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Responsive cross-linked supramolecular polymer network: hierarchical supramolecular polymerization driven by cryptand-based molecular recognition and metal-coordination

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1. Materials and Methods

 2^{S1} was synthesized according to literature procedures. All reagents were commercially available and used as supplied without further purification. Solvents were either employed as purchased or dried according to procedures described in the literature. NMR spectra were recorded with a Bruker Advance DMX 500 spectrophotometer or a Bruker Advance DMX 400 spectrophotometer with the deuterated solvent as the lock and the residual solvent or TMS as the internal reference. ¹H NMR chemical shifts are reported relative to residual solvent signals, and ³¹P{¹H} NMR chemical shifts are referenced to an external unlocked sample of 85% H_3PO_4 (δ 0.0). The two-dimensional diffusion-ordered (2D DOSY) NMR spectra were recorded on a Bruker DRX500 spectrometer. Mass spectra were recorded on a Micromass Quattro II triple-quadrupole mass spectrometer using electrospray ionization with a MassLynx operating system or a Bruker Esquire 3000 plus mass spectrometer (Bruker-Franzen Analytik GmbH, Bremen, Germany) equipped with an ESI interface and an ion trap analyzer. Dynamic light scattering (DLS) was carried out on a Malvern Nanosizer S instrument at room temperature. Scanning electron microscopy (SEM) investigations were carried out on a JEOL 6390LV instrument. The sample for SEM image was prepared by drawing out macroscopic fibers from highly concerntrated CH_3CN/CH_2Cl_2 (1:1, v/v) solution of CSPN ($c \approx 1.00$ M) and then placing them on silicon wafer.





^{3.} Synthesis of tri-cryptand 1

Scheme S1. Synthesis of tri-cryptand host 1



Cryptand **3** (9.59 mg, 15.0 mM) and **2** (6.18 mg, 5.00 mM) were mixed together in CD₂Cl₂ at room temperature for 30 minutes to give host **1**. The ¹H NMR spectrum of **1** is shown in Figure S3. ¹H NMR (400 MHz, CD₂Cl₂, 293K) δ (ppm): 9.29–9.30 (m, 6H), 8.64 (s, 3H), 6.74–6.99 (m, 21H), 5.13–5.22 (m, 12H), 4.01–4.06 (m, 24H), 3.75–3.79 (m, 24H), 3.64–3.67 (m, 24H), 1.75–1.77 (m, 36H), 1.10–1.18 (m, 54H). The ³¹P {¹H} NMR spectrum of **1** is shown in Figure S4. ³¹P {¹H} NMR (s, 161.8 MHz, dichloromethane-*d*₂, 293K) δ (ppm): 15.57 (s, ¹⁹⁵Pt satellites, ¹*J*_{Pt-P} = 2284.6 Hz). ESI-MS for **1** is shown in Figure S5: *m/z* 1754.56 for [**1** – 20Tf]²⁺, 1120.05 for [**1** – 30Tf]³⁺



Figure S3. ¹H NMR spectrum (400 MHz, CD₂Cl₂, 293 K) of 1.



Figure S4. ³¹P{¹H} NMR spectrum (161.8 MHz, CD₂Cl₂, 293 K) of 1.



Figure S5. Electrospray ionization mass spectra of 1.

4. Comparison of ¹H NMR of **1** and **3** and ³¹P NMR spectra of **1** and **2**



Figure S6. Partial ¹H NMR spectra (400 MHz, CD₂Cl₂, 293 K) of (a) tri-cryptand host **1** and (b) cryptand **3**.



Figure S7. ³¹P{¹H} NMR spectra (161.8 MHz, CD₂Cl₂, 293 K) of (a) tri-arm acceptor **2** and (b) tri-cryptand host **1**

5. Partial ¹H NMR spectra of $3_2 - 4$



Figure S8. Partial ¹H NMR spectra (400 MHz, CD₂Cl₂/CD₃CN (1:1, *v/v*), 293 K): (a) 4.00 mM **4**; (b) 8.00 mM **3** + 4.00 mM **4**; (c) 8.00 mM **3**.

- **Display Report** Analysis Info Acquisition Date Operator Instrument 12/06/13 19:37:42 Default esquire3000plus 13120625.d Copy of E3Kp Default.ms WPF-205-BP Analysis Name Method Sample Name Commen Alternating I Scan End Trap Drive Auto MS/MS Ion Polarity Scan Begir Skim 1 off 1500 m/z 154.8 off 50 m/z 40.0 Volt Std/Norma 151.0 Volt 1266 µs +MS, 0.0-0.1min (#1-#8) Intens. x10⁵ 1039.3 $[3_2 \supset 4 - 2PF_6]^{2+}$ 1017.6 102 1122.5 1140 m/z 1100
- 6. LRESI mass spectrum of $3_2 \rightarrow 4$

Figure S9. LRESI mass spectrum of 3₂⊃4.

7. Diffusion coefficient D of 1 + 4, $1 + 4 + KPF_6$, and $1 + 4 + KPF_6 + DB18C6$



Figure S10. Diffusion coefficient *D* (400 MHz, CD_2Cl_2/CD_3CN (1:1, *v*/*v*), 293 K) of 1 + 4, 1 + 4 + KPF₆, and 1 + 4 + KPF₆ + DB18C6 at 5.00 mM.

Reference:

S1. P. J. Stang, B. Olenyuk, J. Fan and A. M. Arif, Organometallics, 1996, 15, 904–908.