Supporting Information

Fluorescent conjugated polymer based on thiocarbonyl quinacridone for sensing mercury ion and bioimaging

Yi Qu,^{a,b} Xinran Zhang,^a Yongquan Wu,^b Fuyou Li^b* and Jianli Hua^a*

^aKey Laboratory for Advanced Materials and Institute of Fine Chemicals, East China University of Science and Technology, 130 Meilong Road, Shanghai 200237, P. R. China

^bDepartment of Chemistry & Laboratory of Advanced Materials, Fudan University, Shanghai, 200433, P. R. China

Emails: jlhua@ecust.edu.cn (Prof. Jianli Hua); fyli@fudan.edu.cn (Prof. Fuyou Li)

Experimental Section

Characterization

The ¹H NMR spectra were recorded on Brucker AM-400 spectrometers with tetramethylsilane (TMS) as the internal standard. The FT-IR spectra were recorded on a Nicolet 380 thermo Electron corporation spectrometer. UV–vis absorption spectra were performed on a Varian Cray 500 spectrophotometer and fluorescence spectra were recorded on a Varian Cray Eclipse fluorescence spectrophotometer. Molecular weights and polydispersity indices of the polymers were estimated in THF by a Waters Associated Gel Permeation Chromatograph (GPC) system. A set of monodisperse polystyrene standards covering the molecular weight range of 10^3 – 10^7 was used for the molecular weight calibration.

Materials and Reagents

Monomer **1** was synthesized according to reference.¹ Other chemicals and reagents were purchased from Sigma-Aldrich. All reagents were used without further purification.

Synthesis

Polymerization of PQA

To a 25 mL Schelenk tube were added (1) (87.3 mg, 0.13 mmol), (2) (82.0 mg, 0.13 mmol), 2 M aqueous potassium carbonate solution (5 mL), and dry THF (10 mL) under argon. After addition of tetrakis(triphenylphosphine)palladium(0) (6.0 mg, 0.006 mmol) as a catalyst, the mixture was heated to 82 °C. The mixture was then stirred for 20 h, after it was poured into methanol-H₂O (220 mL, 10/1, v/v). The precipitate was isolated by filtration and then washed

with methanol and acetone several times. The polymer was obtained after drying to yield a brick red solid of **PQA** 95 mg (77 %).

¹H NMR (400 MHz , CDCl₃ , ppm): δ 0.74 – 0.82 (br, -*CH*₃), 0.89 – 0.97 (br, -*CH*₃), 1.06 – 1.22 (br, -*CH*₂-), 1.31 – 1.47 (br, s, -*CH*₂-), 1.47 – 1.56 (br, -*CH*₂-), 1.64 – 1.78 (br, CH-*CH*₂-), 1.97 – 2.24 (br, NCH₂-*CH*₂-), 4.47 – 4.77 (br, N-*CH*₂-), 7.63 – 7.72 (ArH) , 7.72 – 7.83 (ArH), 7.83 – 7.93 (ArH), 8.08 – 8.22 (ArH), 8.84 – 9.01 (ArH).

General Procedure of Thiolated Reaction Between PQA and Lawesson's Reagent.

To a mixture of **PQA** and Lawesson's reagent with corresponding feed ratios, degassed toluene (10 mL) was added under argon. The mixture was heated to 125 °C and stirred in the dark for 2 h. After workup, the mixture was poured into methanol. The precipitated fibers were collected by filtration and then washed with methanol for four times. The polymer was purified by Soxhlet extraction in acetone for 24 h. The reprecipitation procedure in $CH_2Cl_2/MeOH$ was then repeated for several times. The final product was obtained after drying in vacuum.

Polymer **PTQA1**. **PQA** (98.5 mg, 0.1 mmol), Lawesson's reagent (16.1 mg, 0.04 mmol) were used in the polymerization. Brown **PTQA1** (95.5 mg) was obtained.

Polymer **PTQA2**. **PQA** (98.5 mg, 0.1 mmol), Lawesson's reagent (24.2 mg, 0.06 mmol) were used in the polymerization. Green **PTQA2** (96.0 mg) was obtained.

Polymer **PTQA3**. **PQA** (98.5 mg, 0.1 mmol), Lawesson's reagent (40.4 mg, 0.10 mmol) were used in the polymerization. Greenish black **PTQA3** (96.1 mg) was obtained.

PTQA1~PTQA3 showed similar ¹H NMR results.

For example: **PTQA2**. ¹H NMR (400 MHz , CDCl₃ , ppm): δ 0.62 - 0.99 (br, -*CH*₃), 1.16 (br, -*CH*₂-), 1.31 - 1.80 (br, -*CH*₂-), 2.08 (br, CH-*CH*₂- and NCH₂-*CH*₂-), 4.64 (br, N-*CH*₂-), 7.33 - 8.18 (br, *ArH*), 8.87 (br, C(=O)-*ArH*), 9.45 (br. C(=S)-*ArH*).

Fabrication of PTQA-NPs:

The **PTQA-NPs** were prepared by a modified desolvation method. Briefly, **PTQA2** (1.0 mg) was dissolved in THF (1 mL) and stirred overnight. Then, the polymer solution (200 μ L, 50 ppm) was injected into PBS buffer (20 mL) under ultrasonication using a microtip probe sonicator. THF was evaporated by N₂ flow at 50 °C, and the solution was filtrated through a 0.2 μ m filter.



Figure S1. ¹HNMR spectra of **PQA** and **PTQA1~5** with different thiolation rate in aromatic region. (solvent: CDCl₃)

Table S1. Integrations of ArH adjacent to C=O and C=S moieties

No.	$\int ArH_{C=S}$	$\int ArH_{C=0}$	$\int ArH_{C=S} / \left(\int ArH_{C=S} + \int ArH_{C=O} \right)$	
PTQA1	1.0	2.8	26.3%	
PTQA2	1.0	0.5	66.7%	
PTQA3	1.0	0.0	100%	



Figure S2. FT-IR spectra of PTQA1 and PTQA2.

	1		
No.	M _n	M _w	PDI
POA	4849	5674	1 17
1 211	1019	5071	1.17
ΡΤΟΔ1	4038	5106	1.26
IIQAI	4050	5100	1.20
PTOA2	2070	5147	1 20
I I QA2	5919	5147	1.29
DTOA2	4169	5000	1 41
r i QA3	4108	3888	1.41

Table S2. GPC data of polymers PQA and PTQA1~3



Figure S3. (a) Emission spectra of PTQA2 upon addition of different metal ions in chloroform $([PTQA2]=10 \ \mu g/mL)$ (b) Emission spectra of PTQA-NPs upon addition of different metal ions in PBS solutions ([PTQA-NPs]=10 \ \mu g/mL).



Figure S4. The Z-scan confocal image of the living HeLa cells incubated with PTQA-NPs (10 μ g/mL)