

Organocatalytic Synthesis of Optically Active β -Branched α -Amino Esters via Asymmetric Biomimetic Transamination

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Supporting Information

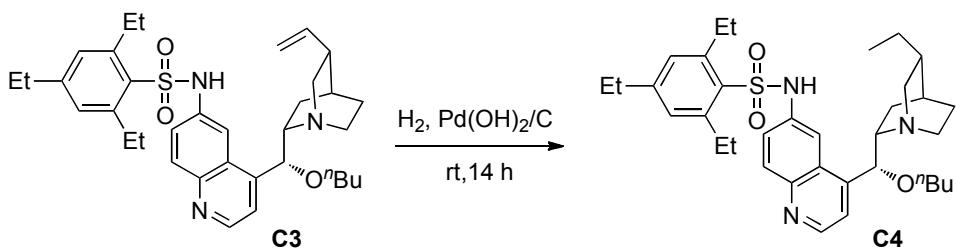
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General Methods. All commercially available reagents were used without further purification. All dry solvents were freshly distilled under nitrogen from appropriate drying agents before use. Toluene, benzene, tetrahydrofuran, and ethyl ether were distilled from sodium-benzophenone. Dichloromethane, 1,2-dichloroethane, and acetonitrile were distilled from CaH₂. N,N-Dimethylformamide was dried over 4 Å molecular sieves (activated at 180 °C under vacuum over 8 h). Column chromatography was performed on silica gel (200-300 mesh). ¹H NMR spectra were recorded on a 400 MHz NMR spectrometer and ¹³C NMR spectra were recorded on a 100 MHz NMR spectrometer. IR spectra were recorded on a FT-IR spectrometer. Melting points were uncorrected. Catalysts **C1-C3** were prepared according to the reported procedures.^{1,2} *t*-Butyl β,β'-disubstituted α-keto esters were generally prepared from the corresponding ketones by olefination with ethyl 2-[(*t*-butyldimethylsilyl)oxy]-2-(diethoxyphosphoryl)acetate,³ hydrolysis of the resulting α-siloxy α,β-unsaturated esters to keto acids,³⁻⁵ and subsequent esterification with isobutene.⁶ α-Keto esters **4i** was prepared from **4h** by ring-closing metathesis (RCM) using Grubbs second-generation ruthenium catalyst.⁷

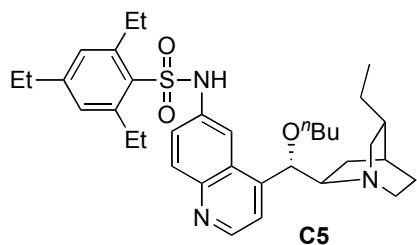
- (1) X. Xiao, Y. Xie, C. Su, M. Liu and Y. Shi, *J. Am. Chem. Soc.*, 2011, **133**, 12914.
- (2) X. Xiao, M. Liu, C. Rong, F. Xue, S. Li, Y. Xie and Y. Shi, *Org. Lett.*, 2012, **14**, 5270.
- (3) S. He, Z. Lai, D. X. Yang, Q. Hong, M. Reibarkh, R. P. Nargund and W. K. Hagmann, *Tetrahedron Lett.*, 2010, **51**, 4361.
- (4) C. H. Senanayake, K. Fang, P. Grover, R. P. Bakale, C. P. Vandenbossche and S. A. Wald, *Tetrahedron Lett.*, 1999, **40**, 819.
- (5) R. M. Archer, S. F. Royer, W. Mahy, C. L. Winn, M. J. Danson and S. D. Bull, *Chem. Eur. J.*, 2013, **19**, 2895.
- (6) H. Molines, M. H. Massoudi, D. Cantacuzene and C. Wakselman, *Synthesis*, 1983, 322.
- (7) H. Hu, J. A. Faraldo and R. M. Coates, *J. Am. Chem. Soc.*, 2009, **131**, 11998.

Preparation of Catalyst C4



To a solution of compound **C3** (1.770 g, 3.0 mmol) in MeOH (30.0 mL) was added Pd(OH)₂/C (20%) (0.300 g).¹ The reaction mixture was stirred at room temperature under hydrogen (1 atm) for 14 h, filtered through Celite, washed with EtOAc, concentrated, and purified by flash column chromatography (silica gel, packed with EtOAc containing 1% Et₃N) (eluent: EtOAc/MeOH = 40/1) to give compound **C4** as a yellow solid (1.699 g, 96% yield). mp. 87-88 °C; $[\alpha]^{20}_{\text{D}} = -83.7$ (*c* 0.73, CHCl₃); IR (film) 3239, 1622, 1457, 1154 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 9.70 (br s, 1H), 8.73 (d, *J* = 4.4 Hz, 1H), 7.93 (d, *J* = 8.8 Hz, 1H), 7.84 (s, 1H), 7.55 (d, *J* = 8.8 Hz, 1H), 7.37 (d, *J* = 4.4 Hz, 1H), 6.95 (s, 2H), 5.20-5.00 (m, 1H), 3.41-2.88 (m, 9H), 2.69-2.52 (m, 3H), 2.44-2.34 (m, 1H), 1.81-1.68 (m, 3H), 1.53-1.40 (m, 3H), 1.40-1.31 (m, 3H), 1.30-1.07 (m, 12H), 0.83-0.72 (m, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 148.9, 148.3, 145.9, 145.8, 145.7, 137.7, 134.2, 131.4, 129.2, 127.4, 124.1, 118.9, 113.2, 79.6, 69.2, 60.3, 58.6, 43.7, 37.3, 32.1, 28.5, 28.4, 28.1, 27.6, 25.4, 22.2, 19.4, 16.7, 14.9, 14.0, 12.1; HRMS Calcd for C₃₅H₅₀N₃O₃S (M+H): 592.3567; Found: 592.3566.

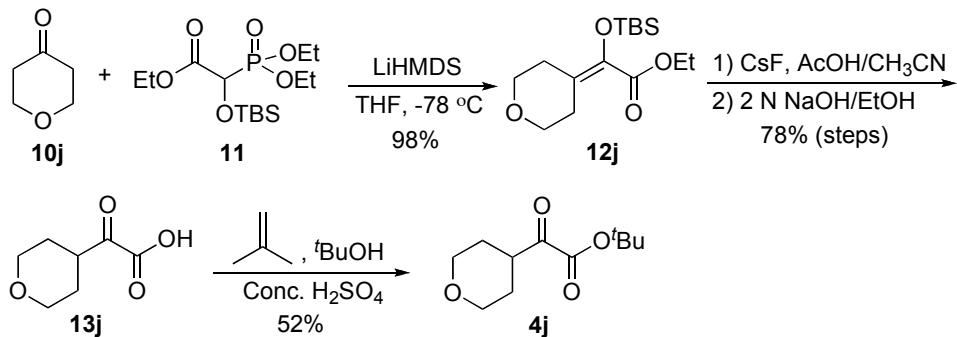
- (1) G. Sabitha, S. Nayak, M. Bhikshapathi, M. Chittapragada and J. S. Yadav, *Synthesis*, 2011, **22**, 3661.



Prepared in a manner similar to **C4**. light yellow solid; mp. 86-87 °C; $[\alpha]^{20}_{\text{D}} =$

+184.2 (*c* 0.84, CHCl₃); IR (film) 3239, 1489, 1457, 1154 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.75 (d, *J* = 4.4 Hz, 1H), 7.94 (d, *J* = 9.2 Hz, 1H), 7.78 (s, 1H), 7.49 (dd, *J* = 9.2, 2.0 Hz, 1H), 7.35 (d, *J* = 4.4 Hz, 1H), 7.05-6.98 (m, 1H), 6.96 (s, 2H), 5.05 (s, 1H), 3.21-3.00 (m, 7H), 2.99-2.79 (m, 3H), 2.79-2.68 (m, 1H), 2.63-2.53 (m, 2H), 2.02-1.91 (m, 1H), 1.72-1.67 (m, 1H), 1.53-1.41 (m, 7H), 1.36-1.17 (m, 12H), 0.94-0.81 (m, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 149.1, 148.4, 146.0, 145.9, 145.7, 137.3, 134.0, 131.4, 129.2, 127.0, 124.9, 119.0, 114.6, 80.4, 69.3, 60.0, 51.0, 50.2, 37.3, 32.2, 28.54, 28.49, 27.0, 26.2, 25.2, 21.5, 19.5, 16.8, 14.9, 14.0, 12.1. HRMS Calcd for C₃₅H₅₀N₃O₃S (M+H): 592.3567; Found: 592.3571.

Representative procedures for the preparation of β,β' -disubstituted α -keto esters



To a solution of phosphonate **11**¹ (38.990 g, 110.0 mmol) in THF (100.0 mL) was added a solution of LiHMDS (1 M solution in THF) (110.0 mL, 110.0 mmol) at -78 °C under N₂.¹ After the reaction mixture was stirred at -78 °C for 30 min, a solution of ketone **10j** (10.011 g, 100.0 mmol) in THF (10.0 mL) was added dropwise over 10 min. The reaction mixture was stirred at -78 °C for 30 min, warmed to room temperature overnight, quenched with saturated aqueous NH₄Cl solution (100.0 mL), extracted with CH₂Cl₂ (3 x 100.0 mL), washed with brine (100.0 mL), dried over MgSO₄, filtered, concentrated, and purified by flash column chromatography (silica gel, eluent: petroleum ether/EtOAc = 10/1) to afford ester **12j** as a light yellow oil (29.446 g, 98%). IR (film) 1712, 1280, 1216, 1097 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 4.21 (q, *J* = 7.2 Hz, 2H), 3.71 (t, , *J* = 5.6 Hz, 2H), 3.67 (t, *J* = 5.6 Hz, 2H), 2.76 (t, *J* = 5.6 Hz, 2H), 2.45 (t, *J* = 5.6 Hz, 2H), 1.33 (t, *J* = 7.2 Hz, 3H), 0.95 (s, 9H), 0.10

(s, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 165.3, 134.9, 129.8, 68.7, 68.2, 60.8, 29.9, 29.4, 25.9, 18.5, 14.4, -4.3; HRMS Calcd for $\text{C}_{15}\text{H}_{28}\text{NaO}_4\text{Si}$ ($\text{M}+\text{Na}$): 323.1649; Found: 323.1654.

(1) S. He, Z. Lai, D. X. Yang, Q. Hong, M. Reibarkh, R. P. Nargund and W. K. Hagmann, *Tetrahedron Lett.*, 2010, **51**, 4361.

To a solution of ester **12j** (29.446 g, 98.0 mmol) and acetic acid (28.0 mL, 490.0 mmol) in CH_3CN (100.0 mL) was added solid cesium fluoride (29.772 g, 196.0 mmol) in one portion at 0 °C.¹ Upon stirring at 0 °C for 30 min and room temperature for 24 h, the reaction mixture was quenched with saturated aqueous NaHCO_3 (100.0 mL), extracted with EtOAc (3 x 100.0 mL), washed with saturated aqueous NaHCO_3 (100.0 mL), and concentrated to give a yellow oil, which was subsequently dissolved in EtOH (200.0 mL), followed by the addition of aqueous 2 N NaOH (100.0 mL).² Upon stirring at room temperature for 17 h, the reaction mixture was diluted with water, washed with n-hexane (3 x 50.0 mL), acidified with concentrated HCl to pH = 2, extracted with CH_2Cl_2 (3 x 50.0 mL), washed with brine (50.0 mL), dried over MgSO_4 , filtered, concentrated, and purified by flash column chromatography (silica gel, eluent: petroleum ether/EtOAc = 10/1 to 0/1) to give α -keto acid **13j** as a white solid (12.010 g, 78% yield). mp. 88-90 °C; IR (film) 3421, 1723, 1273, 1081 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3) δ 4.06 (ddd, J = 11.6, 3.6, 2.4 Hz, 2H), 3.54 (td, J = 11.6, 2.4 Hz, 2H), 3.46 (tt, J = 11.2, 4.0 Hz, 1H), 1.92-1.84 (m, 2H), 1.79-1.67 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 196.4, 161.0, 66.9, 42.5, 27.3; HRMS Calcd for $\text{C}_7\text{H}_9\text{O}_4$ ($\text{M}-\text{H}$): 157.0495; Found: 157.0493.

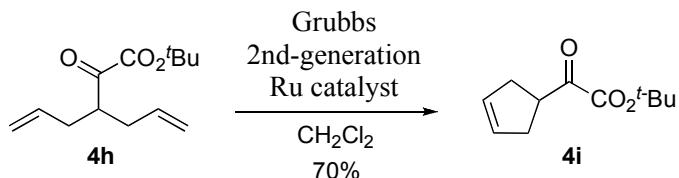
(1) R. M. Archer, S. F. Royer, W. Mahy, C. L. Winn, M. J. Danson and S. D. Bull, *Chem. Eur. J.*, 2013, **19**, 2895.

(2) C. H. Senanayake, K. Fang, P. Grover, R. P. Bakale, C. P. Vandenbossche and S. A. Wald, *Tetrahedron Lett.*, 1999, **40**, 819.

To a screw capped pyrex heavy-walled pressure bottle containing a solution of α -keto acid **13j** (12.012 g, 76.0 mmol), *t*-butanol (12.0 mL), and Et_2O (5.0 mL) at -40 °C, were

added concentrated sulfuric acid (3.0 mL) and liquid isobutene (70.0 mL), respectively.¹ Upon stirring at room temperature for 48 h, the reaction mixture was cooled to -20 °C, quenched with saturated aqueous NaHCO₃ (70.0 mL), extracted with EtOAc (3 x 100.0 mL), washed with brine (100.0 mL), dried over MgSO₄, filtered, concentrated, and purified by flash column chromatography (silica gel, eluent: petroleum ether/EtOAc = 40/1 to 10/1) to give α -keto ester **4j** as a yellow oil (8.424 g, 52% yield). IR (film) 1721, 1371, 1113, 1019 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 4.40-3.95 (m, 2H), 3.48 (td, *J* = 11.2, 1.6 Hz, 2H), 3.24-3.14 (m, 1H), 1.85-1.77 (m, 2H), 1.75-1.62 (m, 2H), 1.54 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 196.6, 161.2, 84.4, 67.0, 43.4, 27.9, 27.3; HRMS Calcd for C₁₁H₁₈NaO₄ (M+Na): 237.1097; Found: 237.1104.

(1) H. Molines, M. H. Massoudi, D. Cantacuzene and C. Wakselman, *Synthesis*, 1983, 322.



A mixture of *t*-butyl 3-allyl-2-oxohex-5-enoate (**4h**) (1.262 g, 5.6 mmol) and Grubbs 2nd-generation Ru catalyst (0.153 g, 0.18 mmol) in dry CH₂Cl₂ (60.0 mL) was stirred at room temperature under N₂ for 18 h,¹ concentrated, and purified by flash column chromatography (silica gel, eluent: petroleum ether/EtOAc = 20/1) to give α -keto ester **4i** as a light yellow oil (0.766 g, 70% yield); IR (film) 1722, 1370, 1161 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 5.62 (br s, 2H), 3.84-3.71 (m, 1H), 2.68-2.62 (m, 4H), 1.54 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 196.3, 161.5, 128.8, 84.1, 45.3, 34.6, 28.0; HRMS Calcd for C₁₁H₁₇O₃ (M+H): 197.1172; Found: 197.1174.

(1) H. Hu, J. A. Faraldo and R. M. Coates, *J. Am. Chem. Soc.*, 2009, **131**, 11998.

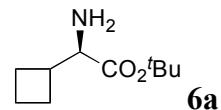
Representative procedure for transamination of β -branched α -keto esters (Table 2, entry 3)

To a well-dried Schlenk tube charged with 4 Å molecular sieves (0.25 g), 4-CNPhCH₂NH₂ (0.198 g, 1.50 mmol), and catalyst **C4** (0.0296 g, 0.050 mmol) under N₂ at room temperature was added a solution of α -keto ester **4c** (0.106 g, 0.50 mmol) in dry toluene (5.0 mL). Upon stirring at 80 °C for 72 h, the reaction mixture was filtered through a short plug of silica gel to remove molecular sieves, washed with EtOAc (containing 1% MeOH), and concentrated. The resulting residue was dissolved in THF (7.5 mL), followed by the addition of aqueous 2 N HCl (7.5 mL). Upon stirring at room temperature for 1 h, the resulting mixture was diluted with water (50 mL) and washed with diethyl ether (3 x 10.0 mL). The organic phase was extracted with 1 N HCl (10.0 mL). The aqueous phases were combined, brought to pH = 8 with solid NaHCO₃, extracted with CH₂Cl₂ (3 x 30.0 mL), dried over MgSO₄, filtered, concentrated, and purified by flash column chromatography (silica gel, eluent: EtOAc/CH₃OH = 40/1) to give α -amino ester **6c** as a yellow oil (0.087 g, 82% yield, 94% ee).

Representative procedure for the preparation of *N*-benzoyl derivative of amino ester for the determination of the enantiomeric excess

To a solution of **6c** (0.021 g, 0.10 mmol) in CH₂Cl₂ (1.0 mL) were added Et₃N (0.018 g, 0.18 mmol) and BzCl (0.021 g, 0.15 mmol). The reaction mixture was stirred at room temperature for 30 min and purified by flash column chromatography (silica gel, eluent: petroleum ether/EtOAc = 10/1) to afford *N*-benzoyl amino ester **14c** as a white solid (0.029 g, 90% yield). The sample was subjected to chiral HPLC (Chiralcel OD-H column) to determine the enantiomeric excess.

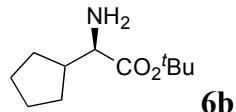
Table 2, entry 1



Yellow oil; [α]²⁰_D = -31.8 (*c* 0.77, CHCl₃) (87% ee); IR (film) 3384, 1727, 1595, 1260, 1154 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 3.20 (d, *J* = 8.4 Hz, 1H), 2.51-2.37 (m, 1H),

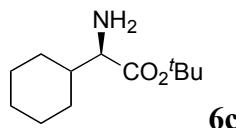
2.10-1.91 (m, 3H), 1.90-1.72 (m, 3H), 1.71-1.55 (m, 2H), 1.43 (s, 9H); ^{13}C NMR (100 MHz, CDCl_3) δ 174.6, 81.0, 59.6, 40.3, 28.3, 25.5, 25.4, 18.1; HRMS Calcd for $\text{C}_{10}\text{H}_{20}\text{NO}_2$ ($\text{M}+\text{H}$): 186.1489; Found: 186.1484.

Table 2, entry 2



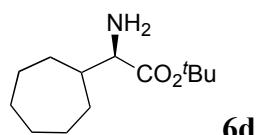
Yellow oil; $[\alpha]^{20}_D = -11.0$ (c 0.84, CHCl_3) (94% ee); IR (film) 3377, 1727, 1367, 1152 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3) δ 3.17 (d, $J = 7.2$ Hz, 1H), 2.12-1.99 (m, 1H), 1.76-1.48 (m, 8H), 1.46 (s, 9H), 1.42-1.28 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 175.4, 80.9, 59.1, 44.6, 29.4, 28.5, 28.3, 25.8, 25.6; HRMS Calcd for $\text{C}_{11}\text{H}_{22}\text{NO}_2$ ($\text{M}+\text{H}$): 200.1645; Found: 200.1642.

Table 2, entry 3



Yellow oil; $[\alpha]^{20}_D = -22.8$ (c 0.87, CHCl_3) (94% ee); IR (film) 3384, 1727, 1367, 1152 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3) δ 3.12 (d, $J = 4.4$ Hz, 1H), 1.79-1.69 (m, 2H), 1.68-1.54 (m, 4H), 1.51-1.35 (m, 2H), 1.45 (s, 9H), 1.31-1.00 (m, 5H); ^{13}C NMR (100 MHz, CDCl_3) δ 175.1, 81.0, 60.3, 42.5, 29.9, 28.3, 28.0, 26.52, 26.51, 26.4; HRMS Calcd for $\text{C}_{12}\text{H}_{24}\text{NO}_2$ ($\text{M}+\text{H}$): 214.1802; Found: 214.1803.

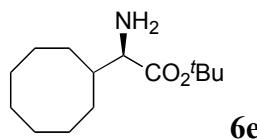
Table 2, entry 4



Yellow oil; $[\alpha]^{20}_D = -23.9$ (c 1.08, CHCl_3) (94% ee); IR (film) 3457, 1738, 1366, 1229 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3) δ 3.22 (d, $J = 4.4$ Hz, 1H), 1.88-1.79 (m, 1H), 1.75-1.63 (m, 3H), 1.63-1.53 (m, 3H), 1.52-1.28 (m, 8H), 1.46 (s, 9H); ^{13}C NMR (100 MHz, CDCl_3)

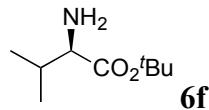
δ 175.2, 81.0, 60.9, 43.7, 31.9, 29.1, 28.5, 28.3, 28.1, 27.4, 27.2; HRMS Calcd for C₁₃H₂₆NO₂ (M+H): 228.1958; Found: 228.1960.

Table 2, entry 5



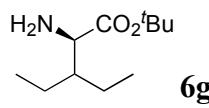
Yellow oil; $[\alpha]^{20}_D = -20.2$ (*c* 1.16, CHCl₃) (95% ee); IR (film) 3381, 1725, 1275, 1154 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 3.18 (d, *J* = 4.8 Hz, 1H), 1.98-1.88 (m, 1H), 1.75-1.64 (m, 2H), 1.63-1.53 (m, 4H), 1.52-1.25 (m, 10H), 1.46 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 175.2, 80.9, 61.4, 41.3, 31.3, 28.3, 27.9, 26.9, 26.8, 26.4, 25.8; HRMS Calcd for C₁₄H₂₈NO₂ (M+H): 242.2115; Found: 242.2112.

Table 2, entry 6

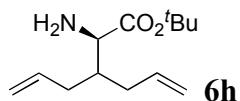


Yellow oil; $[\alpha]^{20}_D = -14.2$ (*c* 0.63, CHCl₃) (92% ee); IR (film) 3403, 1736, 1366, 1218 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 3.16 (d, *J* = 4.4 Hz, 1H), 2.05-1.93 (m, 1H), 1.67 (br s, 2H), 1.45 (s, 9H), 0.96 (d, *J* = 6.8 Hz, 3H), 0.88 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 175.0, 81.0, 60.4, 32.3, 28.3, 19.5, 17.2; HRMS Calcd for C₉H₂₀NO₂ (M+H): 174.1489; Found: 174.1484.

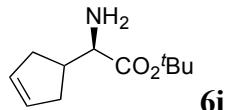
Table 2, entry 7



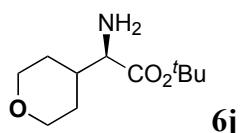
Yellow oil; $[\alpha]^{20}_D = -23.6$ (*c* 0.84, CHCl₃) (91% ee); IR (film) 3386, 1727, 1368, 1154 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 3.40 (d, *J* = 4.0 Hz, 1H), 1.63-1.20 (m, 7H), 1.46 (s, 9H), 0.94 (t, *J* = 7.2 Hz, 3H), 0.90 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 175.7, 80.9, 56.4, 45.5, 28.2, 23.1, 22.1, 12.1, 12.0; HRMS Calcd for C₁₁H₂₄NO₂ (M+H): 202.1802; Found: 202.1799.

Table 2, entry 8

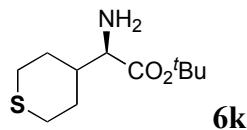
Yellow oil; $[\alpha]^{20}_D = -33.8$ (*c* 1.00, CHCl₃) (92% ee); IR (film) 3384, 1727, 1367, 1217 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 5.85-5.70 (m, 2H), 5.12-4.98 (m, 4H), 3.41 (d, *J* = 3.6 Hz, 1H), 2.18-1.98 (m, 4H), 1.97-1.89 (m, 1H), 1.46 (s, 9H), 1.40 (br s, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 175.2, 137.1, 137.0, 116.9, 116.8, 81.2, 56.3, 41.9, 35.1, 33.9, 28.3; HRMS Calcd for C₁₃H₂₄NO₂ (M+H): 226.1802; Found: 226.1803.

Table 2, entry 9

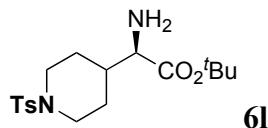
Yellow oil; $[\alpha]^{20}_D = -4.9$ (*c* 1.24, CHCl₃) (92% ee); IR (film) 3462, 1736, 1366, 1218 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 5.65 (br s, 2H), 3.28 (d, *J* = 6.8 Hz, 1H), 2.63-2.52 (m, 1H), 2.48-2.35 (m, 2H), 2.31-2.18 (m, 2H), 1.52 (br s, 2H), 1.45 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 174.8, 129.9, 129.7, 80.9, 58.7, 41.4, 35.7, 34.6, 28.1; HRMS Calcd for C₁₁H₂₀NO₂ (M+H): 198.1489; Found: 198.1485.

Table 2, entry 10

Yellow oil; $[\alpha]^{20}_D = -23.5$ (*c* 0.77, CHCl₃) (91% ee); IR (film) 3383, 1726, 1367, 1152 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 4.03-3.95 (m, 2H), 3.41-3.31 (m, 2H), 3.14 (d, *J* = 6.0 Hz, 1H), 1.87-1.75 (m, 1H), 1.63-1.38 (m, 6H), 1.46 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 174.6, 81.4, 68.2, 68.0, 59.7, 39.9, 29.6, 28.4, 28.3; HRMS Calcd for C₁₁H₂₂NO₃ (M+H): 216.1594; Found: 216.1595.

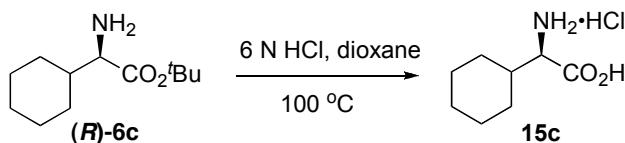
Table 2, entry 11

Yellow oil; $[\alpha]^{20}_D = -21.2$ (*c* 0.91, CHCl₃) (93% ee); IR (film) 3464, 1735, 1366, 1218 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 3.14 (d, *J* = 4.4 Hz, 1H), 2.73-2.57 (m, 4H), 1.96-1.87 (m, 2H), 1.70-1.47 (m, 5H), 1.45 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 174.4, 81.4, 60.2, 42.1, 31.0, 29.4, 28.98, 28.97, 28.3; HRMS Calcd for C₁₁H₂₂NO₂S (M+H): 232.1366; Found: 232.1366.

Table 2, entry 12

Yellow oil; $[\alpha]^{20}_D = -10.4$ (*c* 1.05, CHCl₃) (90% ee); IR (film) 3453, 1737, 1366, 1217 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.63 (d, *J* = 8.0 Hz, 2H), 7.31 (d, *J* = 8.0 Hz, 2H), 3.87-3.77 (m, 2H), 3.14-2.99 (m, 1H), 2.42 (s, 3H), 2.27-2.15 (m, 2H), 1.76-1.68 (m, 1H), 1.66-1.47 (m, 6H), 1.43 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 174.3, 143.6, 133.3, 129.8, 127.9, 81.6, 59.1, 46.5, 46.4, 40.0, 28.3, 27.0, 21.7; HRMS Calcd for C₁₈H₂₈N₂NaO₄S (M+Na): 391.1662; Found: 391.1671.

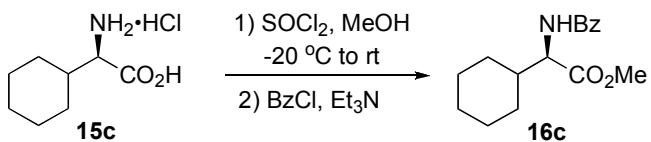
The determination of the absolute configuration of α -amino ester 6c



A solution of α -amino ester (*R*)-6c (0.107 g, 0.50 mmol) (94% ee) in 6 N HCl (7.5 mL) and dioxane (2.5 mL) was stirred at 100 °C for 12 h.¹ The reaction mixture was cooled to room temperature, washed with Et₂O (3 x 5.0 mL), and concentrated to give amino acid hydrochloride 15c as a white solid (0.096 g, 99% yield). decomp. 257 °C; $[\alpha]^{20}_D = -24.9$ (*c* 0.88, 1.5 N HCl) [lit.² $[\alpha]^{20}_D = -33.5$ (1.5 N HCl)]; IR (film) 3331, 1736, 1366, 1217,

1153 cm⁻¹; ¹H NMR (400 MHz, MeOD) δ 3.81 (d, *J* = 4.0 Hz, 1H), 2.00-1.90 (m, 1H), 1.89-1.78 (m, 3H), 1.77-1.65 (m, 2H), 1.41-1.10 (m, 5H); ¹³C NMR (100 MHz, MeOD) δ 171.2, 59.0, 40.2, 29.5, 29.3, 27.0, 26.9, 26.7; HRMS Calcd for C₈H₁₆NO₂ (M-Cl): 158.1176; Found: 158.1172.

- (1) T. Ooi, D. Kato, K. Inamura, K. Ohmatsu and K. Maruoka, *Org. Lett.*, 2007, **9**, 3945.
- (2) C. Toniolo, G. M. Bonora and S. Salardi, *Int. J. Biol. Macromol.*, 1981, **3**, 377.

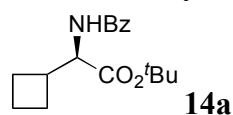


To a solution of amino acid hydrochloride **15c** (0.058 g, 0.30 mmol) in MeOH (2.0 mL) was added SOCl₂ (0.642 g, 5.4 mmol) dropwise at -20 °C. The reaction mixture was stirred at room temperature overnight, concentrated, and dissolved in CH₂Cl₂ (1.0 mL), followed by the addition of Et₃N (0.091 g, 0.90 mmol) and BzCl (0.063 g, 0.45 mmol) successively. Upon stirring at room temperature for 30 min, the reaction mixture was purified by flash column chromatography (silica gel, eluent: petroleum ether/EtOAc = 10/1 to 5/1) to afford *N*-benzoyl amino ester **16c** as a white solid (0.064 g, 78% yield). The sample was subjected to chiral HPLC (Chiralcel OD-H column) to determine the enantiomeric excess. mp. 123-124 °C; [α]²⁰_D = -40.2 (*c* 0.99, CHCl₃) (94% ee); IR (film) 3330, 1740, 1365, 1217 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.81 (d, *J* = 7.6 Hz, 2H), 7.55-7.49 (m, 1H), 7.48-7.42 (m, 2H), 6.62 (d, *J* = 8.0 Hz, 1H), 4.81-4.75 (m, 1H), 3.78 (s, 3H), 1.95-1.86 (m, 1H), 1.81-1.62 (m, 5H), 1.32-1.06 (m, 5H); ¹³C NMR (100 MHz, CDCl₃) δ 172.8, 167.3, 134.3, 131.9, 128.8, 127.2, 57.3, 52.4, 41.5, 29.7, 28.6, 26.1; HRMS Calcd for C₁₆H₂₂NO₃ (M+H): 276.1594; Found: 276.1597.

T. Ooi, D. Kato, K. Inamura, K. Ohmatsu and K. Maruoka, *Org. Lett.*, 2007, **9**, 3945.

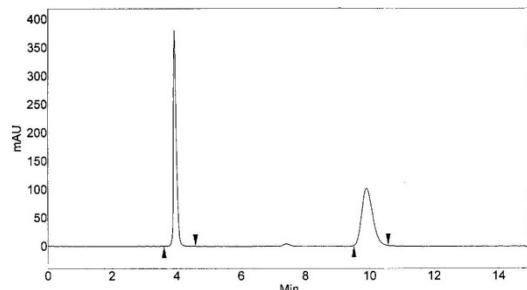
The chromatograms for determination of enantioselectivity

Table 2, entry 1



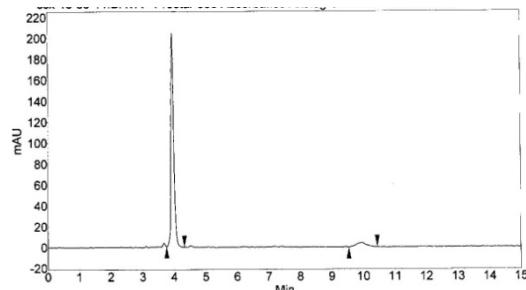
HPLC Condition: Column: Chiralcel OD-H, Daicel Chemical Industries, Ltd.; **Eluent:** Hexanes/IPA (85/15); **Flow rate:** 1.0 mL/min; **Detection:** UV226 nm.

Racemic



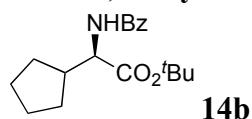
Index	Time [Min]	Area % [%]	Start [Min]	End [Min]
1	3.95	50.043	3.60	4.58
2	9.91	49.957	9.51	10.58
Total		100.000		

Chiral



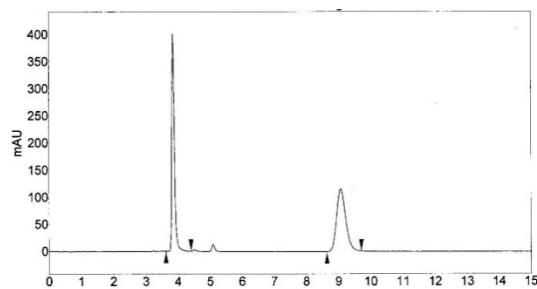
Index	Time [Min]	Area % [%]	Start [Min]	End [Min]
2	3.96	93.589	3.76	4.32
1	9.93	6.411	9.55	10.44
Total		100.000		

Table 2, entry 2



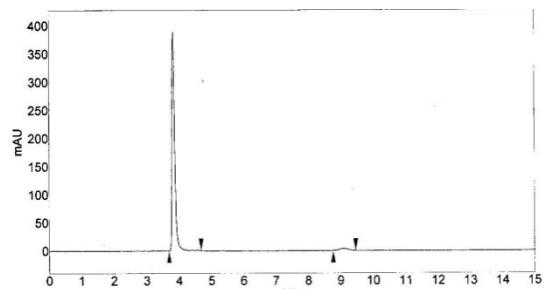
HPLC Condition: Column: Chiralcel OD-H, Daicel Chemical Industries, Ltd.; **Eluent:** Hexanes/IPA (85/15); **Flow rate:** 1.0 mL/min; **Detection:** UV226 nm.

Racemic



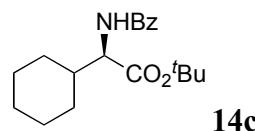
Index	Time [Min]	Area % [%]	Start [Min]	End [Min]
1	3.84	49.842	3.62	4.40
2	9.07	50.158	8.64	9.71
Total		100.000		

Chiral



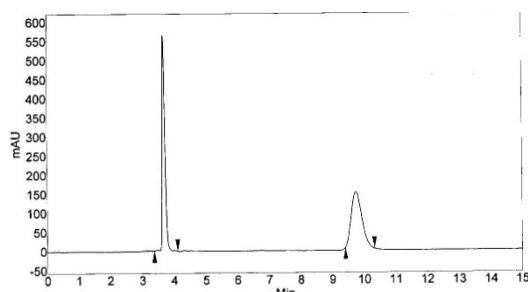
Index	Time [Min]	Area % [%]	Start [Min]	End [Min]
1	3.83	96.834	3.68	4.67
2	9.09	3.166	8.77	9.46
Total		100.000		

Table 2, entry 3



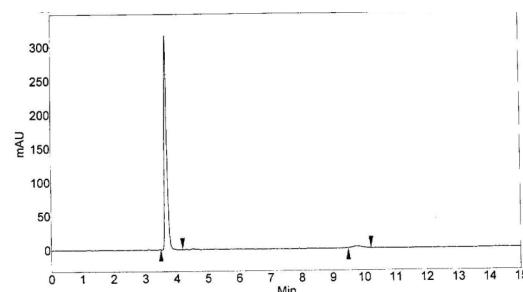
HPLC Condition: **Column:** Chiralcel OD-H, Daicel Chemical Industries, Ltd.; **Eluent:** Hexanes/IPA (85/15); **Flow rate:** 1.0 mL/min; **Detection:** UV226 nm.

Racemic



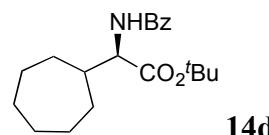
Index	Time [Min]	Area % [%]	Start [Min]	End [Min]
1	3.67	49.822	3.37	4.10
2	9.75	50.178	9.41	10.33
Total		100.000		

Chiral



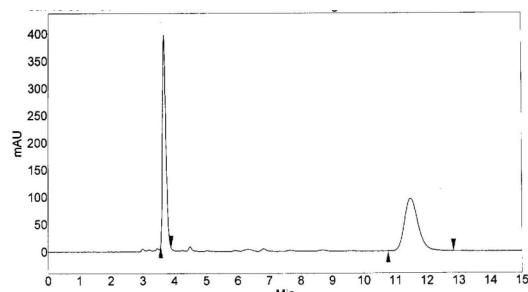
Index	Time [Min]	Area % [%]	Start [Min]	End [Min]
1	3.67	97.100	3.50	4.18
2	9.77	2.900	9.49	10.22
Total		100.000		

Table 2, entry 4



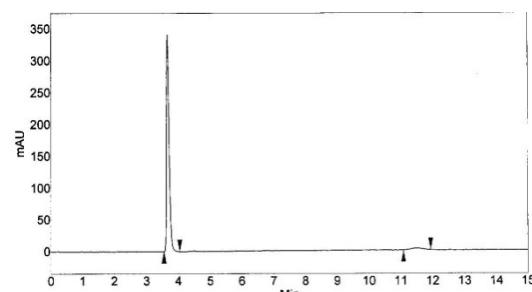
HPLC Condition: **Column:** Chiralcel OD-H, Daicel Chemical Industries, Ltd.; **Eluent:** Hexanes/IPA (85/15); **Flow rate:** 1.0 mL/min; **Detection:** UV226 nm.

Racemic

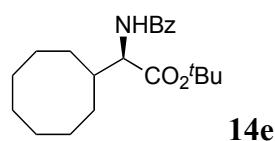


Index	Time [Min]	Area % [%]	Start [Min]	End [Min]
2	3.67	50.052	3.56	3.88
1	11.48	49.948	10.77	12.83
Total		100.000		

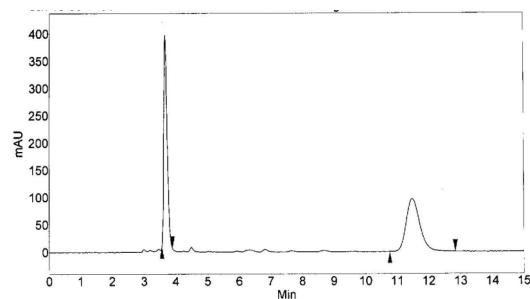
Chiral



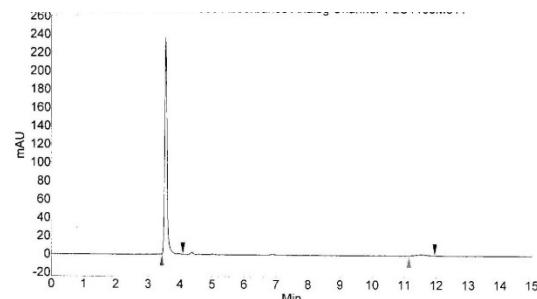
Index	Time [Min]	Area % [%]	Start [Min]	End [Min]
2	3.68	96.938	3.54	4.04
1	11.51	3.062	11.08	11.94
Total		100.000		

Table 2, entry 5

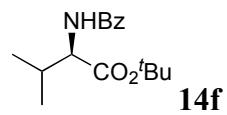
HPLC Condition: **Column:** Chiralcel OD-H, Daicel Chemical Industries, Ltd.; **Eluent:** Hexanes/IPA (85/15); **Flow rate:** 1.0 mL/min; **Detection:** UV226 nm.

Racemic

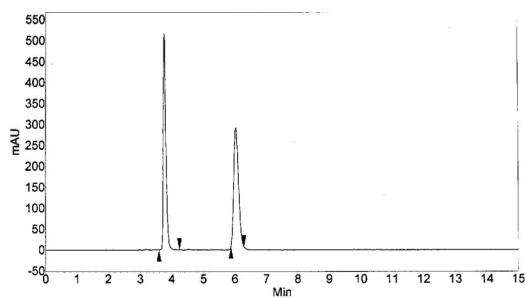
Index	Time [Min]	Area % [%	Start [Min]	End [Min]
2	3.67	50.052	3.56	3.88
1	11.48	49.948	10.77	12.83
Total		100.000		

Chiral

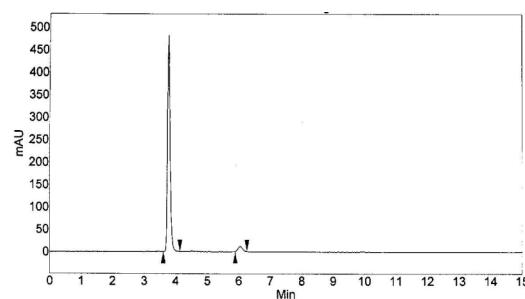
Index	Time [Min]	Area % [%	Start [Min]	End [Min]
1	3.55	97.629	3.44	4.09
2	11.53	2.371	11.15	11.95
Total		100.000		

Table 2, entry 6

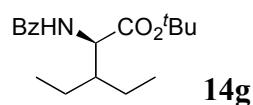
HPLC Condition: **Column:** Chiralcel OD-H, Daicel Chemical Industries, Ltd.; **Eluent:** Hexanes/IPA (85/15); **Flow rate:** 1.0 mL/min; **Detection:** UV226 nm.

Racemic

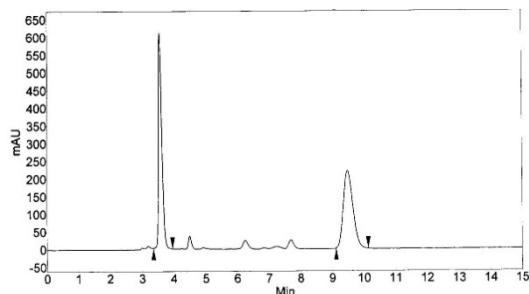
Index	Time [Min]	Area % [%	Start [Min]	End [Min]
1	3.77	49.802	3.59	4.24
2	6.04	50.198	5.87	6.28
Total		100.000		

Chiral

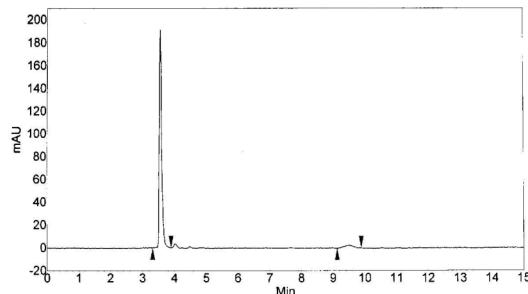
Index	Time [Min]	Area % [%	Start [Min]	End [Min]
1	3.77	96.033	3.61	4.13
2	6.04	3.967	5.88	6.25
Total		100.000		

Table 2, entry 7

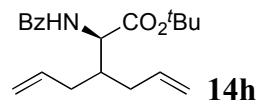
HPLC Condition: Column: Chiralcel OD-H, Daicel Chemical Industries, Ltd.; **Eluent:** Hexanes/IPA (85/15); **Flow rate:** 1.0 mL/min; **Detection:** UV226 nm.

Racemic

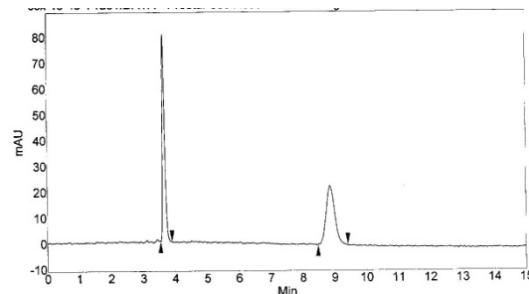
Index	Time [Min]	Area % [%]	Start [Min]	End [Min]
1	3.57	49.988	3.35	3.95
2	9.49	50.012	9.12	10.14
Total		100.000		

Chiral

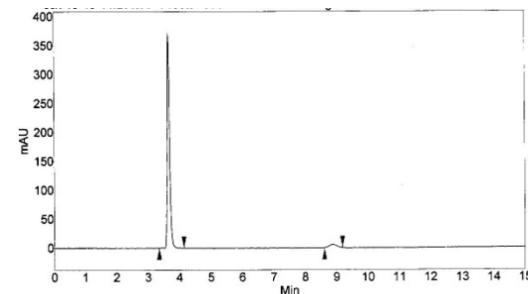
Index	Time [Min]	Area % [%]	Start [Min]	End [Min]
1	3.57	95.302	3.32	3.90
2	9.49	4.698	9.13	9.89
Total		100.000		

Table 2, entry 8

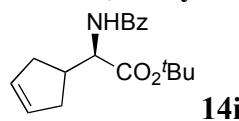
HPLC Condition: Column: Chiralcel OD-H, Daicel Chemical Industries, Ltd.; **Eluent:** Hexanes/IPA (85/15); **Flow rate:** 1.0 mL/min; **Detection:** UV226 nm.

Racemic

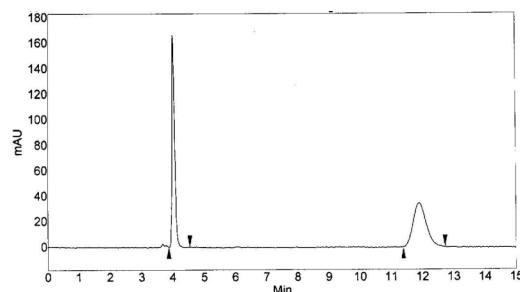
Index	Time [Min]	Area % [%]	Start [Min]	End [Min]
1	3.65	49.930	3.54	3.89
2	8.87	50.070	8.48	9.42
Total		100.000		

Chiral

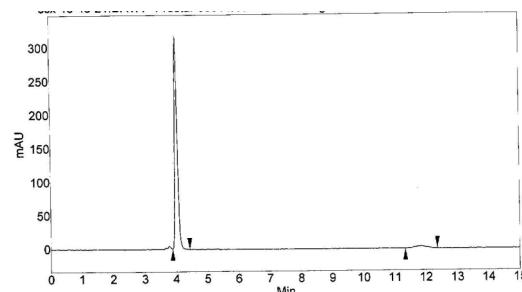
Index	Time [Min]	Area % [%]	Start [Min]	End [Min]
1	3.65	95.756	3.35	4.14
2	8.88	4.244	8.62	9.18
Total		100.000		

Table 2, entry 9

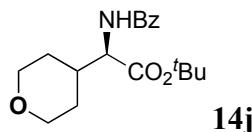
HPLC Condition: Column: Chiralcel OD-H, Daicel Chemical Industries, Ltd.; **Eluent:** Hexanes/IPA (85/15); **Flow rate:** 1.0 mL/min; **Detection:** UV226 nm.

Racemic

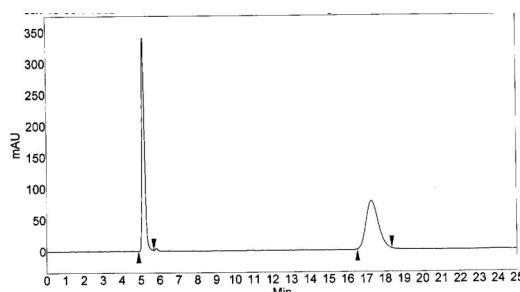
Index	Time [Min]	Area %	Start [Min]	End [Min]
1	4.01	49.945	3.87	4.54
2	11.91	50.055	11.39	12.73
Total		100.000		

Chiral

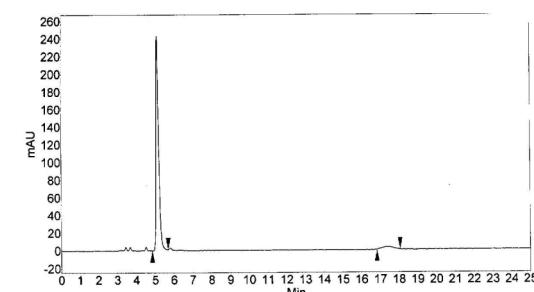
Index	Time [Min]	Area %	Start [Min]	End [Min]
1	4.01	95.802	3.88	4.43
2	11.91	4.198	11.34	12.36
Total		100.000		

Table 2, entry 10

HPLC Condition: Column: Chiralcel OD-H, Daicel Chemical Industries, Ltd.; **Eluent:** Hexanes/IPA (85/15); **Flow rate:** 1.0 mL/min; **Detection:** UV226 nm.

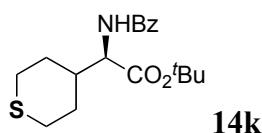
Racemic

Index	Time [Min]	Area %	Start [Min]	End [Min]
1	5.15	49.976	4.88	5.68
2	17.29	50.024	16.54	18.37
Total		100.000		

Chiral

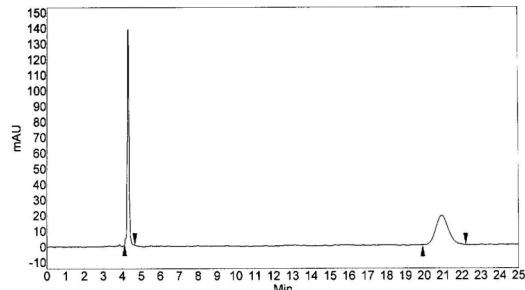
Index	Time [Min]	Area %	Start [Min]	End [Min]
1	5.13	95.501	4.85	5.68
2	17.24	4.499	16.82	18.06
Total		100.000		

Table 2, entry 11



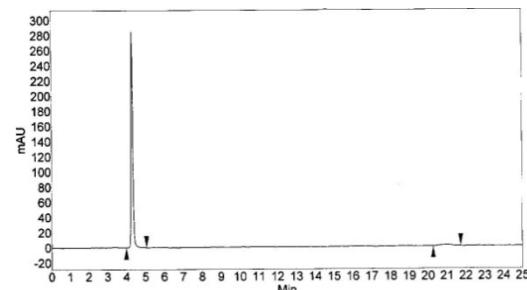
HPLC Condition: **Column:** Chiralcel OD-H, Daicel Chemical Industries, Ltd.; **Eluent:** Hexanes/IPA (85/15); **Flow rate:** 1.0 mL/min; **Detection:** UV226 nm.

Racemic



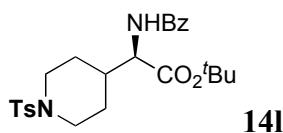
Index	Time [Min]	Area % [%]	Start [Min]	End [Min]
1	4.32	49.953	4.13	4.67
2	20.93	50.047	19.92	22.19
Total		100.000		

Chiral



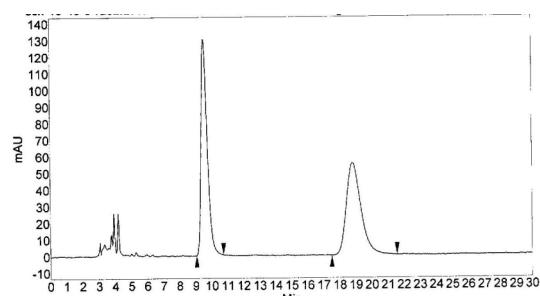
Index	Time [Min]	Area % [%]	Start [Min]	End [Min]
1	4.29	96.272	4.00	5.06
2	21.04	3.728	20.28	21.72
Total		100.000		

Table 2, entry 12



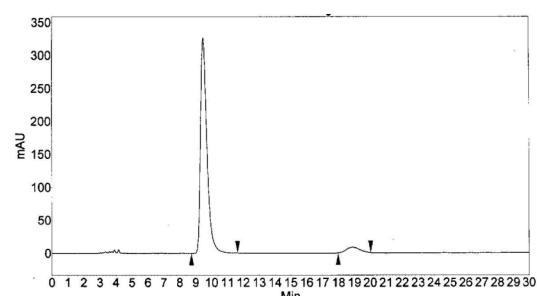
HPLC Condition: **Column:** Chiralcel OD-H, Daicel Chemical Industries, Ltd.; **Eluent:** Hexanes/IPA (85/15); **Flow rate:** 1.0 mL/min; **Detection:** UV226 nm.

Racemic

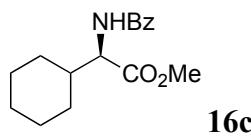


Index	Time [Min]	Area % [%]	Start [Min]	End [Min]
2	9.55	50.088	9.06	10.73
1	18.83	49.912	17.51	21.57
Total		100.000		

Chiral

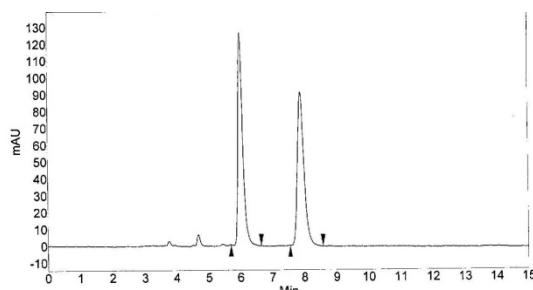


Index	Time [Min]	Area % [%]	Start [Min]	End [Min]
1	9.49	94.924	8.73	11.64
2	18.93	5.076	17.99	20.03
Total		100.000		



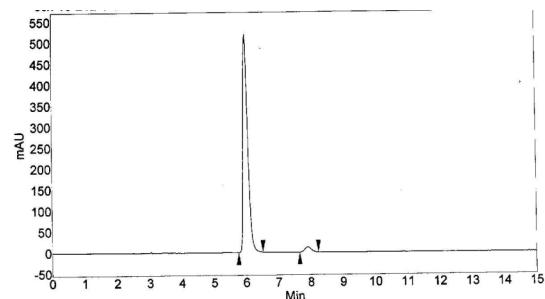
HPLC Condition: **Column:** Chiralcel OD-H, Daicel Chemical Industries, Ltd.; **Eluent:** Hexanes/IPA (90/10); **Flow rate:** 1.0 mL/min; **Detection:** UV226 nm.

Racemic



Index	Time [Min]	Area %	Start [Min]	End [Min]
2	6.00	50.173	5.70	6.64
1	7.88	49.827	7.55	8.58
Total		100.000		

Chiral



Index	Time [Min]	Area %	Start [Min]	End [Min]
2	5.99	96.889	5.76	6.51
1	7.91	3.111	7.65	8.23
Total		100.000		

The X-ray structure of compound 14j

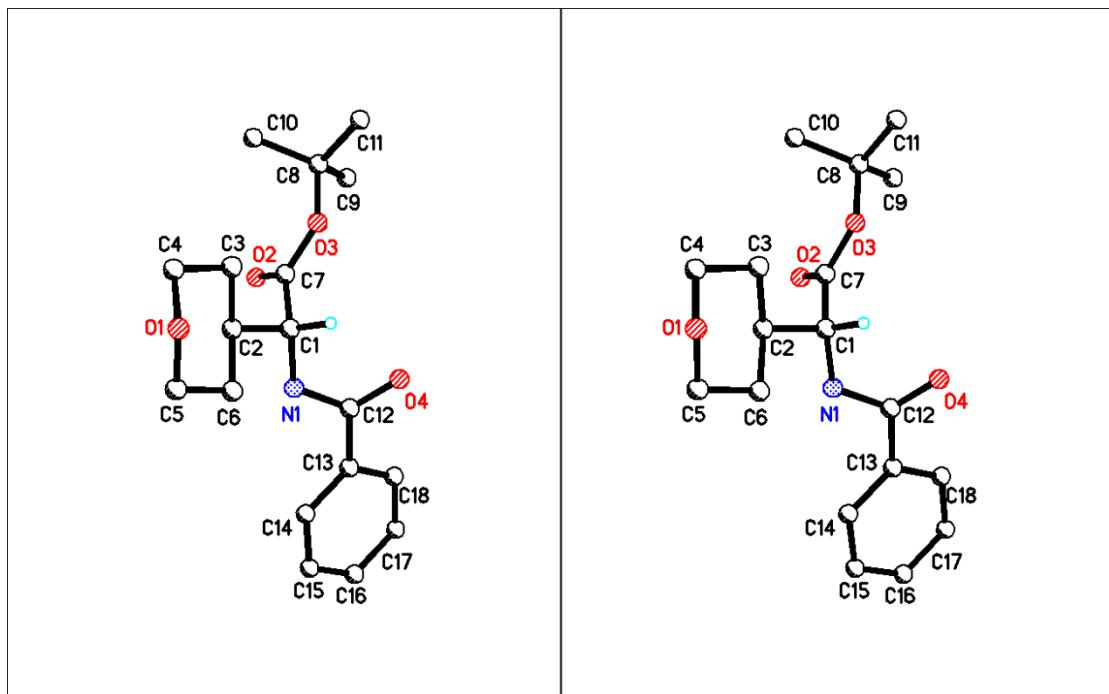
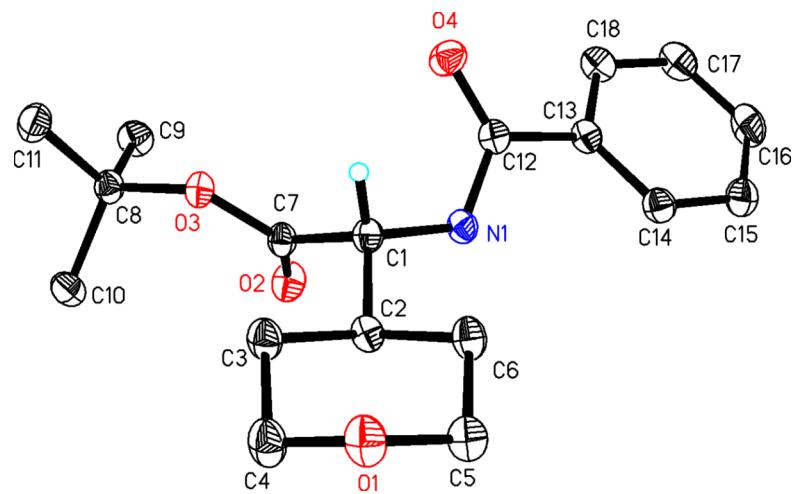


Table 1. Crystal data and structure refinement for **14j**.

Identification code	14j		
Empirical formula	C18 H25 N O4		
Formula weight	319.39		
Temperature	173.1500 K		
Wavelength	0.71073 Å		
Crystal system	Orthorhombic		
Space group	P 21 21 21		
Unit cell dimensions	a = 5.5701(4) Å	b = 9.0120(6) Å	c = 33.683(2) Å
			a = 90°.
			b = 90°.
			g = 90°.
Volume	1690.8(2) Å ³		
Z	4		
Density (calculated)	1.255 Mg/m ³		
Absorption coefficient	0.088 mm ⁻¹		
F(000)	688		
Crystal size	0.34 x 0.14 x 0.1 mm ³		
Theta range for data collection	3.311 to 27.479°.		
Index ranges	-7<=h<=7, -11<=k<=11, -43<=l<=42		
Reflections collected	20256		
Independent reflections	3875 [R(int) = 0.0705]		
Completeness to theta = 26.000°	99.7 %		
Absorption correction	Semi-empirical from equivalents		
Max. and min. transmission	1.0000 and 0.5829		
Refinement method	Full-matrix least-squares on F ²		
Data / restraints / parameters	3875 / 0 / 211		
Goodness-of-fit on F ²	1.116		
Final R indices [I>2sigma(I)]	R1 = 0.0531, wR2 = 0.1152		
R indices (all data)	R1 = 0.0671, wR2 = 0.1257		
Absolute structure parameter	0.2(7)		
Extinction coefficient	n/a		
Largest diff. peak and hole	0.185 and -0.264 e.Å ⁻³		

Table 2. Atomic coordinates ($\times 10^4$) and equivalent isotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for **14j**. U(eq) is defined as one third of the trace of the orthogonalized U^{ij} tensor.

	x	y	z	U(eq)
O1	-2088(4)	3181(3)	7456(1)	43(1)
O2	1300(4)	6221(2)	5964(1)	40(1)
O3	3951(4)	6378(2)	6474(1)	31(1)
O4	5572(4)	2806(3)	5965(1)	42(1)
N1	1579(4)	3120(3)	6052(1)	31(1)
C1	1924(5)	4102(3)	6389(1)	30(1)
C2	-207(5)	4024(3)	6679(1)	30(1)
C3	297(6)	4894(4)	7061(1)	37(1)
C4	-1746(7)	4697(4)	7354(1)	45(1)
C5	-2697(6)	2339(4)	7114(1)	39(1)
C6	-781(6)	2419(3)	6792(1)	35(1)
C7	2338(5)	5680(3)	6244(1)	30(1)
C8	4615(5)	7952(3)	6407(1)	31(1)
C9	5759(6)	8119(4)	6000(1)	41(1)
C10	2463(6)	8948(3)	6461(1)	39(1)
C11	6409(6)	8208(4)	6737(1)	41(1)
C12	3488(6)	2531(3)	5861(1)	31(1)
C13	2973(5)	1501(3)	5525(1)	31(1)
C14	876(6)	675(3)	5504(1)	37(1)
C15	543(7)	-337(4)	5195(1)	46(1)
C16	2263(8)	-487(4)	4905(1)	51(1)
C17	4332(7)	354(4)	4920(1)	45(1)
C18	4706(6)	1341(4)	5229(1)	37(1)

Table 3. Bond lengths [\AA] and angles [$^\circ$] for **14j**.

O1-C4	1.421(4)
O1-C5	1.422(4)
O2-C7	1.209(3)
O3-C7	1.343(3)
O3-C8	1.484(3)
O4-C12	1.238(4)
N1-C1	1.454(3)
N1-C12	1.351(4)
C1-C2	1.539(4)
C1-C7	1.521(4)
C2-C3	1.531(4)
C2-C6	1.529(4)
C3-C4	1.518(5)
C5-C6	1.521(4)
C8-C9	1.520(4)
C8-C10	1.509(4)
C8-C11	1.513(4)
C12-C13	1.491(4)
C13-C14	1.387(4)
C13-C18	1.396(4)
C14-C15	1.396(4)
C15-C16	1.374(5)
C16-C17	1.380(5)
C17-C18	1.385(4)
C4-O1-C5	110.5(2)
C7-O3-C8	121.8(2)
C12-N1-C1	120.5(2)
N1-C1-C2	111.5(2)
N1-C1-C7	109.7(2)
C7-C1-C2	111.3(2)
C3-C2-C1	111.6(2)
C6-C2-C1	111.3(2)
C6-C2-C3	108.2(2)
C4-C3-C2	110.4(3)
O1-C4-C3	111.7(3)

O1-C5-C6	112.5(3)
C5-C6-C2	111.7(2)
O2-C7-O3	125.5(3)
O2-C7-C1	123.7(3)
O3-C7-C1	110.7(2)
O3-C8-C9	109.7(2)
O3-C8-C10	110.7(2)
O3-C8-C11	101.4(2)
C10-C8-C9	112.5(3)
C11-C8-C9	111.8(3)
O4-C12-N1	121.6(3)
O4-C12-C13	121.4(3)
N1-C12-C13	117.0(3)
C14-C13-C12	122.4(3)
C14-C13-C18	119.3(3)
C18-C13-C12	118.3(3)
C13-C14-C15	120.1(3)
C16-C15-C14	120.0(3)
C15-C16-C17	120.2(3)
C16-C17-C18	120.3(3)
C17-C18-C13	120.0(3)

Symmetry transformations used to generate equivalent atoms:

Table 4. Anisotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for **14j**. The anisotropic displacement factor exponent takes the form: $-2p^2 [h^2 a^{*2} U^{11} + \dots + 2 h k a^* b^* U^{12}]$

	U^{11}	U^{22}	U^{33}	U^{23}	U^{13}	U^{12}
O1	61(2)	43(1)	25(1)	-1(1)	6(1)	-5(1)
O2	50(1)	39(1)	33(1)	5(1)	-9(1)	-5(1)
O3	37(1)	27(1)	30(1)	-1(1)	-3(1)	-4(1)
O4	35(1)	55(1)	35(1)	-10(1)	0(1)	-4(1)
N1	35(1)	33(1)	24(1)	-6(1)	0(1)	-1(1)
C1	34(2)	31(1)	24(1)	-1(1)	-2(1)	-1(1)
C2	37(2)	30(1)	23(1)	1(1)	-2(1)	2(1)
C3	45(2)	37(2)	28(1)	-5(1)	3(1)	-3(2)
C4	60(2)	40(2)	34(2)	-6(1)	11(2)	-8(2)
C5	49(2)	38(2)	31(1)	2(1)	0(1)	-2(2)
C6	49(2)	31(2)	26(1)	-1(1)	0(1)	-2(1)
C7	36(2)	29(1)	25(1)	-3(1)	1(1)	-2(1)
C8	35(2)	26(1)	33(1)	0(1)	3(1)	-4(1)
C9	43(2)	42(2)	38(2)	2(1)	11(2)	-2(2)
C10	41(2)	34(2)	42(2)	-1(1)	7(1)	4(1)
C11	41(2)	40(2)	43(2)	-4(1)	-2(2)	-7(2)
C12	37(2)	31(2)	24(1)	1(1)	2(1)	0(1)
C13	39(2)	32(2)	22(1)	-2(1)	0(1)	2(1)
C14	41(2)	37(2)	32(2)	-3(1)	2(1)	1(1)
C15	50(2)	41(2)	47(2)	-11(2)	-4(2)	-3(2)
C16	70(2)	46(2)	37(2)	-17(2)	-8(2)	6(2)
C17	58(2)	50(2)	28(2)	-5(1)	4(2)	9(2)
C18	44(2)	42(2)	26(1)	1(1)	4(1)	4(2)

Table 5. Hydrogen coordinates ($\times 10^4$) and isotropic displacement parameters ($\text{\AA}^2 \times 10^{-3}$) for **14j**.

	x	y	z	U(eq)
H1	150	2913	5973	37
H1A	3368	3780	6531	35
H2	-1620	4455	6550	36
H3A	1782	4545	7179	44
H3B	485	5938	6999	44
H4A	-1403	5261	7593	54
H4B	-3214	5085	7239	54
H5A	-4203	2702	7007	47
H5B	-2927	1311	7190	47
H6A	-1332	1889	6559	42
H6B	669	1938	6887	42
H9A	7046	7418	5973	61
H9B	6377	9108	5971	61
H9C	4576	7938	5798	61
H10A	1298	8736	6259	58
H10B	2955	9966	6441	58
H10C	1766	8776	6717	58
H11A	5636	8065	6989	62
H11B	7015	9203	6721	62
H11C	7712	7518	6711	62
H14	-308	795	5696	44
H15	-845	-910	5185	55
H16	2033	-1157	4699	61
H17	5480	258	4721	54
H18	6112	1896	5239	45

Table 6. Torsion angles [°] for **14j**.

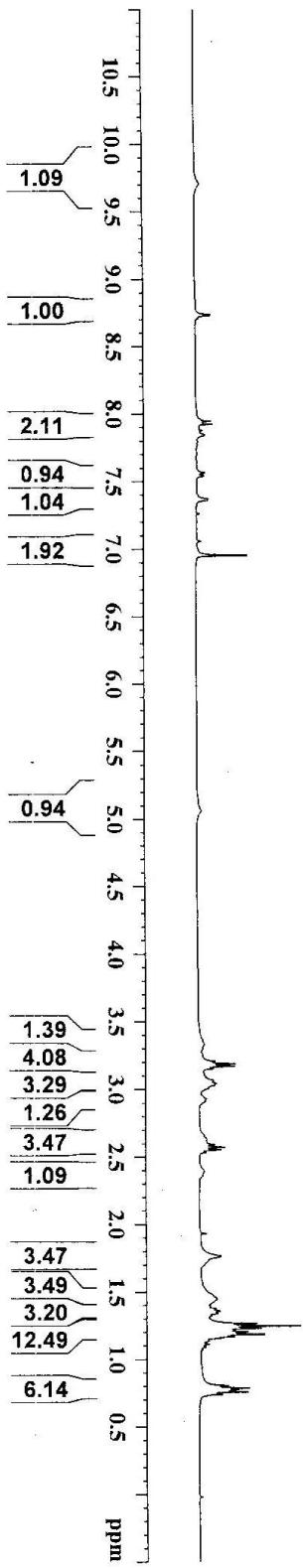
O1-C5-C6-C2	-54.8(3)
O4-C12-C13-C14	151.3(3)
O4-C12-C13-C18	-26.8(4)
N1-C1-C2-C3	172.4(2)
N1-C1-C2-C6	51.4(3)
N1-C1-C7-O2	38.2(4)
N1-C1-C7-O3	-143.3(2)
N1-C12-C13-C14	-27.1(4)
N1-C12-C13-C18	154.8(3)
C1-N1-C12-O4	-0.1(4)
C1-N1-C12-C13	178.3(2)
C1-C2-C3-C4	-175.5(3)
C1-C2-C6-C5	173.7(2)
C2-C1-C7-O2	-85.7(3)
C2-C1-C7-O3	92.8(3)
C2-C3-C4-O1	59.4(4)
C3-C2-C6-C5	50.7(3)
C4-O1-C5-C6	59.0(3)
C5-O1-C4-C3	-61.5(4)
C6-C2-C3-C4	-52.8(3)
C7-O3-C8-C9	-62.4(3)
C7-O3-C8-C10	62.3(3)
C7-O3-C8-C11	179.3(2)
C7-C1-C2-C3	-64.7(3)
C7-C1-C2-C6	174.3(2)
C8-O3-C7-O2	0.9(4)
C8-O3-C7-C1	-177.6(2)
C12-N1-C1-C2	-151.5(3)
C12-N1-C1-C7	84.6(3)
C12-C13-C14-C15	-176.3(3)
C12-C13-C18-C17	177.6(3)
C13-C14-C15-C16	-1.8(5)
C14-C13-C18-C17	-0.6(5)
C14-C15-C16-C17	0.4(5)
C15-C16-C17-C18	0.9(5)
C16-C17-C18-C13	-0.8(5)

C18-C13-C14-C15 1.8(5)

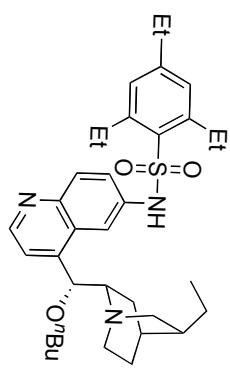
Symmetry transformations used to generate equivalent atoms:

Table 7. Hydrogen bonds for **14j** [Å and °].

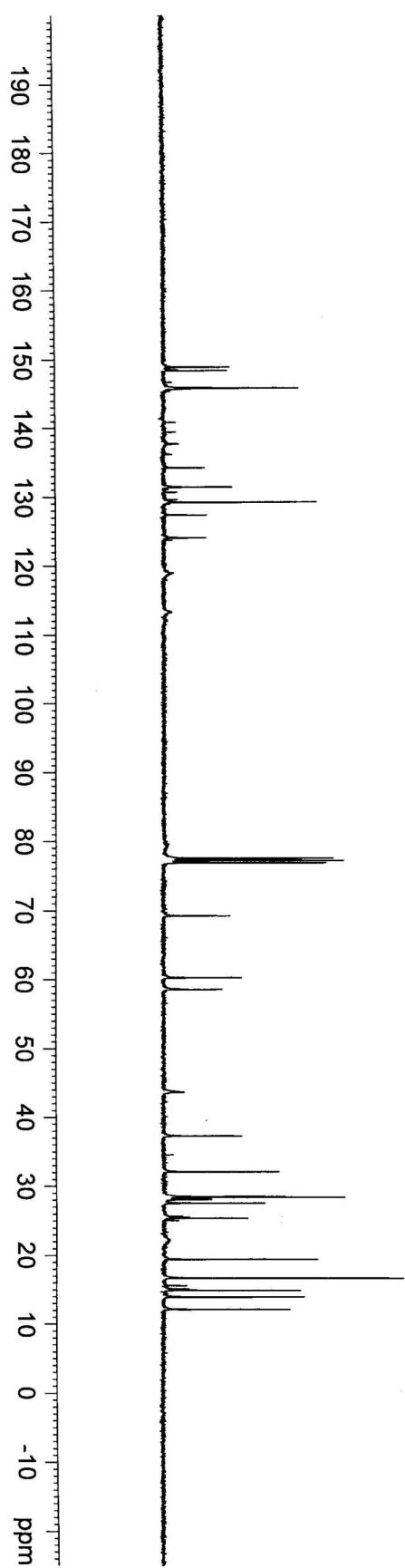
D-H...A	d(D-H)	d(H...A)	d(D...A)	<(DHA)



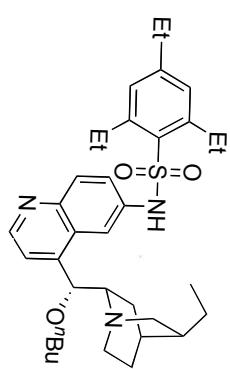
(mixture of rotamers)



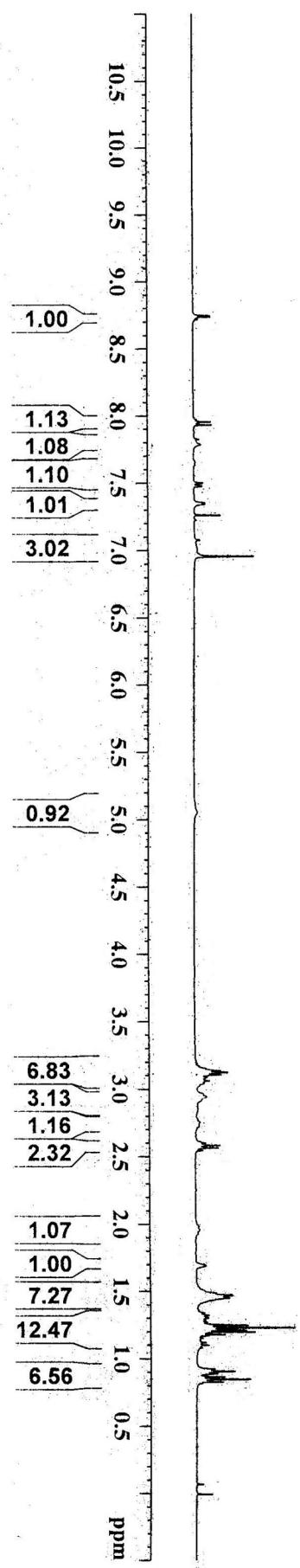
C4



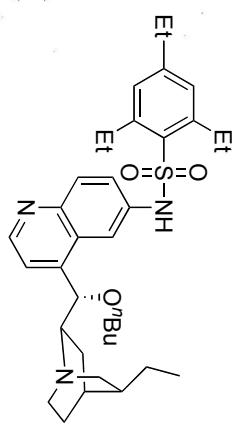
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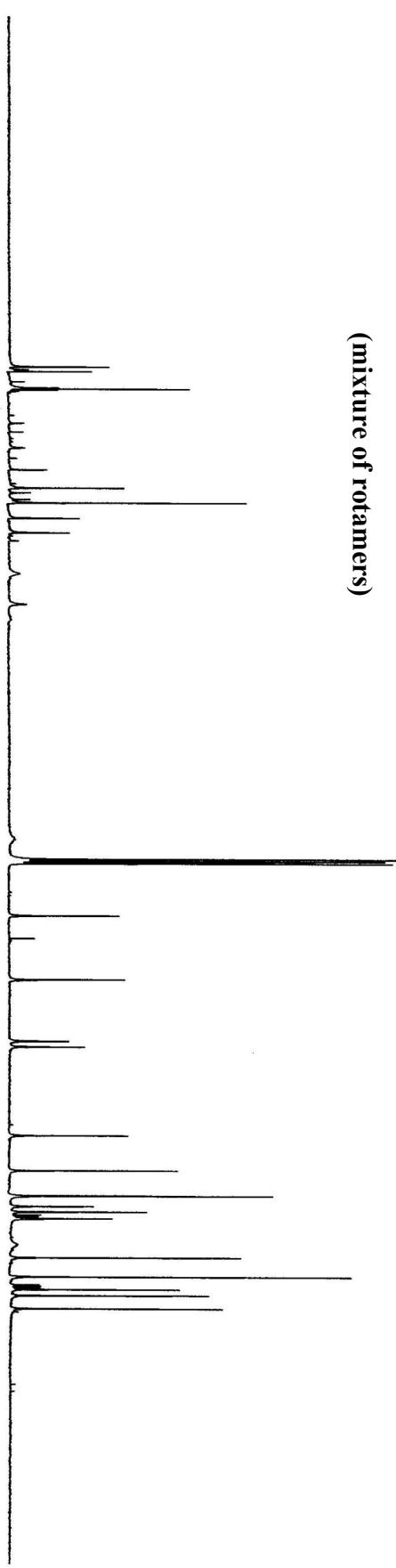
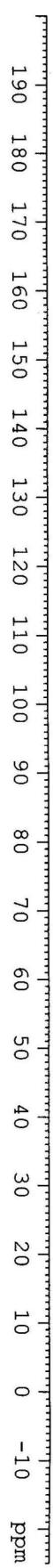
C4



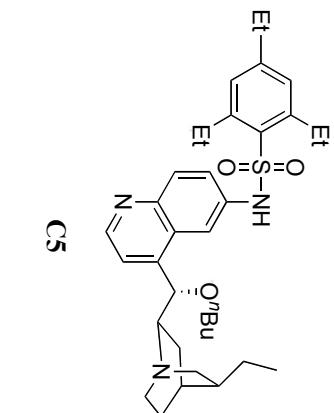
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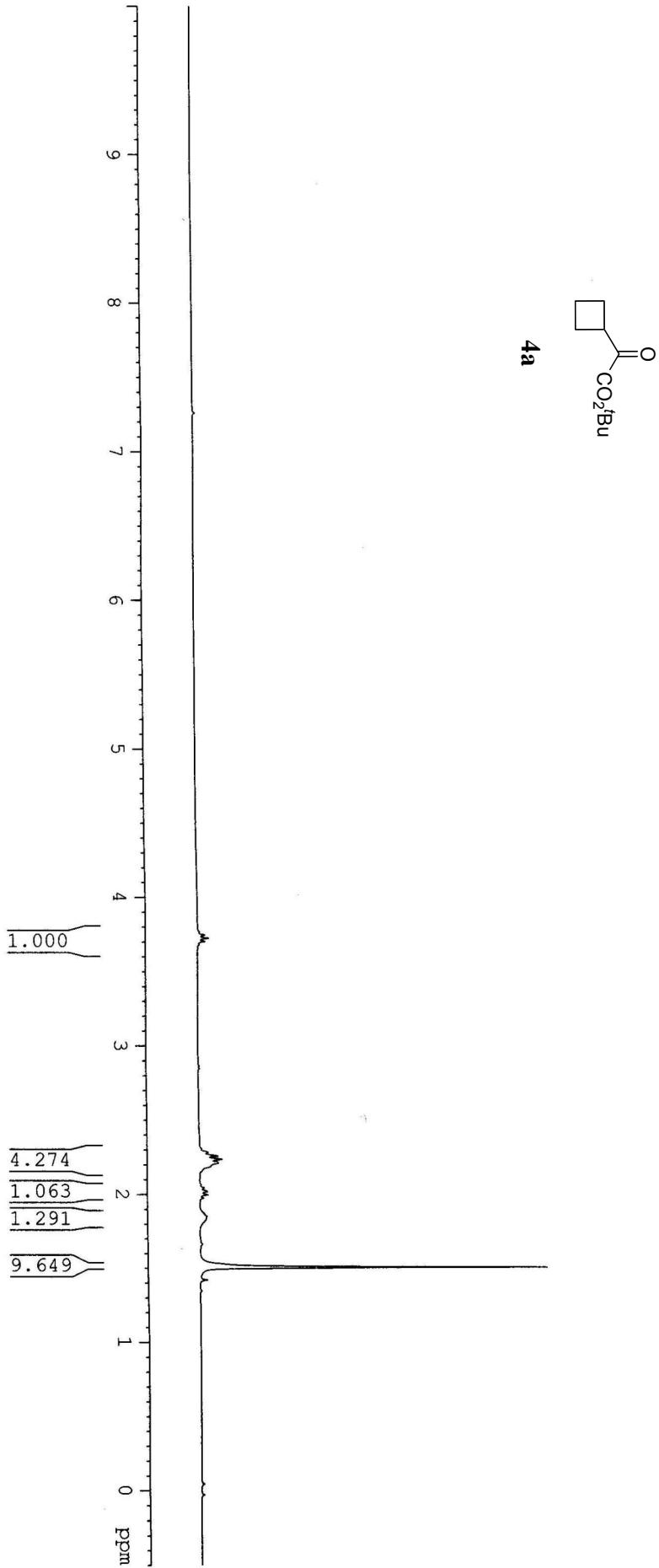
C5

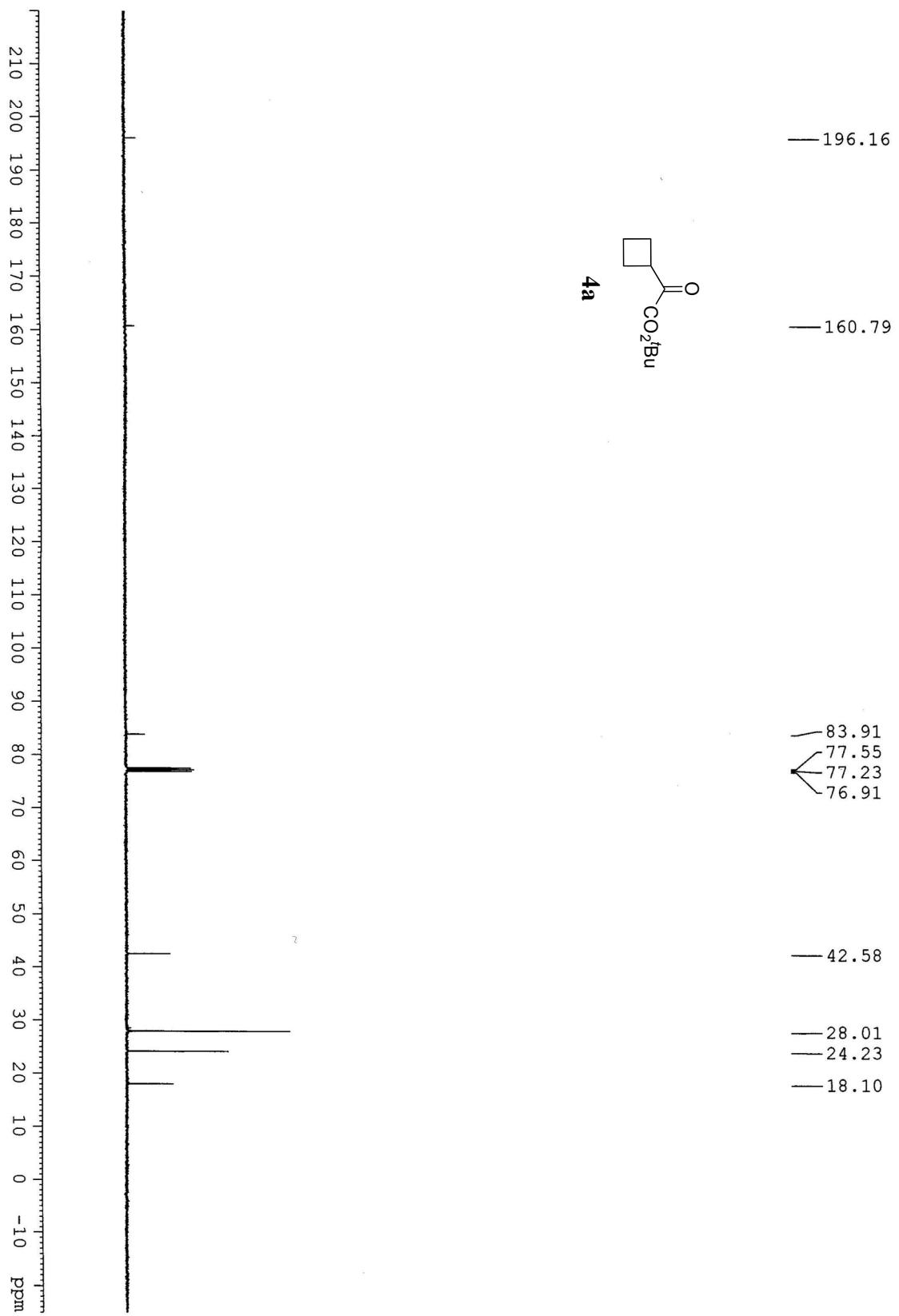


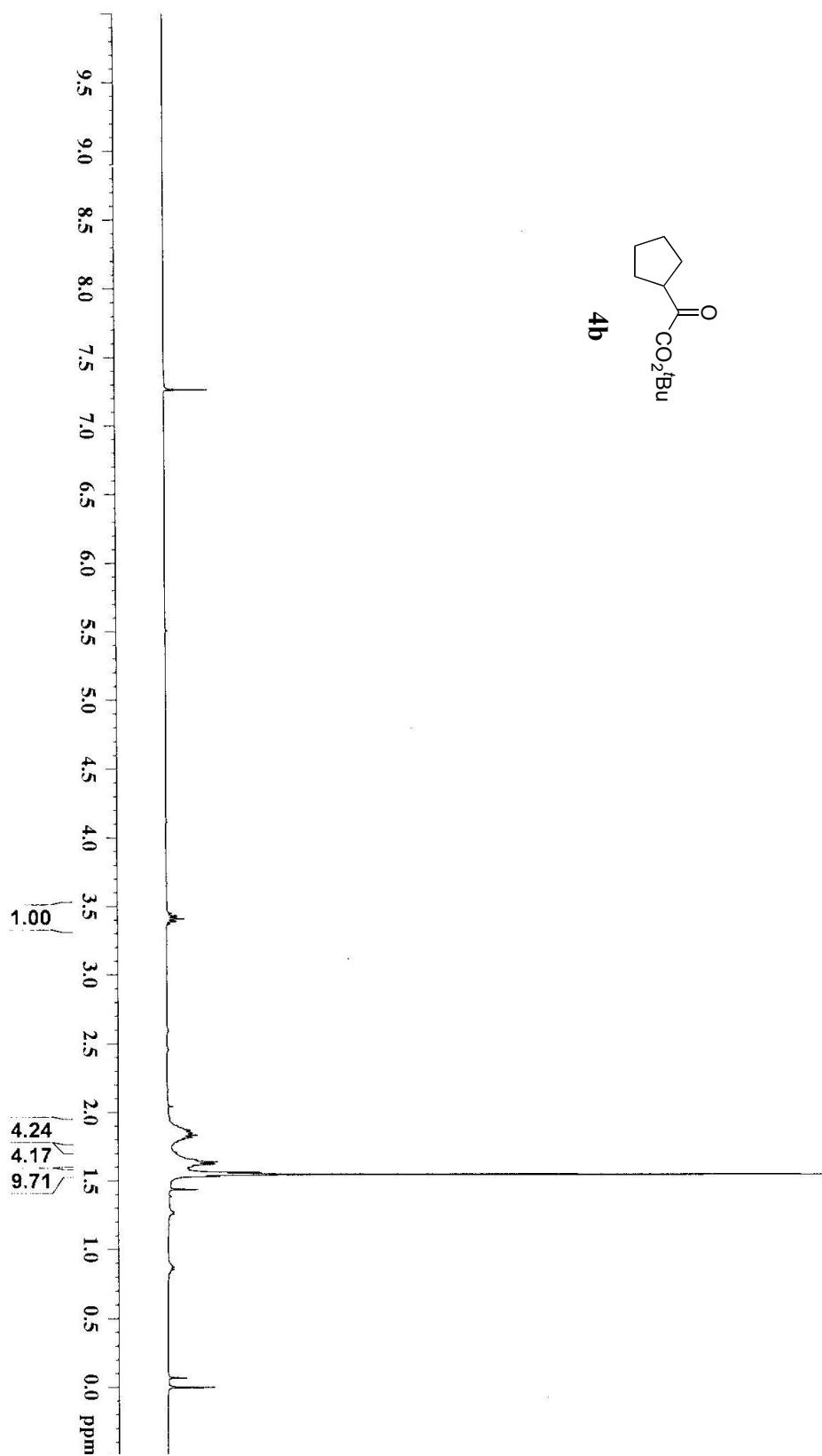
(mixture of rotamers)

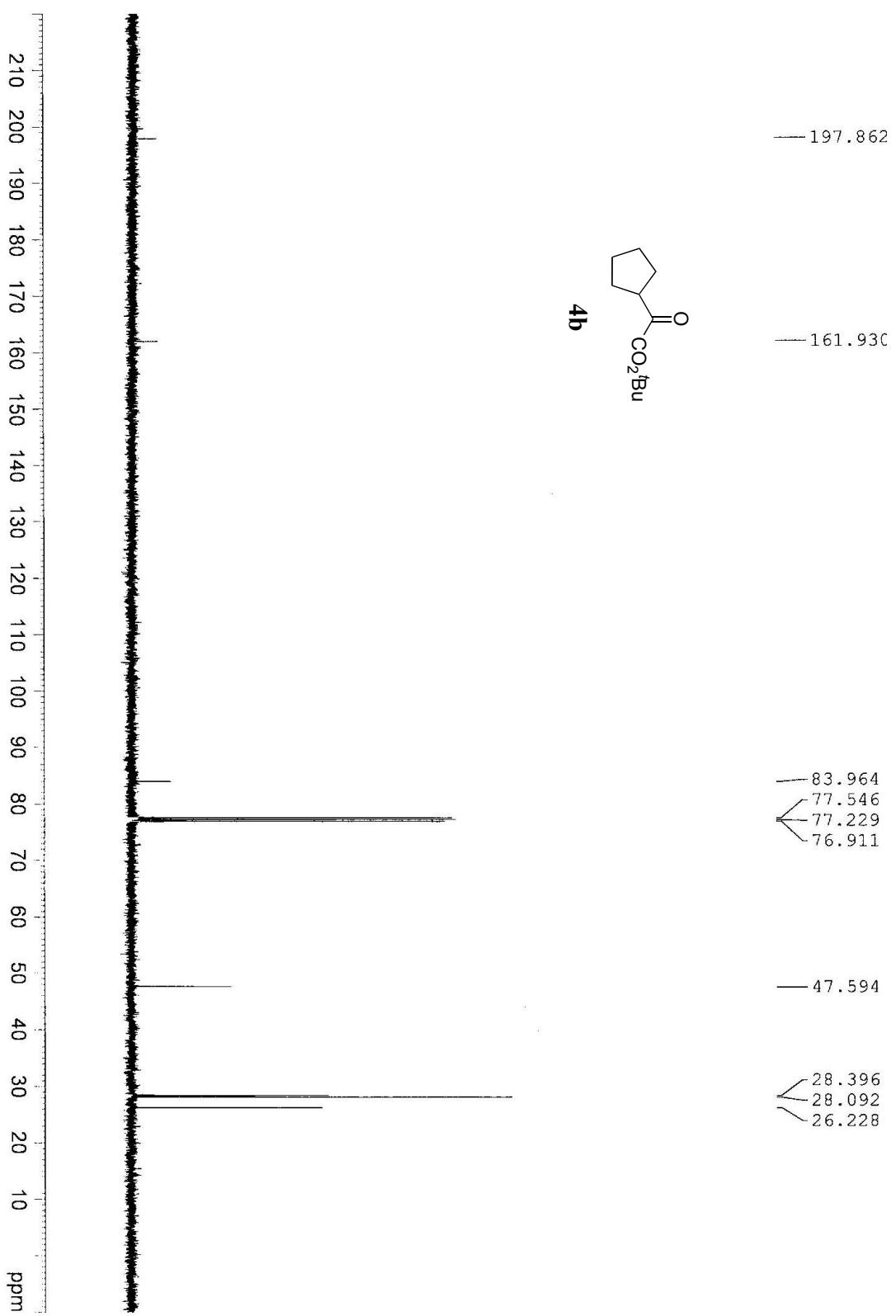


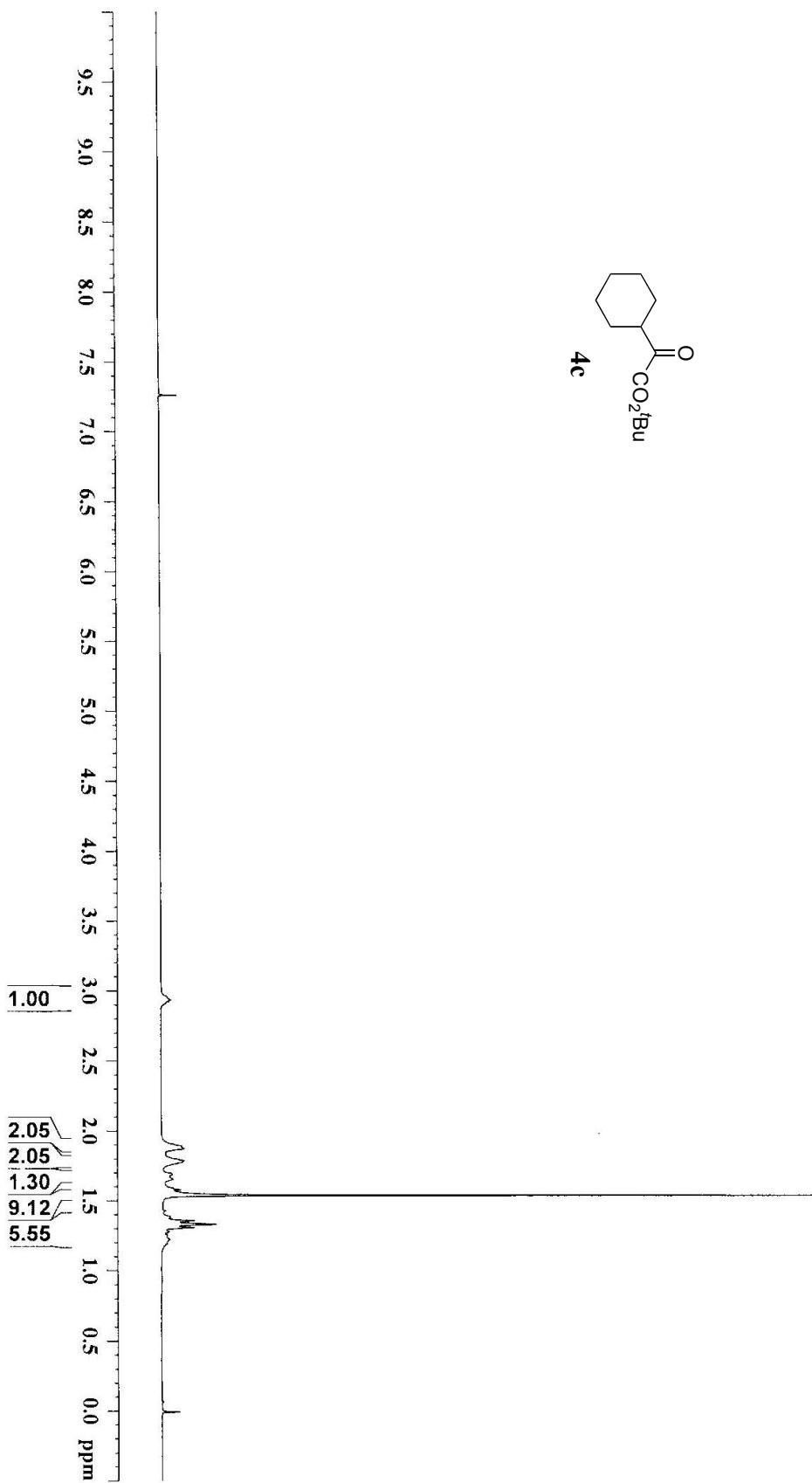
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146.01
145.85
145.72
137.25
134.04
131.39
129.23
127.01
124.91
118.99
114.59
- 80.43
77.54
77.23
76.91
69.30
- 60.01
- 51.02
50.21
37.31
32.19
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27.00
26.20
25.24
21.53
19.54
16.77
14.92
14.03
12.12

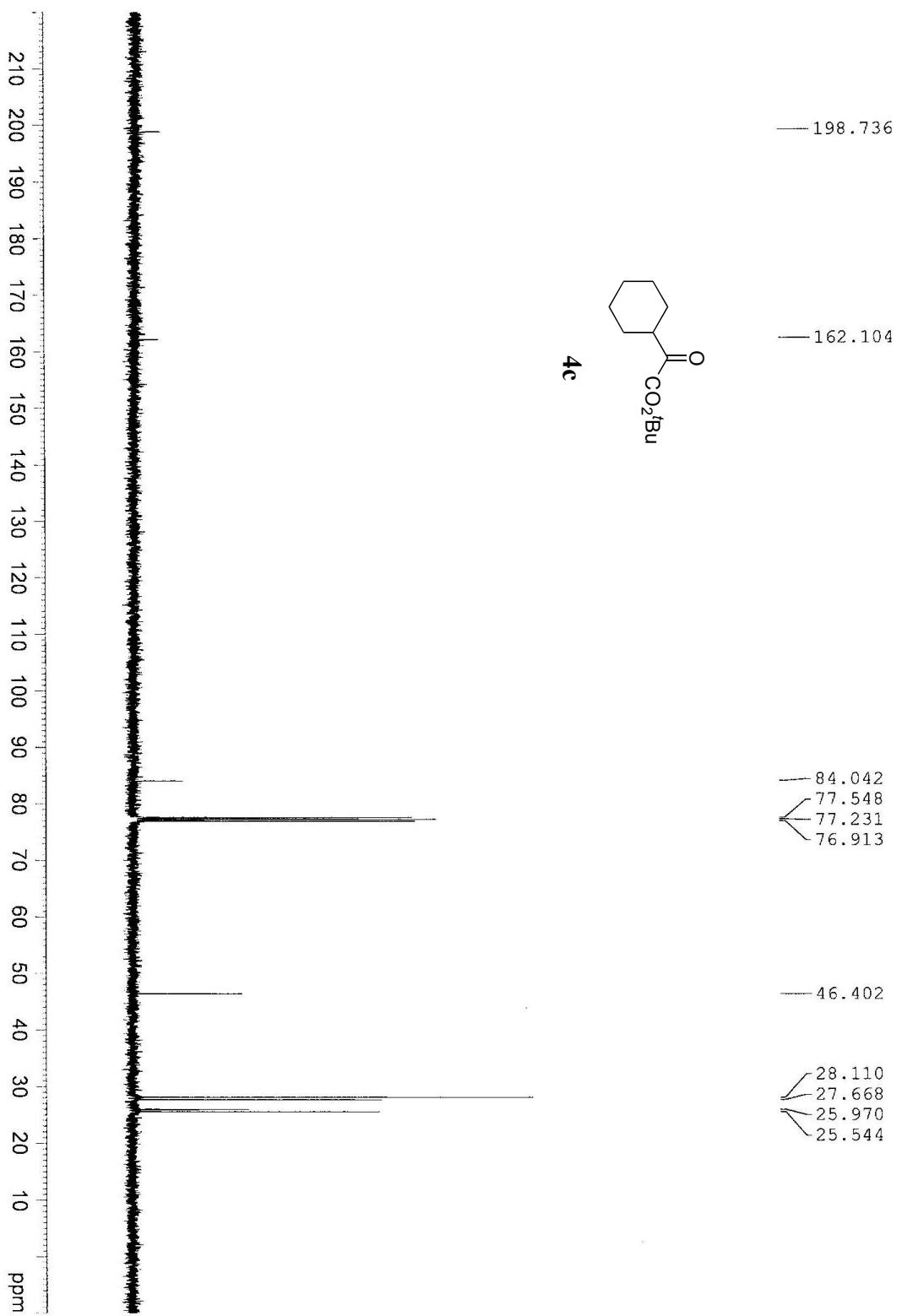


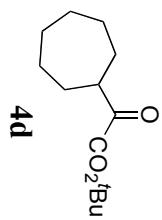
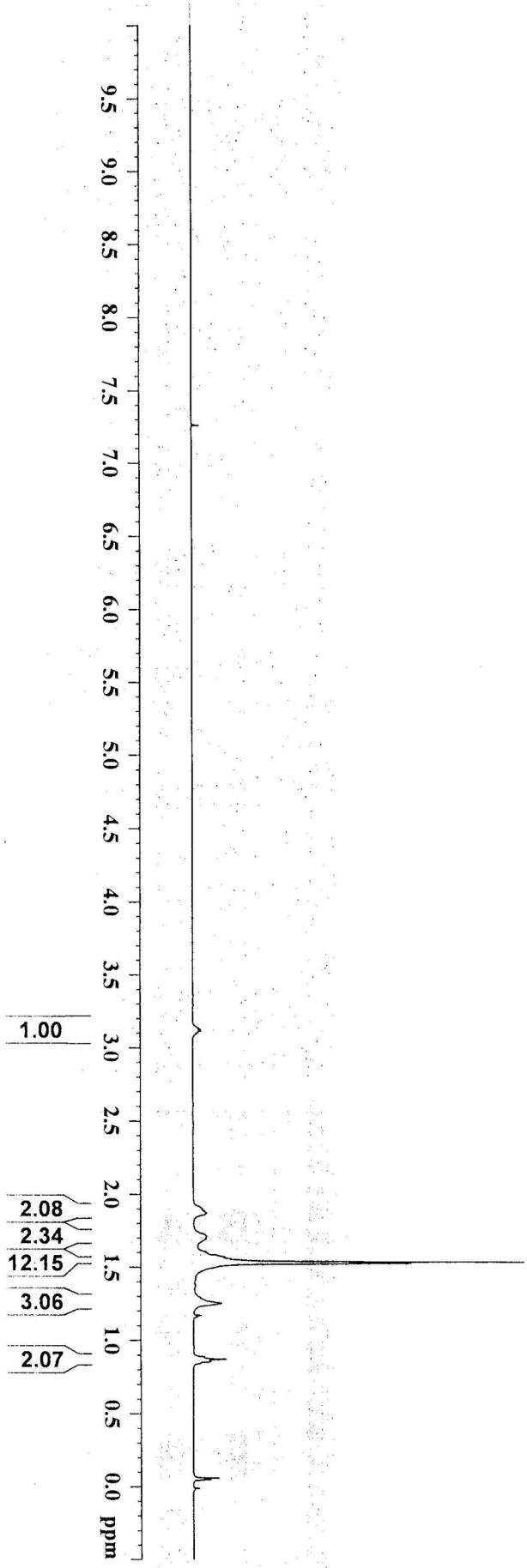


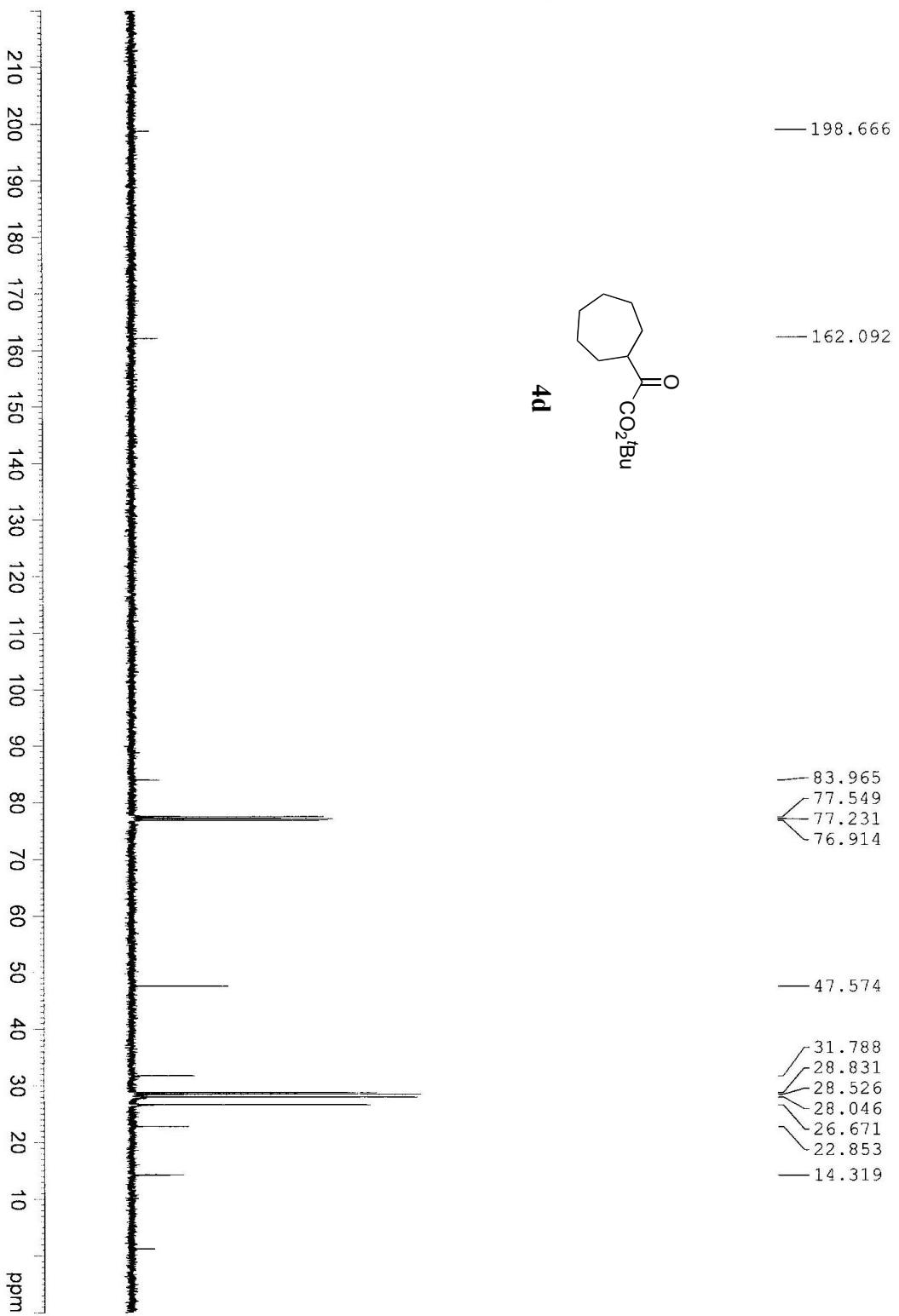


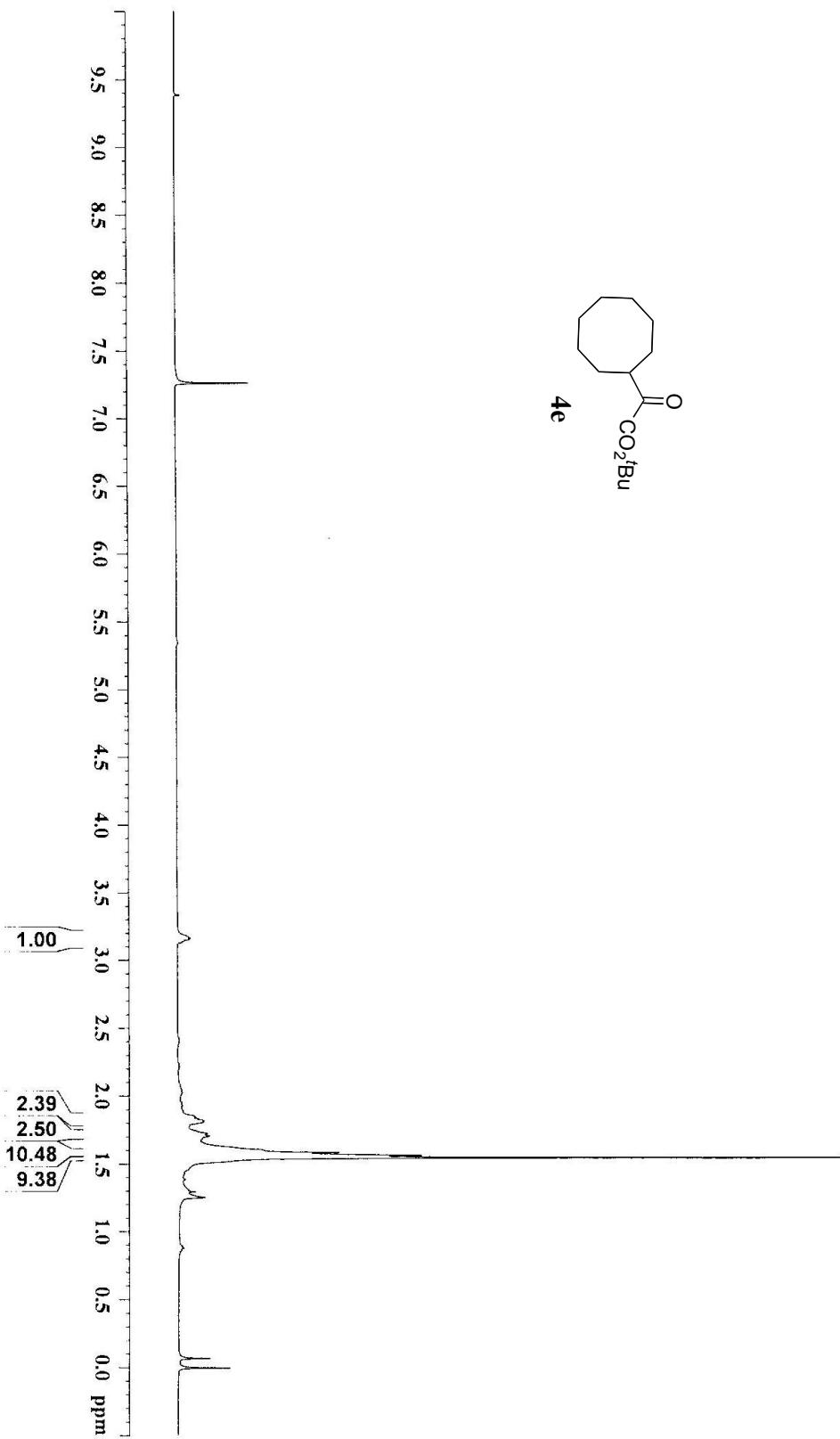


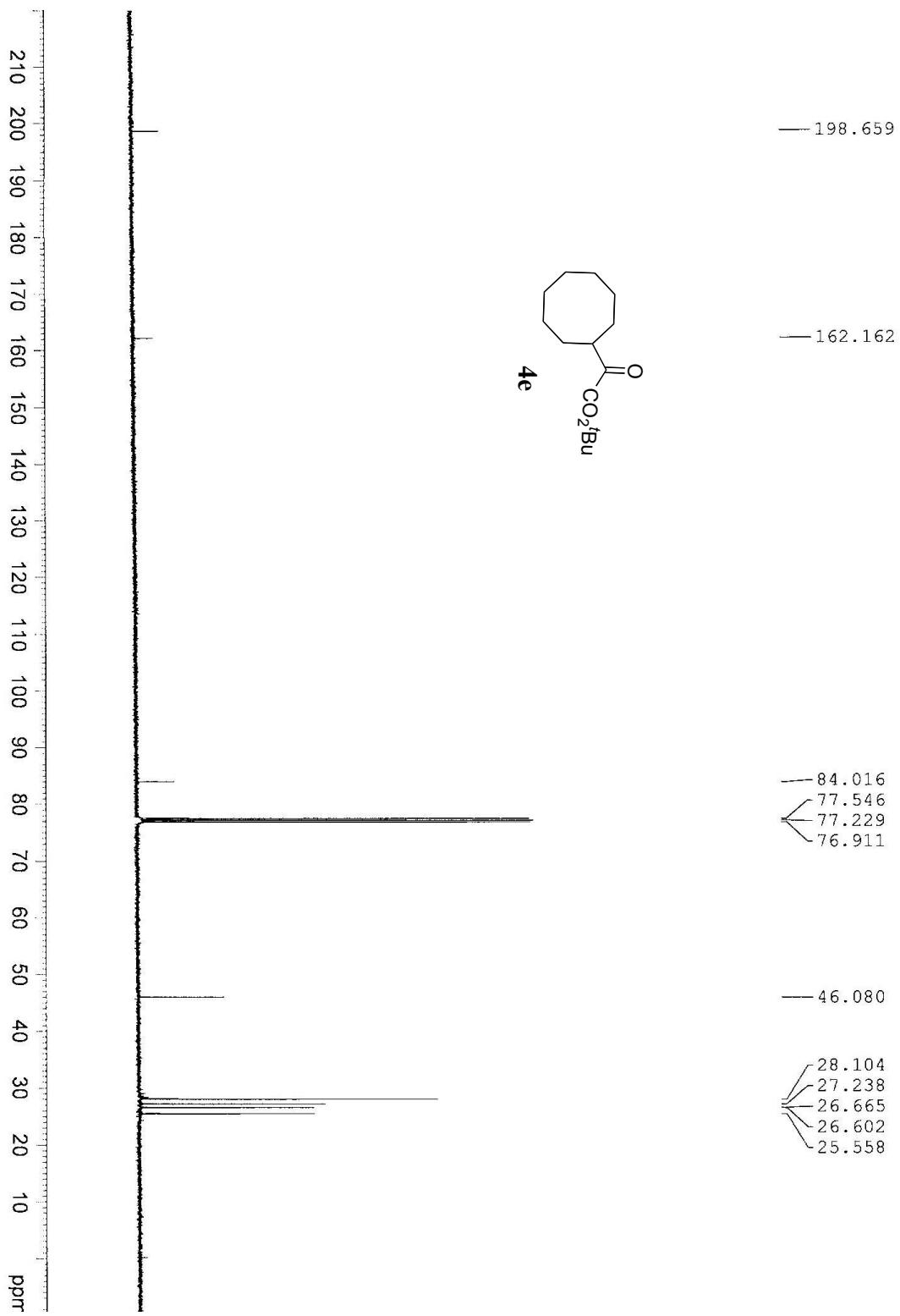


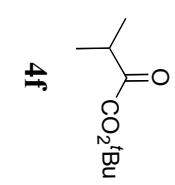
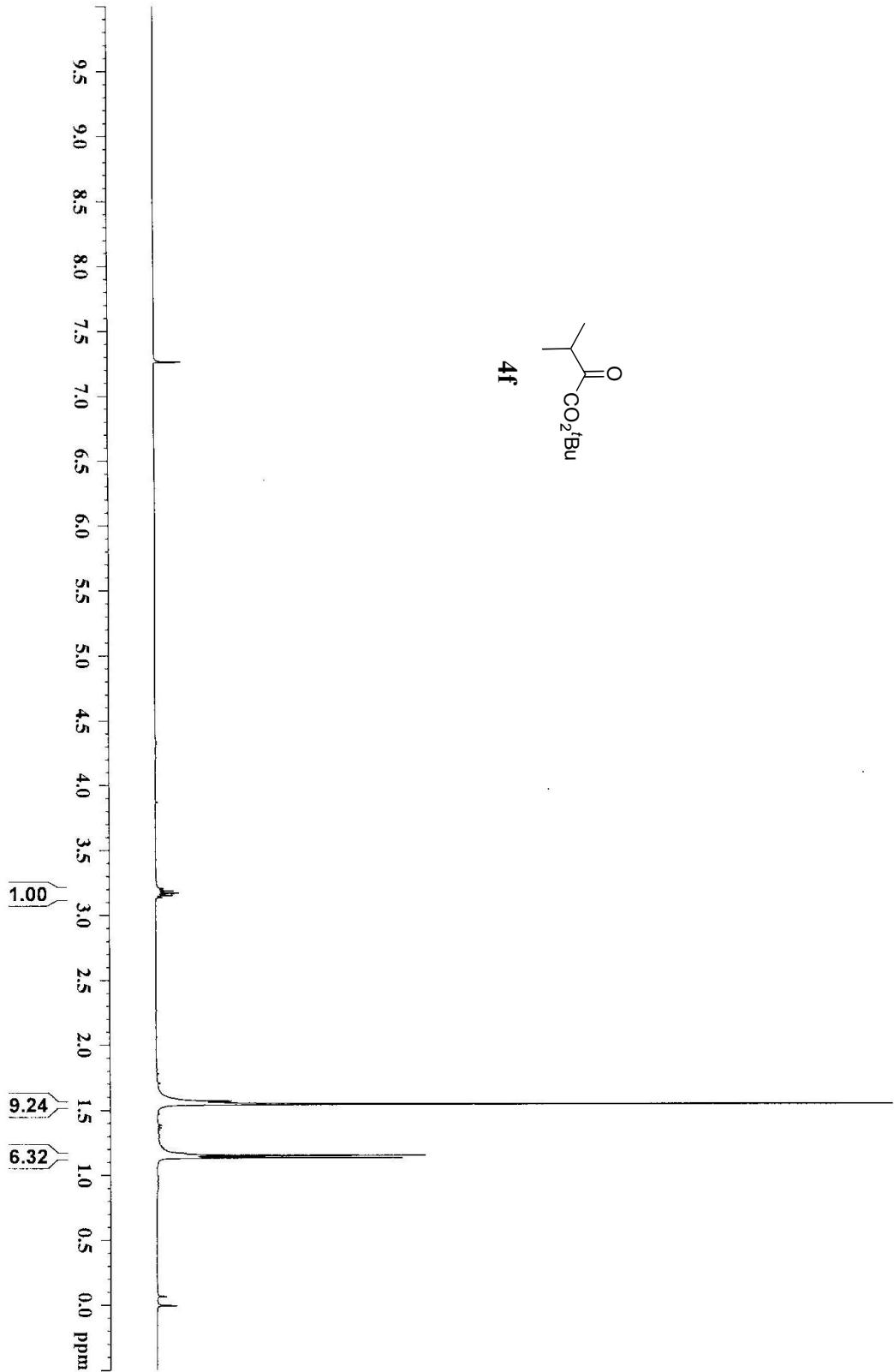


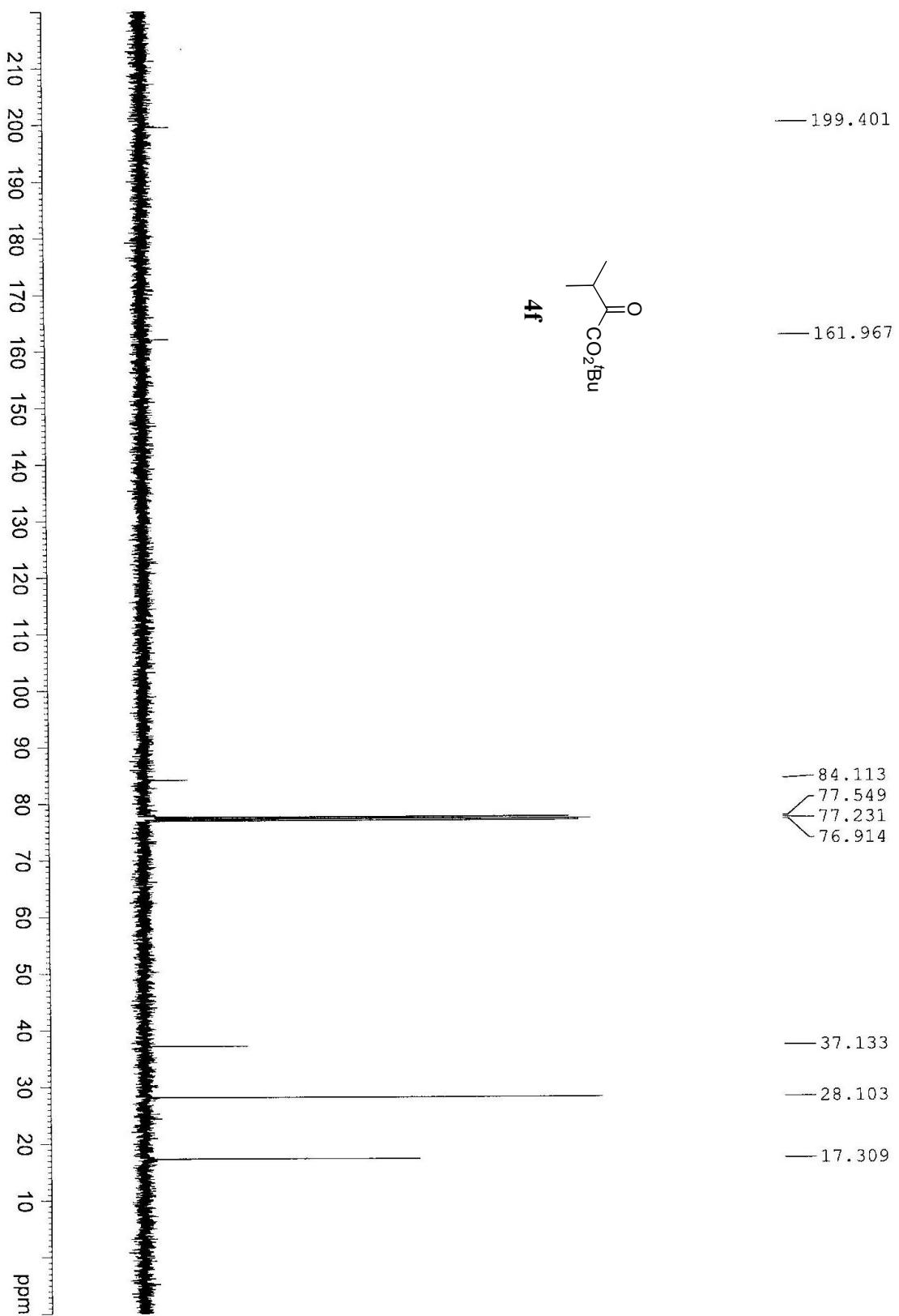


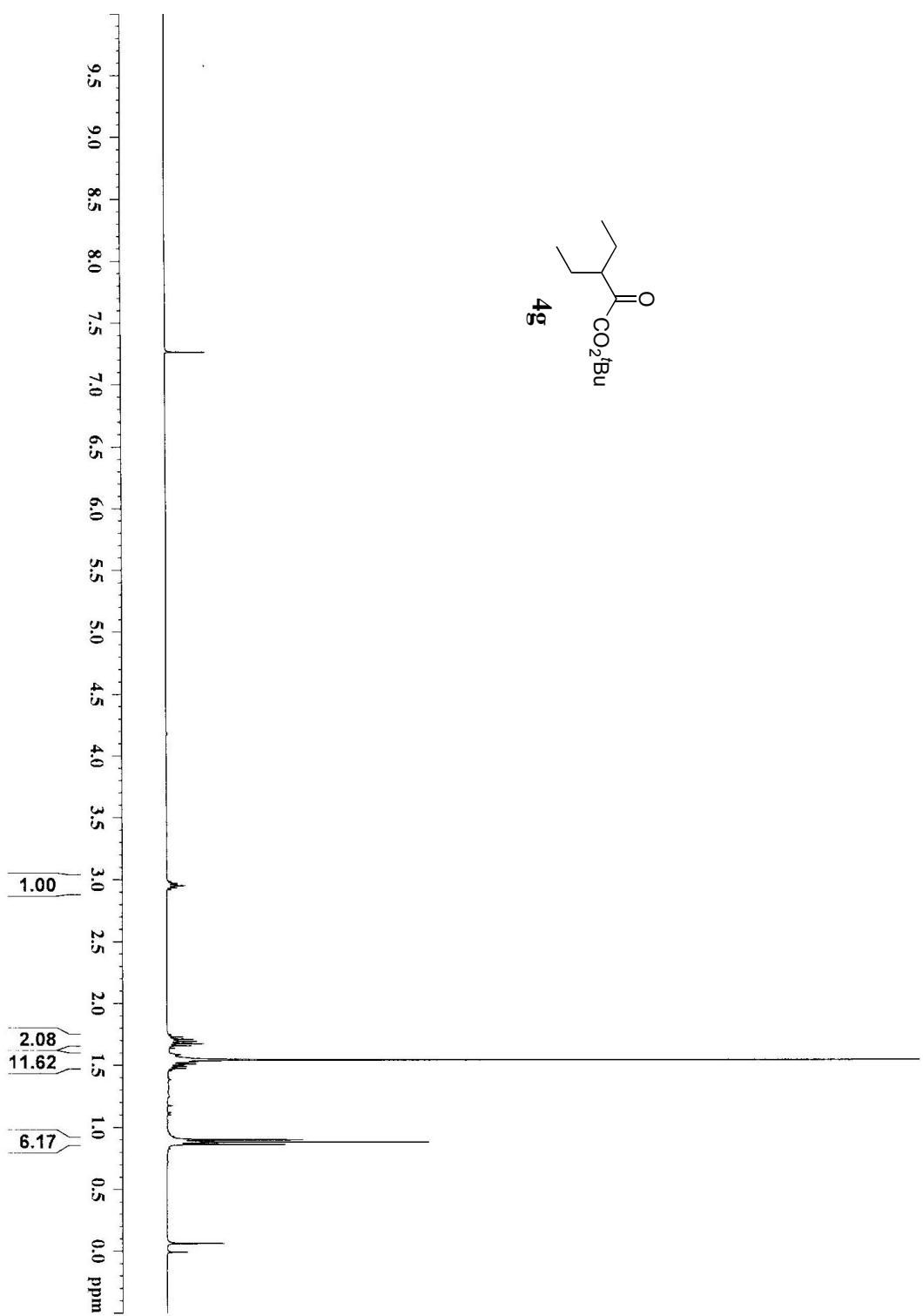


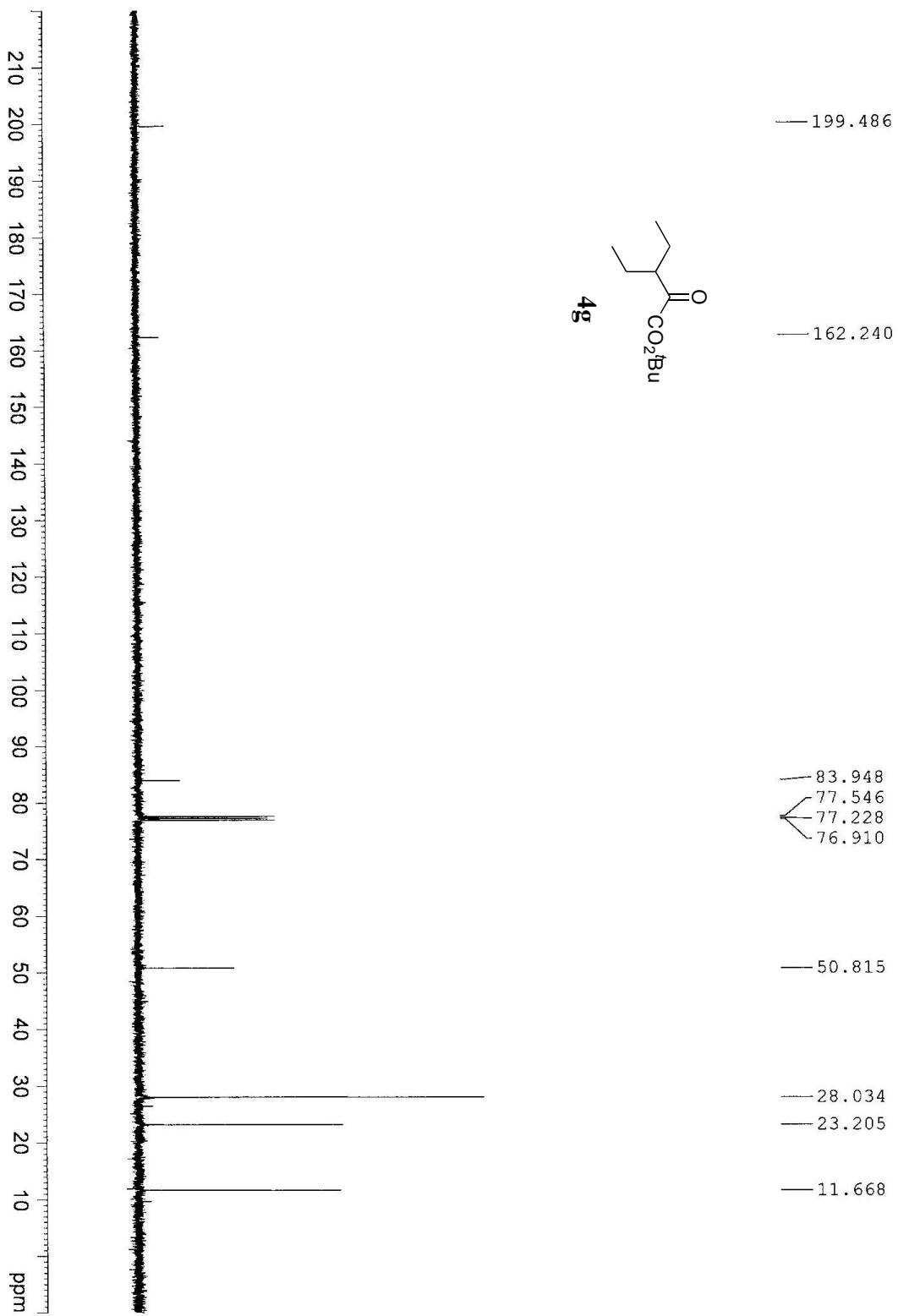


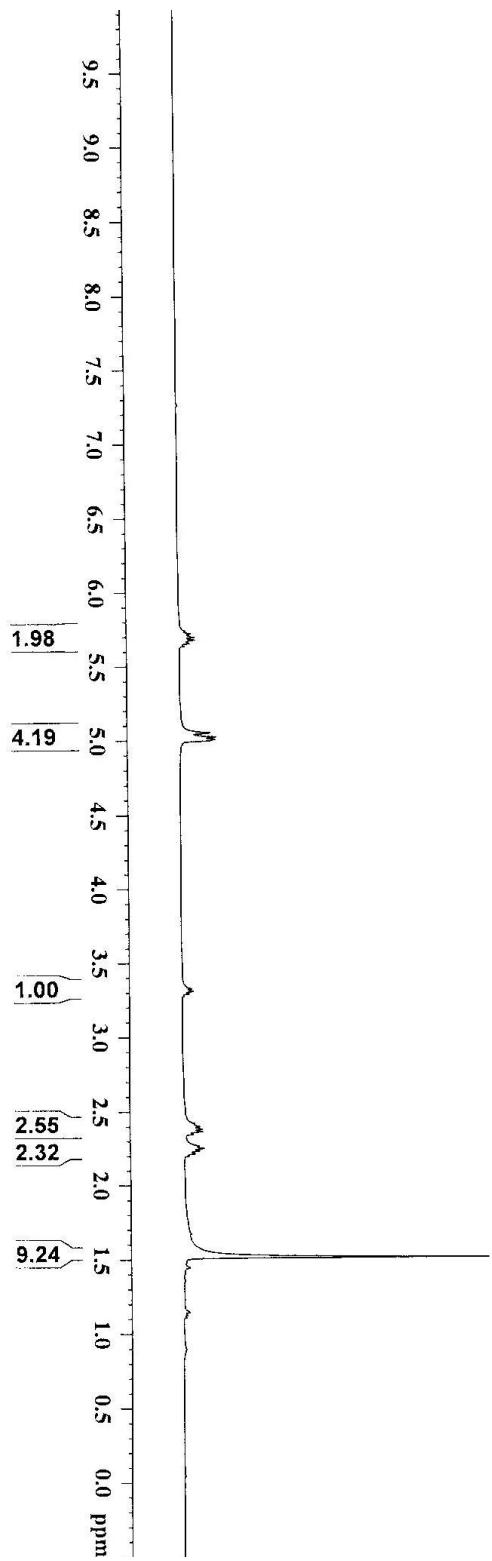
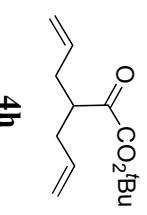


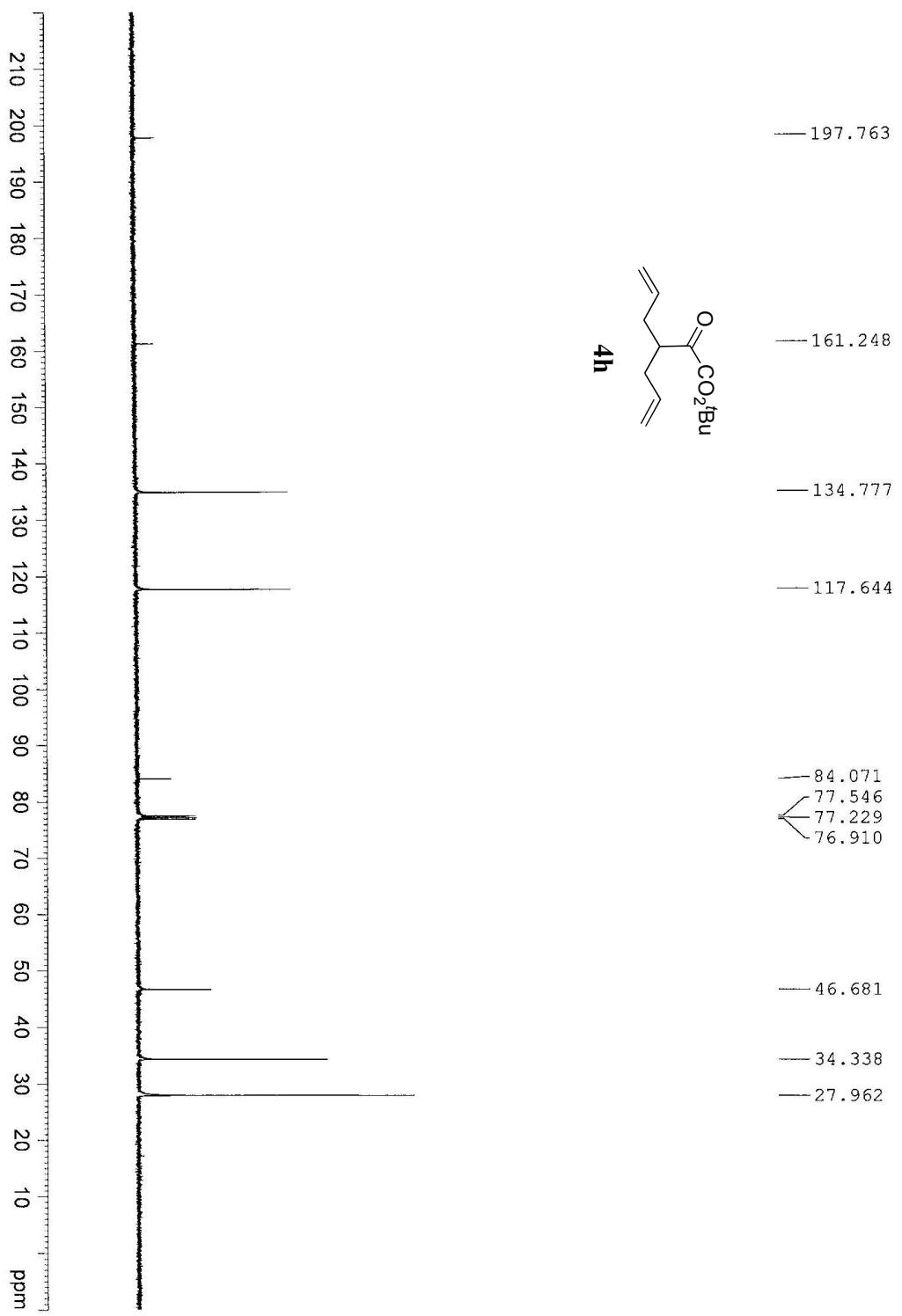


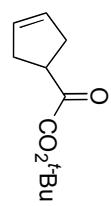




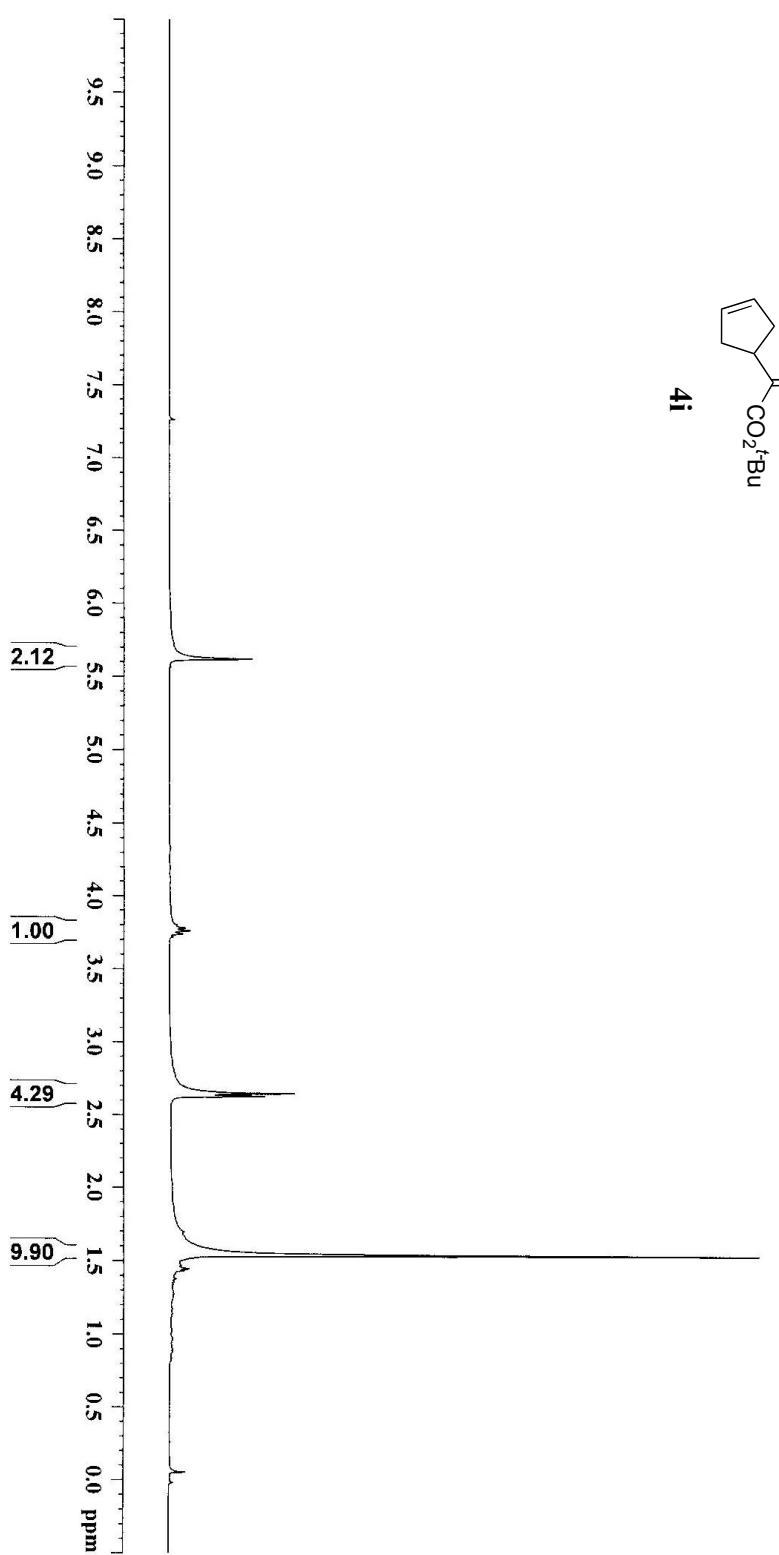


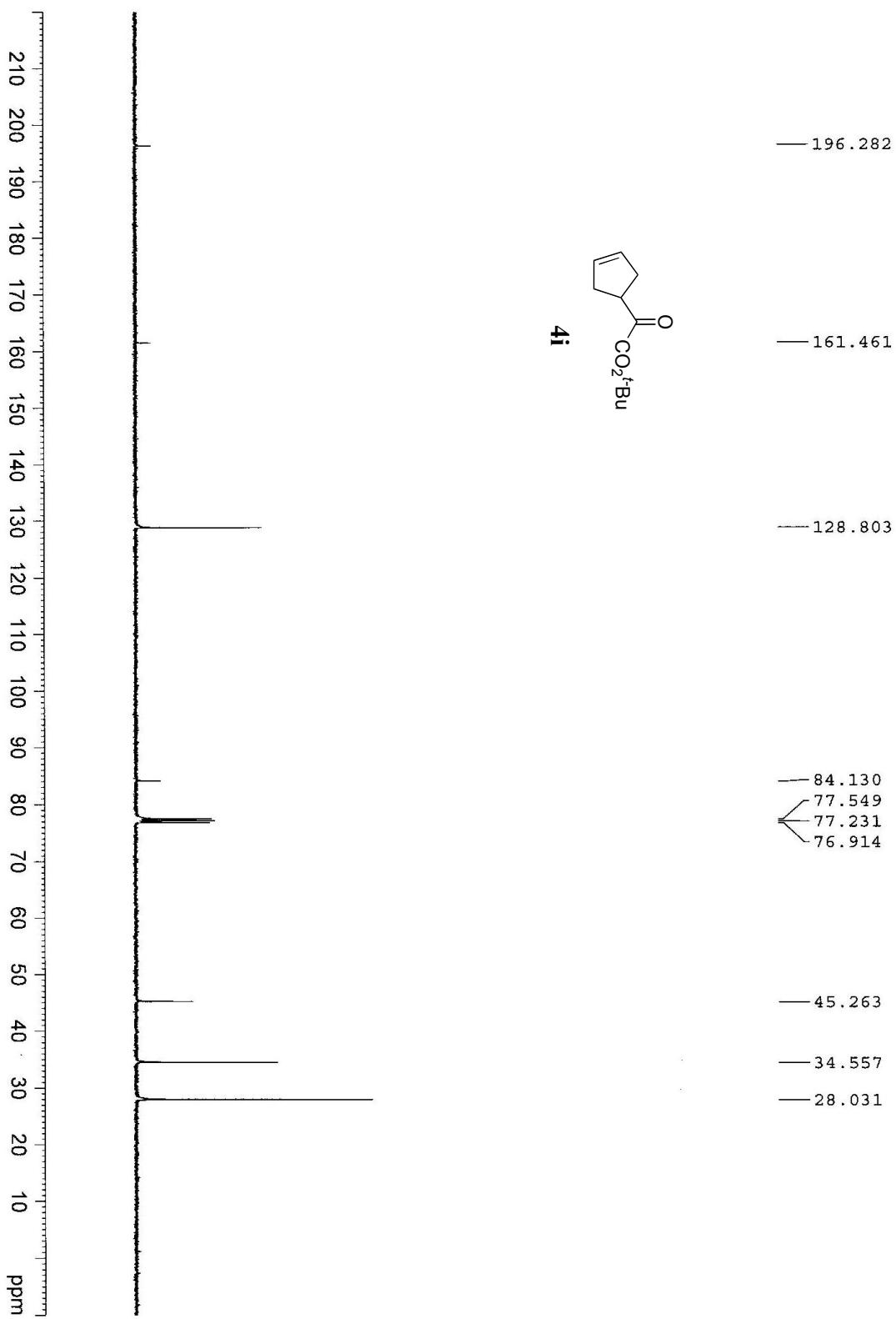


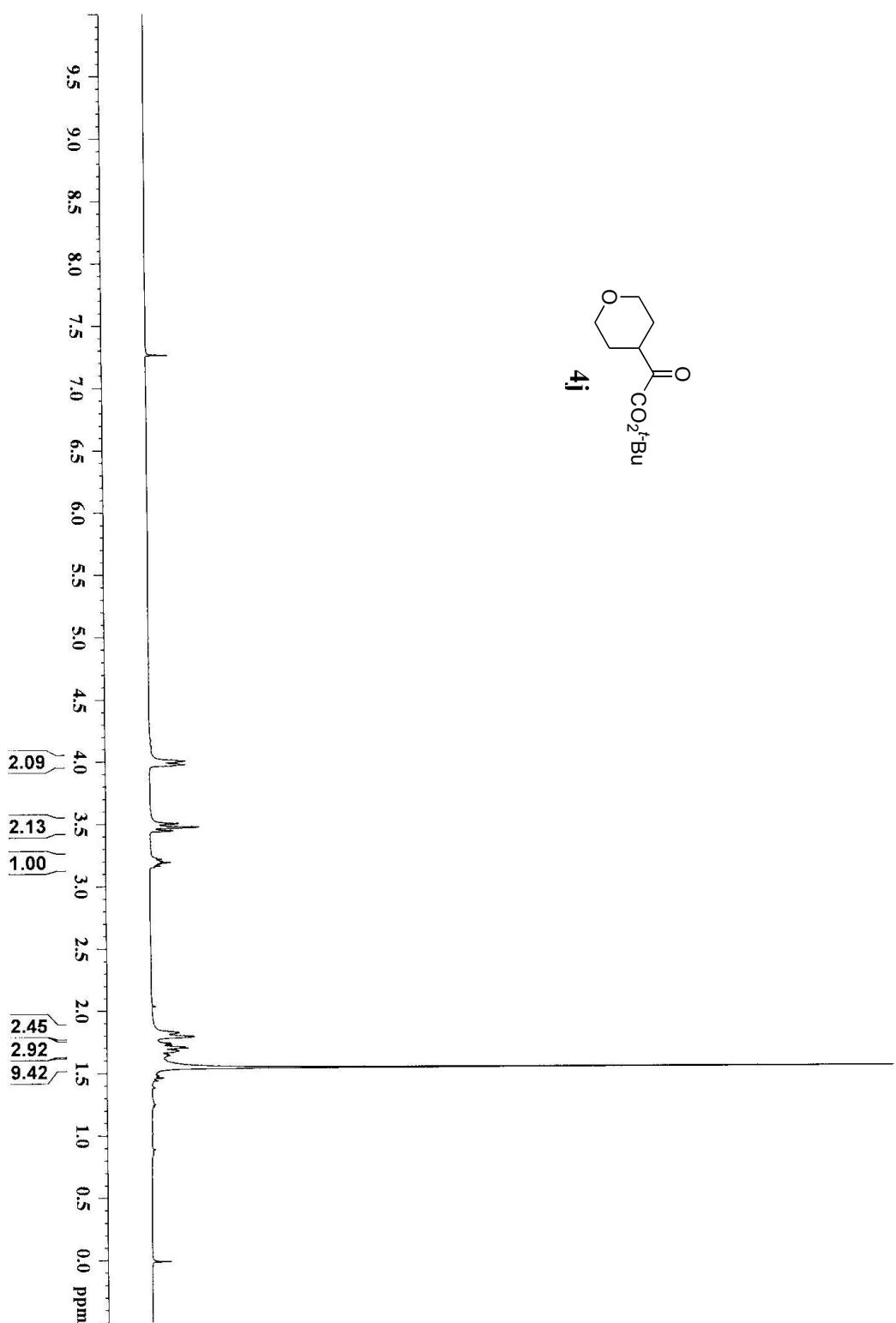
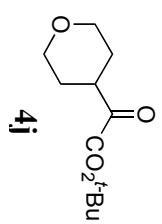


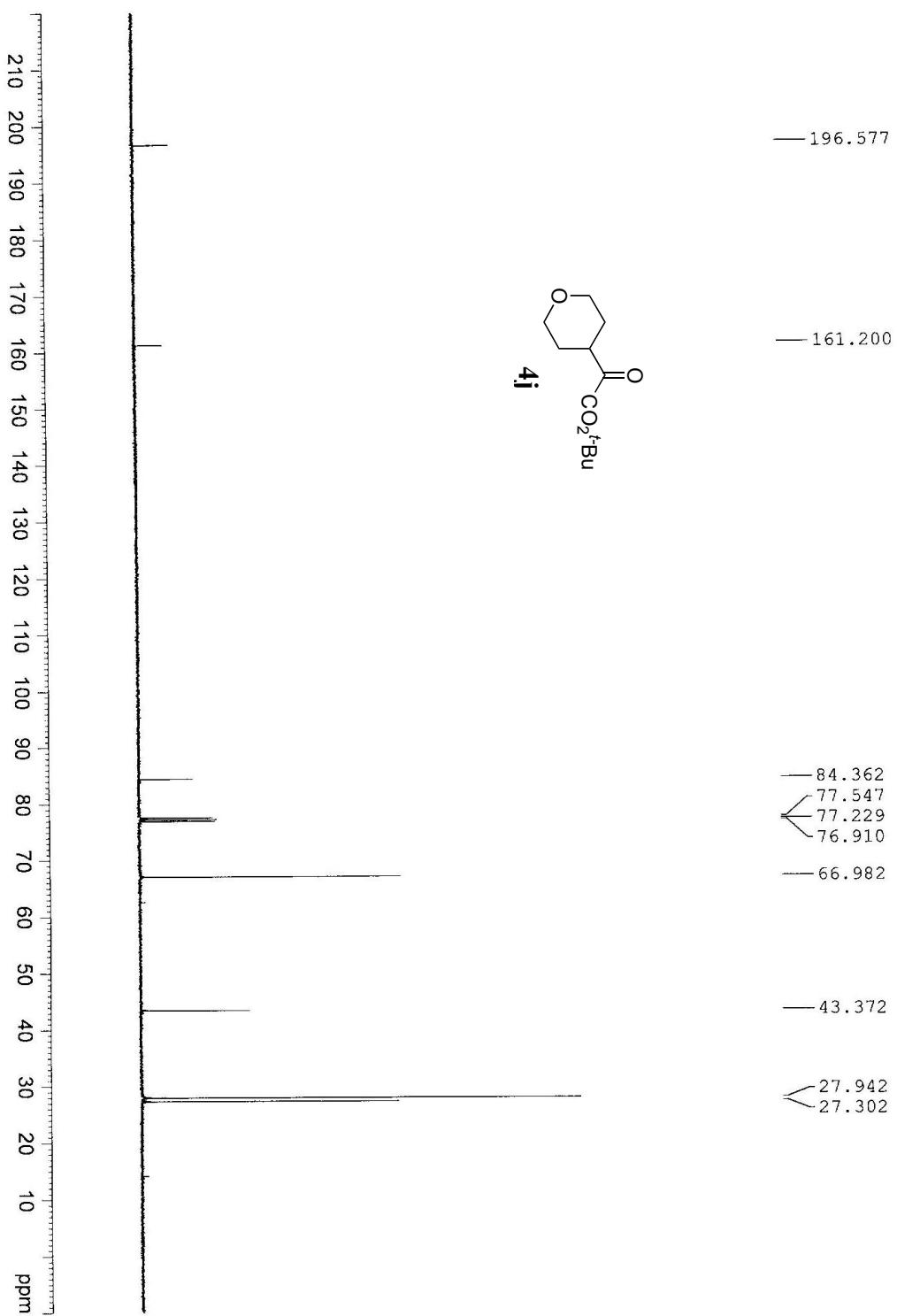


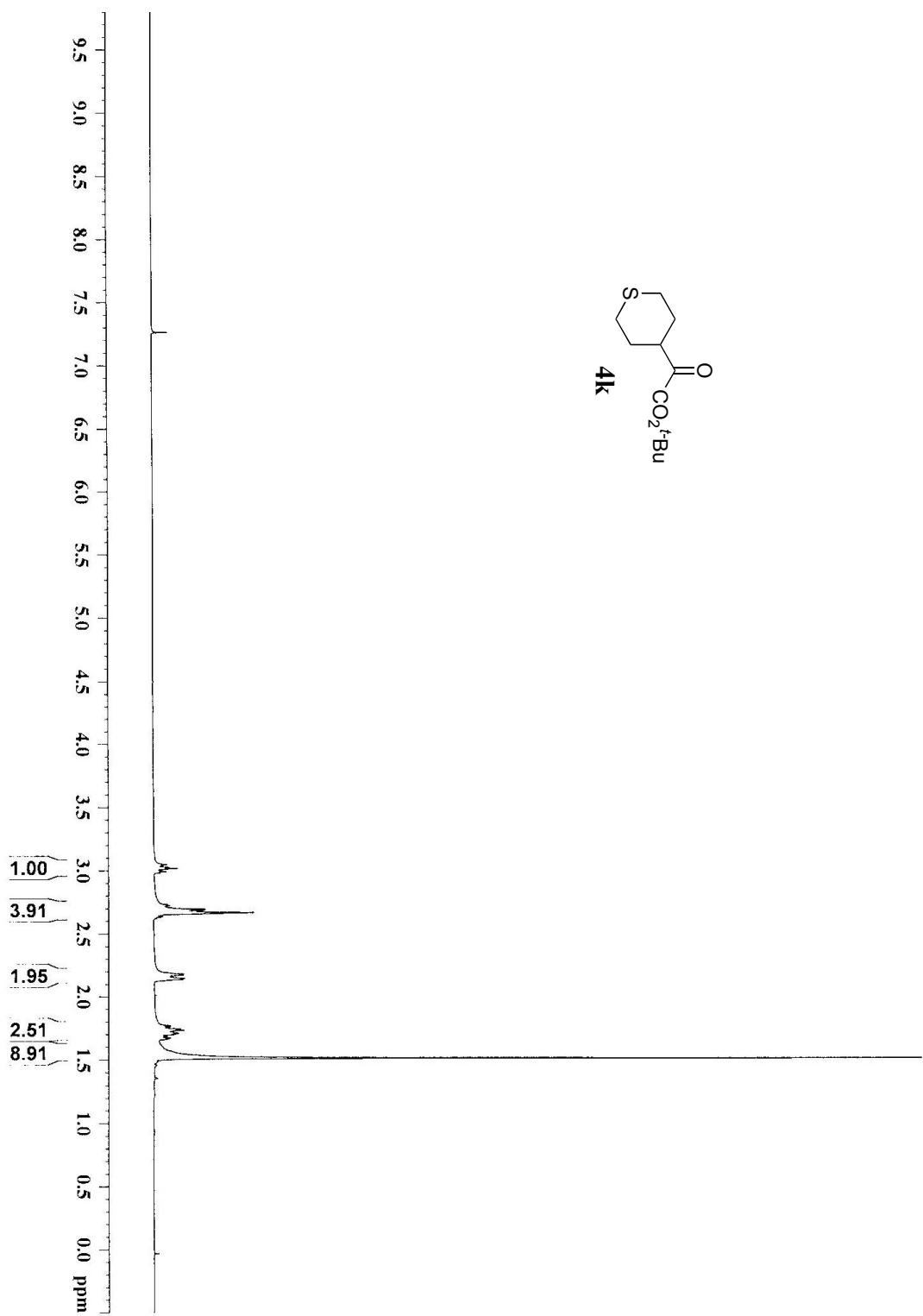
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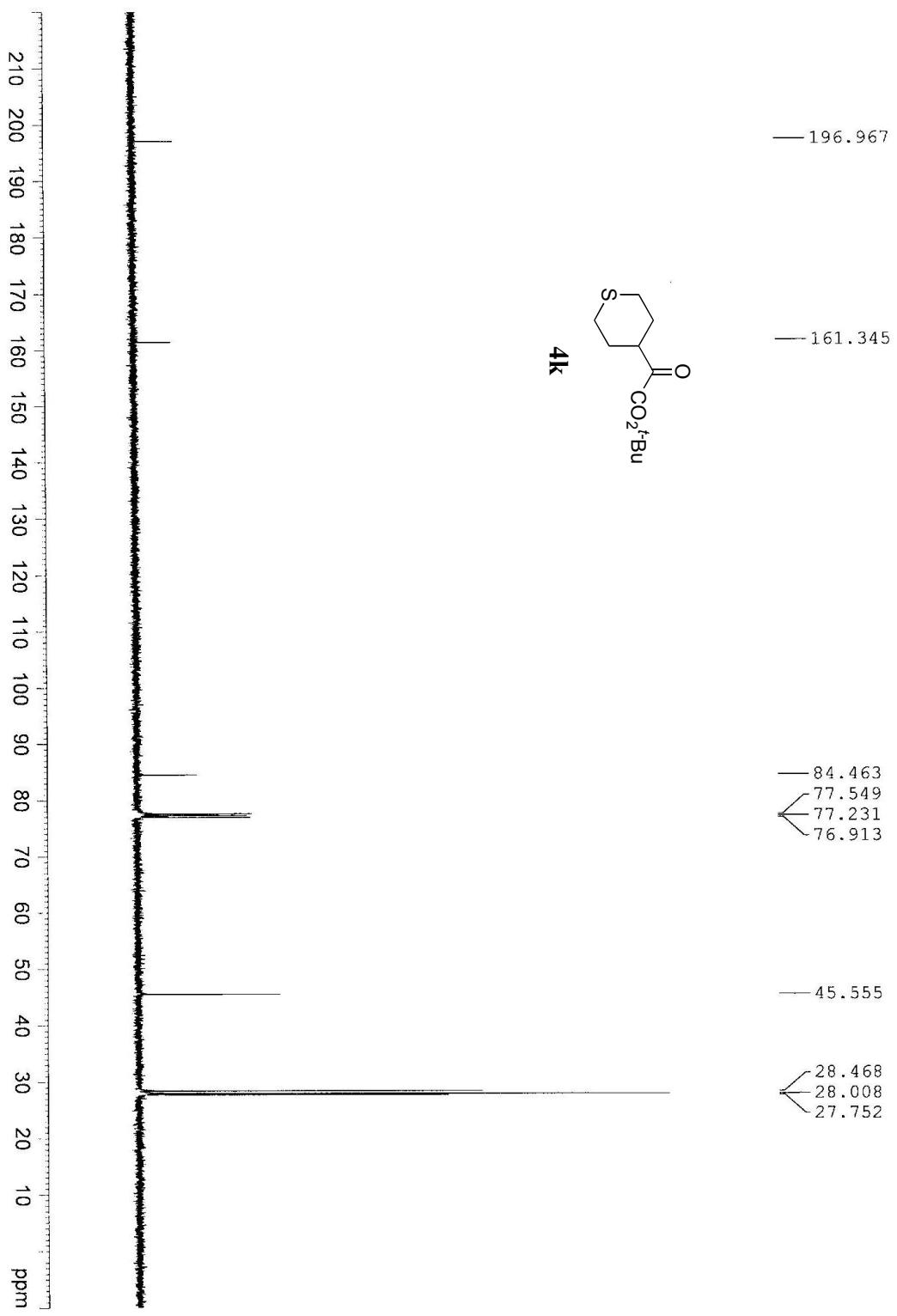


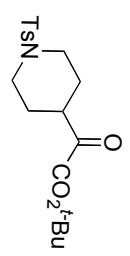




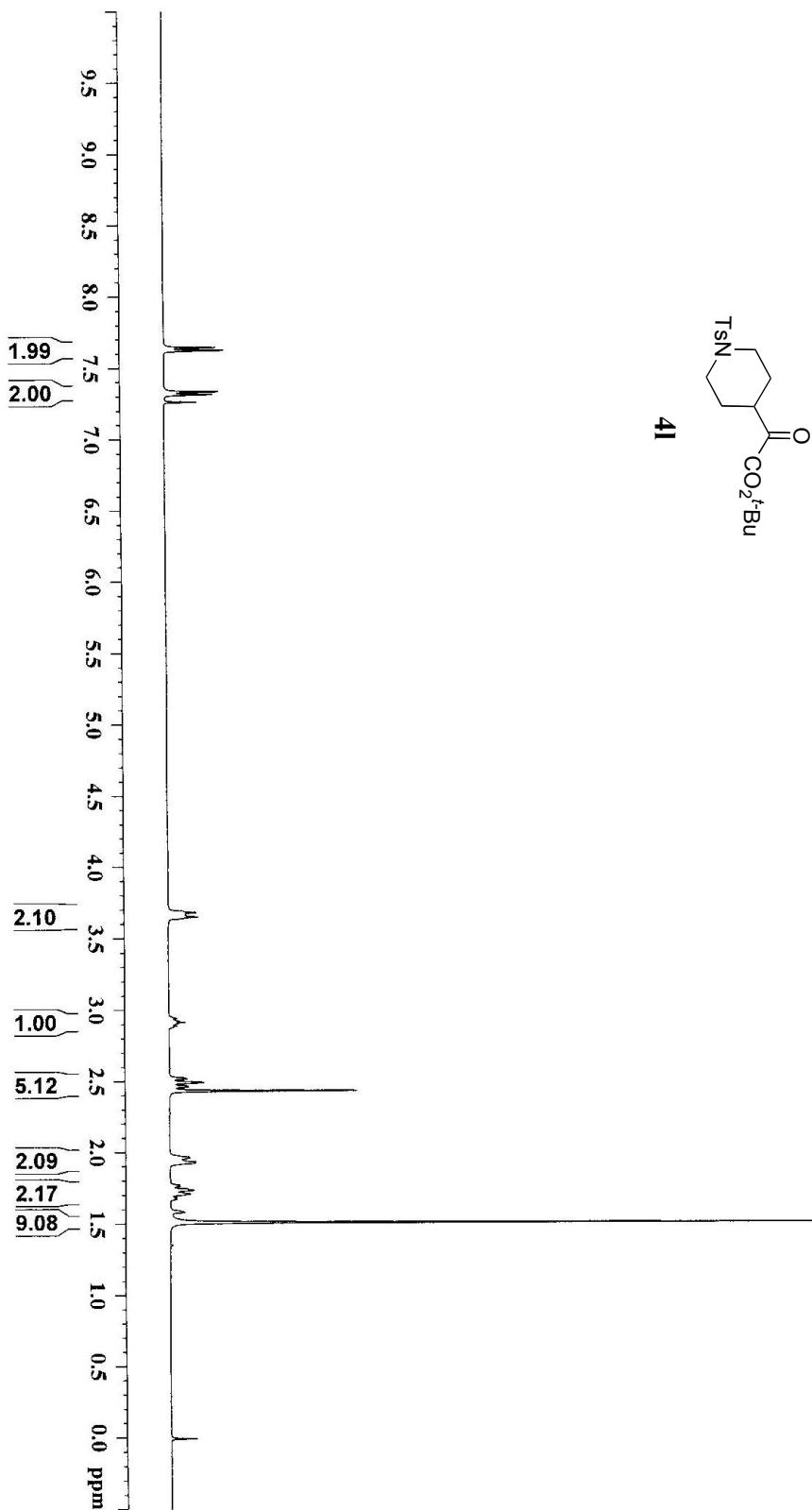








4l



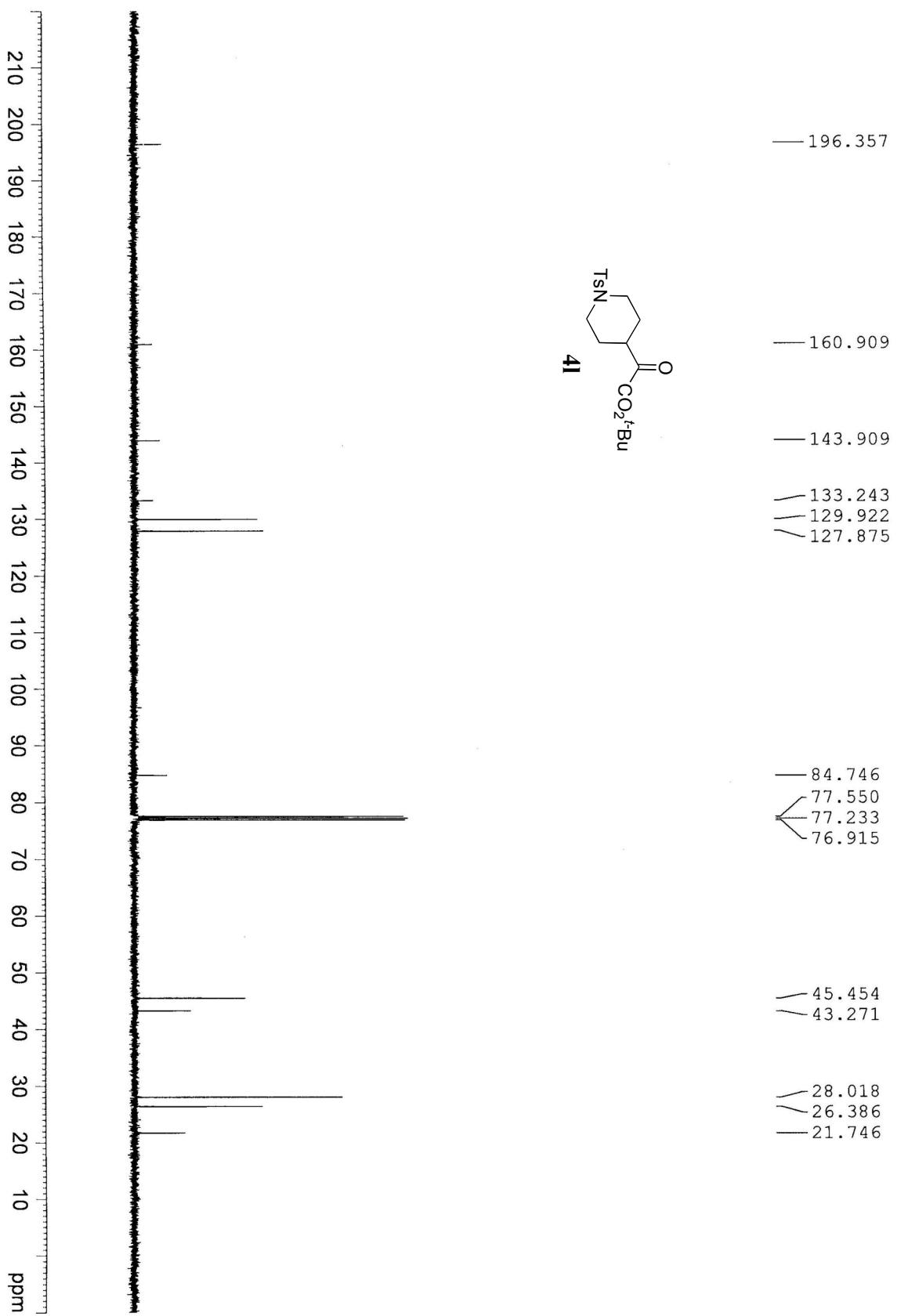


Table 2, entry 1

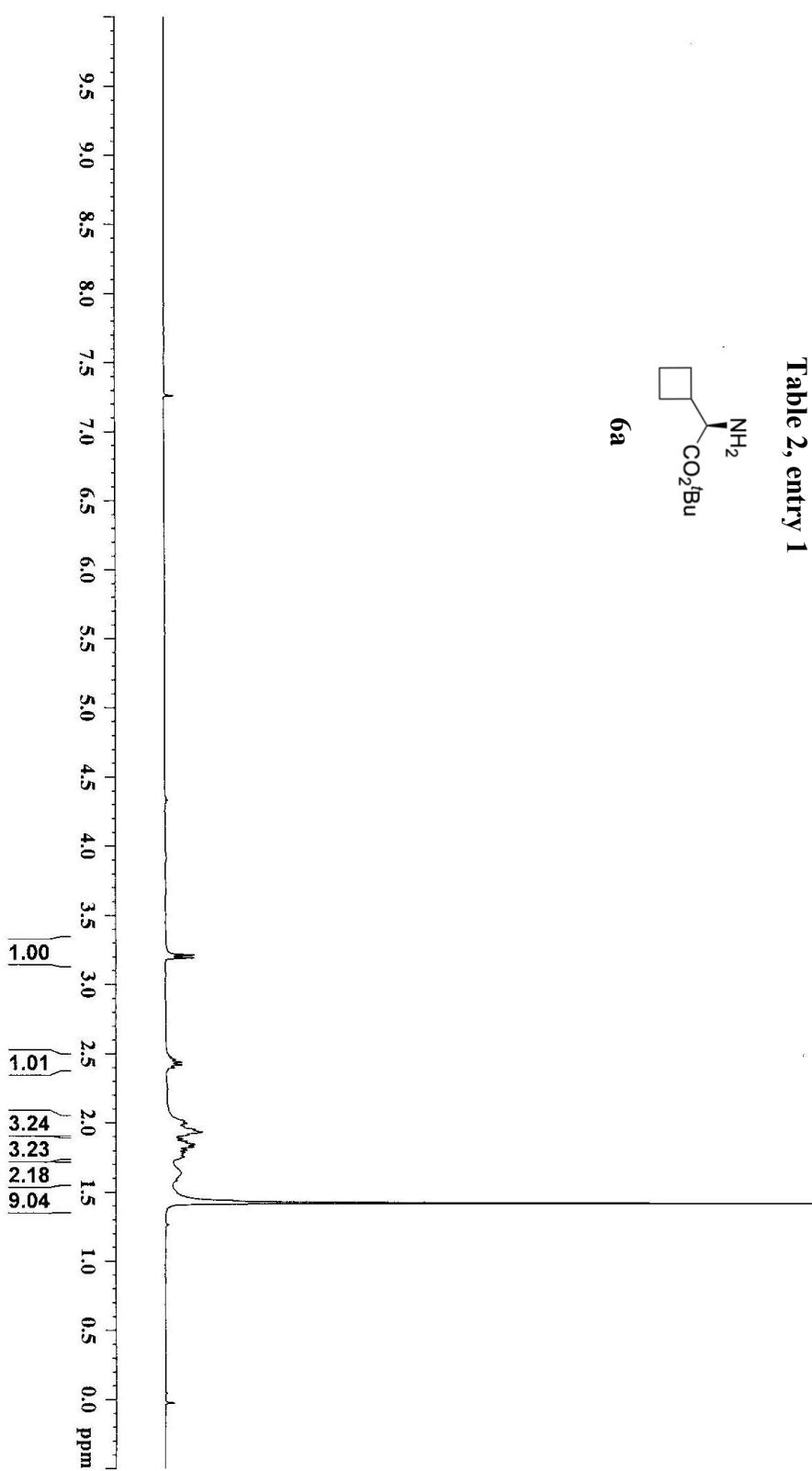
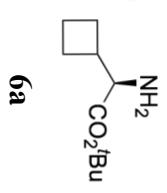


Table 2, entry 1

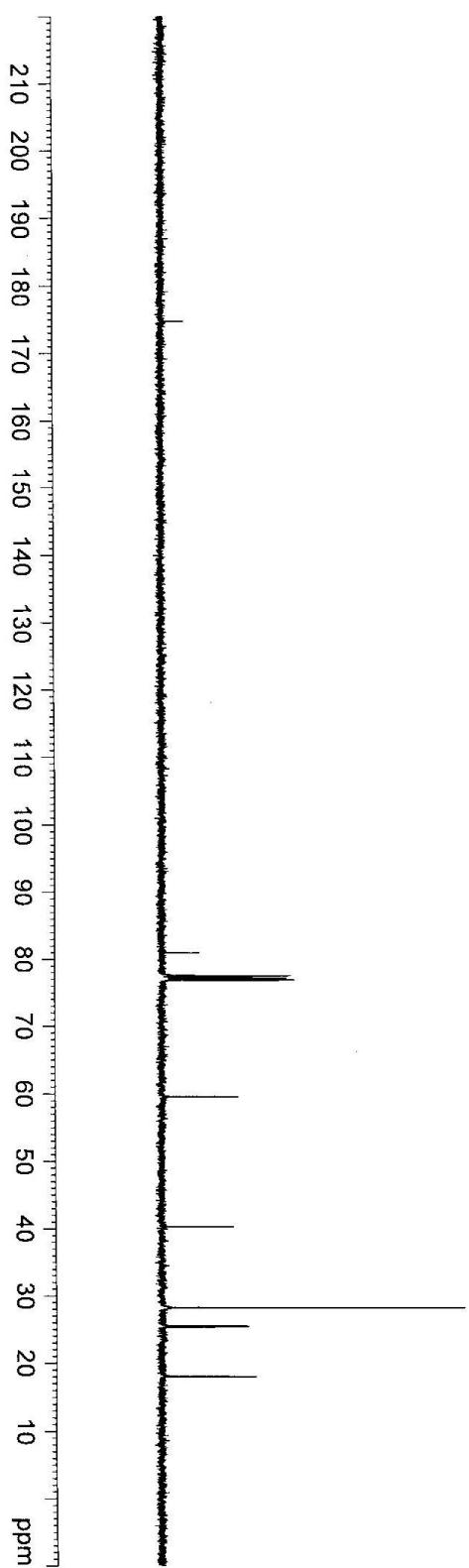
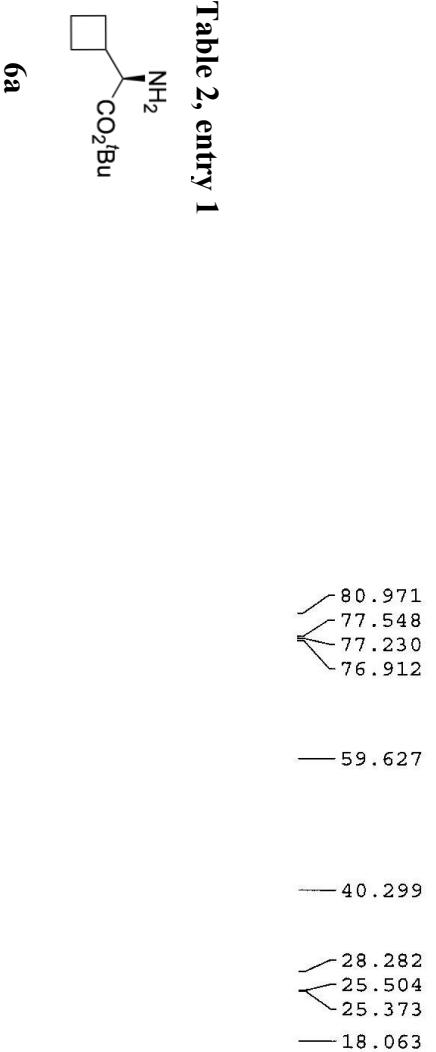
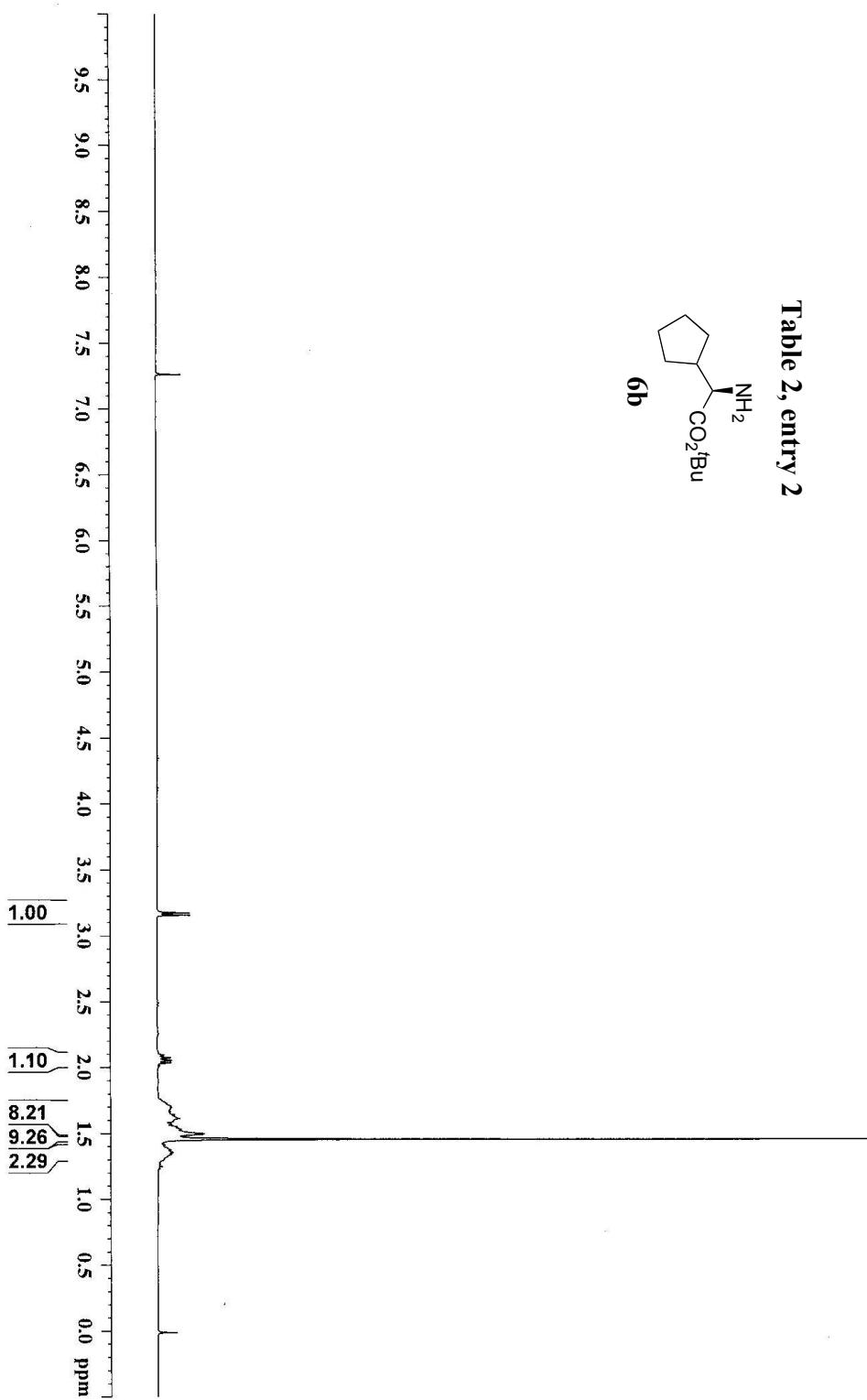
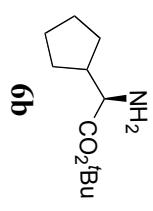


Table 2, entry 2



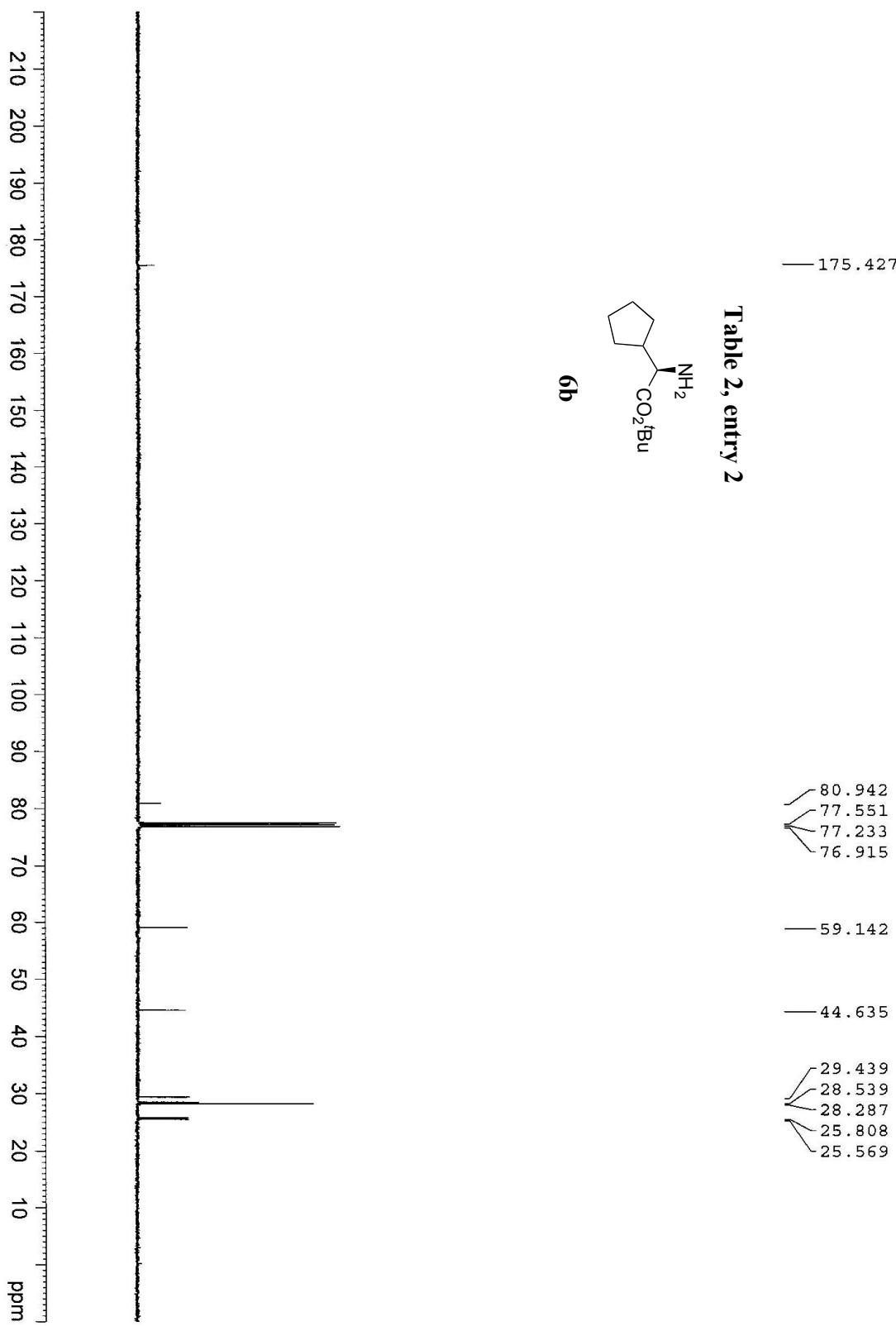
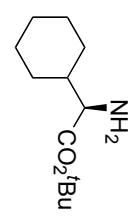


Table 2, entry 2

Table 2, entry 3



6c

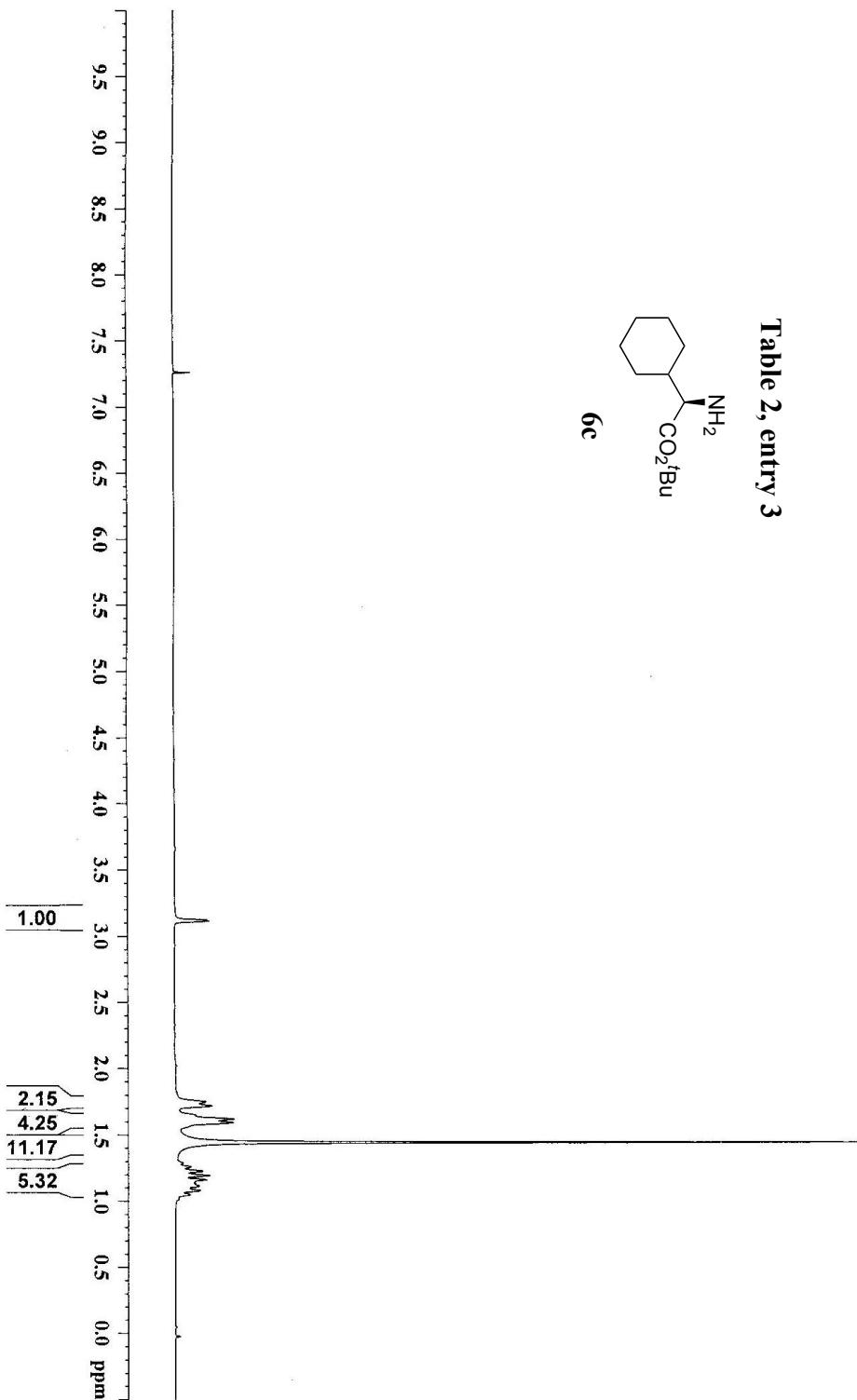


Table 2, entry 3

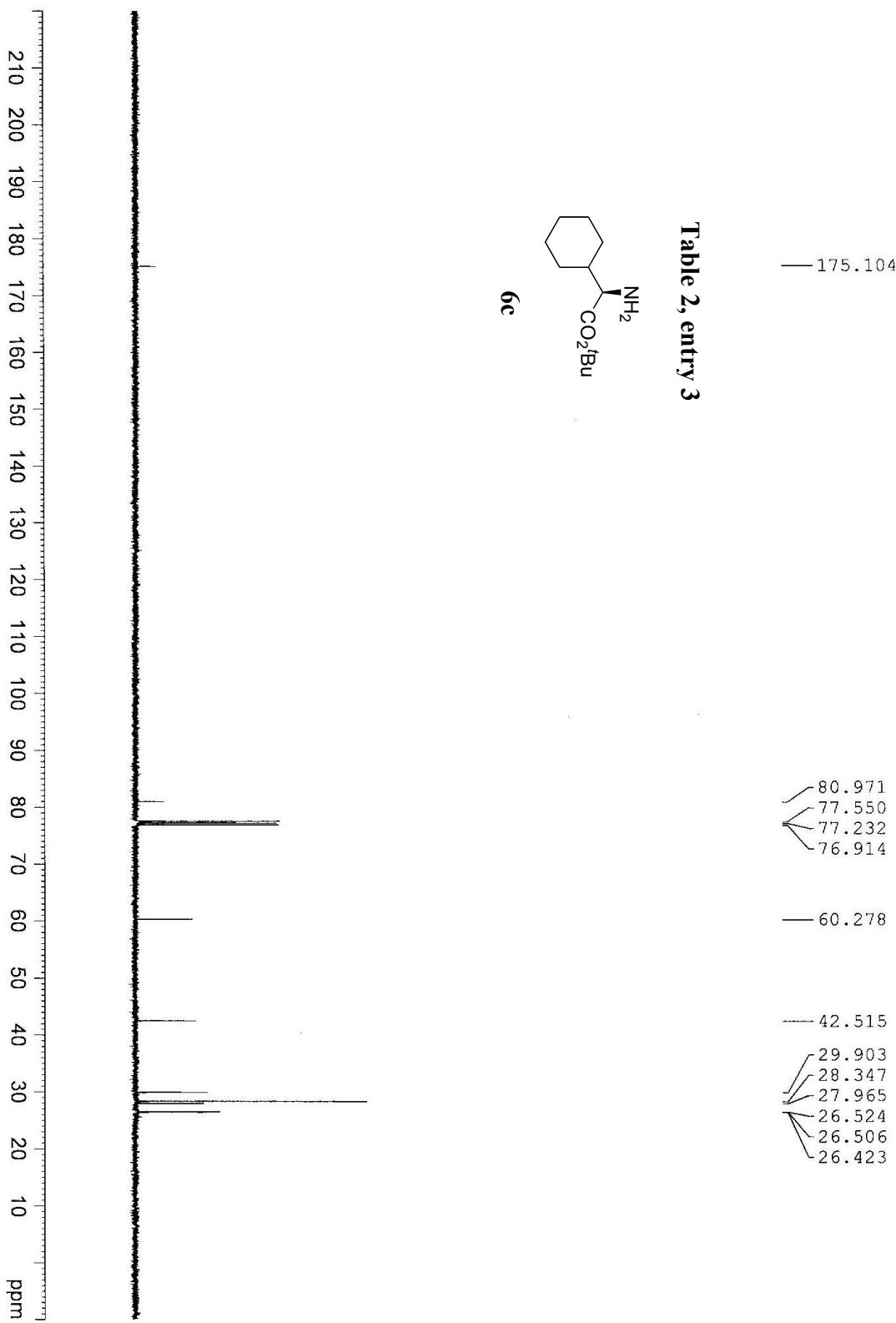


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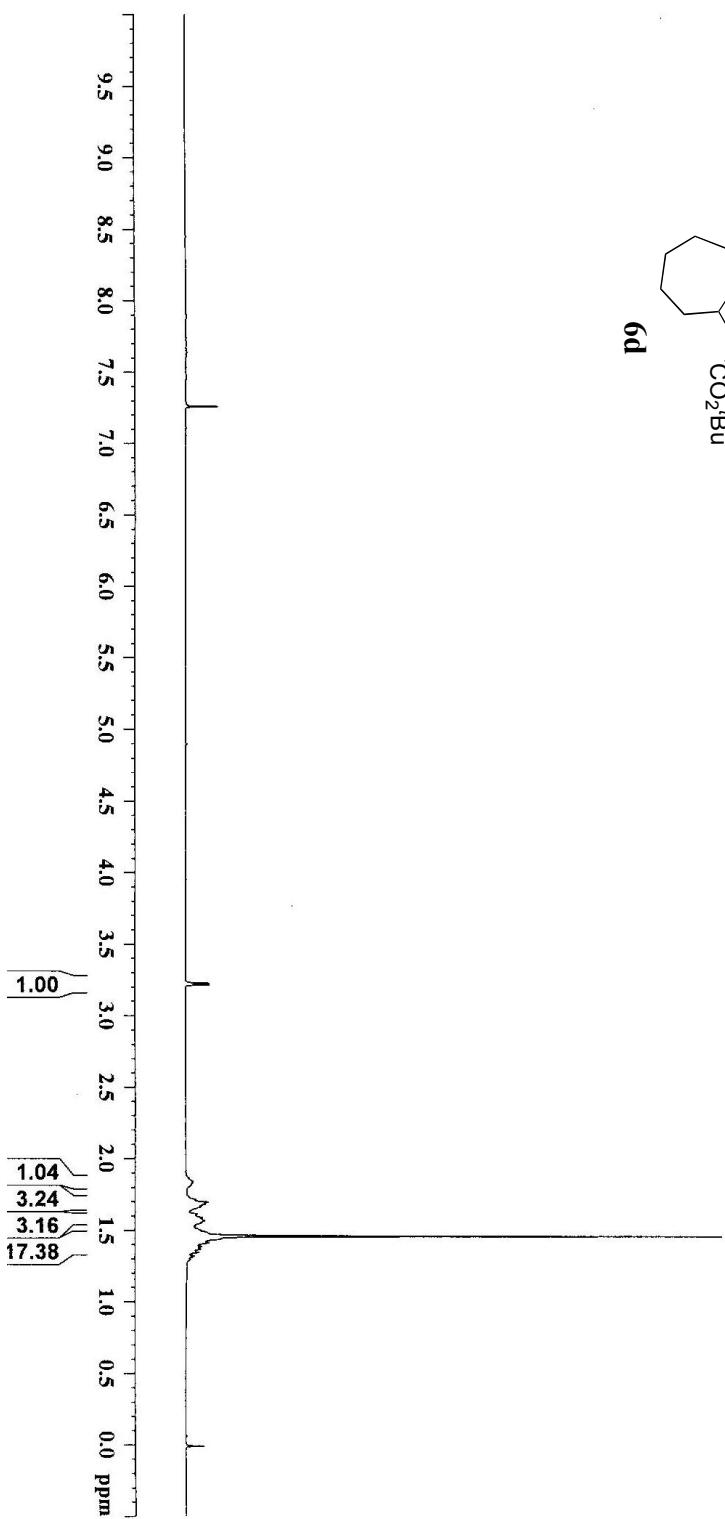
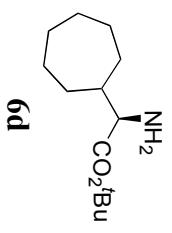


Table 2, entry 4

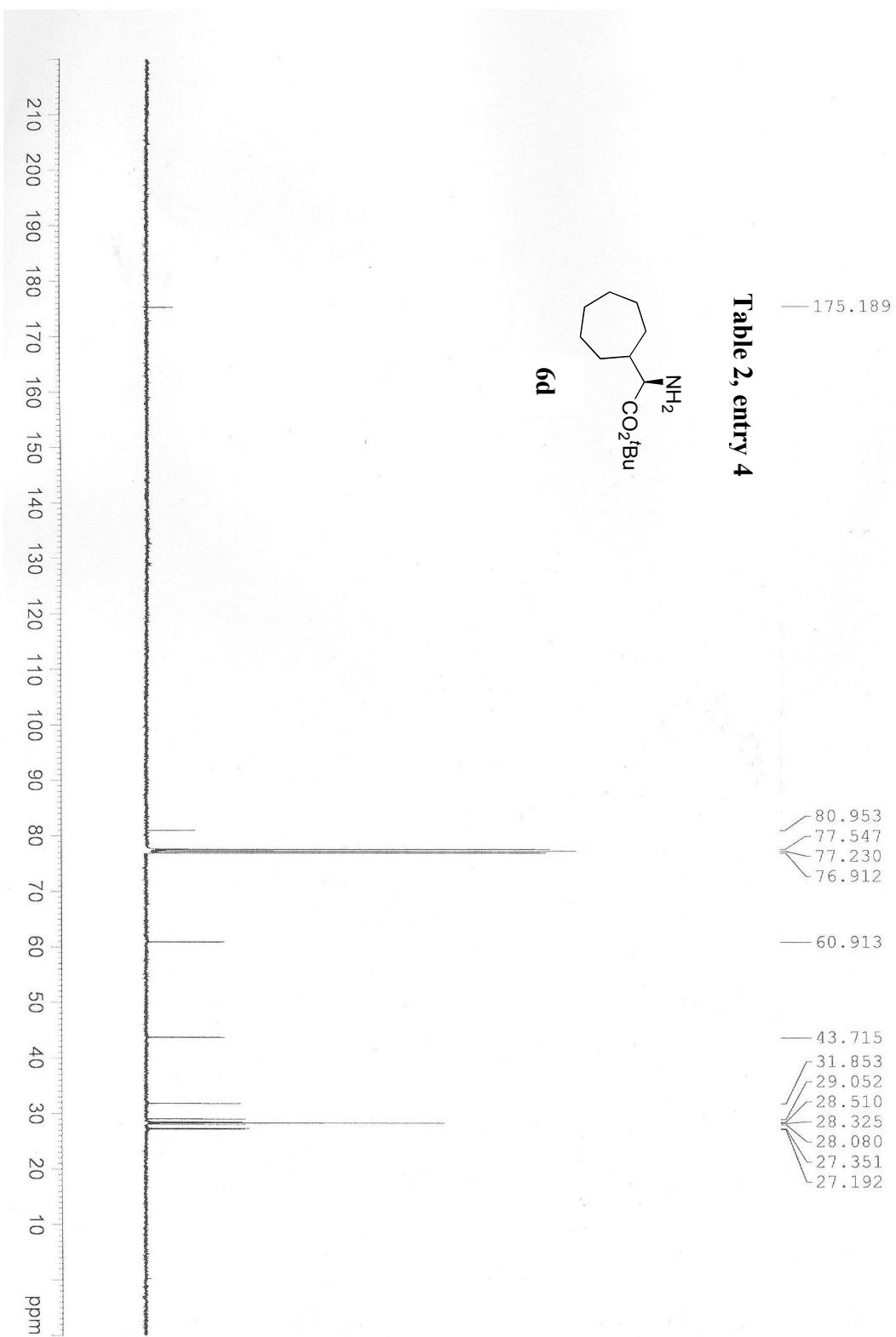
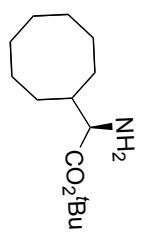
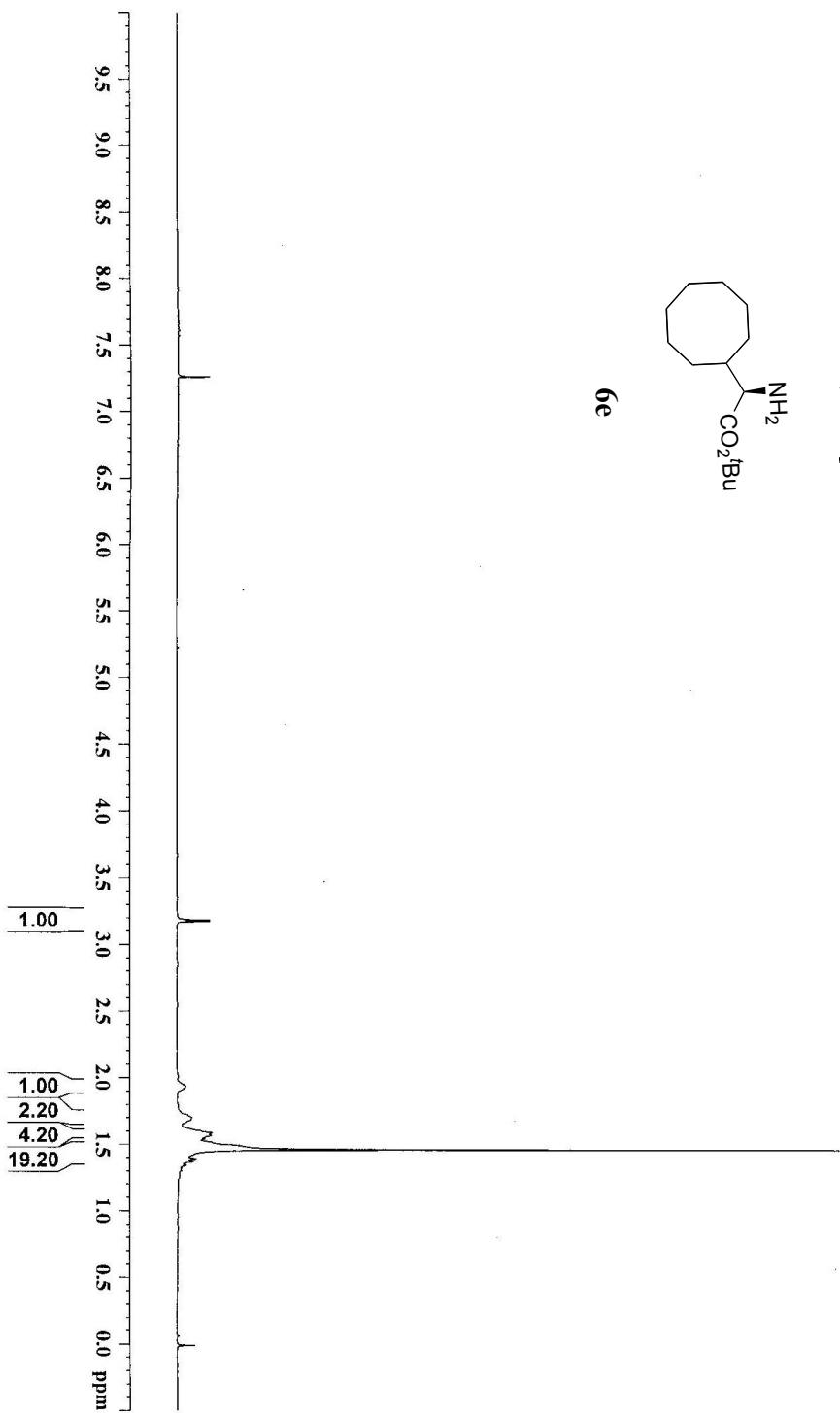


Table 2, entry 5



6e



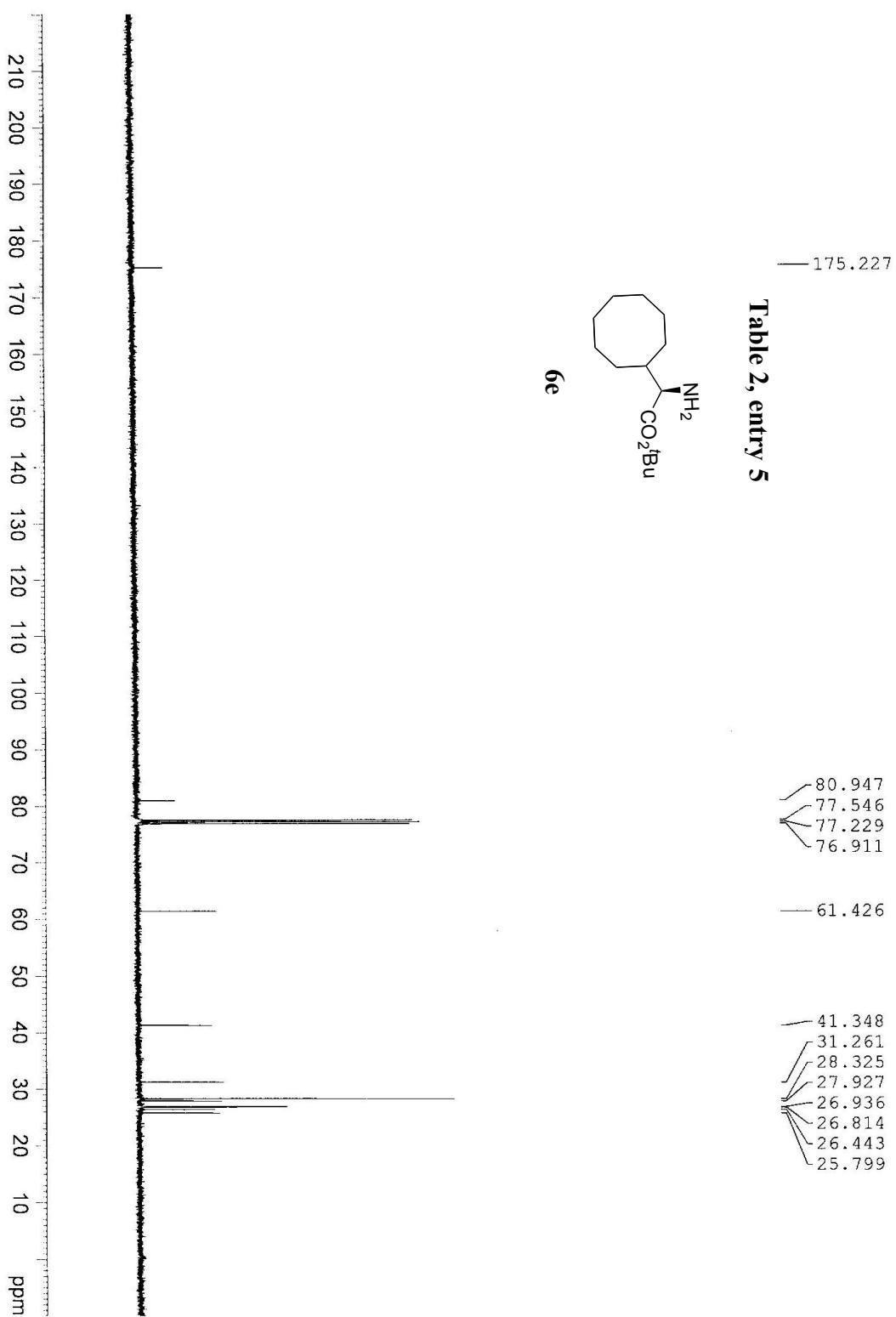
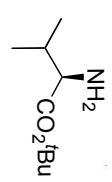


Table 2, entry 5

Table 2, entry 6



6f

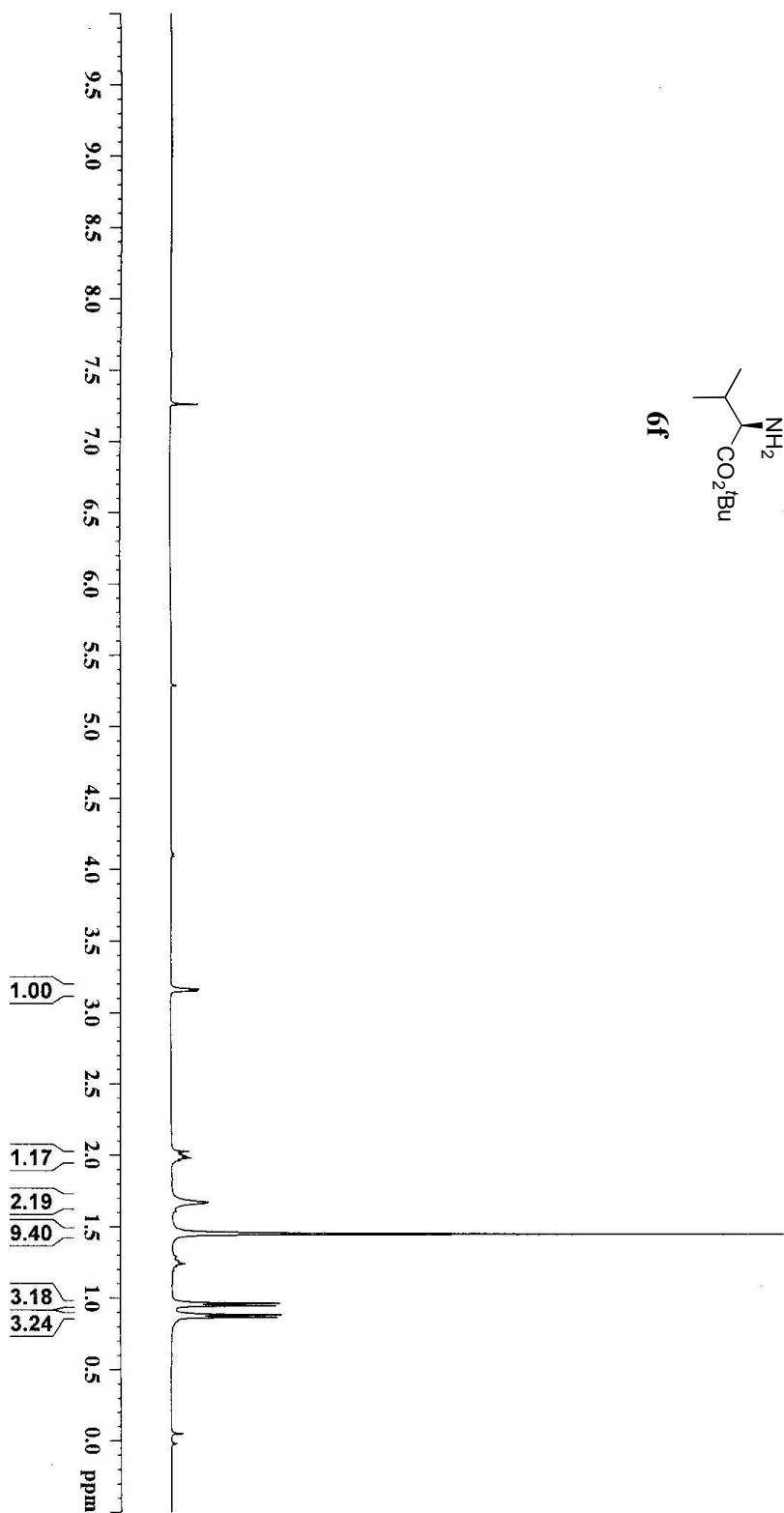


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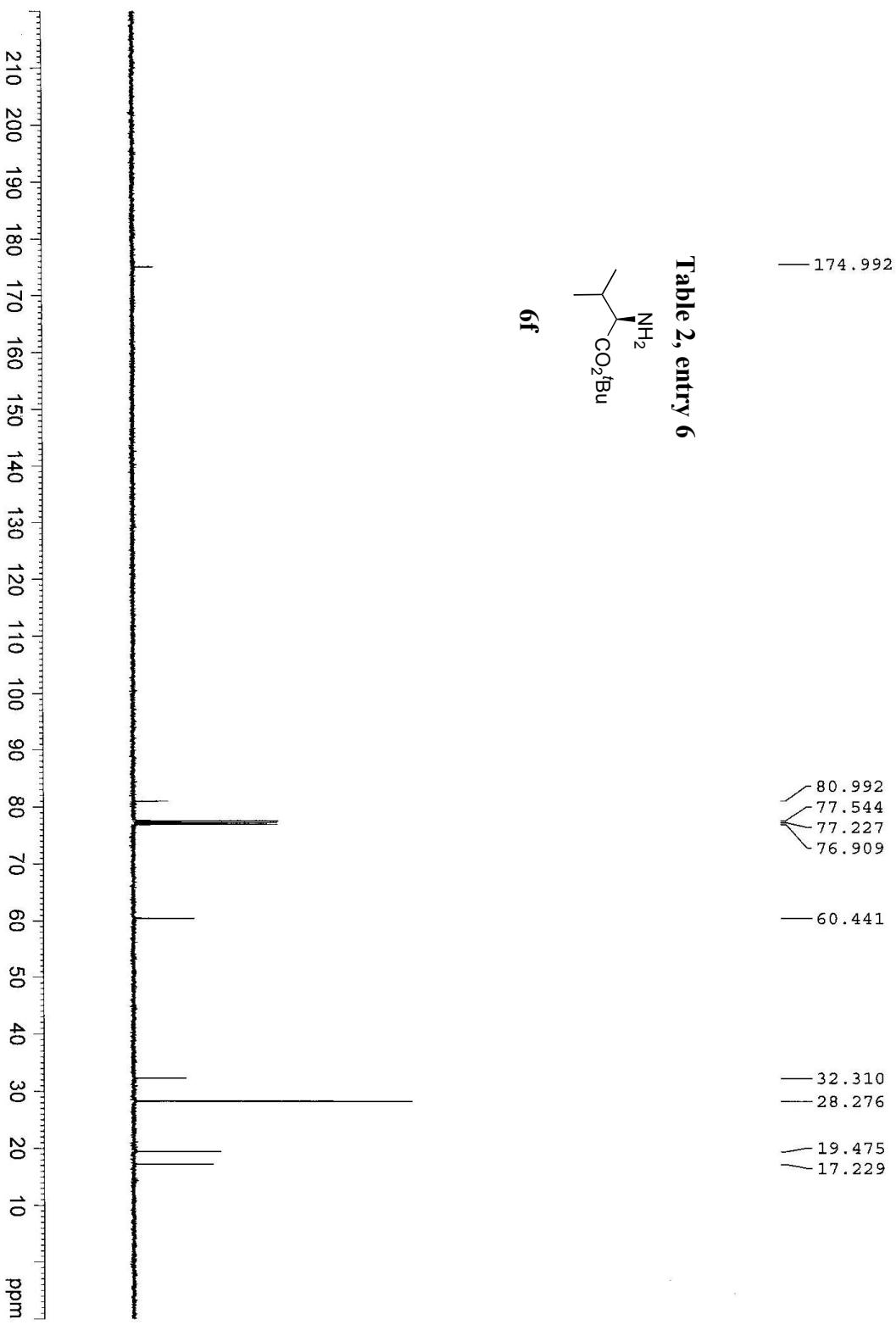
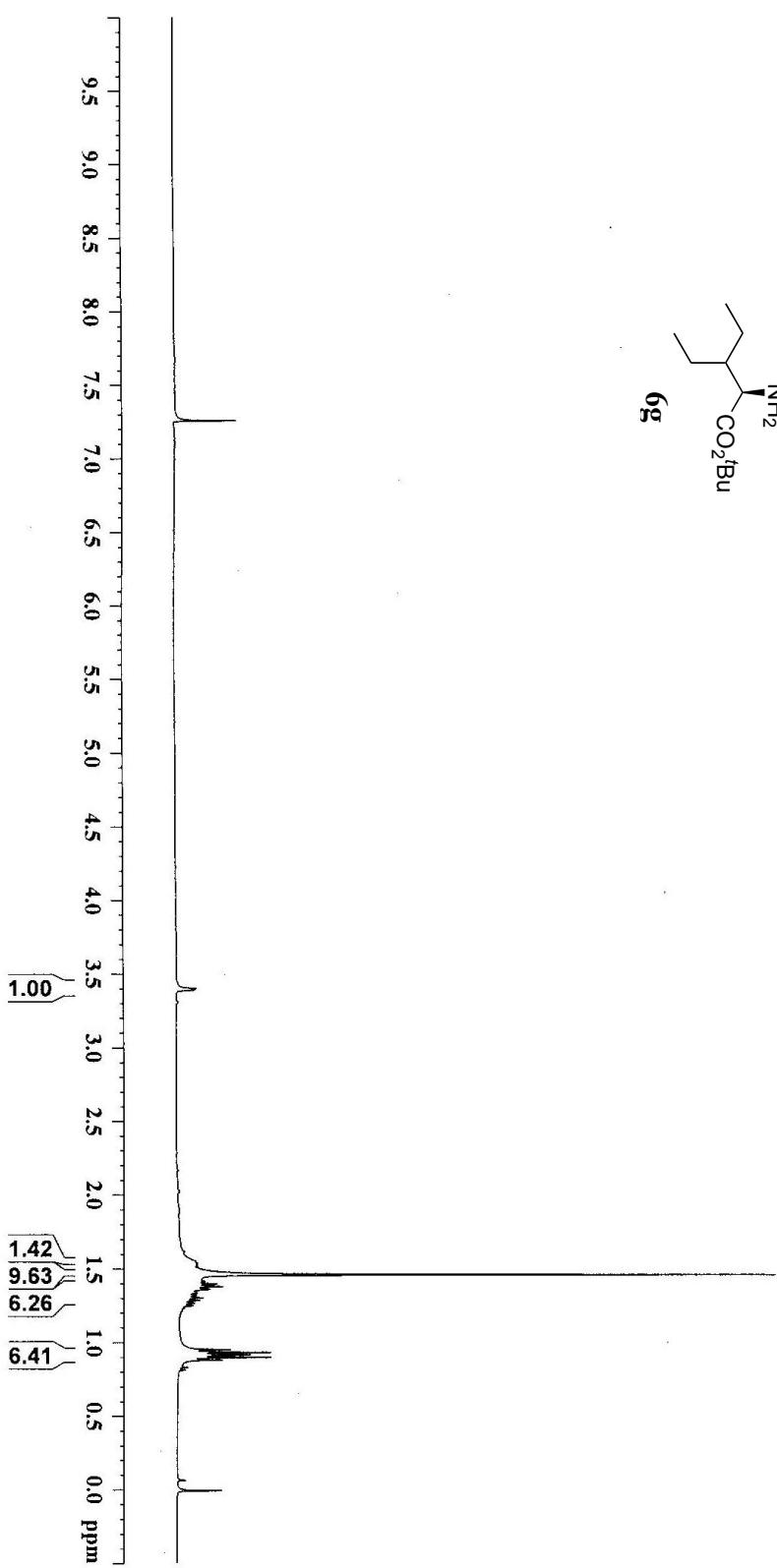
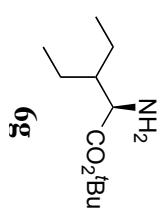
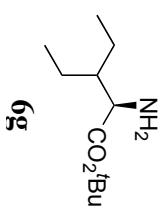


Table 2, entry 7



— 175.741

Table 2, entry 7



80.893
77.551
77.232
76.915

— 56.427

— 45.525

— 28.241
VV 23.064
VV 22.057
VV 12.107
VV 12.007

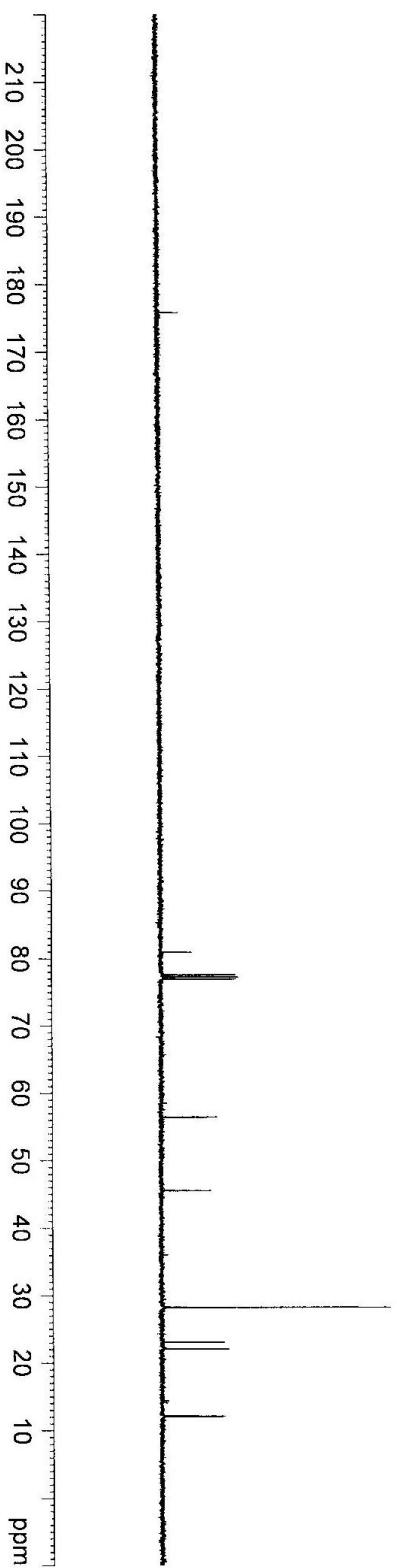
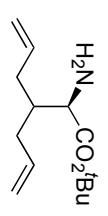
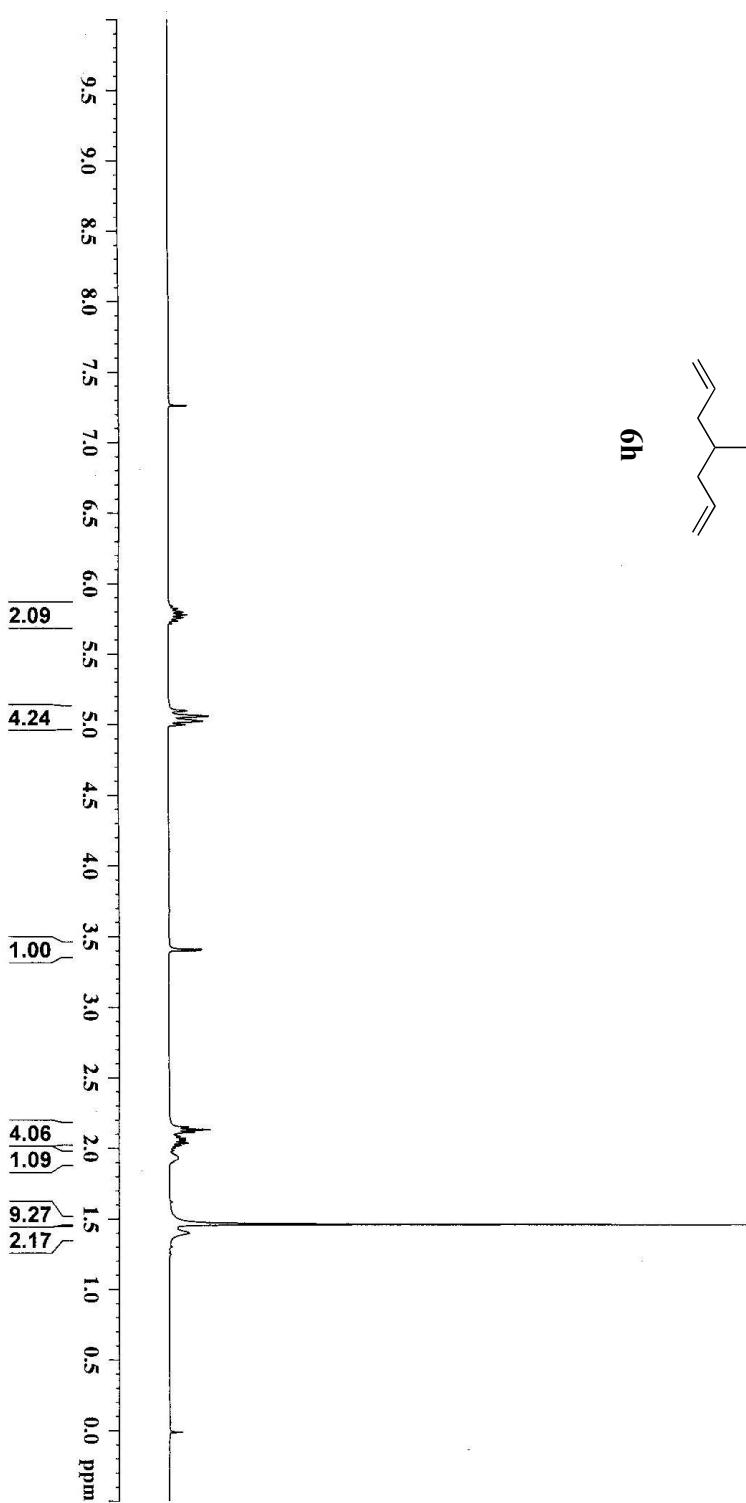


Table 2, entry 8



6h



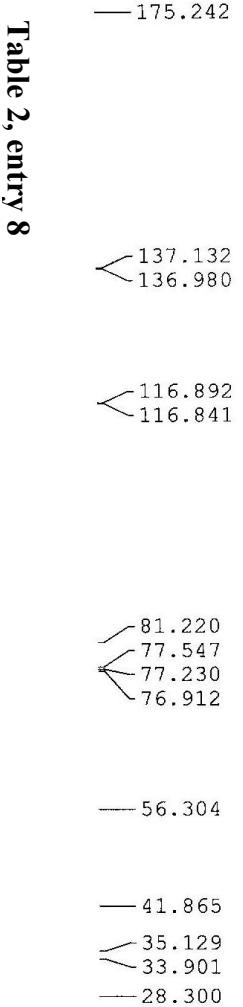


Table 2, entry 8

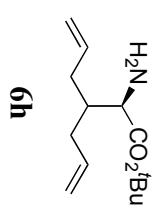


Table 2, entry 9

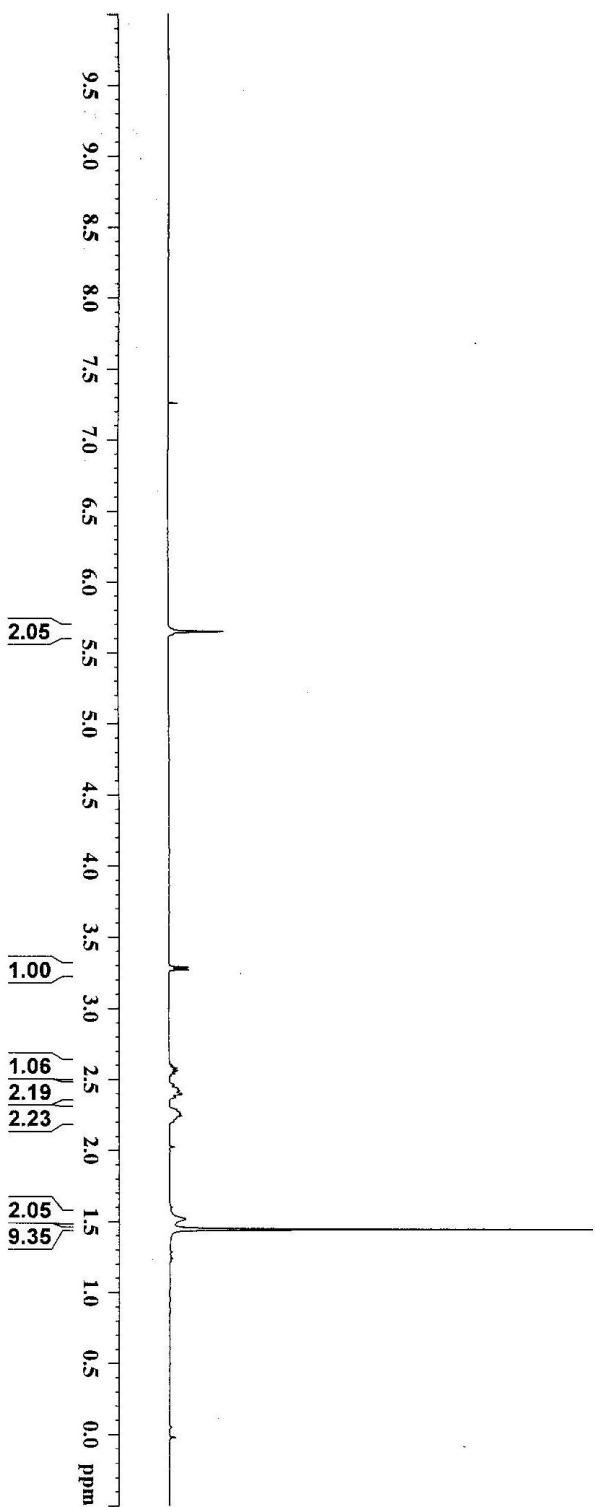
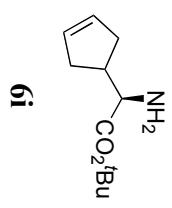


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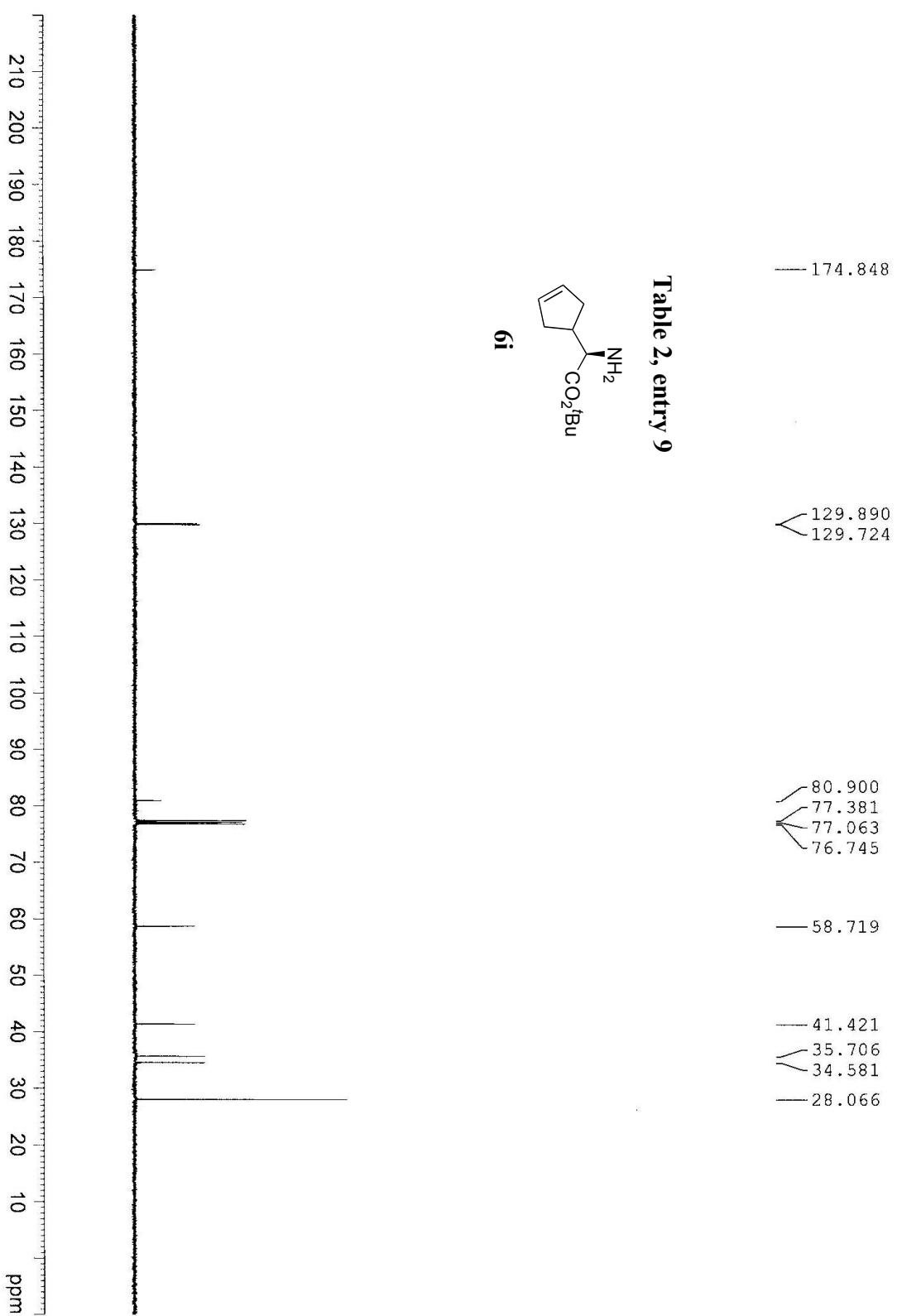
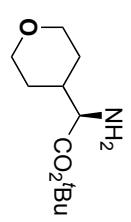
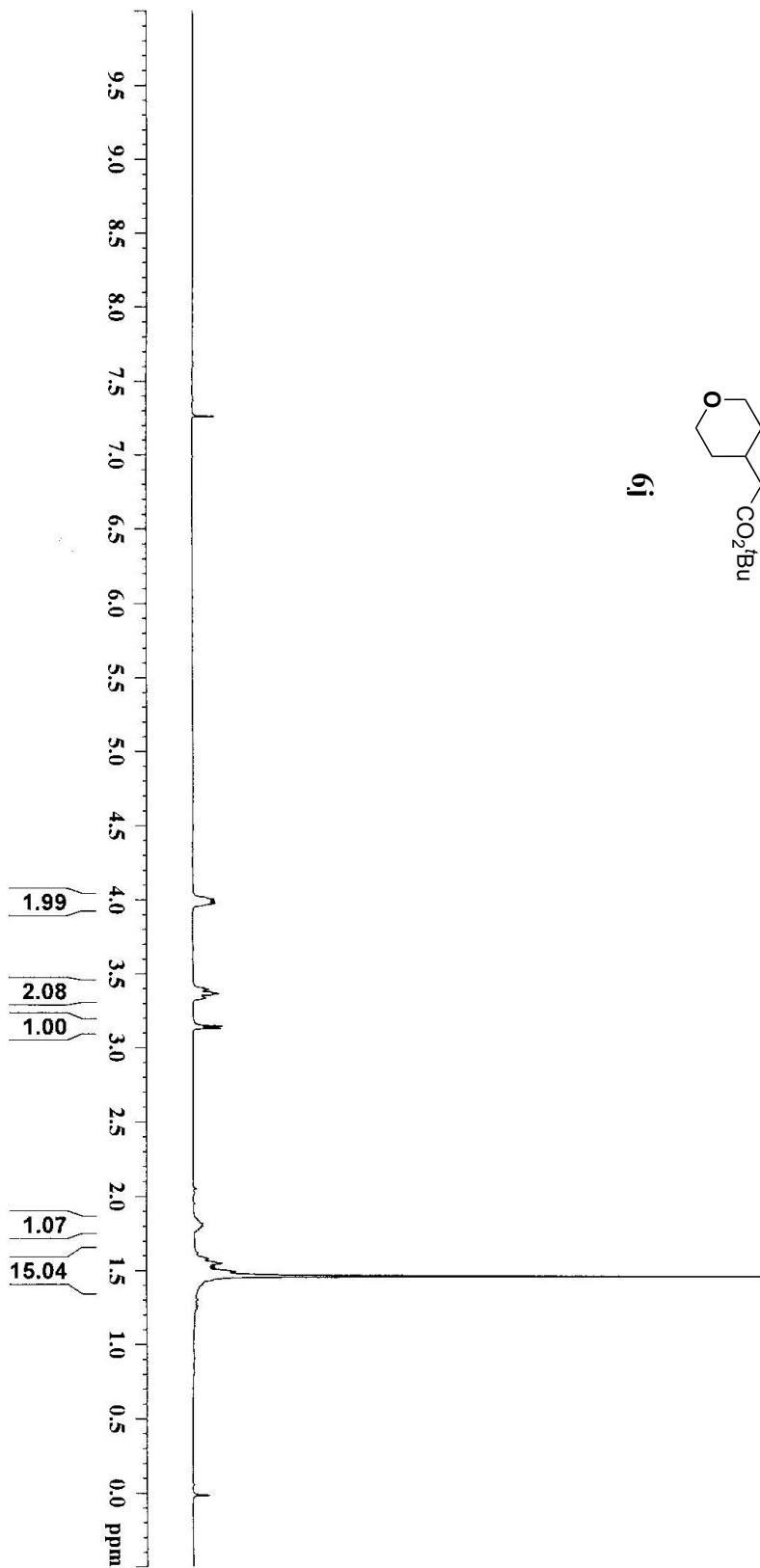


Table 2, entry 10



6j



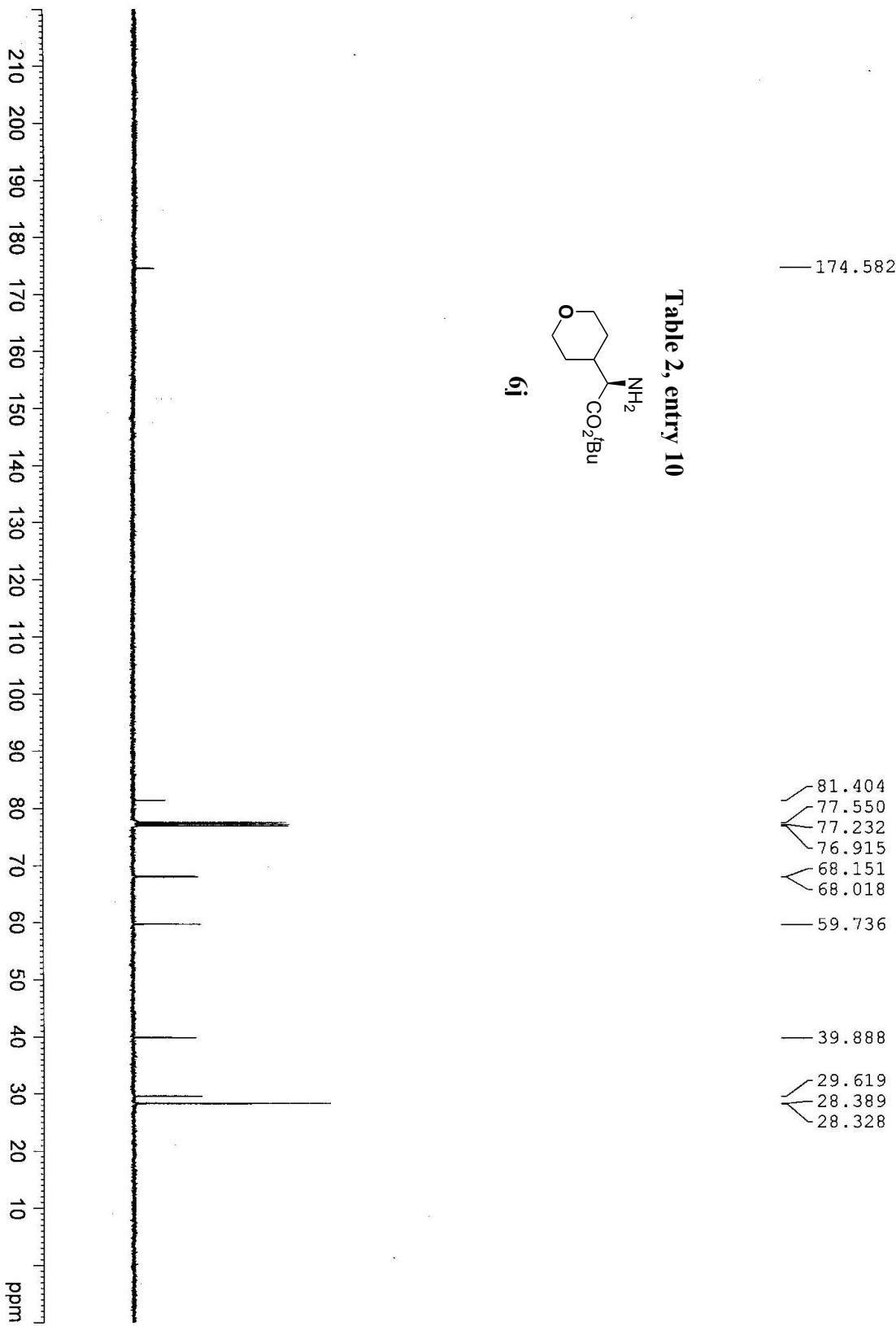
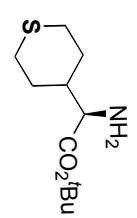


Table 2, entry 10

Table 2, entry 11



6k

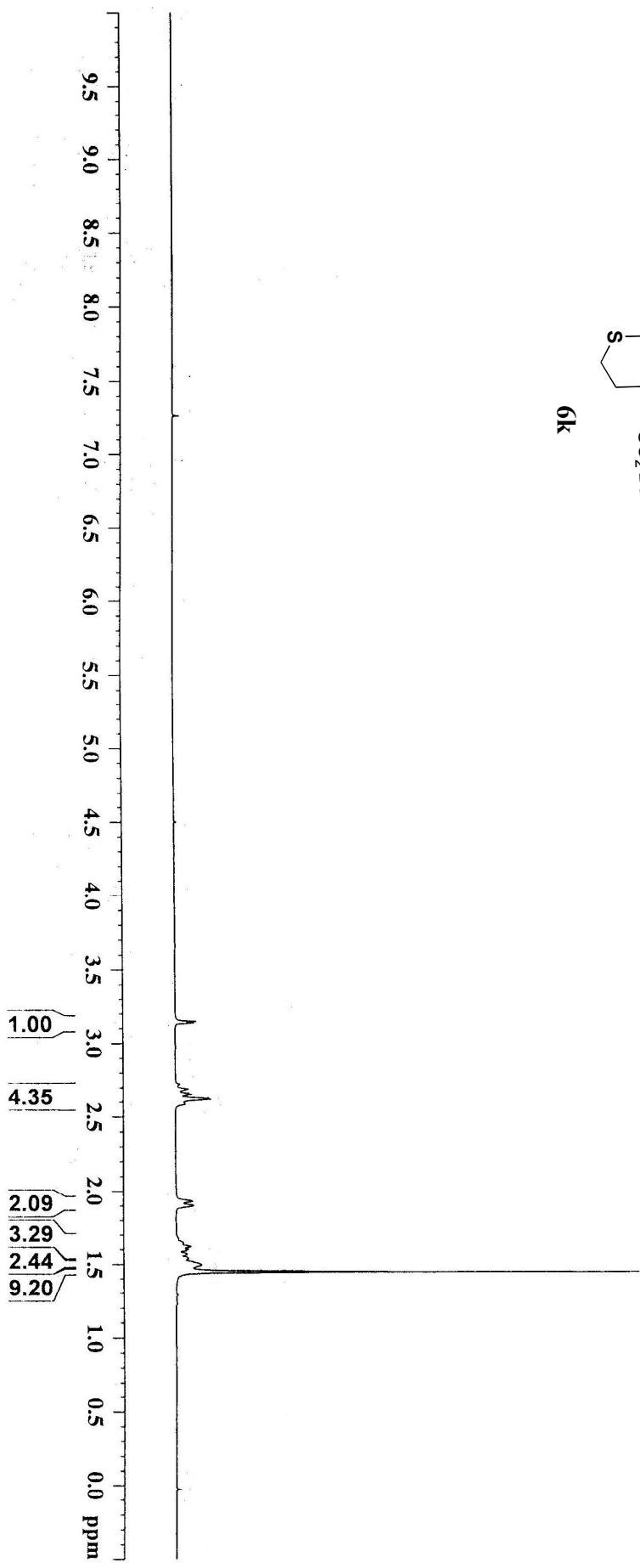


Table 2, entry 11

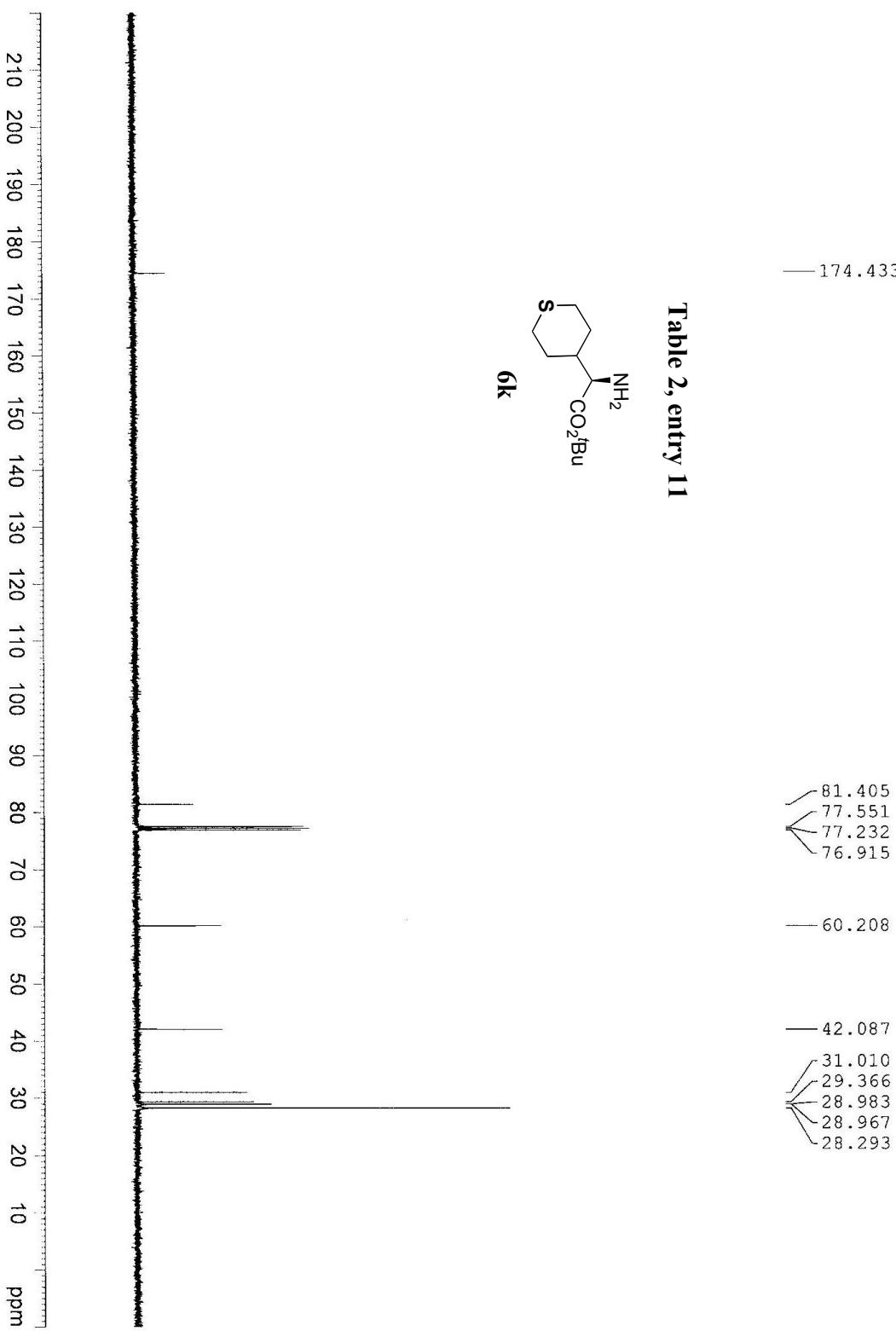
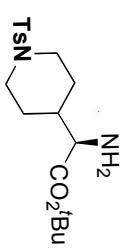


Table 2, entry 12



6l

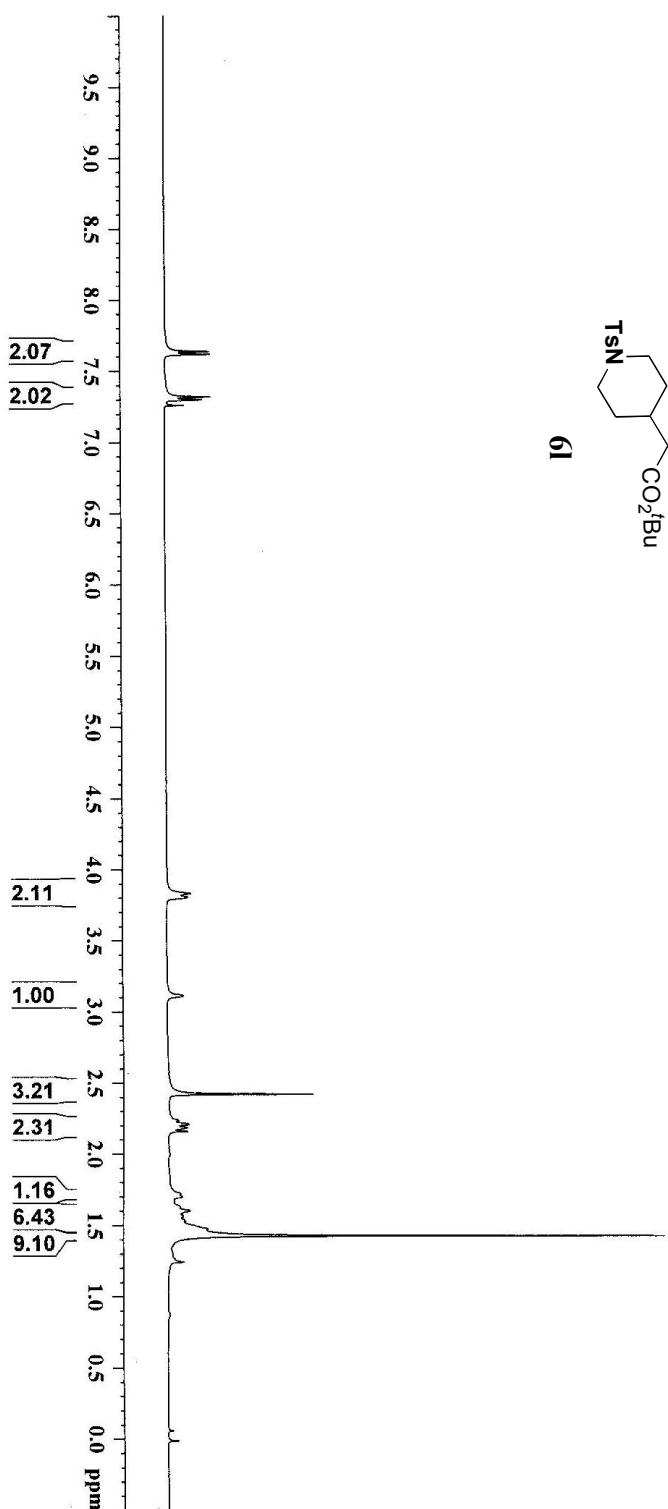
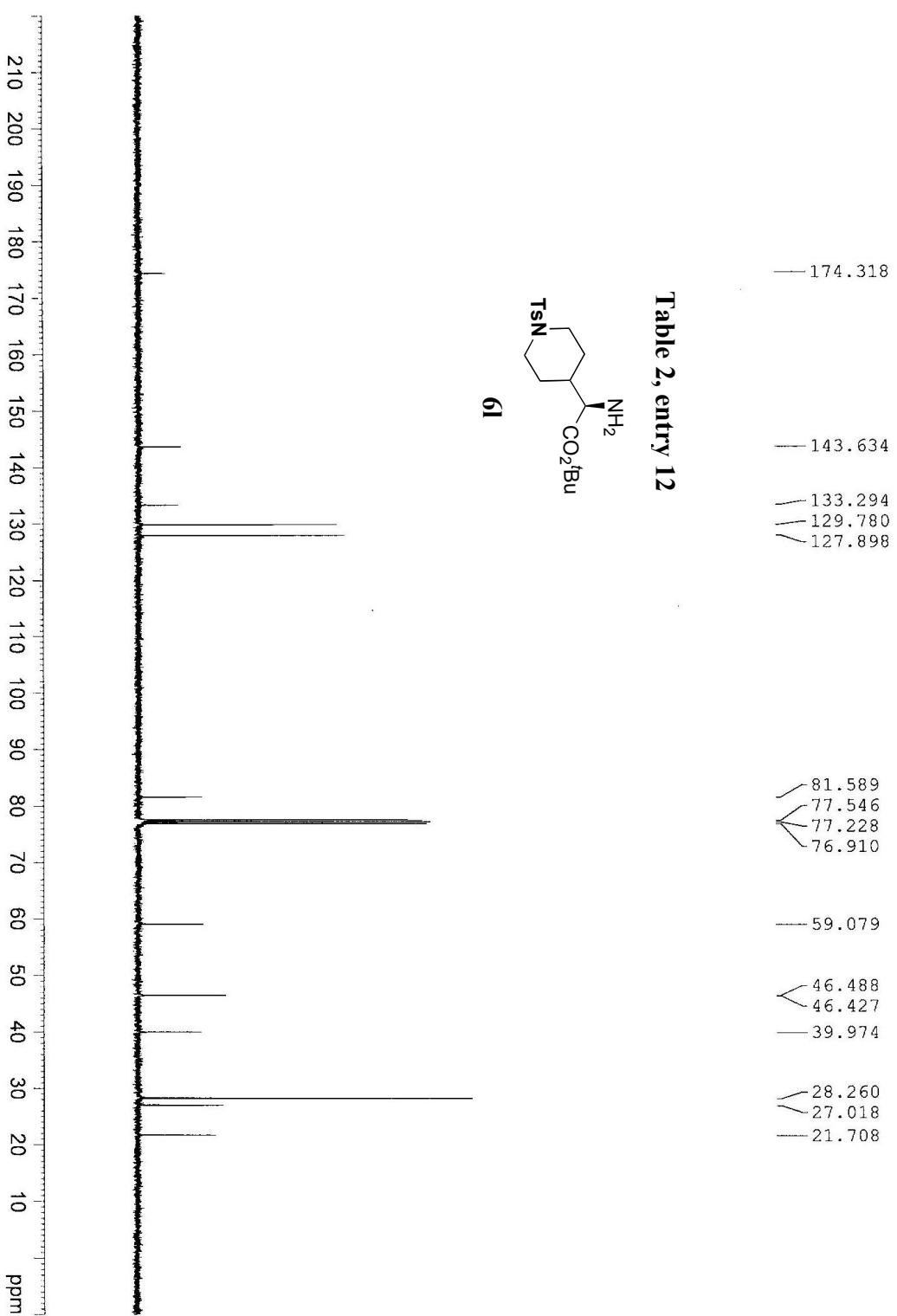
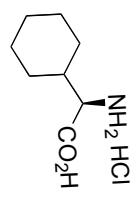
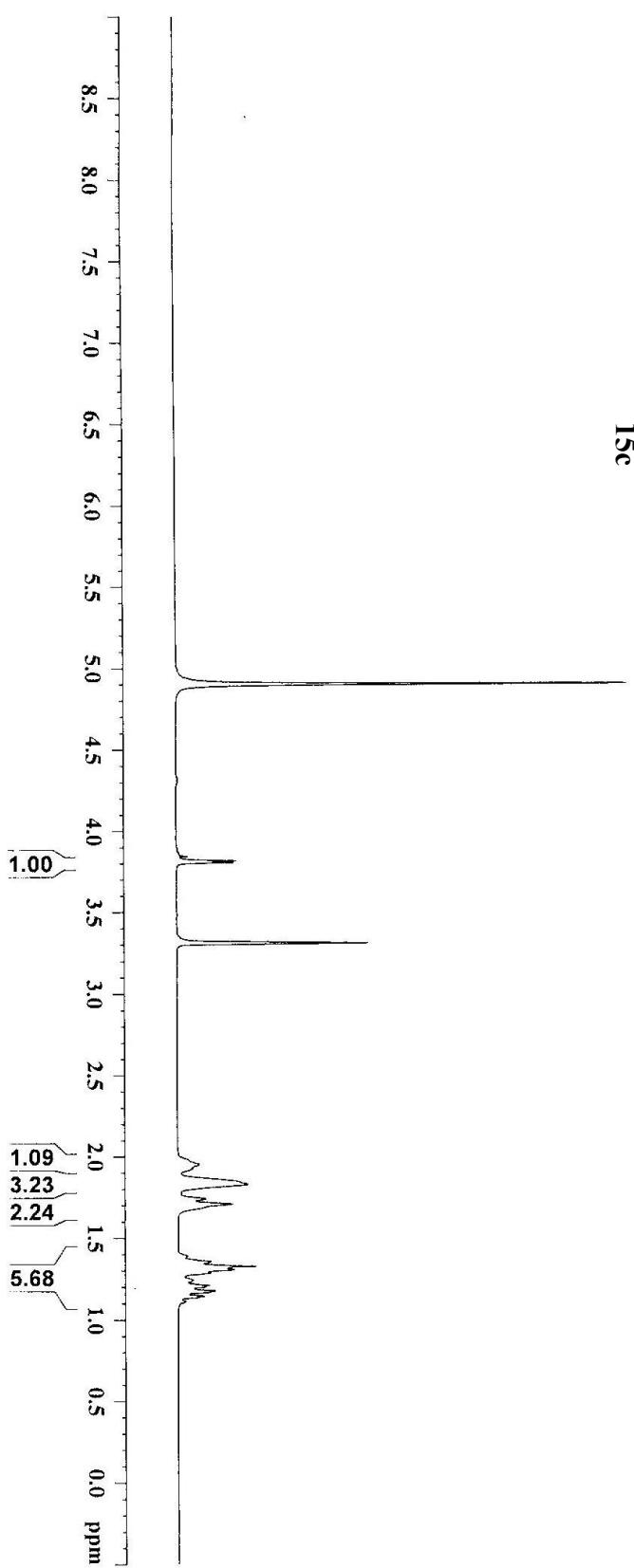


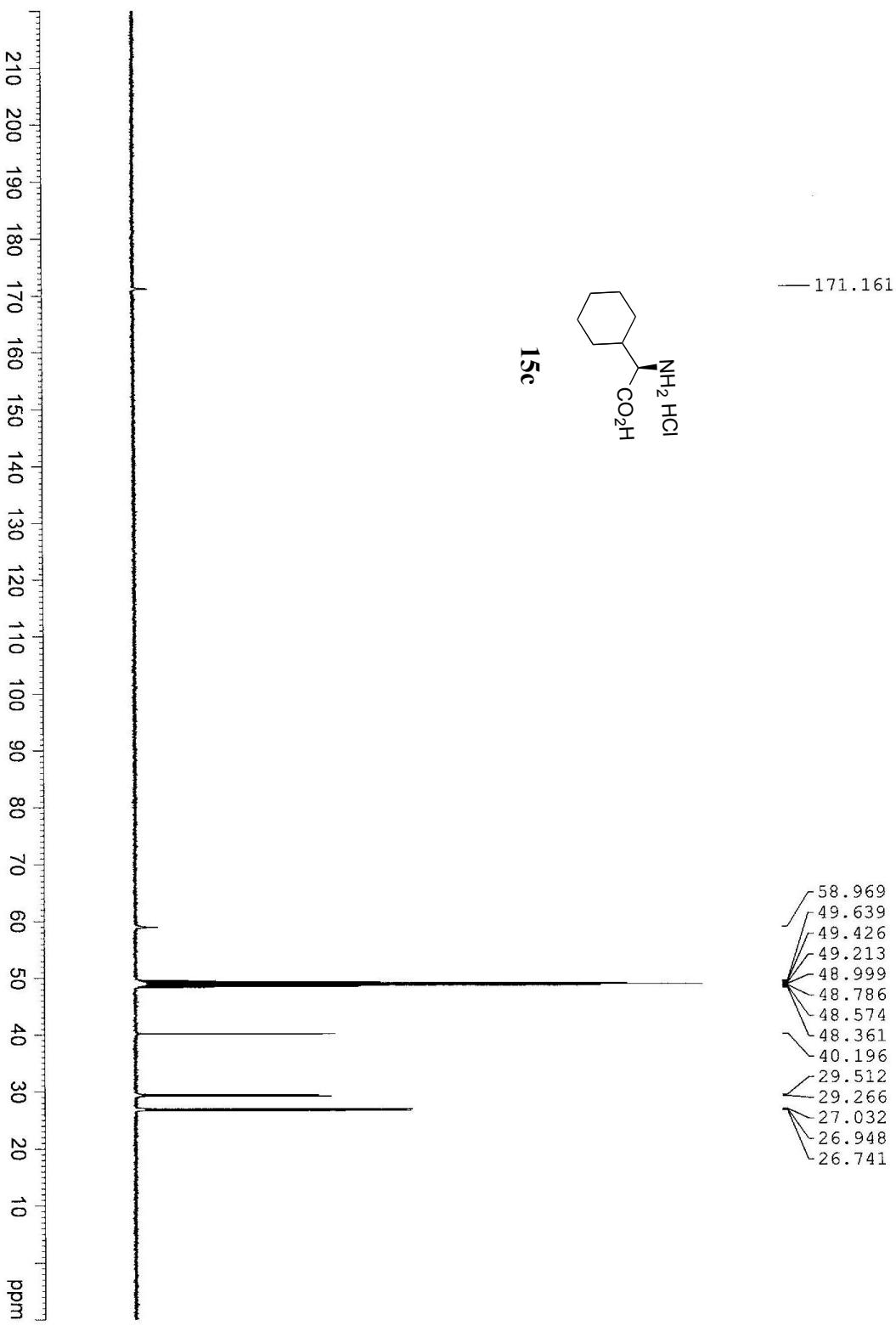
Table 2, entry 12

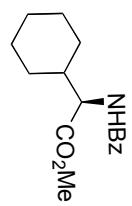
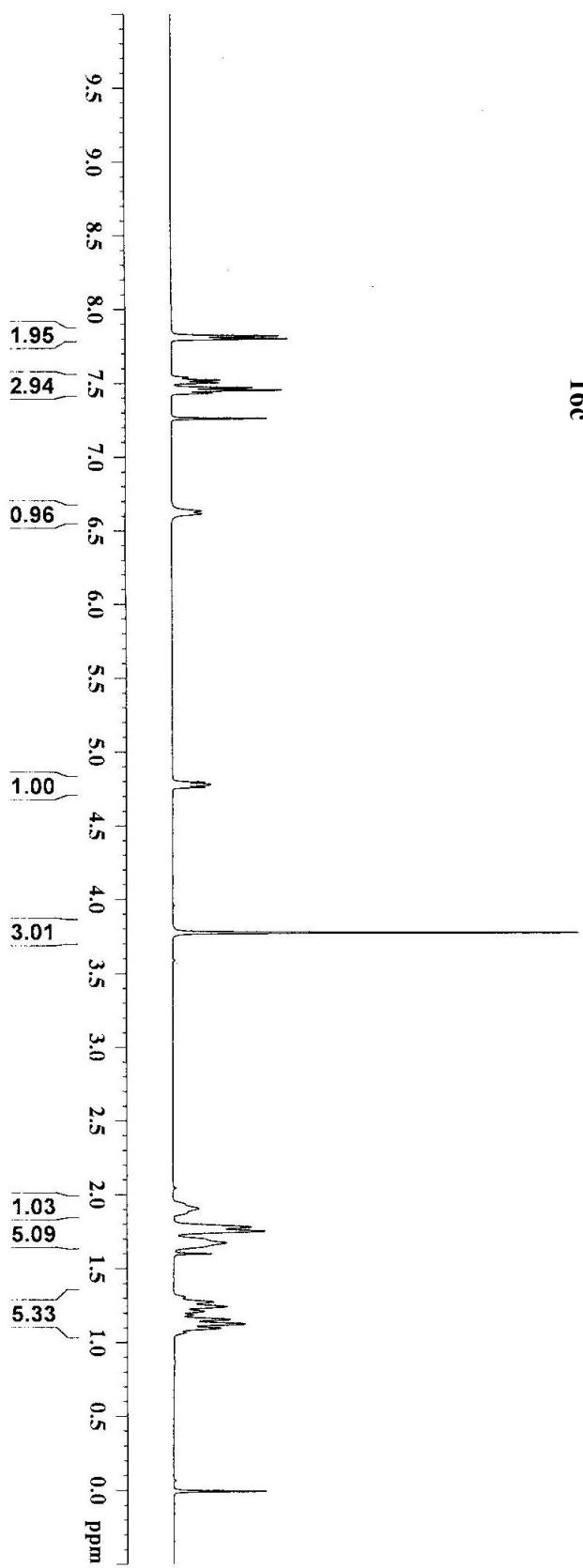




15c







16c

