# Stability and Bioactivity of Thrombin Binding Aptamers Modified with

# **D-/L-Isothymidine in the Loop Regions**

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# The synthesize of the TBA varients

**TBA** and **D-/L-isoT** modified **TBAs** were synthesized on ABI 394 automated RNA/DNA synthesizer (**IsoT** phosphoramidite monomers and **isoT**-modified oligonucleotides were synthesized by our lab according to the literature<sup>1, 2</sup> using standard phosphoramidite chemistry). All the **TBA** variants were purified by C18 reverse high performance liquid chromatography (XBridgeTM OST C18, 2.5  $\mu$ m, 10 mm × 50 mm) using a linear gradient of 5 $\rightarrow$ 30% eluent A in 30 min. Solutions of 0.1 M Et<sub>3</sub>N-CH<sub>3</sub>COOH in water, pH = 7.7, were used as eluent B, and CH<sub>3</sub>CN was used as eluent A. Then the isolated DMT-on oligonucleotides were treated with 80% acetic acid for 10 min at room temperature. After neutralization with Et<sub>3</sub>N, the oligonucleotide solutions were desalted by Sephadex G25

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column respectively. The purity of all oligonucleotides was verified by Capillary Gel Electrophoresis and polypropylene gel electrophoresis to be over 90%, and the oligonucleotide compositions were confirmed by MALDI-TOF-MS spectrometry.



Figure S1. The purity of synthesized oligonucleotides determined by polyacrylamide gel electrophoresis.



**Figure S2**. The purity of synthesized oligonucleotides determined by capillary gel electrophoresis. Instrument and reagents: Beckman PA800 capillary electrophoresis (Beckman, USA), 32 Karat software (Version 5.0, Beckman) workstation; capillary (inner diameter 100 nm, OD 375 nm, the effective length 20 cm); Capillary Coating: 4% polypropylene amide. Separation conditions: 25 mmol / L Tris-boric-EDTA buffer solution (pH 7.94), 20% (w/v) PEG 35000, 25 °C, 25 kV, Detection wavelength was 254 nm.



Figure S3. MALDI-TOF-MS of TBA and its variants. Found: 4728 [M+2H]<sup>+</sup>; Calcd: 4726.



**Figure S4**. The separation results of **TBA** variants with HPLC. Dionex UltiMate 3000 HPLC, XBridgeTM OST C18 column (2.5  $\mu$ m, 10 mm × 50 mm), Gradient program: 5–30% eluent A in 30 min(A: 0.1M Et<sub>3</sub>N-CH<sub>3</sub>COOH in water, pH = 7.7; B: CH<sub>3</sub>CN), column temperature 40 °C.

Table S1. TBA and D/L-isoT modified variants

Name	Sequence(5'-3')	MALDI-TOF-MS [M+2H] <sup>+</sup>	
		Calcd.	Found
ТВА	GGT TGG TGT GGT TGG	4726	4728
TBA-3D	GG <b>T</b> <sub>D</sub> TGG TGT GGT TGG	4726	4728
TBA-3L	$GGT_L$ TGG TGT GGT TGG	4726	4728
TBA-4D	GGT <b>T</b> <sub>D</sub> GG TGT GGT TGG	4726	4728
TBA-4L	GGT $T_L$ GG TGT GGT TGG	4726	4728
TBA-7D	GGT TGG <b>T</b> <sub>D</sub> GT GGT TGG	4726	4728
TBA-7L	GGT TGG $T_L$ GT GGT TGG	4726	4728

TBA-9D	GGT TGG TG <b>T</b> <sub>D</sub> GGT TGG	4726	4728
TBA-9L	GGT TGG TG $T_L$ GGT TGG	4726	4728
TBA-12D	GGT TGG TGT GG <b>T<sub>D</sub> TGG</b>	4726	4728
TBA-12L	GGT TGG TGT $GGT_L$ TGG	4726	4728
TBA-13D	GGT TGG TGT GGT $T_D$ GG	4726	4728
TBA-13L	GGT TGG TGT GGT $T_L$ GG	4726	4728
TBA-3L7D	GG $T_L$ TGG $T_D$ GT GGT TGG	4726	4728
TBA-3L9L	GG $T_L$ TGG TG $T_L$ GGT TGG	4726	4728
TBA-3L 12L	GGT TGG TGT $GGT_L T_L GG$	4726	4728
TBA-7D9L	GGT TGG <b>T</b> <sub>D</sub> G <b>T</b> <sub>L</sub> GGT TGG	4726	4728
TBA-7D 12L	GGT TGG $T_D$ GT GG $T_L$ TGG	4726	4728
<b>TBA-9L 12L</b>	GGT TGG TG $T_L$ GG $T_L$ TGG	4726	4728

Note: The red italics represent D-isoT modification and the blue italics represent L-isoT modification



**Figure S5**. The thermodynamic stability of **D**-/L-isoT modified **TBA**s. The oligonucleotides were dissolved in the Tm buffer (100 mM potassium chloride and 10 mM sodium cacodylate, pH 7.4) and the concentration was  $8 \mu$ M.













Figure S6. SPR results of natural and D-/L-isoT modified TBAs with thrombin



A. TBA/thrombin



## B. TBA-7D/thrombin



#### C. TBA-7L/thrombin



### References

- 1. Z. J. Yang, H. W. Yu, J. M. Min, L. T. Ma, L. H. Zhang, *Tetrahedron: Asymmetry* 1997, 8, 2739.
- 2. J. Chen, L. R. Zhang, J. M. Min, L. H. Zhang, Nucleic Acids Res., 2002, 3005.