## Supporting Information

Thiol-reactive Passerini-methacrylates and Polymorphic Surface Functional Soft Matter Nanoparticles via Ethanolic RAFT Dispersion Polymerization and Postsynthesis Modification

Yiwen Pei, Janina-Miriam Noy, Peter J. Roth, and Andrew B. Lowe\*

School of Chemical Engineering, Centre for Advanced Macromolecular Design, UNSW Australia, University of New South Wales, Kensington, Sydney, NSW 2052, Australia.

profandrewblowe@gmail.com

## **Experimental Section**

**Materials.** All reagents were purchased from the Sigma-Aldrich Chemical Company and used as received unless otherwise noted. 2,2'-Azobis(isobutyronitrile) (AIBN) was purified by two recrystallizations from methanol. 2-(Dimethylamino)ethyl methacrylate (DMAEMA), 3-phenylpropyl methacrylate (PPMA) was passed through a basic Al<sub>2</sub>O<sub>3</sub> column to remove inhibitors prior to use. Novel pentafluorophenyl (PFP)-containing methacrylic monomers, 2-(cyclohexylamino)-2-oxo-1-(perfluorophenyl)ethyl methacrylate (CyAFPEMA), 2-((2-ethoxy-2-oxoethyl)amino)-2-oxo-1-(perfluorophenyl) ethyl methacrylate (EAFPEMA) and 2-(*tert*-butylamino)-2-oxo-1-(perfluorophenyl)ethyl methacrylate (*t*BAFPEMA), were prepared via the Passerini reaction as described recently by Roth *et al.*<sup>1,2</sup> and 4-Cyanopentanoic acid dithiobenzoate (CPADB) was prepared according to a procedure described elsewhere.<sup>3</sup>

## **Copolymer Characterizations**

Size exclusion chromatography (SEC) was performed on a Shimadzu system with four phenogel columns ( $10^2$ ,  $10^3$ ,  $10^4$ ,  $10^6$  Å pore size) in *N*, *N*-dimethylacetamide (DMAc) operating at a flow rate of 1 mL min<sup>-1</sup> at 40 °C using a RID-10A refractive index detector. Chromatograms were analyzed by Cirrus SEC software version 3.0. The system was calibrated with a series of narrow molecular weight distribution polystyrene standards with molecular weights ranging from 0.58-1820 kg mol<sup>-1</sup>.

Transmission electron microscopy (TEM) imaging was conducted at 100 kV on a JEOL1400 TEM instrument. To prepare TEM samples, 5.0  $\mu$ L of a dilute copolymer solution (0.7 w/w %) was deposited onto a copper grid (ProSciTech), stained with uranyl acetate (0.2 w/w % in water), and dried under ambient conditions.

DLS measurements were performed using a Malvern Instrument Zetasizer Nano Series instrument equipped with a 4 mW He-Ne laser operating at 633 nm and an avalanche photodiode (APD) detector. The scattered light was detected at an angle of 173°. For sample preparation, 0.1 mL of the parent RAFTDP solution was diluted with 2.9 mL of ethanol and the solution then stirred for 5 min prior to double filtration through 0.45 µm nylon filters.

Fourier transform infrared (FTIR) spectroscopy was conducted on a Bruker IFS 66/S

instrument under attenuated total reflectance (ATR) and the results were analyzed utilizing OPUS software version 4.0.

NMR characterization analyses were conducted using a Bruker Avance III 300 spectrometer (300.13 MHz for <sup>1</sup>H nuclei and 282 MHz for <sup>19</sup>F nuclei). The internal solvent signal of CDCl<sub>3</sub> was utilized for reference ( $\delta = 7.26$  ppm). The number average molecular weight,  $\overline{M}_{n,NMR}$ , was calculated based on a full conversion of Passerini methacrylate (confirmed by <sup>19</sup>F NMR) and the average degree of polymerization,  $X_n$  of PDMAEMA in RAFT macroCTAs.  $X_n$  of PDMAEMA was estimated based on the integral values of the signals at  $\delta = 4.06$  ppm and 7.30-7.90 ppm, as shown in eqn (1).

$$\bar{X}n (PDMAEMA) = \frac{5 \times I (4.60 \, ppm)}{2 \, \times \, I \, (7.30 - 7.90 \, ppm)} \tag{1}$$

The absolute molecular weight,  $M_{n,NMR}$ , and  $X_n$  of block copolymers were calculated based on  $\bar{X}_n$  of the PDMAEMA-Passerini macro-CTA and the integration ratio of the signal at  $\delta = 7.10$ -7.50 ppm ( $I_{7.10-7.50ppm}$ , aryl protons of PPPMA) and those at 2.30 ppm ( $I_{2.30ppm}$ , methyl protons of PDMAEMA in RAFT macro-CTA), as shown in Figure 2-4 and eqn (2). We also compared the integration ratio between the methylene protons of the PDMAEMA block ( $\delta = 4.20$  ppm) and the methylene protons of the PPPMA block ( $\delta =$ 3.90 ppm) to obtain accurate polymer compositions.

$$Xn (PPPMA) = \frac{6 \times I (7.70 - 7.50 \, ppm)}{5 \times I (2.30 \, ppm)} Xn (PDMAEMA)$$
(2)



**Figure S1.** <sup>1</sup>H NMR spectra of DMAEMA-Passerini macro-CTAs (A)  $P(DMAEMA_{36}-co-CyAFPEMA_2)$ , (B)  $P(DMAEMA_{29}-co-EAFPEMA_2)$  and (C)  $P(DMAEMA_{31}-co-tBAFPEMA_2)$ , recorded in CDCl<sub>3</sub>, with peak assignments. Inset is an expanded region highlighting the presence of the phenyldithioester end group, benzylic and NH hydrogen.



**Figure S2.** (A) <sup>1</sup>H NMR spectrum of PFP-functional copolymer, P(DMAEMA<sub>36</sub>-*co*-CyAFPEMA<sub>2</sub>)-*b*-PPPMA<sub>148</sub> and (B) representative size exclusion chromatograms of the resulting polymer obtained by RAFTDP.



**Figure S3.** (A) <sup>1</sup>H NMR spectrum of PFP-functional copolymer, P(DMAEMA<sub>29</sub>-co-EAFPEMA<sub>2</sub>)-b-PPPMA<sub>44</sub> and (B) representative size exclusion chromatograms of the resulting polymers obtained by RAFTDP.



**Figure S4.** (A) <sup>1</sup>H NMR spectrum of PFP-functional copolymer, P(DMAEMA<sub>31</sub>-*co-t*BAFPEMA<sub>2</sub>)-*b*-PPPMA<sub>95</sub> and (B) representative size exclusion chromatograms of the resulting polymers obtained by RAFTDP.

Table S1. Summary of RAFTDP syntheses with macro-CTAs and PPMA as a comonomer in EtOH at 21 wt% and 70 °C. The NMR-determined average degree of polymerization  $(\bar{X}_n)$  of the PPMA block and absolute molecular weight are given along with the SEC-measured  $\bar{M}_n$  and dispersities, TEM morphology, TEM-measured diameter and DLS-measured hydrodynamic diameter and polydispersities.

	Conv.	PPMA	NMR	SEC	SEC	TEM	TEM	DLS	DLS
Macro-CIA	%	X <sub>n</sub>	MW <sup>a</sup>	${ar M}_{ m n}^{ m b}$	$\boldsymbol{\mathcal{P}}_{\mathrm{M}}^{\mathrm{b}}$	morp. °	D <sup>d</sup>	<b>D</b> <sub>h</sub> <sup>e</sup>	PDI
P(DMAEMA <sub>36</sub> -co-CyAFPEMA <sub>2</sub> )	95	148	36,900	23,400	1.19	S+W	54.5	141.8	0.21
P(DMAEMA <sub>29</sub> -co-EAFPEMA <sub>2</sub> )	96	44	14,300	17,300	1.19	S	41.2	53.4	0.17
P(DMAEMA <sub>29</sub> -co-EAFPEMA <sub>2</sub> )	95	79	21,400	21,200	1.19	S+W	45.6	81.4	0.17
P(DMAEMA <sub>29</sub> -co-EAFPEMA <sub>2</sub> )	92	91	23,900	22,900	1.24	W+V	234.3	215.0	0.21
P(DMAEMA <sub>31</sub> - <i>co</i> - <i>t</i> BAFPEMA <sub>2</sub> )	91	48	15,700	16,200	1.18	S	47.0	45.8	0.18
P(DMAEMA <sub>31</sub> - <i>co</i> - <i>t</i> BAFPEMA <sub>2</sub> )	95	80	22,300	19,600	1.23	S+W	53.2	137.1	0.19
P(DMAEMA <sub>31</sub> - <i>co</i> - <i>t</i> BAFPEMA <sub>2</sub> )	95	87	23,700	19,900	1.21	S+W	54.9	190.1	0.19
P(DMAEMA <sub>31</sub> -co-tBAFPEMA <sub>2</sub> )	92	95	25,200	21,300	1.23	W	55.1	223.4	0.19
P(DMAEMA <sub>31</sub> - <i>co</i> - <i>t</i> BAFPEMA <sub>2</sub> )	87	131	32,700	25,800	1.21	W+V	275.3	230.7	0.79

<sup>a</sup> As determined by end group analysis; <sup>b</sup> as measured in THF on a system calibrated with polystyrene standards; <sup>c</sup> S = spheres, W = worms, V = vesicles; <sup>d</sup> TEM-measured nanoparticle diameter in nm; <sup>e</sup> hydrodynamic diameter in nm.



**Figure S5.** Representative TEM images obtained for (A)  $P(DMAEMA_{36}$ -*co*-CyAFPEMA<sub>2</sub>)-*b*-PPPMA<sub>148</sub> and (B)  $P(DMAEMA_{29}$ -*co*-EAFPEMA<sub>2</sub>)-*b*-PPPMA<sub>y</sub> copolymer nanoparticles synthesized at a total solids concentration of 21 wt% using RAFT dispersion polymerization in ethanol at 70 °C.

**Table S2.** Summary of thiol-*p*-fluoro postpolymerization modification performed in ethanol at 50 °C overnight (up to 2 days for the reaction with worm-like copolymers<sup>c</sup>). Complete substitution was confirmed for all reactions by <sup>19</sup>F NMR spectroscopy.

Entry	<b>PFP-Functional Nano-Objects</b>	Thiol	Base	Thiol/PFP	Thiol/Base	
1a						
1"	p[(DMAEMA <sub>29</sub> -co-EAFPEMA <sub>2</sub> )-PPMA <sub>44</sub> ]	2-mercaptoethanol	DBU	5	1	
2 <sup>a</sup>	p[(DMAEMA <sub>29</sub> -co-EAFPEMA <sub>2</sub> )-PPMA <sub>44</sub> ]	cysteamine hydrochloride	DBU	20	0.5	
20						
3ª	p[(DMAEMA <sub>31</sub> - <i>co-t</i> BAFPEMA <sub>2</sub> )-PPMA <sub>48</sub> ]	1-thio-β-D-glucose tetraacetate	Et <sub>3</sub> N	20	1	
4 <sup>b</sup>	p[(DMAEMA <sub>31</sub> - <i>co-t</i> BAFPEMA <sub>2</sub> )-PPMA <sub>80</sub> ]	Captopril	DBU	5	0.5	
5 <sup>b</sup>	p[(DMAEMA <sub>31</sub> - <i>co-t</i> BAFPEMA <sub>2</sub> )-PPMA <sub>80</sub> ]	thiophenol	Et <sub>3</sub> N	10	1	
6 <sup>c</sup>	p[(DMAEMA <sub>31</sub> - <i>co-t</i> BAFPEMA <sub>2</sub> )-PPMA <sub>95</sub> ]	1-thio-β-D-glucose tetraacetate	Et <sub>3</sub> N	50	1	
			5			

<sup>a</sup> Pure sphere phase; <sup>b</sup> Mixed spheres/worms phase; <sup>c</sup> Pure worm phase.



**Figure S6.** (a) <sup>19</sup>F NMR spectra of the P[(DMAEMA<sub>29</sub>-*co*-EAFPEMA<sub>2</sub>)-*b*-PPMA<sub>44</sub>] exhibiting *ortho*, *meta* and *para* signals associated with the PFP functionality in the Passerini repeat units; (b) after reaction of the spherical nanoparticles with 2-mercaptoethanol, and (c) after purification of the surface modified nano-spheres, with SEC traces before and after reaction shown inset.



**Figure S7.** (a) <sup>19</sup>F NMR spectra of the P[(DMAEMA<sub>29</sub>-*co*-EAFPEMA<sub>2</sub>)-*b*-PPMA<sub>44</sub>] exhibiting *ortho*, *meta* and *para* signals associated with the PFP functionality in the Passerini repeat units; (b) after reaction of the spherical nanoparticles with cysteamine hydrochloride, and (c) after purification of the surface modified nano-spheres, with SEC traces before and after reaction shown inset.



**Figure S8.** <sup>19</sup>F NMR spectra of the (A) unmodified and (B) surface-modified mixed phased sphere/worm nano-objects by reacting  $P[(DMAEMA_{31}-co-tBAFPEMA_2)-b-PPMA_{80}]$  with thiophenol, recorded in CDCl<sub>3</sub>, with peak assignments, with SEC trace before and after reaction shown inset.



**Figure S9.** <sup>19</sup>F NMR spectra of the (A) unmodified and (B) surface-modified mixed phased nano-objects by reacting  $P[(DMAEMA_{31}-co-tBAFPEMA_2)-b-PPMA_{80}]$  with Captopril, recorded in CDCl<sub>3</sub>, with peak assignments, with SEC trace before and after reaction shown inset.



**Figure S10.** (a) <sup>19</sup>F NMR spectra of the P[(DMAEMA<sub>31</sub>-*co-t*BAFPEMA<sub>2</sub>)-*b*-PPMA<sub>95</sub>] exhibiting *ortho*, *meta* and *para* signals associated with the PFP functionality in the Passerini repeat units; (b) after reaction of the spherical nanoparticles with 1-thio- $\beta$ -D-glucose tetraacetate, and (c) after purification of the surface-functional worm-like copolymers, with SEC traces before and after reaction shown inset.



**Figure S11.** FT-IR spectra of unmodified block copolymer,  $P[(DMAEMA_{31}-co-tBAFPEMA_2)-b-PPMA_{95}]$  (**a**, green) and surface modified nano-objects by reaction with 2-mercaptoethanol (**b**, pink), thiophenol (**c**, brown), Captopril (**d**, orange), 1-thio- $\beta$ -D-glucose tetraacetate (**e**, purple), cysteamine hydrochloride (**f**, blue). The occurrence of amidation in the cysteamine-modified copolymer nanoparticles is evident from the disappearance of distinctive ester band (C=O stretch at ~1730 cm<sup>-1</sup>) and clear appearance of the amide bands at ~3290 (N-H stretch), ~1640 (amide C=O stretching) and ~1530 cm<sup>-1</sup> (N-H bend).



**Figure S12.** <sup>1</sup>H NMR spectra of (A) the unmodified and (B) surface-modified spherical nano-objects by reacting P[(DMAEMA<sub>29</sub>-*co*-EAFPEMA<sub>2</sub>)-*b*-PPMA<sub>44</sub>] with 2-mercaptoethanol, recorded in CDCl<sub>3</sub>, with peak assignments.



**Figure S13.** <sup>1</sup>H NMR spectra of (A) the unmodified and (B) surface-modified mixed phased nano-objects by reacting  $P[(DMAEMA_{31}-co-tBAFPEMA_2)-b-PPMA_{80}]$  with Captopril, recorded in CDCl<sub>3</sub>, with peak assignments.



**Figure S14.** <sup>1</sup>H NMR spectra of (A) unmodified and (B) the surface-modified mixed phased nano-objects by reacting  $P[(DMAEMA_{31}-co-tBAFPEMA_2)-b-PPMA_{80}]$  with thiophenol, recorded in CDCl<sub>3</sub>, with peak assignments.



**Figure S15.** <sup>1</sup>H NMR spectra of (A) unmodified and (B) surface-modified spherical nano-objects by reacting P[(DMAEMA<sub>31</sub>-*co-t*BAFPEMA<sub>2</sub>)-*b*-PPMA<sub>48</sub>] with 1-thio- $\beta$ -D-glucose tetraacetate, recorded in CDCl<sub>3</sub>, with peak assignments.



**Figure S16.** <sup>1</sup>H NMR spectra of (A) unmodified and (B) the surface-modified worm-like nano-objects by reacting P[(DMAEMA<sub>31</sub>-*co-t*BAFPEMA<sub>2</sub>)-*b*-PPMA<sub>95</sub>] with 1-thio- $\beta$ -D-glucose tetraacetate, recorded in CDCl<sub>3</sub>, with peak assignments.

## **Reference:**

(1) S. Schmidt, M. Koldevitz, J.-M. Noy, P. J. Roth. *Polym. Chem.*, **2014**, DOI: 10.1039/c4py01147c

(2) J.-M. Noy, M. Koldevitz, P. J. Roth. 2014, DOI: 10.1039/c4py01238k

(3) S. H. Thang, Y. K. Chong, R. T. A. Mayadunne, G. Moad and E. Rizzardo, *Tetrahedron Lett.*, **1999**, 40, 2435–2438