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**Supporting Information** 

# (S)- or (R)- $\alpha$ -(phenyl)ethylamine-derived NH-type ligands: Design, synthesis and applications for chemical resolution of $\alpha$ -amino acids

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## X-Ray data of compound 14a

Physical and c	rystallographic data	Refinement conditions	
Formula	C <sub>34</sub> H <sub>34</sub> Cl N <sub>3</sub> Ni O <sub>4</sub>	Difractometer	Agilent SuperNova Cu
M <sub>r</sub> (g·mol <sup>−1</sup> )	642.8	Detector	CCD (Atlas)
Crystal system	Tetragonal	Temperature (K)	100(1)°
Space group	P4 <sub>1</sub> /n	λ (CuKα) (Å)	1.54184
a (Å)	9.76204(4)	Monochromator	Óptica multicapa
b (Å)	9.76204(4)	Collimator (mm)	0.2
c (Å)	31.9566(2)	Scan mode	Rotation ω
α (°)	90	Scan width (°)	1.0
β (°)	90	Time (s) (Total, h)	2;8 (4)
γ (°)	90	Interval θ (°)	4.53-71.44
V (Å <sup>3</sup> )	3045.357(19)	(hkl) minimum	(–11 –12 –39)
Z	4 (Z'=1)	(hkl) maximun	(12 11 27)
F (000)	1344	Reflections collected	24338
$\mu$ (CuK $\alpha$ ) (mm <sup>-1</sup> )	2.077	Independent reflections (R <sub>int.</sub> )	5213(0.024)
D <sub>x</sub> (g·cm <sup>−3</sup> )	1.402 (1)	Observed reflections $[I>2\sigma(I)]$	5195
Morphology	Cubic prism	Absorption correction	Gaussian
Color	red	Solution	OLEX2
Crystal size (mm)	0.21x0.25x0.30	Refinement	SHELXL97
		Parameters	391
		Restraints	1
		$\Delta/\sigma$ maximum	0.002
		$\Delta/\sigma$ average	0.000
		∆ρ maximum (eÅ <sup>-3</sup> )	0.138
		∆ρ minimum (eÅ <sup>-3</sup> )	-0.147
Friedel coverage	<b>76%</b>	S (GOF)	1.115
Flack <i>x</i>	0.006(12)	Secondary extinction coefficient <sup>b]</sup>	0
Hooft <i>y</i>	0.000(3)	R(F) (I>2σ <sub>ι</sub> , all data)	0.0262, 0.0263
P2(wrong)	<10 <sup>-99</sup>	R <sub>w</sub> (F <sup>2</sup> ) <sup>[a]</sup> (I>2σ₁,all data)	0.0698, 0.0699

[a] Weighting:  $1/[\sigma^2(F_0^2)+(0.0390P)^2]$  where P =  $[Max(F_0^2,0)+2F_c^2]/3$ .

[b] Secondary extinction expression SHELXL:  $F_c^* = kF_c[1+0.001F_c^2\lambda^3/sen(2\theta)]^{-1/4}$ 





Figure S1. ORTEP diagram of 14a. Ellipsoids displayed at 50% probability.









•

ATOMS	LENGHTS (Å)
Ni2A-O1	1 8692(14)
Ni2A-N1	1.8466(16)
Ni2A-N2	1.8404(17)
Ni2A-N3	1.9201(18)
C 1-C 2	1 737(2)
01-012	1 303(2)
02-01	1 213(2)
O3-C16	1 220(3)
N1-C3	1 302(3)
N1-C2	1 475(3)
N2-C16	1.374(3)
N2-C15	1 391(3)
N3-C18	1 510(3)
N3-C17	1 479(3)
C1-C2	1 531(3)
$C_{2}C_{2}C_{2}C_{2}C_{2}C_{2}C_{2}C_{2}$	1 544(3)
$C_{2}$ -C_{10}	1.044(0)
C3-C4	1.501(3)
C4-C9	1 388(3)
C4-C5	1.300(3)
C5-C6	1.387(3)
	1.300(3)
	1.304(3)
CR-C0	1.304(3)
C10-C15	1.000(0)
C10-C11	1.423(3)
C11-C12	1.409(3)
C12 - C12	1 388(3)
C12-C13	1.300(3)
C14 - C15	1.070(0)
C16 C17	1.400(0)
C10-C17	1.510(3)
C18 C10	1.522(3)
C10-C19	1.017(0)
C19-C24 C10 C20	1.395(3)
C19-C20 C20 C21	1.390(3)
C20-C21	1.400(3)
C21-C22	1.300(4)
022-023	1.307(3)
023-024	1.303(3)
020-021	1.317(3)
027-032	1.304(4)
	1.397(3)
020-029	1.391(3)
	1.370(4)
	1.377(4)
031-032	1.395(4)
solvent	
	4 440/9\

001 002	1.000(4)	
solvent		
O1D-C1D C1D-C2D	1.418(3) 1.541(6)	
-		

#### Table S2. Bond angles (°) of 14a

ATOMS	ANGLES (°)
Ni2A-N1-C2	110.80(12)
Ni2A-N1-C.3	127 44(14)
Ni2A-N2-C15	123 89(14)
Ni2A-N2-C16	113 53(14)
Ni2A-N2-C17	103 32(12)
	103.32(12)
NIZA-N3-C10	113.49(13)
NIZA-01-01	114.30(12)
01-01-02	114.88(16)
01-01-02	124.76(18)
01-Ni2A-N1	86.89(7)
01-Ni2A-N2	177.54(8)
01-Ni2A-N3	94.05(7)
O2-C1-C2	120.31(18)
O3-C16-C17	121.24(19)
O3-C16-N2	127.86(19)
N1-C2-C1	107.29(15)
N1-C2-C26	111.51(16)
N1-C3-C10	121.58(17)
N1-C3-C4	120.77(17)
N1-Ni2A-N2	94.53(7)
N1-Ni2A-N3	178.61(8)
N2-C15-C10	120.39(17)
N2-C15-C14	120.86(18)
N2-C16-C17	110,89(17)
N2-Ni2A-N3	84.50(7)
N3-C17-C16	108 50(17)
N3-C18-C19	110 82(17)
N3-C18-C25	109 38(16)
	110 44(16)
	119.44(10)
	119.40(15)
$C_1 - C_2 - C_2 C_2 C_2 C_2 C_2 C_2 C_2 C_2 C_2 C_2$	109.31(17)
02-020-027	114.10(16)
C2-N1-C3	121.72(16)
C3-C10-C11	117.45(17)
C3-C10-C15	123.53(17)
C3-C4-C5	119.68(18)
C3-C4-C9	120.29(19)
C4-C3-C10	117.65(17)
C4-C5-C6	119.74(19)
C4-C9-C8	119.9(2)
C5-C4-C9	119.97(18)
C5-C6-C7	120.1(2)
C6-C7-C8	120.56(19)
C7-C8-C9	119.65(19)
C10-C11-C12	120.25(19)
C10-C15-C14	118,63(19)
C11-C10-C15	119 02(18)
C11-C12-C13	121 15(19)
C12-C13-C14	119 52(18)
C13-C14-C15	121 42(19)
C15-N2-C16	122 58(17)
C17 N2 C19	110 42(16)
	120 00(17)
C10-C19-C2U	120.30(17)
010-019-024	119.92(10)
019-018-025	113.19(10)
019-020-021	120.1(2)
019-024-023	120.58(19)
020-019-024	119.18(19)
C20-C21-C22	120.2(2)
C21-C22-C23	119.7(2)
C22-C23-C24	120.3(2)
C26-C27-C28	120.63(18)
C26-C27-C32	121.3(2)
C27-C28-C29	120.6(2)
C27-C32-C31	120.8(3)
C28-C27-C32	118.1(2)
C28-C29-C30	120.6(2)
C29-C30-C31	119.5(3)́
C30-C31-C32	120.3(3)
solvent	\ - /
	106 2/2)
01D-C1D-C2D	100.3(3)

# Table S3. Torsion angles (°) of 14a

ATOMS	ANGLES (°)		
N1-Ni2A-O1-C1	-11.42(15)	C10-C3-C4-C9	-79.7(2)
N3-Ni2A-O1-C1	167.56(15)	N1-C3-C10-C11	163.56(19)
01-Ni2A-N1-C2	20.73(13)	N1-C3-C10-C15	-16.7(3)
N2-Ni2A-N1-C2	-157.26(14)	C9-C4-C5-C6	1.2(3)
O1-Ni2A-N1-C3	-161.51(17)	C3-C4-C9-C8	176.94(18)
N2-Ni2A-N1-C3	20.50(18)	C3-C4-C5-C6	-176.20(18)
N1-Ni2A-N2-C15	-31.42(17)	C5-C4-C9-C8	-0.4(3)
N3-Ni2A-N2-C15	149.57(17)	C4-C5-C6-C7	-0.6(3)
N1-Ni2A-N2-C16	149.34(15)	C5-C6-C7-C8	-0.8(3)
N3-Ni2A-N2-C16	-29.67(15)	C6-C7-C8-C9	1.5(3)
O1-Ni2A-N3-C17	-138.91(13)	C7-C8-C9-C4	-0.9(3)
N2-Ni2A-N3-C17	39.10(13)	C11-C10-C15-C14	0.5(3)
O1-Ni2A-N3-C18	101.48(13)	C3-C10-C11-C12	-179.81(19)
N2-Ni2A-N3-C18	-80.51(14)	C15-C10-C11-C12	0.5(3)
Ni2A-01-C1-02	-178.42(18)	C3-C10-C15-N2	4.7(3)
Ni2A-01-C1-C2	-1	C3-C10-C15-C14	-179.26(19)
C3-N1-C2-C26	-82.9(2)	C11-C10-C15-N2	-175.55(19)
Ni2A-N1-C2-C1	-24.69(19)	C10-C11-C12-Cl1	177.33(16)
C3-N1-C2-C1	157.41(18)	C10-C11-C12-C13	-1.3(3)
Ni2A-N1-C3-C4	178.55(14)	CI1-C12-C13-C14	-177.50(16)
C2-N1-C3-C10	176.72(18)	C11-C12-C13-C14	1.1(3)
C2-N1-C3-C4	-3	C12-C13-C14-C15	-0.1(3)
Ni2A-N1-C2-C26	94 98(16)	C13-C14-C15-N2	175 4(2)
Ni2A-N1-C3-C10	-0	C13-C14-C15-C10	-0.6(3)
Ni2A-N2-C16-O3	-167 53(18)	N2-C16-C17-N3	21.3(2)
C15-N2-C16-O3	13 2(3)	03-C16-C17-N3	-159 98(19)
C16-N2-C15-C14	26 7(3)	N3-C18-C19-C20	60 1(3)
C16-N2-C15-C10	-157 43(19)	N3-C18-C19-C24	-119 1(2)
Ni2A-N2-C15-C10	23 4(3)	C25-C18-C19-C20	-63 2(3)
Ni2A-N2-C16-C17	11 1(2)	C25-C18-C19-C24	1177(2)
Ni2A-N2-C15-C14	-15252(17)	C18-C19-C20-C21	-1785(2)
C15_NI2_C16_C17	-168 18(18)	$C_{24}$ $C_{19}$ $C_{20}$ $C_{21}$	0.6(3)
Ni2A_N3_C18_C10	-100.10(10) -170.25(12)	C18 - C19 - C20 - C21	178 5(2)
C17_N3_C18_C25	-160.24(16)	$C_{20}$ $C_{10}$ $C_{24}$ $C_{23}$	-0.7(4)
C18-N3-C17-C16	2103.24(10) 80 //3(10)	C10-C20-C21-C23	-0.7(+)
NI2A N3 C17 C16	<i>1</i> 1 26(17)	$C_{20} C_{21} C_{22} C_{23}$	0.2(7)
NI2A-N3-C17-C10	-41.20(17) 53 77(19)	$C_{20}$	-1.0(4)
017 N2 C19 C10	-00.77(10)	$C_{21}$ $C_{22}$ $C_{23}$ $C_{24}$ $C_{10}$	1.0(4)
	104.2(2)	$C_2 C_2 C_2 C_2 C_2 C_2 C_2 C_2 C_2 C_2 $	-0.2(4)
	-104.3(2)		-82.1(2)
02-01-02-N1	-105.71(19)	$C_2 - C_2 - C_2 - C_2 - C_3 $	97.3(Z)
	13.2(3)	020-027-028-029	-1/9.8(2)
01-01-02-N1	16.7(3)	032-027-028-029	0.7(3)
U1-U2-U26-U27	69.8(2)	026-027-032-031	-1/9.6(2)
NT-U2-U26-U27	-48.7(2)	028-027-032-031	-0.1(4)
C4-C3-C10-C11	-15.8(3)	C27-C28-C29-C30	-1.1(4)
C4-C3-C10-C15	163.90(19)	C28-C29-C30-C31	0.8(4)
N1-C3-C4-C5	-81.7(3)	C29-C30-C31-C32	-0.2(4)
N1-C3-C4-C9	100.9(2)	C30-C31-C32-C27	-0.2(4)
C10-C3-C4-C5	97 7(2)		

**Table S4**. Possible hydrogen bonds  $(Å, \circ)$ 

D—H…A	D—H	HA	DA	D—H…A
O1D-H1D···O1 <sup>i</sup>	0.8400	2.0600	2.790(2)	144.00
N3-H3…O1D <sup>ii</sup>	0.9300	2.1100	3.000(3)	161.00
C6-H6…O3 <sup>™</sup>	0.9500	2.4800	3.196(3)	132.00
C14-H14…O3	0.9500	2.3100	2.851(3)	116.00
C17-H17A…CI1 <sup>iv</sup>	0.9900	2.6300	3.432(2)	138.00
C24-H24O2 <sup>v</sup>	0.9500	2.3800	3.283(3)	159.00

Symmetry options: (i)= 2-y,x,1/4+z z; (ii) =- y,2-x,-1/4+z; (iii)= -1-y,-1+x,1/4+z.. (iv)=1+y,2-x,-1/4+z z; (v) =-1+x,y,z

## HPLC analysis data

# General conditions for HPLC analysis of Ni(II) complexes 12-15:

Insturument:	SHIMADZU LC-2010CHT chromatography system and a CLASS-VP <sup>TM</sup> analysis data system (SHIMADZU CORPORATION, Kyoto, Japan).
Column:	Inertsil <sup>TM</sup> ODS-3 (3µm, 150*4.6 mm i.d.)
Eluent:	A = 10mM ammonium formate 0.1% formic acid buffer
	B = acetonitrile
Flow rate:	1.0 mL/min
Temp:	30 °C
Detector:	UV 254 nm

## Reaction of (*R*)-9 with glycine



Peak Number	Retention Time	Area	Area%
1	19.402	3456701	46.22
2	20.330	4022287	53.78
Total			
		7478988	100.00

Gradient A: B = 80:20 to 20:80 (0 to 25 min)

#### Reaction of (R)-9 with phenylalanine



Gradient A: B = 80:20 to 20:80 (0 to 25 min)

## Reaction of (*R*)-9 with phenylglycine



Area%
38.22
7.54
48.44
5.79

Total		
	7748724	100.00

Gradient A: B = 80:20 to 20:80 (0 to 25 min)

#### Reaction of (R)-9 with alanine



Total		
	17941678	100.00

Gradient A: B = 55:45 (0 to 30 min) 55:45 to 10:90 (30 min to 50 min)

#### Reaction of (*R*)-9 with valine



Gradient A: B =	50:50 to 40:60 (0 min to 30 min)
	40:60 to 10:90 (30 min to 50 min)

#### Reaction of (*R*)-9 with leucine



Total		
	18144009	100.00

Gradient A: B = 50:50 to 40:60 (0 min to 30 min) 40:60 to 10:90 (30 min to 50 min)

## Reaction of (*R*)-9 with tyrosine



Detector A -	1	(254nm)
--------------	---	---------

Peak Number	Retention Time	Area	Area%
1	8.286	766992	4.68
2	9.111	7519581	45.91
3	11.272	8091702	49.41

Total		
	16378275	100.00

Gradient A: B = 50:50 to 40:60 (0 min to 30 min) 40:60 to 10:90 (30 min to 50 min)







Peak Number	Retention Time	Area	Area%
1	19.626	6977892	45.65
2	21.664	461378	3.02
3	22.437	570492	3.73
4	25.963	7277190	47.60

Total		
	15286952	100.00

#### Gradient A: B = 55:45 (0 to 30 min) 55:45 to 10:90 (30 min to 50 min)

# HPLC conditions for DMT complex samples

Insturument:	SHIMADZU LC-2010CHT chromatography system and a CLASS-VP <sup>TM</sup> analysis data system (SHIMADZU CORPORATION, Kyoto, Japan).
Column:	Inertsil <sup>TM</sup> ODS-3 ( $3\mu m$ , 150*4.6 mm i.d.)
Eluent:	A = 10mM ammonium formate 0.1% formic acid buffer
	B = acetonitrile
Gradient:	A: $B = 55:45 (0 \text{ to } 30 \text{ min})$
	55:45 to 10:90 (30 min to 50 min)
Flow rate:	1.0 mL/min
Temp:	30 °C
Detector:	UV 254 nm

# Epimerization of 14g



Detector A - 1 (25	4nm)
--------------------	------

Peak Number	Retention Time	Area	Area%
1	22.423	111960	7.60
2	25.896	1360937	92.40
Total			
		1472897	100.00

HPLC conditions for chiral analysis of free DMT

Insturument:SHIMADZU chromatography system, consisting of a LC-10ADvp<br/>high-pressure gradient pump, a SPD-10Avp UV-VIS detector, and a<br/>CLASS-VP<sup>TM</sup> analysis data system (SHIMADZU CORPORATION, Kyoto,<br/>Japan)Column:Astec CHIROBIOTIC<sup>TM</sup> REluent: $H_2O/MeOH = 8: 2$ Flow rate:0.5 mL/minTemp: $25 \,^{\circ}C$ Detector:UV 205 nm $t_R:$  $9.2 \min(R)$ -(-)-DMT<br/> $11.4 \min(S)$ -(+)-DMT

## HPLC of racemic DMT, (±)-18



Detector A - 1	(205nm
----------------	--------

Peak Number	Retention Time	Area	Area%
1	9.243	14451749	49.72
2	11.465	14615997	50.28
Total			
		29067746	100.00

## Disassembly of 14 and 17



Area%	Area	Retention Time	Peak Number
99.24	34001570	9.173	1
0.76	34263359	11.631	2
			Total
100.00	34263359		

#### Disassembly of 15 and 16



## <sup>1</sup>H-NMR of 9 (300 MHz, CDCl<sub>3</sub>)





## <sup>1</sup>H-NMR of 12 (300 MHz, CDCl<sub>3</sub>)





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## <sup>1</sup>H-NMR of 13 (300 MHz, CDCl<sub>3</sub>)





## <sup>1</sup>H-NMR of 14a (300 MHz, CDCl<sub>3</sub>)





# <sup>1</sup>H-NMR of 15a (300 MHz, CDCl<sub>3</sub>)





# <sup>1</sup>H-NMR of 14b (300 MHz, CDCl<sub>3</sub>)





# <sup>1</sup>H-NMR of 15b (300 MHz, CDCl<sub>3</sub>)





## <sup>1</sup>H-NMR of 14c (300 MHz, CDCl<sub>3</sub>)





# <sup>1</sup>H-NMR of 15c (300 MHz, CDCl<sub>3</sub>)





## <sup>1</sup>H-NMR of 14d (300 MHz, CDCl<sub>3</sub>)





# <sup>1</sup>H-NMR of 15d (300 MHz, CDCl<sub>3</sub>)





## <sup>1</sup>H-NMR of 14e (300 MHz, CDCl<sub>3</sub>)





# <sup>1</sup>H-NMR of 15e (300 MHz, CDCl<sub>3</sub>)





## <sup>1</sup>H-NMR of 14f (300 MHz, CDCl<sub>3</sub>)





## <sup>1</sup>H-NMR of 15f (300 MHz, CDCl<sub>3</sub>)





# <sup>1</sup>H-NMR of 14g (300 MHz, CDCl<sub>3</sub>)





# <sup>1</sup>H-NMR of 15g (300 MHz, CDCl<sub>3</sub>)





<sup>1</sup>H-NMR of (-)-18 (300 MHz, D<sub>2</sub>O)



<sup>1</sup>H-NMR of (+)-18 (300 MHz, D<sub>2</sub>O)





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