

## Supplementary Data

# Polymer-Based Biocompatible Fluorescent Sensor for Nano-molar Detection of Zn<sup>2+</sup> in Aqueous Medium and Biological Samples

Kamalpreet Kaur<sup>a</sup>, Manjot Kaur<sup>b</sup>, Amanpreet Kaur<sup>c</sup>, Jasminde Singh<sup>b</sup>, Narinder Singh<sup>a</sup>, Susheel K. Mittal<sup>b,\*</sup>, Navneet Kaur<sup>c,\*</sup>

<sup>a</sup>Department of Chemistry, Indian Institute of Technology Ropar, Rupnagar, Punjab 140001, India

<sup>b</sup>School of Chemistry & Biochemistry, Thapar University, Patiala 147004, India

<sup>c</sup>Centre for Nanoscience & Nanotechnology, Panjab University, Chandigarh, Punjab 160014, India

## List of Contents

**Figure S1.** <sup>1</sup>H NMR spectrum of receptor **2a** in DMSO-*d*<sub>6</sub>.

**Figure S2.** <sup>13</sup>C NMR spectrum of receptor **2a** in DMSO-*d*<sub>6</sub>.

**Figure S3.** IR spectrum of receptor **2a**.

**Figure S4.** <sup>1</sup>H NMR spectrum of receptor **2b** in DMSO-*d*<sub>6</sub>.

**Figure S5.** <sup>13</sup>C NMR spectrum of receptor **2b** in DMSO-*d*<sub>6</sub>.

**Figure S6.** IR spectrum of receptor **2b**.

**Figure S7.** <sup>1</sup>H NMR spectrum of receptor **2c** in DMSO-*d*<sub>6</sub>.

**Figure S8.** <sup>13</sup>C NMR spectrum of receptor **2c** in DMSO-*d*<sub>6</sub>.

**Figure S9.** IR spectrum of receptor **2c**.

**Figure S10.** UV-Vis absorption spectra of (a) receptor **2a** (10 μM) upon addition of 50 μM of Zn<sup>2+</sup> nitrate salt in HEPES buffered (10 mM, pH=7.0) DMF/H<sub>2</sub>O (7:3, v/v) solvent system; (b) receptor **2b** (10 μM) upon addition of 50 μM of Zn<sup>2+</sup> nitrate salt in HEPES buffered (10 mM, pH=7.0) DMF/H<sub>2</sub>O (7:3, v/v) solvent system and (c) receptor **2c** (10 μM) upon addition of 50 μM of Zn<sup>2+</sup> nitrate salt in HEPES buffered (10 mM, pH=7.0) DMF/H<sub>2</sub>O (7:3, v/v) solvent system.

**Figure S11.** Fluorescence spectrum of (a) receptor **2a** (10 μM) recorded in HEPES buffered (10 mM, pH=7.0) DMF/H<sub>2</sub>O (7:3, v/v) solvent system excited at 310 nm; (b) receptor **2b** (10 μM) recorded in HEPES buffered (10 mM, pH=7.0) DMF/H<sub>2</sub>O (7:3, v/v) solvent system excited at 309 nm and (c) receptor **2c** (10 μM) recorded in HEPES buffered (10 mM, pH=7.0) DMF/H<sub>2</sub>O (7:3, v/v) solvent system excited at 370 nm.

**Figure 12.** Changes in the Fluorescence spectra of **2a** upon pH titration of a solution of **2a** (10 μM) in DMF/H<sub>2</sub>O (7:3, v/v) solvent system under: (A) acidic and (B) basic conditions.

**Figure 13.** Changes in the Fluorescence spectra of **2b** upon pH titration of a solution of **2b** (15 μM) in DMF/H<sub>2</sub>O (7:3, v/v) solvent system under: (A) acidic and (B) basic conditions.

**Figure 14.** Changes in the Fluorescence spectra of **2c** upon pH titration of a solution of **2c** (8 μM) in DMF/H<sub>2</sub>O (7:3, v/v) solvent system under: (A) acidic and (B) basic conditions.

**Figure S15.** Fluorescence emission spectra of **2c**. Zn<sup>2+</sup> complex (20 μM) upon successive addition of EDTA (0-40 μM) in HEPES buffered (10 mM, pH=7.0) DMF/H<sub>2</sub>O (7:3, v/v) solvent system.

**Figure S16.** Showing the course of Fluorescence intensity at 418 nm with time (0-20 min) for 20 μM of **2c**. Zn<sup>2+</sup> complex upon addition of 40 μM of EDTA in HEPES buffered (10 mM, pH=7.0) DMF/H<sub>2</sub>O (7:3, v/v) solvent system.

**Figure S17:** Dose-response curve for **2c** for the calculation of IC<sub>50</sub> value

**Figure S18.** Calibration curve for Zn<sup>2+</sup> ion using electrode with ionophore **2a-c**.

**Figure S19.** Working pH range of electrodes with ionophore **2a-c**.

**Figure S20.** Potentiometric titration curves for electrodes with ionophore **2a-c**.

**Table S1.** Total energies of optimized keto and enol forms of receptors **2a**, **2b** and **2c**.

**Table S2.** Slopes of calibration curves for Zn (II) ions using electrodes with ionophores **2a-c**.

**Table S3.** Selectivity coefficients of Zn<sup>2+</sup> ion selective electrodes for different interfering ions ( $1\times10^{-3}$ M) by fixed interference method.

**Table S4.** A comparison of quantum yield of **2c**, **2c** after complexation with Zn<sup>2+</sup>.

**Table S5.** A comparison of results obtained for spiked Zn<sup>2+</sup> in water samples using proposed probe and AAS.

**Scheme S1.** Synthesis of monomer **3**.

**Figure S21.** Mass spectrum of **3**.

**Figure S22.** DFT calculated partial structures of receptor **2a-c**: (A) Enol and Keto forms of receptor **2a**; (B) Enol and Keto forms of receptor **2b**; (C) Enol and Keto forms of receptor **2c**; DFT calculations were performed at B3LYP/6-31G\* level; red, blue and grey spheres refer to O, N and C atoms respectively).

**Figure S23.** (A) Changes in fluorescence intensity of receptor **2a** (10 μM) upon addition of 50 μM of a particular metal nitrate salts in HEPES buffered (10 mM, pH=7.0) DMF/H<sub>2</sub>O (7:3, v/v); with  $\lambda_{\text{ex}} = 310$  nm; (B) Influence of some metal ions on the Zn<sup>2+</sup> based triggering of fluorescence intensity of **2a**.

**Figure S24.** (A) Changes in fluorescence intensity of receptor **2c** (10 μM) upon addition of 50 μM of a particular metal nitrate salts in HEPES buffered (10 mM, pH=7.0) DMF/H<sub>2</sub>O (7:3, v/v); with  $\lambda_{\text{ex}} = 370$  nm; (B) Selectivity of **2c** for Zn<sup>2+</sup> compared to other metal ions: the presence of other metal ions has no effect for Zn<sup>2+</sup> induced fluorescence intensity of **2c**.

**Figure S25.** Changes in fluorescence intensity of receptor **3** (10 μM) upon addition of 50 μM of a particular metal nitrate salts in HEPES buffered (10 mM, pH=7.0) DMF/H<sub>2</sub>O (7:3, v/v); with  $\lambda_{\text{ex}} = 365$  nm.

**Figure S26.** Changes in the Fluorescence spectra of **2c**.Zn<sup>2+</sup> upon pH titration of a solution of **2c**.Zn<sup>2+</sup> in DMF/H<sub>2</sub>O (7:3, v/v) solvent system under: (A) acidic and (B) basic conditions.

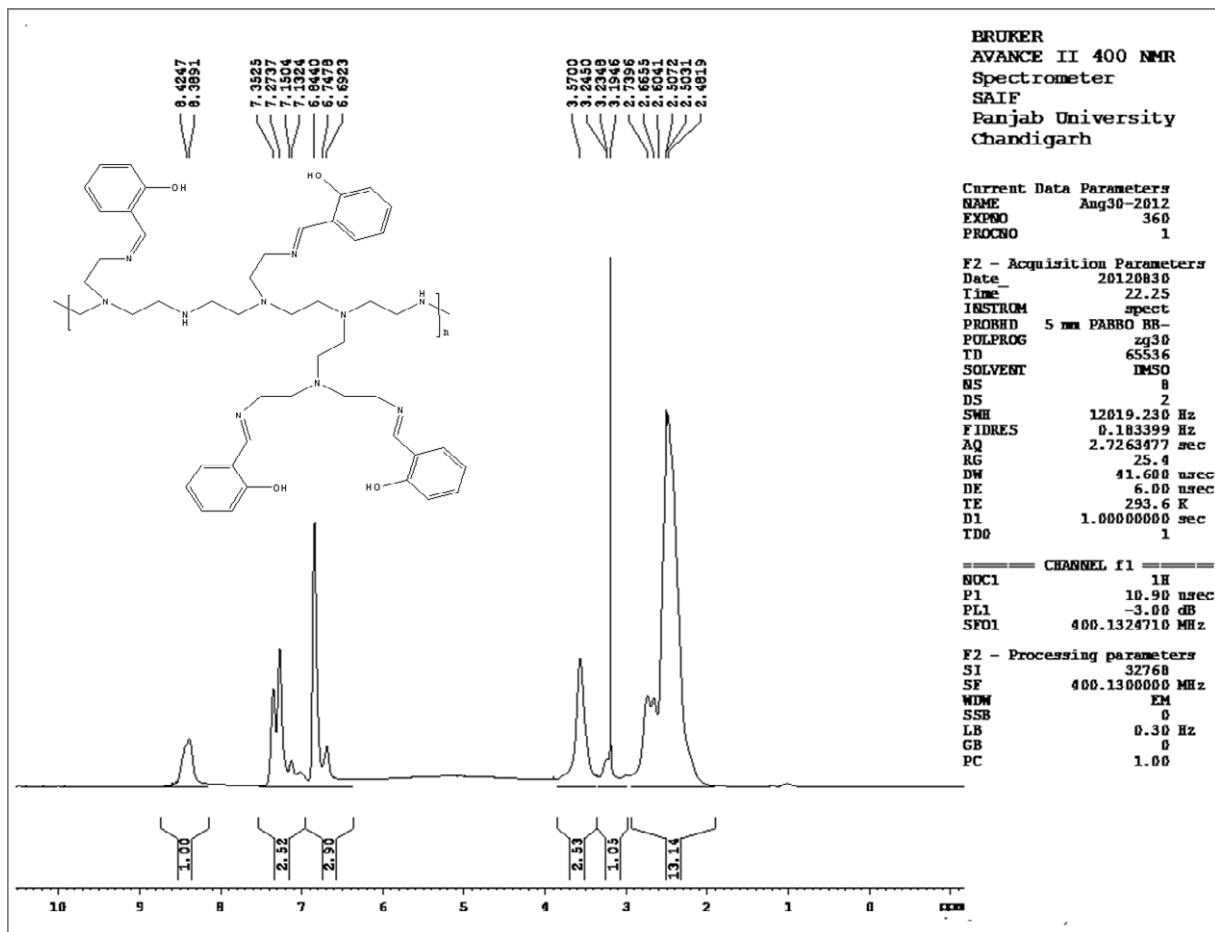
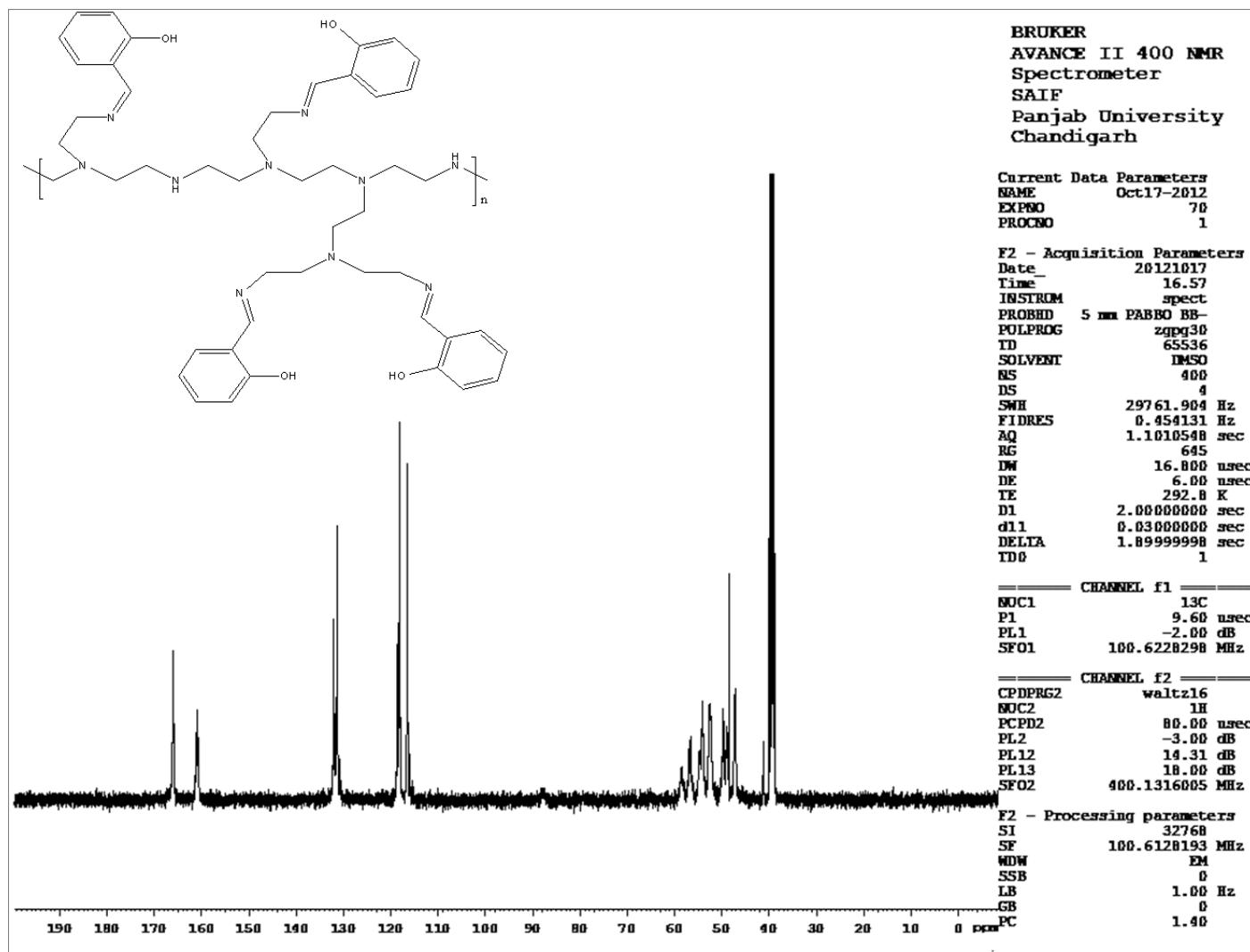
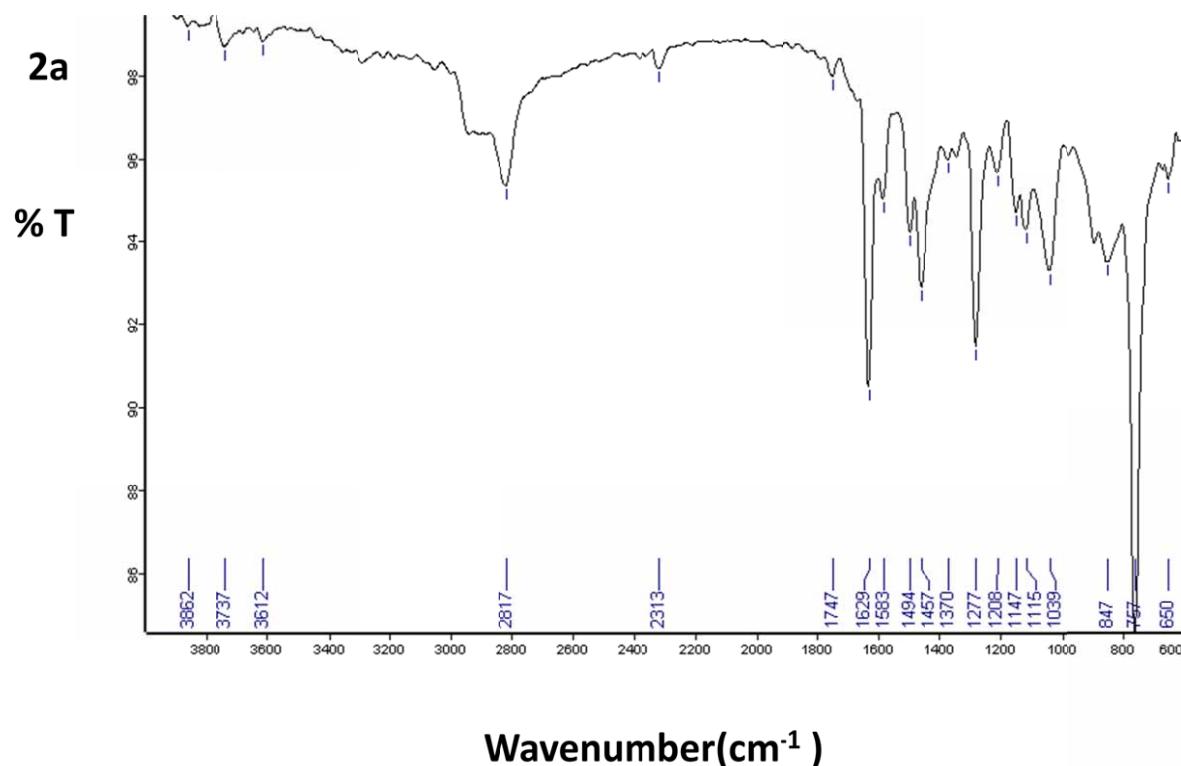


Figure S1.  $^1\text{H}$  NMR spectrum of receptor **2a** in  $\text{DMSO}-d_6$ .



**Figure S2.**  $^{13}\text{C}$  NMR spectrum of receptor **2a** in  $\text{DMSO}-d_6$ .



**Figure S3.** IR spectrum of receptor **2a**.

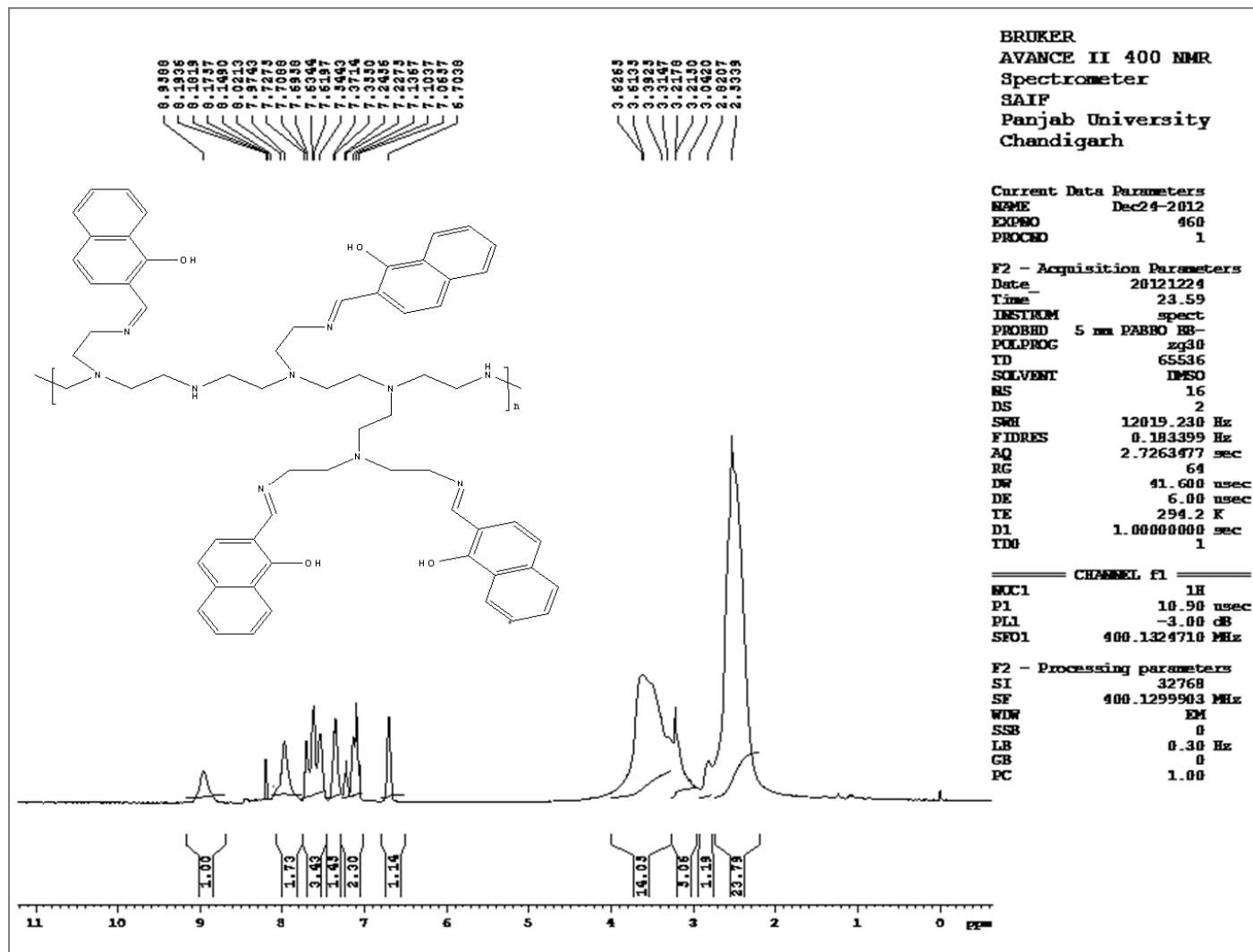
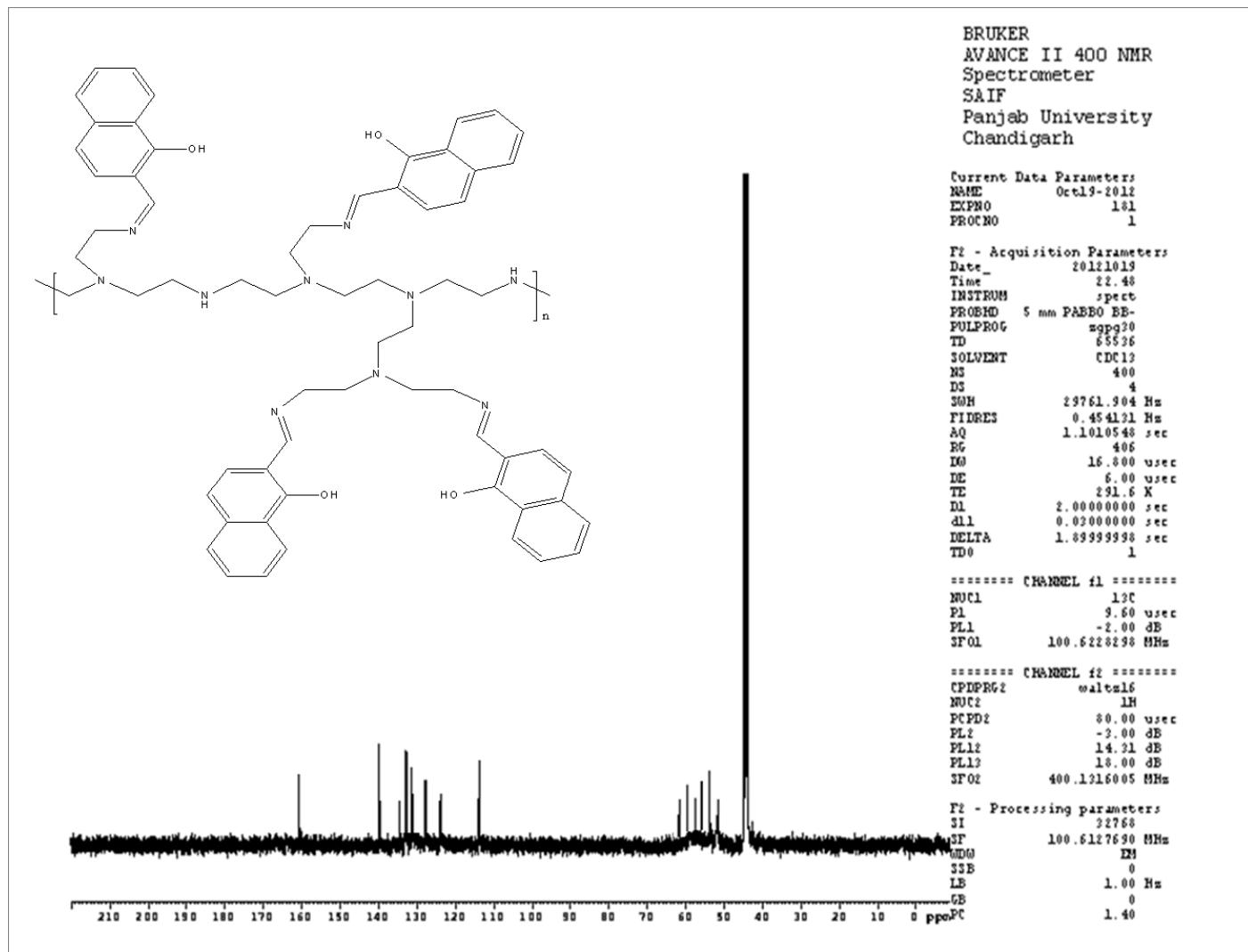
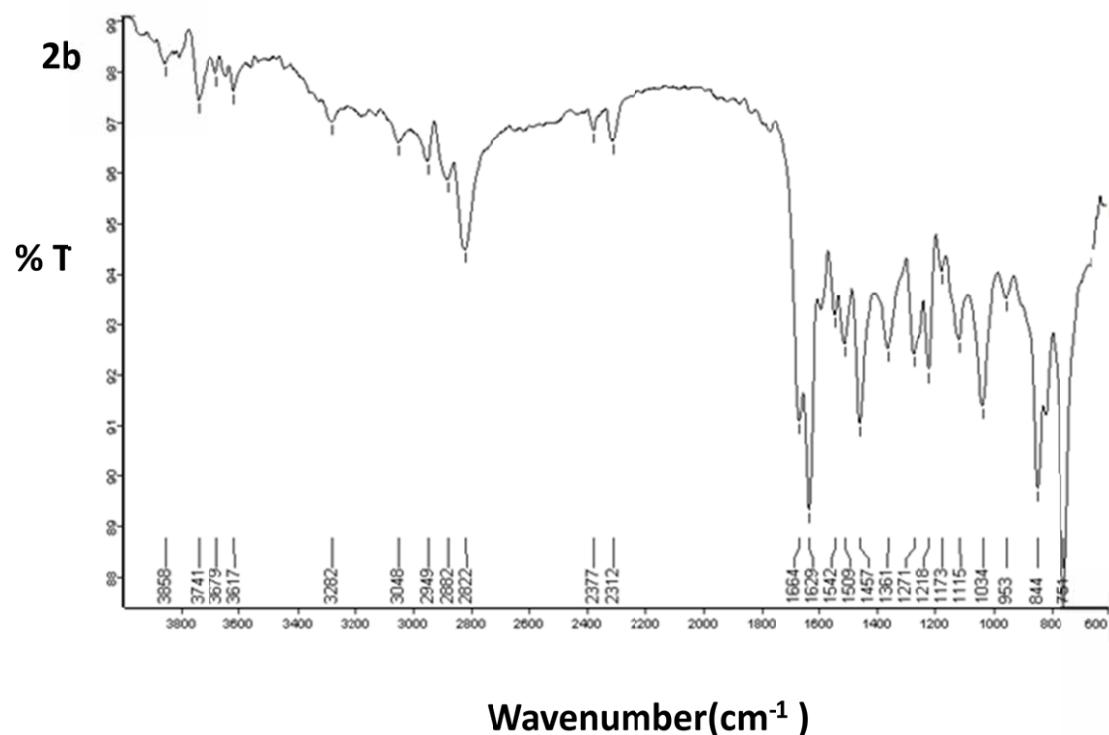


Figure S4. <sup>1</sup>H NMR spectrum of receptor 2b in DMSO-*d*<sub>6</sub>.



**Figure S5.** <sup>13</sup>C NMR spectrum of receptor **2b** in DMSO-*d*<sub>6</sub>.



**Figure S6.** IR spectrum of receptor **2b**.

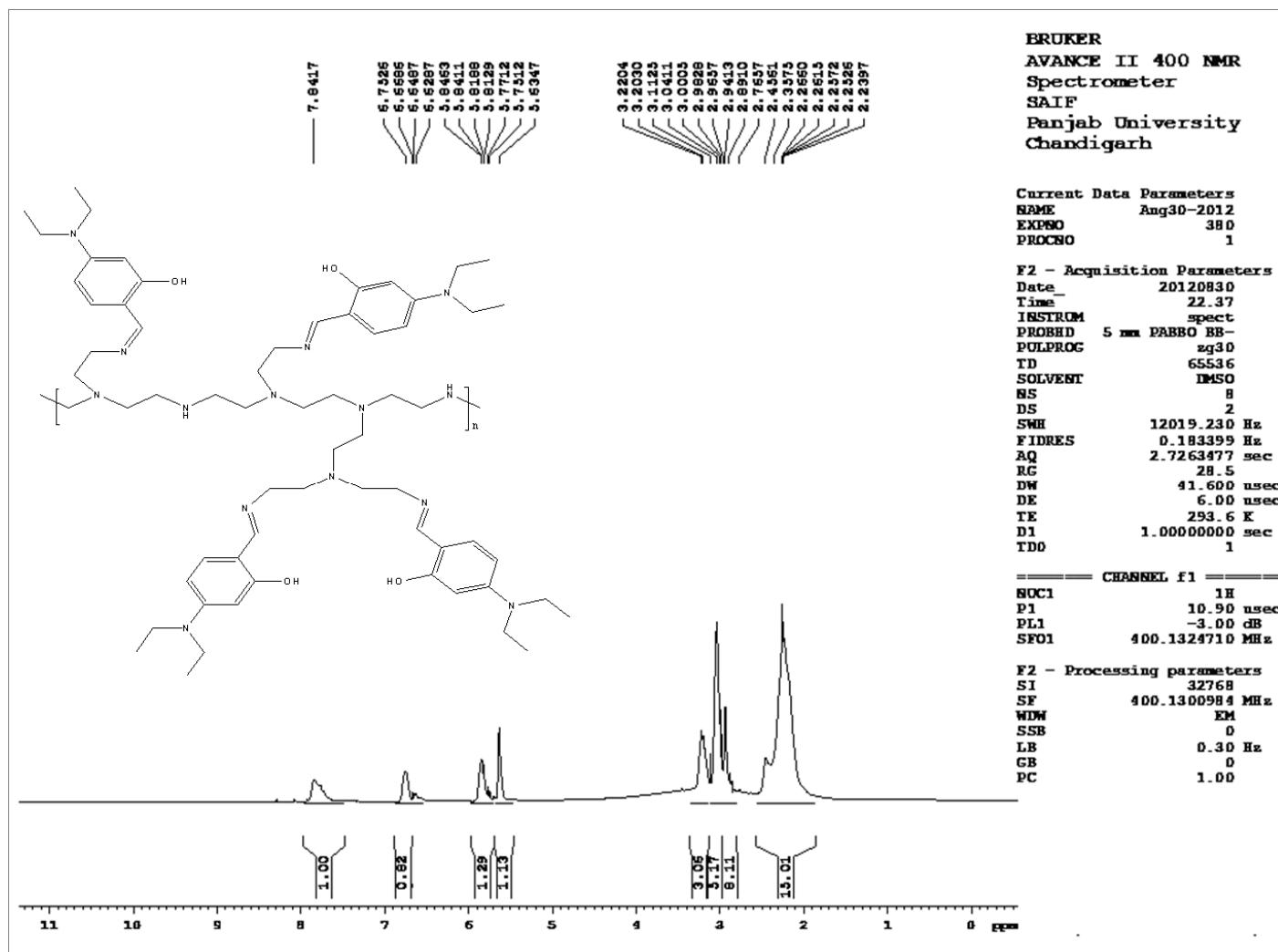
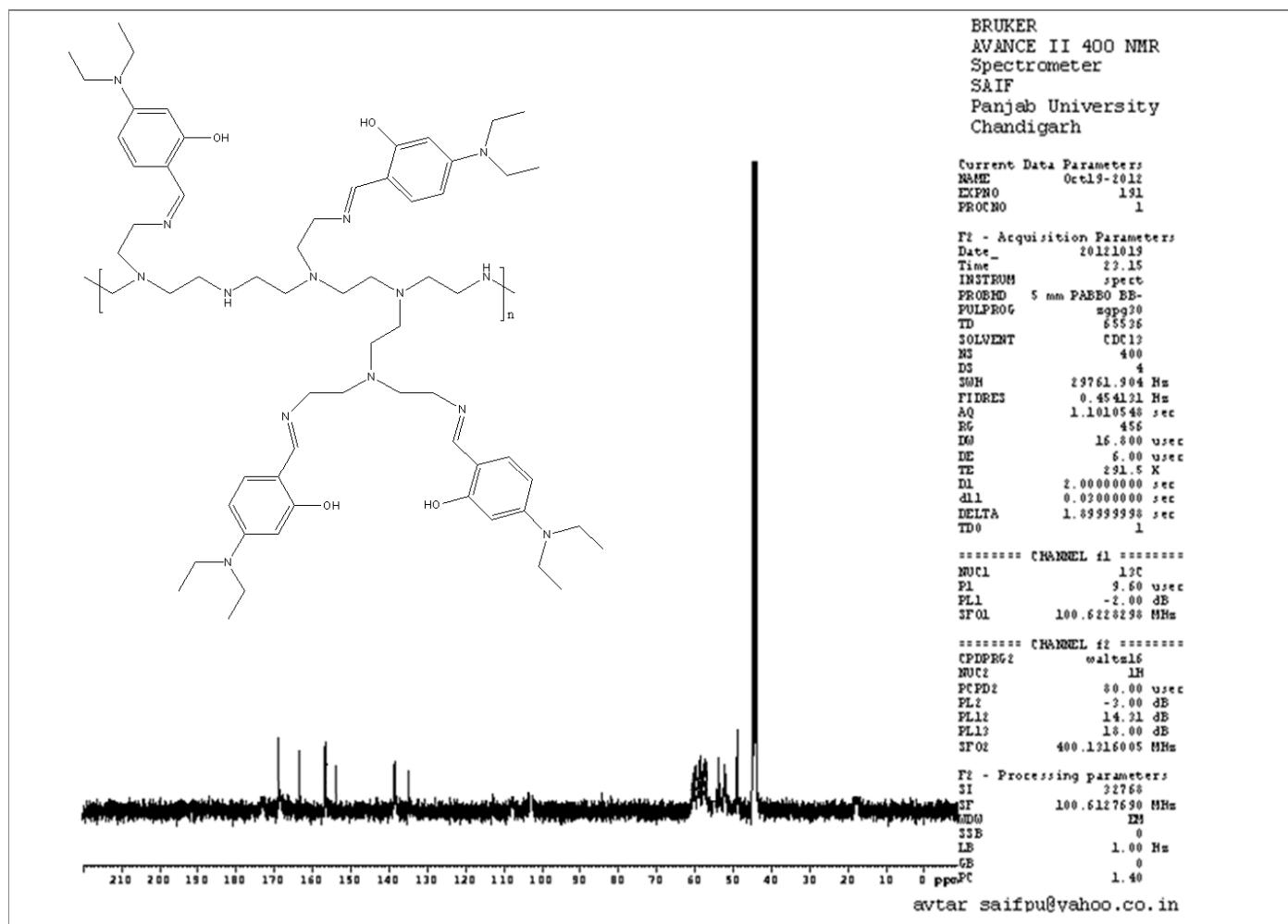
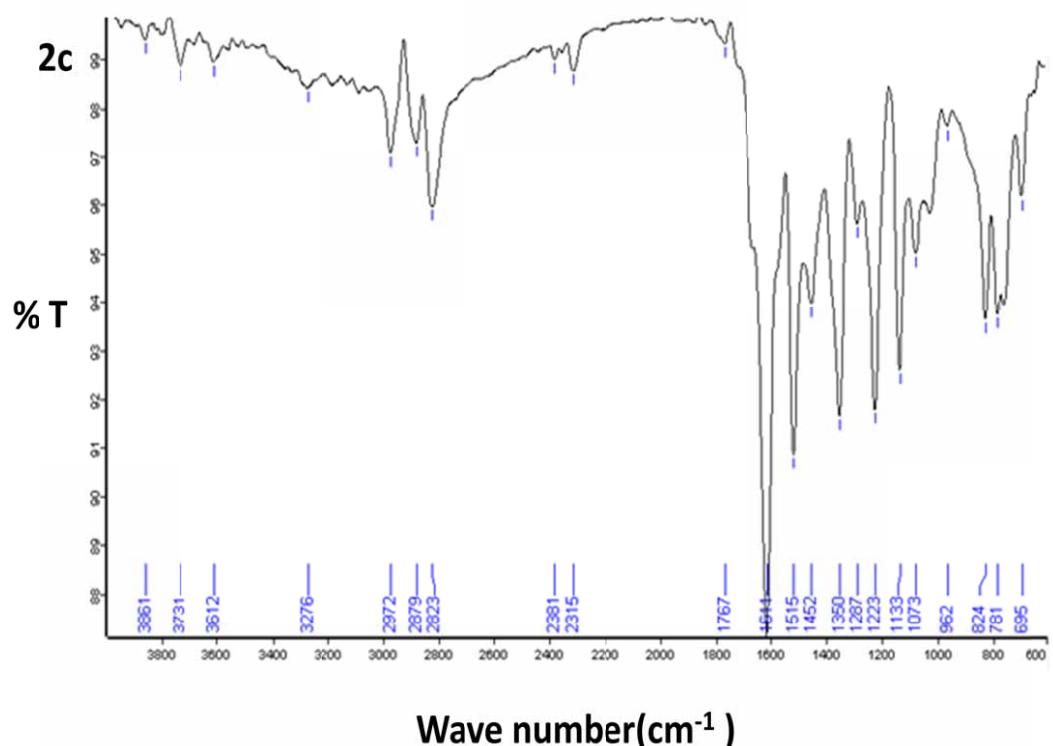


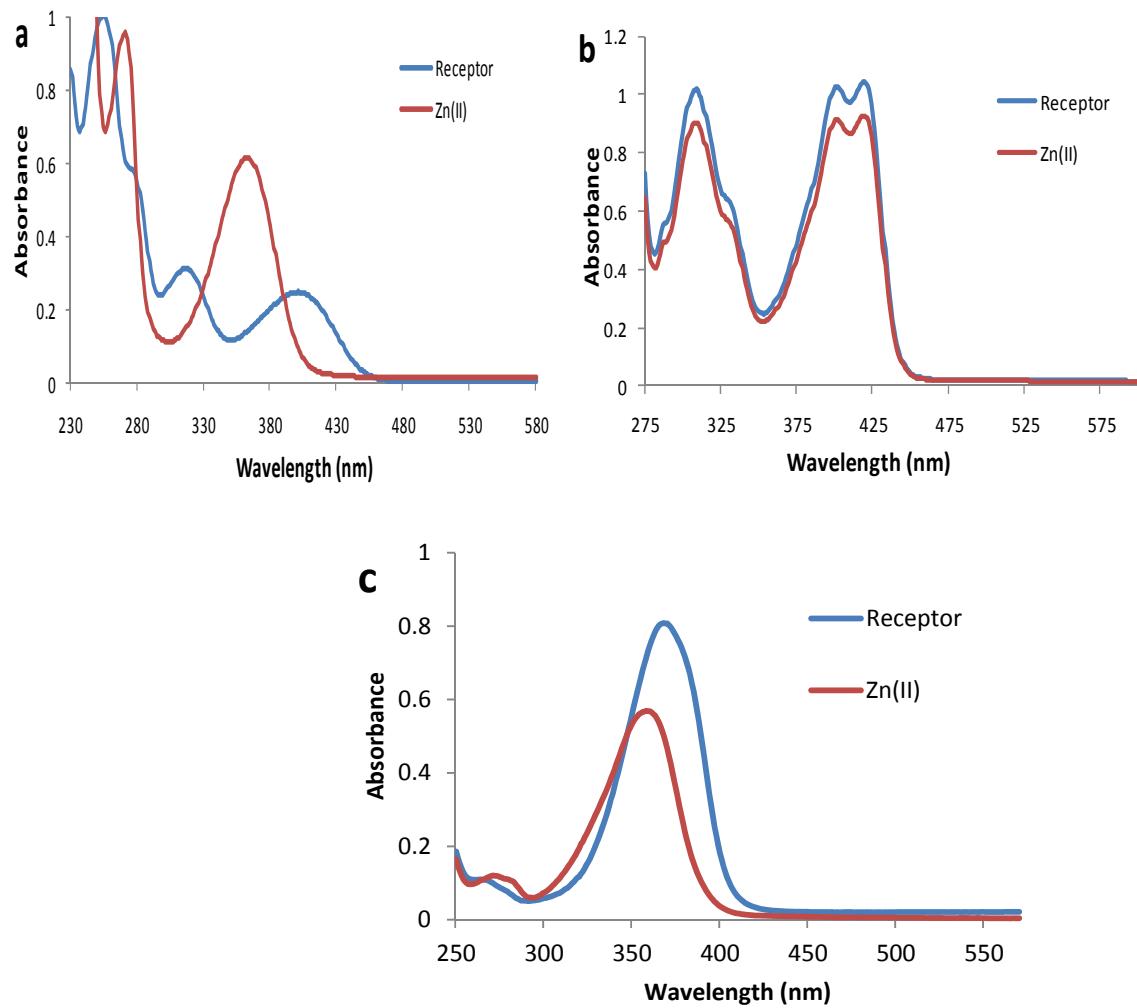
Figure S7.  $^1\text{H}$  NMR spectrum of receptor **2c** in  $\text{DMSO}-d_6$ .



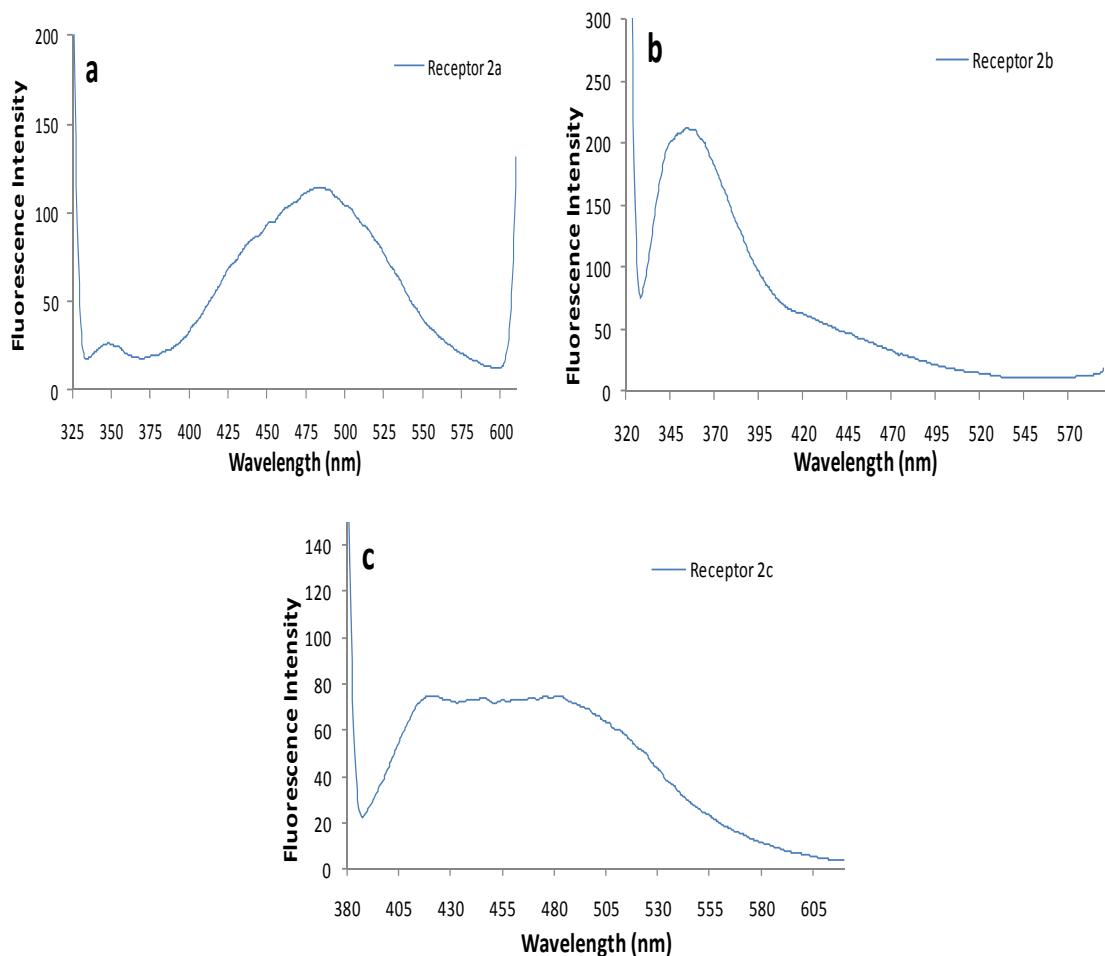
**Figure S8.**  $^{13}\text{C}$  NMR spectrum of receptor **2c** in  $\text{DMSO}-d_6$ .



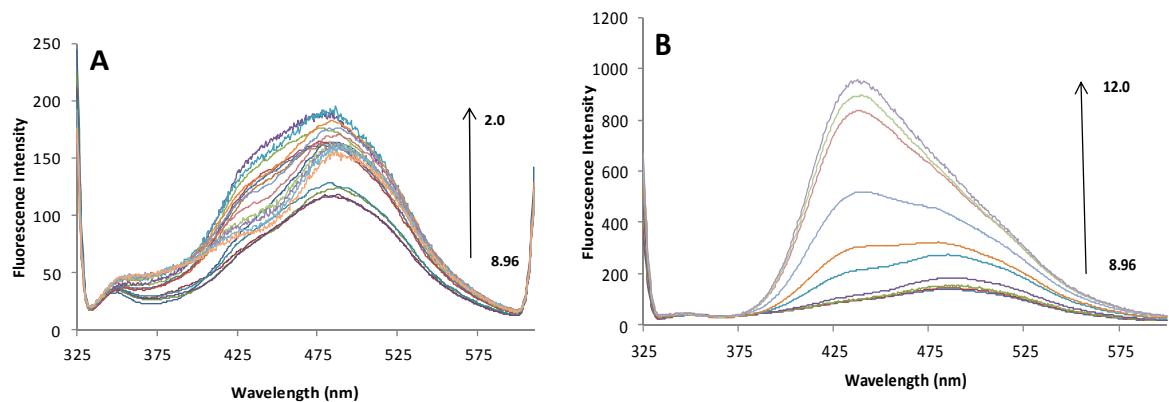
**Figure S9.** IR spectrum of receptor **2c**.



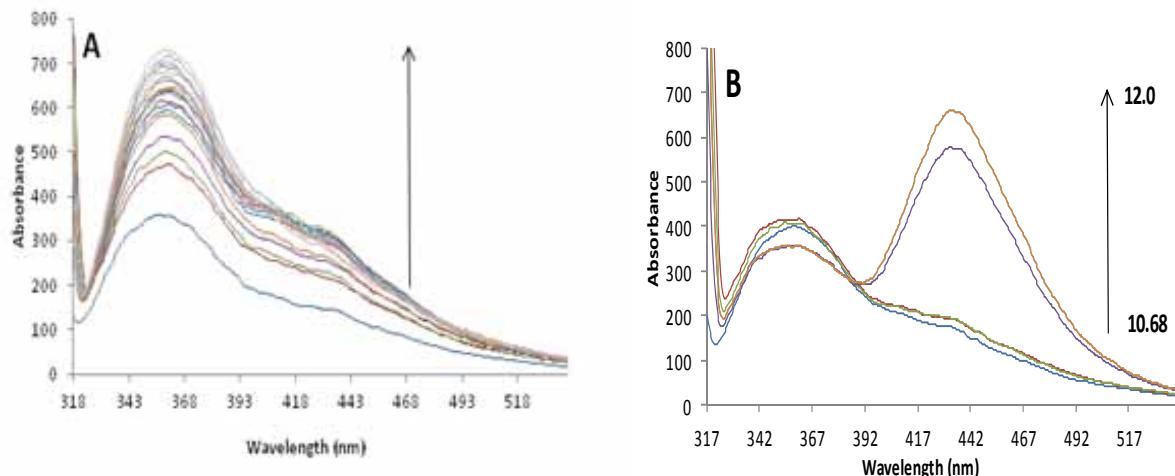
**Figure S10.** UV-Vis absorption spectra of (a) receptor **2a** (10  $\mu$ M) upon addition of 50  $\mu$ M of  $Zn^{2+}$  nitrate salt in HEPES buffered (10 mM, pH=7.0) DMF/H<sub>2</sub>O (7:3, v/v) solvent system; (b) receptor **2b** (10  $\mu$ M) upon addition of 50  $\mu$ M of  $Zn^{2+}$  nitrate salt in HEPES buffered (10 mM, pH=7.0) DMF/H<sub>2</sub>O (7:3, v/v) solvent system and (c) receptor **2c** (10  $\mu$ M) upon addition of 50  $\mu$ M of  $Zn^{2+}$  nitrate salt in HEPES buffered (10 mM, pH=7.0) DMF/H<sub>2</sub>O (7:3, v/v) solvent system.



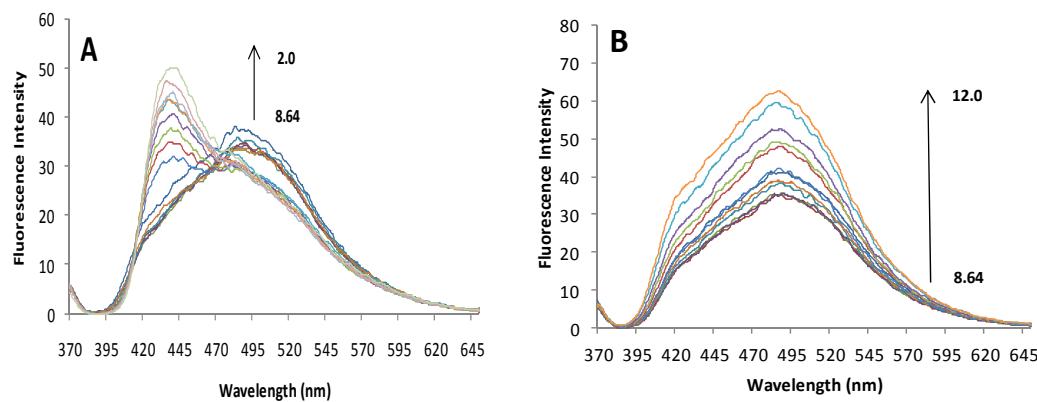
**Figure S11.** Fluorescence spectrum of (a) receptor **2a** ( $10 \mu\text{M}$ ) recorded in HEPES buffered (10 mM, pH=7.0) DMF/H<sub>2</sub>O (7:3, v/v) solvent system excited at 310 nm; (b) receptor **2b** ( $10 \mu\text{M}$ ) recorded in HEPES buffered (10 mM, pH=7.0) DMF/H<sub>2</sub>O (7:3, v/v) solvent system excited at 309 nm and (c) receptor **2c** ( $10 \mu\text{M}$ ) recorded in HEPES buffered (10 mM, pH=7.0) DMF/H<sub>2</sub>O (7:3, v/v) solvent system excited at 370 nm.



**Figure S12.** Changes in the Fluorescence spectra of **2a** upon pH titration of a solution of **2a** (10  $\mu$ M) in DMF/H<sub>2</sub>O (7:3, v/v) solvent system under: (A) acidic and (B) basic conditions.



**Figure S13.** Changes in the Fluorescence spectra of **2b** upon pH titration of a solution of **2b** (15  $\mu$ M) in DMF/H<sub>2</sub>O (7:3, v/v) solvent system under: (A) acidic and (B) basic conditions.



**Figure S14.** Changes in the Fluorescence spectra of **2c** upon pH titration of a solution of **2c** (8  $\mu$ M) in DMF/H<sub>2</sub>O (7:3, v/v) solvent system under: (A) acidic and (B) basic conditions.

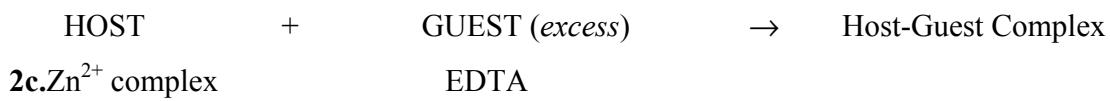
#### Kinetic analysis of the recognition process and determination of activation energy

The first order binding process can be expressed using the following equation:

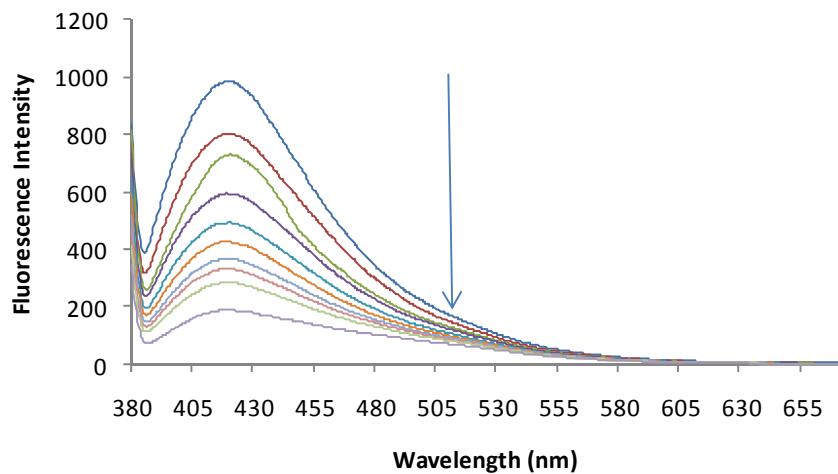
$$\ln[(A_{\infty} - A_t)/(A_{\infty} - A_0)] = kt$$

Where  $A_0$ ,  $A_t$  and  $A_{\infty}$  are the fluorescence intensities of the host-guest complex at time 0, t and infinity respectively.  $K$  is the rate constant.

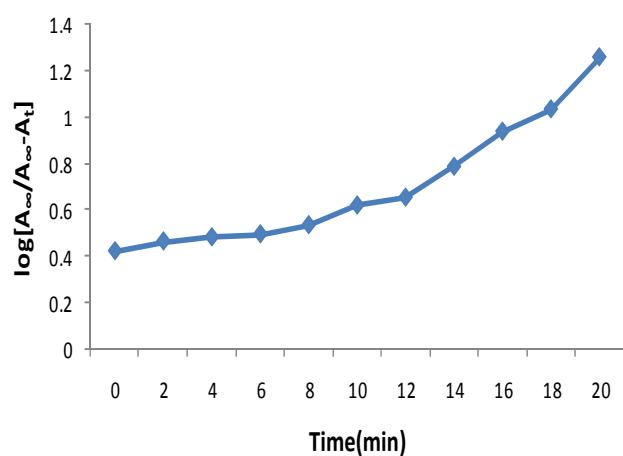
Under present investigations; for kinetic studies, the host guest reaction is progressing in the following way:



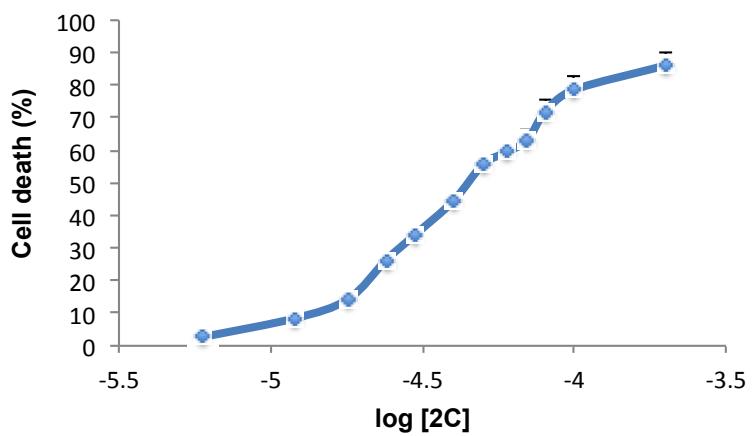
In the above reaction changes in the Fluorescence emission intensity at 418 nm were monitored for the determination of  $K$ . The excess of EDTA was added to the solution of  $\text{2c.Zn}^{2+}$ ; so that the reaction can be regarded as pseudo first order reaction i.e. kinetics depends only on the concentration of host.



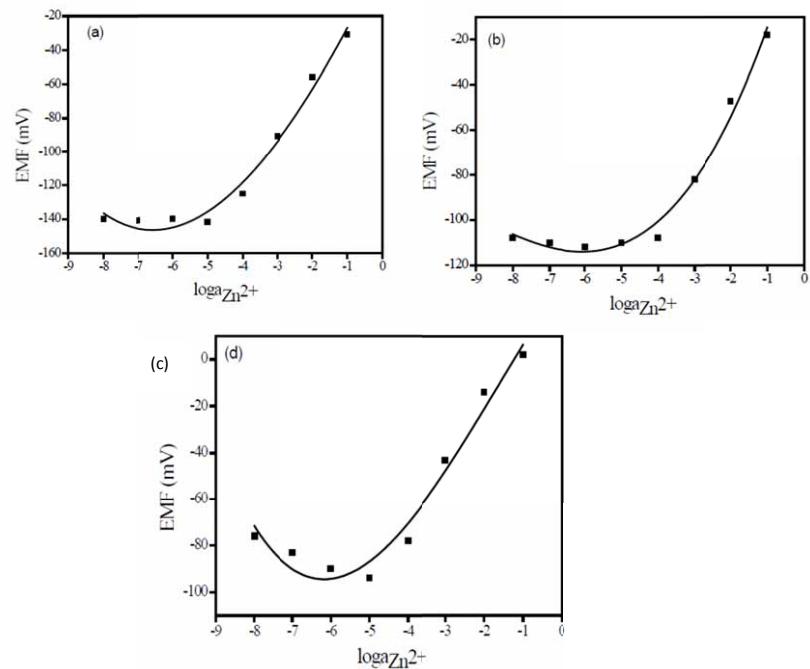
**Figure S15:** Fluorescence emission spectra of **2c**.  $Zn^{2+}$  complex ( $20 \mu M$ ) upon successive addition of EDTA (0- $40 \mu M$ ) in HEPES buffered (10 mM, pH=7.0) DMF/H<sub>2</sub>O (7:3, v/v) solvent system.



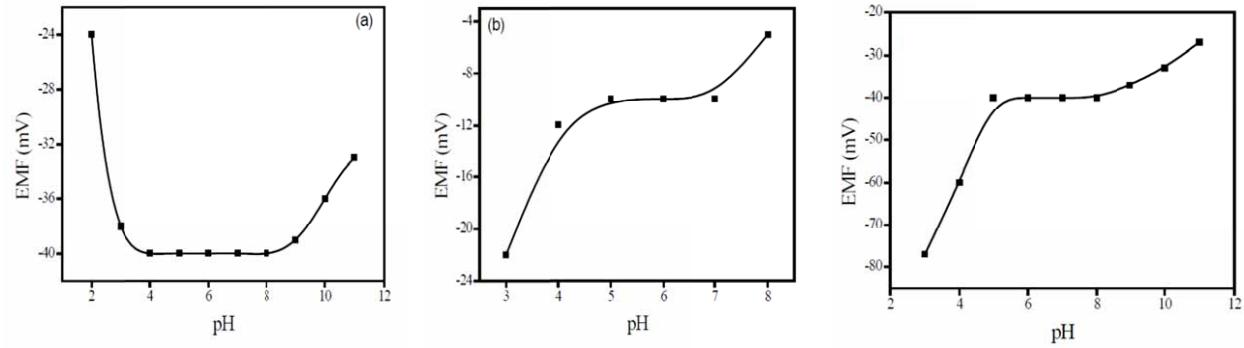
**Figure S16:** Showing the course of Fluorescence intensity at 418 nm with time (0-20 min) for  $20 \mu M$  of **2c**.  $Zn^{2+}$  complex upon addition of  $40 \mu M$  of EDTA in HEPES buffered (10 mM, pH=7.0) DMF/H<sub>2</sub>O (7:3, v/v) solvent system.



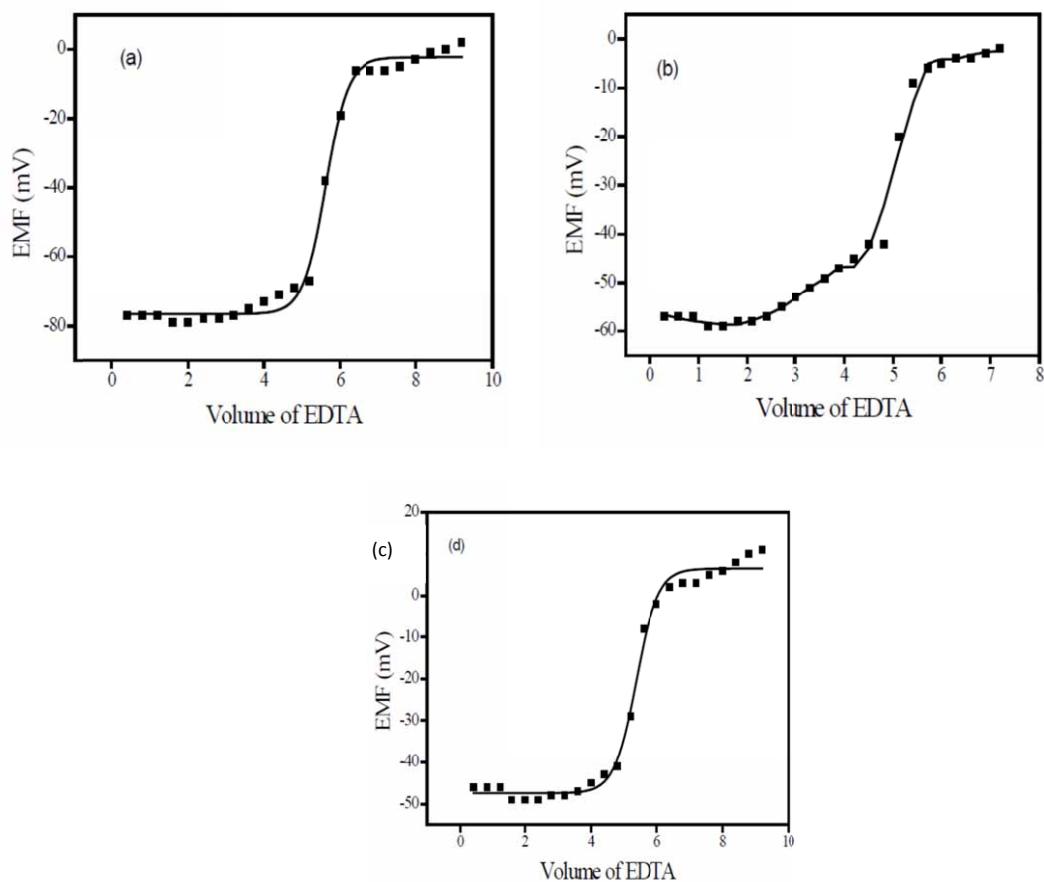
**Figure S17:** Dose-response curve for **2c** for the calculation of IC<sub>50</sub> value



**Figure S18:** Calibration curve for  $\text{Zn}^{2+}$  ion using electrode with ionophore **2a-c**.



**Figure S19:** Working pH range of electrodes with ionophore **2a-c**.



**Figure S20:** Potentiometric titration curves for electrodes with ionophore **2a-c**.

**Table S1.** Total energies of optimized keto and enol forms of receptors **2a**, **2b** and **2c**

Tautomeric forms	Total energy (eV)
Receptor <b>2a</b> (Enol Form)	-14102.943
Receptor <b>2a</b> (Keto Form)	-14105.392
Receptor <b>2b</b> (Enol Form)	-18288.385
Receptor <b>2b</b> (Keto Form)	-18288.657
Receptor <b>2c</b> (Enol Form)	-19897.040
Receptor <b>2c</b> (Keto Form)	-19897.857

**Table S2.** Slopes of calibration curves for Zn (II) ions using electrodes with ionophores **2a-c**.

S No.	Slope (mV/decade)		
	Ionophore, <b>2a</b>	Ionophore, <b>2b</b>	Ionophore, <b>2c</b>
1.	33	32	29
2.	30	30	27
3.	32	29	27
4.	33	30	27
Mean	32 ± 2.0	30.2 ± 1.5	27.5 ± 1.0

**Table S3.** Selectivity coefficients of Zn<sup>2+</sup> ion selective electrodes for different interfering ions ( $1 \times 10^{-3}$ M) by fixed interference method.

S. No.	Interfering ions	Selectivity coefficient, log K		
		<b>2a</b>	<b>2b</b>	<b>2c</b>
1.	Ca <sup>2+</sup>	-1.0	-0.2	-0.5
2.	Mg <sup>2+</sup>	0.0	0.4	-0.2
3.	Fe <sup>2+</sup>	0.5	-	-
4.	Co <sup>2+</sup>	-0.4	-0.3	-0.4
5.	Ni <sup>2+</sup>	0.7	-0.3	-0.4
6.	Cu <sup>2+</sup>	-0.4	-0.6	-0.5

7.	Pb <sup>2+</sup>	-1.3	-0.4	-1.1
8.	Cd <sup>2+</sup>	-0.4	-0.4	-0.4

**Table S4.** A comparison of quantum yield of **2c**, **2c** after complexation with Zn<sup>2+</sup>.

Sr. No.	Polymer <b>2c</b>	Polymer <b>2c</b> + Zn <sup>2+</sup>
1	0.20	0.65
2	0.22	0.67
3	0.21	0.67
Mean	0.21±0.01	0.66±0.01

Fluorescence quantum yield<sup>1</sup> was determined using optically matching standard solutions at a particular excitation wavelength and quantum yield is calculated using the equation:

$$\Phi_{fs} = \Phi_{fr} \times \frac{1 - 10^{-ArLr}}{1 - 10^{-AsLs}} \times \frac{Ns^2}{Nr^2} \times \frac{Ds}{Dr}$$

$\Phi_{fs}$  and  $\Phi_{fr}$  are the quantum yields of sample and the reference respectively,  $A_s$  and  $A_r$  are the absorbance of the sample and the reference respectively,  $D_s$  and  $D_r$  the respective areas of emission for sample and reference.  $L_s$  and  $L_r$  are the lengths of the absorption cells of sample and reference respectively.  $N_s$  and  $N_r$  are the refractive indices of the sample and reference solutions.

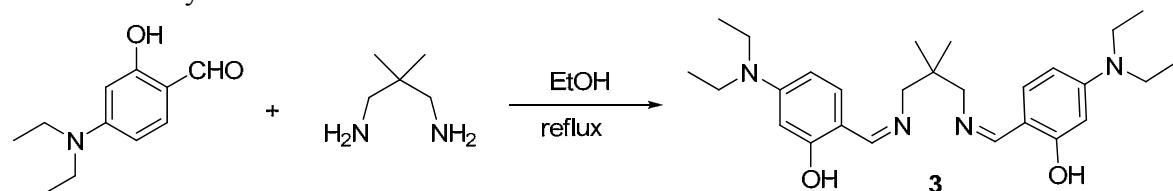
**Table S5.** A comparison of results obtained for spiked Zn<sup>2+</sup> in water samples using proposed probe and AAS.

Sample Code	Zn <sup>2+</sup> (mg/mL) added	Polymer <b>2c</b> (mean <sup>a</sup> )	AAS (mean <sup>a</sup> )
Tap water 1	0.297	0.30	0.299
Tap water 2	1.485	1.49	1.48
Tap water 3	2.97	3.04	3.01
Tap water 4	5.94	5.96	5.97
Tap water 5	14.85	14.92	14.89

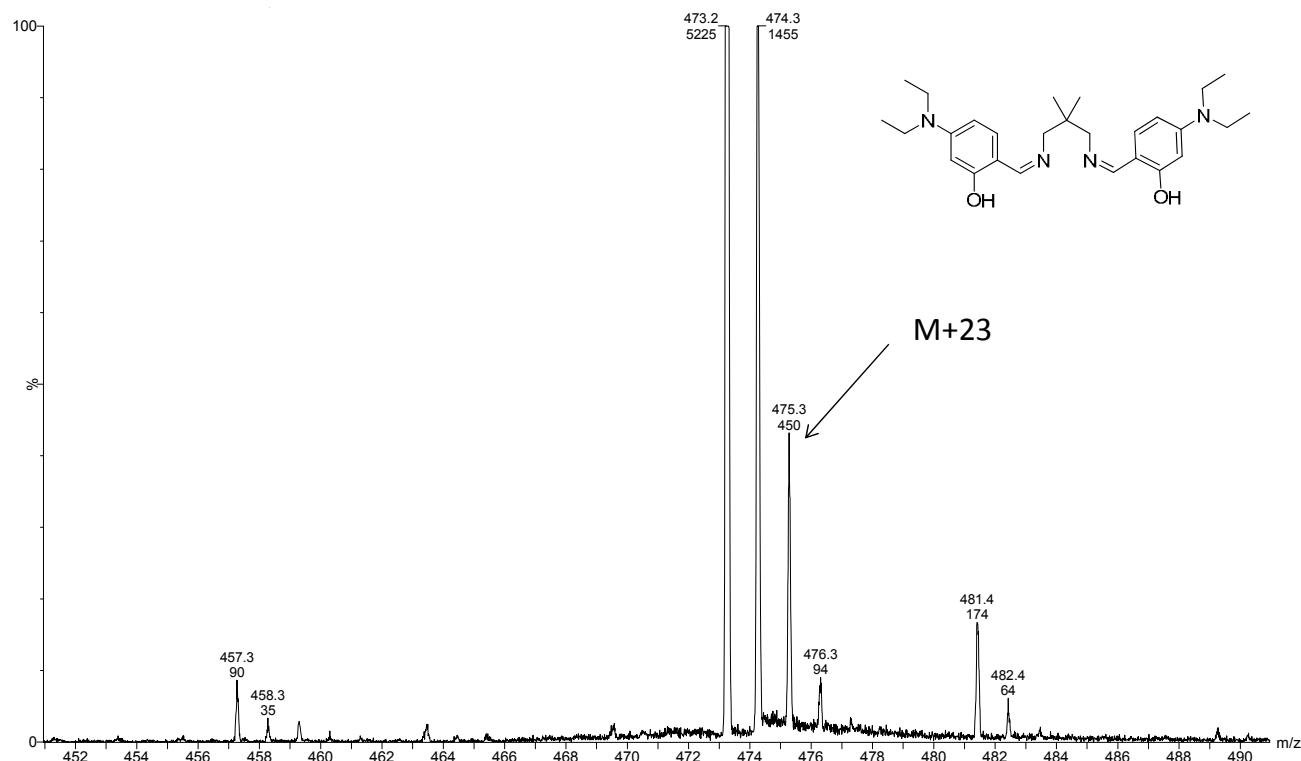
<sup>a</sup> Mean of three determinations.

To evaluate the real practical application of **2c**, the concentration of Zn<sup>2+</sup> was analysed in the environmental water samples taken from tap. The initial AAS studies of sample showed the presence of less than 0.002 µg/mL content of Zn<sup>2+</sup> in the water samples. Therefore, water samples spiked with Zn<sup>2+</sup> were analysed for Zn<sup>2+</sup> content using the **2c** in aqueous medium and AAS. The results show good agreement of spiked Zn<sup>2+</sup> content observed and values determined by AAS. The results revealed that the present probe (**2c**) can work well in environmental samples.

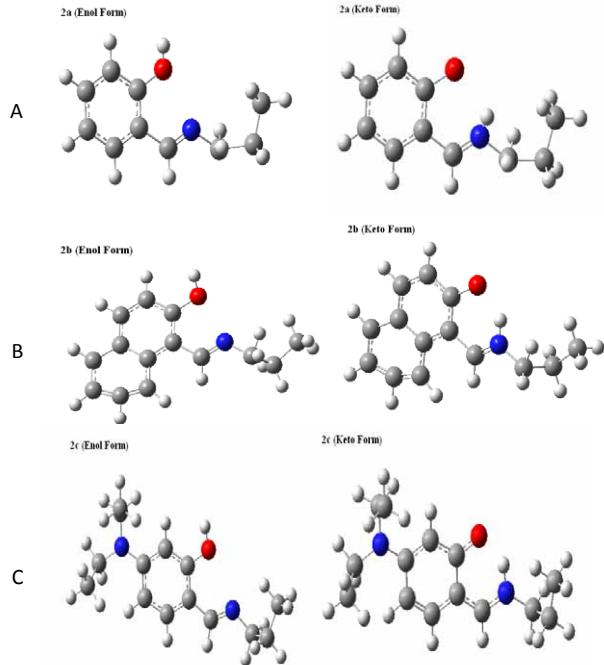
**Scheme S1.** Synthesis of monomer **3**.



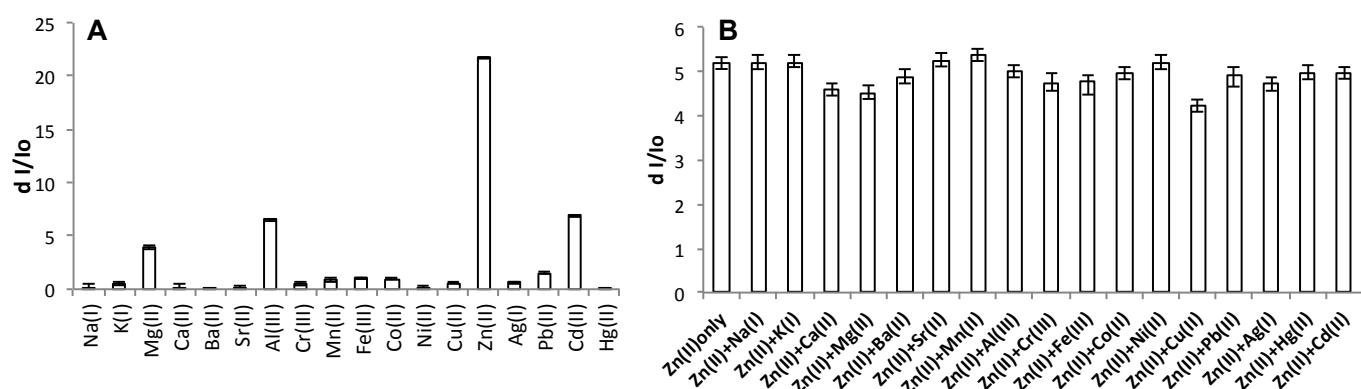
The monomer **3** was synthesized by adding 4-diethylamino-salicylaldehyde (2 mmol) to a solution of 3,3-dimethylpropylenediamine (1 mmol) in ethanol (10 ml). The mixture was refluxed for one hour. The monomer **3** precipitated as yellow solid which was filtered and washed with cold ethanol. % yield = 87%. ESI-MS ( $m/z$ ) 475 ( $M+23$ ); CHN analysis calcd. for ( $C_{27}H_{40}N_4O_2$ ): C, 71.65; H, 8.91; N, 12.38; found: C, 71.68; H, 8.89; N, 12.41.



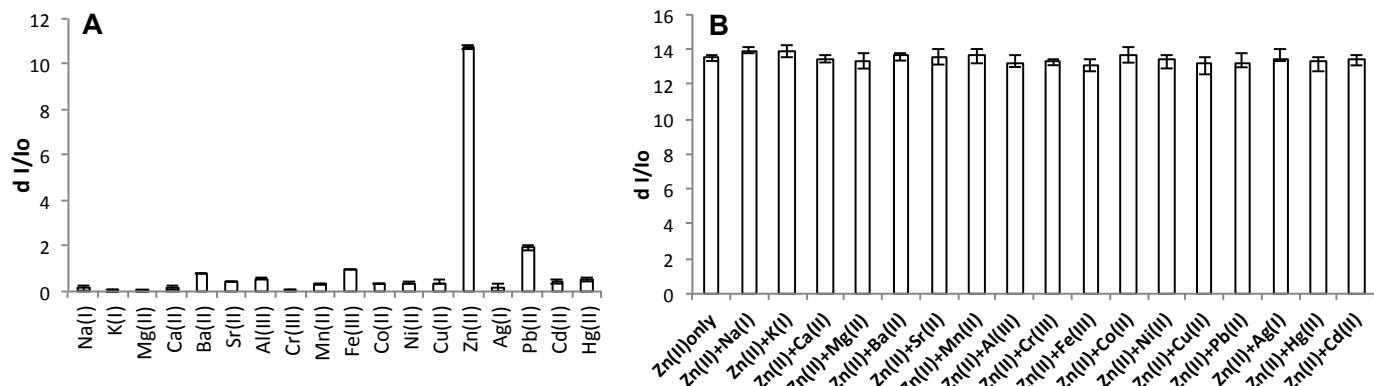
**Figure S21.** Mass spectrum of **3**.



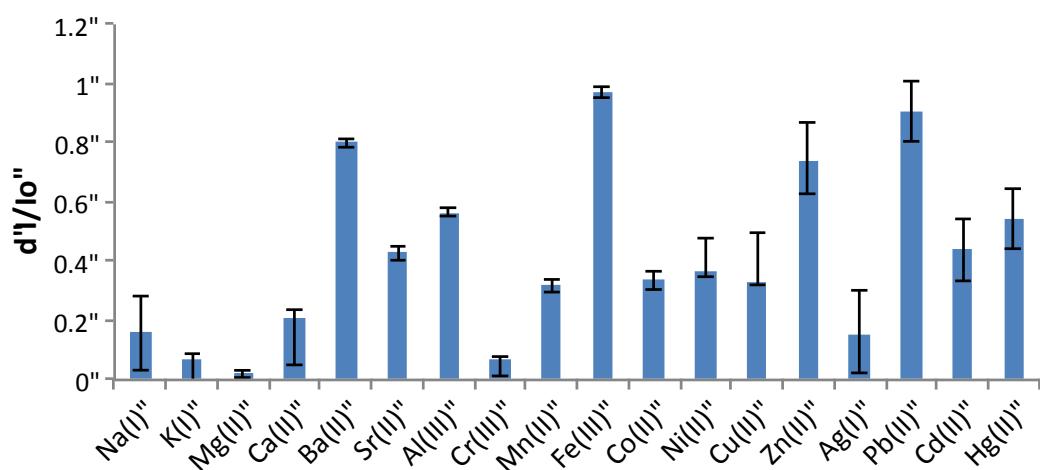
**Figure S22.** DFT calculated partial structures of receptor **2a-c**: (A) Enol and Keto forms of receptor **2a**; (B) Enol and Keto forms of receptor **2b**; (C) Enol and Keto forms of receptor **2c**; DFT calculations were performed at B3LYP/6-31G\* level; red, blue and grey spheres refer to O, N and C atoms respectively).



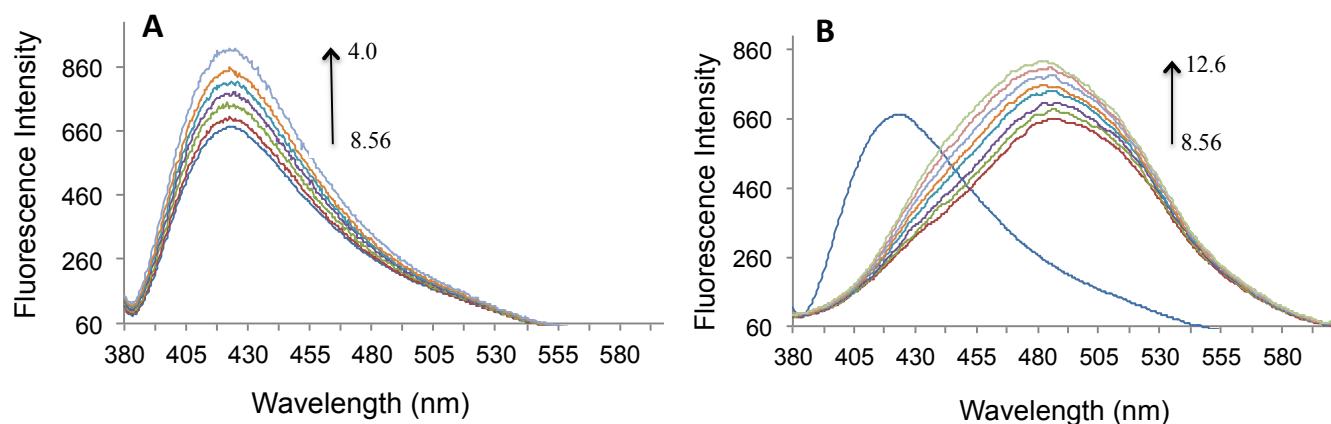
**Figure S23.** (A) Changes in fluorescence intensity of receptor **2a** ( $10 \mu\text{M}$ ) upon addition of  $50 \mu\text{M}$  of a particular metal nitrate salts in HEPES buffered (10 mM, pH=7.0) DMF/H<sub>2</sub>O (7:3, v/v); with  $\lambda_{\text{ex}} = 310 \text{ nm}$ ; (B) Influence of some metal ions on the Zn<sup>2+</sup> based triggering of fluorescence intensity of **2a**.



**Figure S24.** (A) Changes in fluorescence intensity of receptor **2c** ( $10 \mu\text{M}$ ) upon addition of  $50 \mu\text{M}$  of a particular metal nitrate salts in HEPES buffered (10 mM, pH=7.0) DMF/H<sub>2</sub>O (7:3, v/v); with  $\lambda_{\text{ex}} = 370 \text{ nm}$ ; (B) Selectivity of **2c** for Zn<sup>2+</sup> compared to other metal ions: the presence of other metal ions has no effect for Zn<sup>2+</sup> induced fluorescence intensity of **2c**.



**Figure S25.** Changes in fluorescence intensity of receptor **3** ( $10 \mu\text{M}$ ) upon addition of  $50 \mu\text{M}$  of a particular metal nitrate salts in HEPES buffered (10 mM, pH=7.0) DMF/H<sub>2</sub>O (7:3, v/v); with  $\lambda_{\text{ex}} = 365 \text{ nm}$ .



**Figure S26.** Changes in the Fluorescence spectra of **2c**.Zn<sup>2+</sup> upon pH titration of a solution of **2c**.Zn<sup>2+</sup> in DMF/H<sub>2</sub>O (7:3, v/v) solvent system under: (A) acidic and (B) basic conditions.

## References

1. J. N. Demas and G. A. Crosby, J. Phys. Chem., 1971, 75, 991.