Electronic Supplementary Material (ESI) for Organic & Biomolecular Chemistry. This journal is © The Royal Society of Chemistry 2015

Byproduct Promoted Regioselective Sulfenylation of Indoles with Sulfinic Acids

Congrong Liu, * Lianghui Ding

School of Environment Engineering, Nanjing Institute of Technology, Nanjing, Jiangsu 211167, China

Supporting Information

Table of Contents

General Information	S-2
General Procedure for Byproduct Promoted Regioselective Sulfenylation of Indoles with	Sulfinic
Acids (Tables 2 and 3)	S-2
Analytical Data for the Products Shown in Tables 2 and 3	S-2
Sulfenylation of Indoles with Sulfinic Acids in the Abscence of Tetrabutylami	monium
Iodide	S-7
Reduction of Benzylsulfinic Acid 2a	S-7
Reaction of Indole 1a with Diphenyldisulfane 4a in the Optimized Reaction Conditions	S-7
Reaction of Indole 1a with Diphenyldisulfane 4a in 10 mol% I ₂	S-8
References	S-8
Copies of ¹ H and ¹³ C NMR Spectra	S-9

General Information

¹H and ¹³C NMR spectra were recorded on a Bruker AC-400 FT (400 MHz and 100 MHz, respectively) using tetramethylsilane as an internal reference. NMR multiplicities are abbreviated as follows: s = singlet, d = doublet, m = multiplet, br = broad signal. Chemical shifts (δ) and coupling constants (*J*) were expressed in ppm and Hz, respectively.

2a-2m were prepared according to the literature procedure.¹ The rest of chemicals were purchased from the Sinopharm Chemical Reagent Co., Meryer, Acros, Alfa Aesar, and TCI, and used as received.

General Procedure for Byproduct Promoted Regioselective Sulfenylation of Indoles with Sulfinic Acids (Tables 2 and table 3)

To a solution of indole 1 (0.20 mmol) in DCE (1.0 mL) were added sulfinic acid (0.24 mmol), tetrabutylammonium iodide (88.6 mg, 0.24 mmol) and TsOH (6.9 mg, 0.040 mmol). The resulting mixture was stirred at 80 °C for 12 h. The mixture was cooled to room temperature, and purified by flash column chromatography on silica gel, eluting with petroleum ether/ethyl acetate (100:1 to 10:1), to give compound **3**.

Analytical Data for the Products Shown in Tables 2 and 3



3a, ² white solid, m.p. 151–154 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.35 (s, 1H), 7.61 (d, J = 8.0 Hz, 1H), 7.47–7.39 (m, 2H), 7.29–7.22 (m, 1H), 7.19–7.00 (m, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 139.2, 136.5, 130.7, 129.1, 128.7, 125.9, 124.8, 123.1, 120.9, 119.7, 111.6, 102.9.



3b, ² white solid, m.p. 81–83 °C; ¹H NMR (400 MHz, CDCl₃) δ 7.60 (d, *J* = 8.0 Hz, 1H), 7.34 (d, *J* = 8.0 Hz, 1H), 7.30–7.23 (m, 2H), 7.19–7.05 (m, 5H), 7.05–6.97 (m, 1H), 3.76 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 139.7, 137.5, 135.0, 129.8, 128.6, 125.7, 124.6, 122.5, 120.5, 119.7, 109.7, 100.5, 33.0.



3c, ² white solid, m.p. 111–112 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.14 (s, 1H), 7.54 (d, *J* = 7.6 Hz, 1H), 7.30 (d, *J* = 8.0 Hz, 1H), 7.21–7.07 (m, 4H), 7.06–6.97 (m, 3H), 2.46 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 141.1, 139.3, 135.4, 130.3, 128.7, 125.5, 124.5, 122.2, 120.7, 119.0, 110.6, 99.3, 12.1.



3d, ² colorless oil; ¹H NMR (400 MHz, CDCl₃) δ 8.52 (s, 1H), 7.75–7.70 (m, 2H), 7.62 (d, *J* = 7.6 Hz, 1H), 7.43–6.98 (m, 11H); ¹³C NMR (100 MHz, CDCl₃) δ 142.1, 139.3, 135.9, 131.5, 131.2, 128.9, 128.8, 128.7, 128.2, 125.6, 124.7, 123.4, 121.2, 120.0, 111.2.



3e, ² white solid, m.p. 135–137 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.51 (s, 1H), 7.49 (d, J = 2.8 Hz, 1H), 7.40–7.30 (m, 2H), 7.21–7.14 (m, 2H), 7.12–7.02 (m, 4H); ¹³C NMR (100 MHz, CDCl₃) δ 141.0, 137.7, 133.1, 128.7, 126.3, 125.9, 124.7, 124.0, 114.7, 111.1, 103.8.



3f, ² white solid, m.p. 118–120 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.66 (s, 1H), 7.74 (d, J = 1.6 Hz, 1H), 7.43 (d, J = 2.4 Hz, 1H), 7.35–7.23 (m, 2H), 7.19–7.12 (m, 2H), 7.10–7.02 (m, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 138.8, 135.2, 132.0, 131.0, 128.8, 126.0, 125.9, 125.0, 122.1, 114.4, 113.1, 102.5.



3g, ² white solid, m.p. 77–80 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.36 (s, 1H), 7.35 (d, J = 2.8 Hz, 1H), 7.24 (d, J = 8.8 Hz, 1H), 7.18–7.00 (m, 6H), 6.92–6.85 (m, 1H), 3.74 (s, 3H); ¹³C NMR



3h, ² white solid, m.p. 172–175 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.70 (s, 1H), 8.40–8.36 (m, 1H), 8.02–7.94 (m, 1H), 7.56 (d, *J* = 2.8 Hz, 1H), 7.46 (d, *J* = 8.8 Hz, 1H), 7.20–7.12 (m, 2H), 7.12–7.02 (m, 3H), 3.89 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 167.9, 139.1, 138.8, 132.1, 128.9, 128.8, 126.0, 125.0, 124.5, 123.2, 122.5, 111.4, 104.8, 51.9.



3i, ² yellow solid, m.p. 151–154 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.93 (s, 1H), 8.57 (d, J = 2.4Hz, 1H), 8.20–8.14 (m, 1H), 7.66 (d, J = 2.4 Hz, 1H), 7.53–7.46 (m, 1H), 7.23–7.15 (m, 2H), 7.15–7.05 (m, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 142.9, 139.5, 137.8, 133.6, 129.0, 128.9, 126.5, 125.6, 118.7, 116.9, 111.9, 106.7.



3j, ² white solid, m.p. 106–108°C; ¹H NMR (400 MHz, CDCl₃) δ 8.39 (s, 1H), 7.52–7.44 (m, 2H), 7.42 (d, *J* = 1.6 Hz, 1H), 7.20–7.02 (m, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 138.7, 136.8, 131.1, 129.1, 128.8, 127.7, 126.0, 125.0, 121.7, 120.6, 111.6, 103.5.



3k, ² white solid, m.p. 130–133 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.58 (s, 1H), 7.94–7.84 (m, 2H), 7.58–7.52 (m, 2H), 7.50–7.45 (m, 1H), 7.39–7.33 (m, 1H), 7.32–7.25 (m, 2H), 7.22–7.15 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 148.7, 142.6, 136.6, 131.3, 129.3, 128.5, 123.5, 121.3, 120.3, 119.7, 119.2, 111.9, 100.9.



31, ² yellow solid, m.p. 180–183 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.71 (s, 1H), 8.01–7.93 (m, 2H), 7.54–7.46 (m, 3H), 7.34–7.27 (m, 1H), 7.22–7.15 (m, 1H), 7.15–7.08 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 149.9, 144.9, 136.6, 131.3, 128.4, 125.1, 123.9, 123.5, 121.4, 119.2, 112.0, 100.1.



3m, ² white solid, m.p. 130–133 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.36 (s, 1H), 7.59 (d, J = 8.8 Hz, 1H), 7.47–7.39 (m, 2H), 7.29–7.22 (m, 1H), 7.20–7.12 (m, 1H), 7.11–7.04 (m, 2H), 6.90–6.82 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 160.9 (d, J = 243 Hz), 136.5, 134.0 (d, J = 2.9 Hz), 130.5, 128.9, 127.9 (d, J = 7.8 Hz), 123.2, 121.0, 119.6, 115.8 (d, J = 22 Hz), 111.7, 103.4.



3n, ² white solid, m.p. 130–132 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.39 (s, 1H), 7.57 (d, J = 8.0 Hz, 1H), 7.47–7.39 (m, 2H), 7.30–7.22 (m, 1H), 7.20–7.13 (m, 1H), 7.12–7.07 (m, 2H), 7.04–6.97 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 137.9, 136.5, 130.7, 130.6, 128.8, 127.2, 123.2, 121.1, 119.5, 111.7, 102.5.



30, ² white solid, m.p. 144–147 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.40 (s, 1H), 7.59–7.53 (m, 1H), 7.46–7.39 (m, 2H), 7.30–7.21 (m, 3H), 7.20–7.13 (m, 1H), 6.97–6.90 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 138.6, 136.5, 131.7, 130.8, 128.8, 127.5, 123.3, 121.1, 119.5, 118.4, 111.7, 102.3.



3p, ² white solid, m.p. 129–132 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.43 (s, 1H), 7.58–7.53 (m, 1H), 7.48–7.40 (m, 4H), 7.31–7.22 (m, 1H), 7.20–7.13 (m, 1H), 6.85–6.79 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 139.6, 137.5, 136.5, 130.8, 128.8, 127.7, 123.3, 121.1, 119.5, 111.7, 102.1, 89.0.



3q

3q, ² white solid, m.p. 125–127 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.19 (s, 1H), 7.60 (d, J = 7.6 Hz, 1H), 7.35–7.31 (m, 2H), 7.25–7.18 (m, 1H), 7.16–7.10 (m, 1H), 7.02 (d, J = 8.4 Hz, 2H), 6.94 (d, J = 8.4 Hz, 2H), 2.22 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 136.4, 135.4, 134.7, 130.5, 129.5, 129.0, 126.3, 122.9, 120.8, 119.6, 111.6, 103.3, 20.8.



3r, ² yellow solid, m.p. 111–112 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.29 (s, br, 1H), 7.62 (d, J = 8.0 Hz, 1H), 7.39–7.32 (m, 2H), 7.26–7.18 (m, 1H), 7.17–7.08 (m, 3H), 6.75–6.68 (m, 2H), 3.70 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 157.8, 136.5, 130.1, 129.6, 129.0, 128.6, 123.0, 120.8, 119.6, 114.6, 111.6, 104.5, 55.4.



3s, ² white solid, m.p. 140–142 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.45 (s, 1H), 7.74–7.69 (m, 1H), 7.66–7.59 (m, 2H), 7.59–7.52 (m, 2H), 7.50–7.48 (m, 1H), 7.47–7.43 (m, 1H), 7.40–7.31 (m, 2H), 7.30–7.24 (m, 2H), 7.18–7.11 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 136.7, 136.6, 133.8, 131.3, 130.7, 129.1, 128.2, 127.7, 126.9, 126.3, 125.0, 124.8, 123.6, 123.1, 121.0, 119.7, 111.6, 102.9.



3t, ² colorless oil; ¹H NMR (400 MHz, CDCl₃) δ 8.16 (s, 1H), 7.79–7.74 (m, 1H), 7.39–7.34 (m, 1H), 7.29 (d, J = 2.8 Hz, 1H), 7.27–7.17 (m, 2H), 2.37 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 136.3, 128.8, 127.8, 122.7, 120.4, 119.3, 111.5, 108.3, 20.2.



3u, ² yellow oil; ¹H NMR (400 MHz, CDCl₃) δ 8.19 (s, 1H), 7.80–7.75 (m, 1H), 7.40–7.35 (m, 1H), 7.31 (d, J = 2.4 Hz, 1H), 7.27–7.15 (m, 2H), 2.69 (t, J = 7.6 Hz, 2H), 1.59–1.49 (m, 2H), 1.39–1.31 (m, 2H), 1.30–1.21 (m, 8H), 0.86 (t, J = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 136.3, 129.5, 129.2, 122.6, 120.4, 119.5, 111.4, 106.4, 36.5, 31.8, 29.9, 29.2, 28.6, 22.6, 14.1.



3v, ² yellow solid, m.p. 80–82 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.10 (s, 1H), 7.72–7.67 (m, 1H), 7.34 (d, J = 7.6 Hz, 1H), 7.26–7.14 (m, 5H), 7.10–7.04 (m, 2H), 6.98 (d, J = 2.4 Hz, 1H), 3.86 (s, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 139.1, 136.2, 129.9, 129.3, 129.0, 128.2, 126.8, 122.7, 120.5, 119.3, 111.5, 105.3, 41.0.

Sulfenylation of Indoles with Sulfinic Acids in the Abscence of Tetrabutylammonium Iodide



To a solution of indole 1a (23.4 mg, 0.20 mmol) in DCE (1.0 mL) were added benzenesulfinic acid 2a (34.1 mg, 0.24 mmol) and TsOH (6.9 mg, 0.040 mmol). The resulting mixture was stirred at 80 °C for 12 h. The mixture was cooled to room temperature, and detect by TLC, found no compound 3a.

Reduction of Benzenesulfinic Acid 2a

PhSO₂H $\xrightarrow{n-Bu_4NI, TsOH}$ PhS-SPh + I₂ 2a 4a : (96%)

To a solution of benzenesufinic acid 2a (34.1 mg, 0.24 mmol) in DCE (1.0 mL) were added tetrabutylammonium iodide (88.6 mg, 0.24 mmol) and TsOH (6.9 mg, 0.040 mmol). The resulting mixture was stirred at 80 °C for 12 h. The mixture was cooled to room temperature, and purified by flash column chromatography on silica gel, eluting with petroleum ether, to give compound 4a (50.2 mg, 96%).

Reaction of Indole 1a with Diphenyldisulfane 4a in 1.2 eq I₂



To a solution of indole 1a (23.4 mg, 0.20 mmol) in DCE (1.0 mL) were added diphenyldisulfane 4a (52.3 mg, 0.24 mmol), iodine (61.0 mg, 0.24 mmol) and TsOH (6.9 mg, 0.040 mmol). The resulting mixture was stirred at 80 °C for 12 h. The mixture was cooled to room temperature, and purified by flash column chromatography on silica gel, eluting with petroleum ether/ethyl acetate (30:1), to give compound 3a (44.6 mg, 99%).

Reaction of Indole 1a with Diphenyldisulfane 4a under the Optimized Reaction Conditions

To a solution of indole 1a (23.4 mg, 0.20 mmol) in DCE (1.0 mL) were added diphenyldisulfane 4a (52.3 mg, 0.24 mmol), tetrabutylammonium iodide (88.6 mg, 0.24 mmol) and TsOH (6.9 mg, 0.040 mmol). The resulting mixture was stirred at 80 °C for 12 h. The mixture was cooled to room temperature, and detect by TLC, found no compound 3a.

References

[1] M. Conrads, J. Matty, Synthesis. 1991, 11, 2367.

[2] F.- L.Yang, S.-K.Tian, Angew. Chem. 2013, 125, 5029; Angew. Chem. Int. Ed. 2013, 52, 4929.





















S-18























