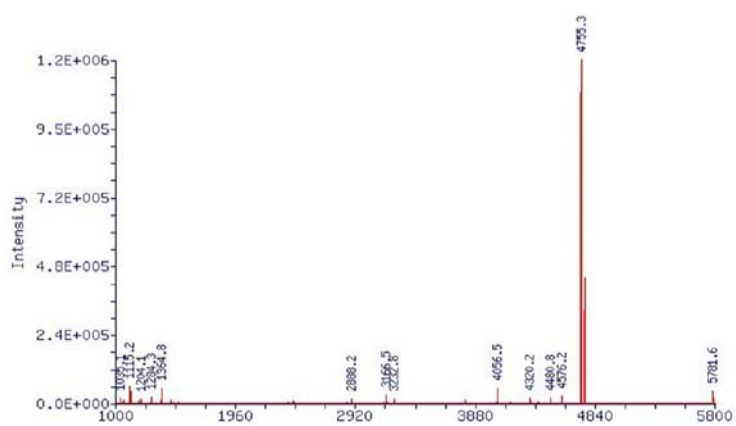
mmpsn-2



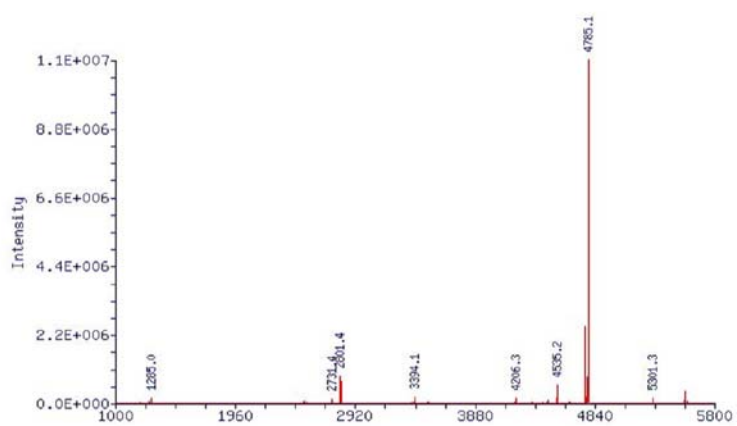
mmpsn-4



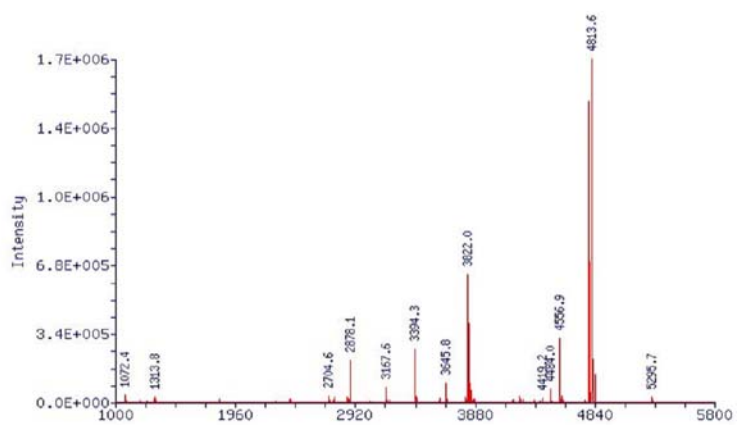
mmpsn-5



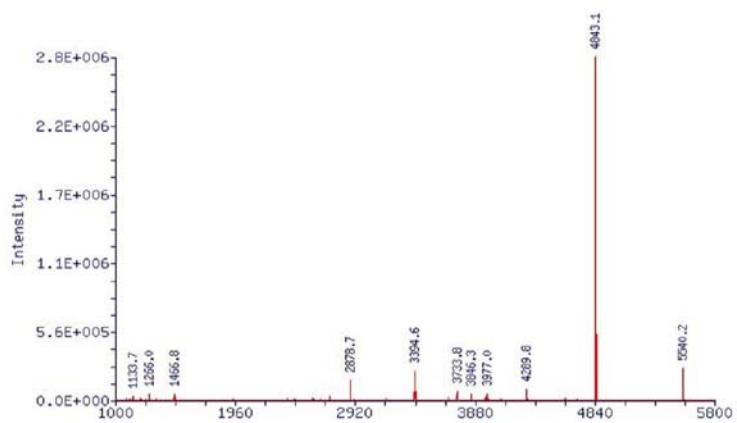
TBA-1



TBA-2



TBA-3



TBA-4

1. (a)Liu, Z.-C.; Shin, D.-S.; Lee, K.-T.; Jun, B.-H.; Kim, Y.-K.; Lee, Y.-S., *Tetrahedron*, 2005, **61**, 7967-7973;(b)Dueholm, K. L.; Egholm, M.; Behrens, C.; Christensen, L.; Hansen, H. F.; Vulpus, T.; Petersen, K. H.; Berg, R. H.; Nielsen, P. E.; Buchardt, O., *J. Org. Chem.* 1994, **59**, 5767-5773;(c)Geen, G. R.; Grinter, T. J.; Kinsey, P. M.; Jarvest, R. L., *Tetrahedron* **1990**, **46**, 6903-6914. (d)Will, D. W.; Breipohl, G.; Langner, D.; Knolle, J.; Uhlmann, E., *Tetrahedron* **1995**, **51**, 12069-12082.