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Supporting Information

Enzyme-triggered delivery of chlorambucil from conjugates

based on the cell-penetrating peptide BP16

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1. Materials and methods

Unless otherwise stated, common chemicals and solvents (HPLC-grade or reagentgrade quality) were purchased from commercial sources and used without further purification. The 9-fluorenylmethoxycarbonyl (Fmoc) derivatives and Fmoc-Rink-4methylbenzhydrylamine (MBHA) resin (0.56 mmol/g) were obtained from Senn Chemicals International (Gentilly, France), NovaBiochem (Schwalbach, Germany) or IRIS Biotech GmbH (Marktredwitz, Germany). Ethyl 2-cyano-2-(hydroxyimino)acetate (Oxyma) was purchased from Novabiochem (Nottingham, UK). Trifluoroacetic acid (TFA), triisopropylsilane (TIS), dimethyl sulfoxide (DMSO), N.N'-diisopropylcarbodiimide (DIPCDI), chlorambucil (CLB), 5(6)-carboxyfluorescein (CF), hydrazine monohydrate and ethylenediaminetetraacetic acid (EDTA) were from Sigma-Aldrich (St. Louis, MO, USA). Cathepsin B (bovine spleen) was purchased from EMD chemicals (San Diego, CA, USA). L-(+)-Cysteine was obtained from Fischer Scientific (Waltham, MA USA). Piperidine and pyridine were purchased from Fluka (Buchs, Switzerland). Acetic anhydride (Ac2O) was purchased from Panreac (Barcelona, Spain). N-Methyl-2pyrrolidinone (NMP), N,N-dimethylformamide (DMF), CH₃OH, CH₂Cl₂, diethyl ether and solvents for high performance liquid chromatography (HPLC) were obtained from (Sentmenat, 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium Scharlau Spain). bromide (MTT), paraformaldehyde and bisbenzimide trihydrochloroacetic acid (Hoechst 33258) were purchased from Sigma-Aldrich (St. Louis, MO, USA). MitoTracker(R)Red CMXRos was from Molecular Probes, Life Technologies (Eugene, Oregon, USA). Dulbecco's modified Eagle's medium (DMEM), phosphate buffered saline (PBS), fetal bovine serum (FBS), penicillin-streptomycin and trypsin were obtained from GIBCO BRL (Grand Island, NY, USA).

Compounds were analyzed under standard analytical HPLC conditions with a Dionex liquid chromatography instrument composed of an UV/Vis Dionex UVD170U detector, a P680 Dionex bomb, an ASI-100 Dionex automatic injector, and CHROMELEON 6.60 software. Detection was performed at 220 nm. Solvent A was 0.1% aq. TFA and solvent B was 0.1% TFA in CH₃CN. Analysis was carried out with a Kromasil 100 C₁₈ (4.6 mm × 40 mm, 3.5 µm) column with a 2-100% B linear gradient over 7 min at a flow rate of 1.0 mL min⁻¹.

Electrospray ionization mass spectrometry (ESI-MS) analyses were performed with an Esquire 6000 ESI ion Trap LC/MS (Bruker Daltonics) instrument equipped with an electrospray ion source (University of Girona). The instrument was operated in the positive ESI(+) ion mode. Samples (5 μ L) were introduced into the mass spectrometer ion source directly through an HPLC autosampler. The mobile phase (80:20 CH₃CN/H₂O at a flow rate of 100 μ L min⁻¹) was delivered by a 1200 Series HPLC pump (Agilent). Nitrogen was employed as both the drying and nebulizing gas.

High-resolution mass spectra (HRMS) were recorded under conditions of ESI with a Bruker MicrOTOF-Q IITM instrument using a hybrid quadrupole time-of-flight mass spectrometer (University of Girona). Samples were introduced into the mass spectrometer ion source by direct infusion through a syringe pump and were externally calibrated using sodium formate. The instrument was operated in the positive ESI(+) ion mode.

HPLC-MS analyses were performed under standard analytical HPLC conditions described above with an Agilent Technologies 1200 HPLC system equipped with a VWD detector and coupled with an Esquire 6000 ESI ion Trap LC/MS (Bruker Daltonics) instrument with an electrospray ion source (University of Girona). The instrument was operated in the positive ESI(+) ion mode.

2. Cell lines

The human breast cancer MCF-7, melanoma SKMEL-28, prostate cancer PC-3 and pancreas cancer CAPAN-1 cell lines were obtained from the American Tissue Culture Collection (ATCC, Rockville, MD, USA). The human skin fibroblasts 1BR3G were obtained from EucellBank (University of Barcelona, Barcelona, Spain). Cells were maintained in DMEM supplemented with 10% FBS and 100 U ml⁻¹ penicillin-streptomycin at 37 °C under a humidified atmosphere containing 5% CO₂. Cells were passaged two times per week.

3. HPLC, ESI-MS and HRMS of peptides and CLB-peptide conjugates

Figure S1: a) HPLC chromatogram (λ = 220 nm), b) HRMS spectrum (*m*/*z*).

BP16



a)



No.	mps retenc min	alçada mAU	Area mAU*min	Area relativa %
1	5,73	29,200	2,392	4,34
2	5,98	560,424	47,628	86,44
3	7,36	43,883	5,076	9,21
Total:		633,507	55.097	100.00

b)









Observed HRMS (top) with the theoretical isotope prediction (bottom).



BP308





No.	mps retenc	alçada	Area	Area relativa
	min	mAU	mAU*min	%
1	6,08	1801,317	177,418	98,51
2	6,98	19,947	1,208	0,67
3	8,49	20,643	1,470	0,82
Total:		1841,907	180,096	100,00



Observed HRMS (top) with the theoretical isotope prediction (bottom).



Observed HRMS (top) with the theoretical isotope prediction (bottom).

Figure S3: a) HPLC chromatogram (λ = 220 nm), b) HRMS spectrum (*m*/*z*).

BP325





No.	mps retenc	alçada	Area	Area relativa
	min	mAU	mAU*min	%
1	6,42	31,278	3,951	3,28
2	7,19	35,817	7,126	5,91
3	7,71	991,334	109,512	90,81
Total:		1058,429	120,589	100,00



Observed HRMS (top) with the theoretical isotope prediction (bottom).



Observed HRMS (top) with the theoretical isotope prediction (bottom).



Observed HRMS (top) with the theoretical isotope prediction (bottom).



Observed HRMS (top) with the theoretical isotope prediction (bottom).

Figure S4: a) HPLC chromatogram (λ = 220 nm), b) HRMS spectrum (*m*/*z*).









No.	mps retenc min	alçada mAU	Area mAU*min	Area relativa %
1	7,24	1460,388	161,177	100,00
Total:		1460,388	161,177	100,00



Observed HRMS (top) with the theoretical isotope prediction (bottom).

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Observed HRMS (top) with the theoretical isotope prediction (bottom).



Observed HRMS (top) with the theoretical isotope prediction (bottom).



Observed HRMS (top) with the theoretical isotope prediction (bottom).

Figure S5: a) HPLC chromatogram (λ = 220 nm), b) HRMS spectrum (*m/z*).

BP332





No.	mps retenc	alçada	Area	Area relativa
	min	mAU	mAU*min	%
1	7,03	44,244	2,866	7,83
2	7,50	377,981	33,741	92,17
Total:		422,225	36,607	100,00



Observed HRMS (top) with the theoretical isotope prediction (bottom).



Observed HRMS (top) with the theoretical isotope prediction (bottom).



Observed HRMS (top) with the theoretical isotope prediction (bottom).

Figure S6: a) HPLC chromatogram (λ = 220 nm), b) HRMS spectrum (*m/z*).







No.	mps retenc min	alçada mAU	Area mAU*min	Area relativa %
1	7,38	742,124	61,018	90,51
2	7,63	104,960	6,398	9,49
Total:		847,085	<mark>67,416</mark>	100,00





Observed HRMS (top) with the theoretical isotope prediction (bottom).



Observed HRMS (top) with the theoretical isotope prediction (bottom).



Figure S7: a) HPLC chromatogram (λ = 220 nm), b) HRMS spectrum (*m/z*).

BP334





No.	Temps retenció min	alçada mAU	Area mAU*min	Area relativa %
1	7,59	1340,298	155,670	100,00
Total:		1340,298	155,670	100,00



Observed HRMS (top) with the theoretical isotope prediction (bottom).





Observed HRMS (top) with the theoretical isotope prediction (bottom).

Observed HRMS (top) with the theoretical isotope prediction (bottom).



Observed HRMS (top) with the theoretical isotope prediction (bottom).

Figure S8: a) HPLC chromatogram (λ = 220 nm), b) HRMS spectrum (*m/z*).







No.	Temps retenció min	alçada mAU	Area mAU*min	Area relativa %
1	7,51	546,652	49,784	100,00
Total:		546,652	49,784	100,00



Observed HRMS (top) with the theoretical isotope prediction (bottom).



Observed HRMS (top) with the theoretical isotope prediction (bottom).



Observed HRMS (top) with the theoretical isotope prediction (bottom).







Observed HRMS (top) with the theoretical isotope prediction (bottom).

Figure S9: a) HPLC chromatogram (λ = 220 nm), b) HRMS spectrum (*m/z*).

BP336





No.	Temps retenció min	alçada mAU	Area mAU*min	Area relativa %
1	7,82	1201,956	133,830	100,00
Total:		1201,956	133,830	100,00



Observed HRMS (top) with the theoretical isotope prediction (bottom).







Observed HRMS (top) with the theoretical isotope prediction (bottom).



Observed HRMS (top) with the theoretical isotope prediction (bottom).

Figure S10: a) HPLC chromatogram (λ = 220 nm), b) HRMS spectrum (*m/z*).







No.	Temps retenció	alçada m∆l l	Area mAl I*min	Area relativa
1	7,67	1534,141	168,935	100,00
Total:		1534,141	168,935	100,00



Observed HRMS (top) with the theoretical isotope prediction (bottom).

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Observed HRMS (top) with the theoretical isotope prediction (bottom).

Observed HRMS (top) with the theoretical isotope prediction (bottom).



Observed HRMS (top) with the theoretical isotope prediction (bottom).

4. HPLC, ESI-MS and HRMS of 5(6)-carboxyfluorescein labeled peptides

Figure S11: a) HPLC chromatogram (λ = 220 nm), b) HRMS spectrum (*m/z*).

CF-BP16



a)



No.	mps retenc min	alçada mAU	Area mAU*min	Area relativa %
1	6,40	60,473	8,138	6,40
2	6,76	473,768	22,057	17,34
3	6,80	981,748	97,024	76,26
Total:		1515,990	127,220	100,00

b)





Observed HRMS (top) with the theoretical isotope prediction (bottom).





Observed HRMS (top) with the theoretical isotope prediction (bottom).

Observed HRMS (top) with the theoretical isotope prediction (bottom).

Figure S12: a) HPLC chromatogram (λ = 220 nm), b) HRMS spectrum (*m/z*).







No.	Temps retenció	alçada	Area	Area relativa
	min	mAU	mAU*min	%
1	6,92	1859,025	98,247	25,25
2	7,00	2323,905	290,924	74,75
Total:		4182,930	389,171	100,00





Observed HRMS (top) with the theoretical isotope prediction (bottom).



Observed HRMS (top) with the theoretical isotope prediction (bottom).



Observed HRMS (top) with the theoretical isotope prediction (bottom).



Observed HRMS (top) with the theoretical isotope prediction (bottom).

Figure S13: a) HPLC chromatogram (λ = 220 nm), b) HRMS spectrum (*m/z*).





No.	mps retenc min	alçada mAU	Area mAU*min	Area relativa %
1	7,56	732,423	77,055	80,59
2	7,91	48,005	3,811	3,99
3	8,48	20,782	3,596	3,76
4	8,80	40,837	4,961	5,19
5	9,17	29,235	3,969	4,15
6	9,58	17,041	2,219	2,32
Total:		888,322	95,611	100,00



Observed HRMS (top) with the theoretical isotope prediction (bottom).

509.0

509.5

510.0

510.5

m/z

507.5

508.0

508.5



Observed HRMS (top) with the theoretical isotope prediction (bottom).



Observed HRMS (top) with the theoretical isotope prediction (bottom).

Figure S14: a) HPLC chromatogram (λ = 220 nm), b) HRMS spectrum (*m/z*).







No.	mps retenc min	alçada mAU	Area mAU*min	Area relativa %
1	6,78	16,216	1,282	1,19
2	7,35	34,135	2,255	2,09
3	7,68	518,041	40,429	37,41
4	7,83	758,095	64,094	59,31
Total:		1326,487	108,061	100,00





Observed HRMS (top) with the theoretical isotope prediction (bottom).



Observed HRMS (top) with the theoretical isotope prediction (bottom).



Observed HRMS (top) with the theoretical isotope prediction (bottom).



Figure S15: a) HPLC chromatogram (λ = 220 nm), b) HRMS spectrum (*m/z*).

BP339







No.	Temps retenció min	alçada mAU	Area mAU*min	Area relativa %
1	7,86	1616,497	156,827	100,00
Total:		1616,497	156,827	100,00



Observed HRMS (top) with the theoretical isotope prediction (bottom).



Observed HRMS (top) with the theoretical isotope prediction (bottom).



Observed HRMS (top) with the theoretical isotope prediction (bottom).



Observed HRMS (top) with the theoretical isotope prediction (bottom).

Figure S16: a) HPLC chromatogram (λ = 220 nm), b) HRMS spectrum (*m/z*).







No.	Temps retenció min	alçada mAU	Area mAU*min	Area relativa %
1	7,47	290,082	25,776	13,11
2	7,61	472,827	37,592	19,13
3	7,70	604,560	47,196	24,01
4	7,79	830,108	85,978	43,75
Total:		2197,577	196,541	100,00



Observed HRMS (top) with the theoretical isotope prediction (bottom).

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Observed HRMS (top) with the theoretical isotope prediction (bottom).



Observed HRMS (top) with the theoretical isotope prediction (bottom).



Observed HRMS (top) with the theoretical isotope prediction (bottom).