

## Supporting Information

### A turn-on fluorescent probe for detection of hydrogen sulfide in aqueous solution and living cells

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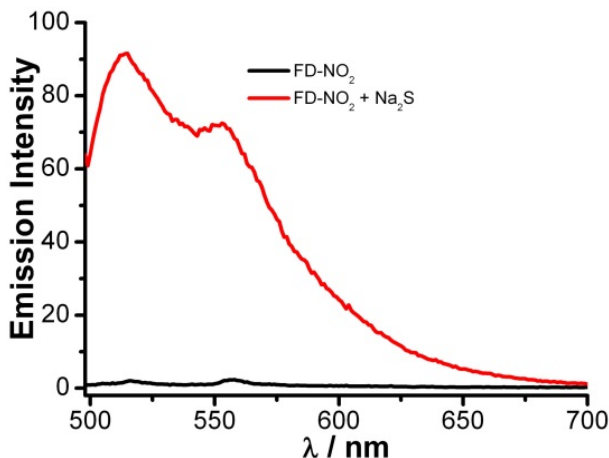
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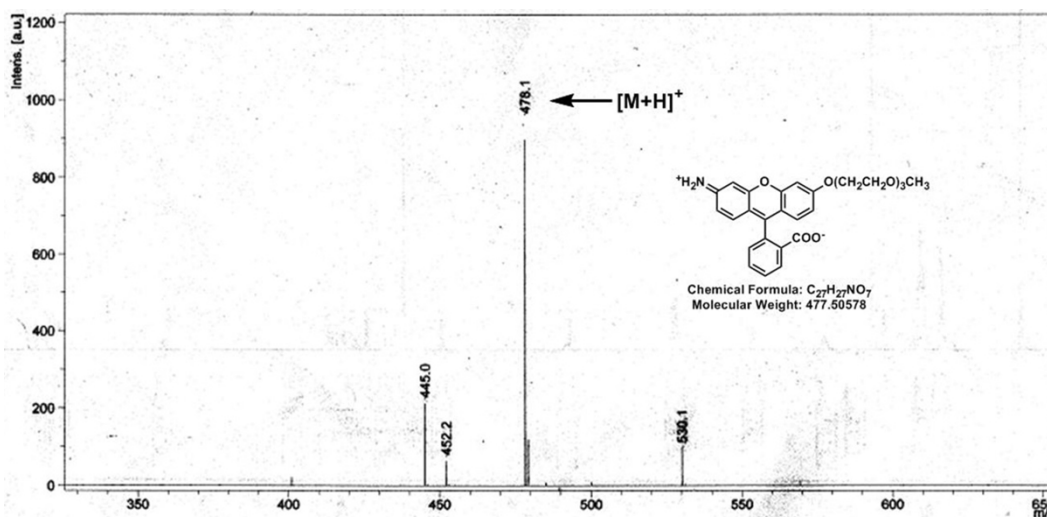
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## 1. Fluorometric and MALDI-TOF Analysis

UV-visible spectra were recorded on SHIMADZU UV-2550 UV-vis spectrometer. Fluorescence spectra were recorded using a HITACHI F-4600 spectrometer. The PMT voltage was 700 V, excitation slit and emission slit were 5 nm. The path length was 1 cm with cell volume of 3.0 mL. The stock solution of FD-NO<sub>2</sub> was prepared in DMSO (2 mM). H<sub>2</sub>O<sub>2</sub>, *tert*-butylhydroperoxide (TBHP), and hypochlorite (NaOCl) were delivered from 30%, 70%, and 14.5% aqueous solutions, respectively. Nitric oxide (NO) was generated from SNP.<sup>S1</sup> H<sub>2</sub>S, N<sub>3</sub><sup>-</sup>, AcO<sup>-</sup>, SCN<sup>-</sup>, SO<sub>3</sub><sup>2-</sup>, S<sub>2</sub>O<sub>3</sub><sup>2-</sup>, O<sub>2</sub><sup>-</sup>, NO<sub>2</sub><sup>-</sup>, and NO<sub>3</sub><sup>-</sup> were prepared from Na<sub>2</sub>S,<sup>S2</sup> NaN<sub>3</sub>, NaOAc, KSCN, Na<sub>2</sub>SO<sub>3</sub>, Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub>, KO<sub>2</sub>,<sup>S3</sup> NaNO<sub>2</sub>, and KNO<sub>3</sub>, respectively.



**Fig. S1** Fluorescence spectra of FD-NO<sub>2</sub> (2 μM) and their reaction solutions with excess Na<sub>2</sub>S (100 μM) in MeOH, 25 °C.

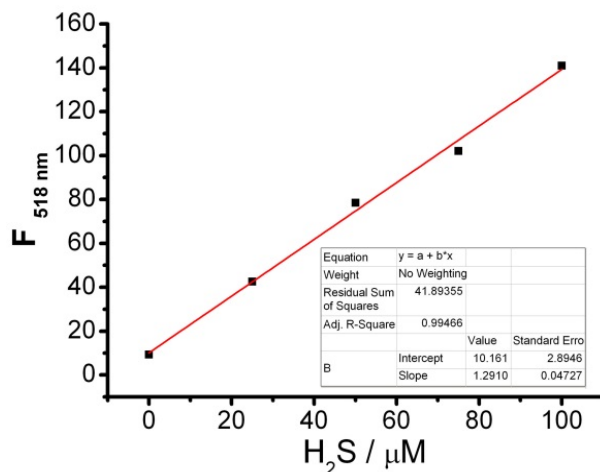


**Fig. S2** MALDI-TOF spectra of FD-NO<sub>2</sub> after addition of 100 μM Na<sub>2</sub>S in MeOH.

## 2. Determination of the detection limit (LOD)

The detection limit was calculated according to the method described in reference.<sup>S4</sup> The emission spectrum of 2 μM FD-NO<sub>2</sub> in PBS buffer was collected for 30 times to determine the background noise  $\sigma$ . A linear regression curve was fitted according to the emission intensity at 518 nm as a function of the concentrations of H<sub>2</sub>S in the range of 0 -

100  $\mu\text{M}$ , and the slope of the curve was obtained. The detection limit ( $3\sigma/\text{slope}$ ) was then determined to be 1  $\mu\text{M}$ .



**Fig. S3** Fluorescence intensities of FD-NO<sub>2</sub> at 518 nm as a function of the concentrations of H<sub>2</sub>S in the range of 0 - 100  $\mu\text{M}$  upon excitation at 480 nm and the calculation of the detection limit of probe FD-NO<sub>2</sub> for H<sub>2</sub>S.

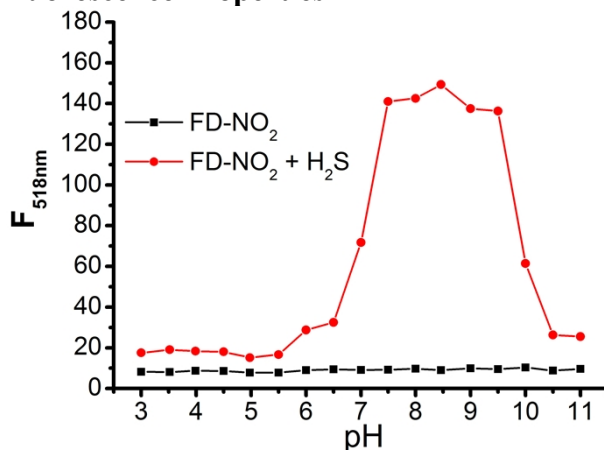
### 3. Determination of quantum yields

Fluorescence quantum yield was determined in the reference of fluorescein ( $\Phi = 0.85$ ) in 0.1 M aqueous NaOH.<sup>S5</sup> The quantum yield of FA is calculated according to following equation.

$$\Phi_x = \Phi_s (A_s S_x) / (A_x S_s)$$

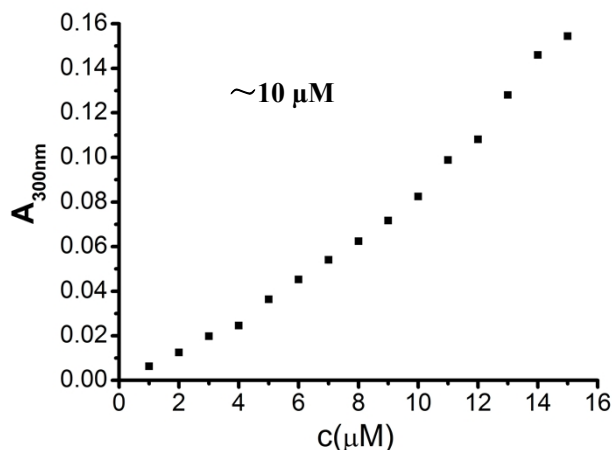
$\Phi_s$  is the fluorescence quantum yield of fluorescein,  $A_x$  and  $A_s$  are the absorbance of FA and the standard.  $S_x$  and  $S_s$  are integrated fluorescence emission corresponding to FA and the standard.

### 4. pH Effect on the Fluorescence Properties

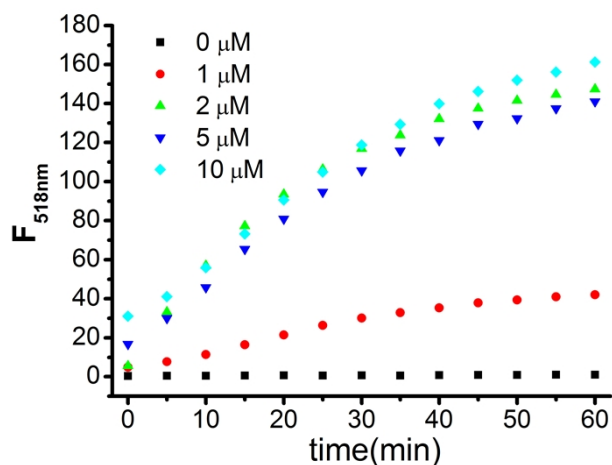


**Fig. S4** Effect of pH on fluorescence intensities at 518 nm for FD-NO<sub>2</sub> (2  $\mu\text{M}$ ) and FD-NO<sub>2</sub> (2  $\mu\text{M}$ ) to H<sub>2</sub>S (100  $\mu\text{M}$ ) for 50 min in PBS buffer upon excitation at 480 nm.

### 5. Solubility



**Fig. S5** Plot of absorbance at 300 nm as a function of FD-NO<sub>2</sub> concentration in PBS buffer (pH 7.4).



**Fig. S6** Plot of fluorescence intensity at 518 nm vs. the reaction time in the presence of varied concentrations of FD-NO<sub>2</sub>: 0 (control), 1, 2, 5 and 10  $\mu\text{M}$ . The measurements were performed at 25 °C in PBS buffer (pH 7.4) with H<sub>2</sub>S (100  $\mu\text{M}$ ) upon excitation at 480 nm.

## 6. Confocal Fluorescence Microscopy

Confocal fluorescence imaging experiments were performed on an Olympus FV-1000 laser scanning microscopy system, based on an IX81 (Olympus, Japan) inverted microscope. The microscope was equipped with multiple visible laser lines (405, 458, 488, 515, 543, 635 nm, CW) and UPLSAPO 60 $\times$ /N.A 1.42 objective.

## 7. Cell Culture Methods

NIH 3T3 cells were obtained from Cell Culture Center, Institute of Basic Medical Sciences Chinese Academy of Medical Sciences; School of Basic Medicine Peking Union Medical College.

NIH 3T3 cells were cultured in Dulbecco's modified Eagle's medium (DMEM, Gibco) supplemented with 10% fetal calf serum (FCS, Gibco), 50  $\mu\text{g}/\text{mL}$  penicillin/streptomycin (Hyclone) at 37°C in a 5/95 CO<sub>2</sub>/air incubator. The cells were cultured 3 days before dye loading on a 35 mm diameter glass-bottomed coverslips. Then the cells were incubated with 10 $\mu\text{M}$  probe FD-NO<sub>2</sub> in serum-free DMEM (0.5 % DMSO, v/v) for 30 min at 37°C under 5% CO<sub>2</sub>, and then washed with PBS three times. The FD-NO<sub>2</sub>-loaded cells were

incubated in serum-free DMEM containing H<sub>2</sub>S (250 μM) for another 30 min at 37 °C, the cells were then washed with PBS three times and bathed in PBS (1 mL) before imaging. As a control, the FD-NO<sub>2</sub>-loaded cells incubated in serum-free DMEM without H<sub>2</sub>S for 30 min at 37 °C were also imaged. To further prove that the fluorescence change of FD-NO<sub>2</sub> in the cells arises from H<sub>2</sub>S, another control experiment was carried out. The cells were pretreated with 500 μM ZnCl<sub>2</sub> (an efficient H<sub>2</sub>S scavenger)<sup>S2</sup> for 30 min, then incubated with 10 μM FD-NO<sub>2</sub> for 30 min, and further treated with 250 μM H<sub>2</sub>S for 30 min at 37 °C. Excitation wavelength of laser was 488 nm. Emissions were collected at 500 - 600 nm. Images were gathered and processed with Olympus FV10-ASW software (Ver. 2.1c)

### Reference

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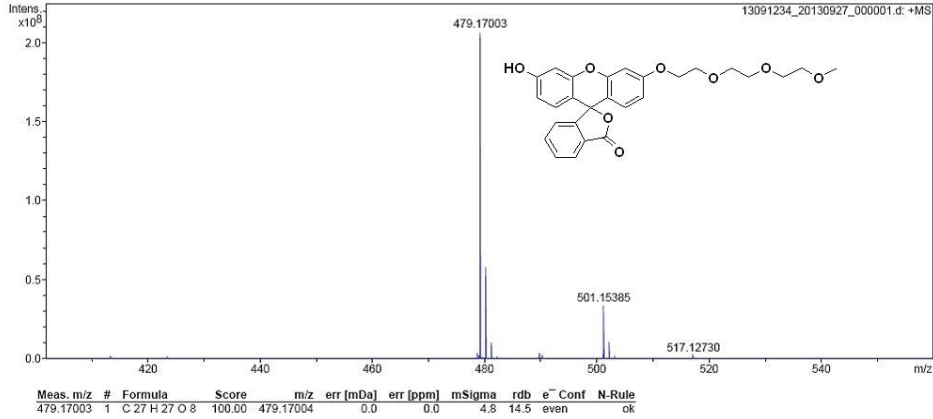
## 8. HRMS-ESI Spectra

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 Comment ESI Positive

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 Operator Peking University

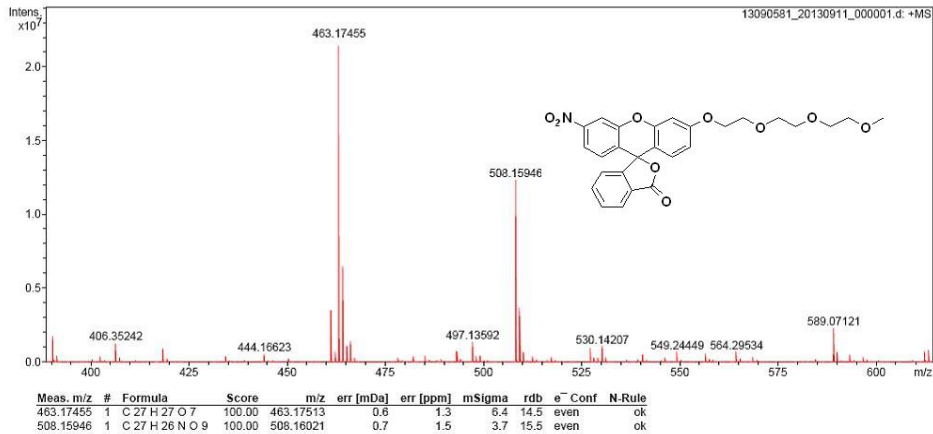


### Peking University Mass Spectrometry Sample Analysis Report

#### Analysis Info

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Acquisition Date 9/11/2013 4:53:40 PM  
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 Operator Peking University



## 9. NMR Data

