## Supporting Information

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- 0.01 M aq.






General procedure for the conjugation of porphyrins to amino-functionalised substrates.
Conjugate 3-poly-L-Lys: A solution of porphyrin $3\left(3 \mathrm{mg}, 4.0 \times 10^{-3} \mathrm{mmol}\right)$ in deionised water was treated with EDC ( $\left.2 \mathrm{mg}, 1.0 \times 10^{-2} \mathrm{mmol}\right)$ and NHS $\left(2 \mathrm{mg}, 1.7 \times 10^{-2} \mathrm{mmol}\right)$ and it was allowed to stand at room temperature for 30 minutes. The solution was then added to a solution of poly-L-lysine ( $30000-70000 \mathrm{MW}$ ) in 0.01 M aqueous $\mathrm{NaHCO}_{3}(20 \mathrm{~mL}$ ). The resulting mixture was shielded from light and shaken for 8 hours on a rotating shaker. The desired conjugate was purified by gel filtration. Methanol was added to the eluate, and the conjugate was recovered by filtration through membrane and dried in vacuo. The desired conjugate was obtained as a light orange-red solid. ( $42 \mathrm{mg}, 84 \%$ recovery). Estimated loading: $398 \mathrm{nmol} / \mathrm{mg}$; UV/vis: ( $\mathrm{H}_{2} \mathrm{O}$ ) $\lambda$ (\%): 401 (100), 517 (13.5), 552 (8.0)

Conjugate 4-poly-L-Lys: The conjugated nanospecies were obtained following the general procedure, using porphyrin $4\left(3 \mathrm{mg}, 3.0 \times 10^{-3} \mathrm{mmol}\right)$. The desired conjugate was obtained as a light red solid. ( $38 \mathrm{mg}, 76 \%$ recovery). Estimated loading: $263 \mathrm{nmol} / \mathrm{mg}$; UV/vis: $\left(\mathrm{H}_{2} \mathrm{O}\right) \lambda$ (\%): 416 (100), 530 (10.0), 568 (3.5).

Conjugate 6-aminodextran: The desired conjugate was obtained following the general procedure, using aminodextran ( $10000 \mathrm{MW}, 100 \mathrm{mg}$ ) and porphyrin $6\left(4 \mathrm{mg}, 3.4 \times 10^{-3}\right.$ mmol ). The desired conjugate was obtained as a dark red solid. ( $72 \mathrm{mg}, 72 \%$ recovery). Estimated loading: $21.2 \mathrm{nmol} / \mathrm{mg}$; UV/vis: ( $\left.\mathrm{H}_{2} \mathrm{O}\right) \lambda(\%): 397$ (100), 513 (11.1), 549 (3.6).

Conjugate 7-aminodextran: The conjugated nanospecies were obtained following the general procedure, using aminodextran (10000 MW, 100 mg ) and porphyrin 7 (4 mg, 3.7 x
$10^{-3} \mathrm{mmol}$ ). The desired conjugate was obtained as a brownish red solid. ( $81 \mathrm{mg}, 81 \%$ recovery). Estimated loading: $17.8 \mathrm{nmol} / \mathrm{mg}$; UV/vis: ( $\left.\mathrm{H}_{2} \mathrm{O}\right) ~ \lambda(\%): 408$ (100), 527 (10), 561 (1.3).

## Spectra




EPSRC National Mass Spectrometry Facility, Swansea
<<LJMGUI004-VM-MAP_0001>> Voyager Spec \#1=>AdvBC(64,0.5,0.1)=>NF0.7[BP = 899.3, 6684]


Dr Giuntini TMPyPPt MW=890?? HFIP PosRef [1:49] (Dith-HFIP)
D:120131Sept13LJMGUIOO4-VM-MAP_0001.dat
EPSRC National Mass Spectrometry Facility, Swansea



## $\stackrel{-82187}{\varepsilon \varepsilon z 8 \dagger}\rangle$




EPSRC National Mass Spectrometry Facility, Swansea
<<LJMGUI005-VM-MAP_0001>> Voyager Spec \#1 $=>$ AdvBC $(64,0.5,0.1)=>$ NF $0.7[B P=810.2,10330]$


EPSRC National Mass Spectrometry Facility, Swansea


0.8

16908 -
0Z91.8~
02L18-
$+91 \varepsilon^{\circ} 8 \sim$
698E8-
ع188:8-
$82 \angle 1 \angle-$
9670.8
$7690 \cdot 8$
029187

$\begin{array}{ll}791 \varepsilon 8 \\ 698.8 & 979 \angle 8 \\ 962 & 729.8-\end{array}$
9t88.8-
$7 \angle 9 \angle 8$
$7 \angle 9 \angle 8=$
$98828=$
EL08.8


FG-6/153 MW=1158?
$(\mathrm{MeOH}) / \mathrm{MeOH}+\mathrm{DEA}$
Francesca Giuntini
05/10/2012 10:19:36
NL:
5.60E5
HULBOY383-OV-HNESN\#2-5
RT: 0.27-0.53 AV: 4 T: FTMS-
p NSI Full ms [120.00-2000.00]


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[^0]FG-6/173 MW = 964 ?
(MeOH)/MeOH + DEA

$\begin{array}{ll}\text { EPSRC National Centre Swansea } & \text { Francesca Giuntini } \\ & 05 / 10 / 201210 \cdot 23: 47\end{array}$






Structures of the porphyrins and nanoconjugates


## Absorption and emission spectra of porphyrins and nanoconjugates

The graphs depicted below represent the absorption (solid lines) and the emission (dashed lines) of the porphyrins (black lines) and the nanoconjugates (dark red lines), as the intensity (normalised arbitrary units) versus wavelength (nanometers).


## $\mathrm{O}_{2}$ calibrations in time-resolved domain

- The samples were held in a four-sided quartz cuvette equipped with a degassing bulb.
- Prior to measurements the sample was completely degassed by a series of freeze-pumpthaw cycles.
- Once degassed the concentration of oxygen in the sample cell was adjusted by backfilling the cell with a known pressure of air, and according to Henry's Law, $\left[\mathrm{O}_{2}\right] \propto \mathrm{p}_{\text {air }}$.
- Phosphorescence lifetime and intensity measurements (in correspondence of the emission $\lambda_{\max }$ ) were performed at six oxygen concentrations within the $0-0.27 \mathrm{M}$ range.
- Phosphorescence lifetimes were recorded using a time-correlated photon counting spectrometer. The samples were excited using the output of a pulsed nitrogen laser (VSL-337i OEM), which provided a 10 Hz pulse train of pulse duration $<5 \mathrm{~ns}$ at 337 nm , with a pulse energy of ca. $5 \mu \mathrm{~J}$. Emission was collected at $90^{\circ}$ to the excitation source and the emission wavelength was selected using a monochromator (Bentham TM300) and the selected light detected using a red-sensitive photon counting photomultiplier module (Hamamatsu H10682-1). The arrival times of multiple photons were recorded from each laser shot using a multi-channel scaler, and the data from a minimum of 10,000 laser shots was acquired to furnish the emission decay.
- Example of calibration: porphyrin 3

| $\mathbf{M}$ | $\mathbf{p}_{\text {air }}(\mathbf{m m H g})$ | $\mathbf{p O}_{\mathbf{2}}(\mathbf{m m H g})^{\mathbf{a}}$ | $\left[\mathbf{O}_{\mathbf{2}}\right](\mathbf{M})^{\mathbf{b}}$ | $\mathbf{I}_{e m}$ | $\mathbf{1 / I _ { e m }}$ | $\boldsymbol{\tau}(\mathbf{s})$ | $\mathbf{1} / \boldsymbol{\tau}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1 | 1.94 | 0.41 | $6.89 \times 10^{-07}$ | 1.00 | 1.00 | $1.03 \times 10^{-05}$ | $9.71 \times 10^{+05}$ |
| 2 | 102.99 | 21.63 | $3.66 \times 10^{-05}$ | 0.49 | 2.03 | $6.04 \times 10^{-06}$ | $1.66 \times 10^{+05}$ |
| 3 | 283.09 | 59.45 | $1.01 \times 10^{-04}$ | 0.24 | 4.14 | $2.74 \times 10^{-06}$ | $3.65 \times 10^{+05}$ |
| 4 | 452.99 | 95.13 | $1.61 \times 10^{-04}$ | 0.17 | 5.85 | $2.09 \times 10^{-06}$ | $4.78 \times 10^{+05}$ |
| 5 | 588.90 | 123.67 | $2.09 \times 10^{-04}$ | 0.15 | 6.63 | $1.56 \times 10^{-06}$ | $6.41 \times 10^{+05}$ |
| 6 | 760.00 | 159.60 | $2.70 \times 10^{-04}$ | 0.10 | 9.65 | $1.15 \times 10^{-06}$ | $8.70 \times 10^{+05}$ |

$\mathbf{M}=$ measurement point
$\mathbf{p}_{\text {air }}=$ air pressure in the sample cell (in mmHg )
$\mathrm{pO}_{2}=$ oxygen pressure in sample cell (in mmHg )
[ $\mathrm{O}_{2}$ ] = molar concentration of oxygen in the sample cell
$\mathbf{I}_{\text {em }}=$ intensity of phosphorescence measured at the emission $\lambda_{\max }(670 \mathrm{~nm}$ for 3)
$\tau=$ triplet lifetime (in seconds)
a.
$\mathrm{pO}_{2}(\mathrm{mmHg})=\mathrm{p}_{\text {air }}(\mathrm{mmHg}) \times 0.21$

$$
\left[\mathrm{O}_{2}\right](\mathrm{M})=\frac{\mathrm{pO}_{2} \mathrm{mmHg}}{\left(760 \mathrm{mmHg} \mathrm{~atm}^{-1}\right) \times\left(757 \mathrm{~atm} \mathrm{M}^{-1}\right)}
$$

0.21: molar fraction of oxygen in air
$760 \mathrm{mmHg} \mathrm{atm}{ }^{-1}$ : conversion factor from Torr to atm
757 atmM $^{-1}$ : Henry's law constant for oxygen in water at $25^{\circ} \mathrm{C}$

- The values $(1 / \tau)$ or ( $1 / \mathrm{I}$ ) were plotted against the oxygen concentration. The $k_{\mathrm{q}}$ values were obtained from the corresponding linear regression equations $\left(1 / \tau=k_{\mathrm{q}}\left[\mathrm{O}_{2}\right]+1 / \tau_{0}\right)$ or $\left(1 / I=k_{q}\left[O_{2}\right]+1 / I_{0}\right)$.
$1 / \tau$ vs. [ $\mathrm{O}_{2}$ ] plots










## $1 / I$ vs. $\left[\mathrm{O}_{2}\right]$ plots










Lifetime and intensity Stern-Volmer plots









Spectrum of the diode lamp used for the photostability experiments



[^0]:    

