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## **Electronic Supplementary Information**

## Design, Synthesis and Biological evaluation of Bicyclic Iminosugar Hybrids: Conformational constraint as an effective tool for tailoring the selectivity of $\alpha$ -glucosidase inhibitors

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## **Abbreviations**

Bn	Benzyl
DCM	Dichloromethane
DIAD	Diisopropylazodicarboxylate
DPPA	Diphenylphosphoryl azide
DMF	Dimethylformamide
EtOAc	Ethyl acetate
EtOH	Ethanol
Eq	Molar equivalent(s)
NMR	Nuclear magnetic resonance
TBAF	Tetrabutylammonium fluoride
t-BuOOH	tert-Butyl hydroperoxide
TBSOTf	tert-Butyldimethylsilyl trifluoromethanesulfonate
THF	Tetrahydrofuran
TLC	Thin Layer chromatogram
TMS	Tetra methyl silane
<i>n</i> -BuLi	<i>n</i> -butyl lithium



<sup>1</sup>H NMR Spectrum of 3a



<sup>13</sup>C NMR Spectrum of 3a

VIP-360



HSQC Spectra of 3a



<sup>1</sup>H NMR Spectrum of 4a



<sup>13</sup>C NMR Spectrum of 4a



HSQC Spectra of 4a



<sup>1</sup>H NMR Spectrum of 5a



<sup>13</sup>C NMR Spectrum of 5a

VIP-361



HSQC Spectra of 5a



<sup>1</sup>H NMR Spectrum of 6a



<sup>13</sup>C NMR Spectrum of 6a



<sup>1</sup>H NMR Spectrum of 7a



<sup>13</sup>C NMR Spectrum of 7a







<sup>1</sup>H NMR Spectrum of 8a



<sup>13</sup>C NMR Spectrum of 8a



HSQC Spectra of 8a



<sup>1</sup>H NMR Spectrum of 9a



<sup>13</sup>C NMR Spectrum of 9a

VIP-384-A



HSQC Spectra of 9a



<sup>1</sup>H NMR Spectrum of 9c



<sup>13</sup>C NMR Spectrum of 9c



HSQC Spectra of 9c



<sup>1</sup>H NMR Spectrum of 3b



<sup>13</sup>C NMR Spectrum of 3b



HSQC Spectra of 3b



<sup>1</sup>H NMR Spectrum of 4b



<sup>13</sup>C NMR Spectrum of 4b



HSQC Spectra of 4b



<sup>1</sup>H NMR Spectrum of 5b



<sup>13</sup>C NMR Spectrum of 5b



HSQC Spectra of 5b

**VIP-367** 



<sup>1</sup>H NMR Spectrum of 6b



<sup>13</sup>C NMR Spectrum of 6b


<sup>1</sup>H NMR Spectrum of 7b



<sup>13</sup>C NMR Spectrum of 7b

VIP-394C



HSQC Spectra of 7b



<sup>1</sup>H NMR Spectrum of 8b



<sup>13</sup>C NMR Spectrum of 8b



HSQC Spectra of 8b



<sup>1</sup>H NMR Spectrum of 9b



<sup>13</sup>C NMR Spectrum of 9b



HSQC Spectra of 9b



<sup>1</sup>H NMR Spectrum of 10a



<sup>13</sup>C NMR Spectrum of 10a



HSQC Spectra of 10a



<sup>1</sup>H NMR Spectrum of 11a



<sup>13</sup>C NMR Spectrum of 11a



HSQC Spectra of 11a







<sup>13</sup>C NMR Spectrum of 12a



HSQC Spectra of 12a



<sup>1</sup>H NMR Spectrum of 13a



<sup>13</sup>C NMR Spectrum of 13a



HSQC Spectra of 13a



<sup>1</sup>H NMR Spectrum of 14a



<sup>13</sup>C NMR Spectrum of 14a

VIP-406LS



HSQC Spectra of 14a



<sup>1</sup>H NMR Spectrum of 10b



<sup>13</sup>C NMR Spectrum of 10b



HSQC Spectra of 10b



<sup>1</sup>H NMR Spectrum of 11b



<sup>13</sup>C NMR Spectrum of 11b



HSQC Spectra of 11b



<sup>1</sup>H NMR Spectrum of 12b



<sup>13</sup>C NMR Spectrum of 12b



HSQC Spectra of 12b



<sup>1</sup>H NMR Spectrum of 13b



<sup>13</sup>C NMR Spectrum of 13b

VIP-409-A



HSQC Spectra of 13b
## 5,1233 5,1205 5,1205 5,1205 5,1205 5,1205 5,1205 6,

VIP-411

Data Parameters VIP 411 01.06.12 100 1 Curre NAME EXPNO NS DS SWH FID AQ RG DW DE TE D1 TD0 NUC1 P1 PL1 SF01 F2 -SI SF WDW SSB LB GB PC 1H 11.60 use -1.00 dB 300.1318534 MHz ang parameters 32768 300.1300139 MHz 6 0.30 Hz 0 1.00 4.0 ppm 3.9 3.8 3.7 3.6 3.5 4 2.02 4.15 VIP-411 - 0.0000 Vata Parameters VIP 411 01.06.12 100 1 CICH<sub>2</sub>SO<sub>2</sub>O QBn BnO SO NS DS SWE FII AQ RG DW DE TE D1 TD0 Θ̈́Bn NUC1 P1 SF01 F2 -SI SF WDW SSB LB GB PC 1H 11.60 -1.00 300.1318534 dB MHz ing paramet 32768 EM 0.30 0.1.00 Hz 0 1 ppm 1.00 14.89 1.02 4.15 1.09 2.02 2.02

<sup>1</sup>H NMR Spectrum of 14b



<sup>13</sup>C NMR Spectrum of 14b



HSQC Spectra of 14b







<sup>13</sup>C NMR Spectrum of 15a



HSQC Spectra of 15a



Proton coupled <sup>13</sup>C Spectra of 15a











<sup>1</sup>H NMR Spectrum of 15b



<sup>13</sup>C NMR Spectrum of 15b

VIP-400



HSQC Spectra of 15b



<sup>1</sup>H NMR Spectrum of 16a



<sup>13</sup>C NMR Spectrum of 16 a



HSQC Spectra of 16a



<sup>1</sup>H NMR Spectrum of 16b



<sup>13</sup>C NMR Spectrum of 16b



HSQC Spectra of 16b



<sup>1</sup>H NMR Spectrum of 17a



<sup>13</sup> C NMR Spectrum of 17a



<sup>1</sup>H NMR Spectrum of 17b



<sup>13</sup>C NMR Spectrum of 17b



<sup>1</sup>H NMR Spectrum of 18a



<sup>13</sup> C NMR Spectrum of 18a





## **Glycosidase Inhibition**

General Methods are mentioned in the manuscript text. Inhibition potencies of compounds were determined according to Gunter and Stefan (1986), Li *et al* (2011) by minute modifications, measuring the residual hydrolytic activities of glycosidase of the corresponding p-nitrophenyl glycosides in presences of compounds specterophometrically. Michaelis-Menten plot of Activity versus Substrate concentration for inhibition and  $K_i$  were determined by nonlinear regression using data to a competitive inhibition model using Graph Pad Prism (version 6.01 for Windows, Graph Pad Software, San Diego California (USA).<sup>1-3</sup>

S.No	Enzyme	Substrate	Buffer	Incubation
				temperature.
1	$\alpha$ -galactosidase (green	<i>p</i> -nitrophenyl $\alpha$ -D-	Citrate	25°C
	coffee bean)	galactopyranoside	phosphate buffer	
-			(50 mM, pH 6.5	
2	$\beta$ -galactosidase (bovine	<i>p</i> -nitrophenyl $\beta$ -D-	Citrate	30°C
	liver)	galactosidase	phosphate buffer	
-			(50 mM, pH 4.5)	
3	Trehalase (porcine kidney)	D-trehalose	Sodium acetate	25 °C
		dihydrate	buffer (50 mM,	
			pH 6.5)	
4	Amyloglucosidase (A.	Starch wheat	Citrate	45 °C
	niger)		phosphate buffer	
			(50 mM, pH 5.5)	
5	$\alpha$ -mannosidase (jack bean)	<i>p</i> -nitrophenyl α-D-	Acetate buffer	25 °C
		mannpyranoside	(50 mM, pH 4.5)	
6	$\beta$ -glucosidase (almond)	<i>p</i> -nitrophenyl $\beta$ -D-	Citrate	37 °C
		glucopyranoside	phosphate buffer	
			(50 mM, pH 5.5)	
7	$\alpha$ -glucosidase (yeast)	<i>p</i> -nitrophenyl $\alpha$ -D-	Citrate	37 °C
		glucopyranoside	phosphate buffer	
			(50 mM, pH 6.8)	
8	Glucosidase (A. niger)	<i>p</i> -nitrophenyl $\alpha$ -D-	Citrate	37 °C
		glucopyranoside	phosphate buffer	
			(50 mM, pH 6.8)	
9	$\alpha$ -glucosidase (rice)	<i>p</i> -nitrophenyl $\alpha$ -D-	Citrate	37 °C
		glucopyranoside	phosphate buffer	
			(50 mM, pH 6.8)	
10	$\beta$ -N-acetyl	4-nitrophenyl N-	Citrate buffer (50	37°C
	glucosaminidase (jack	Acetyl $\beta$ -D	mM, pH 4)	
	bean)	glucosaminide		
11	$\alpha$ -L-fucosidase (bovine	4-nitrophenyl α-L	Citrate	30 °C
	kidney)	fucopyranoside	phosphate buffer	
			(50 mM, pH 5)	

Table 1: List of Enzyme respective substrates, buffer and optimum incubation temperature

## Table 2: % of inhibition of 11 glycosidase by all 4 compounds (17a, 17b, 18a and 18b)at $400\mu$ M and $1000 \mu$ M

	(% of inhibition at $400\mu$ M)				(% of inhibition at $1000\mu$ M)			
ENZYME	17a	17b	<b>18a</b>	18b	17a	17b	<b>18a</b>	18b
<i>a</i> -galactosidase (green coffee bean)	NI (2.78)	NI (1.7)	NI (0.66)	NI (4.44)	NI (6.55)	NI (6.5)	NI (4.76)	NI (5.88)
$\beta$ -galactosidase (bovine liver)	NI (6.58)	NI (6.16)	NI (6.66)	NI (1.43)	NI (6.75)	NI (8.05)	NI (5.56)	NI (3.49)
α-glucosidase (yeast)	NI (1.31)	NI (1.18)	NI (0.64)	NI (0.8)	NI (3.26)	NI (1.66)	NI (0.98)	NI (0.97)
a-glucosidase (rice)	(81.53)	(84.86)	(81.04)	(87.78)	(86.62)	(86.13)	(86.12)	(93.45)
β-glucosidase (almond)	NI (3.44)	NI (3.18)	NI (3.8)	NI (1.81)	NI (1.48)	NI(13.23)	NI (2.7)	NI (2.2)
a-glucosidase (A. niger)	(81.16)	(81.96)	(80.66)	(81.46)	(84.26)	(84.63)	(84.85)	(85.47)
α-mannosidase (jack bean)	NI (2.07)	NI (2.72)	NI (3.51)	NI (3.01)	NI (1.40)	NI (5.95)	NI (4.38)	NI (1.4)
Trehalase (porcine kidney)	NI (0.58)	NI (0.54)	NI (0.85)	NI (1.23)	NI (5.05)	NI (4)	NI (1.14)	NI (2.39)
Amyloglucosidase (A. niger)	NI (0.82)	NI (1.17)	NI (0.35)	NI (2.7)	NI (3.41)	NI (2.74)	NI (3.53)	NI (3.97)
$\beta$ -N- acetyl glucosaminidase (jack bean)	NI (1.57)	NI (2.53)	NI (1.32)	NI (3.83)	NI (2.09)	NI (6.76)	NI (2.14)	NI((1.04)
α -L-fucosidase (Bovine Kidney)	NI (0.55)	NI (0.05)	NI (0.04)	(86.15)	NI (0.63)	NI (0.04)	NI (0.98)	(91.78)

Figures A, B, C, D, E, F, G, H and I represent Michaelis-Menten plot of Activity versus Substrate concentration for inhibition of various enzymes(see methods) by compounds 17a. 17b, 18a and 18b. The  $K_i$  were determined by nonlinear regression using Graph Pad Prism (version 6.01 for Windows, Graph Pad Software, San Diego California USA) These curves show that the inhibition is competitive. Activity represented absorbance of liberated *p*-nitrophenol measured at 405 nm.



















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