# Photorelease of Tyrosine from Alpha-Carboxy-6-nitroveratryl (αCNV) Derivatives

### **Supplementary Information**

Alexander G. Russell,<sup>†</sup> Matthew J. Sadler,<sup>†</sup> Helen J. Laidlaw,<sup>†</sup> Agustín Gutiérrez-Loriente,<sup>†</sup> Christopher W. Wharton,<sup>‡</sup> David Carteau,<sup>§</sup> Dario M. Bassani<sup>\*,§</sup> and John S. Snaith<sup>\*,†</sup>

School of Chemistry, University of Birmingham, Edgbaston, Birmingham, B15 2TT,
U.K., School of Biosciences, University of Birmingham, Edgbaston, Birmingham,
B15 2TT, U.K. and ISM CNRS UMR 5255, Université Bordeaux 1, 33405 Talence,
France.

j.s.snaith@bham.ac.uk

d.bassani@ism.u-bordeaux1.fr

<sup>†</sup> School of Chemistry, University of Birmingham

<sup>‡</sup> School of Biosciences, University of Birmingham

<sup>§</sup> University Bordeaux 1

Index

Details of kinetics and quantum yield measurements	S6-S11
NMR spectra:	
Allyl 2-(4-( <i>tert</i> -butyl)phenoxy)-2-(4,5-dimethoxy-2-nitrophenyl)ac	etate (4):
<sup>1</sup> H NMR (300 MHz, CDCl <sub>3</sub> )	S12
$^{13}$ C NMR (75 MHz, CDCl <sub>3</sub> )	<b>S</b> 13
(2S)-Methyl 3-(4-(2-(allyloxy)-1-(4,5-dimethoxy-2-nitrophenyl)-2-	
oxoethoxy)phenyl)-2-(( <i>tert</i> -butoxycarbonyl)amino)propanoate (5)	:
<sup>1</sup> H NMR (300 MHz, $CDCl_3$ )	S14
<sup>13</sup> C NMR (75 MHz, CDCl <sub>3</sub> )	S15
2-(4-(tert-Butyl)phenoxy)-2-(4,5-dimethoxy-2-nitrophenyl)acetic acid (6):	
<sup>1</sup> H NMR (300 MHz, $d_6$ -acetone)	S16
$^{13}$ C NMR (75 MHz, d <sub>6</sub> -acetone)	S17
(1S)-1-Carboxy-2-(4-(carboxy(4,5-dimethoxy-2-	
nitrophenyl)methoxy)phenyl)ethanaminium 2,2,2-trifluoroacetate	e ( <b>7</b> ):
<sup>1</sup> H NMR (300 MHz, $d_6$ -acetone)	S18
$^{13}$ C NMR (75 MHz, d <sub>6</sub> -acetone)	S19
2-(3,4-Bis(ethoxycarbonylmethoxy)phenyl)acetic acid ethoxycarbonylmethyl	
ester:	
<sup>1</sup> H NMR (300 MHz, $CDCl_3$ )	S20
<sup>13</sup> C NMR (100 MHz, CDCl <sub>3</sub> )	S21
2-(4,5-Bis(ethoxycarbonylmethoxy-2-nitro)phenyl)acetic acid	
ethoxycarbonylmethyl ester (8):	
<sup>1</sup> H NMR (300 MHz, $CDCl_3$ )	S22
$^{13}$ C NMR (75 MHz, CDCl <sub>3</sub> )	

$D_{14}^{1} + 1 = 2 = 2 = 2 = 2 = 2 = 2 = 2 = 2 = 2 =$	
Diethyl 2,2 -((4-(1-diazo-2-(2-ethox)	y-2-oxoethoxy)-2-oxoethyl)-5-nitro-1,2-

#### phenylene)bis(oxy))diacetate:

<sup>1</sup> H NMR (300 MHz, CDCl <sub>3</sub> )	S24

#### 2-(4,5-Bis(ethoxycarbonylmethoxy)-2-nitrophenyl)-2-bromoacetic acid

#### ethoxycarbonylmethyl ester (9):

<sup>1</sup> H NMR (300 MHz, $CDCl_3$ )	S26
	0.07

$$^{13}$$
C NMR (75 MHz, CDCl<sub>3</sub>) S27

Diethyl 2,2'-((4-(1-(4-(tert-butyl)phenoxy)-2-(2-ethoxy-2-oxoethoxy)-2-oxoethyl)-

<sup>1</sup> H NMR (300 MHz, $CDCl_3$ )	S28
---	-----

S29
5

Diethyl 2,2'-((4-(1-(4-((S)-2-((*tert*-butoxycarbonyl)amino)-3-methoxy-3-

oxopropyl)phenoxy)-2-(2-ethoxy-2-oxoethoxy)-2-oxoethyl)-5-nitro-1,2-

phenylene)bis(oxy))diacetate (11):

<sup>1</sup> H NMR (300 MHz, $CDCl_3$ )	S30
---	-----

 $^{13}$ C NMR (75 MHz, CDCl<sub>3</sub>) S31

2,2'-((4-((4-(*tert*-Butyl)phenoxy)(carboxy)methyl)-5-nitro-1,2-

phenylene)bis(oxy))diacetic acid (12):

<sup>1</sup> H NMR (300 MHz, $d_6$ -acetone)	<b>S</b> 32
--	-------------

## (1S)-2-(4-((4,5-Bis(carboxymethoxy)-2-nitrophenyl)(carboxy)methoxy)phenyl)-1-

**S**33

### carboxyethanaminium 2,2,2-trifluoroacetate (13):

\$34

$$^{13}$$
C NMR (75 MHz, D<sub>2</sub>O) S35

<i>tert</i> -Butyl 2-(4,5-dimethoxy-2-nitrophenyl)acetate:	
<sup>1</sup> H NMR (300 MHz, CDCl <sub>3</sub> )	S36
<sup>13</sup> C NMR (75 MHz, CDCl <sub>3</sub> )	S37
tert-Butyl 2-diazo-2-(4,5-dimethoxy-2-nitrophenyl)acetate	
<sup>1</sup> H NMR (300 MHz, CDCl <sub>3</sub> )	S38
<sup>13</sup> C NMR (75 MHz, CDCl <sub>3</sub> )	S39
tert-Butyl 2-bromo-2-(4,5-dimethoxy-2-nitrophenyl)acetate (14):	
<sup>1</sup> H NMR (300 MHz, $CDCl_3$ )	S40
<sup>13</sup> C NMR (125 MHz, CDCl <sub>3</sub> )	S41
(2S)-tert-Butyl 3-(4-(2-(allyloxy)-1-(4,5-dimethoxy-2-nitrophenyl)-	-2-
oxoethoxy)phenyl)-2-(( <i>tert</i> -butoxycarbonyl)amino)propanoate (17	7):
<sup>1</sup> H NMR (300 MHz, $CDCl_3$ )	S42
<sup>13</sup> C NMR (100 MHz, CDCl <sub>3</sub> )	S43
(2S)-Methyl 3-(4-(2-( <i>tert</i> -butoxy)-1-(4,5-dimethoxy-2-nitropheny	l)-2-
oxoethoxy)phenyl)-2-(2,2,2-trifluoroacetamido)propanoate (18):	
<sup>1</sup> H NMR (300 MHz, $d_6$ -acetone)	S44
$^{13}$ C NMR (75 MHz, d <sub>6</sub> -acetone)	S45
(1S)-2-(4-(2-(Allyloxy)-1-(4,5-dimethoxy-2-nitrophenyl)-2-oxoeth	oxy)phenyl)-1-
carboxyethanaminium 2,2,2-trifluoroacetate	
<sup>1</sup> H NMR (300 MHz, $d_6$ -acetone)	S46
(2 <i>S</i> )-2-((((9 <i>H</i> -Fluoren-9-yl)methoxy)carbonyl)amino)-3-(4-(2-(all	yloxy)-1-(4,5-
dimethoxy-2-nitrophenyl)-2-oxoethoxy)phenyl)propanoic acid (19	9)
<sup>1</sup> H NMR (400 MHz, CDCl <sub>3</sub> )	S47
<sup>13</sup> C NMR (100 MHz, CDCl <sub>3</sub> )	S48

#### (2S)-2-((((9H-Fluoren-9-yl)methoxy)carbonyl)amino)-3-(4-(2-(tert-butoxy)-1-(4,5-

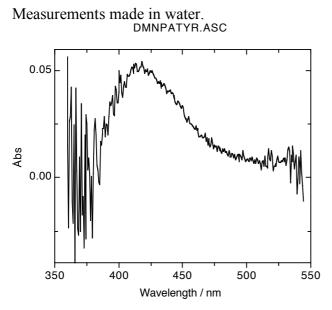
### dimethoxy-2-nitrophenyl)-2-oxoethoxy)phenyl)propanoic acid (20)

$^{1}$ H NMR (300 MHz, CDCl <sub>3</sub> )	S49			
<sup>13</sup> C NMR (75 MHz, CDCl <sub>3</sub> )	S50			
Mosher's amide of +/- tyrosine				
<sup>19</sup> F NMR (282 MHz, d <sub>6</sub> -acetone)	<b>S</b> 51			
Mosher's amide of tyrosine released by photolysis of Mosher's amide of 7				
<sup>19</sup> F NMR (282 MHz, d <sub>6</sub> -acetone)	S52			

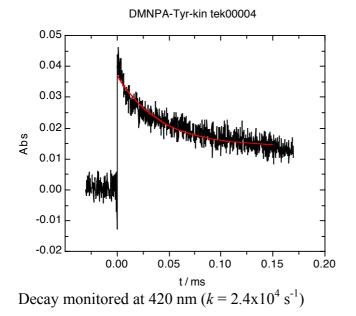
#### **Kinetic measurements**

The measurements were performed in aerated H<sub>2</sub>O or EtOH/H<sub>2</sub>O (4:1) solutions, as indicated, using a pump – probe setup equipped with a frequency - tripled Nd-YAG laser (BMI, 355 nm, 6 ns pulses) as an excitation source. An intensified CCD (Andor Technologies Instaspec V) or a photomultiplier (Hamamatsu R 446 UR connected to a transient digitizer) was used for detection for the spectral and temporal measurements, respectively.

#### **Kinetic Measurements for Compound 7**



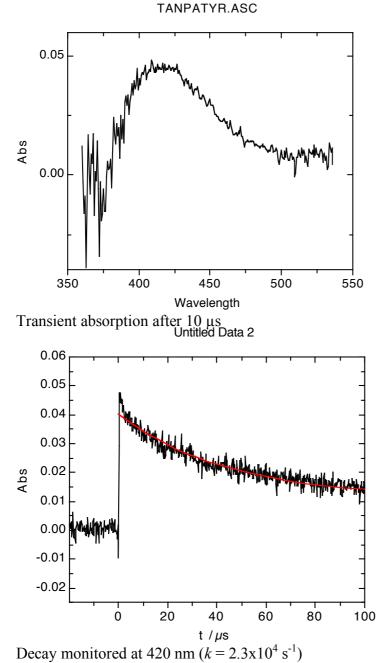
Transient absorption after 10 µs



Chi sq	uared	= 5.2555e-3			
Param	eters:			Standa	ard deviations:
А	=	2.9284e-2	ΔΑ	=	1.4412e+4
x0	=	-9.9136e-6	$\Delta x0$	=	20.1604
t0	=	4.0963e-5	$\Delta t0$	=	1.5494e-6
const	=	1.4108e-2	Δcons	t =	2.0486e-4

#### **Kinetic Measurements for Compound 13**

Measurements made in water.

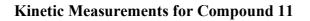


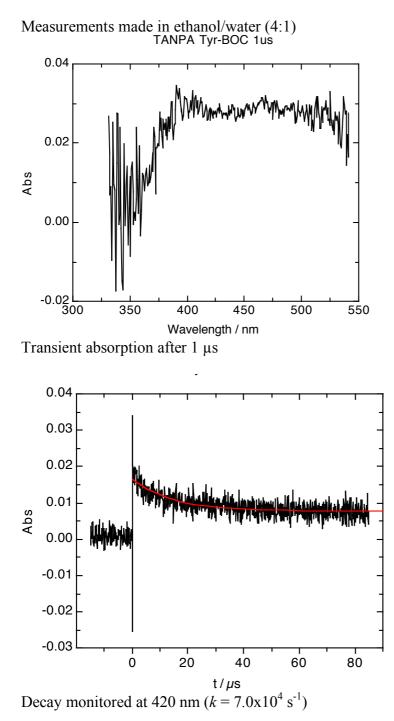
Chi sq	uared	= 3.7	3.7828e-3		
Param	eters:			Standa	ard deviations:
А	=	3.3573e-2	$\Delta A$	=	4436.0902
x0	=	-6.2146e-6	$\Delta x0$	=	5.7719
t0	=	4.3683e-5	$\Delta t0$	=	1.1500e-6
const	=	1.1171e-2	∆cons	t =	1.8916e-4

#### **Kinetic Measurements for Compound 5**

Measurements made interthantolassater (4:1) 0.00 Abs -0.05 350 400 450 500 300 550 Wavelength / nm Transient absorption after 1 µs DMNPA-TyrBOC-kin tek00007 0.10 0.05 Abs 0.00 -0.05 20 40 80 Ó 60 t/µs Decay monitored at 420 nm ( $k = 1.3 \times 10^5 \text{ s}^{-1}$ )

Chi sq	uared	= 1.3074e-2				
Param	eters:				Standa	rd deviations:
А	=	3.7668	le-2	$\Delta A$	=	2.9417e+4
x0	=	-2.037	2e-6	$\Delta x0$	=	5.9850
t0	=	7.6637	'e-6	$\Delta t0$	=	4.0326e-7
const	=	6.4542	le-3	∆const	t =	1.6965e-4





Chi sq	uared	=	3.1589	e-3		
Param	eters:				Standa	rd deviations:
А	=	2.3738	e-2	$\Delta A$	=	3.7685e+4
x0	=	-1.396	5e-5	$\Delta x0$	=	22.5644
t0	=	1.4214	e-5	$\Delta t0$	=	1.0238e-6
const	=	7.7570	e-3	∆const	=	1.1187e-4

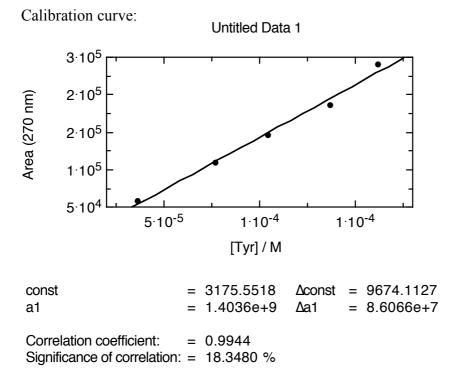
#### Quantum yield determination

Quantum yields were determined using an optical bench consisting of a 0101 Schoeffel lamp housing equipped with a xenon-mercury lamp (150 W) and a 77250 Oriel monochromator. Irradiations were carried out in aerated water at 20 °C at 365 nm (band-width of 10 nm) and followed by HPLC analysis.

The photon flux was determined using potassium ferrioxalate actinometry (Hatchard – Parker actinometer):<sup>1</sup> A 3.0 mL solution of ferrioxalate (0.006 M) in H<sub>2</sub>SO<sub>4</sub> (0.05M) was irradiated and the moles of photoproduct were determined from the change in optical density at 510 nm and the extinction coefficient (11100 M<sup>-1</sup>cm<sup>-1</sup>) 1 h after addition of 0.1 % of 1,10-phenanthroline to the photolyzed solution. The known quantum yield of photoproduct (1.26)<sup>2</sup> was used to calculate the photon flux (I =  $5.48 \times 10^{-7}$  E/min).

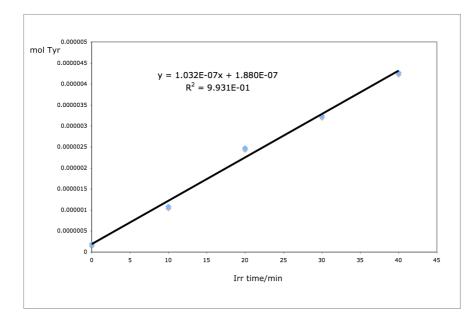
#### 1. HPLC Analysis of Tyr.

Conditions: 70:30 acetonitrile:water (isocratic), 1 mL/min, detector at 270 nm.



#### 2. Quantum yield determination for compound 7

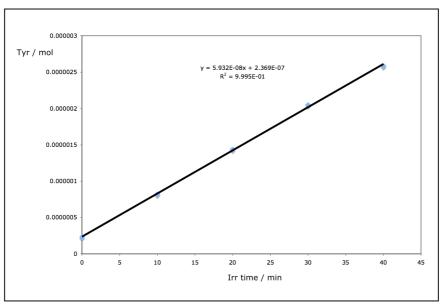
Solution in water at 1.389 mM, irradiated at 365 and analyzed at 10 min intervals.

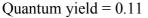


Quantum yield = 0.19

#### 3. Quantum yield determination for compound 13

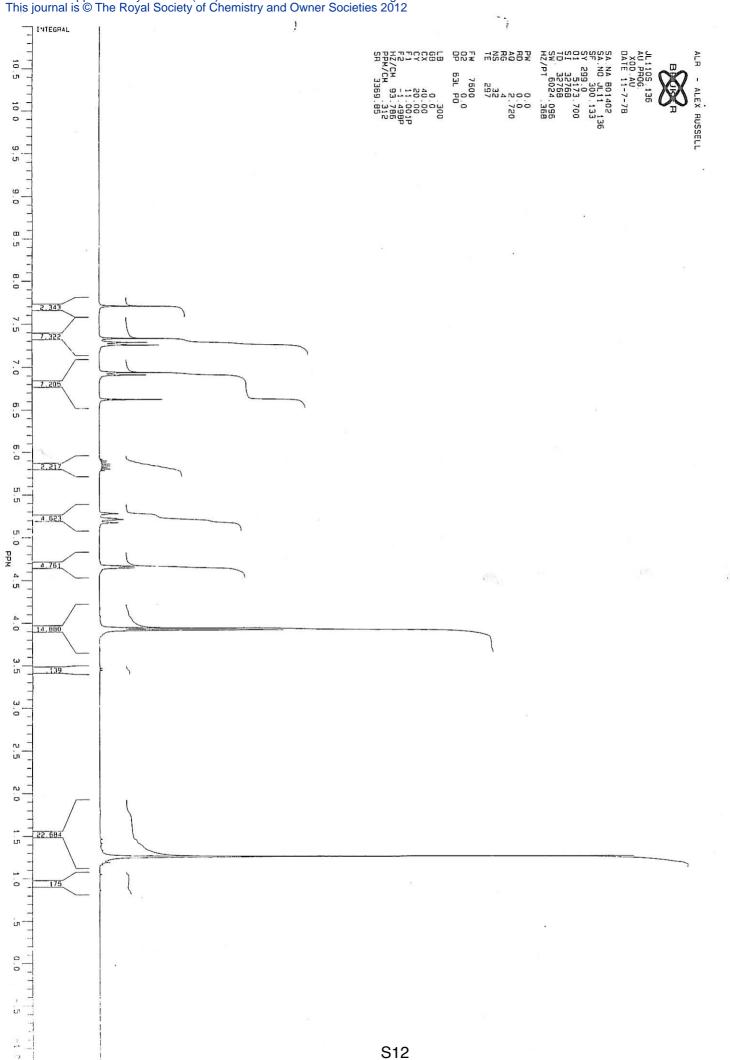
Solution in water at 1.095 mM, irradiated at 365 and analyzed at 10 min intervals.

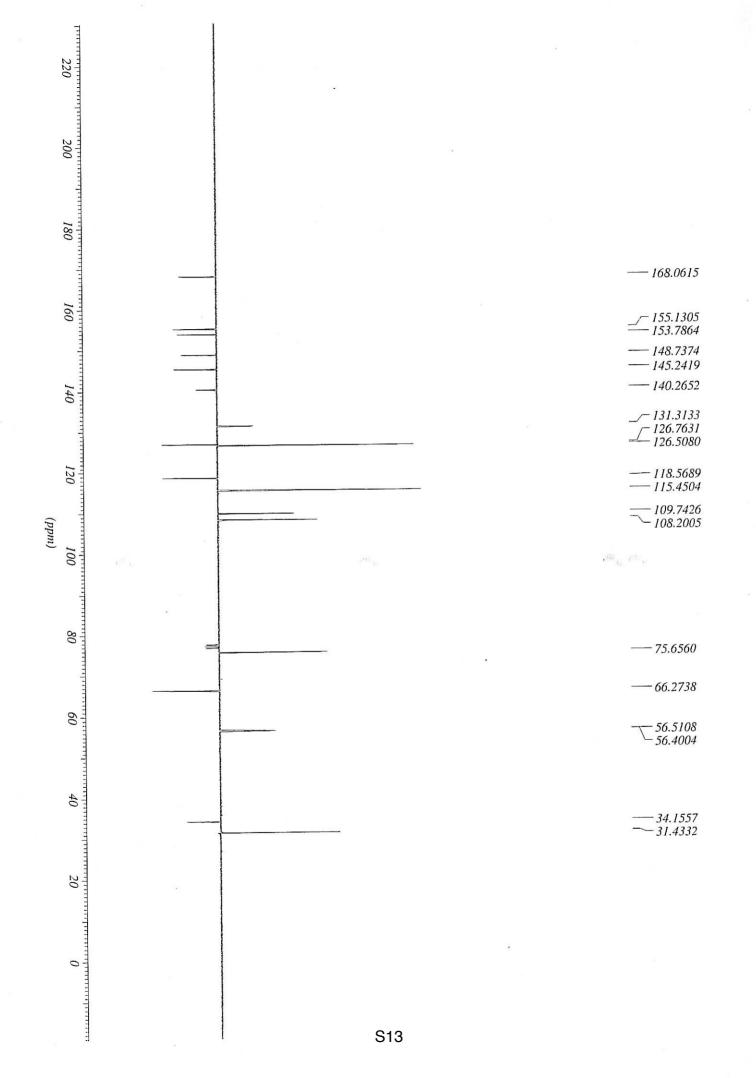


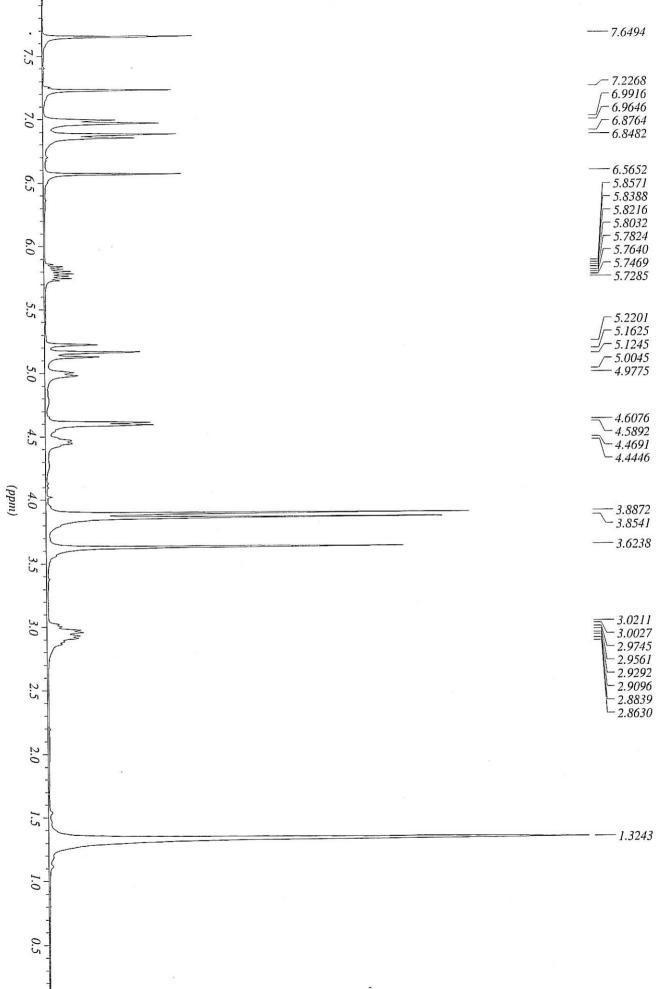


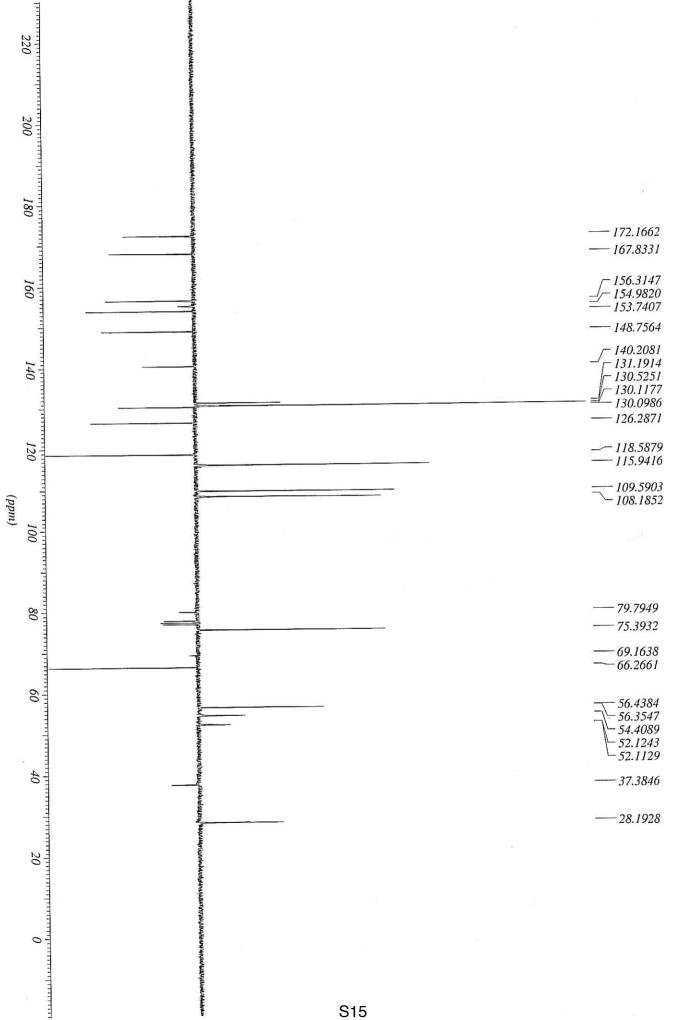
<sup>1</sup> Hatchard, C. G.; Parker, C. A. Proc. R. Soc. **1956**, A235, 518-536.

<sup>2</sup> Scaiano, J. C. *Handbook of Organic Photochemistry*, Vol. I, CRC Press, Florida, **1989**.

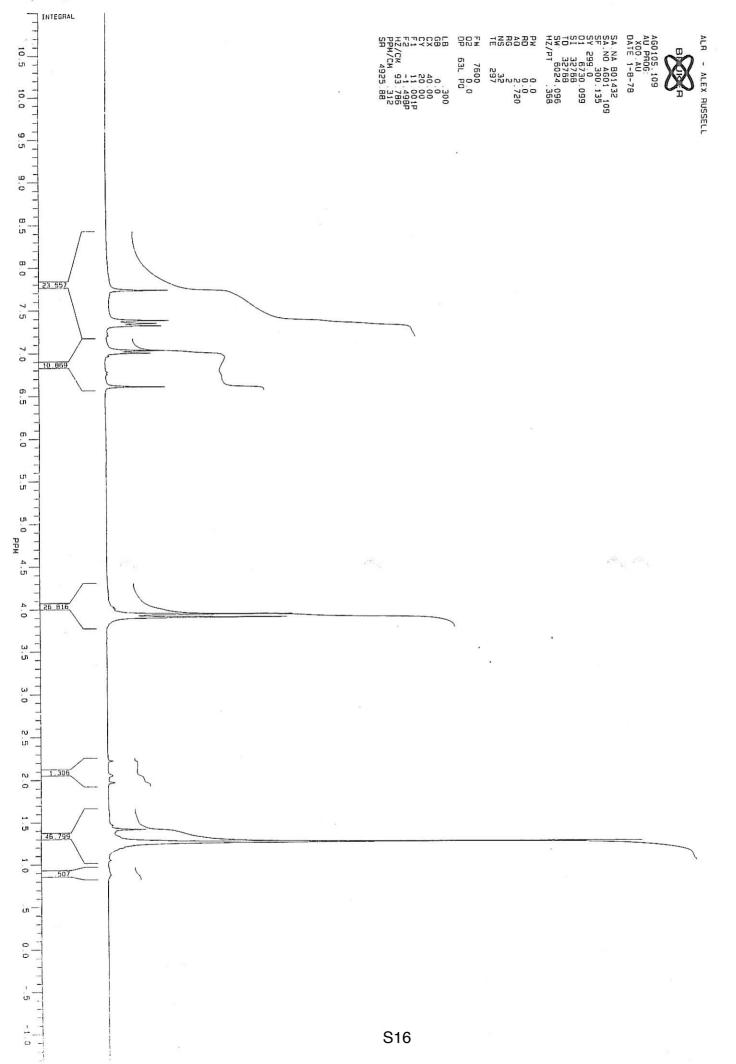


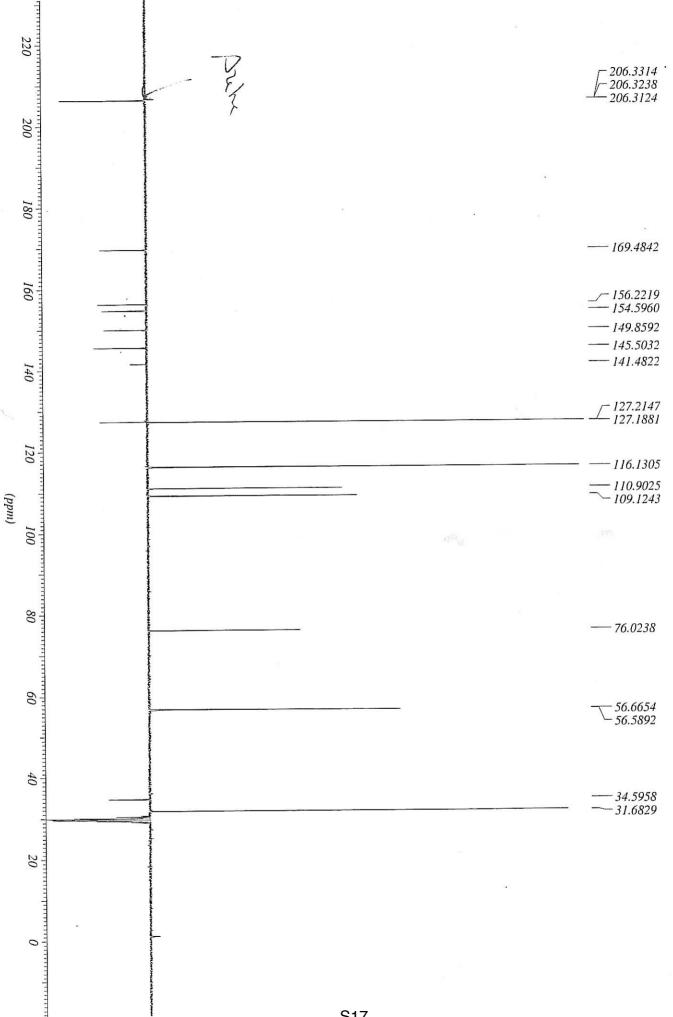


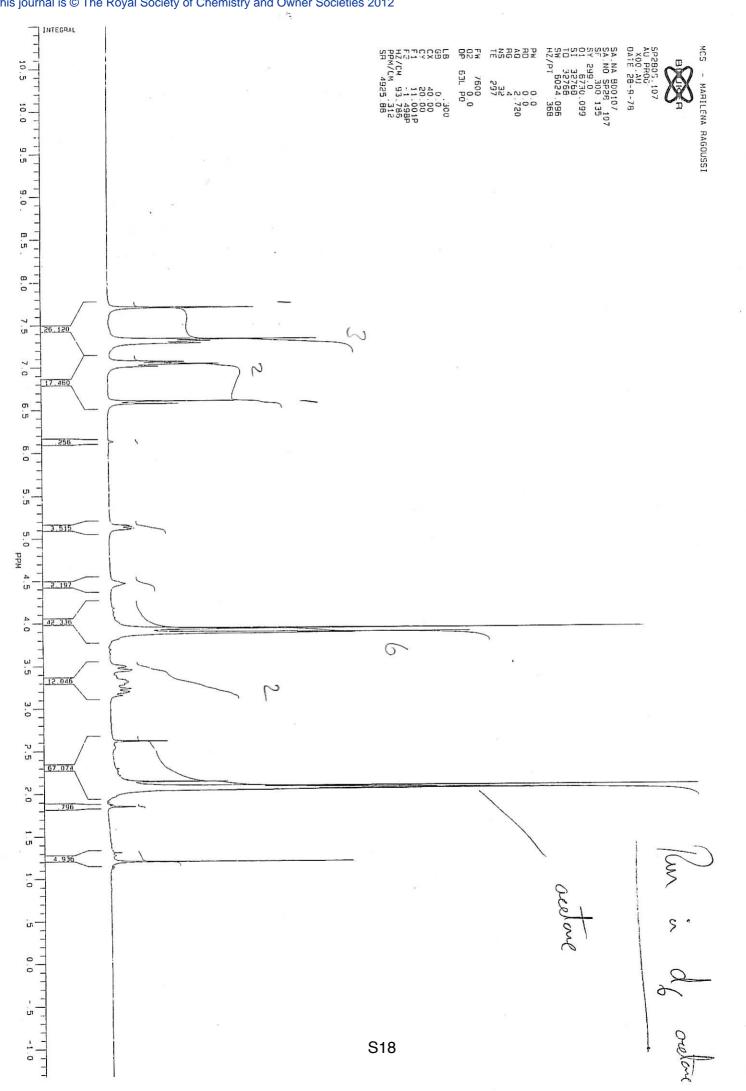




Electronic Supplementary Material (ESI) for Photochemical & Photobiological Science This journal is C The Royal Society of Chemistry and Owner Societies 2012

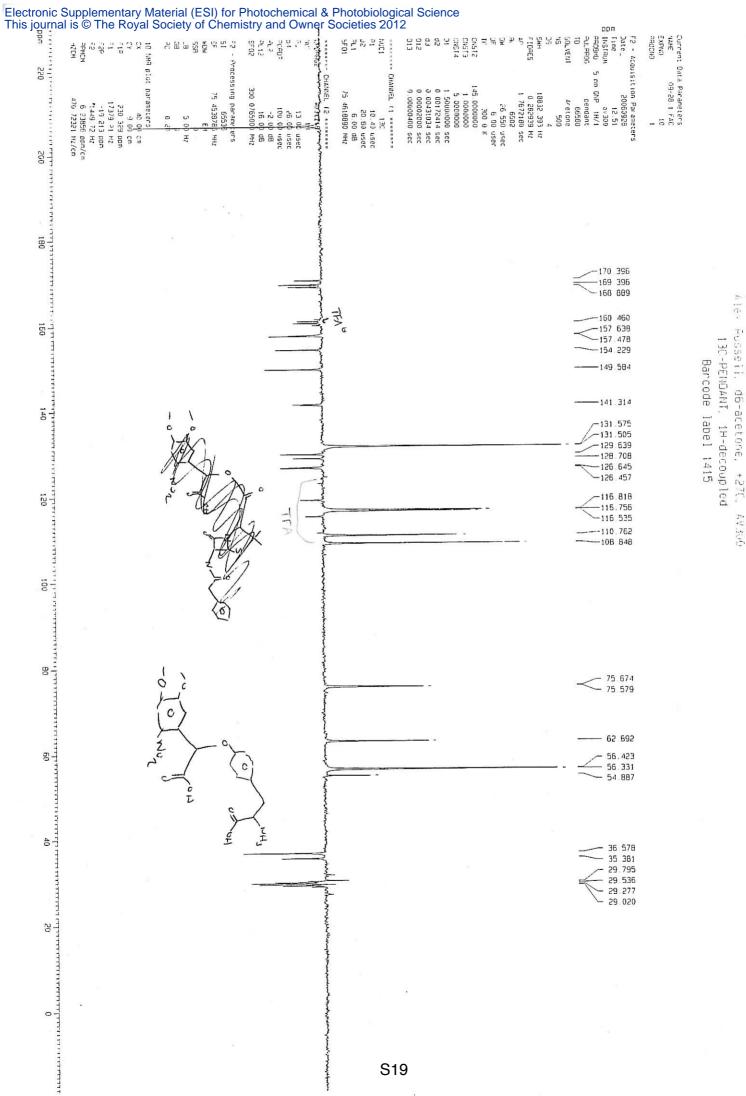




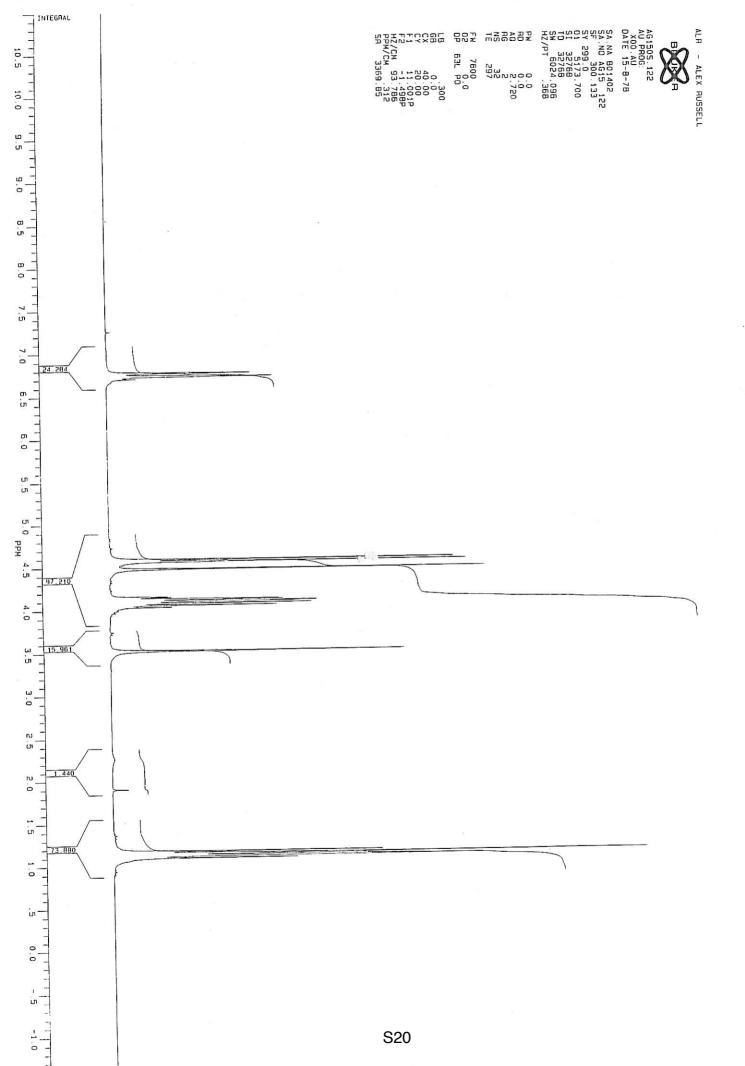


Darry D - Myr.

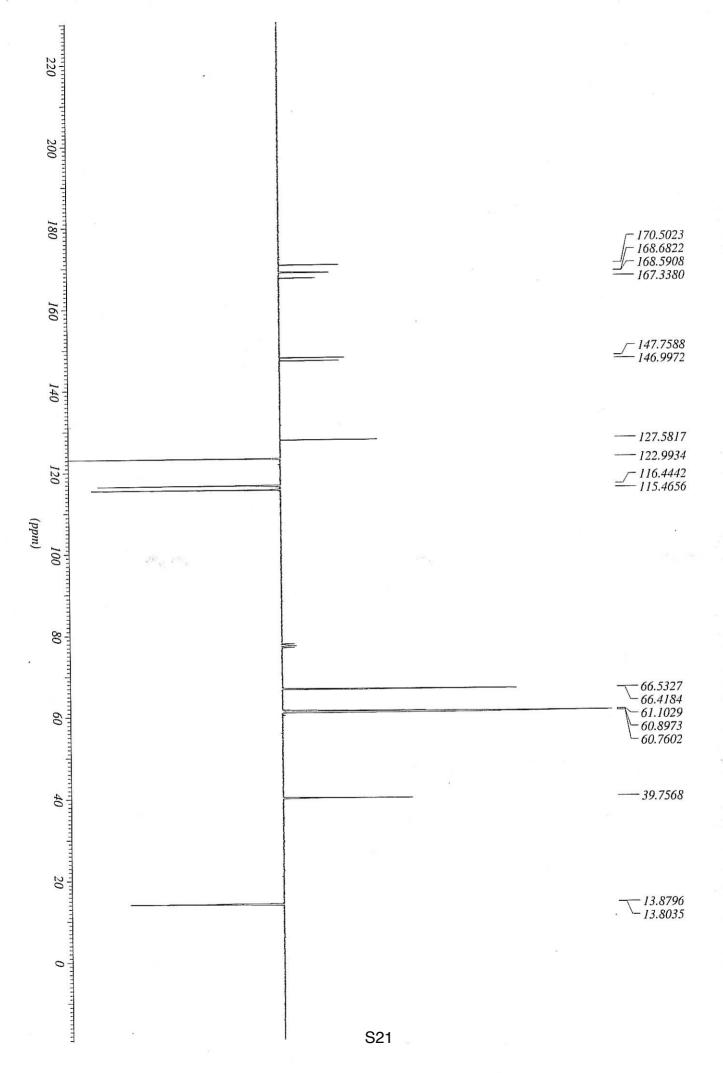
· · · · ·



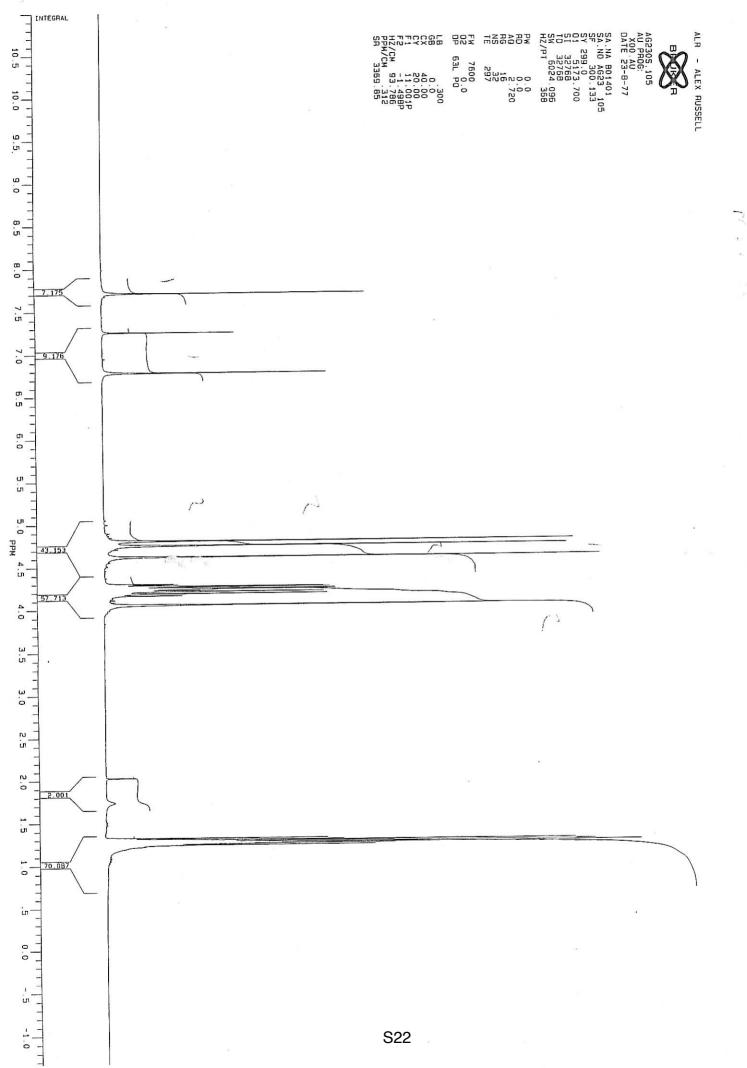
Electronic Supplementary Material (ESI) for Photochemical & Photobiological Science This journal is © The Royal Society of Chemistry and Owner Societies 2012



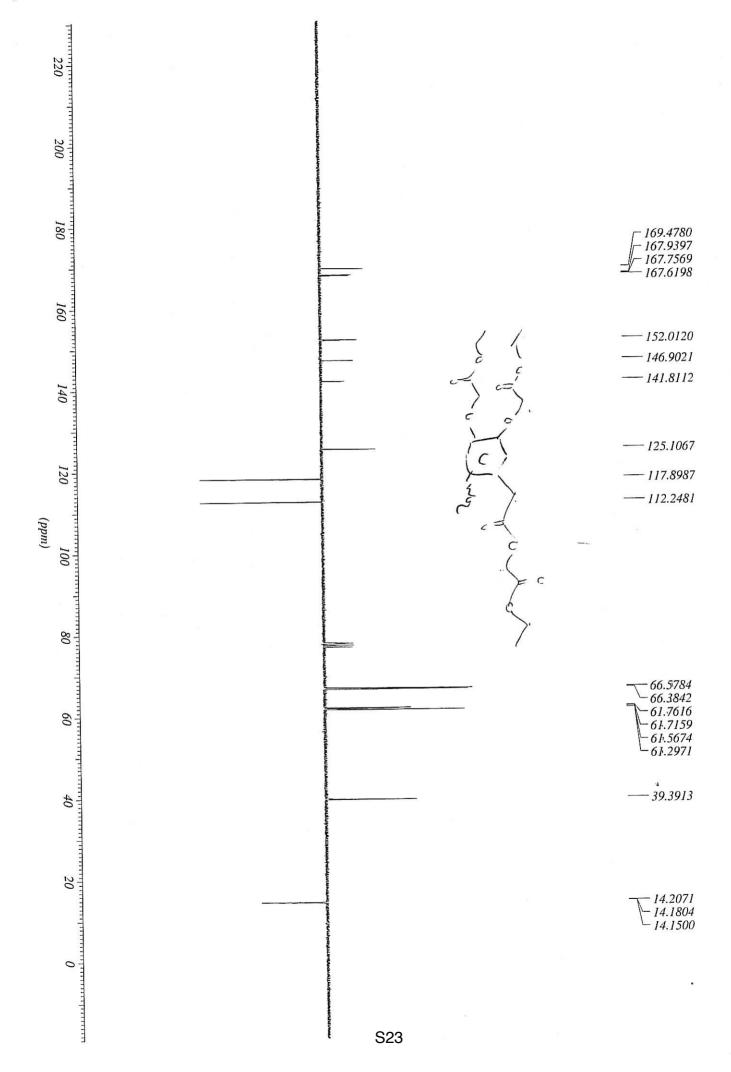
Tet Estar



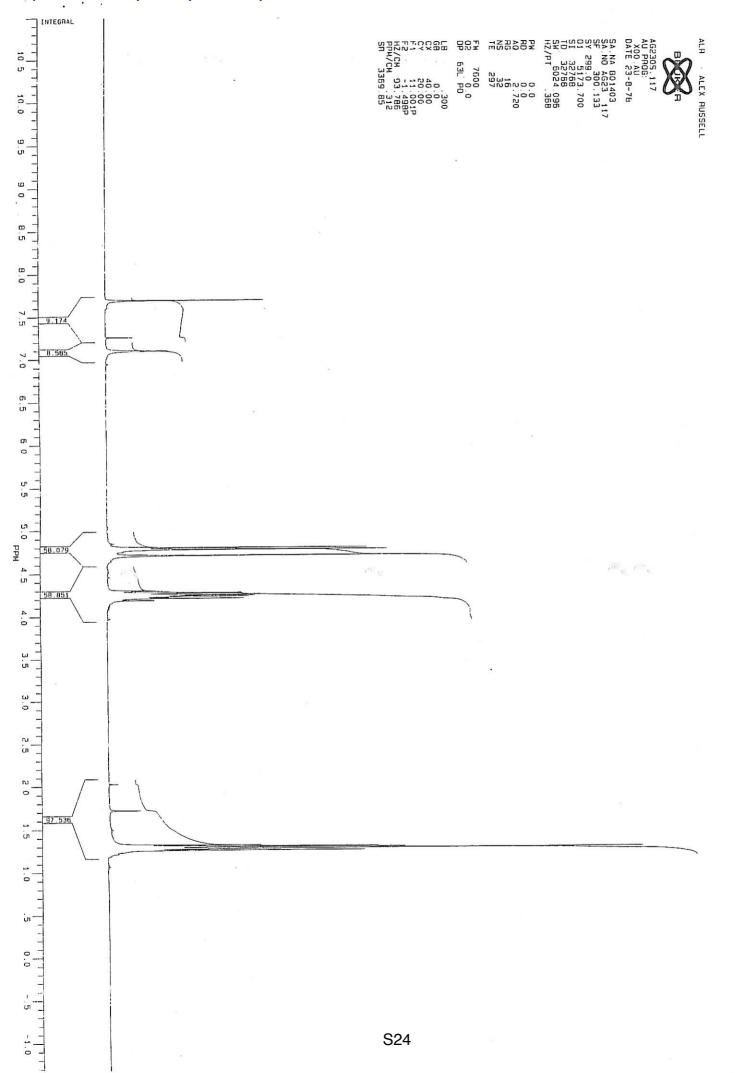
Electronic Supplementary Material (ESI) for Photochemical & Photobiological Science This journal is © The Royal Society of Chemistry and Owner Societies 2012

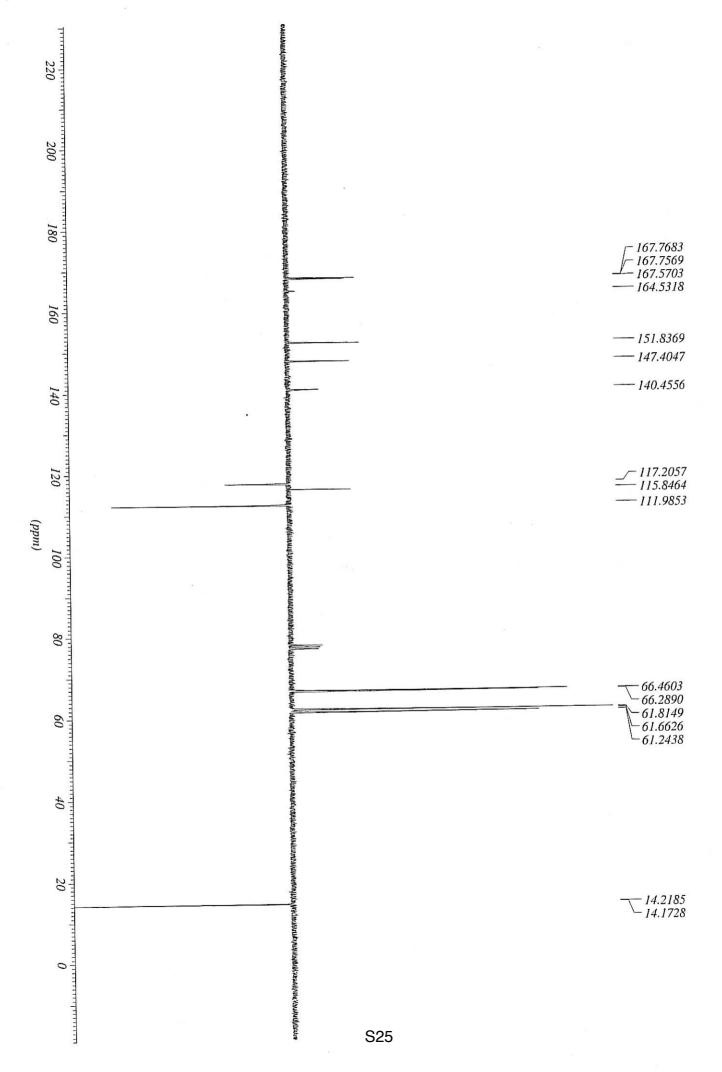


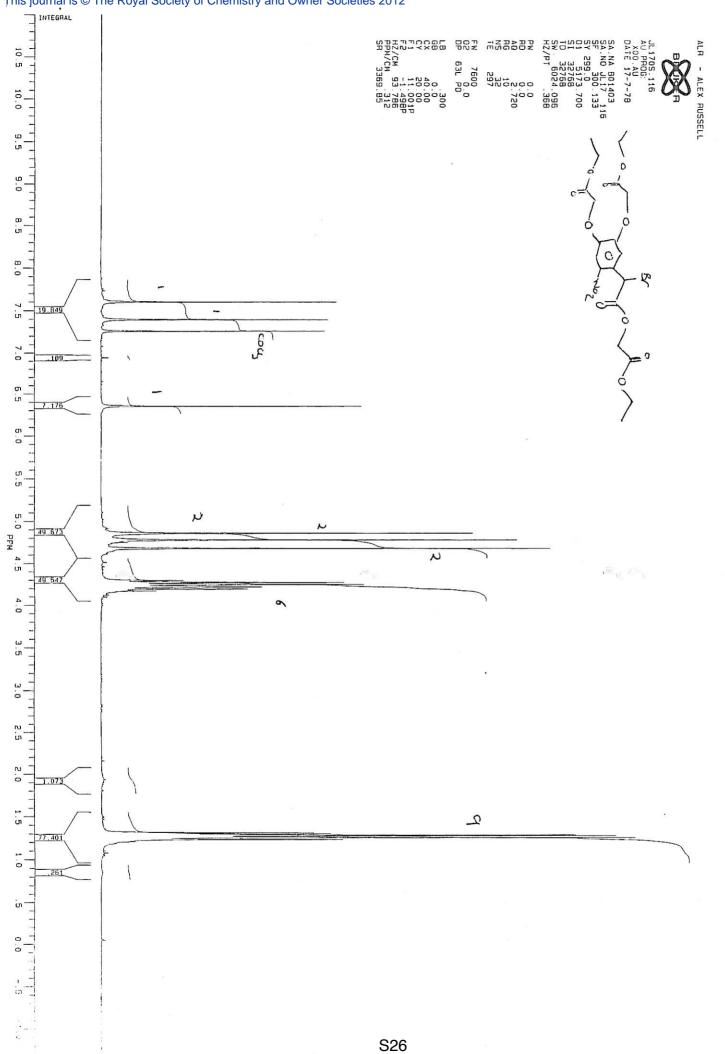
12.0.2

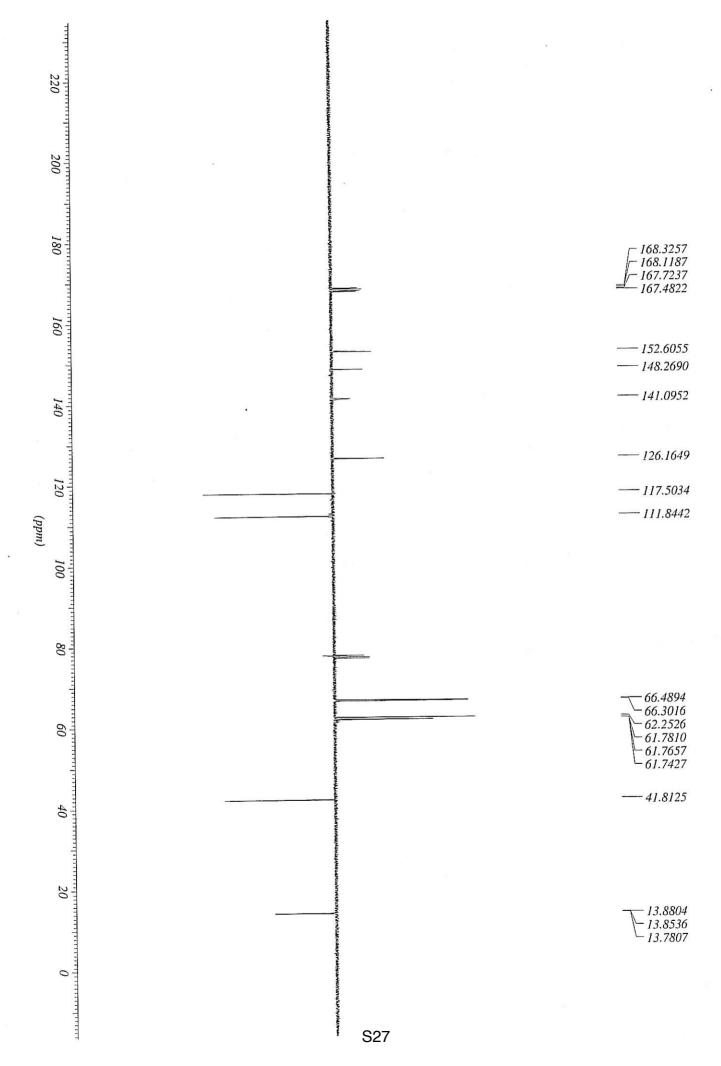


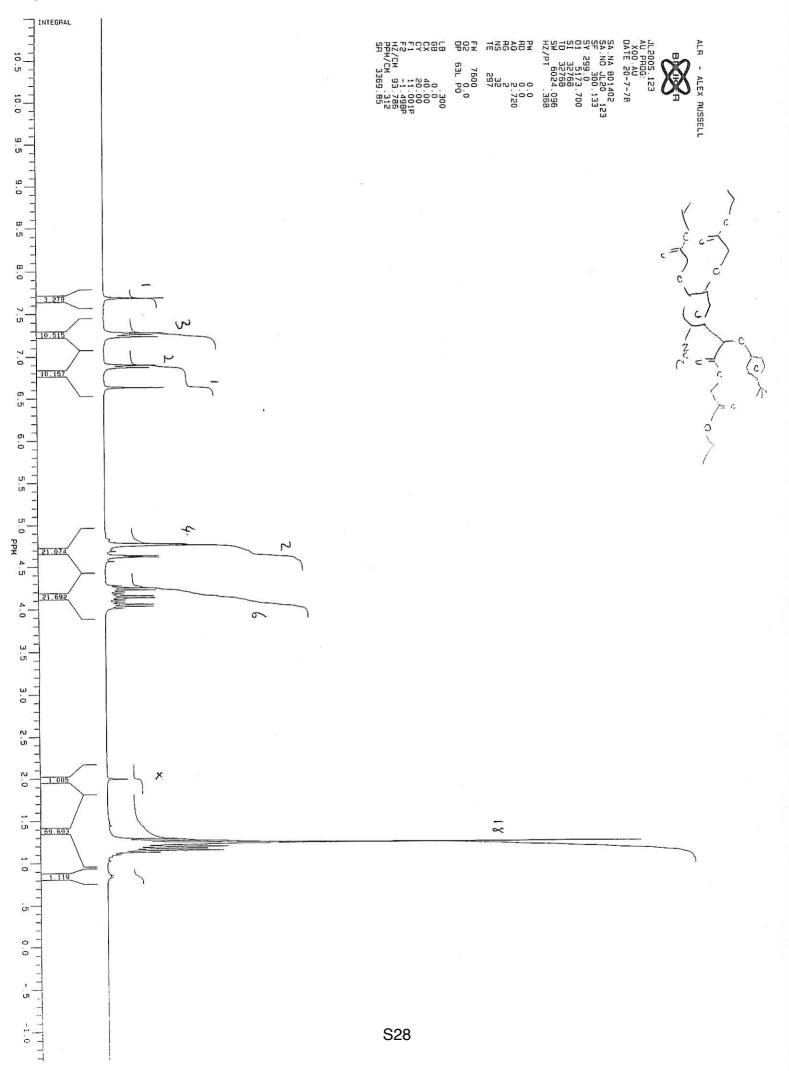
Electronic Supplementary Material (ESI) for Photochemical & Photobiological Science This journal is @ The Royal Society of Chemistry and Owner Societies 2012

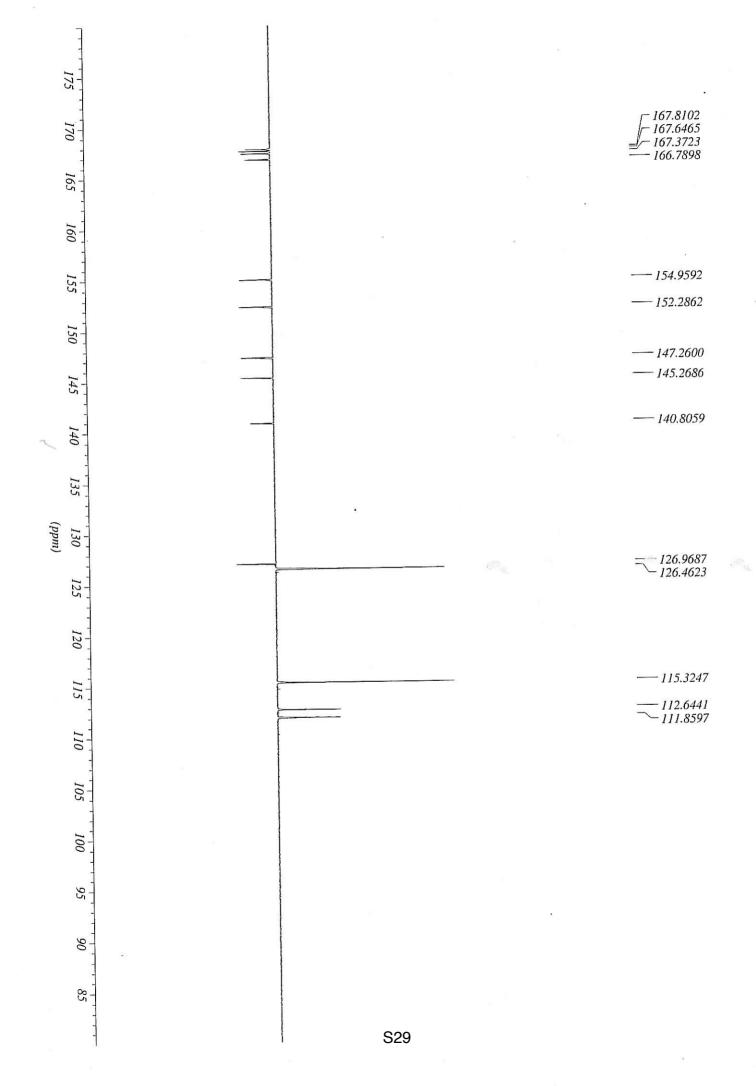


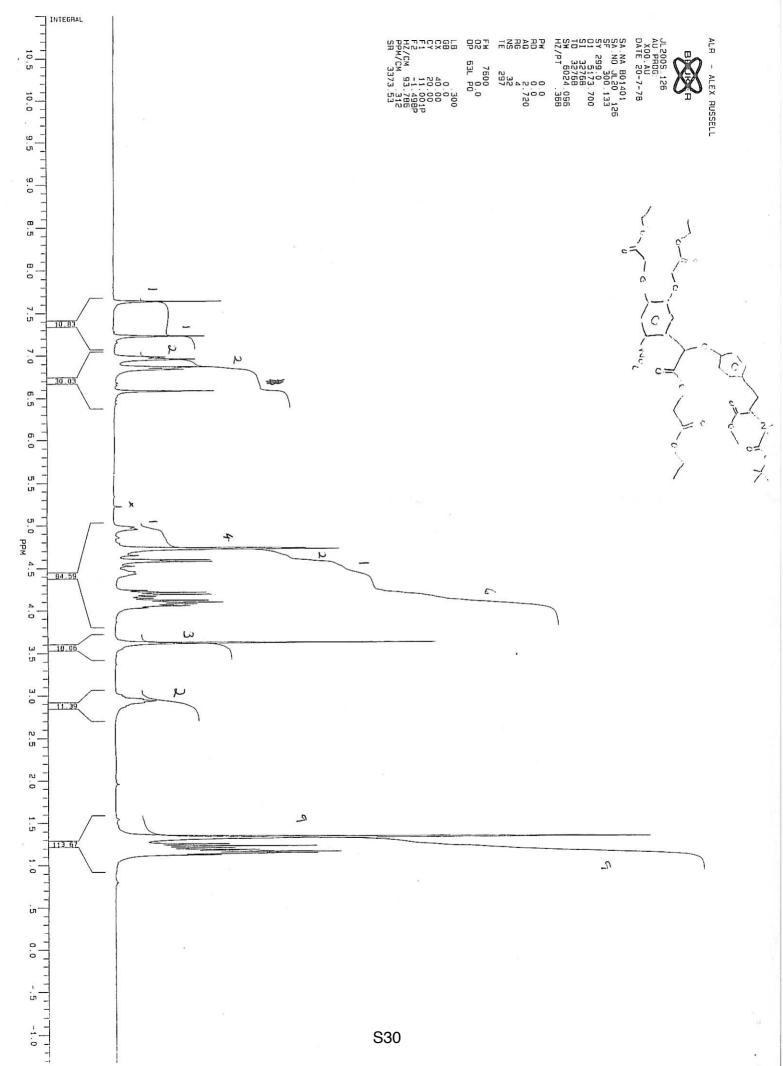


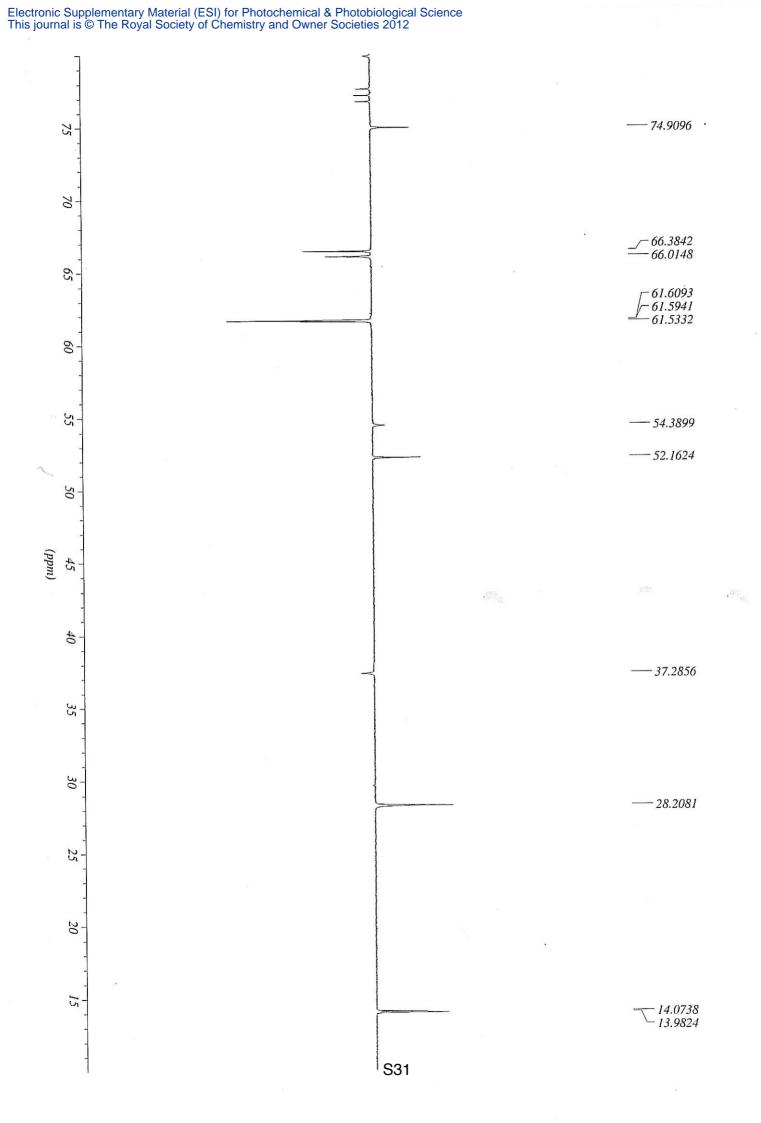


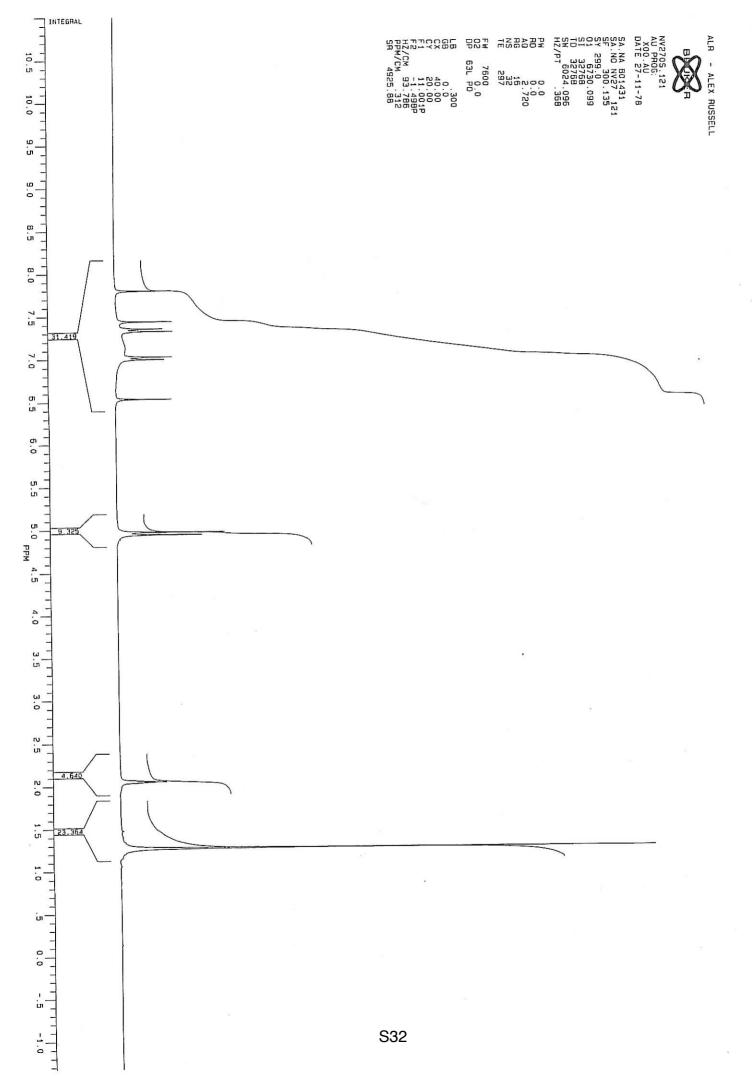




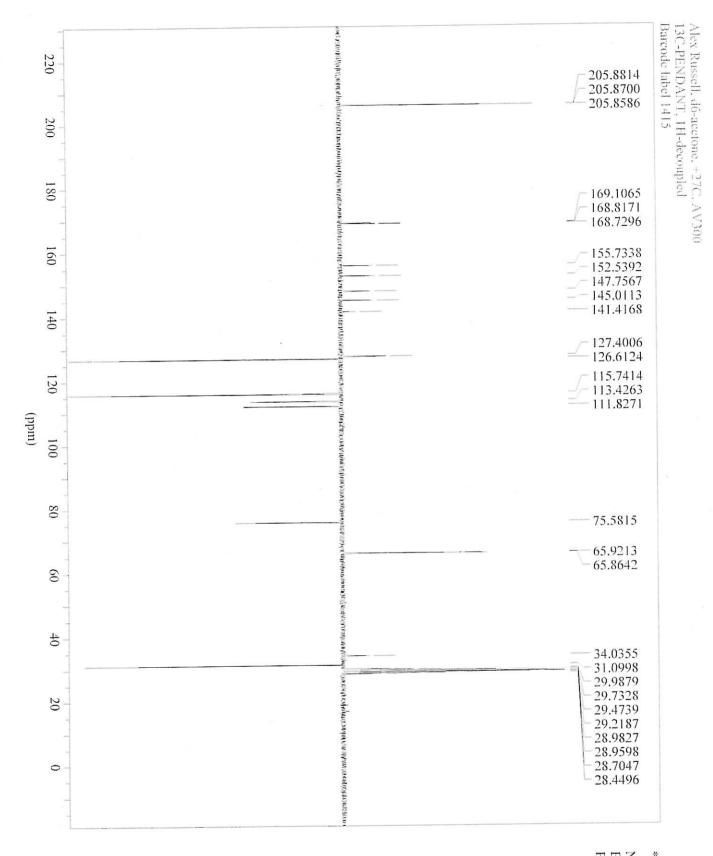




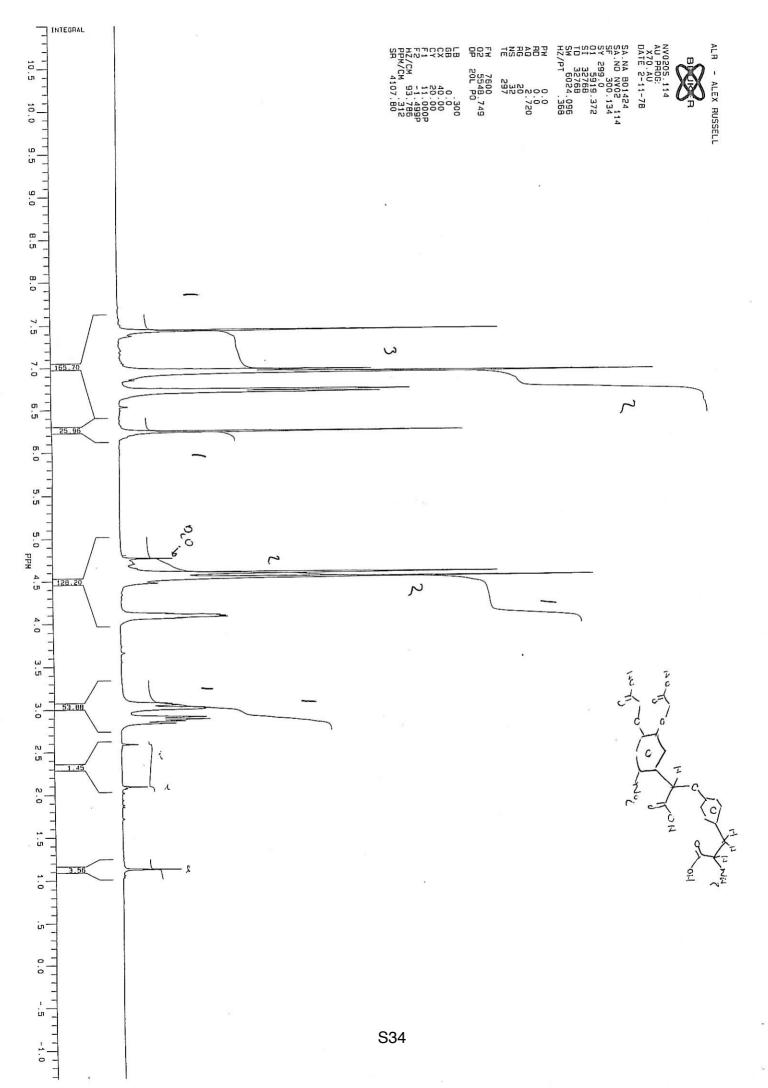


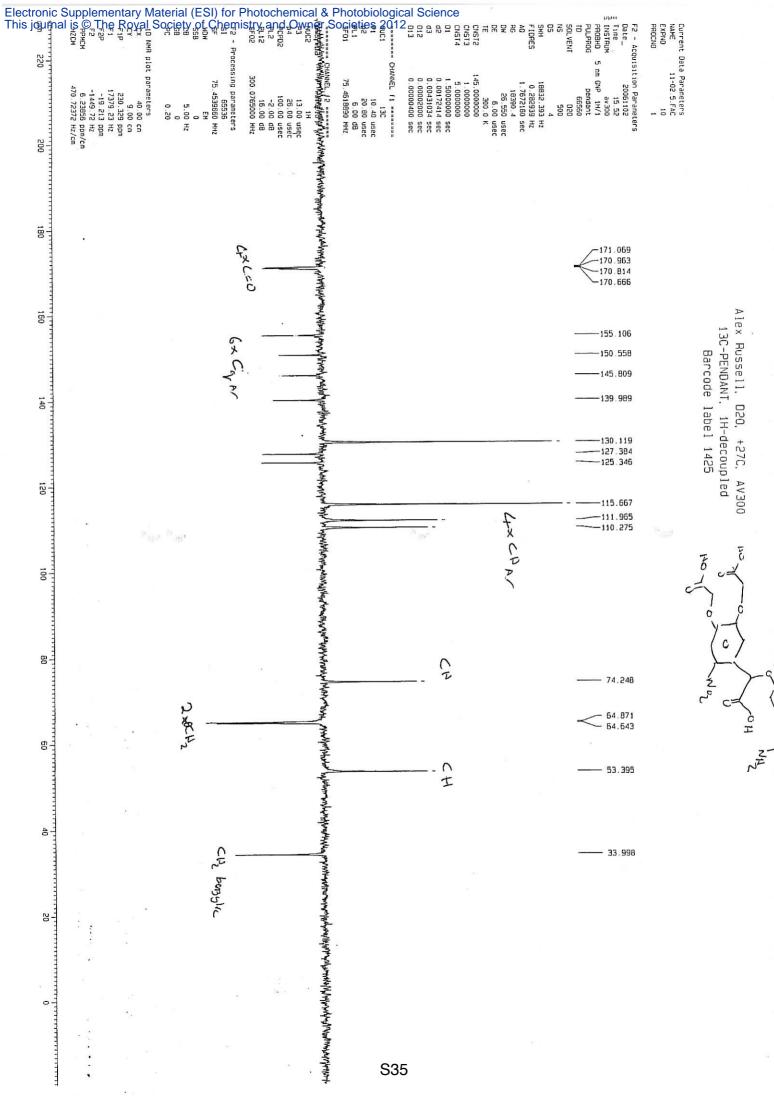


TANKA-Phul Depat

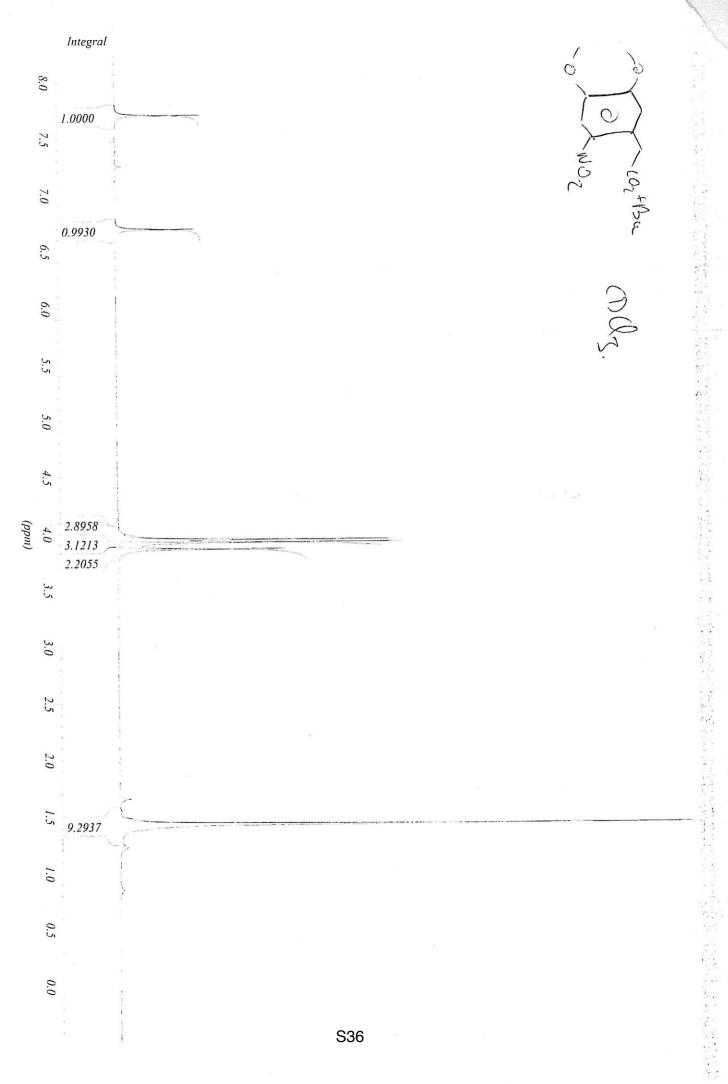


\*\*\* Current Data Parameters \*\*\* NAME : 10 EXPNO : 10 PROCNO : 1

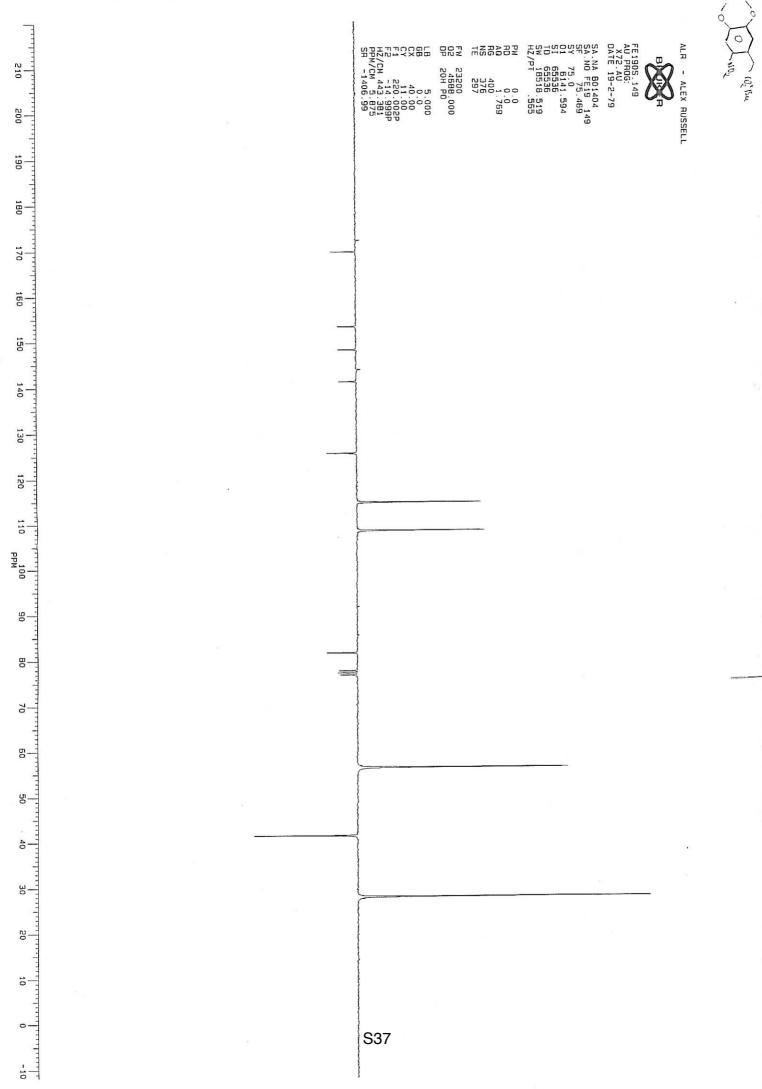


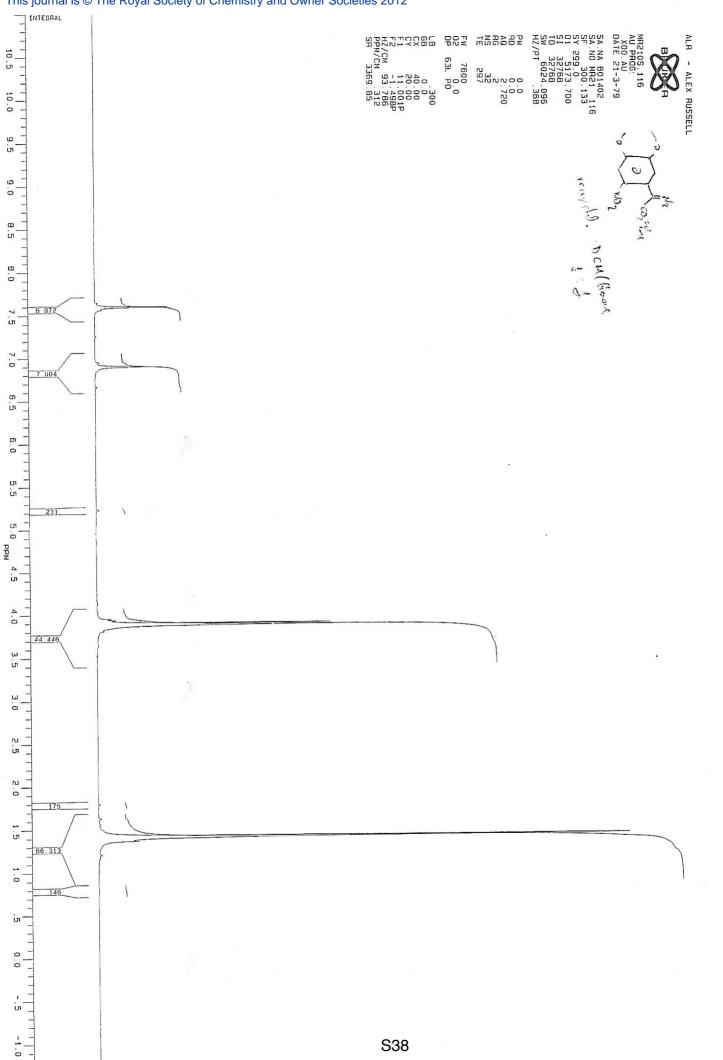


c

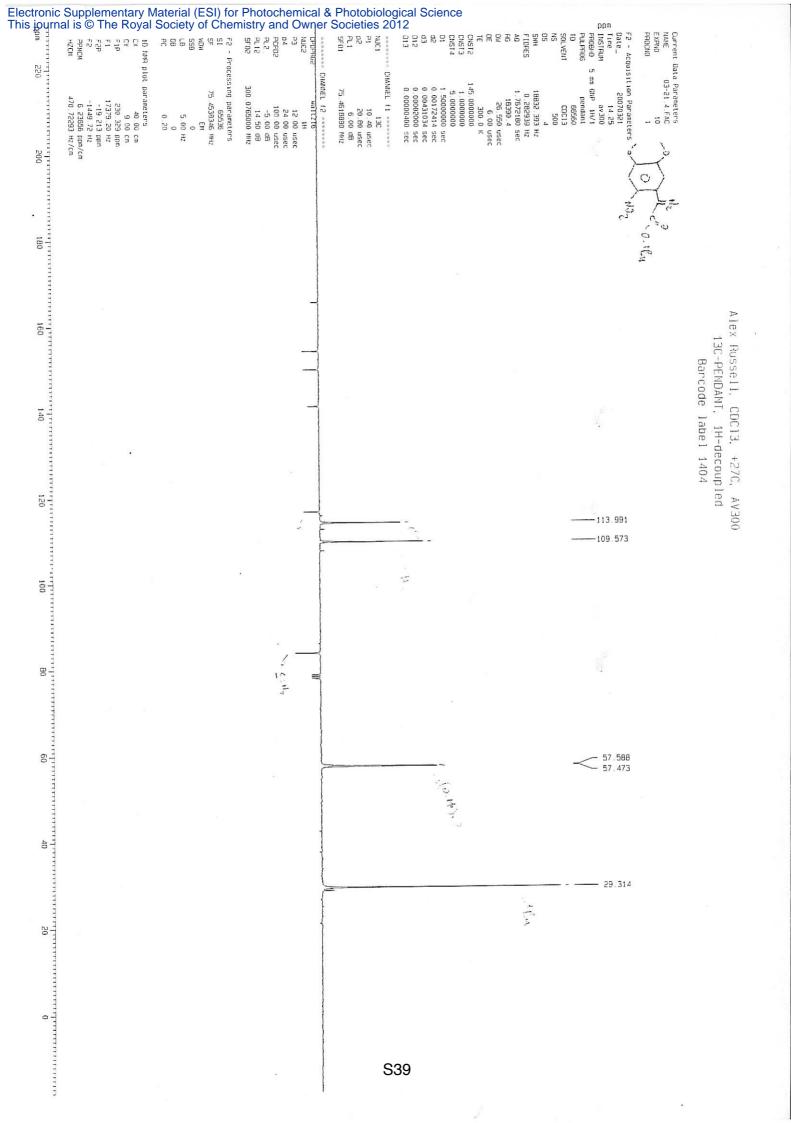


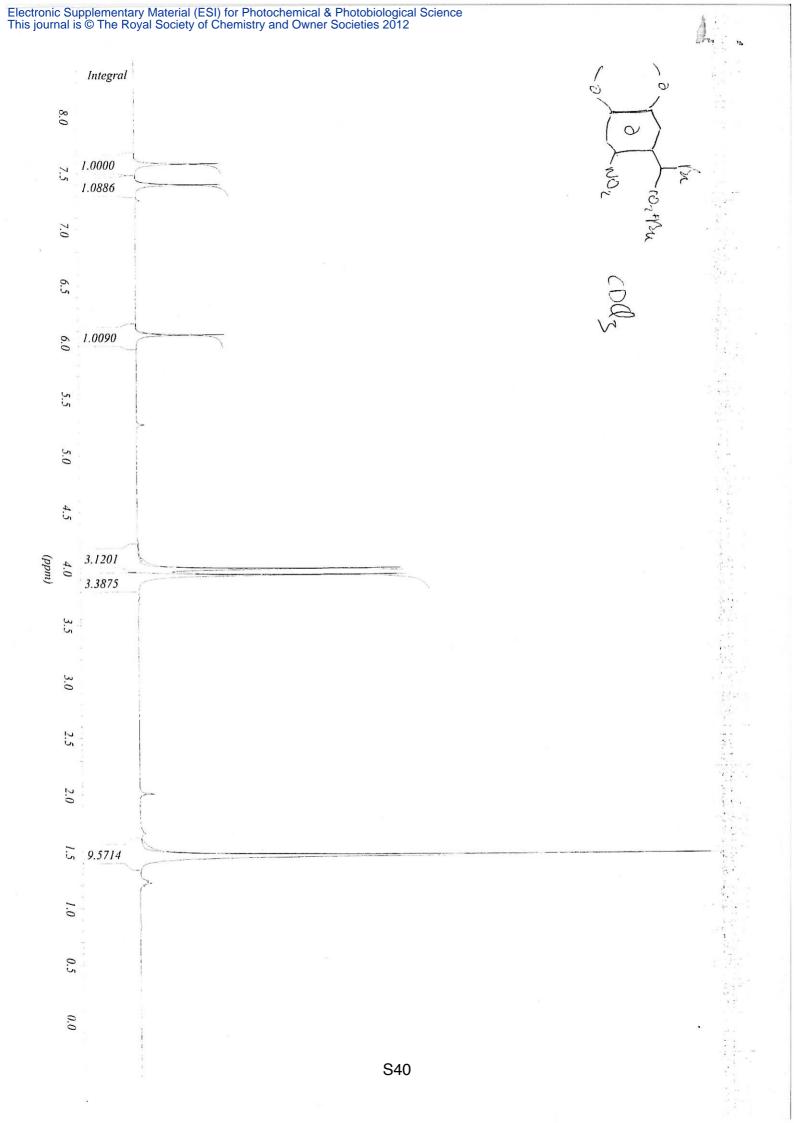
## Electronic Supplementary Material (ESI) for Photochemical & Photobiological Science This journal is © The Royal Society of Chemistry and Owner Societies 2012



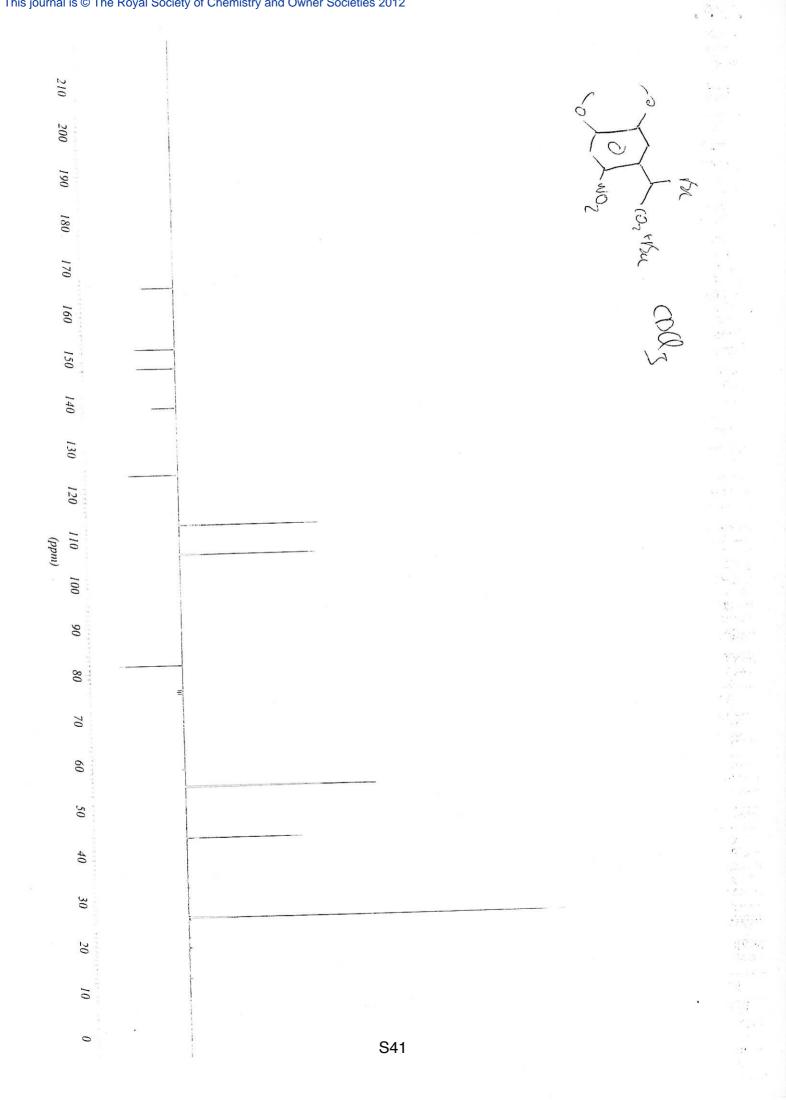


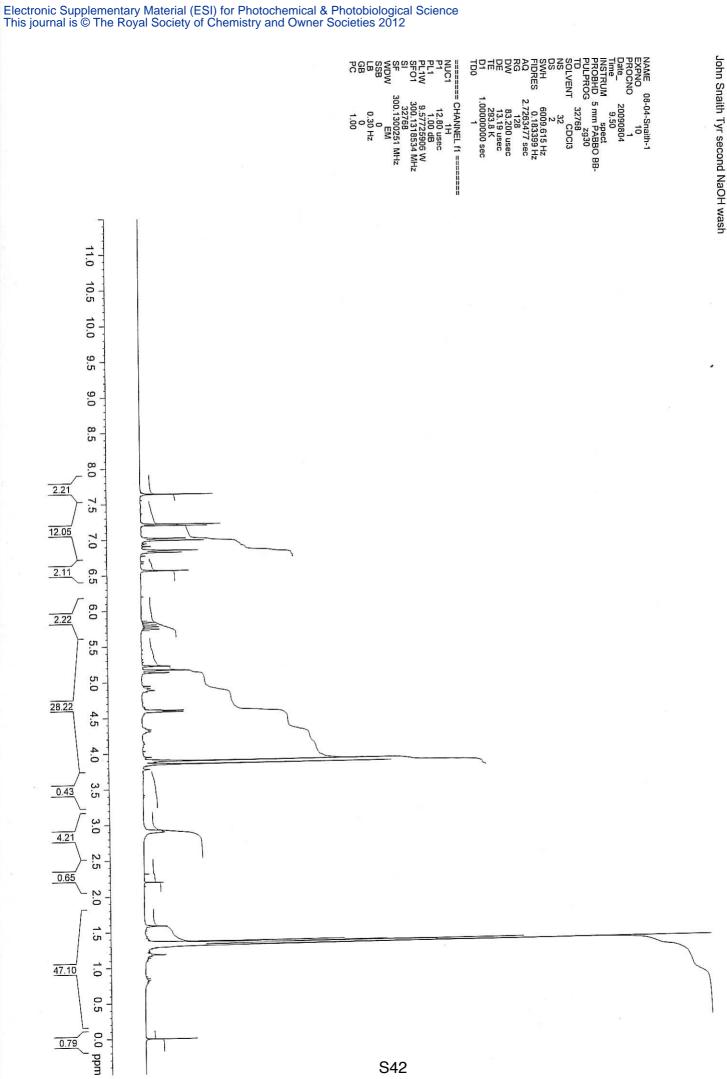
Electronic Supplementary Material (ESI) for Photochemical & Photobiological Science This journal is © The Royal Society of Chemistry and Owner Societies 2012



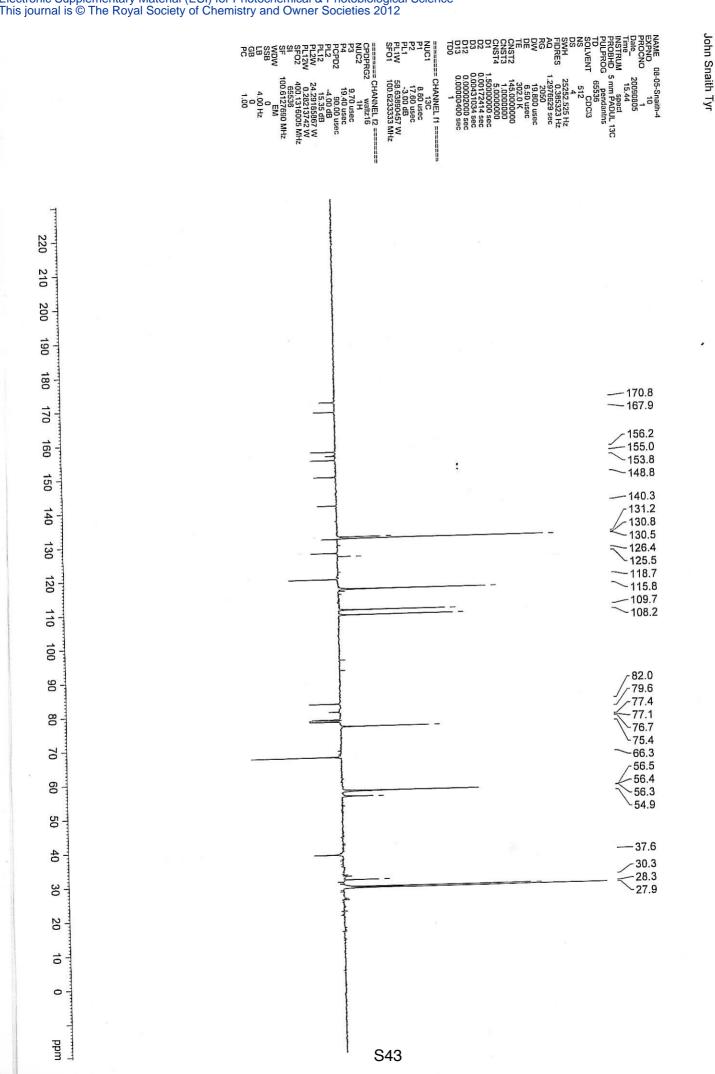


## Electronic Supplementary Material (ESI) for Photochemical & Photobiological Science This journal is O The Royal Society of Chemistry and Owner Societies 2012

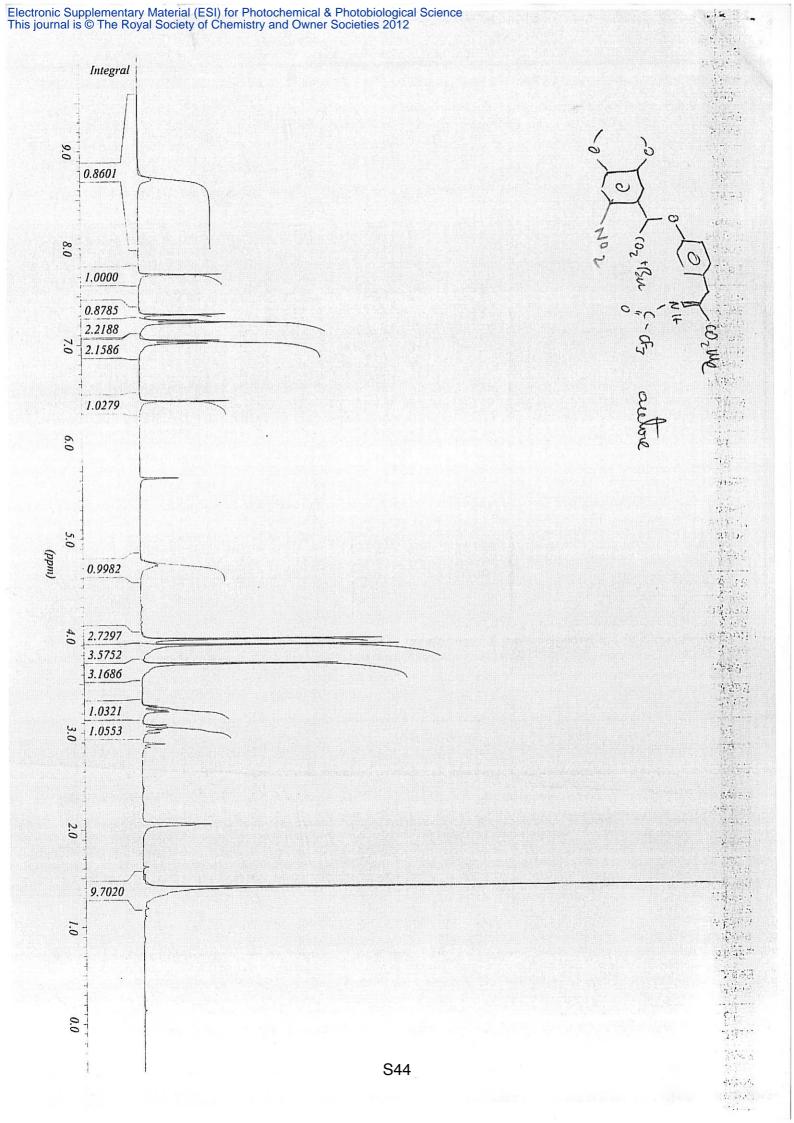


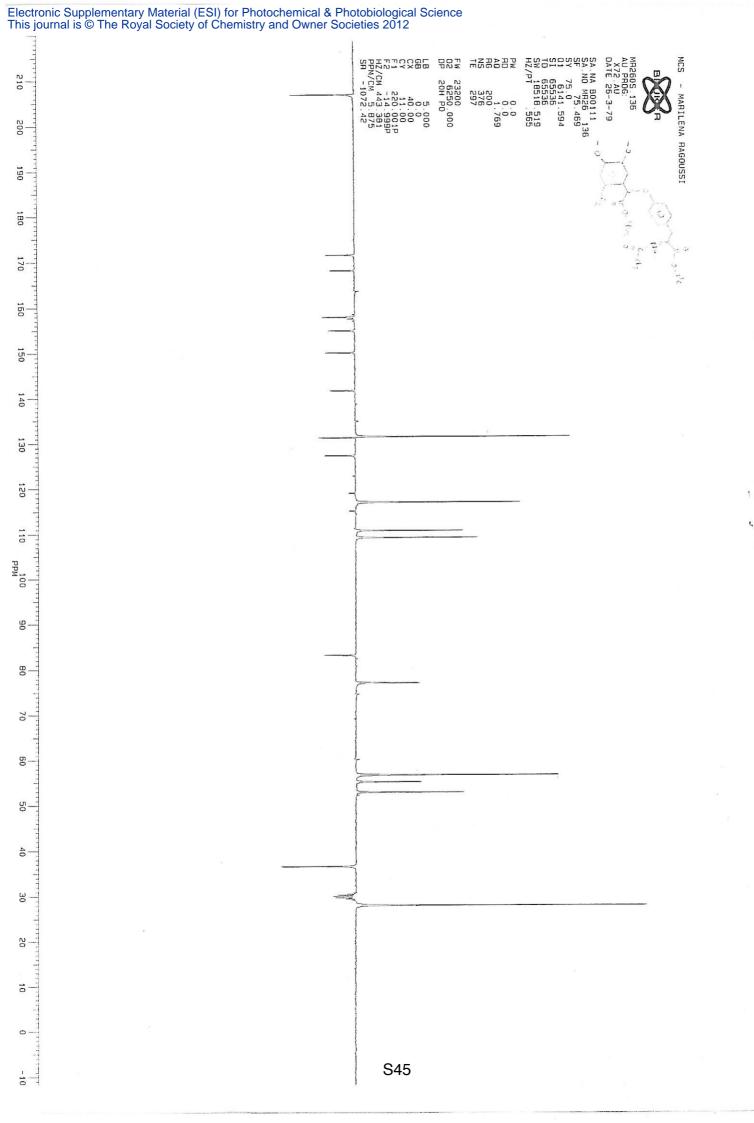


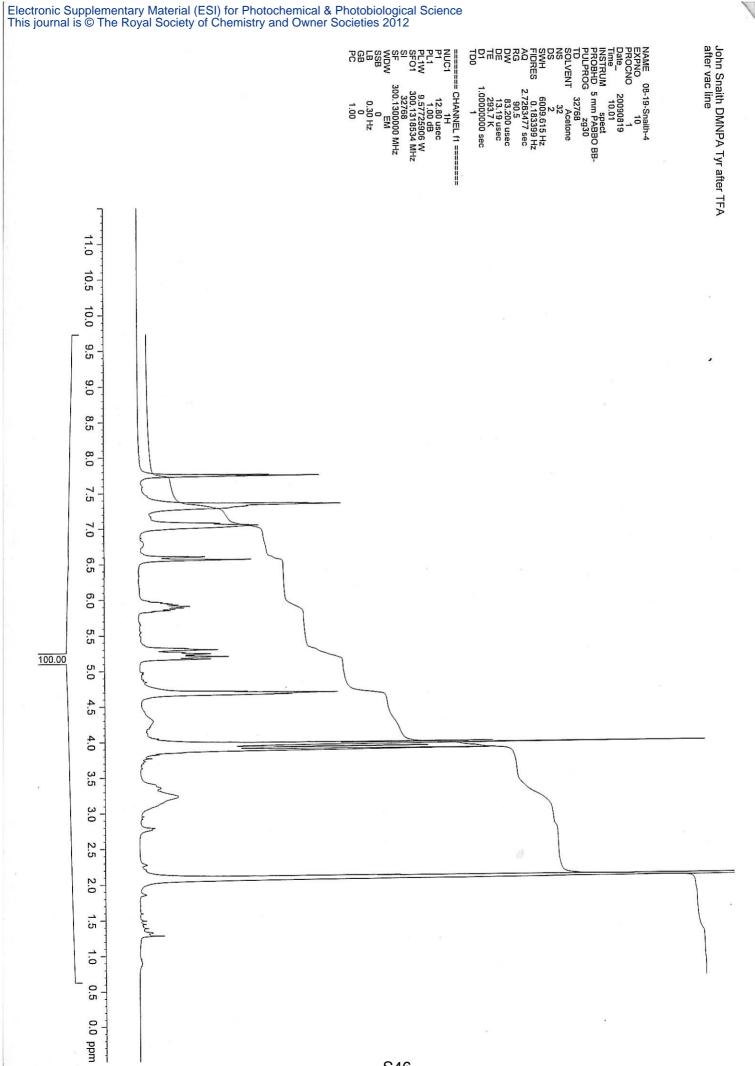
h Twr second NaOH wa



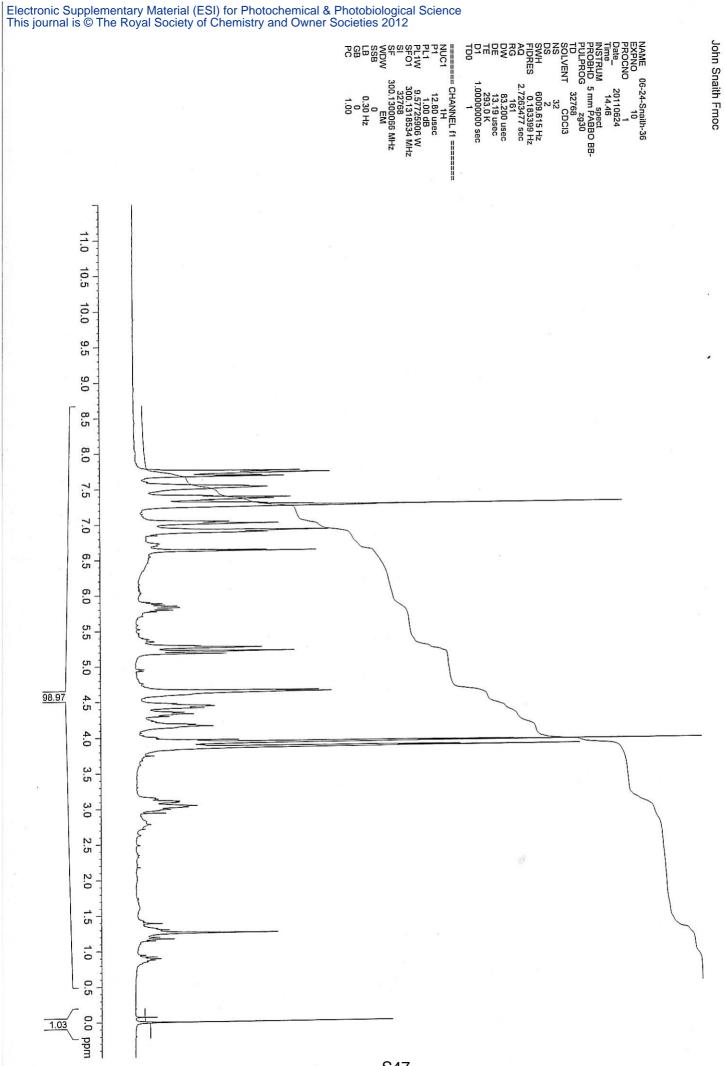
Electronic Supplementary Material (ESI) for Photochemical & Photobiological Science This journal is © The Royal Society of Chemistry and Owner Societies 2012







S46



S47

