

Supporting Information

**Evaluation of the Effect of Fluorination on the Property of
Monofluorinated Dimyristoylphosphatidylcholines**

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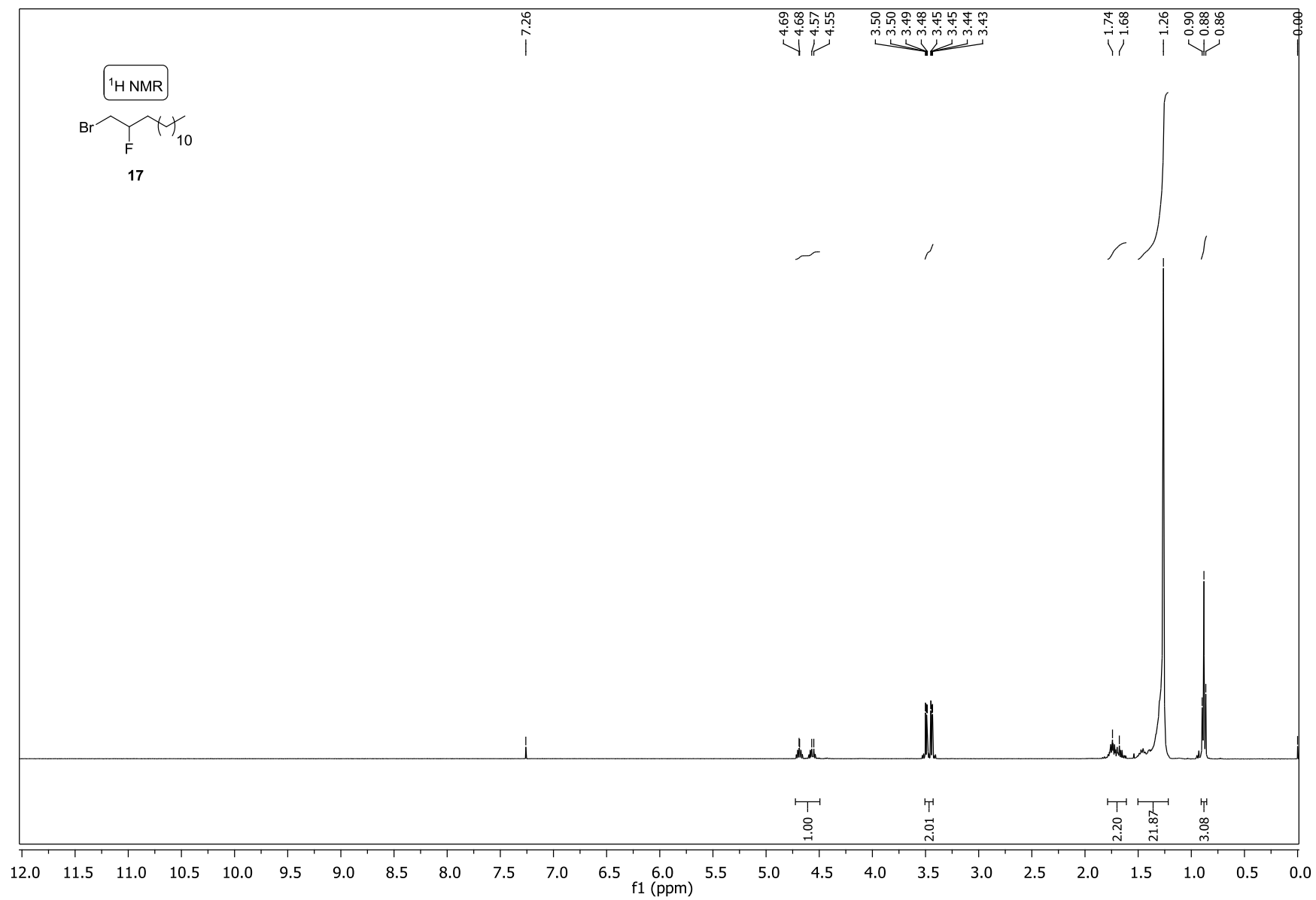
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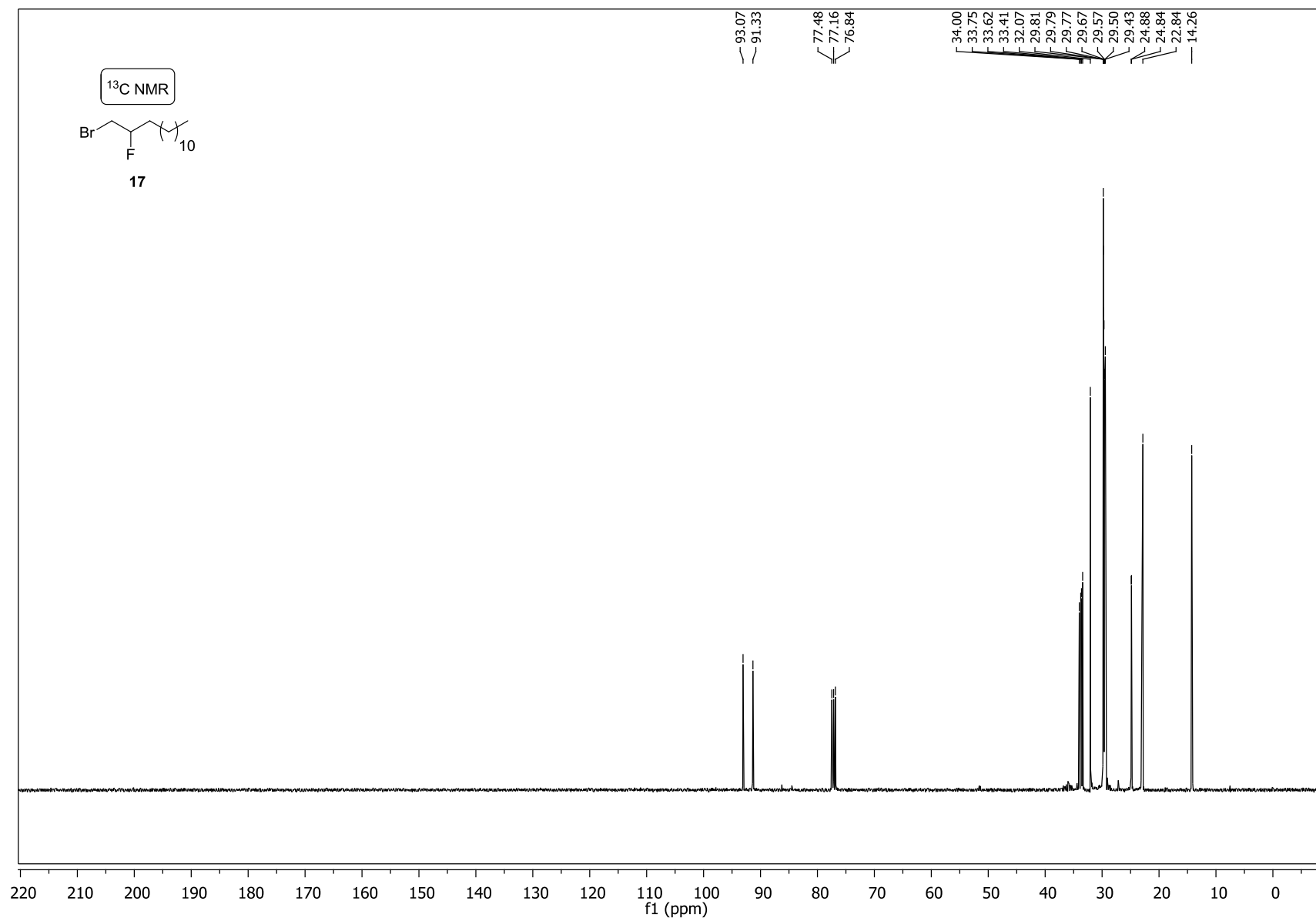
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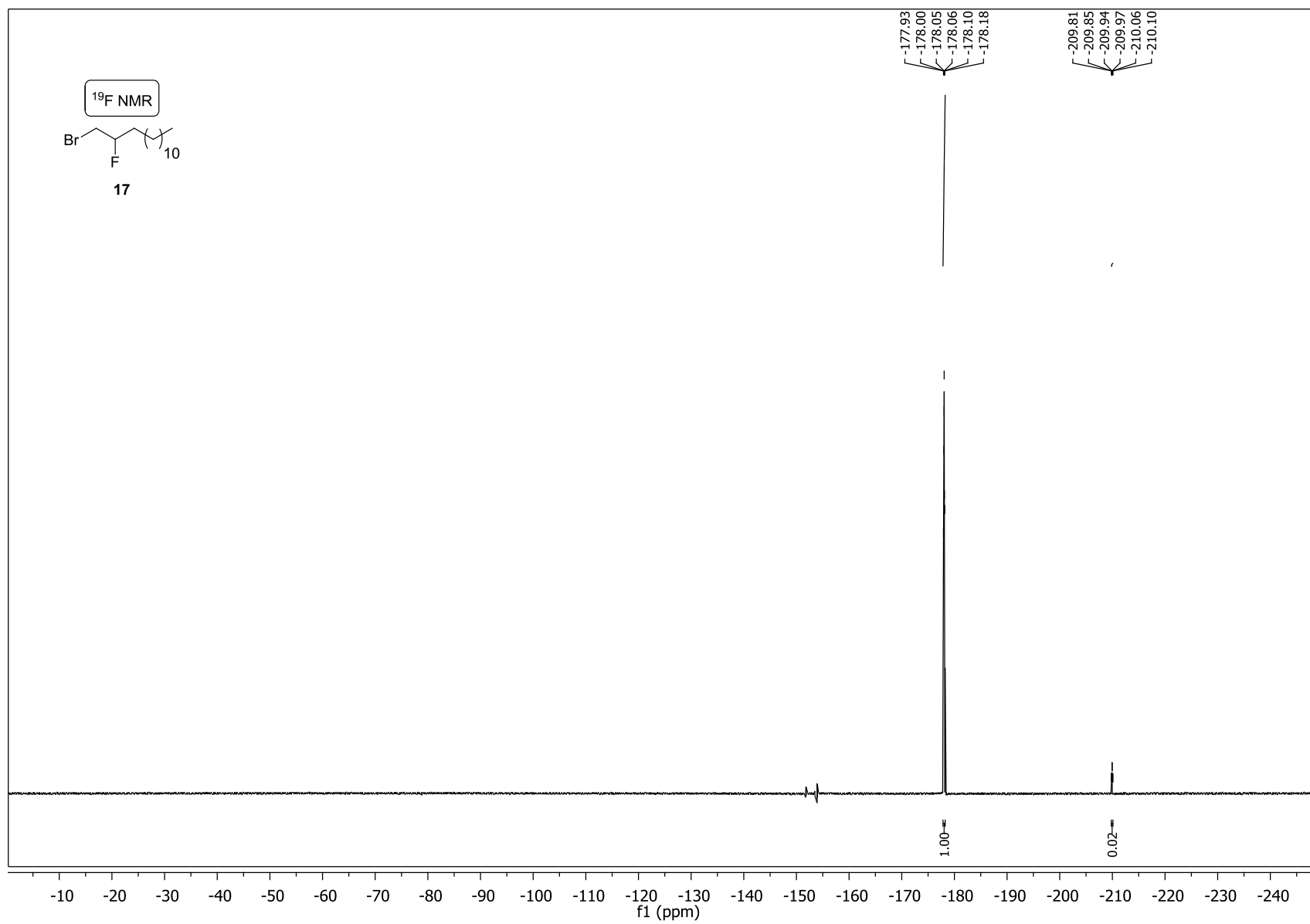
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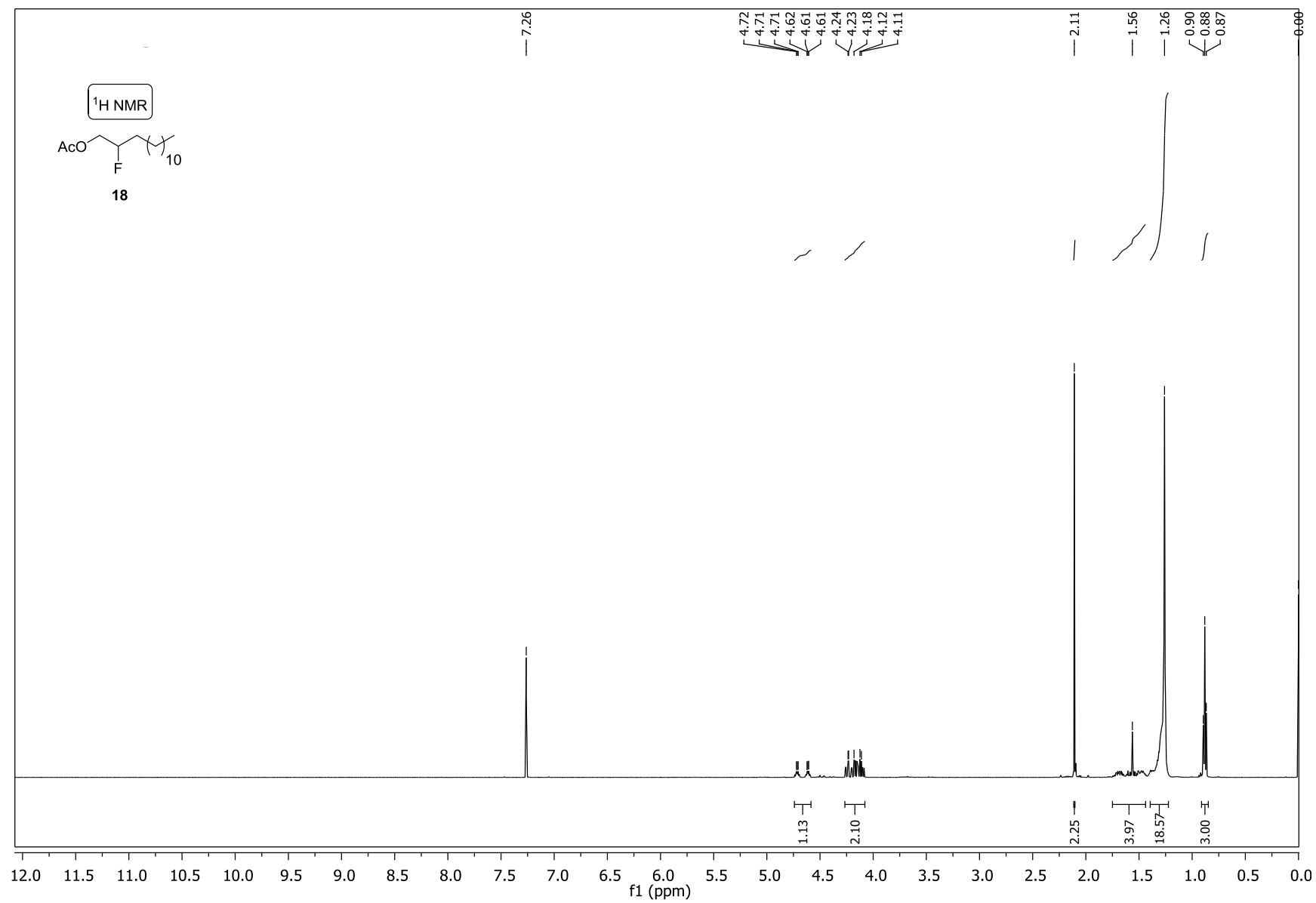
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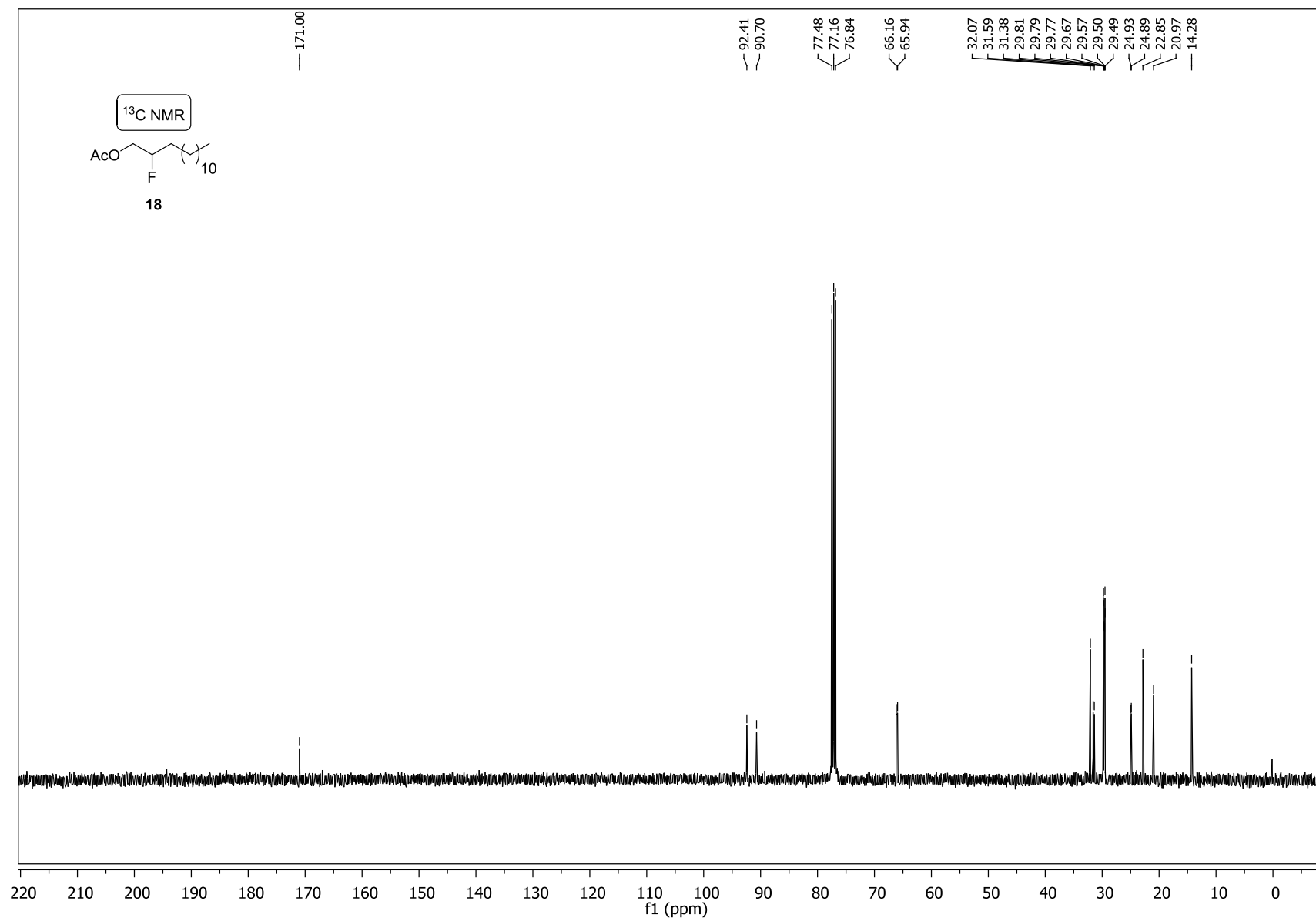
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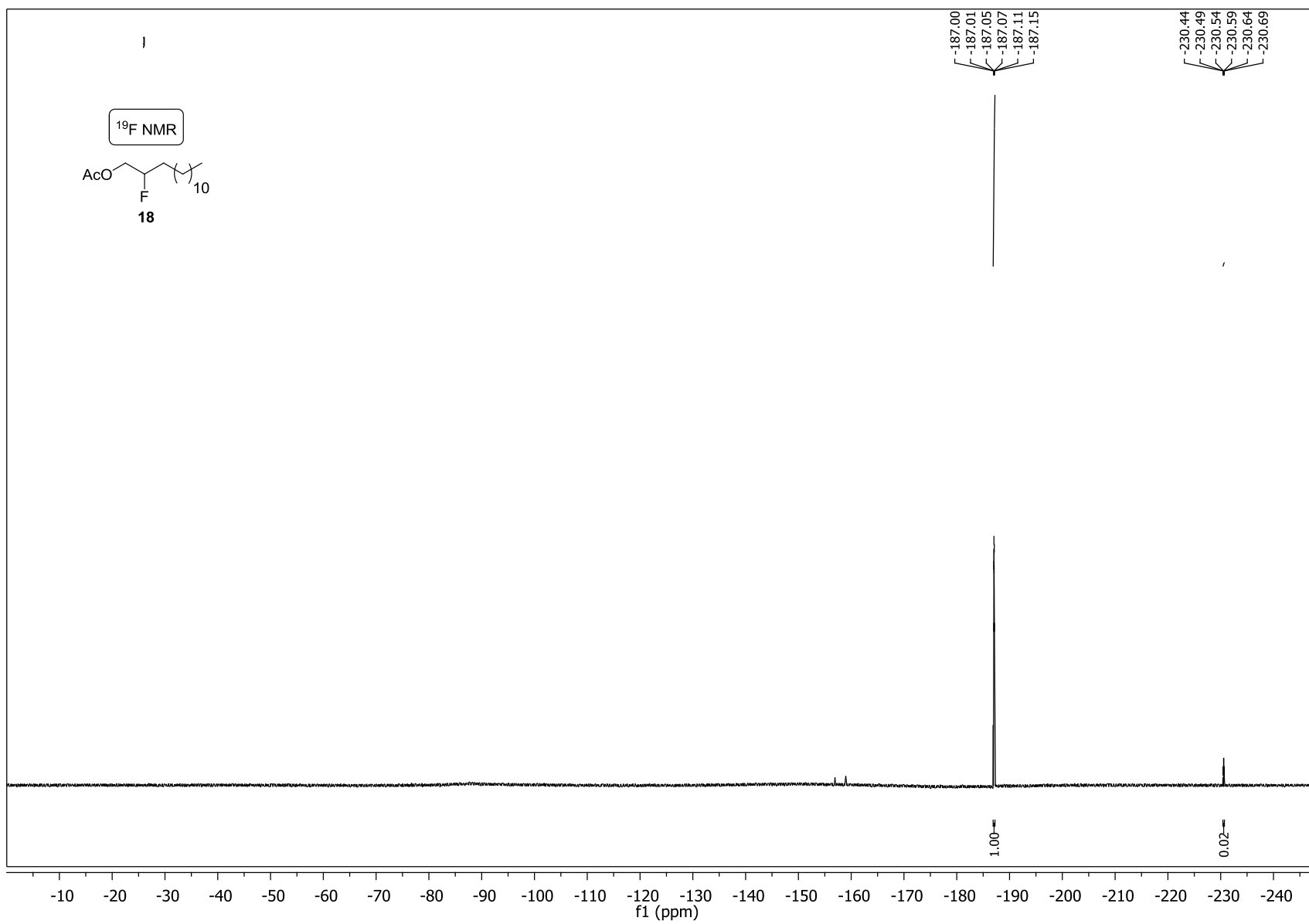


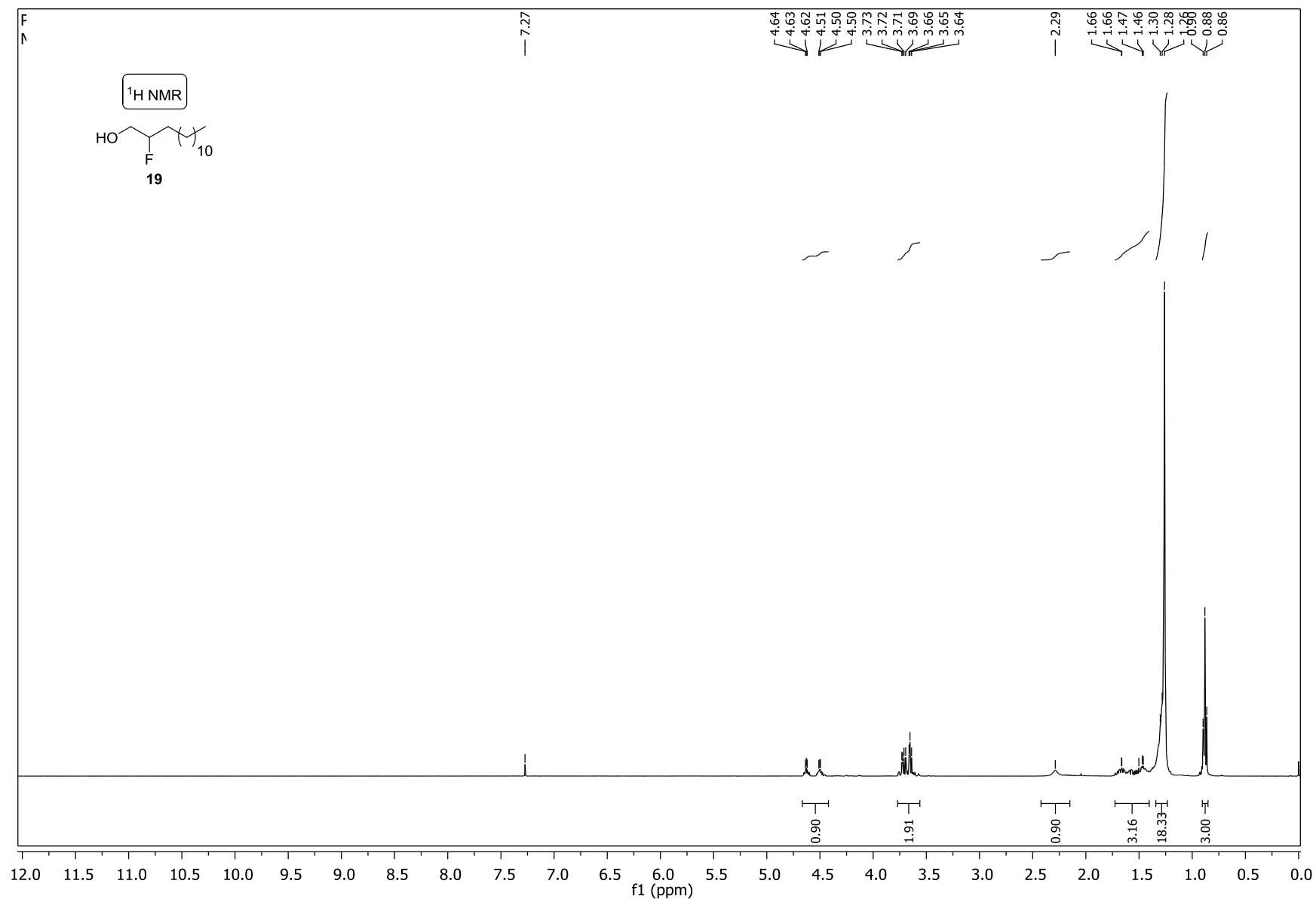


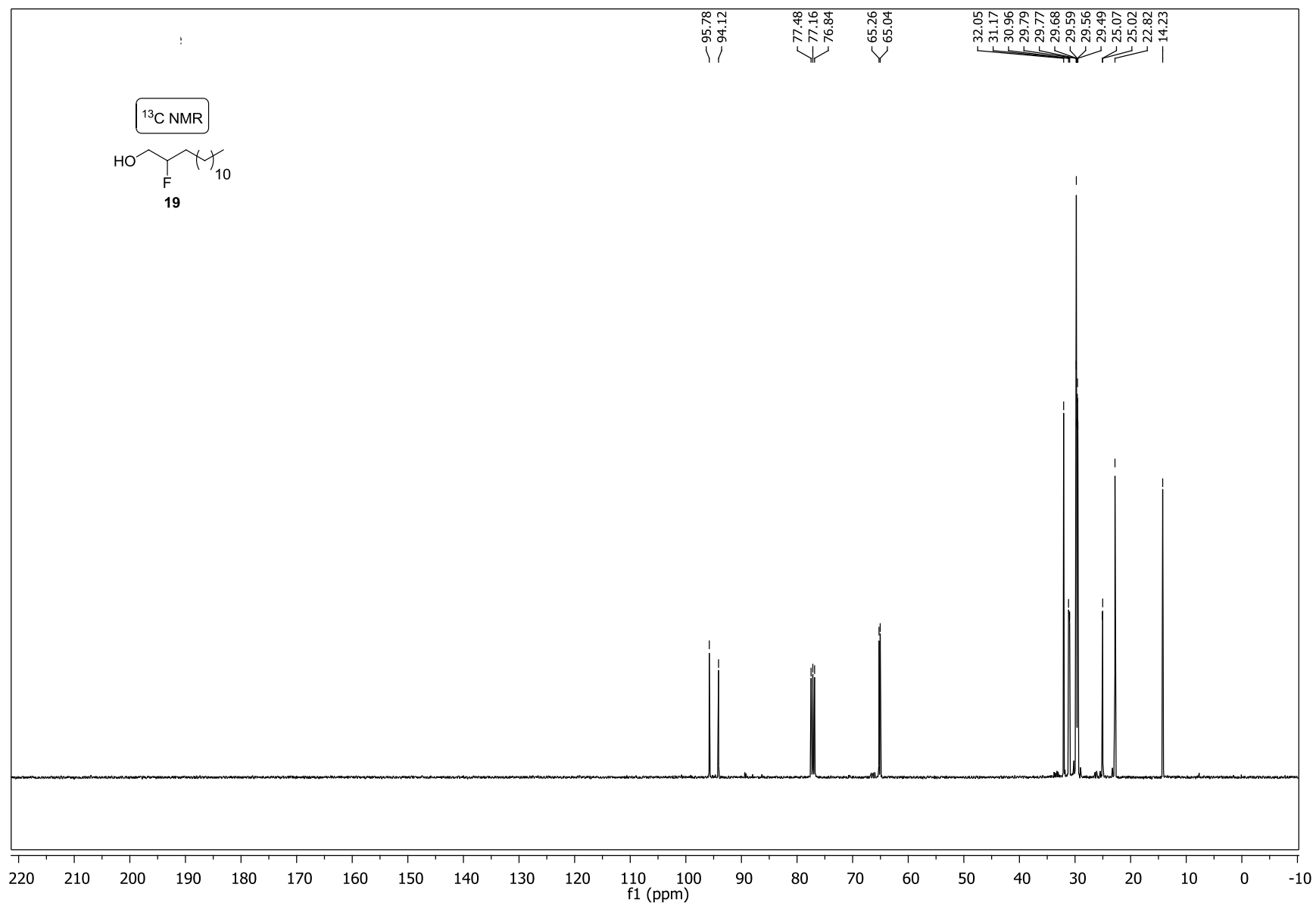


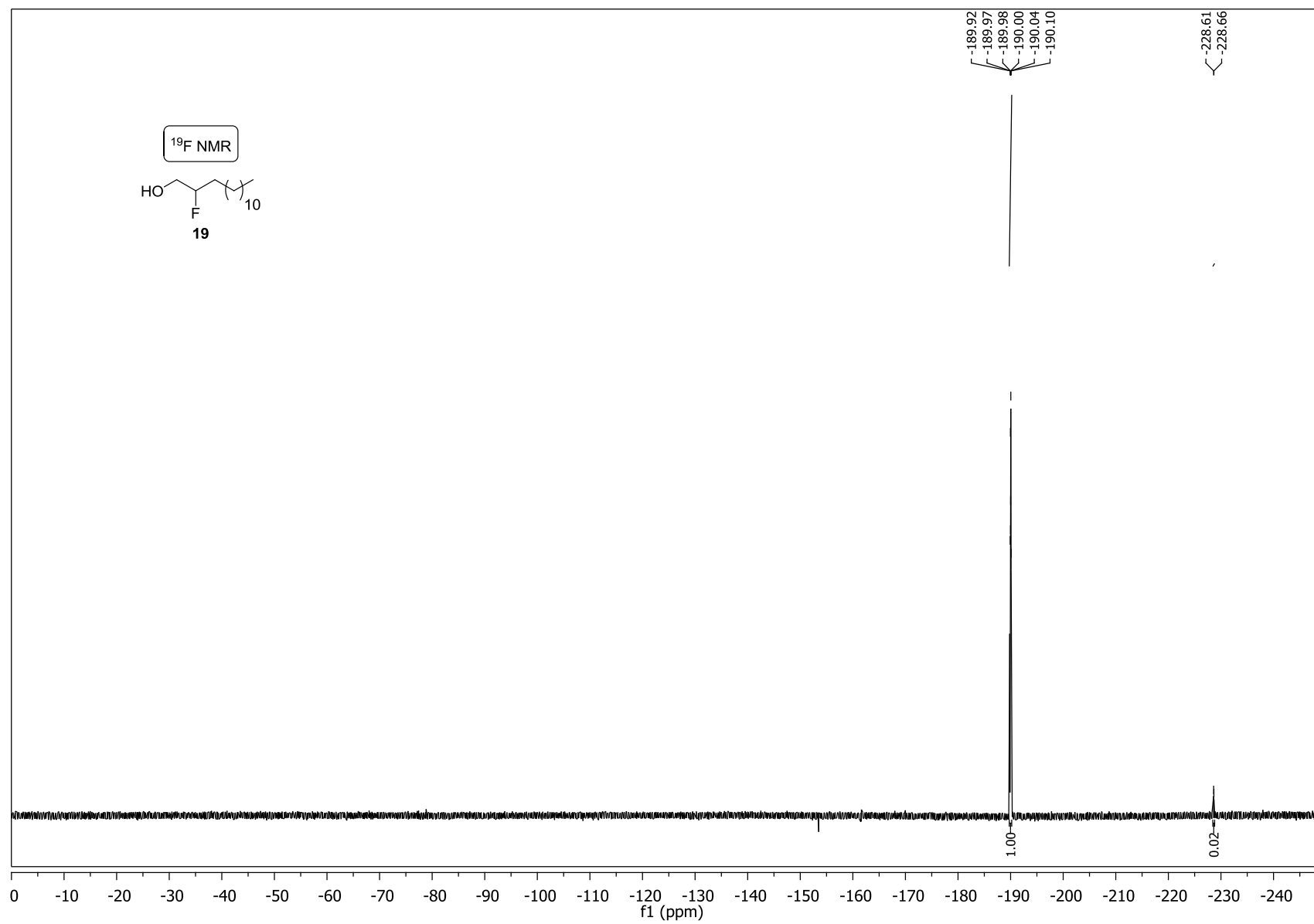


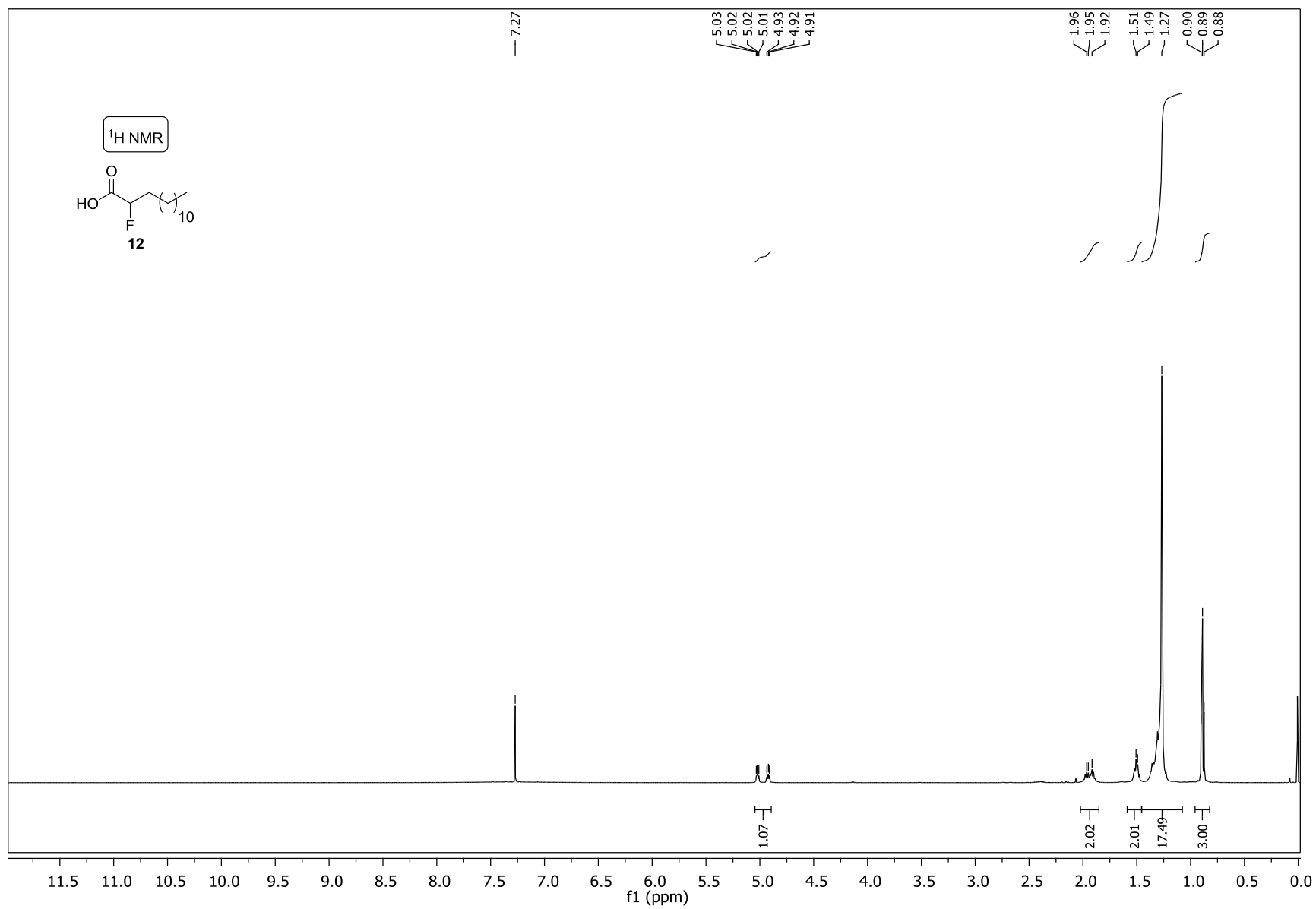


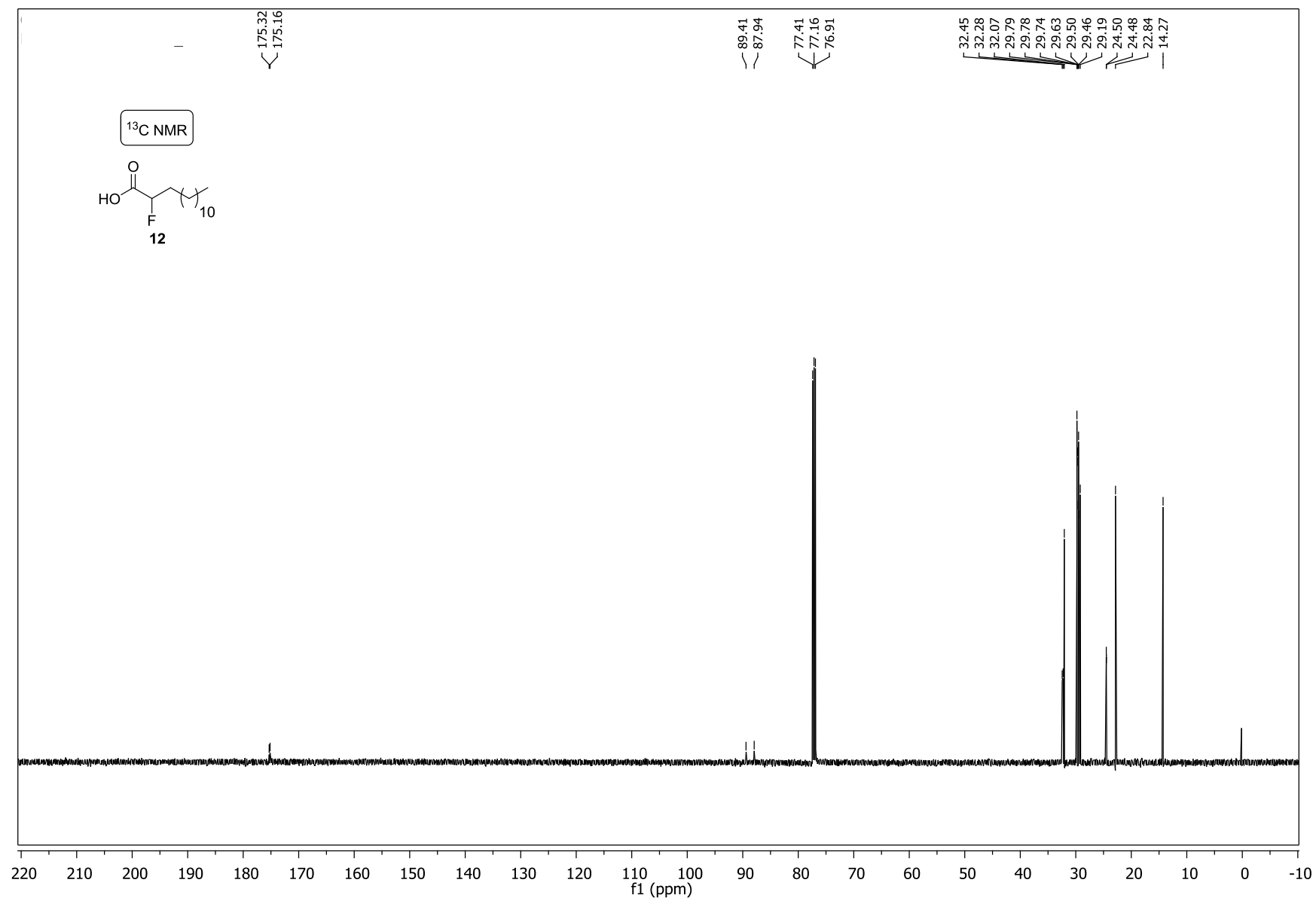


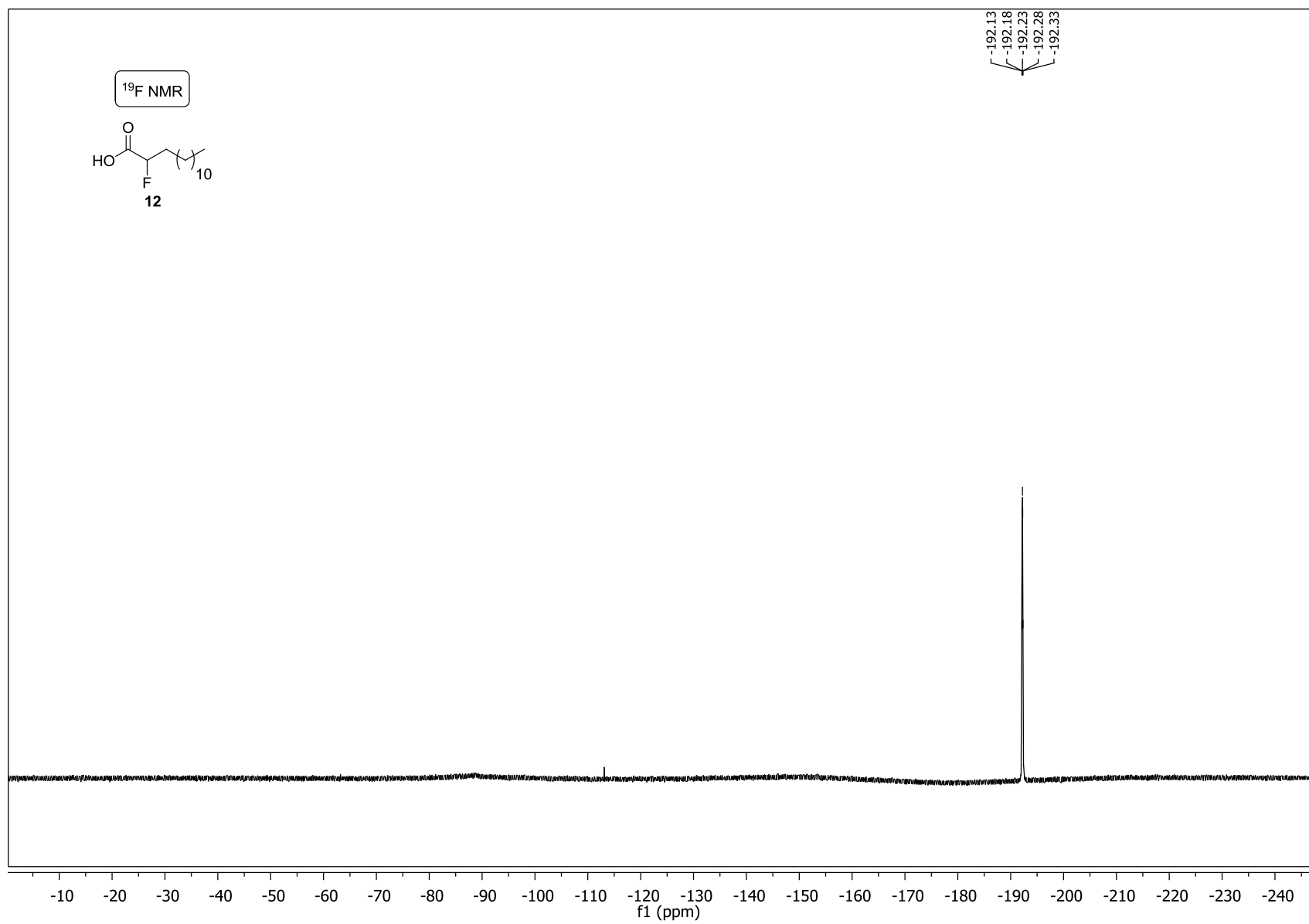


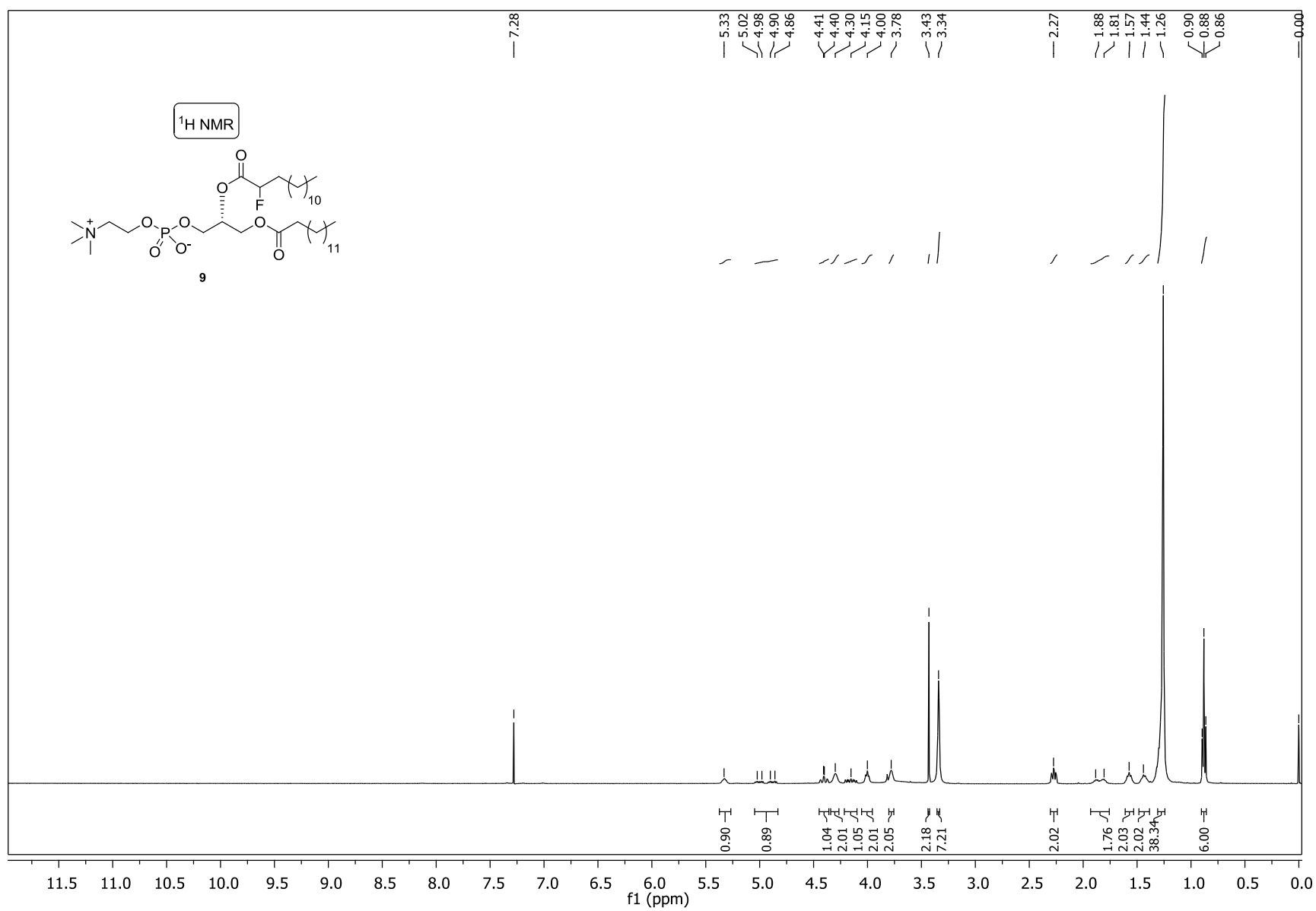


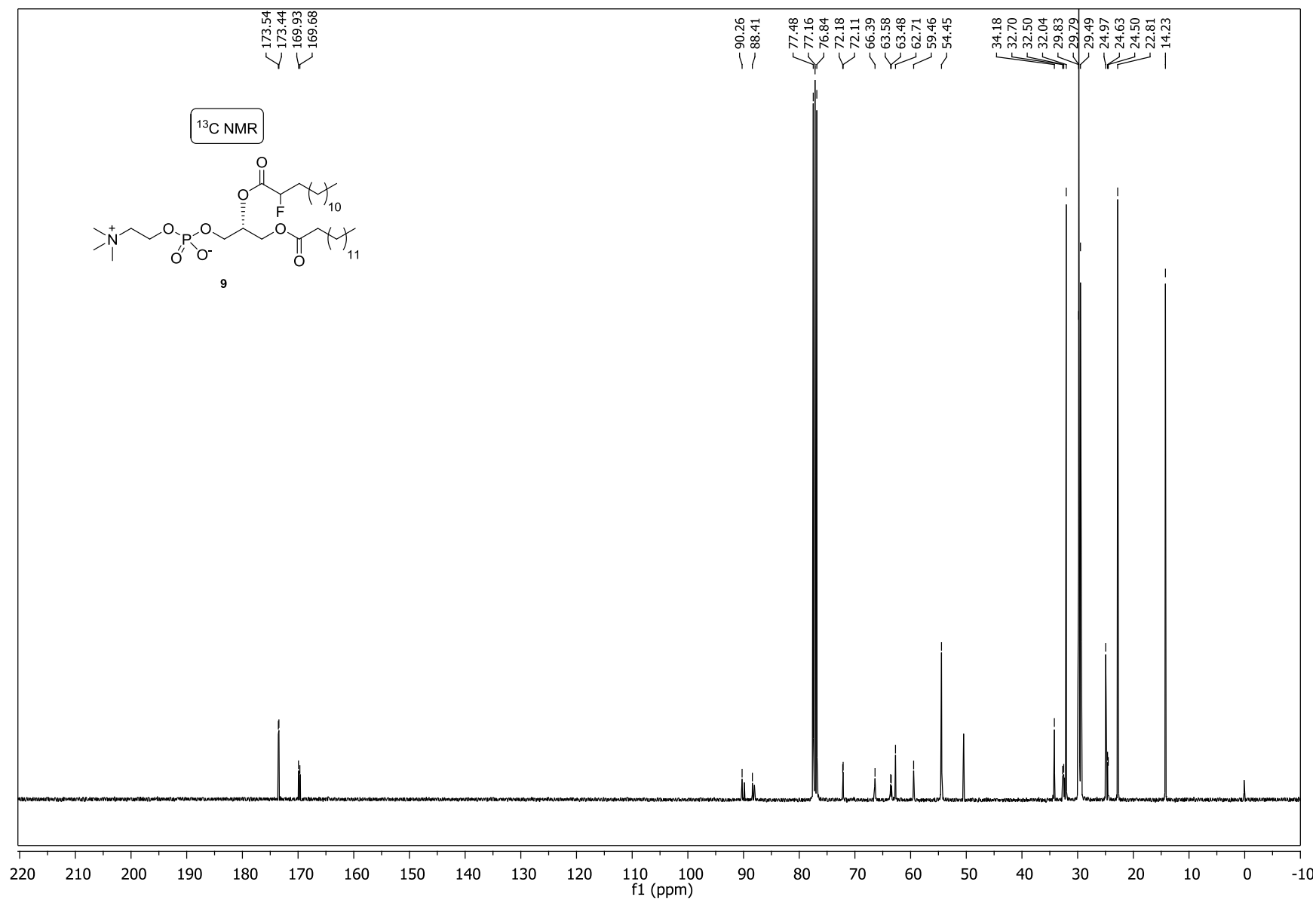


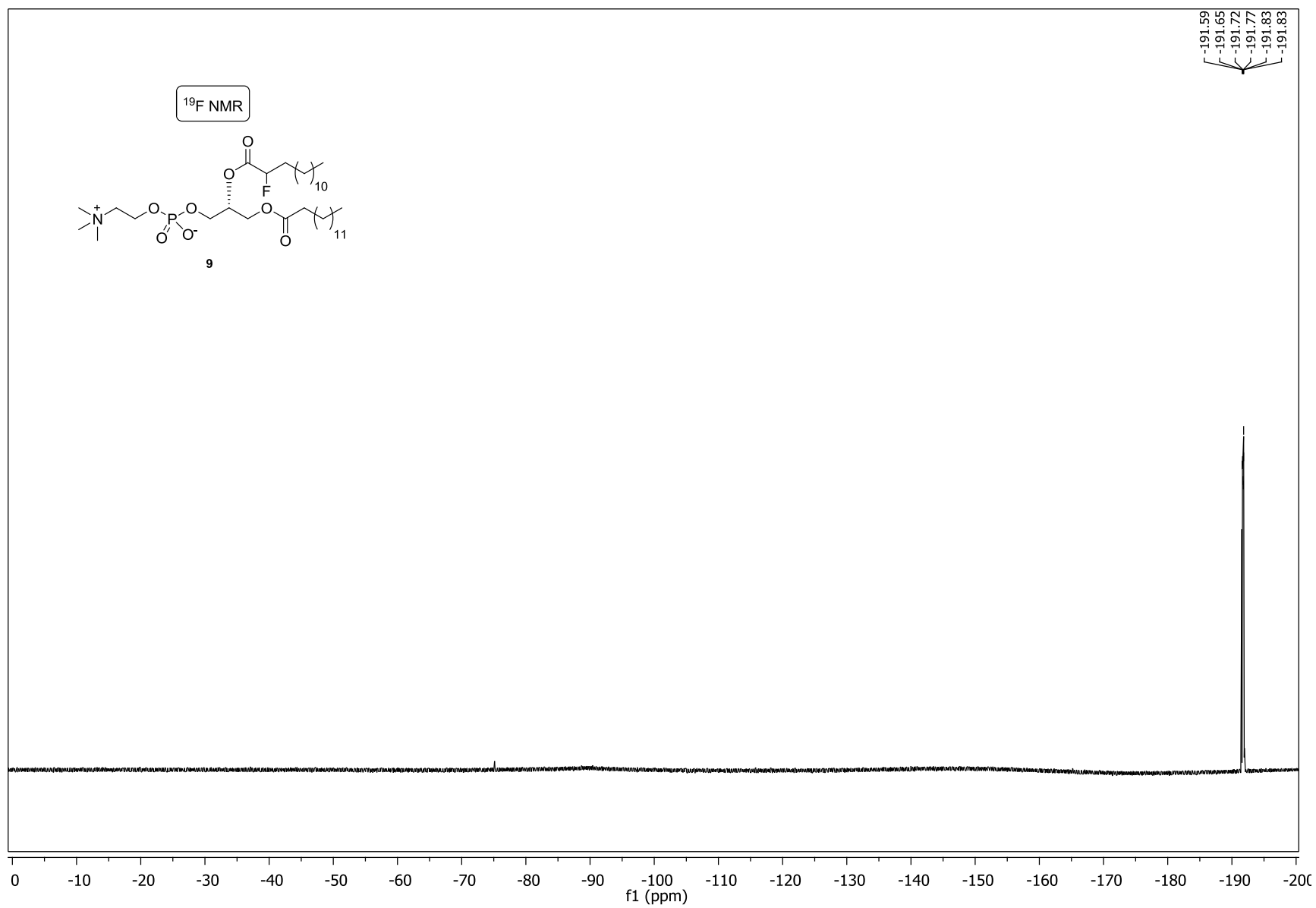


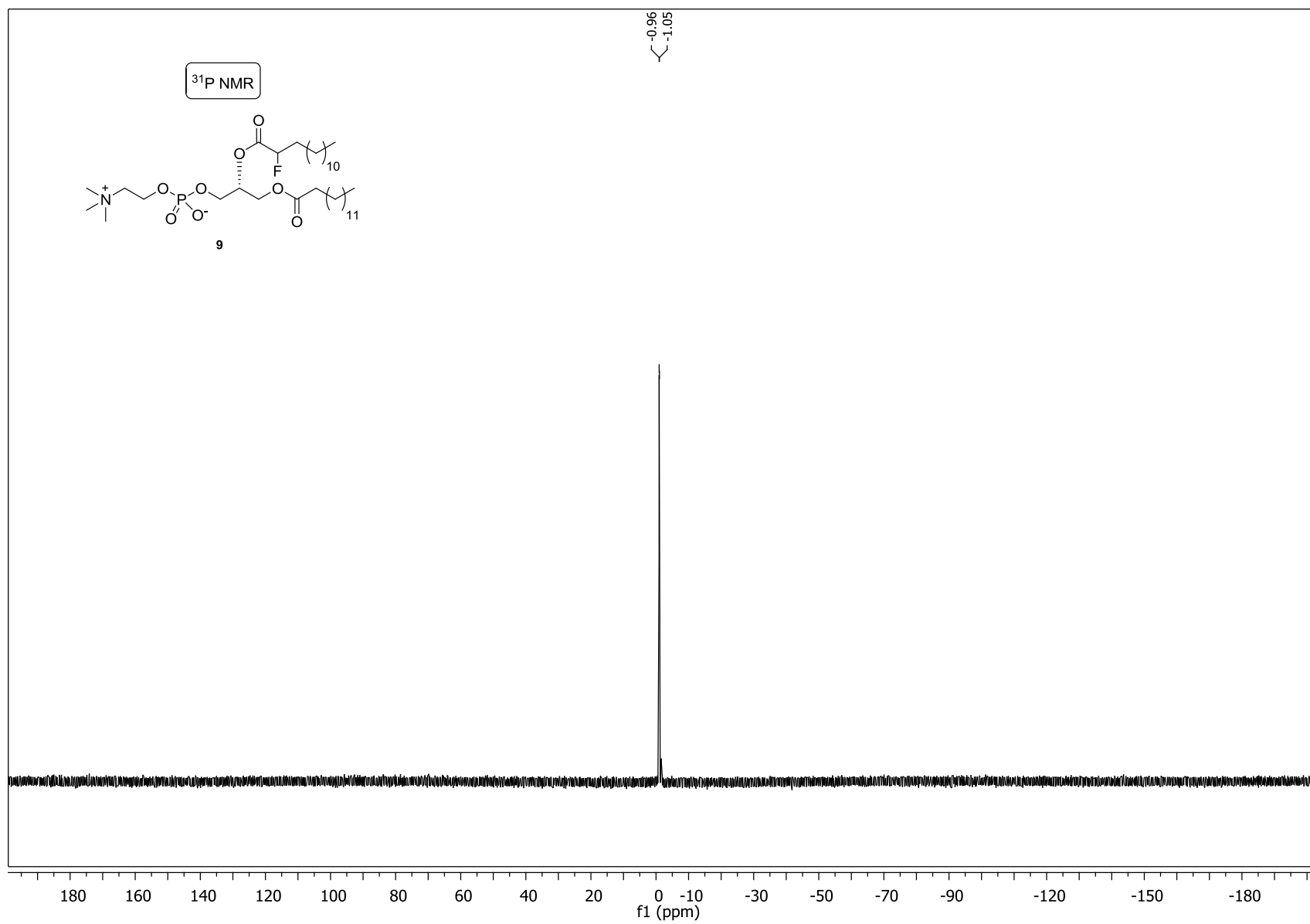


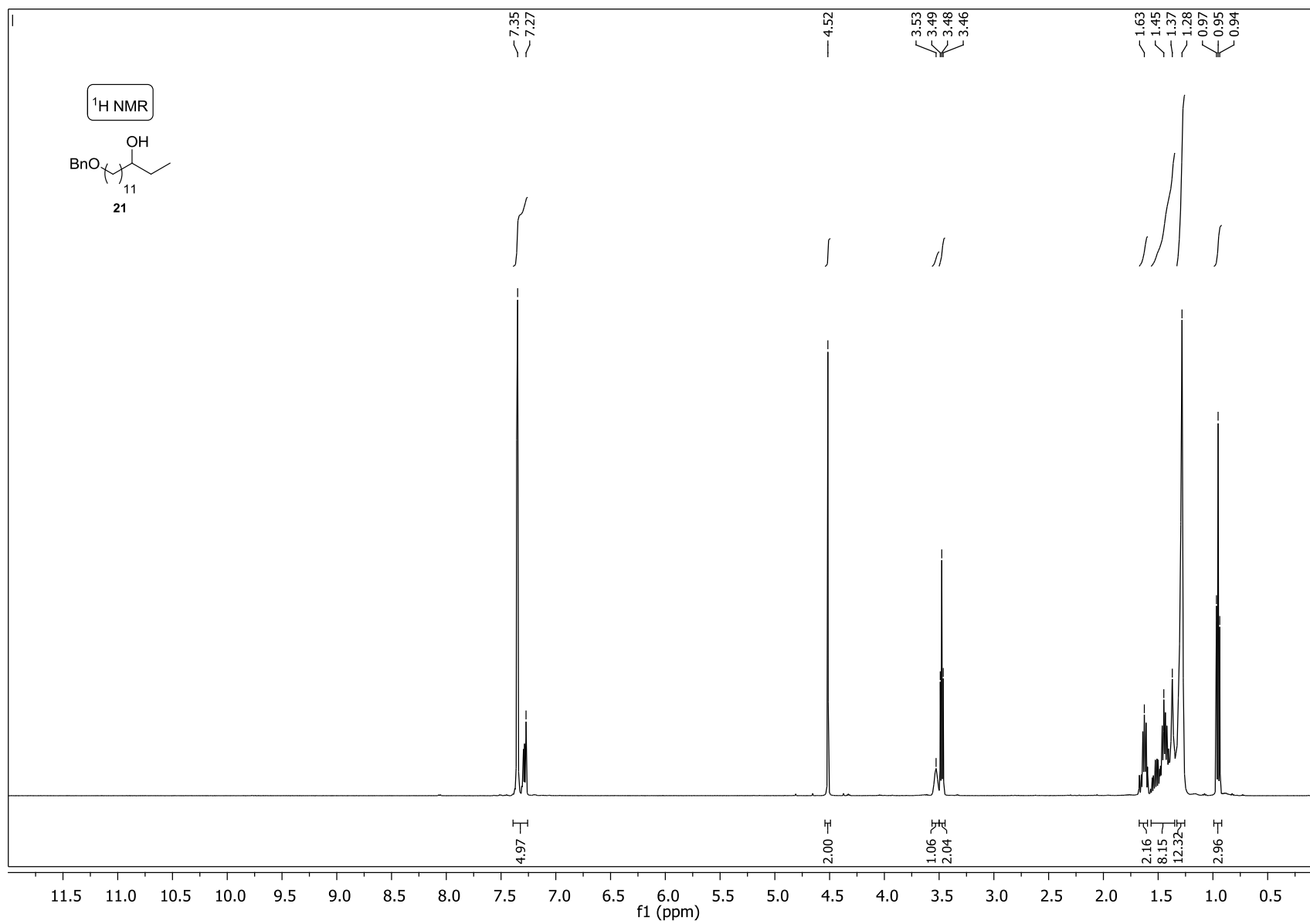


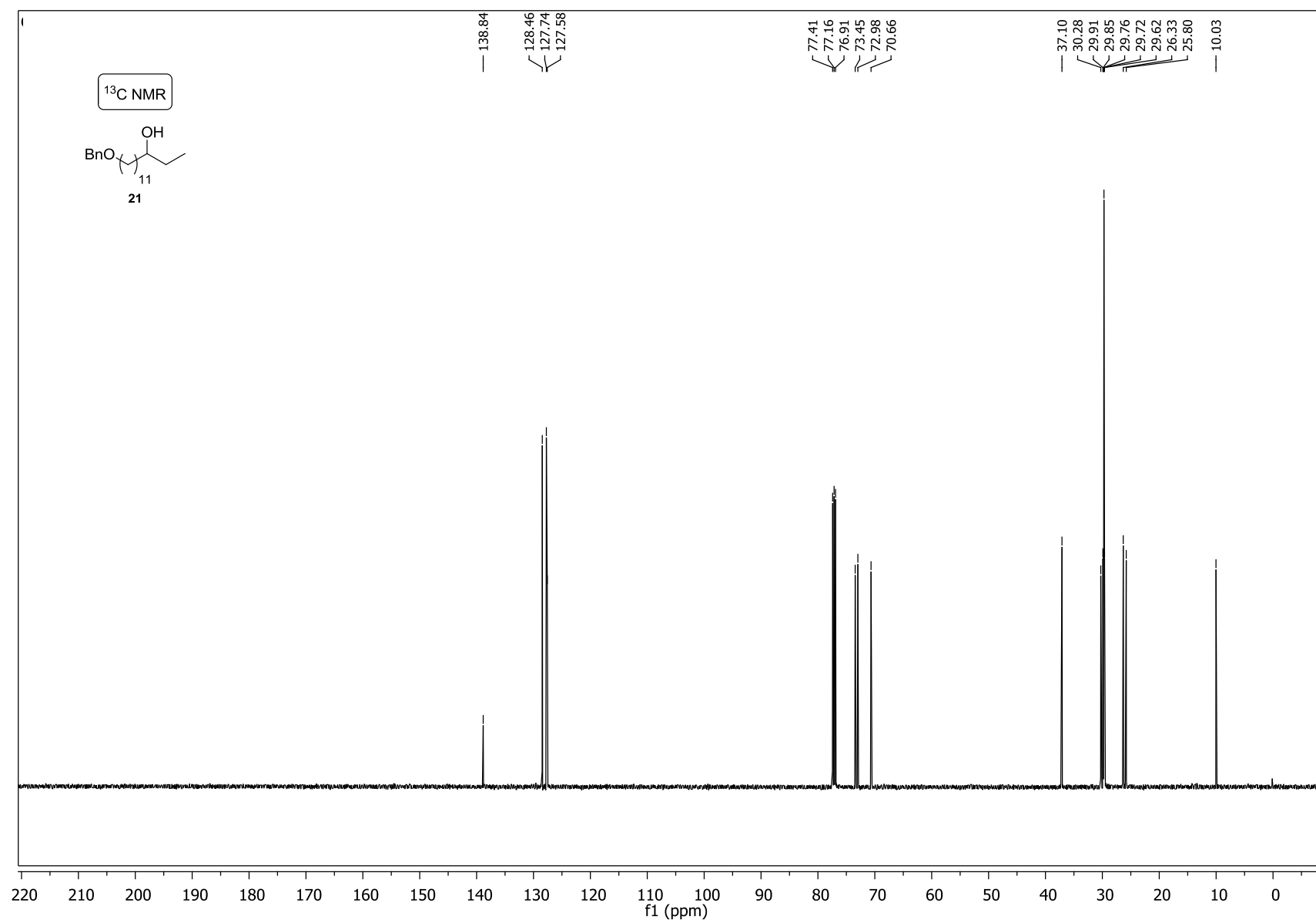


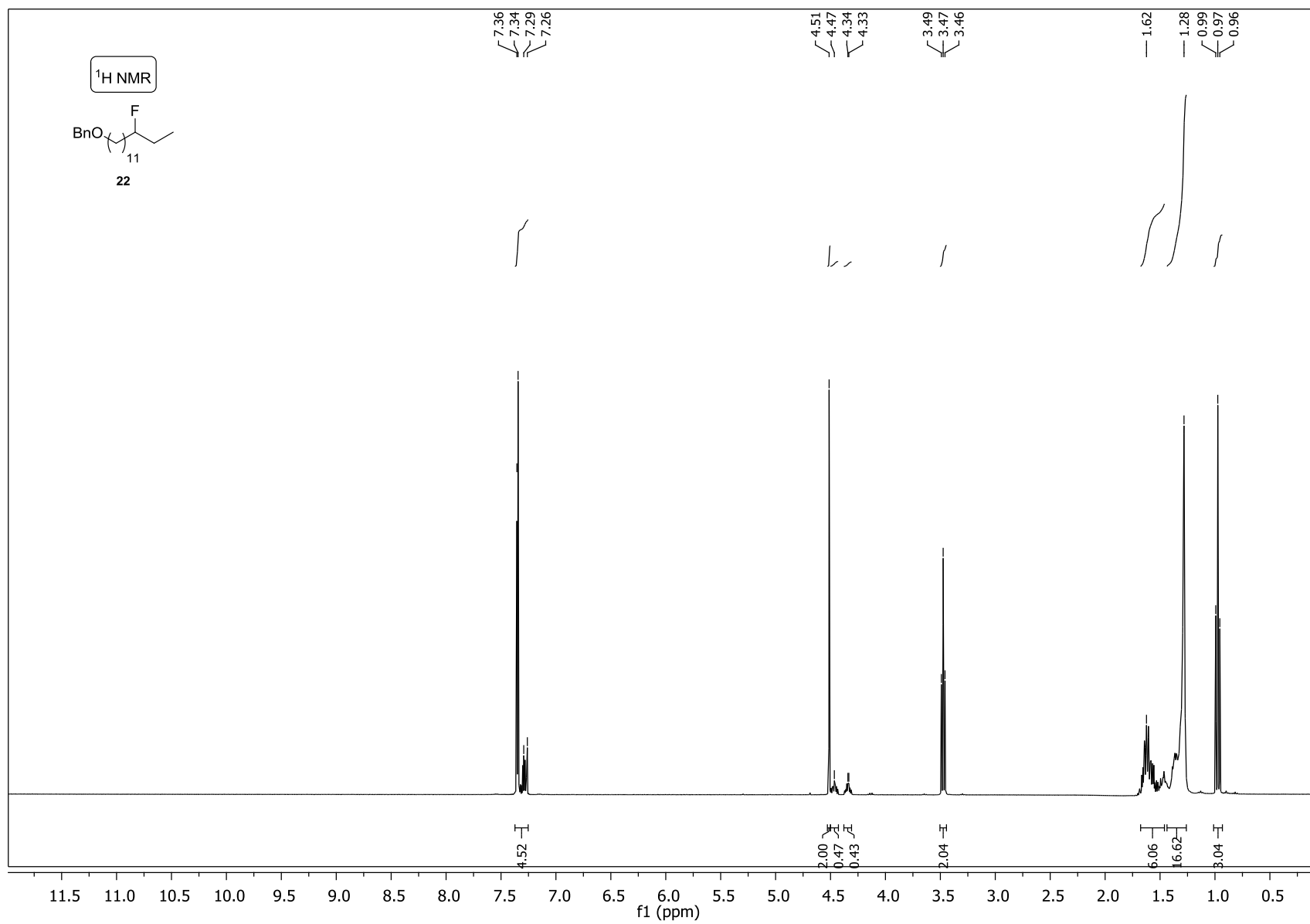


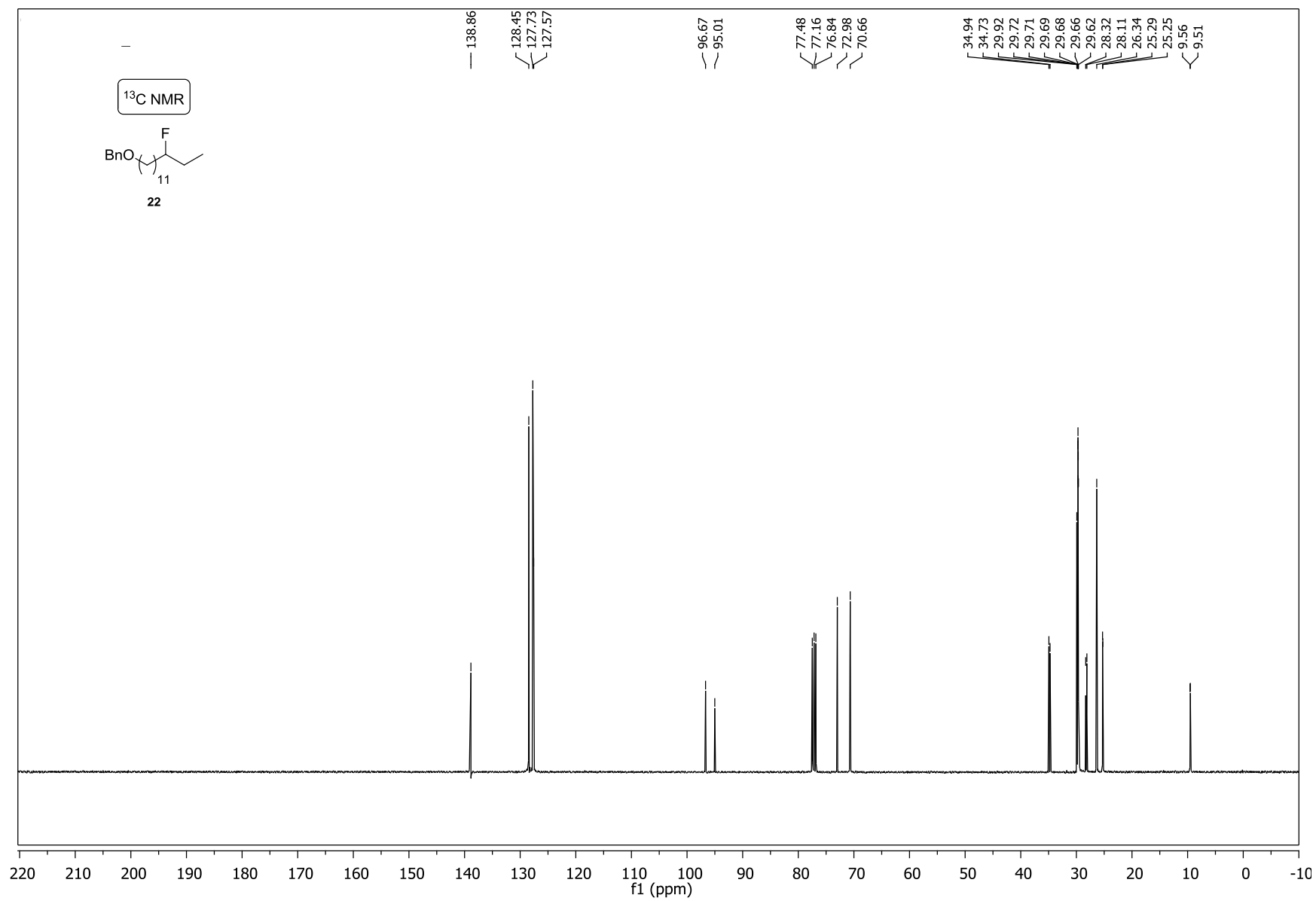


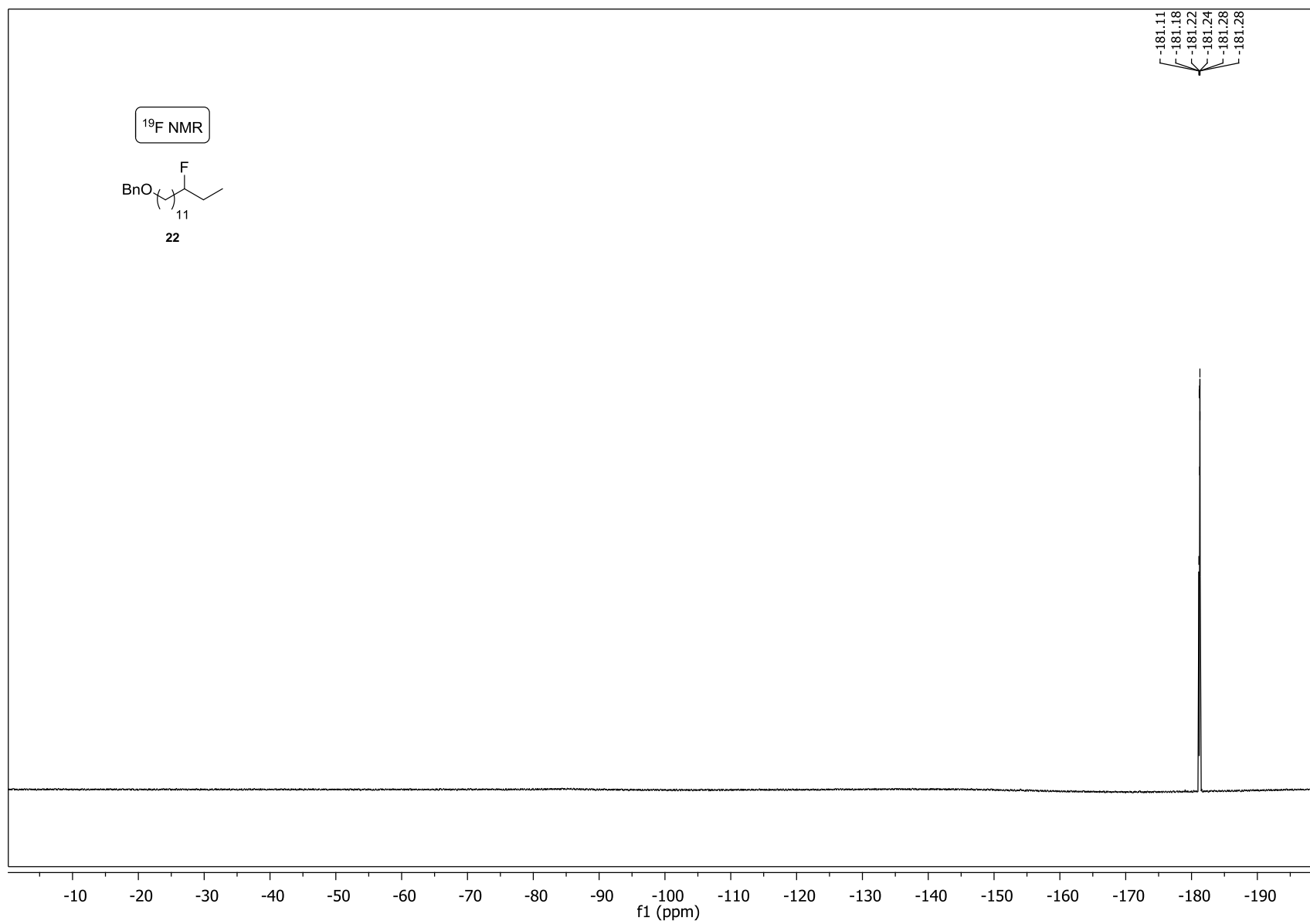


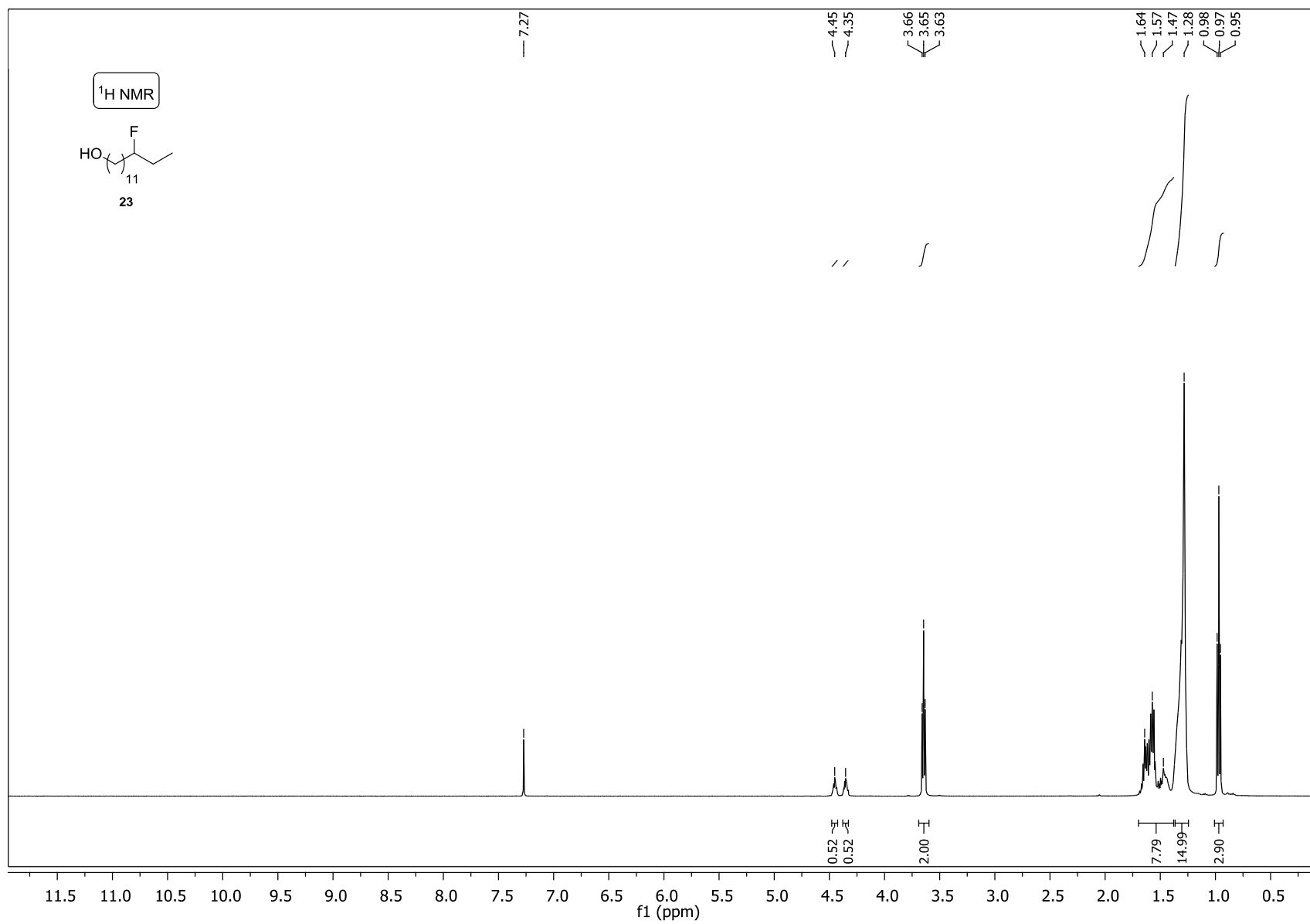


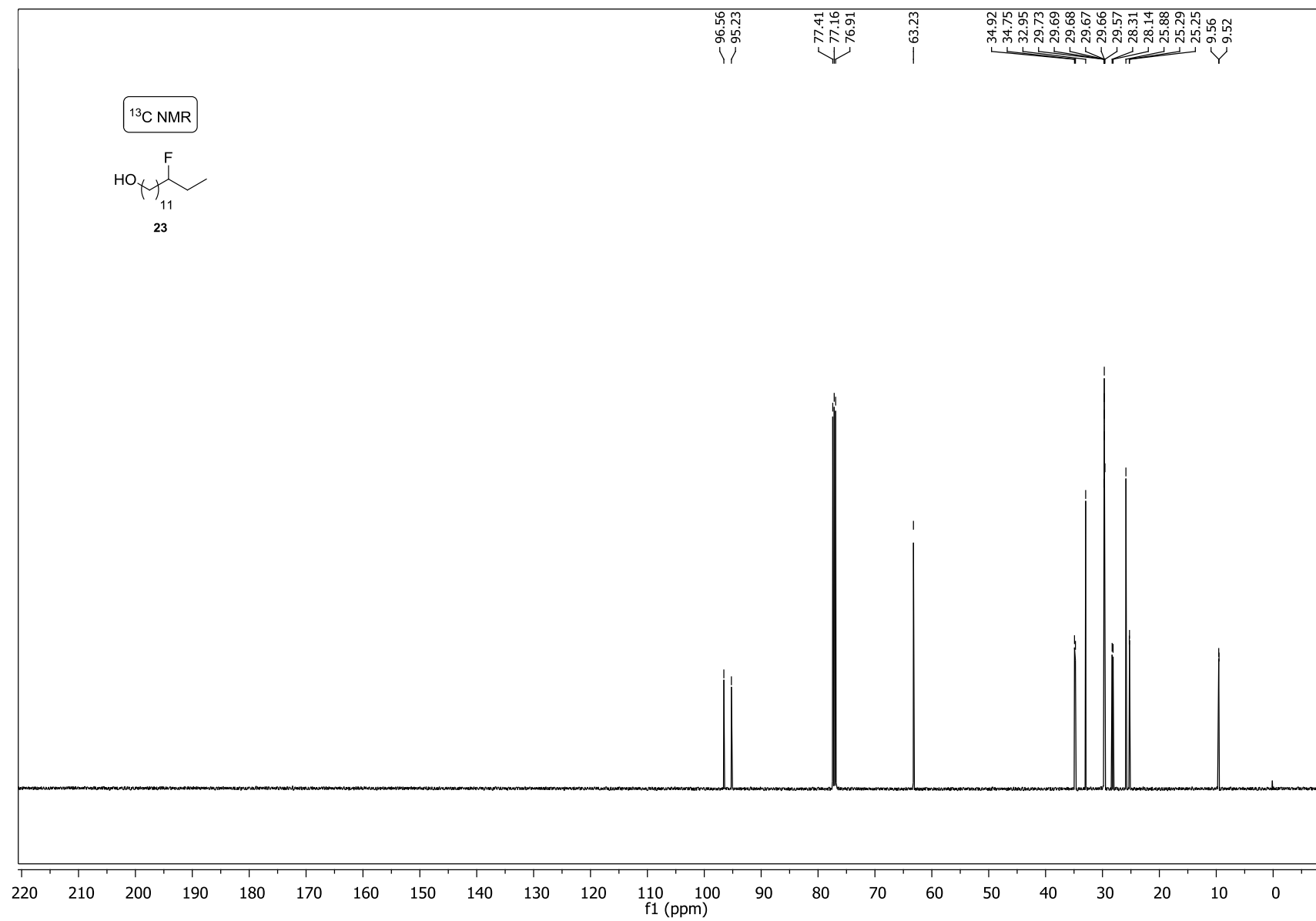


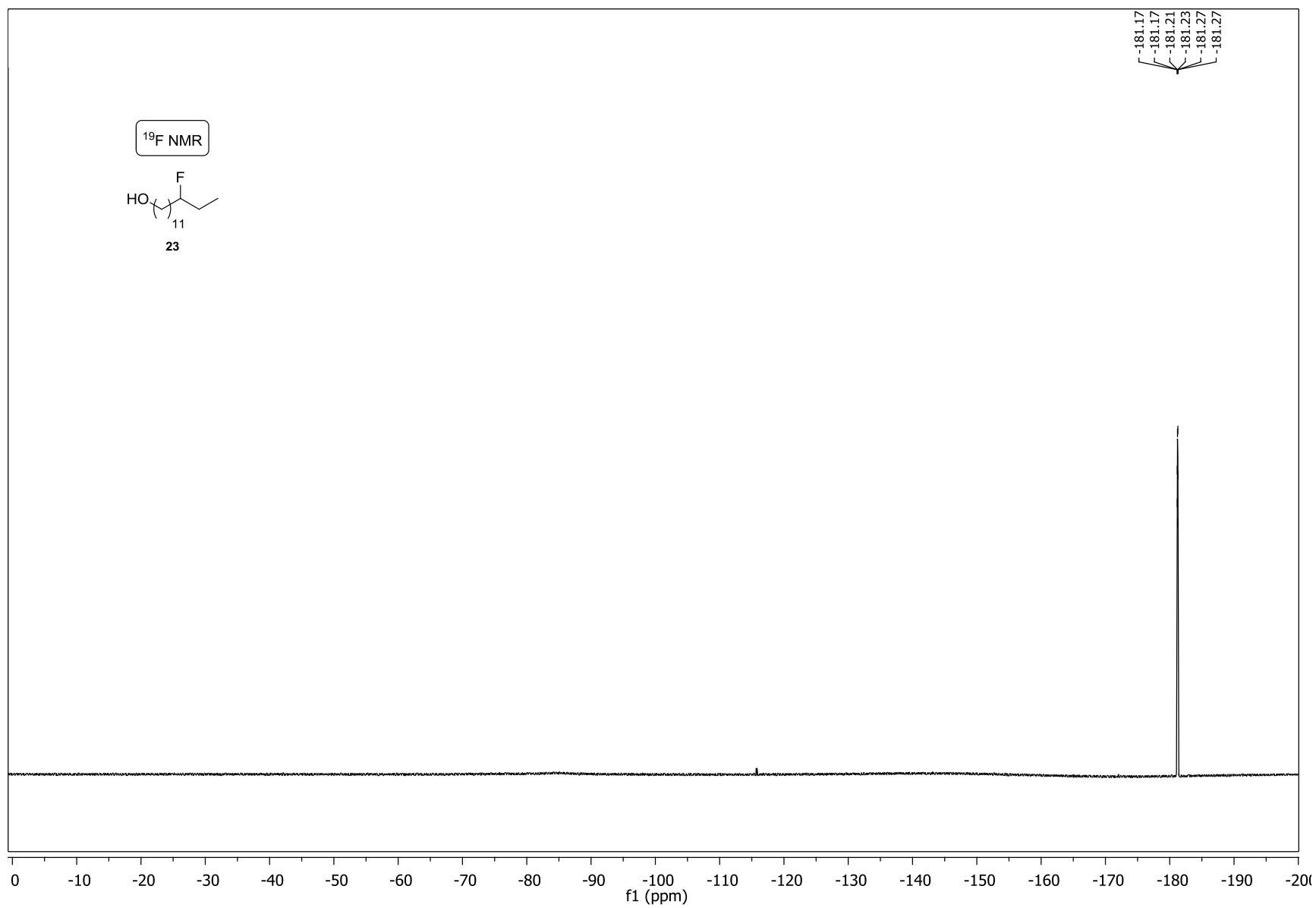


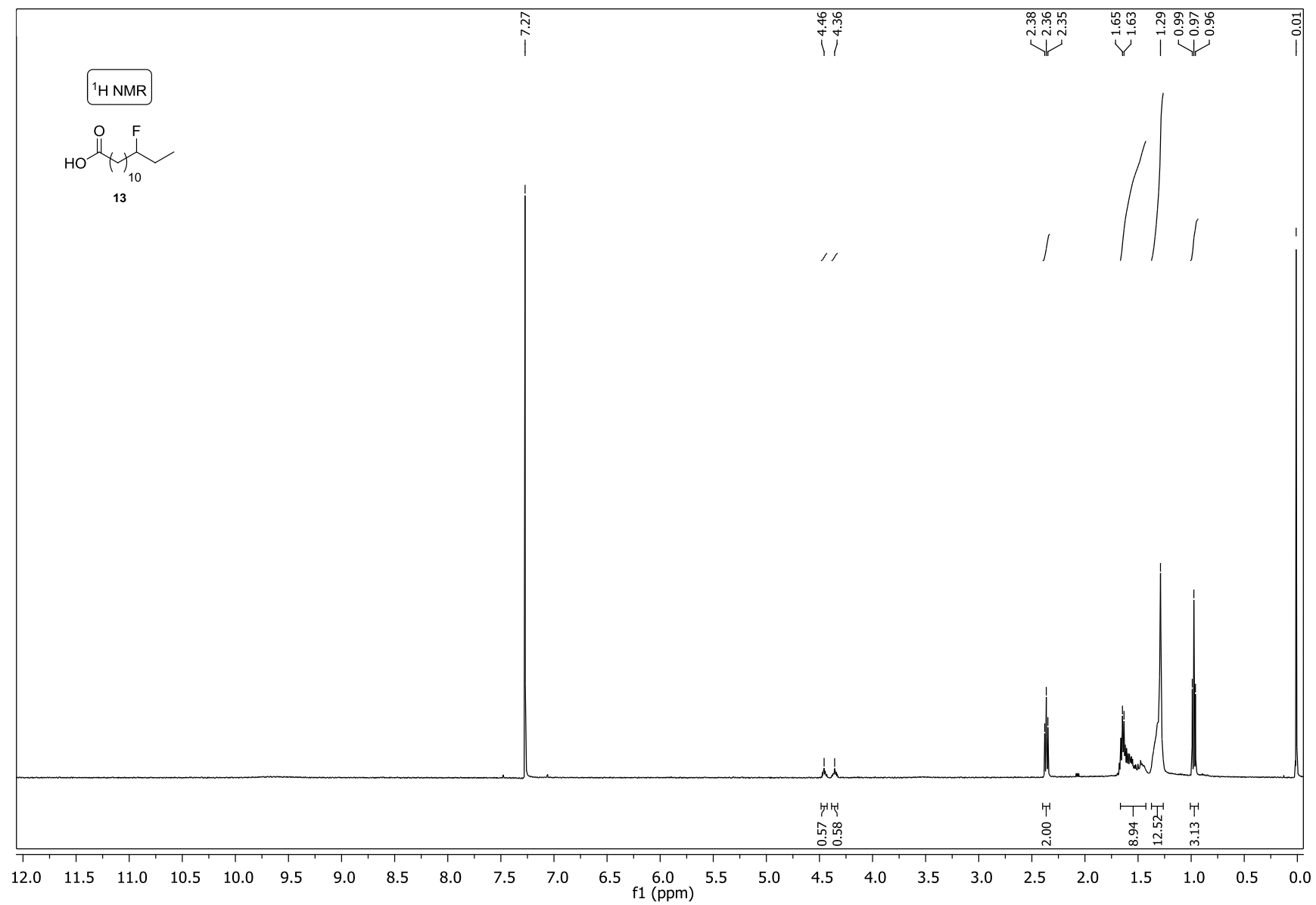


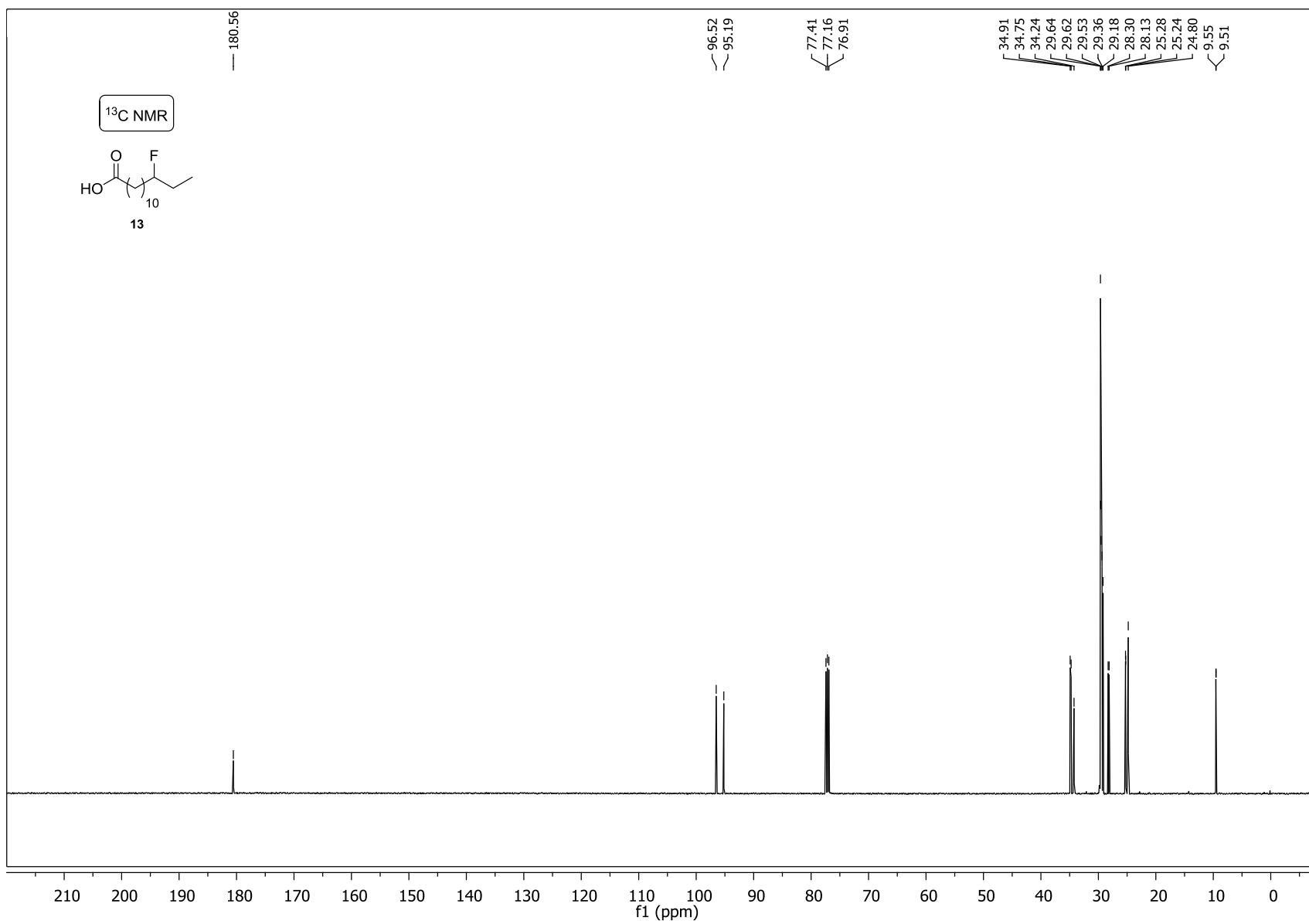


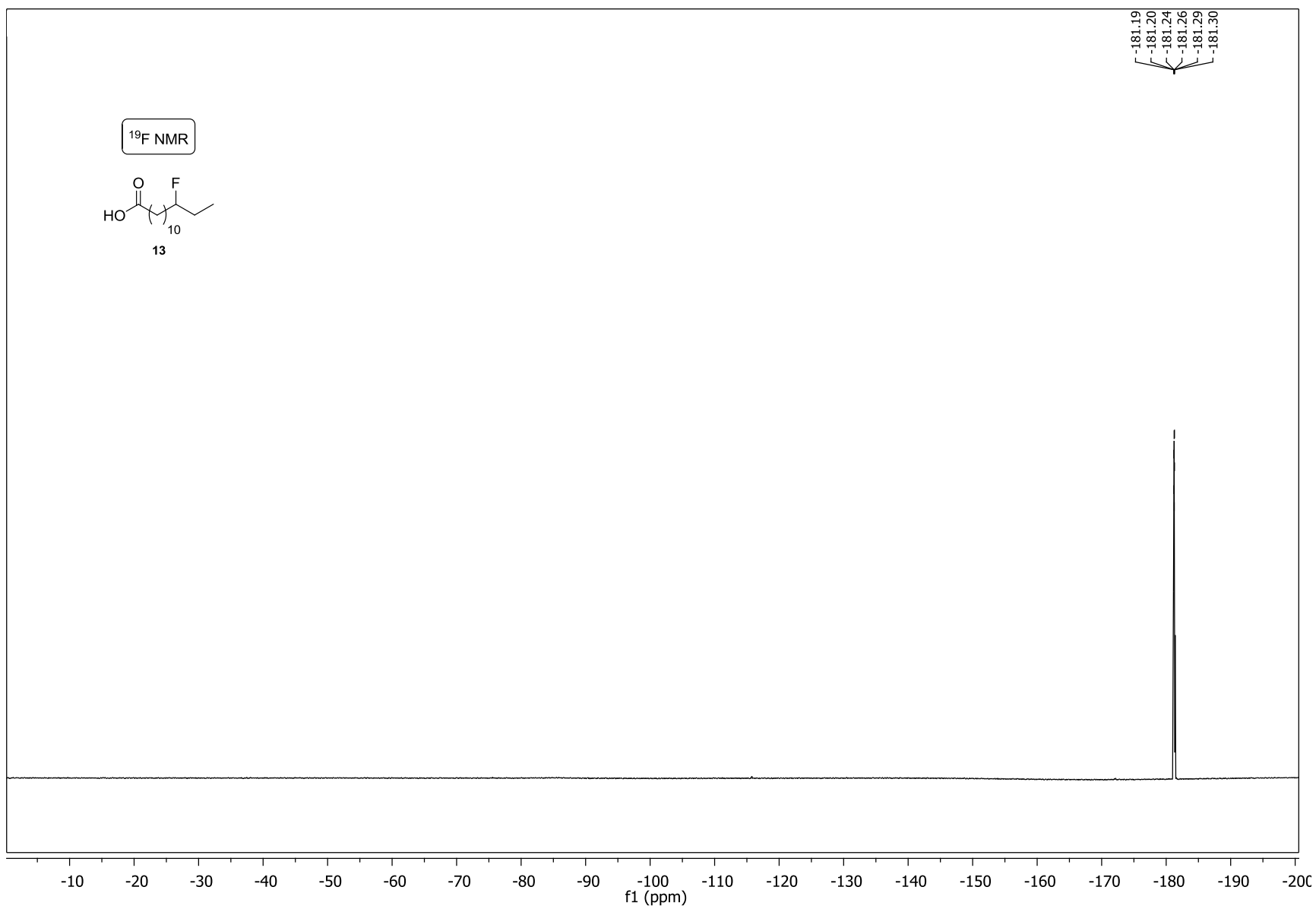


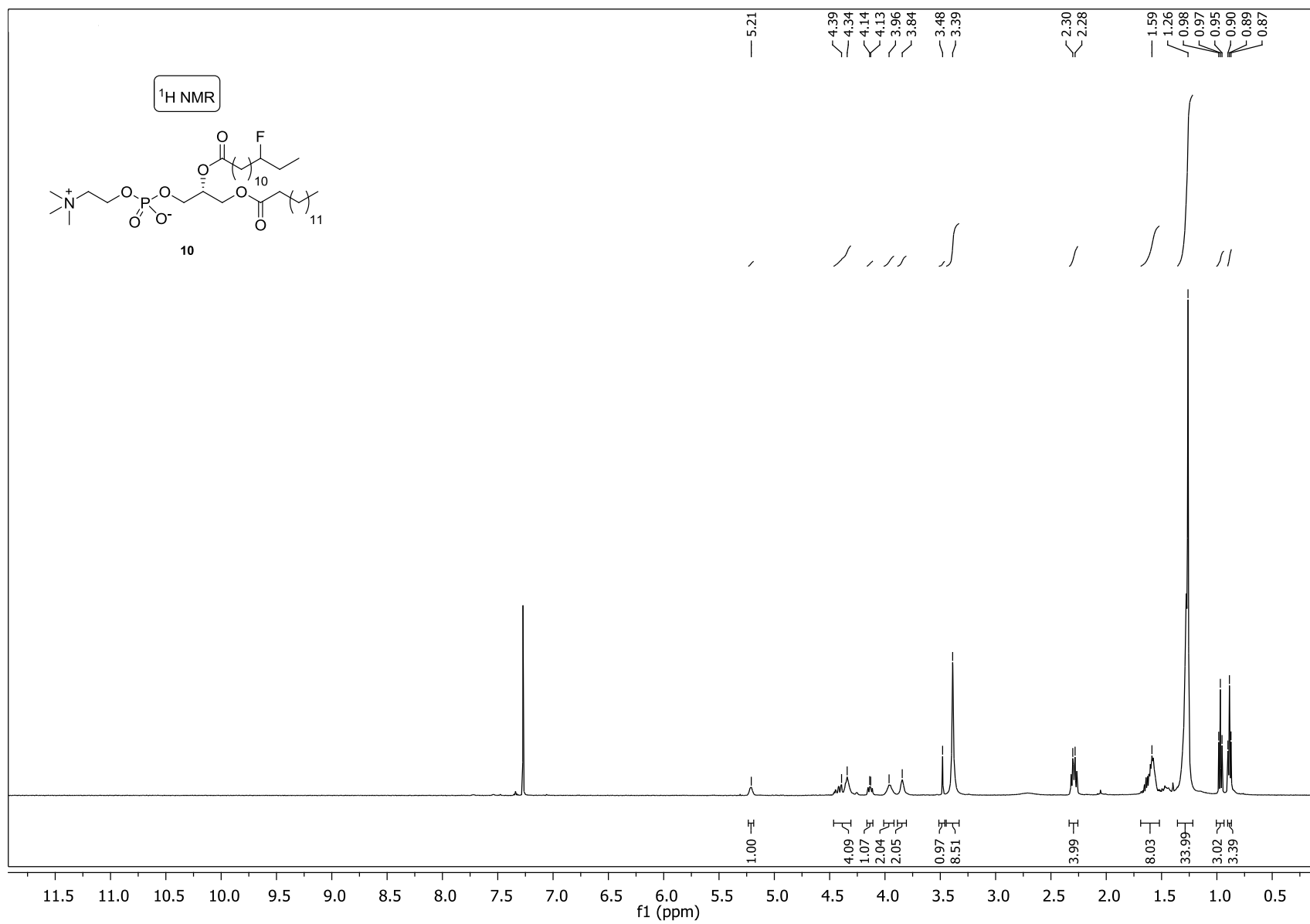


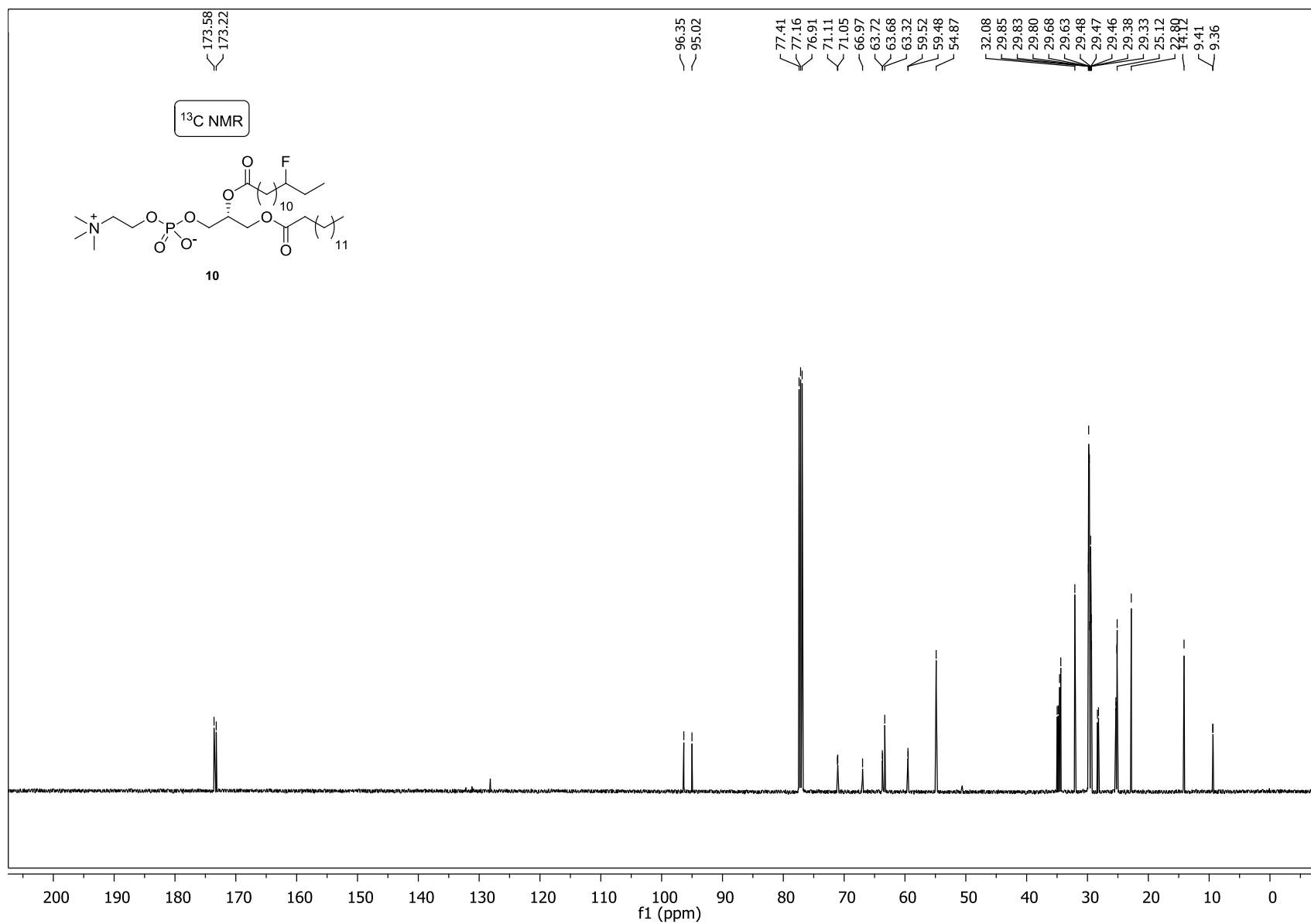


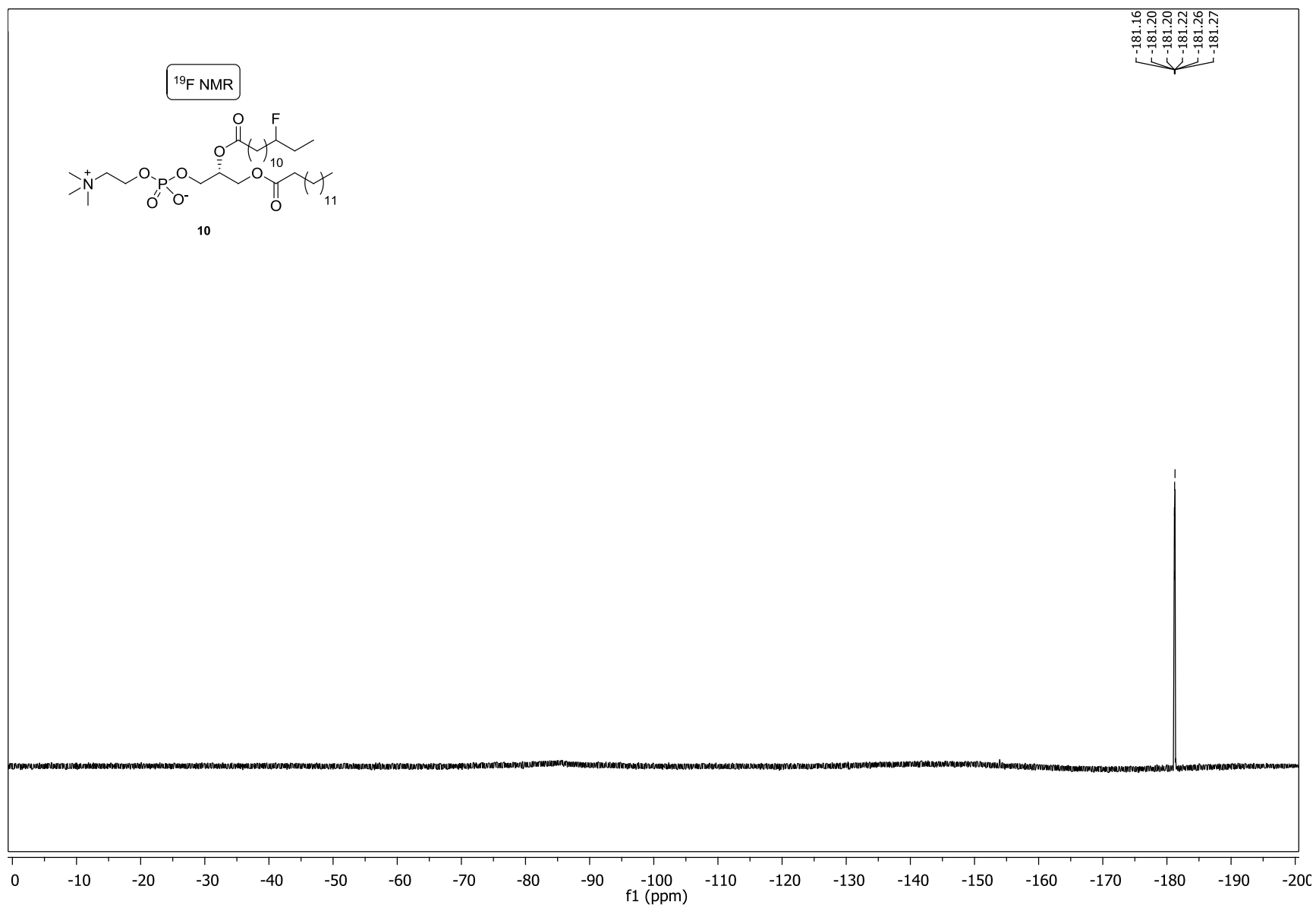


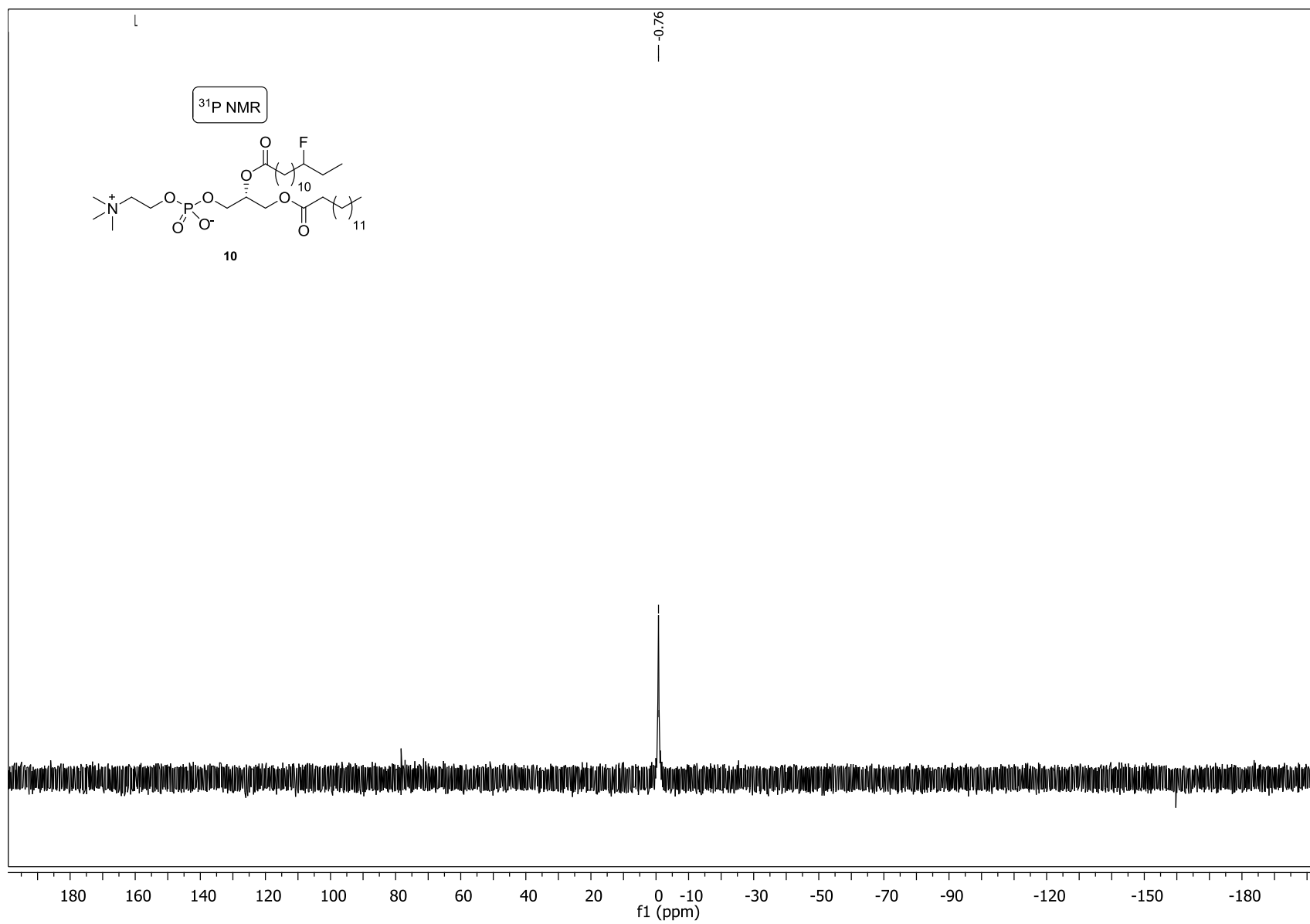


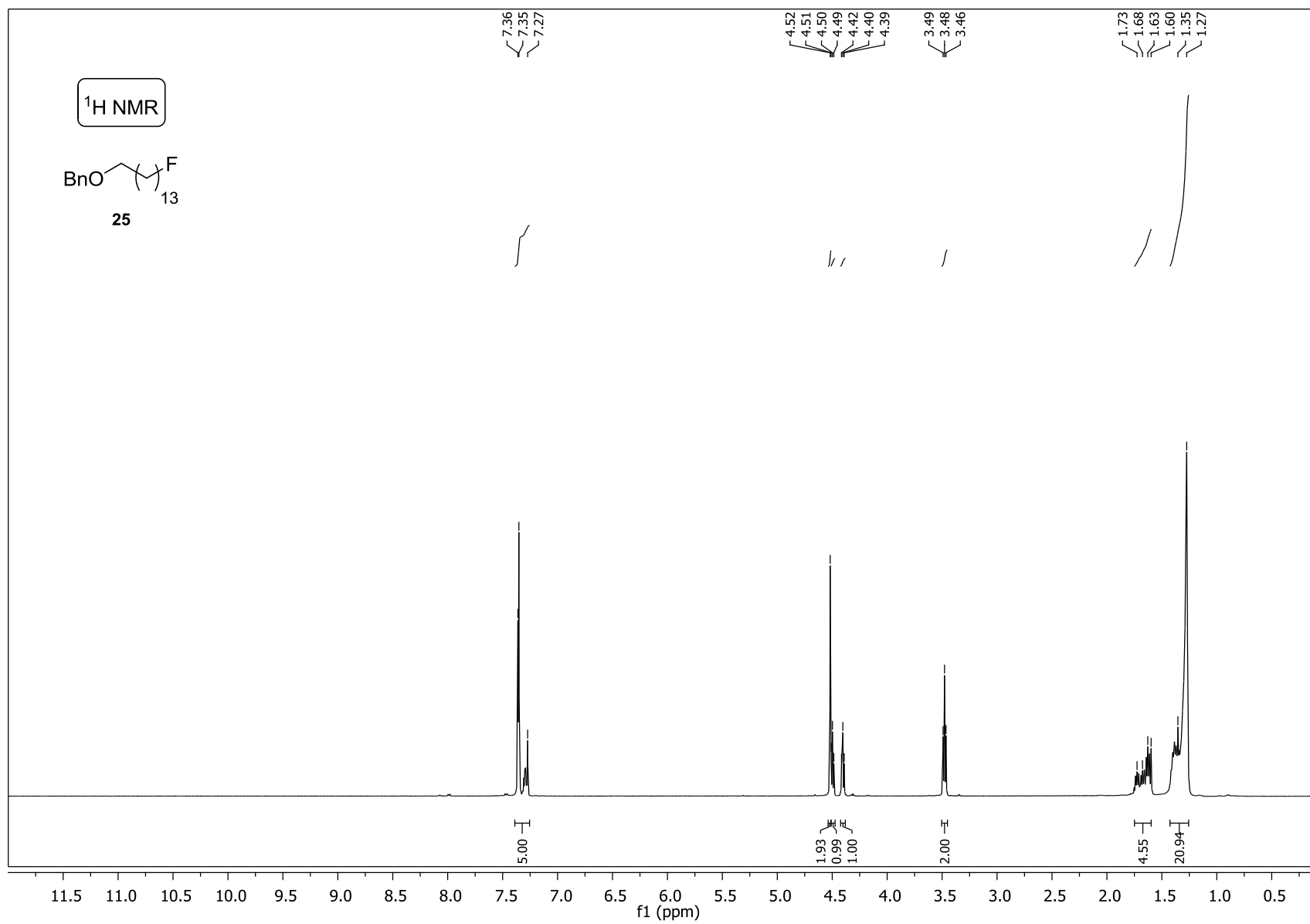


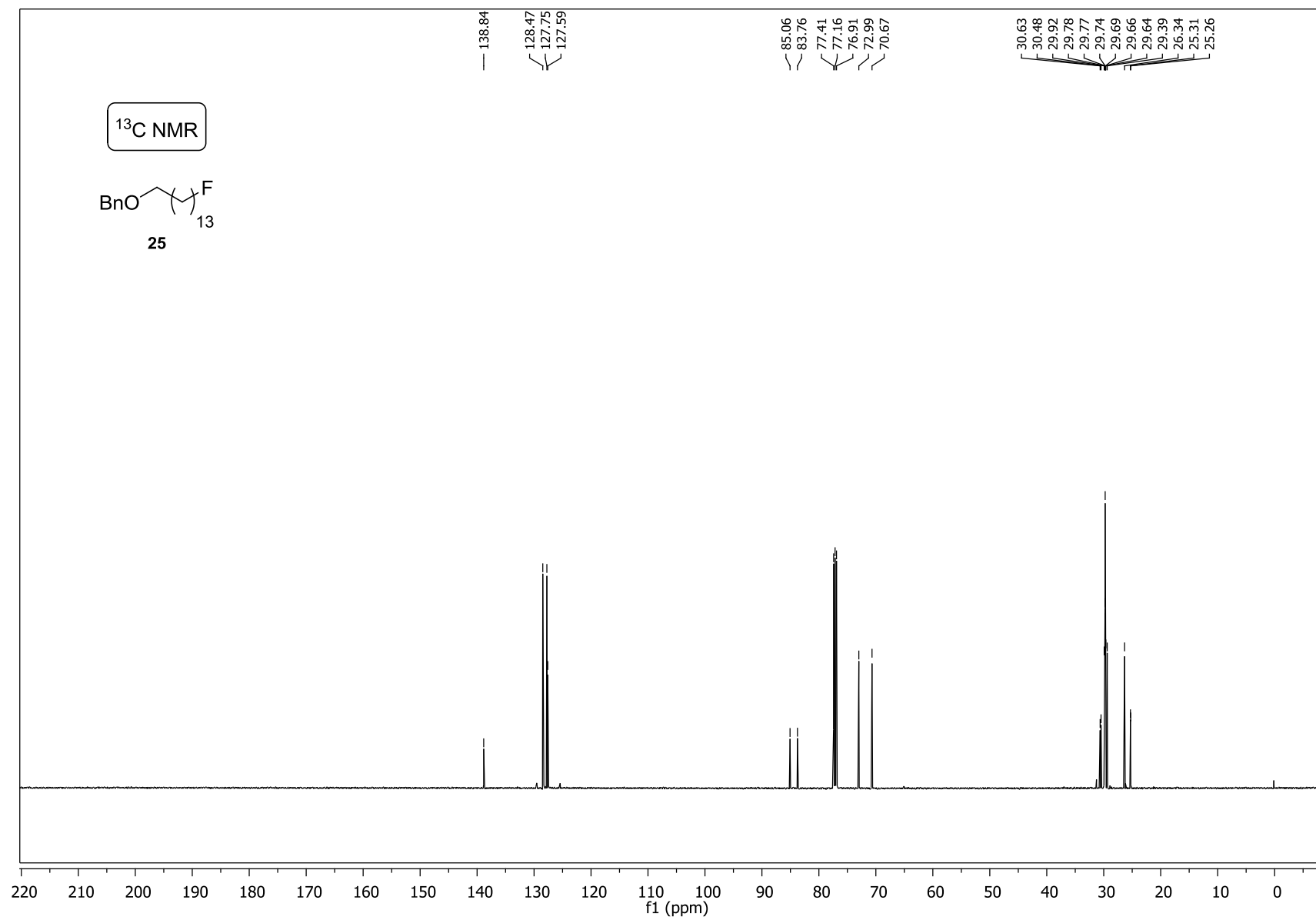


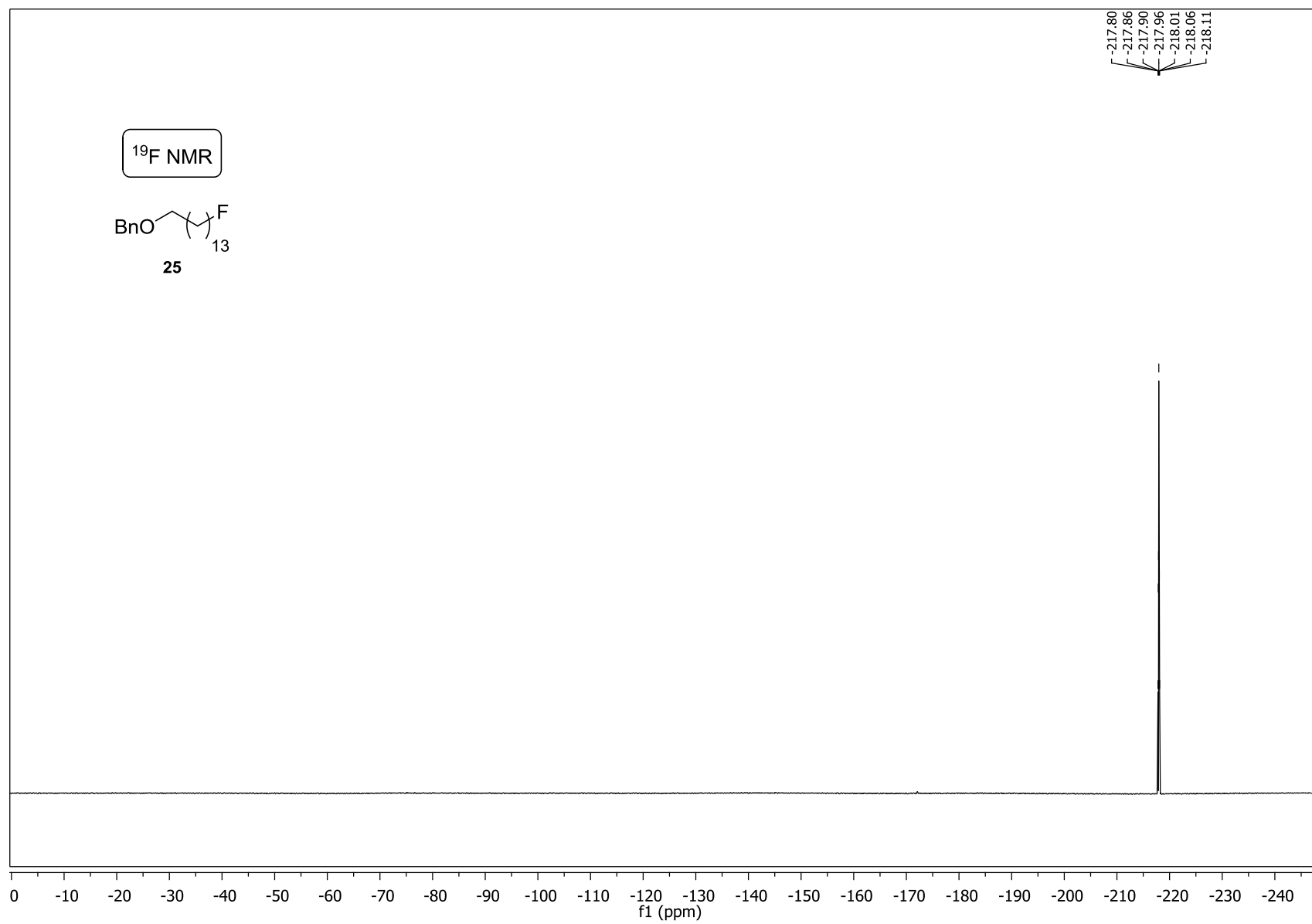


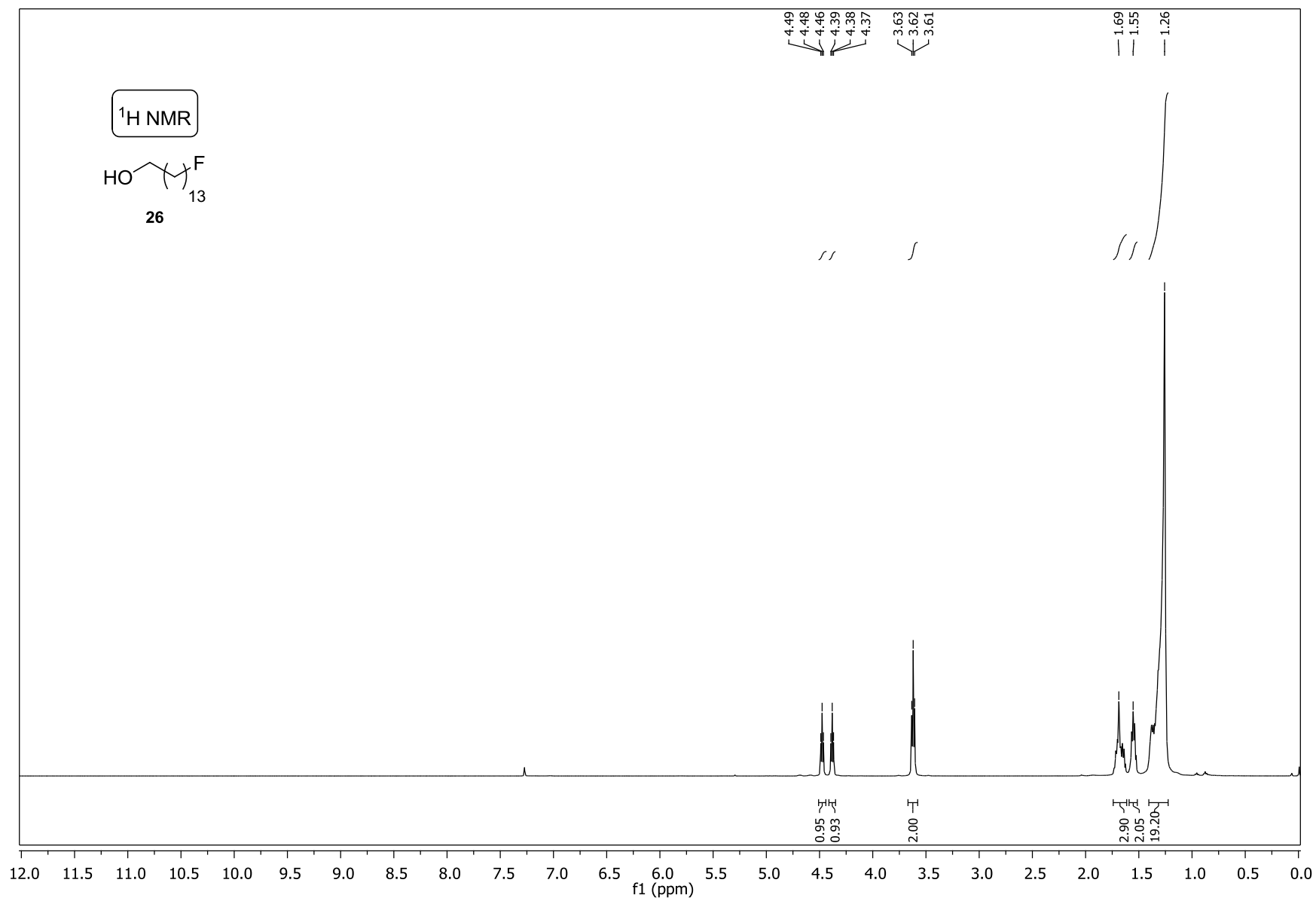


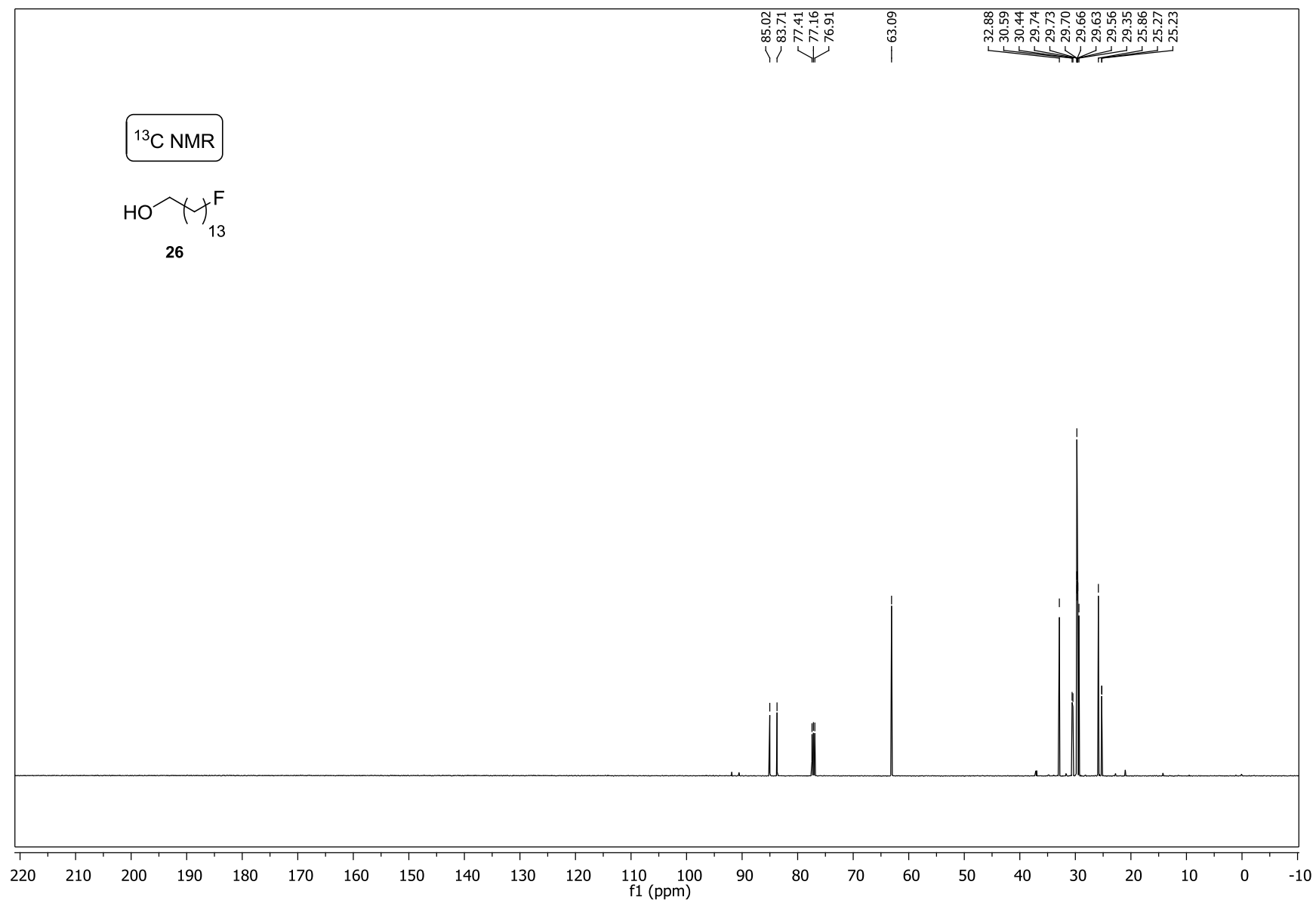


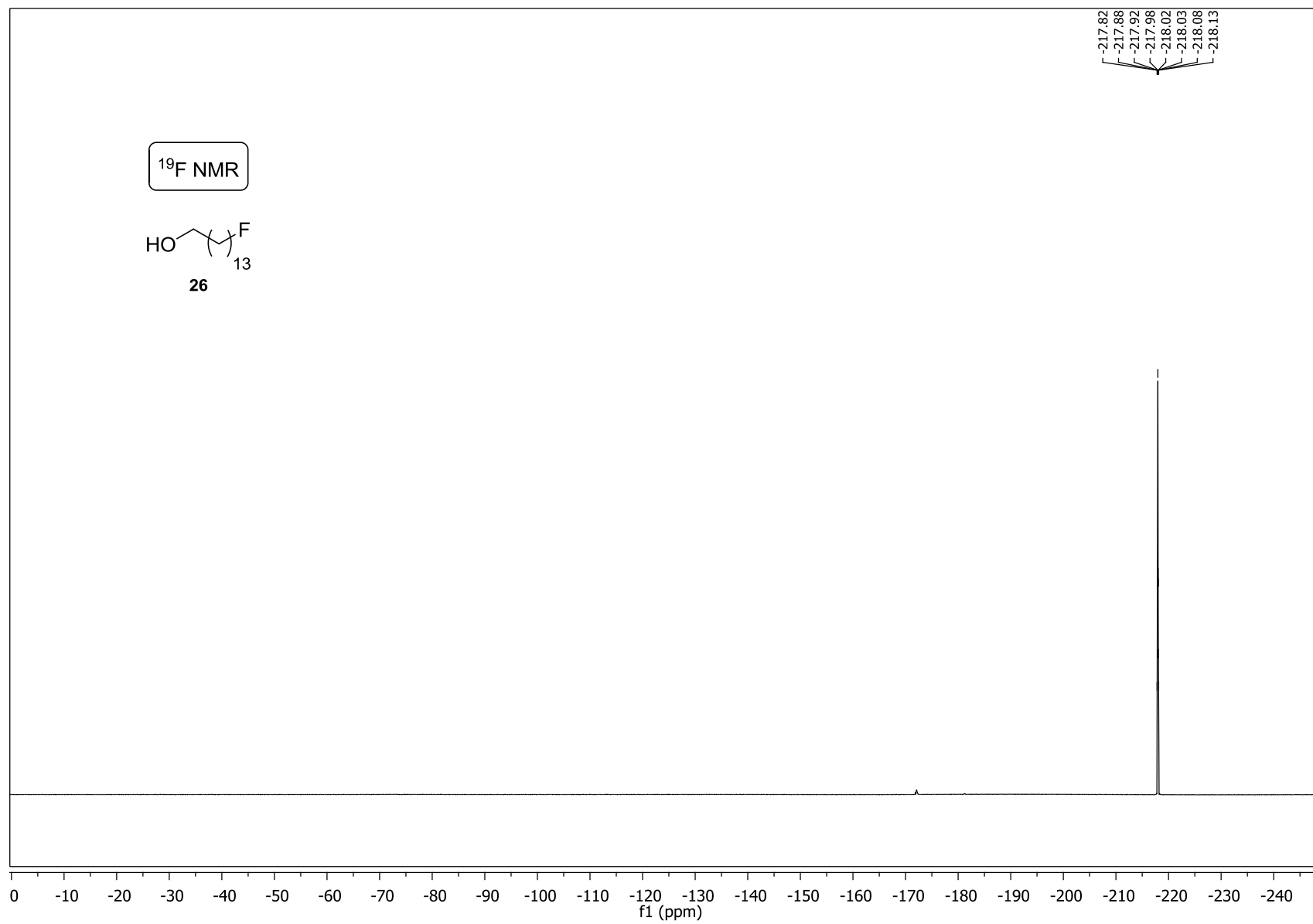


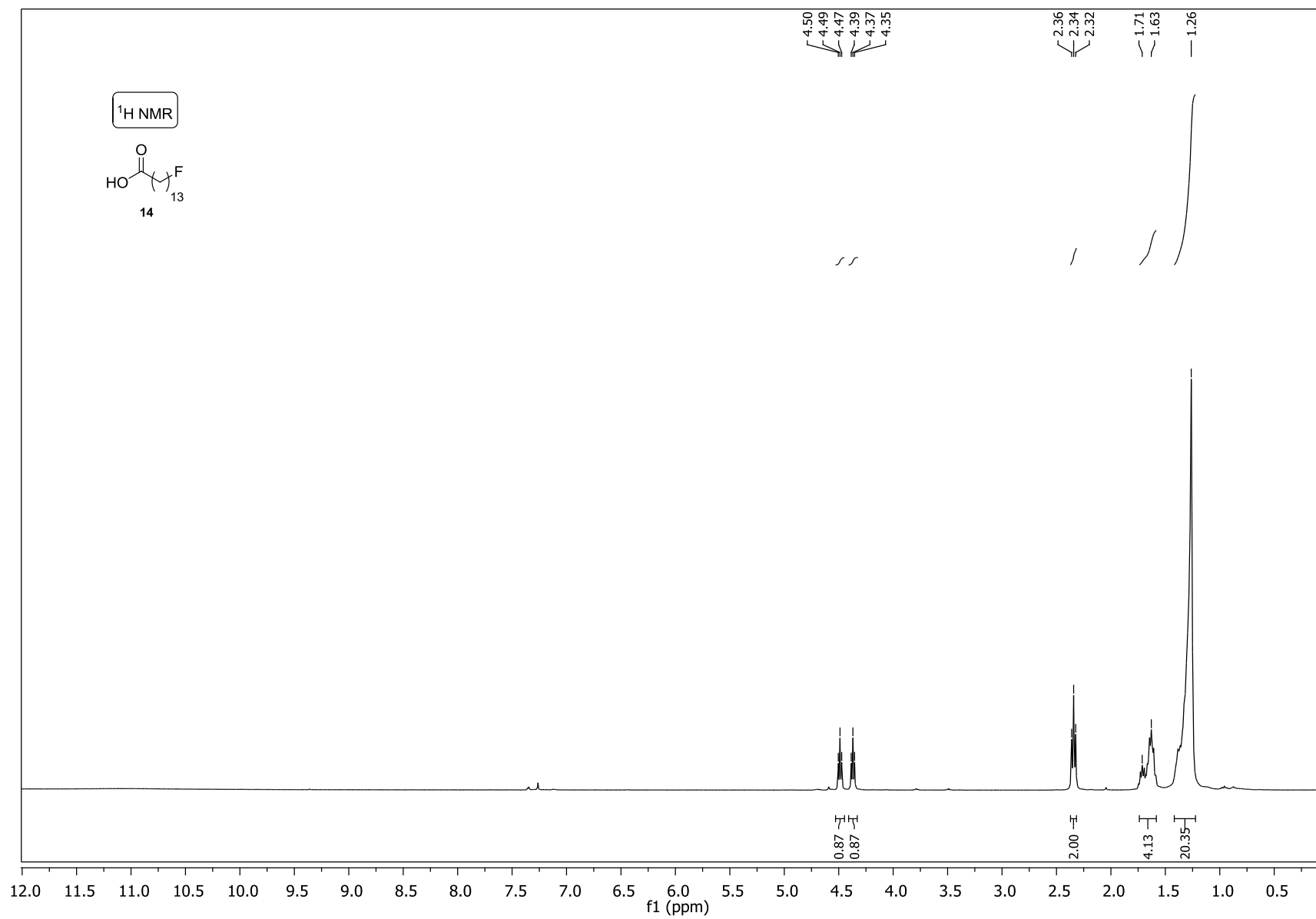


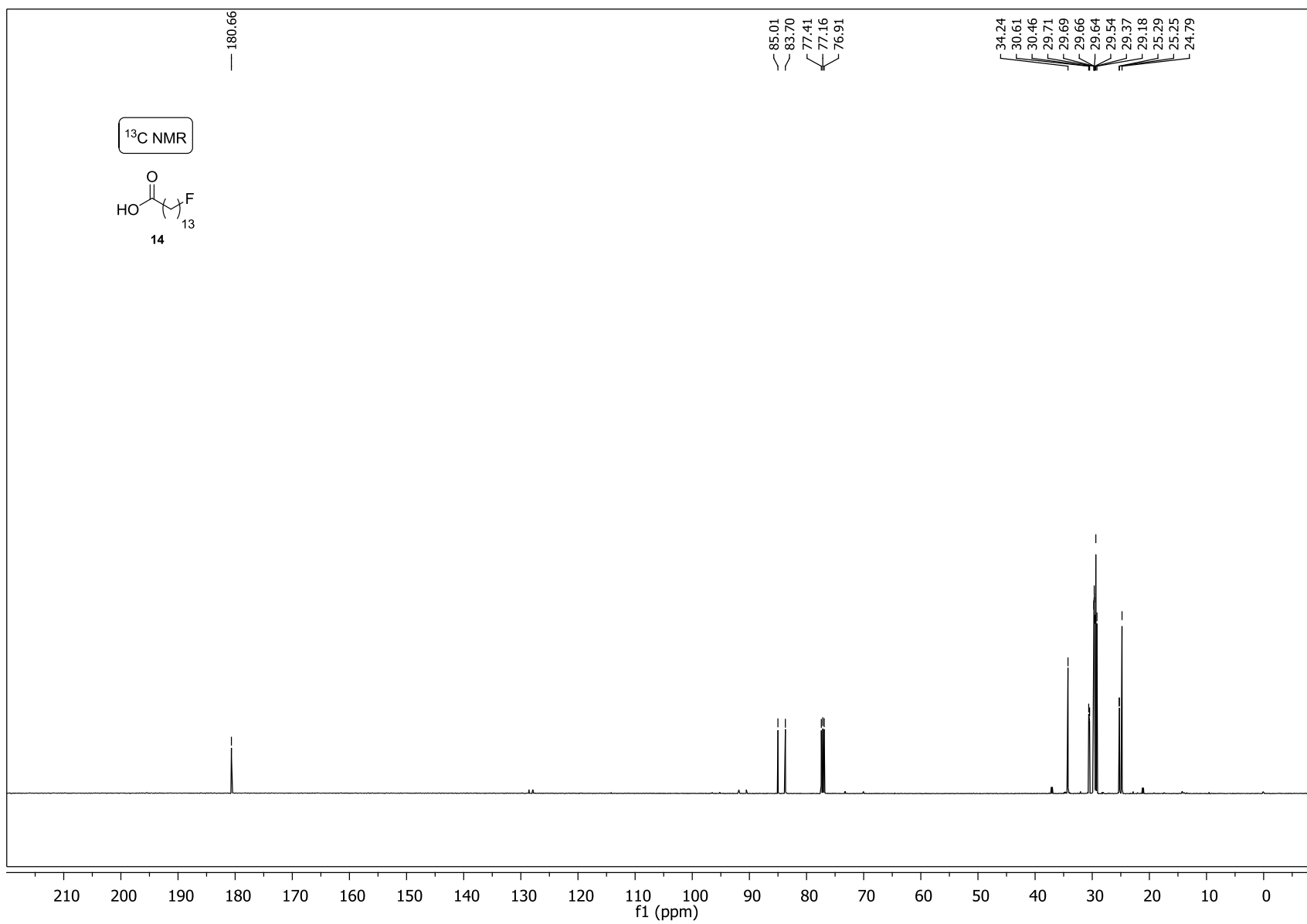


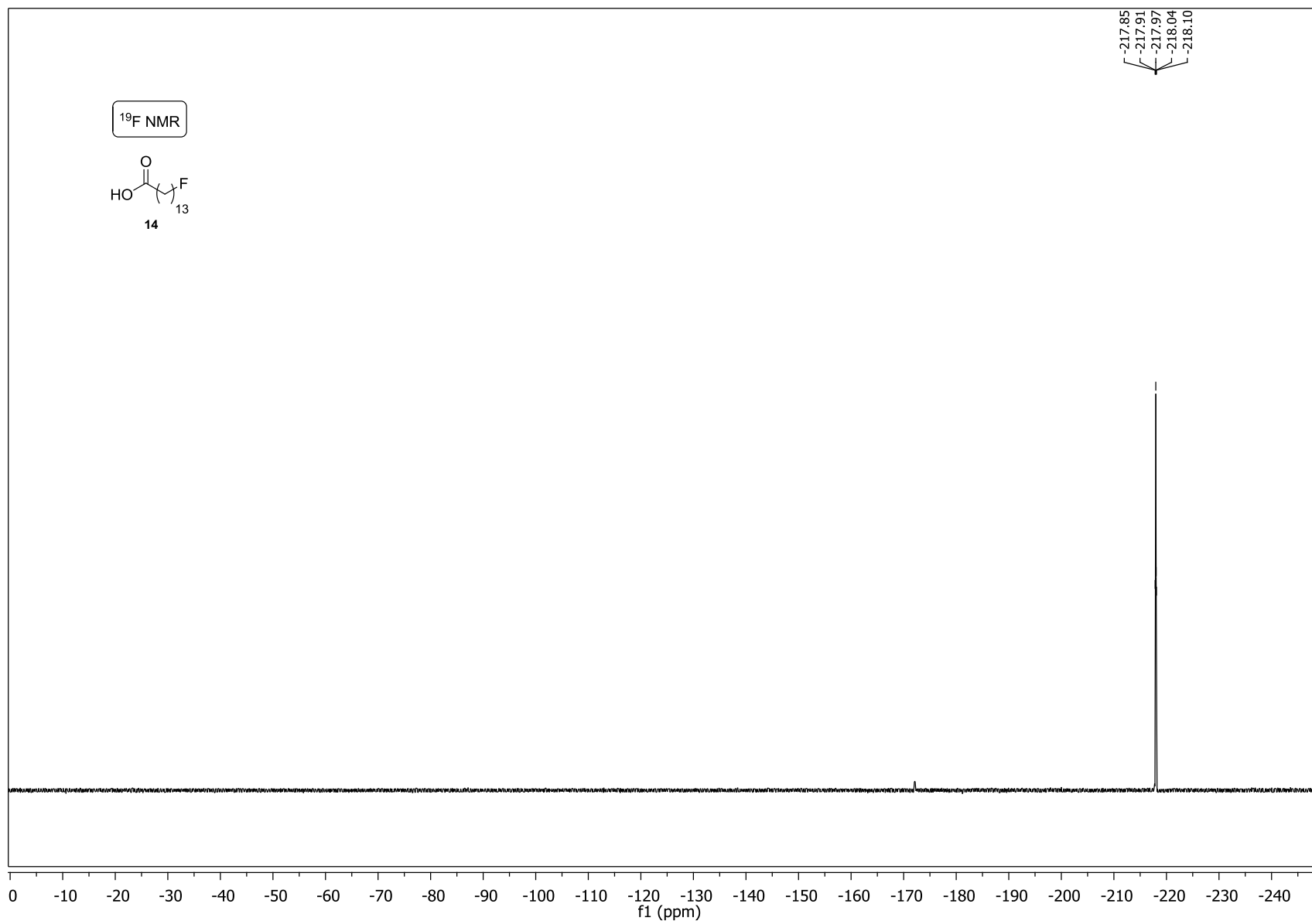


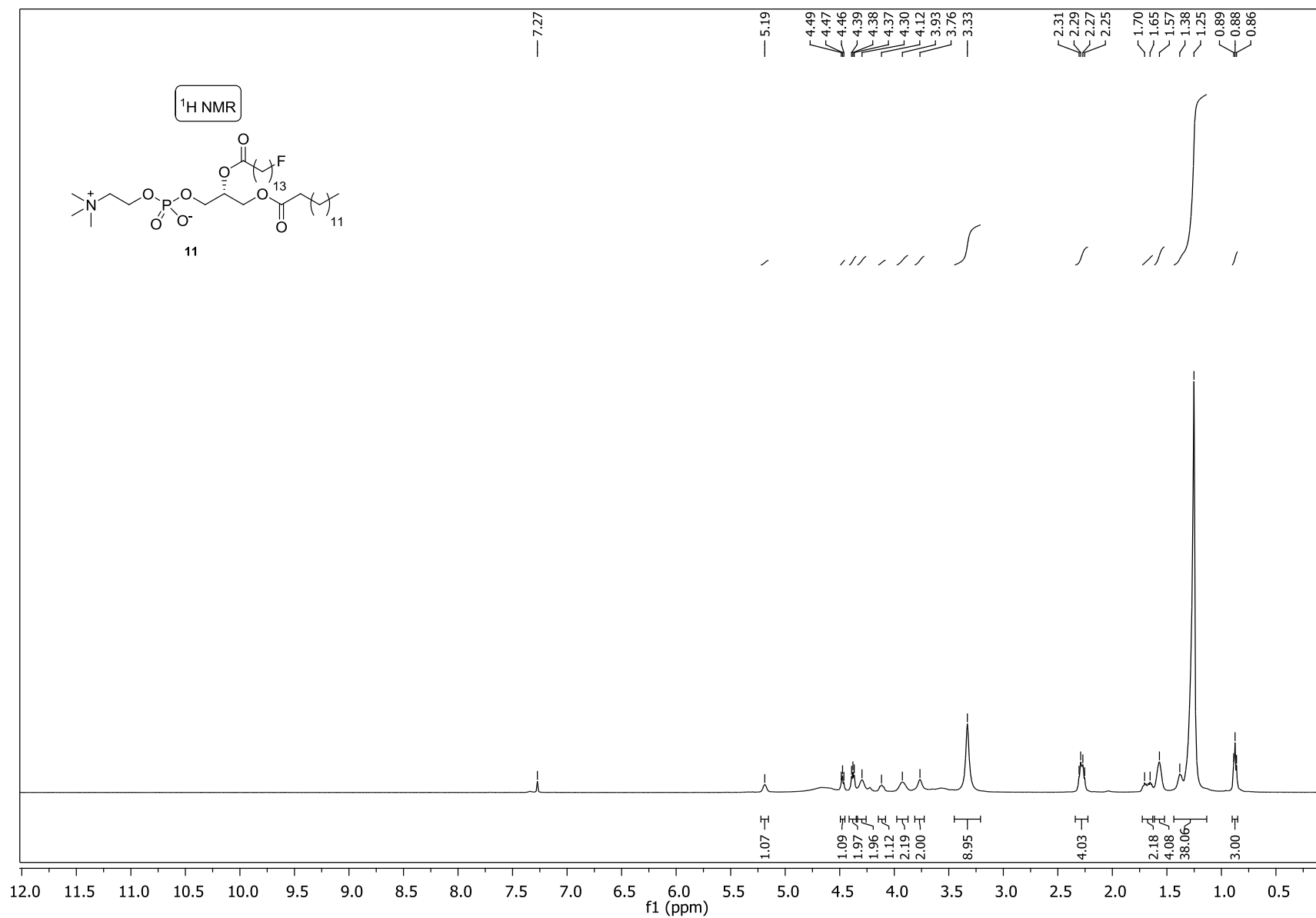


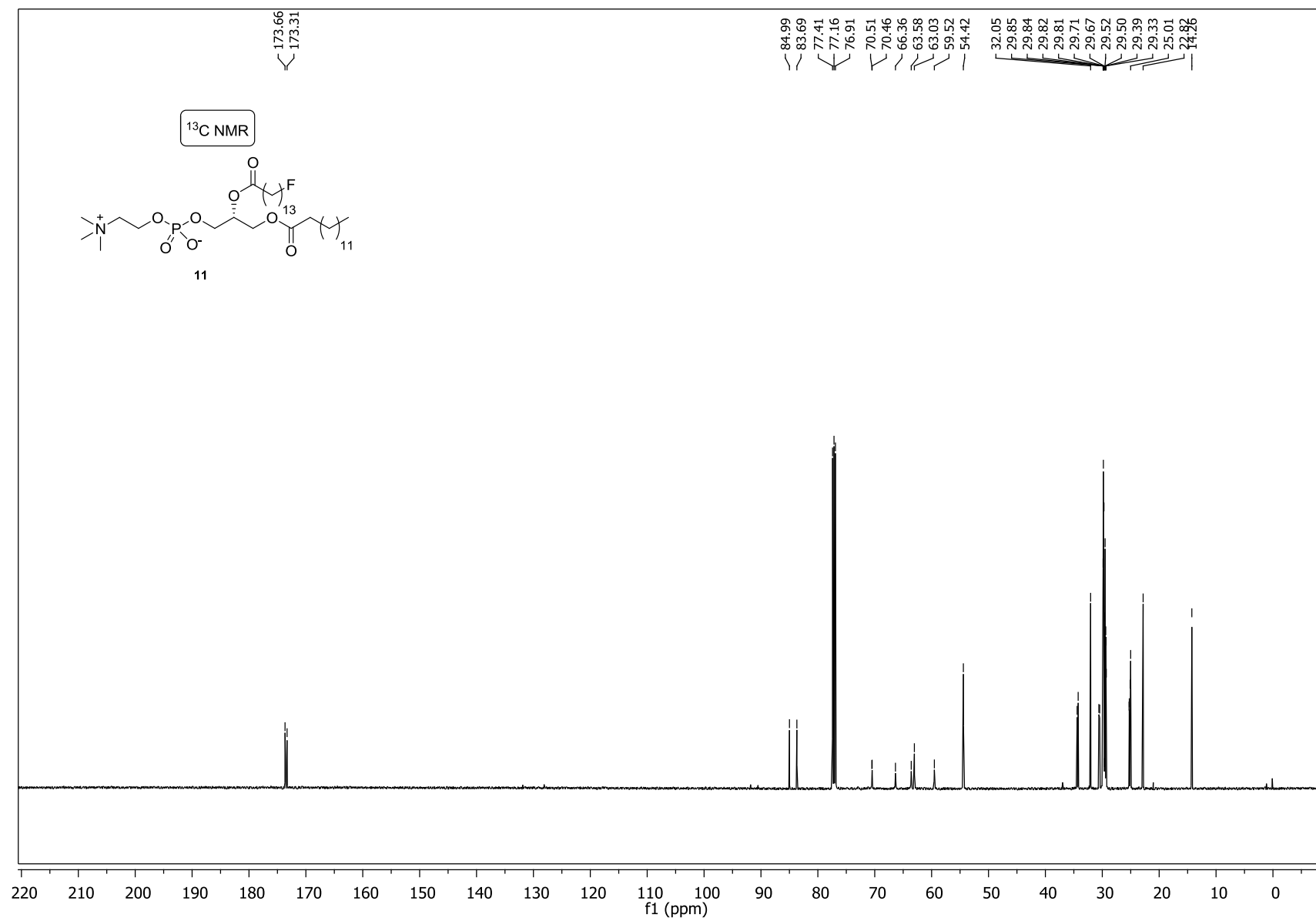


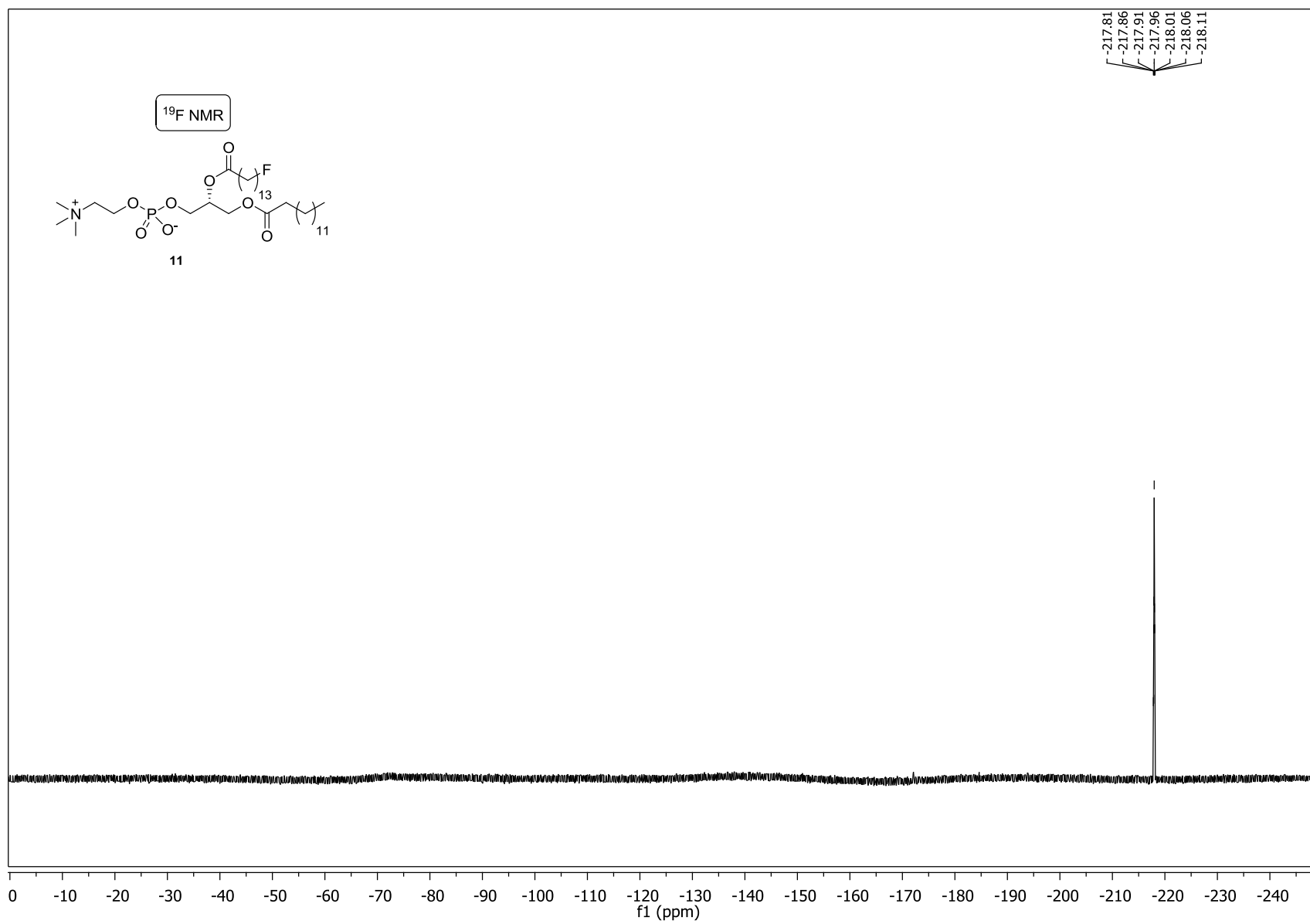


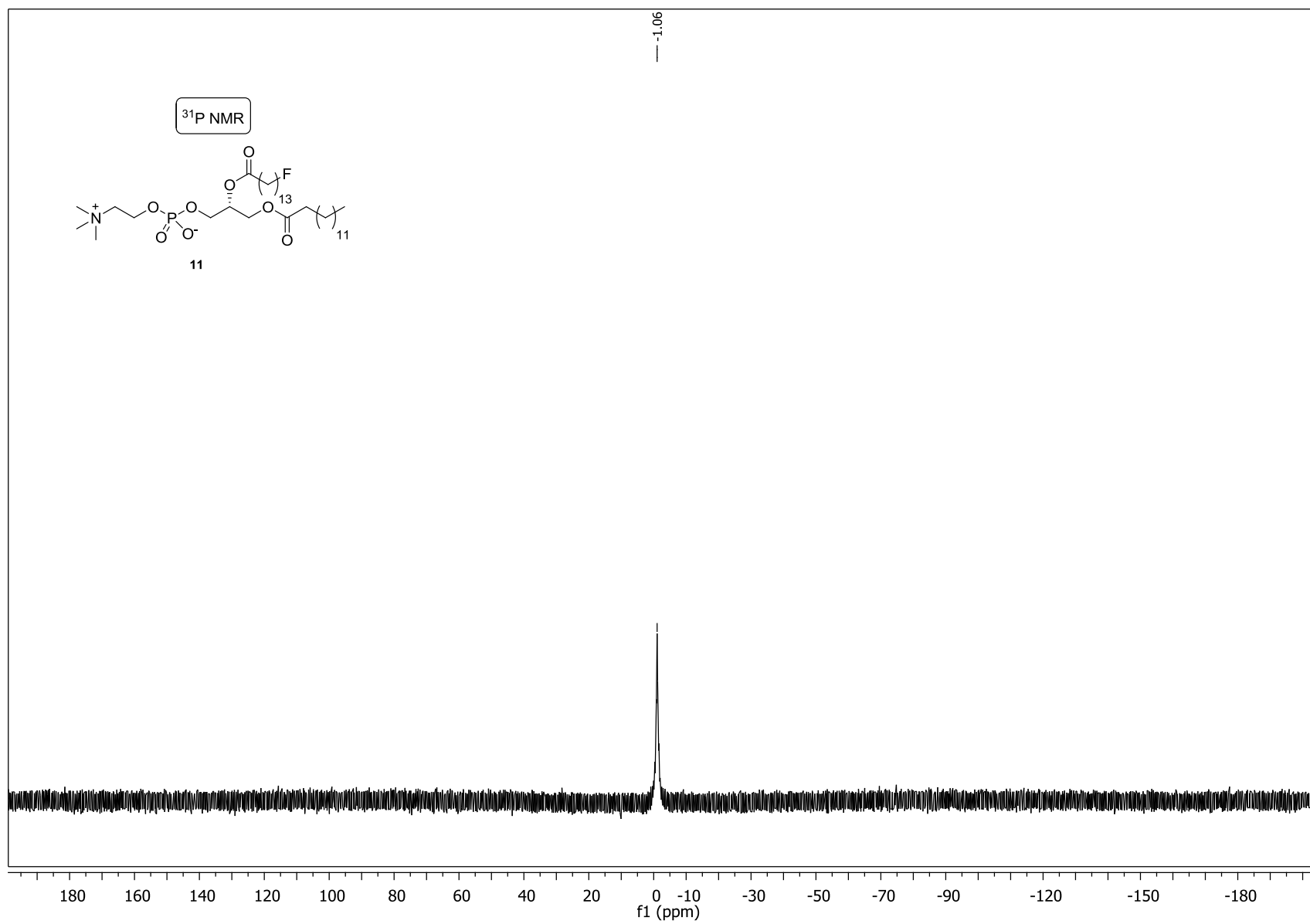












Fourier-transformed Infrared

Sample preparation

The fluorinated lipids were dissolved in chloroform at a concentration known precisely (the source of pure DMPC is also in a solution of chloroform, at a concentration of 25 mg/mL). The lipid multilamellar vesicles were prepared by measuring the necessary volume of the two lipids solutions (fluorinated DMPC and pure DMPC) and combining them before reducing the solvent volume with a stream of nitrogen or argon. The samples are placed in a lyophilizer for at least one night to remove all traces of residual organic solvent. A solution of deuterated water with 0.1 molar of HEPES buffer (2-[4-(2-hydroxyethyl)piperazin-1-yl]ethanesulfonic acid) is added to the samples to obtain a lipid concentration of 2% w/v. Then the samples underwent five thaw (heating 5 minutes at 50°C)/vortex shaking (10 seconds)/freezes (liquid N₂) cycles. After the last cycle, the sample is allowed to cool to room temperature 5 minutes. Table 1 shows the different amounts of lipids and HEPES buffer used for the measurements.

Table 1: Necessary amounts of lipids and buffer for the preparation of FTIR samples

| Fluorinated lipid ratio (%) | Fluorinated lipid mass (mg) | Pure DMPC mass (mg) | HEPES buffer volume (μL) |
|-----------------------------|-----------------------------|---------------------|--------------------------|
| 100 | 1,6 | - | 40 |
| 50 | 0,8 | 0.8 | 40 |
| 25 | 0,4 | 1.2 | 40 |
| 10 | 0,2 | 1.4 | 50 |
| 5 | 0,2 | 3.8 | 100 |
| 2,5 | 0,2 | 7.8 | 200 |

Experiments

Infrared spectra were recorded with a Nicolet Magna 550 Fourier transform spectrometer (Thermo-Nicolet, Madison, WI, USA) equipped with a narrow band mercury–cadmium–telluride (MCT) detector and a germanium-coated KBr beam splitter. Ten microliters of the sample were placed between CaF₂ windows separated by a 6μm Mylar spacer. A total of 128 interferograms were acquired with a resolution of 2 cm⁻¹ in the spectral range of 4000–650 cm⁻¹ at various temperatures ranging from 10 to 70 °C and controlled by a home-made device. The spectra were

corrected for the water vapor and CaF_2 contribution by subtraction of a reference spectrum. The data were processed with the software Grams 386 (Galactic Industries Corporation, Salem, MA, USA). The spectral is baseline-corrected using a cubic function. The methylene symmetric stretching frequency was obtained from the center of gravity calculated at the top 10% of the band. The informations obtained can then be transposed in graph using the Excel software (Microsoft, Redmond, WA). The transition temperatures and the cooperativity indices have been calculated from a custom algorithm in the Matlab program.

Figure 1 shows the type of spectra obtained with these experiments and describe the bands of interest; the CH_2 stretching bands.

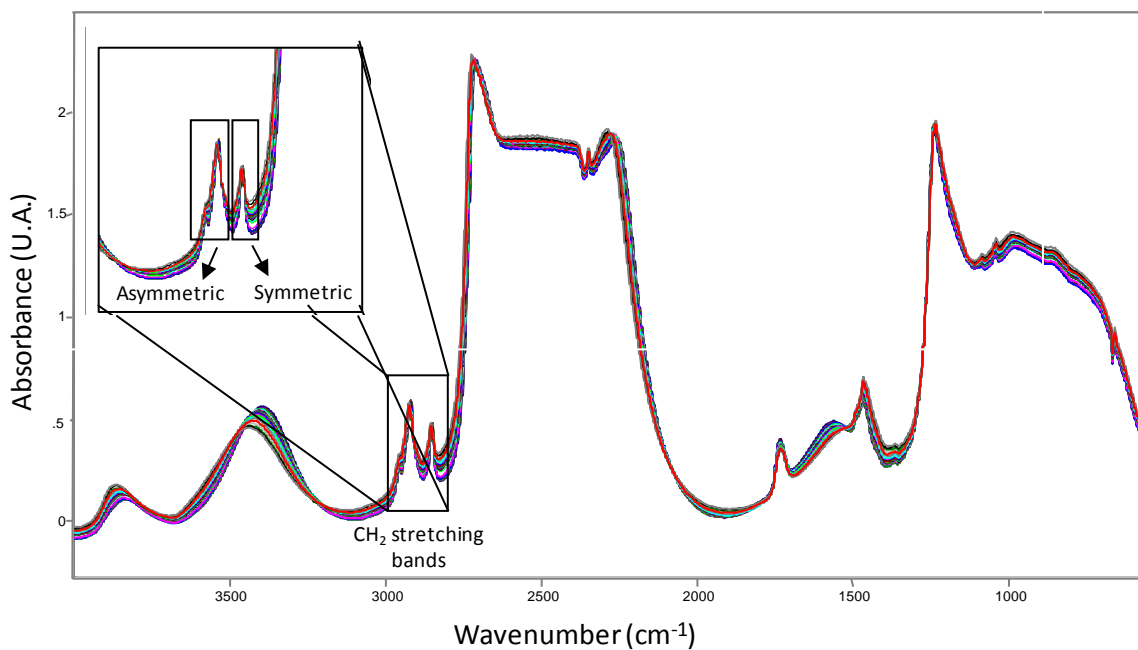


Fig.1 FTIR spectra of 7F-DMPC (10 to 70 °C)