

# A Silver-Catalyzed Transfer Hydrogenation of Aldehydes in Air and Water (Supporting Information)

Mingxin Liu,<sup>a</sup> Feng Zhou,<sup>a</sup> Zhenhua Jia,<sup>a,b</sup> and Chao-Jun Li\*

## Contents

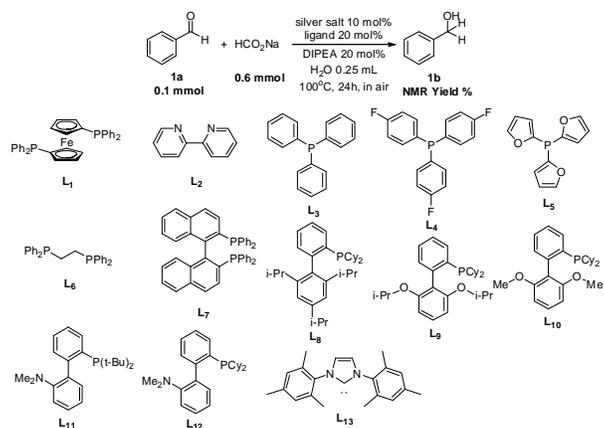
I	General Information
II	Condition Screening
III	General Procedures
IV	Identification of Products
V	NMR Datas

### I. General Information

All transfer hydrogenation reactions were carried out under air. All manipulation and purification procedures were carried out with reagent-grade solvents. Analytical thin-layer chromatography (TLC) was performed using E. Merck silica gel 60 F<sub>254</sub> precoated plates (0.25 mm). Flash chromatography was performed with Biotage Isolera One Flash Purification System, using Biotage SNAP Ultra 25g prepared column. Nuclear magnetic resonance (NMR) spectra were recorded on Varian MERCURY plus-300 spectrometer (<sup>1</sup>H 300 MHz, <sup>13</sup>C 75 MHz) or a Varian MERCURY plus-400 spectrometer (<sup>1</sup>H 400 MHz, <sup>13</sup>C 100 MHz). Chemical shifts for <sup>1</sup>H NMR spectra are reported in parts per million (ppm) from tetramethylsilane with the solvent resonance as the internal standard (CDCl<sub>3</sub>: δ 7.26 ppm). Chemical shifts for <sup>13</sup>C NMR spectra are reported in parts per million (ppm) from tetramethylsilane with the solvent as the internal standard (CDCl<sub>3</sub>: δ 77.0 ppm). Data are reported as following: chemical shift, multiplicity (s = singlet, d = doublet, dd = doublet of doublets, t = triplet, q = quartet, m = multiplet, br = broad signal), and integration.

### II. Condition Screening

**Table 1.** Condition Screening for Aromatic Aldehydes



Silver Salt	Ligand	NMR Yield <sup>a</sup>
AgPF <sub>6</sub>	L1	2 %
AgF	L1	3 %
AgCl	L1	N.D.
AgBr	L1	N.D.
AgI	L1	N.D.
AgOTf	L1	3 %
AgF	L2	N.D.
AgF	L3	N.D.
AgF	L4	6 %
AgF	L5	N.D.
AgF	L6	N.D.
AgF	L7	N.D.
AgF	L8	N.D.
AgF	L9	61 %
AgF	L10	66 %
AgF	L11	50 %
AgF	L12	>99 % (92 %) <sup>b</sup>
AgF	L13	N.D.
AgPF <sub>6</sub>	L12	38 %
AgCl	L12	85 %
AgBr	L12	11 %
AgI	L12	N.D.
//	L12	N.D.
AgF	//	N.D.
AgF	L12	33 % <sup>c</sup>
AgF	L12	26 % <sup>d</sup>
AgF	L12	80 % <sup>e</sup>
AgF	L12	9 % <sup>f</sup>
AgF	L12	N.D. <sup>g</sup>
AgF	L12	N.D. <sup>h</sup>

<sup>a</sup> <sup>1</sup>HNMR yields were determined by using mesitylene as the internal standard;

<sup>b</sup> Isolated Yield;

<sup>c</sup> Reaction was performed without base.

<sup>d</sup> Reaction was carried out without solvent

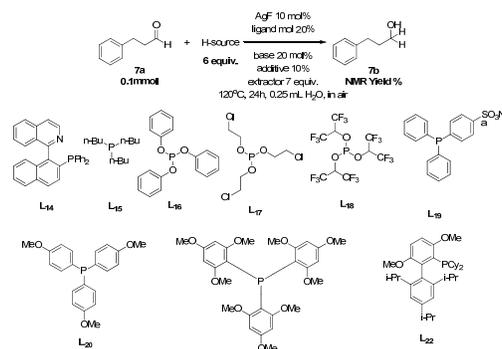
<sup>e</sup> Reaction was carried out in Ethanol

<sup>f</sup> Reaction was carried out in Acetonitrile

<sup>g</sup> Reaction was carried out in Acetone

<sup>h</sup> Reaction was carried out in N-N-dimethylformamide

**Table 2.** Condition Screening for Aliphatic Aldehydes



H-Source	base	Ligand	additive	extractor	NMR Yield
HCO <sub>2</sub> Na	DIPEA	L12	//	//	3 % <sup>a</sup>
HCO <sub>2</sub> H	//	L12	//	//	n.d. <sup>a</sup>
HCO <sub>2</sub> H·DIPEA	DIPEA	L12	//	//	6 % <sup>a</sup>
HCO <sub>2</sub> H·DIPEA	CsF	L12	//	//	7 % <sup>a</sup>
HCO <sub>2</sub> H·DIPEA	CsF	L12	LiF	//	n.d. <sup>a</sup>
HCO <sub>2</sub> H·DIPEA	CsF	L12	//	//	11 %
HCO <sub>2</sub> H·NH <sub>3</sub>	CsF	L12	//	//	n.d.
HCO <sub>2</sub> H·1/2TMEDA	CsF	L12	//	//	n.d.
HCO <sub>2</sub> H·1/2DABCO	CsF	L12	//	//	n.d.
HCO <sub>2</sub> H·DBU	CsF	L12	//	//	10 %
HCO <sub>2</sub> H·DIPEA	CsF	L1	//	//	n.d.
HCO <sub>2</sub> H·DIPEA	CsF	L2	//	//	n.d.
HCO <sub>2</sub> H·DIPEA	CsF	L4	//	//	n.d.
HCO <sub>2</sub> H·DIPEA	CsF	L5	//	//	n.d.
HCO <sub>2</sub> H·DIPEA	CsF	L7	//	//	n.d.
HCO <sub>2</sub> H·DIPEA	CsF	L14	//	//	n.d.
HCO <sub>2</sub> H·DIPEA	CsF	L8	//	//	trace
HCO <sub>2</sub> H·DIPEA	CsF	L13	//	//	n.d.
HCO <sub>2</sub> H·DIPEA	CsF	L15	//	//	n.d.
HCO <sub>2</sub> H·DIPEA	CsF	L16	//	//	trace
HCO <sub>2</sub> H·DIPEA	CsF	L17	//	//	n.d.
HCO <sub>2</sub> H·DIPEA	CsF	L18	//	//	n.d.
HCO <sub>2</sub> H·DIPEA	CsF	L19	//	//	n.d.
HCO <sub>2</sub> H·DIPEA	CsF	L20	//	//	n.d.
HCO <sub>2</sub> H·DIPEA	CsF	L21	//	//	n.d.
HCO <sub>2</sub> H·DIPEA	CsF	L22	//	//	21 %
HCO <sub>2</sub> H·DIPEA	CsF	L10	//	//	15 %
HCO <sub>2</sub> H·DIPEA	CsF	L10	//	DIPEA	30 %
HCO <sub>2</sub> H·DIPEA	CsF	L10	//	PhCl	30 %
HCO <sub>2</sub> H·DIPEA	CsF	L10	TfOH	PhCl	55 %
HCO <sub>2</sub> H·DIPEA	CsF	L10	Benzoic Acid	PhCl	12 %
HCO <sub>2</sub> H·DIPEA	CsF	L10	CF <sub>3</sub> CO <sub>2</sub> H	PhCl	11 %
HCO <sub>2</sub> H·DIPEA	CsF	L10	TfOH	PhCl	75 % <sup>b</sup>
HCO <sub>2</sub> H·DIPEA	CsF	L22	TfOH	PhCl	99 %
HCO <sub>2</sub> H·DIPEA	//	L10	TfOH	PhCl	42 %

### III. General Procedures

**(Synthesis of AgF-DavePhos complex; all the other complexes used in the study were prepared in the same way).** An oven-dried reaction vessel, charged with 12.6 mg silver (I) fluoride (0.1 mmol, 1 equiv) and 78.7 mg 2-dicyclohexylphosphino-2'-(N,N-dimethylamino)biphenyl (DavePhos, 0.2 mmol, 2 equiv), is flushed with argon 3 times. 2.5 mL of dry, air-free methylene chloride (DCM) is added into the vessel. The vessel is then sealed and stirred at room temperature. After stirring overnight (12 h), the mixture is stripped of solvent and the resulting solid is kept under vacuum for 1 h before ready to use.

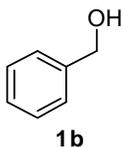
**(General Procedures for the Reduction of Aromatic Aldehydes).** To a stirred solution of 81.6 mg sodium formate (1.2 mmol, 6 equiv) in 0.5 mL distilled H<sub>2</sub>O in air, 9.1 mg of prepared AgF-DavePhos complex (0.02 mmol, 0.1 equiv) is added, along with 20.5  $\mu$ L of benzaldehyde (0.2 mmol, 1 equiv) and 7  $\mu$ L of diisopropylethylamine (DIPEA, 0.04 mmol, 0.2 equiv). The vessel is then sealed and stirred at 100°C for 24h. Then, the reaction mixture is cooled to room temperature, extracted with methylene chloride, and the organic phase is washed with brine. The organic phase is then stripped of solvent and the oily crude product is collected. Further purification can be carried out with flash chromatography to give the product in 19.5 mg (92% yield).

**(General Procedures for the Reduction of Aliphatic Aldehydes).** To a stirred vial of 2 mL H<sub>2</sub>O in air, 45  $\mu$ L of formic acid (1.2 mmol, 6 equiv) and 209  $\mu$ L of diisopropylethylamine (DIPEA, 1.2 mmol, 6 equiv) are added. The mixture is kept stirring until the whole solution is transparent and clear. All the solution is then transferred into a reaction vessel which is charged with 9.5 mg of prepared AgF-SPhos complex (0.02 mmol, 0.1 equiv) and 6.2 mg of cesium fluoride (0.04 mmol, 0.2 equiv) in air. 26.4  $\mu$ L hydrocinnamaldehyde (0.2 mmol, 1 equiv), 1.8  $\mu$ L trifluoromethanesulfonic acid (0.02 mmol, 0.1 equiv) and 142  $\mu$ L chlorobenzene (1.4 mmol, 7 equiv) are then added and the reaction vessel is sealed. The vessel is stirred at 120°C for 24h before cooled down to room temperature. The mixture is extracted with methylene chloride and the resulting organic phase is washed with brine. The solution is then concentrated and subject to flash chromatography to give the desired product in 19.0 mg (71% yield.)

### IV. Identification of Products

All compounds are literature known and the data reported herein are consistent with the literature reports.

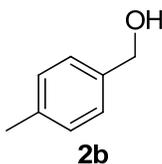
#### Compound 1b:



<sup>1</sup>H-NMR (ppm): 7.38 (m, 5H), 7.30 (m, 1H), 4.70 (s, 2H), 1.62 (br, 1H).

<sup>13</sup>C-NMR (ppm): 140.8, 128.5, 127.7, 127.0, 65.3.

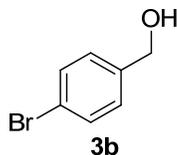
#### Compound 2b:



$^1\text{H-NMR}$  (ppm): 7.27 (m, 2H), 7.18 (m, 2H), 4.65 (s, 2H), 2.36 (s, 3H), 1.63 (br, 1H).

$^{13}\text{C-NMR}$  (ppm): 137.9, 137.4, 129.3, 127.2, 65.2, 21.1.

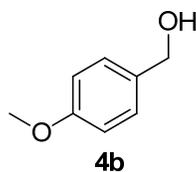
**Compound 3b:**



$^1\text{H-NMR}$  (ppm): 7.48 (m, 2H) 7.23 (m, 2H), 4.64 (s, 2H), 1.86 (br, 1H).

$^{13}\text{C-NMR}$  (ppm): 139.7, 131.7, 128.6, 121.4, 64.5.

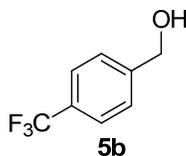
**Compound 4b:**



$^1\text{H-NMR}$  (ppm): 7.29 (m, 2H), 6.89 (m, 2H), 4.63 (s, 2H), 3.81 (s, 3H), 1.59 (br, 1H).

$^{13}\text{C-NMR}$  (ppm): 133.1, 129.4, 128.5, 113.9, 65.1, 55.3.

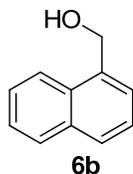
**Compound 5b:**



$^1\text{H-NMR}$  (ppm): 7.62(d, J=8.19Hz, 2H); 7.50 (d, J=8.19Hz, 2H); 4.78(s, 2H), 1.67 (br, 1H).

$^{13}\text{C-NMR}$  (ppm): 144.6, 130.5, 126.8, 125.4(q, J<sub>F-C</sub>=4.02Hz), 122.3, 64.5.

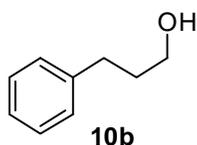
**Compound 6b:**



$^1\text{H-NMR}$  (ppm): 8.07 (m, 1H), 7.89 (m, 1H), 7.80 (m, 1H), 7.53 (m, 2H), 7.46 (m, 2H), 5.06(s, 2H), 2.49 (br, 1H).

$^{13}\text{C-NMR}$  (ppm): 136.3, 133.7, 131.2, 128.6, 128.5, 126.3, 125.8, 125.4, 125.3, 123.6, 63.4.

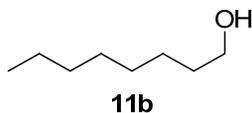
**Compound 10b:**



$^1\text{H-NMR}$  (ppm): 7.29 (m, 2H), 7.20 (m, 3H), 3.68 (t,  $J=6.44\text{Hz}$ , 2H), 2.72 (t,  $J=6.41\text{ Hz}$ , 2H), 1.90 (m, 2H), 1.64 (br, 1H).

$^{13}\text{C-NMR}$  (ppm): 141.8, 128.4 (2 peaks), 125.9, 62.3, 34.2, 32.1.

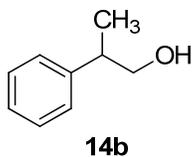
**Compound 11b:**



$^1\text{H-NMR}$  (ppm): 3.64 (t,  $J=6.73\text{Hz}$ , 2H), 1.55 (q,  $J=6.73\text{Hz}$ , 2H), 1.49 (br, 1H), 1.41-1.20 (m, 10H), 0.88 (m, 3H).

$^{13}\text{C-NMR}$  (ppm): 63.1, 32.8, 31.8, 29.3 (2 peaks), 25.6, 22.7, 14.1.

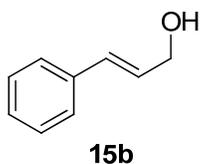
**Compound 14b:**



$^1\text{H-NMR}$  (ppm): 7.34-7.23 (m, 5H), 3.70 (m, 2H), 2.95 (m, 1H), 1.61 (br, 1H), 1.28 (d,  $J=7.03\text{Hz}$ , 3H).

$^{13}\text{C-NMR}$  (ppm): 143.7, 128.6, 127.5, 126.7, 68.7, 42.4, 17.6.

**Compound 15b:**



$^1\text{H-NMR}$  (ppm): 7.43-7.19 (m, 5H), 6.62 (d,  $J=16.09\text{Hz}$ , 1H), 6.36 (dt,  $J=16.09, 5.56\text{Hz}$ , 1H), 4.32 (d,  $J=5.56\text{ Hz}$ , 2H), 1.72 (br, 1H).

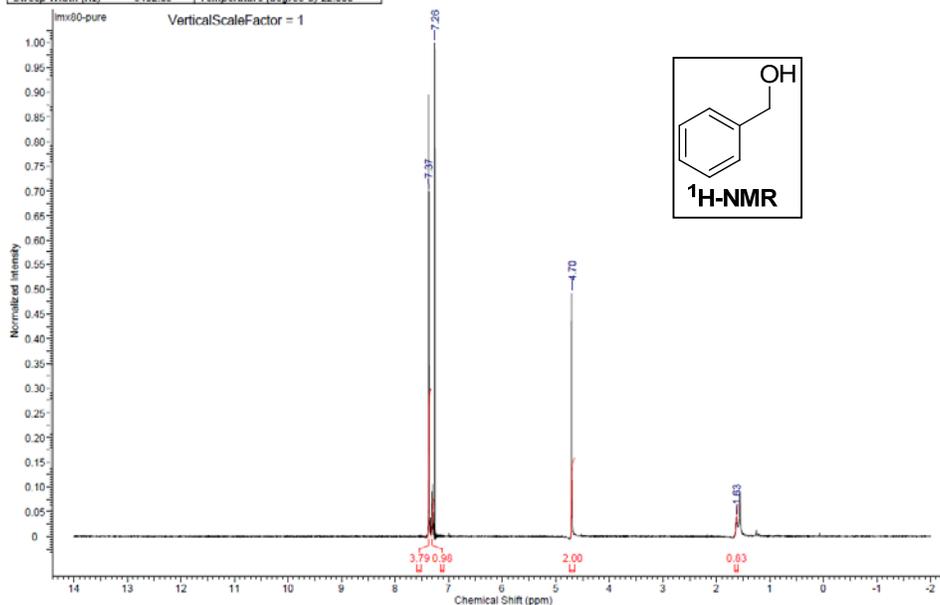
$^{13}\text{C-NMR}$  (ppm): 136.6, 131.1, 128.6, 127.7, 126.4, 63.7.

**V. NMR Datas**

This report was created by ACD/NMR Processor Academic Edition. For more information go to [www.acdlabs.com/nmrproc/](http://www.acdlabs.com/nmrproc/)

2013/12/2 22:35:31

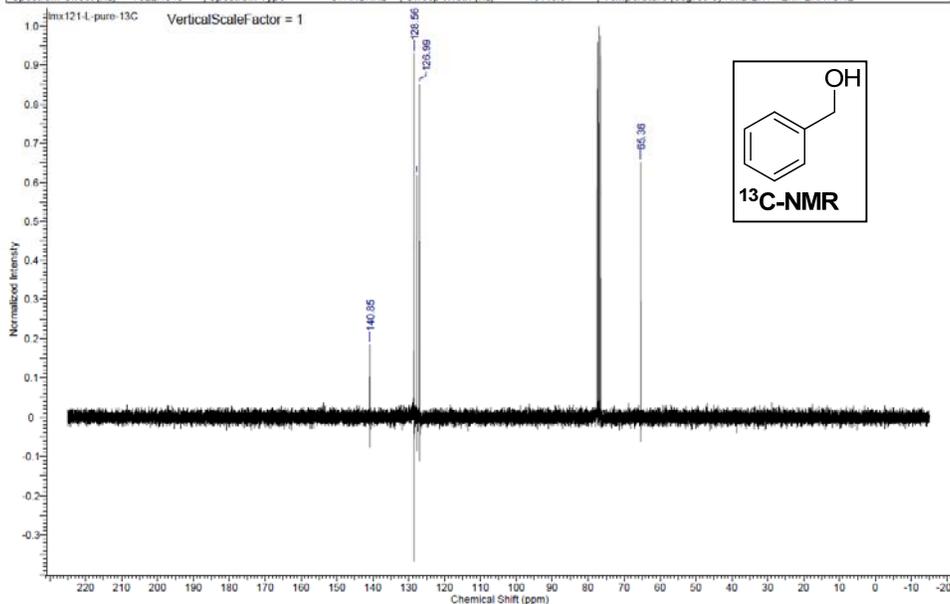
Acquisition Time (sec)	Comment	Std Proton parameters	Date
1.9980			Feb 16 2013
Date Stamp	Feb 16 2013	File Name	H <sup>1</sup> NMR\mx60-pure fidfid
Nucleus	1H	Number of Transients	8
Receiver Gain	20.00	Original Points Count	12791
Sweep Width (Hz)	6402.05	Solvent	CHLOROFORM-d
		Temperature (degree C)	22.000
		Points Count	16384
		Pulse Sequence	s2pul
		Spectrum Offset (Hz)	2400.7075
		Spectrum Type	STANDARD



This report was created by ACD/NMR Processor Academic Edition. For more information go to [www.acdlabs.com/nmrproc/](http://www.acdlabs.com/nmrproc/)

2013/12/2 22:37:19

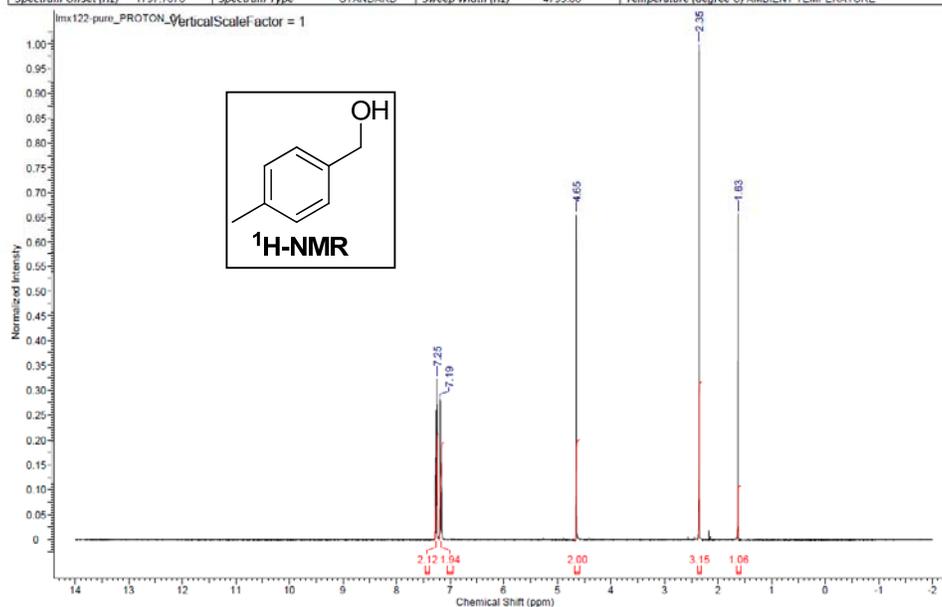
Acquisition Time (sec)	Comment	Std Carbon experiment	Date
1.3005			Mar 5 2013
Date Stamp	Mar 5 2013	File Name	H <sup>13</sup> NMR\mx121-l-pure-13C fidfid
Nucleus	13C	Number of Transients	500
Pulse Sequence	s2pul	Original Points Count	23500
Spectrum Offset (Hz)	7922.1348	Solvent	CHLOROFORM-d
		Temperature (degree C)	AMBIENT TEMPERATURE
		Points Count	32768
		Sweep Width (Hz)	16115.94
		Receiver Gain	20.00
		Spectrum Type	STANDARD



This report was created by ACD/NMR Processor Academic Edition. For more information go to [www.acdlabs.com/nmrproc/](http://www.acdlabs.com/nmrproc/)

2013/12/2 22:39:52

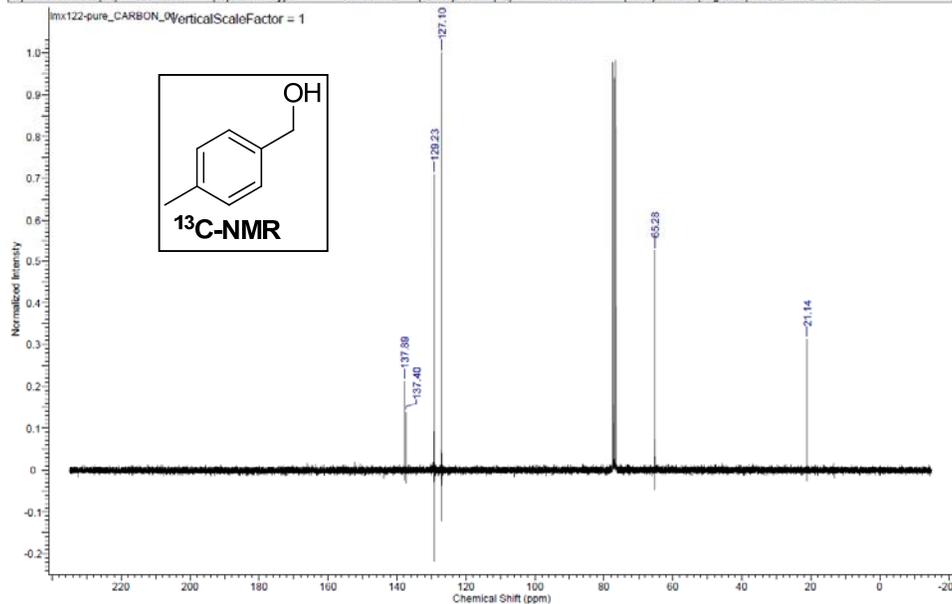
Acquisition Time (sec)	2.0480	Date	Dec. 2. 2013.	Date Stamp	Dec. 2. 2013.
File Name	C:\Users\MML\W\Desktop\NMR\lms122-pure_PROTON_01.fid.tif	Frequency (MHz)	299.83		
Nucleus	<sup>1</sup> H	Number of Transients	8	Original Points Count	9818
Pulse Sequence	s2pul	Receiver Gain	30.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	1797.7676	Spectrum Type	STANDARD	Sweep Width (Hz)	4793.86
				Temperature (degree C)	AMBIENT TEMPERATURE



This report was created by ACD/NMR Processor Academic Edition. For more information go to [www.acdlabs.com/nmrproc/](http://www.acdlabs.com/nmrproc/)

2013/12/2 22:40:46

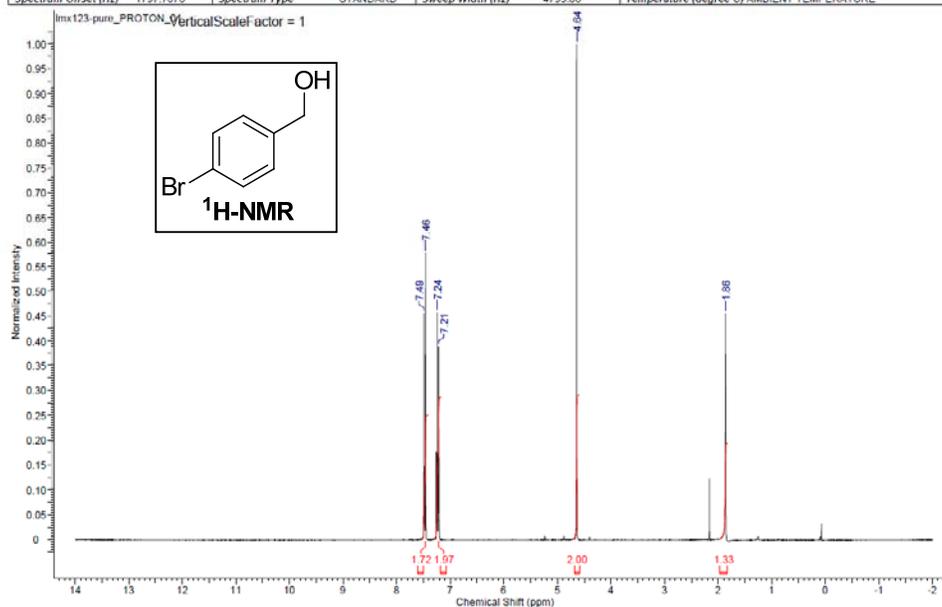
Acquisition Time (sec)	1.0420	Date	Dec. 2. 2013.	Date Stamp	Dec. 2. 2013.
File Name	C:\Users\MML\W\Desktop\NMR\lms122-pure_CARBOON_01.fid.tif	Frequency (MHz)	75.35		
Nucleus	<sup>13</sup> C	Number of Transients	3300	Original Points Count	19624
Pulse Sequence	s2pul	Receiver Gain	30.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	8287.5557	Spectrum Type	STANDARD	Sweep Width (Hz)	18832.36
				Temperature (degree C)	AMBIENT TEMPERATURE



This report was created by ACD/NMR Processor Academic Edition. For more information go to [www.acdlabs.com/nmrproc/](http://www.acdlabs.com/nmrproc/)

2013/12/2 22:41:13

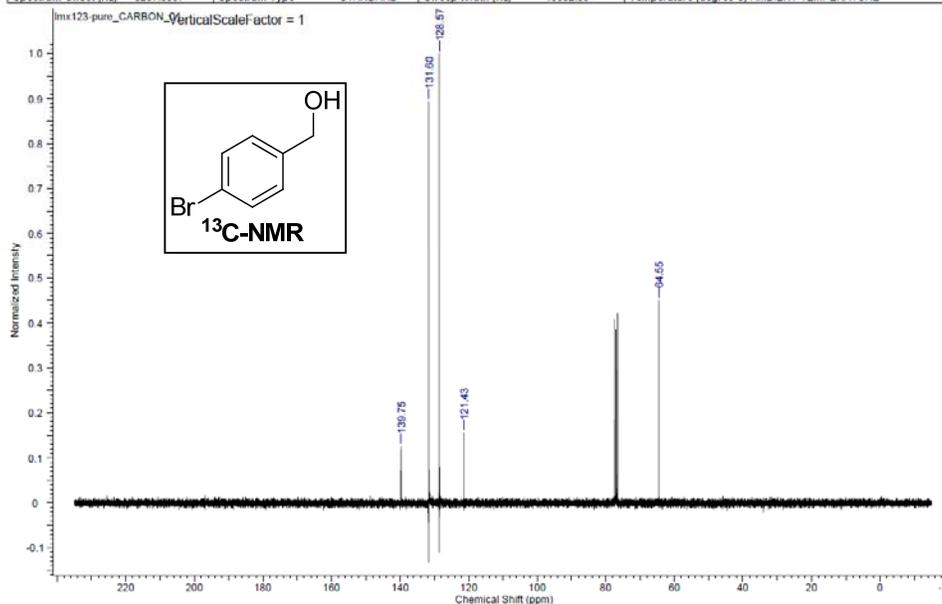
Acquisition Time (sec)	2.0460	Date	Dec. 2 2013	Date Stamp	Dec. 2 2013
File Name	C:\Users\MML\W\Desktop\NMR\lms123_pure_PROTON_01.fid.fid	Frequency (MHz)	299.63		
Nucleus	<sup>1</sup> H	Number of Transients	8	Original Points Count	9818
Pulse Sequence	s2pul	Receiver Gain	30.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	1797.7676	Spectrum Type	STANDARD	Sweep Width (Hz)	4793.86
				Temperature (degree C)	AMBIENT TEMPERATURE



This report was created by ACD/NMR Processor Academic Edition. For more information go to [www.acdlabs.com/nmrproc/](http://www.acdlabs.com/nmrproc/)

2013/12/2 22:41:36

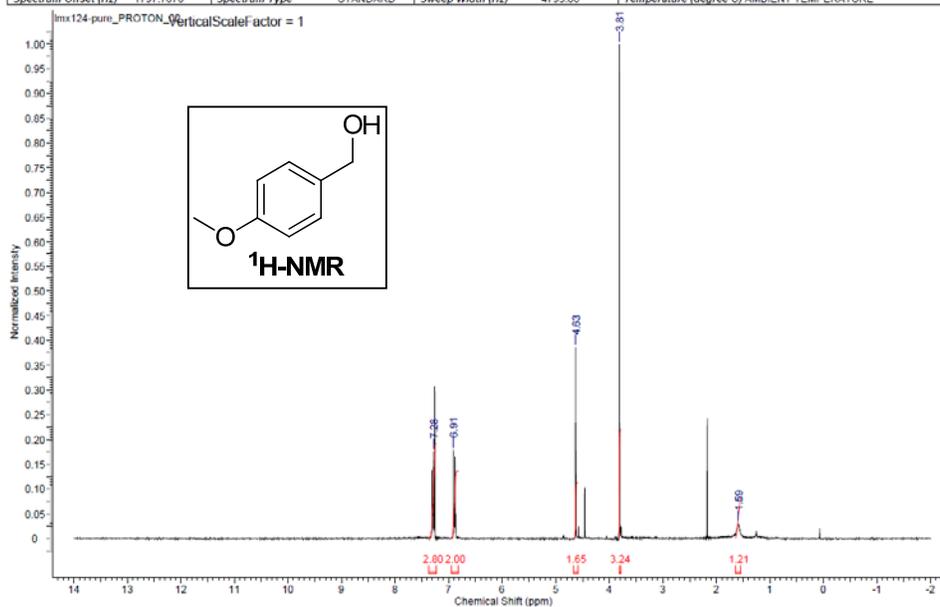
Acquisition Time (sec)	1.0420	Date	Dec. 2 2013	Date Stamp	Dec. 2 2013
File Name	C:\Users\MML\W\Desktop\NMR\lms123_pure_CARRON_01.fid.fid	Frequency (MHz)	75.35		
Nucleus	<sup>13</sup> C	Number of Transients	832	Original Points Count	19624
Pulse Sequence	s2pul	Receiver Gain	30.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	8287.5557	Spectrum Type	STANDARD	Sweep Width (Hz)	16832.39
				Temperature (degree C)	AMBIENT TEMPERATURE



This report was created by ACD/NMR Processor Academic Edition. For more information go to [www.acdlabs.com/nmrproc/](http://www.acdlabs.com/nmrproc/)

2013/12/2 22:42:36

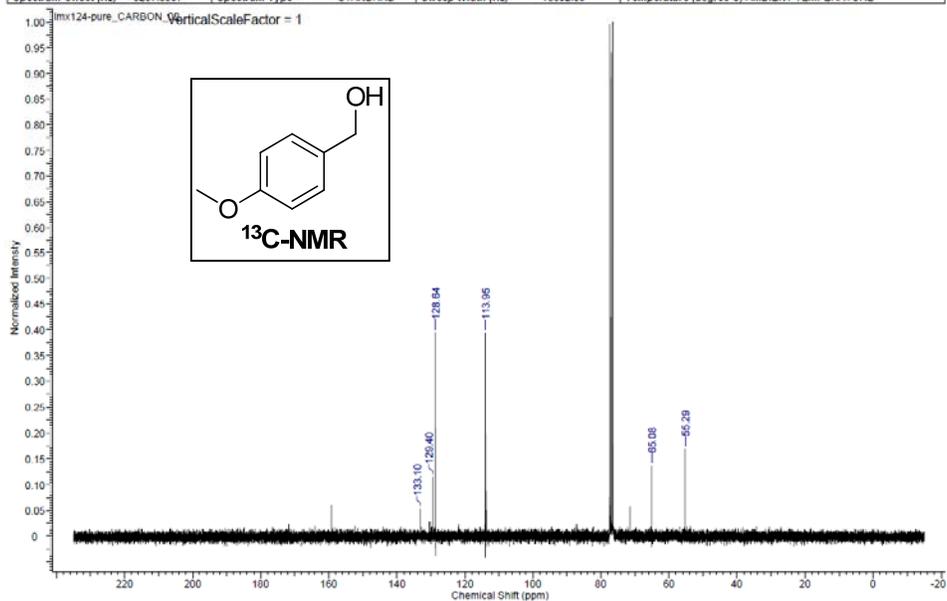
Acquisition Time (sec)	2.0460	Date	Dec. 2. 2013	Date Stamp	Dec. 2. 2013		
File Name	C:\Users\MMI\W\\Desktop\NMR\lms124_pure_PROTON_02.fid.fid	Frequency (MHz)	299.63				
Nucleus	<sup>1</sup> H	Number of Transients	8	Original Points Count	9818	Points Count	16384
Pulse Sequence	s2pul	Receiver Gain	39.00	Solvent	CHLOROFORM-d		
Spectrum Offset (Hz)	1797.7676	Spectrum Type	STANDARD	Sweep Width (Hz)	4793.86	Temperature (degree C)	AMBIENT TEMPERATURE



This report was created by ACD/NMR Processor Academic Edition. For more information go to [www.acdlabs.com/nmrproc/](http://www.acdlabs.com/nmrproc/)

2013/12/22 22:43:10

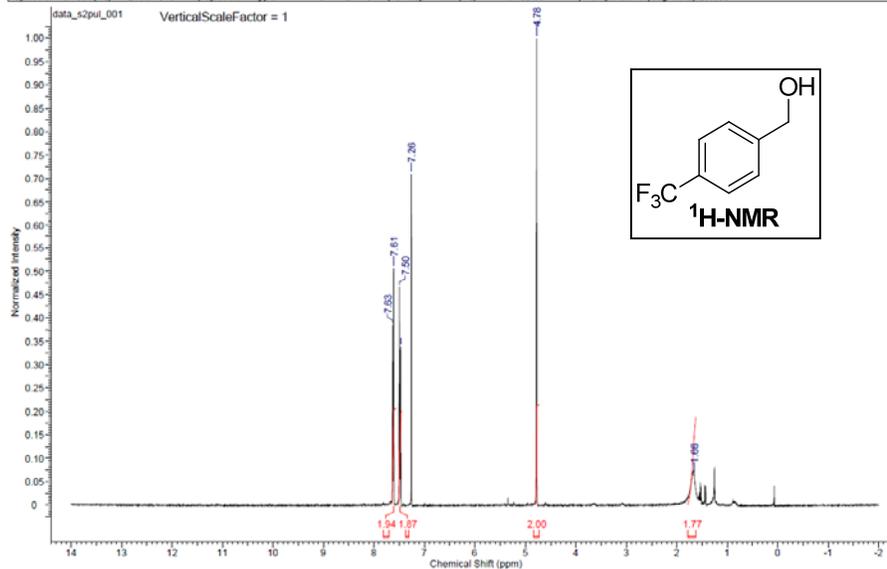
Acquisition Time (sec)	1.0420	Date	Dec. 2 2013	Date Stamp	Dec. 2 2013
File Name	C:\Users\MML\W\Desktop\NMR\lmx124-pure_CARON_02.fid.fid	Frequency (MHz)	75.35		
Nucleus	<sup>13</sup> C	Number of Transients	3300	Original Points Count	19624
Pulse Sequence	s2pul	Receiver Gain	30.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	8287.5557	Spectrum Type	STANDARD	Sweep Width (Hz)	18832.39
				Temperature (degree C)	AMBIENT TEMPERATURE



This report was created by ACD/NMR Processor Academic Edition. For more information go to [www.acdlabs.com/nmrproc/](http://www.acdlabs.com/nmrproc/)

2013/12/3 11:45:33

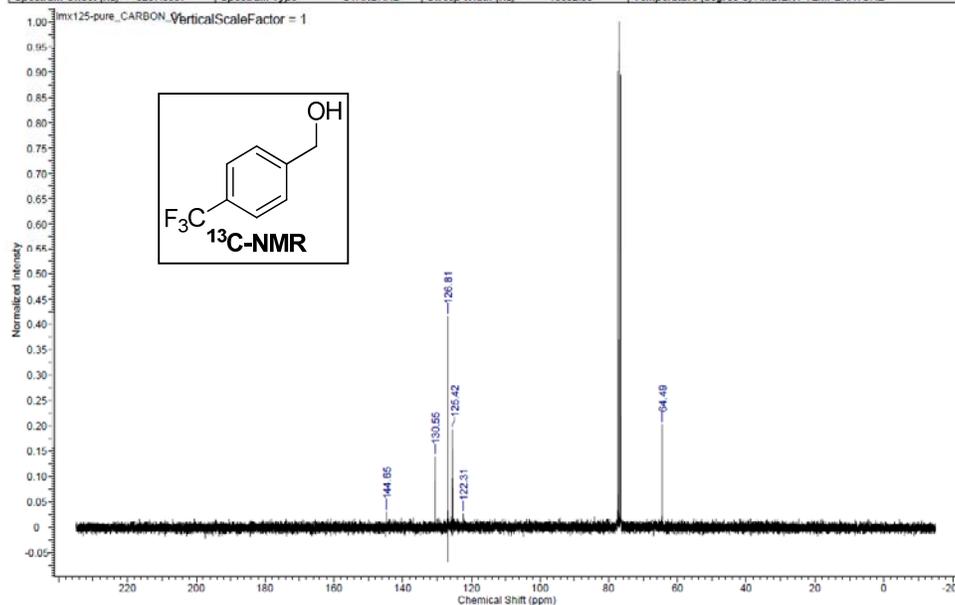
Acquisition Time (sec)	2.5592	Comment	lmx125-new	Date	Dec. 3 2013	Date Stamp	Dec. 3 2013
File Name	C:\Users\MML\W\Desktop\NMR\lmx125-new_20131203_01\data_s2pul_001.fid.fid	Frequency (MHz)	400.12				
Nucleus	<sup>1</sup> H	Number of Transients	8	Original Points Count	16384	Points Count	16384
Pulse Sequence	s2pul	Receiver Gain	24.00	Solvent	CHLOROFORM-d		
Spectrum Offset (Hz)	2400.7075	Spectrum Type	STANDARD	Sweep Width (Hz)	6402.05	Temperature (degree C)	23.000



This report was created by ACD/NMR Processor Academic Edition. For more information go to [www.acdlabs.com/nmrproc/](http://www.acdlabs.com/nmrproc/)

2013/12/3 11:45:57

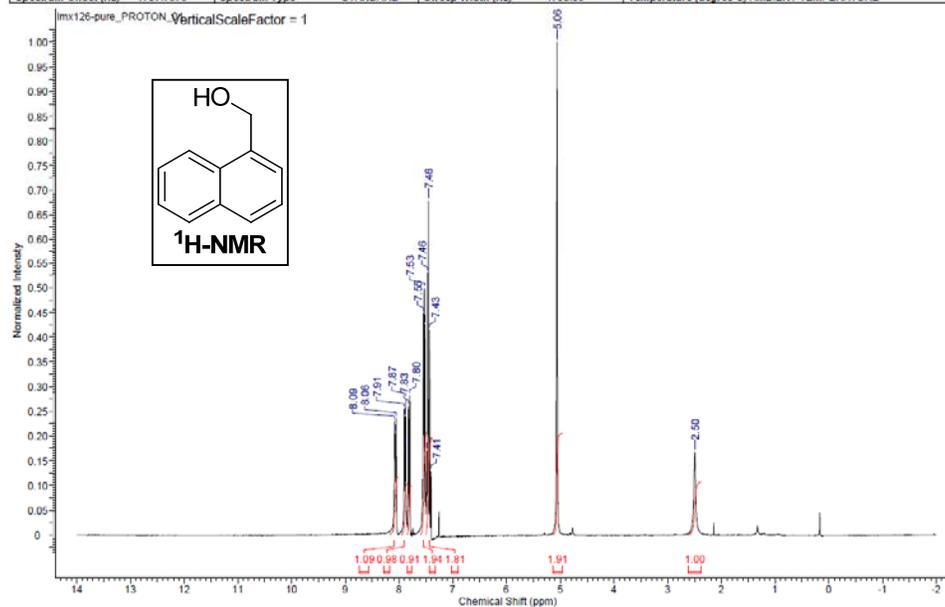
Acquisition Time (sec)	1.0420	Date	Dec. 3 2013	Date Stamp	Dec. 3 2013
File Name	C:\Users\MML\W\Desktop\NMR\imx125-pure_CARBO	Number of Transients	3300	Original Points Count	19624
Nucleus	<sup>13</sup> C	Receiver Gain	30.00	Solvent	CHLOROFORM-d
Pulse Sequence	s2pul	Spectrum Type	STANDARD	Sweep Width (Hz)	18832.39
Spectrum Offset (Hz)	8287.5557			Temperature (degree C)	AMBIENT TEMPERATURE
				Frequency (MHz)	75.35
				Points Count	32768



This report was created by ACD/NMR Processor Academic Edition. For more information go to [www.acdlabs.com/nmrproc/](http://www.acdlabs.com/nmrproc/)

2013/12/2 22:43:49

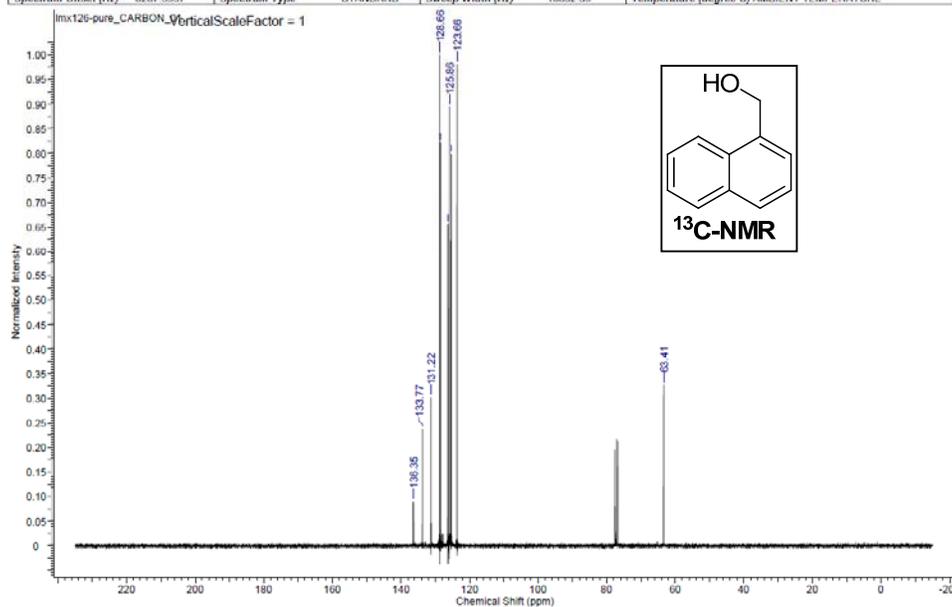
Acquisition Time (sec)	2.0480	Date	Dec. 2 2013	Date Stamp	Dec. 2 2013
File Name	C:\Users\MML\W\Desktop\NMR\imx125-pure_PROTON	Number of Transients	8	Original Points Count	9818
Nucleus	<sup>1</sup> H	Receiver Gain	16.00	Solvent	CHLOROFORM-d
Pulse Sequence	s2pul	Spectrum Type	STANDARD	Sweep Width (Hz)	4793.86
Spectrum Offset (Hz)	1797.7676			Temperature (degree C)	AMBIENT TEMPERATURE
				Frequency (MHz)	299.63
				Points Count	16384



This report was created by ACD/NMR Processor Academic Edition. For more information go to [www.acdlabs.com/nmrproc/](http://www.acdlabs.com/nmrproc/)

2013/12/2 22:44:25

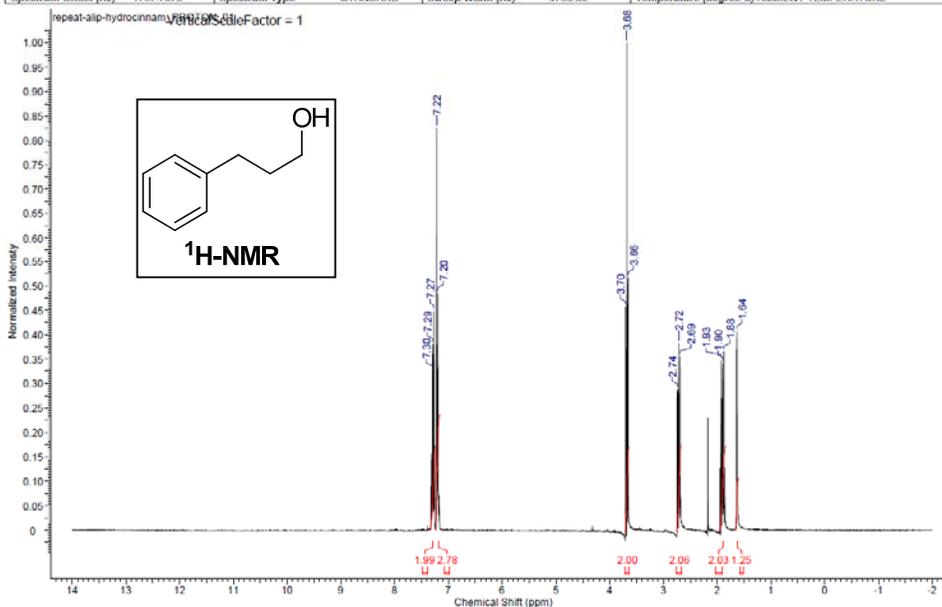
Acquisition Time (sec)	1.0420	Date	Dec. 2 2013	Date Stamp	Dec. 2 2013	Frequency (MHz)	75.35
File Name	C:\Users\MML\W\Desktop\NMR\lms126-pure_CARBO..._01.fid	Number of Transients	2112	Original Points Count	19624	Points Count	32768
Nucleus	<sup>13</sup> C	Receiver Gain	30.00	Solvent	CHLOROFORM-d	Spectrum Type	STANDARD
Pulse Sequence	s2pul	Sweep Width (Hz)	18832.39	Temperature (degree C)	AMBIENT TEMPERATURE		



This report was created by ACD/NMR Processor Academic Edition. For more information go to [www.acdlabs.com/nmrproc/](http://www.acdlabs.com/nmrproc/)

2013/12/2 22:45:55

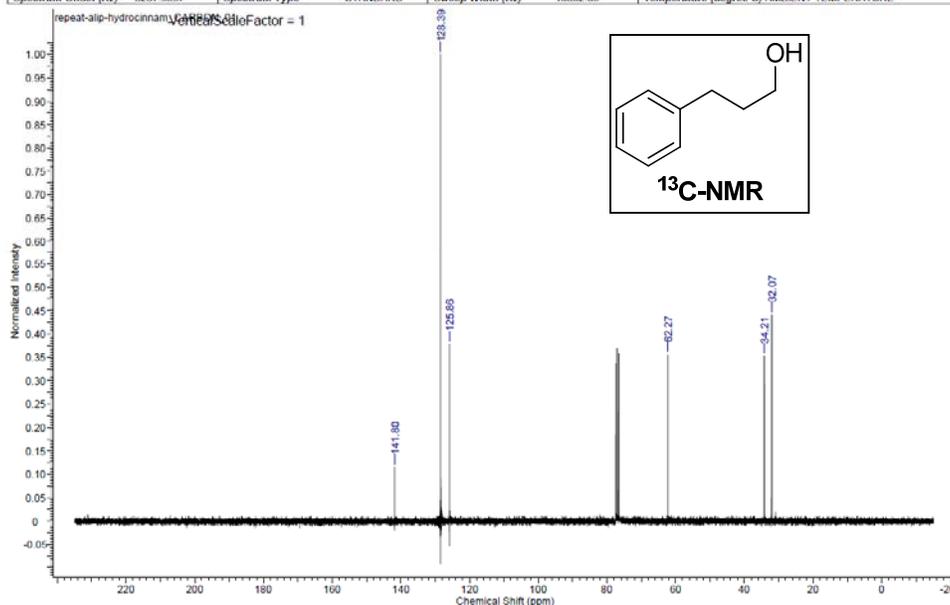
Acquisition Time (sec)	2.0480	Date	Dec. 2 2013	Date Stamp	Dec. 2 2013	Frequency (MHz)	299.63
File Name	C:\Users\MML\W\Desktop\NMR\repeat-alip-hydrocinna..._01.fid	Number of Transients	8	Original Points Count	9818	Points Count	16384
Nucleus	<sup>1</sup> H	Receiver Gain	25.00	Solvent	CHLOROFORM-d	Spectrum Type	STANDARD
Pulse Sequence	s2pul	Sweep Width (Hz)	4793.86	Temperature (degree C)	AMBIENT TEMPERATURE		



This report was created by ACD/NMR Processor Academic Edition. For more information go to [www.acdlabs.com/nmrproc/](http://www.acdlabs.com/nmrproc/)

2013/12/2 22:46:50

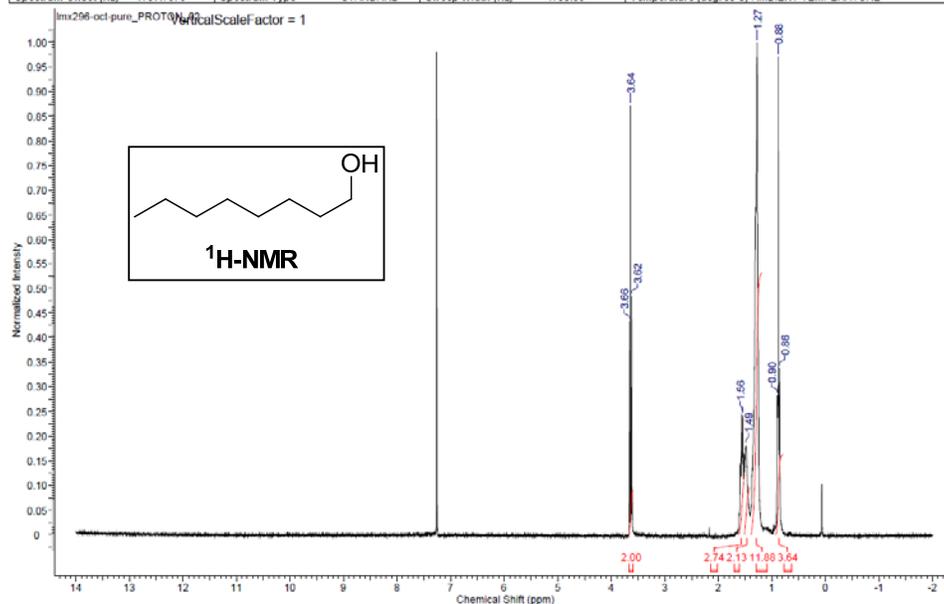
Acquisition Time (sec)	1.0420	Date	Dec 2 2013	Date Stamp	Dec 2 2013	Frequency (MHz)	75.35
File Name	C:\Users\MML\W\Desktop\NMR\repeat-alp-hydrocinna_CARBON_01.fidfid	Number of Transients	1024	Original Points Count	19624	Points Count	32768
Nucleus	<sup>13</sup> C	Receiver Gain	30.00	Solvent	CHLOROFORM-d	Spectrum Type	STANDARD
Pulse Sequence	s2pul	Spectrum Offset (Hz)	8287.5557	Sweep Width (Hz)	18832.39	Temperature (degree C)	AMBIENT TEMPERATURE



This report was created by ACD/NMR Processor Academic Edition. For more information go to [www.acdlabs.com/nmrproc/](http://www.acdlabs.com/nmrproc/)

2013/12/2 22:47:41

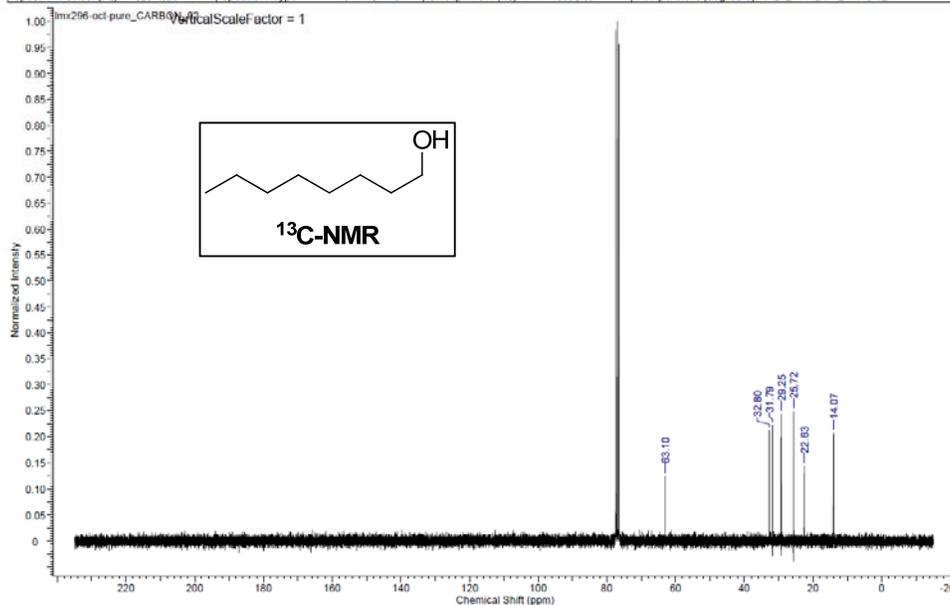
Acquisition Time (sec)	2.0480	Date	Aug 19 2013	Date Stamp	Aug 19 2013	Frequency (MHz)	299.63
File Name	C:\Users\MML\W\Desktop\NMR\lms296-oct-pure_PROTON_02.fidfid	Number of Transients	8	Original Points Count	9818	Points Count	16384
Nucleus	<sup>1</sup> H	Receiver Gain	35.00	Solvent	CHLOROFORM-d	Spectrum Type	STANDARD
Pulse Sequence	s2pul	Spectrum Offset (Hz)	1797.7675	Sweep Width (Hz)	4793.65	Temperature (degree C)	AMBIENT TEMPERATURE



This report was created by ACD/NMR Processor Academic Edition. For more information go to [www.acdlabs.com/nmrproc/](http://www.acdlabs.com/nmrproc/)

2013/12/2 22:49:14

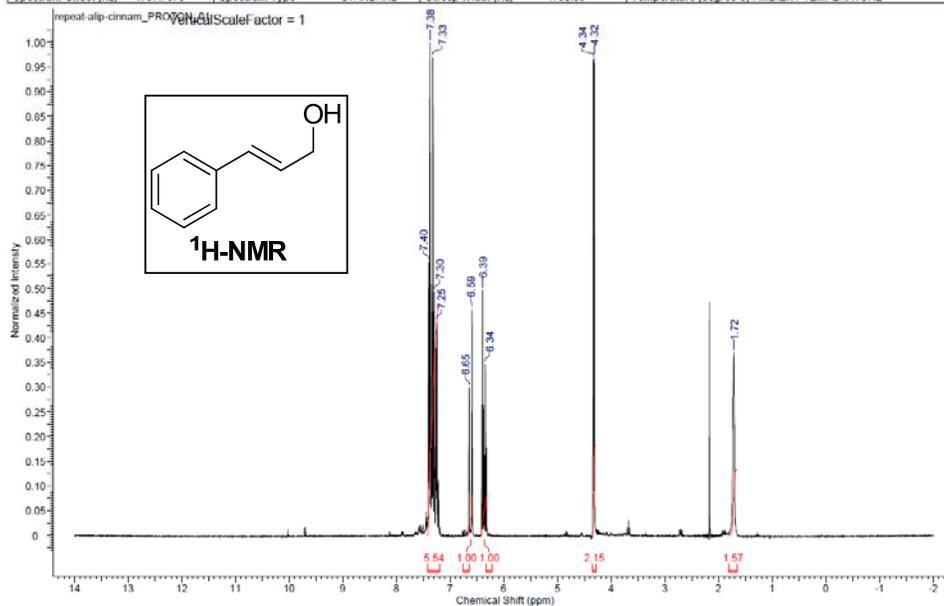
Acquisition Time (sec)	1.0420	Date	Aug 19 2013	Date Stamp	Aug 19 2013
File Name	C:\Users\MML\W\Desktop\NMR\imv296-oct-pure_CARBO	Frequency (MHz)	75.35	Points Count	32768
Nucleus	<sup>13</sup> C	Number of Transients	3300	Original Points Count	19624
Pulse Sequence	s2pul	Receiver Gain	30.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	8287.5557	Spectrum Type	STANDARD	Sweep Width (Hz)	18832.39
				Temperature (degree C)	AMBIENT TEMPERATURE



This report was created by ACD/NMR Processor Academic Edition. For more information go to [www.acdlabs.com/nmrproc/](http://www.acdlabs.com/nmrproc/)

2013/12/2 22:49:56

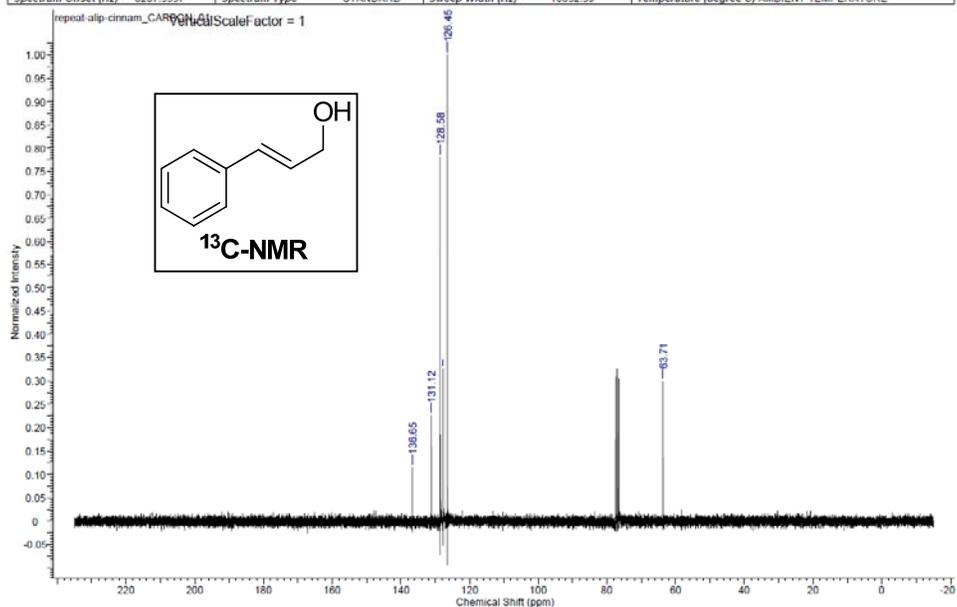
Acquisition Time (sec)	2.0480	Date	Dec 2 2013	Date Stamp	Dec 2 2013
File Name	C:\Users\MML\W\Desktop\NMR\repeat-alip-cinnam_PROTON_01.fid	Frequency (MHz)	290.63	Points Count	16384
Nucleus	<sup>1</sup> H	Number of Transients	8	Original Points Count	9818
Pulse Sequence	s2pul	Receiver Gain	30.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	1797.7675	Spectrum Type	STANDARD	Sweep Width (Hz)	4793.85
				Temperature (degree C)	AMBIENT TEMPERATURE



This report was created by ACD/NMR Processor Academic Edition. For more information go to [www.acdlabs.com/nmrprocl](http://www.acdlabs.com/nmrprocl)

2013/12/2 22:50:19

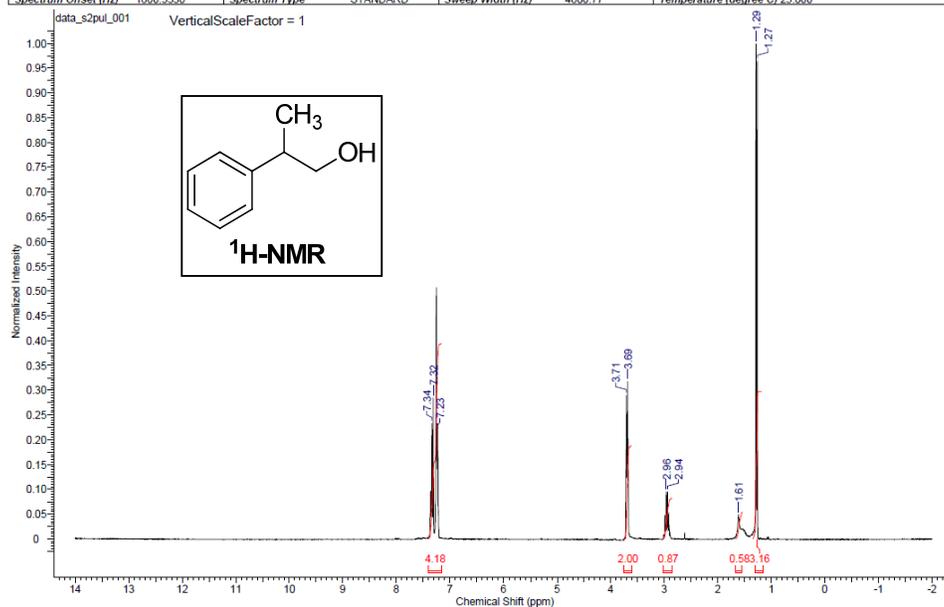
Acquisition Time (sec)	1.0420	Date	Dec 2 2013	Date Stamp	Dec 2 2013
File Name	C:\Users\MML\W\Desktop\NMR\repeat-alip-cinnam_CARBON_01.fid.tif	Frequency (MHz)	75.35		
Nucleus	<sup>13</sup> C	Number of Transients	320	Original Points Count	19624
Pulse Sequence	s2pul	Receiver Gain	30.00	Points Count	32768
Spectrum Offset (Hz)	8287.5557	Spectrum Type	STANDARD	Solvent	CHLOROFORM-d
				Sweep Width (Hz)	18832.39
				Temperature (degree C)	AMBIENT TEMPERATURE



This report was created by ACD/NMR Processor Academic Edition. For more information go to [www.acdlabs.com/nmrprocl](http://www.acdlabs.com/nmrprocl)

2014/1/6 18:32:51

Acquisition Time (sec)	2.0480	Comment	add-alip-phen/propionaldehyde-final	Date	Jan 3 2014
Date Stamp	Jan 3 2014	File Name	C:\Users\MML\W\Desktop\add-alip-phen/propionaldehyde-final	20140103_01\data_s2pul_001.fid.tif	
Frequency (MHz)	300.06	Nucleus	<sup>1</sup> H	Number of Transients	8
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	24.00
Spectrum Offset (Hz)	1800.3330	Spectrum Type	STANDARD	Solvent	CHLOROFORM-d
				Sweep Width (Hz)	4800.77
				Temperature (degree C)	25.000



This report was created by ACD/NMR Processor Academic Edition. For more information go to [www.acdlabs.com/nmrpro/](http://www.acdlabs.com/nmrpro/)

2014/1/16 18:27:17

Acquisition Time (sec)	1.0420	Comment	add-alip-phenylpropionaldehyde-final	Date	Jan. 3.2014
Date Stamp	Jan. 3.2014				
File Name	C:\Users\MML\W\Desktop\add-alip-phenylpropionaldehyde-final_20140103_02\add-alip-phenylpropionaldehyde-final_CARBON_2_20140103_02.fid.tif				
Frequency (MHz)	75.46	Nucleus	<sup>13</sup> C	Number of Transients	1651
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00
Spectrum Offset (Hz)	8299.3770	Spectrum Type	STANDARD	Sweep Width (Hz)	18867.92
				Solvent	CHLOROFORM-d
				Temperature (degree C)	25.000

