

Supporting Information

Sequential Carboxylation/Intramolecular Cyclization Reaction of *o*-Alkynyl Acetophenone Using CO₂

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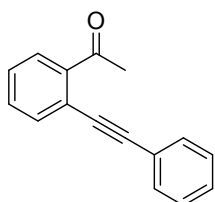
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1. General Information

NMR spectra were recorded on a Bruker AvanceII 400M type (^1H NMR, 400 MHz; ^{13}C NMR, 100 MHz) spectrometer. Their peak frequencies were referenced versus an internal standard (TMS) shifts at 0 ppm for ^1H NMR and against the solvent (CDCl_3 , 77.0 ppm) for ^{13}C NMR, respectively. Multiplicity abbreviations: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet. High resolution mass spectra (HRMS) were recorded on a Q-TOF mass spectrometry (Micromass, Wythenshawe, UK) equipped with Z-spray ionization source. Infrared spectra (IR) were measured using a Nicolet NEXUS FT-IR spectrophotometer.

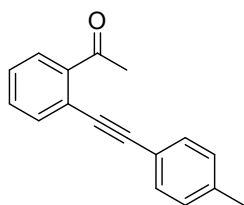
2. Synthesis of *o*-Alkynyl Ketone Substrates

Alkynyl ketones were synthesized through Sonogashira reaction of corresponding 2-bromoketones with terminal alkynes using the following protocol:^[1,2] Under a nitrogen atmosphere, to a solution 2-bromoketone (5.00 mmol) in triethylamine (20 mL), the appropriate terminal alkyne (6.00 mmol) and $\text{Pd}(\text{PPh}_3)_2\text{Cl}_2$ (0.10 mmol) were added. The reaction was stirred at room temperature for 15 min, then CuI (0.05 mmol) was added. The reaction mixture was then stirred at 80 °C until all starting materials was consumed (TLC analysis). The solvent was evaporated under reduced pressure and the crude was purified by flash chromatography over a silica gel column to yield corresponding alkynyl ketone.



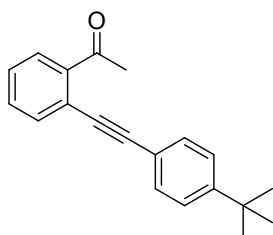
2-(Phenylethynyl)phenylethanone (1a)

^1H NMR (400 MHz, CDCl_3): δ = 7.76-7.35 (m, 9H), 2.79 (s, 3H).



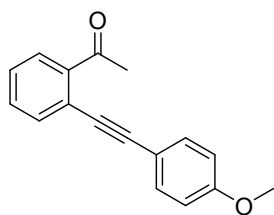
2-(4-Methylphenylethynyl)phenylethanone (1b)

^1H NMR (400 MHz, CDCl_3): δ = 7.75 (d, J = 7.7 Hz, 1H), 7.62 (d, J = 7.6 Hz, 1H), 7.49-7.37 (m, 4H), 7.18 (d, J = 7.8 Hz, 2H), 2.80 (s, 3H), 2.38 (s, 3H).



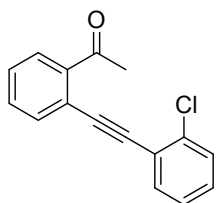
2-(4-*tert*-Butylphenylethynyl)phenylethanone (1c)

^1H NMR (400 MHz, CDCl_3): δ = 7.75 (d, J = 7.7 Hz, 1H), 7.62 (d, J = 7.6 Hz, 1H), 7.53-7.35 (m, 6H), 2.81 (s, 3H), 1.33 (s, 9H).



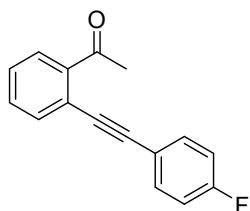
2-(4-Methoxyphenylethynyl)phenylethanone (1d)

$^1\text{H NMR}$ (400 MHz, CDCl_3): $\delta = 7.75$ (d, $J = 7.1$ Hz, 1H), 7.61 (d, $J = 7.2$ Hz, 1H), 7.55-7.43 (m, 3H), 7.38 (t, $J = 7.6$ Hz, 1H), 6.90 (d, $J = 8.7$ Hz, 2H), 3.84 (s, 3H), 2.80 (s, 3H).



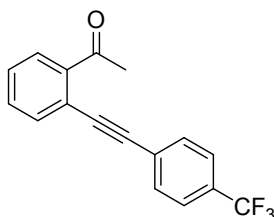
2-(2-Chlorophenylethynyl)phenylethanone (1e)

$^1\text{H NMR}$ (400 MHz, CDCl_3): $\delta = 7.76$ (d, $J = 7.7$ Hz, 1H), 7.68 (d, $J = 7.6$ Hz, 1H), 7.58 (dd, $J = 7.1, 1.9$ Hz, 1H), 7.49 (t, $J = 7.5$ Hz, 1H), 7.42 (t, $J = 7.6$ Hz, 2H), 7.34-7.21 (m, 2H), 2.82 (s, 3H).



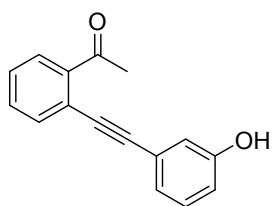
2-(4-Fluorophenylethynyl)phenylethanone (1f)

$^1\text{H NMR}$ (400 MHz, CDCl_3): $\delta = 7.76$ (d, $J = 7.8$ Hz, 1H), 7.62 (d, $J = 7.6$ Hz, 1H), 7.58-7.45 (m, 3H), 7.41 (t, $J = 7.6$ Hz, 1H), 7.07 (t, $J = 8.7$ Hz, 2H), 2.77 (s, 3H).



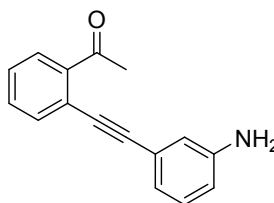
2-(4-Trifluoromethylphenylethynyl)phenylethanone (1g)

$^1\text{H NMR}$ (400 MHz, CDCl_3): $\delta = 7.78$ (dd, $J = 7.7, 1.1$ Hz, 1H), 7.69-7.60 (m, 5H), 7.53-7.43 (m, 2H), 2.76 (s, 3H).



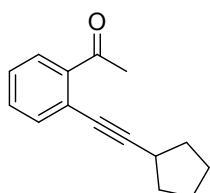
2-(3-Hydroxyphenylethynyl)phenylethanone (1h)

$^1\text{H NMR}$ (400 MHz, CDCl_3): $\delta = 7.77$ (d, $J = 7.6$ Hz, 1H), 7.62 (d, $J = 7.6$ Hz, 1H), 7.48 (t, $J = 7.4$ Hz, 1H), 7.40 (t, $J = 7.7$ Hz, 1H), 7.29-6.99 (m, 3H), 6.93-6.84 (m, 1H), 6.07 (s, 1H), 2.80 (s, 3H).



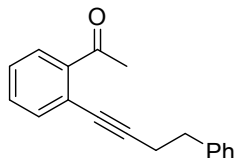
2-(3-Aminophenylethynyl)phenylethanone (1i)

$^1\text{H NMR}$ (400 MHz, CDCl_3): $\delta = 7.75$ (d, $J = 6.1$ Hz, 1H), 7.61 (s, 1H), 7.47-7.40 (m, 2H), 7.15 (s, 1H), 6.96-6.87 (m, 2H), 6.70 (s, 1H), 3.74 (s, 2H), 2.80 (s, 3H).



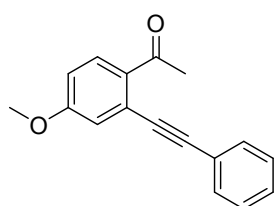
2-(Cyclopentylethynyl)phenylethanone (1j)

$^1\text{H NMR}$ (400 MHz, CDCl_3): δ = 7.66 (dd, J = 7.8, 1.1 Hz, 1H), 7.47 (dd, J = 7.7, 1.0 Hz, 1H), 7.39 (td, J = 7.5, 1.4 Hz, 1H), 7.32 (td, J = 7.6, 1.4 Hz, 1H), 2.92-2.85 (m, 1H), 2.73 (s, 3H), 2.09-1.94 (m, 2H), 1.84-1.53 (m, 6H).



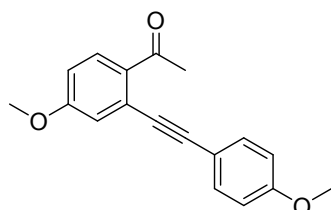
2-(4-Phenyl-1-butynyl)phenylethanone (1k)

$^1\text{H NMR}$ (400 MHz, CDCl_3): δ = 7.64 (d, J = 7.7 Hz, 1H), 7.45 (d, J = 7.6 Hz, 1H), 7.41-7.35 (m, 1H), 7.35-7.20 (m, 6H), 2.94 (t, J = 7.4 Hz, 2H), 2.76 (t, J = 7.4 Hz, 2H), 2.59 (s, 3H).



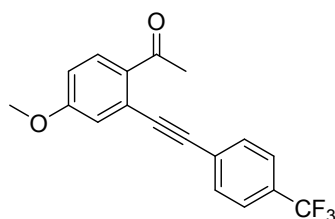
2-(Phenylethynyl)-4-methoxyphenylethanone (1l)

$^1\text{H NMR}$ (400 MHz, CDCl_3): δ = 7.84 (d, J = 8.8 Hz, 1H), 7.60-7.53 (m, 2H), 7.42-7.34 (m, 3H), 7.11 (d, J = 2.6 Hz, 1H), 6.92 (dd, J = 8.8, 2.7 Hz, 1H), 3.88 (s, 3H), 2.77 (s, 3H).



2-(4-Methoxyphenylethynyl)-4-methoxyphenylethanone (1m)

$^1\text{H NMR}$ (400 MHz, CDCl_3): δ = 7.83 (d, J = 8.8 Hz, 1H), 7.55-7.47 (m, 2H), 7.09 (d, J = 2.6 Hz, 1H), 6.93-6.87 (m, 3H), 3.88 (s, 3H), 3.84 (s, 3H), 2.77 (s, 3H).



2-(4-Trifluoromethylphenylethynyl)-4-methoxyphenylethanone (1n)

$^1\text{H NMR}$ (400 MHz, CDCl_3): δ = 7.83 (d, J = 8.8 Hz, 1H), 7.67 (d, J = 8.3 Hz, 2H), 7.61 (d, J = 8.3 Hz, 2H), 7.12 (d, J = 2.6 Hz, 1H), 6.93 (dd, J = 8.8, 2.6 Hz, 1H), 3.87 (s, 3H), 2.72 (s, 3H).

3. Computational Study

DFT calculations were performed using the Gaussian 09 package.³ Firstly, many DFT methods such as B3LYP, B3PW91, TPSSh were used to find the TS which leads to 5-*exo* oxygen cyclization, but all those methods failed to find the TS. So a flexible scan of the potential energy surface (PES) of the formed C-O bond was carried out (Figure S1).

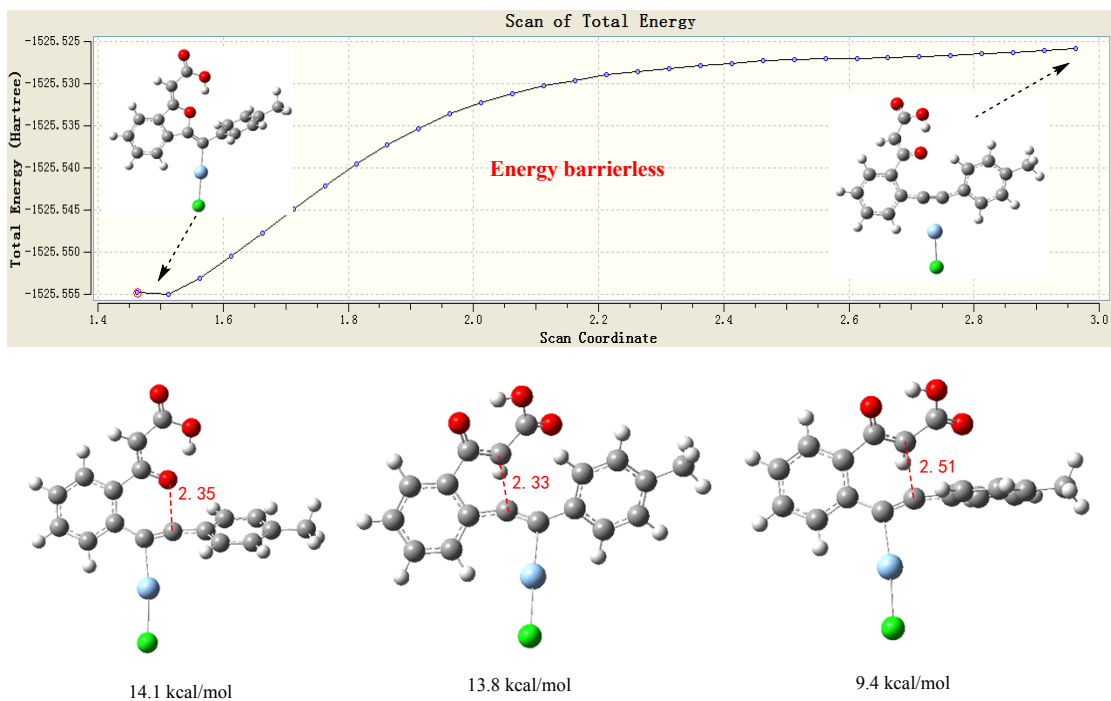


Figure S1. Top: scan curve for the 5-*exo* oxygen cyclization. Below: the other three transition state structures and the solvation free energy computed by the B3LYP method.

4. References

- [1] M. Dell'Acqua, G. Abbiati, A. Arcadi, E. Rossi, *Org. Biomol. Chem.* **2011**, *9*, 7836.
- [2] N. Chernyak, S. I. Gorelsky, V. Gevorgyan, *Angew. Chem. Int. Ed.* **2011**, *50*, 2342.
- [3] Gaussian 09, Revision B.01. Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Scalmani, G.; Barone, V.; Mennucci, B.; Petersson, G. A.; Nakatsuji, H.; Caricato, M.; Li, X.; Hratchian, H. P.; Izmaylov, A. F.; Bloino, J.; Zheng, G.; Sonnenberg, J. L.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Vreven, T.; Montgomery, J. A.; Peralta, J. E.; Ogliaro, F.; Bearpark, M.; Heyd, J. J.; Brothers, E.; Kudin, K. N.; Staroverov, V. N.; Kobayashi, R.; Normand, J.; Raghavachari, K.; Rendell, A.; Burant, J. C.; Iyengar, S. S.; Tomasi, J.; Cossi, M.; Rega, N.; Millam, J. M.; Klene, M.; Knox, J. E.; Cross, J. B.; Bakken, V.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R. E.; Yazyev, O.; Austin, A. J.; Cammi, R.; Pomelli, C.; Ochterski, J. W.; Martin, R. L.; Morokuma, K.; Zakrzewski, V. G.; Voth, G. A.; Salvador, P.; Dannenberg, J. J.; Dapprich, S.; Daniels, A. D.; Farkas, Foresman, J. B.; Ortiz, J. V.; Cioslowski, J.; Fox, D. J. Gaussian Inc., Wallingford CT, 2009.

5. Copies of ^1H NMR and ^{13}C NMR Spectra of Substrates and Products

