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Supporting Information for

Cooperative Hybridization of γPNA Miniprobes to a Repeating Sequence Motif and Application to Telomere Analysis

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Figure S1. UV melting curves for γP_{12} with Telo-2 (black squares) or Telo-4 (red circles) and P_{18} with Telo-3 (blue triangles). Samples contained 2 μ M γP_{12} and either 2 μ M Telo-2 or 1 μ M Telo-4 or 1.3 μ M P_{18} and 1.3 μ M Telo-3. Buffer contained 10 mM Tris-HCI (pH = 7), 0.1 mM EDTA and 100 mM KCI. Important results include (a) higher melting temperature for γP_{12} + Telo-2 versus P_{18} + Telo-3, illustrating higher affinity of γ PNA versus PNA, and (b) higher melting temperature for γP_{12} + Telo-4 versus γP_{12} + Telo-2, illustrating cooperativity of hybridization of γP_{12} + Telo-2 to two adjacent sites on the same DNA complement versus hybridization to two separate DNA oligonucleotides.



Figure S2. UV melting curves for $\gamma P_{6-clamp}$ with **Telo-1-4**. Samples contained 4 μ M $\gamma P_{6-clamp}$ and either 4 μ M **Telo-1**, 2 μ M **Telo-2**, 1.3 μ M **Telo-3** or 1 μ M **Telo-4**. Buffer contained 10 mM Tris-HCI (pH = 7), 0.1 mM EDTA and 100 mM KCI. Significant results include the substantial increase in melting temperature as the number of complementary repeats in the DNA oligonucleotide increases, again illustrating the cooperativity of hybridization to adjacent sites on the DNA complement.



Figure S3. UV melting curve recorded at 295 nM for **Telo-4** in the presence of 100 mM KCI. The hypochromic transition is indicative of a G-quadruplex structure.



Figure S4. SPR sensorgrams for association/dissociation of γ PNA miniprobes with **Telo-4** in the presence of 100 mM KCI (red) or LiCI (black). The large increases in association in the presence of LiCI are consistent with the formation of a DNA quadruplex structure in KCI, which inhibits hybridization of the γ PNAs. Raw data for each γ PNA were normalized to the maximum response unit (RU) signal observed in LiCI.



Figure S5. Telomere staining of Jurkat metaphase chromosome spreads using P_{18} (Left) or γP_{12} (Right). More telomeres are evident after staining with γP_{12} . See Figure 7A for quantitative analysis.



Figure S6. Telomere staining of Jurkat metaphase chromosome spreads using P_{18} (Left) or γP_{12} (Right) under nondenaturing conditions. Metaphases were prepared and stained with a modified protocol that omitted formamide from the hybridization buffer and eliminated the heating step. The exposure time for the Cy3 channel was increased to 800ms for these experiments. Quantitative comparison of denaturing and nondenaturing conditions for telomere staining by γP_{12} is shown in Figure 7.

γP₆ Characterization



Figure S7. MALDI and HPLC traces of γP_6 . m/z = 3032.7 (observed), 3036.2 (calculated).

γP_{6-Clamp} Characterization



Figure S8. MALDI and HPLC traces of purified $\gamma P_{6-Clamp}$. m/z = 3416.1 (observed), 3420.6 (calculated).

γP₁₂ Characterization



Figure S9. MALDI and HPLC traces of γP_{12} . m/z = 5320.9 (observed), 5315.5 (calculated).