

Non-stoichiometric O-acetylation of polysaccharide antigens: convergent synthesis and antibody recognition of acetylated *Shigella flexneri* 2a decasaccharides

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SUPPORTING INFORMATION

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Relevant NMR spectra (^1H , DEPT, HSQC, ^{13}C , including HSQC for $^1J_{\text{C},\text{H}}$ in the case of compound **2**) for new compounds by order of appearance:

S3, S4, 12, 13, 31, S8, 32, S9, 33, 8, 25, S6, 26, S7, 11, 16, 17, 18, 19, 20, 21, 22, 10, 21a, 21b, 27, 29, 7, 34, 35, 23, 24, 6, 5, 36, 38, 39, 40, 41, 1, 2, 3.

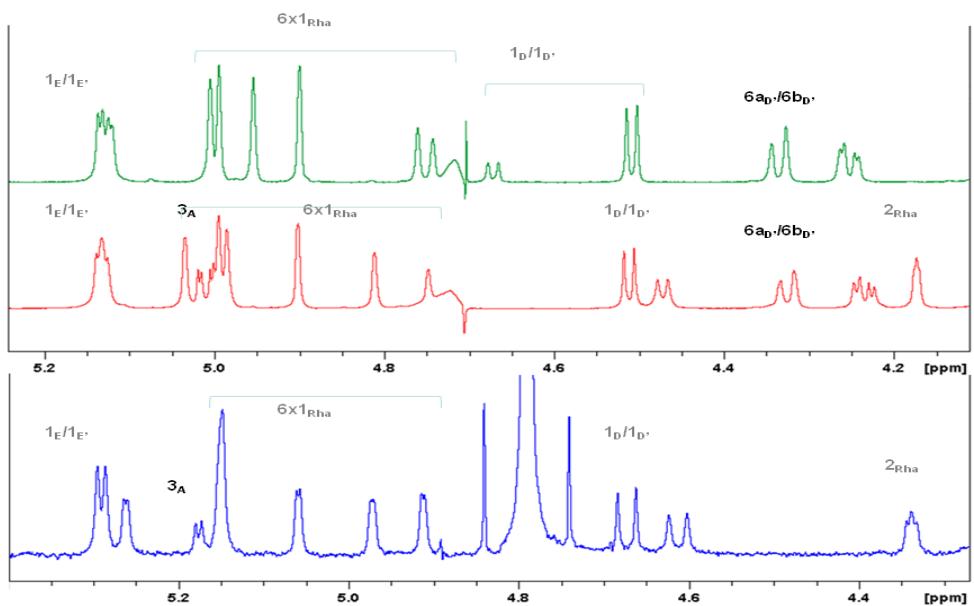
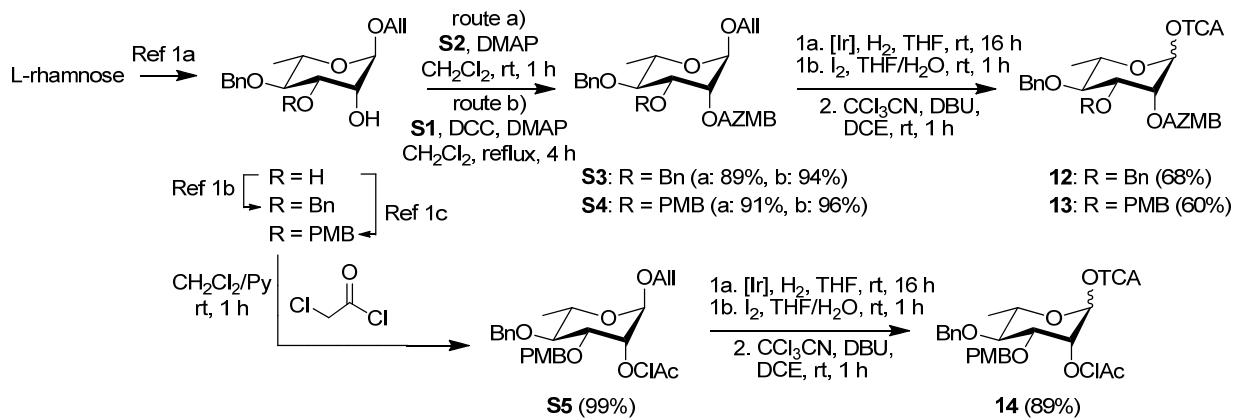
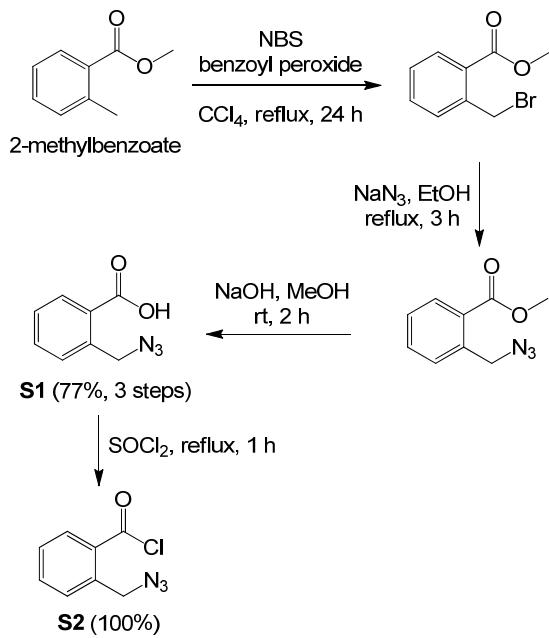


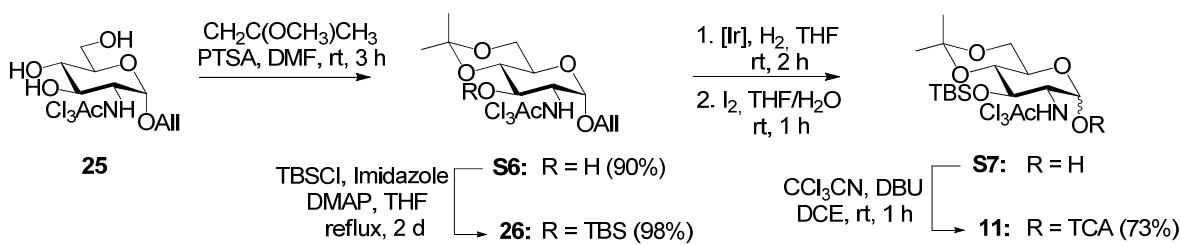
Figure S1. Anomeric region of the ^1H NMR spectra (Bruker AVII700 spectrometer equipped with a 5 mm $^1\text{H}(^{13}\text{C}/^{15}\text{N})$ inverse TCI cryoprobe optimized for ^1H observation, 700 MHz, D_2O , 303 K, water suppression) of decasaccharides **2** (red) and **3** (green) to that (Bruker Avance spectrometer equipped with a 5 mm BBO probe, 400 MHz, D_2O , 303 K) of decasaccharide **1** (Blue). The protons geminal to the acetylated hydroxyls are indicated.



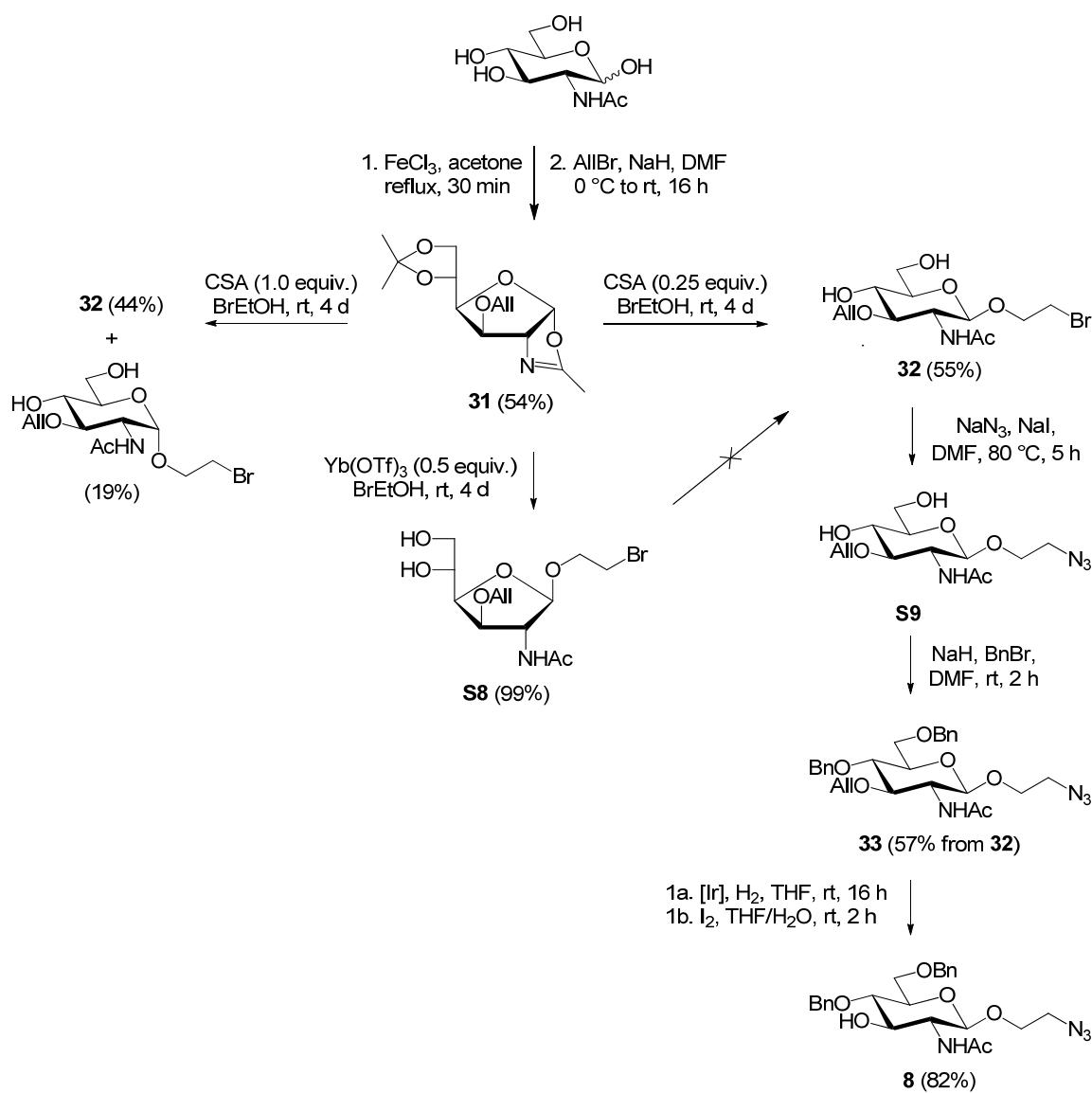
Scheme S1. Synthesis of L-rhamnosyl trichloroacetimidate donors (**12-14**).¹⁻³
[Ir]: Ir(COD){PCH₃(C₆H₅)₂}₂⁺.PF₆⁻



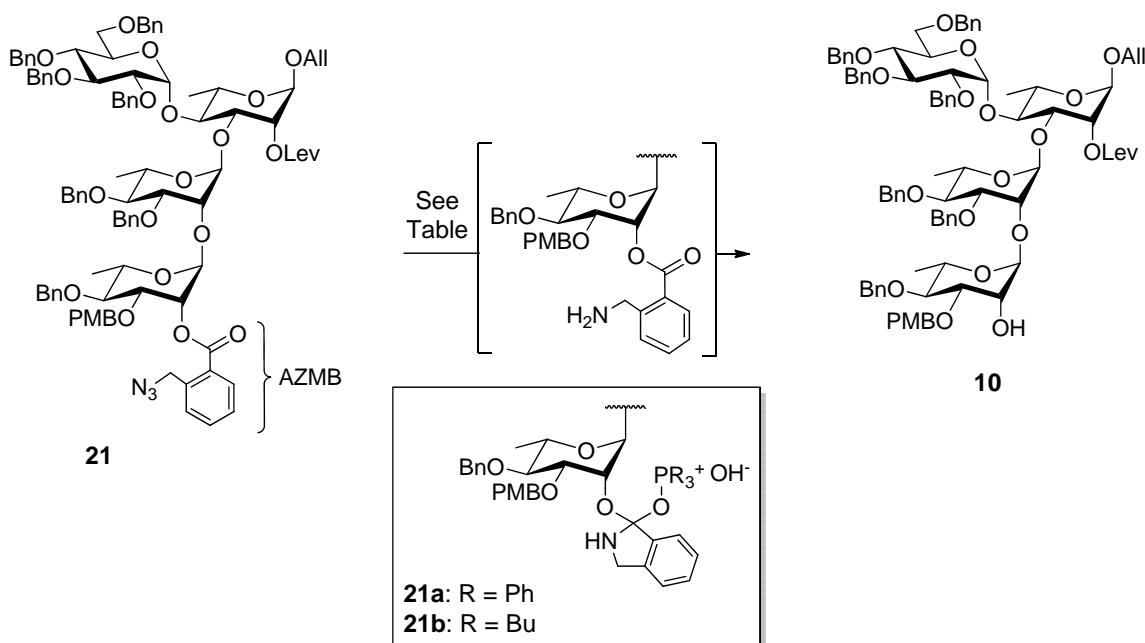
Scheme S2. Synthesis of AZMBOH (**S1**) and AZMBCl (**S2**) reagents inspired from a combination of known protocols.^{4, 5}



Scheme S3. Synthesis of the D-glucosaminyl donor **11** showing all isolated intermediates.



Scheme S4. Synthesis of *N*-acetyl-D-glucosamine acceptor **8** via acid-catalyzed oxazoline-opening. [Ir]: $\text{Ir}(\text{COD})\{\text{PCH}_3(\text{C}_6\text{H}_5)_2\}_2^+\text{PF}_6^-$



Scheme S5. Side products observed during attempted AZMB removal in compound **21** using Staudinger conditions.

Table S1. Attempts to remove the AZMB group on tetrasaccharide **21**.

Entry	Reagents (equiv.)	Solvent	Temp. Time	10 Yield (%) ^a
1 ⁵	PBu ₃ (3.0) H ₂ O (5.0)	THF	rt 4 h	47% 53% ^b
2 ⁶	PBu ₃ (3.0) then H ₂ O (5.0)	THF	rt 2 h	trace ^c
3 ⁷	PPh ₃ (10.) SiO ₂	THF/H ₂ O 9:1	rt 2 d	61% 66% ^b
4	PPh ₃ (10) SiO ₂	THF/H ₂ O 9:1	reflux 6 h	trace ^c
5 ⁸	H ₂ S	Py/H ₂ O 4:1	reflux 24 h	trace ^d
6	H ₂ S	Py/H ₂ O 4:1	140 °C (μW) ^[e] 4 h	44% ^d
7 ⁹	HS(CH ₂) ₃ SH (50) Et ₃ N (10)	CH ₂ Cl ₂ /MeOH 1:1	reflux 2 d	degradation
8 ⁷	NaBH ₄ (3.0) NiCl ₂ .6H ₂ O (0.1)	CH ₂ Cl ₂ /EtOH 1:1	rt 1 h	27%

^a Isolated yield. ^b Based on recovery starting material. ^c The phosphonium derivative (**21a** or **21b**) was the major product. These polar side-products were unambiguously identified from ¹H, ¹³C and ³¹P NMR analysis as well as by HR-ESI-TOF-MS (**21a**: δ (³¹P NMR, CDCl₃) 40.1 ppm (s, OPPh₃), m/z 1900.8340 [M]⁺, calcd for C₁₁₅H₁₂₃NO₂₂P, 1900.8274; **21b**: m/z 1840.9080 [M]⁺, calcd for C₁₀₉H₁₃₅NO₂₂P, 1840.9208). ^d H₂S was bubbled for 1 h through the solution before heating. ^e μW: microwave.

Materials and Methods

Antigenicity analysis

Panel of protective anti-SF2a mIgGs: The protective mIgGs specific for the SF2a O-Ag used in this study – A2-1, D15-7, C1-7, E4-1, F22-4 – were described previously.¹⁰

Panel of synthetic decasaccharides: Decasaccharide **4** was described previously.¹¹ For each one of the four decasaccharides (**1-4**), the concentration of the mother solution was measured by a method adapted from amino acid analysis. The method allows hexosamine quantification¹² and was found appropriate to overcome the inherent difficulties associated in accurately measuring small amounts of low molecular weight antigens.¹³ Briefly, the amino sugar compositions of the decasaccharide mother solutions ready for antigenicity analysis were determined with a L-8800 Hitachi automatic amino acid analyzer (classical post column derivatization with ninhydrin after ion-exchange chromatography separation), after hydrolysis in 1% phenol 6N HCl in glass tubes at 95 °C for 16 h, and evaporation of the volatiles.

Inhibition ELISA for IC₅₀ measurement: The binding of the available mIgGs to decasaccharides **1-4** was measured according to a known protocol.¹⁴ The mIgG concentration to be used was defined in the first step. To do so, a standard curve was established for each mAb. The mIgGs were incubated at different concentrations, overnight at 4 °C, on microtiter plates coated with purified SF2a LPS at a concentration of 2.5 µg/mL in a carbonate buffer (pH 9.6), then with PBS-BSA 1% for 30 min at 4°C. After washing with PBS-Tween 20 (0.05%), alkaline phosphatase-conjugated anti-mouse IgG was added at a dilution of 1/5,000 (Sigma-Aldrich) for 1 h at 37 °C. After washing with PBS-Tween 20 (0.05%), the substrate was added (12 mg of *p*-nitrophenylphosphate in 1.2 mL of 1 M Tris-HCl buffer (pH 8.8) and 10.8 mL of 5 M NaCl). Once the color developed, the plate was read at 405 nm Dynatech MR400 microplate reader). A standard curve $OD = f([Ab])$ was fitted to the quadratic equation $Y = aX^2 + bX + c$, where Y is the OD and X is the Ab concentration. A correlation factor (r^2) of 0.99 was routinely obtained.

The IC₅₀, defined as the concentration of oligosaccharides required to inhibit 50% of mIgG binding to LPS, were measured in a second step. Each mIgG at a given concentration (chosen as the minimal concentration of mAb which gives the maximal OD on the standard curve mentioned above) was incubated overnight at 4 °C with each of decasaccharides **1-4** at various concentrations in PBS-BSA 1%. The maximum concentration tested was 0.5 mM for all decasaccharides. Measurement of unbound mIgG was performed as described above using microtiter plates coated with purified SF2a LPS. The mAb concentration was deduced from the standard curve.

NMR analysis (Fig. 2)

The ^1H NMR spectra of decasaccharide **4** and pentadecasaccharide [AB(E)CD]₃ shown in Fig. 2 were run on a Varian Unity Inova 600 MHz spectrometer equipped with a cryogenically-cooled triple resonance $^1\text{H}\{^{13}\text{C}/^{15}\text{N}\}$ PFG probe (D_2O , 308 K).

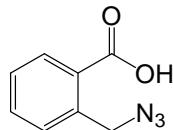
Synthesis: General methods.

Chemical reagents were used as received. Air and water sensitive reactions were performed in dried glassware under Ar atmosphere. Moisture sensitive reagents were introduced via a dry syringe. Anhydr. toluene (Tol), diethyl ether (Et_2O), dichloromethane (CH_2Cl_2), 1,2-dichloroethane (DCE), tetrahydrofuran (THF), *N,N*-dimethylformamide (DMF), acetonitrile (CH_3CN), ethanol (EtOH), methanol (MeOH) and pyridine (Py) were supplied on MS and were used as received. Additional solvents commonly cited in the text are abbreviated as Chex (cyclohexane), DMSO (dimethyl sulfoxide) and CCl_4 (carbon tetrachloride). Powered 4 Å MS (MS) and powered 4 Å acid-washed molecular sieves (AW-MS) were activated before use by heating at ≥ 250 °C under vacuum. Analytical thin-layer chromatography was performed with silica gel 60 F₂₅₄, 0.25 mm pre-coated TLC plates. Compounds were visualized using UV₂₅₄ and/or orcinol (1 mg·mL⁻¹) in 10% aq. H_2SO_4 with charring. Flash column chromatography was carried out using silica gel (particle size 0.040–0.063 mm). Nuclear magnetic resonance (NMR) spectra of all intermediates were recorded at 303 K on a Bruker Avance spectrometer equipped with a BBO probe at 400 MHz (^1H) and 100 MHz (^{13}C). NMR spectra of decasaccharides **2–3** were recorded at 303 K on a Bruker AVII700 equipped with a 5 mm $^1\text{H}(^{13}\text{C}/^{15}\text{N})$ inverse TCI cryoprobe optimised for ^1H observation and with a cold ^{13}C channel at 700 MHz (^1H) and 175 MHz (^{13}C). Elucidations of chemical structures were based on ^1H , COSY, DEPT-135, HSQC, ^{13}C , ^{13}C gated decoupling and HMBC experiments. Signals are reported as m (multiplet), s (singlet), d (doublet), t (triplet), dd (doublet of doublet), dq (doublet of quadruplet), ddd (doublet of doublet of doublet), ddt (doublet of doublet of triplet), br s (broad singlet), br d (broad doublet) and coupling constants are reported in hertz (Hz). The chemical shifts are reported in ppm (δ) relative to residual solvent peak. Of the two magnetically non-equivalent geminal protons at C-6, the one resonating at lower field is denoted H-6a, and the one at higher field is denoted H-6b. Interchangeable assignments are marked with an asterisk in the listing of signal assignments. Sugar residues are serially lettered according to the lettering of the repeating unit of the *S. flexneri* 2a O-Ag and identified by a subscript in the listing of signal assignments. The values of the $^1J_{\text{C},\text{H}}$ constants of the anomeric carbons/protons were measured from the HSQC for $^1J_{\text{C},\text{H}}$ spectrum of decasaccharide **2**, only. This ascertained that target **2** had the required structure. Since all novel decasaccharides described in the manuscript are issued from decasaccharide **5**, this result ensured that all residues pertaining to all synthesized new oligosaccharides have the correct anomery. High resolution electrospray ionisation/time-of-flight mass spectra (HR-ESI-TOF-MS) were recorded in the positive-ion mode with 1:1 $\text{CH}_3\text{CN}/\text{H}_2\text{O}$ containing 0.1% formic acid as the ESI-TOF spectrometer solution. High resolution matrix-assisted laser desorption-ionisation/time-of-flight mass spectra (HR-MALDI-TOF-MS) were recorded in the positive-ion mode. The former were measured at the UMR CNRS 3523, Institut Pasteur, Paris, France and the

later were obtained at the Institut de Chimie des Substances Naturelles, Gif sur Yvettes, France. Optical rotations were obtained using sodium D line at rt (22 °C) on a Bellingham + Stanley Ltd. ADP220 polarimeter. $[\alpha]_D$ values are given in $10^{-1} \text{ deg cm}^2 \text{ g}^{-1}$.

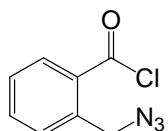
Synthesis: Experimental procedures

2-(Azidomethyl)benzoic acid⁴ (S1).



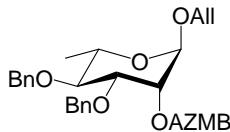
NBS (102 g, 572 mmol, 1.01 equiv.) and benzoyl peroxide (1.37 g, 6.66 mmol, 0.01 equiv) were added to methyl 2-methylbenzoate (85.0 g, 566 mmol) in anhydrous CCl_4 (1.0 L). CAUTION! This reaction is very exothermic, care must be taken in order to minimize the risks of runaway reaction. The mixture was stirred for 24 h at reflux under Ar. After cooling to 0 °C, the precipitate was removed by filtration. The organic phase was evaporated to dryness. NaN_3 (37.2 g, 572 mmol, 1.01 equiv.) was added to the residue in anhydrous EtOH (1.6 L) and the mixture was stirred for 3 h at reflux under Ar. Brine (250 mL) was added and the mixture was concentrated (to 1/3 volume) under reduced pressure. The aq. phase was extracted with CH_2Cl_2 (3×500 mL), the organic layer was dried over anhydrous Na_2SO_4 , filtered, and concentrated to dryness. NaOH (51.0 g) was added to the residue in MeOH/H₂O (1.2 L, 5:2 v/v) and. The mixture was stirred for 2 h at rt, and concentrated (to 1/3 volume) under reduced pressure. The aq. phase was extracted with CH_2Cl_2 (3×500 mL) and acidified with 6 N aq. HCl. The precipitated acid was then extracted with CH_2Cl_2 (3×250 mL), the organic phases were pooled and concentrated to dryness to give a yellow solid residue. The crude material was recrystallized from Chex to give S1 (77.3 g, 77%, 3 steps) as a white powder. Physical and analytical data were in agreement with those published.⁴

2-(Azidomethyl)benzoyl chloride⁵ (S2).



Compound S1 (7.00 g, 41.2 mmol) was dissolved in SOCl_2 (6.0 mL, 82 mmol, 2.0 equiv.) and the mixture was stirred at reflux for 1 h under Ar. Volatiles were evaporated under reduced pressure and co-evaporated with toluene to dryness to give crude acyl chloride S2 as a yellow oil, which was used for the next step without purification. Physical and analytical data were in agreement with those published.⁵

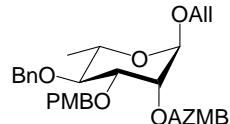
Allyl 2-*O*-(2-(azidomethyl)benzoyl)-3,4-di-*O*-benzyl- α -L-rhamnopyranoside (S3).



Route a. DMAP (3.18 g, 26.0 mmol, 2.0 equiv.) and acyl chloride **S2** (3.02 g, 26.0 mmol, 2.0 equiv.) were added to allyl 3,4-di-*O*-benzyl- α -L-rhamnopyranoside² (5.00 g, 13.0 mmol) in anhydrous CH₂Cl₂ (130 mL). The mixture was stirred for 1 h at rt under Ar. The organic phase was washed with satd aq. NaHCO₃ (2×50 mL) and brine (1×50 mL). The solution was dried over anhydrous Na₂SO₄, filtered and solvents were evaporated. The residue was purified by FC (Chex/EtOAc 95:5 to 9:1) to give allyl glycoside **S3** (6.78 g, 94%) as a colorless oil.

Route b. DMAP (10.9 g, 88.8 mmol, 1.0 equiv), DCC (36.7 g, 178 mmol, 2.0 equiv.) and benzoic acid **S1** (23.6 g, 133 mmol, 1.5 equiv) were added to allyl 3,4-di-*O*-benzyl- α -L-rhamnopyranoside² (34.2 g, 88.9 mmol) in anhydrous CH₂Cl₂ (910 mL). The mixture was stirred for 4 h at reflux under Ar, and cooled to rt. The mixture was filtered over a pad of Celite®, and the solvents were removed under reduced pressure. The residue was purified by FC (Chex/EtOAc 95:5 to 7:3) to give allyl glycoside **S3** (43.1 g, 89%) as a light yellow oil. *R*_f 0.84 (Chex/EtOAc 7:3); [α]_D²⁵ +16° (c 1.0, CHCl₃); ¹H NMR (CDCl₃, 400 MHz) δ: 8.14-8.11 (m, 1H, CH_{Ph}), 7.64-7.29 (m, 13H, CH_{Ph}), 5.98 (m, 1H, H-2_{All}), 5.71 (dd, *J*_{2,3} = 3.2 Hz, *J*_{1,2} = 1.8 Hz, 1H, H-2) 5.38 (ddd, *J* = 17.2, 3.1, 1.5 Hz, 1H, H-3a_{All}), 5.29 (ddd, *J* = 10.4, 2.5, 1.3 Hz, 1H, H-3b_{All}), 5.02 (d, *J* = 10.9 Hz, 1H, CH₂Ph), 5.02 (d, *J*_{1,2} = 1.8 Hz, 1H, H-1), 4.89-4.82 (m, 3H, CH₂Ph, CH₂N₃), 4.74 (d, *J* = 10.9 Hz, 1H, CH₂Ph), 4.69 (d, *J* = 11.3 Hz, 1H, CH₂Ph), 4.26 (ddt, *J* = 12.9, 5.3, 1.4 Hz, 1H, H-1a_{All}), 4.18 (dd, *J*_{3,4} = 9.4 Hz, *J*_{2,3} = 3.3 Hz, 1H, H-3), 4.09 (ddt, *J* = 12.9, 6.0, 1.3 Hz, 1H, H-1b_{All}), 3.94 (dq, 1H, H-5), 3.64 (pt, *J*_{3,4} = *J*_{4,5} = 9.4 Hz, 1H, H-4), 1.45 (d, *J*_{5,6} = 6.1 Hz, 3H, H-6). ¹³C NMR (CDCl₃, 100 MHz) δ: 166.0 (C=O), 138.5 (C_{Ph}), 138.1 (C_{Ph}), 137.3 (C_{Ph}) 133.6 (C-2_{All}), 132.8-127.7 (CH_{Ph}), 117.7 (C-3_{All}), 96.8 (C-1), 80.2 (C-4), 78.2 (C-3), 75.5 (CH₂Ph), 71.8 (CH₂Ph), 70.0 (C-2), 68.2 (C-1_{All}), 67.9 (C-5), 53.0 (CH₂N₃), 18.2 (C-6). HR-ESI-TOF-MS *m/z* 566.2254 [M + Na]⁺ (calcd for C₃₁H₃₃N₃O₆Na, 566.2267).

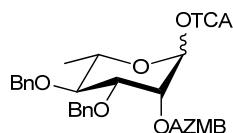
Allyl 2-*O*-(2-(azidomethyl)benzoyl)-4-*O*-benzyl-3-*O*-*para*-methoxybenzyl- α -L-rhamnopyranoside (S4**).**



Route a. DMAP (2.95 g, 24.1 mmol, 2.0 equiv.) and acyl chloride **S2** (2.80 g, 24.1 mmol, 2.0 equiv) were added to allyl 4-*O*-benzyl-3-*O*-*para*-methoxybenzyl- α -L-rhamnopyranoside³ (5.00 g, 12.1 mmol) in anhydrous CH₂Cl₂ (121 mL). The mixture was stirred for 1 h at rt under Ar. The organic phase was washed with satd aq. NaHCO₃ (2×50 mL) and brine (1×50 mL). The solution was dried over anhydrous Na₂SO₄, filtered, and solvents were evaporated. The residue was purified by FC (Chex/EtOAc 95:5 to 8:2) to give allyl glycoside **S4** (6.26 g, 91%) as a colorless oil.

Route b. DMAP (4.13 g, 33.8 mmol, 1.0 equiv), DCC (13.9 g, 67.6 mmol, 2.0 equiv.) and the benzoic acid **S1** (8.98 g, 50.7 mmol, 1.5 equiv) were added to allyl 4-*O*-benzyl-3-*O*-*para*-methoxybenzyl- α -L-rhamnopyranoside¹⁵ (14.0 g, 33.8 mmol) in anhydrous CH₂Cl₂ (338 mL). The mixture was stirred for 4 h at reflux under Ar, and cooled to rt. The mixture was filtered over a pad of Celite®, washed with several portions of CH₂Cl₂, and solvents were evaporated. The residue was purified by FC (Chex/EtOAc 95:5 to 7:3) to give allyl glycoside **S4** (18.6 g, 96%) as a light yellow oil. *R*_f 0.79 (Chex/EtOAc 7:3); [α]_D²⁵ +32° (*c* 1.0, CHCl₃); ¹H NMR (CDCl₃, 400 MHz) δ: 8.05-8.02 (m, 1H, CH_{Ph}), 7.60-7.56 (m, 1H, CH_{Ph}), 7.53-7.50 (m, 1H, CH_{Ph}), 7.45-7.40 (m, 1H, CH_{Ph}), 7.36-7.22 (m, 7H, CH_{Ph}), 6.83-6.78 (m, 2H, CH_{Ph}), 5.91 (m, 1H, H-2_{All}), 5.60 (dd, 1H, H-2), 5.30 (ddd, *J* = 17.2, 3.2, 1.6 Hz, 1H, H-3a_{All}), 5.22 (ddd, *J* = 10.4, 2.8, 1.3 Hz, 1H, H-3b_{All}), 4.92 (d, *J* = 10.9 Hz, 1H, CH₂Ph), 4.92 (d, *J*_{1,2} = 1.7 Hz, 1H, H-1), 4.78 (br s, 2H, CH₂N₃), 4.71 (d, *J* = 10.9 Hz, 1H, CH₂Ph), 4.64 (d, *J* = 11.0 Hz, 1H, CH₂Ph), 4.54 (d, 1H, CH₂Ph), 4.19 (ddt, *J* = 12.8, 5.1, 1.5 Hz, 1H, H-1a_{All}), 4.07 (dd, *J*_{3,4} = 9.3 Hz, *J*_{2,3} = 3.4 Hz, 1H, H-3), 4.01 (ddt, *J* = 12.8, 6.1, 1.3 Hz, 1H, H-1b_{All}), 3.84 (dq, 1H, H-5), 3.77 (s, 3H, OCH₃), 3.52 (pt, *J*_{3,4} = *J*_{4,5} = 9.4 Hz, 1H, H-4), 1.35 (d, *J*_{5,6} = 6.3 Hz, 3H, H-6). ¹³C NMR (CDCl₃, 100 MHz) δ: 166.1 (C=O), 159.4 (C_{Ph}), 138.6 (C_{Ph}), 137.4 (C_{Ph}), 133.7 (C-2_{All}), 132.9-127.8 (CH_{Ph}), 117.8 (C-3_{All}), 113.9 (CH_{Ph}), 96.9 (C-1), 80.3 (C-4), 78.0 (C-3), 75.5 (CH₂Ph), 71.6 (CH₂Ph), 70.1 (C-2), 68.3 (C-1_{All}), 67.9 (C-5), 55.4 (OCH₃), 53.1 (CH₂N₃), 18.2 (C-6). HR-ESI-TOF-MS *m/z* 596.2415 [M + Na]⁺ (calcd for C₃₂H₃₅N₃O₇Na, 596.2372).

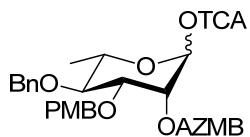
2-*O*-(2-(Azidomethyl)benzoyl)-3,4-di-*O*-benzyl- α -L-rhamnopyranosyl trichloroacetimidate (12).



[Ir] (467 mg, 0.55 mmol, 0.05 equiv.) was dissolved in anhydrous THF (81 mL) and the resulting red solution was degassed under Ar. Hydrogen was bubbled through the solution for 5 min, and the resulting yellow solution was degassed under Ar. Allyl glycoside **S3** (6.00 g, 11.0 mmol) in anhydrous THF (110 mL) was added, and the mixture was stirred for 2 h at rt under Ar. Iodine (5.60 g, 22.1 mmol, 2.0 equiv.) in THF/H₂O (72 mL, 4:1 v/v) was added to the mixture, which was stirred for 1 h at rt. Freshly prepared 10% aq. sodium bisulfite (75 mL) was added. The mixture was concentrated (to 1/3 volume) under reduced pressure and the aq. phase was extracted with CH₂Cl₂ (3×150 mL). The organic layer was washed with H₂O (2×100 mL), brine (100 mL), dried over anhydrous Na₂SO₄, filtered and evaporated. CCl₃CN (5.5 mL, 55 mmol, 5.0 equiv.) and DBU (0.46 mL, 3.1 mmol, 0.28 equiv.) were added to the residue in anhydrous DCE (99 mL). The mixture was stirred for 1 h at rt under Ar, and solvents were evaporated. The residue was purified by FC (Chex/EtOAc 95:5 to 8:2 + 0.5% Et₃N) to give trichloroacetimidate **12** (4.84 g, 68%) as a yellow oil. *R*_f 0.81 (Chex/EtOAc 7:3); ¹H NMR (CDCl₃, 400 MHz) δ: 8.70 (s, 1H, NH), 8.09-8.06 (m, 1H, CH_{Ph}), 7.64-7.59 (m, 1H, CH_{Ph}), 7.56-7.53 (m, 1H, CH_{Ph}), 7.49-7.44 (m, 1H, CH_{Ph}),

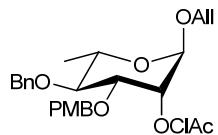
7.37-7.25 (m, 10H, CH_{Ph}), 6.33 (d, $J_{1,2} = 2.0$ Hz, 1H, H-1), 5.71 (dd, 1H, H-2), 4.95 (d, $J = 10.9$ Hz, 1H, CH_2Ph), 4.80 (d, $J = 11.4$ Hz, 1H, CH_2Ph), 4.78 (br s, 2H, CH_2N_3), 4.68 (d, 1H, CH_2Ph), 4.66 (d, 1H, CH_2Ph), 4.12 (dd, $J_{3,4} = 9.4$ Hz, $J_{2,3} = 3.2$ Hz, 1H, H-3), 4.02 (m, 1H, H-5), 3.64 (pt, $J_{4,5} = 9.6$ Hz, 1H, H-4), 1.38 (d, $J_{5,6} = 5.7$ Hz, 3H, H-6). ^{13}C NMR ($CDCl_3$, 100 MHz) δ : 165.7 ($C=O$), 160.3 ($C=NH$), 138.2 (C_{Ph}), 137.7 ($2\times C_{Ph}$), 133.3-128.1 (CH_{Ph}), 95.3 (C-1), 91.0 (CCl_3), 79.6 (C-4), 77.4 (C-3), 75.8 (CH_2Ph), 72.2 (CH_2Ph), 70.9 (C-5), 68.7 (C-2), 53.1 (CH_2N_3), 18.3 (C-6).

2-O-(2-(Azidomethyl)benzoyl)-4-O-benzyl-3-O-*para*-methoxybenzyl- α -L-rhamnopyranosyl trichloroacetimidate (13).



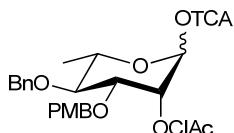
[Ir] (663 mg, 0.784 mmol, 0.03 equiv.) was dissolved in anhydrous THF (131 mL) and the resulting red solution was degassed under Ar. Hydrogen was bubbled through the solution for 5 min, and the resulting yellow solution was degassed under Ar. Allyl glycoside S4 (15.0 g, 26.2 mmol) in anhydrous THF (131 mL) was added and the mixture was stirred for 2 h at rt under Ar. Iodine (13.3 g, 52.3 mmol, 2.0 equiv.) in THF/H₂O (157 mL, 4:1 v/v) was added to the mixture, which was stirred for 3 h at rt. Freshly prepared 10% aq. sodium bisulfite (100 mL) was added. The mixture was concentrated (to 1/3 volume) under reduced pressure and the aq. phase was extracted with CH_2Cl_2 (3×250 mL). The organic layer was washed with H₂O (100 mL), brine (250 mL), dried over anhydrous Na₂SO₄, filtered and evaporated. CCl_3CN (13.1 mL, 131 mmol, 5.0 equiv.) and DBU (1.1 mL, 7.3 mmol, 0.28 equiv.) were added to the residue in anhydrous DCE (114 mL). The mixture was stirred for 1 h at rt under Ar, and the solvents were evaporated. The crude material was purified by FC ($CH_2Cl_2 + 0.5\%$ Et₃N) to give 13 (10.7 g, 60%) as a yellow oil. R_f 0.76 (Chex/EtOAc 7:3); $[\alpha]^{25}_D -3^\circ$ (*c* 1.0, CHCl₃); 1H NMR ($CDCl_3$, 400 MHz) δ : 8.71 (s, 1H, NH), 8.09-8.06 (m, 1H, CH_{Ph}), 7.65-7.61 (m, 1H, CH_{Ph}), 7.58-7.55 (m, 1H, CH_{Ph}), 7.50-7.45 (m, 1H, CH_{Ph}), 7.39-7.25 (m, 7H, CH_{Ph}), 6.85-6.81 (m, 2H, CH_{Ph}), 6.33 (d, $J_{1,2} = 1.9$ Hz, 1H, H-1), 5.70 (dd, 1H, H-2), 4.95 (d, $J = 10.8$ Hz, 1H, CH_2Ph), 4.81 (br s, 2H, CH_2N_3), 4.74 (d, $J = 11.0$ Hz, 1H, CH_2Ph), 4.68 (d, 1H, CH_2Ph), 4.61 (d, $J = 11.0$ Hz, 1H, CH_2Ph), 4.11 (dd, $J_{3,4} = 9.5$ Hz, $J_{2,3} = 3.3$ Hz, 1H, H-3), 4.01 (dq, 1H, H-5), 3.80 (s, 3H, OCH_3), 3.63 (t, $J_{4,5} = 9.6$ Hz, 1H, H-4), 1.39 (d, $J_{5,6} = 6.0$ Hz, 3H, H-6). ^{13}C NMR ($CDCl_3$, 100 MHz) δ : 165.7 ($C=O$), 160.3 ($C=NH$), 159.6 (C_{Ph}), 138.3 (C_{Ph}), 137.7 (C_{Ph}), 133.2-128.0 (CH_{Ph}), 114.0 (C_{Ph}), 95.3 (C-1), 91.0 (CCl_3), 79.5 (C-4), 77.0 (C-3), 75.8 (CH_2Ph), 71.8 (CH_2Ph), 70.9 (C-5), 68.7 (C-2), 55.4 (OCH_3), 53.2 (CH_2N_3), 18.3 (C-6).

Allyl 4-O-benzyl-2-O-chloroacetyl-3-O-*para*-methoxybenzyl- α -L-rhamnopyranoside (S5).



Chloroacetyl chloride (192 μ L, 2.41 mmol, 4.0 equiv.) was carefully added to allyl 4-*O*-benzyl-3-*O*-*para*-methoxybenzyl- α -L-rhamnopyranoside¹⁵ (250 mg, 0.603 mmol) in cold (0 °C) anhyd. CH₂Cl₂/Py (5.4 mL, 8:1 *v/v*). The mixture was stirred for 3 h at rt under Ar, then diluted with CH₂Cl₂ (100 mL). The organic phase was washed with 10% aq HCl (50 mL), satd NaHCO₃ (50 mL) and brine (50 mL), then dried over anhyd. Na₂SO₄. Volatiles were evaporated and the residue was purified on a short pad of silica gel (CH₂Cl₂, then Chex/EtOAc 7:3) to give the fully protected **S5** (296 mg, 100%) as a yellow oil. R_f 0.75 (Chex/EtOAc 7:3); $[\alpha]^{25}_D$ -22° (*c* 1.0, CHCl₃); ¹H NMR (CDCl₃, 400 MHz) δ : 7.37-7.23 (m, 7H, CH_{Ph}), 6.87-6.83 (m, 2H, CH_{Ph}), 5.88 (m, 1H, H-2_{All}), 5.44 (dd, $J_{2,3}$ = 3.3 Hz, 1H, H-2), 5.28 (ddd, J = 17.2, 3.2, 1.5 Hz, 1H, H-3a_{All}), 5.21 (ddd, J = 10.3, 2.6, 1.3 Hz, 1H, H-3a_{All}), 4.89 (d, J = 10.8 Hz, 1H, CH₂Ph), 4.80 (d, $J_{1,2}$ = 1.7 Hz, 1H, H-1), 4.64 (d, J = 10.8 Hz, 1H, CH₂Ph), 4.60 (d, 1H, CH₂Ph), 4.47 (d, 1H, CH₂Ph), 4.16 (ddt, J = 12.7, 5.3, 1.5 Hz, 1H, H-1a_{All}), 4.16 (s, 2H, CH₂Cl), 4.01-3.95 (m, 2H, H-1b_{All}, H-3), 3.82-3.75 (dq, 1H, H-5), 3.80 (s, 3H, OCH₃), 3.39 (pt, $J_{3,4}$ = $J_{4,5}$ = 9.6 Hz, 1H, H-4), 1.32 (d, $J_{5,6}$ = 6.1 Hz, 3H, H-6). ¹³C NMR (CDCl₃, 100 MHz) δ : 167.0 (C=O), 159.5 (C_{Ph}), 138.5 (C_{Ph}), 133.5 (C-2_{All}), 129.9-127.8 (CH_{Ph}), 117.9 (C-3_{All}), 114.0 (CH_{Ph}), 96.5 (C-1), 80.0 (C-4), 77.8 (C-3), 75.5 (CH₂Ph), 71.8 (CH₂Ph), 71.1 (C-2), 68.2 (C-1_{All}), 67.9 (C-5), 55.4 (OCH₃), 41.1 (CH₂Cl), 18.0 (C-6). HR-ESI-TOF-MS *m/z* 513.1653 [M + Na]⁺ (calcd for C₂₆H₃₁ClO₇Na, 513.1656).

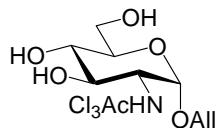
4-*O*-Benzyl-2-*O*-chloroacetyl-3-*O*-*para*-methoxybenzyl- α -L-rhamnopyranosyl trichloro-acetimidate (14).



[Ir] (122 mg, 0.14 mmol, 0.05 equiv.) was dissolved in anhyd. THF (14 mL) and the resulting red solution was degassed under Ar. Hydrogen was bubbled through the solution for 15 min, and the resulting yellow solution was degassed under Ar. The allyl glycoside **S5** (1.42 g, 2.89 mmol) in anhyd. THF (14 mL) was then added. The mixture was stirred overnight at rt under Ar. Iodine (1.47 g, 5.78 mmol, 2.0 equiv.) in THF/H₂O (6.0 mL, 4:1 *v/v*) was added to the mixture, which was stirred for 1 h at rt. Freshly prepared 10% aq sodium bisulfite was added. The mixture was concentrated (to 1/3 volume) under reduced pressure and the aq phase was extracted three times with CH₂Cl₂. The organic layer was washed twice with H₂O, once with brine, dried over anhyd. Na₂SO₄, filtered and evaporated. CCl₃CN (1.5 mL, 14 mmol, 5.0 equiv.) and DBU (130 μ L, 0.87 mmol, 0.30 equiv.) were added to the residue in anhyd. DCE (5.8 mL). The mixture was stirred for 1 h at rt under Ar, and directly purified by FC (Chex/EtOAc 9:1 to 8:2 + 1% Et₃N) to give **14** (1.53 g, 89%) as a yellow oil. R_f 0.73 (Chex/EtOAc 7:3); ¹H NMR (CDCl₃, 400 MHz) δ (α): 8.68

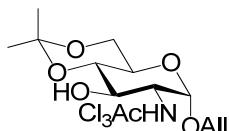
(s, 1H, NH), 7.38-7.22 (m, 7H, CH_{Ph}), 6.87-6.82 (m, 2H, CH_{Ph}), 6.20 (br s, 1H, H-1), 5.51 (br s, 1H, H-2), 4.91 (d, *J* = 10.8 Hz, 1H, CH₂Ph), 4.66 (d, *J* = 11.9 Hz, 1H, CH₂Ph), 4.63 (d, 1H, CH₂Ph), 4.52 (d, 1H, CH₂Ph), 4.19 (s, 2H, CH₂Cl), 4.02-3.91 (m, 2H, H-3, H-5), 3.80 (s, 3H, OCH₃), 3.48 (pt, *J*_{3,4} = *J*_{4,5} = 9.6 Hz, 1H, H-4), 1.34 (d, *J*_{5,6} = 6.1 Hz, 3H, H-6). ¹³C NMR (CDCl₃, 100 MHz) δ (α): 166.7 (C=O), 160.1 (C=NH), 159.6 (C_{Ph}), 138.1 (C_{Ph}), 130.2-128.1 (CH_{Ph}), 114.0 (CH_{Ph}), 94.8 (C-1), 90.9 (CCl₃), 79.1 (C-4), 76.7 (C-3), 75.8 (CH₂Ph), 72.0 (CH₂Ph), 70.9 (C-5), 69.6 (C-2), 55.4 (OCH₃), 40.9 (CH₂Cl), 18.0 (C-6).

Allyl 2-deoxy-2-trichloroacetamido- α -D-glucopyranoside (**25**).



D-Glucosamine hydrochloride (10.0 g, 46.4 mmol) was dissolved in anhydrous MeOH (199 mL) and the mixture was stirred for 1 h at rt under Ar. After 15 min, trichloroacetic anhydride (12.7 mL, 69.6 mmol, 1.5 equiv.) was added dropwise at 0 °C and the mixture was stirred for 2 h at this temperature. The reaction was neutralized with Dowex resin (H⁺ form), filtered and solvents were evaporated. Acetyl chloride (0.37 mL, 5.2 mmol, 3.4 equiv.) was slowly added to the residue in allylic alcohol (3.1 mL) at 0 °C. The mixture was stirred for 3 h at 70 °C, and diluted with MeOH. Solid NaHCO₃ was added until neutrality was reached. After filtration, solvents were evaporated and the residue was purified by FC (CH₂Cl₂/MeOH 95:5 to 9:1) to give triol **25** (7.53 g, 45%, 2 steps) as an amorphous brownish solid. *R*_f 0.16 (CH₂Cl₂/MeOH 95:5); [α]²⁵_D +113° (c 1.0, MeOH); ¹H NMR (CDCl₃, 400 MHz) δ: 7.14 (d, *J*_{NH,2} = 8.8 Hz, 1H, NH), 5.85 (m, 1H, H-2_{All}), 5.29 (ddd, 1H, *J* = 17.1, 3.0, 1.5 Hz, 1H, H-3a_{All}), 5.21 (ddd, 1H, *J* = 10.4, 2.5, 1.1 Hz, 1H, H-3b_{All}), 4.95 (d, *J*_{1,2} = 3.6 Hz, 1H, H-1), 4.19 (ddt, *J* = 13.0, 5.2, 1.4 Hz, 1H, H-1a_{All}), 4.08 (ddd, *J*_{2,3} = 9.5 Hz, 1H, H-2), 4.00 (ddt, *J* = 13.0, 6.3, 1.2 Hz, 1H, H-1b_{All}), 3.94 (dd, *J*_{6a,6b} = 12.5 Hz, *J*_{5,6a} = 2.3 Hz, 1H, H-6a), 3.88-3.81 (m, 2H, H-6b, H-3), 3.77 (pt, *J*_{3,4} = 9.3 Hz, 1H, H-4), 3.64 (dt, *J*_{4,5} = 9.6 Hz, *J*_{5,6a} = *J*_{5,6b} = 2.5 Hz, 1H, H-5). ¹³C NMR (CDCl₃, 100 MHz) δ: 162.9 (C=O), 133.3 (C-2_{All}), 118.5 (C-3_{All}), 96.1 (C-1), 92.6 (CCl₃), 72.8 (C-3), 72.0 (C-5), 70.1 (C-4), 68.7 (C-1_{All}), 61.3 (C-6), 55.5 (C-2). HR-ESI-TOF-MS *m/z* 364.0120 [M + H]⁺ (calcd for C₁₂H₁₇Cl₃NO₆, 364.0121).

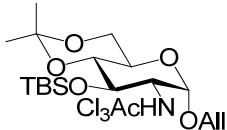
Allyl 2-deoxy-4,6-O-isopropylidene-2-trichloroacetamido- α -D-glucopyranoside (**S6**).



2-Methoxypropene (4.2 mL, 44.4 mmol, 2.0 equiv.) and *p*TSA (422 mg, 2.22 mmol, 0.1 equiv.) were added to triol **25** (8.07 g, 22.1 mmol) in anhydrous DMF (75 mL). The mixture was stirred for

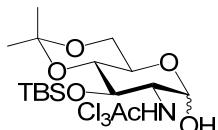
1 h at rt under Ar. Et₃N (3.0 mL) was added and solvents were evaporated. The residue was purified by FC (Chex/EtOAc 8:2 to 7:3) to give alcohol **S6** (8.09 g, 90%) as a white amorphous powder. *R*_f 0.36 (Chex/EtOAc 7:3); [α]²⁵_D +95° (*c* 1.0, CHCl₃); ¹H NMR (Py-d₅, 400 MHz) δ: 10.35 (d, *J*_{NH,2} = 8.1 Hz, 1H, NH), 7.69 (d, *J*_{O_H,3} = 5.3 Hz, 1H, OH), 6.01 (m, 1H, H-2_{AII}), 5.36 (ddd, *J* = 17.2, 3.2, 1.6 Hz, 1H, H-3a_{AII}), 5.36 (d, *J*_{1,2} = 3.8 Hz, 1H, H-1), 5.18 (ddd, *J* = 10.3, 2.8, 1.3 Hz, 1H, H-3b_{AII}), 4.72 (ddd, *J*_{2,3} = 10.2 Hz, 1H, H-2), 4.50 (m, 1H, H-3), 4.28 (ddt, *J* = 13.0, 5.3, 1.4 Hz, 1H, H-1a_{AII}), 4.09 (ddt, *J* = 13.0, 6.2, 1.3 Hz, 1H, H-1b_{AII}), 4.04-3.90 (m, 4H, H-4, H-5, H-6a, H-6b), 1.49 (s, 3H, C(CH₃)₂), 1.44 (s, 3H, C(CH₃)₂). ¹³C NMR (Py-d₅, 100 MHz) δ: 163.6 (C=O), 134.9 (C-2_{AII}), 118.2 (C-3_{AII}), 100.2 (C(CH₃)₂), 97.8 (C-1), 94.5 (CCl₃), 76.6 (C-4), 69.3 (C-1_{AII}), 68.8 (C-3), 65.0 (C-5), 63.0 (C-6), 58.1 (C-2), 29.7 (C(CH₃)₂), 19.6 (C(CH₃)₂). HR-ESI-TOF-MS *m/z* 404.0441 [M + H]⁺ (calcd for C₁₅H₂₁Cl₃NO, 404.0435).

Allyl 3-*O*-tert-butylidemethylsilyl-2-deoxy-4,6-*O*-isopropylidene-2-trichloroacetamido- α -D-glucopyranoside (26).



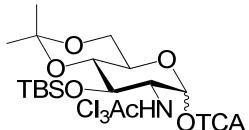
DMAP (241 mg, 2.0 mmol, 0.1 equiv.), imidazole (3.36 g, 49.3 mmol, 2.5 equiv.) and TBSCl (8.92 g, 59.2 mmol, 3.0 equiv.) were added to alcohol **S6** (7.98 g, 19.7 mmol) in anhydrt. THF (59 mL). The mixture was stirred for 2 d at reflux under Ar. After cooling to rt, the reaction was filtered and the filtrate was evaporated. The residue was purified by FC (Chex to Chex/EtOAc 9:1) to give **26** (10.0 g, 98%) as a white amorphous powder. *R*_f 0.82 (Chex/EtOAc 7:3); [α]²⁵_D +73° (*c* 1.0, CHCl₃); ¹H NMR (Py-d₅, 400 MHz) δ: 8.91 (d, *J*_{NH,2} = 9.1 Hz, 1H, NH), 5.90 (m, 1H, H-2_{AII}), 5.27 (br dd, *J* = 17.2, 2.9 Hz, 1H, H-3a_{AII}), 5.16-5.11 (m, 2H, H-1, H-3b_{AII}), 4.54 (pdt, *J*_{2,3} = 9.5 Hz, *J*_{1,2} = 3.8 Hz, 1H, H-2), 4.26 (pt, *J*_{3,4} = 9.3 Hz, 1H, H-3), 4.22 (ddt, *J* = 13.0, 5.5, 1.3 Hz, 1H, H-1a_{AII}), 4.06-3.99 (m, 2H, H-1b_{AII}, H-6a), 3.97-3.85 (m, 2H, H-5, H-6b), 3.71 (pt, *J*_{4,5} = 8.9 Hz, 1H, H-4), 1.52 (s, 3H, C(CH₃)₂), 1.49 (s, 3H, C(CH₃)₂), 1.00 (s, 9H, C(CH₃)₃), 0.16 (s, 3H, Si(CH₃)₂), 0.14 (s, 3H, Si(CH₃)₂). ¹³C NMR (Py-d₅, 100 MHz) δ: 163.1 (C=O), 134.5 (C-2_{AII}), 118.8 (C-3_{AII}), 100.1 (C(CH₃)₂), 97.5 (C-1), 94.0 (CCl₃), 75.8 (C-4), 71.4 (C-3), 69.3 (C-1_{AII}), 64.5 (C-5), 62.8 (C-6), 57.9 (C-2), 29.7 (C(CH₃)₂), 26.6 (C(CH₃)₃, 3C), 19.5 (C(CH₃)₂), 18.9 (C(CH₃)₃), -3.2 (Si(CH₃)₂), -4.3 (Si(CH₃)₂). HR-ESI-TOF-MS *m/z* 518.1277 [M + H]⁺ (calcd for C₂₀H₃₅Cl₃NO₆Si, 518.1299).

3-*O*-tert-Butyldemethylsilyl-2-deoxy-4,6-*O*-isopropylidene-2-trichloroacetamido- α / β -D-glucopyranose (S7).



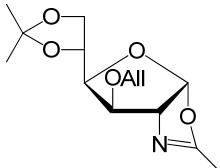
[Ir] (815 mg, 0.96 mmol, 0.05 equiv.) was dissolved in anhydrous THF (96 mL) and the resulting red solution was degassed under Ar. Hydrogen was bubbled through the solution for 5 min, and the resulting yellow solution was degassed under Ar. Allyl glycoside **26** (10.0 g, 19.3 mmol) in anhydrous THF (96 mL) was added and the mixture was stirred overnight at rt under Ar. Iodine (9.78 g, 38.5 mmol, 2.0 equiv.) in THF/H₂O (115 mL, 4:1 v/v) was added to the mixture, which was stirred for 1 h at rt. Freshly prepared 10% aq sodium bisulfite (75 mL) was added. The mixture was concentrated (to 1/3 volume) under reduced pressure and the aq phase was extracted three times with CH₂Cl₂ (3×250 mL). The organic layer was washed with H₂O (2×250 mL), brine (250 mL), dried over anhydrous Na₂SO₄, filtered and evaporated. The residue was purified by FC (Chex/EtOAc 9:1 to 6:4) to give hemiacetal **S7** (7.99 g, 87%, ratio α/β ≈ 7:1) as a yellow amorphous powder. *R*_f 0.46 (Chex/EtOAc 7:3); ¹H NMR (Py-d₅, 400 MHz) δ (α): 9.31 (br s, 1H, OH), 8.37 (br d, 1H, NH), 5.65 (pt, *J*_{1,OH} = 3.4 Hz, 1H, H-1), 4.55 (td, *J*_{2,3} = *J*_{2,NH} = 9.5 Hz, *J*_{1,2} = 3.7 Hz, 1H, H-2), 4.39 (pt, *J*_{3,4} = 9.0 Hz, 1H, H-3), 4.34 (dt, *J*_{5,6a} = *J*_{5,6b} = 5.1 Hz, 1H, H-5), 4.02 (dd, *J*_{6a,6b} = 10.7 Hz, *J*_{5,6a} = 5.4 Hz, 1H, H-6a), 3.92 (pt, *J*_{5,6a} = 10.6 Hz, 1H, H-6b), 3.79 (pt, *J*_{4,5} = 9.2 Hz, 1H, H-4), 1.55 (s, 3H, C(CH₃)₂), 1.53 (s, 3H, C(CH₃)₂), 1.06 (s, 9H, C(CH₃)₃), 0.25 (s, 6H, Si(CH₃)₂). ¹³C NMR (Py-d₅, 100 MHz) δ (α): 163.0 (C=O), 100.1 (C(CH₃)₂), 94.2 (CCl₃), 92.7 (C-1), 76.2 (C-4), 71.6 (C-3), 64.1 (C-5), 63.3 (C-6), 58.7 (C-2), 29.7 (C(CH₃)₂), 26.6 (C(CH₃)₃), 19.5 (C(CH₃)₂), 19.0 (C(CH₃)₃), -3.2 (Si(CH₃)₂), -4.2 (Si(CH₃)₂). HR-ESI-TOF-MS *m/z* 478.0997 [M + H]⁺ (calcd for C₁₇H₃₁Cl₃NO₆Si, 478.0968).

3-O-tert-Butyldimethylsilyl-2-deoxy-4,6-O-isopropylidene-2-trichloroacetamido- α/β -D-glucopyranosyl trichloroacetimidate (11).



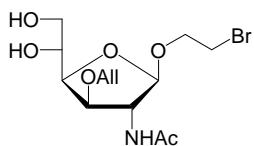
CCl₃CN (8.3 mL, 82 mmol, 5.0 equiv.) and DBU (0.74 mL, 4.9 mmol, 0.3 equiv.) were added to hemiacetal **S7** (7.88 g, 16.5 mmol) in anhydrous DCE (33 mL). The mixture was stirred for 1 h at rt under Ar, and directly purified by FC (Chex/EtOAc 9:1 + 1% Et₃N) to give trichloroacetimidate **11** (8.62 g, 84%, α/β 7:1) as a white foam. *R*_f 0.74 (Chex/EtOAc 7:3); [α]²⁵_D +63° (*c* 1.0, CHCl₃); ¹H NMR (Py-d₅, 400 MHz) δ (α): 10.48 (s, 1H, C=NH), 8.26 (d, *J*_{NH,2} = 8.7 Hz, 1H, NH), 6.97 (d, *J*_{1,2} = 3.9 Hz, 1H, H-1), 4.78 (td, *J*_{2,3} = 9.4 Hz, 1H, H-2), 4.48 (t, *J*_{3,4} = 9.3 Hz, 1H, H-3), 4.22 (dq, 1H, H-5), 4.06 (dd, *J*_{6a,6b} = 10.7 Hz, *J*_{5,6a} = 5.1 Hz, 1H, H-6a), 3.95-3.87 (m, 2H, H-6b, H-4), 1.55 (s, 3H, C(CH₃)₂), 1.51 (s, 3H, C(CH₃)₂), 1.00 (s, 9H, C(CH₃)₃), 0.21 (s, 3H, Si(CH₃)₂), 0.18 (s, 3H, Si(CH₃)₂). ¹³C NMR (Py-d₅, 100 MHz) δ (α): 163.0 (C=O), 160.1 (C=NH), 100.5 (C(CH₃)₂), 95.6 (C-1), 93.5 (CCl₃), 91.7 (CCl₃), 74.9 (C-4), 71.6 (C-3), 67.1 (C-5), 62.5 (C-6), 57.2 (C-2), 29.5 (C(CH₃)₂), 26.5 (C(CH₃)₃, 3C), 19.5 (C(CH₃)₂), 18.9 (C(CH₃)₃), -3.2 (Si(CH₃)₂), -4.3 (Si(CH₃)₂).

(2-Methyl-(3-*O*-allyl-1,2-dideoxy-5,6-*O*-isopropylidene- α -D-glucofuranosyl)-[2,1-d]-2-oxazoline (31).



NaH (60% oil dispersion, 414 mg, 12.3 mmol, 3.0 equiv.) was added to the crude oxazoline **30**¹⁶ (1.00 g, 4.11 mmol) in anhydrous DMF (12.3 mL) at 0 °C. After 1 h at this temperature, allyl bromide (1.07 mL, 12.3 mmol, 3.0 equiv.) was added dropwise and the mixture was stirred overnight at rt under Ar. After cooling to 0 °C, MeOH was carefully added. H₂O (50 mL) was added and the aq. phase was extracted with CH₂Cl₂ (3×100 mL). The organic layer was dried over anhydrous Na₂SO₄, filtered and evaporated to dryness. The residue was purified by FC (Chex/EtOAc 7:3 to 2:8) to give oxazoline **31** (892 mg, 77% from *N*-acetyl-D-glucosamine) as a yellow oil. *R*_f 0.28 (CH₂Cl₂/MeOH 95:5); [α]²⁵_D -46° (*c* 1.0, CHCl₃); ¹H NMR (CDCl₃, 400 MHz) δ: 6.10 (d, *J*_{1,2} = 5.1 Hz, 1H, H-1), 5.90 (m, 1H, H-2_{AII}), 5.31 (ddd, *J* = 17.3, 3.3, 1.5 Hz, 1H, H-3a_{AII}), 5.20 (ddd, *J* = 10.5, 2.9, 1.4 Hz, 1H, H-3b_{AII}), 4.48 (m, 1H, H-2), 4.32 (dt, *J*_{5,6a} = *J*_{5,6b} = 6.0 Hz, 1H, H-5), 4.18 (ddt, *J* = 12.9, 5.3, 1.4 Hz, 1H, H-1a_{AII}), 4.14-4.04 (m, 4H, H-1b_{AII}, H-6a, H-3, H-6b), 3.81 (dd, *J*_{4,5} = 7.1 Hz, *J*_{3,4} = 3.1 Hz, 1H, H-4), 2.01 (d, *J*_{2,Me} = 1.5 Hz, 3H, CH₃), 1.40 (s, 3H, C(CH₃)₂), 1.34 (s, 3H, C(CH₃)₂). ¹³C NMR (CDCl₃, 100 MHz) δ: 167.0 (C=N), 134.3 (C-2_{AII}), 117.5 (C-3_{AII}), 109.1 (C(CH₃)₂), 107.1 (C-1), 81.6 (C-4), 81.5 (C-3), 75.8 (C-2), 72.7 (C-5), 71.2 (C-1_{AII}), 67.0 (C-6), 26.9 (C(CH₃)₂), 25.5 (C(CH₃)₂), 14.3 (CH₃). HR-ESI-TOF-MS *m/z* 284.1468 [M + H]⁺ (calcd for C₁₄H₂₂NO₅, 284.1498).

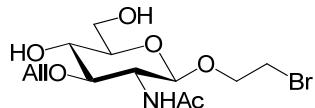
2-Bromoethyl 2-acetamido-3-*O*-allyl-2-deoxy- β -D-glucofuranoside (S8).



Yb(OTf)₃ (5.47 g, 8.82 mmol, 0.5 equiv.) was added to oxazoline **31** (5.00 g, 17.6 mmol) in anhydrous 2-bromoethanol (29 mL). The mixture was stirred overnight at rt under Ar. Solvents were evaporated and the residue was purified by FC (CH₂Cl₂/MeOH 95:5 to 85:15) to give **S8** (6.47 g, 99%) as a brownish foam. *R*_f 0.12 (CH₂Cl₂/MeOH 95:5); [α]²⁵_D -71° (*c* 1.0, MeOH); ¹H NMR (CDCl₃, 400 MHz) δ: 6.43 (d, *J*_{NH,2} = 7.7 Hz, 1H, NH), 5.89 (m, 1H, H-2_{AII}), 5.32 (br d, *J* = 17.3 Hz, 1H, H-3a_{AII}), 5.19 (br d, *J* = 10.5 Hz, 1H, H-3b_{AII}), 4.95 (s, 1H, H-1), 4.41 (d, 1H, H-2), 4.32 (br dd, *J* = 13.0, 4.9 Hz, 1H, H-1a_{AII}), 4.26 (dd, *J*_{4,5} = 8.7 Hz, *J*_{3,4} = 6.1 Hz, 1H, H-4), 4.09-4.02 (m, 3H, H-1b_{AII}, H-5, H-3), 3.93 (dt, *J*_{a,b} = 11.1, *J*_{vic} = 6.0 Hz, 1H, OCH₂a), 3.82 (m, *J*_{vic} = 3.1 Hz, 1H, H-6a), 3.76-3.67 (m, 2H, H-6b, OCH₂b), 3.44 (m, *J* = 6.1 Hz, 2H, CH₂Br), 2.98 (br s, 2H, OH), 1.98 (s, 3H, C(O)CH₃). ¹³C NMR (CDCl₃, 100 MHz) δ: 170.2 (C=O), 133.8 (C-2_{AII}), 118.1

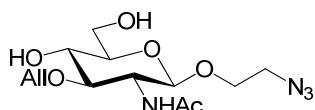
(C-3_{AlI}), 107.0 (C-1), 83.0 (C-3), 80.1 (C-4), 70.9 (C-1_{AlI}), 70.8 (C-5), 68.2 (OCH₂), 64.0 (C-6), 59.7 (C-2), 30.2 (CH₂Br), 23.2 (C(O)CH₃). HR-ESI-TOF-MS *m/z* 390.0522 [M + Na]⁺ (calcd for C₁₃H₂₂BrNO₆Na, 390.0528).

2-Bromoethyl 2-acetamido-3-*O*-allyl-2-deoxy- β -D-glucopyranoside (32).



CSA (20 mg, 86 μ mol, 0.25 equiv.) was added to oxazoline **31** (100 mg, 353 μ mol) in anhyd. 2-bromoethanol (0.71 mL). The mixture was stirred for 4 d at rt under Ar, and neutralized by adding solid NaHCO₃, then filtered. Solvents were evaporated and the residue was purified by FC (CH₂Cl₂/MeOH 95:5 to 9:1) to give diol **32** (70 mg, 55%) as a white amorphous powder. *R*_f 0.13 (CH₂Cl₂/MeOH 95:5); $[\alpha]^{25}_D$ -19° (*c* 1.0, MeOH); ¹H NMR (DMSO-d₆, 400 MHz) δ : 7.78 (d, *J*_{NH,2} = 9.1 Hz, 1H, NH), 5.84 (m, 1H, H-2_{AlI}), 5.18 (ddd, *J* = 17.2, 3.8, 1.8 Hz, 1H, H-3a_{AlI}), 5.04 (ddd, *J* = 10.4, 3.5, 1.5 Hz, 1H, H-3b_{AlI}), 4.41 (d, *J*_{1,2} = 8.5 Hz, 1H, H-1), 4.21 (ddt, *J* = 13.1, 5.3, 1.5 Hz, 1H, H-1a_{AlI}), 4.05 (ddt, *J* = 13.1, 5.3, 1.5 Hz, 1H, H-1b_{AlI}), 3.95 (dt, *J* = 11.6, 5.8 Hz, 1H, OCH₂a), 3.76 (dt, *J* = 11.7, 6.0 Hz, 1H, OCH₂b), 3.68 (br d, *J*_{6a,6b} = 10.7 Hz, 1H, H-6a), 3.60-3.43 (m, 4H, CH₂Br, H-6b, H-2), 3.32 (br s₀, 2H, OH), 3.29 (pt₀, 1H, H-3), 3.20 (pdt, *J*_{4,5} = 9.5 Hz, *J*_{5,6a} = *J*_{5,6b} = 5.9 Hz, 1H, H-5), 3.13 (ddd, *J*_{3,4} = 9.4 Hz, *J*_{4,OH} = 2.0 Hz, 1H, H-4), 1.80 (s, 3H, C(O)CH₃). ¹³C NMR (DMSO-d₆, 100 MHz) δ : 168.9 (C=O), 136.0 (C-2_{AlI}), 115.4 (C-3_{AlI}), 100.7 (C-1), 82.3 (C-3), 77.0 (C-4), 72.3 (C-1_{AlI}), 69.9 (C-5), 68.3 (OCH₂), 60.8 (C-6), 54.0 (C-2), 31.9 (CH₂Br), 23.0 (C(O)CH₃). HR-ESI-TOF-MS *m/z* 390.0540 [M + Na]⁺ (calcd for C₁₃H₂₂BrNO₆Na, 390.0528).

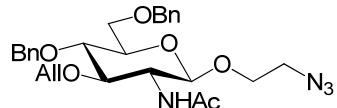
2-Azidoethyl 2-acetamido-3-*O*-allyl-2-deoxy- β -D-glucopyranoside (S9).



To a solution of diol **32** (3.19 g, 8.67 mmol) in anhyd. DMF (43.4 mL) were sequentially added NaI (6.50 g, 43.4 mmol, 5.0 equiv.) and NaN₃ (2.82 g, 43.4 mmol, 5.0 equiv). The mixture was stirred overnight at 80 °C, and the solvents were removed under reduced pressure. The residue was adsorbed on silica gel (22 g) and purified by FC (CH₂Cl₂/MeOH 95:5 to 9:1) to give the azidoethyl glycoside **S9** contaminated with sodium salts (6.78 g). Available analytical data for diol **S9**: *R*_f 0.11 (CH₂Cl₂/MeOH 95:5); $[\alpha]^{25}_D$ -19° (*c* 1.0, MeOH); ¹H NMR (Py-d₅, 400 MHz) δ : 10.56 (bd, 1H, NH), 7.15 (m, 1H, H-2_{AlI}), 6.42 (*J*_{1,2} = 8.7), 6.38 (m, 1H, H-3_{AlI}), 6.10 (br d, *J* = 10.4 Hz, *J* = 1.4 Hz, 1H, H-3b_{AlI}), 5.78 (m, 2H, H-1a_{AlI}, H-1b_{AlI}), 5.60 (p, *J*_{1,2} = 9.9 Hz, *J*_{2,3} = 9.2 Hz, 9.0 1H, H-2), 5.50-5.40 (m, 2H, H-3, H-6a), 5.35 (dd, *J*_{6a,6b} = 11.8 Hz, *J*_{5,6b} = 4.5 Hz, 1H, H-6b), 4.19-4.10 (m, 3H, CH₂N₃a, H-4, OH), 5.04-4.90 (m, 2H, CH₂N₃b, H-5), 4.64 (m, 1H,

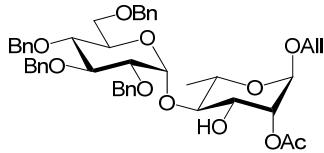
OCH₂a), 4.50 (m, 1H, OCH₂b), 3.31 (s, 3H, C(O)CH₃). ¹³C NMR (Py-d₅, 100 MHz) δ: 172.5 (C=O), 136.4 (C-2_{All}), 116.6 (C-3_{All}), 102.0 (C-1), 83.7 (C-3), 78.0 (C-5), 74.5 (C-1_{All}), 71.9 (C-4), 68.8 (CH₂N₃), 62.2 (C-6), 56.1 (C-2), 51.3 (OCH₂), 23.9 (C(O)CH₃). HR-ESI-TOF-MS *m/z* 353.1406 [M + Na]⁺ (calcd for C₁₃H₂₂N₄O₆Na, 353.1437).

2-Azidoethyl 2-acetamido-3-*O*-allyl-4,6-di-*O*-benzyl-2-deoxy-β-D-glucopyranoside (33).



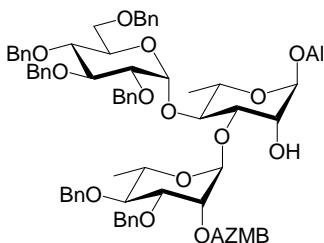
NaI (6.50 g, 43.4 mmol, 5.0 equiv.) and NaN₃ (2.82 g, 43.4 mmol, 5.0 equiv.) were sequentially added to diol **32** (3.19 g, 8.67 mmol) in anhyd. DMF (43.4 mL). The mixture was stirred overnight at 80 °C, and solvents were removed under reduced pressure. The residue was adsorbed on silica gel (22 g) and purified by FC (CH₂Cl₂/MeOH 95:5 to 9:1) to give the azidoethyl glycoside (6.78 g) contaminated with sodium salts. BnBr (3.91 mL, 32.9 mmol, 4.0 equiv.) was slowly added to the above crude material (2.72 g) in anhyd. DMF (165 mL) stirred at 0 °C. NaH (858 mg, 25.5 mmol, 3.0 equiv.) was added portionwise and stirring went on for 2 h at 0 °C. MeOH was added and the mixture was diluted with EtOAc (250 mL). H₂O was added carefully (250 mL) and the aq. phase was extracted with EtOAc (3×250 mL). The organic layer was washed with H₂O (1×250 mL), dried (anhyd. Na₂SO₄), filtered and solvents were removed under reduced pressure. The residue was purified by FC (Chex/EtOAc 6:4 to 0:10) to give fully protected **33** (2.39 g, 57%, 2 steps from **32**) as a white amorphous powder. *R*_f 0.06 (Chex/EtOAc 7:3); [α]²⁵_D +1° (c 1.0, CHCl₃); ¹H NMR (CDCl₃, 400 MHz) δ: 7.35-7.21 (m, 10H, CH_{Ph}), 5.89 (m, 1H, H-2_{All}), 5.73 (d, *J*_{NH,2} = 7.3 Hz, 1H, NH), 5.24 (ddd, *J* = 17.3, 3.2, 1.6 Hz, 1H, H-3a_{All}), 5.15 (br d, *J* = 10.4 Hz, 1H, H-3b_{All}), 4.96 (d, *J*_{1,2} = 7.8 Hz, 1H, H-1), 4.78 (d, *J* = 11.1 Hz, 1H, CH₂Ph), 4.60 (d, *J* = 12.2 Hz, 1H, CH₂Ph), 4.57 (d, 1H, CH₂Ph), 4.54 (d, 1H, CH₂Ph), 4.27 (br dd, *J* = 12.6, 5.5 Hz, 1H, H-1a_{All}), 4.15 (br dd, *J* = 12.6, 5.8 Hz, 1H, H-1b_{All}), 4.11-4.00 (m, 2H, H-3, OCH₂a), 3.76-3.66 (m, 3H, OCH₂b, H-6a, H-6b), 3.60-3.52 (m, 2H, H-4, H-5), 3.48 (ddd, *J* = 13.2, 8.0, 3.4 Hz, 1H, CH₂N₃a), 3.32-3.22 (m, 2H, H-2, CH₂N₃b), 1.98 (s, 3H, C(O)CH₃). ¹³C NMR (CDCl₃, 100 MHz) δ: 170.8 (C=O), 138.1 (2C, C_{Ph}), 134.9 (C-2_{All}), 128.4-127.7 (CH_{Ph}), 117.0 (C-3_{All}), 99.7 (C-1), 80.3 (C-3), 78.4 (C-4), 74.8 (C-5), 74.7 (CH₂Ph), 73.8 (C-1_{All}), 73.5 (CH₂Ph), 68.9 (C-6), 68.3 (OCH₂), 57.4 (C-2), 50.7 (CH₂N₃), 23.7 (C(O)CH₃). HR-ESI-TOF-MS *m/z* 511.2566 [M + H]⁺ (calcd for C₂₇H₃₅N₄O₆, 511.2556).

Allyl 2,3,4,6-tetra-*O*-benzyl-α-D-glucopyranosyl-(1→4)-2-*O*-acetyl-α-L-rhamnopyranoside (16).



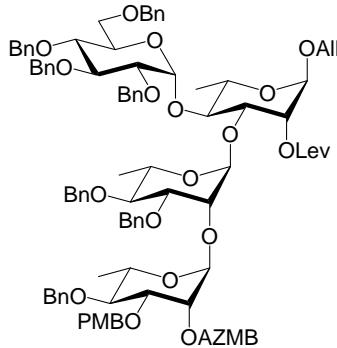
MeC(OMe)_3 (1.04 mL, 8.25 mmol, 2.0 equiv.) and *p*TSA (14 mg, 0.081 mmol, 0.02 equiv.) were added to diol **9**¹⁷ (2.96 g, 4.07 mmol) in anhydrous CH_3CN (8.3 mL). The mixture was stirred for 1 h at rt, then 80% aq. HOAc (8.3 mL) was added at 0 °C. The mixture was stirred for another 1 h at rt, then cold H_2O (50 mL) was added dropwise. The aq. phase was extracted with CH_2Cl_2 (3×100 mL), and the organic layer was washed with brine (50 mL), dried over anhydrous Na_2SO_4 and evaporated to dryness. Crude acetate **16** was obtained as a colorless oil and used without further purification. Available analytical data for acceptor **16**: ^1H NMR (CDCl_3 , 400 MHz) δ : 7.38-7.12 (m, 20H, CH_{Ph}), 5.90 (m, 1H, H-2_{All}), 5.30 (ddd, $J = 17.2, 3.2, 1.6$ Hz, 1H, H-3a_{All}), 5.21 (ddd, $J = 10.4, 2.8, 1.3$ Hz, 1H, H-3b_{All}), 5.14 (dd, $J_{2,3} = 3.7$ Hz, 1H, H-2_C), 4.97 (d, 1H, $J_{1,2} = 3.8$ Hz, H-1_E), 4.95 (d, 1H, $J = 11.0$ Hz, CH_2Ph), 4.82 (d, $J = 10.9$ Hz, 1H, CH_2Ph), 4.81 (d, $J = 10.9$ Hz, 1H, CH_2Ph), 4.77 (d, $J_{1,2} = 1.6$ Hz, 1H, H-1_C), 4.71 (d, 1H, $J = 11.7$ Hz, CH_2Ph), 4.68 (d, 1H, $J = 11.7$ Hz, CH_2Ph), 4.56 (d, 1H, $J = 12.2$ Hz, CH_2Ph), 4.47 (d, 1H, $J = 12.2$ Hz, CH_2Ph), 4.46 (d, $J = 10.9$ Hz, 1H, CH_2Ph), 4.15 (ddt, $J = 12.6, 5.1, 1.5$ Hz, 1H, H-1a_{All}), 4.08 (ddd, $J_{4,5} = 10.1$ Hz, $J_{5,6b} = 4.9$ Hz, 1H, H-5_E), 4.01-3.93 (m, 3H, H-1b_{All}, H-3_C, H-3_E), 3.78 (dq, 1H, $J_{4,5} = 9.5$ Hz, H-5_C), 3.64 (dd, $J_{5,6a} = 2.4$ Hz, $J_{6a,6b} = 10.2$ Hz, 1H, H-6a_E), 3.63-3.56 (m, 2H, H-6b_E, H-2_E), 3.54 (dd, $J_{3,4} = 9.0$ Hz, 1H, H-4_E), 3.37 (pt, $J_{3,4} = 9.2$ Hz, 1H, H-4_C), 2.11 (s, 3H, C(O)CH_3), 1.39 (d, $J_{5,6} = 6.4$ Hz, 3H, H-6_C). ^{13}C NMR (CDCl_3 , 100 MHz) δ : 170.6 (C=O), 138.8 (C_{Ph}), 138.2 (C_{Ph}), 138.1 (C_{Ph}), 137.8 (C_{Ph}), 133.8 (C-2_{All}), 129.2-127.8 (CH_{Ph}), 117.7 (C-3_{All}), 98.8 (C-1_E), 96.7 (C-1_C), 85.4 (C-4_C), 81.9 (C-3_C), 80.2 (C-2_E), 78.1 (C-4_E), 75.8, 75.3 (2C, CH_2Ph), 73.8 (2C, CH_2Ph), 72.4 (C-2_C), 71.4 (C-5_E), 68.9 (C-6_E), 68.4 (C-1_{All}), 68.4 (C-3_E), 66.8 (C-5_C), 21.2 (C(O)CH_3), 17.9 (C-6_C). HR-ESI-TOF-MS m/z 769.3573 [$\text{M} + \text{H}]^+$ (calcd for $\text{C}_{45}\text{H}_{53}\text{O}_{11}$, 769.3588), m/z 791.3387 [$\text{M} + \text{Na}]^+$ (calcd for $\text{C}_{45}\text{H}_{52}\text{O}_{11}\text{Na}$, 791.3408).

Allyl (2-*O*-(2-(azidomethyl)benzoyl)-3,4-di-*O*-benzyl- α -L-rhamnopyranosyl)-(1→3)-[2,3,4,6-tetra-*O*-benzyl- α -D-glucopyranosyl-(1→4)]- α -L-rhamnopyranoside (18).



Route b. Bu_2SnO (24 mg, 0.096 mmol, 2.0 equiv) was added to trisaccharide **17** (60 mg, 0.048 mmol) in anhydrous MeOH (4.0 mL). The mixture was refluxed overnight under Ar. Solvents were evaporated under reduced pressure and the residue was purified by FC (Tol/EtOAc 9:1 to 8:2) to give alcohol **18** (36 mg, 62%) as a colorless oil. See manuscript for *route a* and analytical data.

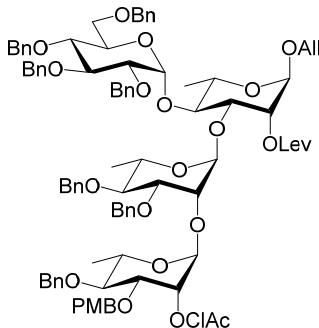
Allyl (2-O-(2-(azidomethyl)benzoyl)-4-O-benzyl-3-O-*para*-methoxybenzyl- α -L-rhamnopyranosyl)-(1 \rightarrow 2)-(3,4-di-O-benzyl- α -L-rhamnopyranosyl)-(1 \rightarrow 3)-[2,3,4,6-tetra-O-benzyl- α -D-glucopyranosyl-(1 \rightarrow 4)]-2-O-levulinoyl- α -L-rhamnopyranoside (21).



Acceptor **20** (5.36 g, 4.66 mmol) and donor **13** (4.74 g, 6.99 mmol, 1.5 equiv.) in anhydrous Et₂O (70 mL) containing 4 Å AW-MS (7.0 g) were stirred for 30 min at -10 °C under Ar. TMSOTf (84 µL, 0.47 mmol, 0.1 equiv.) was added keeping rigorously anhydrous conditions. After stirring for 1 h at -10 °C to rt, Et₃N (300 µL) was added. The suspension was filtered and solvents were evaporated. The residue was purified by FC (Tol/EtOAc 95:5 to 9:1) to give tetrasaccharide **21** as a white sticky solid slightly contaminated with trichloroacetamide (6.18 g), along with unreacted **20** (1.35 g, 20%). Available analytical data for tetrasaccharide **21**: *R*_f 0.45 (Chex/EtOAc 7:3); ¹H NMR (CDCl₃, 400 MHz) δ: 7.94-7.91 (m, 1H, CH_{Ph}), 7.55-7.47 (m, 2H, CH_{Ph}), 7.36-7.02 (m, 38H, CH_{Ph}), 6.80-6.76 (m, 2H, CH_{Ph}), 5.88 (m, 1H, H-2_{All}), 5.74 (dd, *J*_{2,3} = 3.2 Hz, 1H, H-2_A), 5.27 (ddd, *J* = 17.3, 3.2, 1.5 Hz, 1H, H-3a_{All}), 5.19 (ddd, *J* = 10.4, 2.6, 1.2 Hz, 1H, H-3b_{All}), 5.19 (d, *J*_{1,2} = 1.7 Hz, 1H, H-1_A), 5.11 (dd, *J*_{2,3} = 3.5 Hz, *J*_{1,2} = 1.9 Hz, 1H, H-2_C), 4.98 (d, *J*_{1,2} = 3.2 Hz, 1H, H-1_E), 4.96 (d, *J*_{1,2} = 1.6 Hz, 1H, H-1_B), 4.93-4.89 (m, 3H, CH₂Ph), 4.81 (d, *J* = 10.9 Hz, 1H, CH₂Ph), 4.76-4.69 (m, 5H, CH₂N₃a, H-1_C, CH₂Ph (3H)), 4.67-4.44 (m, 8H, CH₂Ph, CH₂N₃b), 4.44 (m, 1H, H-2_B), 4.38 (d, *J* = 10.8 Hz, 1H, CH₂Ph), 4.28 (d, *J* = 12.1 Hz, 1H, CH₂Ph), 4.11 (m, *J* = 13.0, 5.3, 1.5 Hz, 1H, H-1a_{All}), 4.05 (dd, *J*_{3,4} = 9.4 Hz, *J*_{2,3} = 3.1 Hz, 1H, H-3_A), 4.01-3.90 (m, 5H, H-1b_{All}, H-5_A, H-2_E, H-3_C, H-5_E), 3.79-3.67 (m, 7H, OCH₃, H-5_C, H-6a_E, H-6b_E, H-3_B), 3.64 (pt, *J*_{3,4} = *J*_{4,5} = 9.6 Hz, 1H, H-4_E), 3.60-3.49 (m, 3H, H-4_A, H-5_B, H-4_C), 3.47-3.38 (m, 2H, H-3_E, H-4_B), 2.74-2.54 (m, 4H, CH₂Lev), 2.08 (s, 3H, C(O)CH₃), 1.36 (d, *J*_{5,6} = 6.2 Hz, 3H, H-6_A), 1.30 (d, *J*_{5,6} = 6.3 Hz, 3H, H-6_C), 1.20 (d, *J*_{5,6} = 6.2 Hz, 3H, H-6_B). ¹³C NMR (CDCl₃, 100 MHz) δ: 206.3 (C(O)CH₂Lev), 172.3 (C=O), 165.5 (C=O), 159.3 (C_{Ph}), 138.8 (C_{Ph}), 138.8 (C_{Ph}), 138.7 (C_{Ph}), 138.6 (C_{Ph}), 138.3 (C_{Ph}), 138.3 (C_{Ph}), 138.1 (C_{Ph}), 137.7 (C_{Ph}), 133.7 (C-2_{All}), 132.9 (CH_{Ph}), 131.4 (CH_{Ph}), 130.2 (CH_{Ph}), 129.8 (CH_{Ph}), 129.5 (CH_{Ph}), 128.7-127.4 (CH_{Ph}), 117.7 (C-3_{All}), 113.8 (CH_{Ph}), 101.4 (C-1_B), 99.3 (C-1_A), 97.8 (C-1_E), 95.9 (C-1_C), 81.8 (C-5_E), 81.3 (C-3_E), 80.3 (C-4_C), 80.1 (C-4_B), 79.9 (C-3_C), 79.1 (C-3_B), 77.7 (C-4_A), 77.4 (C-3_A, C-4_E), 75.7, 75.4, 75.1 (4C, CH₂Ph), 74.8 (C-2_B), 73.9, 73.0 (2C, CH₂Ph), 72.5 (C-2_C), 71.4 (CH₂Ph), 71.4 (C-5_A), 71.0 (CH₂Ph), 70.1 (C-2_A), 68.9 (C-5_B), 68.5 (C-1_{All}), 68.4 (C-2_E), 68.3 (C-6_E), 67.6

(C-5_C), 55.3 (OCH₃), 53.1 (CH₂N₃), 38.1 (CH₂Lev), 31.0 (C(O)CH₃), 28.3 (CH₂Lev), 18.7 (C-6_C), 18.6 (C-6_A), 18.2 (C-6_B). HR-ESI-TOF-MS *m/z* 1688.7185 [M + Na]⁺ (calcd for C₉₇H₁₀₇N₃O₂₂Na, 1688.7244).

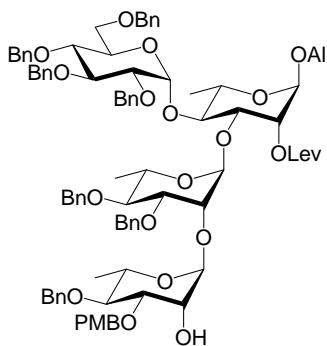
Allyl (4-*O*-benzyl-2-*O*-chloroacetyl-3-*O*-methoxybenzyl- α -L-rhamnopyranosyl)-(1 \rightarrow 2)-(3,4-di-*O*-benzyl- α -L-rhamnopyranosyl)-(1 \rightarrow 3)-[2,3,4,6-tetra-*O*-benzyl- α -D-glucopyranosyl-(1 \rightarrow 4)]-2-*O*-levulinoyl- α -L-rhamnopyranoside (22).



Acceptor **20** (769 mg, 668 μ mol) and donor **14** (791 mg, 1.34 mmol, 2.0 equiv.) in anhydrt. Et₂O (10 mL) containing 4 Å AW-MS were stirred for 30 min at -10 °C under Ar. TMSOTf (12 μ L, 67 μ mol, 0.1 equiv.) was added keeping rigorously anhydrt. conditions. After stirring for 1 h at -10 °C to rt, Et₃N (100 μ L) was added. The suspension was filtered and solvents were evaporated. The residue was purified by FC (Tol/EtOAc 95:5 to 8:2) to give tetrasaccharide **22** (1.03 g) as a white sticky solid slightly contaminated with trichloroacetamide. *R*_f 0.46 (Chex/EtOAc 7:3); ¹H NMR (CDCl₃, 400 MHz) δ : 7.38-7.09 (m, 37H, CH_{Ph}), 6.84-6.80 (m, 2H, CH_{Ph}), 5.89 (m, 1H, H-2_{All}), 5.57 (dd, *J*_{2,3} = 2.9 Hz, *J*_{1,2} = 1.8 Hz, 1H, H-2_A), 5.29 (ddd, *J* = 17.3, 3.1, 1.5 Hz, 1H, H-3a_{All}), 5.20 (ddd, *J* = 10.4, 2.6, 1.3 Hz, 1H, H-3b_{All}), 5.11 (dd, *J*_{2,3} = 3.0 Hz, *J*_{1,2} = 2.1 Hz, 1H, H-2_C), 5.06 (d, *J*_{1,2} = 1.3 Hz, 1H, H-1_A), 5.01 (d, *J*_{1,2} = 3.1 Hz, 1H, H-1_E), 4.95 (br s, 1H, H-1_B), 4.94 (d, *J* = 11.1 Hz, 1H, CH₂Ph), 4.89 (d, *J* = 11.0 Hz, 1H, CH₂Ph), 4.88 (d, *J* = 10.9 Hz, CH₂Ph), 4.85-4.74 (m, 4H, CH₂Ph, H-1_C), 4.67-4.52 (m, 7H, CH₂Ph), 4.47 (d, *J* = 11.0 Hz, 1H, CH₂Ph), 4.45 (d, *J* = 11.0 Hz, 1H, CH₂Ph), 4.40 (bdd, *J*_{2,3} = *J*_{1,2} = 2.0 Hz, 1H, H-2_B), 4.36 (d, *J* = 12.0 Hz, 1H, CH₂Ph), 4.13 (m, *J* = 12.8, 5.3, 1.6 Hz, 1H, H-1a_{All}), 4.04-3.90 (m, 7H, CH₂Cla, H-5_A, H-1b_{All}, H-5_E, H-3_C, H-3_A, H-3_E), 3.84 (d, *J* = 15.0 Hz, 1H, CH₂Clb), 3.81-3.66 (m, 8H, H-6a_E, H-6b_E, H-4_E, H-3_B, H-5_C, OCH₃), 3.62-3.53 (m, 2H, H-4_A, H-5_B), 3.49 (dd, *J*_{3,4} = 9.7 Hz, *J*_{3,4} = 3.3 Hz, 1H, H-2_E), 3.37 (pt, *J*_{3,4} = *J*_{4,5} = 9.5 Hz, 2H, H-4_B, H-4_C), 2.77-2.53 (m, 4H, CH₂Lev), 2.08 (s, 3H, C(O)CH₃), 1.33 (d, *J*_{5,6} = 6.1 Hz, 6H, H-6_A, H-6_C), 1.20 (d, *J*_{5,6} = 6.1 Hz, 3H, H-6_B). ¹³C NMR (CDCl₃, 100 MHz) δ : 206.2 (C(O)CH₂Lev), 172.3 (C=O_{Lev}), 166.5 (C=O_{AcCl}), 159.3 (C_{Ph}), 138.8-137.9 (C_{Ph}), 133.7 (C-2_{All}), 130.1-125.4 (CH_{Ph}), 117.8 (C-3_{All}), 113.9 (CH_{Ph}), 101.4 (C-1_B), 99.0 (C-1_A), 97.7 (C-1_E), 95.9 (C-1_C), 81.8 (C-3_E), 81.1 (C-2_E), 80.0 (C-4_B*), 79.9 (C-4_C*), 79.7 (C-3_C), 79.0 (C-3_B), 77.6 (C-4_A), 77.4 (C-4_E), 77.2 (C-3_A), 75.7, 75.4, 75.3, 75.1 (4C, CH₂Ph), 74.8 (C-2_B), 73.9, 72.9 (2C, CH₂Ph), 72.5 (C-2_C), 71.7 (CH₂Ph), 71.3 (C-5_A), 71.1 (C-2_A), 70.9 (CH₂Ph), 68.8 (C-5_B), 68.5 (C-1_{All}), 68.4 (C-5_E), 68.2 (C-6_E), 67.5 (C-5_C), 55.3 (OCH₃), 41.0

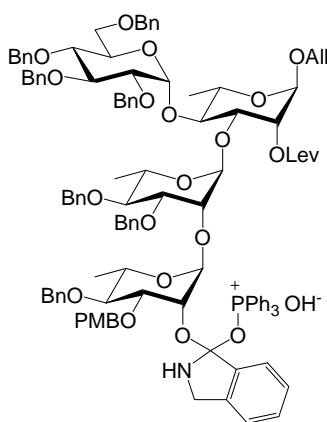
(CH₂Cl), 38.0 (CH₂_{Lev}), 29.9 (C(O)CH₃), 28.2 (CH₂_{Lev}), 18.7 (C-6_A*), 18.4 (C-6_C*), 18.1 (C-6_B). HR-ESI-TOF-MS *m/z* 1583.6742 [M + H]⁺ (calcd for C₉₁H₁₀₄ClO₂₂, 1583.6708), *m/z* 1605.6541 [M + Na]⁺ (calcd for C₉₁H₁₀₃ClO₂₂Na, 1605.6527).

Allyl (4-*O*-benzyl-3-*O*-*para*-methoxybenzyl- α -L-rhamnopyranosyl)-(1 \rightarrow 2)-(3,4-di-*O*-benzyl- α -L-rhamnopyranosyl)-(1 \rightarrow 3)-[2,3,4,6-tetra-*O*-benzyl- α -D-glucopyranosyl-(1 \rightarrow 4)]-2-*O*-levulinoyl- α -L-rhamnopyranoside (10).



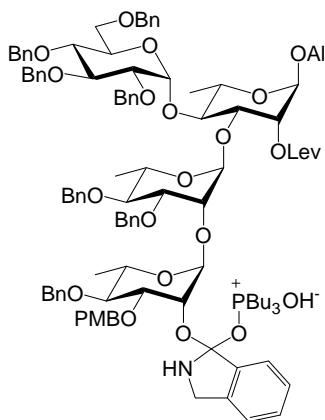
Route b. PPh₃ (4.84 g, 18.4 mmol, 5.0 equiv.) and SiO₂ (1.54 g) were added to tetrasaccharide **21** (6.15 g) in THF/H₂O (36.9 mL, 9:1 v/v). The mixture was stirred for 24 h at rt, then additional portions of PPh₃ (4.84 g, 18.4 mmol, 5.0 equiv.) and SiO₂ (1.54 g) were added. The mixture was stirred overnight at rt, filtered, washed with several portions of CH₂Cl₂ and solvents were evaporated. The residue was purified by FC (Tol/EtOAc 95:5 to 8:2) to give acceptor **10** (3.39 g, 60% from **20**) as a white foam along with unreacted **21** (476 mg, 8%). See manuscript for *route a* and analytical data.

Allyl (4-*O*-benzyl-3-*O*-*para*-methoxybenzyl-2-*O*-(1-*O*-(triphenylphosphonium)isoindol-1-yl)- α -L-rhamnopyranosyl)-(1 \rightarrow 2)-(3,4-di-*O*-benzyl- α -L-rhamnopyranosyl)-(1 \rightarrow 3)-[2,3,4,6-tetra-*O*-benzyl- α -D-glucopyranosyl-(1 \rightarrow 4)]-2-*O*-levulinoyl- α -L-rhamnopyranoside (21a).



Eluting the chromatographic column of the precedent reaction (*route a*, see manuscript) using CH₂Cl₂/MeOH 95:5 to 100% MeOH gave **21a** (632 mg, 9%) as a white foam. *R*_f 0.16 (CH₂Cl₂/MeOH 95:5); [α]_D²⁵ +15° (c 1.0, CHCl₃); ¹H NMR (CDCl₃, 400 MHz) δ: 9.51 (m, *J* = 7.4 Hz, 1H, NH), 8.16-8.12 (m, 1H, CH_{Ph}), 7.82-7.03 (m, 55H, CH_{Ph}), 6.79-6.75 (m, 2H, CH_{Ph}), 5.89 (m, 1H, H-2_{AII}), 5.54 (dd, *J*_{2,3} = 3.3 Hz, *J*_{1,2} = 2.0 Hz, 1H, H-2_A), 5.29 (m, *J* = 17.2, 3.0, 1.5 Hz, 1H, H-3a_{AII}), 5.20 (m, *J* = 10.4, 2.6, 1.3 Hz, 1H, H-3b_{AII}), 5.13-5.10 (m, 2H, H-2_C, H-1_A), 5.01-4.72 (m, 11H, CH₂NH, H-1_E, H-1_B, H-1_C, CH₂Ph), 4.66-4.37 (m, 10H, CH₂Ph (9H), H-2_B), 4.29 (d, *J* = 11.8 Hz, 1H, CH₂Ph), 4.13 (m, *J* = 12.9, 5.3, 1.4 Hz, 1H, H-1a_{AII}), 4.02-3.91 (m, 6H, H-5_A, H-1b_{AII}, H-3_A, H-3_C, H-5_E, H-2_E), 3.81-3.69 (m, 7H, OCH₃, H-5_C, H-6a_E, H-6b_E, H-3_B), 3.68-3.52 (m, 3H, H-4_E, H-4_A, H-5_B), 3.50-3.42 (m, 2H, H-4_C, H-3_E), 3.67 (pt, *J*_{3,4} = *J*_{4,5} = 9.3 Hz, 1H, H-4_B), 2.76-2.54 (m, 4H, CH₂Lev), 2.08 (s, 3H, CH₃), 1.35 (d, *J*_{5,6} = 6.2 Hz, 3H, C-6_A), 1.33 (d, *J*_{5,6} = 6.2 Hz, 3H, C-6_C), 1.20 (d, *J*_{5,6} = 6.2 Hz, 3H, C-6_B). ¹³C NMR (CDCl₃, 100 MHz) δ: 205.9 (C(O)CH₂Lev), 172.2 (C(O)CH₂Lev), 165.5 (COPPh₃⁺), 159.3 (C_{Ph}), 141.4 (C_{Ph}), 141.3 (C_{Ph}), 138.8 (C_{Ph}), 138.8 (C_{Ph}), 138.6 (C_{Ph}), 138.6 (C_{Ph}), 138.4 (C_{Ph}), 138.3 (C_{Ph}), 138.0 (C_{Ph}), 134.6 (C-2_{AII}), 133.7-126.9 (CH_{Ph}), 117.6 (C-3_{AII}), 113.9 (CH_{Ph}), 101.3 (C-1_B), 99.1 (C-1_A), 97.8 (C-1_E), 96.0 (C-1_C), 81.8 (C-5_E), 81.3 (C-3_E), 80.3 (C-4_C), 80.1 (C-4_B), 79.8 (C-3_C), 79.1 (C-3_B), 78.0 (C-4_A), 77.8 (C-4_E), 77.0 (C-3_A), 75.7, 75.2, 75.1 (4C, CH₂Ph), 74.4 (C-2_B), 73.9, 73.0 (2C, CH₂Ph), 72.5 (C-2_C), 71.4 (C-5_A), 71.2, 70.9 (2C, CH₂Ph), 69.9 (C-2_A), 68.9 (C-5_B), 68.5 (C-1_{AII}), 68.5 (C-2_E), 68.3 (C-6_E), 67.6 (C-5_C), 55.4 (OCH₃), 43.2 (CH₂NH), 38.1 (CH₂Lev), 29.8 (C(O)CH₃), 28.3 (CH₂Lev), 18.7 (C-6_A), 18.6 (C-6_C), 18.1 (C-6_B). ³¹P NMR (CDCl₃, 100 MHz) δ: 40.1 ppm (s, PPh₃). HR-ESI-TOF-MS *m/z* 1900.8340 [M]⁺ (calcd for C₁₁₅H₁₂₃NO₂₂P, 1900.8274).

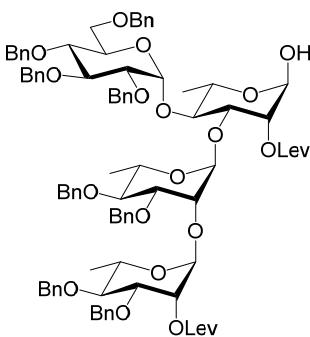
Allyl (4-*O*-benzyl-3-*O*-para-methoxybenzyl-2-*O*-(1-*O*-(tributylphosphonium)isoindol-1-yl)- α -L-rhamnopyranosyl)-(1→2)-(3,4-di-*O*-benzyl- α -L-rhamnopyranosyl)-(1→3)-[2,3,4,6-tetra-*O*-benzyl- α -D-glucopyranosyl-(1→4)]-2-*O*-levulinoyl- α -L-rhamnopyranoside (21b).



PBu₃ (225 μL, 0.90 mmol, 3.0 equiv.) was added to tetrasaccharide **21** (500 mg) in anhydrous THF (10 mL). The mixture was stirred for 2 h at rt, then H₂O (27 μL, 1.5 mmol, 5.0 equiv.) was added. The mixture was stirred overnight at rt and solvents were evaporated. The residue was purified by

FC ($\text{CH}_2\text{Cl}_2/\text{MeOH}$ 95:5 to 9:1) to give **21b** (245 mg, 49% from **21**) as a white foam. R_f 0.15 ($\text{CH}_2\text{Cl}_2/\text{MeOH}$ 95:5); $[\alpha]^{25}_{\text{D}} +13^\circ$ (c 1.0, CHCl_3); ^1H NMR (Py-d₅, 400 MHz) δ : 9.41 (m, $J_{2,\text{NH}} = 7.6$ Hz, 1H, NH), 8.34-8.30 (m, 1H, CH_{Ph}), 7.63-7.20 (m, 40H, CH_{Ph}), 7.04-6.99 (m, 2H, CH_{Ph}), 6.28 (br s, 1H, H-2_A), 5.85 (m, 1H, H-2_{All}), 5.81 (br s, 1H, H-1_A), 5.71 (br s, 1H, H-2_C), 5.64 (br s, 1H, H-1_B), 5.49 (d, 1H, H-1_E), 5.28-4.63 (m, 22H, CH_2Ph , H-3a_{All}, H-1_C, H-3b_{All}, H-2_B, CH_2NHa , CH_2NHb), 4.57 (dd, $J_{3,4} = 8.9$ Hz, $J_{2,3} = 2.4$ Hz, 1H, H-3_C), 4.49 (dd, $J_{3,4} = 9.3$ Hz, $J_{2,3} = 2.9$ Hz, 1H, H-3_A), 4.46-4.38 (m, 2H, H-5_A, H-5_C), 4.36-4.27 (m, 2H, H-3_E, H-3_B), 4.22-4.04 (m, 6H, H-1a_{All}, H-6a_E, H-4_C, H-5_B, H-5_E, H-6b_E), 4.03-3.90 (m, 4H, H-1b_{All}, H-4_E, H-4_A, H-4_B), 3.82 (dd, $J_{2,3} = 9.8$ Hz, $J_{1,2} = 3.0$ Hz, 1H, H-2_E), 3.66 (s, 3H, OCH_3), 2.82-2.69 (m, 4H, CH_2Lev), 2.47-2.37 (m, 6H, CH_2Bu), 2.03 (s, 3H, CH_3Lev), 1.64 (d, $J_{5,6} = 6.2$ Hz, 3H, H-6_B), 1.63-55 (m, 6H, CH_2Bu), 1.52 (d, 3H, $J_{5,6} = 6.2$ Hz, H-6_C), 1.50 (d, 3H, $J_{5,6} = 6.2$ Hz, H-6_A), 1.38-1.26 (m, 6H, CH_2Bu), 0.82-0.77 (m, 9H, CH_3Bu). ^{13}C NMR (Py-d₅, 100 MHz) δ : 206.1 ($\text{C}(\text{O})\text{CH}_2\text{Lev}$), 173.1 ($\text{C}(\text{O})\text{CH}_2\text{Lev}$), 167.1 (COPBu_3^+), 160.2 (C_{Ph}), 143.0-139.4 (C_{Ph}), 134.7 (C-2_{All}), 133.9-128.2 (CH_{Ph}), 117.7 (C-3_{All}), 114.8 (CH_{Ph}), 102.4 (C-1_B), 100.1 (C-1_A), 98.6 (C-1_E), 96.8 (C-1_C), 82.6 (C-3_E), 82.3 (C-2_E), 81.2 (C-4_A), 80.9 (C-4_B), 80.7 (C-3_C), 80.2 (C-3_B), 79.0 (C-4_C), 78.8 (C-4_E), 78.3 (C-3_A), 76.1, 75.8, 75.6, 75.5 (4C, CH_2Ph), 75.5 (C-2_B), 74.3, 73.7 (2C, CH_2Ph), 73.4 (C-2_C), 72.4 (C-5_A), 72.0, 71.8 (2C, CH_2Ph), 71.1 (C-2_A), 69.8 (C-5_B), 69.7 (C-6_E), 69.4 (C-5_C), 68.8 (C-1_{All}), 68.6 (C-5_E), 55.7 (OCH_3), 42.4 (CH_2NH), 38.5 (CH_2Lev), 29.9 ($\text{C}(\text{O})\text{CH}_3$), 29.1 (CH_2Lev), 24.5 (CH_2Bu), 24.4 (CH_2Bu), 23.8 (CH_2Bu), 22.6 (CH_2Bu), 22.0 (CH_2Bu), 19.7, 19.0, 19.0 (3C, C-6_A, C-6_B, C-6_C), 13.9 (CH_3Bu). HR-ESI-TOF-MS m/z 1840.9080 [M]⁺ (calcd for $\text{C}_{109}\text{H}_{135}\text{NO}_{22}\text{P}$, 1841.9247).

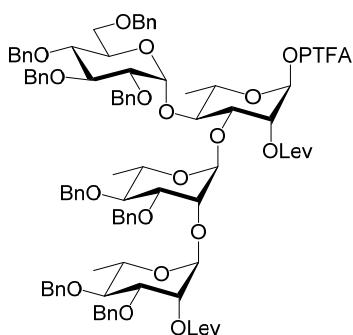
(3,4-Di-O-benzyl-2-O-levulinyl- α -L-rhamnopyranosyl)-(1 \rightarrow 2)-(3,4-di-O-benzyl- α -L-rhamnopyranosyl)-(1 \rightarrow 3)-[2,3,4,6-tetra-O-benzyl- α -D-glucopyranosyl-(1 \rightarrow 4)]-2-O-levulinyl- α -L-rhamnopyranose (24).



[Ir] (91 mg, 0.11 mmol, 0.1 equiv.) was dissolved in anhydrous THF (5.4 mL) and the red solution was degassed. Hydrogen was bubbled through the solution for 15 min, and the resulting yellow solution was once again degassed. Tetrasaccharide **23** (1.69 g, 1.07 mmol) in anhydrous THF (5.4 mL) was added and the mixture was stirred for 20 h at rt under Ar. Iodine (544 mg, 2.14 mmol, 2.0 equiv.) in THF/H₂O (6.4 mL, 4:1 v/v) was added and the mixture was stirred for 1 h at rt. Freshly prepared 10% aq. NaHSO₃ was added and the mixture was diluted with CH₂Cl₂ (250 mL)

and water (100 mL). The aq. phase was extracted with CH_2Cl_2 (3×250 mL) and the organic layer was dried over anhydr. Na_2SO_4 , filtered and evaporated to dryness. The residue was purified by FC (Tol/EtOAc 8:2 to 7:3) to give hemiacetal **24** (1.52 g, 92%, ratio $\alpha/\beta > 20:1$) as a light yellow foam. R_f 0.19 (Chex/EtOAc 7:3); $[\alpha]^{25}_D +22^\circ$ (c 1.0, CHCl_3); ^1H NMR (CDCl_3 , 400 MHz) δ (α): 7.37-7.11 (m, 40H, CH_{Ph}), 5.56 (dd, $J_{2,3} = 3.3$ Hz, $J_{1,2} = 1.8$ Hz, 1H, H- 2_A), 5.12 (br d, 1H, H- 2_C), 5.10-5.06 (m, 2H, H- 1_C , H- 1_A), 5.00-4.90 (m, 5H, H- 1_E , H- 1_B , CH_2Ph), 4.85 (d, $J = 11.0$ Hz, 1H, CH_2Ph), 4.81 (d, $J = 11.0$ Hz, 1H, CH_2Ph), 4.75 (d, $J = 11.7$ Hz, 1H, CH_2Ph), 4.72 (d, $J = 11.4$ Hz, 1H, CH_2Ph), 4.67-4.51 (m, 7H, CH_2Ph), 4.48 (d, $J = 10.8$ Hz, 1H, CH_2Ph), 4.41-4.36 (m, 2H, H- 2_B , CH_2Ph), 4.05-3.91 (m, 6H, H- 3_C , H- 5_A , H- 5_E , H- 3_A , H- 3_E , H- 5_C), 3.82-3.68 (m, 4H, H- 6_a_E , H- 6_b_E , H- 3_B , H- 4_E), 3.62-3.53 (m, 2H, H- 5_B , H- 4_A), 3.48 (dd, $J_{2,3} = 9.7$ Hz, $J_{1,2} = 3.1$ Hz, 1H, H- 2_E), 3.43 (pt, $J_{3,4} = J_{4,5} = 9.3$ Hz, 1H, H- 4_C), 3.40 (pt, $J_{3,4} = J_{4,5} = 9.3$ Hz, 1H, H- 4_B), 3.06 (d, $J_{OH,1} = 4.2$ Hz, 1H, OH- 1_C), 2.76-2.47 (m, 8H, $\text{CH}_{2\text{Lev}}$), 2.08 (s, 6H, $\text{CH}_{3\text{Lev}}$), 1.35 (d, $J_{5,6} = 6.2$ Hz, 3H, H- 6_A), 1.32 (d, $J_{5,6} = 6.3$ Hz, 3H, H- 6_C), 1.21 (d, $J_{5,6} = 6.2$ Hz, 3H, H- 6_B). ^{13}C NMR (CDCl_3 , 100 MHz) δ (α): 206.0 (2C, $C(\text{O})\text{CH}_{2\text{Lev}}$), 172.2 ($C=O$), 171.8 ($C=O$), 138.9-138.3 (C_{Ph}), 128.7-127.4 (CH_{Ph}), 101.1 (C- 1_B), 99.4 (C- 1_A), 97.8 (C- 1_E), 91.3 (C- 1_C), 81.9 (C- 3_E), 81.4 (C- 2_E), 80.2 (C- 4_C), 80.1 (C- 4_B), 79.0 (C- 3_C , C- 3_B), 78.2 (C- 4_A), 77.8 (2C, C- 3_A , C- 4_E), 75.7, 75.4, 75.4, 75.2 (4C, CH_2Ph), 74.9 (C- 2_B), 73.9, 73.1 (4C, CH_2Ph), 72.8 (C- 2_C), 71.7 (CH_2Ph), 71.5 (C- 5_A), 70.9 (CH_2Ph), 69.5 (C- 2_A), 68.9 (C- 5_B), 68.5 (C- 6_E), 68.4 (C- 5_E), 67.6 (C- 5_C), 38.2 ($\text{CH}_{2\text{Lev}}$), 38.1 ($\text{CH}_{2\text{Lev}}$), 29.8 ($C(\text{O})\text{CH}_3$), 29.8 ($C(\text{O})\text{CH}_3$), 28.3 ($\text{CH}_{2\text{Lev}}$), 28.2 ($\text{CH}_{2\text{Lev}}$), 18.9 (C- 6_A), 18.5 (C- 6_C), 18.1 (C- 6_B). HR-ESI-TOF-MS m/z 1535.6907 [$M + \text{H}]^+$ (calcd for $\text{C}_{90}\text{H}_{103}\text{O}_{22}$, 1535.6941), m/z 1557.6749 [$M + \text{Na}]^+$ (calcd for $\text{C}_{90}\text{H}_{102}\text{O}_{22}\text{Na}$, 1557.6760).

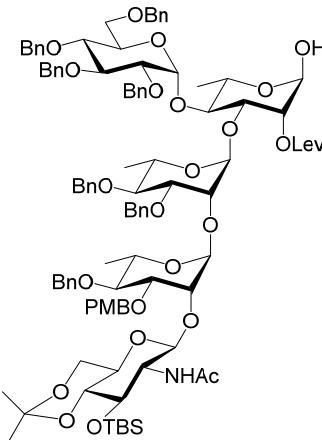
(3,4-Di-O-benzyl-2-O-levulinyl- α -L-rhamnopyranosyl)-(1 \rightarrow 2)-(3,4-di-O-benzyl- α -L-rhamnopyranosyl)-(1 \rightarrow 3)-[2,3,4,6-tetra-O-benzyl- α -D-glucopyranosyl-(1 \rightarrow 4)]-2-O-levulinyl- α -L-rhamnopyranosyl N-(phenyl)trifluoroacetimidate (6).



PTFACl¹⁸ (254 mg, 1.22 mmol, 2.0 equiv.) and Cs_2CO_3 (219 mg, 673 μmol , 1.1 equiv.) were added to hemiacetal **24** (940 mg, 612 μmol) in acetone (6.1 mL). The mixture was stirred for 3 h at rt, filtered and evaporated to dryness. The residue was purified by FC (Tol/EtOAc 95:5 to 9:1 + 1% Et_3N) to furnish PTFA **6** (971 mg, 93%) as a white foam. R_f 0.47 (Chex/EtOAc 7:3); $[\alpha]^{25}_D +11^\circ$ (c 1.0, CHCl_3); ^1H NMR (Py-d_5 , 400 MHz) δ : 7.67-7.10 (m, 45H, CH_{Ph}), 7.00 (bs, 1H, H- 1_C), 6.09 (br s, 1H, H- 2_A), 5.89 (br s, 1H, H- 2_C), 5.72-5.68 (m, 2H, H- 1_A , H- 1_B), 5.50 (bd, $J_{1,2} =$

3.0 Hz, 1H, H-1_E), 5.25-5.07 (m, 5H, CH₂Ph), 5.02-4.77 (m, 11H, CH₂Ph, H-2_B), 4.73 (d, *J* = 12.0 Hz, 1H, CH₂Ph), 4.70-4.65 (m, 1H, H-3_C), 4.61 (d, *J* = 11.2 Hz, 1H, CH₂Ph), 4.45-4.08 (m, 10H, H-5_E, H-5_A, H-3_A, H-3_E, H-3_B, H-4_C, H-5_C, H-5_B, H-6a_E, H-6b_E), 4.01 (pt, *J*_{3,4} = *J*_{4,5} = 9.5 Hz, 1H, H-4_E), 3.95 (pt, *J*_{3,4} = *J*_{4,5} = 9.3 Hz, 1H, H-4_B), 3.90-3.83 (m, 2H, H-4_A, H-2_E), 2.90-2.70 (m, 8H, CH₂Lev), 2.03 (s, 3H, CH₃Lev), 2.02 (s, 3H, CH₃Lev), 1.64 (d, *J*_{5,6} = 5.5 Hz, 3H, H-6_C), 1.53 (d, *J*_{5,6} = 6.1 Hz, 3H, H-6_B), 1.52 (d, *J*_{5,6} = 6.0 Hz, 3H, H-6_A). ¹³C NMR (Py-d₅, 100 MHz) δ: 206.3 (C(O)CH₂Lev), 206.0 (C(O)CH₂Lev), 172.9 (C=O), 172.8 (C=O), 144.3 (C=NPh), 140.2-139.2 (C_{Ph}), 129.7-128.2 (CH_{Ph}), 120.4 (CH_{Ph}), 102.4 (C-1_B), 100.5 (C-1_A), 99.0 (C-1_E), 82.5 (C-3_E), 82.1 (C-2_E), 81.0 (C-4_A), 80.6 (C-4_B), 79.9 (C-3_C), 79.8 (C-3_B), 78.8 (C-3_A), 78.5 (C-4_E), 77.9 (C-4_C), 76.0, 75.9 (2C, CH₂Ph), 75.8 (C-2_B), 75.6, 75.5, 74.3, 73.8 (4C, CH₂Ph), 72.5 (C-5_E), 72.0 (CH₂Ph), 71.6 (C-2_C), 71.6 (CH₂Ph), 71.4 (C-5_C), 70.1 (C-2_A), 70.0 (C-5_B), 69.5 (C-6_E), 69.3 (C-5_A), 38.4 (CH₂Lev), 38.3 (CH₂Lev), 29.9 (CH₃Lev), 29.8 (CH₃Lev), 28.9 (CH₂Lev), 28.8 (CH₂Lev), 19.6 (C-6_C), 18.9 (C-6_B*), 18.7 (C-6_A*).

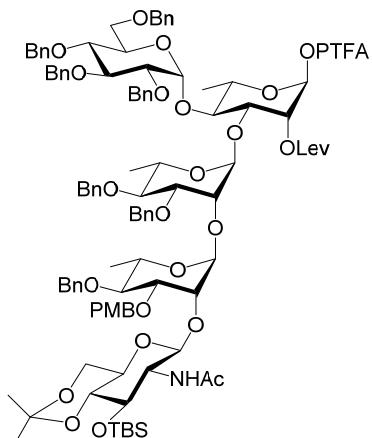
(2-Acetamido-3-*O*-*tert*-butyldimethylsilyl-2-deoxy-4,6-*O*-isopropylidene- β -D-glucopyranosyl)-(1→2)-(4-*O*-benzyl-3-*O*-*para*-methoxybenzyl- α -L-rhamnopyranosyl)-(1→2)-(3,4-di-*O*-benzyl- α -L-rhamnopyranosyl)-(1→3)-[2,3,4,6-tetra-*O*-benzyl- α -D-glucopyranosyl-(1→4)]-2-*O*-levulinyl- α -L-rhamnopyranose (29).



Bu₃SnH (2.05 mL, 7.62 mmol, 10.0 equiv.) and AIBN (125 mg, 0.762 mmol, 1.0 equiv.) were added to allyl glycoside **27** (1.50 g, 0.762 mmol) in anhydrous toluene (30.5 mL). After stirring for 2 h at reflux under Ar, solvents were evaporated. The residue was purified by FC (Tol/EtOAc 95:5 to 9:1 + 1% Et₃N) to give acetamide **28** contaminated with tin salts (1.42 g) as a white foam. [Ir] (64 mg, 76 μmol, 0.10 equiv.) was dissolved in anhydrous THF (3.8 mL) and the red solution was degassed. Hydrogen was bubbled through the solution for 15 min and the resulting yellow solution was once again degassed. The crude **28** in anhydrous THF (3.8 mL) was added. After stirring for 16 h at rt under Ar, iodine (387 mg, 1.52 mmol, 2.0 equiv.) in THF/H₂O (4.6 mL, 4:1 v/v) was added. After an additional 1 h at rt, freshly prepared 10% aq. NaHSO₃ was added. The mixture was partitioned between CH₂Cl₂ (100 mL) and water (50 mL) and the aq.

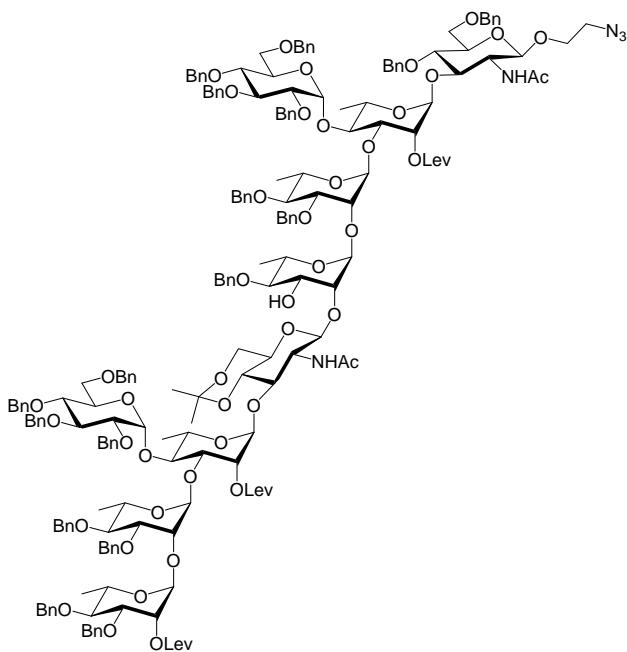
phase was extracted with CH_2Cl_2 (3×100 mL). The organic layer was dried over anhydrous Na_2SO_4 , filtered and evaporated to dryness. The residue was purified by FC (Tol/EtOAc 9:1 to 6:4 + 1% Et_3N) to give hemiacetal **29** (810 mg, 58% from **27**) as a white foam. R_f 0.15 (Chex/EtOAc 7:3); $[\alpha]^{25}_{\text{D}} +12^\circ$ (c 0.5, CHCl_3); ^1H NMR (Py-d₅, 400 MHz) δ : 9.27 (br s, 1H, NH), 7.68-7.26 (m, 37H, CH_{Ph}), 7.01-6.96 (m, 2H, CH_{Ph}), 5.85 (br s, 1H, H-2_C), 5.76-5.72 (m, 2H, H-1_A, H-1_C), 5.65 (d, $J_{1,2} = 7.2$ Hz, 1H, H-1_{D'}), 5.58 (br s, 1H, H-1_B), 5.48 (d, $J_{1,2} = 2.7$ Hz, 1H, H-1_E), 5.29-4.97 (m, 8H, CH_2Ph , H-2_B), 4.96-4.62 (m, 12H, CH_2Ph , H-3_C, H-2_A, H-3_{D'}), 4.55 (dq, 1H, H-5_C), 4.43 (br d, $J = 9.9$ Hz, 1H, H-5_E), 4.39-4.02 (m, 9H, H-3_E, H-3_A, H-3_B, H-5_A, H-6a_E, H-4_C, H-5_B, H-6b_E, H-4_E), 3.98 (pt, $J_{3,4} = J_{4,5} = 9.4$ Hz, 1H, H-4_A), 3.88 (pt, $J_{3,4} = J_{4,5} = 9.4$ Hz, 1H, H-4_B), 3.85-3.73 (m, 3H, H-2_E, H-2_{D'}, H-6a_{D'}), 3.58 (s, 3H, OCH_3), 3.56-3.42 (m, 3H, H-6b_{D'}, H-5_{D'}, H-4_{D'}), 2.83-2.65 (m, 4H, $\text{CH}_{2\text{Lev}}$), 2.28 (s, 3H, C(O) CH_3Ac), 1.99 (s, 3H, C(O) CH_3Lev), 1.68 (d, $J_{5,6} = 6.1$ Hz, 3H, H-6_C), 1.52 (d, $J_{5,6} = 6.0$ Hz, 3H, H-6_B), 1.46 (s, 3H, CH_3Iso), 1.39 (d, $J_{5,6} = 6.0$ Hz, 3H, H-6_A), 1.29 (s, 3H, CH_3Iso), 1.02 (s, 9H, C(CH_3)₃), 0.24 (s, 3H, Si(CH_3)₂), 0.21 (s, 3H, Si(CH_3)₂). ^{13}C NMR (Py-d₅, 100 MHz) δ : 206.0 (C(O) $\text{CH}_{2\text{Lev}}$), 173.2 (C=O), 170.7 (C(O) CH_3Ac), 160.3 (C_{Ph}), 140.3-139.7 (C_{Ph}), 131.6-128.2 (CH_{Ph}), 114.7 (CH_{Ph}), 102.9 (C-1_B), 102.8 (C-1_{D'}), 102.5 (C-1_A), 99.8 (C(CH_3)₂), 98.6 (C-1_E), 92.1 (C-1_C), 82.6 (C-3_E), 82.6 (C-2_E), 81.7 (C-4_A), 81.3 (C-3_C, C-4_B), 80.2 (C-3_A), 79.7 (C-4_C), 79.6 (C-3_B), 78.4 (C-4_E), 76.7 (C-2_A), 76.1 (2C, CH_2Ph), 75.8 (C-4_{D'}), 75.8 (CH_2Ph), 75.5 (C-2_B), 75.3 (CH_2Ph), 75.0 (C-2_C), 74.2, 74.0, 72.8 (3C, CH_2Ph), 72.7 (C-3_{D'}), 72.5 (C-5_E), 71.8 (CH_2Ph), 69.8 (C-5_B), 69.7 (C-5_A), 69.6 (C-6_E), 67.9 (C-5_C), 67.5 (C-5_{D'}), 62.9 (C-6_{D'}), 60.4 (C-2_{D'}), 55.5 (OCH_3), 38.6 ($\text{CH}_{2\text{Lev}}$), 29.9 (C(O) CH_3Lev), 29.7 (CH₃Iso), 29.2 ($\text{CH}_{2\text{Lev}}$), 26.5 (C(CH_3)₃), 24.5 (C(O) CH_3Ac), 20.1 (C-6_C), 19.6 (CH₃Iso), 19.0 (C-6_B), 18.9 (C(CH_3)₃), 18.8 (C-6_A), -3.4 (Si(CH_3)₂), -4.2 (Si(CH_3)₂). HR-ESI-TOF-MS m/z 1824.8639 [M + H]⁺ (calcd for C₁₀₃H₁₃₀NO₂₆Si, 1824.8650), m/z 1846.8492 [M + Na]⁺ (calcd for C₁₀₃H₁₂₉NO₂₆SiNa, 1846.8469).

(2-Acetamido-3-*O*-*tert*-butyldimethylsilyl-2-deoxy-4,6-*O*-isopropylidene- β -D-glucopyranosyl)-(1 \rightarrow 2)-(4-*O*-benzyl-3-*O*-*para*-methoxybenzyl- α -L-rhamnopyranosyl)-(1 \rightarrow 2)-(3,4-di-*O*-benzyl- α -L-rhamnopyranosyl)-(1 \rightarrow 3)-[2,3,4,6-tetra-*O*-benzyl- α -D-glucopyranosyl-(1 \rightarrow 4)]-2-*O*-levulinyl- α -L-rhamnopyranosyl *N*-(phenyl)trifluoroacetimidate (7).



N-(Phenyl)trifluoroacetimidoyl chloride¹⁸ (177 mg, 0.851 mmol, 2.0 equiv.) and Cs₂CO₃ (153 mg, 468 µmol, 1.1 equiv.) were added to a solution of hemiacetal **29** (777 mg, 0.426 mmol) in acetone (2.1 mL). The mixture was stirred for 2 h at rt, filtered and evaporated to dryness. The residue was purified by flash chromatography (Tol/EtOAc 95:5 to 9:1 + 1% Et₃N) to furnish donor **7** (753 mg, 89%) as a white foam. *R*_f 0.48 (Chex/EtOAc 7:3); [α]²⁵_D +4° (*c* 0.5, CHCl₃); ¹H NMR (Py-d₅, 400 MHz) δ: 7.67-7.09 (m, 42H, CH_{Ph}), 7.03-6.97 (m, 4H, CH_{Ph}, NH, H-1_C), 5.86 (br s, 1H, H-2_C), 5.73 (br s, 1H, H-1_A), 5.65-5.59 (m, 2H, H-1_{D'}, H-1_B), 5.46 (d, 1H, *J*_{1,2} = 2.7 Hz, H-1_E), 5.23-4.57 (m, 18H, CH₂Ph, H-2_B, H-2_A), 4.67-4.53 (m, 2H, H-3_{D'}, H-3_C), 4.38 (br d, *J* = 10.0 Hz, 1H, H-5_E), 4.35-4.13 (m, 7H, H-3_A, H-5_A, H-3_E, H-5_C, H-3_B, H-6a_E, H-4_C), 4.13-3.95 (m, 4H, H-5_B, H-6b_E, H-4_E, H-4_B), 3.88 (pt, *J*_{3,4} = *J*_{4,5} = 9.1 Hz, 1H, H-4_A), 3.85-3.77 (m, 3H, H-2_E, H-2_{D'}, H-6a_{D'}), 3.63 (s, 3H, OCH₃), 3.61-3.44 (m, 3H, H-6b_{D'}, H-5_{D'}, H-4_{D'}), 2.75 (br s, 4H, CH₂Lev), 2.26 (s, 3H, C(O)CH₃_{Ac}), 2.00 (s, 3H, C(O)CH₃_{Lev}), 1.63 (d, *J*_{5,6} = 5.6 Hz, 3H, H-6_C), 1.53 (d, *J*_{5,6} = 6.2 Hz, 3H, H-6_B), 1.49 (d, *J*_{5,6} = 6.1 Hz, 3H, H-6_A), 1.48 (s, 3H, CH₃Iso), 1.34 (s, 3H, CH₃Iso), 1.04 (s, 9H, C(CH₃)₃), 0.25 (s, 3H, Si(CH₃)₂), 0.23 (s, 3H, Si(CH₃)₂). ¹³C NMR (Py-d₅, 100 MHz) δ: 205.9 (C(O)CH₂Lev), 172.9 (C=O), 170.7 (C(O)CH₃_{Ac}), 160.3 (C_{Ph}), 144.3 (C=NPh), 140.2-138.4 (C_{Ph}), 131.6-126.2 (CH_{Ph}), 102.9 (2C, C-1_D, C-1_B), 102.7 (C-1_A), 99.8 (C(CH₃)₂), 99.3 (C-1_E), 82.6 (C-3_E), 82.4 (C-2_E), 81.7 (C-4_B), 81.2 (C-4_A), 80.3 (C-3_C), 80.2 (C-3_A), 79.6 (C-3_B), 79.0 (C-4_C), 78.5 (C-4_E), 76.7 (C-2_A), 76.2, 76.1 (2C, CH₂Ph), 75.9 (C-4_{D'}), 75.9 (CH₂Ph), 75.8 (C-2_B), 75.5, 74.5, 74.1 (3C, CH₂Ph), 72.8 (3C, CH₂Ph, C-3_{D'}, C-5_E), 71.8 (C-2_C), 71.4 (C-5_C), 71.3 (CH₂Ph), 70.1 (C-5_B), 69.8 (C-5_A), 69.5 (C-6_E), 67.6 (C-5_{D'}), 63.0 (C-6_{D'}), 60.4 (C-2_{D'}), 55.6 (OCH₃), 38.4 (CH₂Lev), 29.8 (C(O)CH₃_{Lev}), 29.7 (CH₃Iso), 28.8 (CH₂Lev), 26.5 (C(CH₃)₃), 24.5 (C(O)CH₃_{Ac}), 19.6 (C-6_C), 19.5 (CH₃Iso), 19.0 (C-6_B), 18.9 (C(CH₃)₃), 18.7 (C-6_A), -3.4 (Si(CH₃)₂), -4.2 (Si(CH₃)₂).

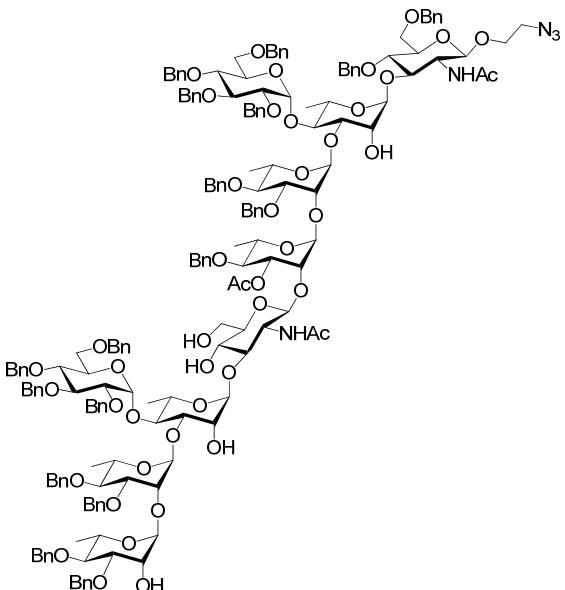
2-Azidoethyl (3,4-di-O-benzyl-2-O-levulinoyl- α -L-rhamnopyranosyl)-(1→2)-(3,4-di-O-benzyl- α -L-rhamnopyranosyl)-(1→3)-[2,3,4,6-tetra-O-benzyl- α -D-glucopyranosyl-(1→4)]-(2-O-levulinoyl- α -L-rhamnopyranosyl)-(1→3)-(2-acetamido-2-deoxy-4,6-O-isopropylidene- β -D-glucopyranosyl)-(1→2)-(4-O-benzyl- α -L-rhamnopyranosyl)-(1→2)-(3,4-di-O-benzyl- α -L-rhamnopyranosyl)-(1→3)-[2,3,4,6-tetra-O-benzyl- α -D-glucopyranosyl-(1→4)]-(2-O-levulinoyl- α -L-rhamnopyranosyl)-(1→3)-2-acetamido-4,6-di-O-benzyl-2-deoxy- β -D-glucopyranoside (36).



H_2O (425 μL) and DDQ (39 mg, 170 μmol , 2.0 equiv.) were added to decasaccharide **5** (313 mg, 85.0 μmol) in anhydr. CH_2Cl_2 (4.3 mL). The mixture was stirred for 1.5 h at rt under Ar. Satd aq $NaHCO_3$ (25 mL) was added and the aq phase was extracted with CH_2Cl_2 (3×25 mL) and dried over anhydr. Na_2SO_4 . Solvents were evaporated and the residue was purified by FC ($CH_2Cl_2/MeOH$ 99:1 to 98:2 + 0.5% Et_3N) to give alcohol **36** (211 mg, 70%) as a white foam along with unreacted **5** (40 mg, 13%). R_f 0.61 ($CH_2Cl_2/MeOH$ 95:5); $[\alpha]^{25}_D -22^\circ$ (*c* 0.32, $CHCl_3$); 1H NMR (Py-d₅, 400 MHz) δ : 9.44 (d, *J* = 7.6 Hz, 1H, NH), 9.35 (br s, 1H, NH), 7.70-7.25 (m, 85H, CH_{Ph}), 6.63 (br s, 1H, OH), 6.09 (dd, *J*_{2,3} = 2.7 Hz, *J*_{1,2} = 1.8 Hz, 1H, H-2_{A'}), 5.78-5.40 (11H, H-1_A, H-1_{A'}, H-1_B, H-1_{B'}, H-1_C, H-1_{C'}, H-1_D', H-1_E, H-1_{E'}, H-2_C, H-2_{C'}), 5.36-3.63 (m, 79H, H-2_A, H-3_A, H-3_{A'}, H-4_A, H-4_{A'}, H-5_A, H-5_{A'}, H-2_B, H-2_{B'}, H-3_B, H-3_{B'}, H-4_B, H-4_{B'}, H-5_B, H-5_{B'}, H-3_C, H-3_{C'}, H-4_C, H-4_{C'}, H-5_C, H-5_{C'}, H-1_D, H-2_D, H-2_{D'}, H-3_D, H-3_{D'}, H-4_D, H-5_D, H-6a_D, H-6b_D, H-6a_{D'}, H-2_E, H-2_{E'}, H-3_E, H-3_{E'}, H-4_E, H-4_{E'}, H-5_E, H-5_{E'}, H-6a_E, H-6b_E, H-6a_{E'}, H-6b_{E'}, OCH_2a , OCH_2b , CH_2Ph), 3.54-3.32 (m, 5H, H-4_D', H-5_D', H-6b_{D'}, CH_2N_3a , CH_2N_3b), 2.87-2.62 (m, 12H, 6× CH_2Lev), 2.50 (s, 3H, $C(O)CH_3Ac$), 2.36 (s, 3H, $C(O)CH_3Ac$), 2.02 (s, 3H, $C(O)CH_3Lev$), 2.02 (s, 3H, $C(O)CH_3Lev$), 2.00 (s, 3H, $C(O)CH_3Lev$), 1.68 (d, *J*_{5,6} = 6.1 Hz, 3H, CH_3Rha), 1.53 (d, *J*_{5,6} = 6.2 Hz, 3H, CH_3Rha), 1.51-1.43 (m, 12H, 3× CH_3Rha , CH_3Iso), 1.41 (d, *J*_{5,6} = 6.1 Hz, 3H, CH_3Rha), 1.26 (s, 3H, CH_3Iso). ^{13}C NMR (Py-d₅, 100 MHz) δ (partial): 206.3, 206.1, 206.0 (3× $C(O)CH_2Lev$), 173.1, 173.0, 172.7 (3C, $C=O_{Lev}$), 171.9 (2C, $C(O)CH_3Ac$), 140.2-139.3 (C_{Ph}), 129.3-128.0 (CH_{Ph}), 103.4, 102.5 (2C), 102.4, 101.0, 100.3, 99.2, 98.4, 98.1, 97.8 (C-1_A, C-1_{A'}, C-1_B, C-1_{B'}, C-1_C, C-1_{C'}, C-1_D, C-1_{D'}, C-1_E, C-1_{E'}), 83.2, 82.7, 82.6, 82.5, 82.2, 81.4, 81.0, 80.7, 80.4, 80.2, 79.9, 79.6, 78.7, 78.6, 78.5, 78.4, 77.0, 76.1, 75.9, 75.8, 75.7, 75.5, 75.4, 75.0, 74.2, 74.0, 73.9, 73.7, 73.6, 72.8, 72.4, 72.2, 71.9, 71.6, 70.7, 70.0, 69.7, 69.4, 69.2, 68.6, 68.5, 68.1, 67.8 (C-2_A, C-2_{A'}, C-3_A, C-3_{A'}, C-4_A, C-4_{A'}, C-5_A, C-5_{A'}, C-2_B, C-2_{B'}, C-3_B, C-3_{B'}, C-4_B, C-4_{B'}, C-5_B, C-5_{B'}, C-2_C, C-2_{C'}, C-3_C, C-3_{C'}, C-4_C, C-4_{C'}, C-5_C, C-5_{C'}, C-3_D, C-3_{D'}, C-4_D, C-4_{D'}, C-

δ 5_D, C-5_{D'}, C-6_D, C-2_E, C-2_{E'}, C-3_E, C-3_{E'}, C-4_E, C-4_{E'}, C-5_E, C-5_{E'}, C-6_E, C-6_{E'}, 17 \times CHPh₂), 62.9 (C-6_{D'}), 58.9 (C-2_{D'}), 58.2 (C-2_D), 51.5 (CH₂N₃), 38.4, 38.4, 38.3 (3C, CH₂Lev), 29.9, 29.8, 29.8 (3C, C(O)CH₃Lev), 29.0, 28.9, 28.9 (3C, CH₂Lev), 24.4, 24.2 (2C, C(O)CH₃Ac), 19.8, 19.6, 19.5, 19.0, 18.9 (3C), 18.8 (8C, 6 CH₃Rha, 2 CH₃Iso). HR-ESI-TOF-MS m/z 1780.38 [M + 2H]²⁺ (calcd for C₂₀₃H₂₃₇N₅O₅₁, 1780.31).

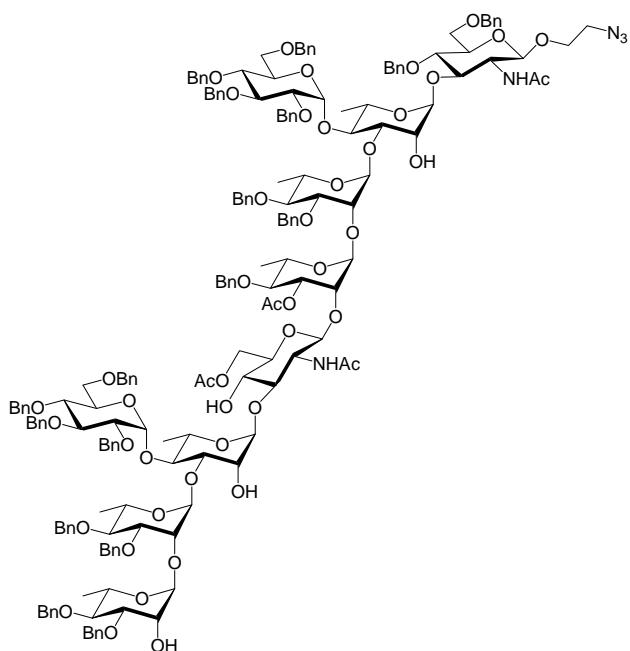
2-Azidoethyl (3,4-di-O-benzyl- α -L-rhamnopyranosyl)-(1 \rightarrow 2)-(3,4-di-O-benzyl- α -L-rhamnopyranosyl)-(1 \rightarrow 3)-[2,3,4,6-tetra-O-benzyl- α -D-glucopyranosyl-(1 \rightarrow 4)]-(α -L-rhamnopyranosyl)-(1 \rightarrow 3)-(2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 2)-(3-O-acetyl-4-O-benzyl- α -L-rhamnopyranosyl)-(1 \rightarrow 2)-(3,4-di-O-benzyl- α -L-rhamnopyranosyl)-(1 \rightarrow 3)-[2,3,4,6-tetra-O-benzyl- α -D-glucopyranosyl-(1 \rightarrow 4)]-(α -L-rhamnopyranosyl)-(1 \rightarrow 3)-2-acetamido-4,6-di-O-benzyl-2-deoxy- β -D-glucopyranoside (38).



Ac₂O (531 μ L, 5.62 mmol, 100 equiv.) and DMAP (7.0 mg, 56 μ mol, 1.0 equiv) were added to alcohol **36** (200 mg, 56.2 μ mol) in anhydrous Py (5.6 mL). The mixture was stirred for 1 h at rt under Ar. Solvents were evaporated and co-evaporated with toluene (3 \times). Aq TFA (50% v/v, 1.8 mL) was added dropwise to the residue (202 mg, 56.2 μ mol) in CH₂Cl₂ (8.4 mL) at 0 °C. The mixture was stirred for 2 h from 0 °C to rt. The solution was diluted with CH₂Cl₂ (10 mL) and excess TFA was neutralized with satd aq NaHCO₃ (25 mL). The aq phase was extracted with CH₂Cl₂ (3 \times 25 mL). The pooled organic layers were washed with brine (25 mL), dried over anhydrous Na₂SO₄, filtered, and solvents were evaporated. Hydrazine hydrate (35 μ L, 1.12 mmol, 20 equiv.) was added to the residue (200 mg, 56.2 μ mol) in anhydrous Py/AcOH (8.5 mL, 3:2 v/v). The mixture was stirred for 2 h at rt under Ar. The solution was diluted with CH₂Cl₂ (25 mL). The organic layer was washed with H₂O (25 mL) and the aq phase was extracted with CH₂Cl₂

(3×25 mL). Solvents were evaporated and co-evaporated with toluene (3×). The residue was purified by FC (CH₂Cl₂/MeOH 99:1 to 98:2) to give acetate **38** (145 mg, 79% over 3 steps) as a white powder. *R*_f 0.54 (CH₂Cl₂/MeOH 95:5); [α]²⁵_D -55° (*c* 0.20, CHCl₃); ¹H NMR (Py-d₅, 400 MHz) δ: 9.53 (d, *J*_{NH,2} = 7.2 Hz, 1H, NH), 9.40 (d, *J*_{NH,2} = 8.5 Hz, 1H, NH), 7.70-7.14 (m, 85H, CH_{Ph}), 6.01 (dd, *J*_{3,4} = 9.8 Hz, *J*_{2,3} = 2.4 Hz, 1H, H-3_A), 5.95-5.40 (9H, H-1_A, H-1_{A'}, H-1_B, H-1_{B'}, H-1_C, H-1_{C'}, H-1_{D'}, H-1_E, H-1_{E'}), 5.34-3.62 (m, 84H, H-2_A, H-2_{A'}, H-3_{A'}, H-4_A, H-4_{A'}, H-5_A, H-5_{A'}, H-2_B, H-2_{B'}, H-3_B, H-3_{B'}, H-4_B, H-4_{B'}, H-5_B, H-5_{B'}, H-2_C, H-2_{C'}, H-3_C, H-3_{C'}, H-4_C, H-4_{C'}, H-5_C, H-5_{C'}, H-1_D, H-2_D, H-2_{D'}, H-3_D, H-3_{D'}, H-4_D, H-4_{D'}, H-5_D, H-5_{D'}, H-6a_D, H-6a_{D'}, H-6b_D, H-6b_{D'}, H-2_E, H-2_{E'}, H-3_E, H-3_{E'}, H-4_E, H-4_{E'}, H-5_E, H-5_{E'}, H-6a_E, H-6a_{E'}, H-6b_E, H-6b_{E'}, OCH₂a, OCH₂b, CH₂Ph), 3.49-3.41 (m, 1H, CH₂N₃a), 3.34-3.27 (m, 1H, CH₂N₃b), 2.21 (s, 3H, C(O)CH₃Ac), 2.15, 2.09 (2s, 6H, C(O)CH₃NHAc), 1.68 (d, *J*_{5,6} = 6.1 Hz, 3H, CH₃Rha), 1.54-1.42 (m, 15H, CH₃Rha). ¹³C NMR (Py-d₅, 100 MHz) δ (partial): 171.1, 171.0, 170.9 (3×C(O)CH₃), 140.3-139.2 (C_{Ph}), 129.4-127.9 (CH_{Ph}), 103.9, 103.1, 102.8, 102.5, 102.3 (2C), 102.2, 101.5, 98.9, 98.4 (C-1_A, C-1_{A'}, C-1_B, C-1_{B'}, C-1_C, C-1_{C'}, C-1_D, C-1_{D'}, C-1_E, C-1_{E'}), 83.6, 83.2, 82.6, 82.5, 82.3, 82.2, 81.5, 81.4, 81.3, 81.1, 80.5, 80.3, 79.8, 78.9, 78.7, 78.5, 78.0, 77.8, 76.9, 76.0, 75.8, 75.7, 75.5, 75.4, 75.3, 73.9, 73.8, 73.8, 73.7, 73.5, 72.7, 72.3, 72.1, 71.8, 71.7, 71.5, 71.0, 70.9, 69.9, 69.6, 69.5, 69.4, 69.2, 69.1, 68.9, 68.2 (C-2_A, C-3_A, C-4_A, C-5_A, C-2_B, C-3_B, C-4_B, C-5_B, C-2_C, C-3_C, C-4_C, C-5_C, C-3_D, C-4_D, C-5_D, C-6_D, C-2_E, C-3_E, C-4_E, C-5_E, C-6_E, C-2_{A'}, C-3_{A'}, C-4_{A'}, C-5_{A'}, C-2_{B'}, C-3_{B'}, C-4_{B'}, C-5_{B'}, C-2_{C'}, C-3_{C'}, C-4_{C'}, C-5_{C'}, C-3_{D'}, C-4_{D'}, C-5_{D'}, C-2_{E'}, C-3_{E'}, C-4_{E'}, C-5_{E'}, C-6_{E'}, OCH₂, 17×CH₂Ph), 63.1 (C-6_{D'}), 59.2 (C-2_{D'}), 57.5 (C-2_D), 51.4 (CH₂N₃), 24.0 (C(O)CH₃NHAc), 23.8 (C(O)CH₃NHAc), 21.4 (C(O)CH₃Ac), 19.7, 19.6, 19.1, 18.9, 18.8, 18.7 (6×CH₃Rha). HR-ESI-TOF-MS *m/z* 1634.33 [M + 2H]²⁺ (calcd for C₁₈₇H₂₁₇N₅O₄₆, 1634.24).

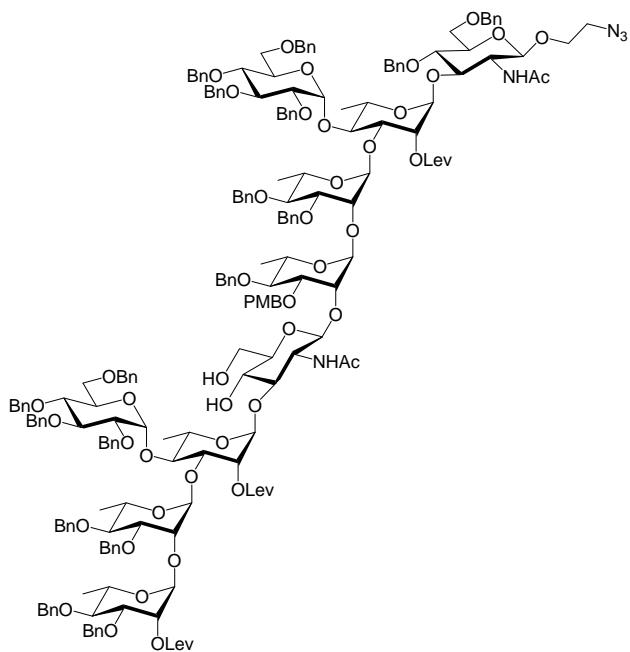
2-Azidoethyl (3,4-di-O-benzyl- α -L-rhamnopyranosyl)-(1→2)-(3,4-di-O-benzyl- α -L-rhamnopyranosyl)-(1→3)-[2,3,4,6-tetra-O-benzyl- α -D-glucopyranosyl-(1→4)]-(α -L-rhamnopyranosyl)-(1→3)-(2-acetamido-6-O-acetyl-2-deoxy- β -D-glucopyranosyl)-(1→2)-(3-O-acetyl-4-O-benzyl- α -L-rhamnopyranosyl)-(1→2)-(3,4-di-O-benzyl- α -L-rhamnopyranosyl)-(1→3)-[2,3,4,6-tetra-O-benzyl- α -D-glucopyranosyl-(1→4)]-(α -L-rhamnopyranosyl)-(1→3)-2-acetamido-4,6-di-O-benzyl-2-deoxy- β -D-glucopyranoside (39).



Ac_2O (536 μL , 5.67 mmol, 100 equiv.) and DMAP (7.0 mg, 56 μmol , 1.0 equiv) were added to alcohol **36** (202 mg, 56.7 μmol) in anhydrous Py (2.8 mL). The mixture was stirred for 1 h at rt under Ar, and solvents were evaporated and co-evaporated with toluene (3 \times). Aq TFA (50% v/v, 1.8 mL) was added dropwise to the residue (204 mg, 56.7 μmol) in CH_2Cl_2 (8.5 mL) at 0 °C. The mixture was stirred for 2 h from 0 °C to rt. The solution was diluted with CH_2Cl_2 (10 mL) and excess TFA was neutralized with satd aq NaHCO_3 (25 mL). The aq phase was extracted with CH_2Cl_2 (3 \times 25 mL). The pooled organic layers were washed with brine (25 mL), dried over anhydrous Na_2SO_4 , filtered, and solvents were evaporated. Acetyl chloride (113 μmol , 100 μL , 2.0 equiv.) was added from a fresh stock solution (80 μL of acetyl chloride in 1.0 mL anhydrous CH_2Cl_2) to the residue (202 mg, 56.7 μmol) in *sym*-collidine (2.2 mL) at -45 °C ($\text{CH}_3\text{CN}/\text{ice CO}_2$ bath) under Ar. Additional acetyl chloride (57 μmol , 50 μL of stock solution, 1.0 equiv.) was added after 60, 120, 180 and 240 min. During this time, the temperature was gradually raised from -45 °C to rt. After reaction completion (6 h), excess acetyl chloride was quenched with MeOH, CH_2Cl_2 was added and the organic phase was washed with 5% aq HCl, satd NaHCO_3 and brine, and dried over anhydrous Na_2SO_4 . Solvents were evaporated and co-evaporated with toluene (3 \times). Hydrazine hydrate (35 μL , 1.12 mmol, 20 equiv.) was added to the residue (204 mg, 56.7 μmol) in anhydrous Py/AcOH (8.5 mL, 3:2 v/v). The mixture was stirred for 2 h at rt under Ar. The solution was diluted with CH_2Cl_2 (25 mL). The organic layer was washed with H_2O (25 mL) and the aq phase was extracted with CH_2Cl_2 (3 \times 25 mL). Solvents were evaporated and co-evaporated with toluene (3 \times). The residue was purified by FC ($\text{CH}_2\text{Cl}_2/\text{MeOH}$ 99:1 to 98:2) to give diacetate **39** (138 mg, 74%, 4 steps) as a white powder. R_f 0.63 ($\text{CH}_2\text{Cl}_2/\text{MeOH}$ 95:5); $[\alpha]^{25}_D +15^\circ$ (c 0.40, CHCl_3); ^1H NMR (Py-d₅, 400 MHz) δ : 9.59 (d, $J_{\text{NH},2} = 7.3$ Hz, 1H, NH), 9.25 (d, $J_{\text{NH},2} = 8.5$ Hz, 1H, NH), 7.69-7.17 (m, 85H, CH_{Ph}), 5.98 (dd, $J_{3,4} = 9.9$ Hz, $J_{2,3} = 2.7$ Hz, 1H, H-3_A), 5.86-5.47 (m, 9H, H-1_A, H-1_{A'}, H-1_B, H-1_{B'}, H-1_C, H-1_{C'}, H-1_D', H-1_E, H-1_{E'}), 5.30-3.56 (m, 83H, H-2_A, H-

$2_{\text{A}'}, \text{H}-3_{\text{A}'}, \text{H}-4_{\text{A}'}, \text{H}-5_{\text{A}'}, \text{H}-2_{\text{B}'}, \text{H}-3_{\text{B}'}, \text{H}-4_{\text{B}'}, \text{H}-5_{\text{B}'}, \text{H}-5_{\text{B}'}, \text{H}-2_{\text{C}'}, \text{H}-2_{\text{C}'}, \text{H}-3_{\text{C}'}, \text{H}-4_{\text{C}'}, \text{H}-4_{\text{C}'}, \text{H}-5_{\text{C}'}, \text{H}-5_{\text{C}'}, \text{H}-1_{\text{D}'}, \text{H}-2_{\text{D}'}, \text{H}-3_{\text{D}'}, \text{H}-3_{\text{D}'}, \text{H}-4_{\text{D}'}, \text{H}-4_{\text{D}'}, \text{H}-5_{\text{D}'}, \text{H}-5_{\text{D}'}, \text{H}-6_{\text{aD}'}, \text{H}-6_{\text{aD}'}, \text{H}-6_{\text{bD}'}, \text{H}-6_{\text{bD}'}, \text{H}-2_{\text{E}'}, \text{H}-2_{\text{E}'}, \text{H}-3_{\text{E}'}, \text{H}-3_{\text{E}'}, \text{H}-4_{\text{E}'}, \text{H}-4_{\text{E}'}, \text{H}-5_{\text{E}'}, \text{H}-5_{\text{E}'}, \text{H}-6_{\text{aE}'}, \text{H}-6_{\text{aE}'}, \text{H}-6_{\text{bE}'}, \text{H}-6_{\text{bE}'}, \text{OCH}_2\text{a}, \text{OCH}_2\text{b}, \text{CH}_2\text{Ph}), 3.60 (\text{m}, J_{1,2} = 8.3 \text{ Hz}, 1\text{H}, \text{H}-2_{\text{D}'}), 3.48-3.41 (\text{m}, 1\text{H}, \text{CH}_2\text{N}_3\text{a}), 3.32 (\text{ddd}, J = 13.2, 5.3, 3.5 \text{ Hz}, 1\text{H}, \text{CH}_2\text{N}_3\text{b}), 2.26 (\text{s}, 3\text{H}, \text{C(O)CH}_3\text{Ac}), 2.18, 2.09 (2\text{s}, 6\text{H}, \text{C(O)CH}_3\text{NHAc}), 1.73 (\text{s}, 3\text{H}, \text{C(O)CH}_3\text{Ac}), 1.65 (\text{d}, J_{5,6} = 6.1 \text{ Hz}, 3\text{H}, \text{CH}_3\text{Rha}), 1.54-1.40 (\text{m}, 15\text{H}, \text{CH}_3\text{Rha})$. ^{13}C NMR (Py-d₅, 100 MHz) δ (partial): 171.2 ($\text{C(O)CH}_3\text{Ac}$), 171.0 (3× $\text{C(O)CH}_3\text{Ac}$), 140.4-139.3 (C_{Ph}), 129.4-127.9 (CH_{Ph}), 104.0, 103.2, 102.9, 102.6, 102.3, 102.2, 101.6 (2C), 99.0, 98.6 ($\text{C}-1_{\text{A}'}, \text{C}-1_{\text{A}'}, \text{C}-1_{\text{B}'}, \text{C}-1_{\text{B}'}, \text{C}-1_{\text{C}'}, \text{C}-1_{\text{C}'}, \text{C}-1_{\text{D}'}, \text{C}-1_{\text{D}'}, \text{C}-1_{\text{E}'}, \text{C}-1_{\text{E}'}$), 83.4, 82.6, 82.6, 82.3, 81.9, 81.6, 81.6, 81.2, 80.7, 80.5, 80.3, 79.6, 78.9, 78.8, 78.6, 77.2, 77.0, 76.8, 76.1, 76.1, 76.1, 76.0, 75.8, 75.8, 75.7, 75.6, 75.5, 75.1, 74.9, 74.5, 74.4, 74.0, 73.9, 73.9, 73.8, 73.7, 72.9, 72.4, 72.2, 71.9, 71.8, 71.6, 71.0, 70.4, 70.0, 69.6, 69.5, 69.4, 69.3, 69.2, 69.1, 68.3 ($\text{C}-2_{\text{A}'}, \text{C}-2_{\text{A}'}, \text{C}-3_{\text{A}'}, \text{C}-3_{\text{A}'}, \text{C}-4_{\text{A}'}, \text{C}-4_{\text{A}'}, \text{C}-5_{\text{A}'}, \text{C}-5_{\text{A}'}, \text{C}-2_{\text{B}'}, \text{C}-2_{\text{B}'}, \text{C}-3_{\text{B}'}, \text{C}-3_{\text{B}'}, \text{C}-4_{\text{B}'}, \text{C}-4_{\text{B}'}, \text{C}-5_{\text{B}'}, \text{C}-5_{\text{B}'}, \text{C}-2_{\text{C}'}, \text{C}-2_{\text{C}'}, \text{C}-3_{\text{C}'}, \text{C}-3_{\text{C}'}, \text{C}-4_{\text{C}'}, \text{C}-4_{\text{C}'}, \text{C}-5_{\text{C}'}, \text{C}-5_{\text{C}'}, \text{C}-3_{\text{D}'}, \text{C}-3_{\text{D}'}, \text{C}-4_{\text{D}'}, \text{C}-4_{\text{D}'}, \text{C}-5_{\text{D}'}, \text{C}-5_{\text{D}'}, \text{C}-6_{\text{D}'}, \text{C}-2_{\text{E}'}, \text{C}-2_{\text{E}'}, \text{C}-3_{\text{E}'}, \text{C}-3_{\text{E}'}, \text{C}-4_{\text{E}'}, \text{C}-4_{\text{E}'}, \text{C}-5_{\text{E}'}, \text{C}-5_{\text{E}'}, \text{C}-6_{\text{E}'}, \text{C}-6_{\text{E}'}, \text{OCH}_2$), 59.7 ($\text{C}-2_{\text{D}'}$), 57.5 ($\text{C}-2_{\text{D}}$), 51.5 (CH_2N_3), 24.1, 23.9 (2C, $\text{C(O)CH}_3\text{NHAc}$), 21.5, 21.0 (2C, $\text{C(O)CH}_3\text{Ac}$), 19.7, 19.7, 19.2, 18.9 (2C), 18.8 (6C, CH_3Rha). HR-MALDI-TOF-MS m/z 3331.47489 [$\text{M} + \text{Na}$]⁺ (calcd for $\text{C}_{187}\text{H}_{217}\text{N}_5\text{O}_{47}\text{Na}$, 3331.46361).

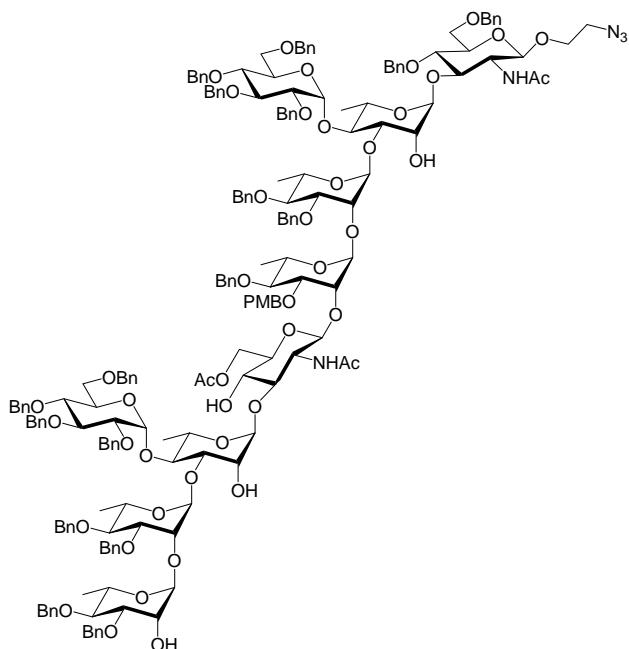
2-Azidoethyl (3,4-di-O-benzyl-2-O-levulinoyl- α -L-rhamnopyranosyl)-(1→2)-(3,4-di-O-benzyl- α -L-rhamnopyranosyl)-(1→3)-[2,3,4,6-tetra-O-benzyl- α -D-glucopyranosyl-(1→4)]-(2-O-levulinoyl- α -L-rhamnopyranosyl)-(1→3)-(2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1→2)-(4-O-benzyl-3-O-para-methoxybenzyl- α -L-rhamnopyranosyl)-(1→2)-(3,4-di-O-benzyl- α -L-rhamnopyranosyl)-(1→3)-[2,3,4,6-tetra-O-benzyl- α -D-glucopyranosyl-(1→4)]-(2-O-levulinoyl- α -L-rhamnopyranosyl)-(1→3)-2-acetamido-4,6-di-O-benzyl-2-deoxy- β -D-glucopyranoside (40).



Aq TFA (50% v/v, 1.8 mL) was added to decasaccharide **5** (210 mg, 57.0 μ mol) in CH_2Cl_2 (8.6 mL) at 0 °C. The mixture was stirred for 2 h from 0 °C to rt, then diluted with CH_2Cl_2 (10 mL) and washed with satd NaHCO_3 (25 mL). The aq phase was extracted with CH_2Cl_2 (3×25 mL), the pooled organic layers were washed with brine (25 mL), dried over anhydr. Na_2SO_4 , filtered and solvents were evaporated. The residue was purified by FC ($\text{CH}_2\text{Cl}_2/\text{MeOH}$ 99:1 to 98:2) to give diol **40** (187 mg, 90%) as a white powder. $[\alpha]^{25}_{\text{D}} -36^\circ$ (*c* 0.22, CHCl_3); ^1H NMR (Py-d₅, 400 MHz) δ : 9.54 (br s, 1H, NH), 7.77-7.09 (m, 87H, CH_{Ph}), 6.98-6.94 (m, 2H, CH_{Ph}), 6.12 (br s, 1H, H-2_A), 5.86-5.25 (m, 12H, H-2_C, H-2_{C'}, H-1_A, H-1_{A'}, H-1_B, H-1_{B'}, H-1_C, H-1_{C'}, H-1_D, H-1_{D'}, H-1_E, H-1_{E'}), 5.24-3.53 (m, 86H, H-2_A, H-3_A, H-3_{A'}, H-4_A, H-4_{A'}, H-5_A, H-5_{A'}, H-2_B, H-2_{B'}, H-3_B, H-3_{B'}, H-4_B, H-4_{B'}, H-5_B, H-5_{B'}, H-3_C, H-3_{C'}, H-4_C, H-4_{C'}, H-5_C, H-5_{C'}, H-2_D, H-2_{D'}, H-3_D, H-3_{D'}, H-4_D, H-4_{D'}, H-5_D, H-5_{D'}, H-6a_D, H-6a_{D'}, H-6b_D, H-6b_{D'}, H-2_E, H-2_{E'}, H-3_E, H-3_{E'}, H-4_E, H-4_{E'}, H-5_E, H-5_{E'}, H-6a_E, H-6a_{E'}, H-6b_E, H-6b_{E'}, OCH_3 , OCH_2a , OCH_2b , CH_2Ph), 3.53-3.44 (m, 1H, $\text{CH}_2\text{N}_3\text{a}$), 3.41-3.33 (m, 1H, $\text{CH}_2\text{N}_3\text{b}$), 2.87-2.58 (m, 12H, CH_2Lev), 2.46, 2.40 (2s, 6H, C(O) CH_3NHAc), 2.04, 2.01, 1.99 (3s, 9H, C(O) CH_3Lev), 1.71 (d, $J_{5,6} = 5.9$ Hz, 3H, CH_3Rha), 1.57 (d, $J_{5,6} = 6.3$ Hz, 3H, CH_3Rha), 1.51 (d, $J_{5,6} = 6.0$ Hz, 3H, CH_3Rha), 1.43-1.36 (m, 6H, CH_3Rha), 1.30 (d, $J_{5,6} = 5.8$ Hz, 3H, CH_3Rha). ^{13}C NMR (Py-d₅, 100 MHz) δ (partial): 206.2 (C(O) CH_2Lev), 206.0 (2C, C(O) CH_2Lev), 173.0 (2C, C(O) CH_3Lev), 172.7 (C(O) CH_3Lev), 172.1, 172.0 (2C, C(O) CH_3NHAc), 160.2 (C_{Ph}), 140.3-139.2 (C_{Ph}), 130.9 (CH_{Ph}), 129.4-127.8 (CH_{Ph}), 114.6 (CH_{Ph}), 102.2 (5C), 101.1, 100.6, 99.2, 98.8, 98.5 (C-1_A, C-1_{A'}, C-1_B, C-1_{B'}, C-1_C, C-1_{C'}, C-1_D, C-1_{D'}, C-1_E, C-1_{E'}), 82.8, 82.6, 82.5, 81.6, 81.1, 80.9, 80.0, 79.8, 79.6, 79.4, 79.3, 78.7, 78.6, 78.5, 78.1, 77.1, 76.4, 76.1, 76.0, 75.9, 75.8, 75.6, 75.0, 74.4, 74.2, 74.0, 73.8, 72.8, 72.5, 72.3, 72.0, 71.5, 71.3, 70.2, 70.0, 69.7, 69.5, 69.5, 69.2, 68.5, 68.4 (C-2_A, C-2_{A'}, C-3_A, C-3_{A'}, C-4_A, C-4_{A'}, C-5_A, C-5_{A'}, C-2_B, C-2_{B'}, C-3_B, C-3_{B'}, C-4_B, C-4_{B'}, C-5_B, C-5_{B'}, C-2_C, C-2_{C'}, C-3_C, C-3_{C'}, C-4_C, C-4_{C'}, C-5_C, C-5_{C'}, C-3_D, C-3_{D'}, C-4_D, C-4_{D'}, C-5_D, C-5_{D'}, C-6_D, C-2_E, C-2_{E'}, C-3_E, C-3_{E'}, C-4_E, C-4_{E'}.

$4_{\text{E}'}$, $\text{C}-5_{\text{E}'}$, $\text{C}-5_{\text{E}}$, $\text{C}-6_{\text{E}'}$, OCH_2 , CH_2Ph), 63.6 ($\text{C}-6_{\text{D}'}$), 59.6 ($\text{C}-2_{\text{D}'}$), 58.2 ($\text{C}-2_{\text{D}}$), 55.5 (OCH_3), 51.6 (CH_2N_3), 38.5, 38.5, 38.4 (3C, CH_2Lev), 29.9, 29.9, 29.8 (3C, $\text{C}(\text{O})\text{CH}_3\text{Lev}$), 29.1, 29.1, 29.0 (6C, CH_2Lev), 24.5, 24.2 (2C, $\text{C}(\text{O})\text{CH}_3\text{NHAc}$), 19.5–18.8 (6C, $\text{C}-6_{\text{A}}$, $\text{C}-6_{\text{A}'}$, $\text{C}-6_{\text{B}}$, $\text{C}-6_{\text{B}'}$, $\text{C}-6_{\text{C}}$, $\text{C}-6_{\text{C}'}$). HR-MALDI-TOF-MS m/z 3661.61385 [$\text{M} + \text{Na}$]⁺ (calcd for $\text{C}_{208}\text{H}_{239}\text{N}_5\text{O}_{52}\text{Na}$, 3661.61034).

2-Azidoethyl (3,4-di-*O*-benzyl- α -L-rhamnopyranosyl)-(1 \rightarrow 2)-(3,4-di-*O*-benzyl- α -L-rhamnopyranosyl)-(1 \rightarrow 3)-[2,3,4,6-tetra-*O*-benzyl- α -D-glucopyranosyl-(1 \rightarrow 4)]-(α -L-rhamnopyranosyl)-(1 \rightarrow 3)-(2-acetamido-6-*O*-acetyl-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 2)-(4-*O*-benzyl-3-*O*-para-methoxybenzyl- α -L-rhamnopyranosyl)-(1 \rightarrow 2)-(3,4-di-*O*-benzyl- α -L-rhamnopyranosyl)-(1 \rightarrow 3)-[2,3,4,6-tetra-*O*-benzyl- α -D-glucopyranosyl-(1 \rightarrow 4)]-(α -L-rhamnopyranosyl)-(1 \rightarrow 3)-2-acetamido-4,6-di-*O*-benzyl-2-deoxy- β -D-glucopyranoside (41).



Acetyl chloride (75.8 μmol , 100 μL , 2.0 equiv.) from a fresh stock solution (108 μL of acetyl chloride in 2.0 mL anhydrous CH_2Cl_2) was added to diol **40** (138 mg, 37.9 μmol) in *sym*-collidine (1.5 mL) at -45 °C ($\text{CH}_3\text{CN}/\text{ice CO}_2$ bath) under Ar. Additional acetyl chloride (190 μmol , 250 μL of stock solution, 5.0 equiv.) was added after 60, 120, 180, 240, 300 and 360 min. During this time, the temperature was gradually raised from -45 °C to rt. After reaction completion (8 h), excess acetyl chloride was quenched with MeOH, CH_2Cl_2 was added and the organic phase was washed with 10% aq HCl. The aq layer was extracted with CH_2Cl_2 (3 \times) and the pooled organic phases were washed with satd NaHCO_3 and brine, then dried over anhydrous Na_2SO_4 . Solvents were evaporated and co-evaporated with toluene (3 \times). Hydrazine hydrate (24 μL , 0.76 mmol, 20 equiv.) was added to the residue (140 mg, 37.9 μmol) in anhydrous Py/AcOH (5.7 mL, 3:2 v/v). The mixture was stirred for 2 h at rt under Ar. The solution was diluted with CH_2Cl_2 (25 mL), the

organic layer was washed with H₂O (25 mL) and the aq phase was extracted with CH₂Cl₂ (3×25 mL). Solvents were evaporated and co-evaporated with toluene (3×). The residue was purified by FC (CH₂Cl₂/MeOH 99:1 to 98.5:1.5) to give acetate **41** (94 mg, 81%, 2 steps) as a white amorphous solid. $[\alpha]^{25}_D -4^\circ$ (*c* 0.50, CHCl₃); ¹H NMR (Py-d₅, 400 MHz) δ: 9.20 (d, *J*_{NH,2} = 8.6 Hz, 1H, NH), 9.05 (d, *J*_{NH,2} = 7.7 Hz, 1H, NH), 7.72-7.20 (m, 87H, CH_{Ph}), 7.01-6.97 (m, 2H, CH_{Ph}), 5.87-5.48 (m, 9H, H-1_A, H-1_{A'}, H-1_B, H-1_{B'}, H-1_C, H-1_{C'}, H-1_D', H-1_E, H-1_{E'}), 5.27-3.70 (m, 87 H, H-2_A, H-2_{A'}, H-3_A, H-3_{A'}, H-4_A, H-4_{A'}, H-5_A, H-5_{A'}, H-2_B, H-2_{B'}, H-3_B, H-3_{B'}, H-4_B, H-4_{B'}, H-5_B, H-5_{B'}, H-2_C, H-2_{C'}, H-3_C, H-3_{C'}, H-4_C, H-4_{C'}, H-5_C, H-5_{C'}, H-1_D, H-2_D, H-2_{D'}, H-3_D, H-3_{D'}, H-4_D, H-4_{D'}, H-5_D, H-5_{D'}, H-6a_D, H-6a_{D'}, H-6b_D, H-6b_{D'}, H-2_E, H-2_{E'}, H-3_E, H-3_{E'}, H-4_E, H-4_{E'}, H-5_E, H-5_{E'}, H-6a_E, H-6a_{E'}, H-6b_E, H-6b_{E'}, OCH₂a, OCH₂b, CH₂Ph), 3.65 (s, 3H, OCH₃), 3.49-3.41 (m, 1H, CH₂N₃a), 3.33 (ddd, *J* = 13.1, 5.1, 3.7 Hz, 1H, CH₂N₃b), 2.10, 2.09 (2s, 6H, C(O)CH₃NHAc), 1.79 (s, 3H, C(O)CH₃Ac), 1.66 (d, *J*_{5,6} = 6.1 Hz, 3H, CH₃Rha), 1.54-1.39 (m, 15H, CH₃Rha). ¹³C NMR (Py-d₅, 100 MHz) δ (partial): 171.1, 171.0, 170.9 (3C, C(O)CH₃Ac), 160.2 (C_{Ph}), 140.4-139.2 (C_{Ph}), 130.9 (CH_{Ph}), 129.5-127.8 (CH_{Ph}), 114.7 (CH_{Ph}), 104.0, 103.2, 102.8, 102.7, 102.5, 102.2, 102.0, 101.7, 99.0, 98.5 (C-1_A, C-1_{A'}, C-1_B, C-1_{B'}, C-1_C, C-1_{C'}, C-1_D, C-1_{D'}, C-1_E, C-1_{E'}), 84.0, 83.2, 82.6, 82.3, 81.9, 81.7, 81.6, 81.3, 80.6, 80.2, 79.5, 78.8, 78.7, 78.6, 77.0, 76.3, 76.1, 76.0, 75.9, 75.8, 75.6, 75.3, 73.9, 73.9, 73.8, 73.7, 72.9, 72.6, 72.3, 72.2, 71.9, 71.7, 70.9, 70.4, 70.1, 69.7, 69.5, 69.5, 69.3, 69.2, 68.3 (C-2_A, C-2_{A'}, C-3_A, C-3_{A'}, C-4_A, C-4_{A'}, C-5_A, C-5_{A'}, C-2_B, C-2_{B'}, C-3_B, C-3_{B'}, C-4_B, C-4_{B'}, C-5_B, C-5_{B'}, C-2_C, C-2_{C'}, C-3_C, C-3_{C'}, C-4_C, C-4_{C'}, C-5_C, C-5_{C'}, C-3_D, C-3_{D'}, C-4_D, C-4_{D'}, C-5_D, C-5_{D'}, C-6_D, C-2_E, C-2_{E'}, C-3_E, C-3_{E'}, C-4_E, C-4_{E'}, C-5_E, C-5_{E'}, C-6_E, C-6_{E'}, OCH₂, CH₂Ph), 64.7 (C-6_{D'}), 59.0 (C-2_{D'}), 57.4 (C-2_D), 55.6 (OCH₃), 51.5 (CH₂N₃), 24.1, 23.9 (2C, C(O)CH₃NHAc), 21.0 (C(O)CH₃Ac), 19.9, 19.7, 19.3, 19.0, 18.9 (6C, C-6_A, C-6_{A'}, C-6_B, C-6_{B'}, C-6_C, C-6_{C'}). HR-MALDI-TOF-MS *m/z* 3409.50588 [M + Na]⁺ (calcd for C₁₉₅H₂₂₃N₅O₄₇Na, 3409.51056).

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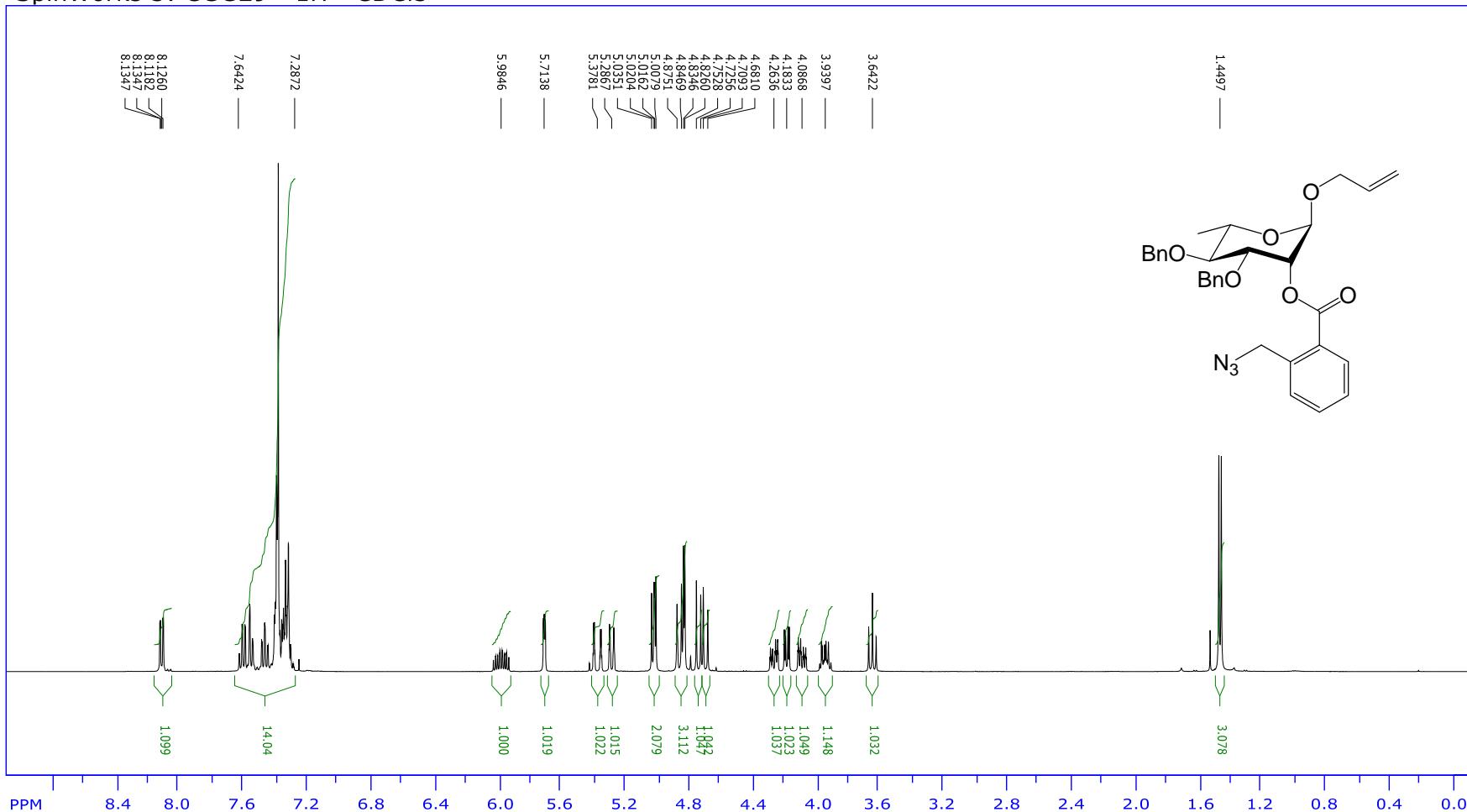
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Relevant NMR spectra (^1H , DEPT, HSQC, ^{13}C , including HSQC for $^1\text{J}_{\text{C},\text{H}}$ for compound 2 only) for new compounds listed below by order of appearance

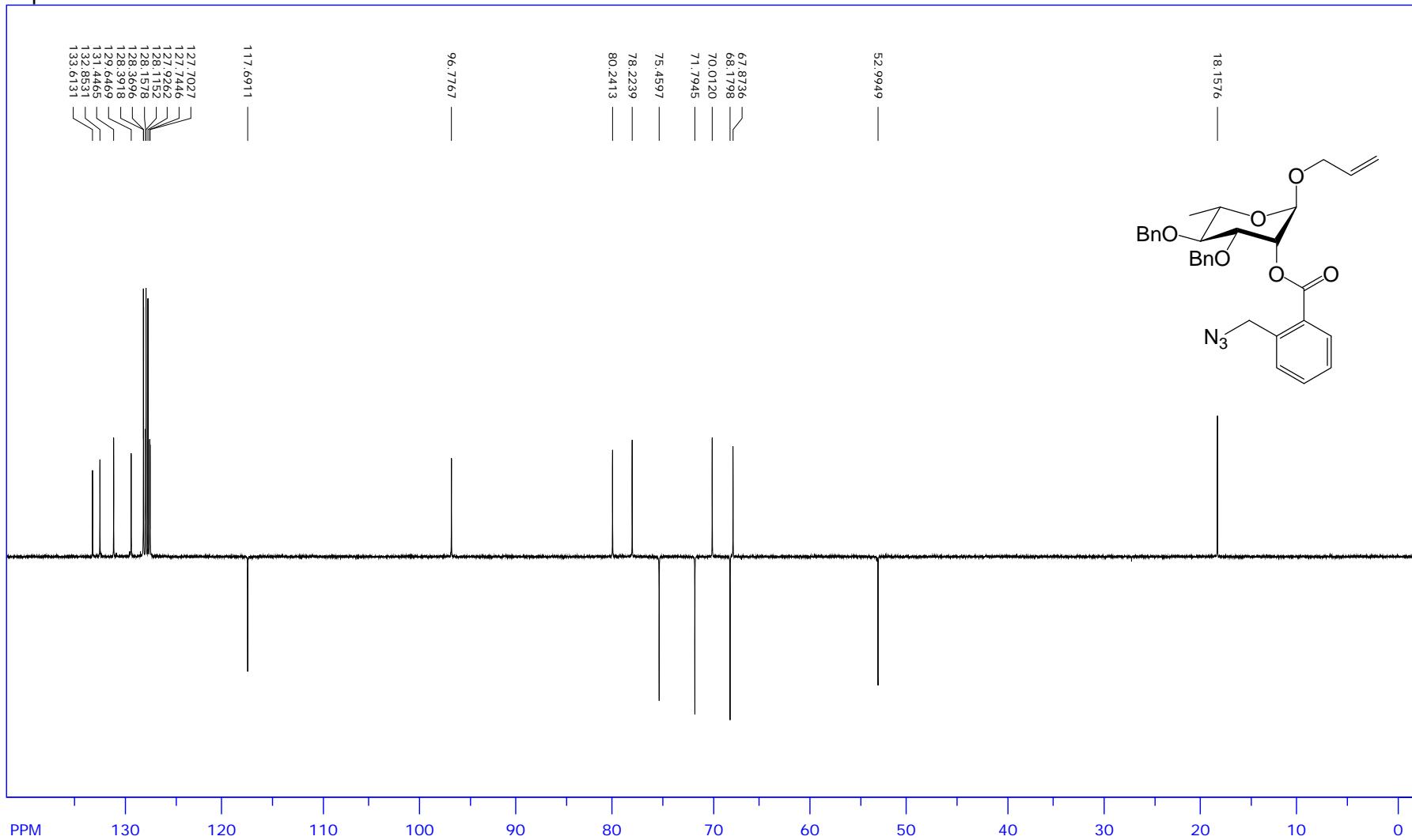
S3, S4, S6-S9, 5-8, 10-14, 16-24, 25-27, 29, 31-36, 38-41, 1-3.

Compound S3

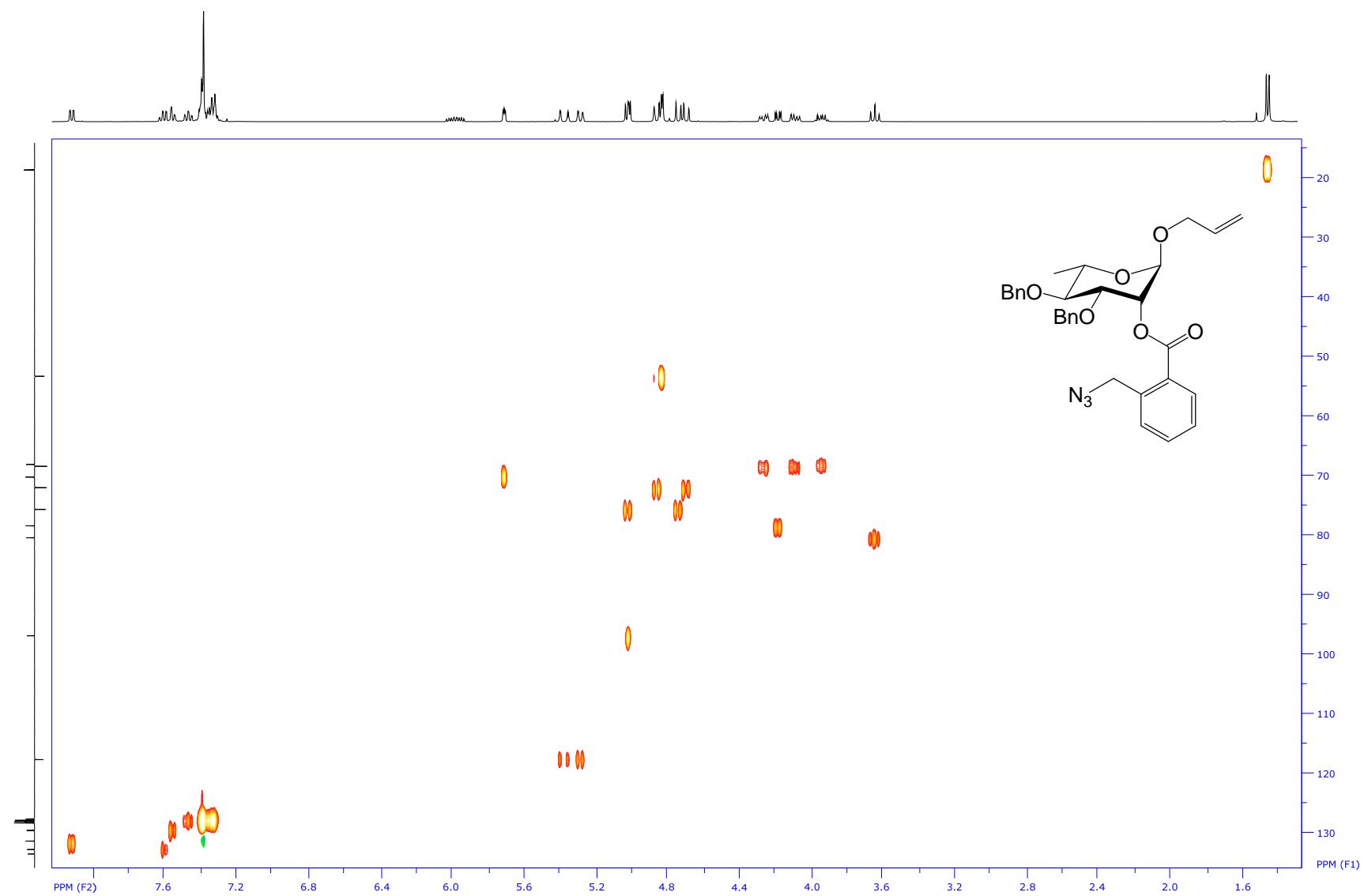
SpinWorks 3: CGG29 - 1H - CDCl₃



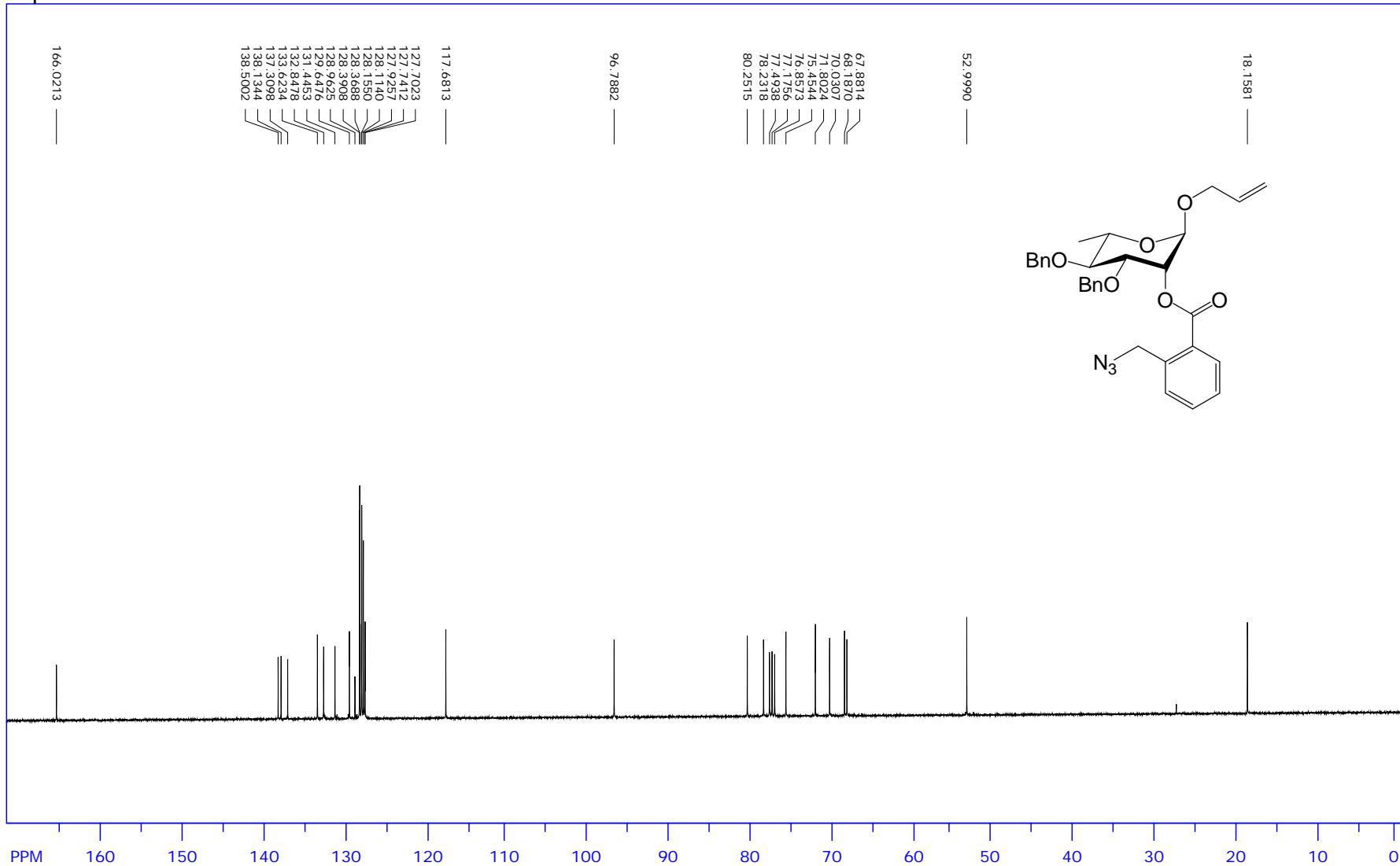
SpinWorks 3: CGG29 - DEPT135 - CDCl₃



SpinWorks 3: CGG29 - COSY - CDCl₃

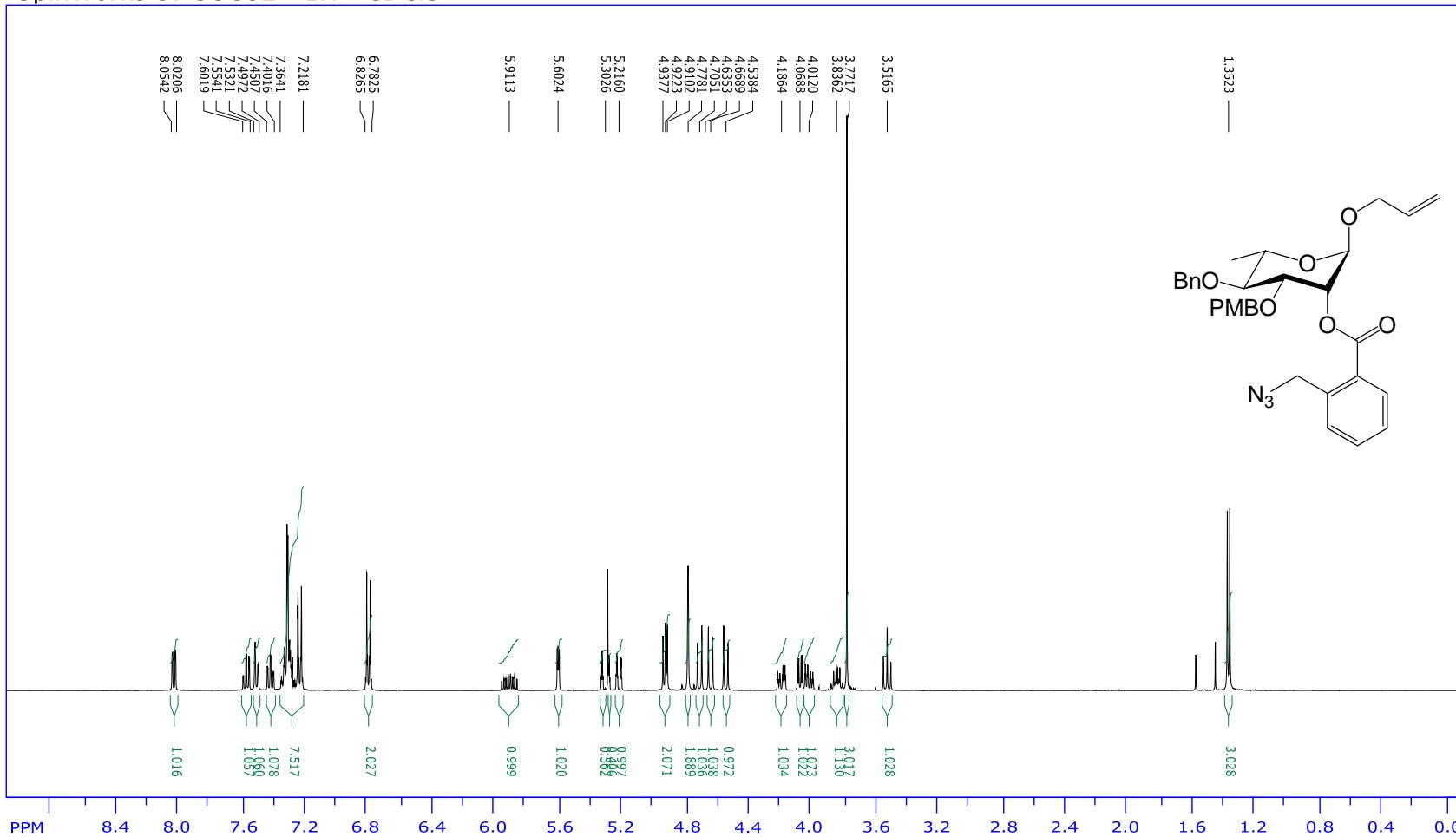


SpinWorks 3: CGG29 - 13C - CDCl3

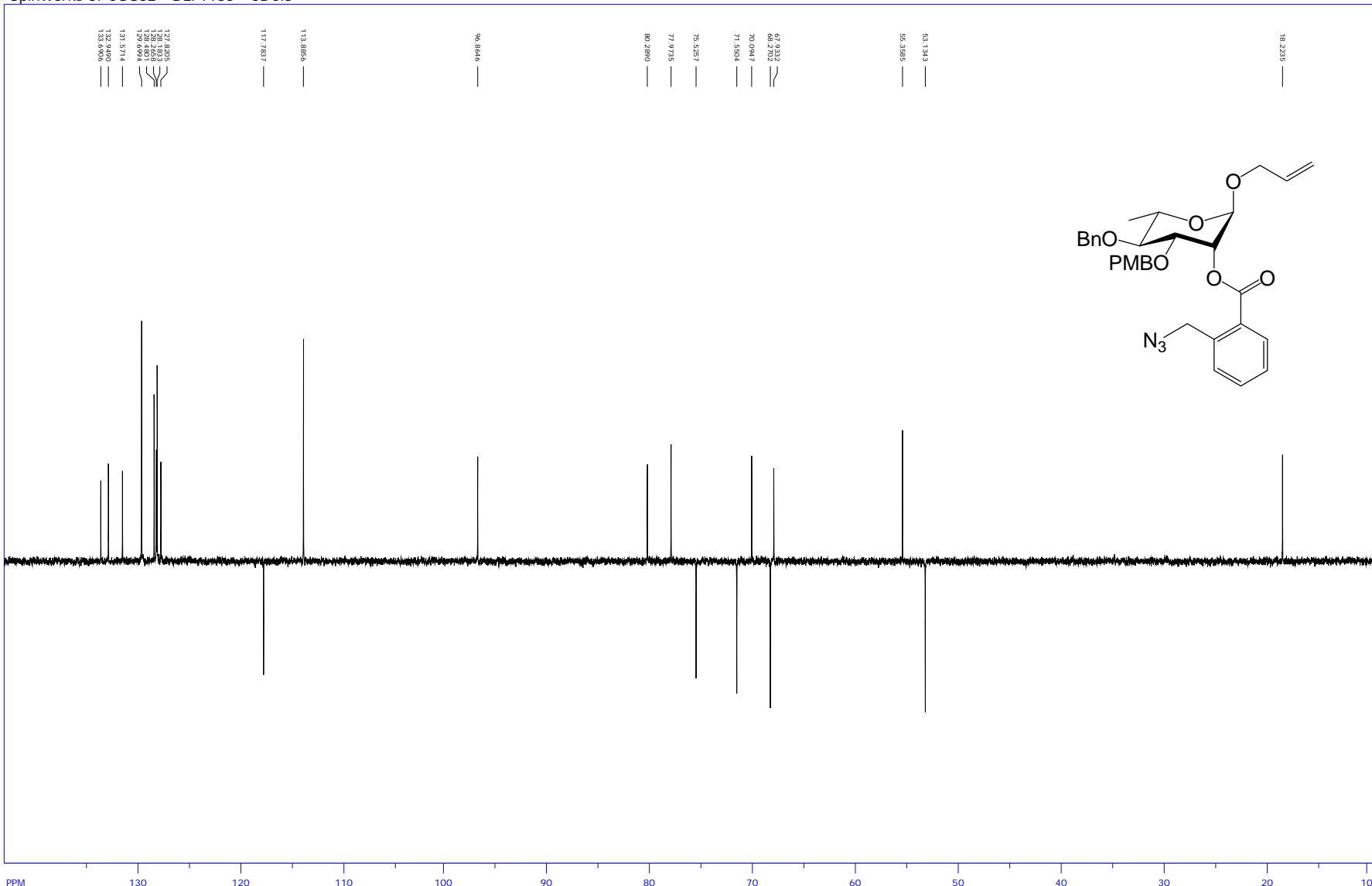


Compound S4

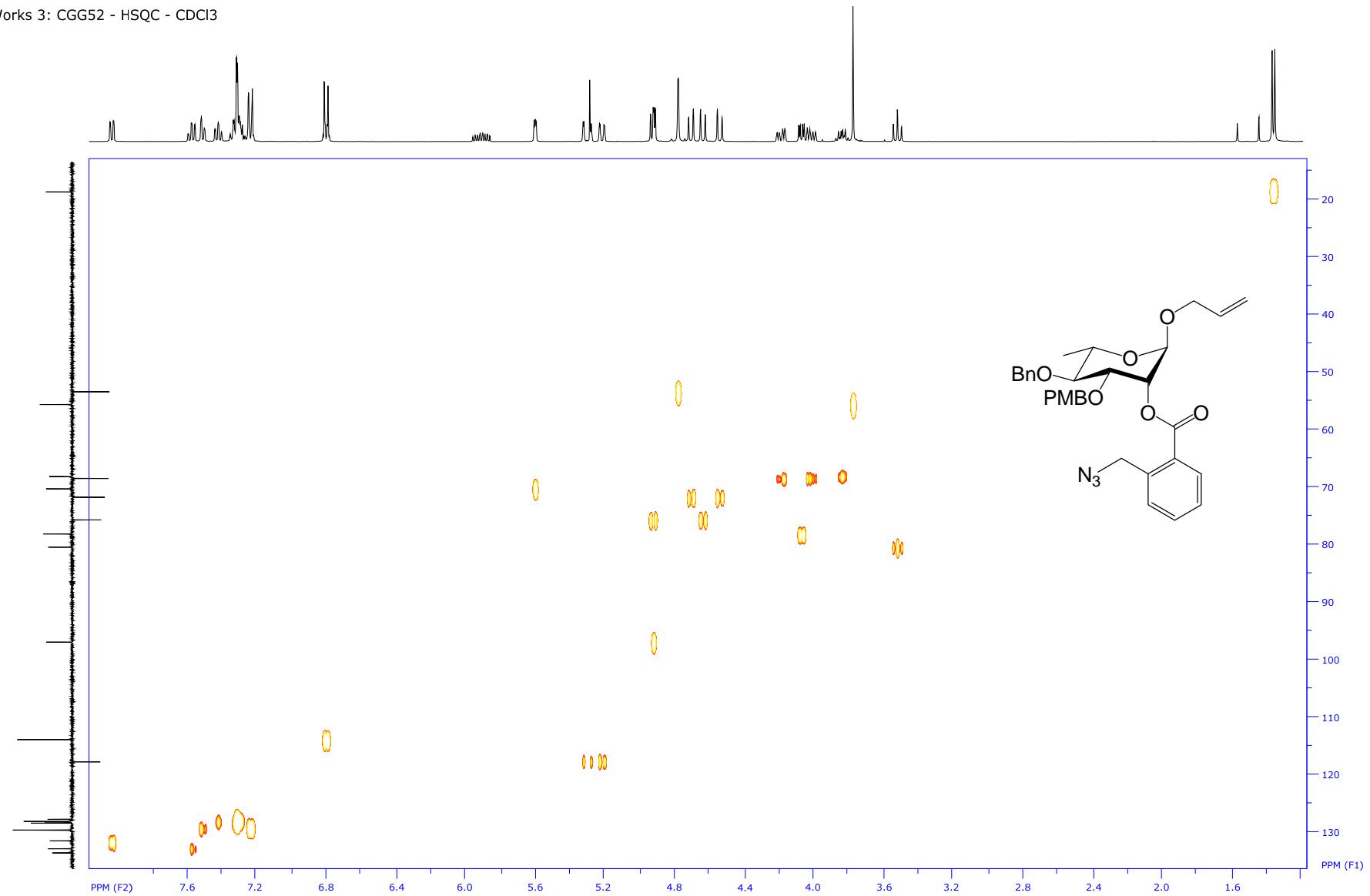
SpinWorks 3: CGG52 - 1H - CDCl₃



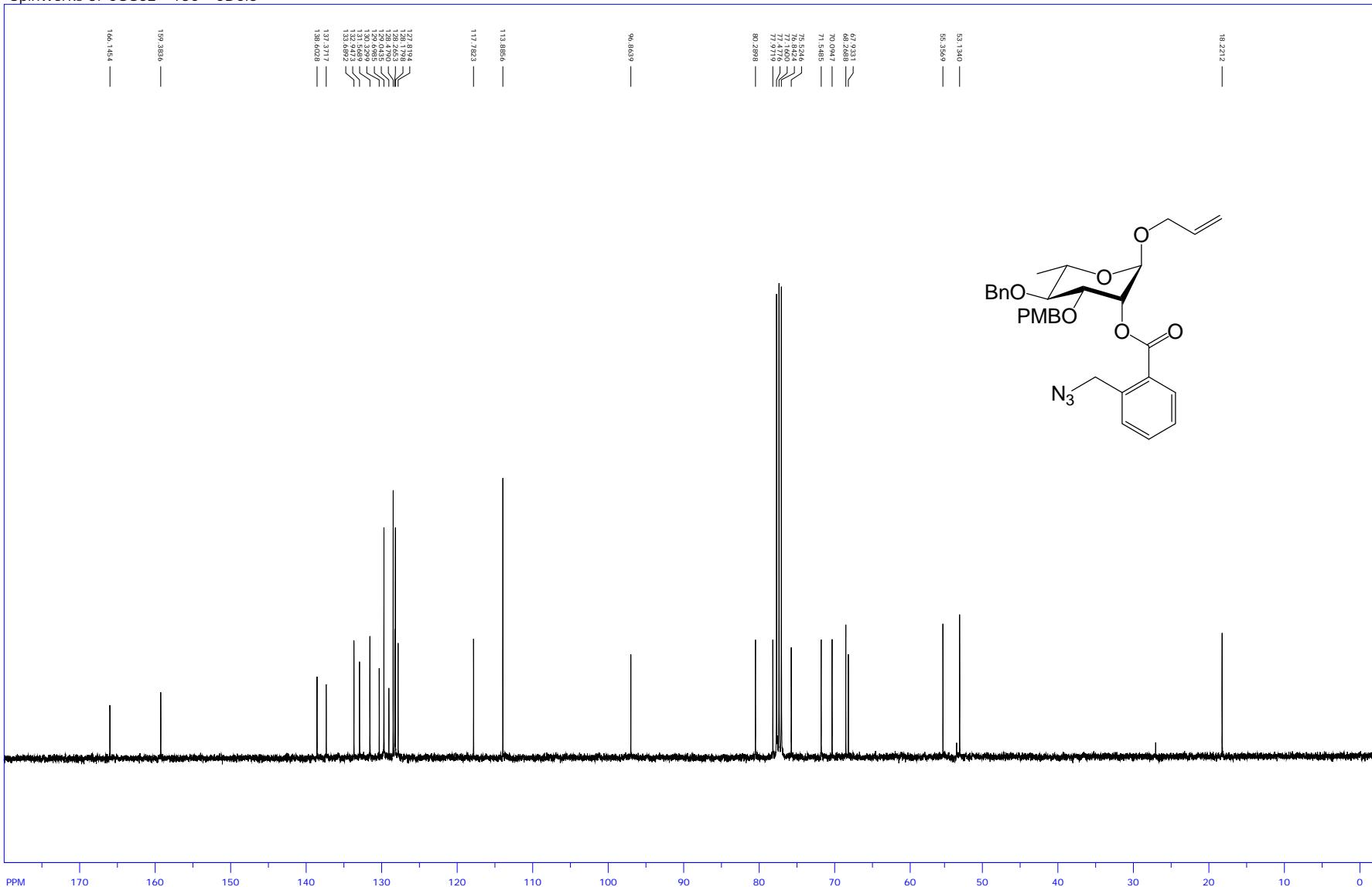
SpinWorks 3: CGG52 - DEPT135 - CDCl₃



SpinWorks 3: CGG52 - HSQC - CDCl₃

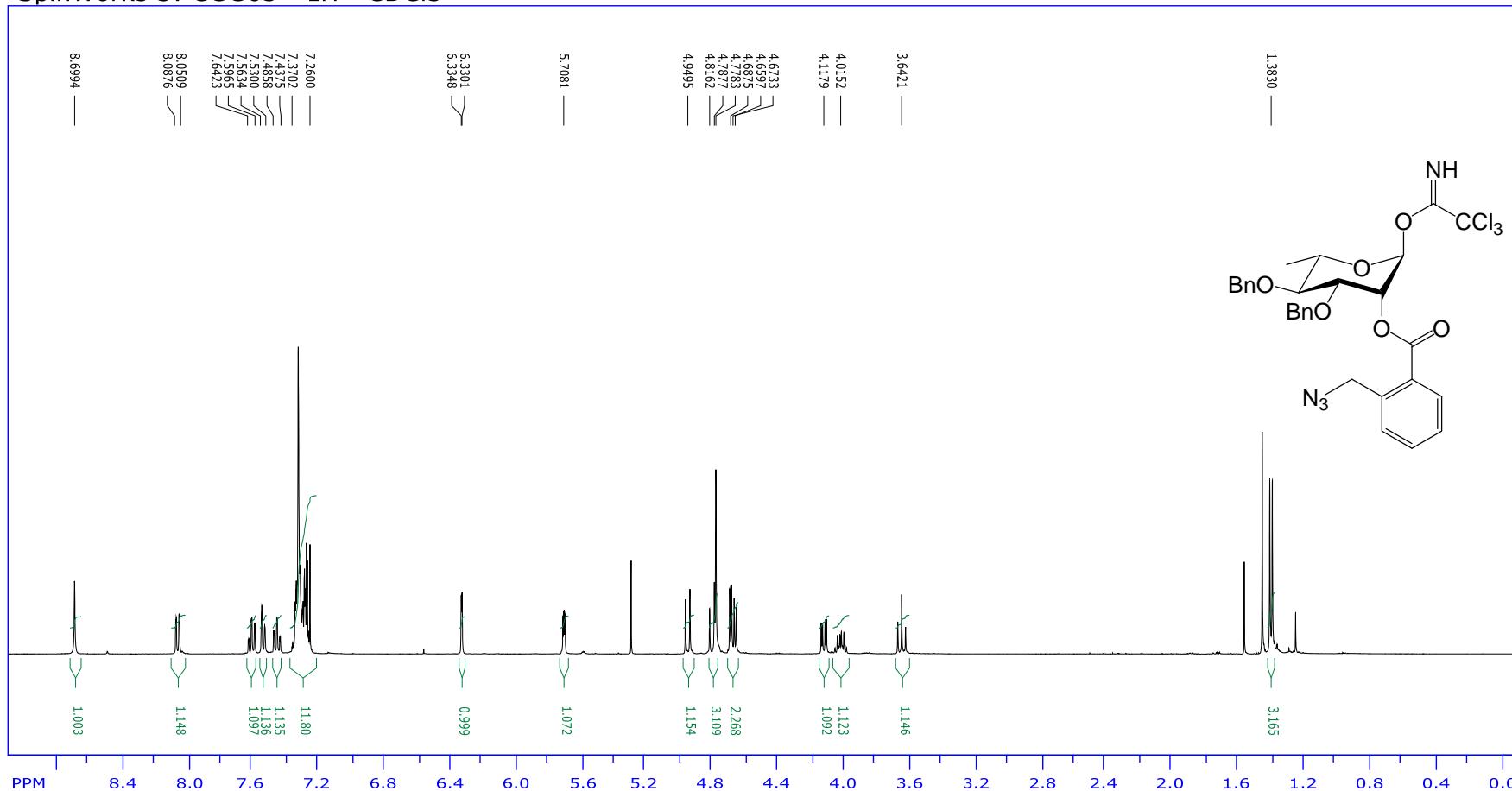


SpinWorks 3: CGG52 - 13C - CDCl3

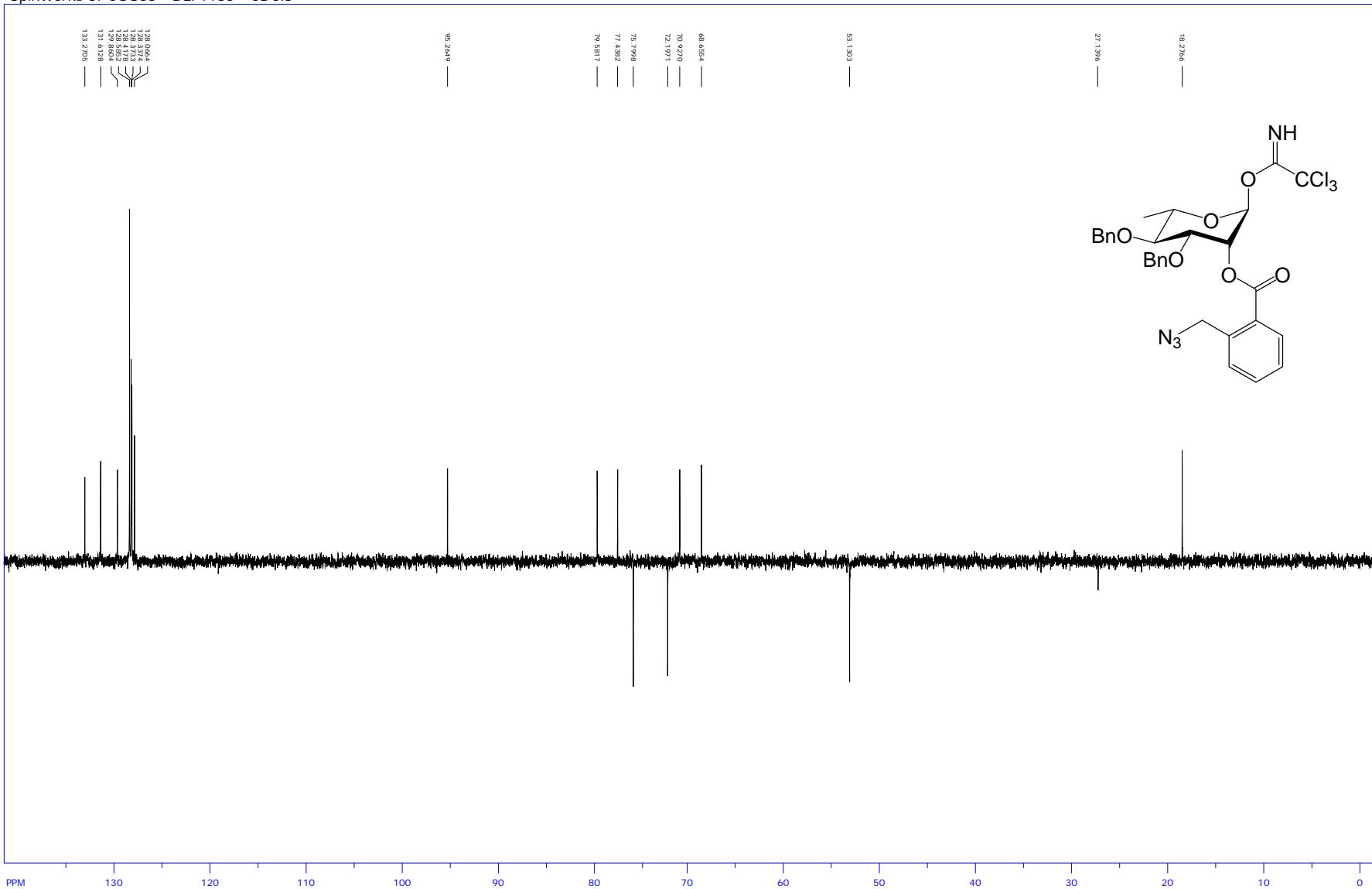


Compound 12

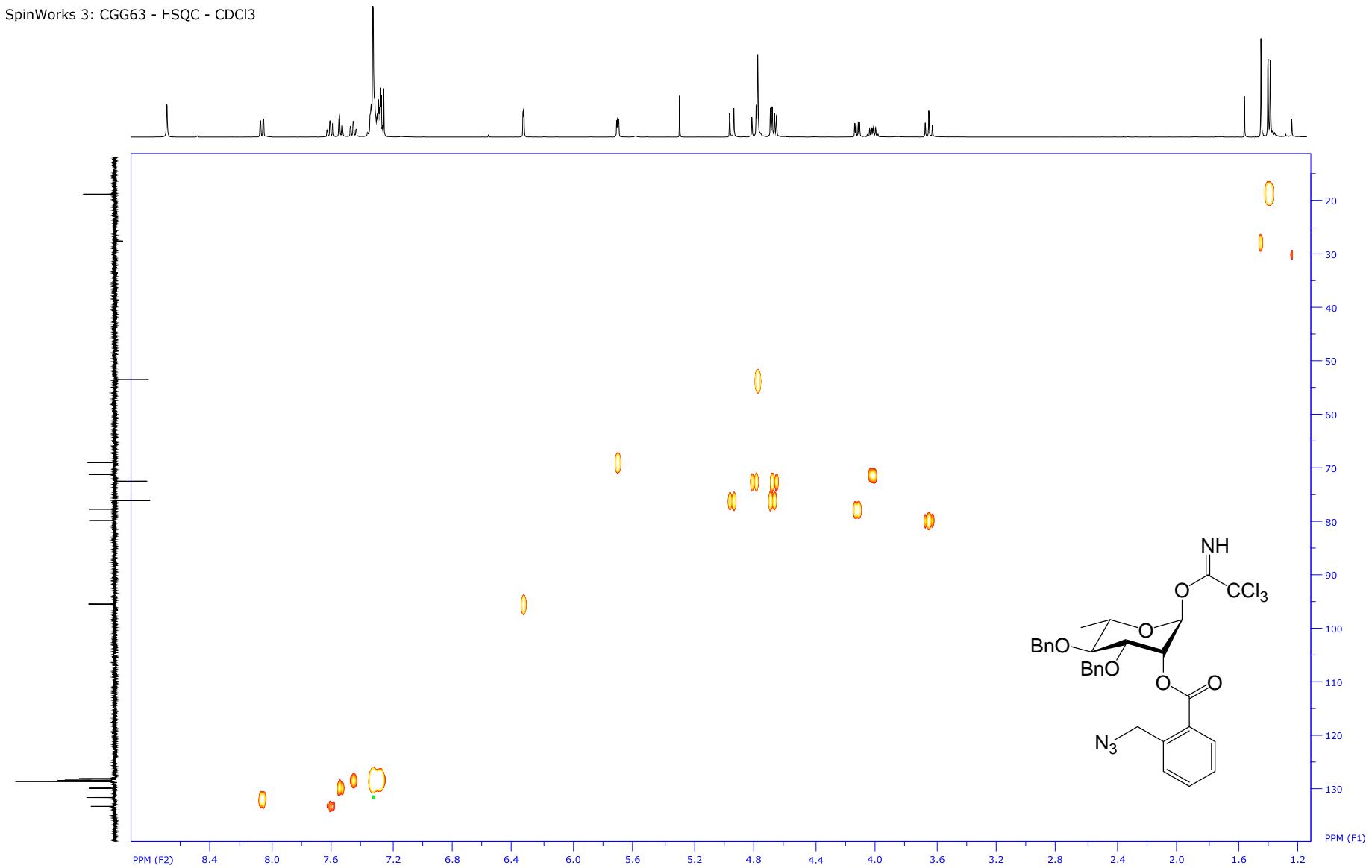
SpinWorks 3: CGG63 - 1H - CDCl₃

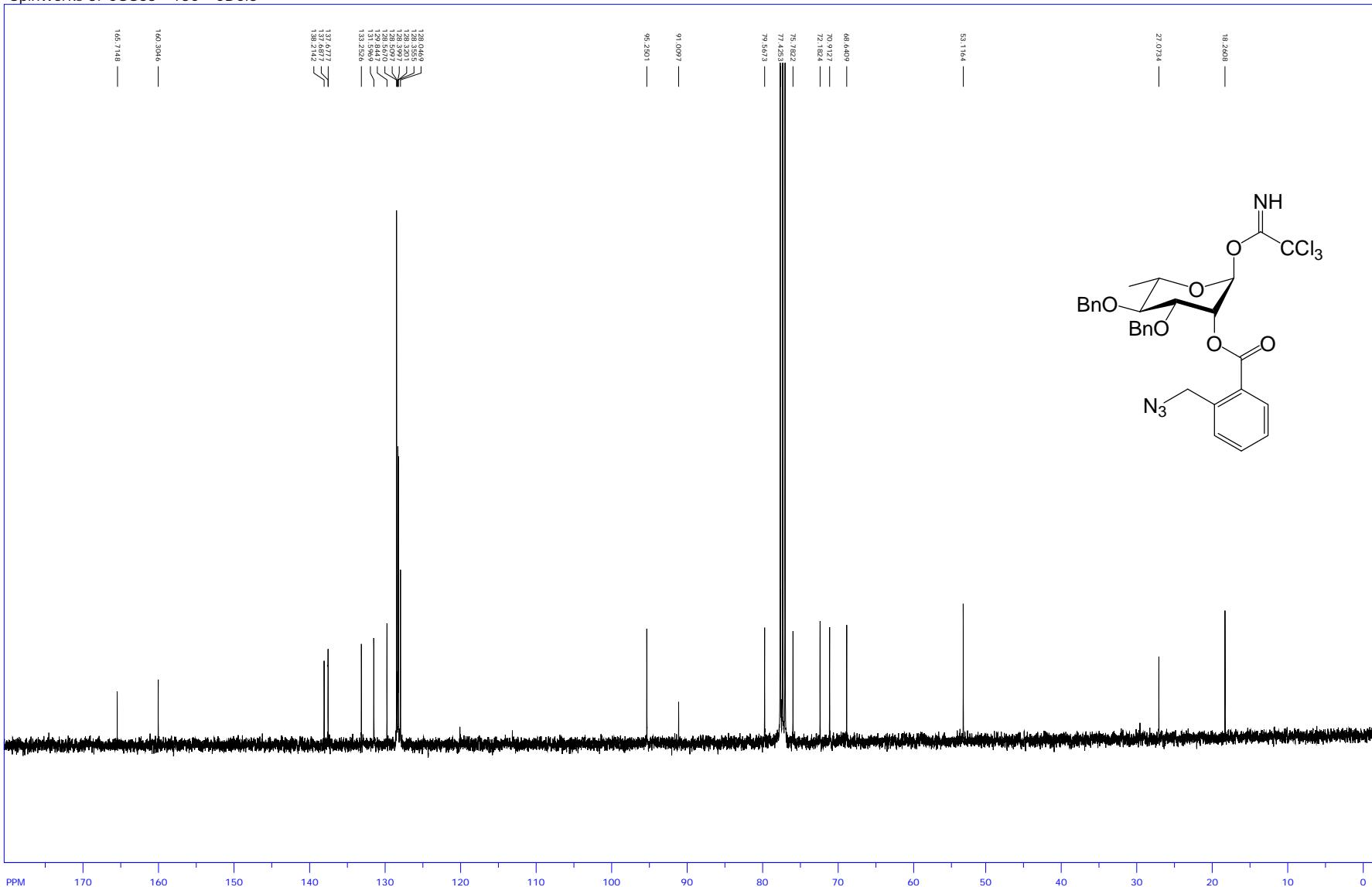


SpinWorks 3: CGG63 - DEPT135 - CDCl₃



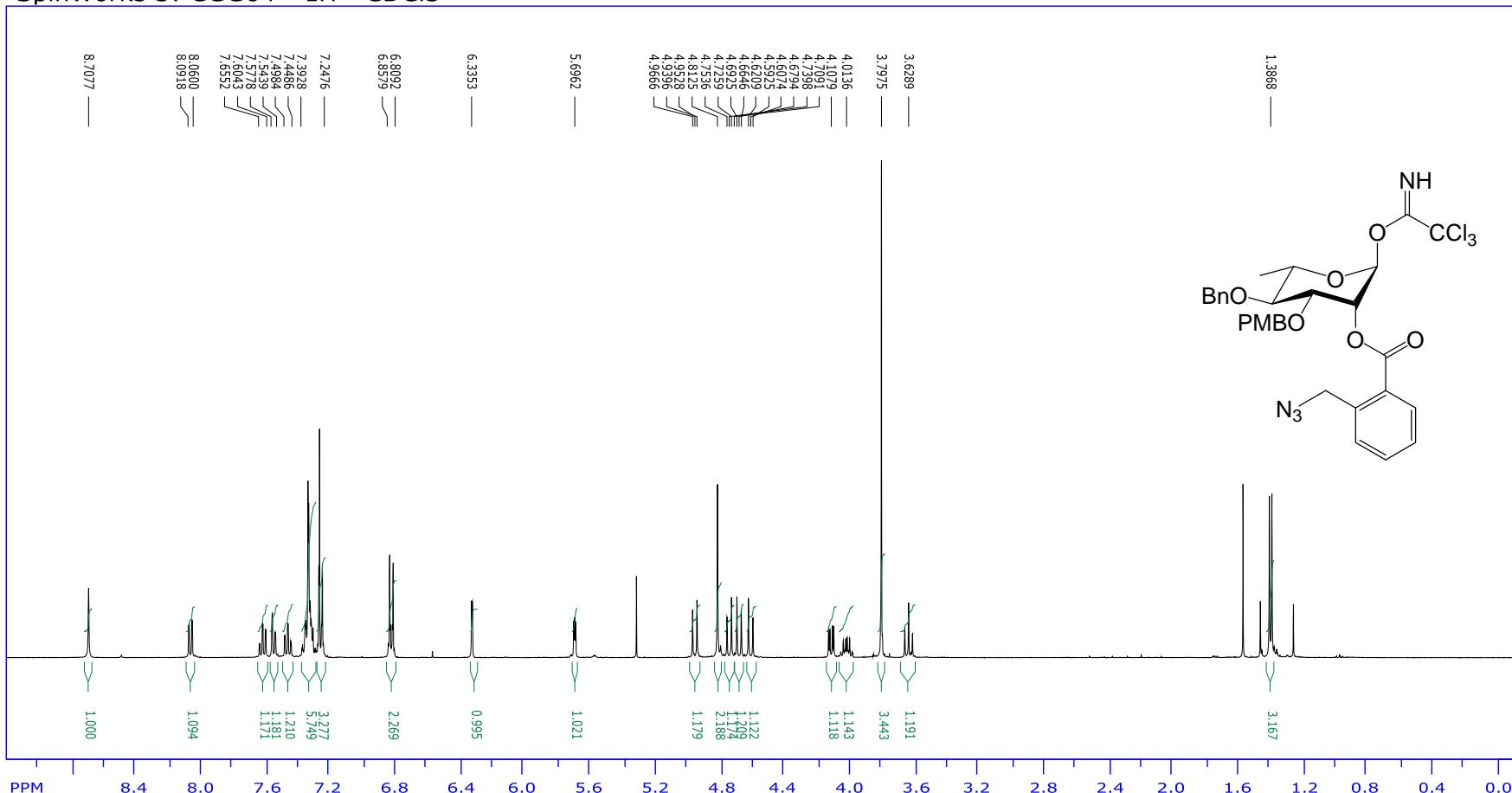
SpinWorks 3: CGG63 - HSQC - CDCl₃



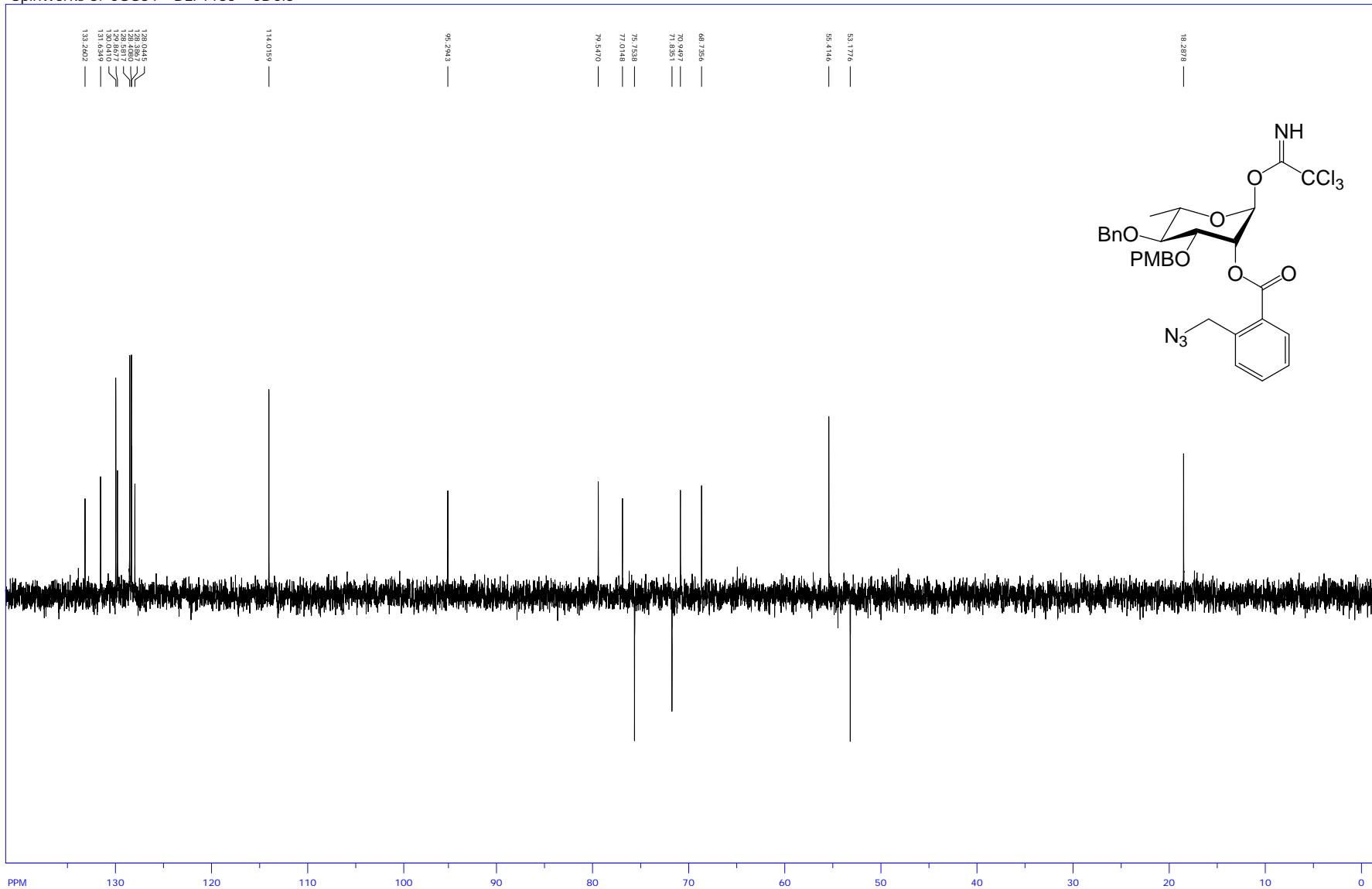


Compound 13

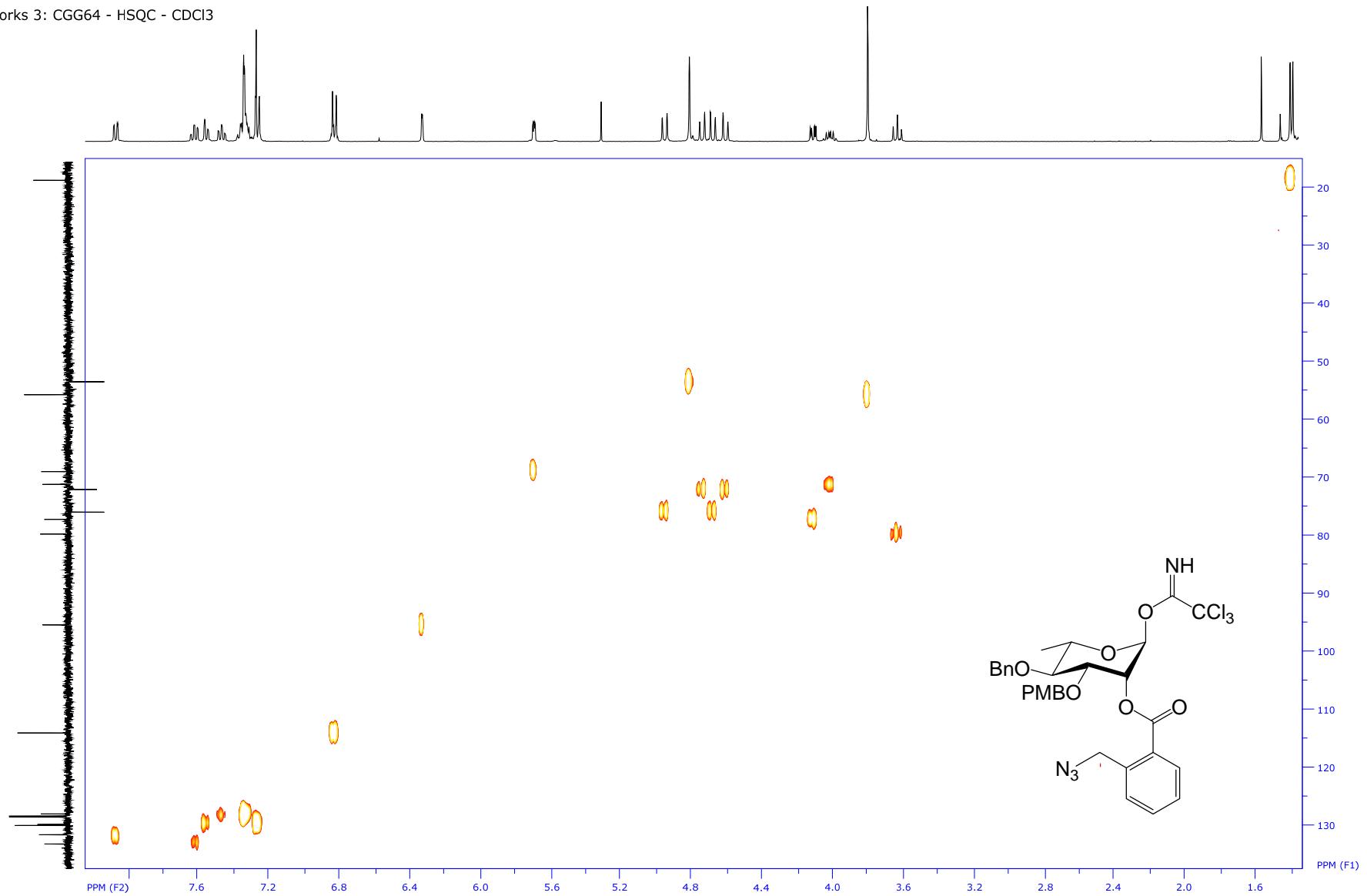
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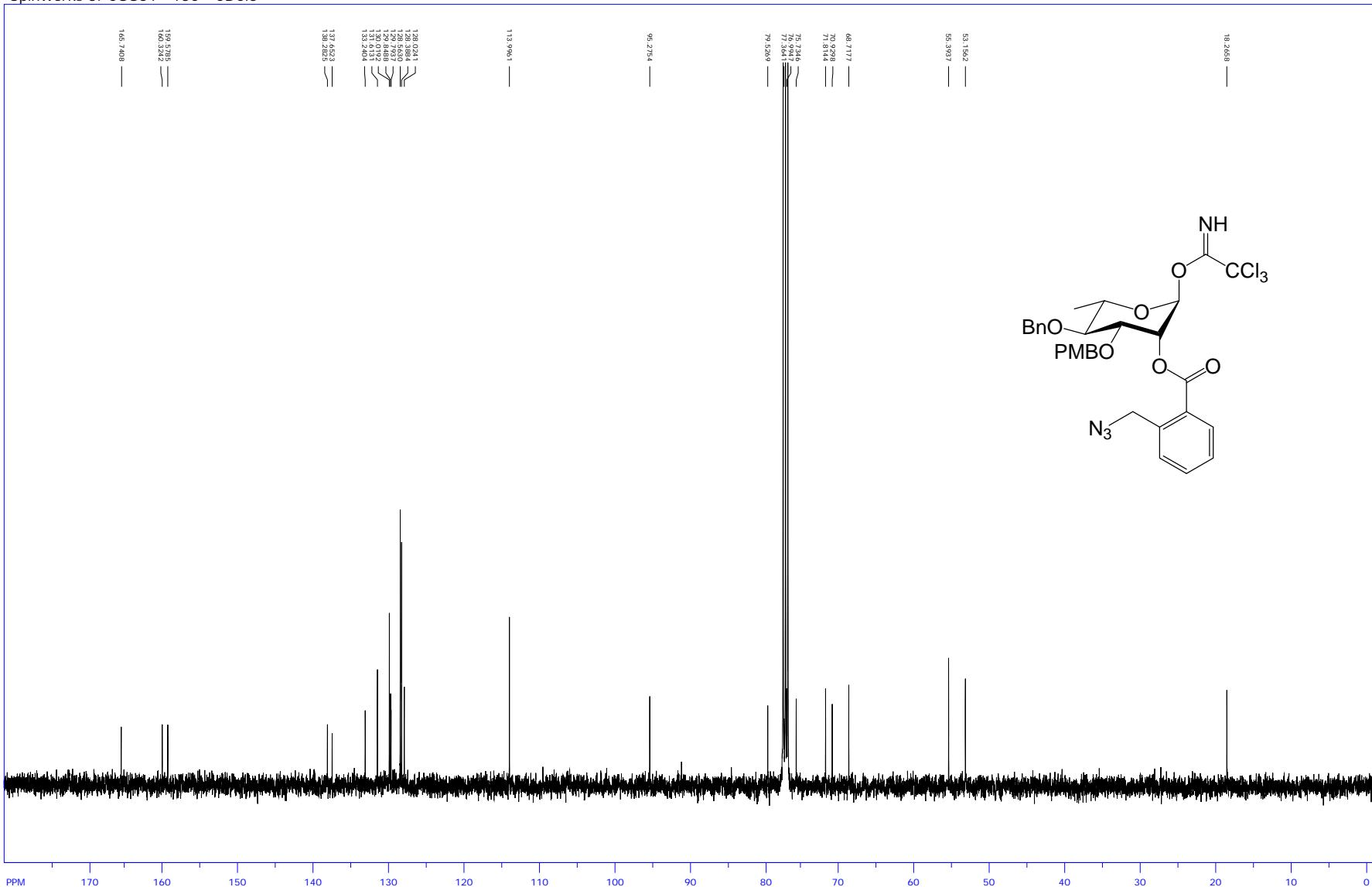


SpinWorks 3: CGG64 - DEPT135 - CDCI3



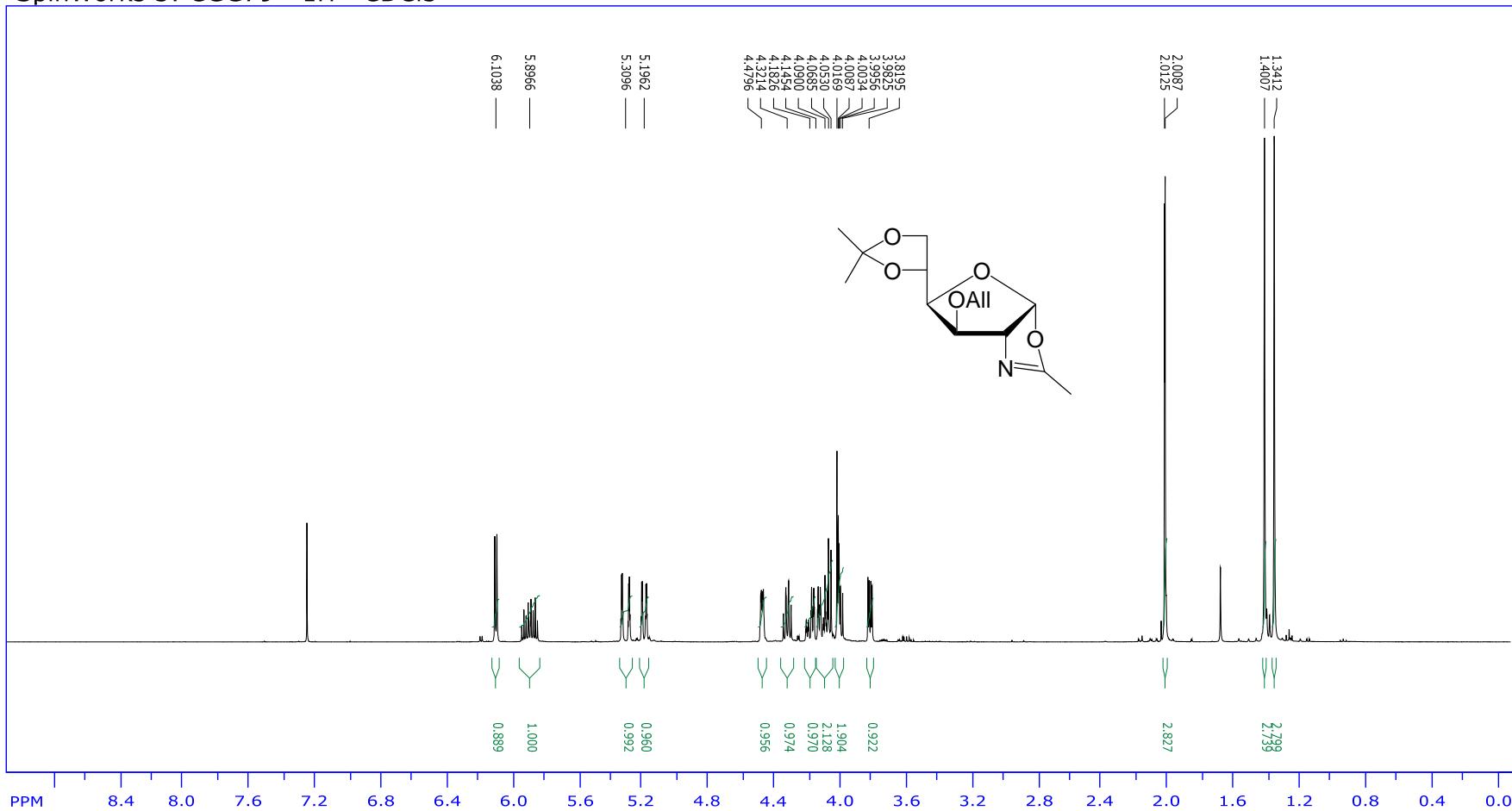
SpinWorks 3: CGG64 - HSQC - CDCl₃



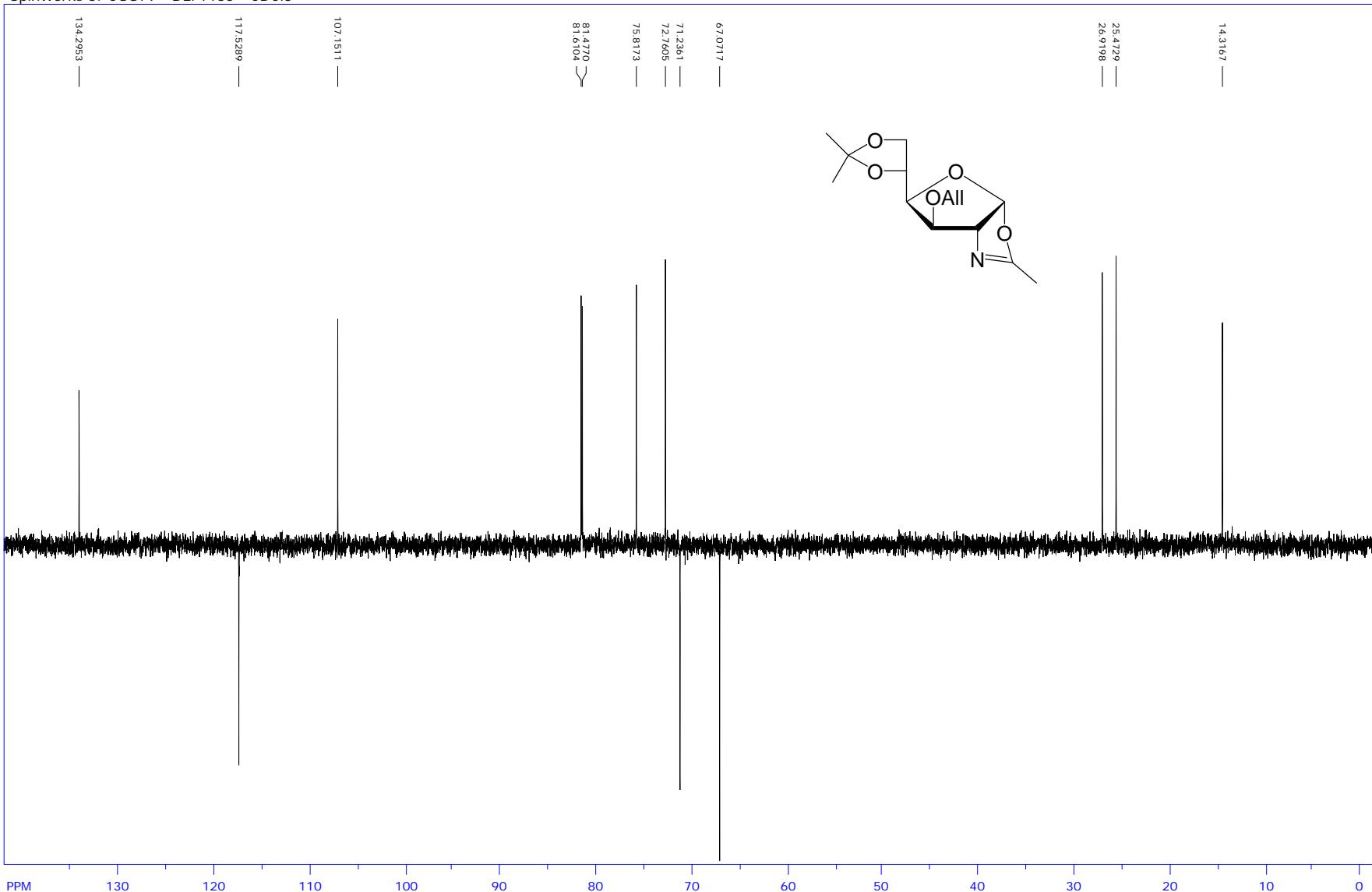
SpinWorks 3: CGG64 - 13C - CDCl₃

Compound 31

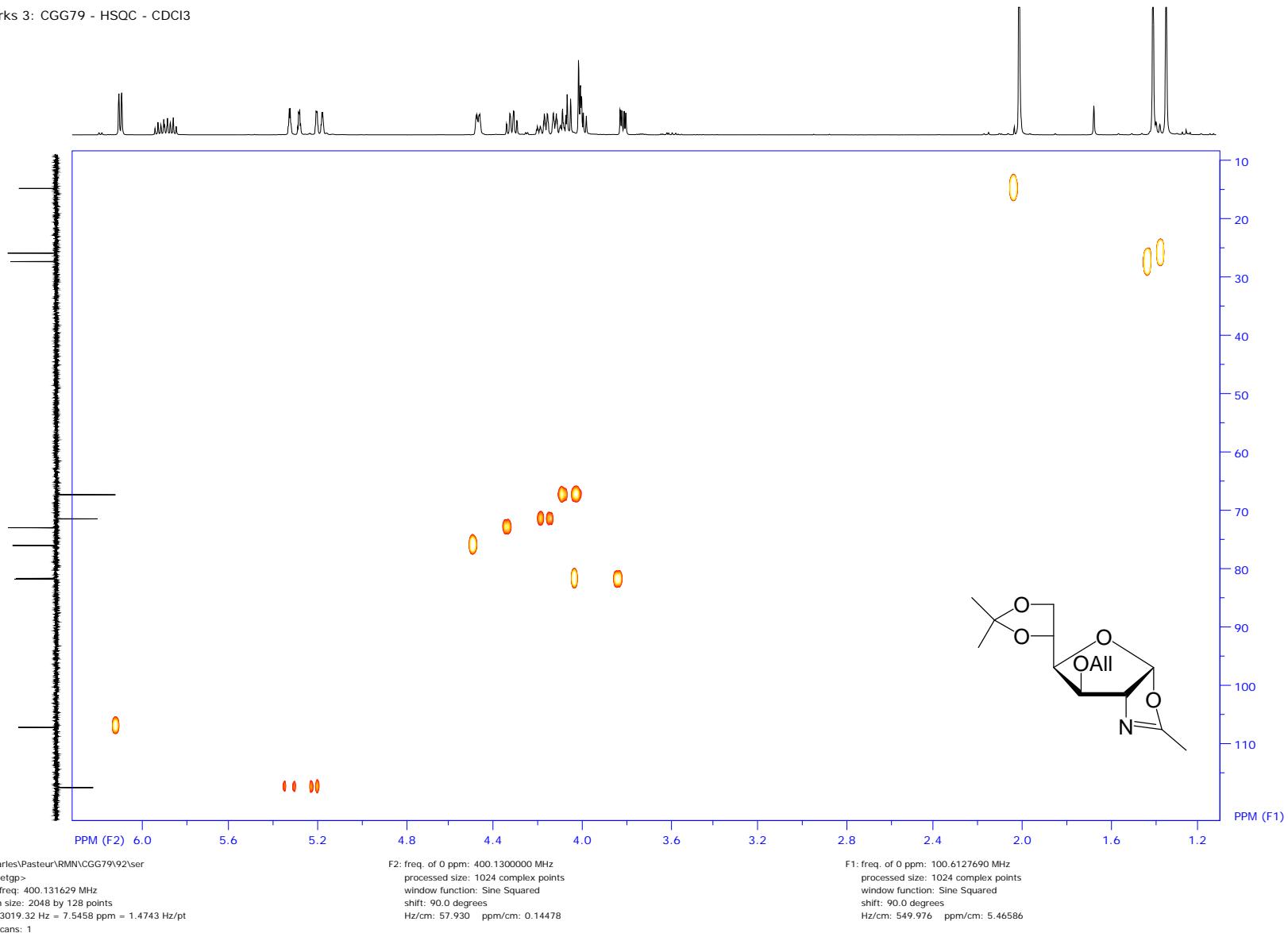
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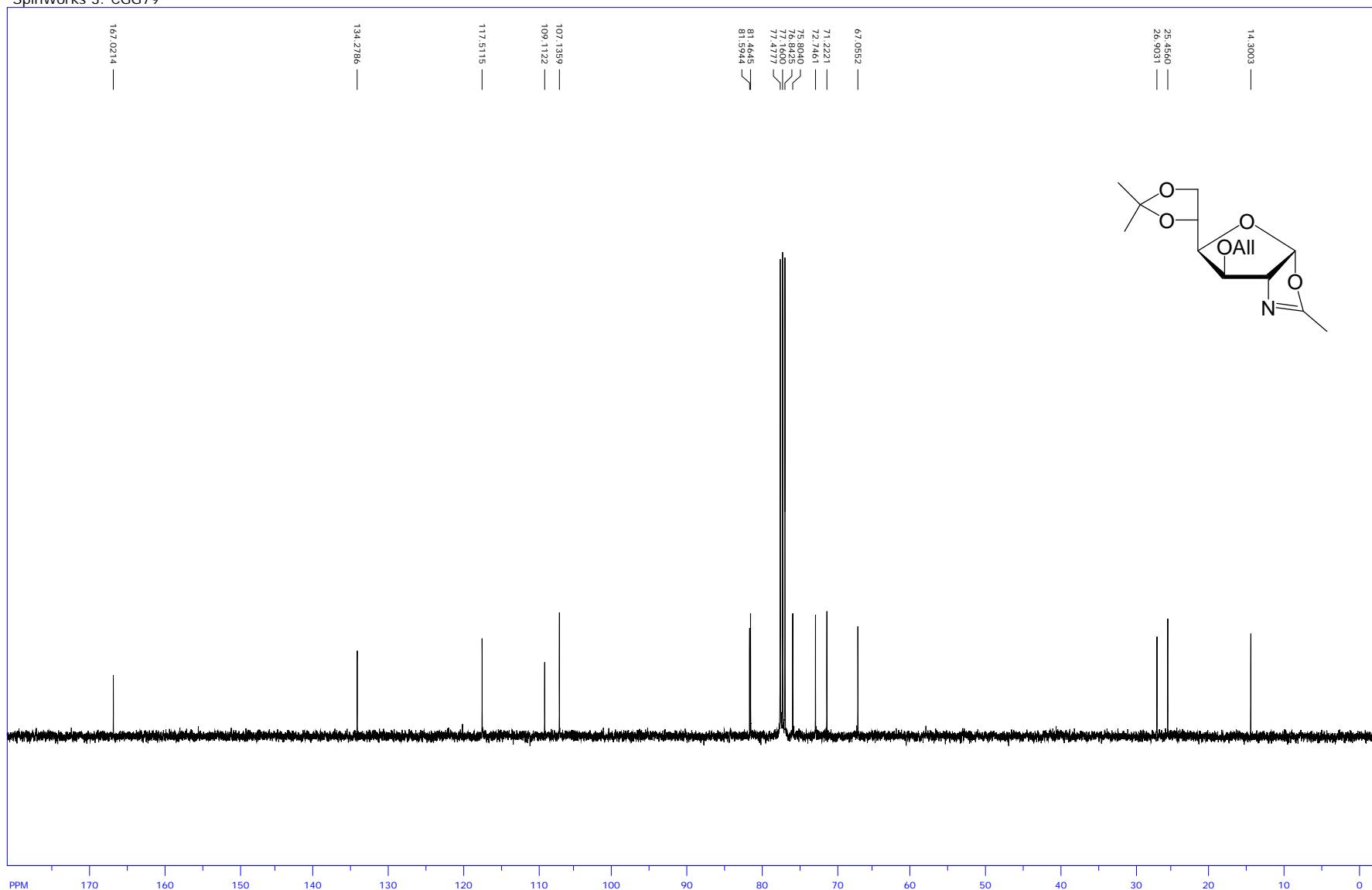


SpinWorks 3: CGG79 - DEPT135 - CDCl₃

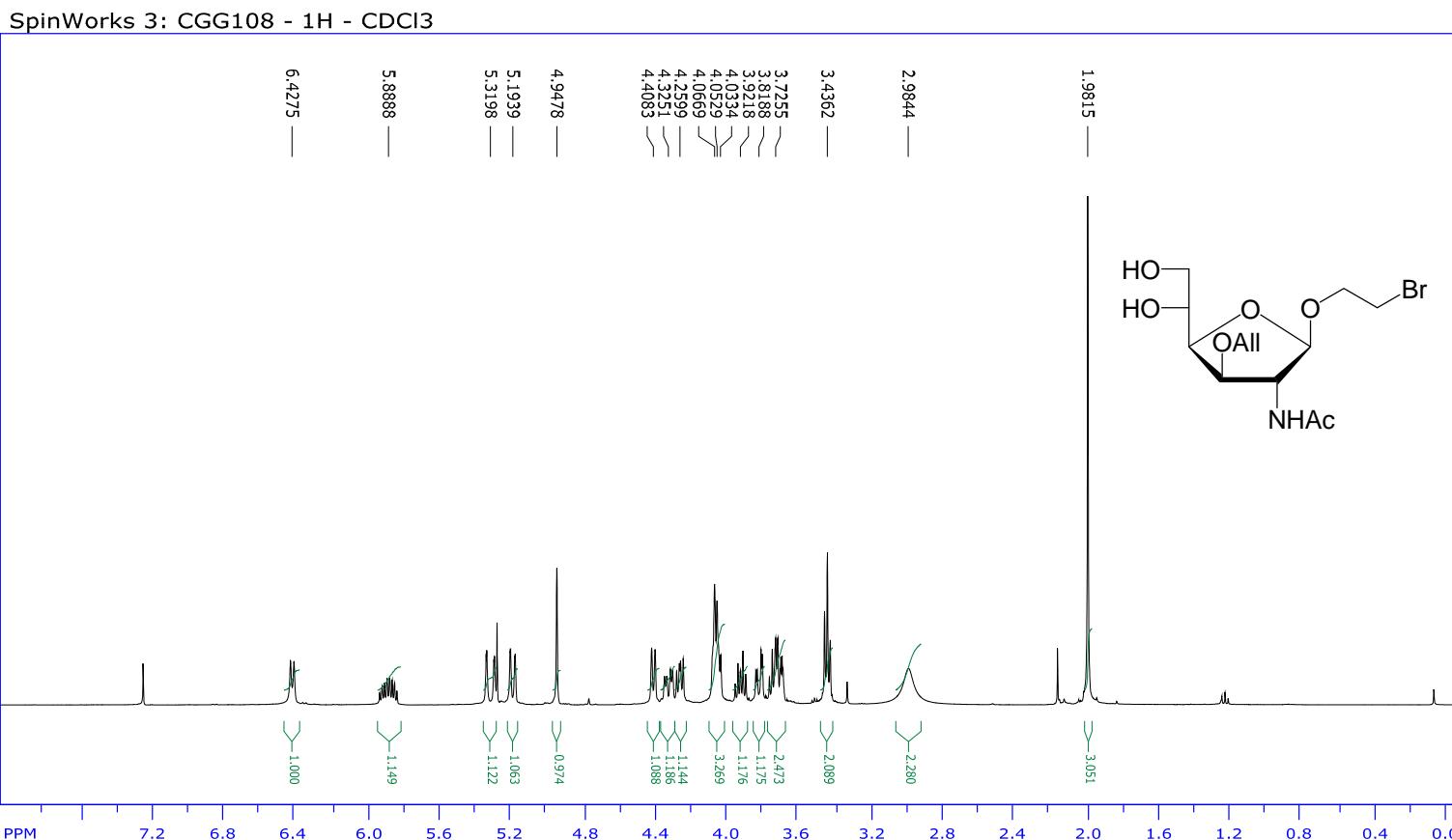


SpinWorks 3: CGG79 - HSQC - CDCl₃

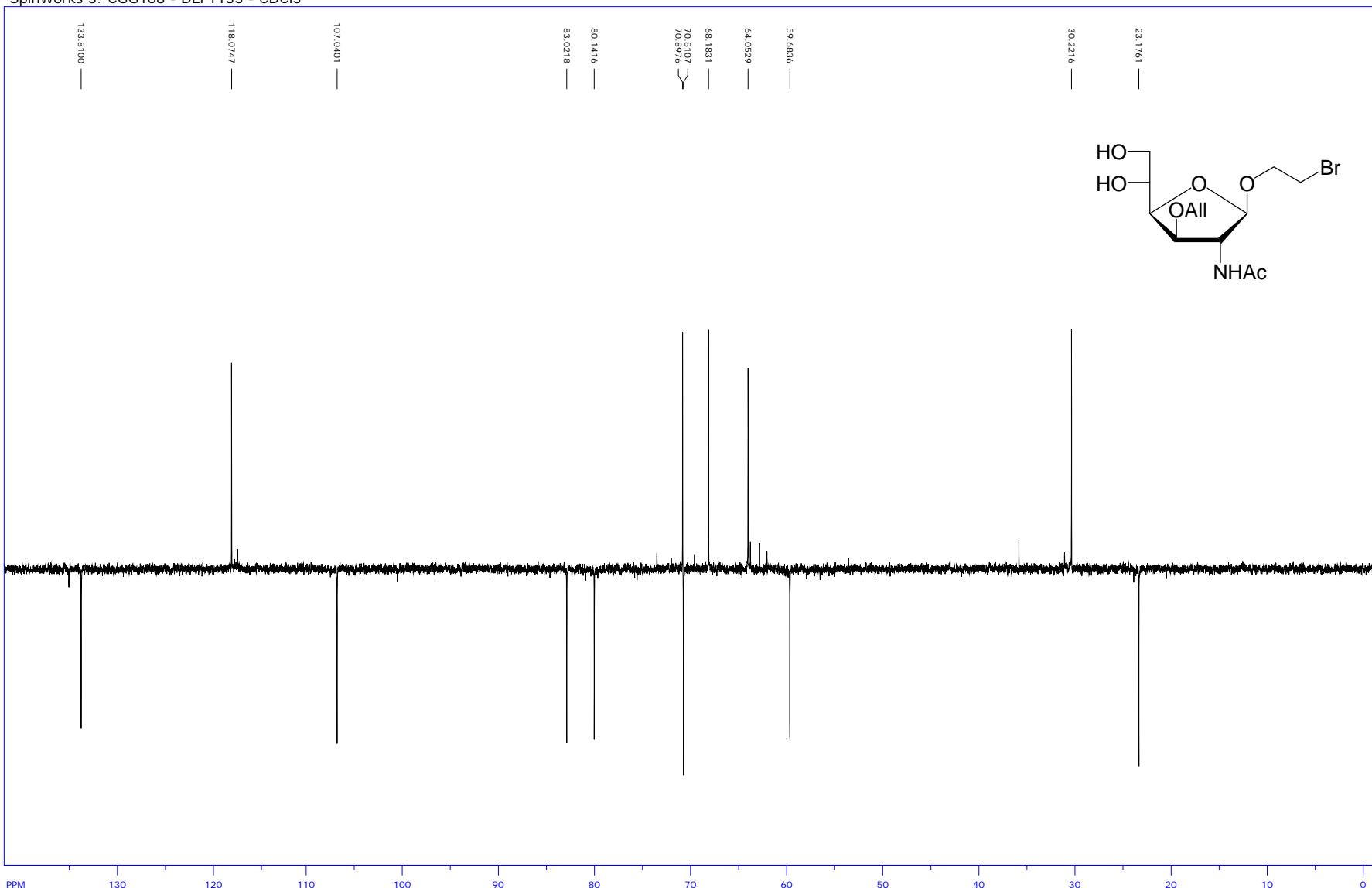




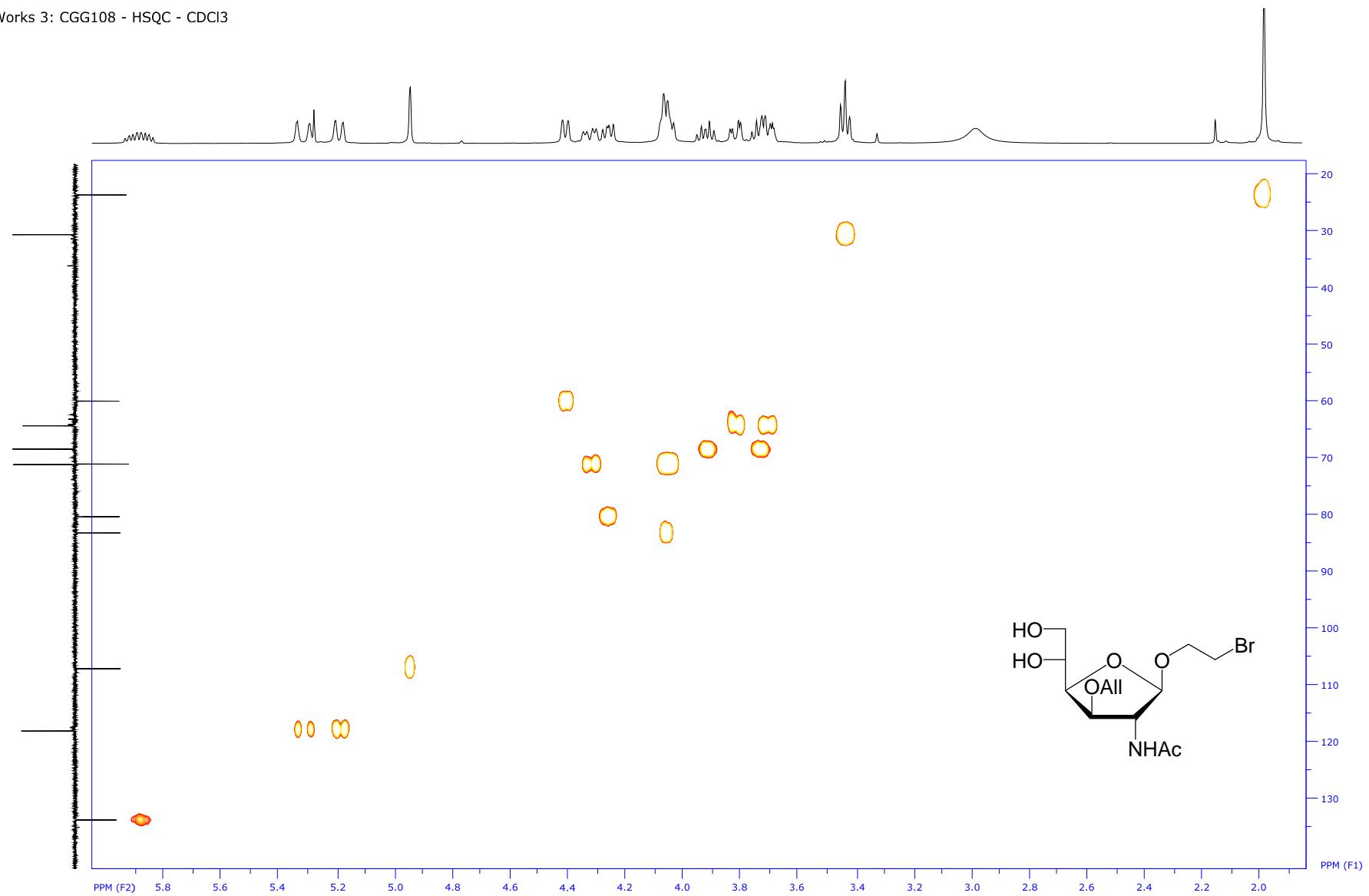
Compound S8

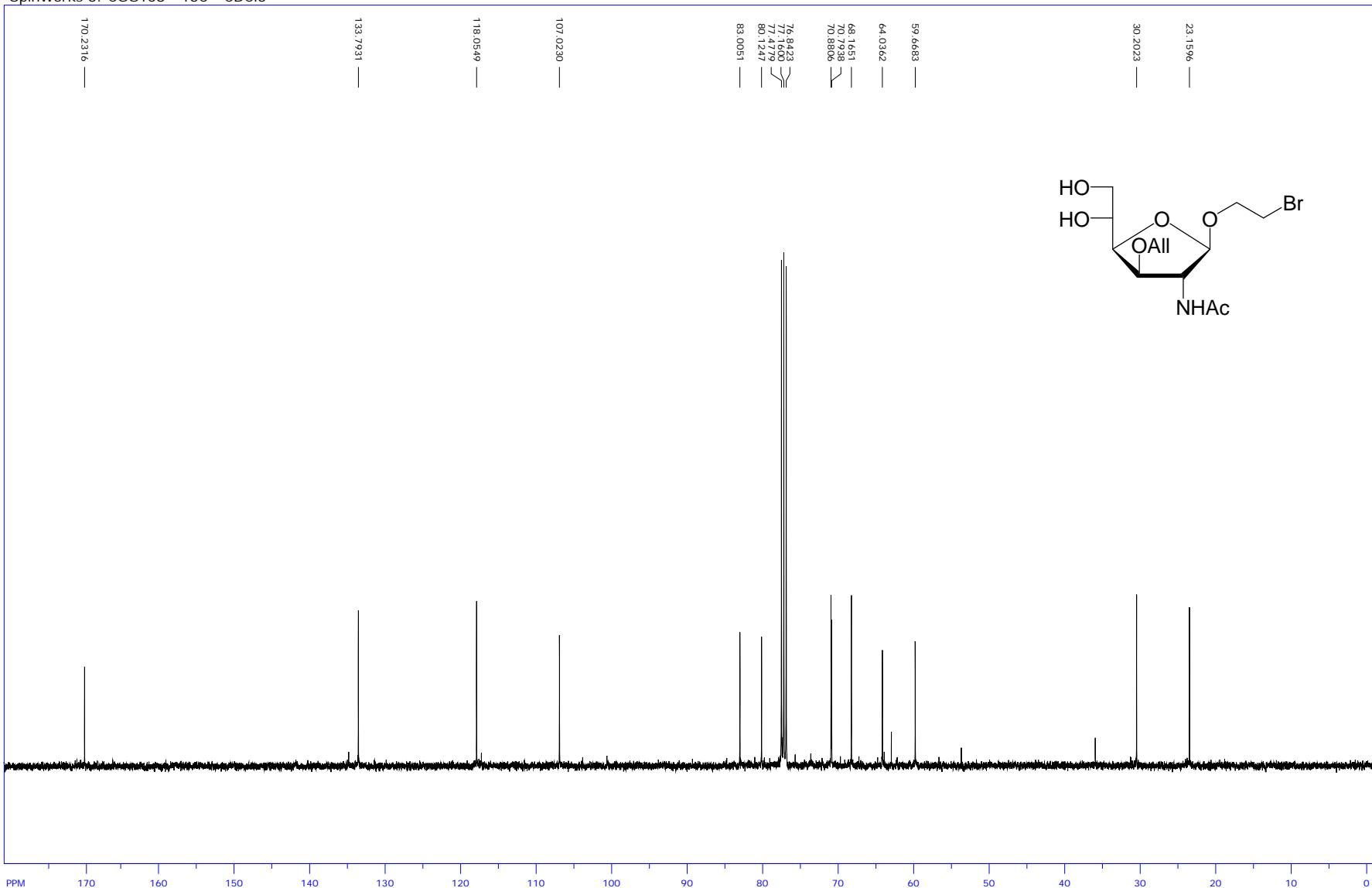


SpinWorks 3: CGG108 - DEPT135 - CDCl₃



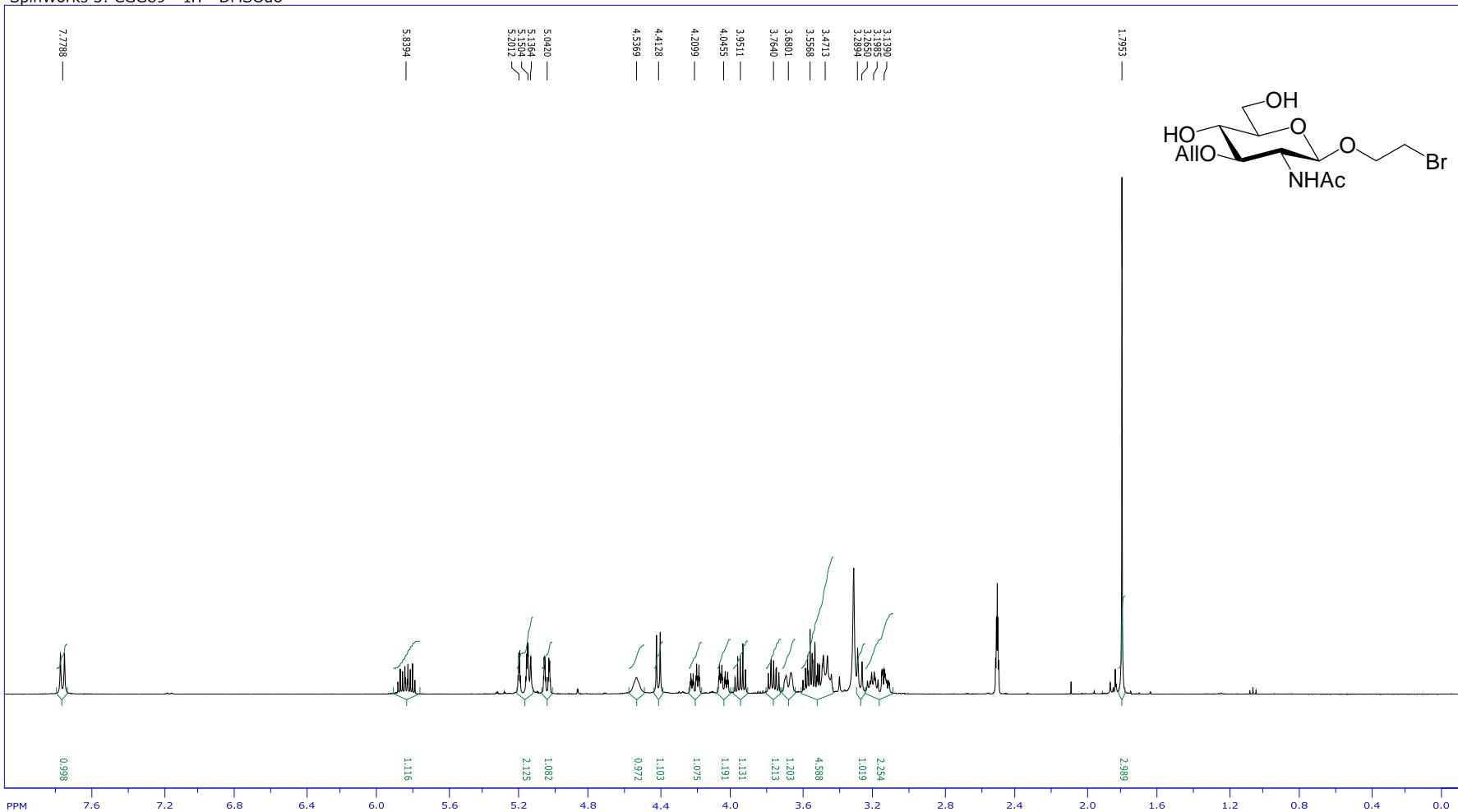
SpinWorks 3: CGG108 - HSQC - CDCl₃



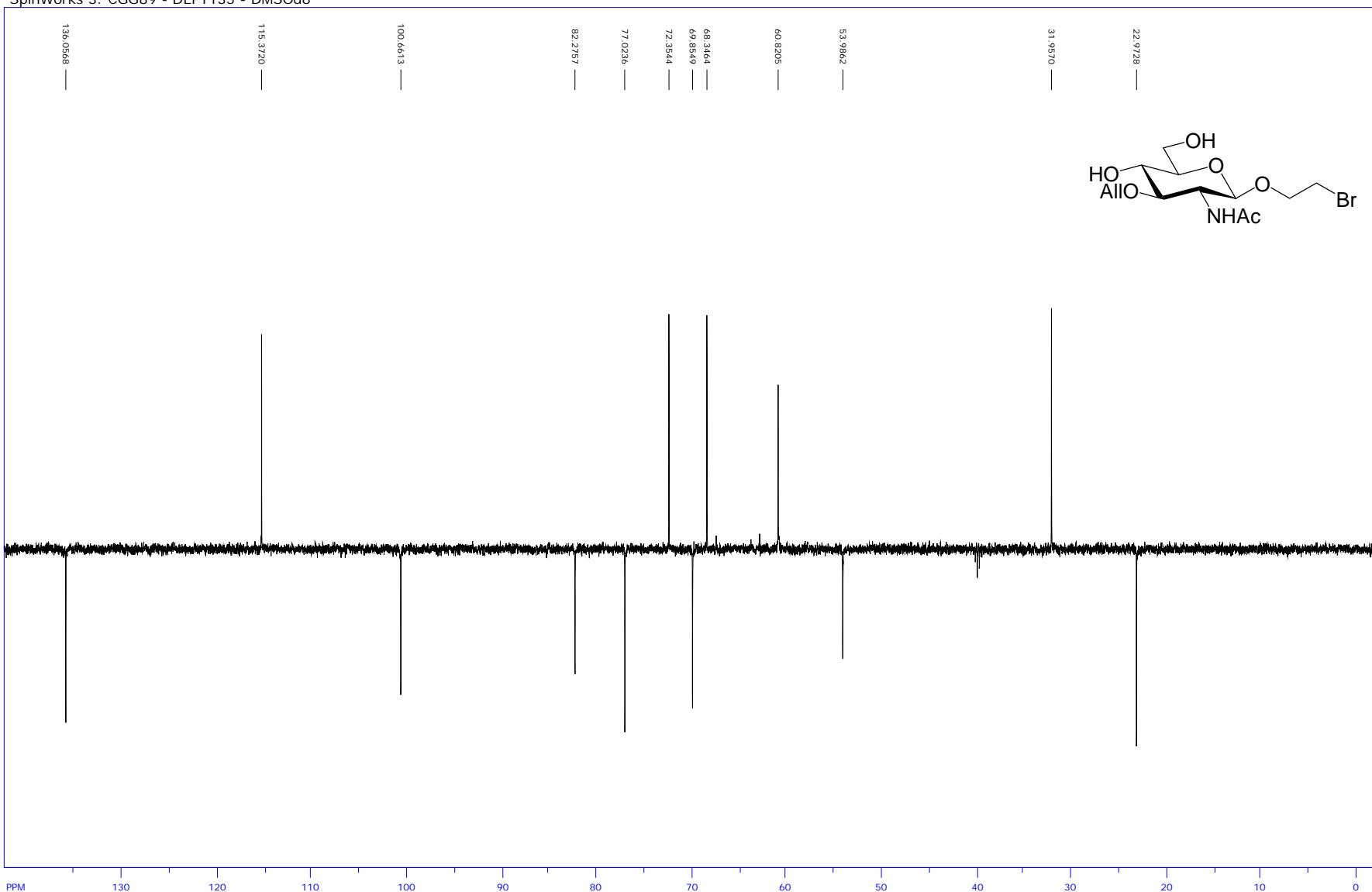


Compound 32

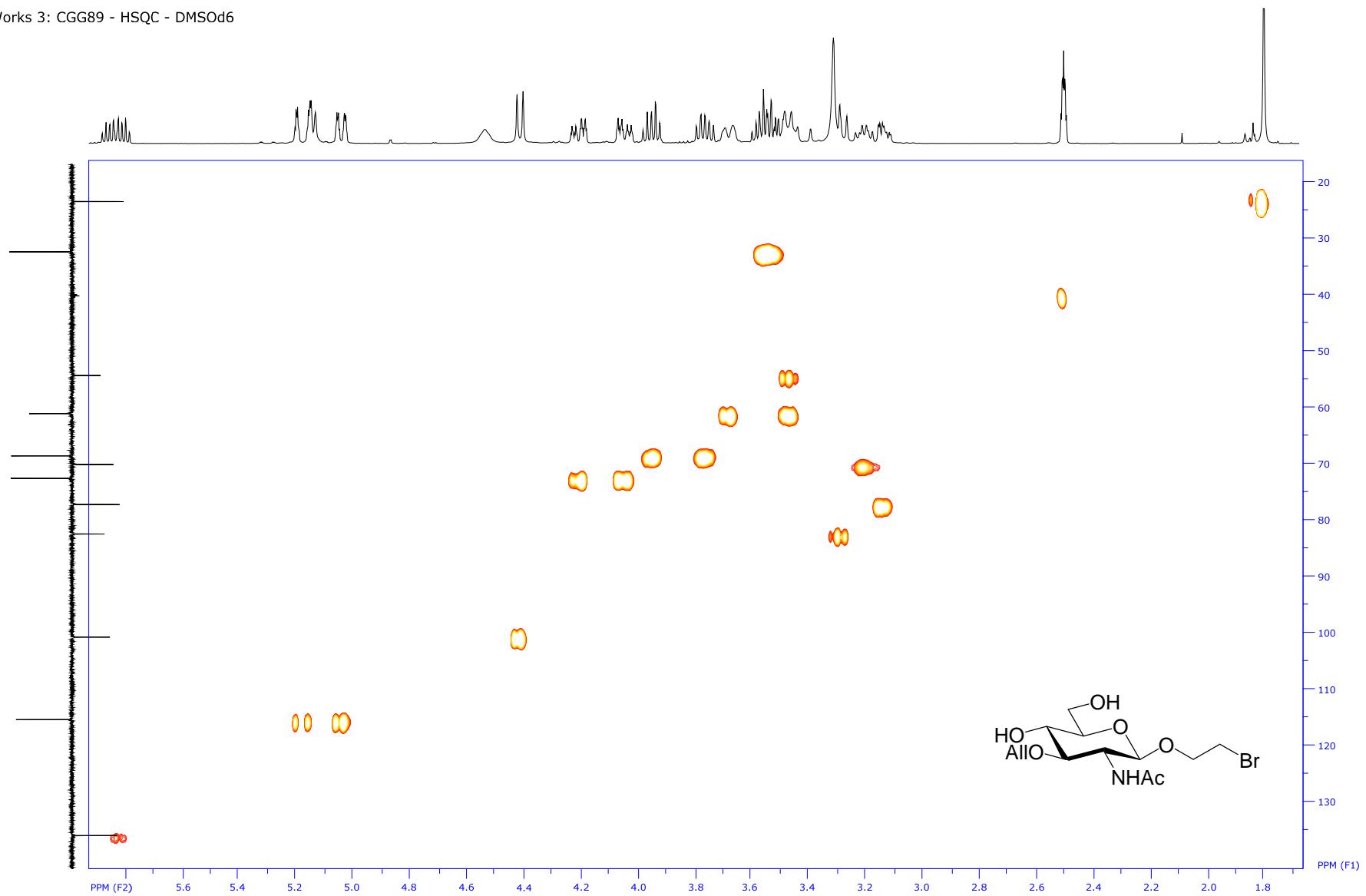
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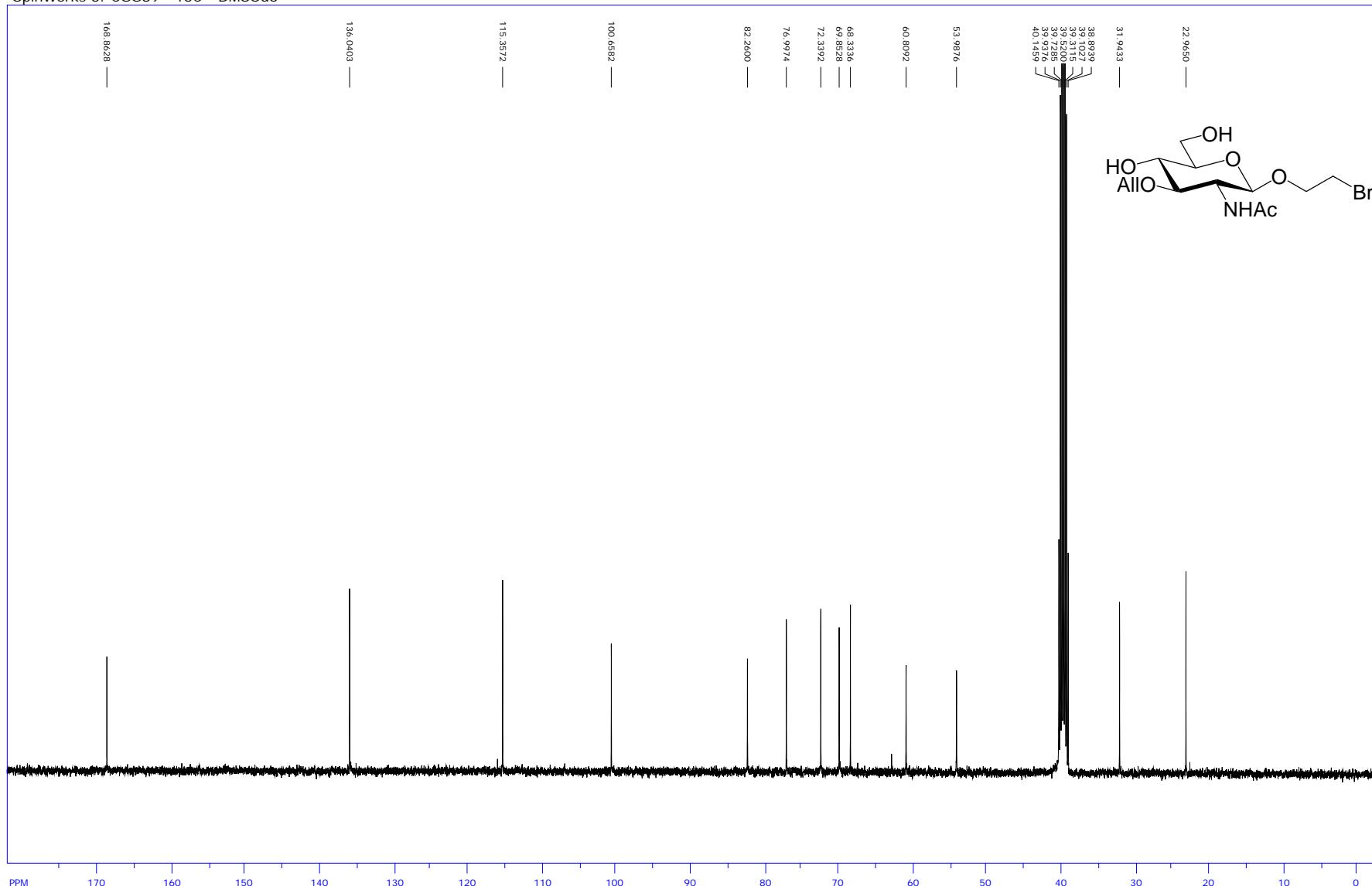
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SpinWorks 3: CGG89 - HSQC - DMSOd6

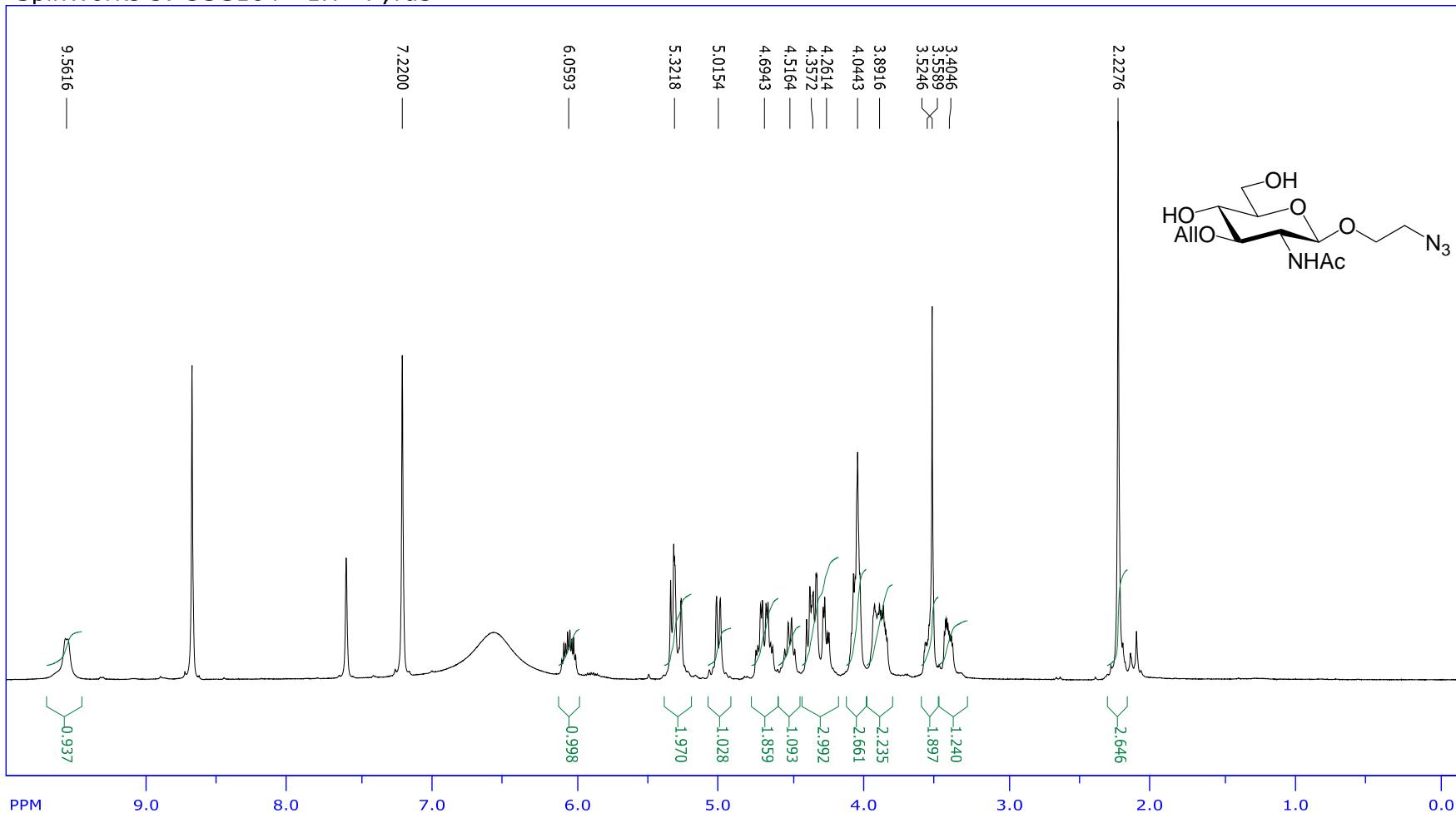


SpinWorks 3: CGG89 - 13C - DMSOd6

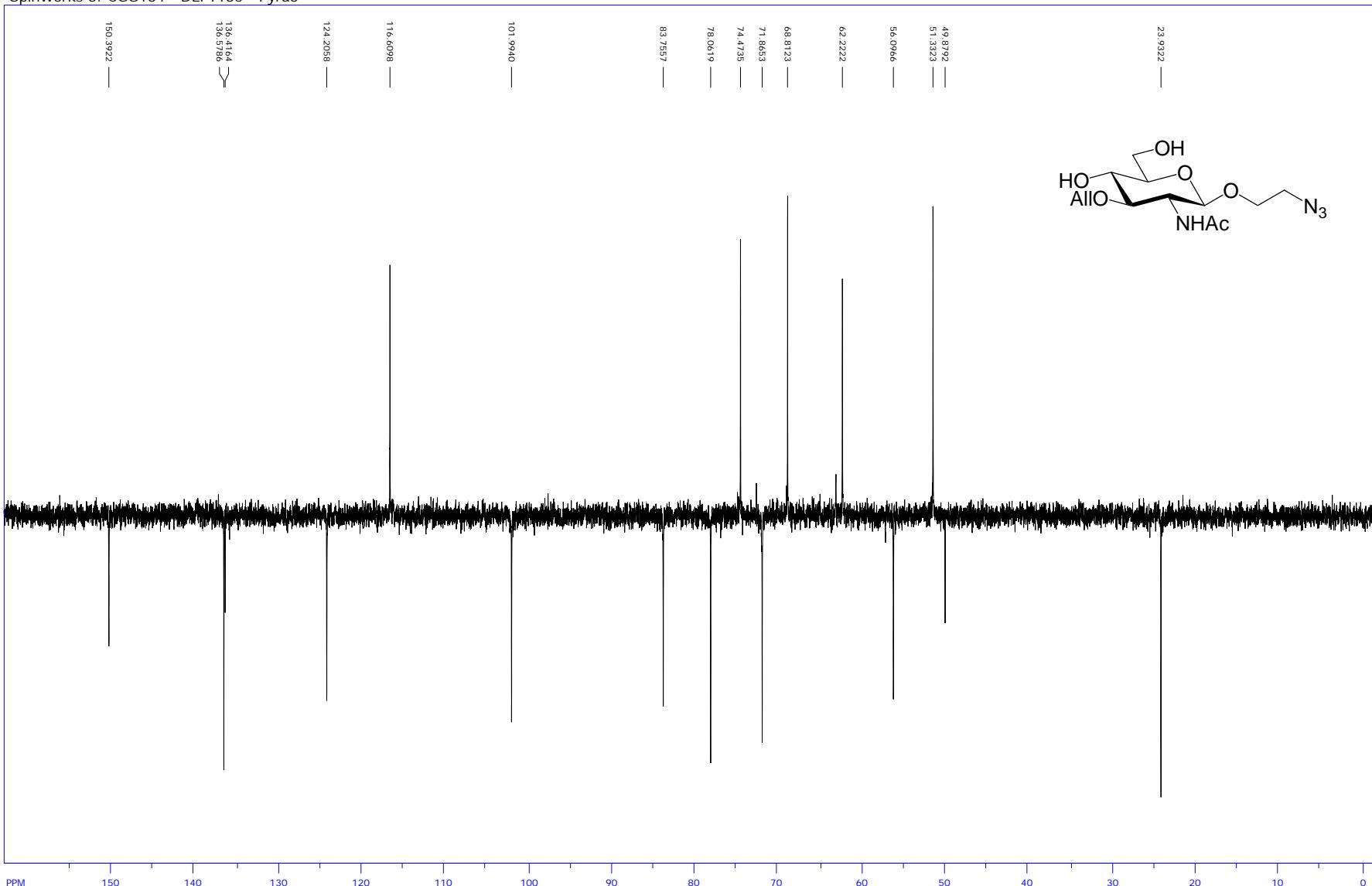


Compound S9

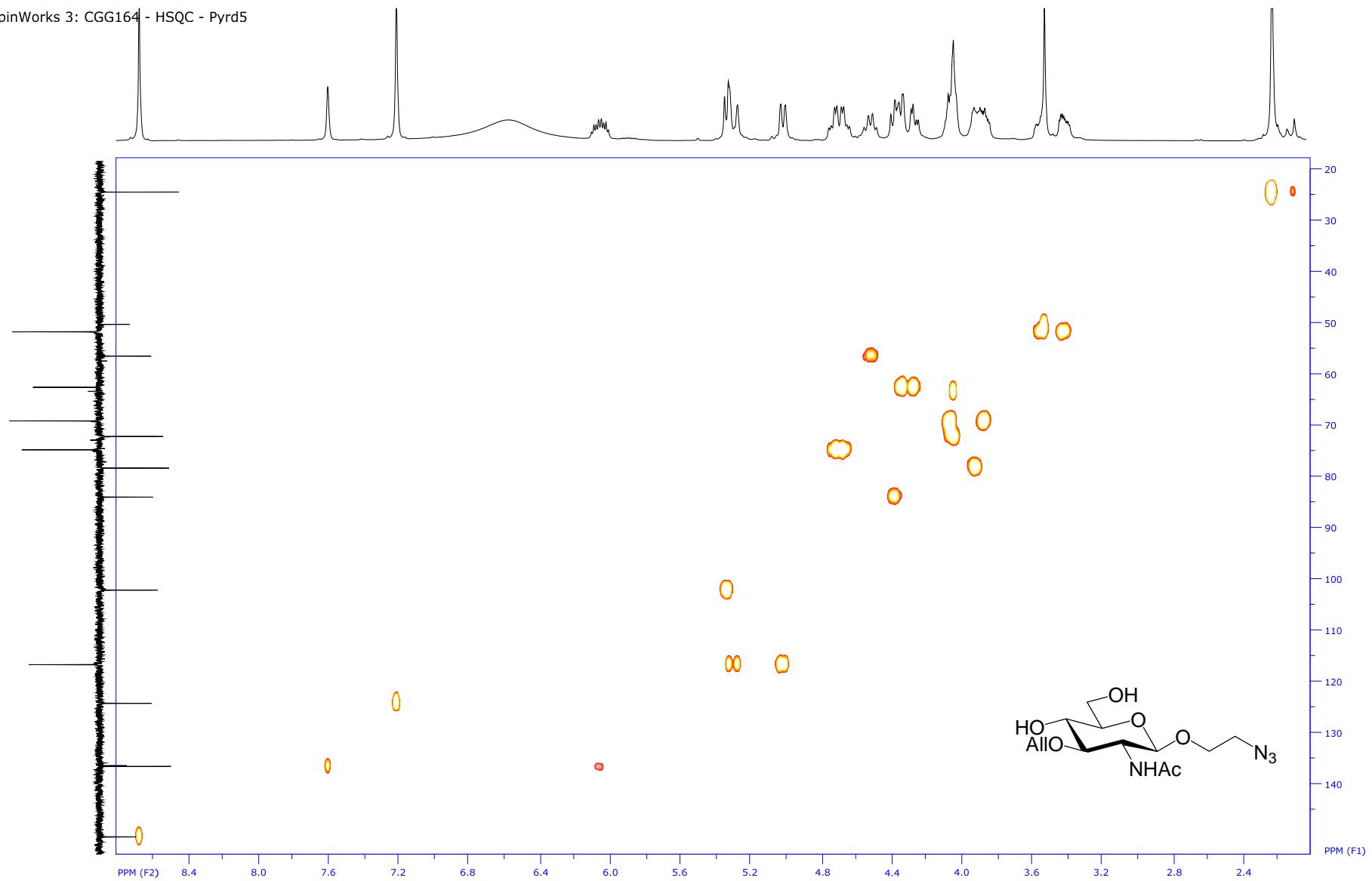
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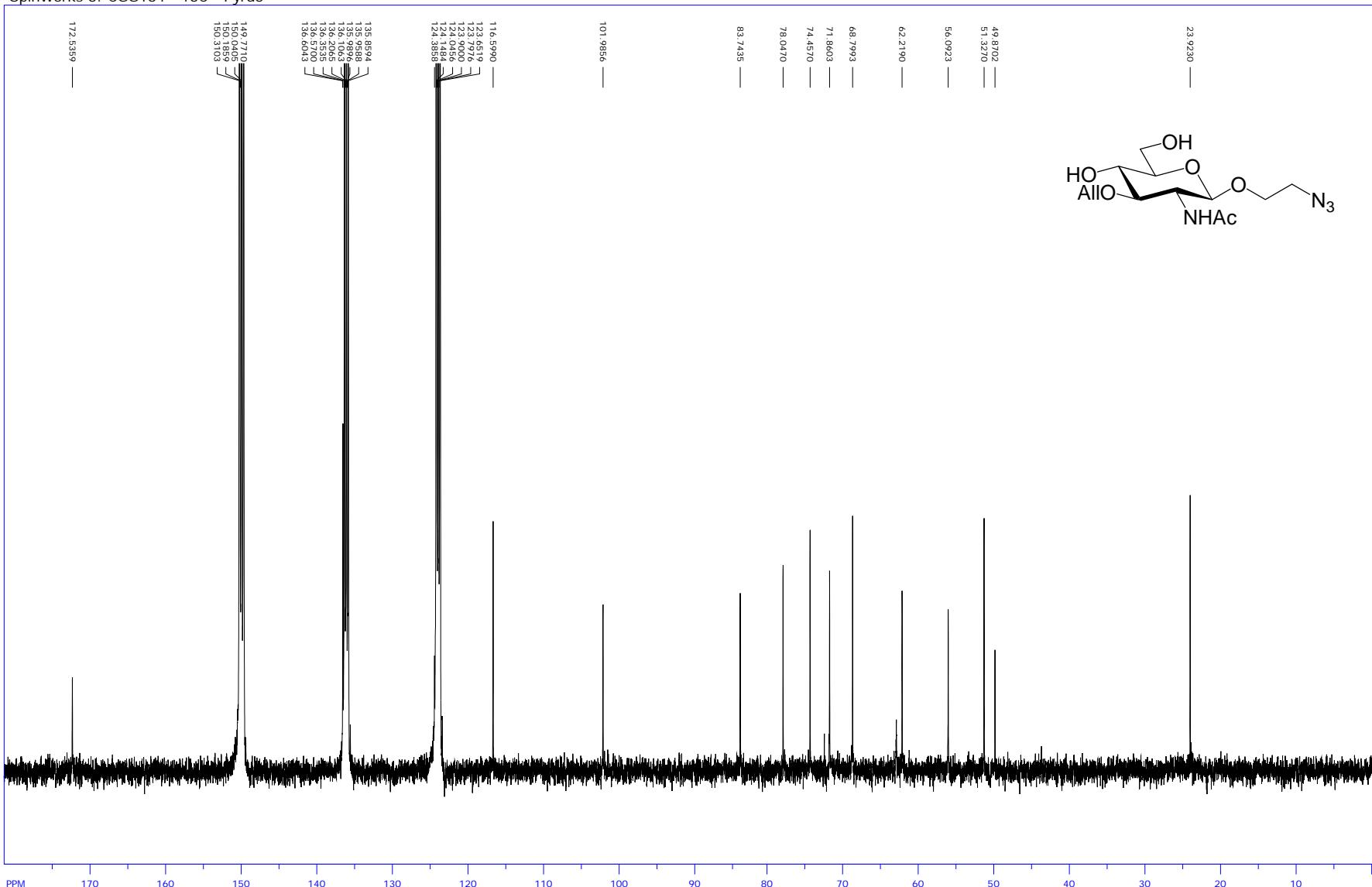
SpinWorks 3: CGG164 - DEPT135 - Pyrd5



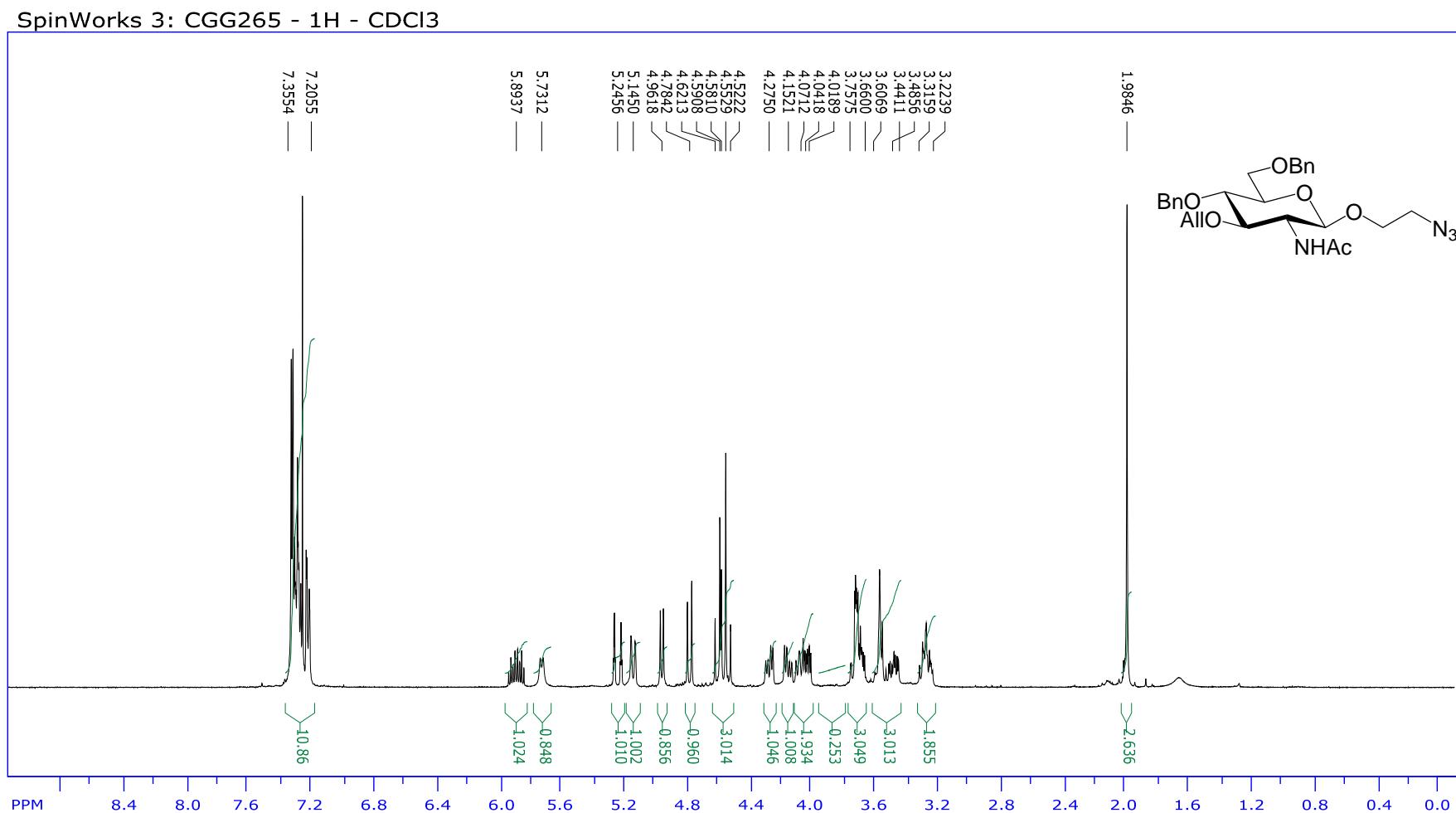
SpinWorks 3: CGG164 - HSQC - Pyrd5

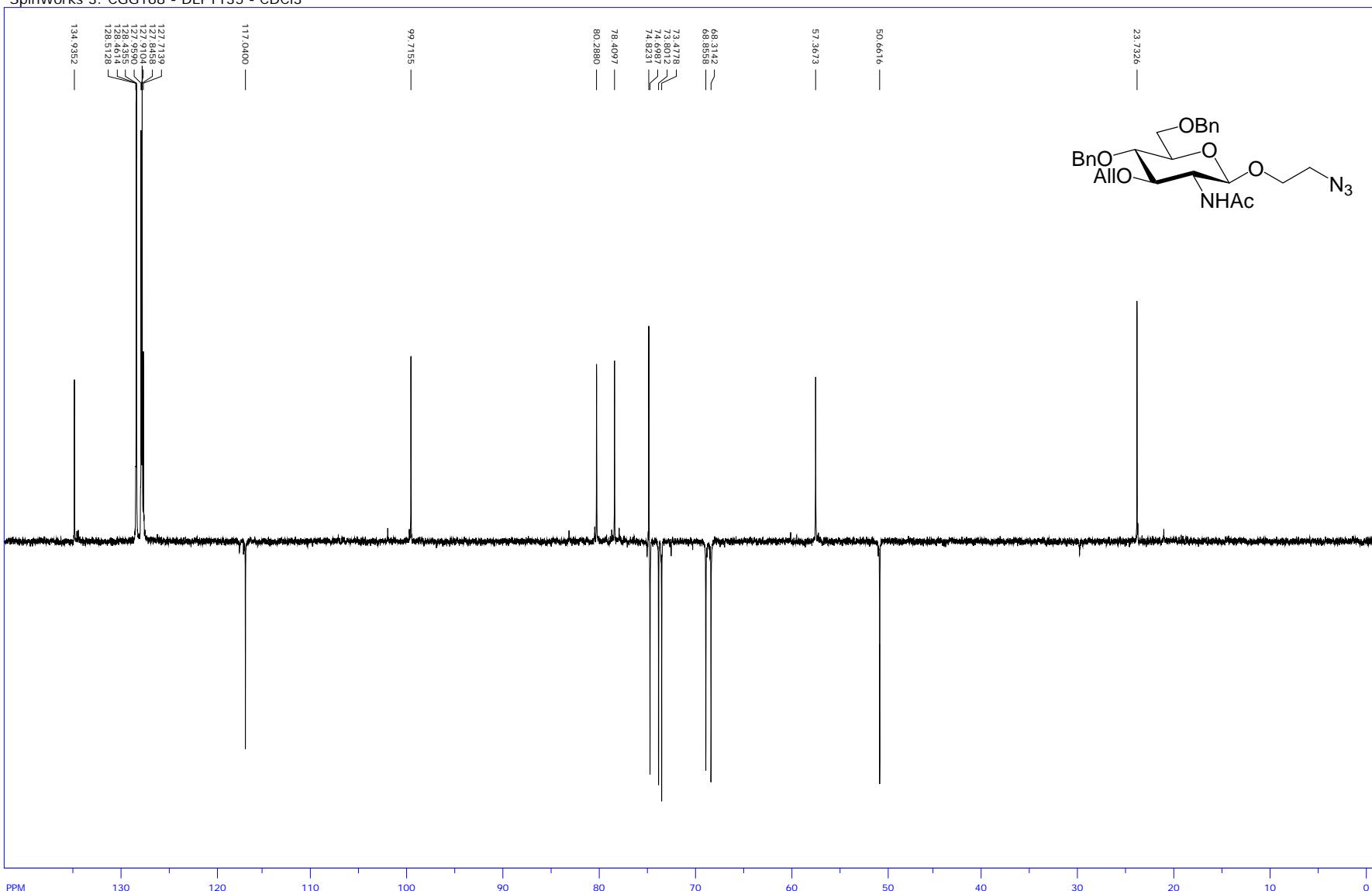


SpinWorks 3: CGG164 - 13C - Pyrd5

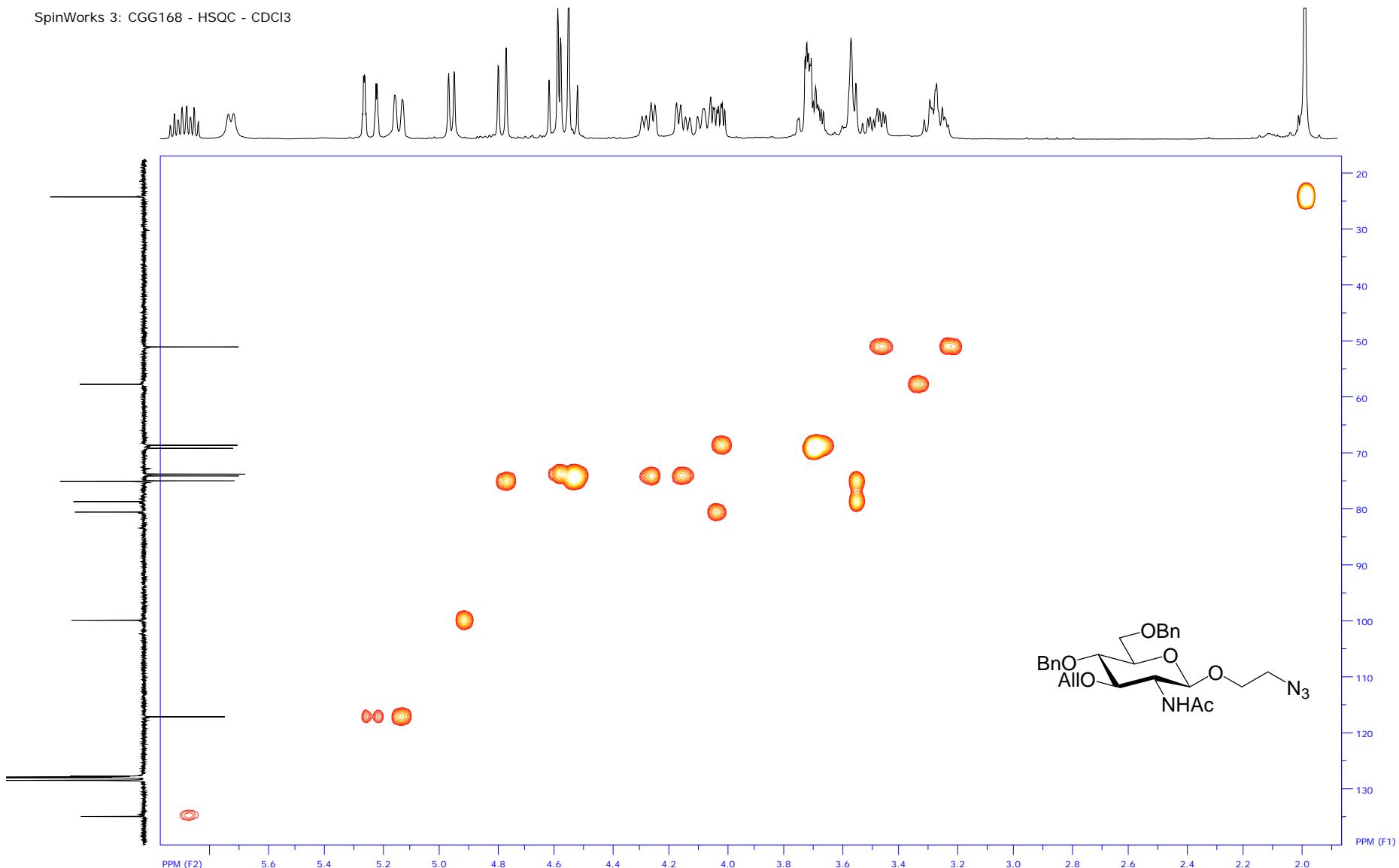


Compound 33

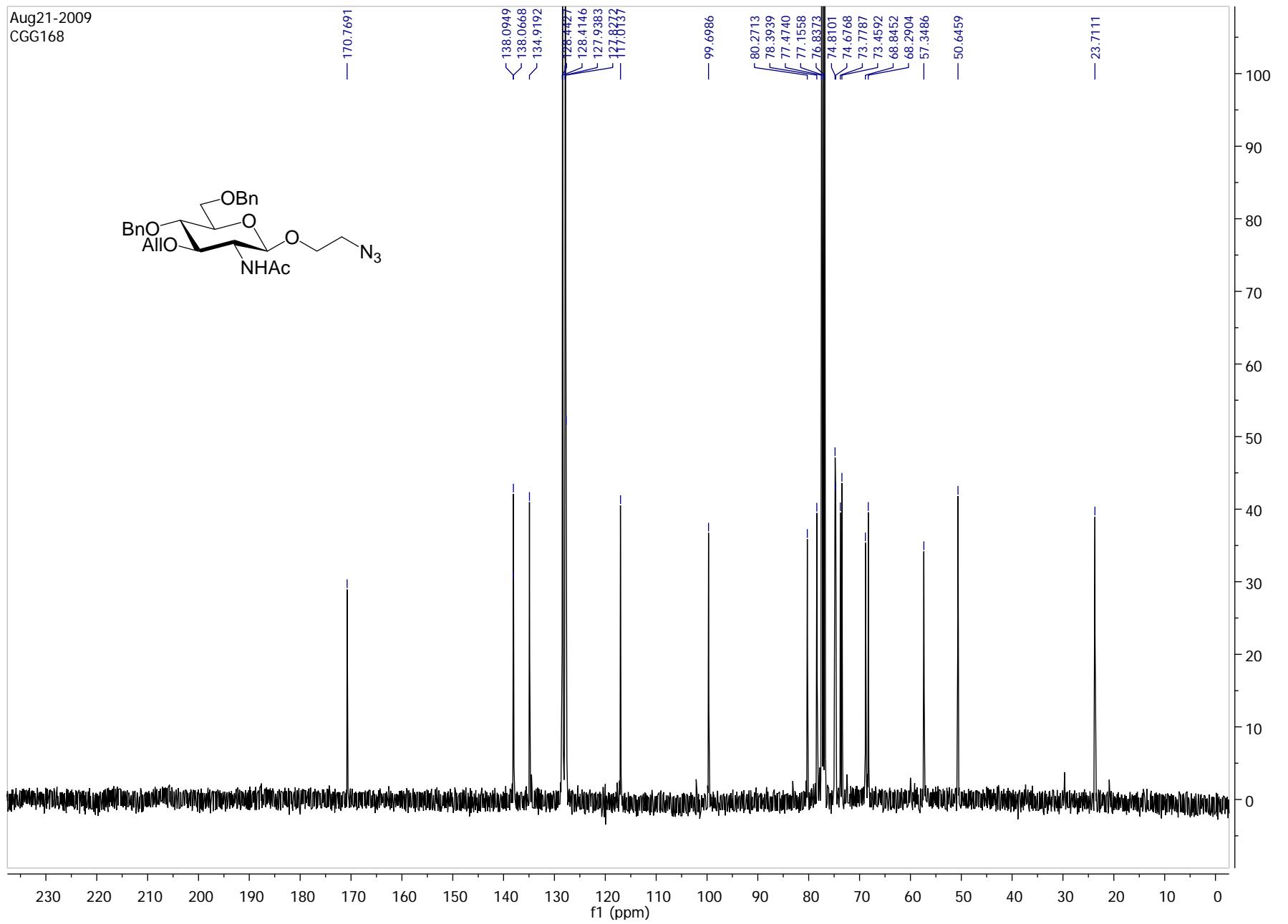
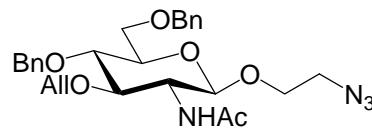


SpinWorks 3: CGG168 - DEPT135 - CDCl₃

SpinWorks 3: CGG168 - HSQC - CDCl3

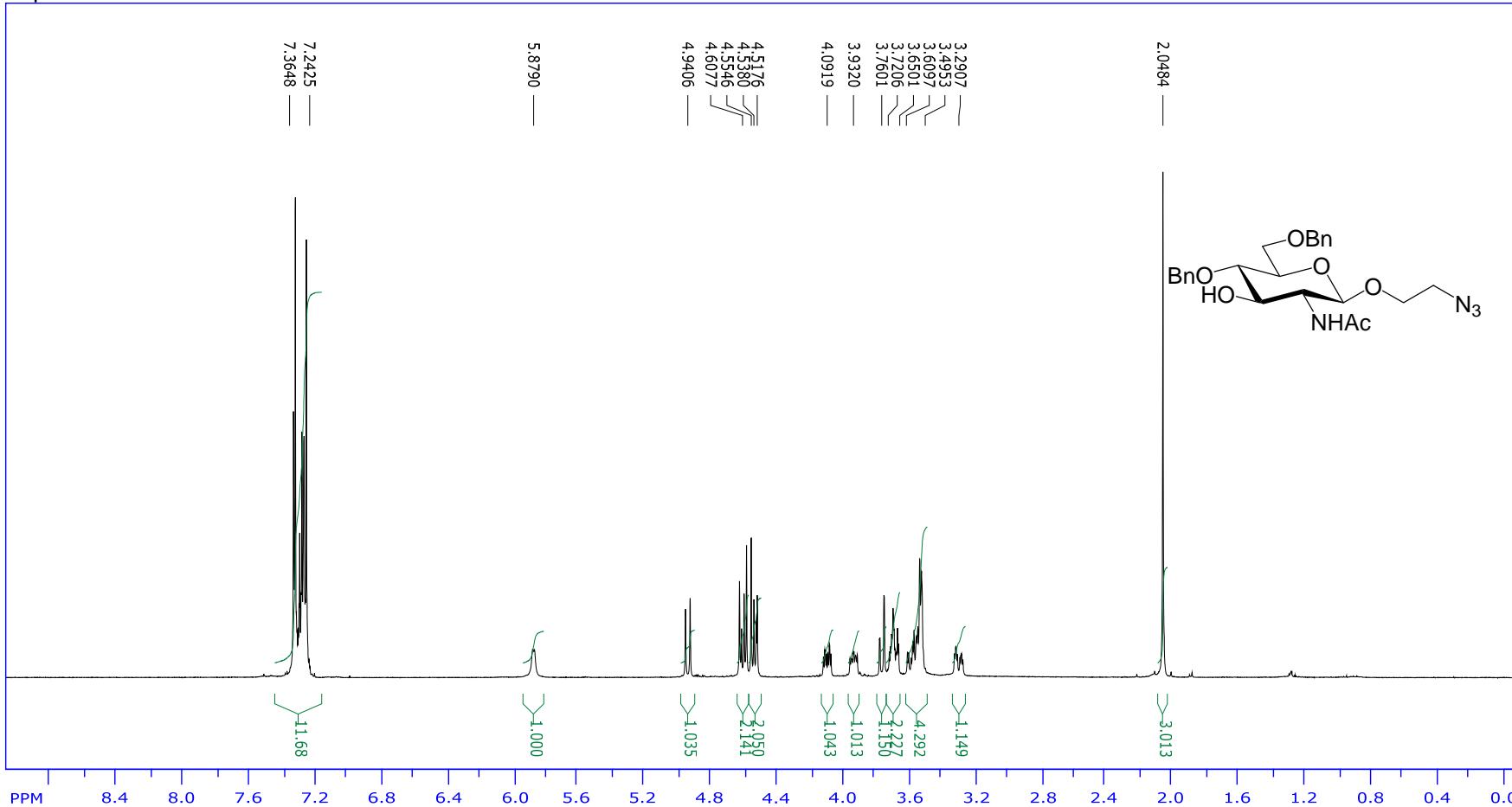


Aug21-2009
CGG168

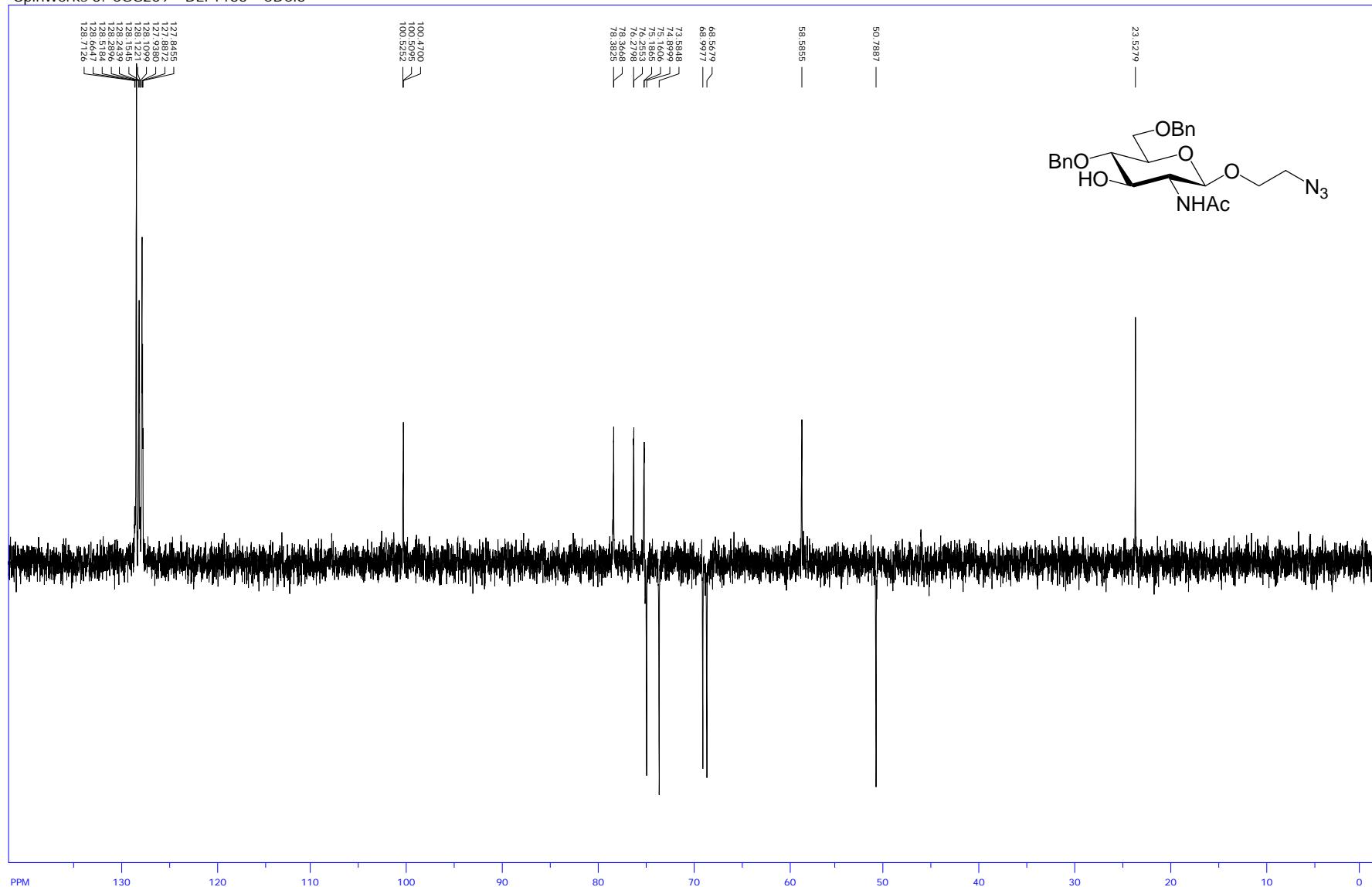


Compound 8

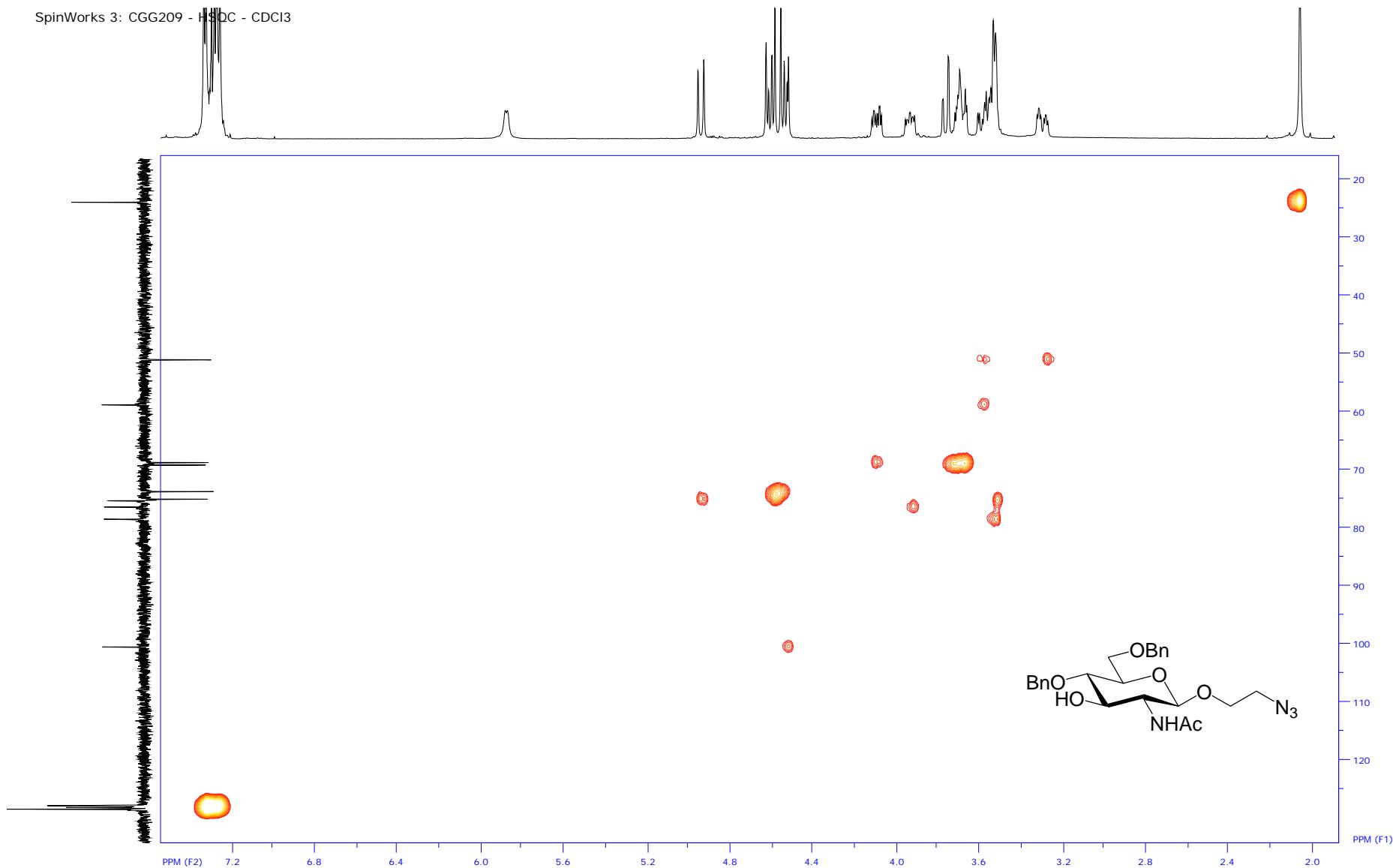
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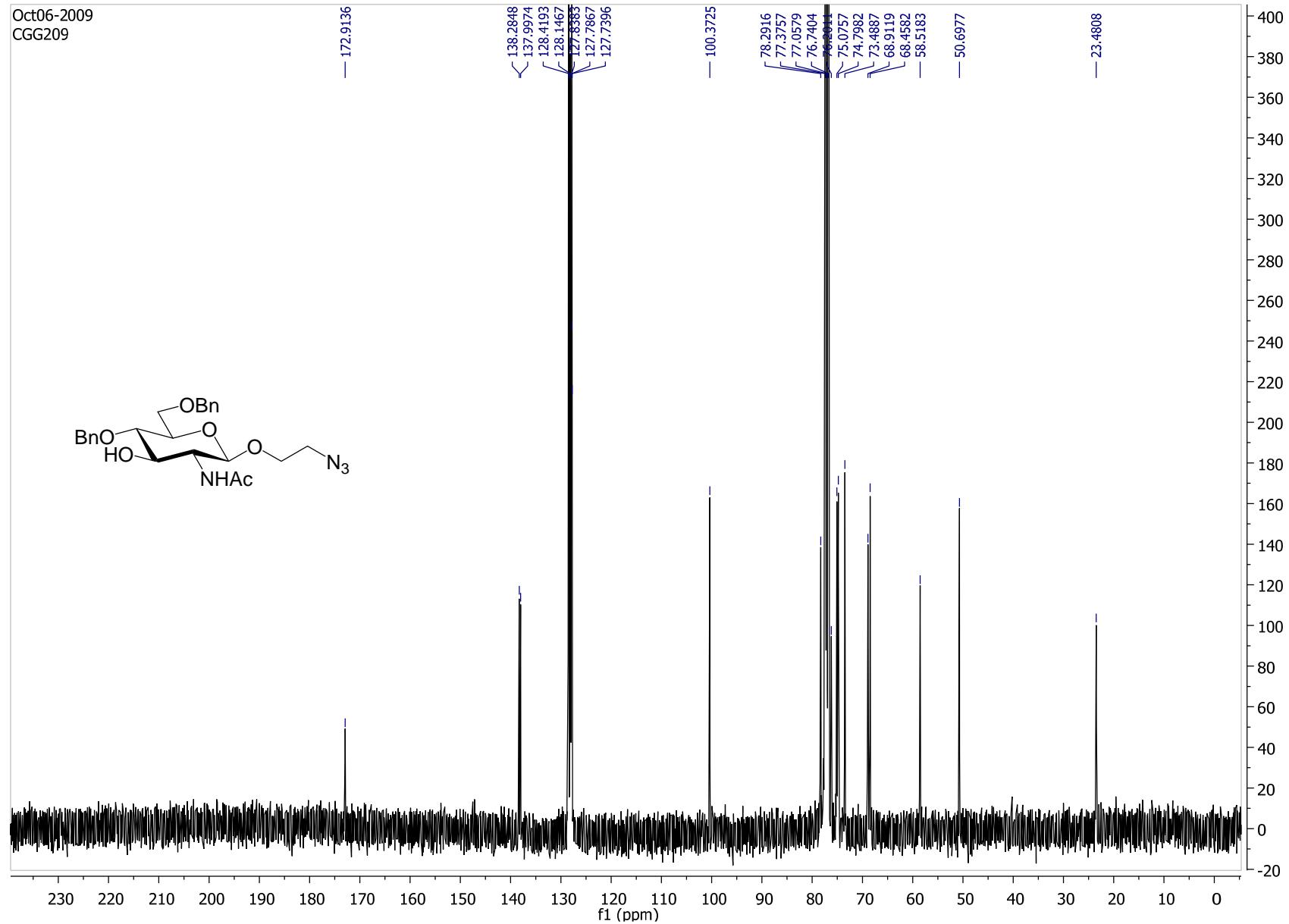


SpinWorks 3: CGG209 - DEPT135 - CDCl₃



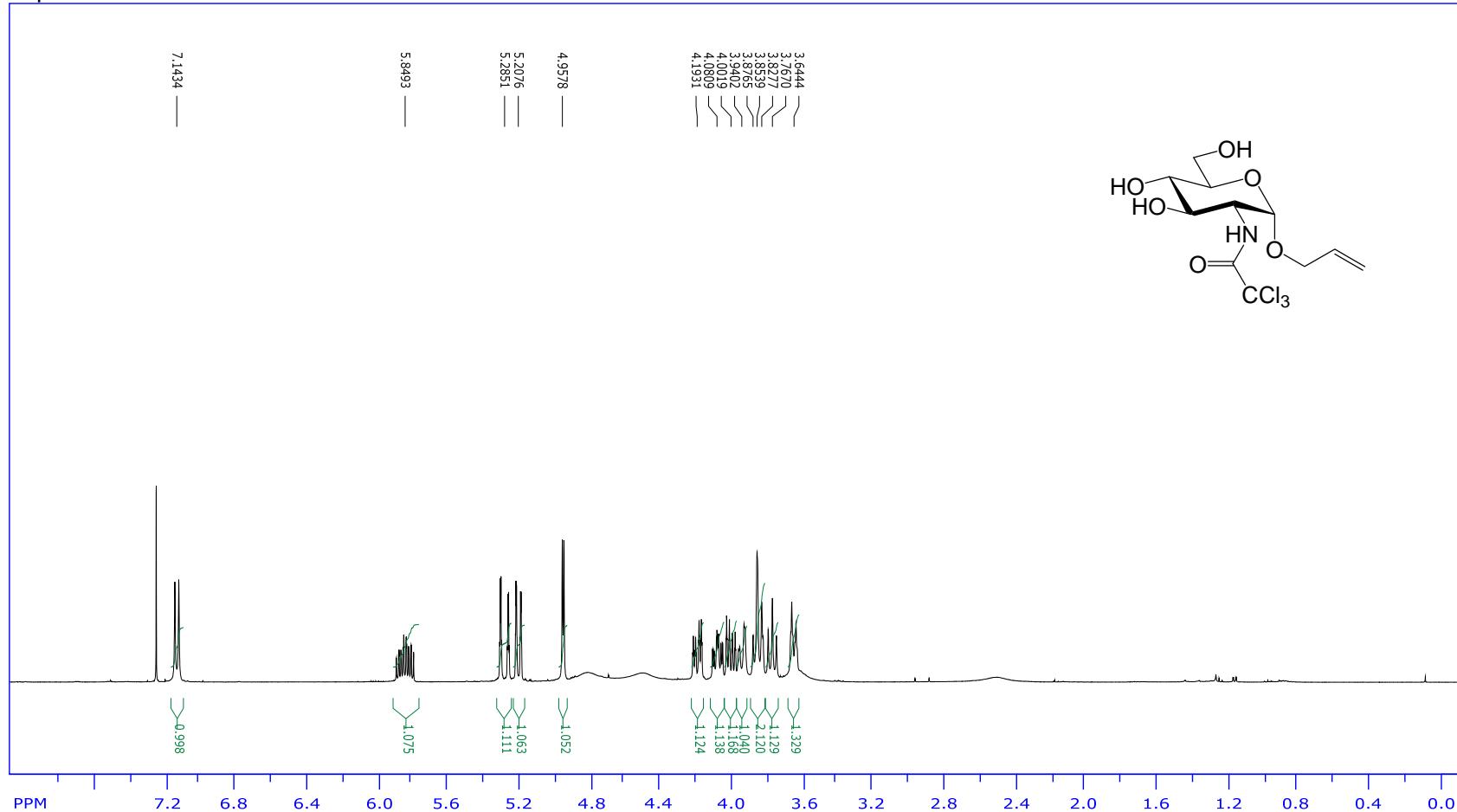
SpinWorks 3: CGG209 - HSQC - CDCl₃



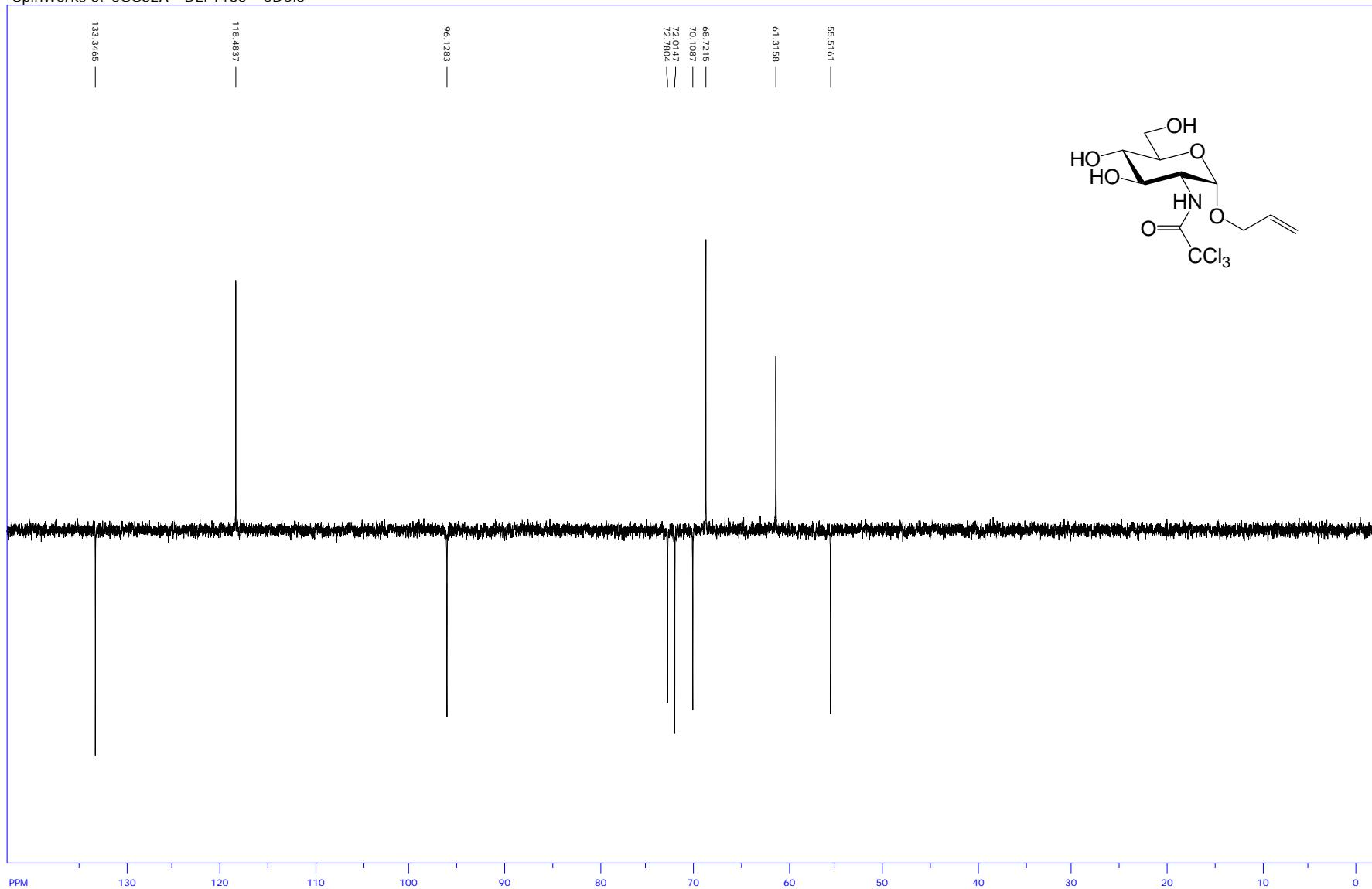


Compound 25

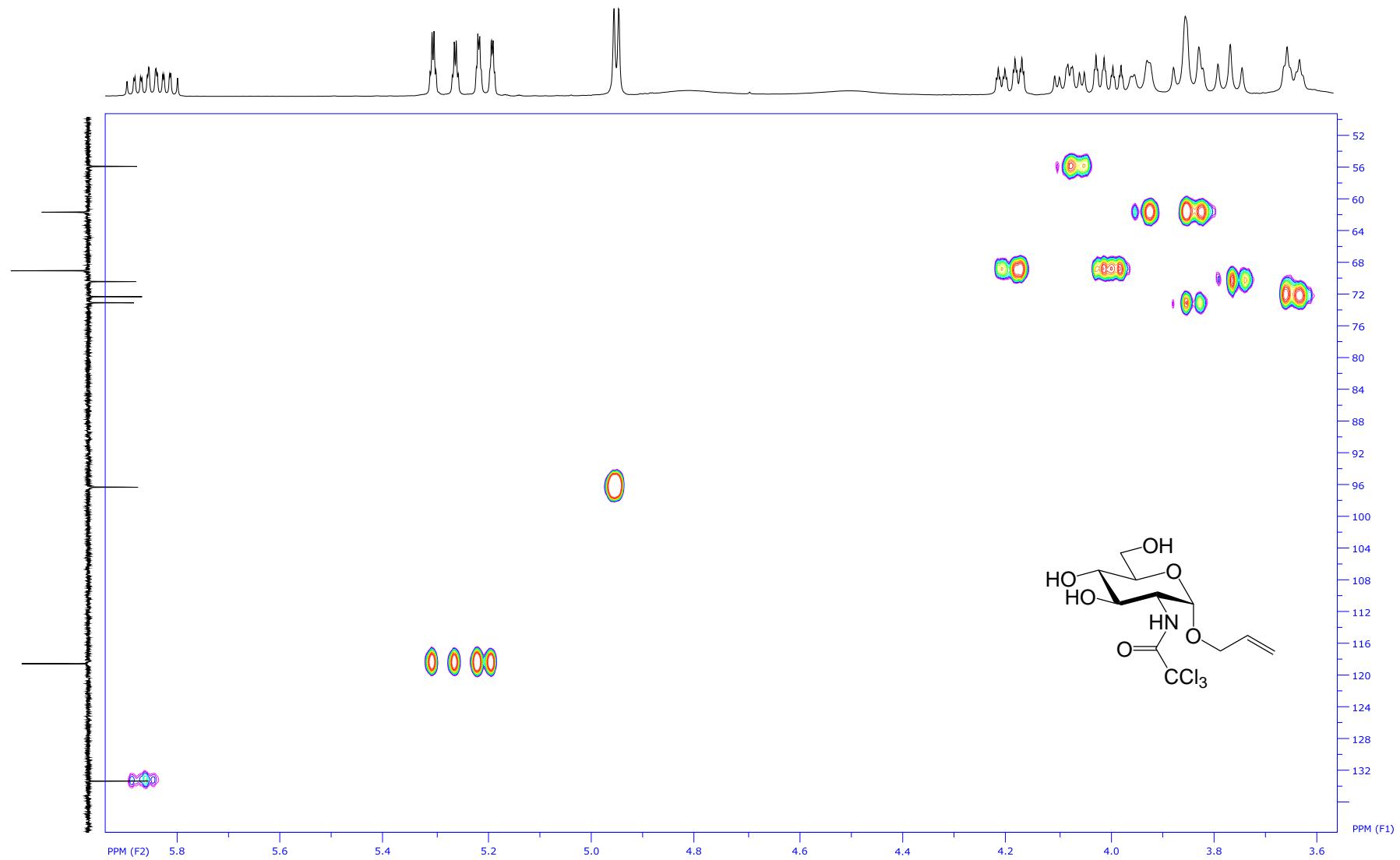
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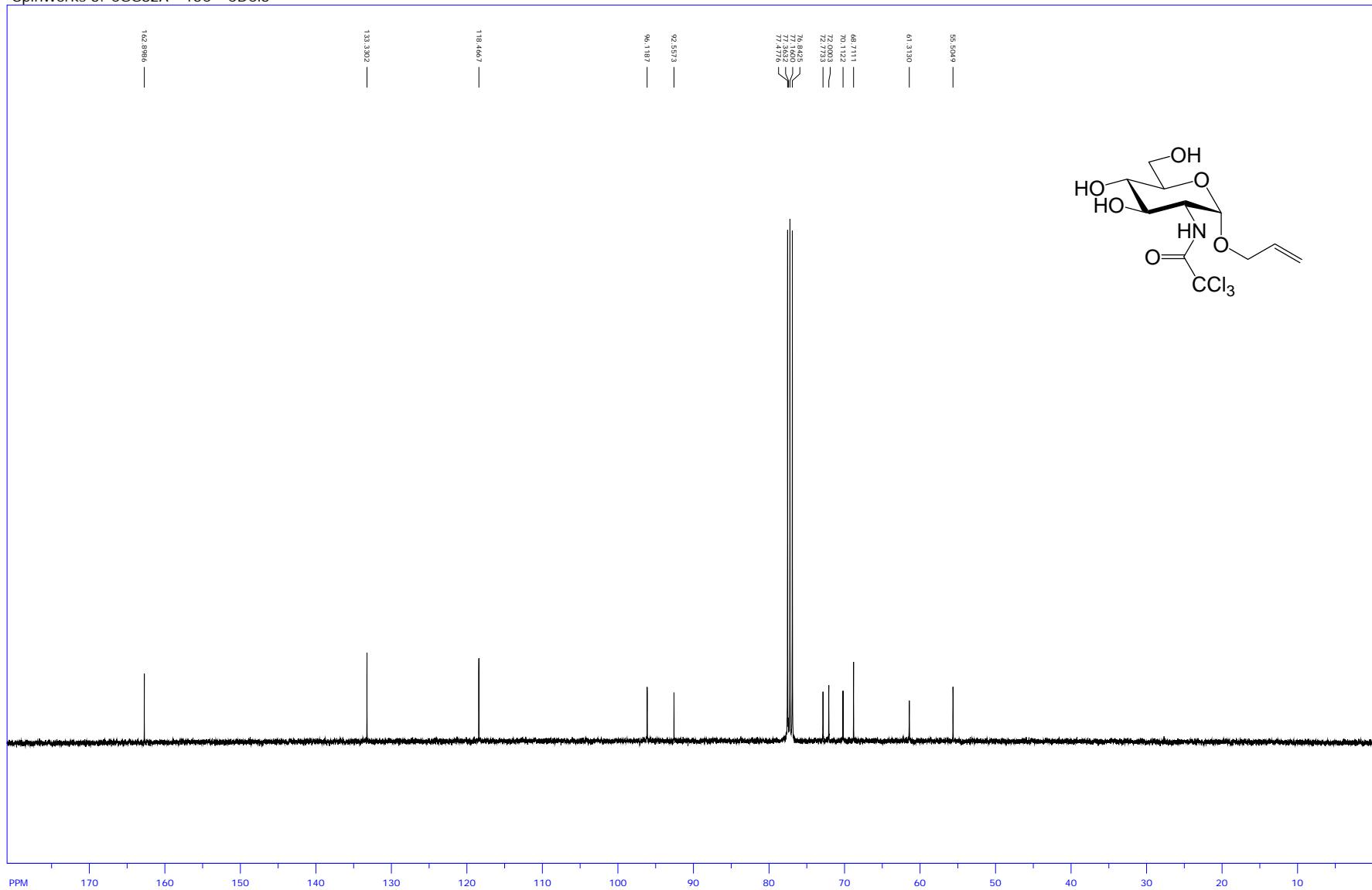
SpinWorks 3: CGG82A - DEPT135 - CDCl₃



SpinWorks 3: CGG82A - HSQC - CDCl3

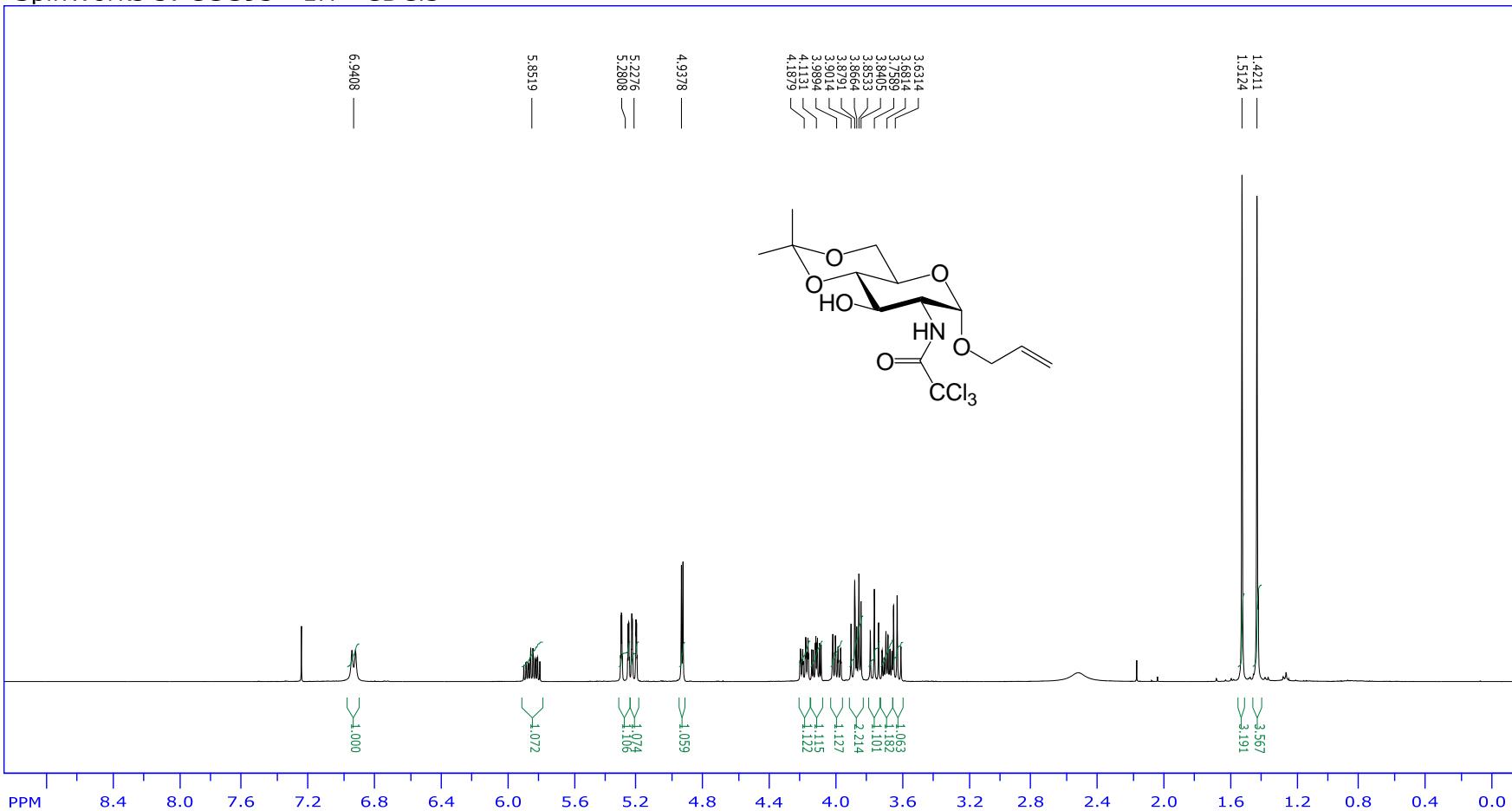


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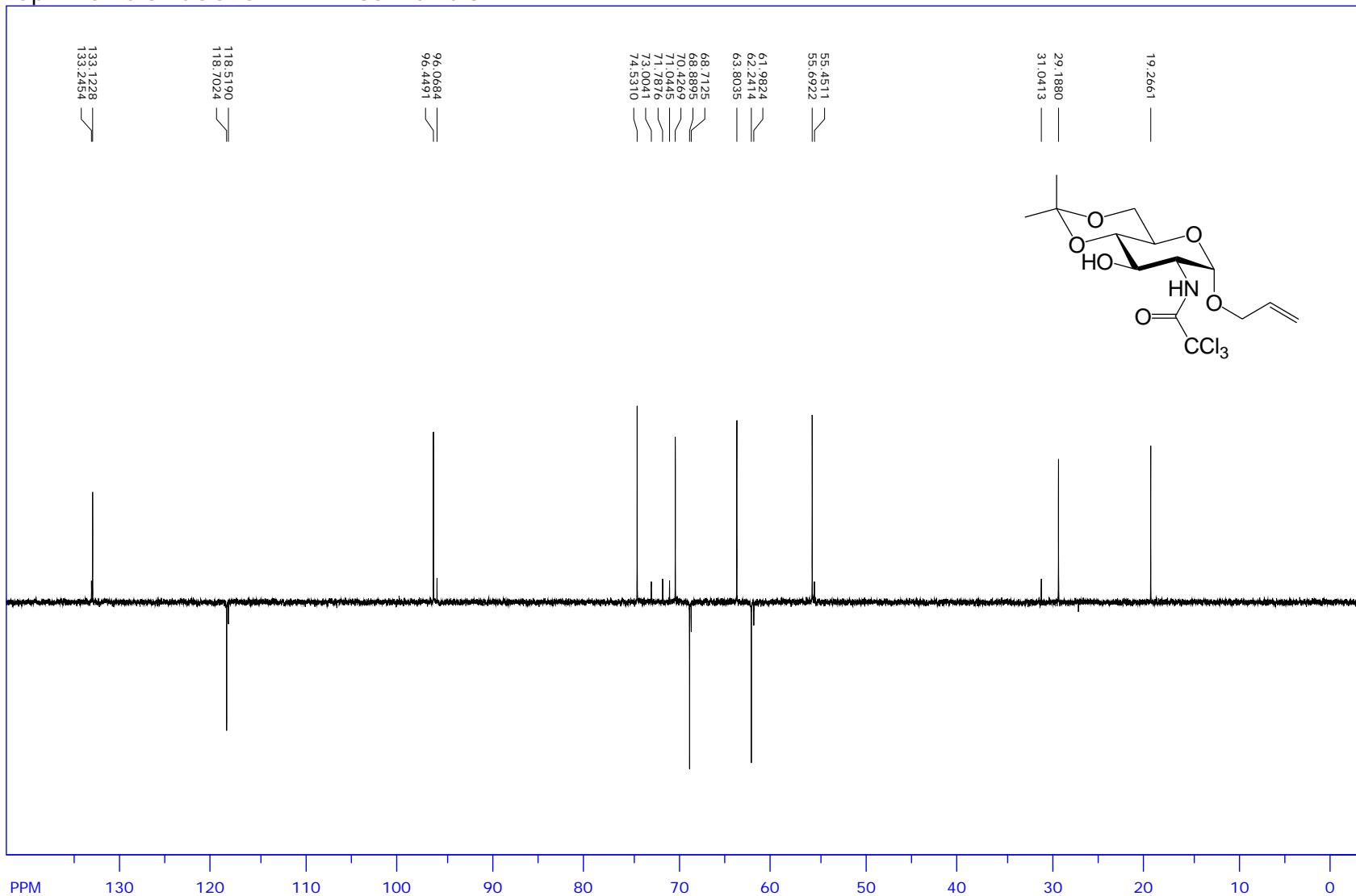


Compound S6

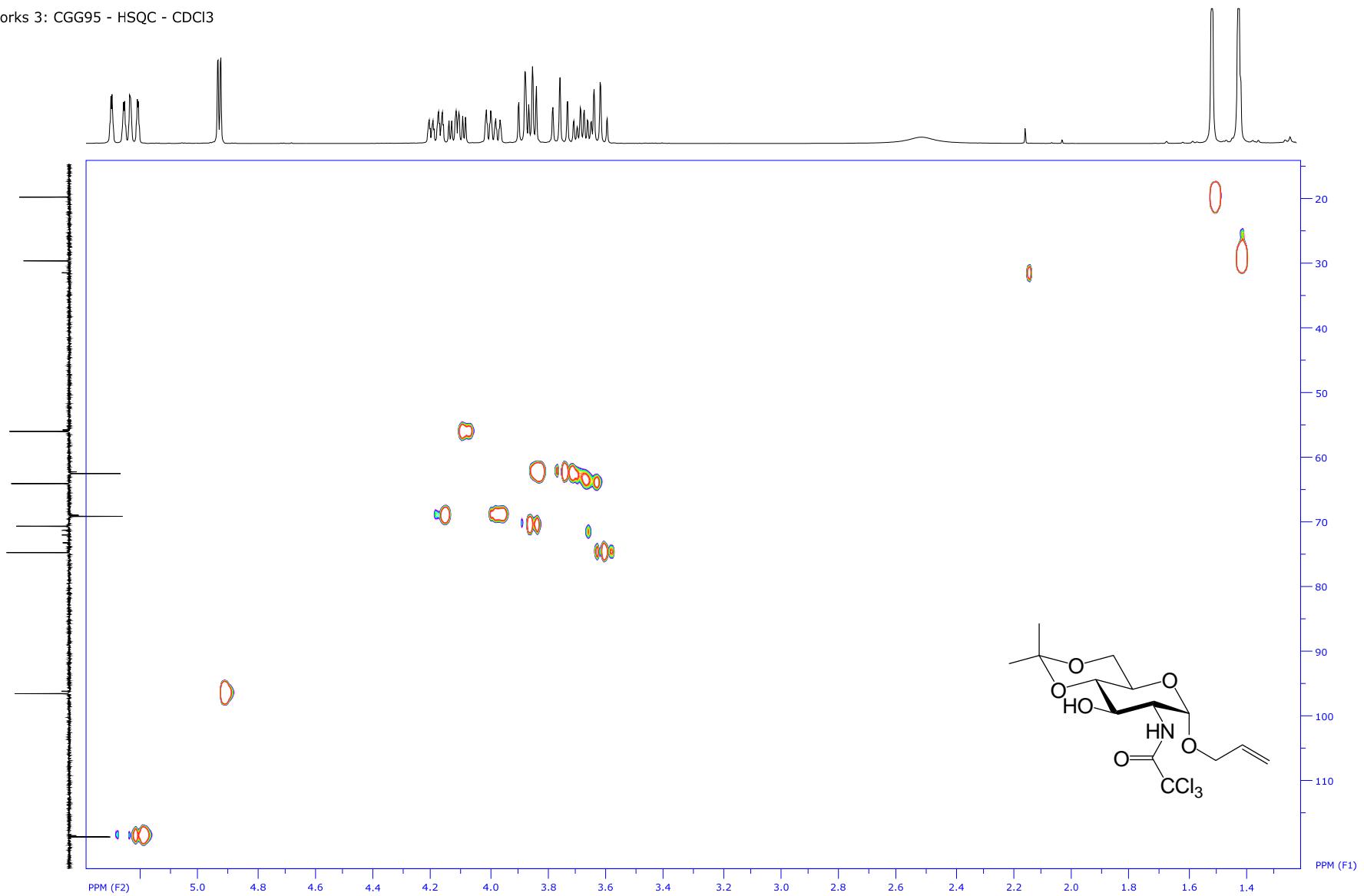
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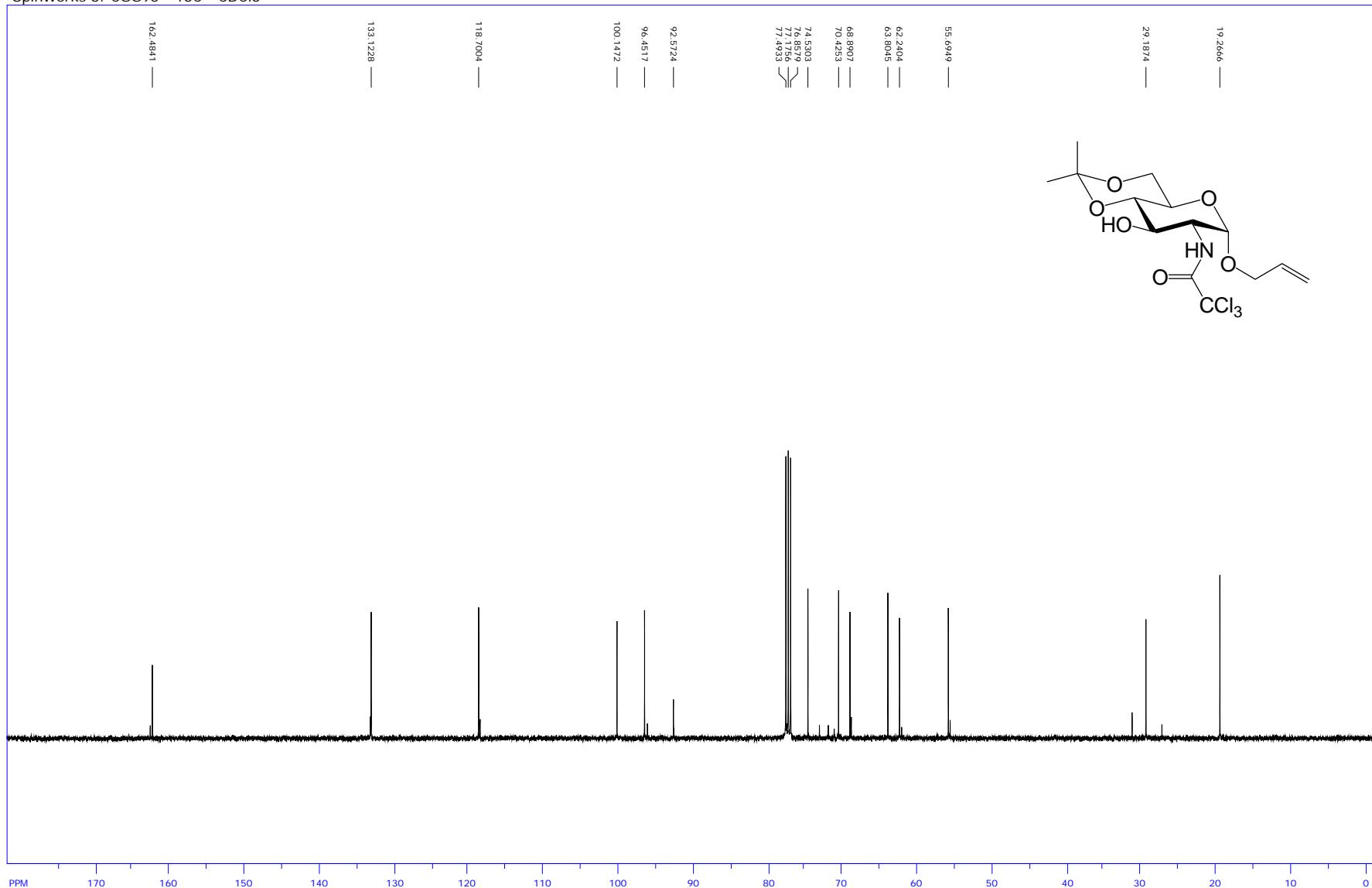
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SpinWorks 3: CGG95 - HSQC - CDCl₃

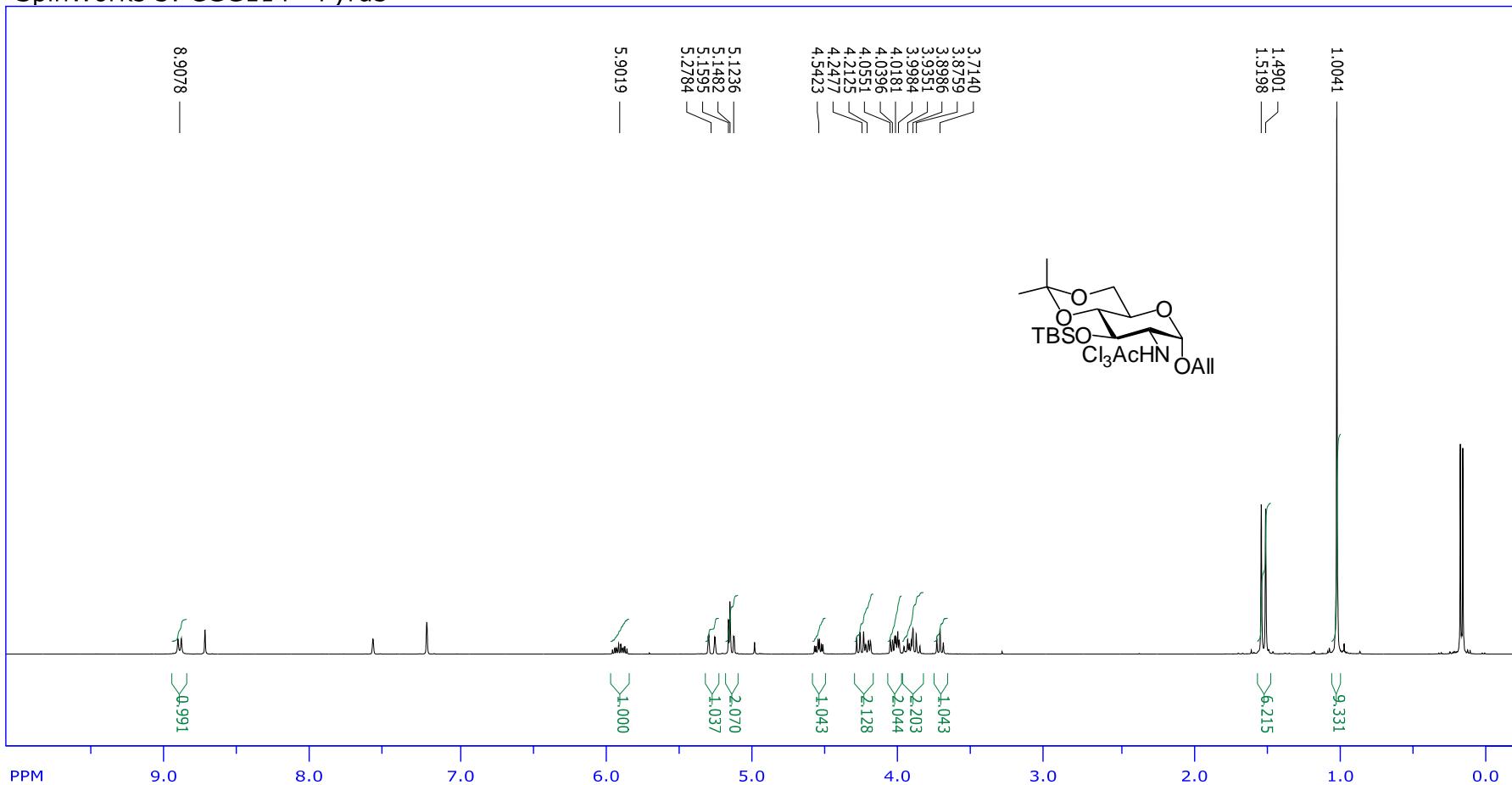


SpinWorks 3: CGG95 - 13C - CDCl₃

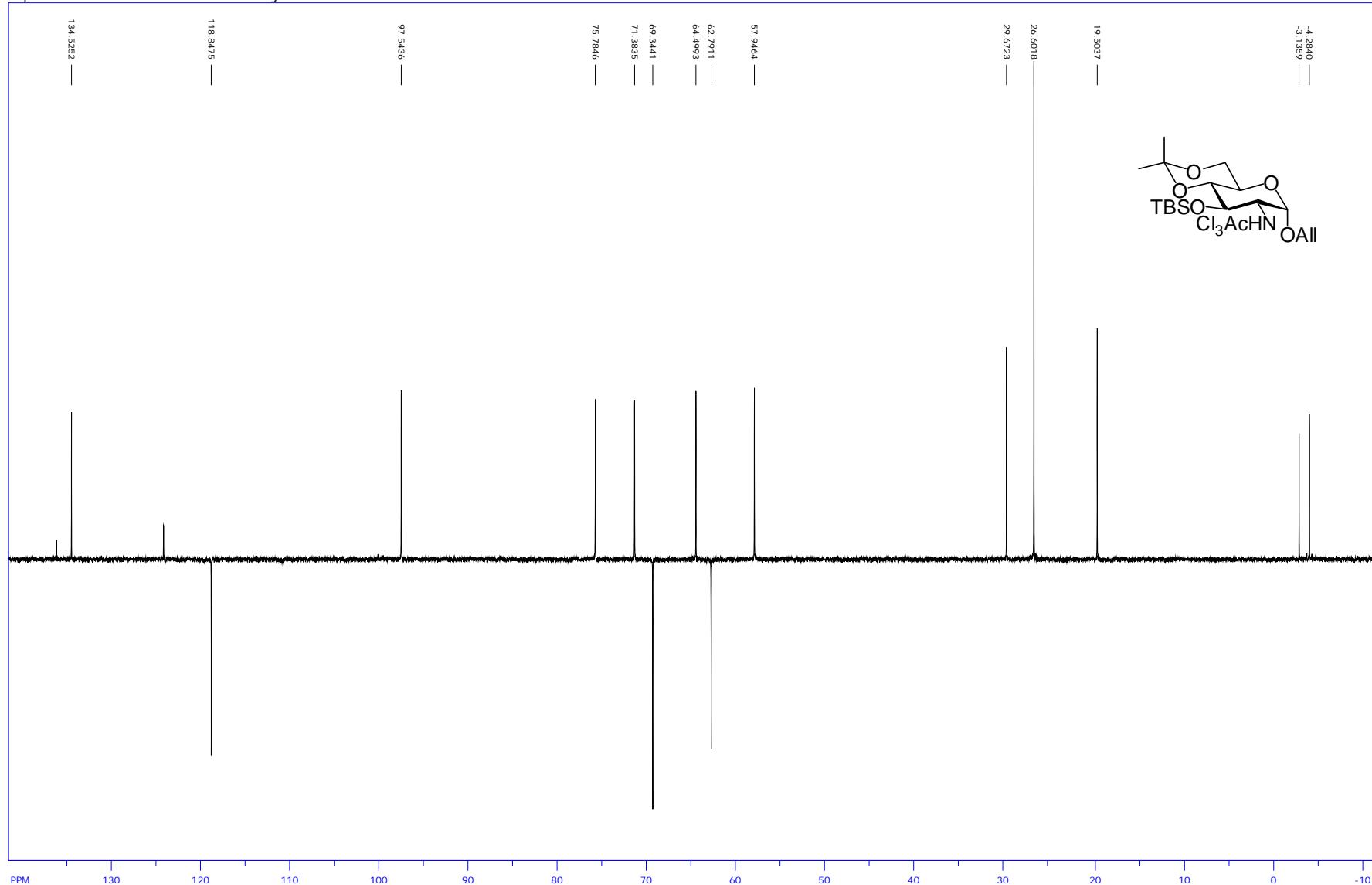


Compound 26

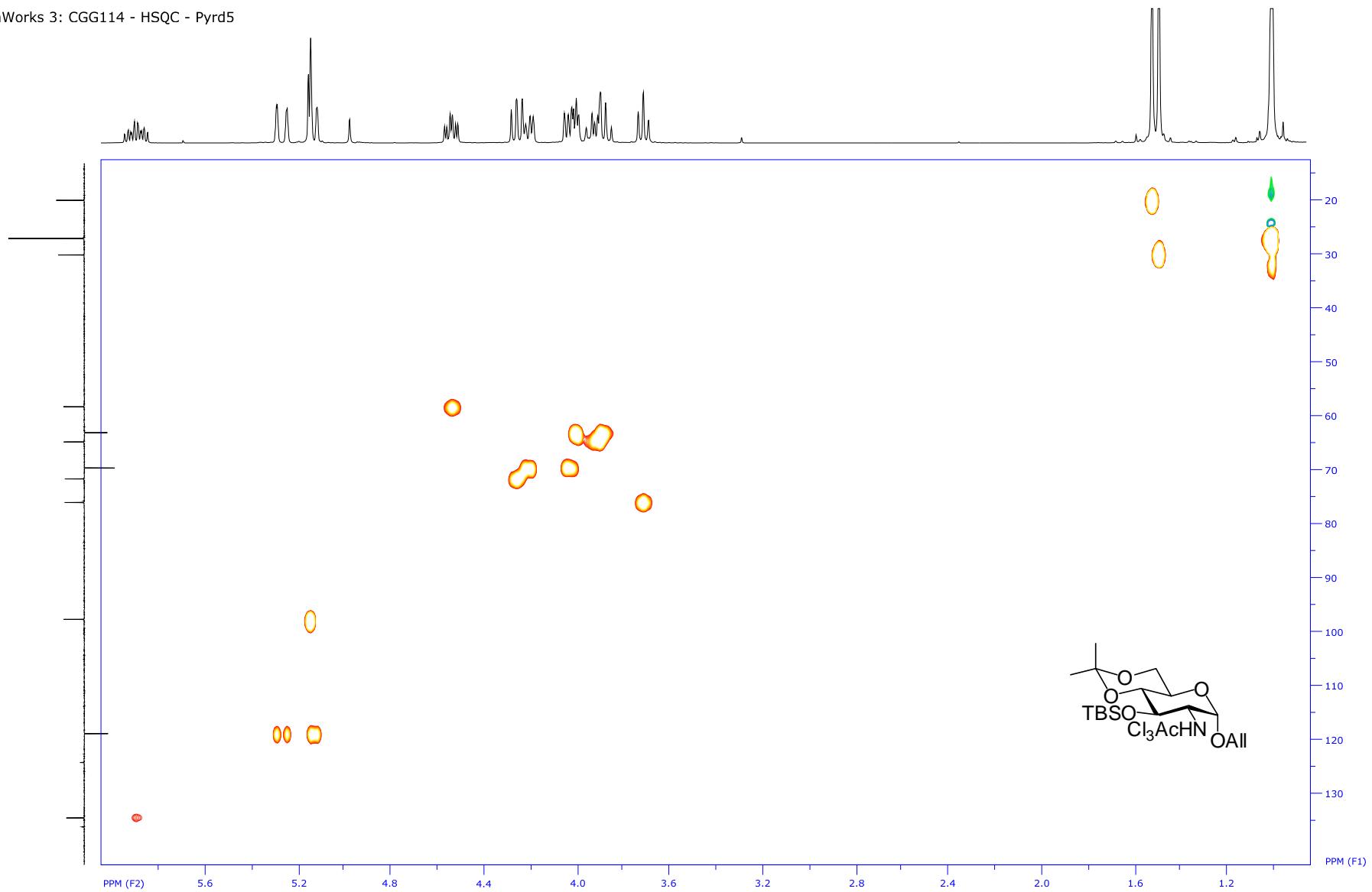
SpinWorks 3: CGG114 - Pyrd5



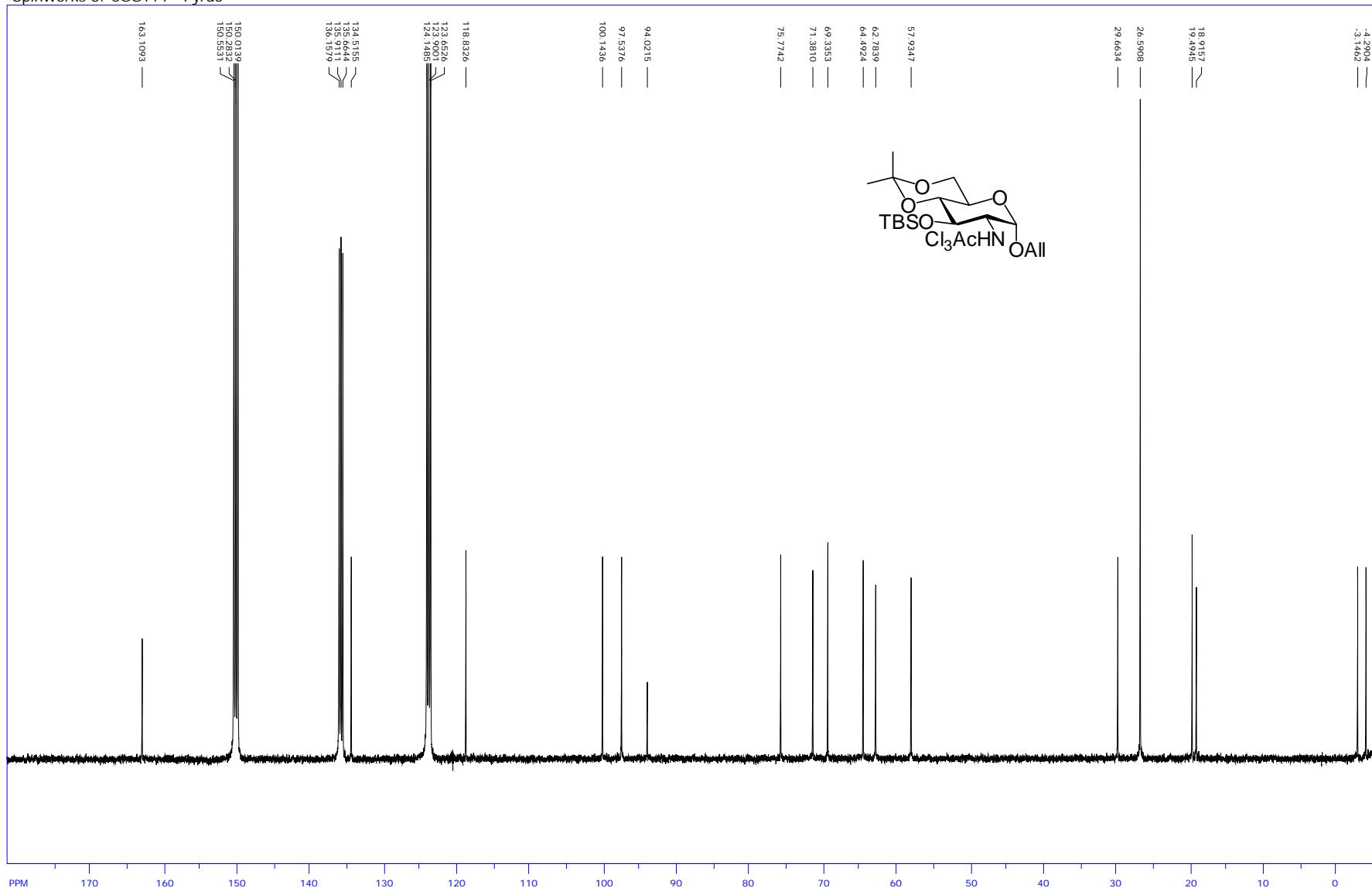
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SpinWorks 3: CGG114 - HSQC - Pyrd5



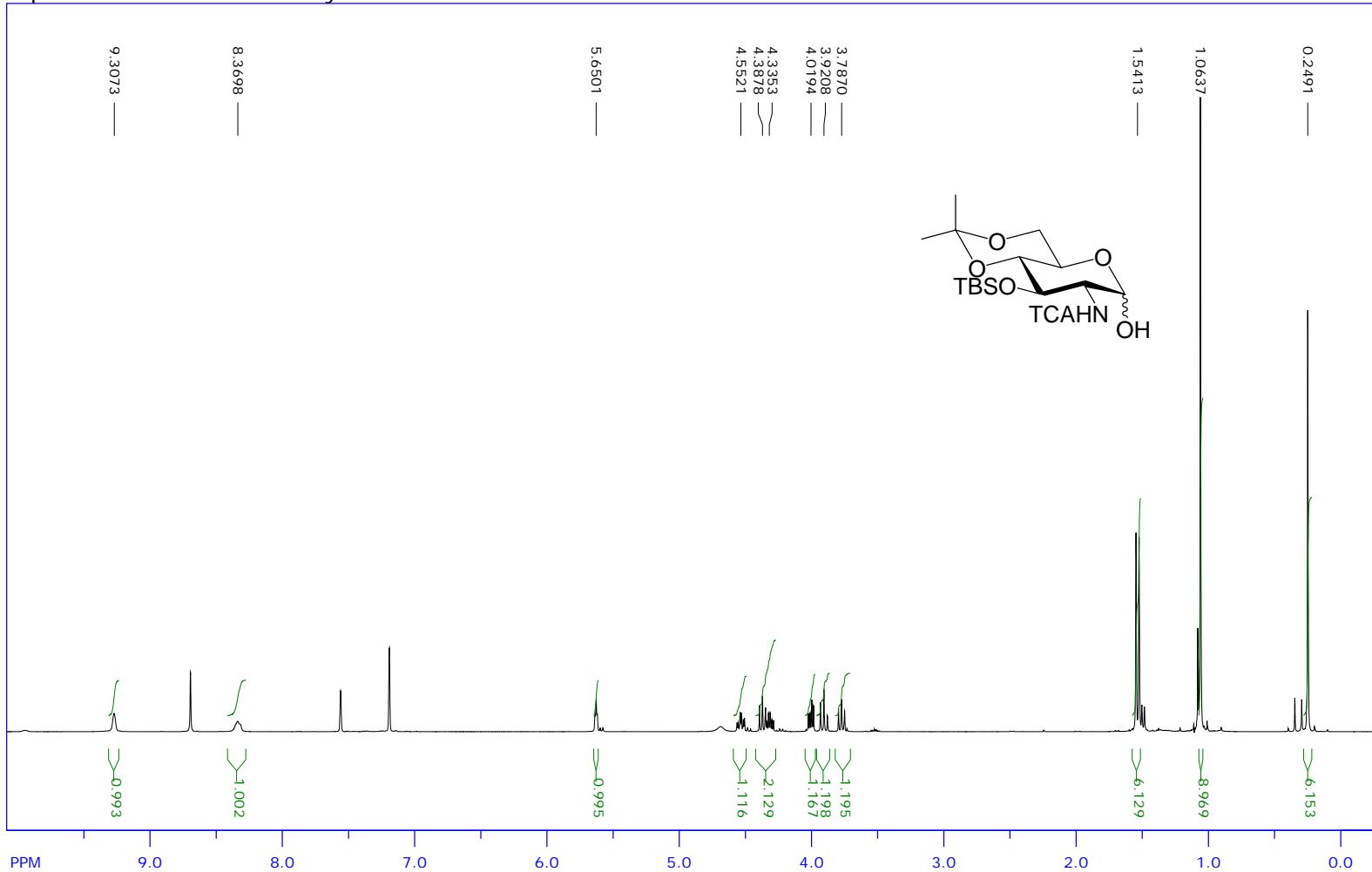
SpinWorks 3: CGG114 - Pyrd5



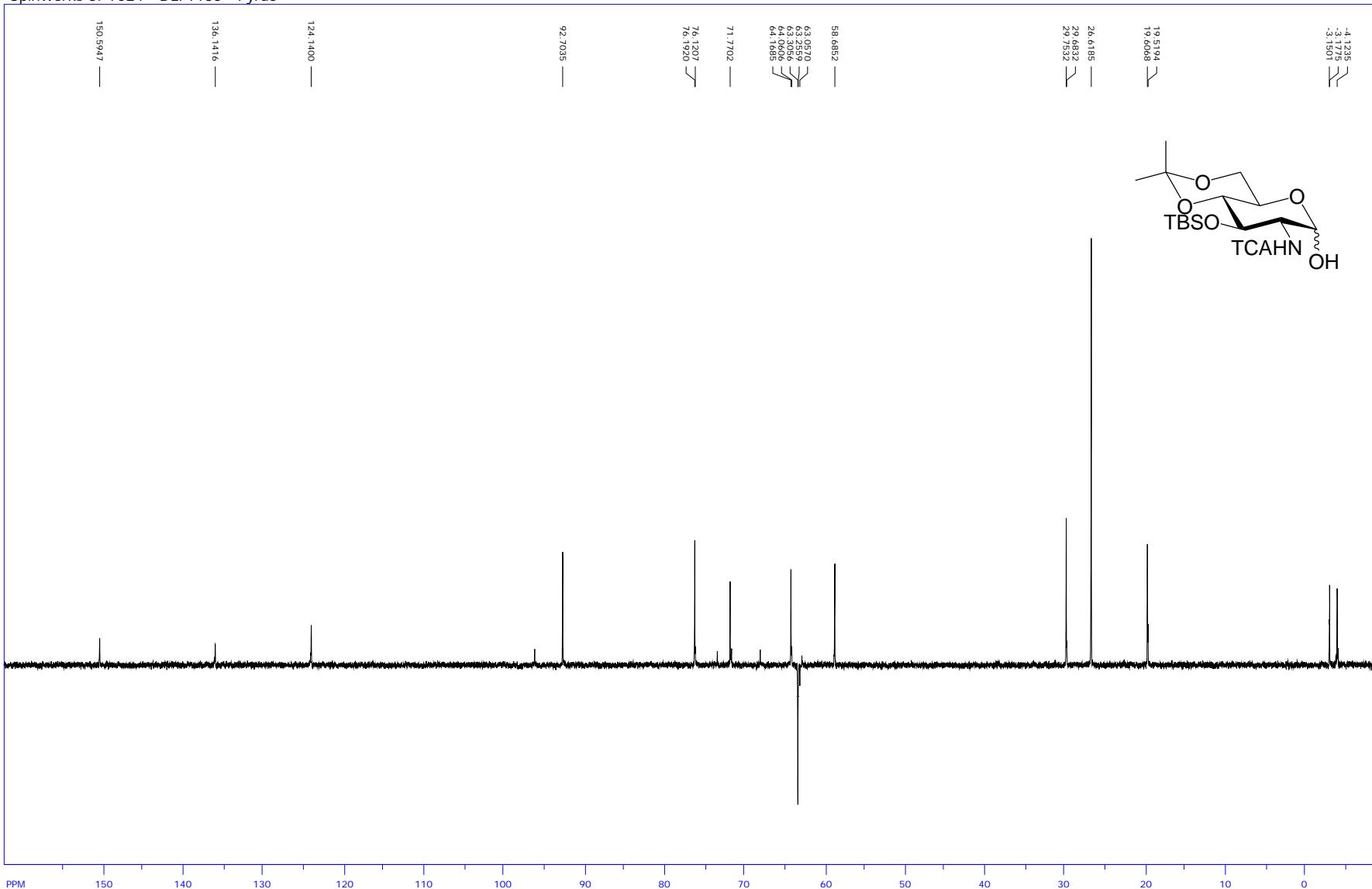
S90

Compound S7

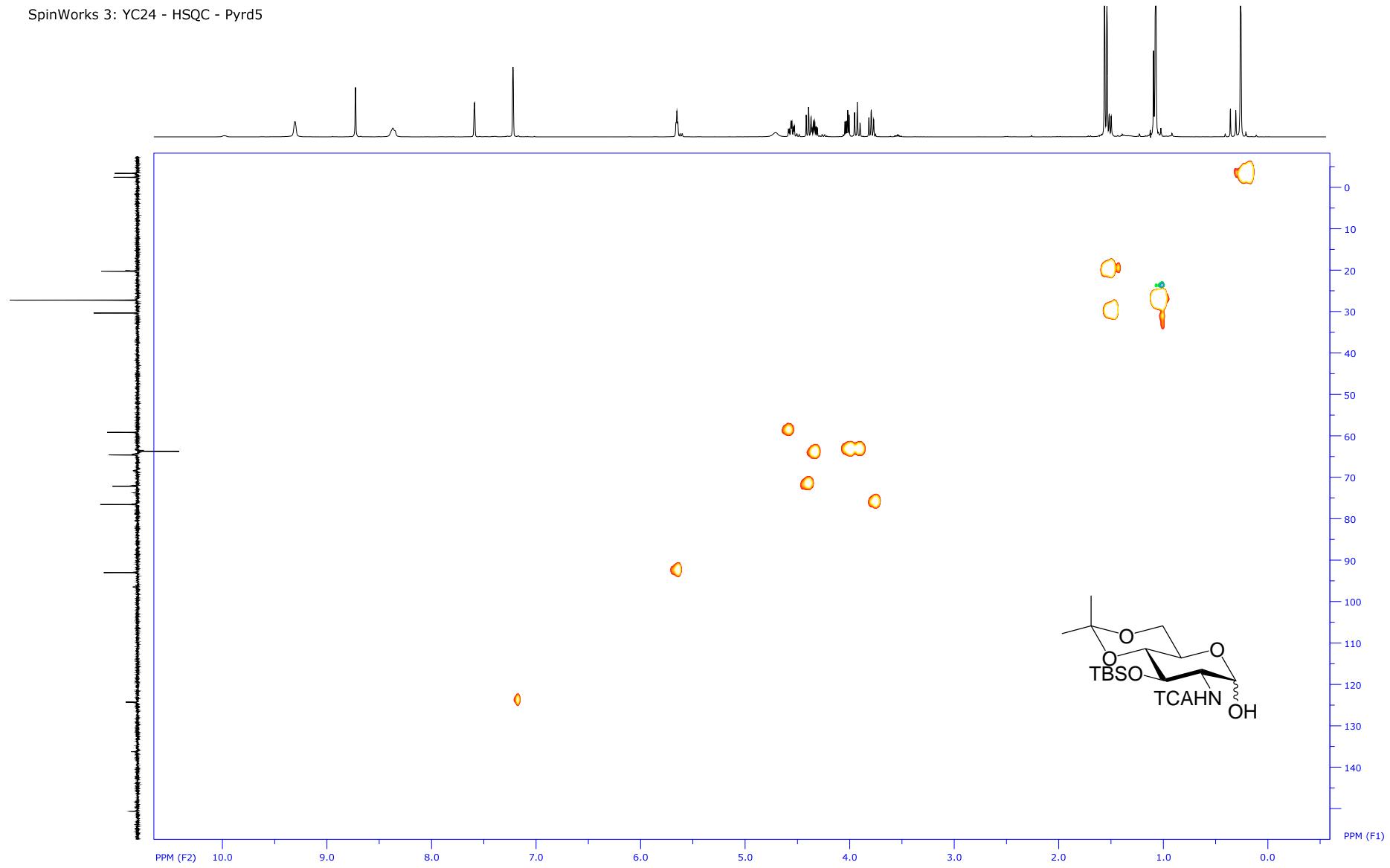
SpinWorks 3: YC24 - 1H - Pyr-d5



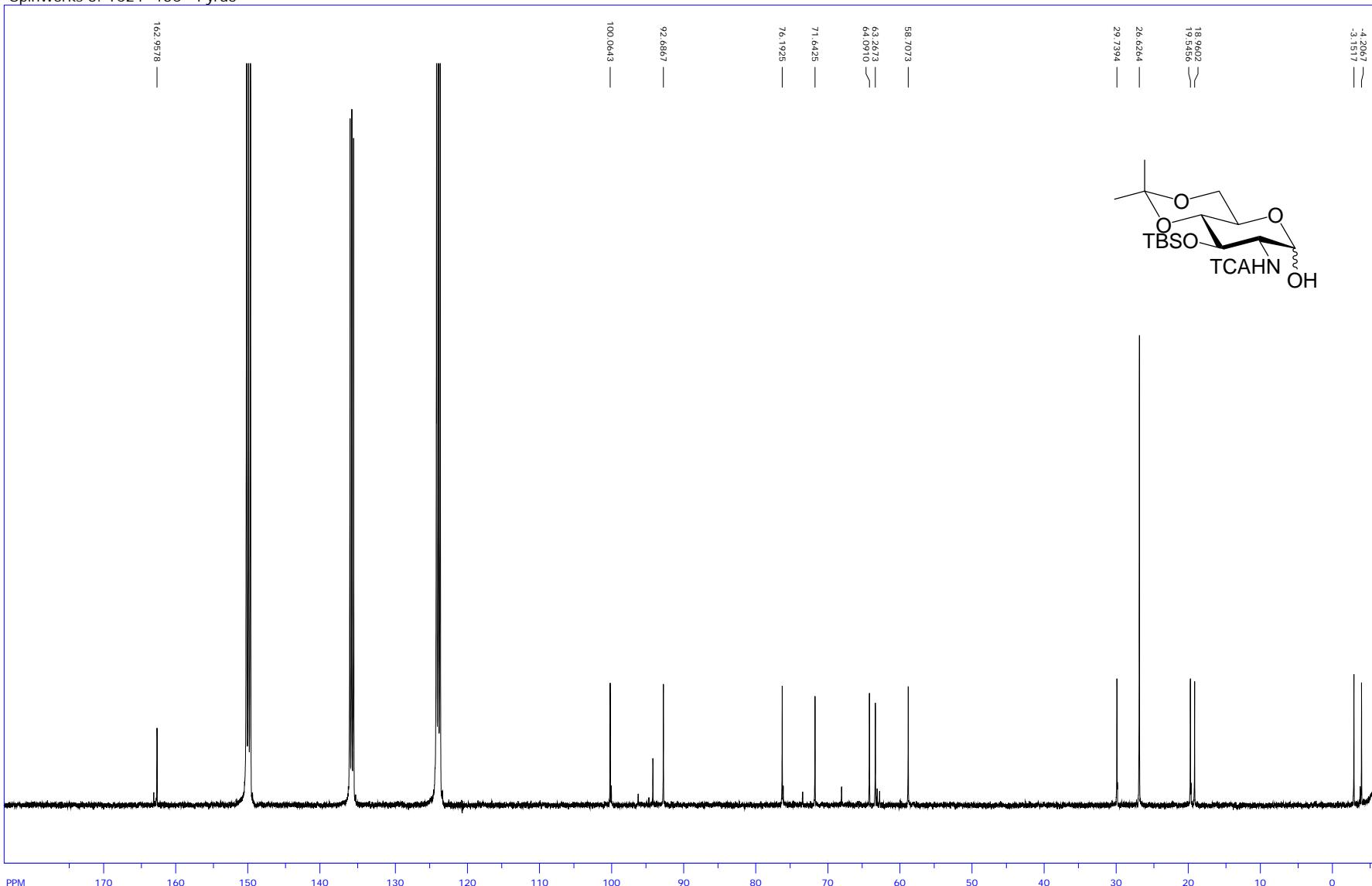
SpinWorks 3: YC24 - DEPT135 - Pyrd5



SpinWorks 3: YC24 - HSQC - Pyrd5

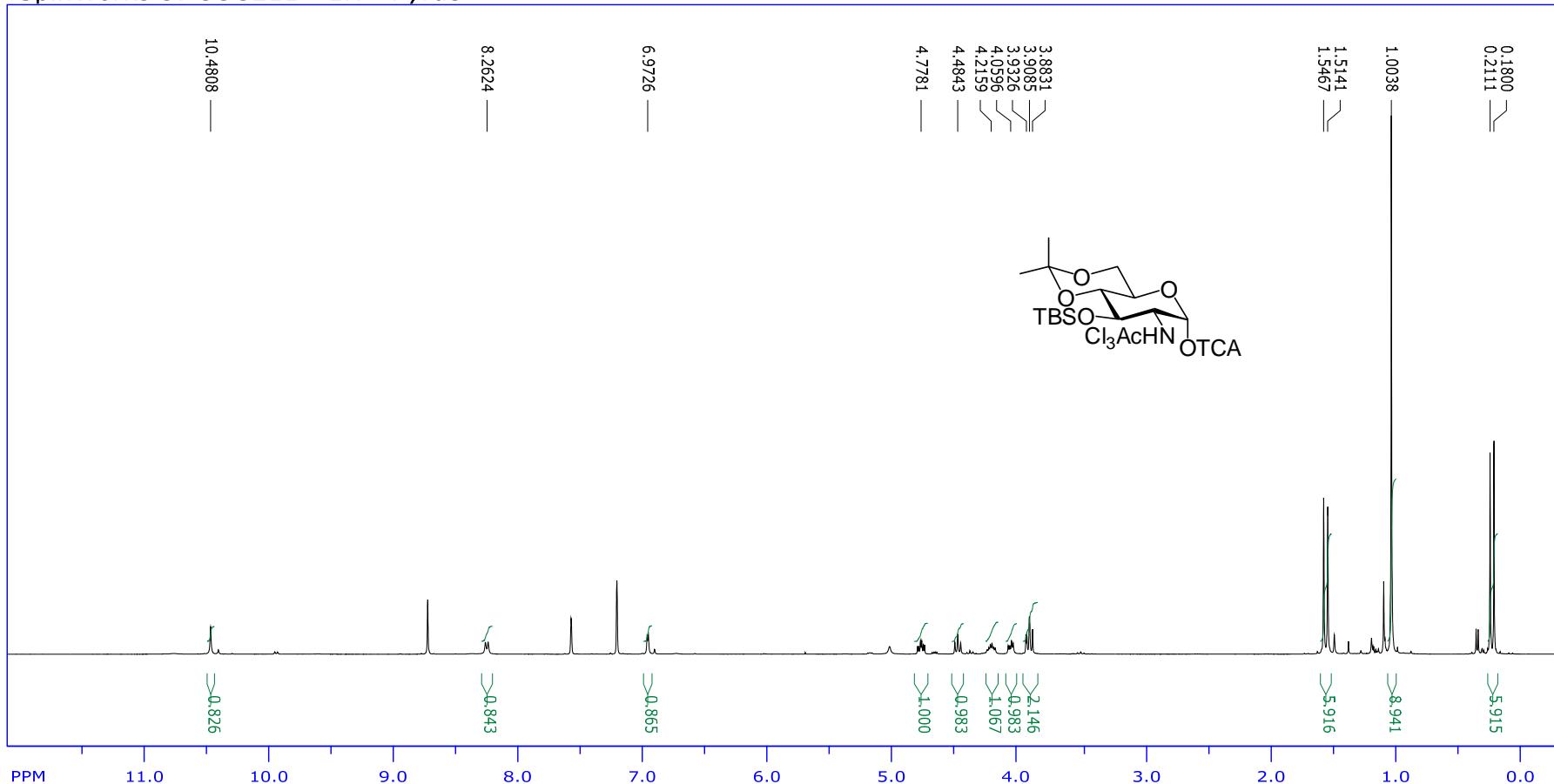


SpinWorks 3: YC24 -13C - Pyrd5

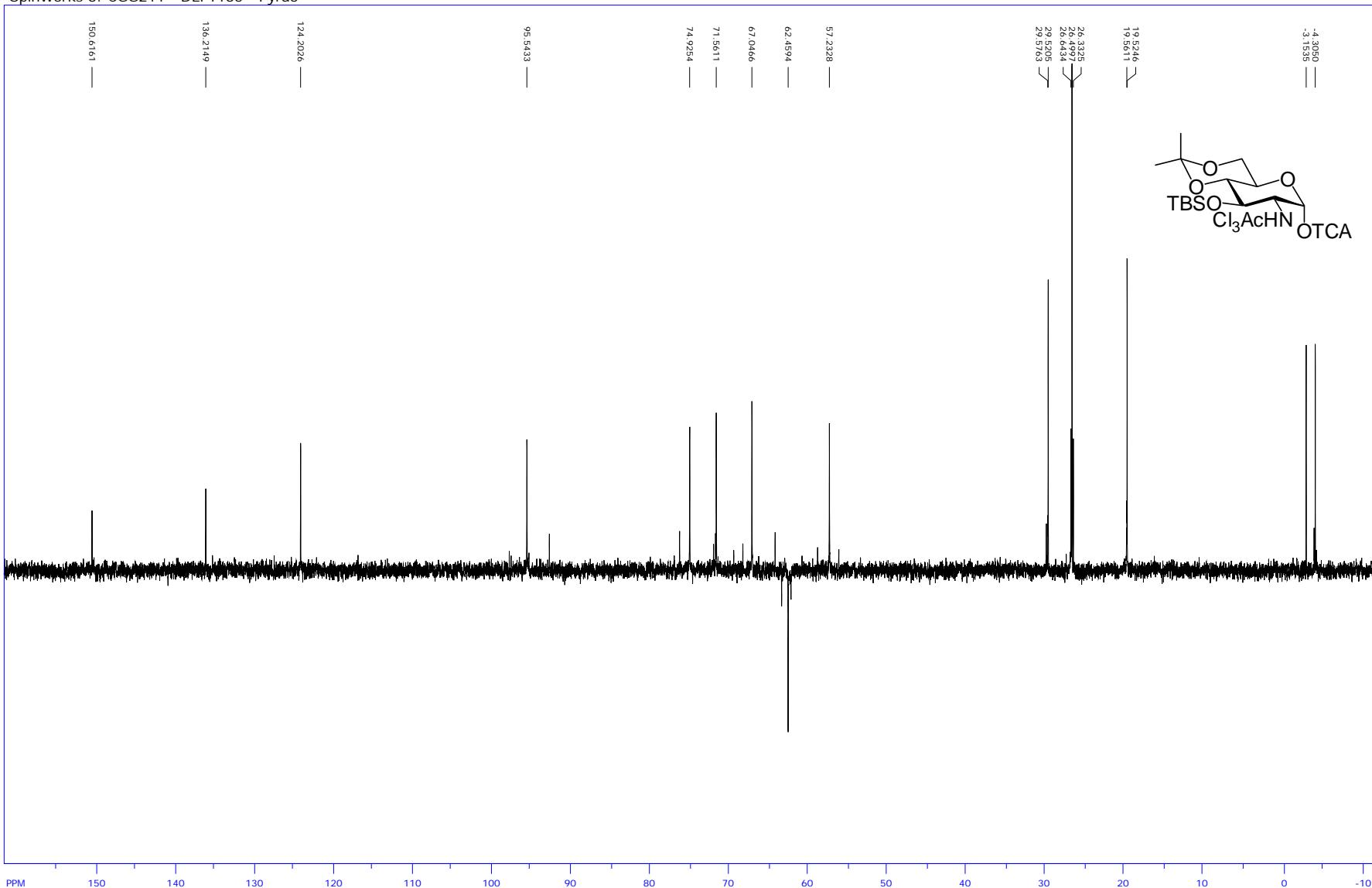


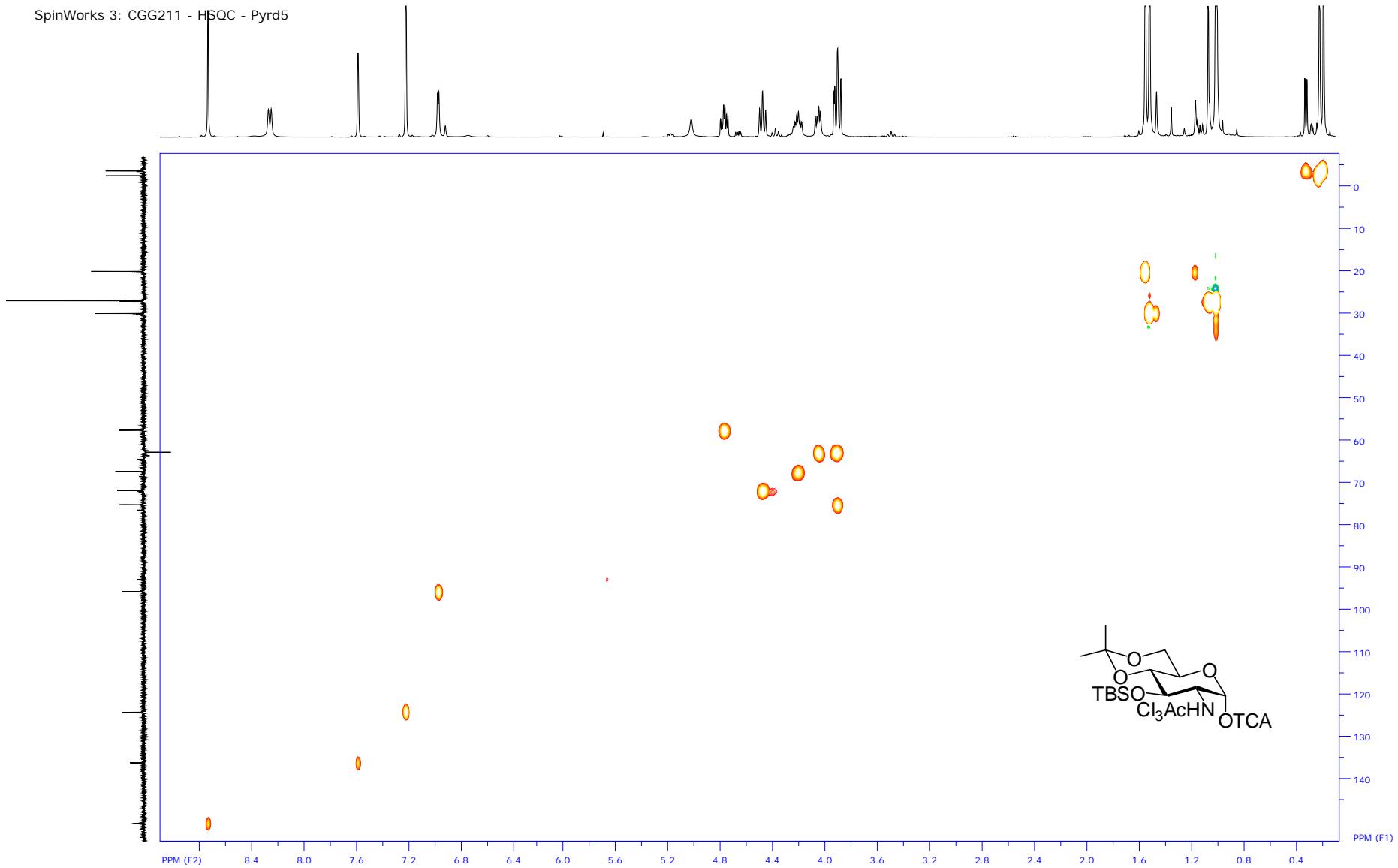
Compound 11

SpinWorks 3: CGG211 - 1H - Pyrd5

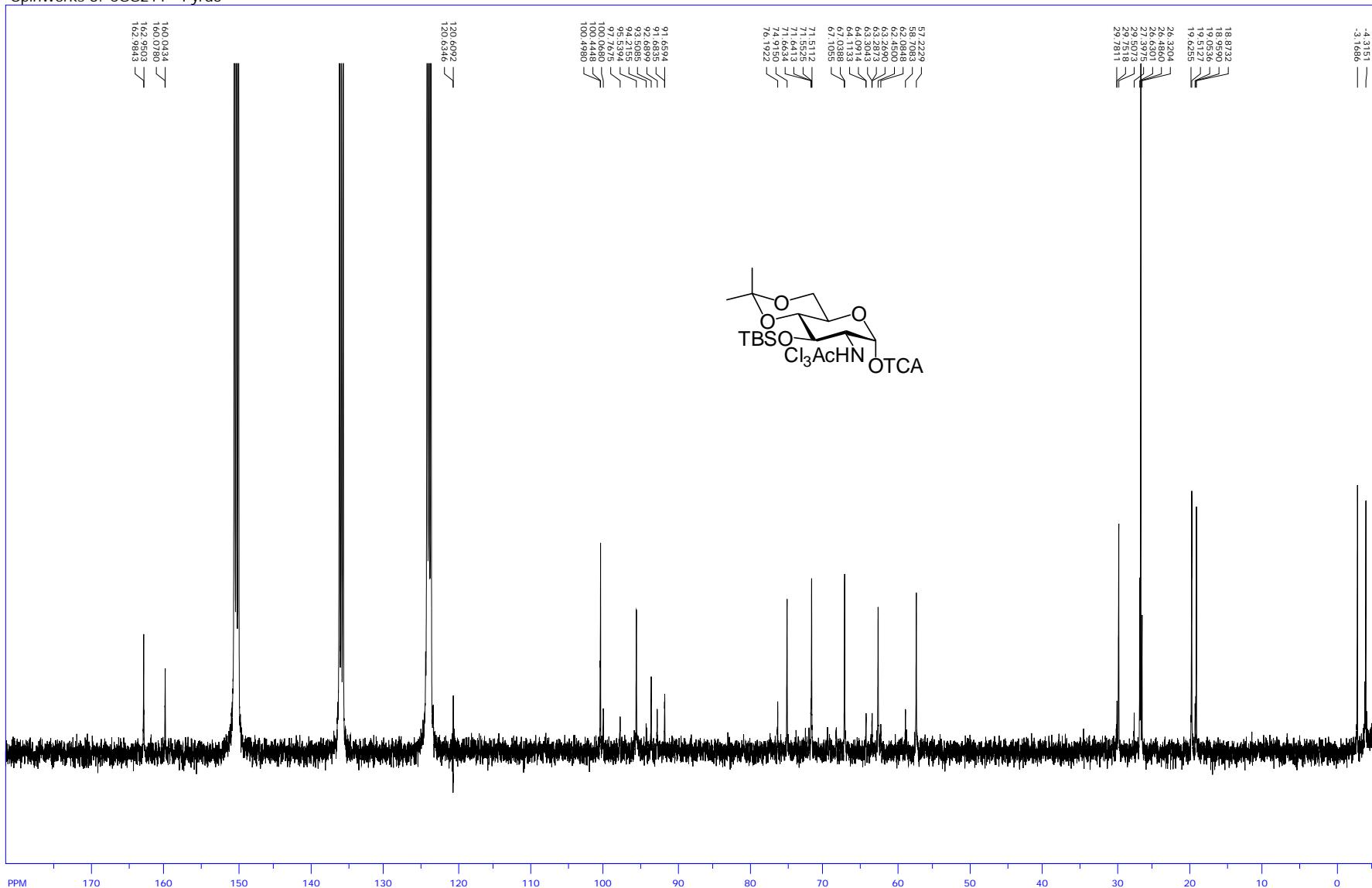


SpinWorks 3: CGG211 - DEPT135 - Pyrd5



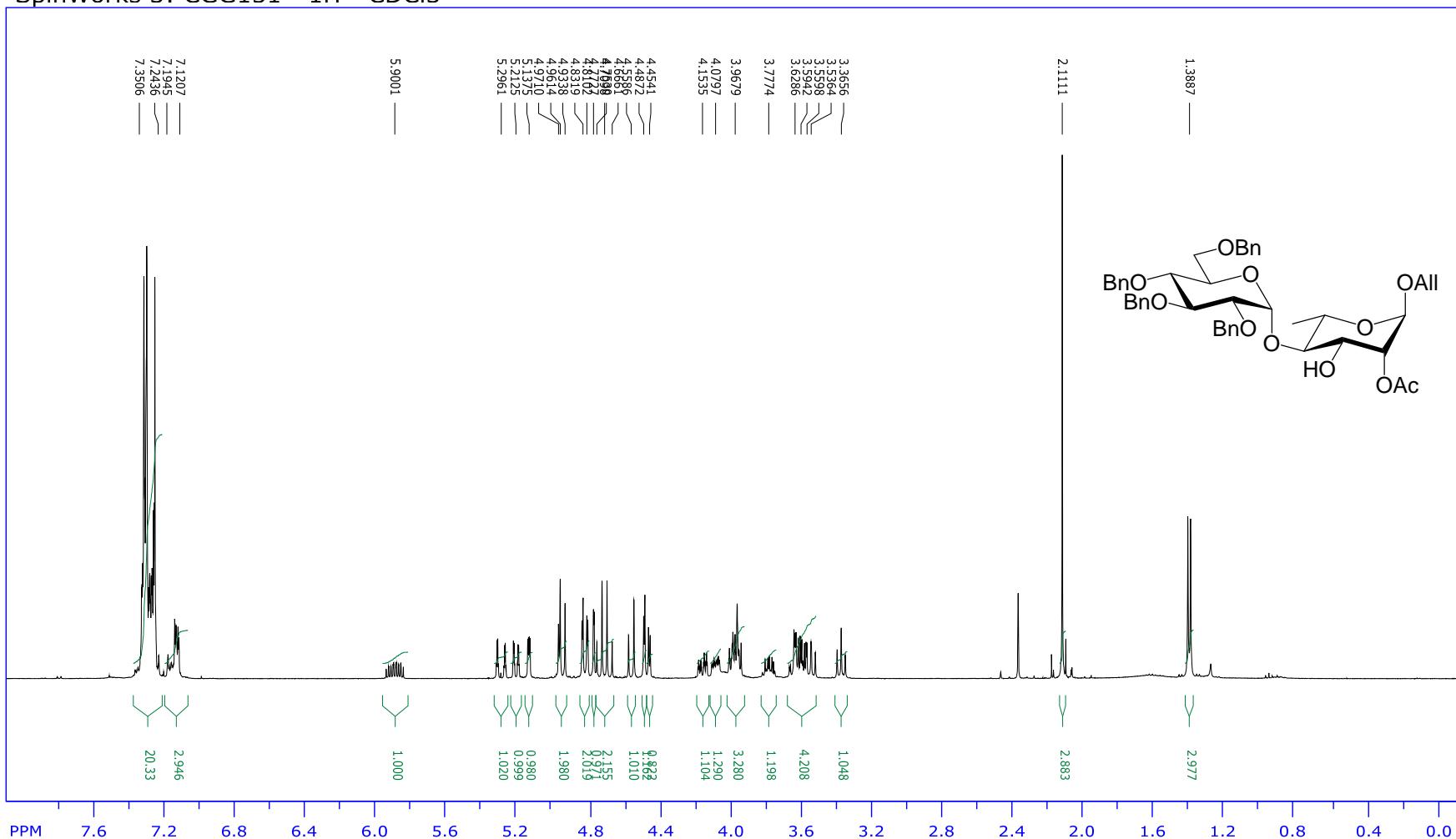


SpinWorks 3: CGG211 - Pyrd5

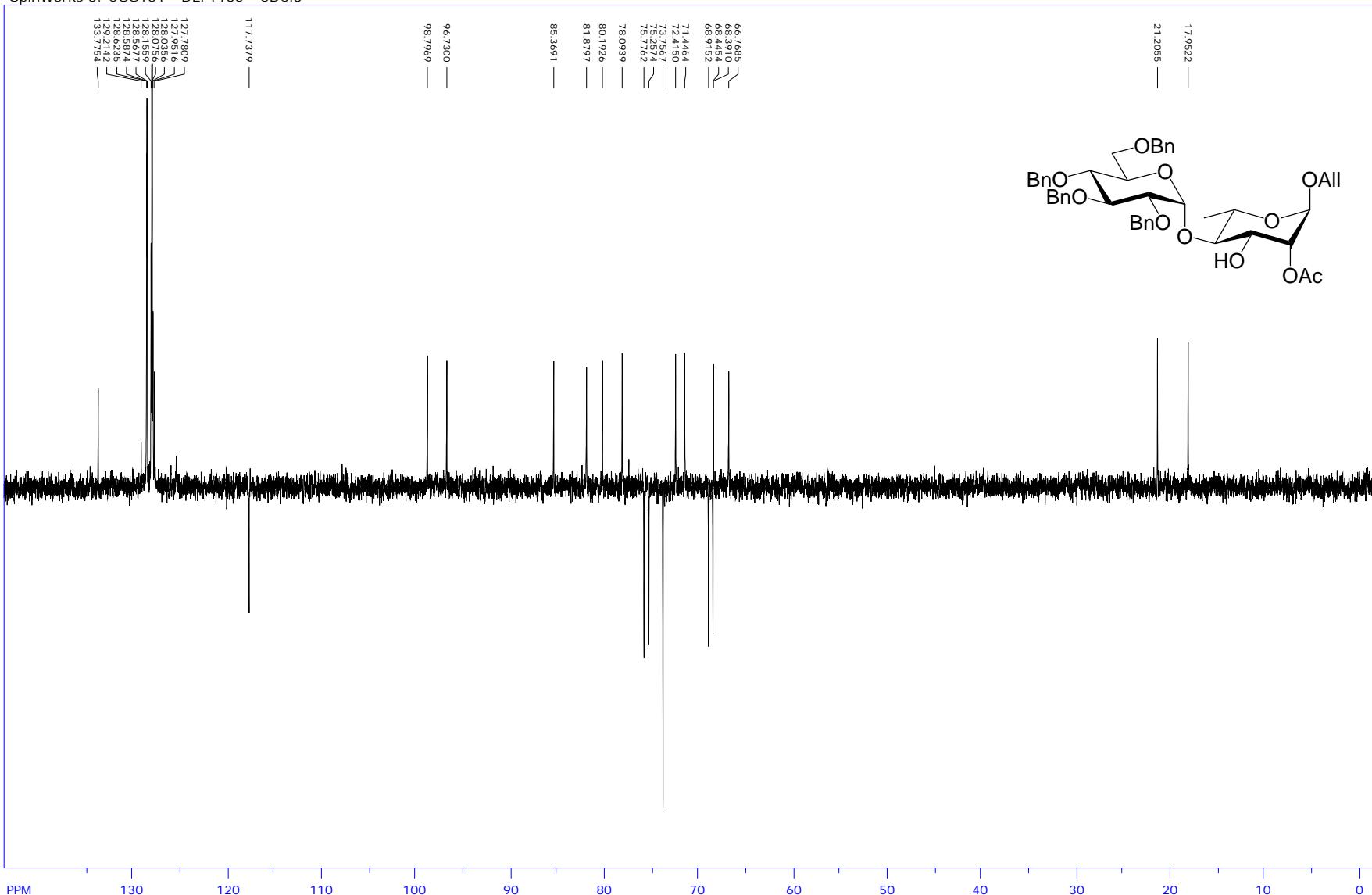


Compound 16

SpinWorks 3: CGG151 - 1H - CDCl₃

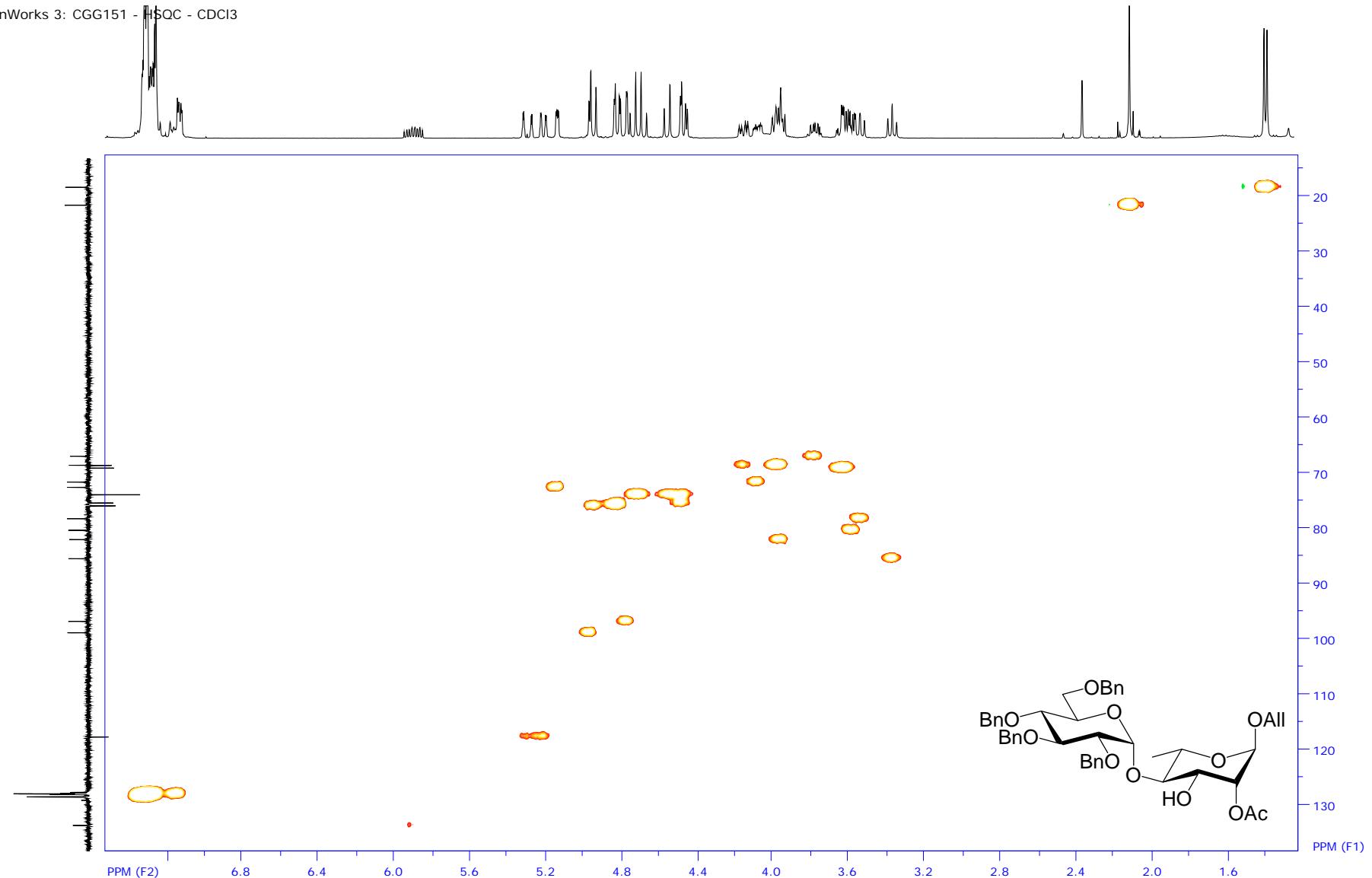


SpinWorks 3: CGG151 - DEPT135 - CDCI3

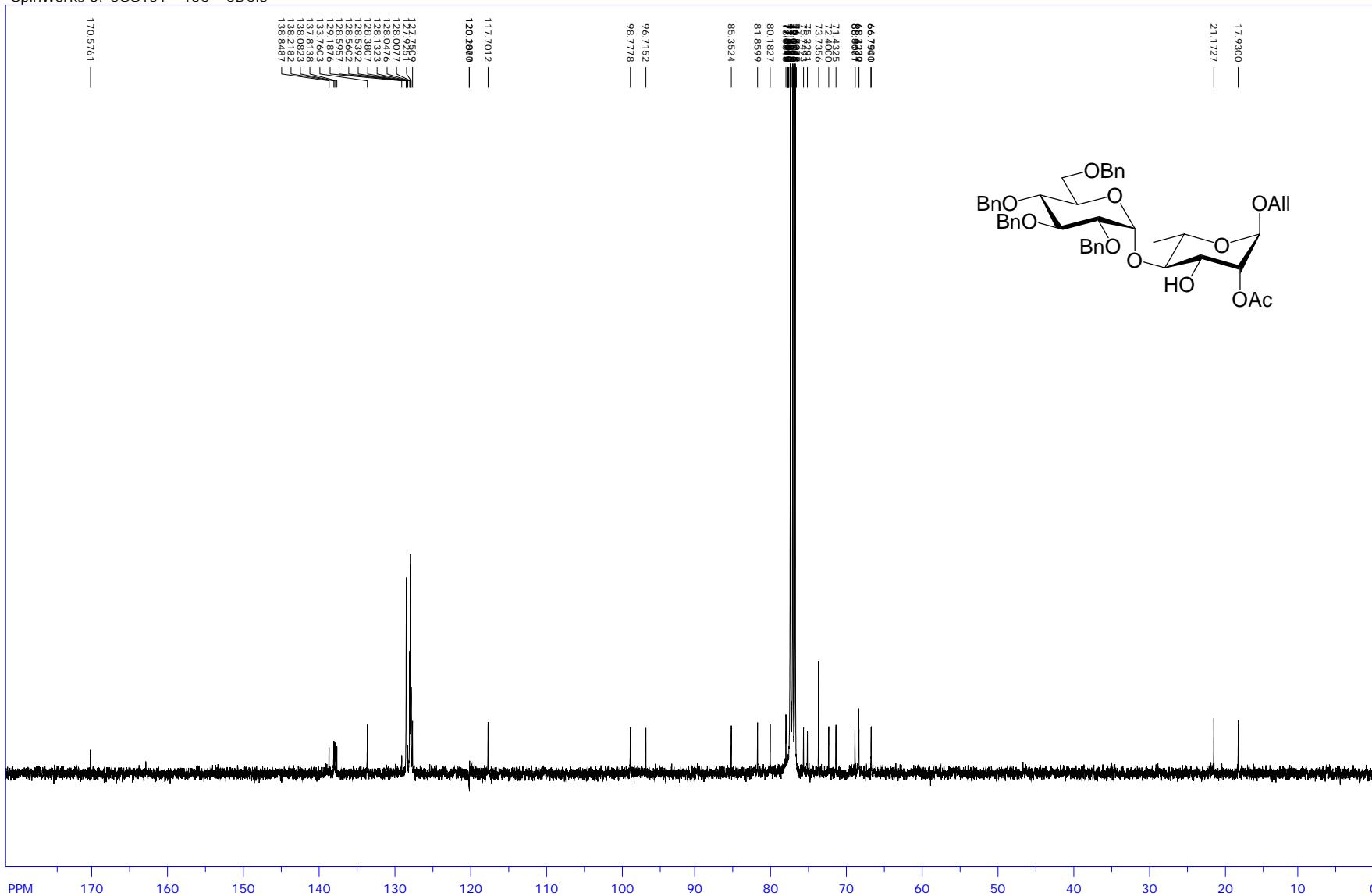


S100

SpinWorks 3: CGG151 - HSQC - CDCl₃



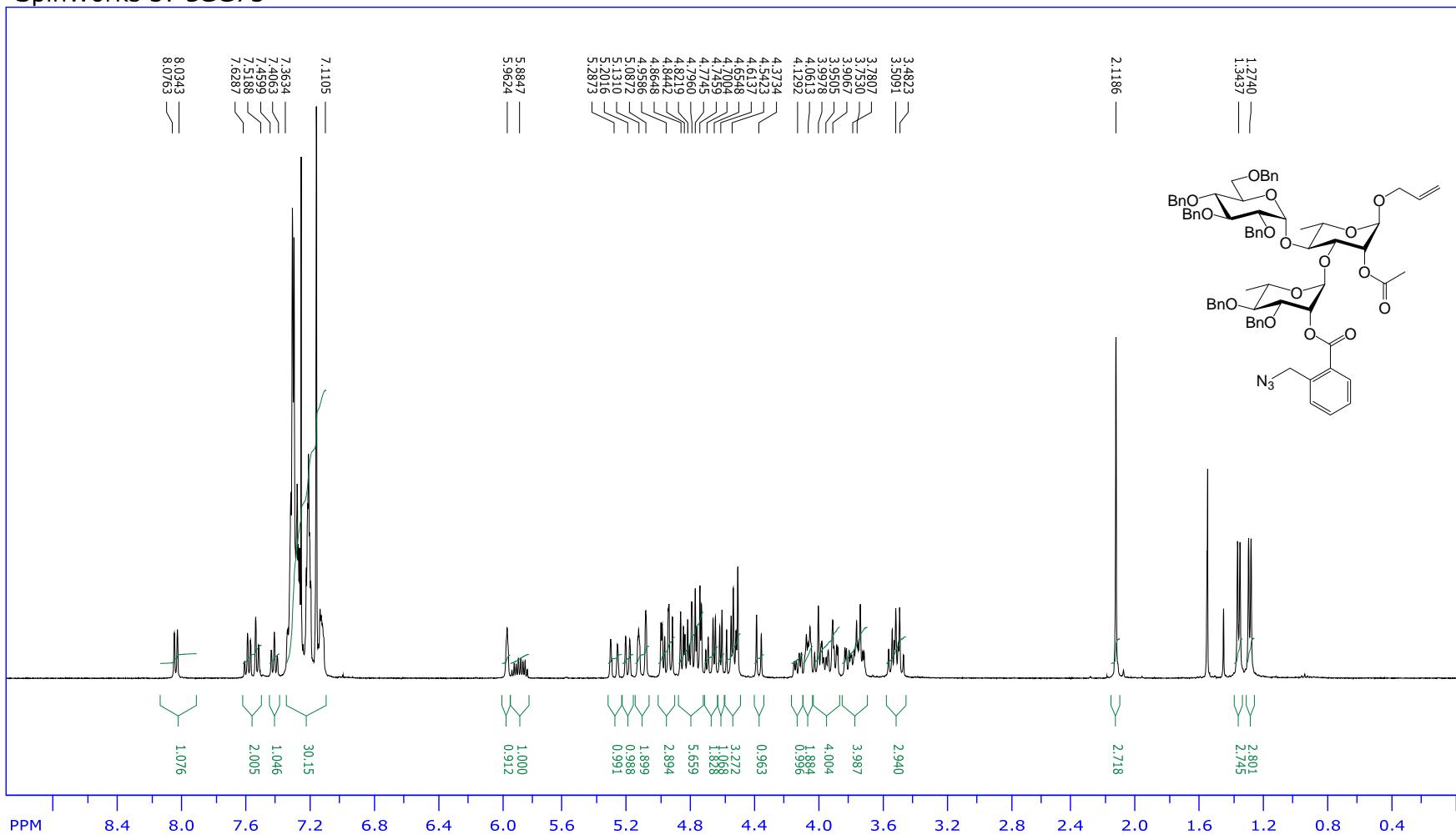
SpinWorks 3: CGG151 - 13C - CDCI3

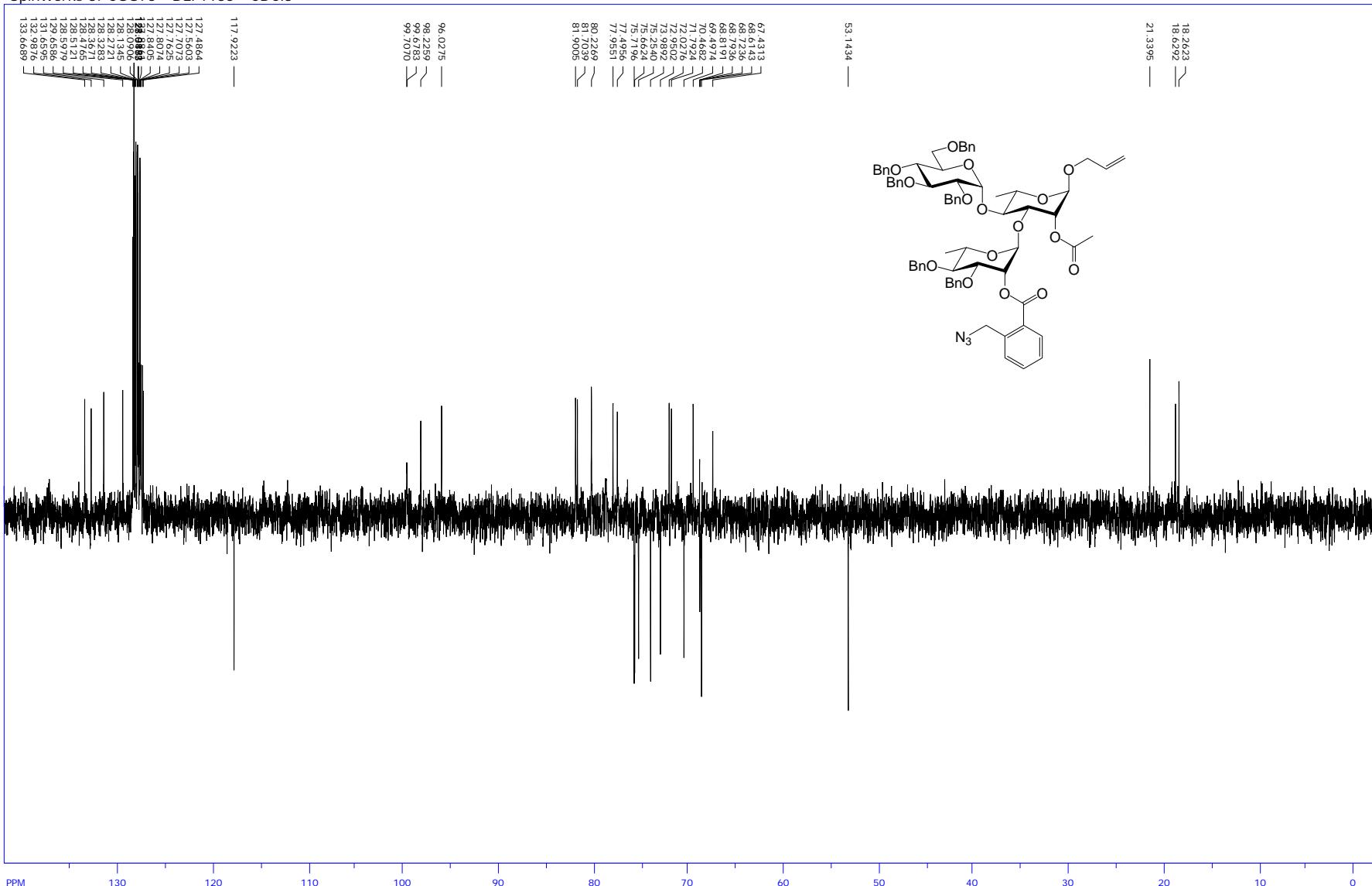


S102

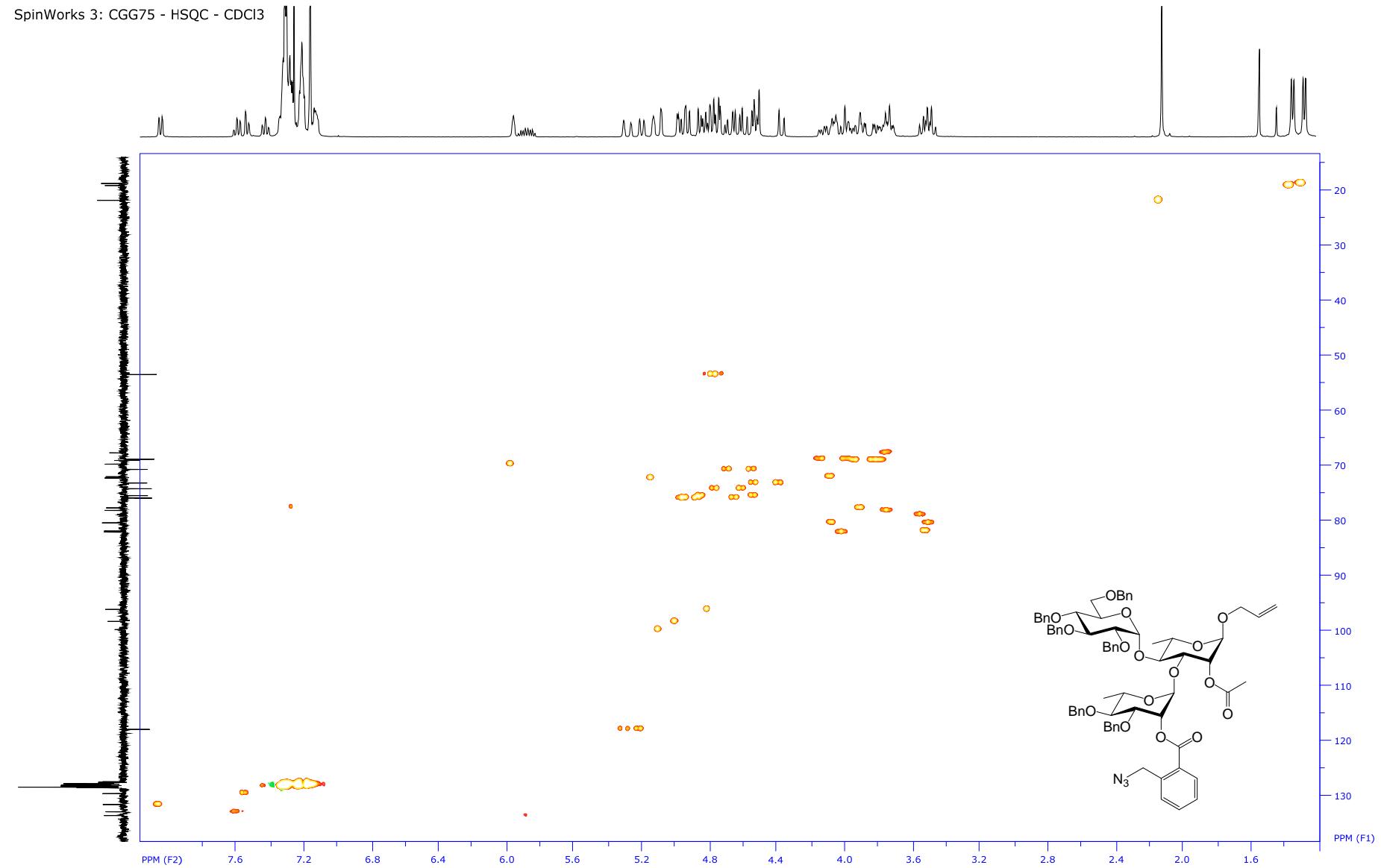
Compound 17

SpinWorks 3: CGG75

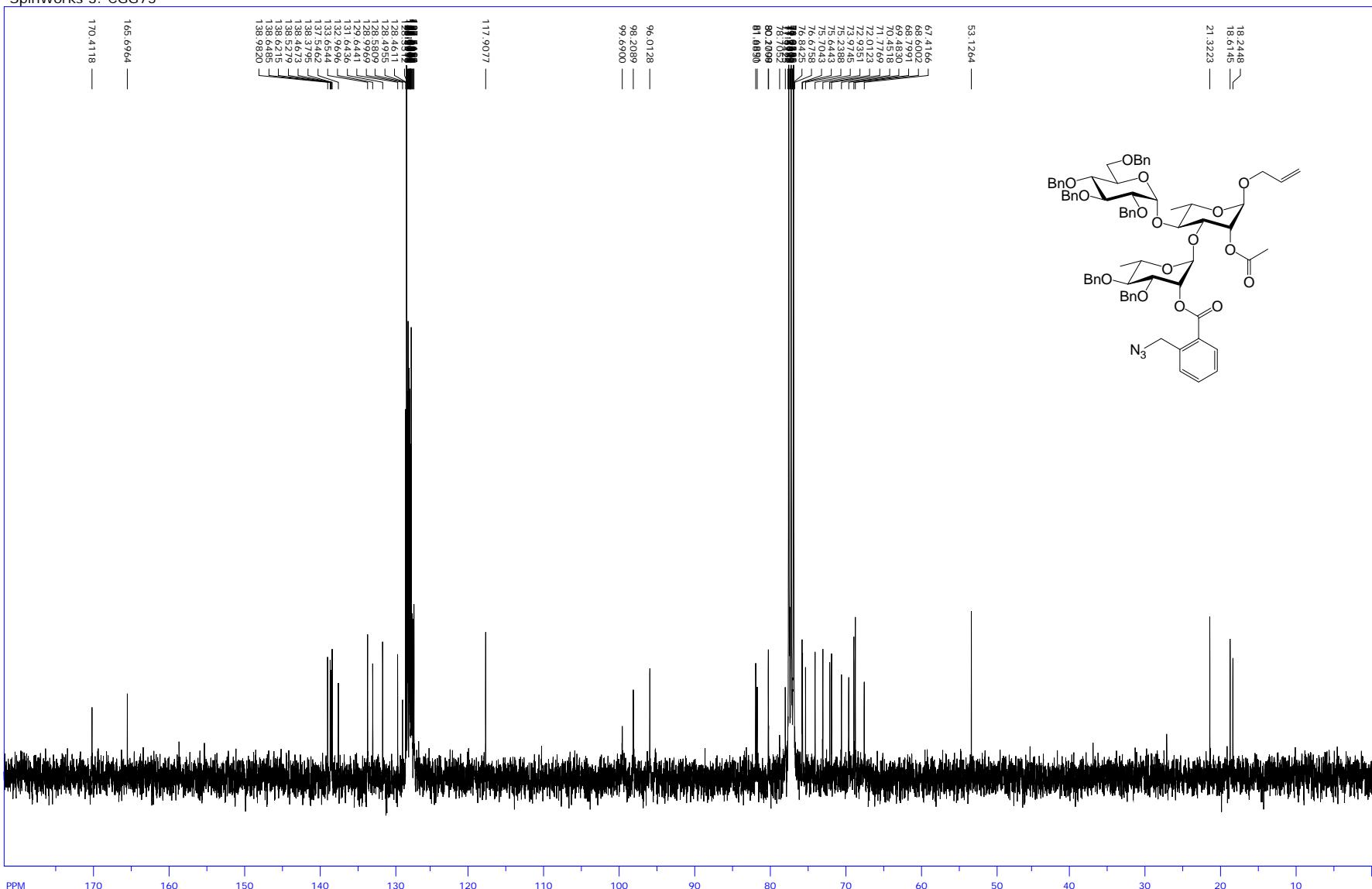


SpinWorks 3: CGG75 - DEPT135 - CDCl₃

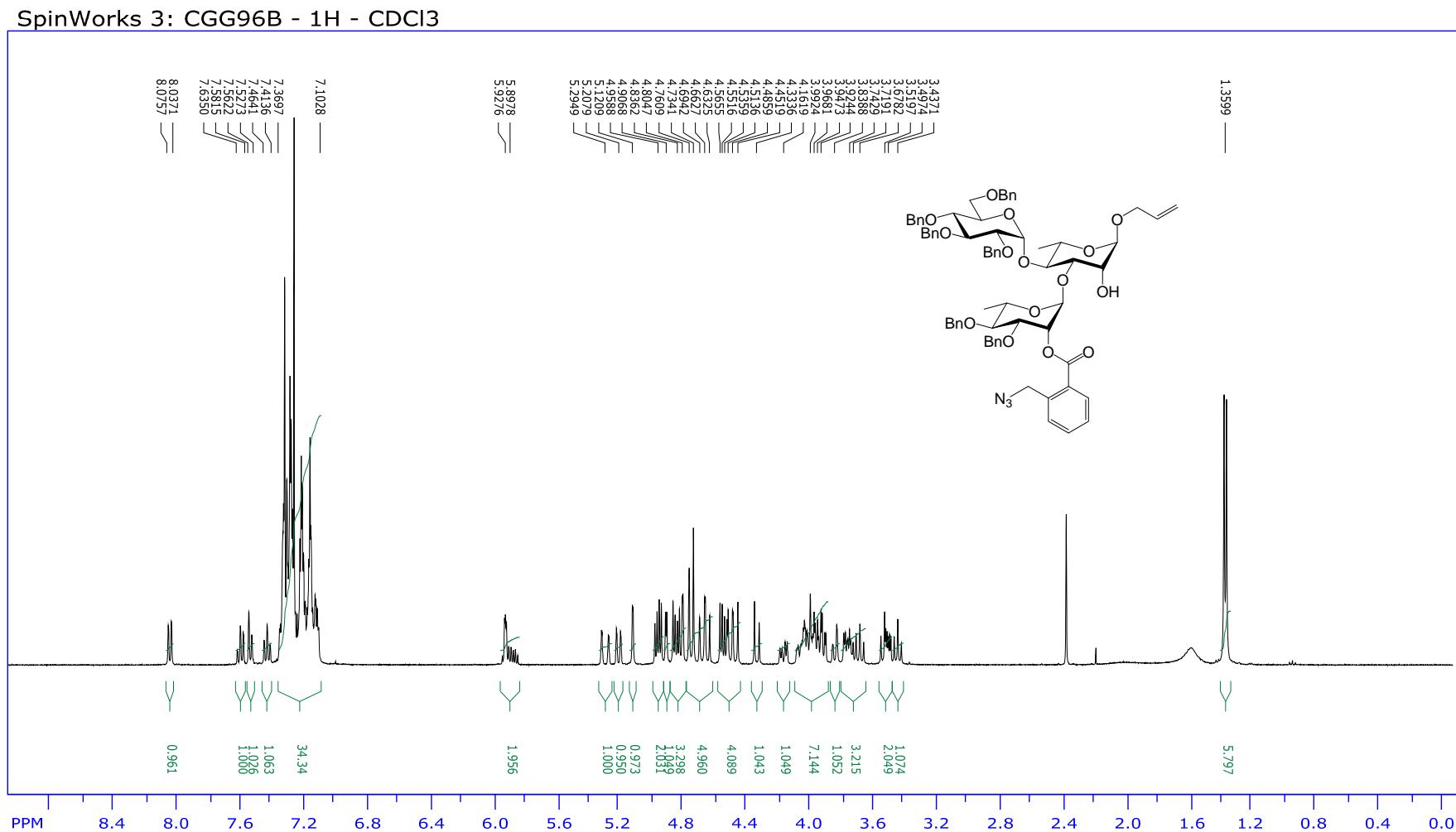
SpinWorks 3: CGG75 - HSQC - CDCl3

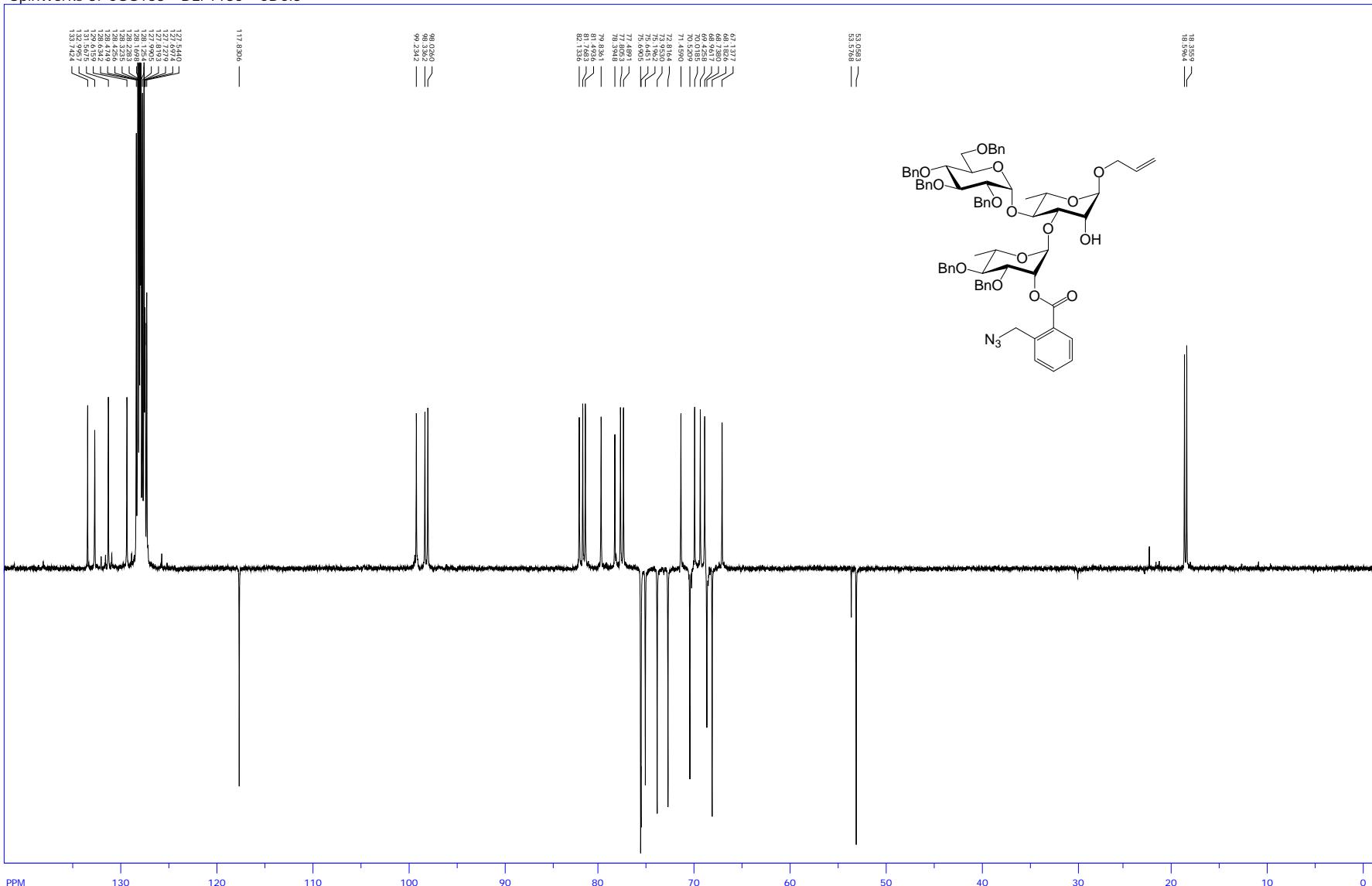


SpinWorks 3: CGG75

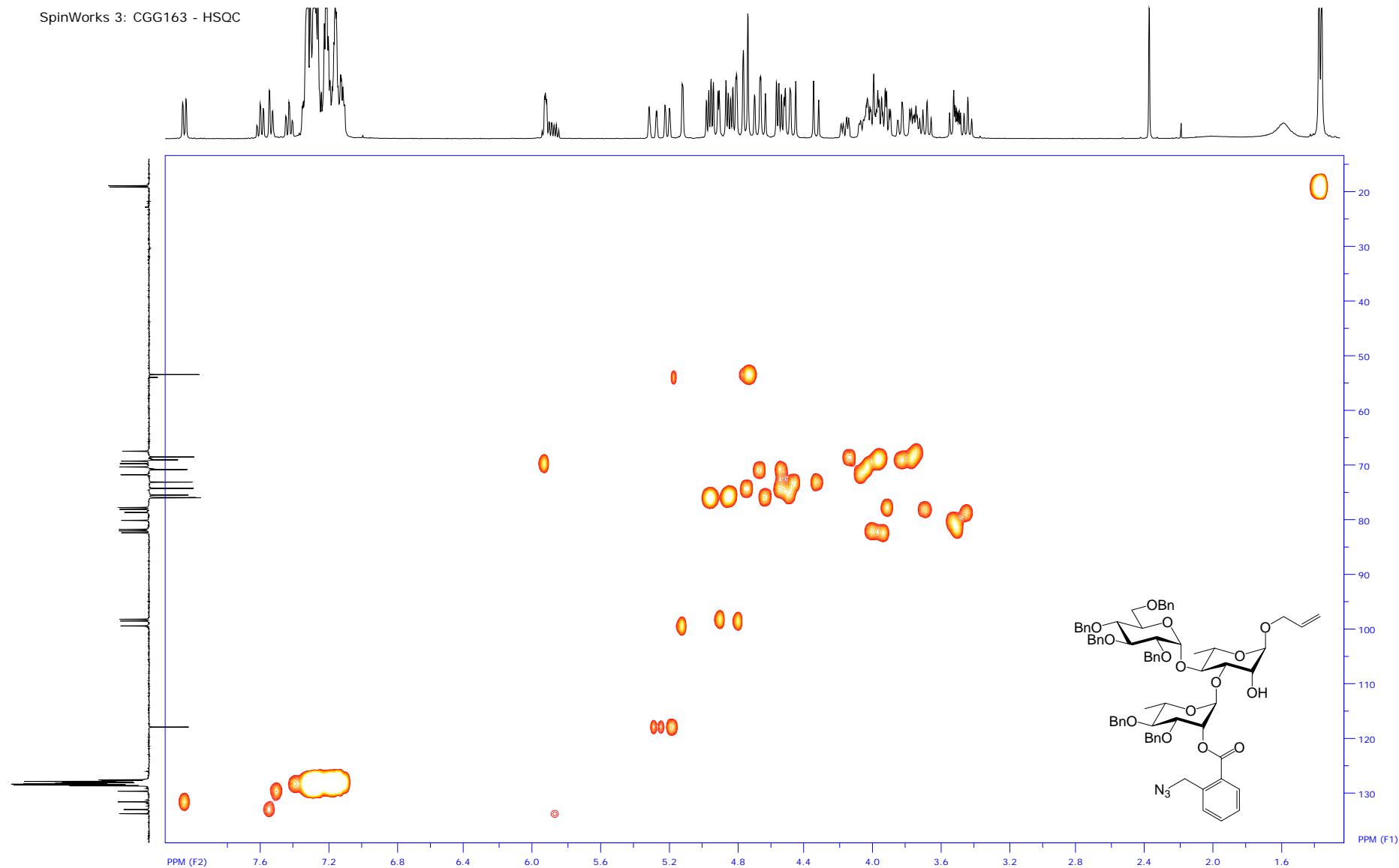


Compound 18

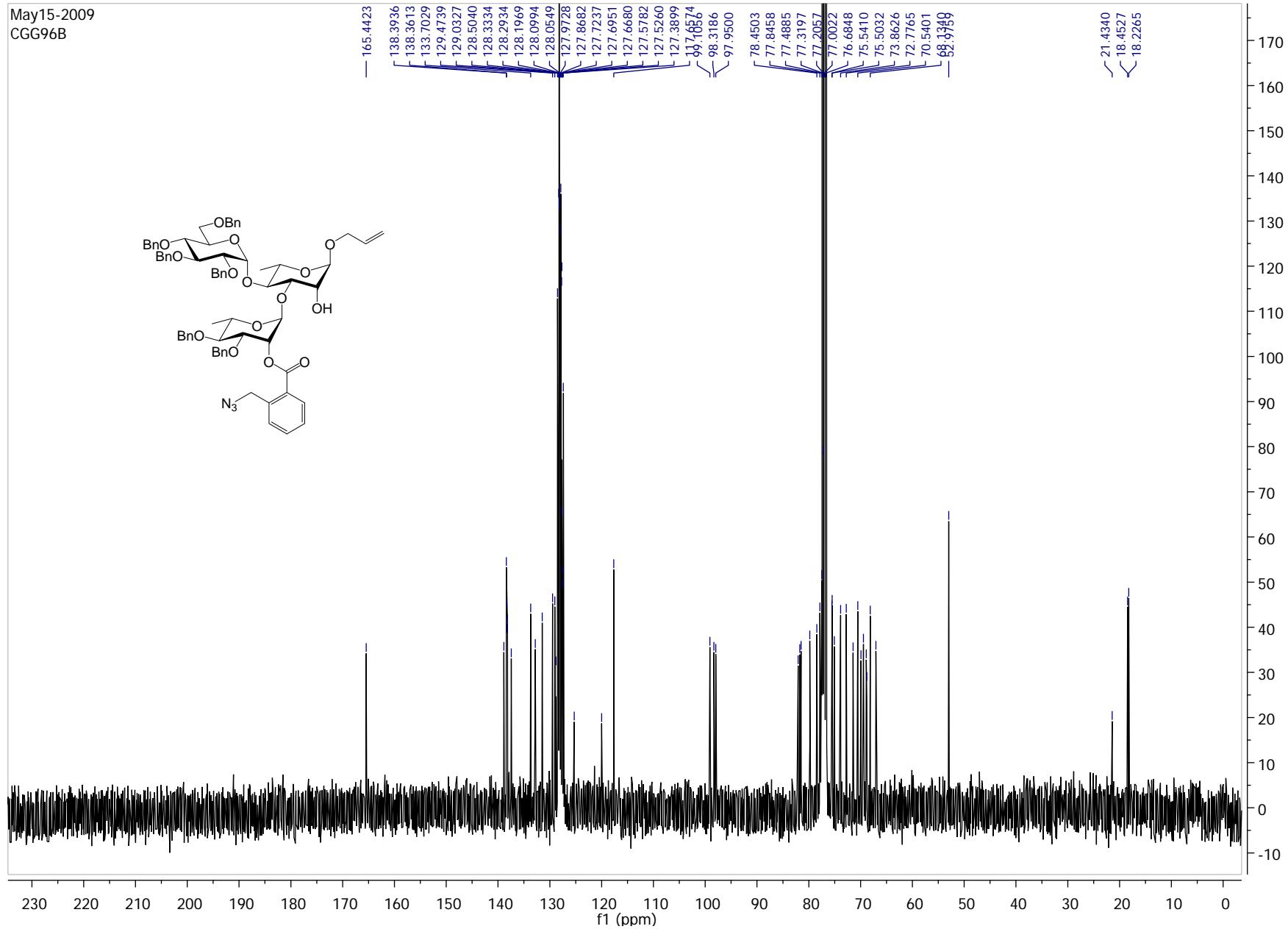


SpinWorks 3: CGG163 - DEPT135 - CDCl₃

SpinWorks 3: CGG163 - HSQC

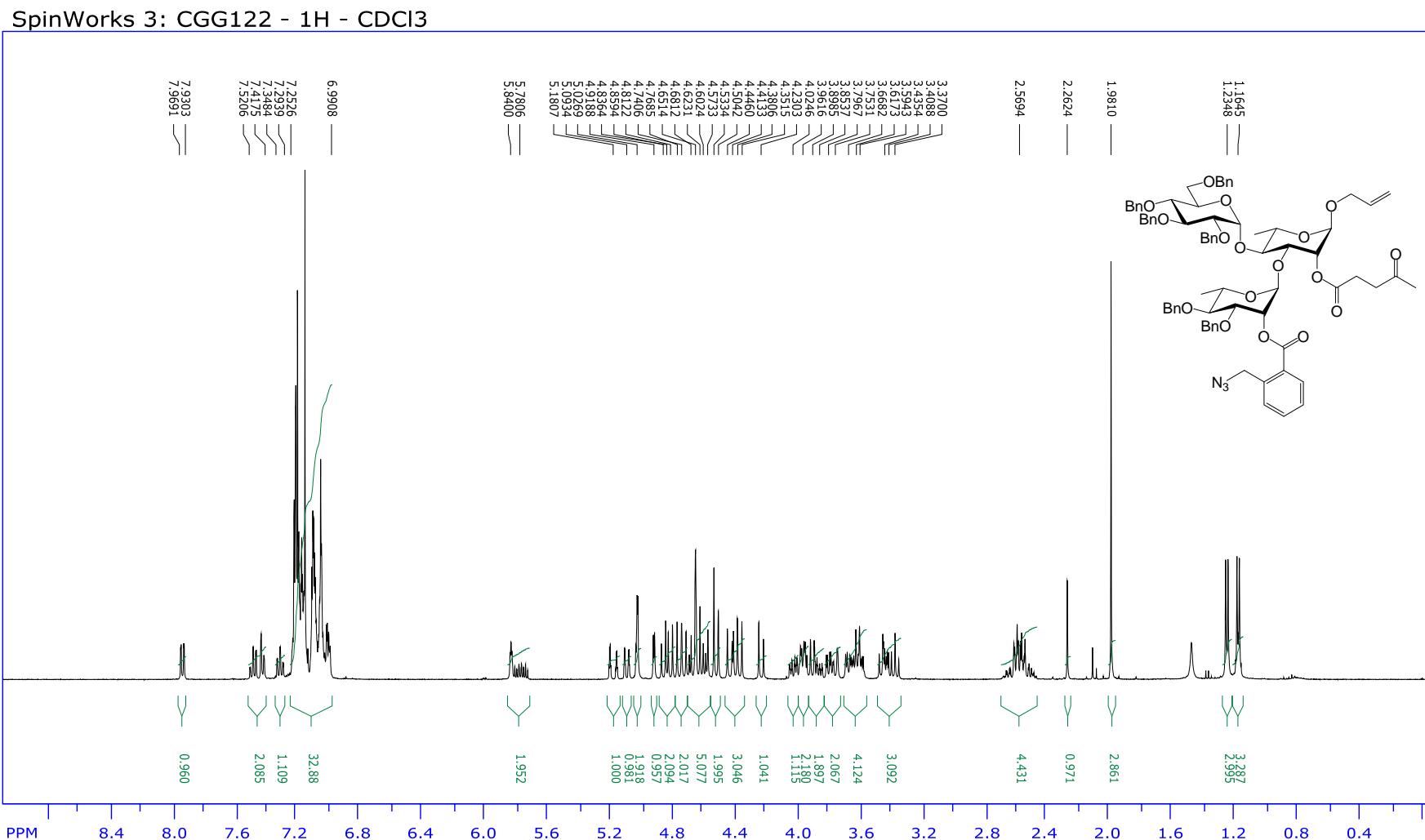


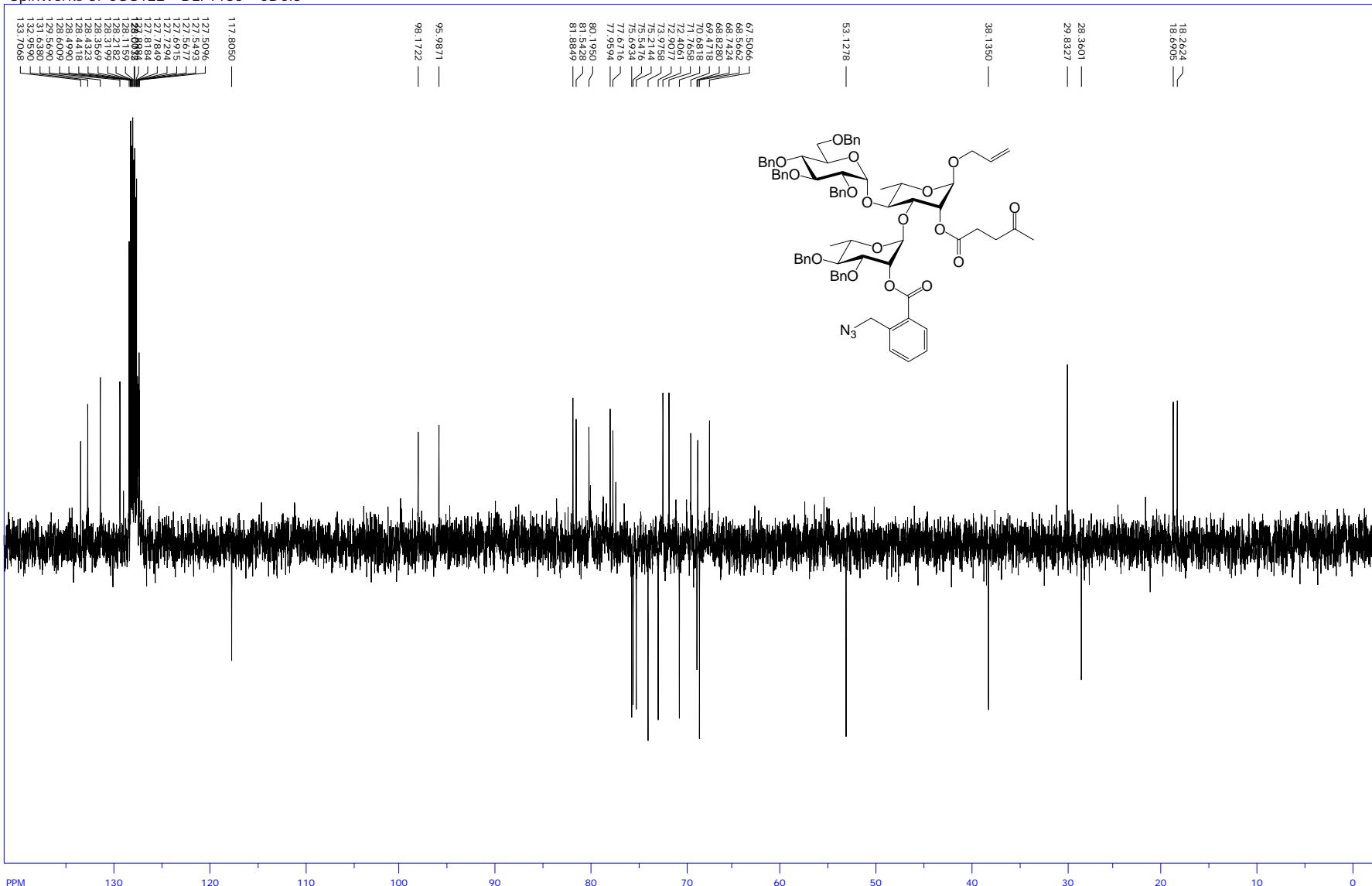
S109



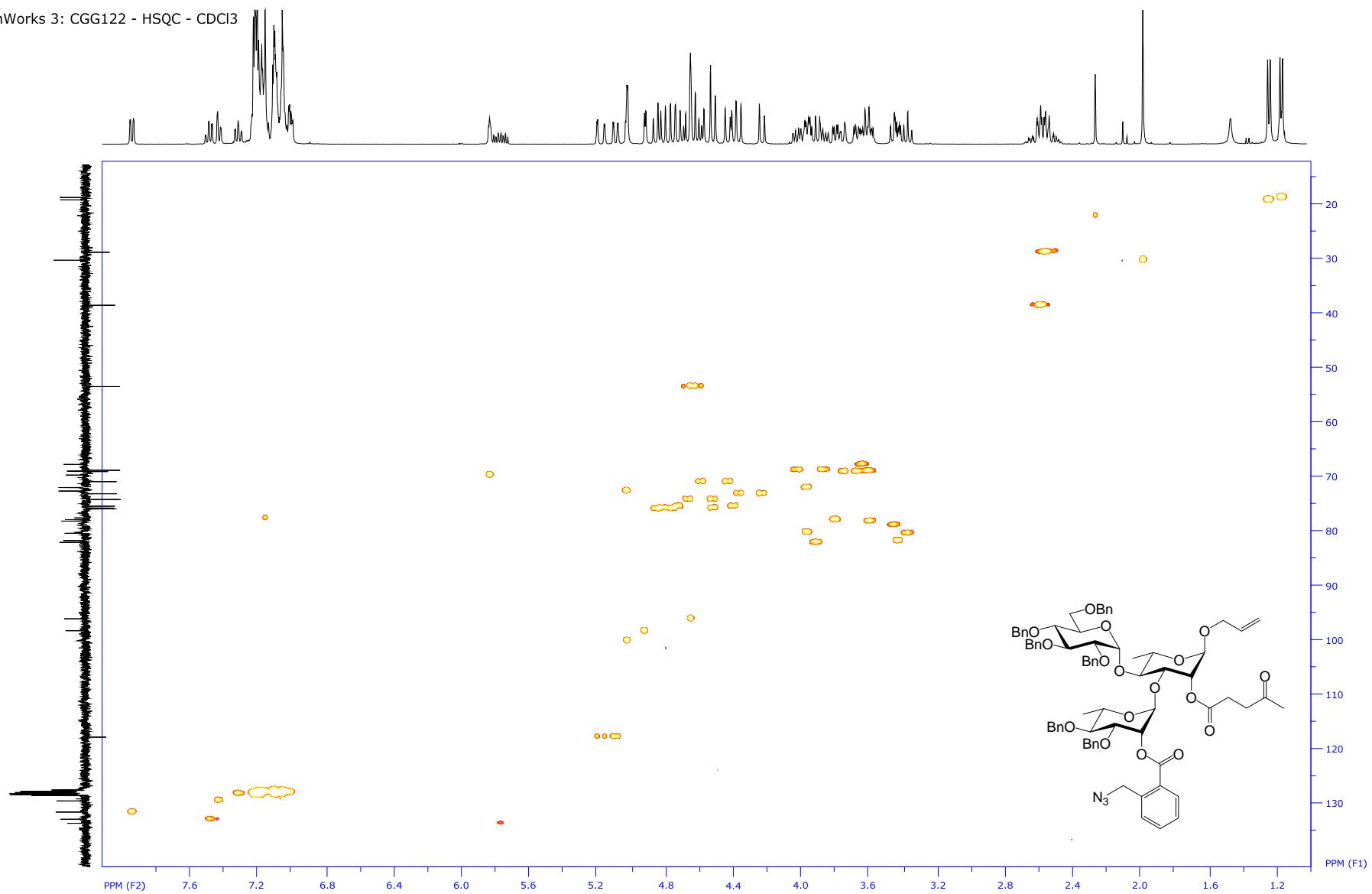
S110

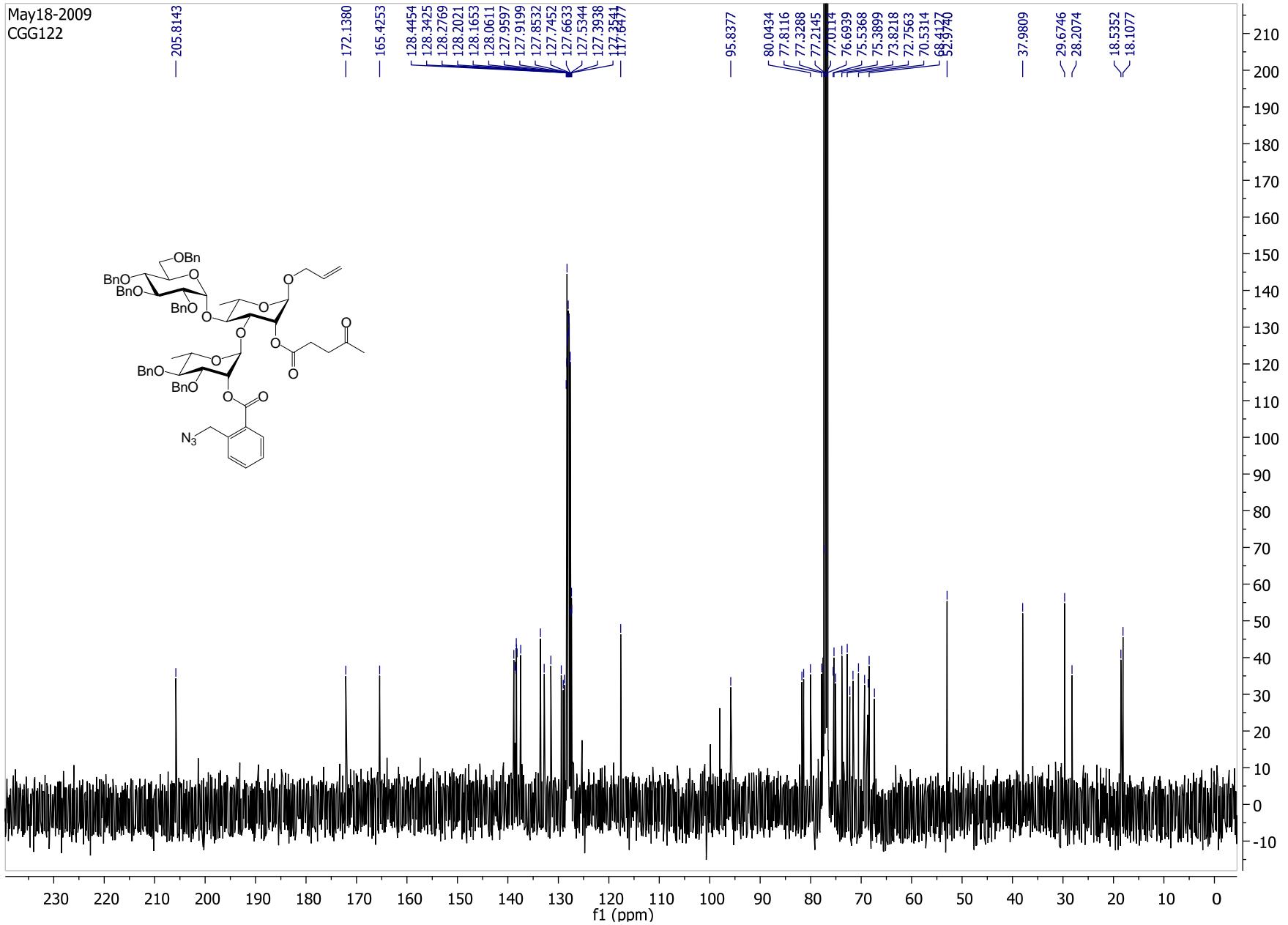
Compound 19



SpinWorks 3: CGG122 - DEPT135 - CDCl₃

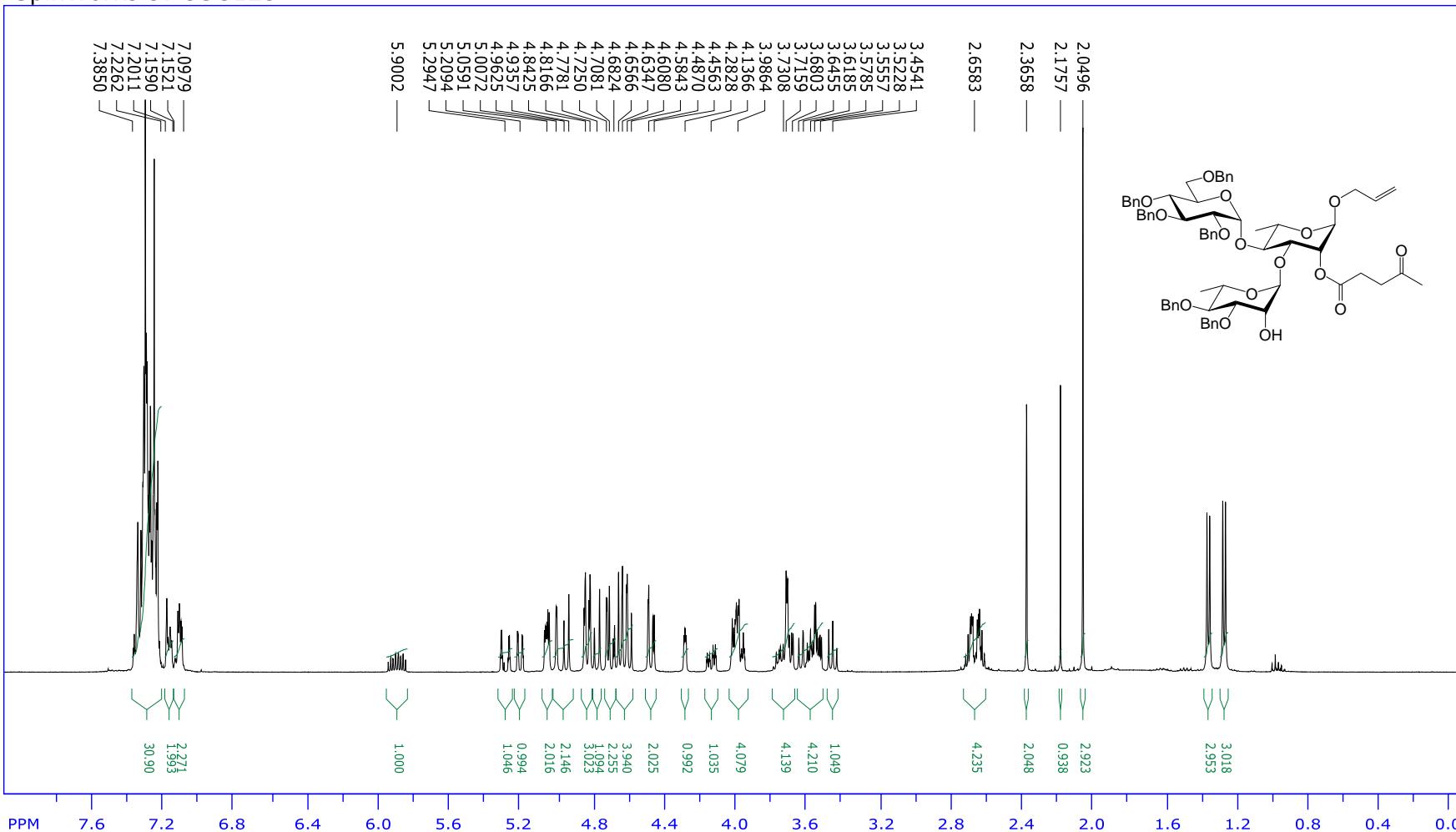
SpinWorks 3: CGG122 - HSQC - CDCl₃



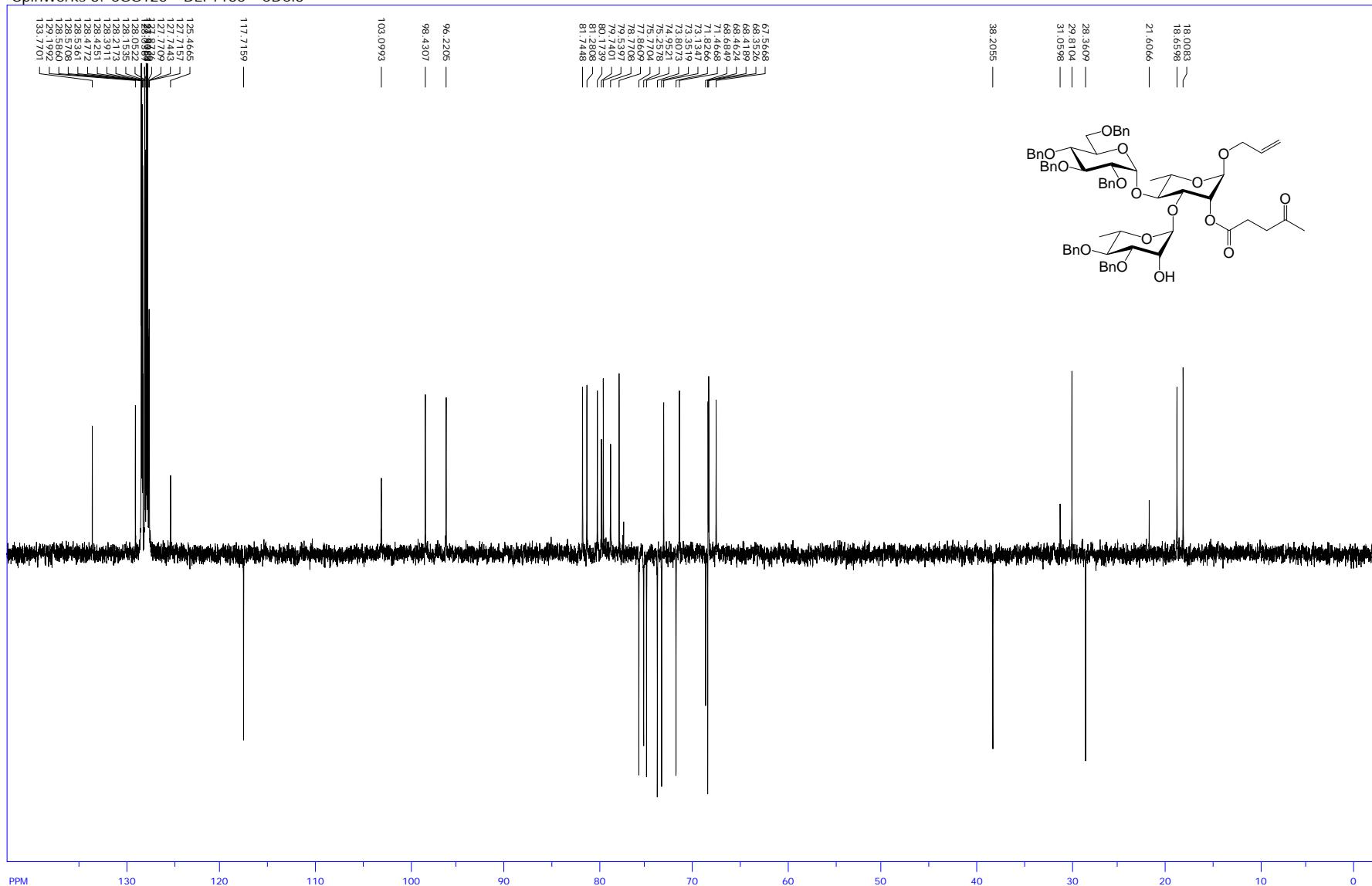


Compound 20

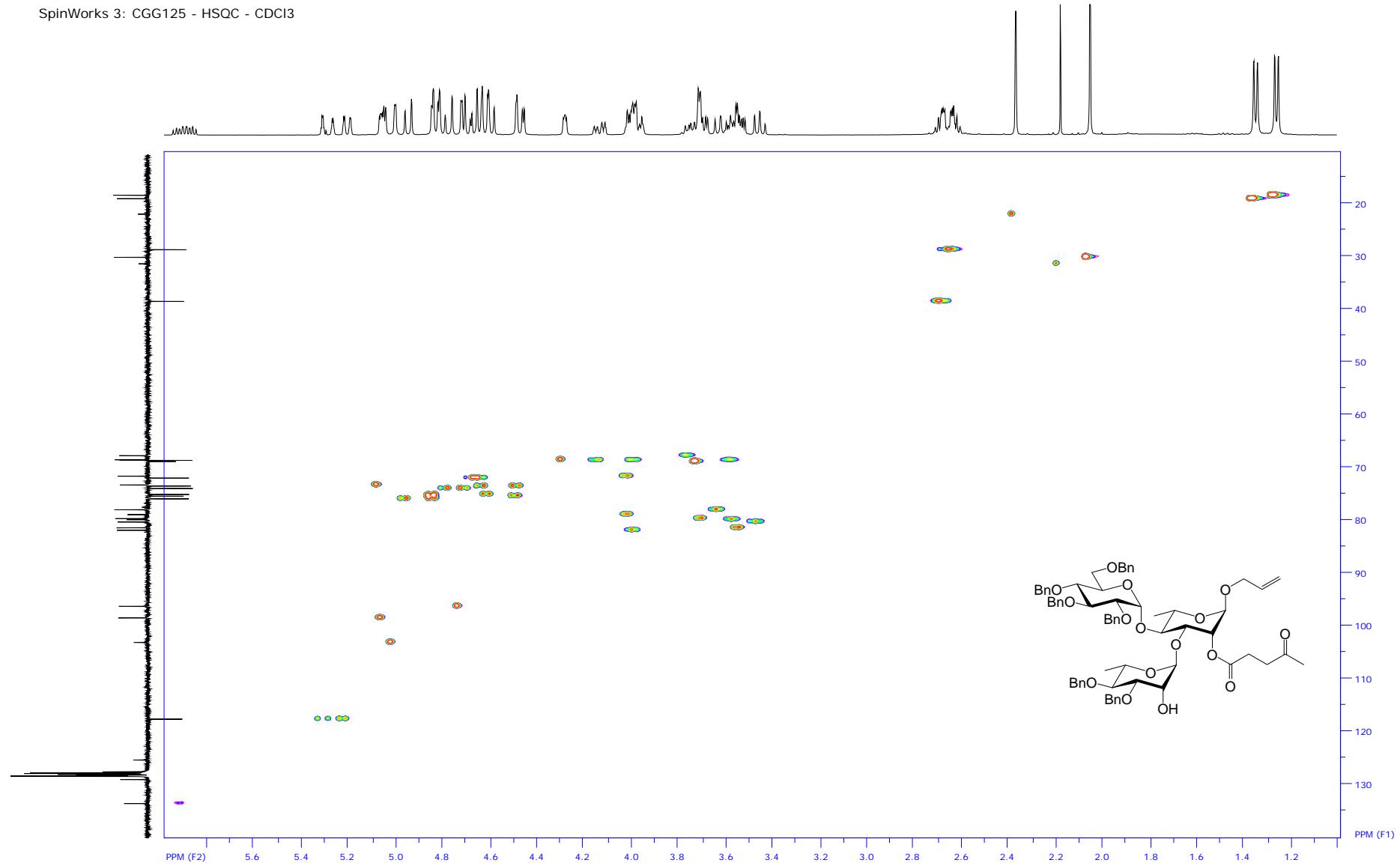
SpinWorks 3: CGG125

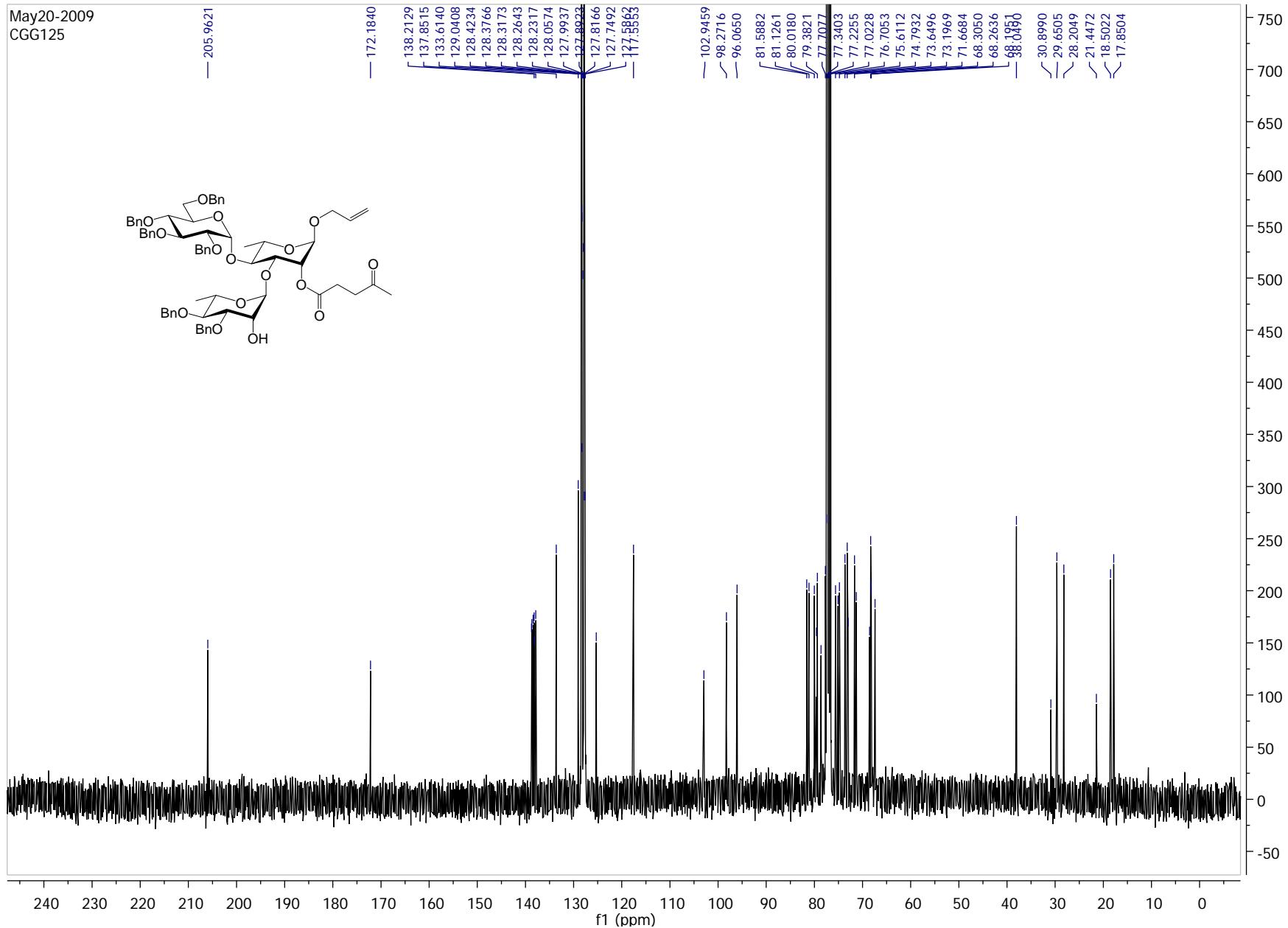


SpinWorks 3: CGG125 - DEPT135 - CDCl₃

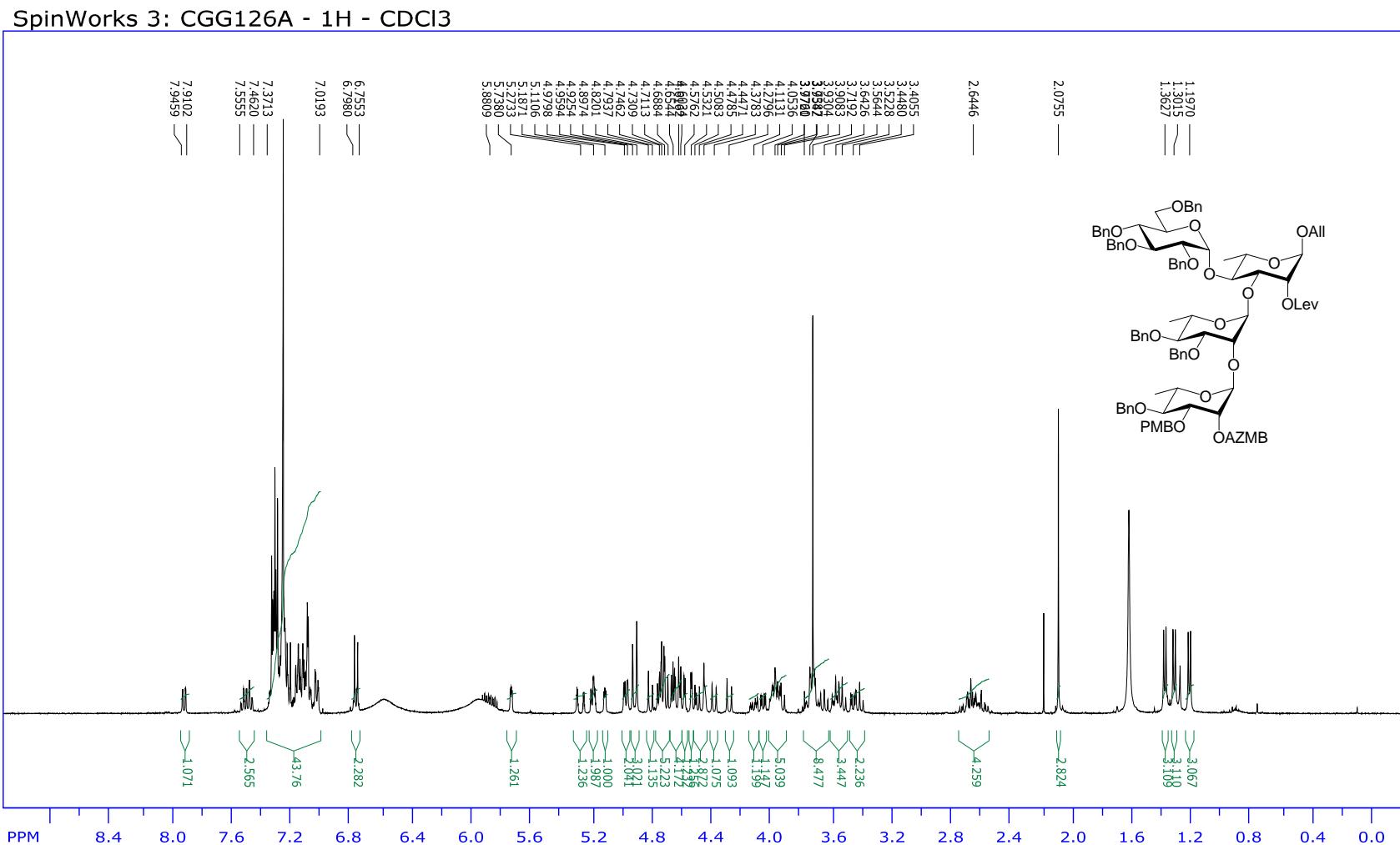


SpinWorks 3: CGG125 - HSQC - CDCl3

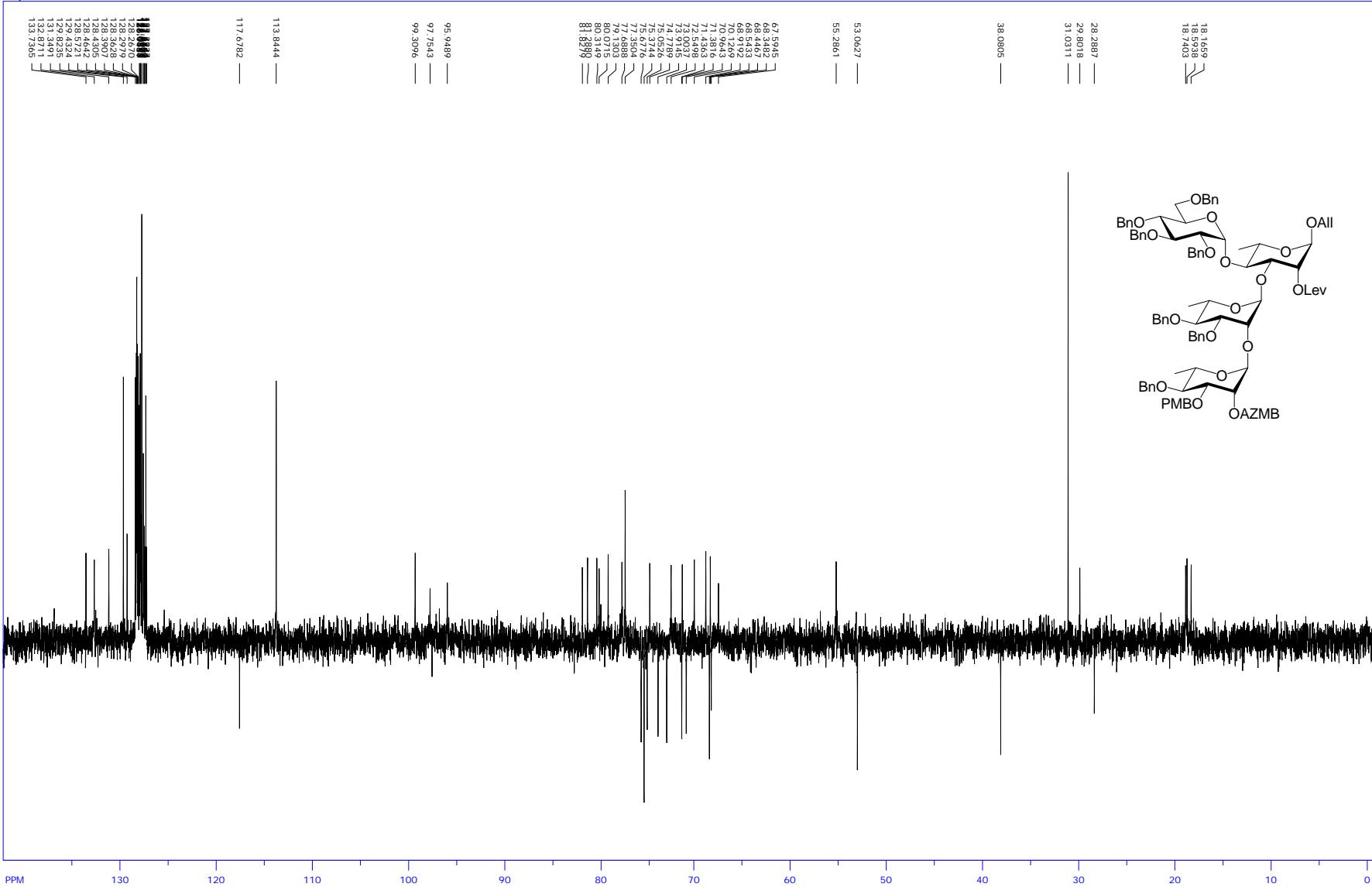




Compound 21

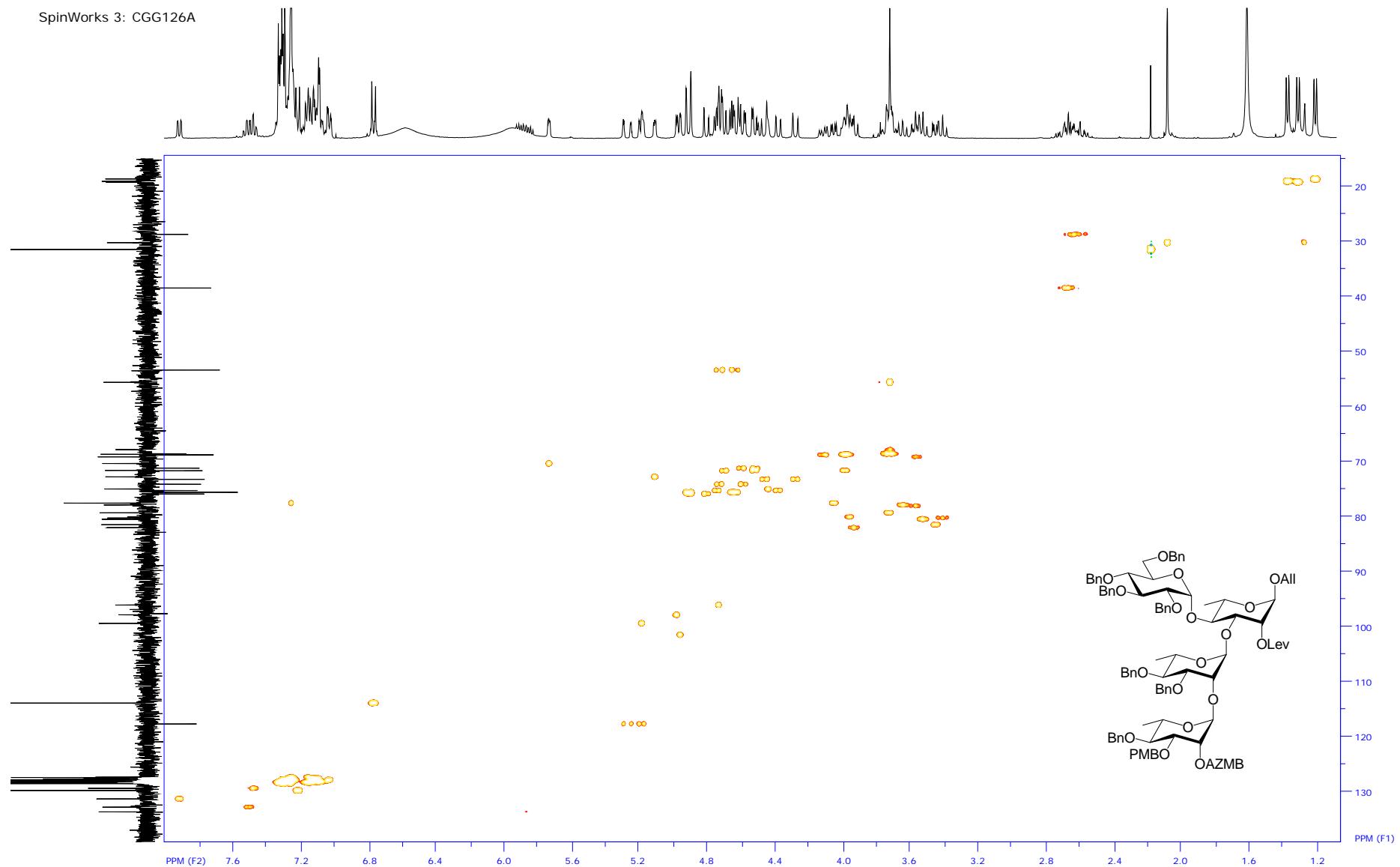


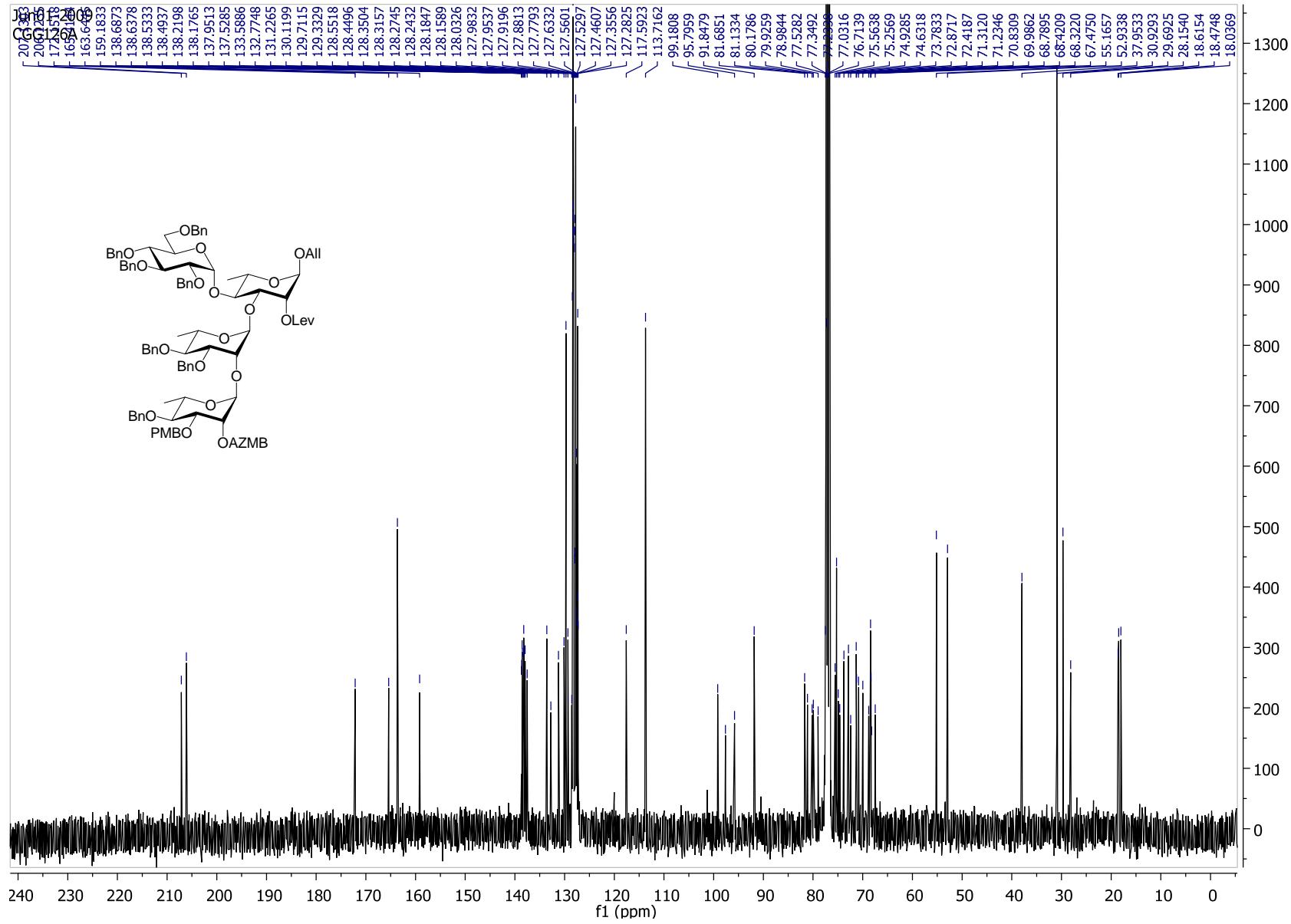
SpinWorks 3: CGG126A



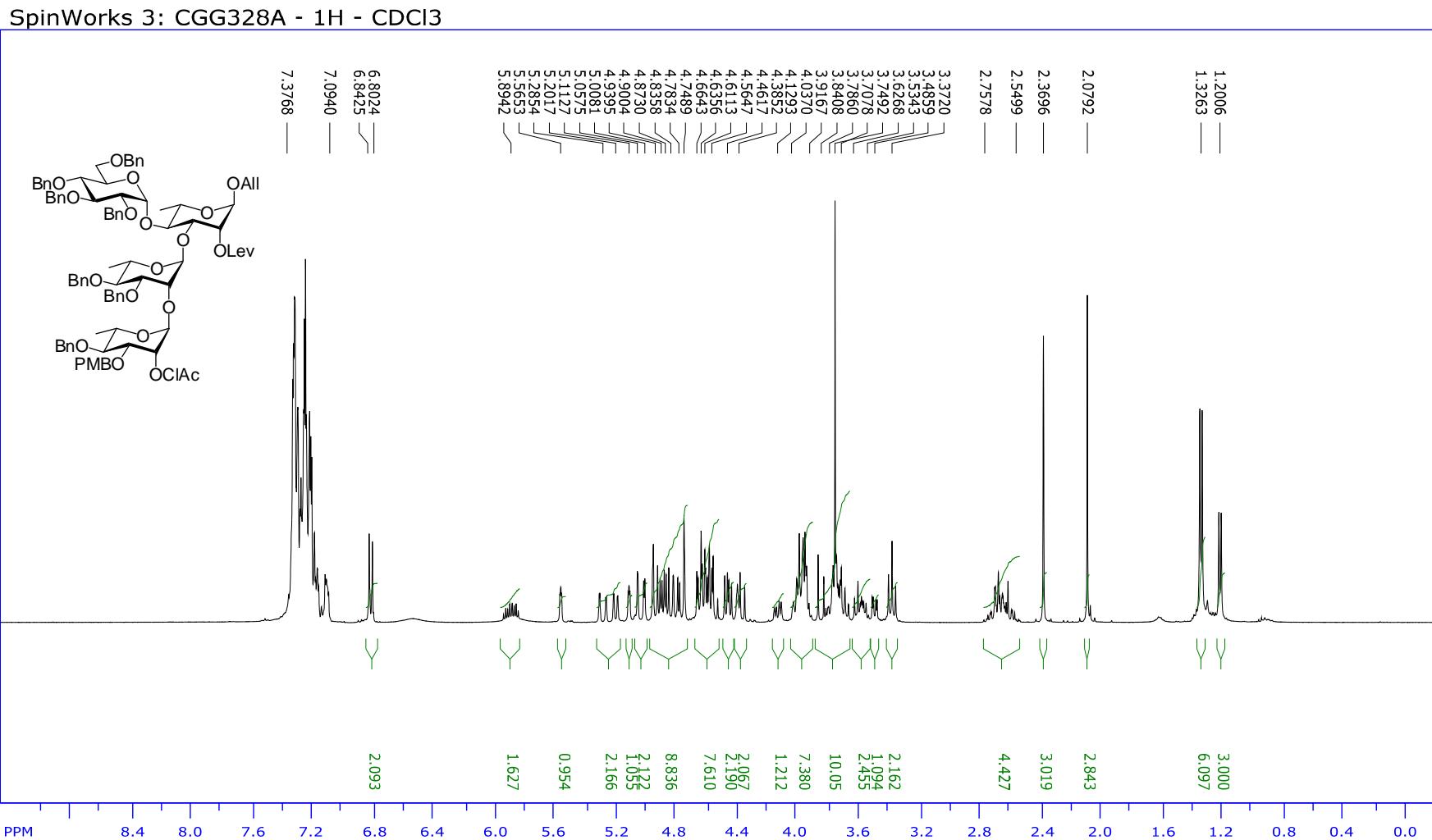
S120

SpinWorks 3: CGG126A

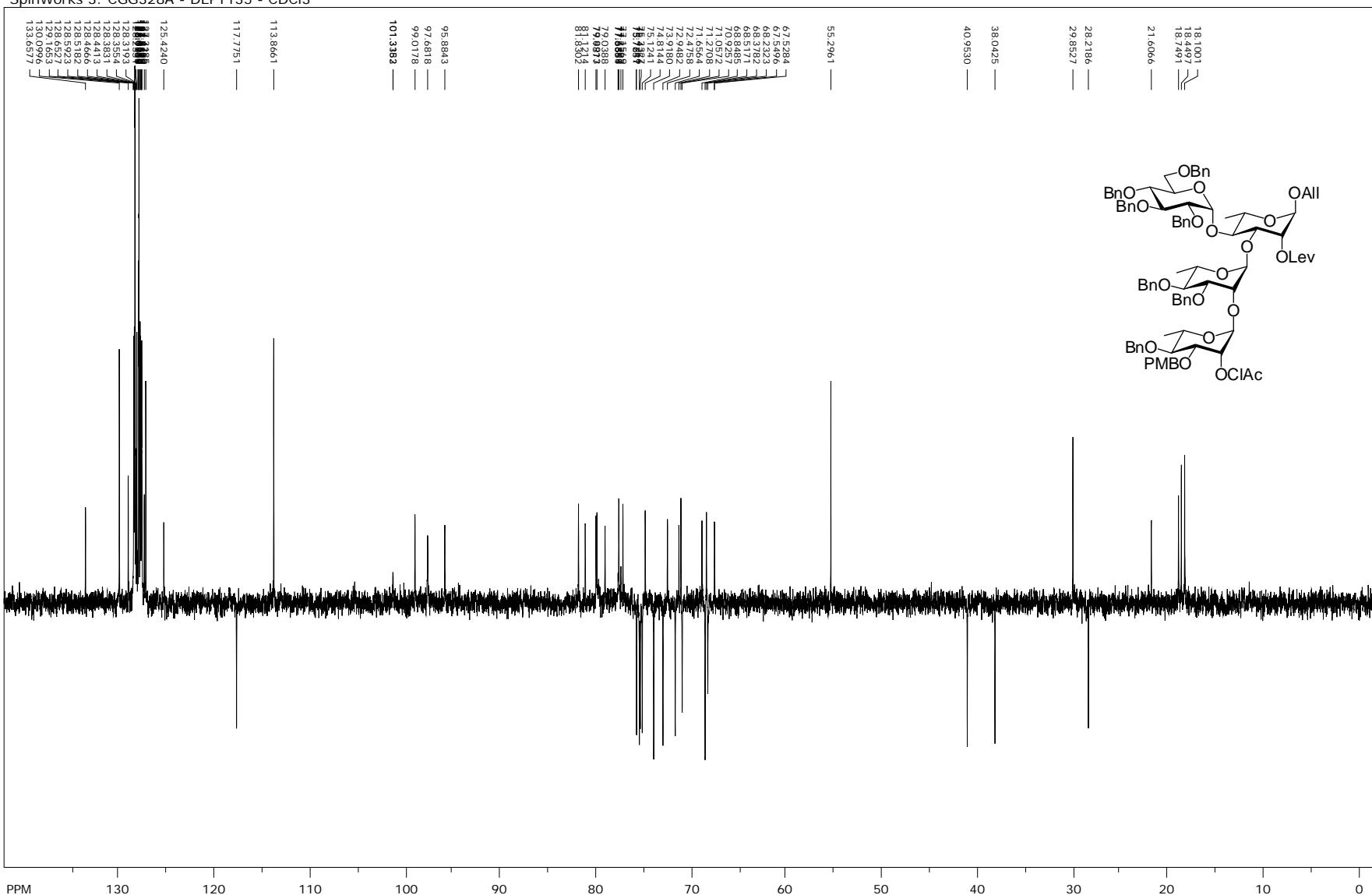




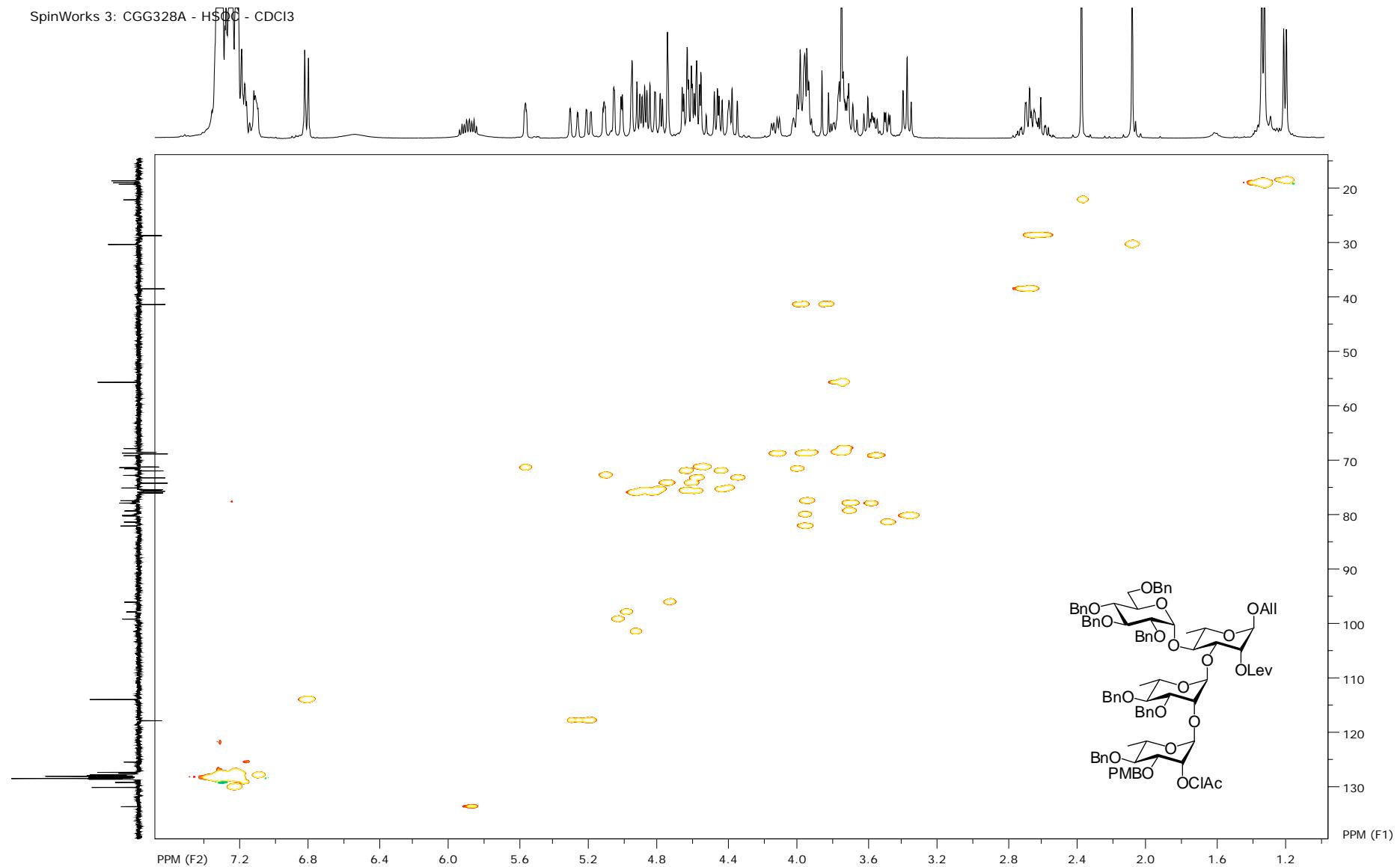
Compound 22



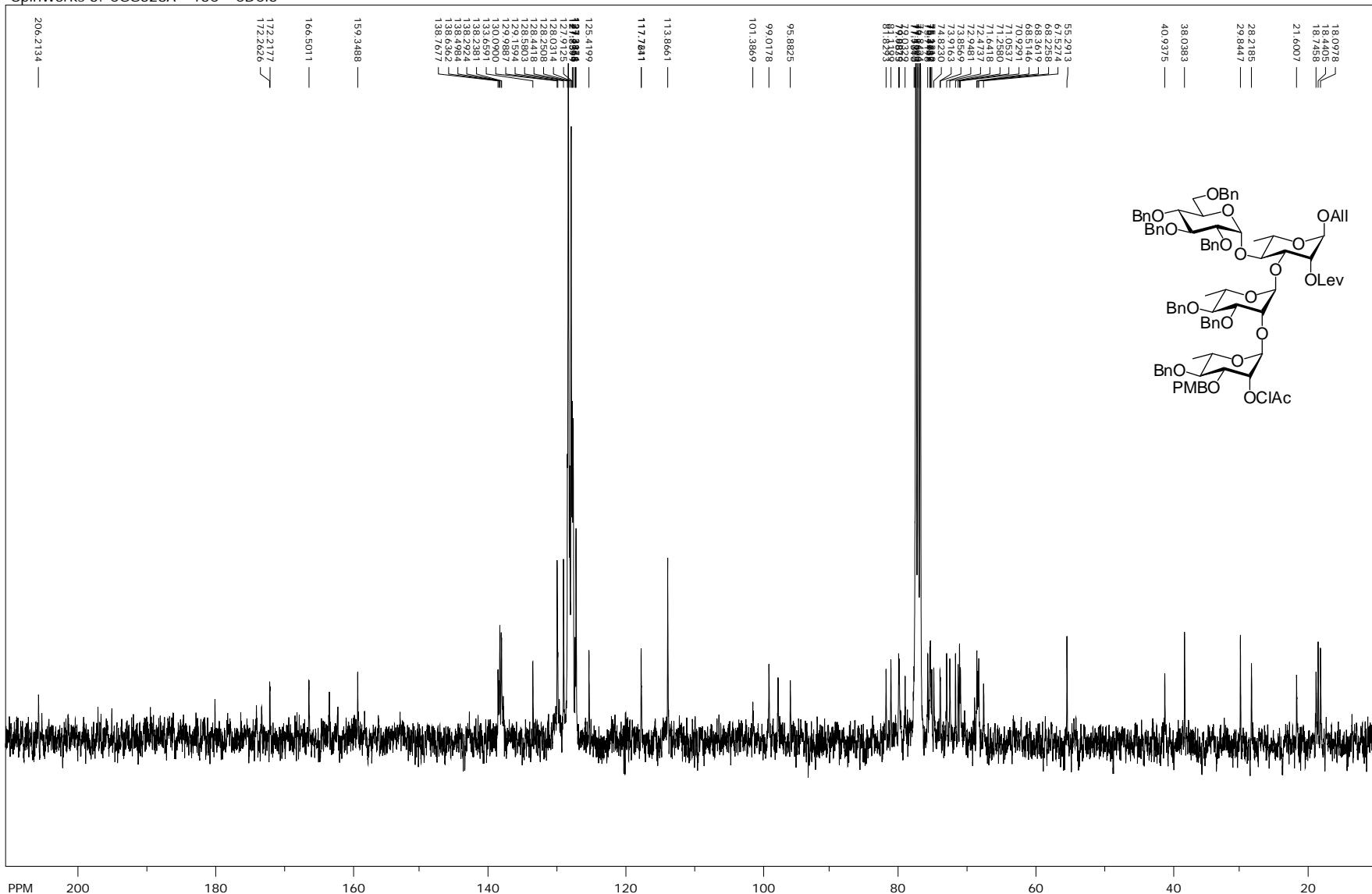
SpinWorks 3: CGG328A - DEPT135 - CDCl₃



SpinWorks 3: CGG328A - HSQC - CDCl₃

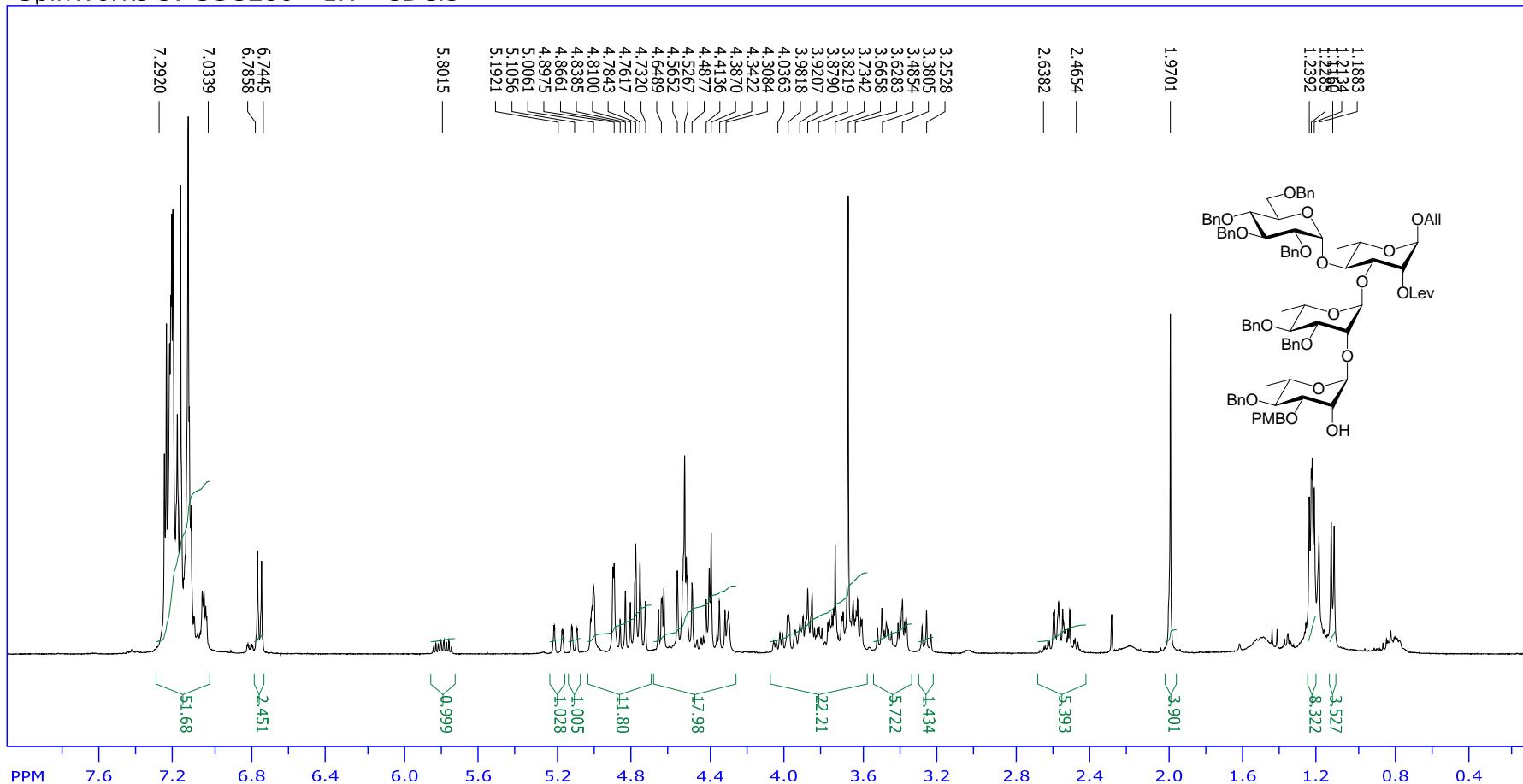


SpinWorks 3: CGG328A - 13C - CDCl3

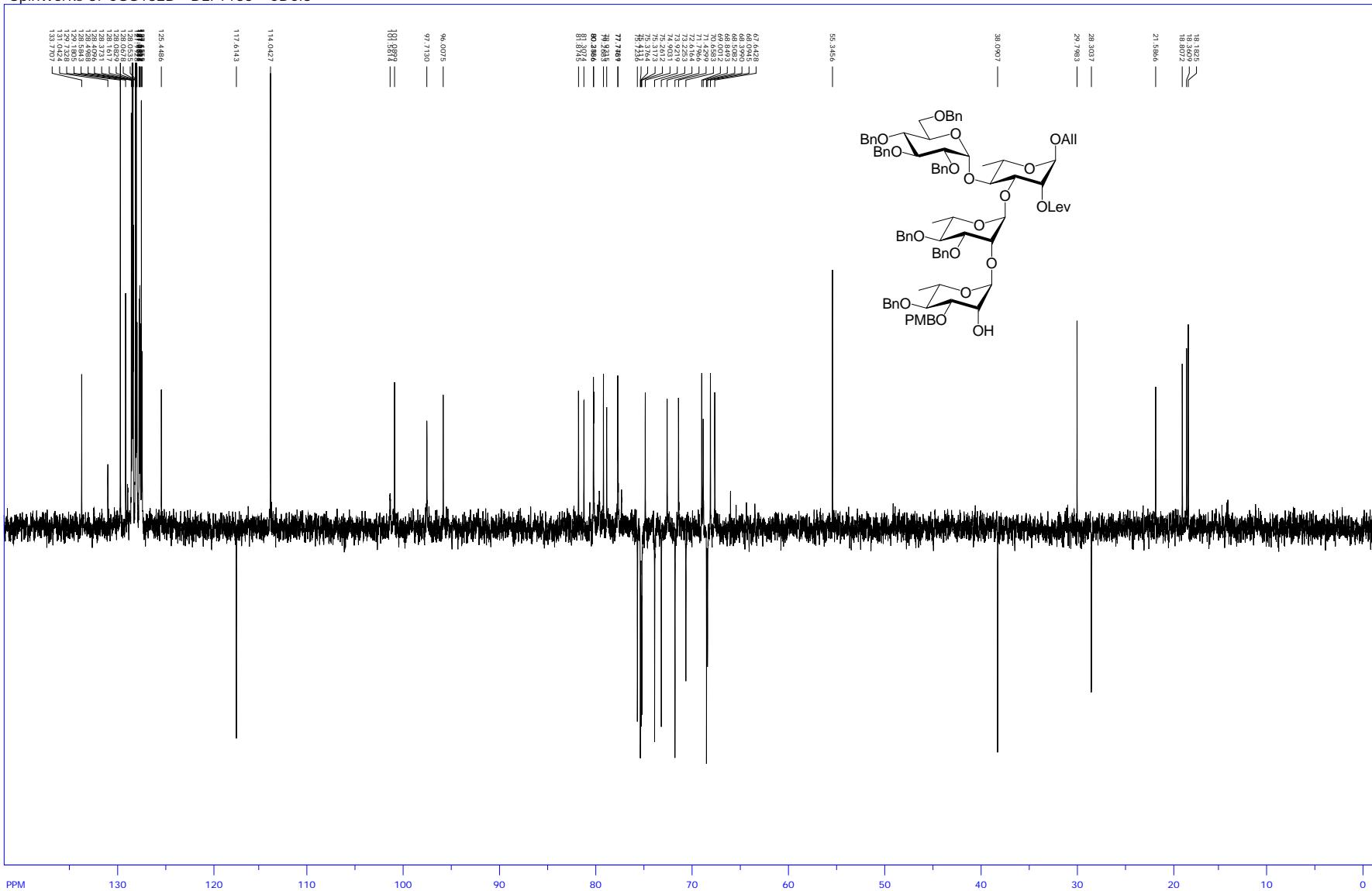


Compound 10

SpinWorks 3: CGG280 - 1H - CDCl₃

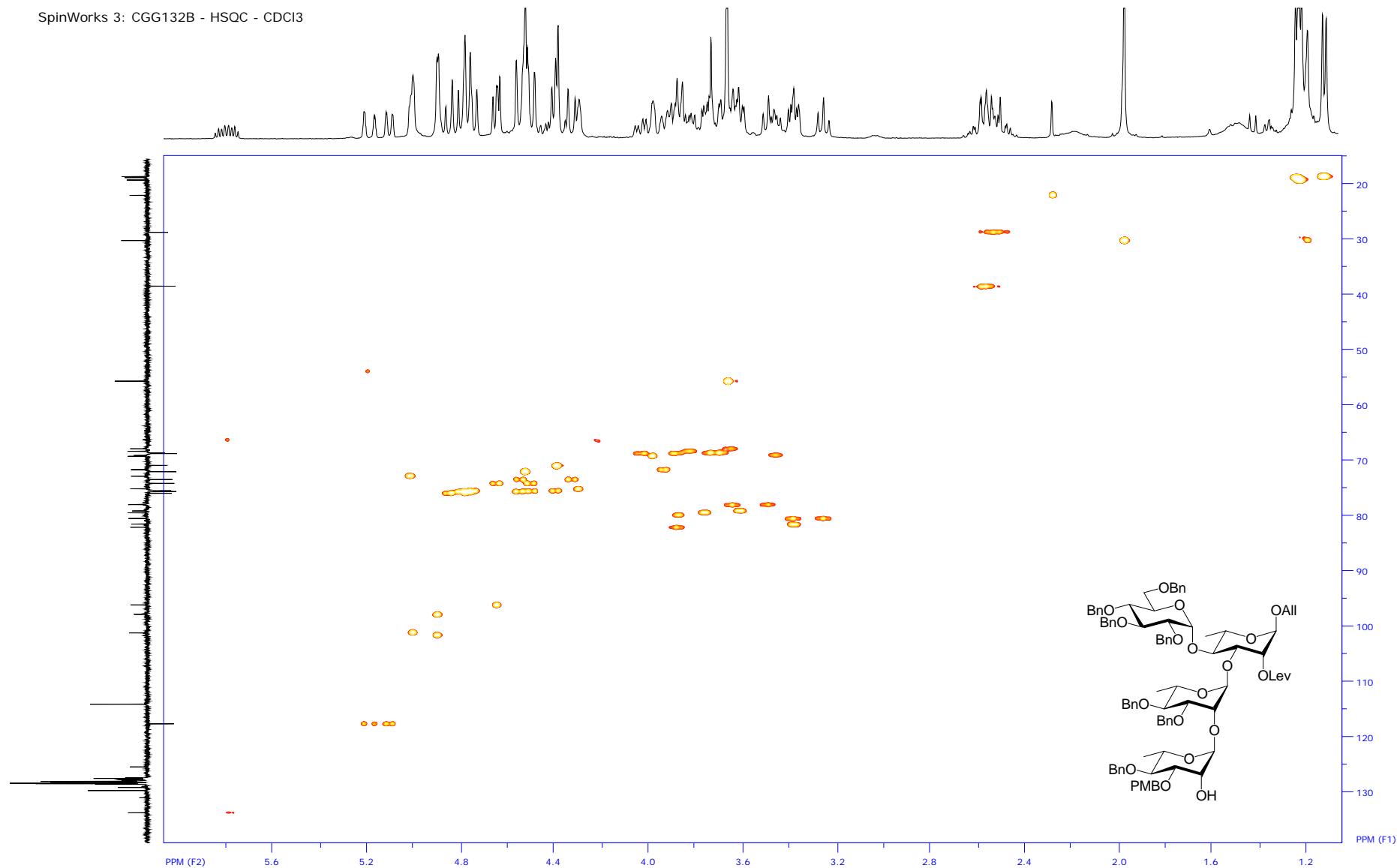


SpinWorks 3: CGG132B - DEPT135 - CDCI3

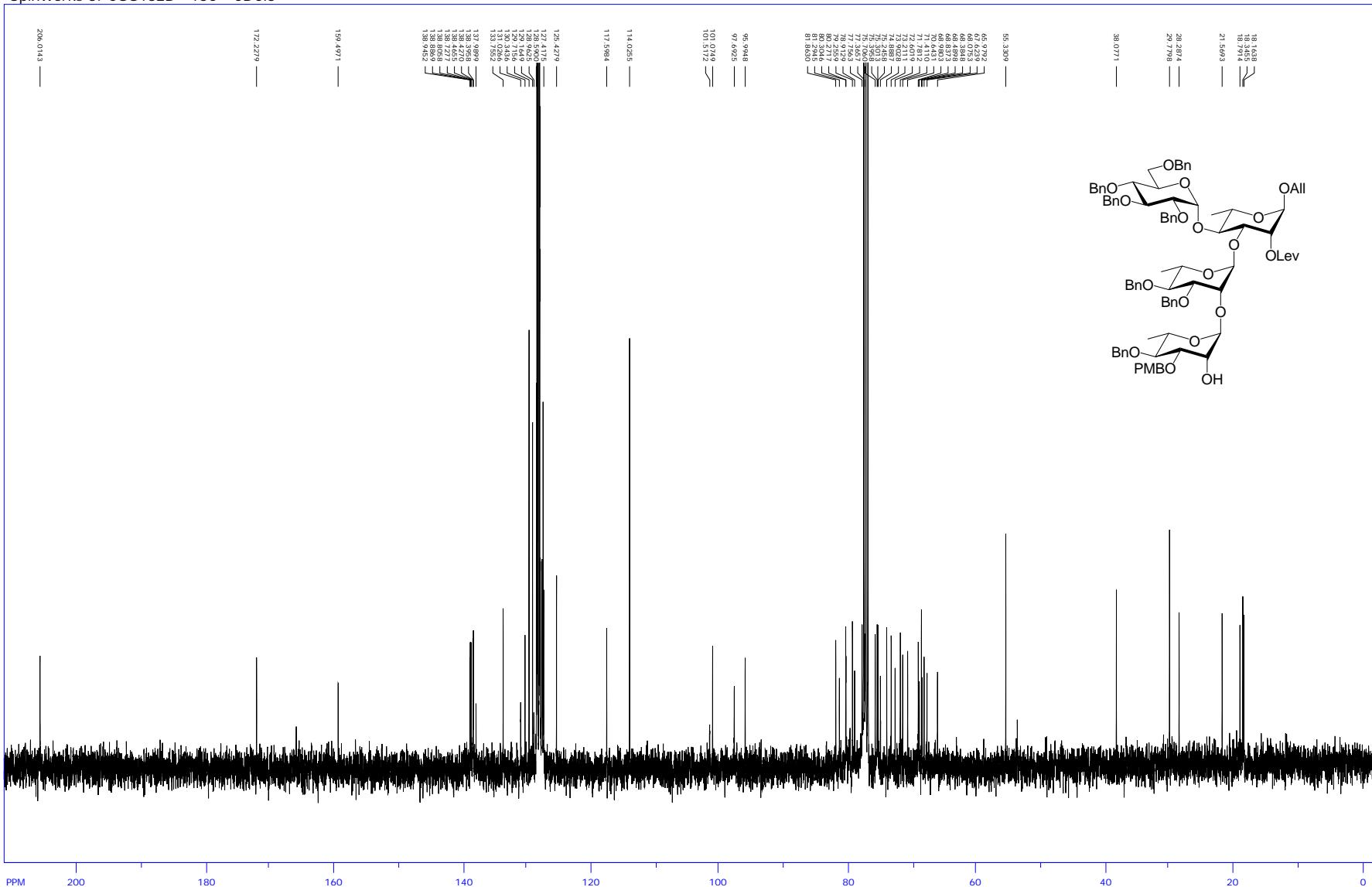


S128

SpinWorks 3: CGG132B - HSQC - CDCl₃

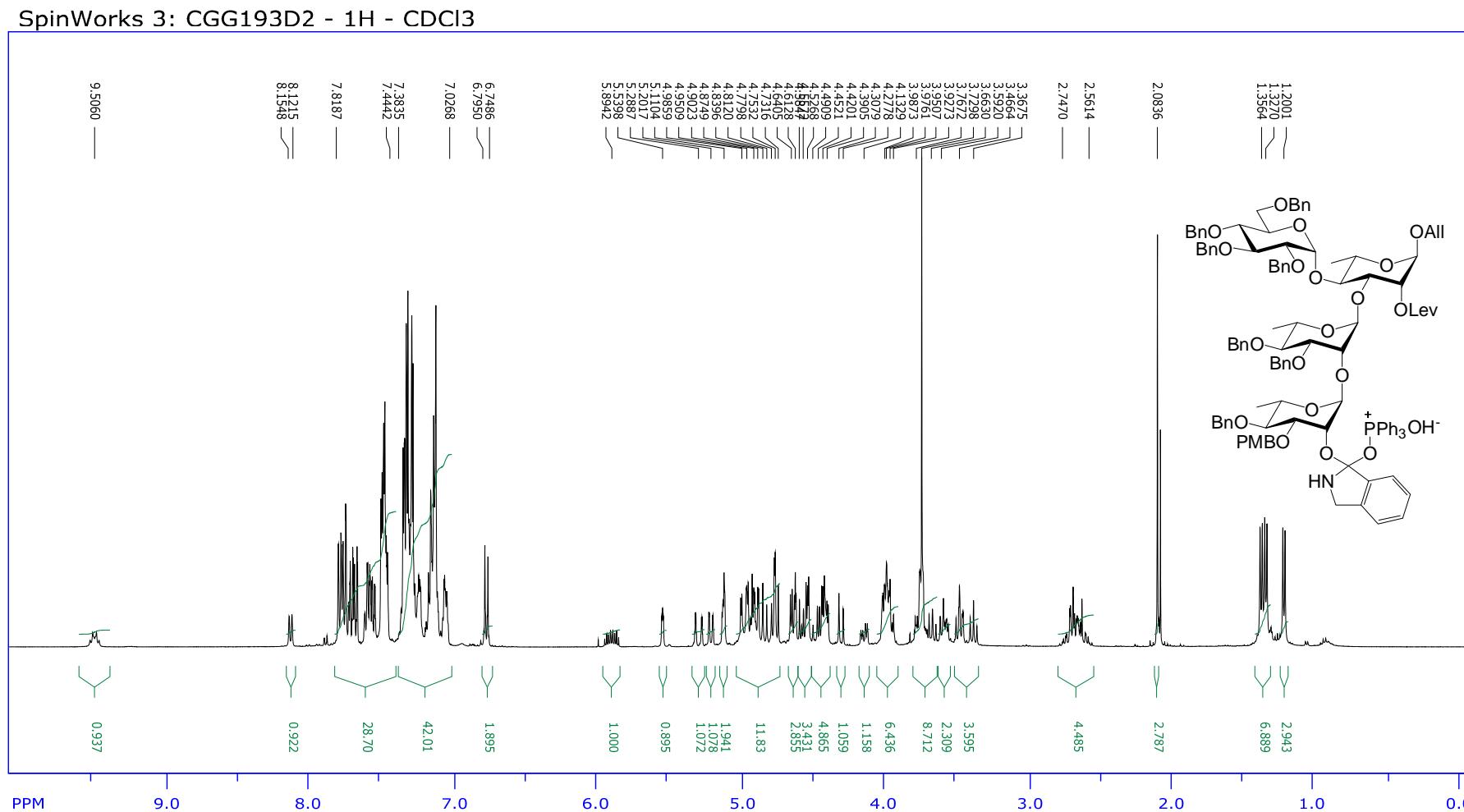


SpinWorks 3: CGG132B - 13C - CDCl₃

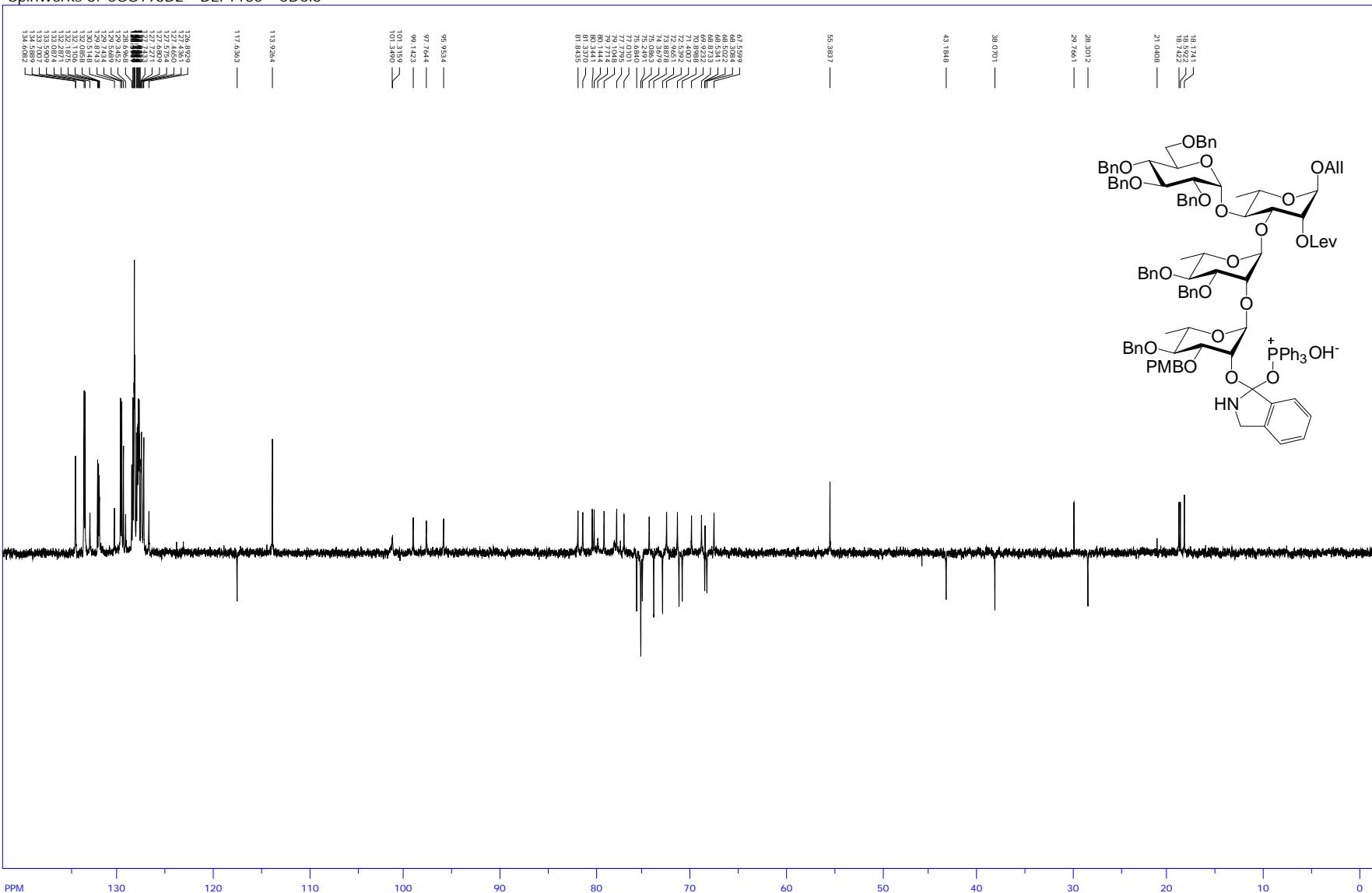


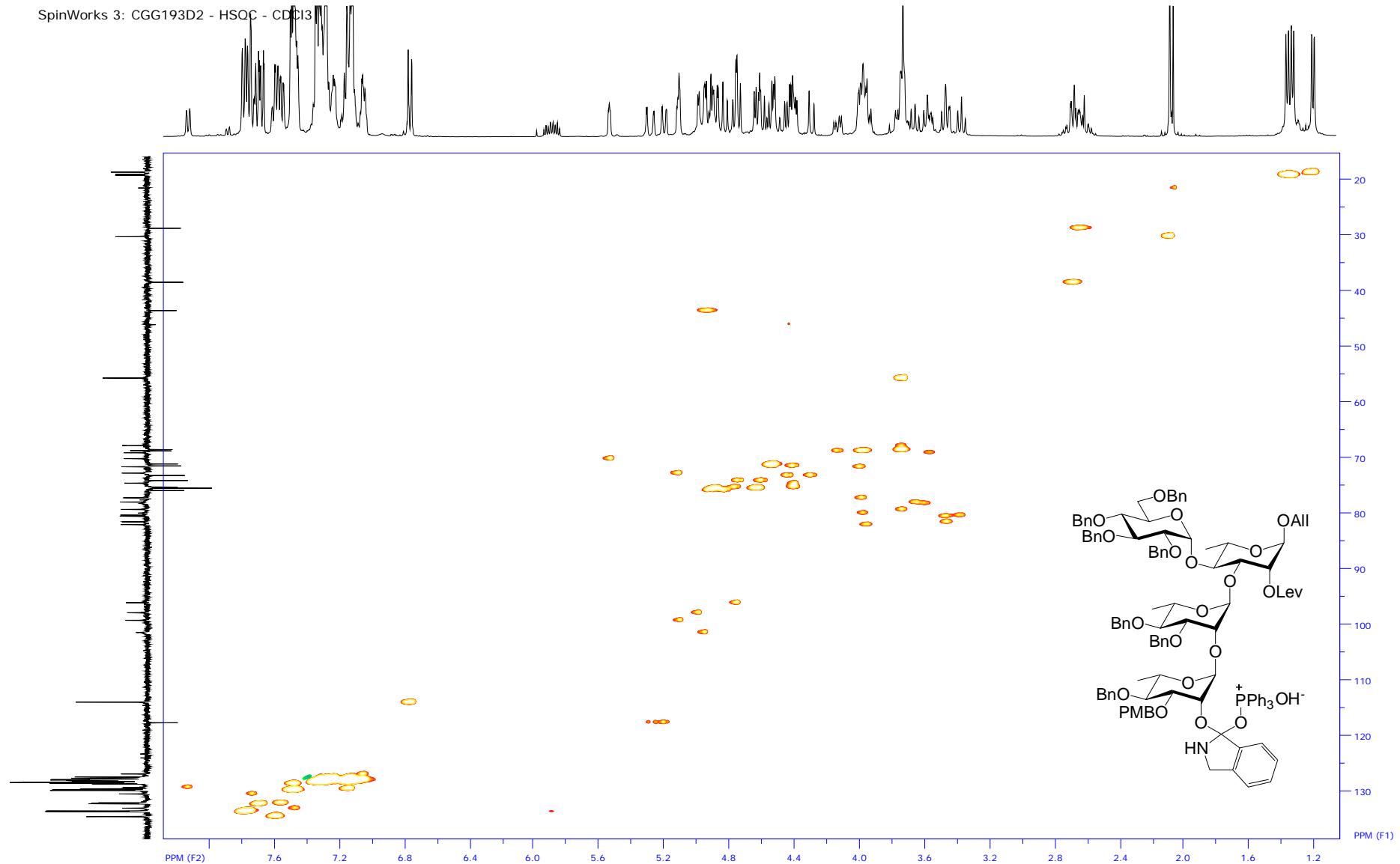
S130

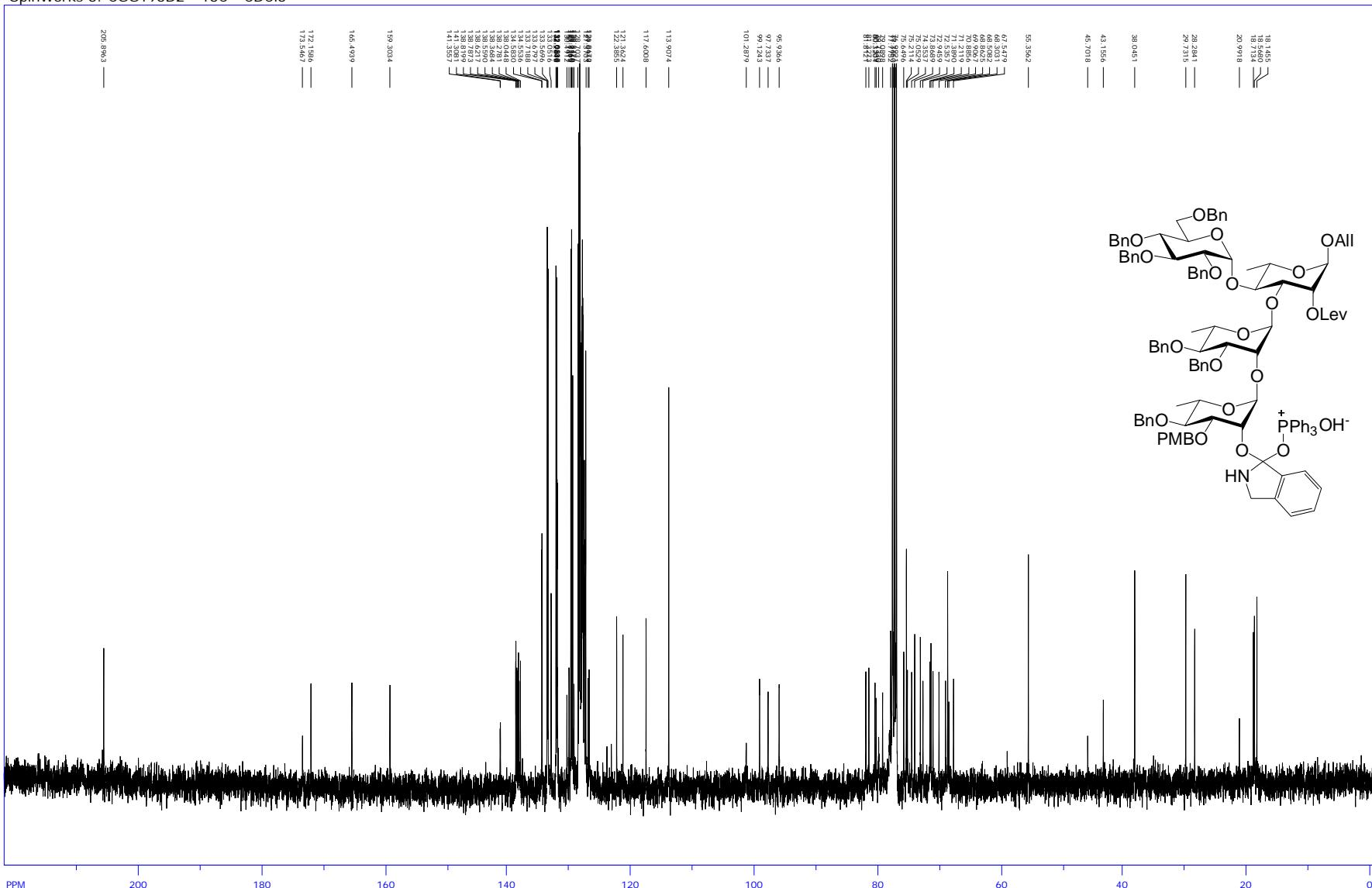
Compound 21a



SpinWorks 3: CGG193D2 - DEPT135 - CDCl₃

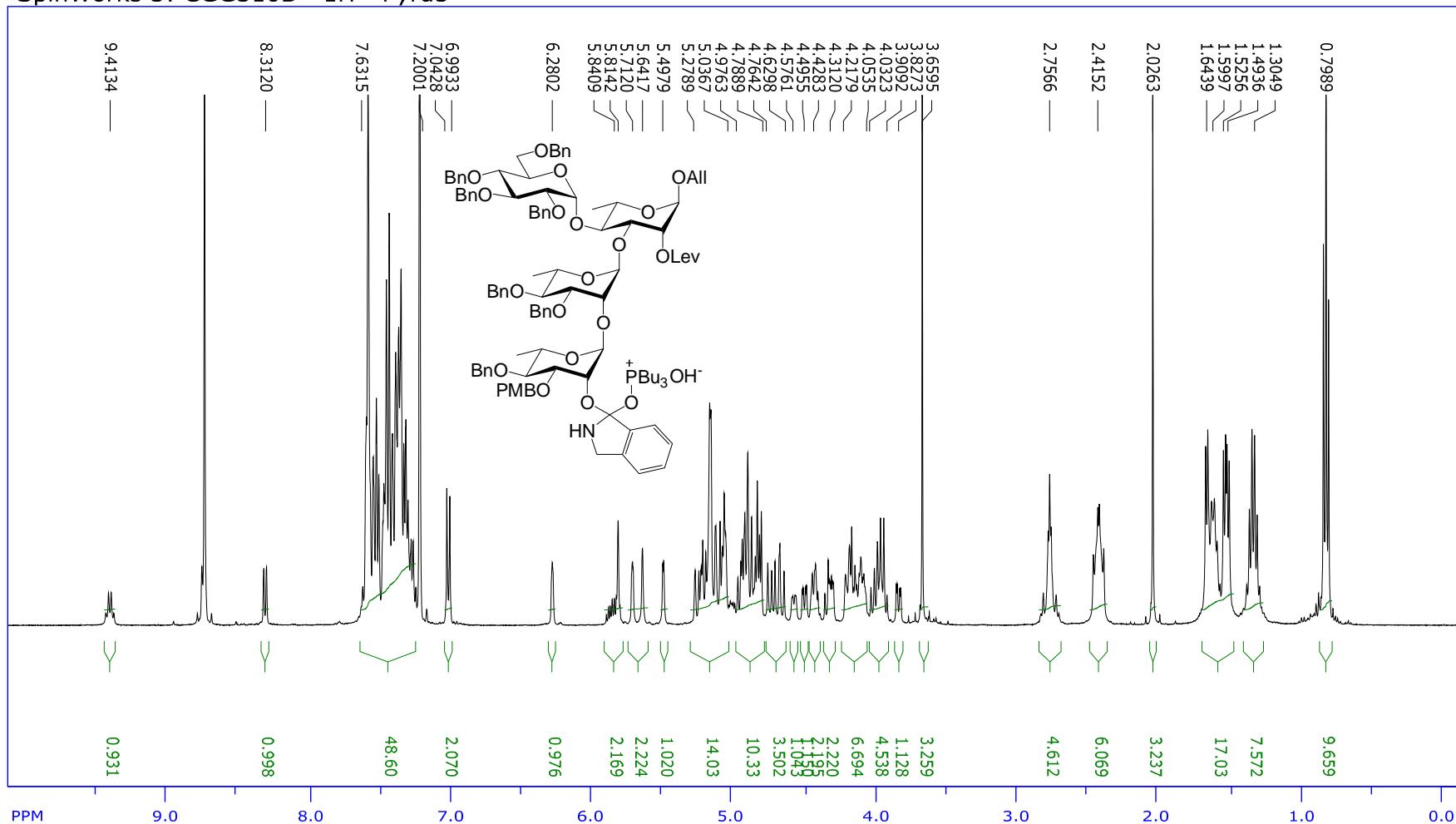




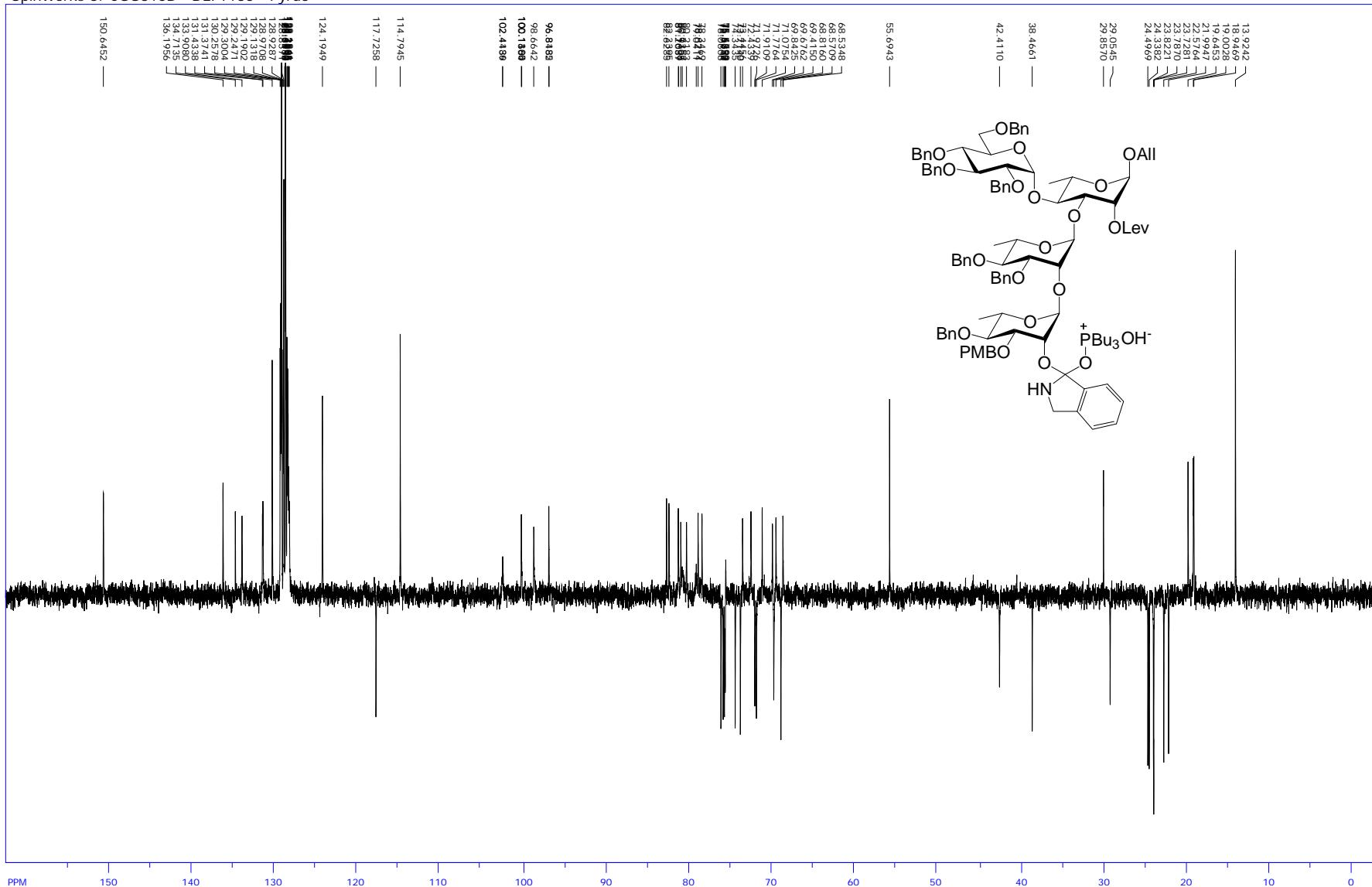
SpinWorks 3: CGG193D2 - 13C - CDCl₃

Compound 21b

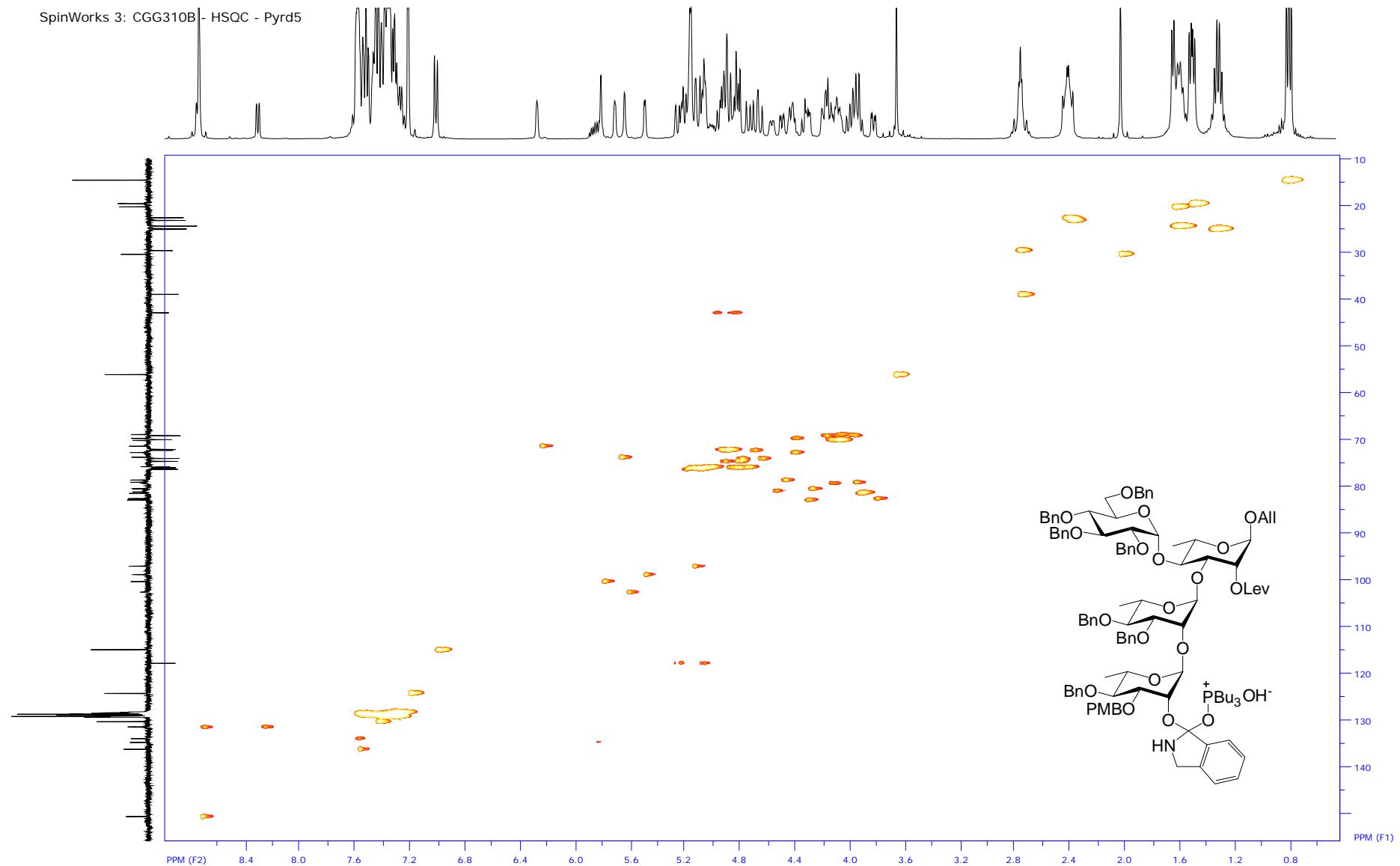
SpinWorks 3: CGG310B - 1H - Pyrd5



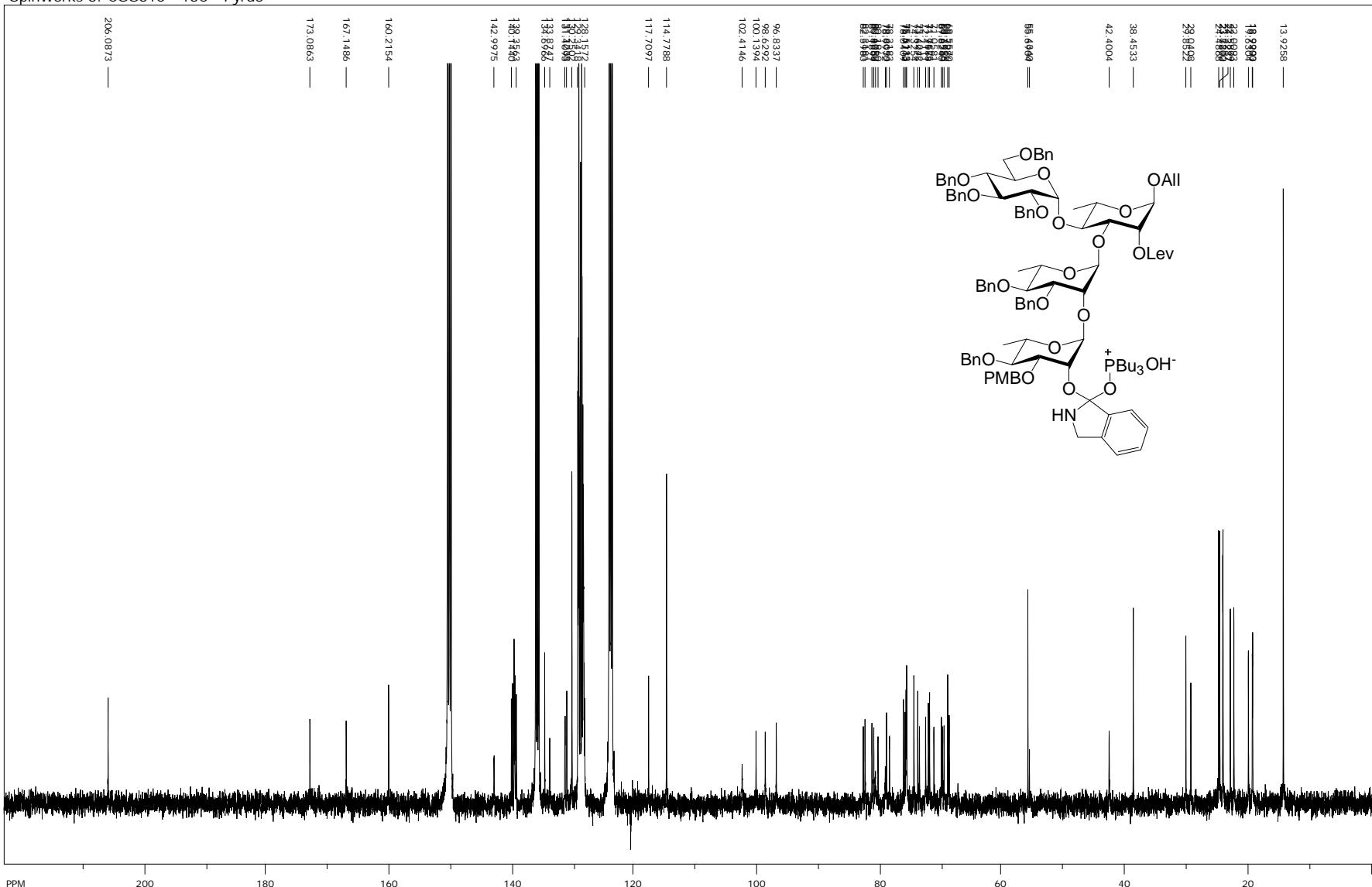
SpinWorks 3: CGG310B - DEPT135 - Pyrd5



SpinWorks 3: CGG310B - HSQC - Pyrd5

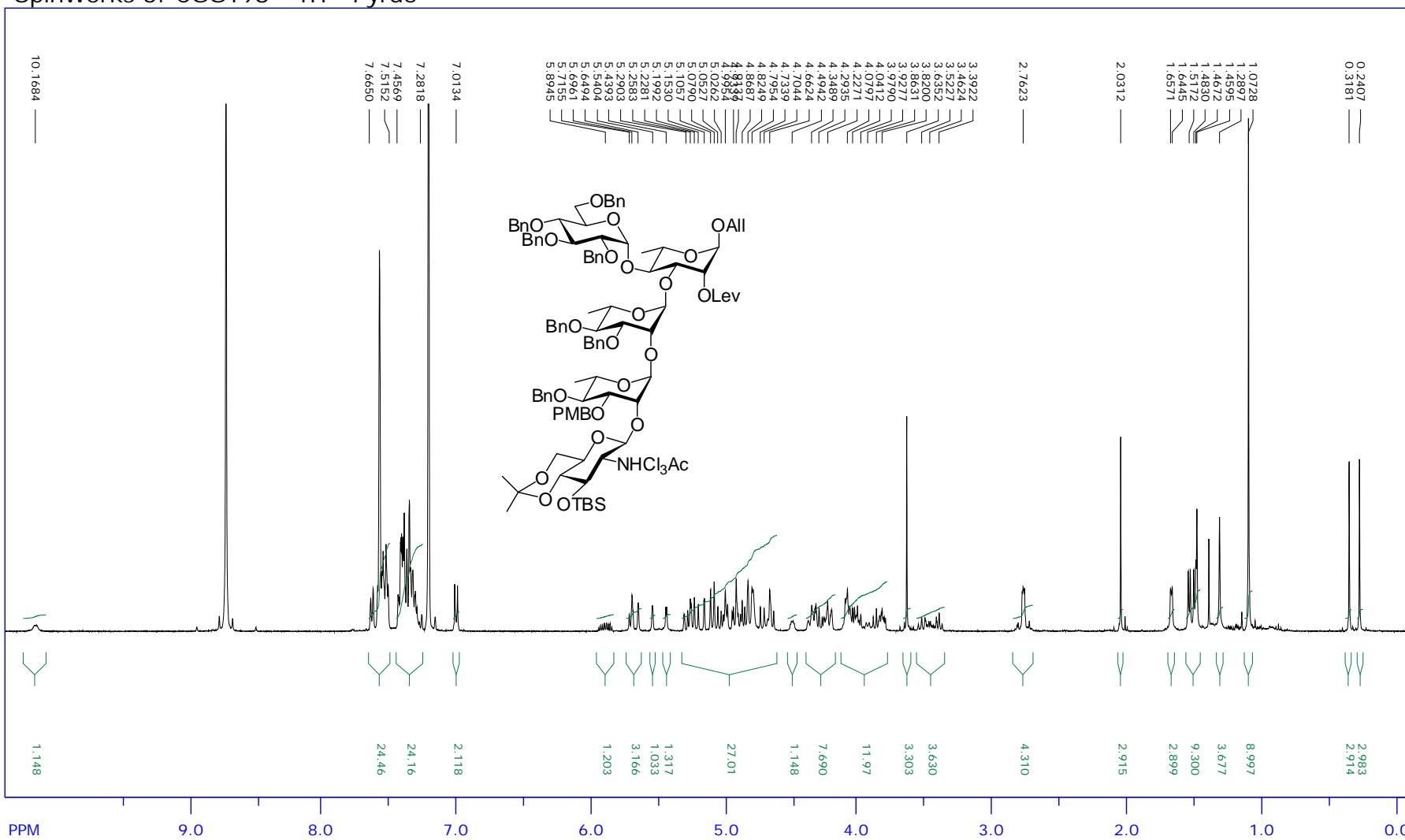


SpinWorks 3: CGG310 - 13C - Pyrd5

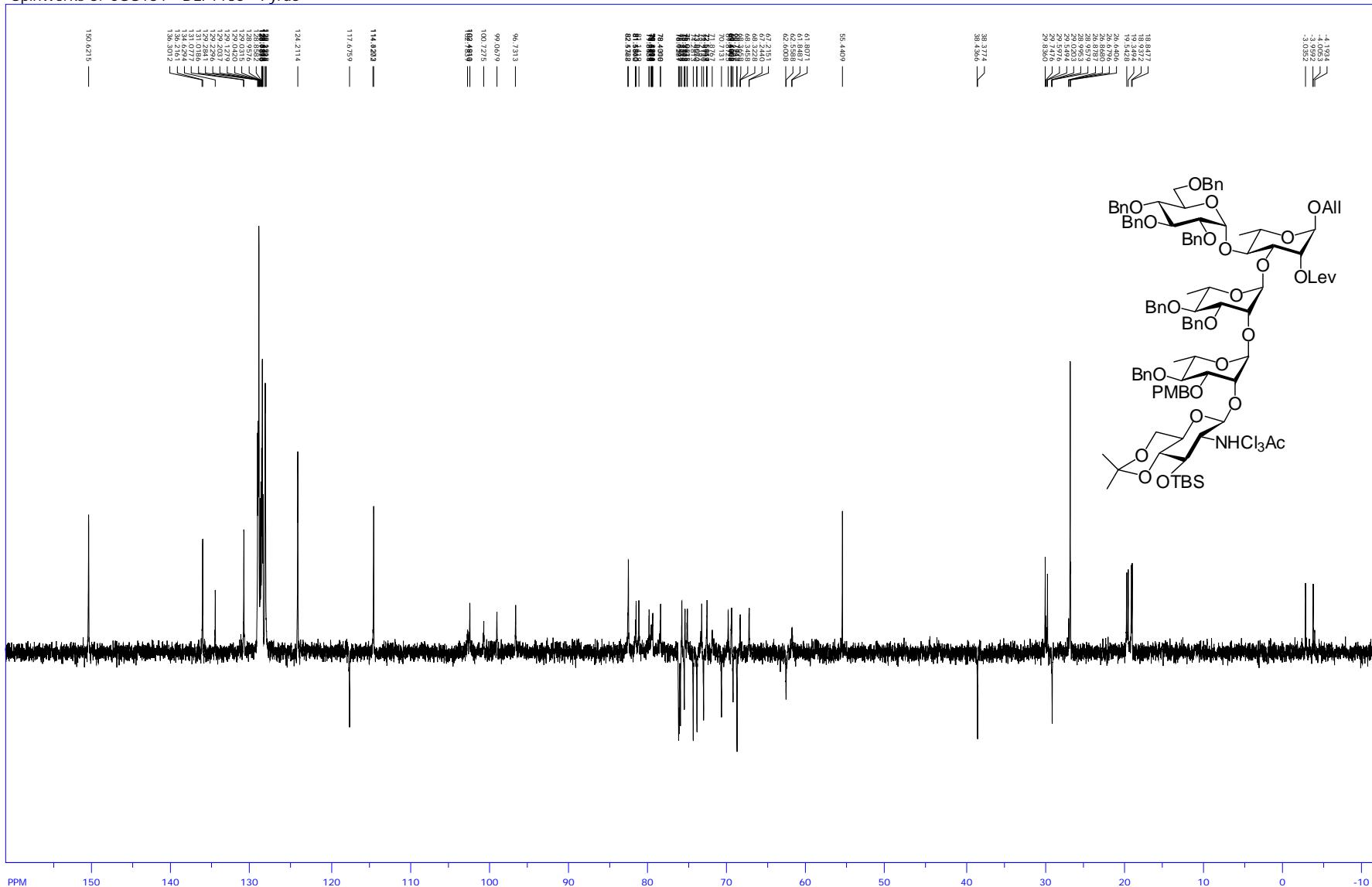


Compound 27

SpinWorks 3: CGG198 - 1H - Pyrd5

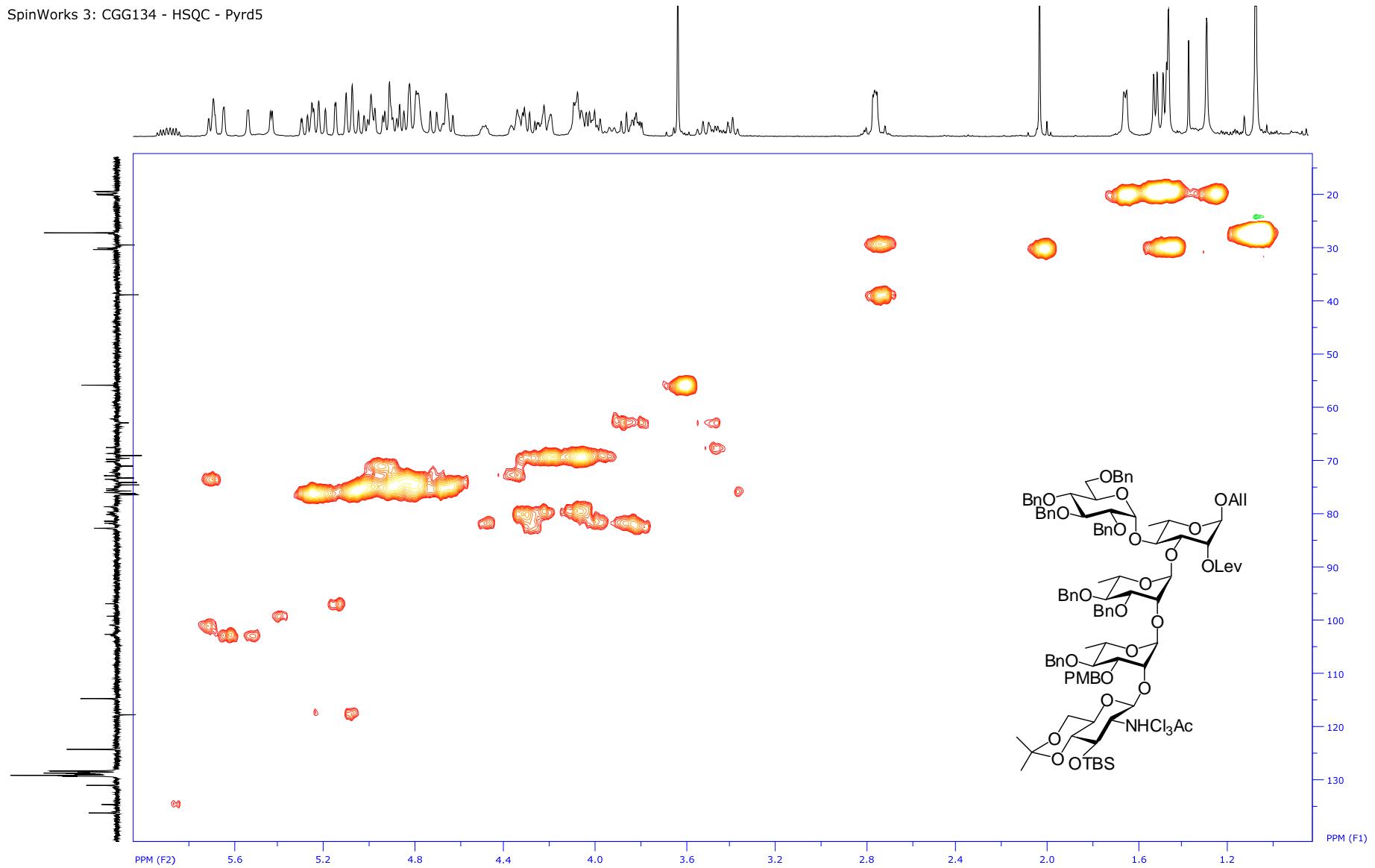


SpinWorks 3: CGG134 - DEPT135 - Pyrd5

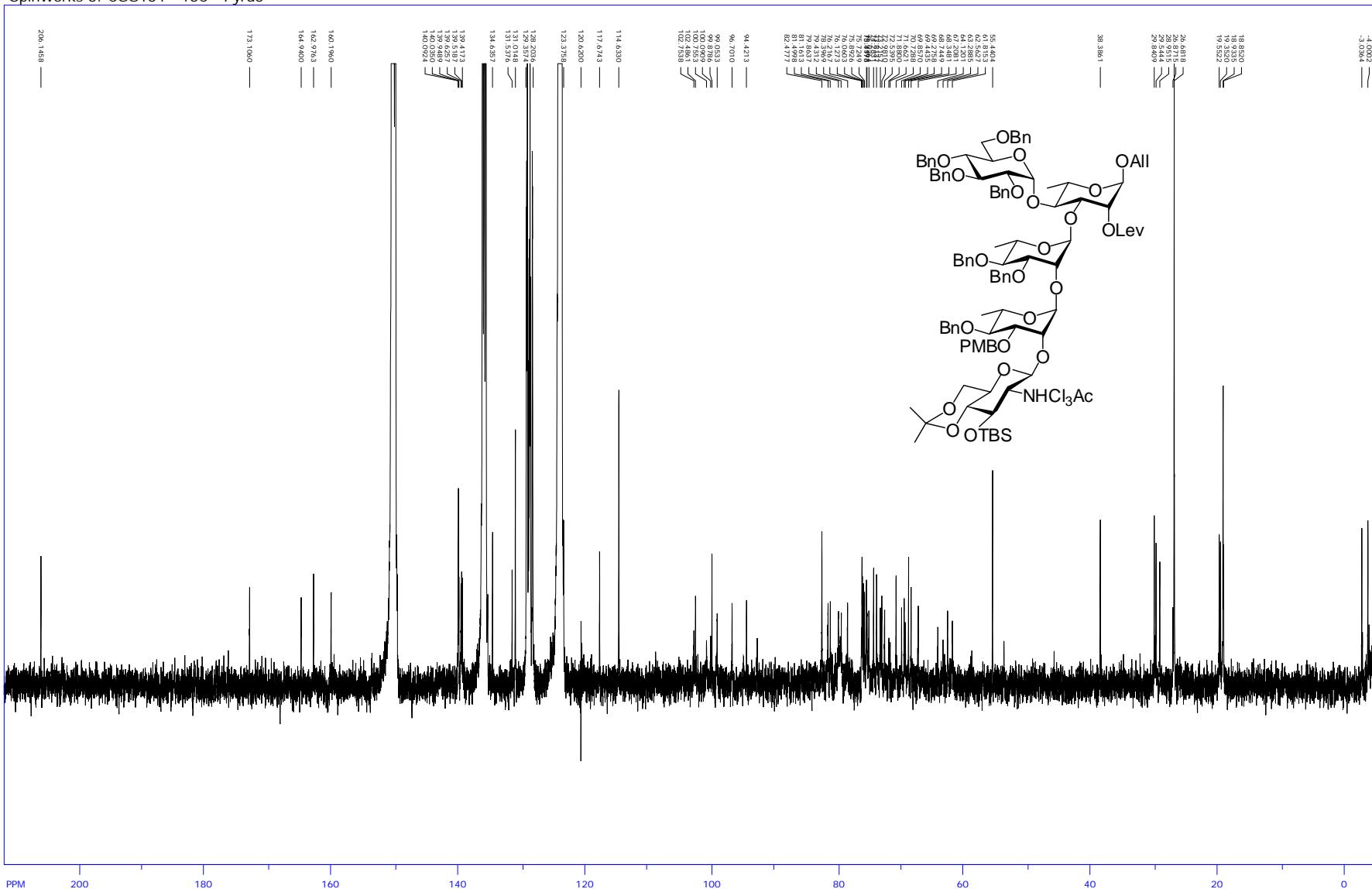


S140

SpinWorks 3: CGG134 - HSQC - Pyrd5

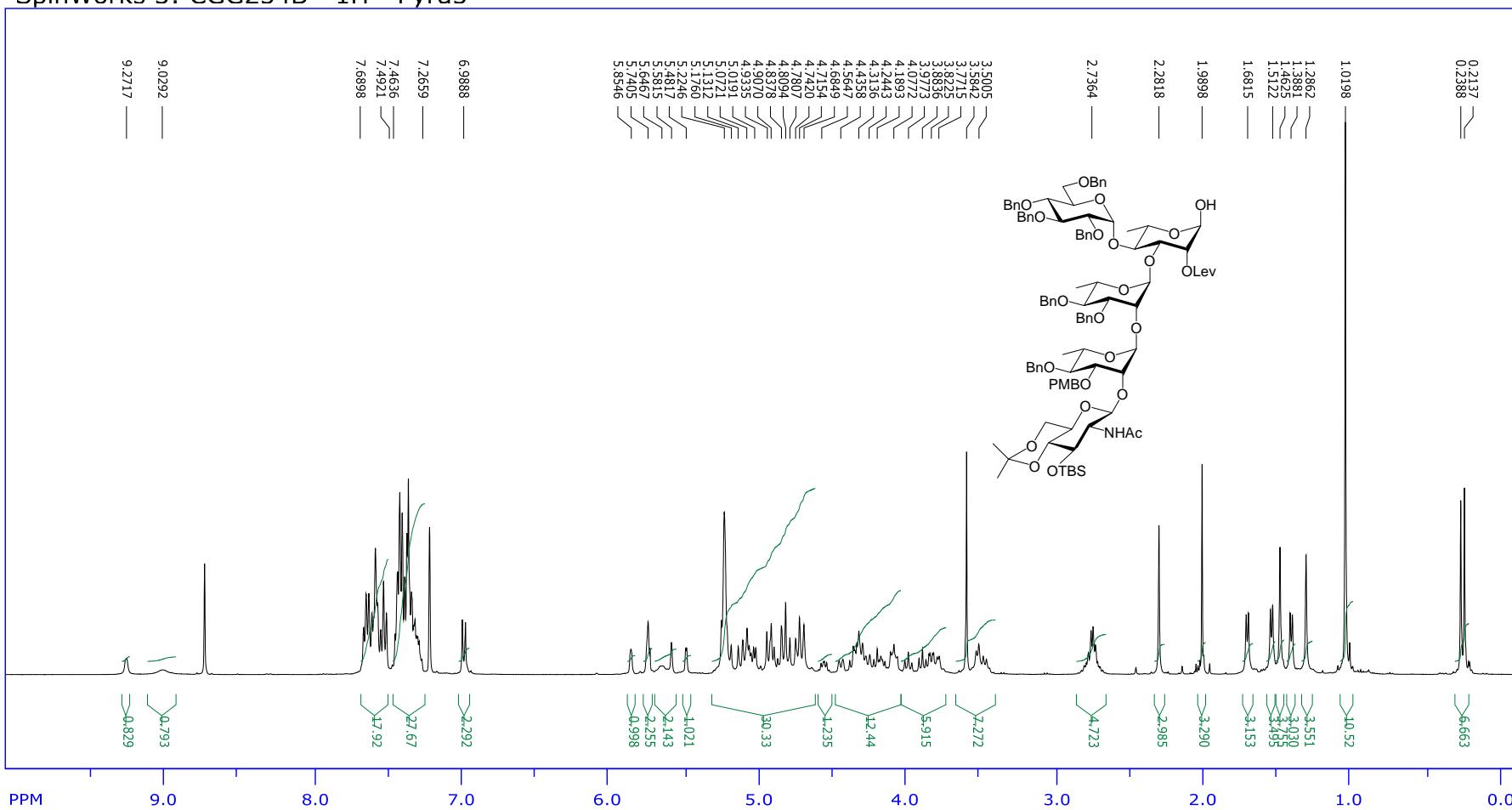


SpinWorks 3: CGG134 - 13C - Pyrd5

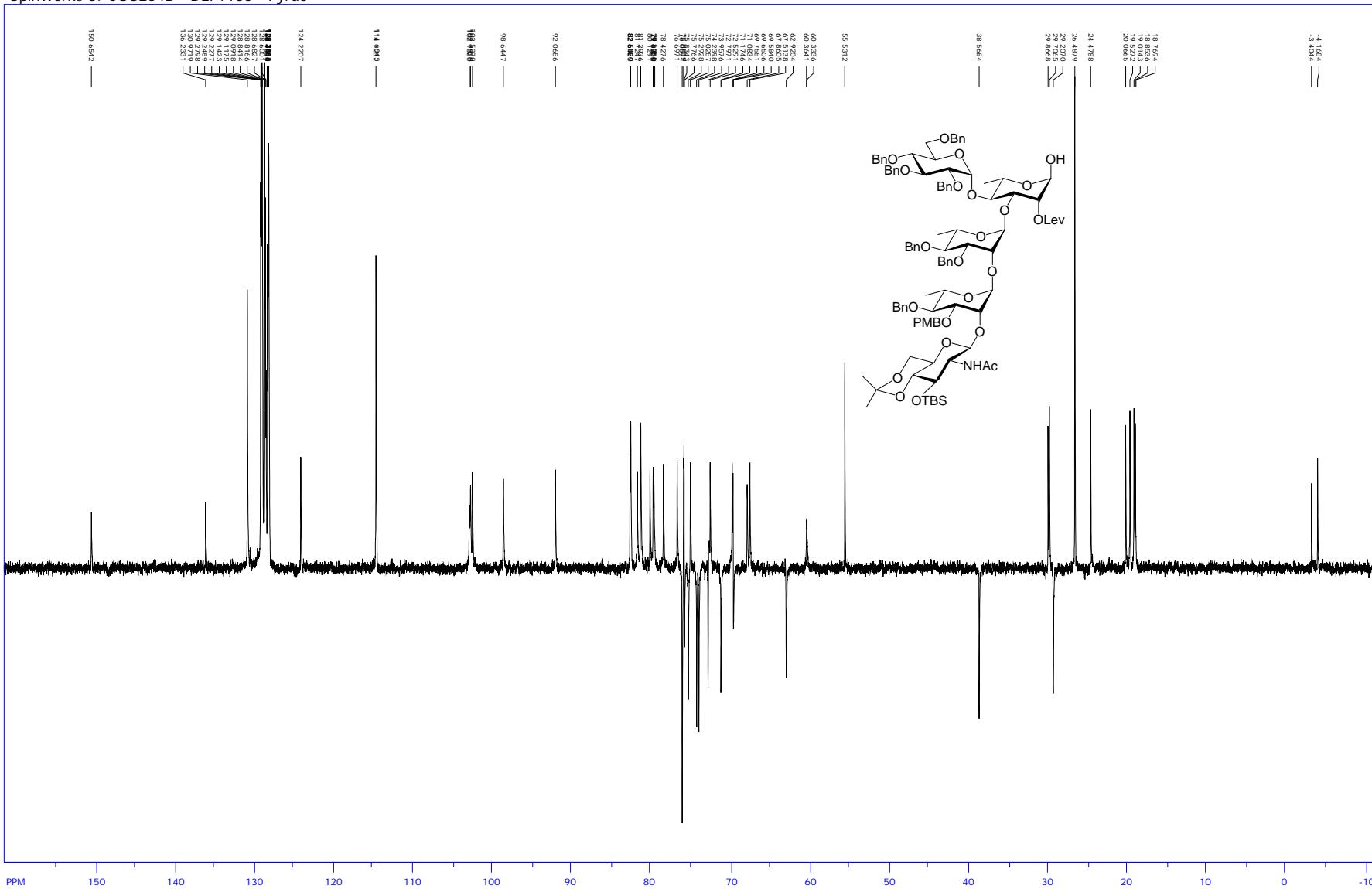


Compound 29

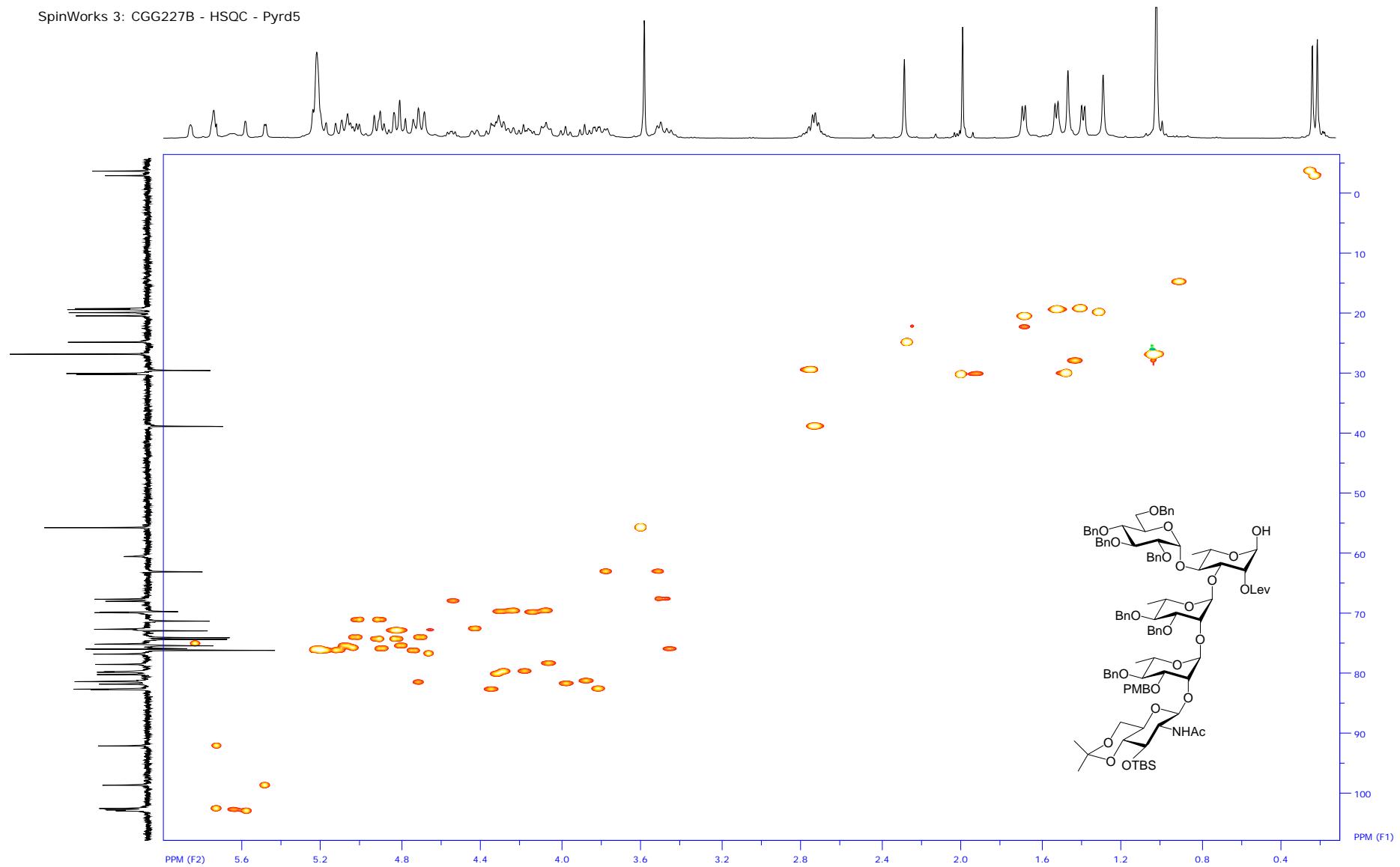
SpinWorks 3: CGG254B - 1H - Pyrd5



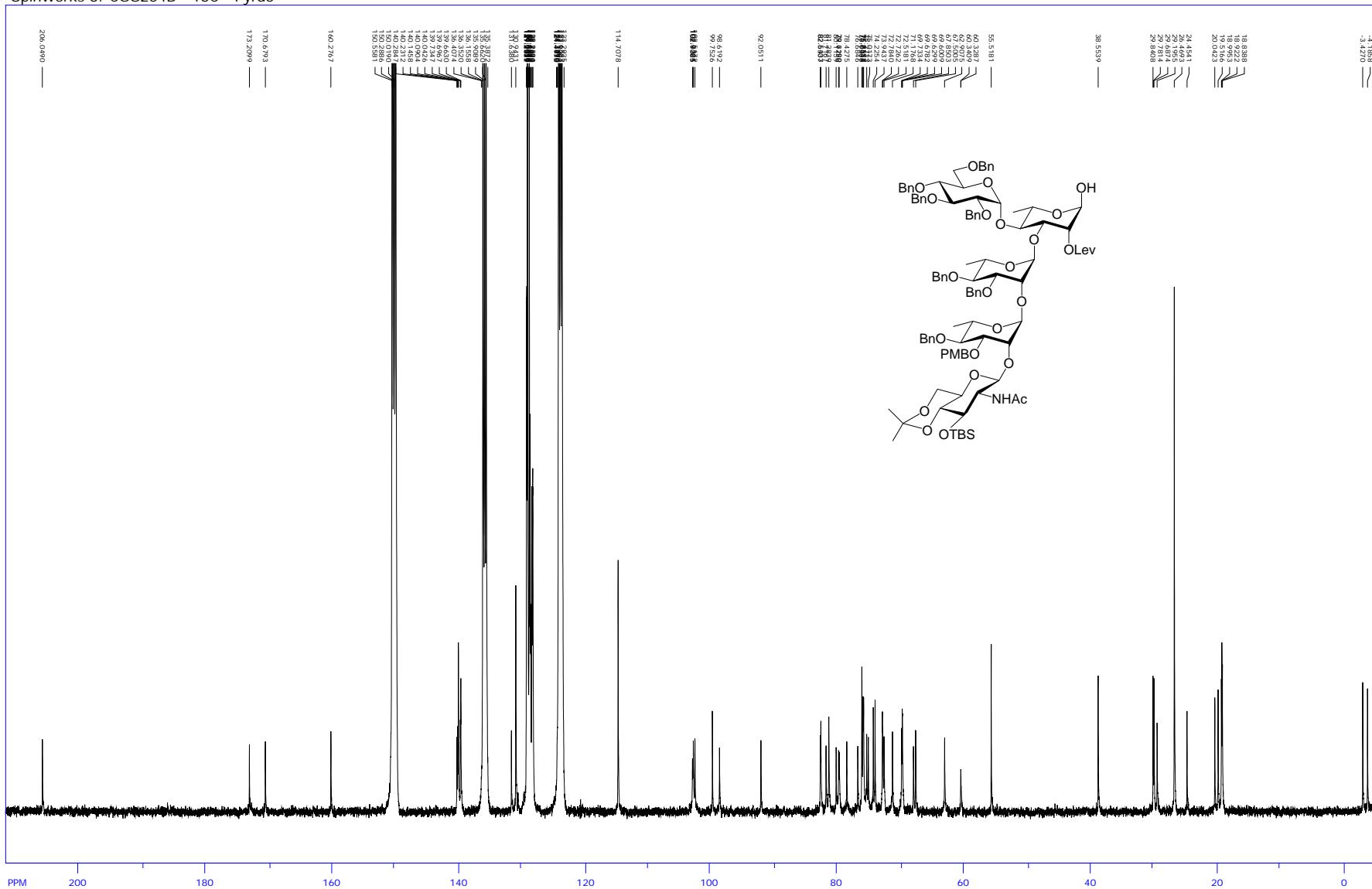
SpinWorks 3: CGG254B - DEPT135 - Pyrd5



SpinWorks 3: CGG227B - HSQC - Pyrd5

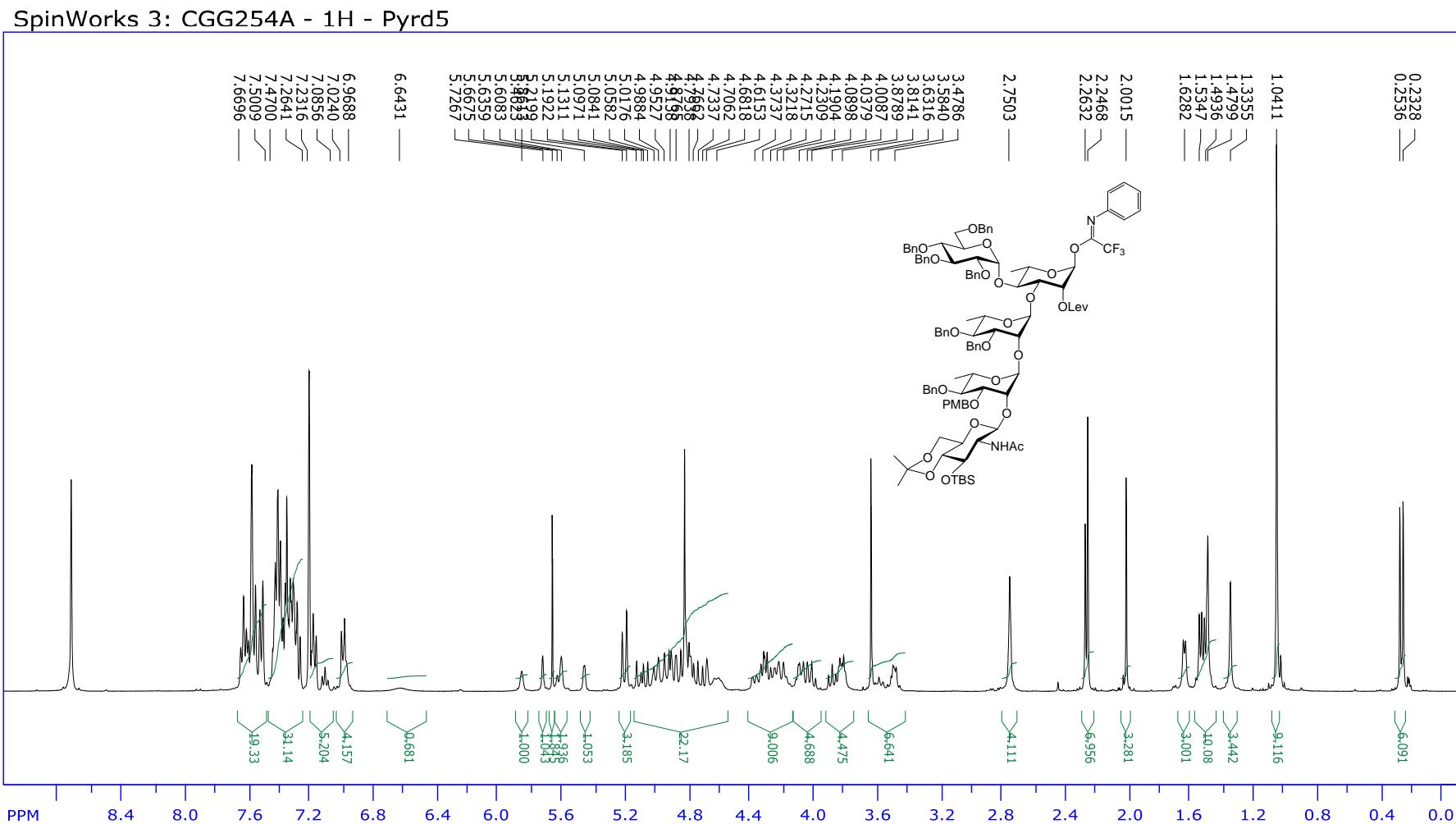


SpinWorks 3: CGG254B - 13C - Pyrd5

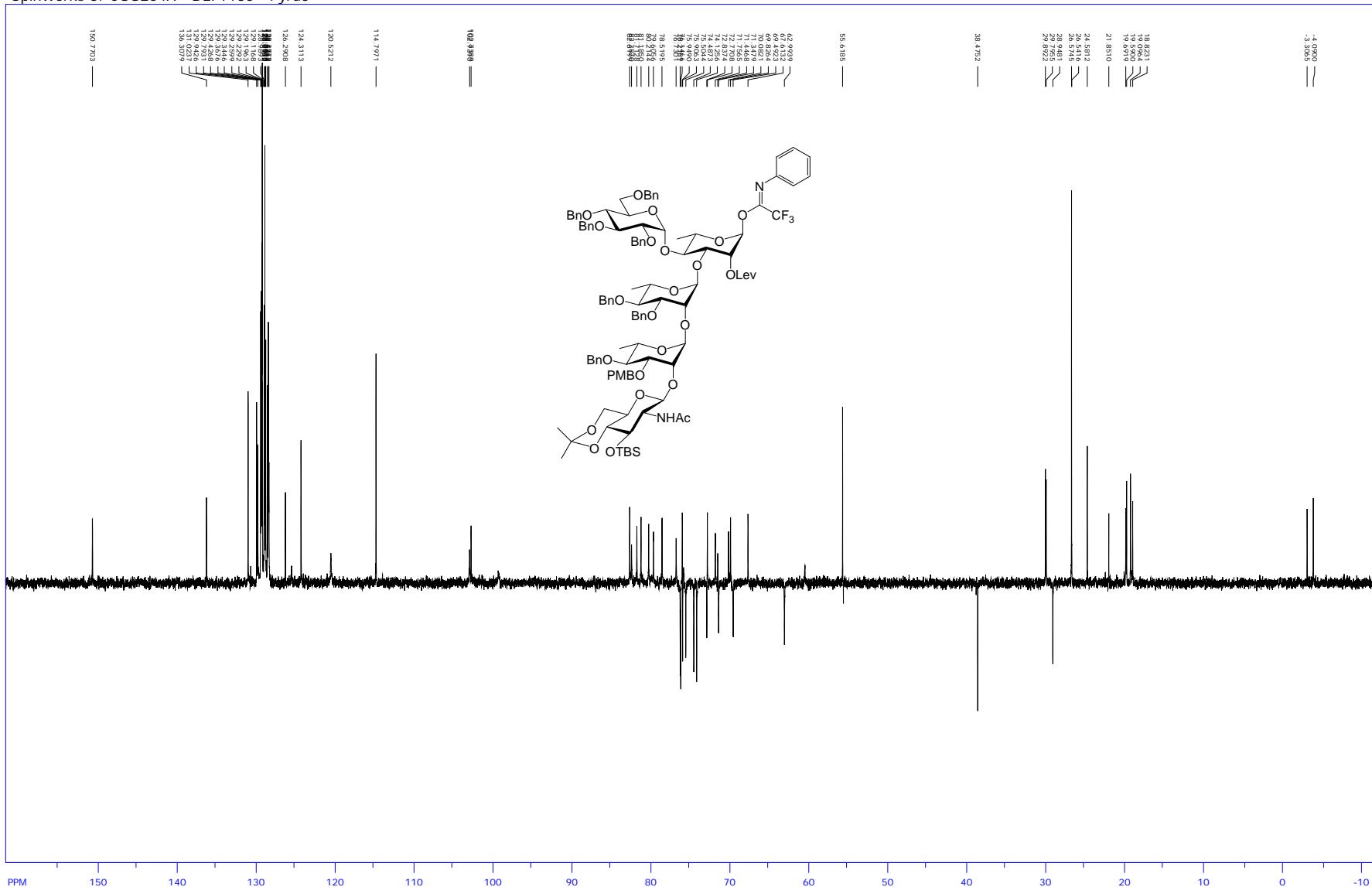


S146

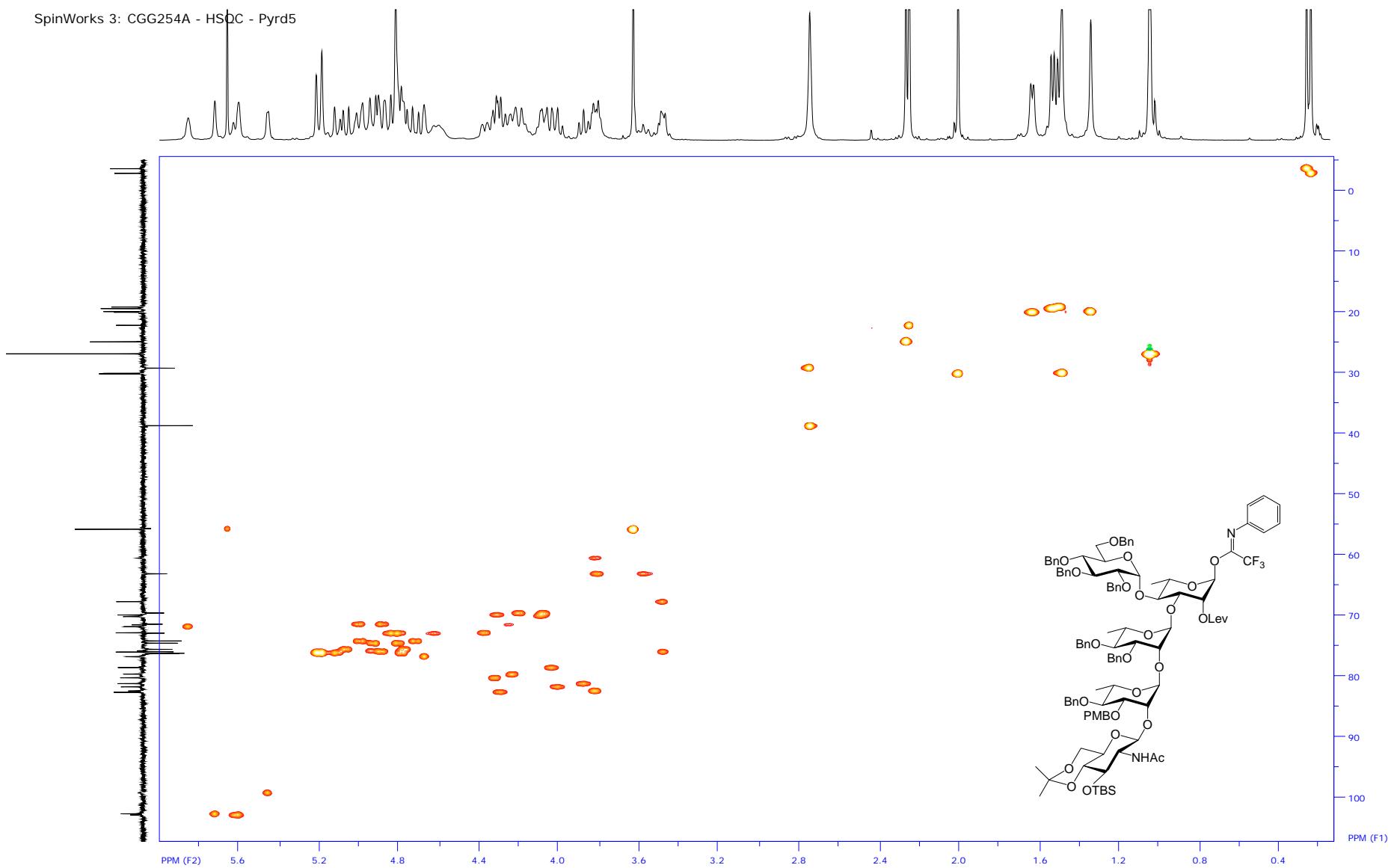
Compound 7



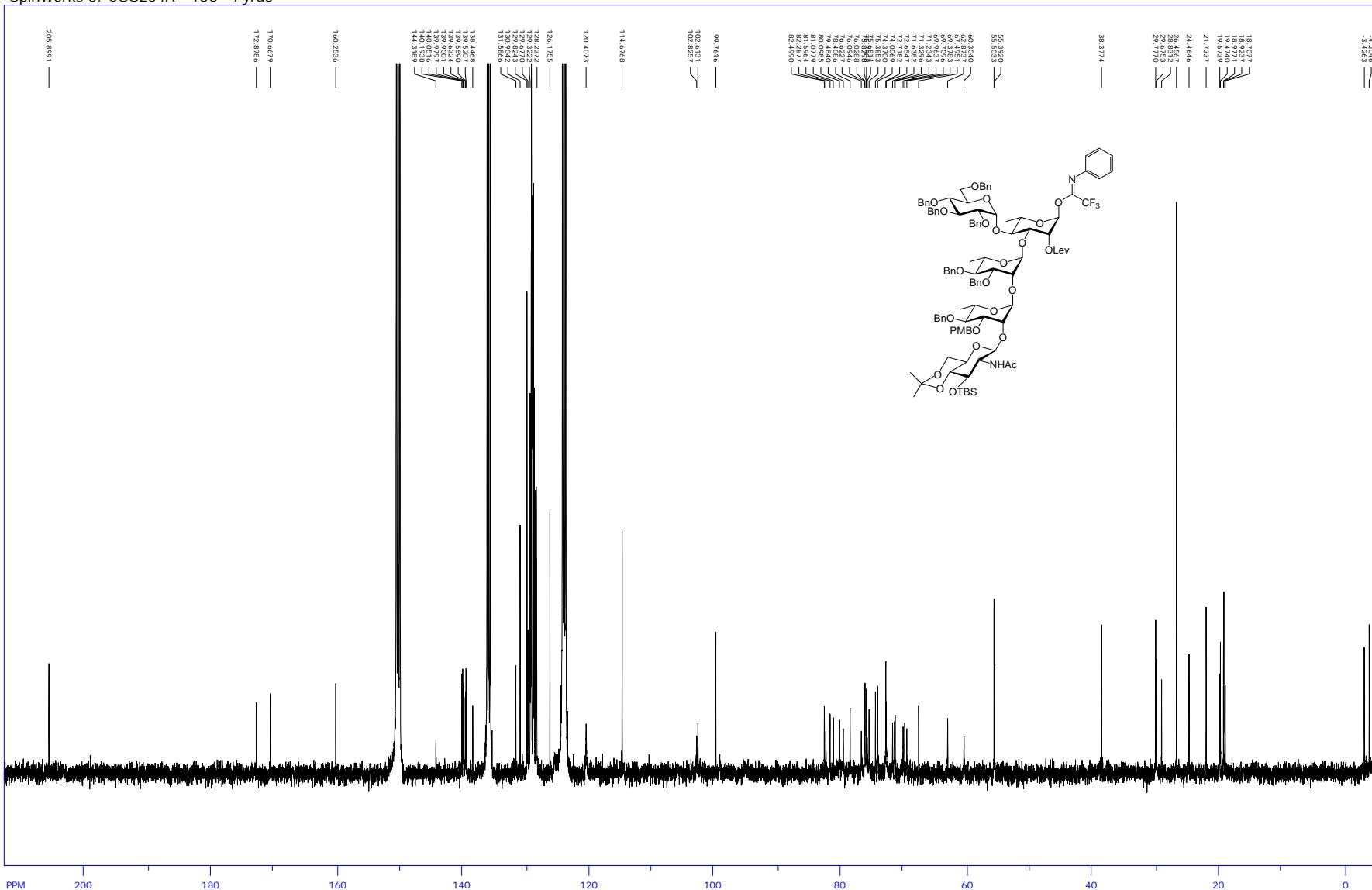
SpinWorks 3: CGG254A - DEPT135 - Pyrd5



SpinWorks 3: CGG254A - HSQC - Pyrd5



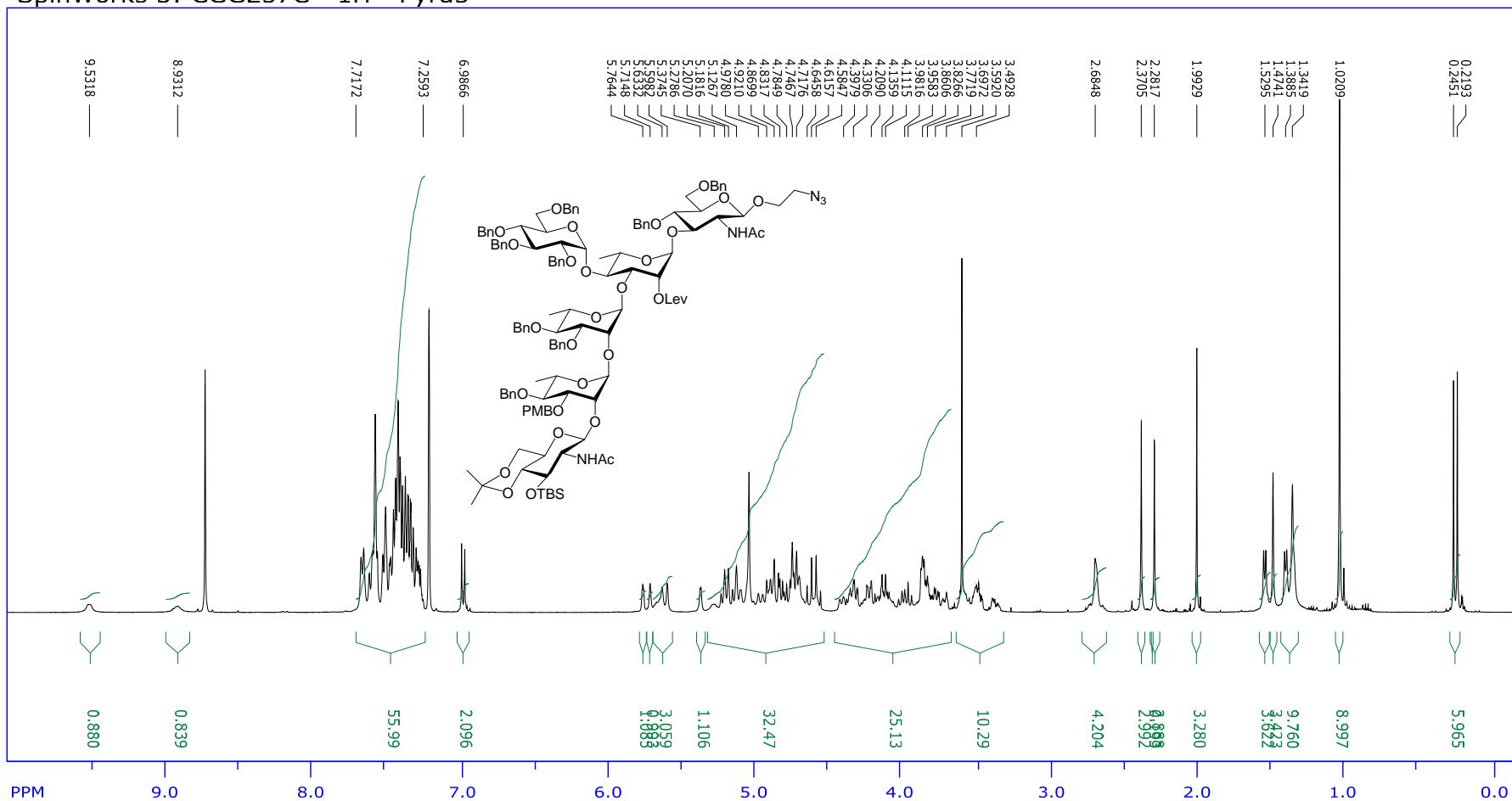
SpinWorks 3: CGG254A - 13C - Pyrd5



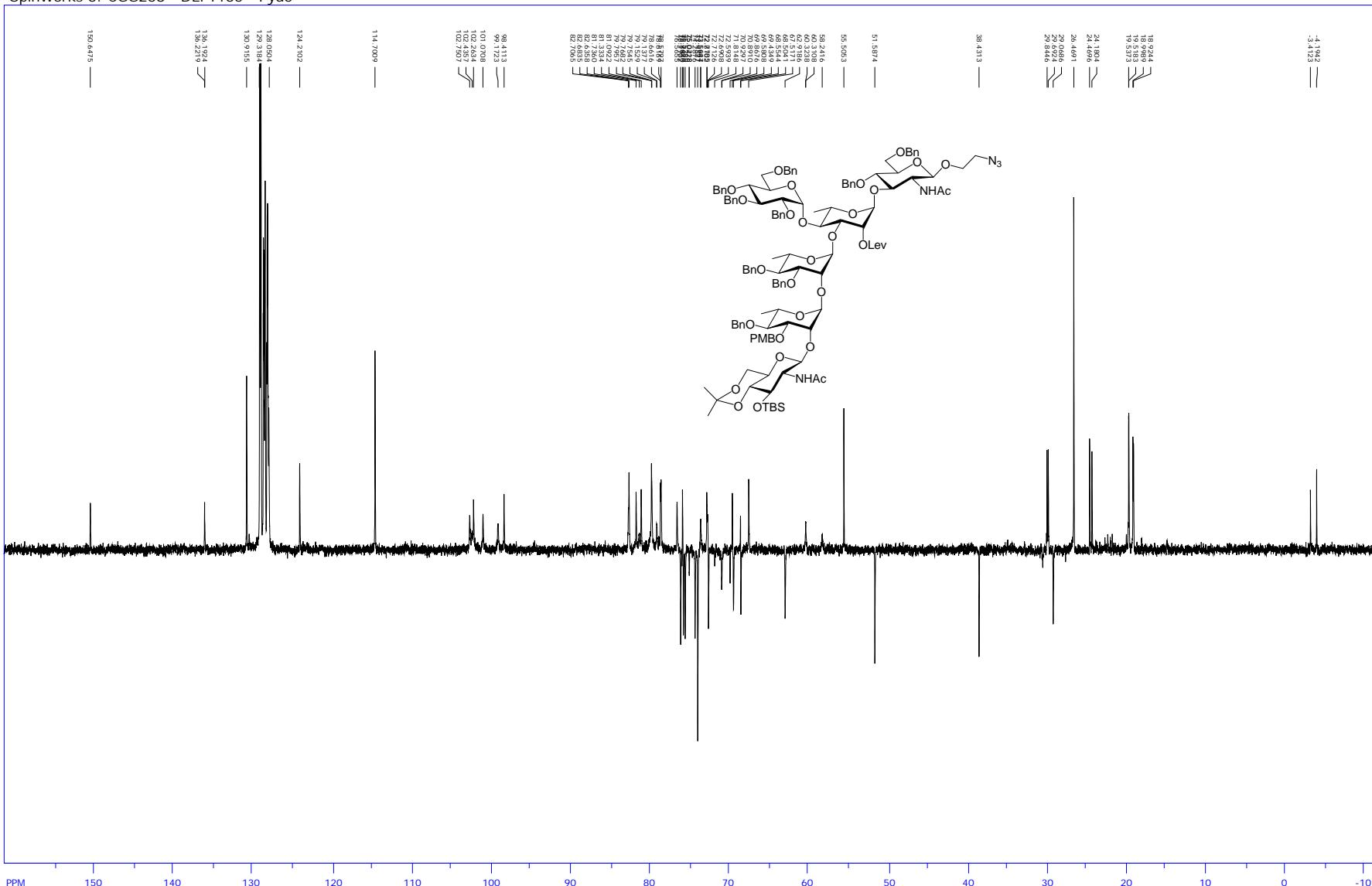
S150

Compound 34

SpinWorks 3: CGG257C - 1H - Pyrd5

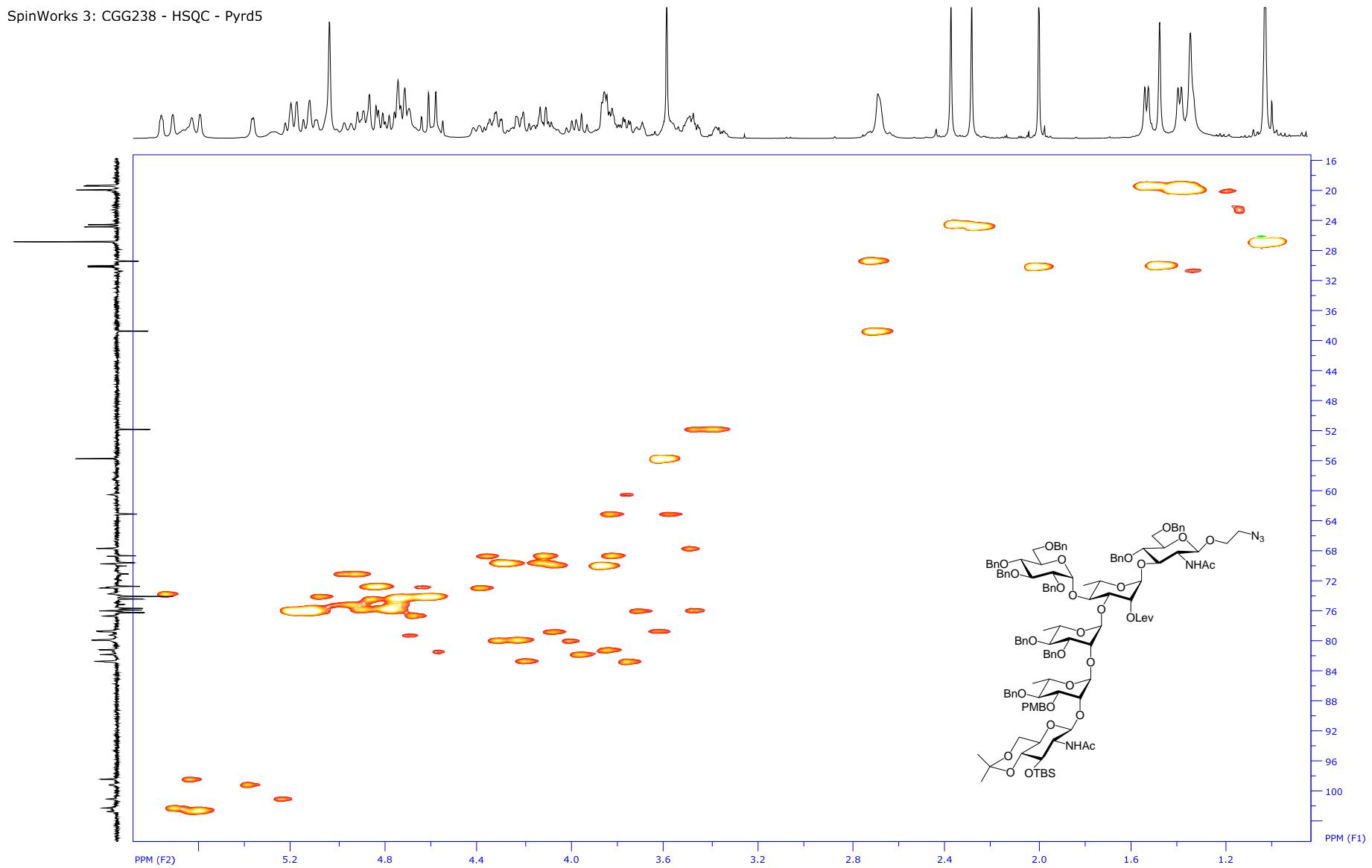


SpinWorks 3: CGG238 - DEPT135 - Pyd5

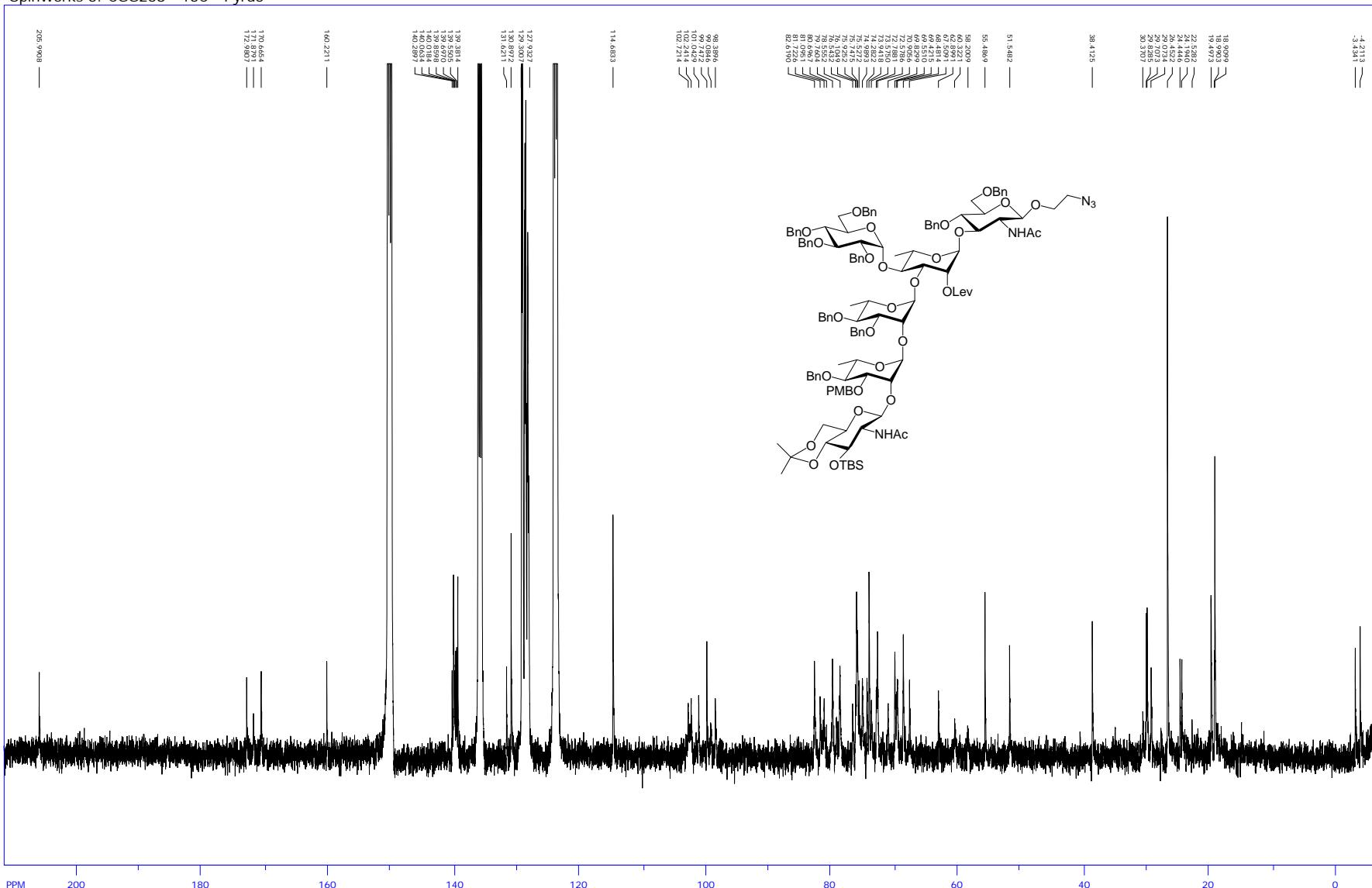


S152

SpinWorks 3: CGG238 - HSQC - Pyrd5

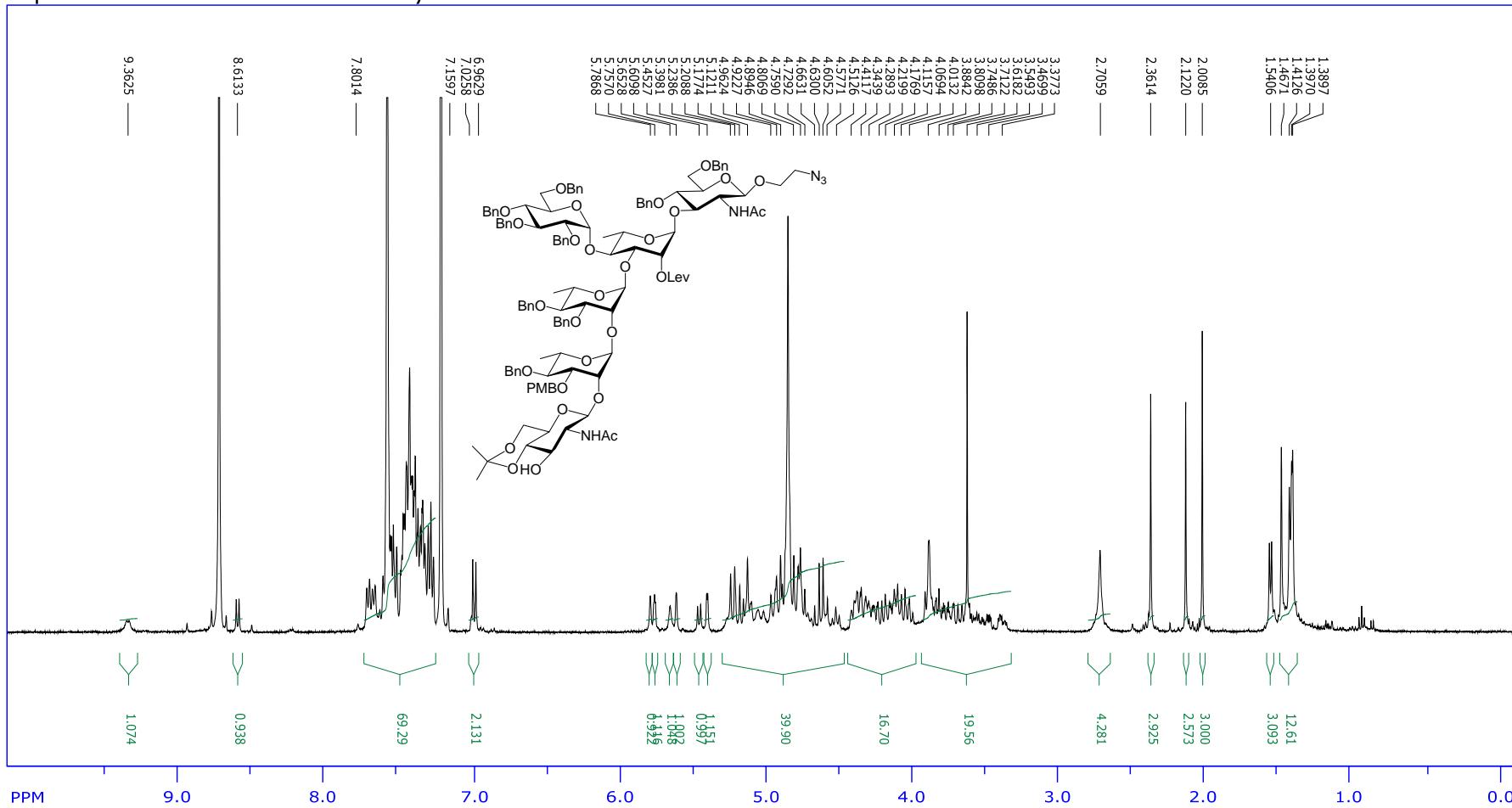


SpinWorks 3: CGG238 - 13C - Pyrd5

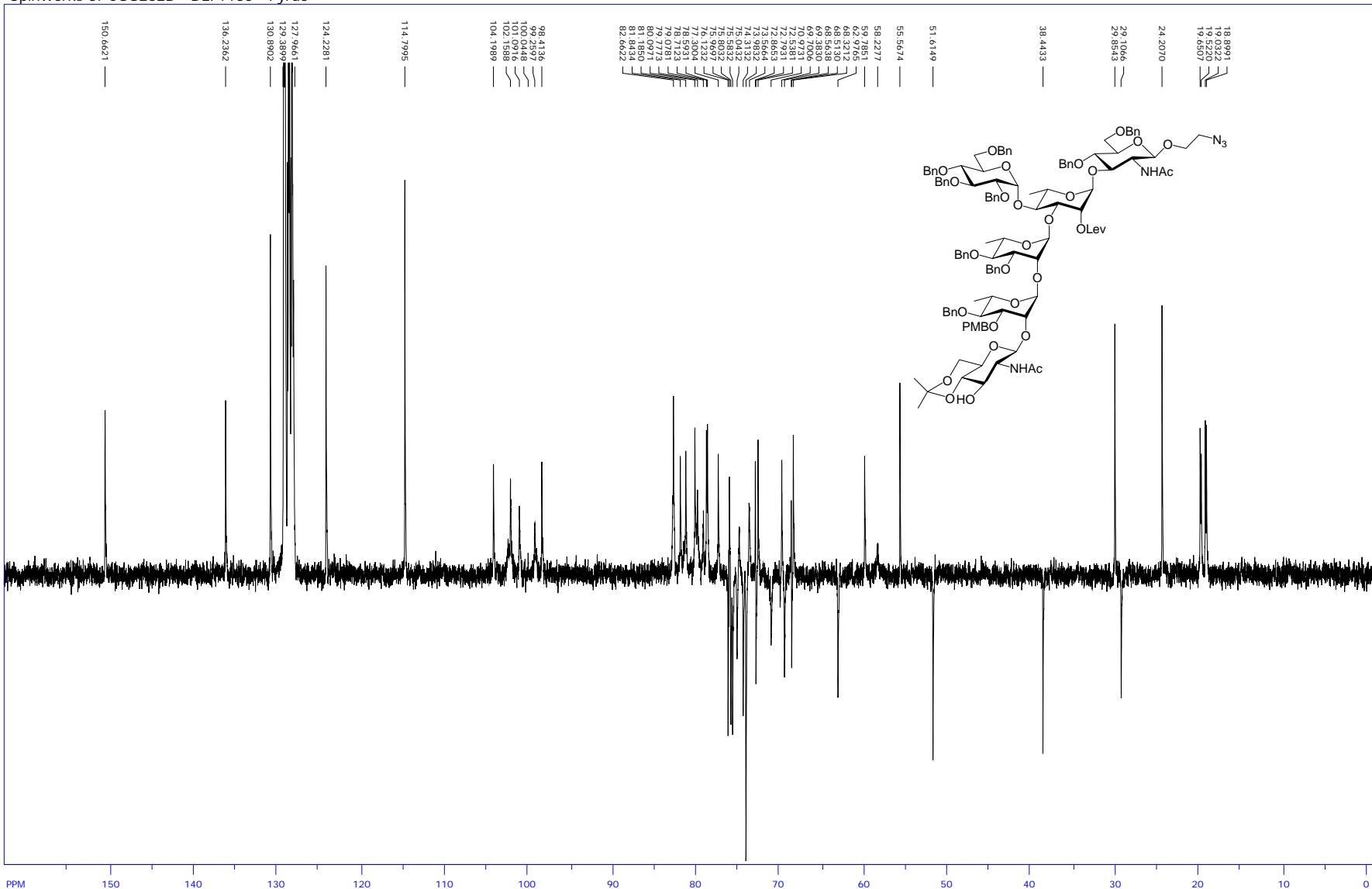


Compound 35

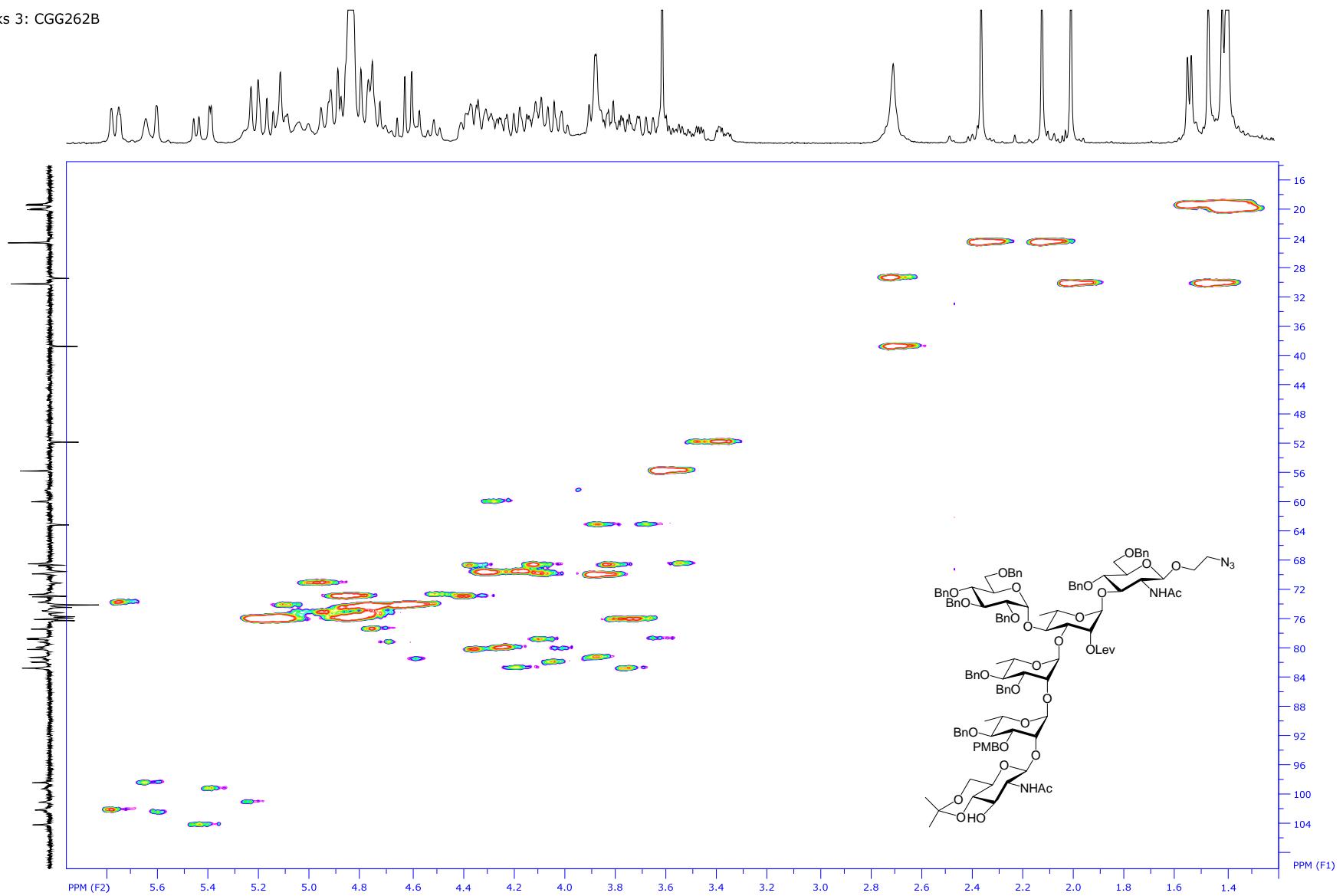
SpinWorks 3: CGG281B - 1H - Pyrd5



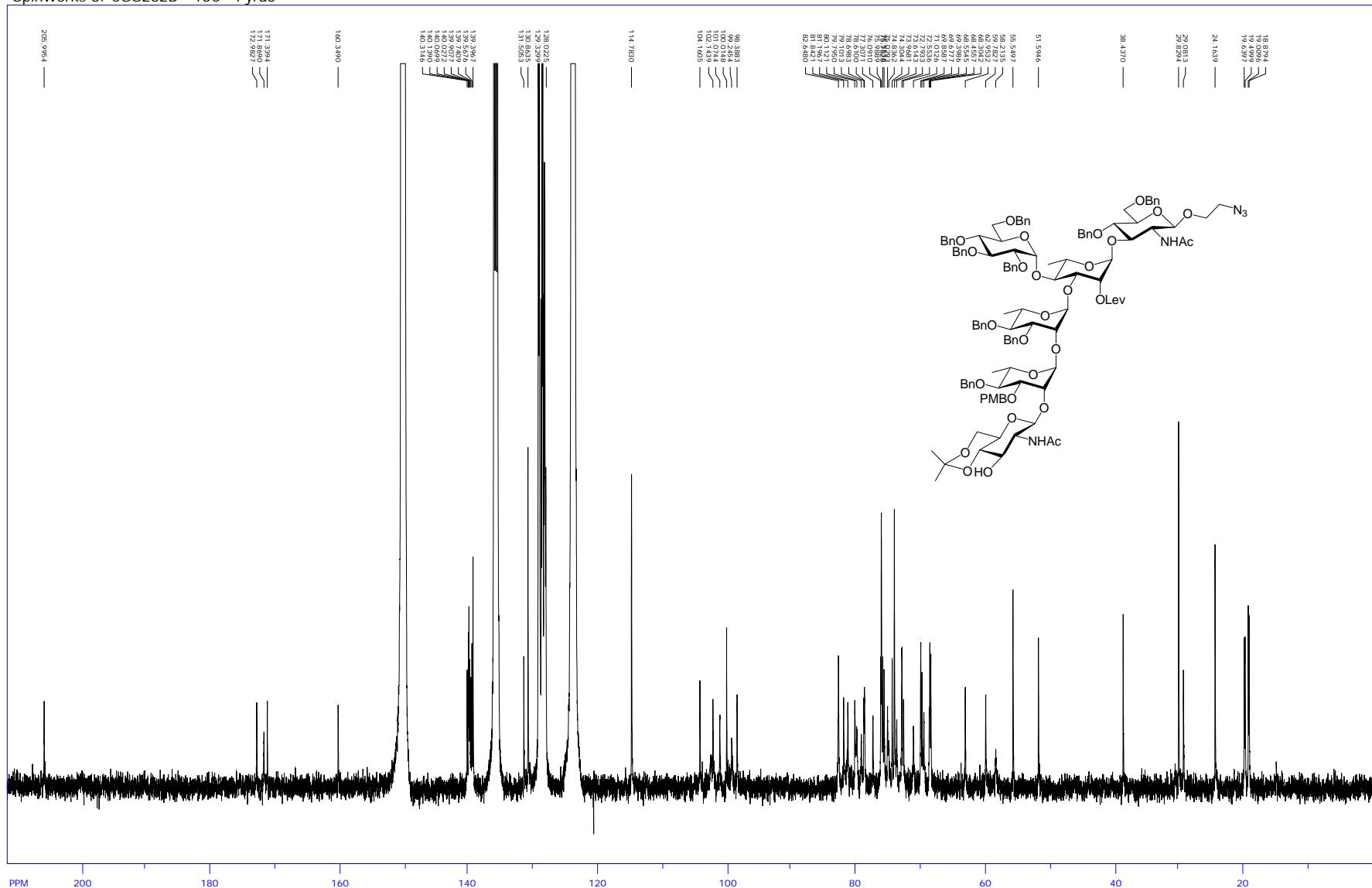
SpinWorks 3: CGG262B - DEPT135 - Pyrd5



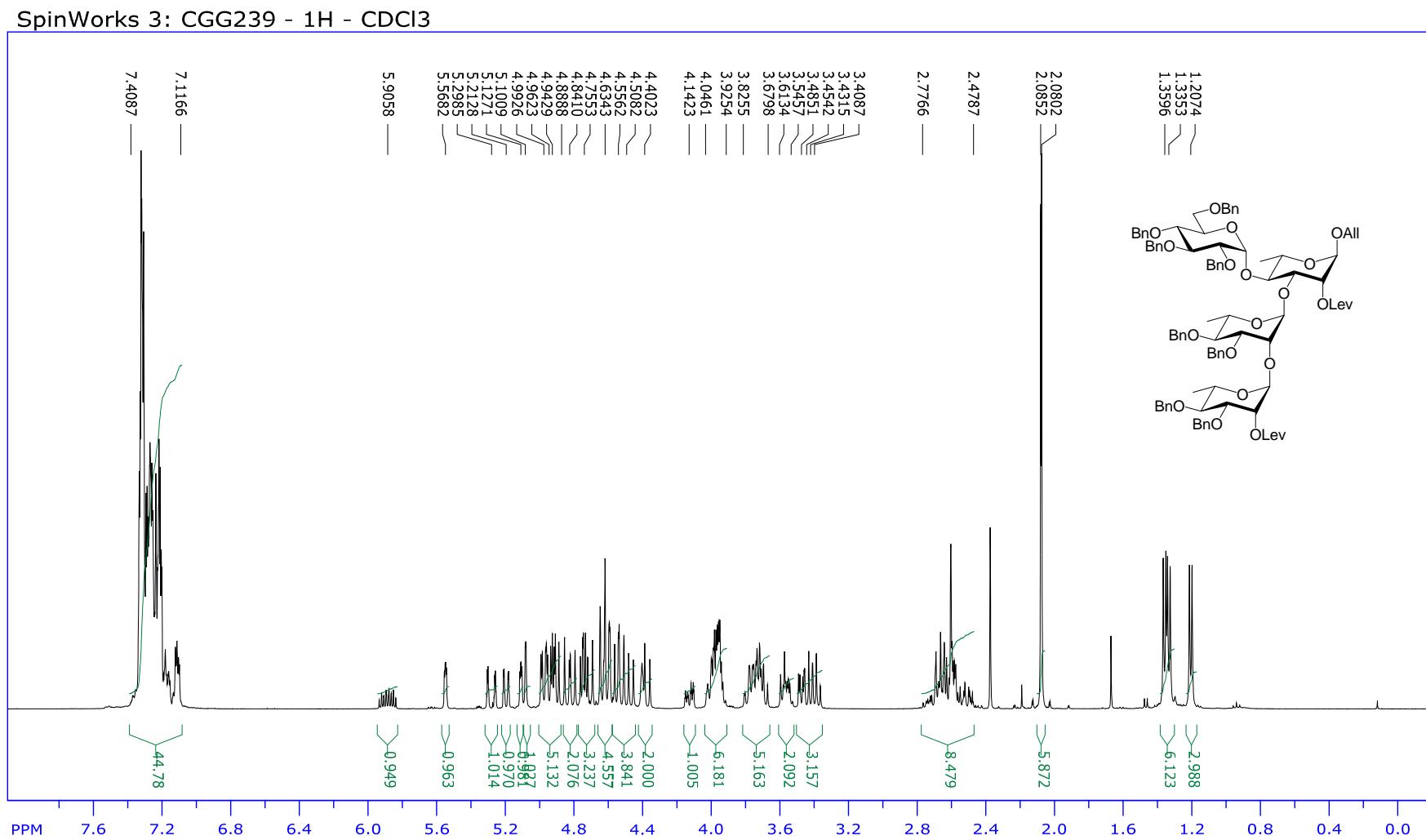
SpinWorks 3: CGG262B



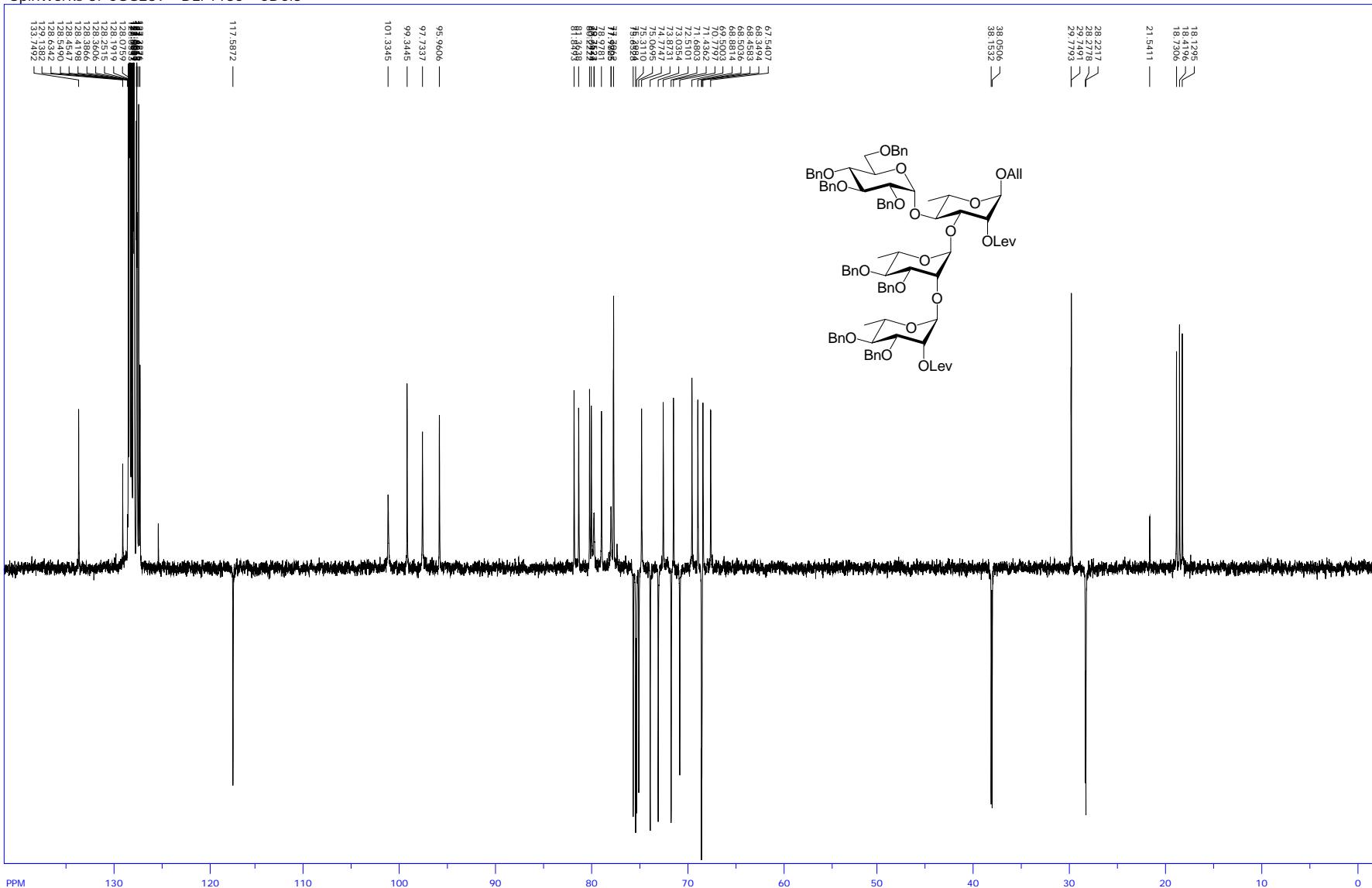
SpinWorks 3: CGG262B - 13C - Pyrd5



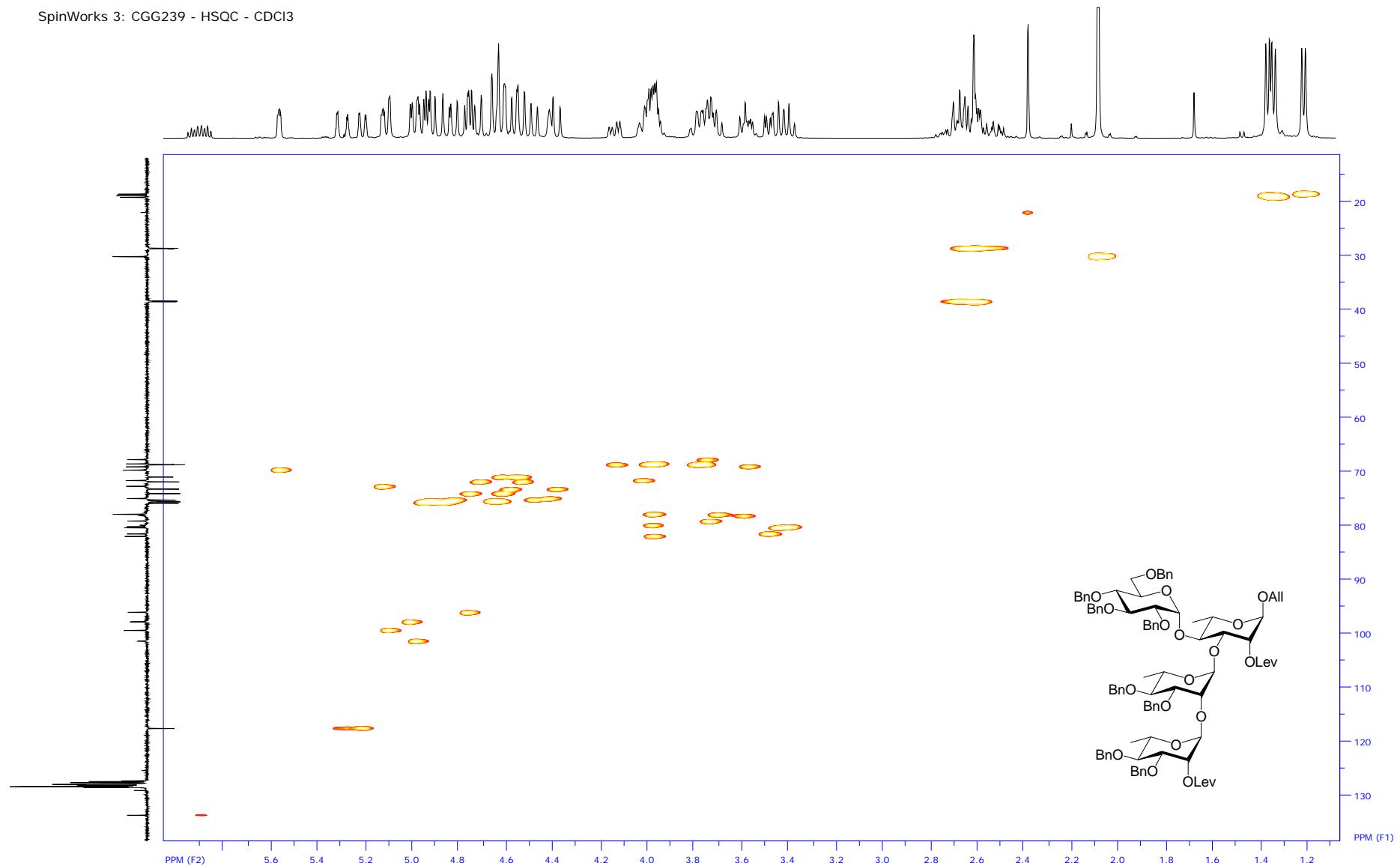
Compound 23



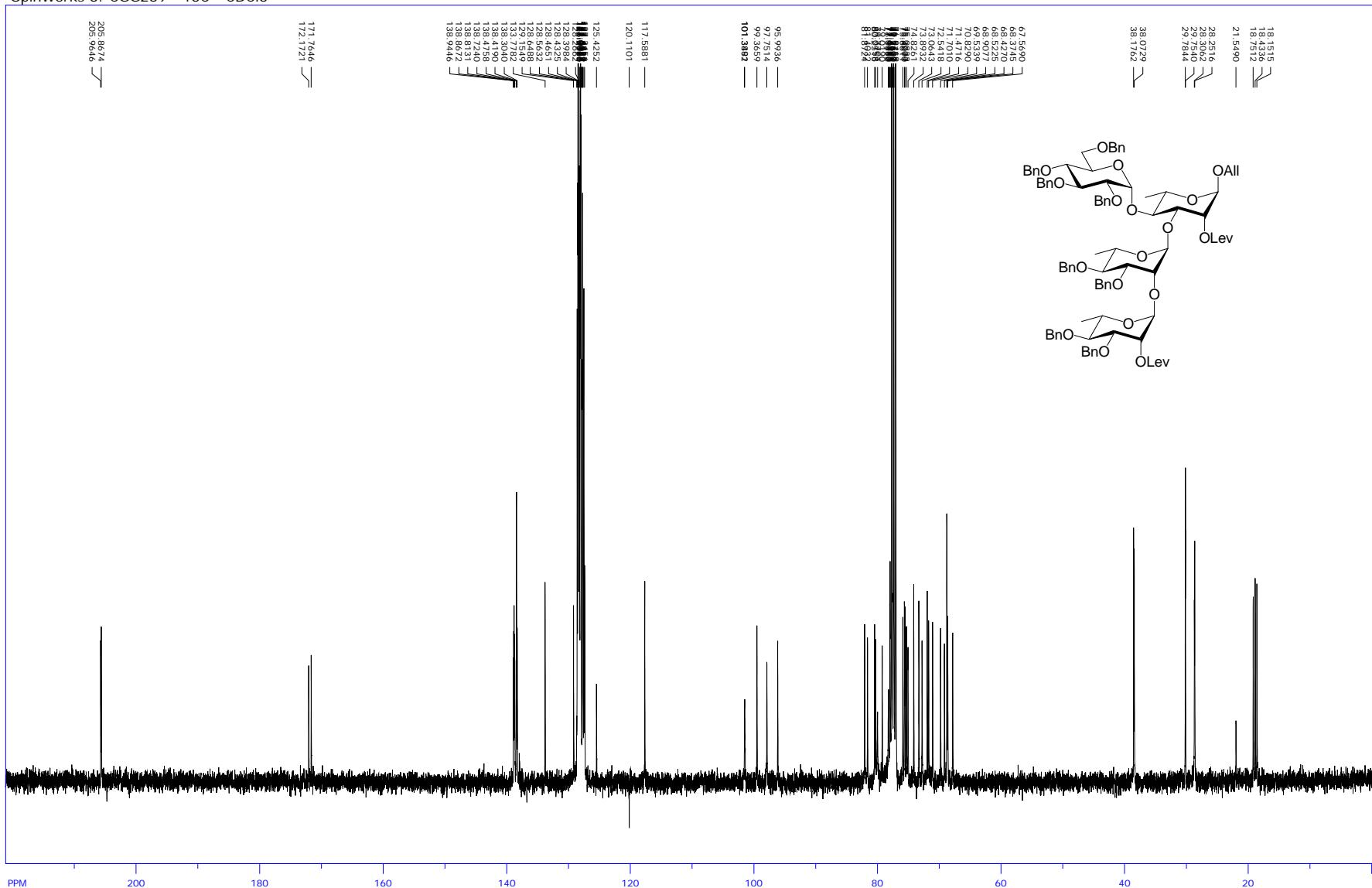
SpinWorks 3: CGG239 - DEPT135 - CDCl₃



SpinWorks 3: CGG239 - HSQC - CDCl3

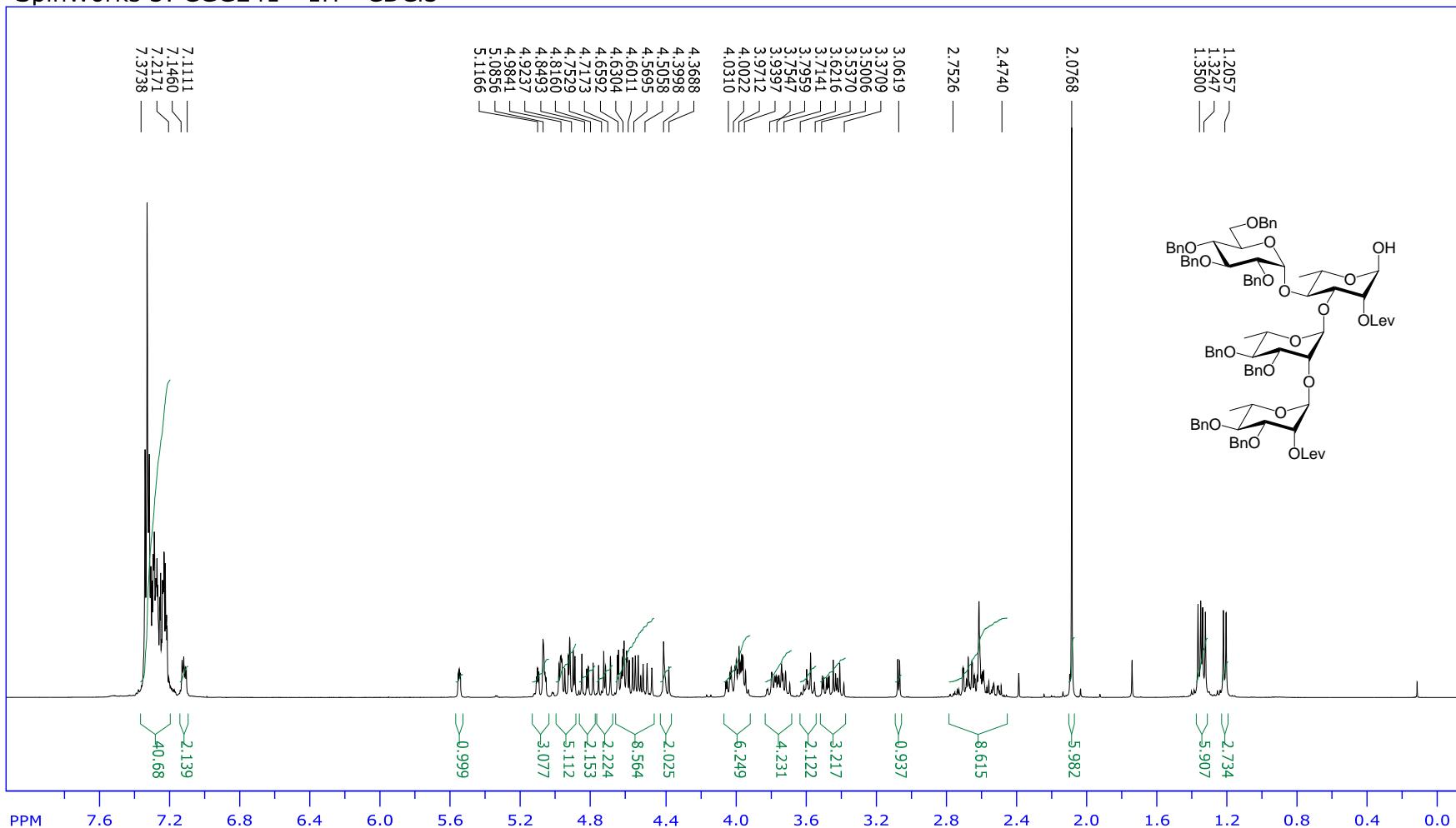


SpinWorks 3: CGG239 - 13C - CDCl3

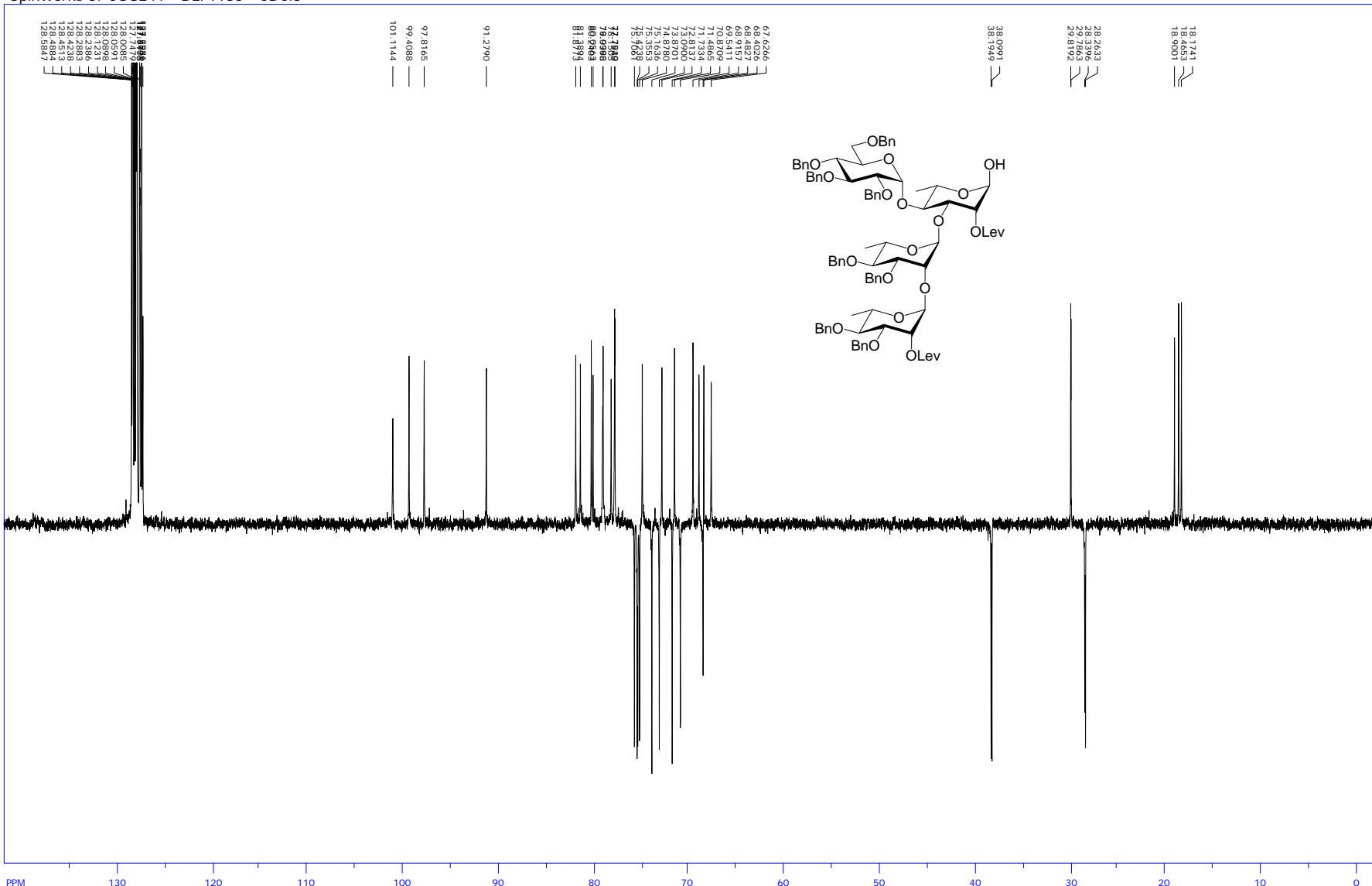


Compound 24

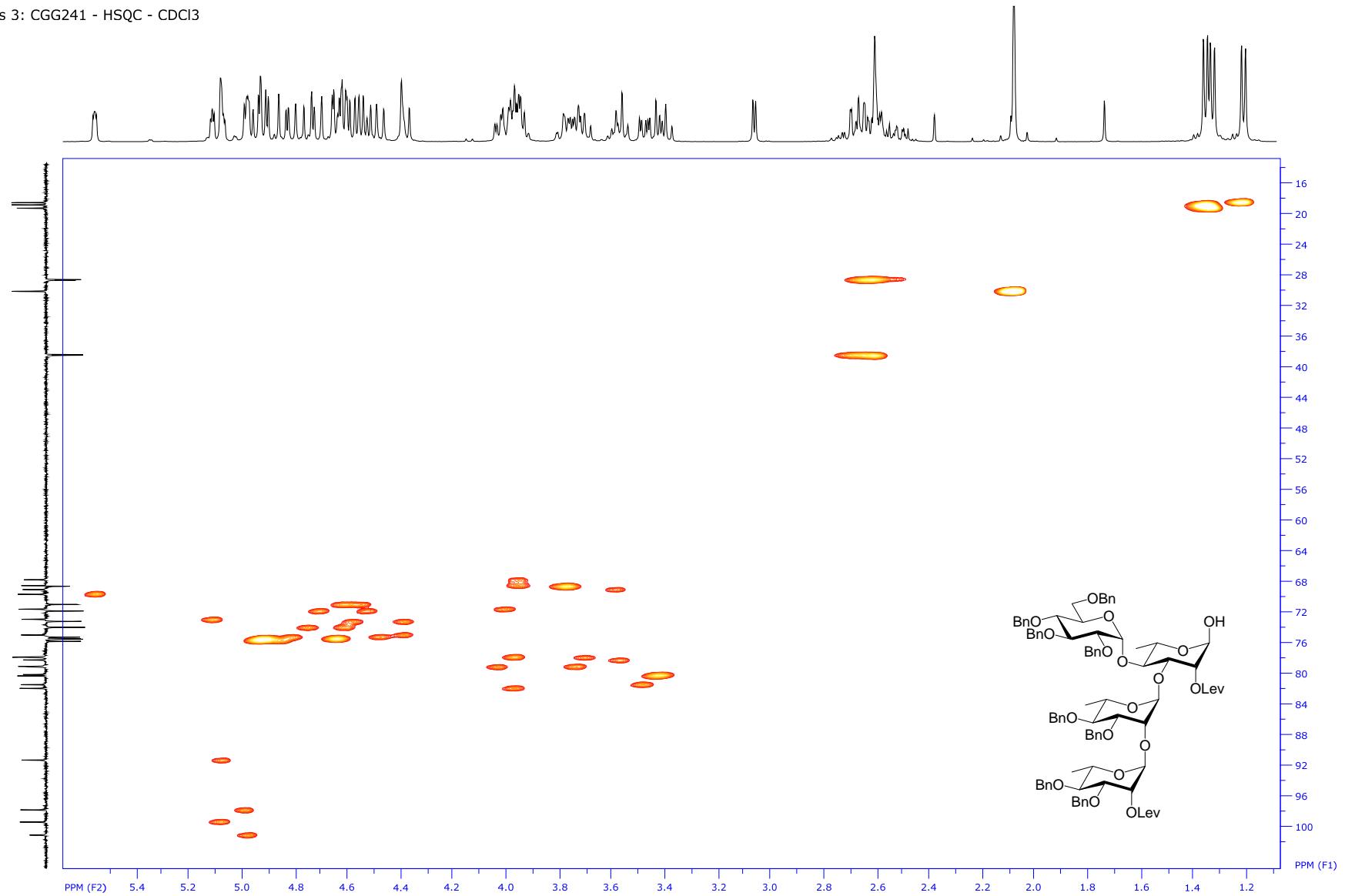
SpinWorks 3: CGG241 - 1H - CDCl₃



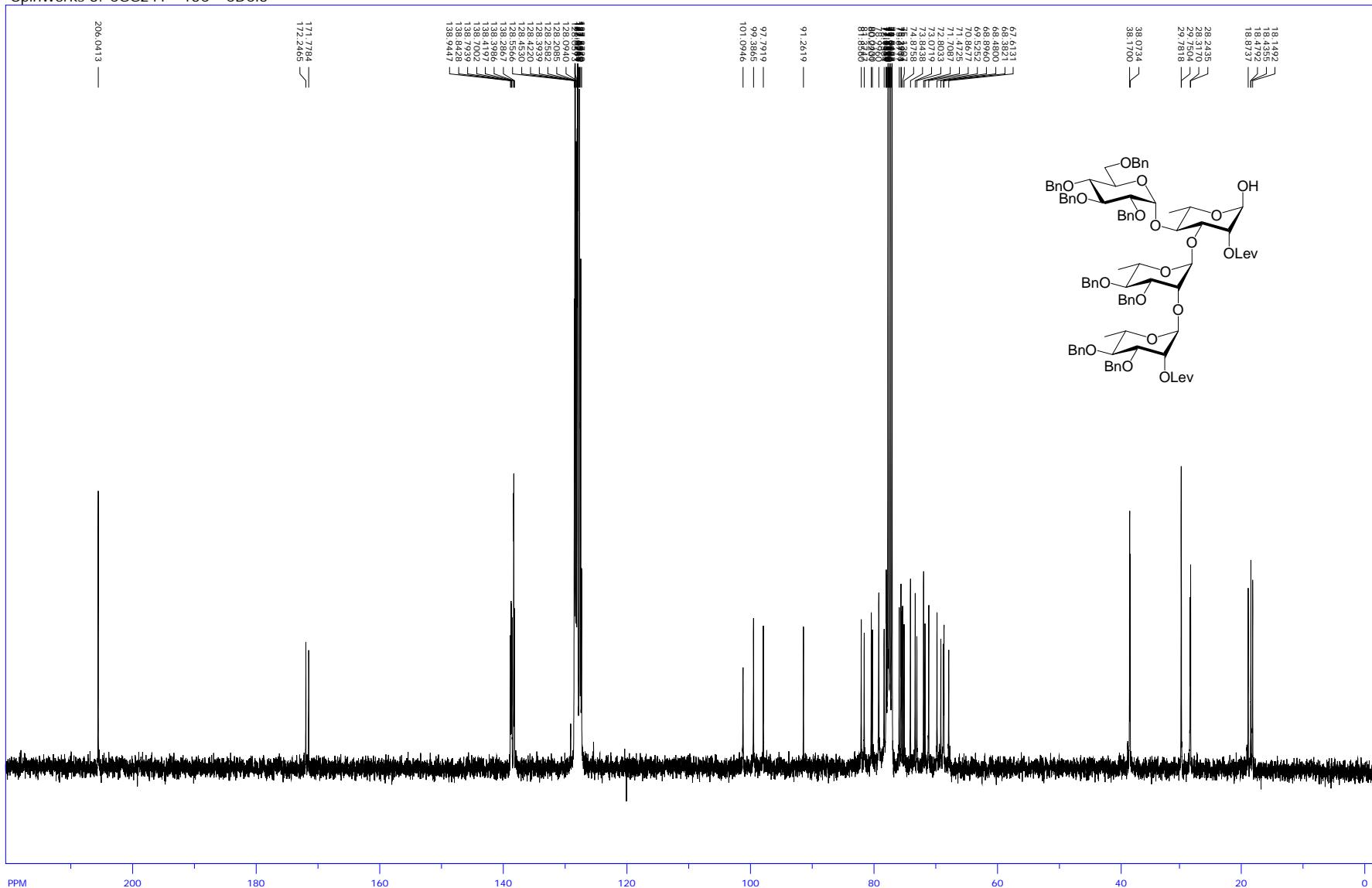
SpinWorks 3: CGG241 - DEPT135 - CDCl₃



SpinWorks 3: CGG241 - HSQC - CDCl₃



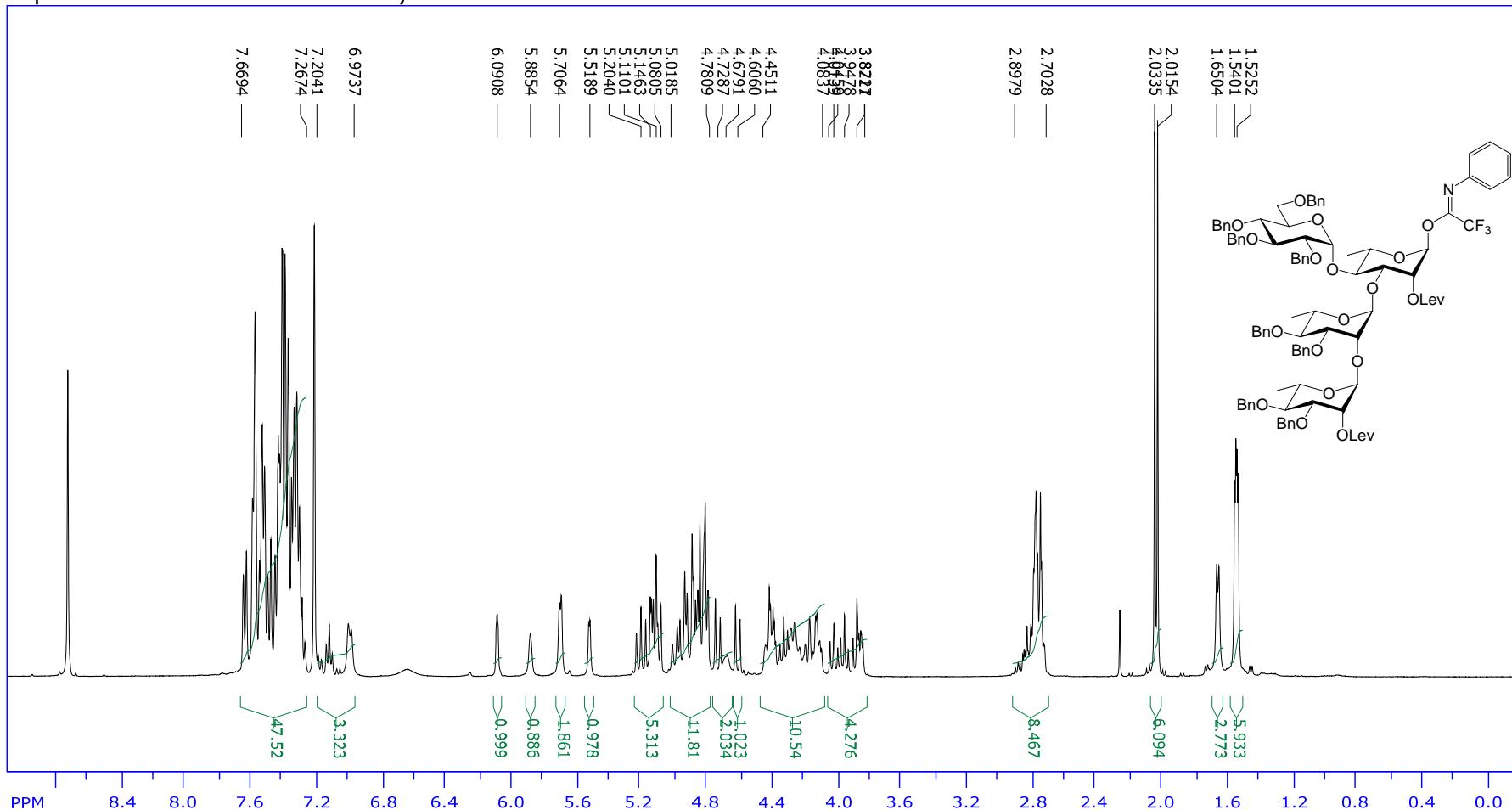
SpinWorks 3: CGG241 - 13C - CDCl3



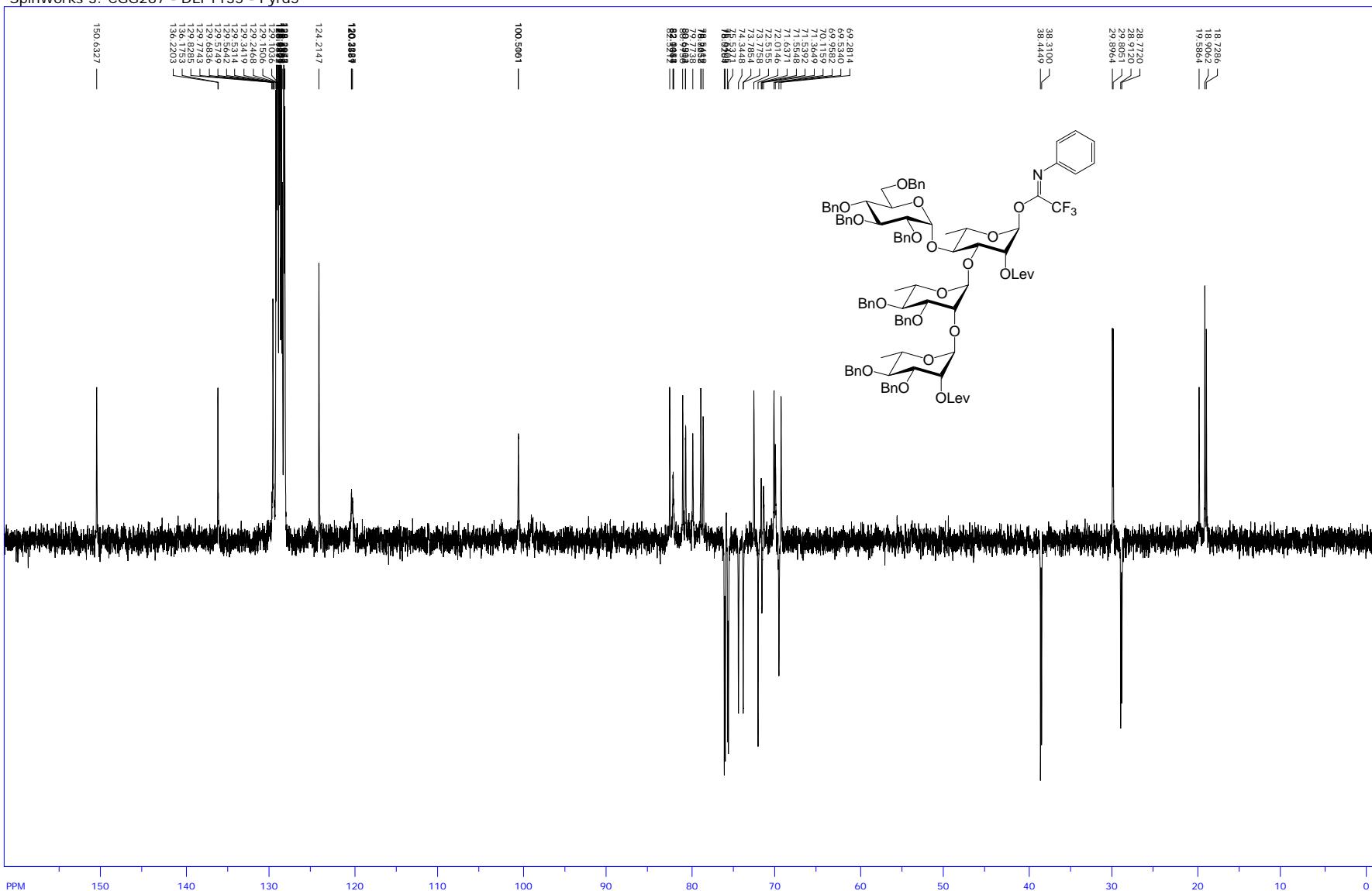
S166

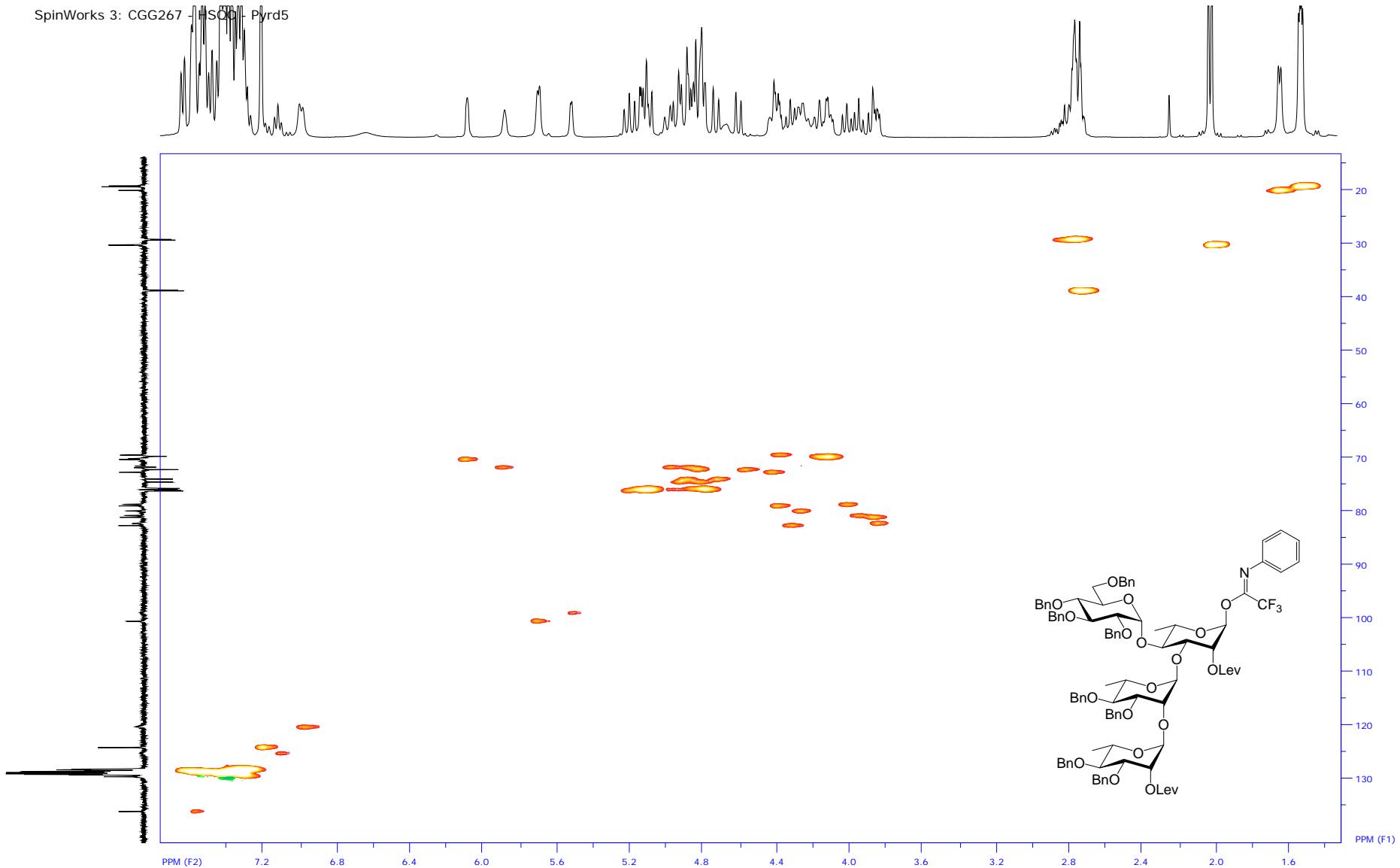
Compound 6

SpinWorks 3: CGG267 - 1H - Pyrd5

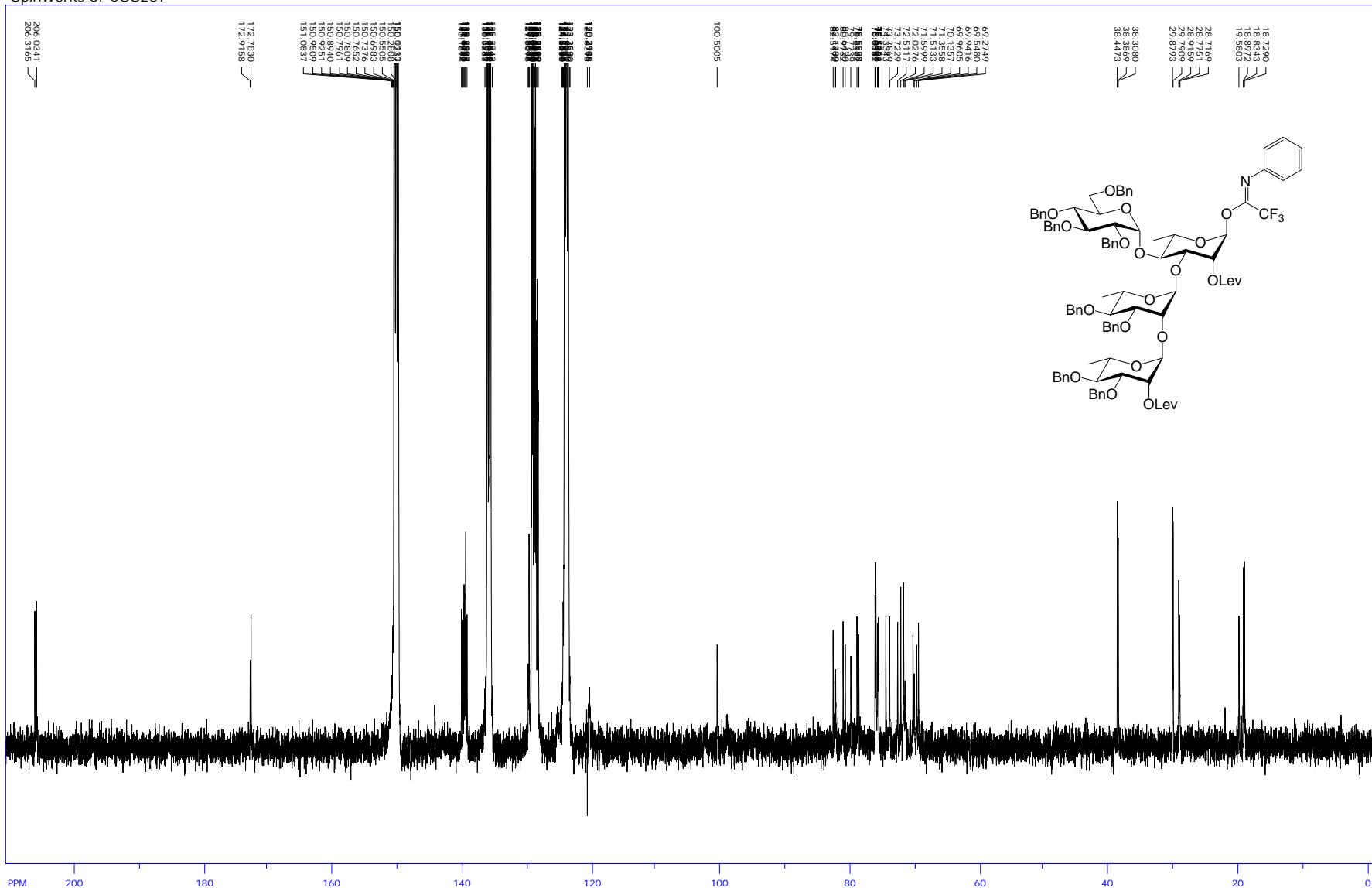


SpinWorks 3: CGG267 - DEPT135 - Pyrd5



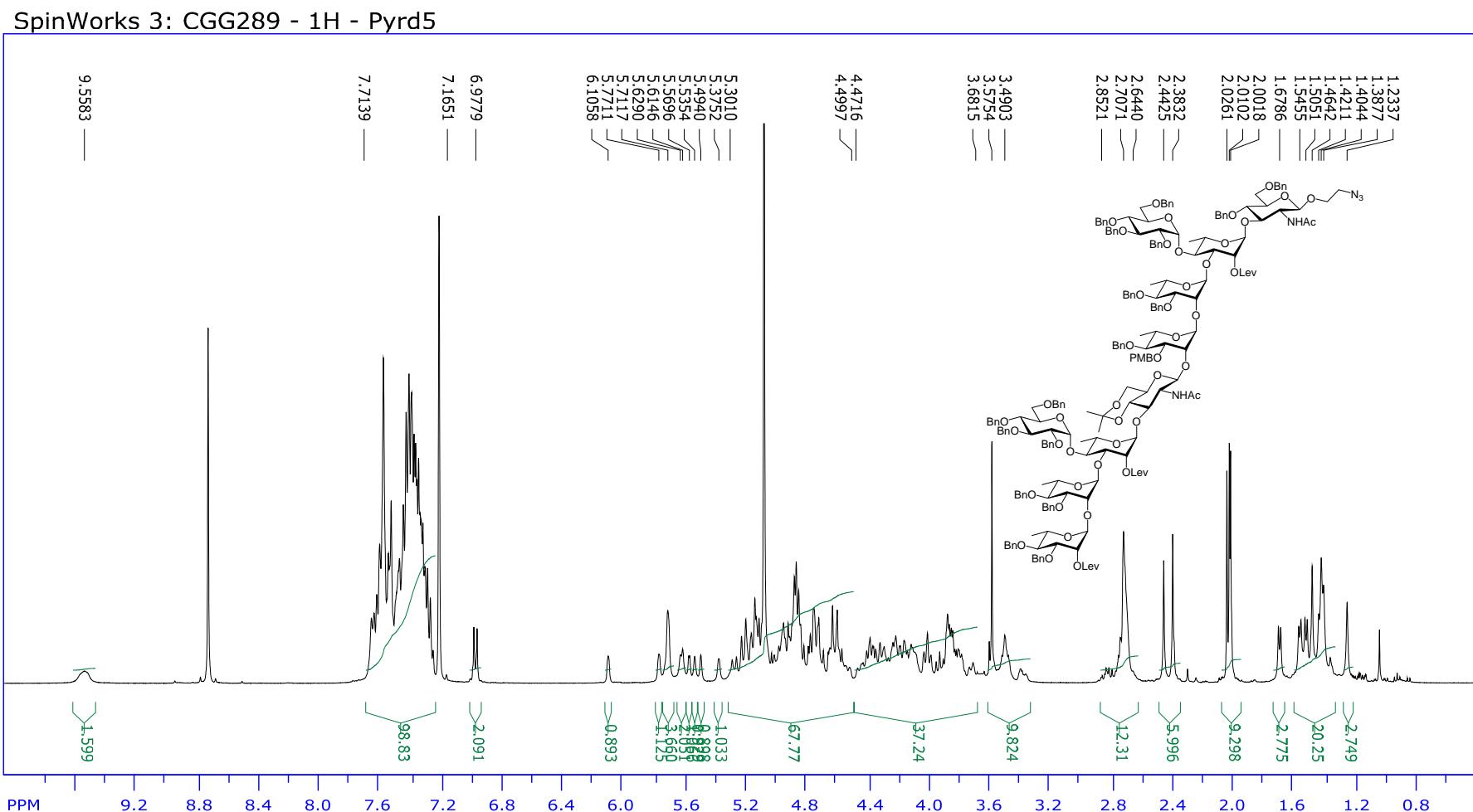


SpinWorks 3: CGG267

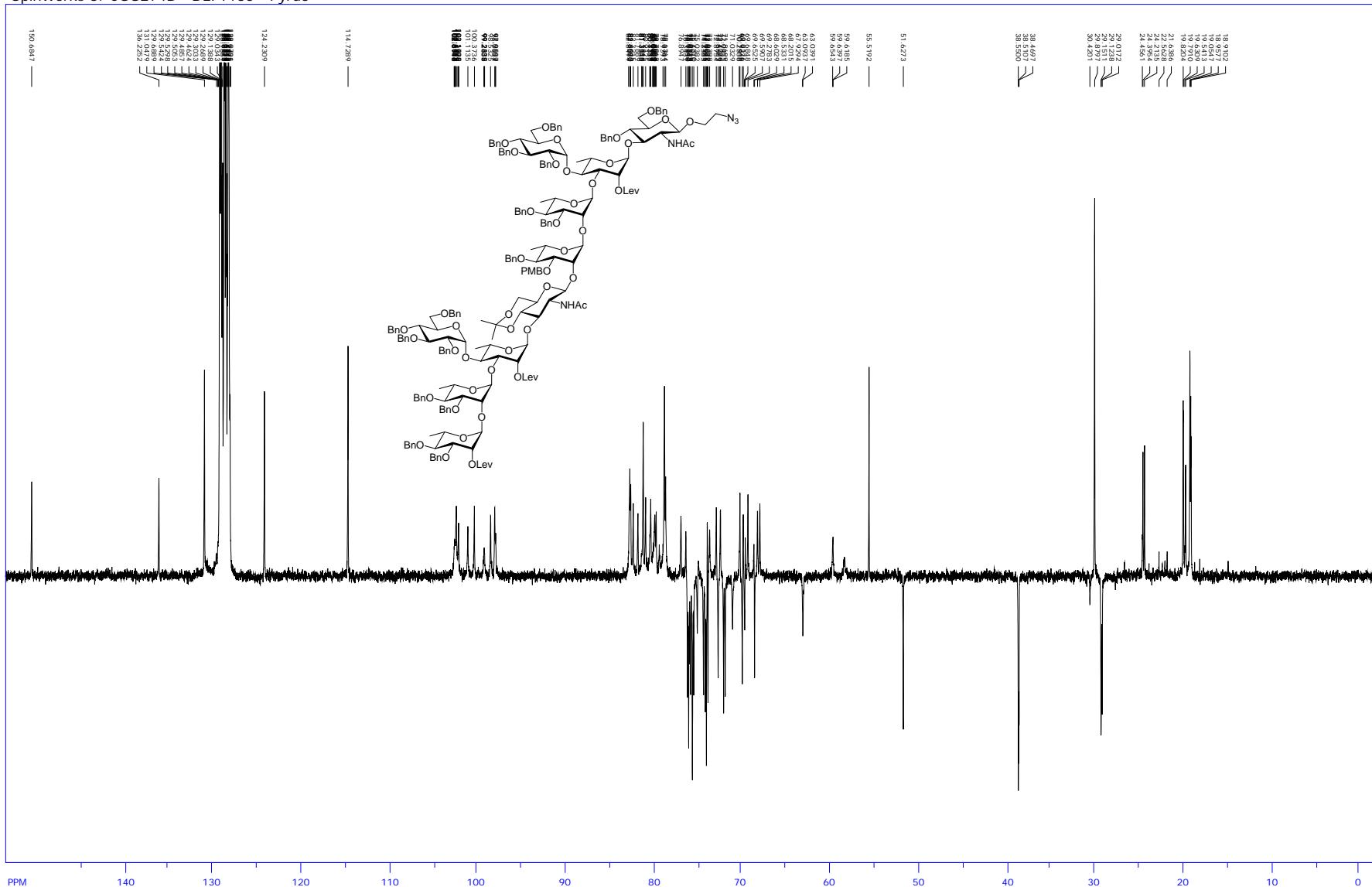


S170

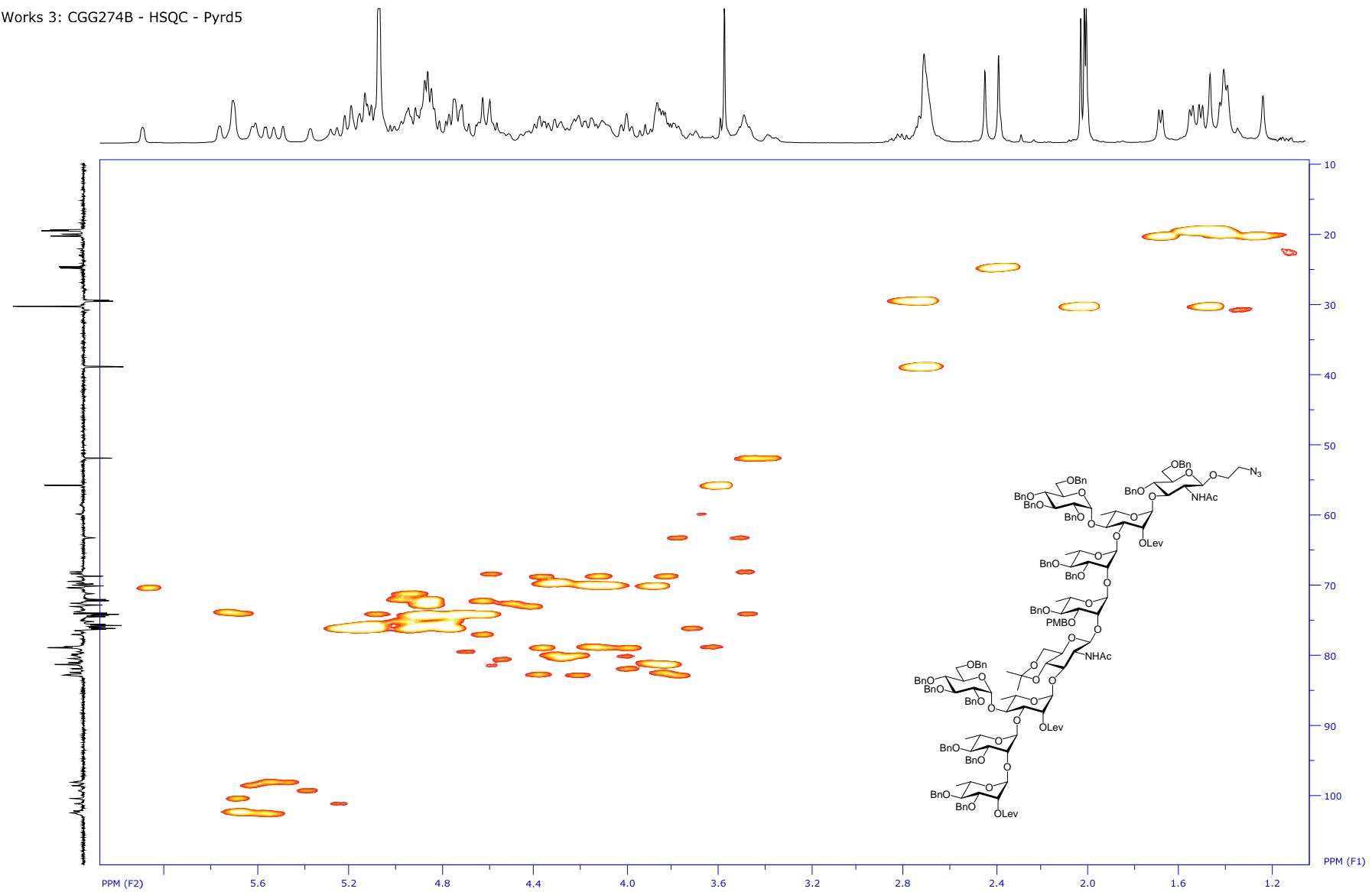
Compound 5



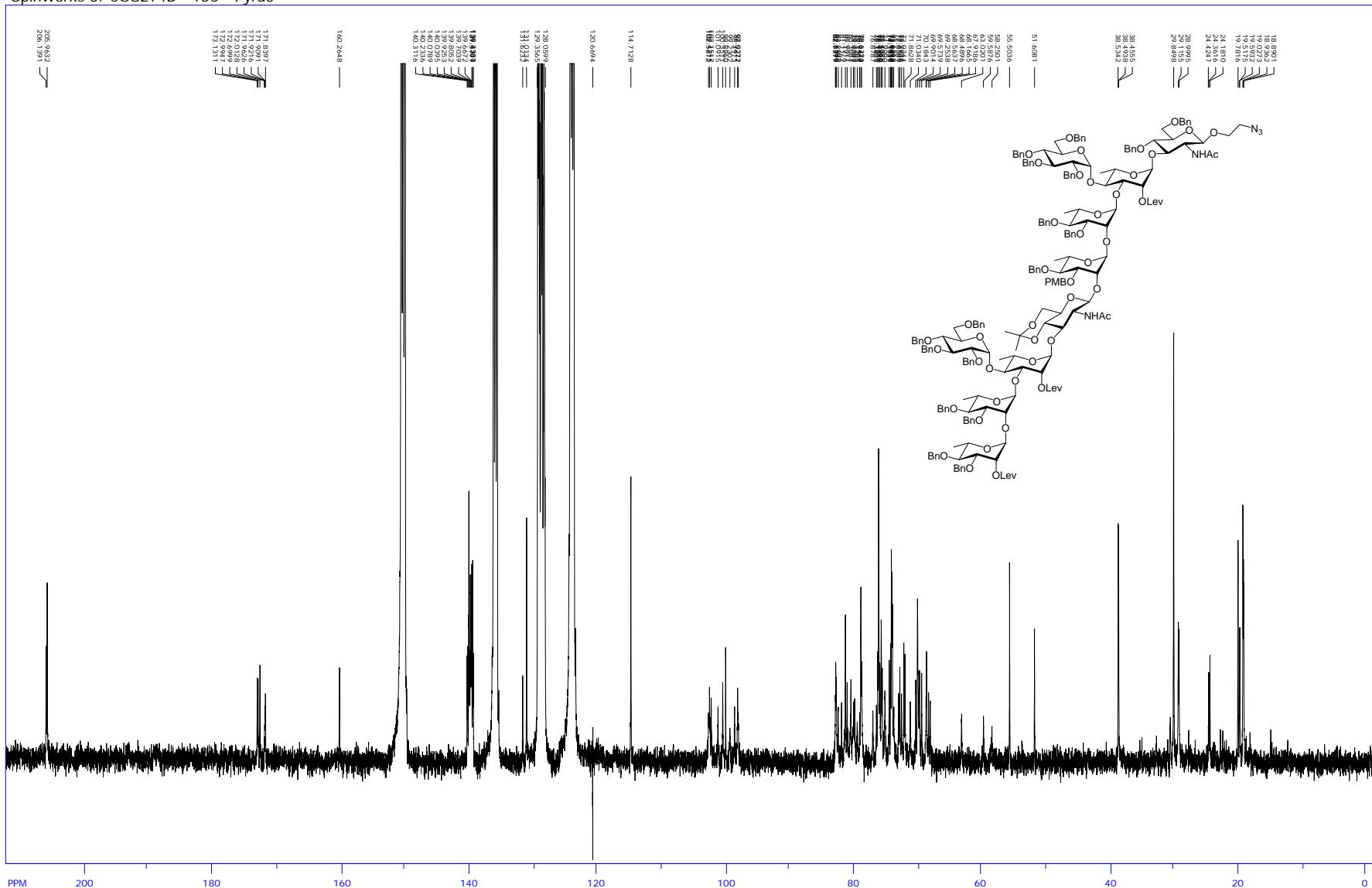
SpinWorks 3: CGG274B - DEPT135 - Pyrd5



SpinWorks 3: CGG274B - HSQC - Pyrd5

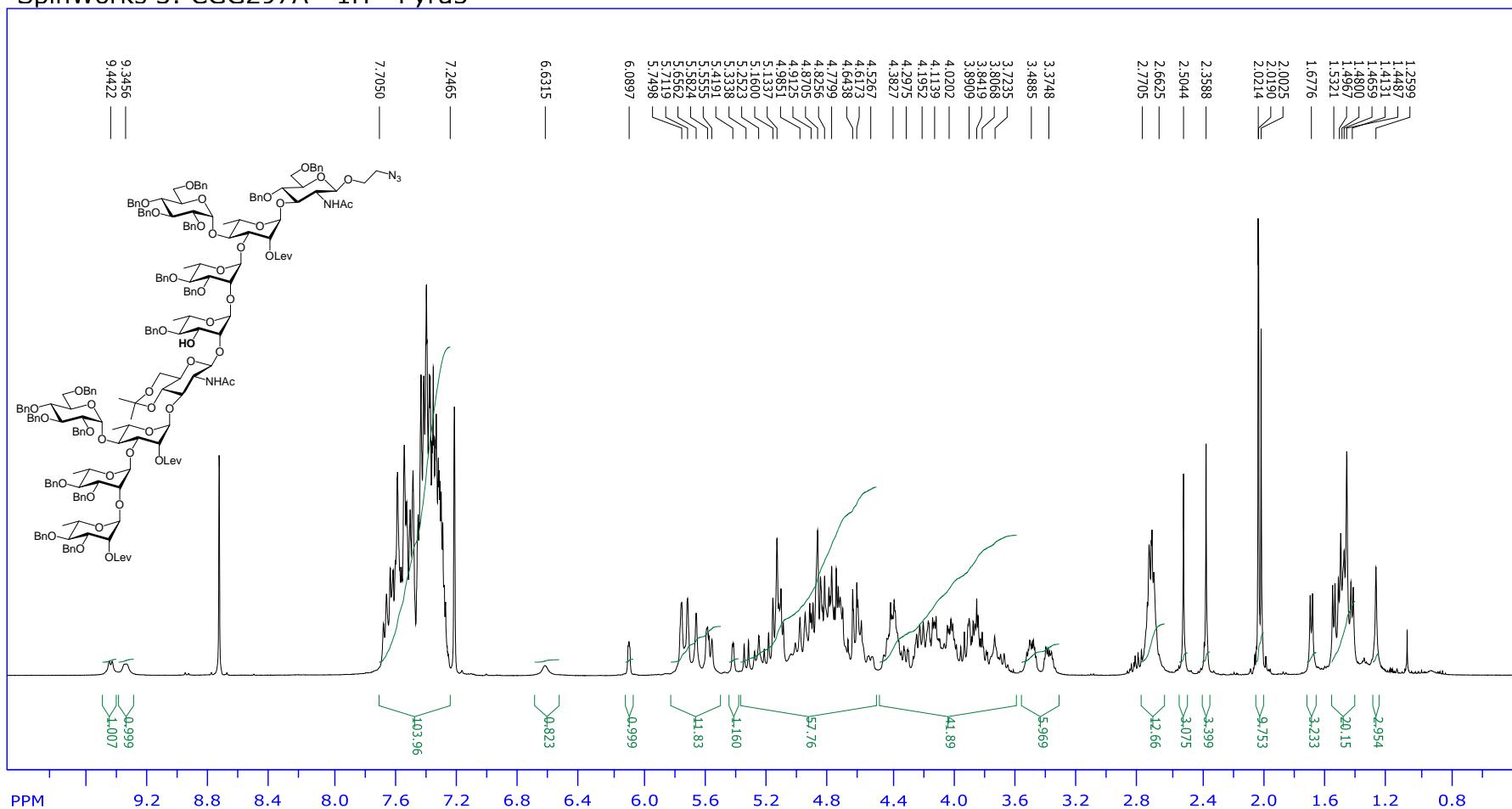


SpinWorks 3: CGG274B - 13C - Pyrd5

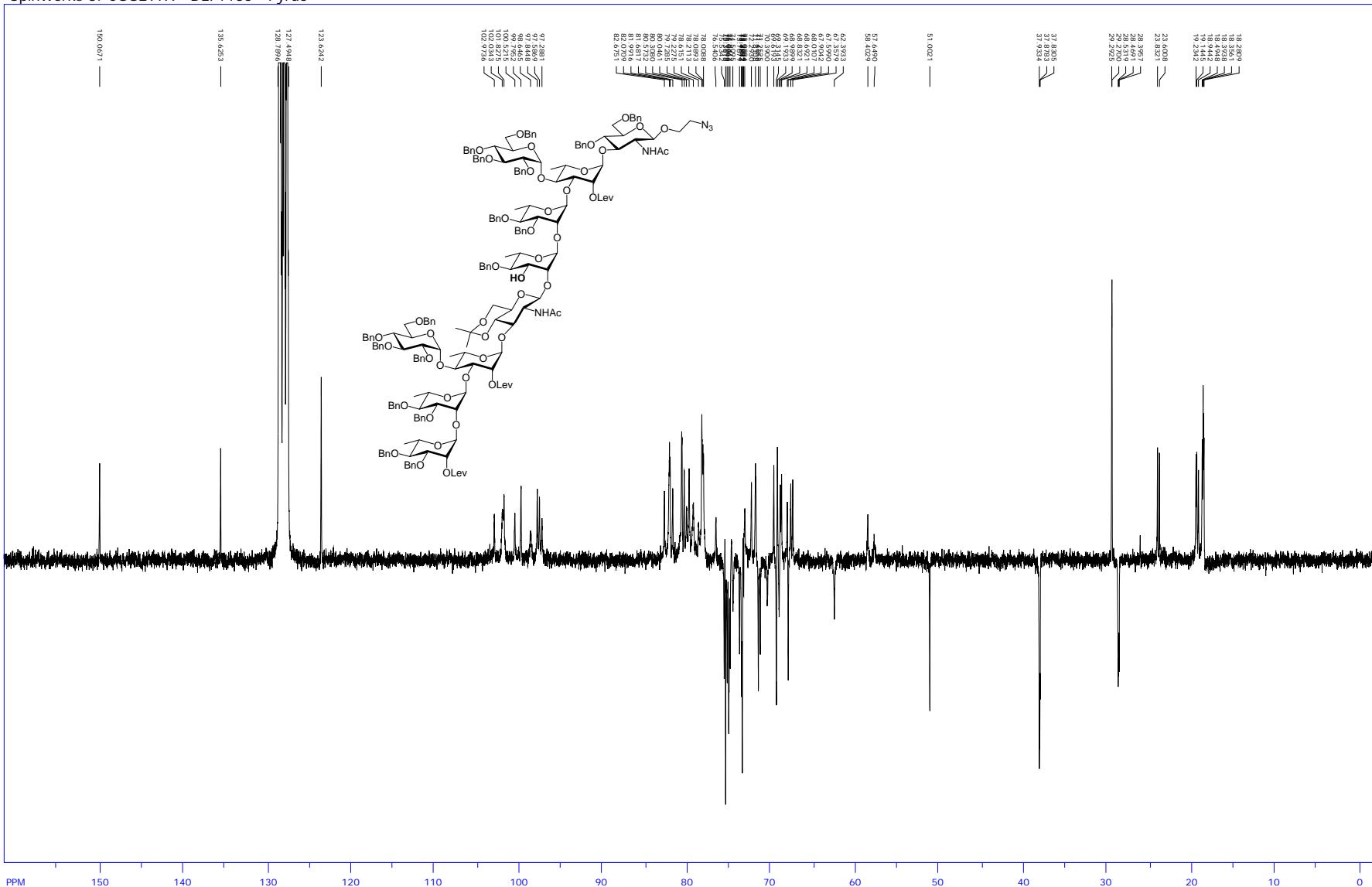


Compound 36

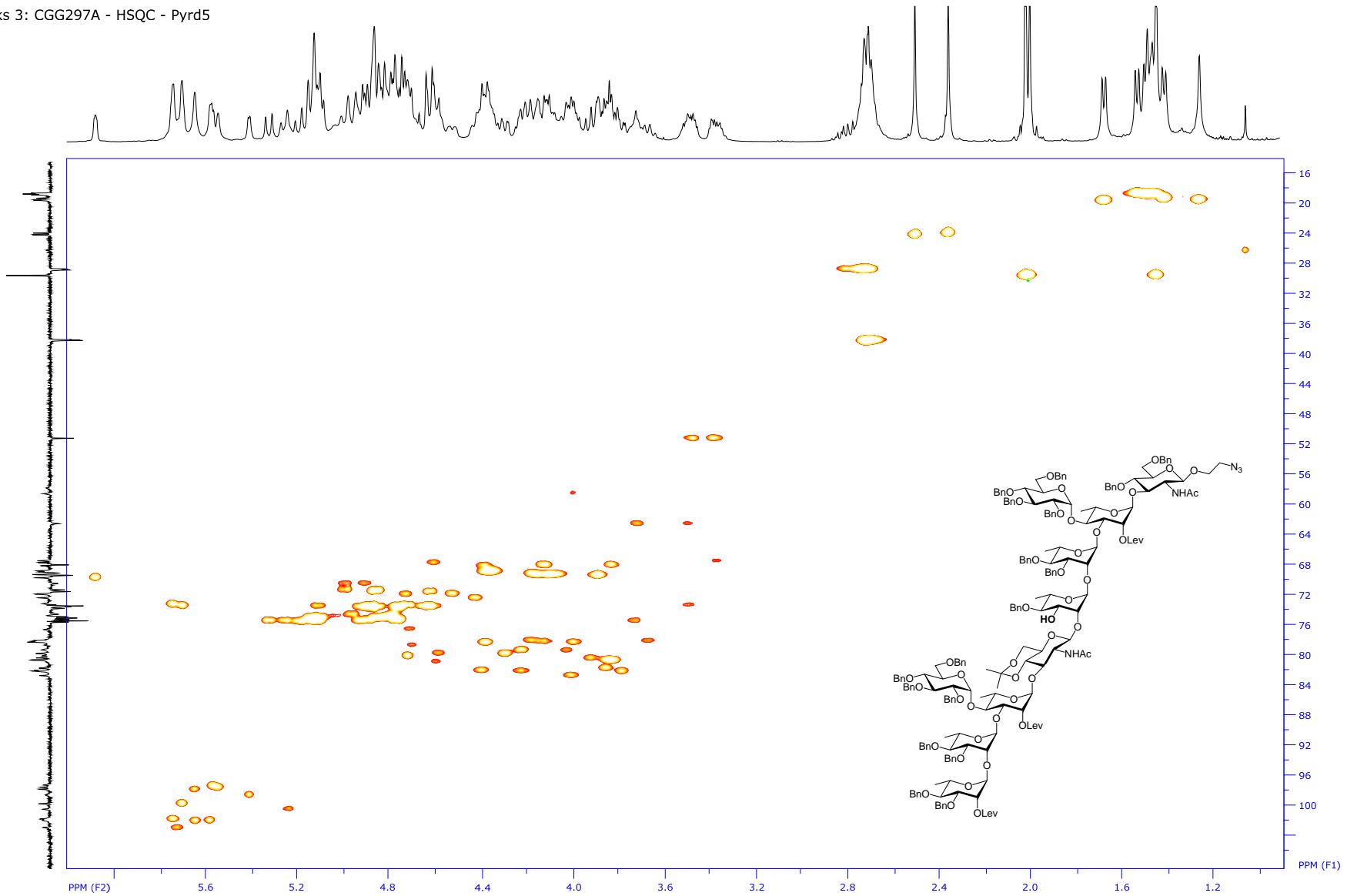
SpinWorks 3: CGG297A - 1H - Pyrd5



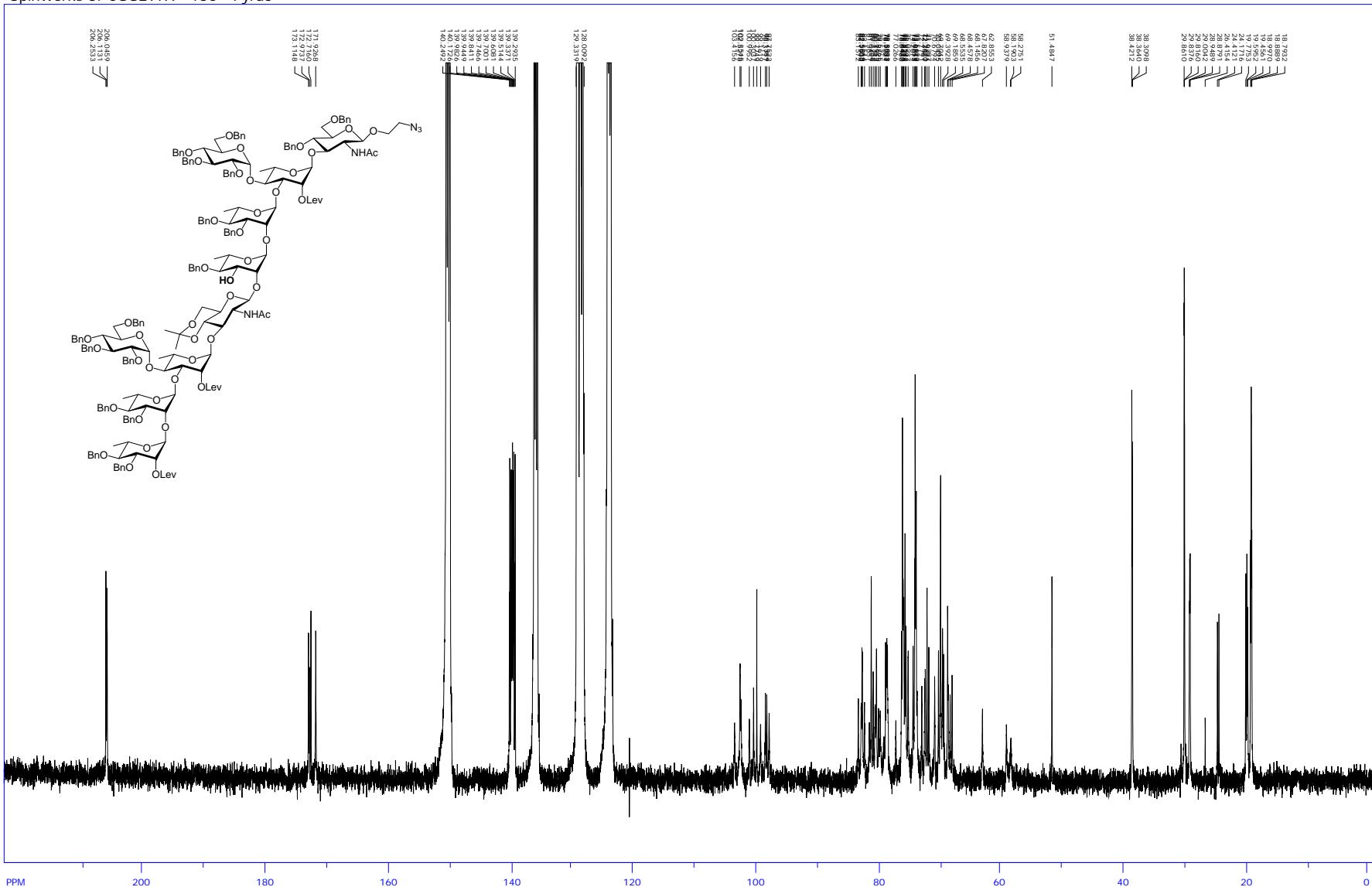
SpinWorks 3: CGG297A - DEPT135 - Pyrd5



SpinWorks 3: CGG297A - HSQC - Pyrd5

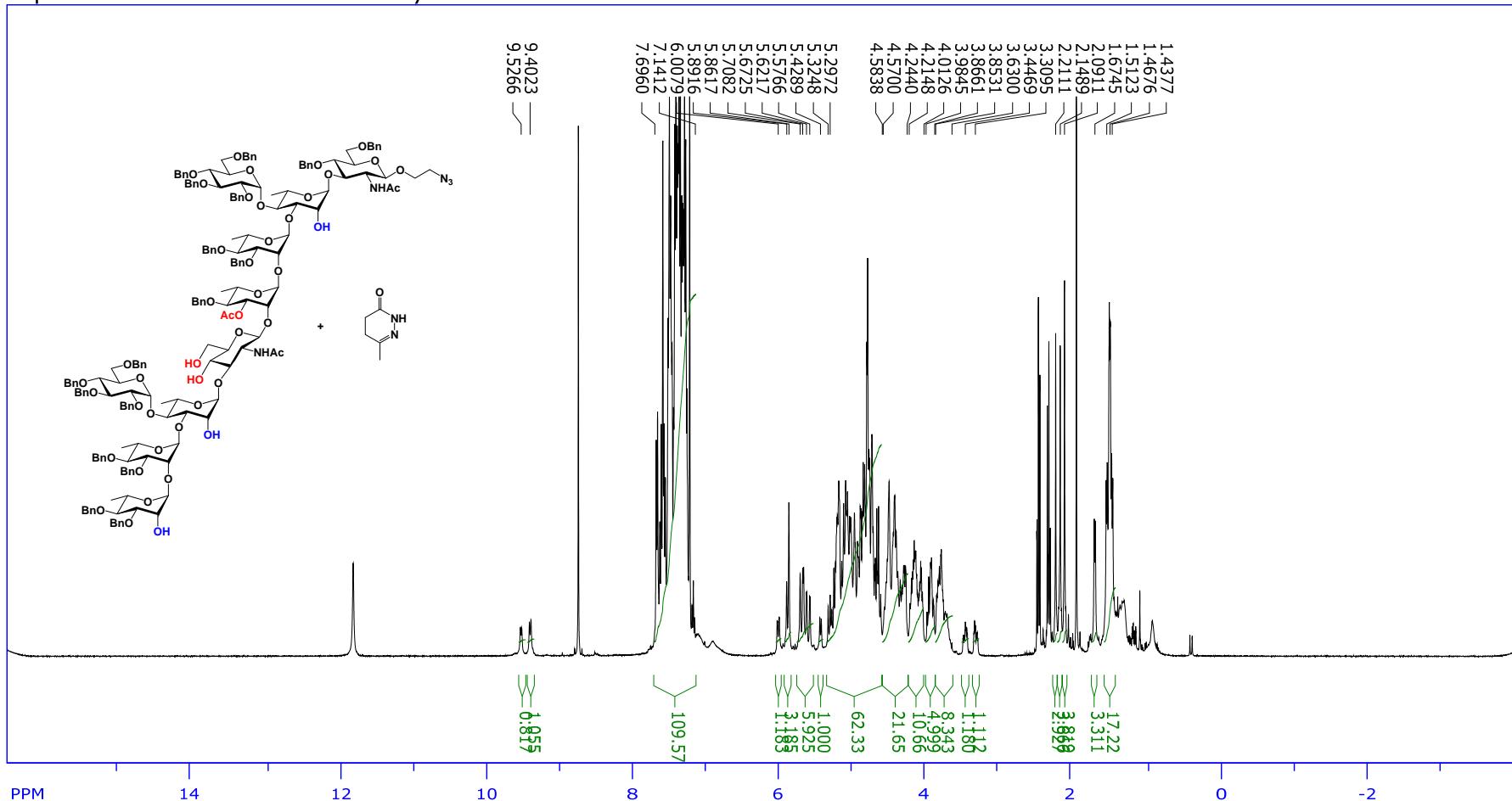


SpinWorks 3: CGG297A - 13C - Pyrd5

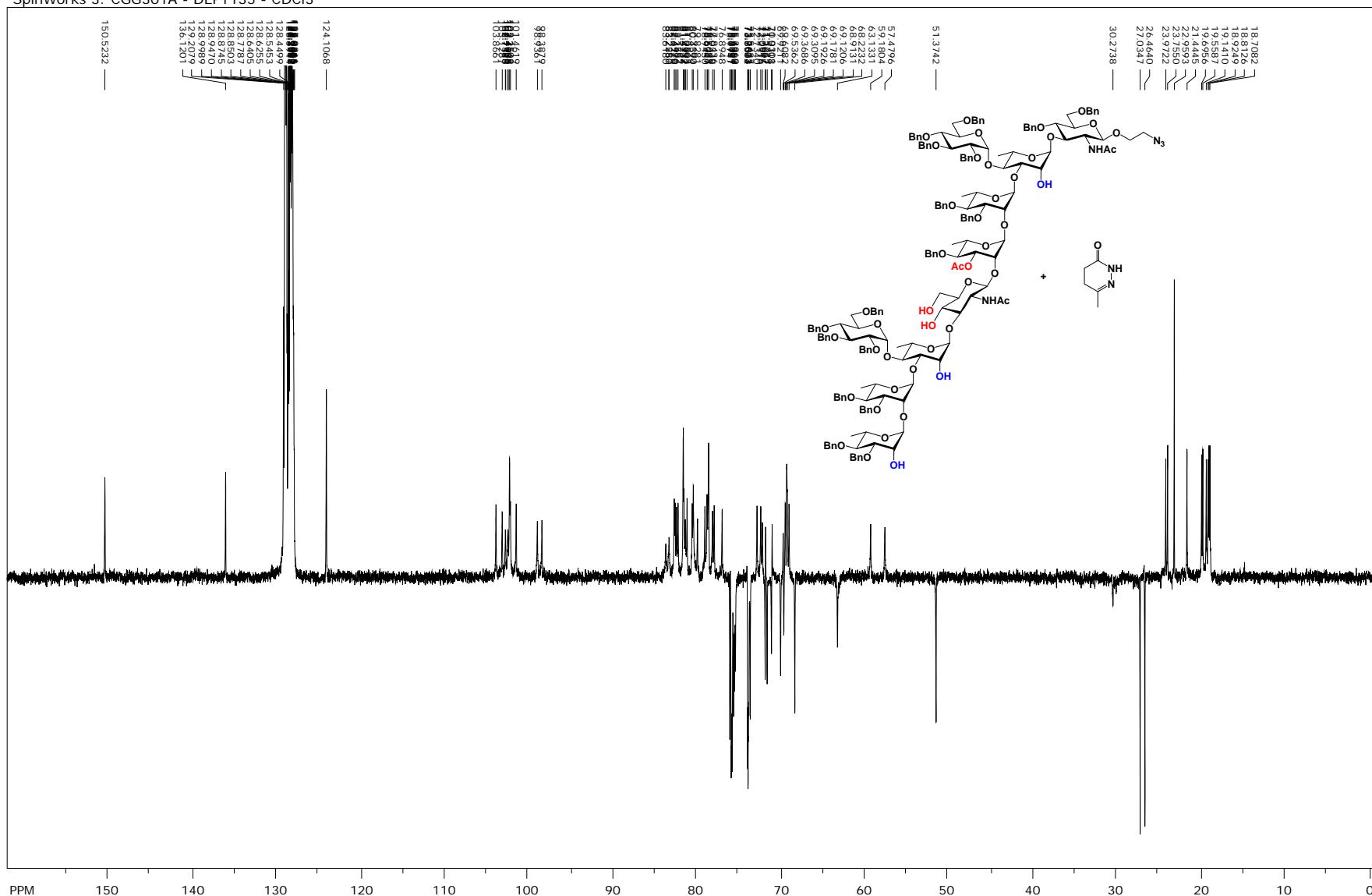


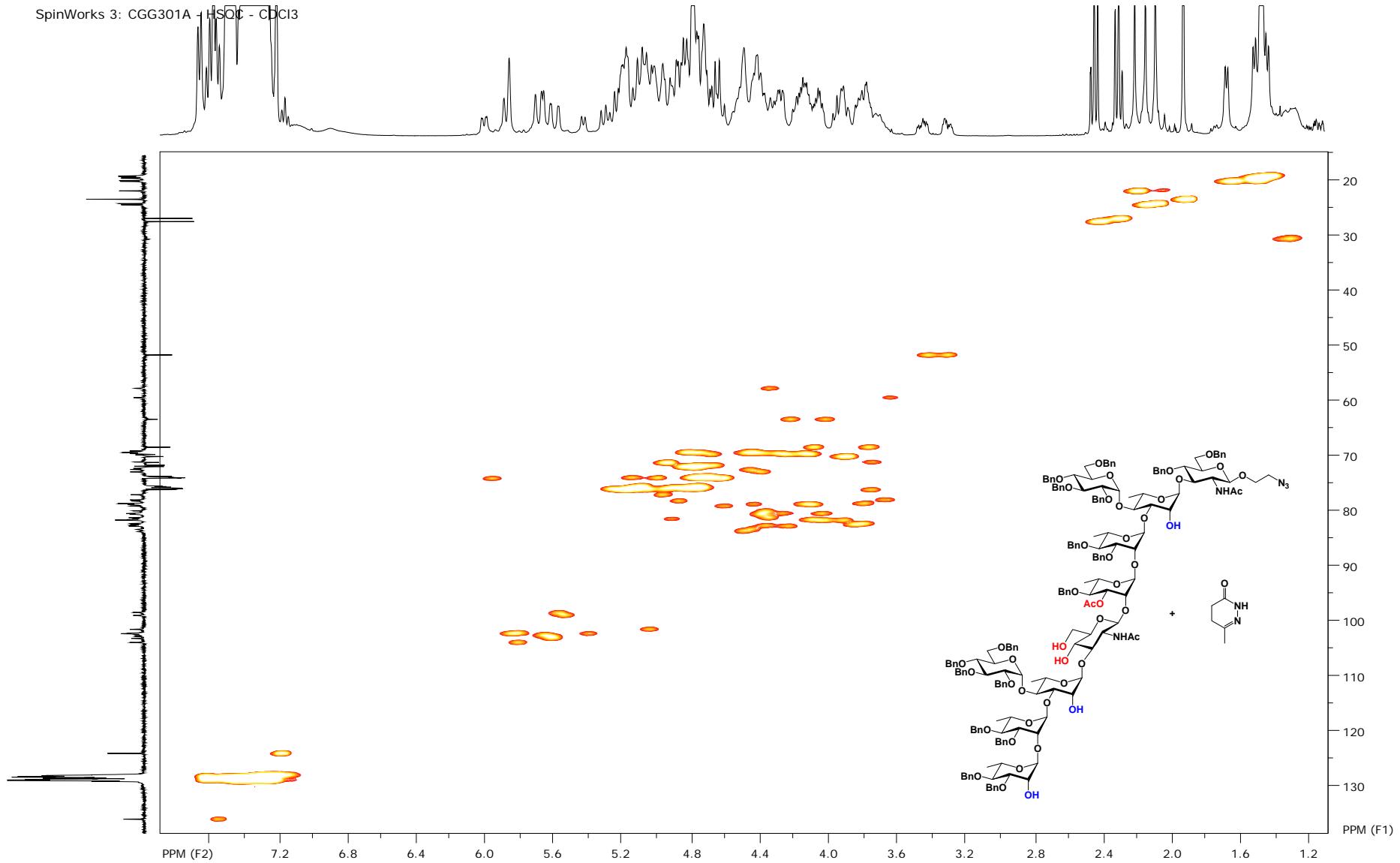
Compound 38

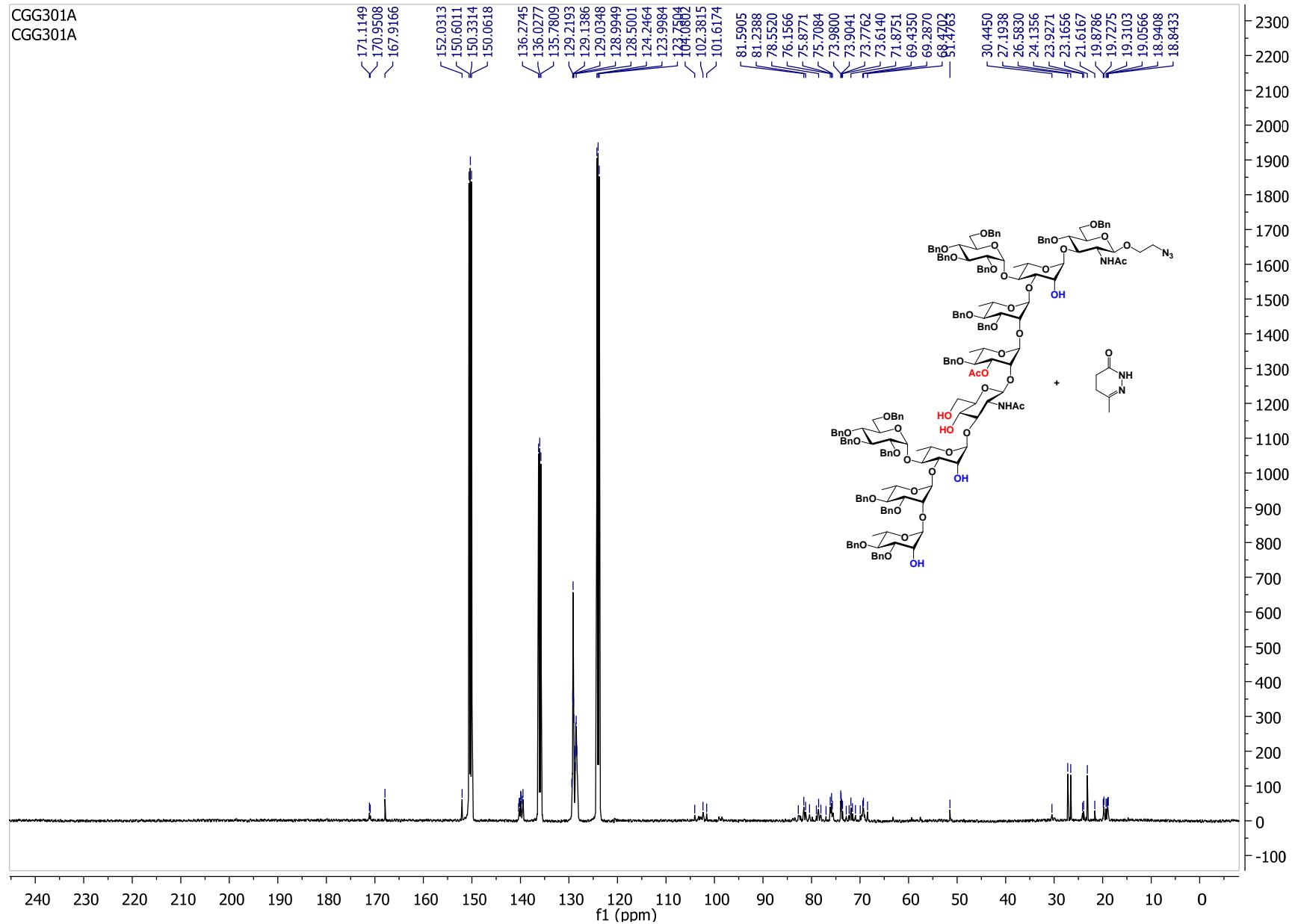
SpinWorks 3: CGG301A - 1H - Pyrd5



SpinWorks 3: CGG301A - DEPT135 - CDCI3

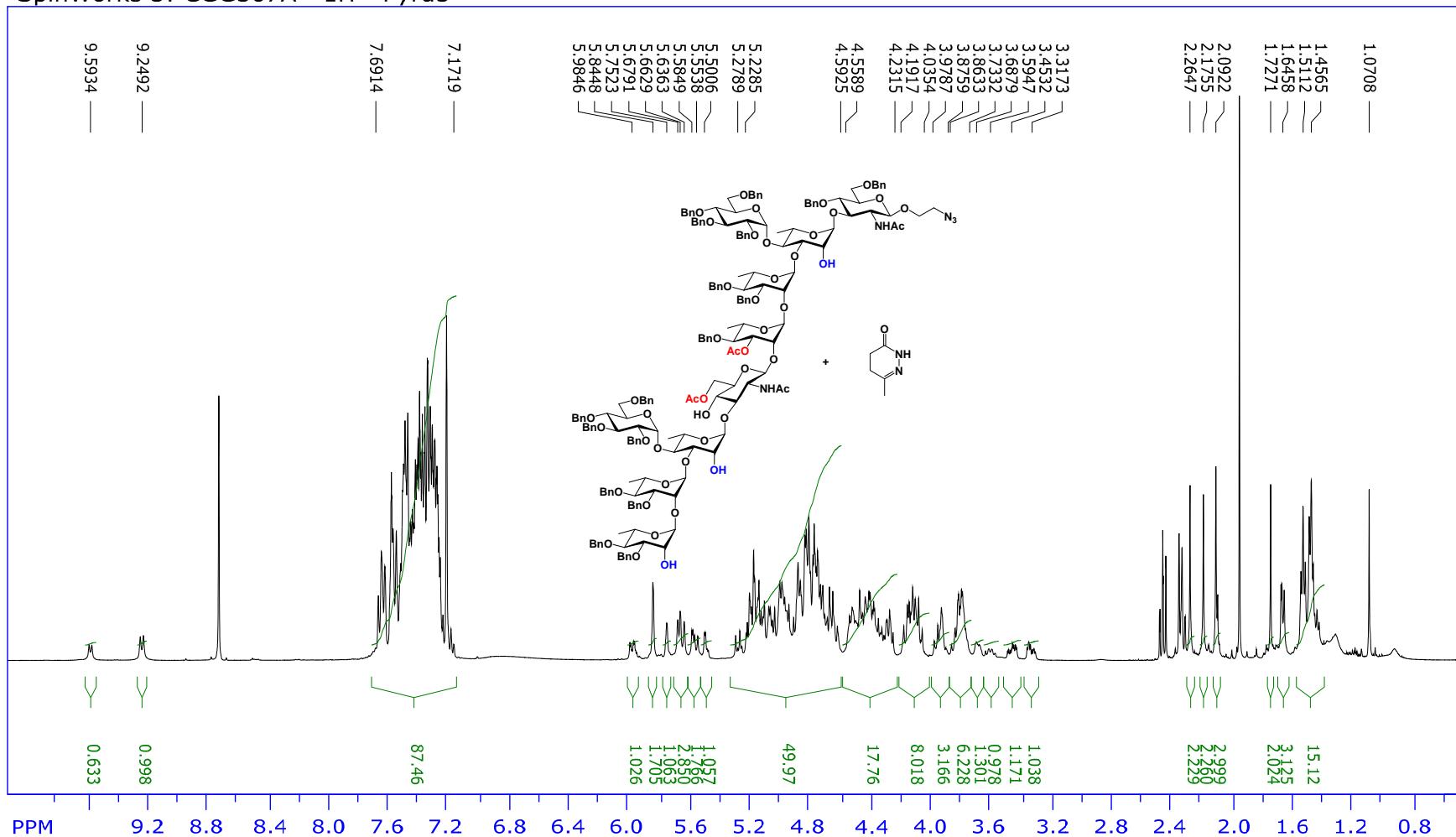




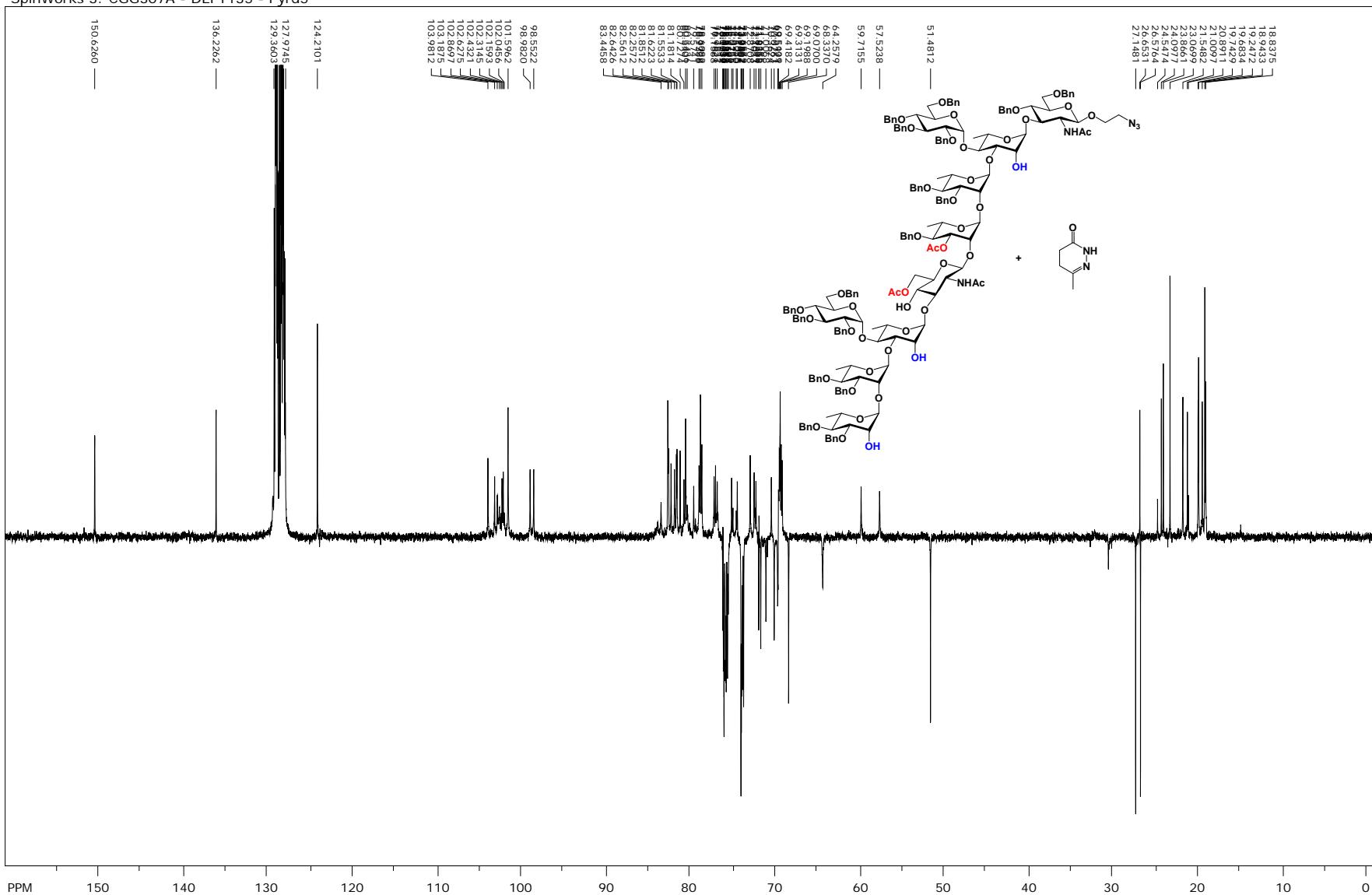


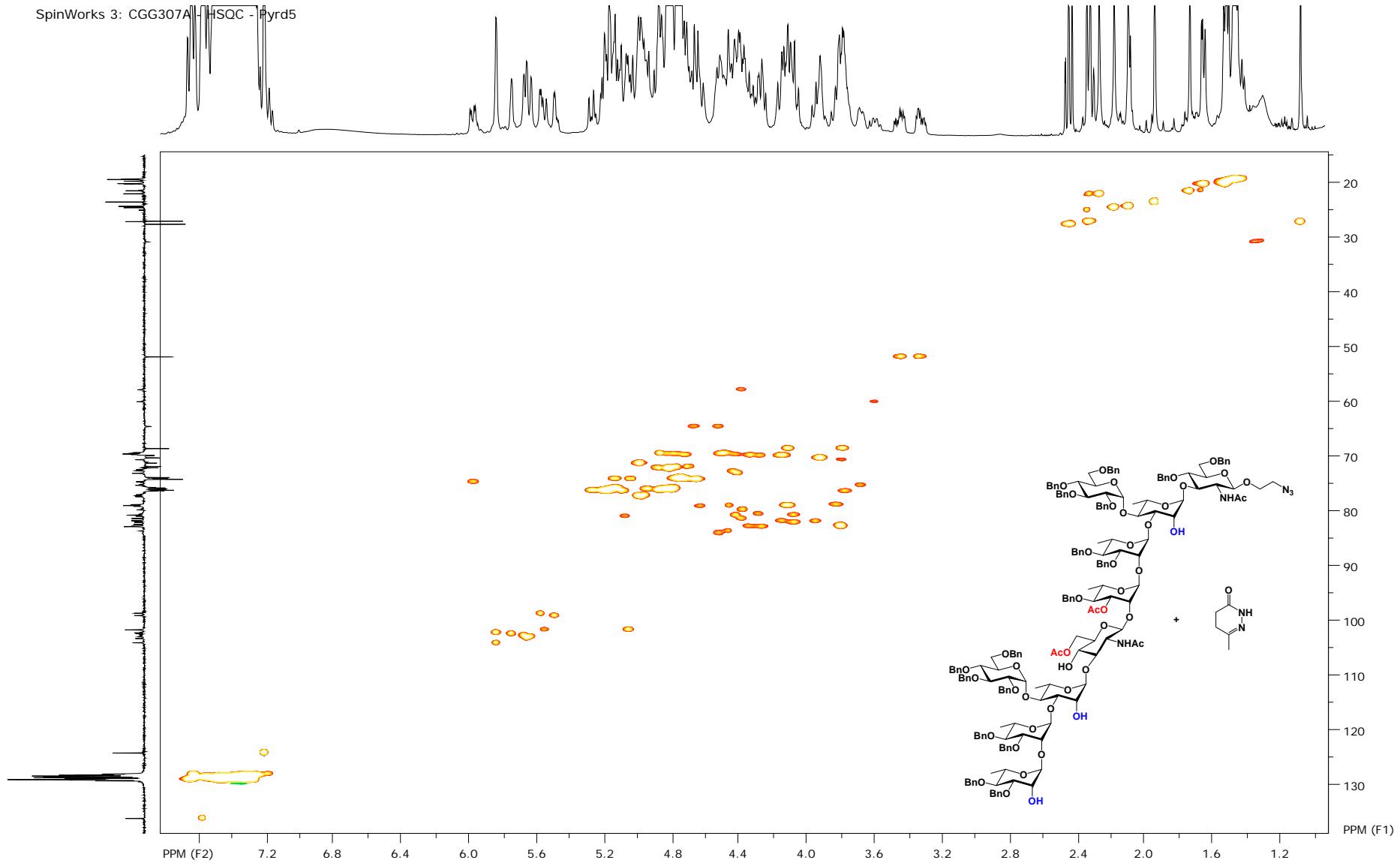
Compound 39

SpinWorks 3: CGG307A - 1H - Pyrd5

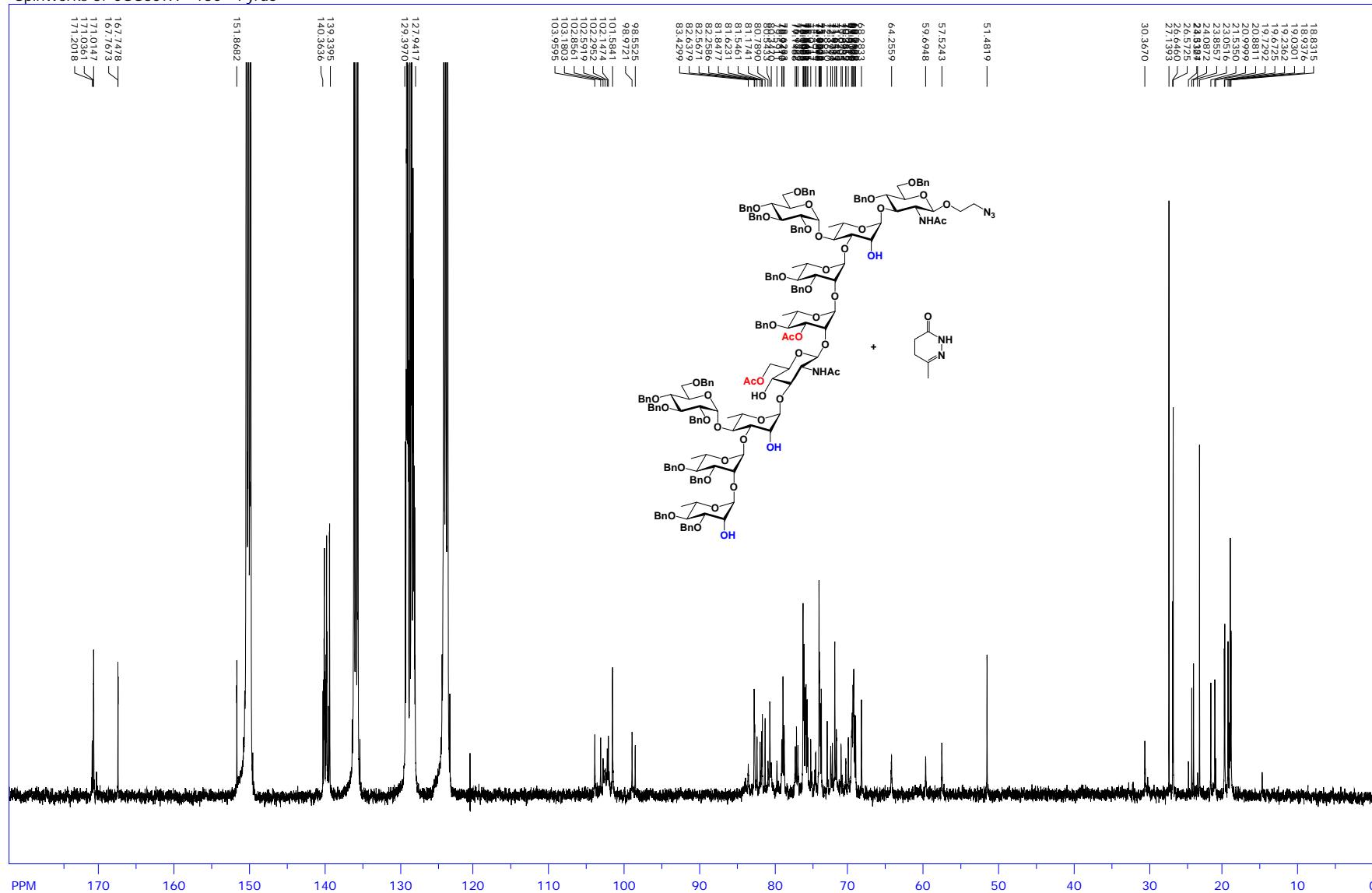


SpinWorks 3: CGG307A - DEPT135 - Pyrd5



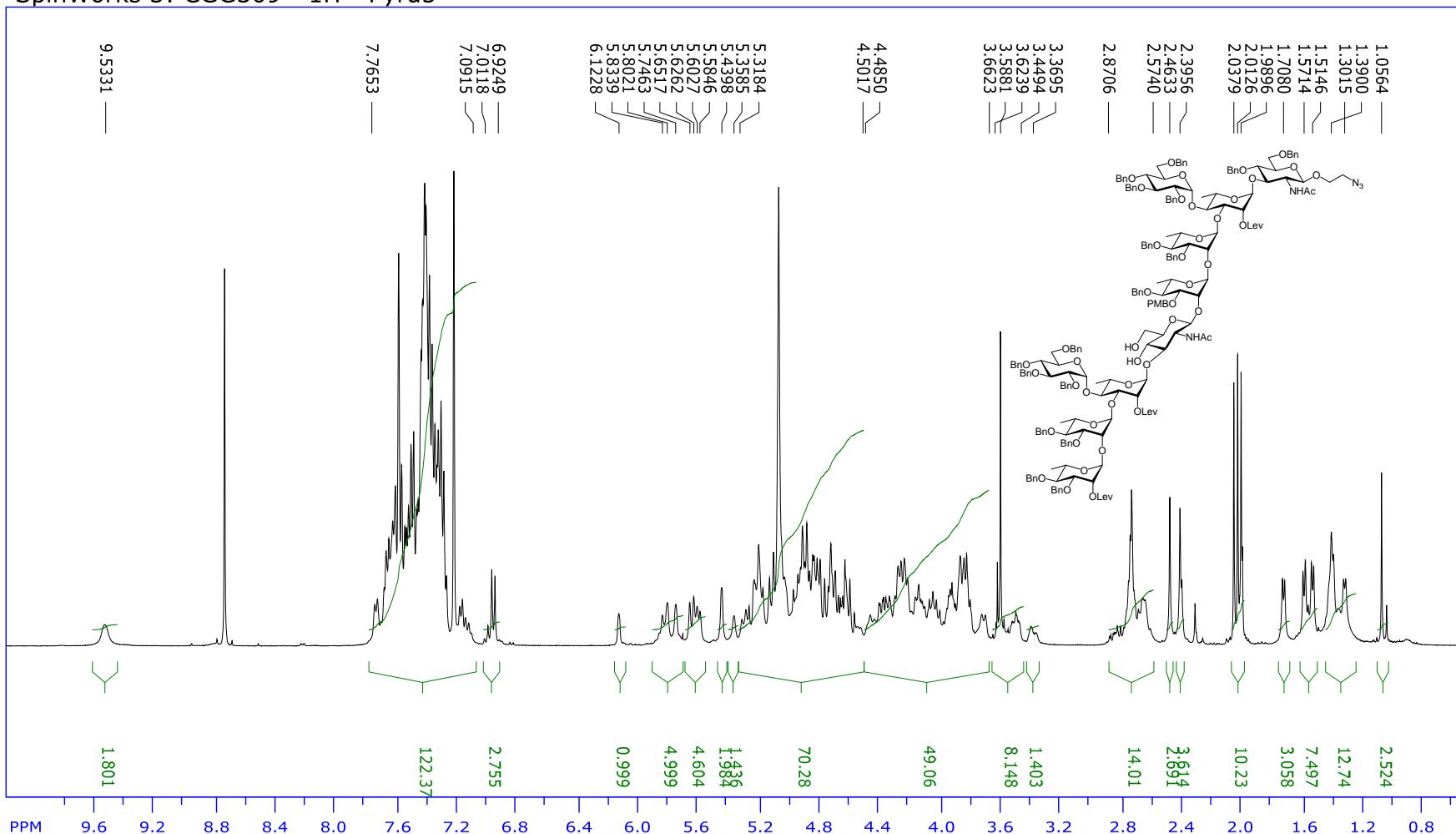


SpinWorks 3: CGG307A - 13C - Pyrd5

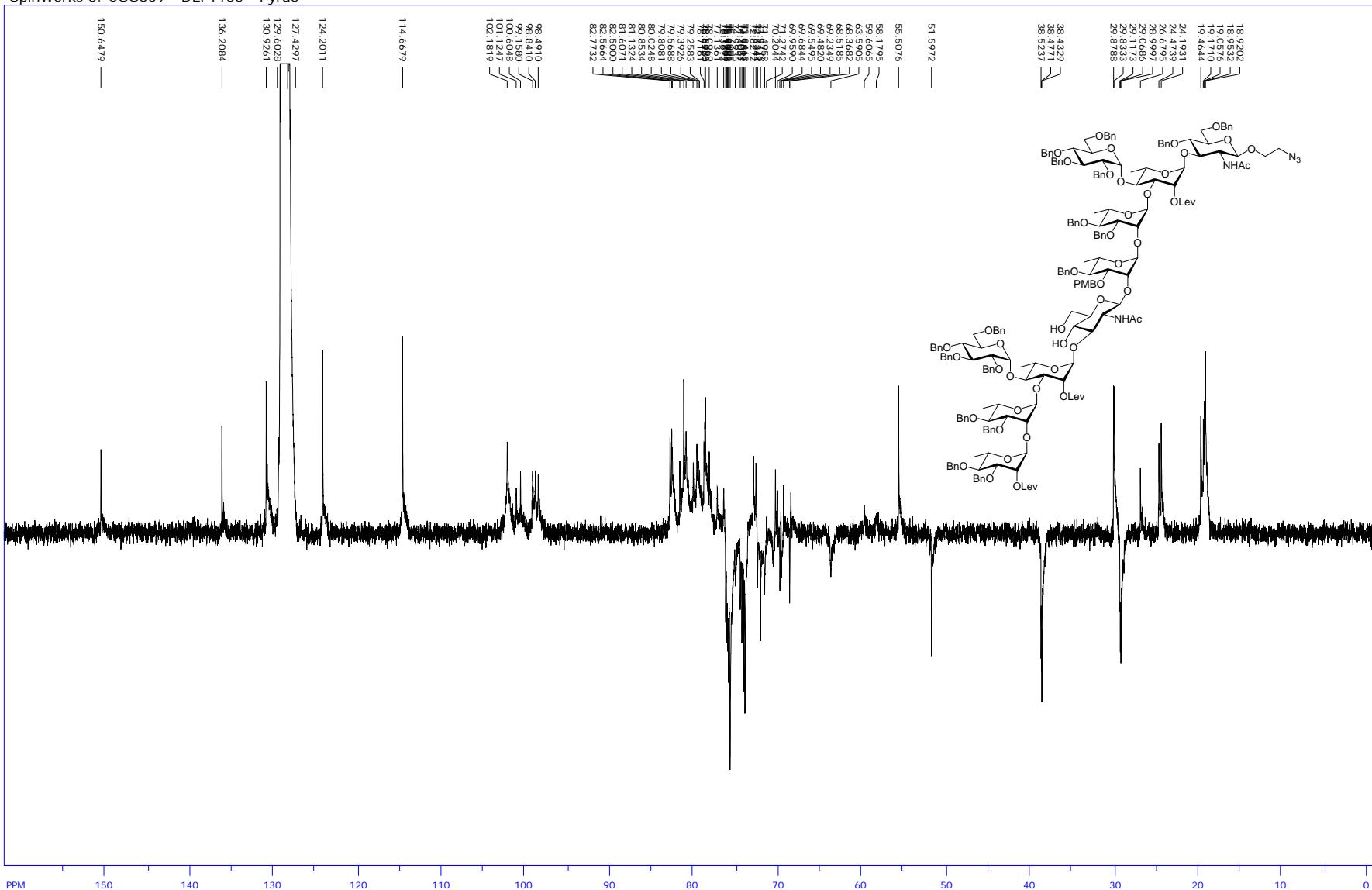


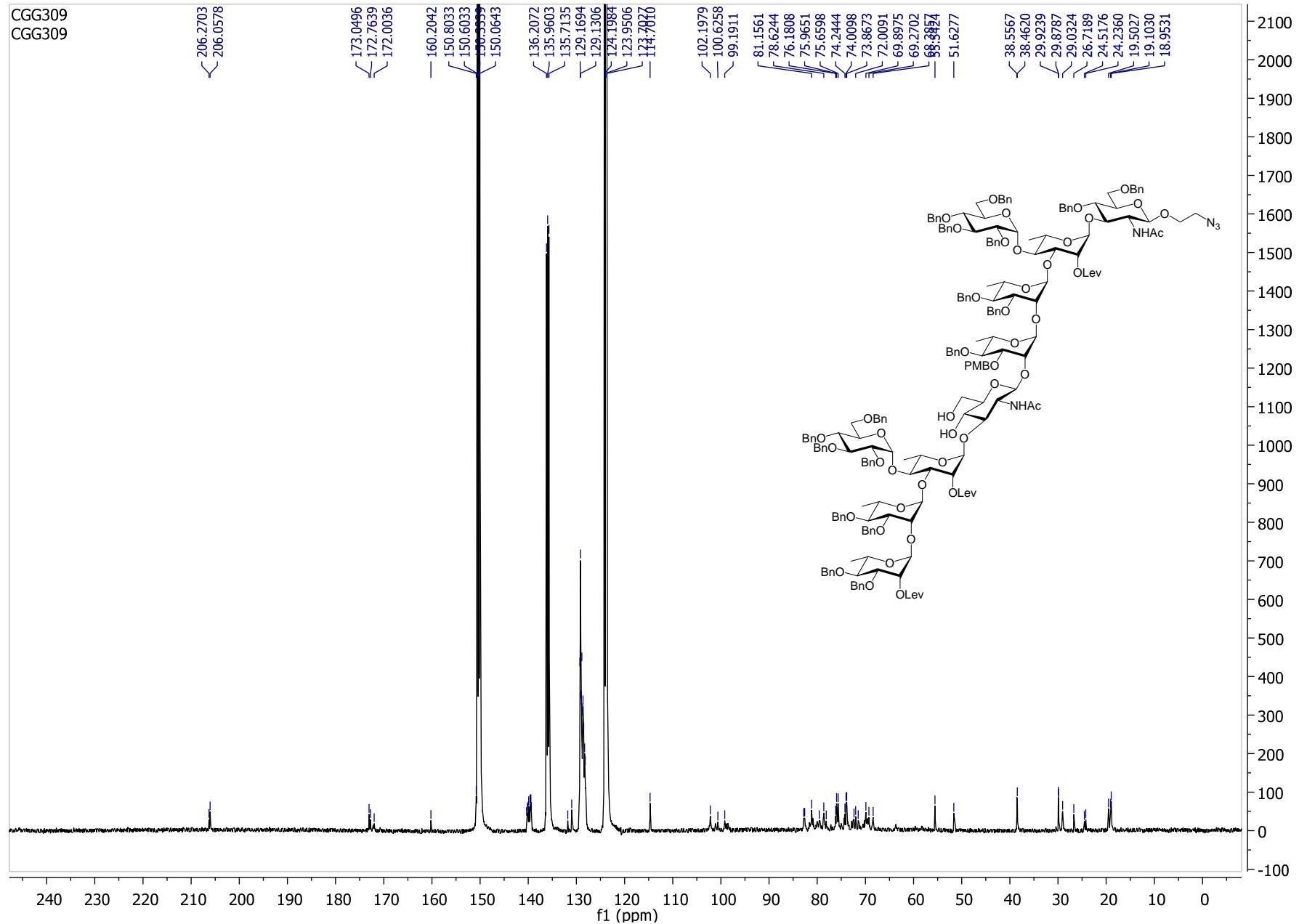
Compound 40

SpinWorks 3: CGG309 - 1H - Pyrd5

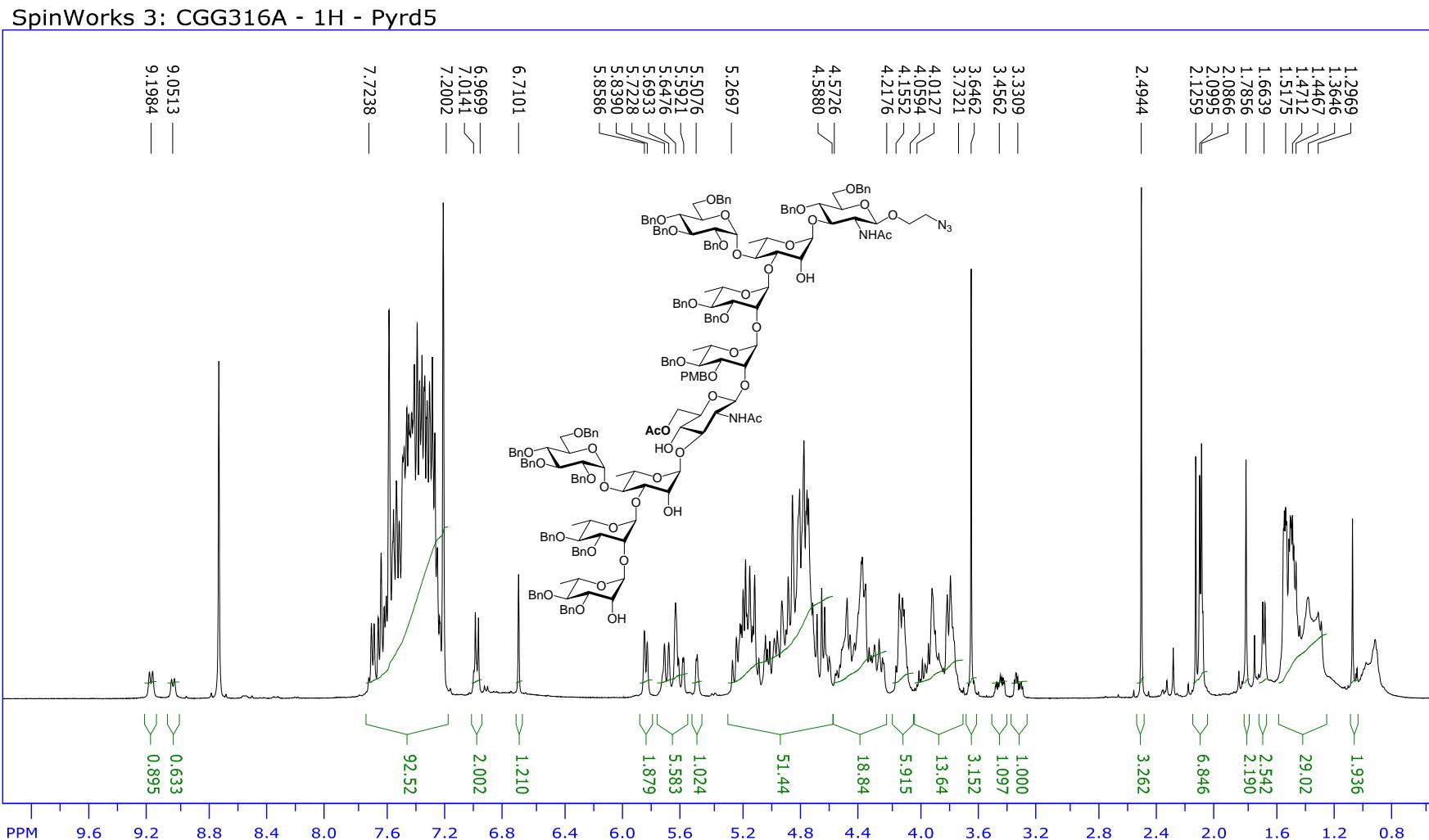


SpinWorks 3: CGG309 - DEPT135 - Pyrd5

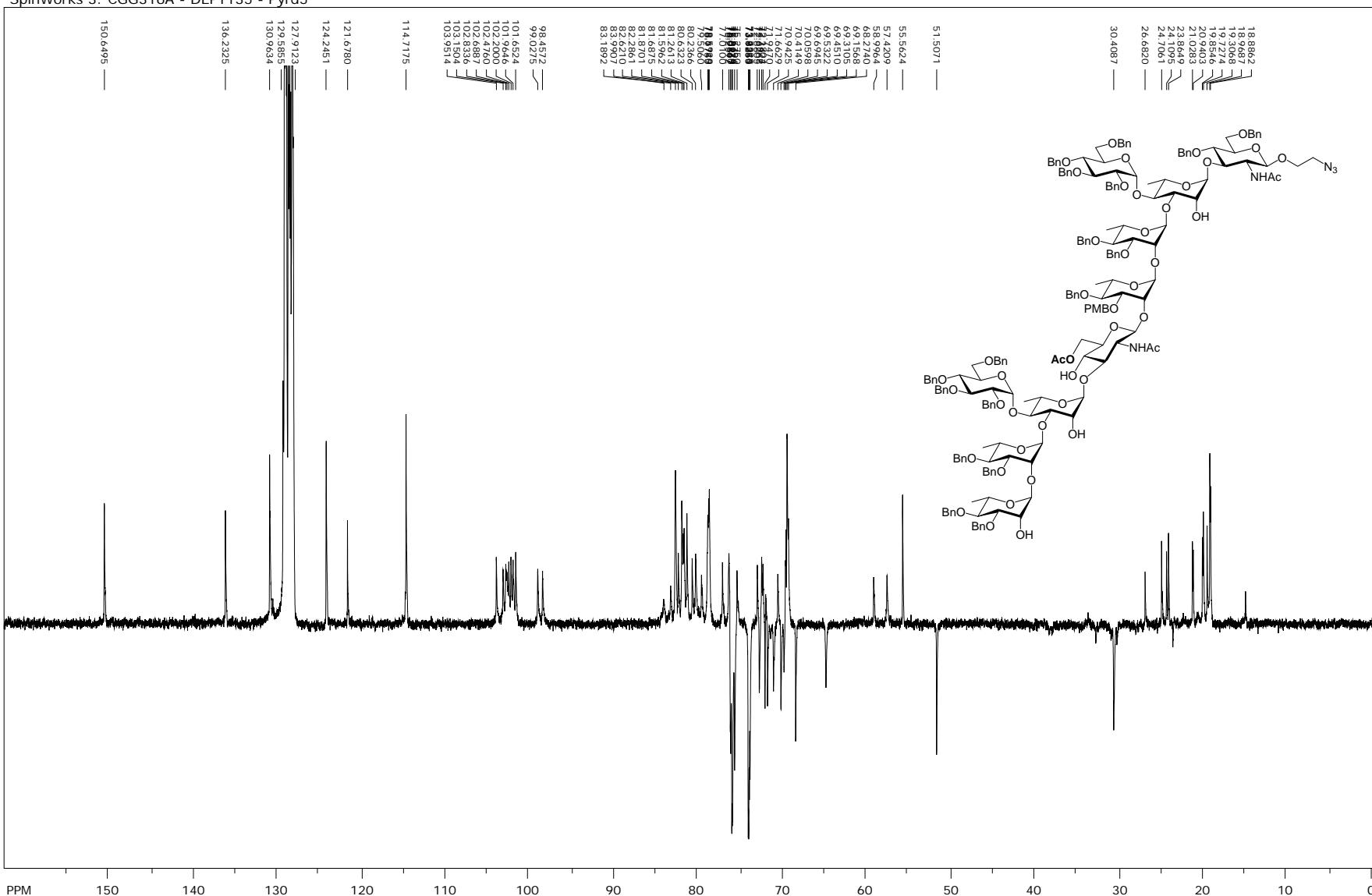


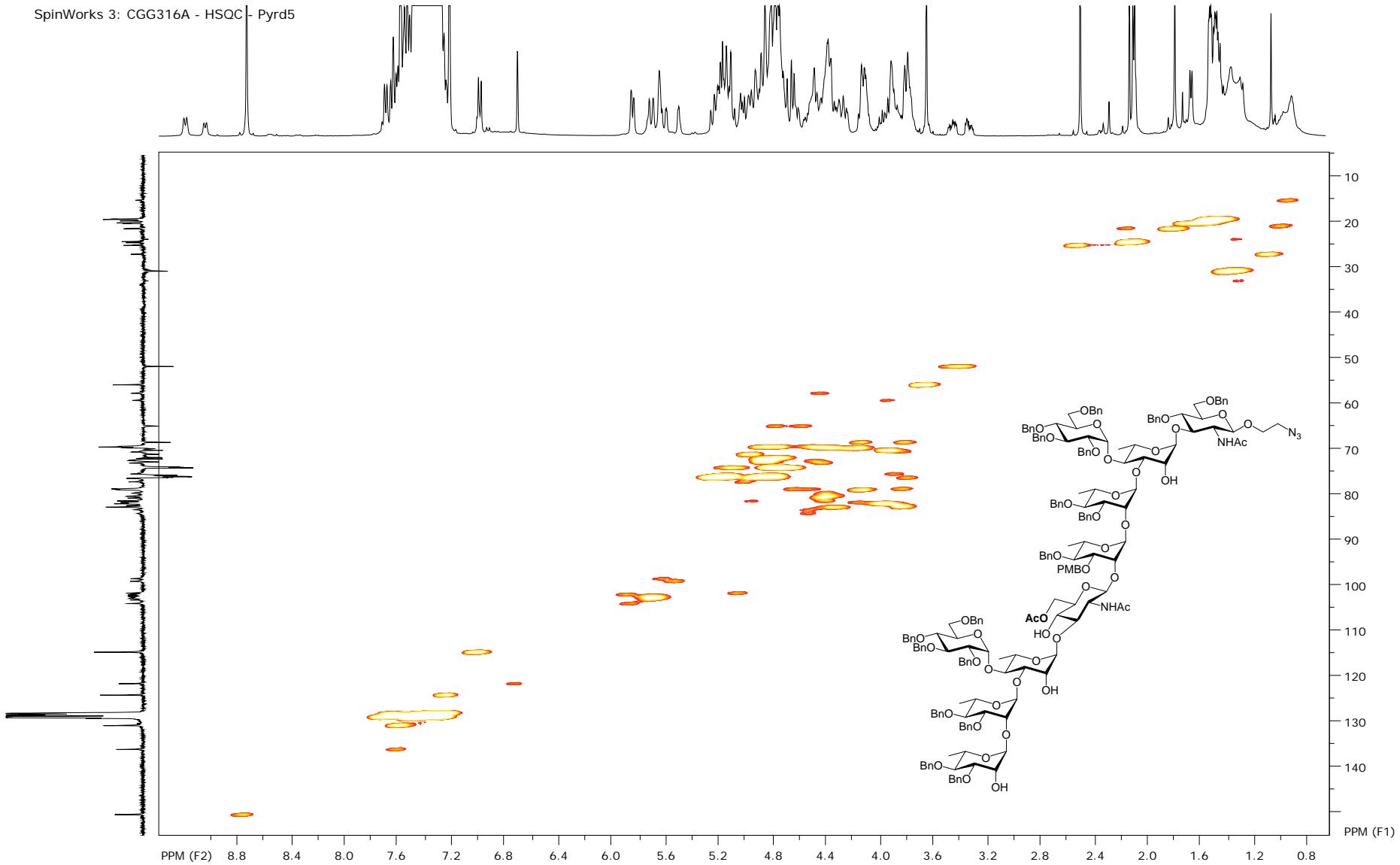


Compound 41

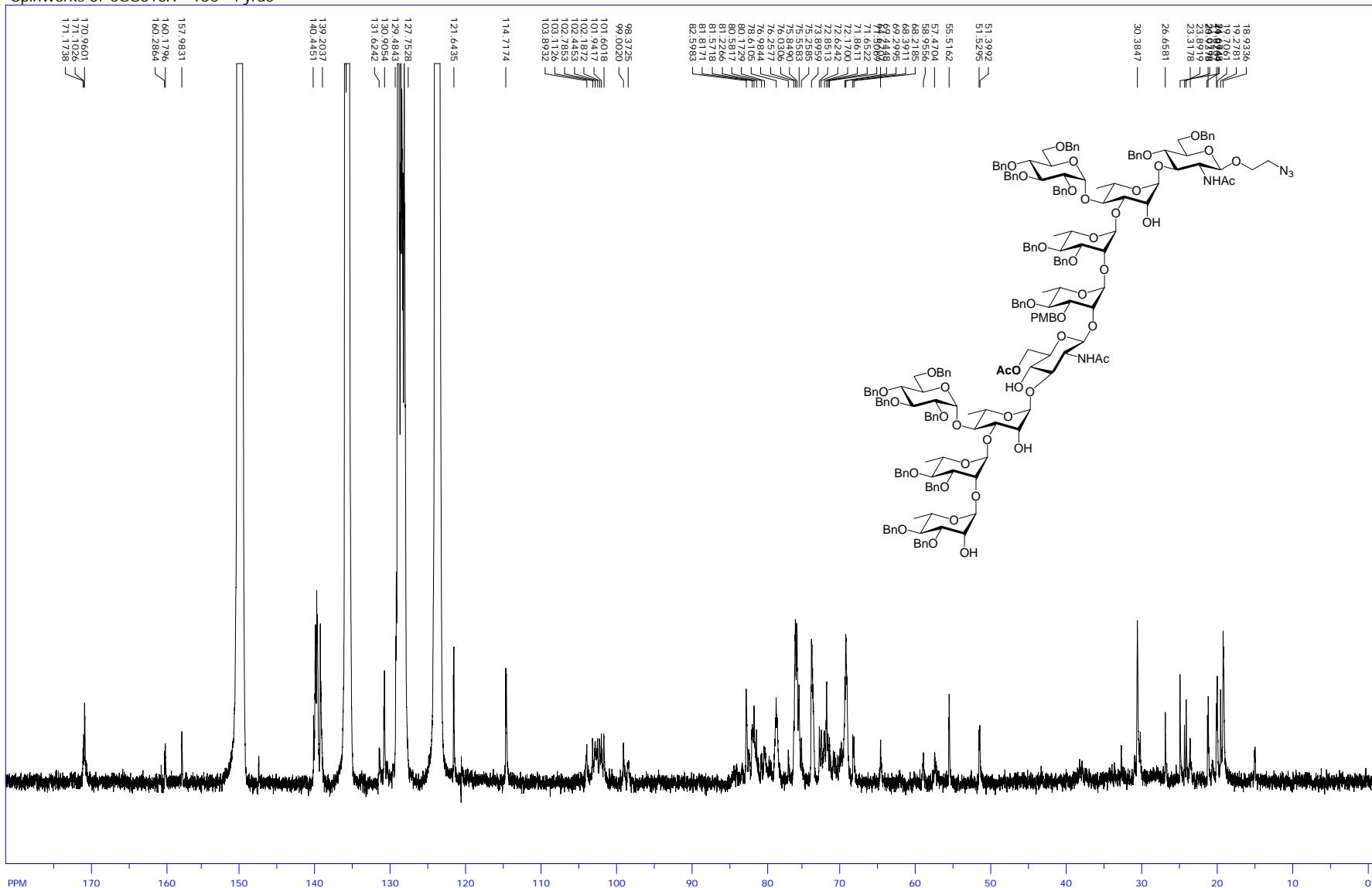


SpinWorks 3: CGG316A - DEPT135 - Pyrd5

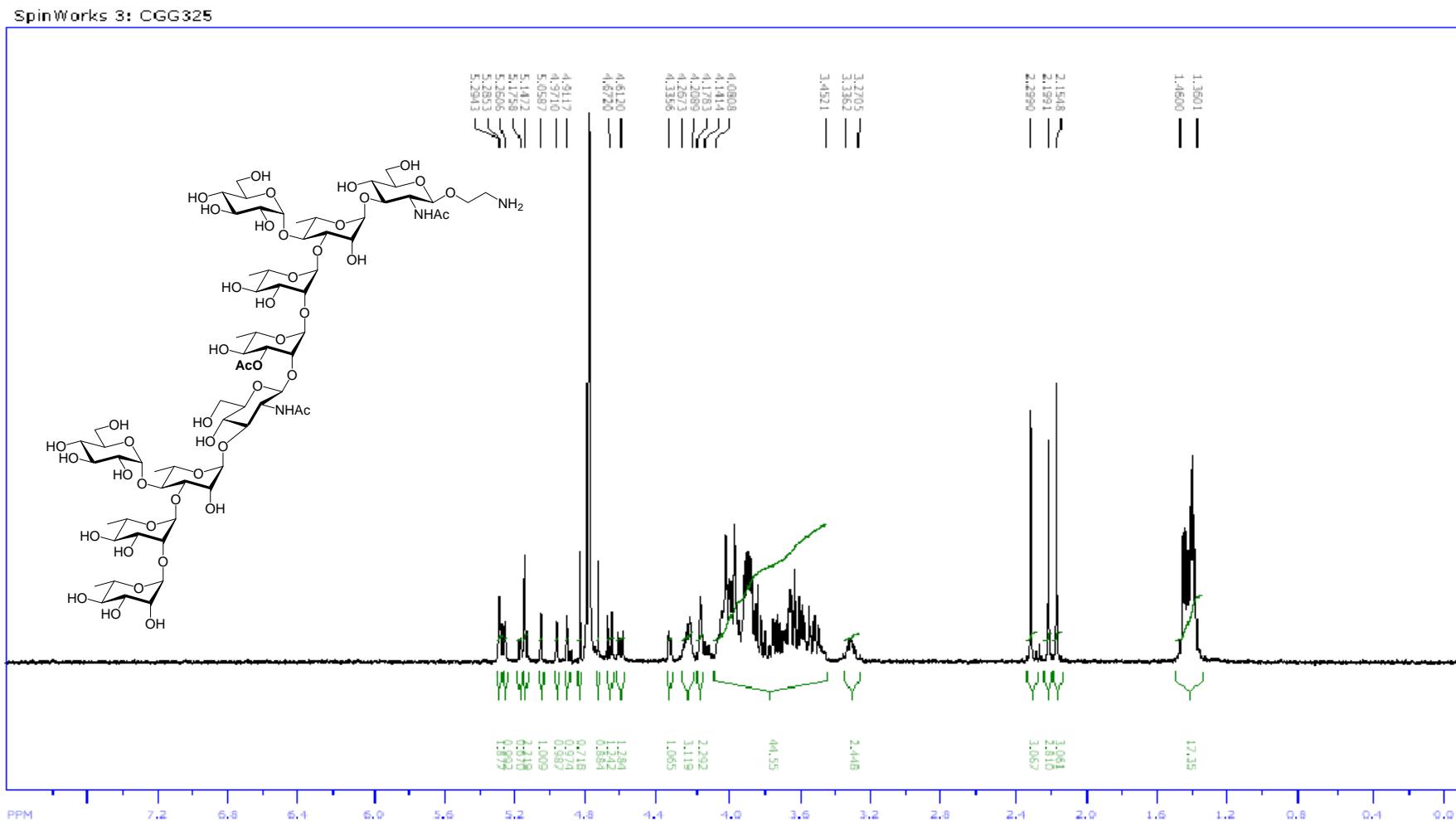




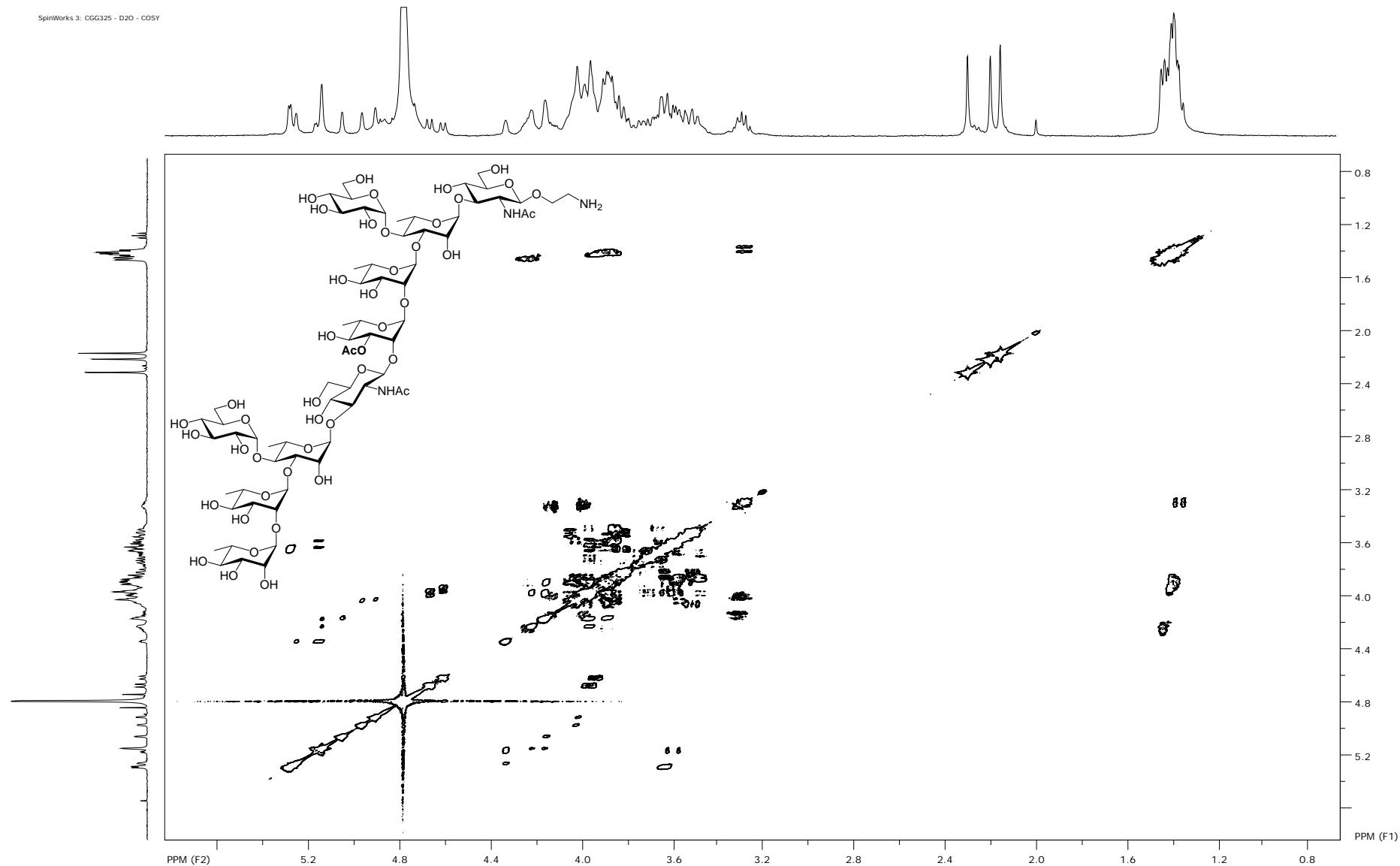
SpinWorks 3: CGG316A - 13C - Pyrd5



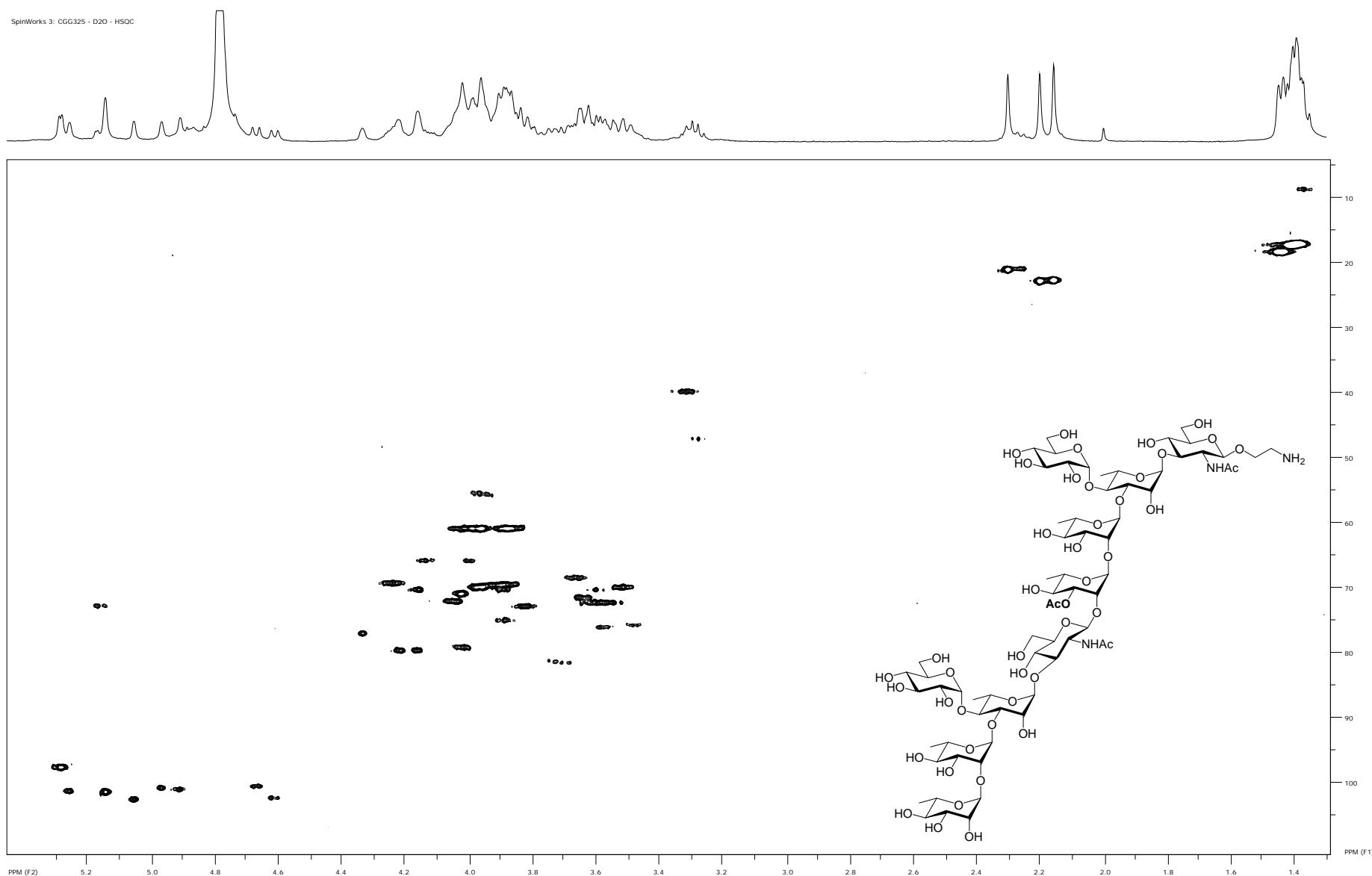
Compound 1



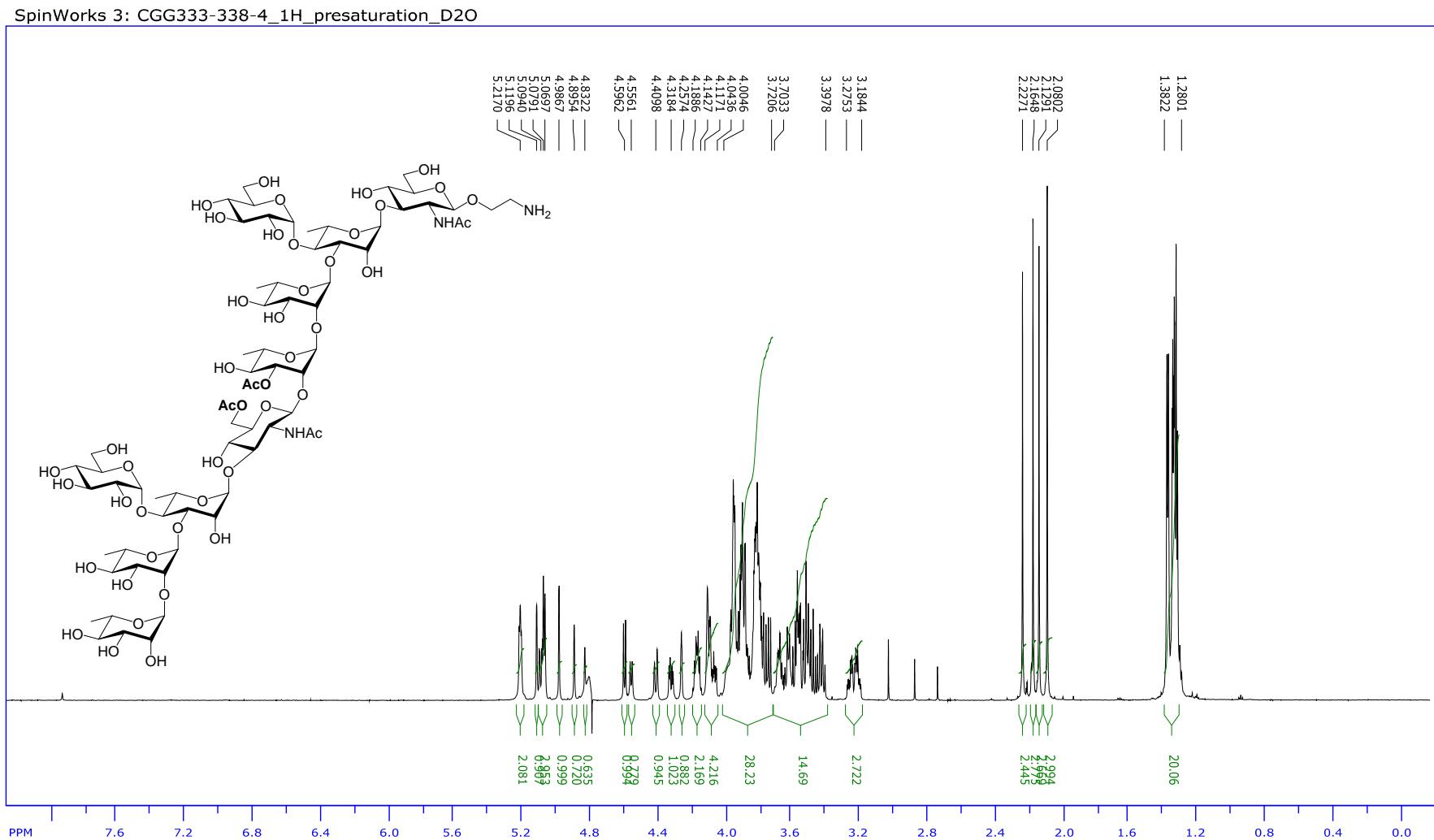
SpinWorks 3: CGG325 - D₂O - COSY



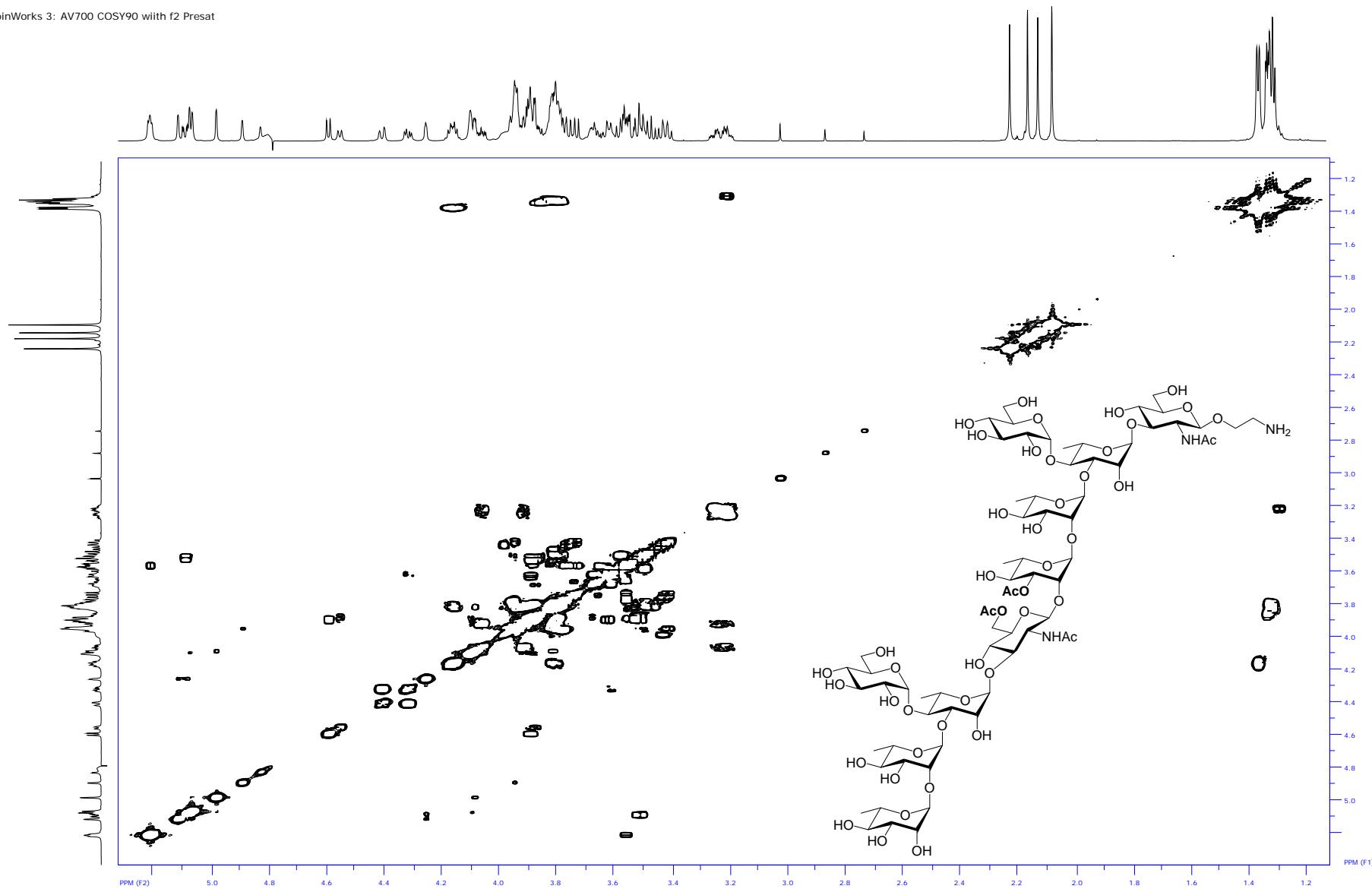
SpinWorks 3: CGG325 - D2O - HSQC



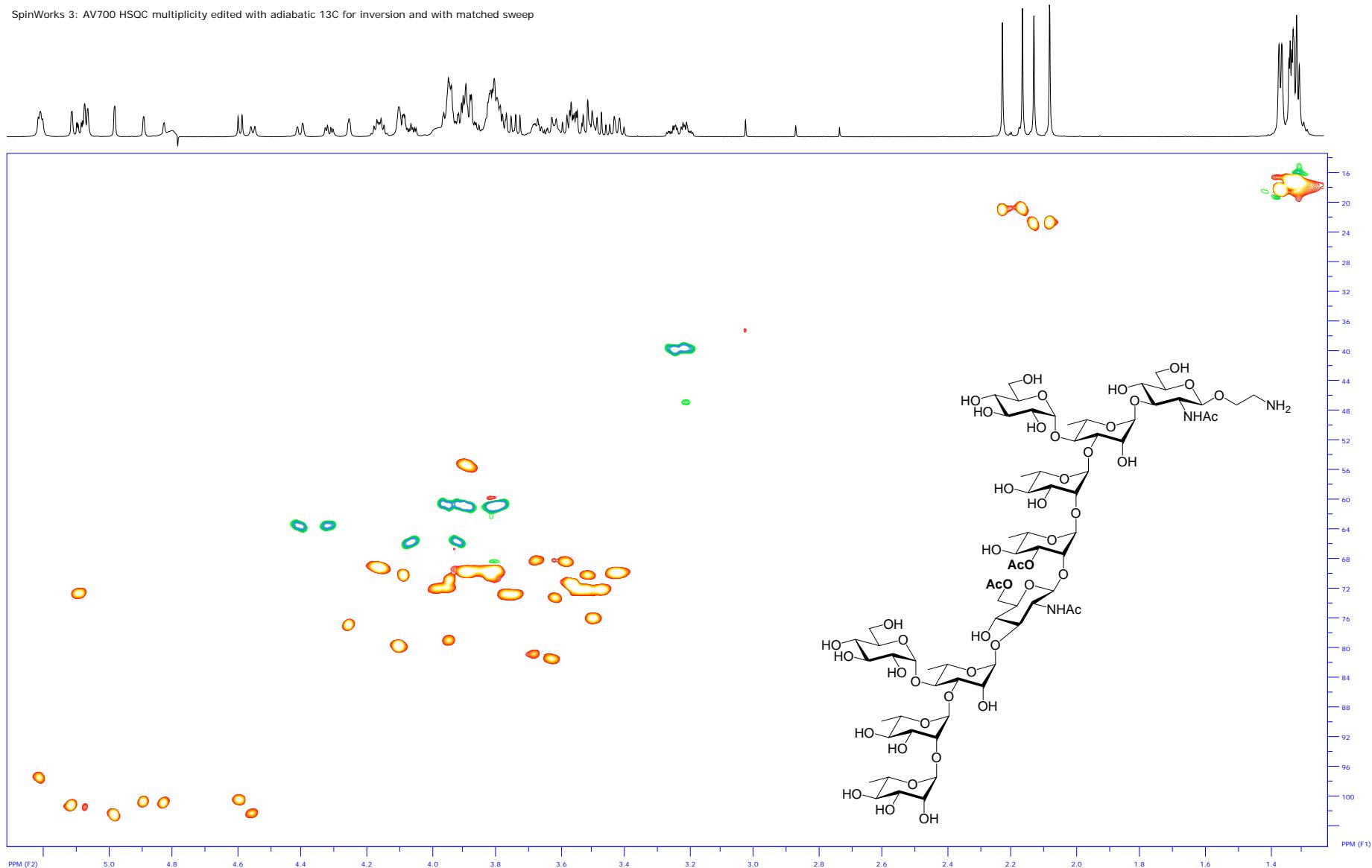
Compound 2



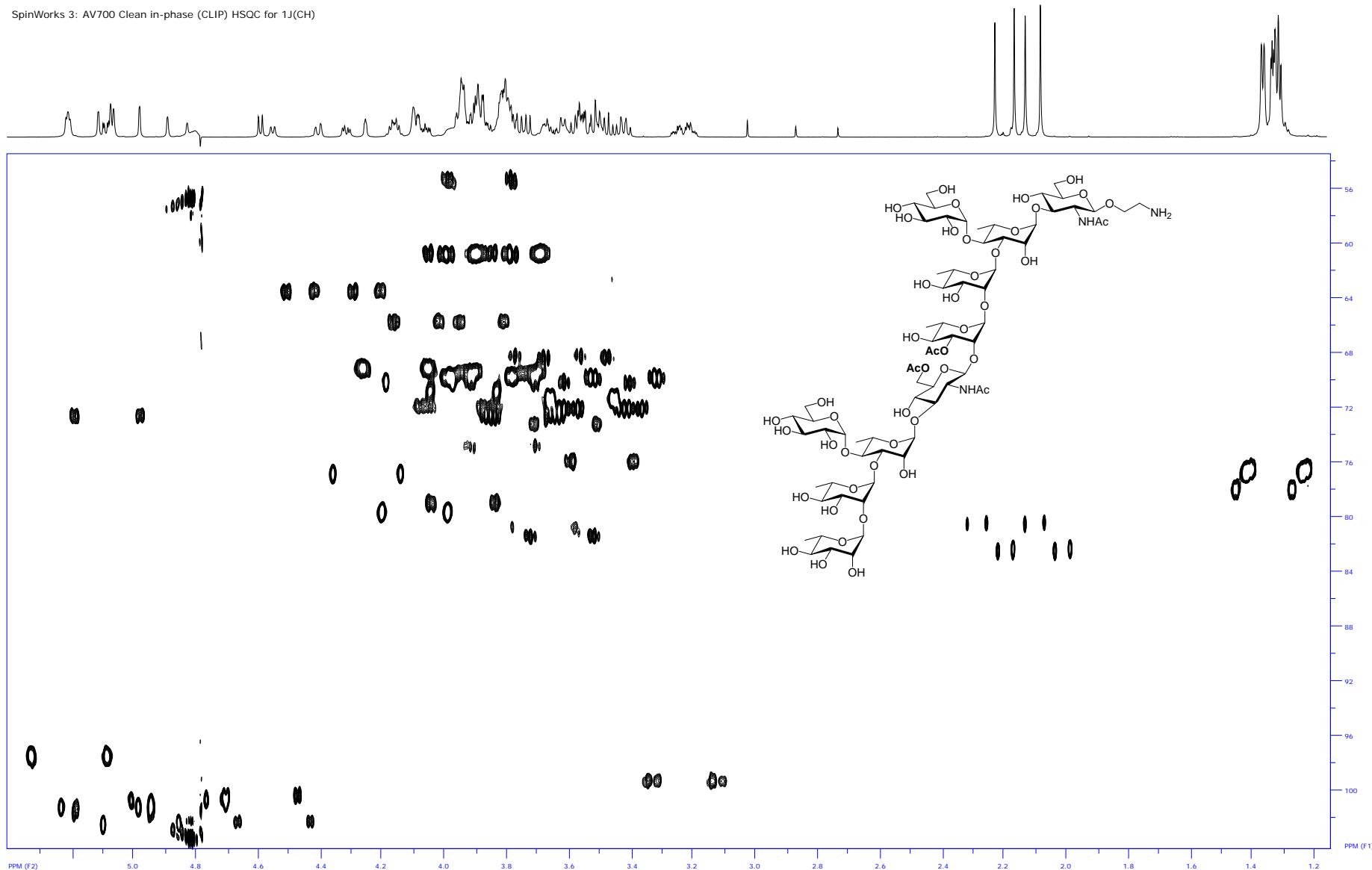
SpinWorks 3: AV700 COSY90 with f2 Presat



SpinWorks 3: AV700 HSQC multiplicity edited with adiabatic ^{13}C for inversion and with matched sweep

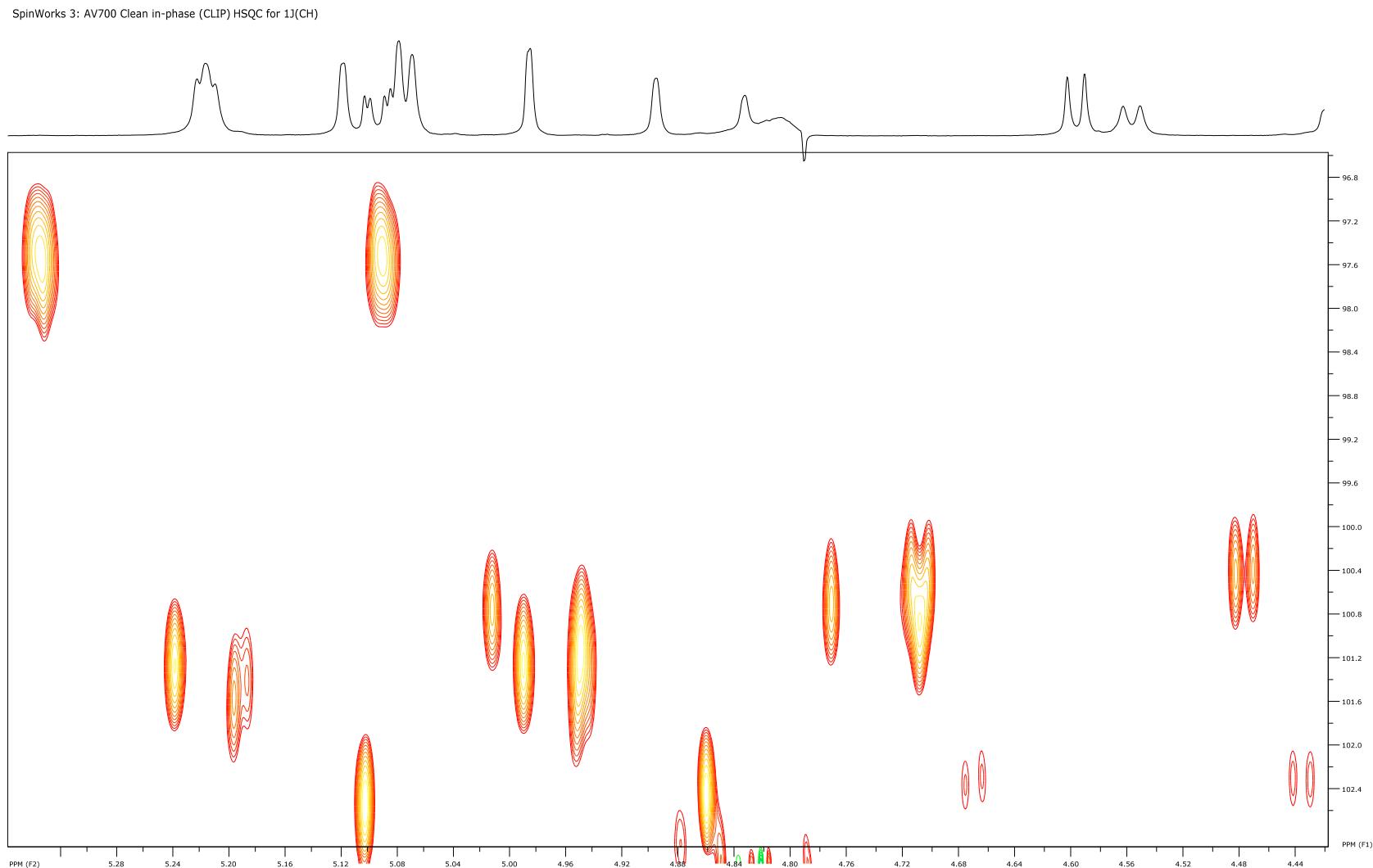


SpinWorks 3: AV700 Clean in-phase (CLIP) HSQC for 1J(CH)

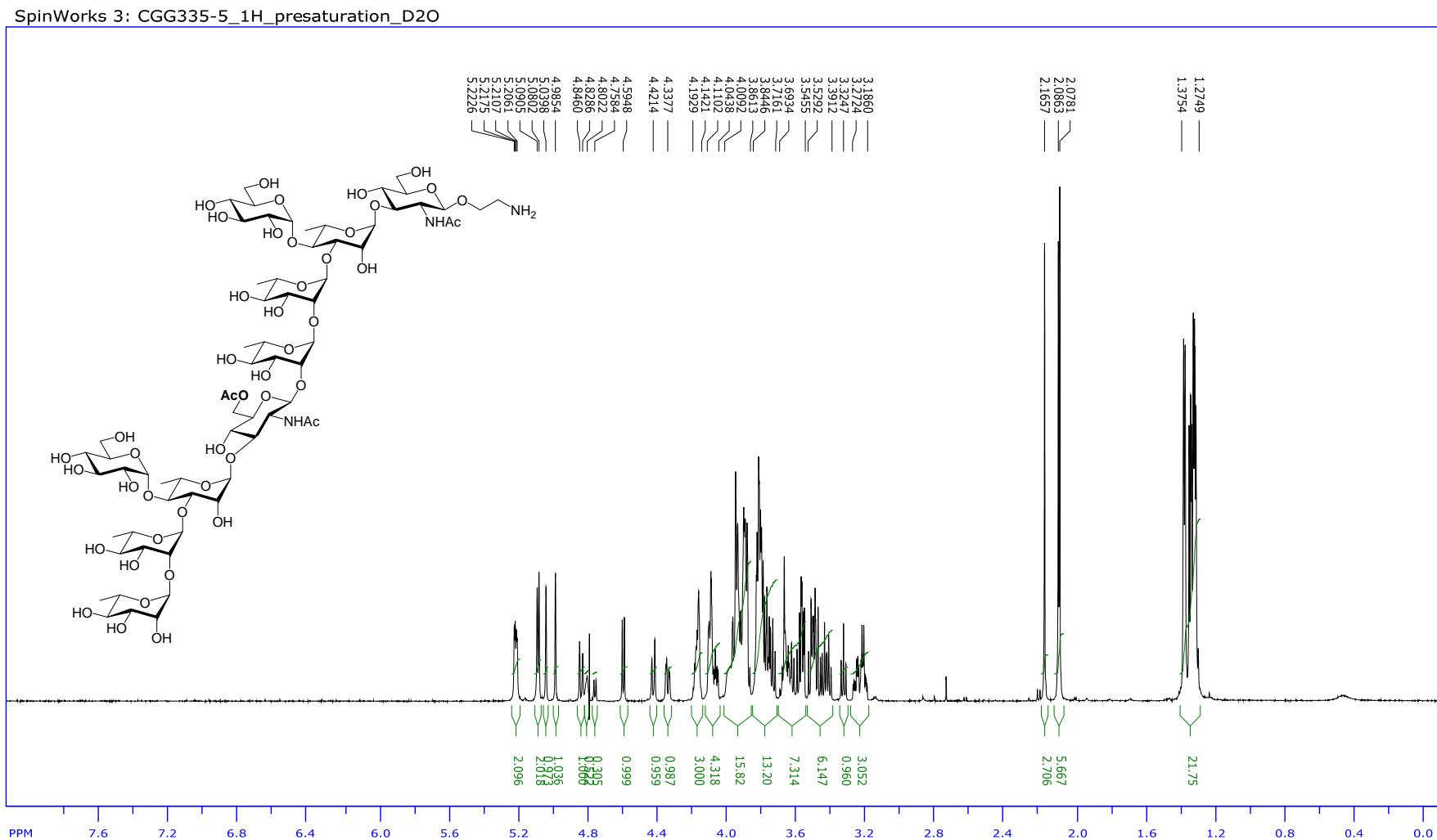


S200

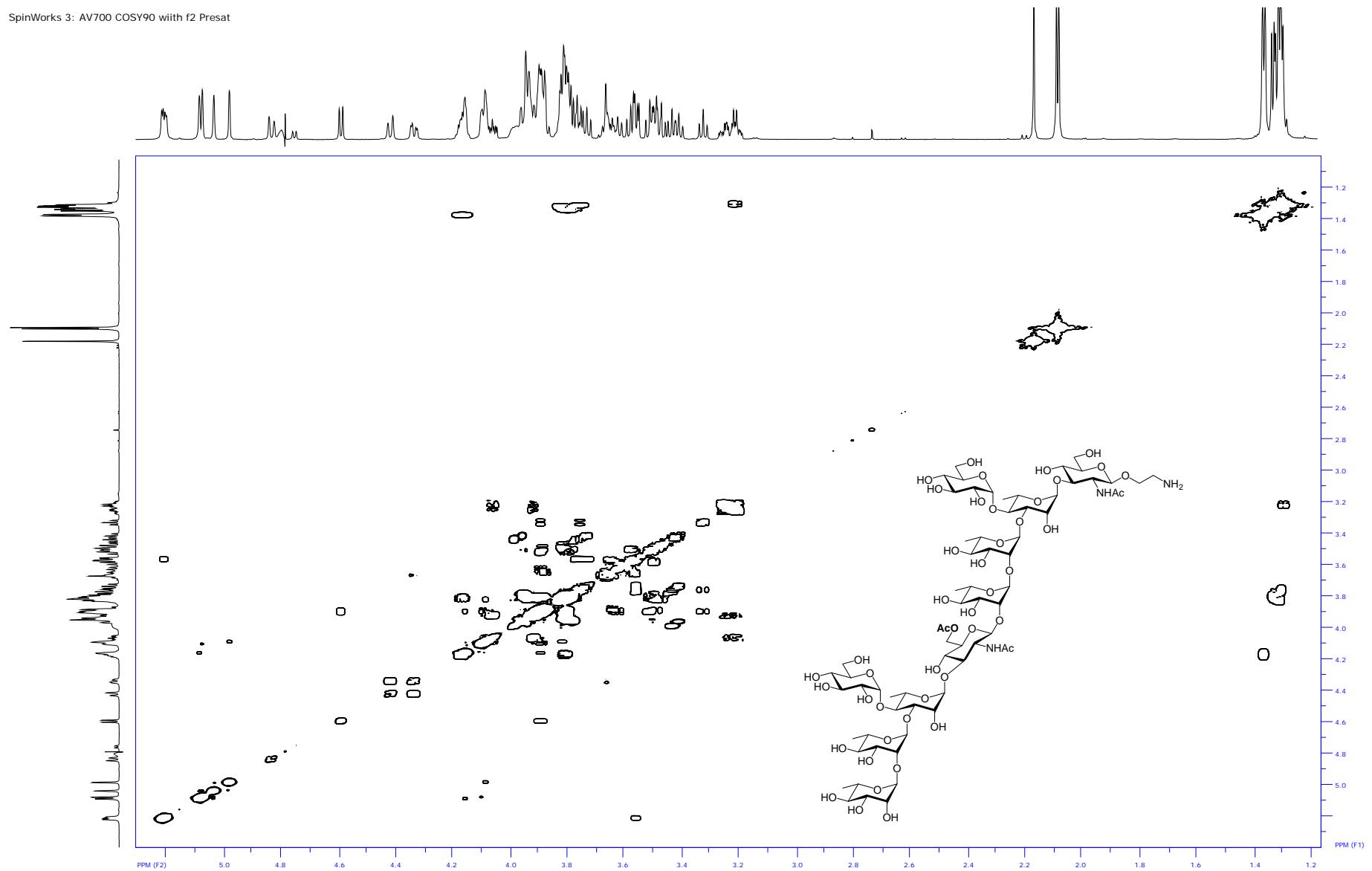
Enlargement of the above spectrum (HSQC for $^1J_{C,H}$) showing the anomeric region



Compound 3



SpinWorks 3: AV700 COSY90 with f2 Presat



SpinWorks 3: AV700 HSQC multiplicity edited with adiabatic ^{13}C for inversion and with matched sweep

