Supporting Information for

Poly(*N*-isopropylacrylamide)-poly(ferrocenylsilane) dualresponsive hydrogels: Synthesis, characterization and antimicrobial applications †

Xiaofeng Sui,^a Xueling Feng,^a Andrea Di Luca,^b Clemens A. van Blitterswijk,^b Lorenzo Moroni,^b Mark A. Hempenius,^{*a} G. Julius Vancso^{*a}

^a Materials Science and Technology of Polymers, MESA⁺ Institute for Nanotechnology, University of Twente, 7500 AE Enschede, The Netherlands. Fax: +31 (0)53 489 3823; E-mail: m.a.hempenius@utwente.nl; <u>g.j.vancso@utwente.nl</u>

^b Department of Tissue Regeneration, MIRA Institute for Biomedical Technology and Technical Medicine, University of Twente, P.O. Box 217, 7500 AE Enschede, The Netherlands.



Fig. S1
FTIR spectra of (a) PFS 2, (b) PNIPAM-2% PFS hydrogel 3 s, (c) PNIPAM-2% PFS hydrogel 60 s, (d) PNIPAM-2% PFS hydrogel 180 s, (e) PNIPAM-2% PFS hydrogel 30 min and (f) PNIPAM hydrogel.



Fig. S2 Photographs of the formed gels: (a) PNIPAM-1% PFS, (b) PNIPAM-2% PFS and (c) PNIPAM-5% PFS.



Fig. S3 Frequency dependence of storage moduli (G') and loss moduli (G") for the obtained PNIPAM-2% PFS hydrogel.



Fig. S4 DSC measurements of the various hydrogels: (a) PNIPAM, (b) PNIPAM-1% PFS-, (c)
PNIPAM-2% PFS, (d) PNIPAM-5% PFS, (e) PNIPAM-1% PFS after oxidation, (f)
PNIPAM-2% PFS after oxidation and (g) PNIPAM-5% PFS after oxidation.



Fig. S5 SEM image and EDS spectrum of a 2% PFS-PNIPAM/silver composite hydrogel.



Fig. S6 UV-VIS absorption spectra of silver nanoparticles synthesized inside the hydrogel.



Fig. S7 Optical microscopy images of MC-3T3 growing in contact with gels after 3 days. Scale bar = 1 mm.