

Supporting Information

Glycosylation of *N*-Acetyl Glycosamine Using Catalytic Iron(III) Triflate: from a Microwave Batch Chemistry to a Scalable Continuous-Flow Process

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Experimental procedures for compounds **1β**, **1α**, **4**, **6β**, **8**, **10**, **12-24**, **29**, **30**, **32**, **34-49** and **55**
Characterization data and NMR spectra (¹H, ¹³C) of compounds **8**, **10**, **13**, **16-18**, **21-24**, **34-38** and **40**.

General considerations

Commercial chemicals were obtained from Aldrich, Acros Organics, Alfa Aesar or Carbosynth and were used without further purification. All non-aqueous reactions were run under inert atmosphere (argon), by using standard techniques for manipulating air-sensitive compounds. All the glassware was stored into the oven or was flame-dried before using. Anhydrous solvents were obtained by filtration through drying columns. Dichloromethane and chloroform were stabilized under amylene.

Batch reactions were generally monitored by analytical thin-layer chromatography performed on silica gel 60 F₂₅₄ precoated plates and were visualised under UV (254 nm) and with Vanillin as revelator.

Microwave irradiation experiments were carried out in a CEM Discover instrument or in an Anton Paar Monowave 300 instrument with internal fiber-optic or IR temperature control. The vials used are in Pyrex and were sealed with Teflon-coated septums. With the Anton Paar instrument, the homogeneity and the good magnetic stirring of the reaction were controlled with a camera which directly focuses on the reaction vial.

Flow reactions were performed in a Vapourtec R-series system which is a high-temperature (up to 250 °C), high-pressure (up to 42 bar) instrument for performing homogeneous reactions. This R-series system is constituted by a R2C+ pump module (module with acid-resistant pumps and injection loops), a R4 reactor module and a fraction collector. The reaction mixture was pumped into the system with one HPLC pump, passed through a stainless steel 1 mm i.d. reactor, through a back-pressure regulator (which controls the pressure into the whole system) and was finally collected in the fraction collector. Perfluoroalkoxy (PFA) polymer tubing (1 mm i.d.) was used for interconnecting lines. Reaction parameters like temperature, residence time (or flow rate) and collection volume were monitored by using the instrument interface and the Flow Commander software.

Flash chromatography was performed on an Isco Combiflash Companion instrument using 50 µm silica columns. Preparative thin-layer chromatography was performed on silica gel 60 F₂₅₄ 0.5 mm 20×20 cm plates and visualised under UV (254 nm). Semi-preparative HPLC was performed using a Waters 600 instrument, combined with a 2424 Evaporating Light Scattering Detector (ELSD), a 2996 Photodiode Array Detector (PDA) and a 2767 sample manager. The column used was a Waters Sunfire C18, 19×150 mm, 5 µm.

Deuterated chloroform used for NMR analyses was neutralised by addition of anhydrous and granular K₂CO₃. ¹H NMR spectra were recorded on Bruker 300 or 500 MHz instruments. ¹³C NMR spectra were recorded on the same instruments at 75 or 125 MHz. Chemical shifts δ are expressed in parts per million relative to residual chloroform as an internal standard (δ = 7.26 ppm for ¹H NMR and 77.4 ppm for ¹³C NMR). For ¹H NMR spectra, data are reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quadruplet, m = multiplet, brs = broad singlet, dt = doublet of triplets), coupling constant (in Hz) and integration. Interpretations were obtained using DEPT 135, ¹H-¹H COSY, ¹H-¹³C HMQC and HMBC experiments. Low-resolution mass spectra were obtained on a Waters Acquity UPLC system by electrospray ionization (ESI), combined to a Photodiode Array Detector (PDA), an Evaporating Light Scattering Detector (ELSD) and a Tandem Quadrupole Detector (TQD). High-resolution mass spectra were obtained with a Waters Acquity UPLC (by direct injection or with a BEH C18 2.1×50 mm, 1.7 µm column) combined with a Diode Array Detector (DAD) and a Waters LCT Premier XE mass instrument (electrospray ionization with a time-of-flight (ToF) analyzer). IR spectra were recorded on a PerkinElmer Spectrum 100 FT-IR spectrometer, in reciprocal centimeters (cm⁻¹). Melting points were determined using a Büchi

B-540 apparatus. Optical rotations were determined using an Anton Paar MCP 300 polarimeter with a 1 dm-long cell and data are reported as follows: $[\alpha]_D^{\text{temperature}}$ (in 10^{-1} deg.cm²/g), concentration (c in g/100 mL) and solvent. Elemental analyses were performed with a CHNOS Perkin-Elmer analyser (Gif-sur-Yvette, ICSN).

General procedures

A. General procedure for microwave-assisted glycosylation

The peracetylated glycosamine (2 eq.), TTBP (2 eq.) and the iron triflate catalyst (15 mol-%) are added to the acceptor (1 eq.) in an oven-dried, argon-purged, 4, 10 or 30 mL (filling volume) microwave vial equipped with a magnetic stirring bar. Everything is flushed under argon and dry CH₂Cl₂ is added ([acceptor] = 0.065 M). Liquid acceptor (benzyl alcohol) is placed into the tube after the solvent.

Anton Paar Monowave 300 instrument: after sealing the vial, the reaction mixture is heated to 110 °C under microwave irradiation for 30 minutes to 3 hours (1 minute ramp time from room temperature to 110 °C and 30 minutes to 3 hours hold time at 110 °C, stirring set at 800 rpm). CEM Discover instrument: after sealing the vial, the reaction mixture is heated to 80 °C under microwave irradiation for 30 minutes to 3 hours.

The reaction mixture is then diluted in CH₂Cl₂ and washed with a saturated aqueous solution of NaHCO₃. The aqueous layer is extracted with CH₂Cl₂ (×4) and the combined organic layers are washed with brine, dried over Na₂SO₄, filtered and concentrated under reduced pressure. The crude product is purified by flash chromatography on silica gel (heptane/EtOAc 5:5 to 0:1) to afford the pure product.

B. General procedure for setting up and cleaning up the flow system

Before each experiment, the desired assembly (including sample loop for small-scale reactions) is purged by pumping dichloromethane or chloroform (depending on the reaction solvent) at a flow rate of 5 mL/min, through both solvent and reagent needles for 10 min. Then, the needles are inserted through a septum-sealed, argon-overpressured flask (using the built-in gas manifold) of dry reaction solvent mixture (dichloromethane/acetonitrile 7:3 or chloroform/acetonitrile 7:3). The whole system is then dried by pumping this mixture at a flow rate of 5 mL/min for 10 minutes.

After each experiment, the whole system is washed with the reaction solvent mixture at a flow rate of 5 mL/min for 10 minutes and then with isopropanol with the same flow rate for another 10 minutes.

C. General procedure for glycosylation under continuous flow conditions

C1: small-scale reactions

As the glycosylation reaction is slow at room temperature, all the reagents and the catalyst are mixed together before injection into the flow system. The *N*-acetyl-D-glucosamine **1β** (0.300 mmol, 2 eq.) and the iron triflate catalyst (0.023 mmol, 15 mol-%) are added to the acceptor (0.150 mmol, 1 eq.) in an oven-dried, argon-purged vial equipped with a magnetic stirring bar. Dry mixture of dichloromethane/acetonitrile 7:3 (2 mL, [acceptor] = 0.075 M) is added and the reaction mixture is stirred and sonicated for a few minutes (until complete homogenisation). Liquid acceptor (benzyl alcohol) is placed into the tube after the solvent.

The variation between the solvent volume and the reaction mixture volume was controlled and was not significant.

After setting up and drying the whole flow system (Figure 1) with dry dichloromethane/acetonitrile 7:3 (procedure B), the pump is primed and the reaction mixture is injected with a syringe into the system *via* a 2 mL-sample loop. The reaction is then fully automated and the reaction parameters (temperature, residence time, collection volume) are controlled using Flow Commander software. The dry solvent mixture (dichloromethane/acetonitrile 7:3) is pumped and pushes the reaction mixture, which is in the sample loop, into the 10mL-stainless steel reactor heated at 110 °C with a fixed flow rate (corresponding to the desired residence time, 45 or 70 min depending on the acceptor). The system pressure (33 bar) is controlled with a back pressure regulator and the reaction mixture is finally collected into a fraction collector.

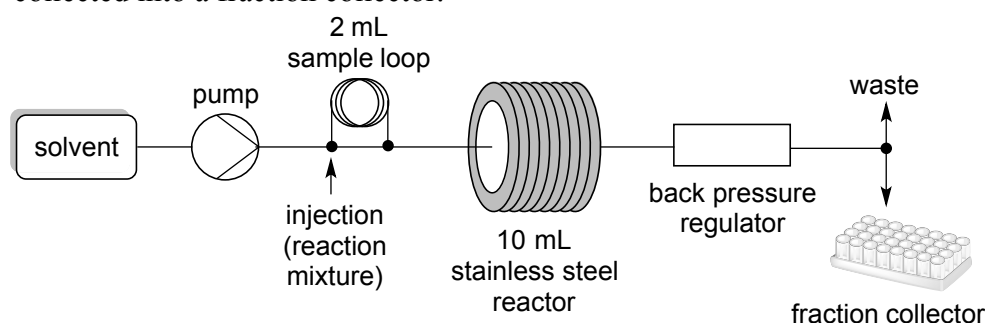


Figure 1

At the end of the reaction, the reaction mixture is diluted with 20 mL of dichloromethane and washed with a saturated aqueous solution of NaHCO_3 (20 mL). The aqueous layer is extracted with CH_2Cl_2 (4×20 mL) and the combined organic layers are washed with brine (20 mL), dried over Na_2SO_4 , filtered and evaporated under reduced pressure. The crude product is purified by flash chromatography on silica gel (heptane/EtOAc 5:5 to 0:1) to give the pure product.

C2: scale-up

As the glycosylation reaction is slow at room temperature, all the reagents and catalyst are mixed together before injection into the flow system. The *N*-acetyl-D-glucosamine **1 β** (6.50 mmol, 2 eq.) and the iron triflate catalyst (0.49 mmol, 15 mol-%) are added to the acceptor (3.25 mmol, 1 eq.) in an oven-dried, argon-purged vial equipped with a magnetic stirring bar. Dry mixture of chloroform/acetonitrile 7:3 (45 mL, [acceptor] = 0.072 M) is added and the reaction mixture is stirred and sonicated for a few minutes (until complete homogenisation).

After setting up and drying the whole flow system (Figure 2) with dry mixture of chloroform/acetonitrile 7:3 (procedure B), the reagent needle is transferred into the septum-sealed, argon-overpressured vial containing the reaction mixture. This solution is maintained under argon atmosphere during the whole reaction. The pump is then primed to bring the reaction mixture to the solvent/reagent switch valve. The reaction is fully automated and the reaction parameters (temperature, residence time, collection volume) are controlled using Flow Commander software. The reaction mixture is pumped into two 10 mL-stainless steel reactors in series, heated at 110 °C with a fixed flow rate (corresponding to the desired residence time of 70 min). Once all the reaction mixture is loaded into the reactor, the liquid stream is changed back to solvent (mixture of dry chloroform/acetonitrile 7:3) with the same flow rate and temperature until the end of the reaction. The system pressure (33 bar) is controlled with a back pressure regulator and the reaction mixture is finally collected into a single receptor.

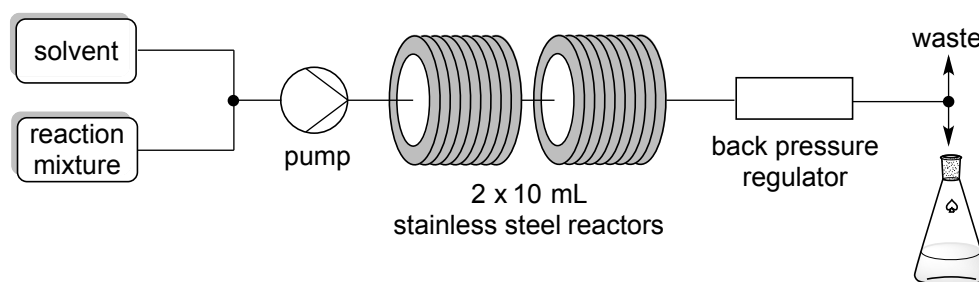


Figure 2

At the end of the reaction, the reaction mixture is diluted with CH_2Cl_2 (50 mL) and washed with a saturated aqueous solution of NaHCO_3 (50 mL). The aqueous layer is extracted with CH_2Cl_2 (4×50 mL) and the combined organic layers are washed with brine (100 mL), dried over Na_2SO_4 , filtered and evaporated under reduced pressure. The crude product is purified by flash chromatography on silica gel (heptane/EtOAc 5:5 to 0:1) to give the pure product.

Products description

Donor **2**¹ and acceptors **3**,² **7**,³ **11**,⁴ **25**⁵ and **27**⁶ were prepared according to known procedures. Donor **33** and acceptor **5** are commercially available.

Iron triflate catalysts

Fe(OTf)₃·6.2DMSO and **Fe(NTf₂)₃·6.3DMSO** were prepared according to the procedure of Antoniotti *et al.*⁷

Fe(OTf)₃·6.2DMSO: Elemental analysis: calculated %C = 18.73, %H = 3.80, %F = 17.32, %Fe = 5.66 and %S = 29.87 and experimental found %C = 18.61, %H = 3.77, %F = 16.97, %Fe = 5.22 and %S = 30.09.

Fe(NTf₂)₃·6.3DMSO: Elemental analysis: calculated %C = 16.09, %H = 2.74, %F = 24.63, %Fe = 4.02 and %S = 28.40 and experimental found %C = 16.00, %H = 2.69, %F = 24.97, %Fe = 3.72 and %S = 29.05.

¹ V. K. Srivastava, *Carbohydr. Res.* **1982**, *103*, 286-292.

² T. Ishikawa, Y. Shimizu, T. Kudoh, S. Saito, *Org. Lett.* **2003**, *5*, 3879-3882.

³ *WO Pat.*, 106403 A1, 2003.

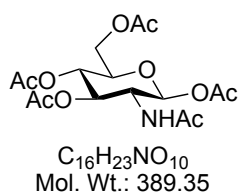
⁴ A. Stévenin, F.-D. Boyer, J.-M. Beau, *J. Org. Chem.* **2010**, *75*, 1783-1786.

⁵ D. Crich, A. U. Vinod, *J. Org. Chem.* **2005**, *70*, 1291-1296.

⁶ D. Tailler, J.-C. Jacquinet, A.-M. Noirot, J.-M. Beau, *J. Chem. Soc., Perkin Trans. I* **1992**, 3163-3164.

⁷ S. Antoniotti, E. Duñach, *Chem. Commun.* **2008**, *8*, 993-995.

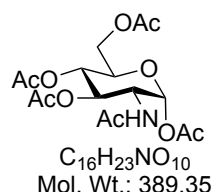
2-Acetamido-1,3,4,6-tetra-*O*-acetyl-2-deoxy- β -D-glucopyranose **1 β**



solid). **1 β** is also commercially available.

1,3,4,6-Tetra-*O*-acetyl-2-amino-2-deoxy- β -D-glucopyranose hydrochloride⁸ (5 g, 13.02 mmol, 1 eq.), acetic anhydride (7.3 mL, 78.17 mmol, 6 eq.) and pyridine (50 mL) were stirred at room temperature for 5 hours under argon. The volatiles were evaporated under reduced pressure and the crude product was purified by chromatography on silica gel (heptane/EtOAc 3:7 to 0:1) to afford **1 β** ⁹ (4.40 g, 87 %, white amorphous

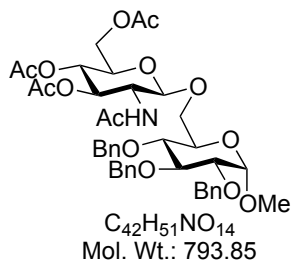
2-Acetamido-1,3,4,6-tetra-*O*-acetyl-2-deoxy- α -D-glucopyranose **1 α**



to 3:7) to give the desired product **1 α** ¹⁰ (1.4 g, 80 %, white amorphous solid). **1 α** is also commercially available.

N-Acetyl-D-glucosamine (1 g, 4.52 mmol) and sodium acetate (990 mg, 12.07 mmol, 2.7 eq.) in acetic anhydride (14 mL) were stirred at 150 °C for 8 hours under argon. The reaction mixture was poured into ice and water, extracted with CH₂Cl₂ (3×50 mL), washed with water (20 mL), brine (10 mL), dried over Na₂SO₄ and evaporated under reduced pressure. The crude product was purified by chromatography on silica gel (heptane/EtOAc 1:1

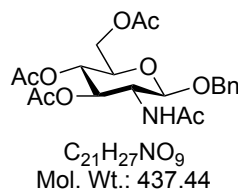
Methyl (2-acetamido-3,4,6-tri-*O*-acetyl-2-deoxy- β -D-glucopyranosyl)-(1→6)-2,3,4-tri-*O*-benzyl- α -D-glucopyranoside **4**



0.49 mmol, 15 mol-%) and acceptor **3** (1.51 g, 3.25 mmol, 1 eq.) in a mixture of dry CHCl₃/CH₃CN 7:3 (45 mL) using general procedure C2 (Vapourtec instrument, 110 °C, 33 bar, 70 min) (2.00 g, 78 %, white amorphous solid).

Microwave conditions: **4**¹¹ was obtained under microwave conditions using donor **1 β** (50 mg, 0.128 mmol, 2 eq.), TTBP (32 mg, 0.129 mmol, 2 eq.), Fe(OTf)₃·6.2DMSO (10 mg, 0.010 mmol, 15 mol-%) and acceptor **3**² (30 mg, 0.065 mmol, 1 eq.) in dry CH₂Cl₂ (1 mL), according to general procedure A (Anton Paar Monowave 300 instrument, 110 °C, 45 min) (46 mg, 89 %, white amorphous solid). Flow conditions: **4** was also obtained under flow conditions from donor **1 β** (2.53 g, 6.50 mmol, 2 eq.), Fe(OTf)₃·6.2DMSO (484 mg,

Benzyl 2-acetamido-3,4,6-tri-*O*-acetyl-2-deoxy- β -D-glucopyranoside **6 β**



also obtained under flow conditions from donor **1 β** (117 mg, 0.301 mmol, 2 eq.), Fe(OTf)₃·6.2DMSO (23 mg, 0.023 mmol, 15 mol-%) and benzyl alcohol **5** (16 μ L,

Microwave conditions: **6 β** ¹² was obtained under microwave conditions using donor **1 β** (50 mg, 0.128 mmol, 2 eq.), TTBP (32 mg, 0.129 mmol, 2 eq.), Fe(OTf)₃·6.2DMSO (10 mg, 0.010 mmol, 15 mol-%) and benzyl alcohol **5** (7 μ L, 0.068 mmol, 1 eq.) in dry CH₂Cl₂ (1 mL) according to general procedure A (Anton Paar Monowave 300 instrument, 110 °C, 45 min) (28 mg, 95 %, white amorphous solid). Flow conditions: **6 β** was

⁸ H. Myszk, D. Bednarczyk, M. Najder, W. Kaca, *Carbohydr. Res.* **2003**, 338, 133-141.

⁹ *US Pat.*, 281395 A1, 2013.

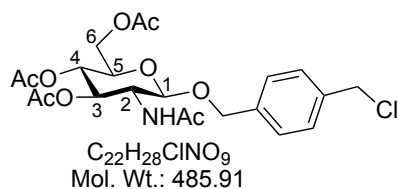
¹⁰ J. E. Heidlas, W. J. Lees, P. Pale, G. M. Whitesides, *J. Org. Chem.* **1992**, 57, 146-151.

¹¹ W. Dullenkopf, J. C. Castro-Palomino, L. Manzoni, R. R. Schmidt *Carbohydr. Res.* **1996**, 296, 135-147.

¹² A. Lubineau, H. Bienaymé, *Carbohydr. Res.* **1991**, 212, 267-271.

0.150 mmol, 1 eq.) in a mixture of dry CH₂Cl₂/CH₃CN 7:3 (2 mL), using general procedure C1 (Vapourtec instrument, 110 °C, 33 bar, 45 min) (51 mg, 77 %, white amorphous solid).

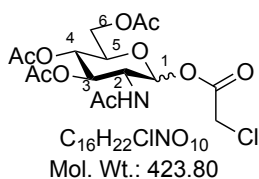
4-(Chloromethyl)benzyl 2-acetamido-3,4,6-tri-*O*-acetyl-2-deoxy-β-D-glucopyranoside 8



Microwave conditions: **8** was obtained under microwave conditions using donor **1β** (50 mg, 0.128 mmol, 2 eq.), TTBP (32 mg, 0.129 mmol, 2 eq.), Fe(OTf)₃·6.2DMSO (10 mg, 0.010 mmol, 15 mol-%) and 4-(chloromethyl)benzyl alcohol **7**³ (10 mg, 0.064 mmol, 1 eq.) in dry CH₂Cl₂ (1 mL), according to general procedure A (Anton Paar Monowave

300 instrument, 110 °C, 30 min) (30 mg, 95%, white amorphous solid). **Flow conditions:** **8** was also obtained under flow conditions using donor **1β** (117 mg, 0.300 mmol, 2 eq.), Fe(OTf)₃·6.2DMSO (23 mg, 0.023 mmol, 15 mol-%) and 4-(chloromethyl)benzyl alcohol **7** (24 mg, 0.153 mmol, 1 eq.) in a mixture of dry CH₂Cl₂/CH₃CN 7:3 (2 mL), according to general procedure C1 (Vapourtec instrument, 110 °C, 33 bar, 45 min) (56 mg, 75 %, white amorphous solid). [α]_D²¹: -36.0 (*c* 1.0 in CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ: 7.36 (d, *J* = 8.0 Hz, 2H, Ar), 7.29 (d, *J* = 8.0 Hz, 2H, Ar), 5.48 (d, *J*_{NH,2} = 9.0 Hz, 1H, NH), 5.21 (dd, *J*_{3,2} = 10.5 Hz and *J*_{3,4} = 9.5 Hz, 1H, H3), 5.09 (t, *J*_{4,3} = *J*_{4,5} = 9.5 Hz, 1H, H4), 4.88 (d, *J* = 12.0 Hz, 1H, OCH₂Ar), 4.65 (d, *J*_{1,2} = 9.0 Hz, 1H, H1), 4.61-4.58 (m, 3H, OCH₂Ar and ArCH₂Cl), 4.27 (dd, *J*_{6,6'} = 12.5 Hz and *J*_{6,5} = 4.5 Hz, 1H, H6), 4.16 (dd, *J*_{6',6} = 12.5 Hz and *J*_{6',5} = 2.5 Hz, 1H, H6'), 3.96 (dt, *J*_{2,3} = 10.5 Hz and *J*_{2,1} = *J*_{2,NH} = 9.0 Hz, 1H, H2), 3.67 (ddd, *J*_{5,4} = 9.5 Hz, *J*_{5,6} = 4.5 Hz and *J*_{5,6'} = 2.5 Hz, 1H, H5), 2.10 (s, 3H, OCOCH₃), 2.02 (s, 3H, OCOCH₃), 2.01 (s, 3H, OCOCH₃), 1.91 (s, 3H, NHCOCH₃). ¹³C NMR (75 MHz, CDCl₃) δ: 171.3 (C, COCH₃), 171.1 (C, COCH₃), 170.5 (C, COCH₃), 169.8 (C, COCH₃), 137.7 (C, OCH₂Ar), 137.6 (C, ArCH₂Cl), 129.1 (2CH, Ar), 128.6 (2CH, Ar), 100.0 (CH, C1), 72.7 (CH, C3), 72.3 (CH, C5), 70.6 (CH₂, OCH₂Ar), 68.9 (CH, C4), 62.5 (CH₂, C6), 54.9 (CH, C2), 46.2 (CH₂, ArCH₂Cl), 23.7 (CH₃, NHCOCH₃), 21.2 (CH₃, OCOCH₃), 21.1 (CH₃, OCOCH₃), 21.0 (CH₃, OCOCH₃). IR ν (film, cm⁻¹): 3278 (N-H), 3103 (=C-H), 2954 and 2877 (C-H), 1738 (C=O), 1661 (NH-C=O). MS (ESI): *m/z* = 486 ([M+H]⁺, 100%), 508 ([M+Na]⁺, 40%), 971 ([2M+H]⁺, 15%), 993 ([2M+Na]⁺, 50%). HRMS (ESI): Calcd for C₂₂H₂₉ClNO₉ [M+H]⁺: 486.1531. Found: 486.1531.

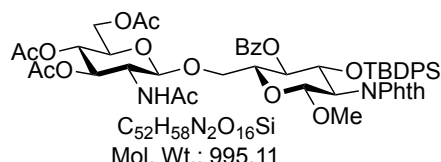
2-Chloroacetyl 2-acetamido-3,4,6-tri-*O*-acetyl-2-deoxy-α/β-D-glucopyranose 10



2-chloroacetic acid **9** (0.6 g, 6.42 mmol, 2.5 eq.) and commercial Fe(OTf)₃ (260 mg, 0.51 mmol, 20 mol-%) are added to donor **1β** (1 g, 2.57 mmol, 1 eq.) in an oven-dried, argon-purged microwave vial equipped with a magnetic stirring bar. Everything is flushed under argon and dry CH₂Cl₂ is added (1 mL). After sealing the vial, the reaction mixture is heated to 80 °C under microwave irradiation for 45 minutes (CEM Discover instrument). Then, the reaction mixture is diluted in CH₂Cl₂ (20 mL) and washed with a saturated aqueous solution of NaHCO₃ (20 mL). The aqueous layer is extracted with CH₂Cl₂ (4×20 mL) and the combined organic layers are washed with brine (20 mL), dried over Na₂SO₄, filtered and concentrated under reduced pressure. The crude product is purified by flash chromatography on silica gel (heptane/EtOAc 5:5 to 0:1) to afford pure product **10** (230 mg, α/β mixture 8/2, 21 %, colorless oil). ¹H NMR (300 MHz, CDCl₃) δ: 6.22 (d, *J*_{1α,2α} = 3.5 Hz, 0.8H, H1α), 5.92 (d, *J*_{NHβ,2β} = 9.5 Hz, 0.2H, NH), 5.75 (d, *J*_{NHα,2α} = 9.0 Hz, 0.8H, NH), 5.74 (d, *J*_{1β,2β} = 9.0 Hz, 0.2H, H1β), 5.25-5.12 (m, 1.2H), 4.50-4.43 (ddd, *J*_{2α,3α} = 10.5 Hz, *J*_{NHα,2α} = 9.0 Hz and *J*_{1α,2α} = 3.5 Hz, 0.8H, H2α), 4.28-4.18 (m, 1H), 4.13 (d, *J* = 1.0 Hz, 1.6H, CH₂Clα), 4.11-3.98 (m, 3.2H), 3.85-3.79 (dddd, *J* = 9.5 Hz, *J* = 7.0 Hz, *J* = 5.0 Hz and *J* = 2.5

Hz, 0.2H, H5 β), 2.05 (s, 3H, Me), 2.02, 2.01, 2.006, 2.00 (4s, 6H, Me), 1.90 (s, 3H, Me). ^{13}C NMR (75 MHz, CDCl_3) α isomer δ : 171.8 (C, CO), 170.8 (C, CO), 170.3 (C, CO), 169.3 (C, CO), 165.5 (C, CO), 92.4 (CH, C1), 70.5, 70.4, 67.4 (3CH, C3, C4, C5), 61.5 (CH₂, C6), 51.5 (CH, C2), 40.8 (CH₂Cl), 23.1 (CH₃), 20.9 (CH₃), 20.8 (CH₃), 20.7 (CH₃). MS (ESI): m/z = 330 ($[\text{M}-\text{CO}_2\text{HCH}_2\text{Cl}]^+$, 100%), 441 ($[\text{M}+\text{NH}_4]^+$, 50%). HRMS (ESI): Calcd for $\text{C}_{16}\text{H}_{26}\text{ClN}_2\text{O}_{10}$ $[\text{M}+\text{NH}_4]^+$: 441.1276. Found: 441.1296.

Methyl (2-acetamido-3,4,6-tri-*O*-acetyl-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 6)-4-*O*-benzoyl-3-*O*-(*tert*-butyldiphenylsilyloxy)-2-deoxy-2-phthalimido- β -D-glucopyranoside 12



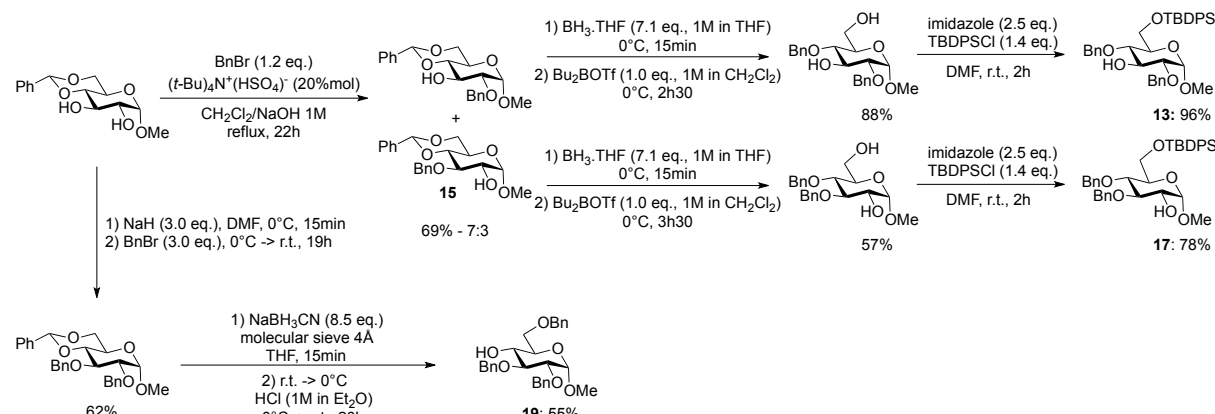
12⁴ was obtained under microwave conditions using donor **1b** (33 mg, 0.084 mmol, 2 eq.), TTBP (21 mg, 0.084 mmol, 2 eq.), $\text{Fe}(\text{OTf})_3 \cdot 6.2\text{DMSO}$ (6 mg, 0.006 mmol, 15 mol-%) and acceptor **11⁴** (28 mg, 0.042 mmol, 1 eq.) in dry CH_2Cl_2 (1 mL), according to general procedure A with a lower concentration (CEM Discover instrument, 80 °C, 45 min) (32 mg, 76 %, white amorphous solid).

Methyl 3-*O*-benzyl-4,6-*O*-benzylidene- α -D-glucopyranoside 15

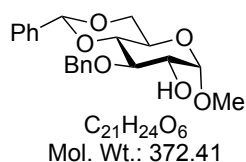
Methyl 2,4-di-*O*-benzyl-6-*O*-*tert*-butyldiphenylsilyl- α -D-glucopyranoside 13

Methyl 3,4-di-*O*-benzyl-6-*O*-*tert*-butyldiphenylsilyl- α -D-glucopyranoside 17

Methyl 2,3,6-tri-*O*-benzyl- α -D-glucopyranoside 19



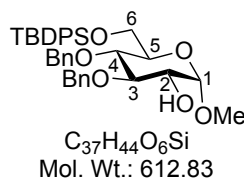
Scheme 1. Synthesis of acceptors 15, 13, 17 and 19



To a mixture of methyl 4,6-*O*-benzylidene- α -D-glucopyranoside (600 mg, 2.13 mmol, 1 eq.) in CH_2Cl_2 (20 mL), was added tetrabutylammonium hydrogensulfate (144 mg, 0.43 mmol, 20 mol-%) and benzyl bromide (0.30 mL, 2.55 mmol, 1.2 eq.). 1 M aqueous NaOH (7 mL) is added and the reaction mixture is stirred under reflux for 22 hours.¹³ Then, the reaction mixture is diluted with CH_2Cl_2 , the organic layer is separated and the water phase is extracted twice with CH_2Cl_2 . The combined organic layers are washed with aqueous saturated NaHCO_3 , brine, dried over Na_2SO_4 , filtered, and concentrated under reduced pressure. The crude product is purified by chromatography on silica gel (heptane/EtOAc 7:3 to 5:5) to afford pure methyl 2-*O*-benzyl-4,6-*O*-benzylidene- α -D-

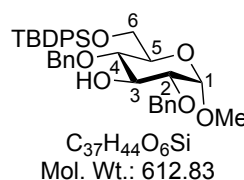
¹³ D. Crich, W. Li, H. Li *J. Am. Chem. Soc.* **2004**, *126*, 15081-15086.

glucopyranoside¹⁴ (21 %, 166 mg) and methyl 3-*O*-benzyl-4,6-*O*-benzylidene- α -D-glucopyranoside **15**¹⁴ (48 %, 380 mg).



Methyl 3-*O*-benzyl-4,6-*O*-benzylidene- α -D-glucopyranoside **15** (169 mg, 0.45 mmol, 1 eq.) is concentrated twice from toluene in a round-bottom flask. Under argon and with magnetic stirring, the flask is almost entirely submerged in an ice-water bath for 10 min and a solution of borane·THF (3.2 mL, 1 M in THF, 3.2 mmol, 7.1 eq.) is added slowly with a syringe along the sides of the flask. After stirring for 15 minutes, a 1 M solution of dibutylboron triflate in CH₂Cl₂ (0.45 mL, 0.45 mmol, 1.0 equiv) is added dropwise and the resulting solution is stirred for 3.5 hours at 0 °C under argon.¹⁵ Then, triethylamine (0.25 mL) is added, methanol (20 mL) is added slowly and the mixture is concentrated under reduced pressure. The mixture is coevaporated three more times with methanol (3×20 mL) and the crude product is purified by column chromatography on silica gel (heptane/EtOAc 7:3 to 4:6) to afford pure methyl 3,4-di-*O*-benzyl- α -D-glucopyranoside¹⁶ (57%, 97 mg).

Methyl 3,4-di-*O*-benzyl- α -D-glucopyranoside (97 mg, 0.26 mmol, 1 eq.), imidazole (44 mg, 0.65 mmol, 2.5 eq.) and *tert*-butyl(chloro)diphenylsilane (9.5 μ L, 0.36 mmol, 1.4 eq.) in dry DMF (1.2 mL) were stirred at room temperature for 2 hours under argon. The reaction mixture was poured into a saturated aqueous solution of NaHCO₃ (20 mL), extracted with Et₂O (3×10 mL), washed with brine (20 mL), dried over Na₂SO₄ and evaporated under reduced pressure. The crude product was purified by chromatography on silica gel (heptane/EtOAc 9:1 to 6:4) to give the desired product **17** (125 mg, 78 %, colorless oil). [α]_D²⁰: +41.8 (*c* 0.9 in CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ : 7.74-7.68 (m, 4H, Ph), 7.45-7.26 (m, 14H, Ph), 7.19-7.16 (m, 2H, Ph), 4.92 (d, *J* = 11.0 Hz, 1H, CH₂Ph), 4.90-4.85 (m, 2H, CH₂Ph), 4.79 (d, *J*_{1,2} = 3.5 Hz, 1H, H1), 4.64 (d, *J* = 10.5 Hz, 1H, CH₂Ph), 3.91 (m, 2H, H6 and H6'), 3.78 (t, *J*_{3,2} = *J*_{3,4} = 9.0 Hz, 1H, H3), 3.77-3.62 (m, 3H, H2, H4 and H5), 3.40 (s, 3H, OCH₃), 2.14 (brs, 1H, OH), 1.06 (s, 9H, C(CH₃)₃). ¹³C NMR (75 MHz, CDCl₃) δ : 139.0 (C, Ph), 138.6 (C, Ph), 136.2 (CH, Ph), 136.0 (CH, Ph), 134.0 (C, Ph), 133.6 (C, Ph), 130.0 (CH, Ph), 130.0 (CH, Ph), 128.9 (CH, Ph), 128.8 (CH, Ph), 128.4 (CH, Ph), 128.2 (CH, Ph), 128.1 (CH, Ph), 128.1 (CH, Ph), 127.9 (CH, Ph), 99.5 (CH, C1), 83.8 (CH, C3), 78.0 (CH, C4), 75.9 (CH₂, CH₂Ph), 75.4 (CH₂, CH₂Ph), 73.6 (CH, C2), 72.1 (CH, C5), 63.2 (CH₂, C6), 55.3 (CH₃, OCH₃), 27.2 (3CH₃, C(CH₃)₃), 19.7 (C, C(CH₃)₃). IR ν (film, cm⁻¹): 3465 (O-H), 3068 (=C-H), 2931 and 2857 (C-H). MS (ESI): *m/z* = 630 ([M+NH₄]⁺, 5%), 635 ([M+Na]⁺, 100%), 1248 ([2M+Na]⁺, 20%). HRMS (ESI): Calcd for C₃₇H₄₄NaO₆Si [M+Na]⁺: 635.2805. Found: 635.2814.



Methyl 2-*O*-benzyl-4,6-*O*-benzylidene- α -D-glucopyranoside (282 mg, 0.76 mmol, 1 eq.) is concentrated twice by coevaporation with toluene in a round-bottom flask. Under argon and with magnetic stirring, the flask is almost entirely submerged in an ice-water bath for 10 min and a solution of borane·THF (5.5 mL, 1 M in THF, 5.5 mmol, 7.1 eq.) is added slowly with a syringe along the sides of the flask. After stirring for 15 minutes, a solution of dibutylboron triflate (0.76 mL, 1 M in CH₂Cl₂, 0.76 mmol, 1.0 equiv) is added dropwise and the resulting solution is stirred for 2.5 hours at 0 °C under argon.¹⁵ Then, triethylamine (0.4 mL) is added, methanol (35 mL) is added slowly and the mixture is concentrated under reduced pressure. The mixture is coevaporated three more times with methanol (3×35 mL) and the crude product is purified by column chromatography on

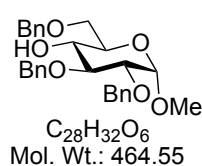
¹⁴ A. G. M. Barrett, R. W. Read, D. H. R. Barton, *J. Chem. Soc., Perkin Trans. 1* **1980**, 2184-2190.

¹⁵ V. Y. Dudkin, J. S. Miller, A. S. Dudkina, C. Antczak, D. A. Scheinberg, S. J. Danishefsky, *J. Am. Chem. Soc.* **2008**, *130*, 13598-13607.

¹⁶ A. François, D. Urban, J.-M. Beau, *Angew. Chem., Int. Ed.* **2007**, *46*, 8662-8665.

silica gel (heptane/EtOAc 7:3 to 4:6) to afford pure methyl 2,4-di-*O*-benzyl- α -D-glucopyranoside¹⁷ (88%, 250 mg).

Methyl 2,4-di-*O*-benzyl- α -D-glucopyranoside (250 mg, 0.67 mmol, 1 eq.), imidazole (113 mg, 1.66 mmol, 2.5 eq.) and *tert*-butyl(chloro)diphenylsilane (240 μ L, 0.93 mmol, 1.4 eq.) in dry DMF (3 mL) were stirred at room temperature for 2 hours under argon. The reaction mixture was poured into a saturated aqueous solution of NaHCO₃ (30 mL), extracted with Et₂O (3 \times 20 mL), washed with brine (30 mL), dried over Na₂SO₄ and evaporated under reduced pressure. The crude product was purified by chromatography on silica gel (heptane/EtOAc 9:1 to 7:3) to give the desired product **13** (390 mg, 96 %, colorless oil). $[\alpha]_{\text{D}}^{20}$: +47.1 (*c* 1.0 in CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ : 7.72-7.67 (m, 4H, Ph), 7.45-7.25 (m, 14H, Ph), 7.24-7.20 (m, 2H, Ph), 4.89 (d, *J* = 11.0 Hz, 1H, CH₂Ph), 4.73 (s, 2H, CH₂Ph), 4.69 (d, *J*_{1,2} = 3.5 Hz, 1H, H1), 4.63 (d, *J* = 11.0 Hz, 1H, CH₂Ph), 4.11 (t, *J*_{3,2} = *J*_{3,4} = 9.5 Hz, 1H, H3), 3.92 (dd, *J*_{6,6'} = 11.0 Hz and *J*_{6,5} = 2.5 Hz, 1H, H6), 3.87 (dd, *J*_{6',6} = 11.0 Hz and *J*_{6',5} = 4.0 Hz, 1H, H6'), 3.69 (ddd, *J*_{5,4} = 9.5 Hz, *J*_{5,6'} = 4.0 Hz and *J*_{5,6} = 2.5 Hz, 1H, H5), 3.54 (t, *J*_{4,3} = *J*_{4,5} = 9.5 Hz, 1H, H4), 3.42 (dd, *J*_{2,3} = 9.5 Hz and *J*_{2,1} = 3.5 Hz, 1H, H2), 3.34 (s, 3H, OCH₃), 2.37 (brs, 1H, OH), 1.06 (s, 9H, C(CH₃)₃). ¹³C NMR (75 MHz, CDCl₃) δ : 138.8 (C, Ph), 138.5 (C, Ph), 136.2 (CH, Ph), 136.0 (CH, Ph), 134.0 (C, Ph), 133.7 (C, Ph), 130.0 (CH, Ph), 129.9 (CH, Ph), 128.9 (CH, Ph), 128.7 (CH, Ph), 128.4 (CH, Ph), 128.2 (CH, Ph), 128.0 (CH, Ph), 128.0 (CH, Ph), 127.9 (CH, Ph), 97.6 (CH, C1), 80.3 (CH, C2), 78.1 (CH, C4), 75.0 (CH₂, CH₂Ph), 74.0 (CH, C3), 73.4 (CH₂, CH₂Ph), 71.5 (CH, C5), 63.5 (CH₂, C6), 55.2 (CH₃, OCH₃), 27.2 (3CH₃, C(CH₃)₃), 19.7 (C, C(CH₃)₃). IR ν (film, cm⁻¹): 3461 (O-H), 3068 (=C-H), 2930 and 2857 (C-H). MS (ESI): *m/z* = 630 ([M+NH₄]⁺, 10%), 635 ([M+Na]⁺, 100%), 1248 ([2M+Na]⁺, 85%). HRMS (ESI): Calcd for C₃₇H₄₄NaO₆Si [M+Na]⁺: 635.2805. Found: 635.2808.



Methyl 4,6-*O*-benzylidene- α -D-glucopyranoside (3.00 g, 10.63 mmol, 1 eq.) in dry DMF (35 mL) is cooled to 0 °C under argon. NaH (60% dispersion in mineral oil, 960 mg, 31.88 mmol, 3 eq.), is added by portion and the reaction mixture is stirred at 0 °C for 15 minutes under argon. BnBr (3.8 mL, 31.88 mmol, 3 eq.) is added slowly and the reaction mixture is allowed to warm to room temperature and is stirred at this temperature for 19 hours. MeOH (3 mL) and water (20 mL) are successively added and the reaction mixture is extracted with EtOAc (3 \times 10 mL). The combined organic layers are washed with NaCl sat. (30 mL), dried over Na₂SO₄ and evaporated under reduced pressure. The crude product is purified by chromatography on silica gel (heptane/EtOAc 90:10 to 5:5) to afford methyl 2,3-di-*O*-benzyl-4,6-*O*-benzylidene- α -D-glucopyranoside¹⁴ (3.05 g, 62 %).

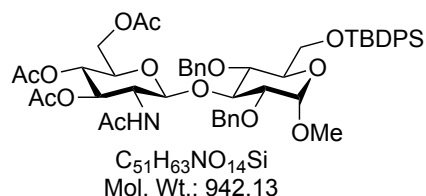
To a solution of methyl 2,3-di-*O*-benzyl-4,6-*O*-benzylidene- α -D-glucopyranoside (1.50 g, 3.24 mmol, 1 eq.) in dry THF (35 mL) is added powdered molecular sieves (4 Å, 1.70 g), methyl orange (3 mg) and NaBH₃CN (1.70 g, 27.57 mmol, 8.5 eq.) under argon. After 15 minutes at room temperature, the yellow solution is cooled to 0 °C and HCl in Et₂O (1 M) is added slowly until the solution turns pink and gas evolution ceased completely (danger, release of HCN!).¹⁸ The solution is then warmed to room temperature and stirred at this temperature for 20 hours under argon. The reaction mixture is poured into a cold saturated aqueous solution of NaHCO₃ (60 mL) and the aqueous layer is extracted with EtOAc (3 \times 60 mL). The combined organic layers are washed with water (3 \times 20 mL), brine (60 mL), dried over Na₂SO₄ and evaporated under reduced pressure. The crude product is purified by

¹⁷ T. Ogawa, T. Kaburagi, *Carbohydr. Res.* **1982**, 103, 53-64.

¹⁸ S. Norsikian, A. Lubineau, *Org. Biomol. Chem.* **2005**, 3, 4089-4094.

chromatography on silica gel (heptane/EtOAc 9:1 to 6:4) to give the desired product **19**¹⁹ (830 mg, 55 %, white amorphous solid).

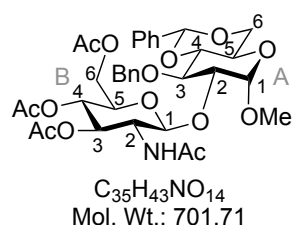
Methyl (2-acetamido-3,4,6-tri-*O*-acetyl-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 3)-2,4-di-*O*-benzyl-6-*O*-*tert*-butyldiphenylsilyl- α -D-glucopyranoside **14**



14⁴ was obtained under microwave conditions using donor **1b** (50 mg, 0.128 mmol, 2 eq.), TTBP (32 mg, 0.129 mmol, 2 eq.), Fe(OTf)₃·6.2DMSO (10 mg, 0.010 mmol, 15 mol-%) and acceptor **13** (42 mg, 0.068 mmol, 1 eq.) in dry CH₂Cl₂ (1 mL), according to general procedure A (Anton Paar Monowave 300 instrument, 110 °C, 45 min) (47 mg,

74 %, white amorphous solid).

Methyl (2-acetamido-3,4,6-tri-*O*-acetyl-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 2)-3-*O*-benzyl-4,6-*O*-benzylidene- α -D-glucopyranoside **16**

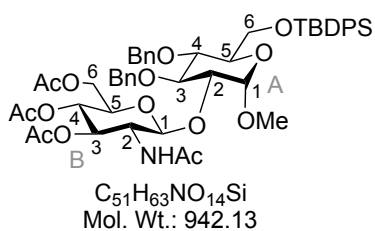


16 was obtained under microwave conditions using donor **1b** (50 mg, 0.128 mmol, 2 eq.), TTBP (32 mg, 0.129 mmol, 2 eq.), Fe(OTf)₃·6.2DMSO (10 mg, 0.010 mmol, 15 mol-%) and acceptor **15** (23 mg, 0.062 mmol, 1 eq.) in dry CH₂Cl₂ (1 mL), according to general procedure A (Anton Paar Monowave 300 instrument, 110 °C, 60 min) (27 mg, 61 %, white amorphous solid). $[\alpha]_D^{22}$: +15.9 (*c* 0.9 in CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ : 7.44-7.40 (m, 2H, Ph),

7.36-7.24 (m, 8H, Ph), 5.53 (s, 1H, CHPh), 5.23 (dd, $J_{B3,B2}$ = 10.5 Hz and $J_{B3,B4}$ = 9.5 Hz, 1H, HB-3), 5.16 (d, $J_{NH,B2}$ = 8.5 Hz, 1H, NH), 5.01 (t, $J_{B4,B3}$ = $J_{B4,B5}$ = 9.5 Hz, 1H, HB-4), 4.92 (d, J = 12.0 Hz, 1H, CH₂Ph), 4.90 (d, $J_{B1,B2}$ = 8.5 Hz, 1H, HB-1), 4.85 (d, $J_{A1,A2}$ = 3.5 Hz, 1H, HA-1), 4.63 (d, J = 12.0 Hz, 1H, CH₂Ph), 4.27 (dd, $J_{A6,A6'}$ = 10.0 Hz and $J_{A6,A5}$ = 4.5 Hz, 1H, HA-6), 4.17 (d, $J_{B6,B5}$ = $J_{B6',B5}$ = 4.0 Hz, 2H, HB-6 and HB-6'), 3.99 (t, $J_{A3,A2}$ = $J_{A3,A4}$ = 9.5 Hz, 1H, HA-3), 3.88-3.79 (m, 2H, HB-2 and HA-5), 3.75-3.62 (m, 3H, HB-5, HA-2 and HA-6'), 3.59 (t, $J_{A4,A3}$ = $J_{A4,A5}$ = 9.5 Hz, 1H, HA-4), 3.40 (s, 3H, OCH₃), 2.07 (s, 3H, OCOCH₃), 2.00 (s, 3H, OCOCH₃), 1.97 (s, 3H, OCOCH₃), 1.51 (s, 3H, NHCOCH₃). ¹³C NMR (75 MHz, CDCl₃) δ : 171.0 (C, COCH₃), 171.0 (C, COCH₃), 170.7 (C, COCH₃), 169.8 (C, COCH₃), 139.3 (C, Ph), 137.6 (C, Ph), 129.4 (CH, Ph), 128.9 (CH, Ph), 128.6 (CH, Ph), 128.1 (CH, Ph), 127.4 (CH, Ph), 126.4 (CH, Ph), 102.4 (CH, CB-1), 101.8 (CH, CHPh), 100.4 (CH, CA-1), 82.8 (CH, CA-4), 81.0 (CH, CA-2), 78.0 (CH, CA-3), 75.1 (CH₂, CH₂Ph), 72.8 (CH, CB-3), 72.2 (CH, CB-5), 69.5 (CH₂, CA-6), 69.0 (CH, CB-4), 62.6 (CH₂, CB-6), 62.5 (CH, CA-5), 55.9 (CH₃, OCH₃), 55.3 (CH, CB-2), 23.3 (CH₃, NHCOCH₃), 21.2 (CH₃, OCOCH₃), 21.0 (CH₃, OCOCH₃). IR ν (film, cm⁻¹): 3280 (N-H), 3092, 3062 and 3030 (=C-H), 2920 and 2871 (C-H), 1744 (C=O), 1666 (NH-C=O). MS (ESI): *m/z* = 702 ([M+H]⁺, 100%), 724 ([M+Na]⁺, 55%), 1404 ([2M+H]⁺, 10%), 1426 ([2M+Na]⁺, 75%). HRMS (ESI): Calcd for C₃₅H₄₄NO₁₄ [M+H]⁺: 702.2762. Found: 702.2766.

Methyl (2-acetamido-3,4,6-tri-*O*-acetyl-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 2)-3,4-di-*O*-benzyl-6-*O*-*tert*-butyldiphenylsilyl- α -D-glucopyranoside **18**

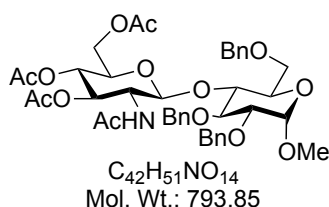
¹⁹ C.-R. Shie, Z.-H. Tzeng, S. S. Kulkarni, B.-J. Uang, C.-Y. Hsu, S.-C. Hung *Angew. Chem., Int. Ed.* **2005**, *44*, 1665-1668.



18 was obtained under microwave conditions using donor **1β** (50 mg, 0.128 mmol, 2 eq.), TTBP (32 mg, 0.129 mmol, 2 eq.), Fe(OTf)₃·6.2DMSO (10 mg, 0.010 mmol, 15 mol-%) and acceptor **17** (42 mg, 0.068 mmol, 1 eq.) in dry CH₂Cl₂ (1 mL), according to general procedure A (Anton Paar Monowave 300 instrument, 110 °C, 60 min) (34 mg, 53 %, white amorphous solid). $[\alpha]_D^{20}$: +24.2 (*c* 1.0 in CHCl₃). ¹H NMR (300 MHz,

CDCl₃) δ : 7.73-7.67 (m, 4H, Ph), 7.43-7.27 (m, 12H, Ph), 7.22-7.20 (m, 2H, Ph), 7.06-7.03 (m, 2H, Ph), 5.35 (dd, $J_{B3,B2}$ = 10.5 Hz and $J_{B3,B4}$ = 9.5 Hz, 1H, HB-3), 5.20 (d, $J_{NH,B2}$ = 8.5 Hz, 1H, NH), 5.03 (t, $J_{B4,B3}$ = $J_{B4,B5}$ = 9.5 Hz, 1H, HB-4), 4.99 (d, $J_{B1,B2}$ = 8.5 Hz, 1H, HB-1), 4.91 (d, $J_{A1,A2}$ = 3.5 Hz, 1H, HA-1), 4.83 (s, 2H, CH₂Ph), 4.76 (d, J = 10.5 Hz, 1H, CH₂Ph), 4.58 (d, J = 10.5 Hz, 1H, CH₂Ph), 4.23-4.21 (m, 2H, HB-6 and HB-6'), 3.97 (dd, J = 9.5 Hz and J = 8.5 Hz, 1H, HA-3), 3.88 (d, $J_{A6,A5}$ = $J_{A6',A5}$ = 2.5 Hz, 2H, HA-6 and HA-6'), 3.84-3.61 (m, 5H, HB-2, HB-5, HA-2, HA-5 and HA-4), 3.37 (s, 3H, OCH₃), 2.09 (s, 3H, OCOCH₃), 2.03 (s, 3H, OCOCH₃), 1.99 (s, 3H, OCOCH₃), 1.45 (s, 3H, NHCOCH₃), 1.07 (s, 9H, C(CH₃)₃). ¹³C NMR (75 MHz, CDCl₃) δ : 171.0 (C, COCH₃), 171.0 (C, COCH₃), 170.7 (C, COCH₃), 169.9 (C, COCH₃), 139.5 (C, Ph), 138.4 (C, Ph), 136.2 (CH, Ph), 136.0 (CH, Ph), 133.9 (C, Ph), 133.6 (C, Ph), 130.0 (CH, Ph), 130.0 (CH, Ph), 128.9 (CH, Ph), 128.7 (CH, Ph), 128.2 (CH, Ph), 128.1 (CH, Ph), 128.0 (CH, Ph), 128.0 (CH, Ph), 127.9 (CH, Ph), 127.0 (CH, Ph), 102.2 (CB-1), 99.4 (CH, CA-1), 82.5 (CH, CA-2), 81.4 (CH, CA-3), 78.4 (CH, CA-4), 75.4 (CH₂, CH₂Ph), 75.4 (CH₂, CH₂Ph), 72.7 (CH, CB-3), 72.1 (CH, CB-5), 71.5 (CH, CA-5), 69.2 (CB-4), 63.1 (CH₂, CA-6), 62.8 (CH₂, CB-6), 55.7 (CH, CB-2), 55.3 (CH₃, OCH₃), 27.2 (CH₃, C(CH₃)₃), 23.2 (CH₃, NHCOCH₃), 21.1 (CH₃, OCOCH₃), 21.0 (CH₃, OCOCH₃), 19.7 (C, C(CH₃)₃). IR ν (film, cm⁻¹): 3265 (N-H), 3071 and 3032 (=C-H), 2959, 2931 and 2857 (C-H), 1745 (C=O), 1656 (NH-C=O). MS (ESI): *m/z* = 964 ([M+Na]⁺, 100%). HRMS (ESI): Calcd for C₅₁H₆₃NNaO₁₄Si [M+Na]⁺: 964.3916. Found: 964.3923.

Methyl (2-acetamido-3,4,6-tri-*O*-acetyl-2-deoxy-β-D-glucopyranosyl)-(1→4)-2,3,6-tri-*O*-benzyl-α-D-glucopyranoside **20**

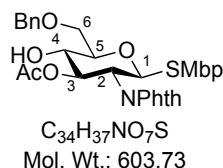


20²⁰ was obtained under microwave conditions using donor **1β** (500 mg, 1.284 mmol, 2 eq.), TTBP (320 mg, 1.288 mmol, 2 eq.), Fe(OTf)₃·6.2DMSO (96 mg, 0.097 mmol, 15 mol-%) and acceptor **19** (300 mg, 0.646 mmol, 1 eq.) in dry CH₂Cl₂ (1 mL), according to general procedure A with a higher concentration (Anton Paar Monowave 300 instrument, 110 °C, 3 hours) (191

mg, 37 %, white amorphous solid).

²⁰ H. Christensen, M. S. Christiansen, J. Petersen, H. H. Jensen, *Org. Biomol. Chem.* **2008**, 6, 3276-3283.

(2-Methyl-5-*tert*-butylphenyl) 3-*O*-acetyl-6-*O*-benzyl-2-deoxy-2-phthalimido-1-thio-β-D-glucopyranoside 21

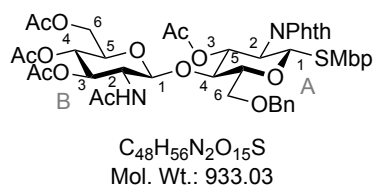


(2-Methyl-5-*tert*-butylphenyl) 3-*O*-acetyl-4,6-*O*-benzylidene-2-deoxy-2-phthalimido-1-thio-β-D-glucopyranoside²¹ (1.4 g, 2.33 mmol, 1 eq.) and trifluoroacetic acid (864 μL, 11.63 mmol, 5 eq.) in dry CH₂Cl₂ (23 mL) were cooled to 0 °C under argon. Et₃SiH (1.85 mL, 11.63, 5 eq.) was added dropwise to the reaction mixture which was then warmed to room temperature and stirred for 8 hours under argon. The reaction mixture was poured into a saturated aqueous solution of NaHCO₃ (30 mL), extracted with CH₂Cl₂ (3×40 mL), dried over Na₂SO₄ and evaporated under reduced pressure. The crude product was purified by chromatography on silica gel (heptane/EtOAc 8:2 to 6:4) to give the desired product **21** (1.07 g, 76 %, white amorphous solid). [α]_D²⁵: +32.3 (*c* 1.0, CHCl₃). Mp: 68.2-73.5 °C (from heptane/EtOAc). ¹H NMR (500 MHz, CDCl₃) δ: 7.87-7.80 (m, 2H, NPhth), 7.75-7.69 (m, 2H, NPhth), 7.46 (d, *J* = 1.5 Hz, 1H, Ph), 7.36-7.27 (m, 5H, Ph), 7.16 (dd, *J* = 8.0 Hz, *J* = 1.5 Hz, 1H, Ph), 7.01 (d, *J* = 8.0 Hz, 1H, Ph), 5.66 (dd, *J*_{3,2} = 10.0 Hz, *J*_{3,4} = 9.0 Hz, 1H, H3), 5.62 (d, *J*_{1,2} = 10.0 Hz, 1H, H1), 4.58 (AB system, *J* = 12.0 Hz, 2H, CH₂Ph), 4.34 (t, *J*_{2,1} = *J*_{2,3} = 10.0 Hz, 1H, H2), 3.88-3.81 (m, 2H, H4 and H6), 3.77 (dd, *J*_{6,6'} = 10.0 Hz, *J*_{6,5} = 5.0 Hz, 1H, H6'), 3.71 (dd, *J*_{5,4} = 9.5 Hz, *J*_{5,6'} = 5.0 Hz, 1H, H5), 2.91 (d, *J*_{OH,4} = 3.5 Hz, 1H, OH), 2.13 (s, 3H, Me), 1.90 (s, 3H, Ac), 1.22 (s, 9H, *t*-Bu). ¹³C NMR (75 MHz, CDCl₃) δ: 171.2 (C, CO), 168.0 (C, NCO), 167.5 (C, NCO), 149.7 (C, Ph), 137.7 (C, Ph), 137.4 (C, Ph), 134.6 (CH, NPhth), 134.4 (CH, NPhth), 131.9 (C, NPhth), 131.5 (C, NPhth), 131.3 (C, Ph), 130.9 (CH, Ph), 130.1 (CH, Ph), 128.7 (2CH, Ph), 128.1 (3CH, Ph), 125.6 (CH, Ph), 123.8 (2CH, NPhth), 84.4 (CH, C1), 78.1 (CH, C5), 74.5 (CH, C3), 74.0 (CH₂, CH₂Ph), 71.7 (CH, C4), 70.5 (CH₂, C6), 54.0 (CH, C2), 34.6 (C, *t*-Bu), 31.4 (3CH₃, *t*-Bu), 20.9 (CH₃, Ac), 20.5 (CH₃, Me). IR ν (film, cm⁻¹): 3472 (O-H), 3063 and 3036 (=C-H), 2964, 2908 and 2865 (C-H), 1776 (N-C=O), 1742 (C=O), 1715 (N-C=O). MS (ESI): *m/z* = 621 ([M+NH₄]⁺, 60%). HRMS (ESI): Calcd for C₃₄H₄₁N₂O₇S [M+NH₄]⁺: 621.2634. Found: 621.2618.

(2-Methyl-5-*tert*-butylphenyl)

(2-acetamido-3,4,6-tri-*O*-acetyl-2-deoxy-β-D-

glucopyranosyl)-(1→4)-3-*O*-acetyl-6-*O*-benzyl-2-deoxy-2-phthalimido-1-thio-β-D-glucopyranoside 22



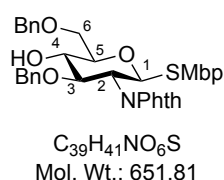
Acceptor **21** (155 mg, 0.257 mmol, 2 eq.), TTBP (32 mg, 0.129 mmol, 2 eq.) and Fe(OTf)₃·6.2DMSO (19 mg, 0.019 mmol, 15 mol-%) are added to donor **1β** (50 mg, 0.128 mmol, 1 eq.) in an oven-dried, argon-purged microwave vial equipped with a magnetic stirring bar. Everything is flushed under argon and dry CH₂Cl₂ is added (1 mL). After sealing the vial, the reaction mixture is heated to 70 °C under microwave irradiation for 11 hours (CEM Discover instrument). Then, the reaction mixture is diluted in CH₂Cl₂ (20 mL) and washed with a saturated aqueous solution of NaHCO₃ (20 mL). The aqueous layer is extracted with CH₂Cl₂ (4×20 mL) and the combined organic layers are washed with brine (20 mL), dried over Na₂SO₄, filtered and concentrated under reduced pressure. The crude product was purified by chromatography on silica gel (heptane/EtOAc 5:5 to 0:1) to afford pure product **22** (28 mg, 23 %, white amorphous solid). [α]_D²⁵: +42.4 (*c* 0.5, CHCl₃). ¹H NMR (500 MHz, CDCl₃) δ: 7.85-7.78 (m, 2H, NPhth), 7.74-7.67 (m, 2H, NPhth), 7.49-7.39 (m, 6H, Ph), 7.16 (dd, *J* = 8.0 Hz, *J* = 2.0 Hz, 1H, Ph), 7.03 (d, *J* = 8.0 Hz, 1H, Ph), 5.68 (t, *J*_{A3,A2} = *J*_{A3,A4} = 9.5 Hz, 1H, HA-3), 5.58 (d, *J*_{A1,A2} = 10.5 Hz, 1H, HA-1), 5.05 (t, *J*_{B3,B2} = *J*_{B3,B4} = 9.5 Hz, 1H, HB-3), 4.90 (t, *J*_{B4,B3}

²¹ M. Collot, J. Savreux, J.-M. Mallet, *Tetrahedron* **2008**, *64*, 1523-1535.

$= J_{B4,B5} = 9.5$ Hz, 1H, HB-4), 4.83 (d, $J = 12.0$ Hz, 1H, CH₂Ph), 4.77 (d, $J_{NH,B2} = 9.0$ Hz, 1H, NH), 4.57 (d, $J_{B1,B2} = 8.5$ Hz, 1H, HB-1), 4.45 (d, $J = 12.0$ Hz, 1H, CH₂Ph), 4.37-4.27 (m, 2H, HB-6 and HA-2), 4.03 (t, $J_{A4,A3} = J_{A4,A5} = 9.5$ Hz, 1H, HA-4), 3.95 (d, $J_{B6',B6} = 12.0$ Hz, 1H, HB-6'), 3.69-3.57 (m, 4H, HB-2, HA-5, HA-6 and HA-6'), 3.52-3.44 (m, 1H, HB-5), 2.15 (s, 3H, Me), 2.01 (s, 3H, Ac), 1.99 (s, 3H, Ac), 1.96 (s, 3H, Ac), 1.83 (s, 3H, Ac), 1.72 (s, 3H, Ac), 1.25 (s, 9H, *t*-Bu). ¹³C NMR (75 MHz, CDCl₃) δ : 170.8 (2C, 2CO), 170.3 (C, CO), 170.0 (C, CO), 169.6 (C, CO), 167.8 (C, NCO), 167.4 (C, NCO), 149.6 (C, Ph), 138.0 (C, Ph), 137.6 (C, Ph), 134.5 (CH, NPhth), 134.3 (CH, NPhth), 131.9 (C, NPhth), 131.5 (C, NPhth), 131.2 (CH, Ph), 131.1 (CH, Ph), 130.1 (CH, Ph), 129.1 (2CH, Ph), 129.0 (2CH, Ph), 128.8 (CH, Ph), 125.7 (CH, Ph), 123.8 (CH, NPhth), 123.7 (CH, NPhth), 100.0 (CH, CB-1), 84.4 (CH, CA-1), 78.5 (CH, CA-5), 75.3 (CH, CA-4), 73.9 (CH₂, CH₂Ph), 72.6 (CH, CB-3), 71.8 (CH, CA-3), 71.7 (CH, CB-5), 68.6 (CH, CB-4), 67.8 (CH₂, CA-6), 62.0 (CH₂, CB-6), 55.0 (CH, CB-2), 54.4 (CH, CA-2), 34.6 (C, *t*-Bu), 31.4 (3CH₃, *t*-Bu), 23.3 (CH₃, Ac), 20.8 (3CH₃, 2Ac and Me), 20.6 (CH₃, Ac), 20.5 (CH₃, Ac). IR ν (film, cm⁻¹): 3024 (N-H), 2964 (C-H), 1745 (C=O), 1719 (N-C=O). MS (ESI): $m/z = 955$ ([M+Na]⁺, 100%). HRMS (ESI): Calcd for C₄₈H₅₆N₂NaO₁₅S [M+Na]⁺: 955.3299. Found: 955.3287.

(2-Methyl-5-*tert*-butylphenyl)

3,6-di-*O*-benzyl-2-deoxy-2-phthalimido-1-thio- β -D-glucopyranoside **23**

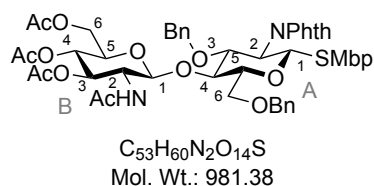


To a suspension of (2-Methyl-5-*tert*-butylphenyl) 4,6-*O*-benzylidene-2-deoxy-2-phthalimido-1-thio- β -D-glucopyranoside²¹ (4.09 g, 7.31 mmol) and NaH (60 % dispersion in mineral oil, 366 mg, 9.14 mmol, 1.25 eq.) in dry DMF (40 mL), was added dropwise BnBr (1.31 mL, 10.96 mmol, 1.5 eq.). The reaction mixture was stirred at room temperature for 4 hours under argon. MeOH (6 mL) and water (100 mL) were successively added and the reaction mixture was extracted with EtOAc (3 \times 50 mL), dried over Na₂SO₄ and evaporated under reduced pressure. The crude product was purified by chromatography on silica gel (heptane/EtOAc 1:0 to 7:3) to give (2-methyl-5-*tert*-butylphenyl) 3-*O*-benzyl-4,6-*O*-benzylidene-2-deoxy-2-phthalimido-1-thio- β -D-glucopyranoside (3 g, 63 %, white amorphous solid). 3-*O*-benzyl-4,6-*O*-benzylidene-2-deoxy-2-phthalimido-1-thio- β -D-glucopyranoside (1.5 g, 2.31 mmol) and NaBH₃CN (1.99 g, 31.63 mmol, 13.7 eq.) in dry CH₂Cl₂ (23 mL) were cooled to 0 °C under argon. A solution of HCl in Et₂O (2 M, 23 mL) was added dropwise to the reaction mixture (danger, release of HCN!). It was then warmed to room temperature and stirred at this temperature for 8 hours under argon. The reaction mixture was poured into a saturated aqueous solution of NaHCO₃ (50 mL), extracted with CH₂Cl₂ (3 \times 70 mL), washed with a saturated aqueous solution of NaHCO₃ (40 mL), a 1 M aqueous solution of HCl (40 mL), brine (20 mL), dried over Na₂SO₄ and evaporated under reduced pressure. The crude product was purified by chromatography on silica gel (heptane/EtOAc 8:2 to 6:4) to give the desired product **23** (1.24 g, 82 %, white amorphous solid). $[\alpha]_D^{25}$: +81.2 (*c* 1.0, CHCl₃). ¹H NMR (500 MHz, CDCl₃) δ : 7.82 (d, $J = 6.5$ Hz, 1H, NPhth), 7.73-7.61 (m, 3H, NPhth), 7.41 (d, $J = 1.5$ Hz, 1H, Ph), 7.36-7.27 (m, 5H, Ph), 7.12 (dd, $J = 8.0$ Hz, $J = 2.0$ Hz, 1H, Ph), 7.05-7.01 (m, 2H, Ph), 6.98 (d, $J = 8.0$ Hz, 1H, Ph), 6.96-6.90 (m, 3H, Ph), 5.44 (d, $J_{1,2} = 10.0$ Hz, 1H, H1), 4.72 (d, $J = 12.0$ Hz, 1H, CH₂Ph), 4.63-4.49 (m, 3H, CH₂Ph), 4.32-4.22 (m, 2H, H2 and H3), 3.88-3.79 (m, 2H, H4 and H6), 3.75 (dd, $J_{6',6} = 10.0$ Hz, $J_{6',5} = 5.5$ Hz, 1H, H6'), 3.67-3.60 (m, 1H, H5), 2.88 (d, $J_{OH,4} = 2.0$ Hz, 1H, OH), 2.09 (s, 3H, Me), 1.2 (s, 9H, *t*-Bu). ¹³C NMR (75 MHz, CDCl₃) δ : 168.2 (C, NCO), 167.5 (C, NCO), 149.6 (2C, NPhth), 138.3 (C, Ph), 137.7 (C, Ph), 137.1 (C, Ph), 134.2 (CH, NPhth), 134.0 (CH, NPhth), 131.9 (2C, Ph), 130.4 (CH, Ph), 130.0 (CH, Ph), 128.7 (2CH, Ph), 128.4 (2CH, Ph), 128.2 (3CH, Ph), 128.1 (2CH, Ph), 127.7 (CH, Ph), 125.3 (CH, Ph), 123.7 (CH, NPhth), 123.4 (CH, NPhth), 84.7 (CH, C1), 79.8 (CH, C3), 77.7 (CH, C5),

74.6 (CH and CH₂, C4 and CH₂Ph), 74.0 (CH₂, CH₂Ph), 71.0 (CH₂, C6), 54.8 (CH, C2), 34.6 (C, *t*-Bu), 31.4 (3CH₃, *t*-Bu), 20.4 (CH₃, Me). IR ν (film, cm⁻¹): 3476 (O-H), 3059 and 3028 (=C-H), 2956, 2909 and 2865 (C-H), 1774 (N-C=O), 1713 (N-C=O). MS (ESI): m/z = 674 ([M+Na]⁺, 100%). HRMS (ESI): Calcd for C₃₉H₄₁NNaO₆S [M+Na]⁺: 674.2553. Found: 674.2576. Elementary Analysis: Calcd for C₃₉H₄₁NO₆S: C, 71.86; H, 6.34; N, 2.15; S, 4.92. Found: C, 71.45; H, 6.49; N, 2.22; S, 4.87.

(2-Methyl-5-*tert*-butylphenyl)

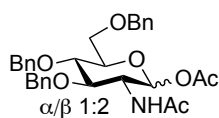
(2-acetamido-3,4,6-tri-*O*-acetyl-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 4)-3,6-di-*O*-benzyl-2-deoxy-2-phthalimido-1-thio- β -D-glucopyranoside **24**



Acceptor **23** (417 mg, 0.640 mmol, 5 eq.) and Fe(OTf)₃·6.2DMSO (62 mg, 0.063 mmol, 50 mol%) were added to donor **1b** (50 mg, 0.128 mmol, 1 eq.) in an oven-dried, argon-purged, microwave vial equipped with a magnetic stirring bar. Everything is flushed under argon and dry CH₂Cl₂ is added (1 mL). After sealing the vial, the reaction

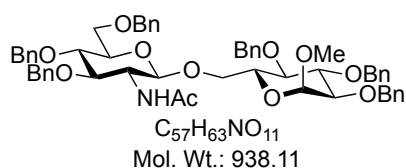
mixture is heated to 70 °C under microwave irradiation for 3 hours (CEM Discover instrument). Then, the reaction mixture is diluted in CH₂Cl₂ (20 mL) and washed with a saturated aqueous solution of NaHCO₃ (20 mL). The aqueous layer is extracted with CH₂Cl₂ (4×20 mL) and the combined organic layers are washed with brine (20 mL), dried over Na₂SO₄, filtered and concentrated under reduced pressure. The crude product was purified by chromatography on silica gel (heptane/EtOAc 5:5 to 0:1) to afford pure product **24** (30 mg, 25 %, white amorphous solid). [α]_D²⁵: +44.1 (*c* 0.5, CHCl₃). ¹H NMR (500 MHz, CDCl₃) δ : 7.78-7.70 (d, *J* = 7.0 Hz, 1H, NPhth), 7.68-7.57 (m, 2H, NPhth), 7.55-7.38 (m, 7H, NPhth and Ph), 7.12 (d, *J* = 7.5 Hz, 1H, Ph), 6.99 (d, *J* = 7.5 Hz, 1H, Ph), 6.94 (d, *J* = 7.5 Hz, 2H, Ph), 6.80-6.68 (m, 3H, Ph), 5.35 (d, *J*_{A1,A2} = 10.0 Hz, 1H, HA-1), 4.96 (t, *J*_{B4,B3} = *J*_{B4,B5} = 9.5 Hz, 1H, HB-4), 4.92-4.82 (m, 2H, CH₂Ph and HB-3), 4.75 (d, *J* = 12.5 Hz, 1H, CH₂Ph), 4.68 (d, *J*_{NH,B2} = 9.5 Hz, 1H, NH), 4.47 (d, *J*_{B1,B2} = 8.5 Hz, 1H, HB-1), 4.38 (d, *J* = 12.0 Hz, 1H, CH₂Ph), 4.35 (d, *J* = 12.5 Hz, 1H, CH₂Ph), 4.25-4.18 (m, 3H, HB-6, HA-2 and HA-3), 4.04-3.92 (m, 3H, HB-2, HB-6' and HA-4), 3.67 (dd, *J*_{A6,A6'} = 10.5 Hz, *J*_{A6,A5} = 2.0 Hz, 1H, HA-6), 3.59 (d, *J*_{A6',A6} = 10.5 Hz, *J*_{A6',A5} = 1.0 Hz, 1H, HA-6'), 3.56-3.46 (m, 2H, HB-5 and HA-5), 2.10 (s, 3H, Me), 2.00 (s, 3H, Ac), 1.97 (s, 3H, Ac), 1.94 (s, 3H, Ac), 1.70 (s, 3H, Ac), 1.23 (s, 9H, *t*-Bu). ¹³C NMR (75 MHz, CDCl₃) δ : 171.0 (C, CO), 170.9 (C, CO), 169.9 (C, CO), 169.5 (C, CO), 167.9 (C, NCO), 167.4 (C, NCO), 149.5 (C, Ph), 138.8 (C, Ph), 137.8 (C, Ph), 137.2 (C, Ph), 133.9 (CH, NPhth), 133.7 (CH, NPhth), 131.9 (C, Ph), 131.8 (2C, NPhth), 130.6 (CH, Ph), 130.0 (CH, Ph), 129.3 (2CH, Ph), 129.2 (2CH, Ph), 129.1 (CH, Ph), 128.1 (2CH, Ph), 128.0 (2CH, Ph), 127.1 (CH, Ph), 125.3 (CH, Ph), 123.5 (CH, NPhth), 123.4 (CH, NPhth), 101.0 (CH, CB-1), 84.6 (CH, CA-1), 78.9 (CH, CA-4), 78.5 (CH, CA-5), 78.2 (CH, CA-3), 75.0 (CH₂, CH₂Ph), 74.2 (CH₂, CH₂Ph), 73.3 (CH, CB-3), 71.6 (CH, CB-5), 68.8 (CH, CB-4), 68.0 (CH₂, CA-6), 62.1 (CH₂, CB-6), 55.2 (CH, CA-2), 54.4 (CH, CB-2), 34.6 (C, *t*-Bu), 31.4 (3CH₃, *t*-Bu), 23.3 (CH₃, Ac), 20.8 (3CH₃, 2Ac and Me), 20.4 (CH₃, Ac). IR ν (film, cm⁻¹): 3288 (N-H), 2944, 2927 and 2861 (C-H), 1778 (N-C=O), 1746 (C=O), 1714 (N-C=O), 1663 (NH-C=O). MS (ESI): m/z = 1003 ([M+Na]⁺, 100%). HRMS (ESI): Calcd for C₅₃H₆₀N₂NaO₁₄S [M+Na]⁺: 1003.3663. Found: 1003.3691. Elementary Analysis: Calcd for C₅₃H₆₀N₂O₁₄S: C, 64.88; H, 6.16; N, 2.86; S, 3.27. Found: C, 65.27; H, 6.56; N, 2.51; S, 2.89.

2-Acetamido-1-*O*-acetyl-3,4,6-tri-*O*-benzyl-2-deoxy- α/β -D-glucopyranose **29**



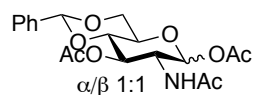
2-acetamido-3,4,6-tri-*O*-benzyl-2-deoxy-D-glucopyranose²² (510 mg, 1.03 mmol, 1 eq.) and pyridine hydrochloride (153 mg, 1.32 mmol, 1.3 eq.) in pyridine (5 mL) are stirred at 100 °C for 1 hour under argon. To this solution, acetic anhydride (255 μ L, 2.70 mmol, 2.6 eq.) is added and the reaction mixture is stirred at room temperature for 8 hours. The volatiles are evaporated under reduced pressure and the crude product is purified by chromatography on silica gel (heptane/EtOAc 6:4 to 0:1) to afford pure product **29**²³ as a mixture of two anomers (470 mg, 88 %, α/β 1:2, white amorphous solid).

Methyl (2-acetamido-3,4,6-tri-*O*-benzyl-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 6)-2,3,4-tri-*O*-benzyl- α -D-glucopyranoside **30**

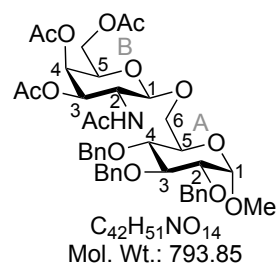


30²⁴ was obtained under microwave conditions using donor **29** (69 mg, 0.129 mmol, 2 eq.), TTBP (32 mg, 0.129 mmol, 2 eq.), $Fe(OTf)_3 \cdot 6.2DMSO$ (9 mg, 0.010 mmol, 15 mol-%) and acceptor **32** (30 mg, 0.065 mmol, 1 eq.) in dry CH_2Cl_2 (1 mL), according to general procedure A (CEM Discover instrument, 80 °C, 45 min) (62 mg, 86 %, white amorphous solid).

2-Acetamido-1,3-di-*O*-acetyl-4,6-*O*-benzylidene-2-deoxy- α/β -D-glucopyranose **32**



N-acetyl-D-glucosamine (5 g, 22.6 mmol, 1 eq.), benzaldehyde (13.7 mL, 135.6 mmol, 6 eq.) and $ZnCl_2$ (3.1 g, 22.6 mmol, 1 eq.) were stirred at room temperature for 12 hours under argon. The precipitate was filtered off, washed with petroleum ether (2 \times 40 mL), washed with water (2 \times 40 mL) and dried over reduced pressure. The crude 4,6-*O*-benzylidene-D-glucosamine (6.99 g, quant.) was used in the next step without further purification. This intermediate (500 mg, 1.62 mmol, 1 eq.) and pyridine hydrochloride (243 mg, 2.10 mmol, 1.3 eq.) in pyridine (5 mL) were stirred at 100 °C for 1 hour. To this solution, acetic anhydride (772 μ L, 8.24 mmol, 5.1 eq.) was added and the reaction mixture was stirred at room temperature for 8 hours.²⁵ The volatiles were evaporated under reduced pressure and the crude product was purified by chromatography on silica gel (heptane/EtOAc 6:4 to 0:1) to afford pure product **32**²⁶ (374 mg, 59 %, α/β 1:1, white amorphous solid).



Methyl (2-acetamido-3,4,6-tri-*O*-acetyl-2-deoxy- β -D-galactopyranosyl)-(1 \rightarrow 6)-2,3,4-tri-*O*-benzyl- α -D-glucopyranoside **34**

34 was obtained under microwave conditions using donor **33** (50 mg, 0.128 mmol, 2 eq.), TTBP (32 mg, 0.129 mmol, 2 eq.), $Fe(OTf)_3 \cdot 6.2DMSO$ (10 mg, 0.010 mmol, 15 mol-%) and acceptor **3**² (30 mg, 0.065 mmol, 1 eq.) in dry CH_2Cl_2 (1 mL), according to general procedure A (Anton Paar Monowave 300 instrument, 110 °C, 30 min) (49 mg, 95 %, amorphous white solid). $[\alpha]_D^{20}$: +1.70 (*c* 1.0 in $CHCl_3$). 1H NMR (300 MHz, $CDCl_3$) δ : 7.38-7.28 (m, 15H, 3Ph), 5.33 (dd, $J_{B4,B3}$ = 3.5 Hz and $J_{B4,B5}$ = 1.0 Hz, 1H, HB-4), 5.28 (d, $J_{NH,B2}$ =

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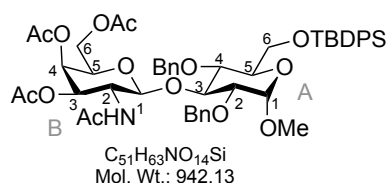
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²⁵ P. Rollin, P. Sinaÿ, *J. Chem. Soc., Perkin Trans. 1* **1977**, 2513-2517.

²⁶ F. Dasgupta, L. Anderson, *Carbohydr. Res.* **1990**, 202, 239-255.

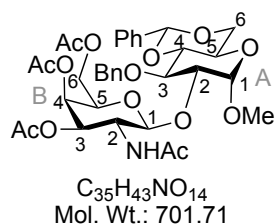
9.0 Hz, 1H, NH), 5.26 (dd, $J_{B3,B2} = 11.0$ Hz and $J_{B3,B4} = 3.5$ Hz, 1H, HB-3), 4.99 (d, $J = 11.0$ Hz, 1H, CH_2Ph), 4.85 (d, $J = 11.0$ Hz, 1H, CH_2Ph), 4.80 (d, $J = 11.0$ Hz, 1H, CH_2Ph), 4.79 (d, $J = 12.0$ Hz, 1H, CH_2Ph), 4.67 (d, $J_{B1,B2} = 8.5$ Hz, 1H, HB-1), 4.65 (d, $J = 12.0$ Hz, 1H, CH_2Ph), 4.60 (d, $J_{A1,A2} = 3.5$ Hz, 1H, HA-1), 4.57 (d, $J = 11.0$ Hz, 1H, CH_2Ph), 4.11-4.06 (m, 3H, HB-6, HB-6' and HA-6), 4.04-3.94 (m, 2H, HA-3 and HB-2), 3.87 (dt, $J_{B5,B6} = J_{B5,B6'} = 6.5$ Hz and $J_{B5,B4} = 1.0$ Hz, 1H, HB-5), 3.80-3.75 (m, 1H, HA-5), 3.72 (dd, $J_{A6',A6} = 10.5$ Hz and $J_{A6',A5} = 4.0$ Hz, 1H, HA-6'), 3.53 (dd, $J_{A2,A3} = 9.5$ Hz and $J_{A2,A1} = 3.5$ Hz, 1H, HA-2), 3.48 (t, $J_{A4,A3} = J_{A4,A5} = 9.5$ Hz, 1H, HA-4), 3.37 (s, 3H, OCH_3), 2.10 (s, 3H, $COCH_3$), 2.00 (s, 3H, $COCH_3$), 1.99 (s, 3H, $COCH_3$), 1.83 (s, 3H, $COCH_3$). ^{13}C NMR (75 MHz, $CDCl_3$) δ : 170.8 (C, $COCH_3$), 170.8 (C, $COCH_3$), 170.6 (C, $COCH_3$), 170.4 (C, $COCH_3$), 139.1 (C, Ph), 138.7 (C, Ph), 138.5 (C, Ph), 128.9 (CH, Ph), 128.8 (CH, Ph), 128.8 (CH, Ph), 128.5 (CH, Ph), 128.3 (CH, Ph), 128.3 (CH, Ph), 128.3 (CH, Ph), 128.1 (CH, Ph), 128.0 (CH, Ph), 101.3 (CH, CB-1), 98.4 (CH, CA-1), 82.4 (CH, CA-3), 80.1 (CH, CA-2), 77.8 (CH, CA-4), 76.1 (CH_2 , CH_2Ph), 75.0 (CH_2 , CH_2Ph), 73.7 (CH_2 , CH_2Ph), 71.1 (CH, CB-5), 70.2 (CH, CB-3), 69.9 (CH, CA-5), 68.1 (CH_2 , CA-6), 67.1 (CH, CB-4), 61.7 (CH_2 , CB-6), 55.6 (CH_3 , OCH_3), 51.8 (CH, CB-2), 23.8 (CH_3 , $COCH_3$), 21.1 (CH_3 , $COCH_3$), 21.0 (CH_3 , $COCH_3$). IR ν (film, cm^{-1}): 3286 (N-H), 3092, 3065 and 3031 ($=C-H$), 2921 (C-H), 1743 (C=O), 1661 (NH-C=O). MS (ESI): $m/z = 794$ ($[M+H]^+$, 85%), 816 ($[M+Na]^+$, 100%). HRMS (ESI): Calcd for $C_{42}H_{52}NO_{14}$ $[M+H]^+$: 794.3388. Found: 794.3381.

Methyl (2-acetamido-3,4,6-tri-*O*-acetyl-2-deoxy- β -D-galactopyranosyl)-(1 \rightarrow 3)-2,4-di-*O*-benzyl-6-*O*-*tert*-butyldiphenylsilyl- α -D-glucopyranoside **35**



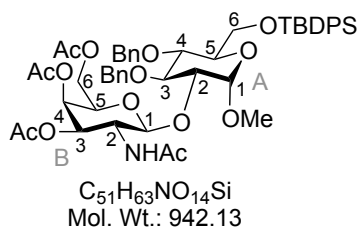
35 was obtained under microwave conditions using donor **33** (50 mg, 0.128 mmol, 2 eq.), TTBP (32 mg, 0.129 mmol, 2 eq.), $Fe(OTf)_3 \cdot 6.2DMSO$ (10 mg, 0.010 mmol, 15 mol-%) and acceptor **13** (40 mg, 0.065 mmol, 1 eq.) in dry CH_2Cl_2 (1 mL), according to general procedure A (Anton Paar Monowave 300 instrument, 110 $^{\circ}C$, 30 min) (46 mg, 75 %, white amorphous solid). $[\alpha]_D^{20}$: +16.1 (c 1.0 in $CHCl_3$). 1H NMR (300 MHz, $CDCl_3$) δ : 7.72-7.68 (m, 4H, Ph), 7.48-7.33 (m, 13H, Ph), 7.28-7.26 (m, 3H, Ph), 5.30 (br d, $J_{B4,B3} = 3.0$ Hz, 1H, HB-4), 5.06 (d, $J = 10.5$ Hz, 1H, CH_2Ph), 5.02 (d, $J_{NH,B2} = 9.5$ Hz, 1H, NH), 4.94 (d, $J_{B1,B2} = 8.5$ Hz, 1H, HB-1), 4.93 (dd, $J_{B3,B2} = 11.0$ Hz and $J_{B3,B4} = 3.0$ Hz, 1H, HB-3), 4.80 (d, $J = 12.0$ Hz, 1H, CH_2Ph), 4.75 (d, $J_{A1,A2} = 3.5$ Hz, 1H, HA-1), 4.58 (d, $J = 12.0$ Hz, 1H, CH_2Ph), 4.48 (d, $J = 10.5$ Hz, 1H, CH_2Ph), 4.26 (t, $J_{A3,A2} = J_{A3,A4} = 9.0$ Hz, 1H, HA-3), 4.22-4.09 (m, 2H, HB-6 and HB-2), 3.94 (dd, $J_{B6',B6} = 11.0$ Hz and $J_{B6',B5} = 6.0$ Hz, 1H, HB-6'), 3.87-3.82 (m, 3H, HA-6, HA-6' and HB-5), 3.71-3.65 (m, 1H, HA-5), 3.57-3.50 (m, 2H, HA-2 and HA-4), 3.35 (s, 3H, OCH_3), 2.11 (s, 3H, $OCOCH_3$), 1.99 (s, 3H, $OCOCH_3$), 1.96 (s, 3H, $OCOCH_3$), 1.71 (s, 3H, $NHCOCH_3$), 1.07 (s, 9H, $C(CH_3)_3$). ^{13}C NMR (75 MHz, $CDCl_3$) δ : 171.0 (C, $COCH_3$), 170.7 (C, $COCH_3$), 170.7 (C, $COCH_3$), 170.4 (C, $COCH_3$), 138.8 (C, Ph), 138.4 (C, Ph), 136.1 (CH, Ph), 136.0 (CH, Ph), 133.9 (C, Ph), 133.7 (C, Ph), 130.0 (CH, Ph), 129.9 (CH, Ph), 129.4 (CH, Ph), 128.8 (CH, Ph), 128.5 (CH, Ph), 128.0 (CH, Ph), 127.9 (CH, Ph), 127.5 (CH, Ph), 102.2 (CH, CB-1), 97.1 (CH, CA-1), 81.7 (CH, CA-2), 79.8 (CH, CA-3), 76.1 (CH, CA-4), 75.2 (CH_2 , CH_2Ph), 72.7 (CH_2 , CH_2Ph), 71.8 (CH, CA-5), 71.5 (CH, CB-3), 70.9 (CH, CB-5), 66.9 (CH, CB-4), 63.4 (CH_2 , CA-6), 61.4 (CH_2 , CB-6), 55.2 (CH_3 , OCH_3), 51.6 (CH, CB-2), 27.2 ($3CH_3$, $C(CH_3)_3$), 23.6 (CH_3 , $NHCOCH_3$), 21.1 (CH_3 , $OCOCH_3$), 21.1 (CH_3 , $OCOCH_3$), 19.7 (C, $C(CH_3)_3$). IR ν (film, cm^{-1}): 3285 (N-H), 2958, 2929, 2899 and 2861 (C-H), 1750 (C=O), 1661 (NH-C=O). MS (ESI): $m/z = 942$ ($[M+H]^+$, 35%), 964 ($[M+Na]^+$, 100%). HRMS (ESI): Calcd for $C_{51}H_{63}NNaO_{14}Si$ $[M+Na]^+$: 964.3916. Found: 964.3917.

Methyl (2-acetamido-3,4,6-tri-*O*-acetyl-2-deoxy- β -D-galactopyranosyl)-(1 \rightarrow 2)-3-*O*-benzyl-4,6-*O*-benzylidene- α -D-glucopyranoside **36**



36 was obtained under microwave conditions using donor **33** (46 mg, 0.118 mmol, 2 eq.), TTBP (29 mg, 0.117 mmol, 2 eq.), $Fe(OTf)_3 \cdot 6.2DMSO$ (9 mg, 0.009 mmol, 15 mol-%) and acceptor **15** (22 mg, 0.059 mmol, 1 eq.) in dry CH_2Cl_2 (1 mL), according to general procedure A (Anton Paar Monowave 300 instrument, 110 °C, 60 min) (26 mg, 63 %, amorphous white solid). $[\alpha]_D^{20}$: +5.43 (c 0.7 in $CHCl_3$). 1H NMR (300 MHz, $CDCl_3$) δ : 7.47-7.44 (m, 2H, Ph), 7.38-7.29 (m, 8H, Ph), 5.56 (s, 1H, $CHPh$), 5.36-5.29 (m, 2H, HB-3 and HB-4), 5.06 (d, $J_{NH,B2}$ = 8.5 Hz, 1H, NH), 5.00 (d, $J_{B1,B2}$ = 8.5 Hz, 1H, HB-1), 4.96 (d, J = 12.0 Hz, 1H, CH_2Ph), 4.87 (d, $J_{A1,A2}$ = 3.5 Hz, 1H, HA-1), 4.67 (d, J = 12.0 Hz, 1H, CH_2Ph), 4.30 (dd, $J_{A6,A6'}$ = 10.0 Hz and $J_{A6,A5}$ = 4.5 Hz, 1H, HA-6), 4.14 (d, $J_{B6,B5}$ = $J_{B6',B5}$ = 6.5 Hz, 2H, HB-6 and HB-6'), 4.04 (t, $J_{A3,A2}$ = $J_{A3,A4}$ = 9.5 Hz, 1H, HA-3), 3.96-3.83 (m, 3H, HB-2, HB-5, HA-5), 3.78-3.71 (m, 1H, HA-6'), 3.71-3.66 (dd, $J_{A2,A3}$ = 9.5 Hz and $J_{A2,A1}$ = 3.5 Hz, 1H, HA-2), 3.62 (t, $J_{A4,A3}$ = $J_{A4,A5}$ = 9.5 Hz, 1H, HA-4), 3.43 (s, 3H, OCH_3), 2.15 (s, 3H, $COCH_3$), 2.06 (s, 3H, $COCH_3$), 1.97 (s, 3H, $COCH_3$), 1.49 (s, 3H, $COCH_3$). ^{13}C NMR (75 MHz, $CDCl_3$) δ : 170.9 (C, $COCH_3$), 170.8 (C, $COCH_3$), 170.5 (C, $COCH_3$), 137.7 (C, Ph), 129.4 (CH, Ph), 128.9 (CH, Ph), 128.6 (CH, Ph), 128.1 (CH, Ph), 127.6 (CH, Ph), 126.4 (CH, Ph), 102.5 (CH, $CHPh$), 101.8 (CH, CB-1), 100.4 (CH, CA-1), 82.9 (CH, CA-4), 81.4 (CH, CA-2), 78.0 (CH, CA-3), 75.2 (CH_2 , CH_2Ph), 71.3 (CH, CB-5), 70.2 (CH, CB-3), 69.5 (CH_2 , CA-6), 67.2 (CH, CB-4), 62.6 (CH, CA-5), 62.2 (CH_2 , CB-6), 55.8 (CH_3 , OCH_3), 52.5 (CH, CB-2), 23.4 ($COCH_3$), 21.1 ($COCH_3$), 21.0 ($COCH_3$). IR ν (film, cm^{-1}): 3288 (N-H), 3095, 3071 and 3033 (=C-H), 2921 and 2851 (C-H), 1745 (C=O), 1661 (NH-C=O). MS (ESI): m/z = 702 ($[M+H]^+$, 100%), 724 ($[M+Na]^+$, 85%), 1425 ($[2M+Na]^+$, 90%). HRMS (ESI): Calcd for $C_{35}H_{44}NO_{14}$ $[M+H]^+$: 702.2762. Found: 702.2774.

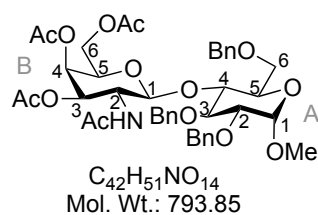
Methyl (2-acetamido-3,4,6-tri-*O*-acetyl-2-deoxy- β -D-galactopyranosyl)-(1 \rightarrow 2)-3,4-di-*O*-benzyl-6-*O*-*tert*-butyldiphenylsilyl- α -D-glucopyranoside **37**



37 was obtained under microwave conditions using donor **33** (50 mg, 0.128 mmol, 2 eq.), TTBP (32 mg, 0.129 mmol, 2 eq.), $Fe(OTf)_3 \cdot 6.2DMSO$ (10 mg, 0.010 mmol, 15 mol-%) and acceptor **17** (40 mg, 0.065 mmol, 1 eq.) in dry CH_2Cl_2 (1 mL), according to general procedure A (Anton Paar Monowave 300 instrument, 110 °C, 45 min) (34 mg, 55 %, amorphous white solid). $[\alpha]_D^{20}$: +18.8 (c 1.0 in $CHCl_3$). 1H NMR (300 MHz, $CDCl_3$) δ : 7.73-7.68 (m, 4H, Ph), 7.43-7.27 (m, 11H, Ph), 7.23-7.19 (m, 3H, Ph), 7.07-7.04 (m, 2H, Ph), 5.42 (dd, $J_{B3,B2}$ = 10.5 Hz and $J_{B3,B4}$ = 3.5 Hz, 1H, HB-3), 5.37 (dd, $J_{B4,B3}$ = 3.5 Hz and $J_{B4,B5}$ = 1.0 Hz, 1H, HB-4), 5.12 (d, $J_{NH,B2}$ = 8.0 Hz, 1H, NH), 5.06 (d, $J_{B1,B2}$ = 8.5 Hz, 1H, HB-1), 4.91 (d, $J_{A1,A2}$ = 3.5 Hz, 1H, HA-1), 4.86 (d, J = 2.0 Hz, 2H, CH_2Ph), 4.79 (d, J = 10.5 Hz, 1H, CH_2Ph), 4.59 (d, J = 10.5 Hz, 1H, CH_2Ph), 4.18-4.16 (m, 2H, HB-6 and HB-6'), 4.02-3.97 (m, 2H, HB-5, HA-3), 3.93-3.84 (m, 3H, HB-2, HA-6 and HA-6'), 3.73-3.62 (m, 3H, HA-2, HA-4 and HA-5), 3.38 (s, 3H, OCH_3), 2.15 (s, 3H, $COCH_3$), 2.07 (s, 3H, $COCH_3$), 1.97 (s, 3H, $COCH_3$), 1.42 (s, 3H, $COCH_3$), 1.07 (s, 9H, $C(CH_3)_3$). ^{13}C NMR (75 MHz, $CDCl_3$) δ : 170.9 (C, $COCH_3$), 170.8 (C, $COCH_3$), 170.7 (C, $COCH_3$), 170.5 (C, $COCH_3$), 139.5 (C, Ph), 138.4 (C, Ph), 136.2 (CH, Ph), 136.0 (CH, Ph), 133.9 (C, Ph), 133.6 (C, Ph), 130.0 (CH, Ph), 130.0 (CH, Ph), 128.9 (CH, Ph), 128.7 (CH, Ph), 128.3 (CH, Ph), 128.1 (CH, Ph), 128.0 (CH, Ph), 127.9 (CH, Ph), 127.1 (CH, Ph), 102.2 (CH, CB-1), 99.3 (CH, CA-1), 82.9 (CH, CA-2), 81.3 (CA-3), 78.4 (CH, CA-4), 75.5 (CH_2 , CH_2Ph), 75.5 (CH_2 , CH_2Ph),

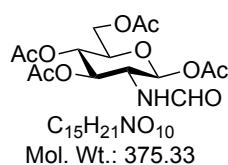
71.6 (CH, CA-5), 71.2 (CH, CB-5), 69.9 (CH, CB-3), 67.2 (CH, CB-4), 63.1 (CH₂, CA-6), 62.3 (CH₂, CB-6), 55.2 (CH₃, OCH₃), 52.7 (CH, CB-2), 27.2 (CH₃, C(CH₃)₃), 23.3 (CH₃, COCH₃), 21.1 (CH₃, COCH₃), 21.0 (CH₃, COCH₃), 19.7 (C, C(CH₃)₃). IR ν (film, cm⁻¹): 3280 (N-H), 3068 and 3029 (=C-H), 2931 and 2858 (C-H), 1747 (C=O), 1656 (NH-C=O). MS (ESI): m/z = 964 ([M+Na]⁺, 100%). HRMS (ESI): Calcd for C₅₁H₆₃NNaO₁₄Si [M+Na]⁺: 964.3916. Found: 964.3915.

Methyl (2-acetamido-3,4,6-tri-*O*-acetyl-2-deoxy- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-*O*-benzyl- α -D-glucopyranoside **38**



38 was obtained under microwave conditions using donor **33** (50 mg, 0.128 mmol, 2 eq.), TTBP (32 mg, 0.129 mmol, 2 eq.), Fe(OTf)₃·6.2DMSO (10 mg, 0.010 mmol, 15 mol-%) and acceptor **19** (29 mg, 0.064 mmol, 1 eq.) in dry CH₂Cl₂ (1 mL), according to general procedure A (Anton Paar Monowave 300 instrument, 110 °C, 3 hours) (13 mg, 26 %, amorphous white solid). $[\alpha]_D^{20}$: -27.2 (*c* 0.7 in CHCl₃). ¹H NMR (500 MHz, CDCl₃) δ : 7.48-7.24 (m, 15H, Ph), 5.21 (dd, J = 3.0 Hz, 1H, HB-4), 4.97 (d, J = 11.0 Hz, 1H, CH₂Ph), 4.84 (d, J = 12.0 Hz, 1H, CH₂Ph), 4.81-4.76 (m, 3H, CH₂Ph, HB-3), 4.63 (d, J = 12.0 Hz, 1H, CH₂Ph), 4.57 (d, $J_{A1,A2}$ = 4.0 Hz, 1H, HA-1), 4.40 (d, $J_{B1,B2}$ = 8.5 Hz, 1H, HB-1), 4.38 (d, $J_{NH,B2}$ = 10.0 Hz, 1H, NH), 4.36 (d, J = 12.0 Hz, 1H, CH₂Ph), 3.97 (dt, $J_{B2,NH}$ = 10.0 Hz and $J_{B2,B1}$ = $J_{B2,B3}$ = 8.5 Hz, 1H, HB-2), 3.91-3.81 (m, 4H, HA-3, HA-4, HB-6, HB-6'), 3.66-3.58 (m, 3H, HA-6, HA-5, HB-5), 3.50 (dd, $J_{A6',A6}$ = 10.5 Hz and $J_{A6',A5}$ = 2.0 Hz, 1H, HA-6'), 3.49-3.47 (m, 1H, HA-2), 3.37 (s, 3H, OCH₃), 2.06 (s, 3H, OCOCH₃), 2.00 (s, 3H, OCOCH₃), 1.96 (s, 3H, OCOCH₃), 1.74 (s, 3H, NHCOCH₃). ¹³C NMR (125 MHz, CDCl₃) δ : 170.8 (C, COCH₃), 170.6 (C, COCH₃), 170.6 (C, COCH₃), 170.1 (C, COCH₃), 139.9 (C, Ph), 138.7 (C, Ph), 138.2 (C, Ph), 129.5 (CH, Ph), 129.3 (CH, Ph), 129.2 (CH, Ph), 128.8 (CH, Ph), 128.5 (CH, Ph), 128.5 (CH, Ph), 128.2 (CH, Ph), 127.7 (CH, Ph), 127.6 (CH, Ph), 101.0 (CH, CB-1), 98.9 (CH, CA-1), 80.2 (CH, CA-3), 79.2 (CH, CA-2), 77.1 (CH, CA-4), 75.4 (CH₂, CH₂Ph), 74.2 (CH₂, CH₂Ph), 74.0 (CH₂, CH₂Ph), 70.8 (CH, CB-3), 70.6 (CH, CB-5), 69.8 (CH, CA-5), 67.9 (CH₂, CA-6), 66.5 (CH, CB-4), 61.2 (CH₂, CB-6), 55.7 (CH₃, OCH₃), 51.6 (CH, CB-2), 23.7 (CH₃, NHCOCH₃), 21.1 (CH₃, OCOCH₃), 21.0 (CH₃, OCOCH₃), 21.0 (CH₃, OCOCH₃). IR ν (film, cm⁻¹): 3326 (N-H), 3089, 3062 and 3030 (=C-H), 2918 (C-H), 1749 (C=O), 1670 (NH-C=O). MS (ESI): m/z = 794 ([M+H]⁺, 100%), 816 ([M+Na]⁺, 50%). HRMS (ESI): Calcd for C₄₂H₅₂NO₁₄ [M+H]⁺: 794.3388. Found: 794.3399.

1,3,4,6-Tetra-*O*-acetyl-2-deoxy-2-formamido- β -D-glucopyranose **39**



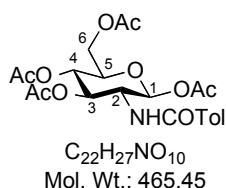
To a solution of 1,3,4,6-tetra-*O*-acetyl-2-amino-2-deoxy- β -D-glucopyranose hydrochloride⁸ (1 g, 2.61 mmol) in a 1:1 CH₂Cl₂-saturated aqueous solution of NaHCO₃ (40 mL) was added dropwise acetoformic anhydride²⁷ (495 μ L, 7.82 mmol, 3 eq.) at 0 °C.

The reaction was then warmed to room temperature and stirred at this temperature for 3 hours and then the organic layer was separated and the aqueous layer was extracted with CH₂Cl₂. The combined organic layers were dried over Na₂SO₄ and evaporated under reduced pressure. The crude product was purified by chromatography on silica gel

²⁷ L. I. Krimen, J. Savage, P. Yates, *Org. Synth.* **1970**, *50*, 1-1.

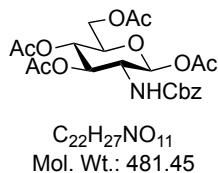
(heptane/EtOAc 1:1 to 0:1) to give the desired product **39**²⁸ (912 mg, 93 %, white amorphous solid).

1,3,4,6-Tetra-*O*-acetyl-2-deoxy-2-(4-methylbenzamido)- β -D-glucopyranose **40**



To a solution of 1,3,4,6-tetra-*O*-acetyl-2-amino-2-deoxy- β -D-glucopyranose hydrochloride⁸ (1 g, 2.61 mmol) in pyridine (11 mL) was added dropwise 4-methylbenzoyl chloride (2.1 mL, 15.6 mmol, 6 eq.) at 0 °C under argon. The reaction was warmed to room temperature and stirred at this temperature for 8 hours. Then, the reaction mixture was poured into water (30 mL), extracted with CH₂Cl₂ (3×40 mL) and the combined organic layers were washed with brine (10 mL), dried over Na₂SO₄ and evaporated under reduced pressure. The crude product was purified by chromatography on silica gel (heptane/EtOAc 1:1 to 0:1) to afford pure product **40** (910 mg, 75 %, white solid). Mp: 205.4-210.3 °C (from EtOAc). [α]_D²²: +42.8 (*c* 1.0, CHCl₃). ¹H NMR (500 MHz, CDCl₃) δ : 7.56 (d, *J* = 8.0 Hz, 2H, Ph), 7.20 (d, *J* = 8.0 Hz, 2H, Ph), 6.11 (br d, *J*_{NH,2} = 9.5 Hz, 1H, NH), 5.78 (d, *J*_{1,2} = 8.5 Hz, 1H, H1), 5.27-5.17 (m, 2H, H3 and H4), 4.59-4.48 (m, 1H, H2), 4.28 (d, *J*_{6,6'} = 12.0 Hz, *J*_{6,5} = 4.5 Hz, 1H, H6), 4.14 (d, *J*_{6',6} = 12.0 Hz, *J*_{6',5} = 2.0 Hz, 1H, H6'), 3.83 (ddd, *J*_{5,4} = 9.5 Hz, *J*_{5,6} = 4.5 Hz, *J*_{5,6'} = 2.0 Hz, 1H, H5), 2.37 (s, 3H, Me), 2.09 (s, 3H, Ac), 2.05 (s, 3H, Ac), 2.04 (s, 3H, Ac), 1.96 (s, 3H, Ac). ¹³C NMR (75 MHz, CDCl₃) δ : 171.8 (C, CO), 170.9 (C, CO), 169.8 (C, CO), 169.5 (C, CO), 167.4 (C, CO), 142.7 (C, Ph), 130.9 (C, Ph), 129.6 (2CH, Ph), 127.1 (2CH, Ph), 93.1 (CH, C1), 73.4 (CH, C5), 73.0 (CH, C3), 68.0 (CH, C4), 62.0 (CH₂, C6), 53.4 (CH, C2), 21.6 (CH₃, Ac), 21.1 (CH₃, Ac), 21.0 (CH₃, Ac), 20.8 (2CH₃, 2Ac). IR ν (film, cm⁻¹): 3027 (N-H), 2952 (C-H), 1750 (C=O), 1647 (NH-C=O). MS (ESI): *m/z* = 406 ([M-Ac]⁺, 100%), 488 ([M+Na]⁺, 25%). HRMS (ESI): Calcd for C₂₂H₂₇NNaO₁₀ [M+Na]⁺: 488.1533. Found: 488.1529.

1,3,4,6-Tetra-*O*-acetyl-2-benzyloxycarbonylamino-2-deoxy- β -D-glucopyranose **41**

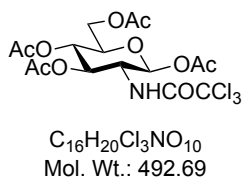


To a solution 1,3,4,6-tetra-*O*-acetyl-2-amino-2-deoxy- β -D-glucopyranose hydrochloride⁸ (10 g, 26.1 mmol) in a 1:2 CH₂Cl₂-saturated aqueous solution of NaHCO₃ (380 mL) was added dropwise CbzCl (4.1 mL, 28.6 mmol, 1.1 eq.). The reaction was stirred at room temperature for 2 hours and then the organic layer was separated and the aqueous layer was extracted with CH₂Cl₂. The combined organic layers were washed with brine, dried over Na₂SO₄ and evaporated under reduced pressure. The crude product was purified by chromatography on silica gel (heptane/EtOAc 1:1 to 0:1) to give the desired product **41**²⁹ (10.5 g, 84 %, white amorphous solid).

²⁸ (a) C. G. Greig, D. H. Leaback, P. G. Walker, *J. Chem. Soc.* **1961**, 879-883; (b) D. H. R. Barton, G. Bringmann, G. Lamotte, W. B. Motherwell, R. S. Hay Motherwell, A. E. A. Porter, *J. Chem. Soc., Perkin Trans. I* **1980**, 2657-2664.

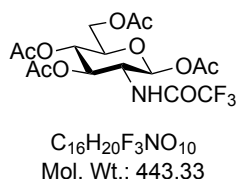
²⁹ (a) P. Boullanger, M. Jouineau, B. Bouammali, D. Lafont, G. Descotes, *Carbohydr. Res.* **1990**, 202, 151-164; (b) N. S. Simpkins, S. Stokes, A. J. Whittle, *J. Chem. Soc., Perkin Trans. I*, **1992**, 2471-2478.

1,3,4,6-Tetra-*O*-acetyl-2-deoxy-2-trichloroacetamido- β -D-glucopyranose 42



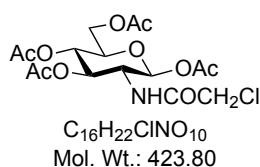
1,3,4,6-Tetra-*O*-acetyl-2-amino-2-deoxy- β -D-glucopyranose hydrochloride⁸ (10 g, 26.06 mmol) and trichloroacetic anhydride (14.3 mL, 78.2 mmol, 3 eq.) in pyridine (11 mL) were stirred at room temperature for 5 hours under argon. The volatiles were evaporated under reduced pressure and the crude product was purified by chromatography on silica gel (heptane/EtOAc 9:1 to 1:1) to afford pure product **42**³⁰ (12 g, 93 %, white amorphous solid).

1,3,4,6-Tetra-*O*-acetyl-2-deoxy-2-trifluoroacetamido- β -D-glucopyranose 43



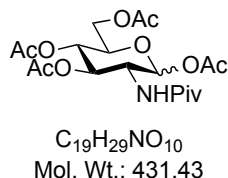
To a solution 1,3,4,6-tetra-*O*-acetyl-2-amino-2-deoxy- β -D-glucopyranose hydrochloride⁸ (1 g, 2.61 mmol) and pyridine (425 μ L, 5.21 mmol, 2 eq.) in dry CH_2Cl_2 (10 mL) was added dropwise trifluoroacetic anhydride (753 μ L, 3.91 mmol, 1.5 eq.) at 0 °C under argon. The reaction was then warmed to room temperature and stirred at this temperature for 5 hours. The volatiles were evaporated under reduced pressure and the crude product was purified by chromatography on silica gel (heptane/EtOAc 8:2 to 7:3) to give the desired product **43**³¹ (1.11 g, 96 %, white amorphous solid).

1,3,4,6-Tetra-*O*-acetyl-2-deoxy-2-chloroacetamido- β -D-glucopyranose 44



To a solution of 1,3,4,6-tetra-*O*-acetyl-2-amino-2-deoxy- β -D-glucopyranose hydrochloride⁸ (1 g, 2.61 mmol), DMAP (382 mg, 3.13 mmol, 1.2 eq.) and pyridine (850 μ L, 10.4 mmol, 4 eq.) in dry CH_2Cl_2 (10 mL) was added dropwise chloroacetyl chloride (415 μ L, 5.22 mmol, 2 eq.) at 0 °C under argon. The reaction was then warmed to room temperature and stirred at this temperature for 8 hours. The volatiles were evaporated under reduced pressure and the crude product was purified by chromatography on silica gel (heptane/EtOAc 8:2 to 7:3) to give the desired product **44**²⁶ (1.06 g, 96 %, white amorphous solid).

1,3,4,6-Tetra-*O*-acetyl-2-deoxy-2-pivalamido- α/β -D-glucopyranose 45

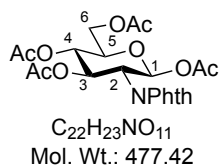


To a solution of 1,3,4,6-tetra-*O*-acetyl-2-amino-2-deoxy- β -D-glucopyranose hydrochloride⁸ (1 g, 2.61 mmol), DMAP (382 mg, 3.13 mmol, 1.2 eq.) and pyridine (850 μ L, 10.42 mmol, 4 eq.) in dry CH_2Cl_2 (10 mL) was added dropwise pivalic anhydride (1.06 mL, 5.21 mmol, 2 eq.) at 0 °C under argon. The reaction was then warmed to room temperature and stirred at this temperature for 2 days. The volatiles were evaporated under reduced pressure and the crude product was purified by chromatography on silica gel (heptane/EtOAc 1:1 to 0:1) to give the product **45** (392 mg, 35 %, α/β 1.5/1, white amorphous solid).

1,3,4,6-Tetra-*O*-acetyl-2-deoxy-2-phthalimido- β -D-glucopyranose 46

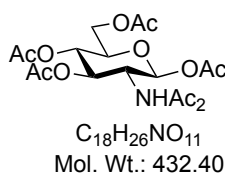
³⁰ G. Blatter, J.-M. Beau, J.-C. Jacquinet, *Carbohydr. Res.* **1994**, 260, 189-202.

³¹ (a) M. L. Wolfrom, H. B. Bhat, *J. Org. Chem.* **1967**, 32, 1821-1823; (b) I. R. Greig, M. S. Macauley, I. H. Williams, D. J. Vocadlo, *J. Am. Chem. Soc.* **2009**, 131, 13415-13422.



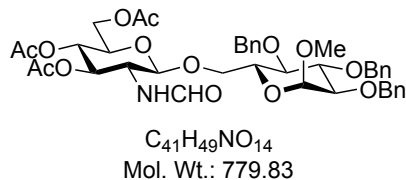
To a solution of D-glucosamine hydrochloride (40 g, 185.6 mmol, 1 eq.) in water (300 mL) are successively added phthalic anhydride (55 g, 372 mmol, 2 eq.) and NaHCO₃ (46 g, 548 mmol, 2.95 eq.). Then, the reaction mixture is stirred at 40 °C for 12 hours. The volatiles are evaporated under reduced pressure by coevaporation with toluene and the residue obtained was used without any further purification. Acetic anhydride (229 mL, 2445 mmol, 13.2 eq.) was added to a solution of crude intermediate in pyridine (350 mL) and the reaction mixture was stirred at 40 °C for 10 hours. The volatiles were evaporated under reduced pressure by coevaporation with toluene. The residue was diluted with CH₂Cl₂ (600 mL), washed with water (3×120 mL), brine (100 mL), dried over Na₂SO₄, filtered and evaporated under reduced pressure. The crude product was purified by chromatography on silica gel (heptane/EtOAc 7:3 to 3:7) to give the desired product as a mixture of two anomers (83 g, 93%, α/β: 1/2.3, white amorphous solid). This mixture was separated by chromatography on silica gel (toluene/acetone 95:05) to afford pure β-product **46**³² (white amorphous solid).

1,3,4,6-Tetra-*O*-acetyl-2-acetylacetamido-2-deoxy-β-D-glucopyranose **47**



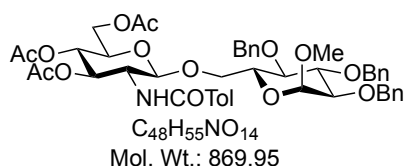
1β (1.2 g, 3.08 mmol) and TsOH monohydrate (276 mg, 1.45 mmol, 0.47 eq.) in isopropenylacetate (35 mL) were stirred at 65 °C for 4.5 hours under argon. Et₃N (8 mL) was added and the volatiles were evaporated under reduced pressure. The crude product was purified by chromatography on silica gel (heptane/EtOAc 8:2 to 1:1) to afford pure product **47**³³ (1.12 g, 84 %, white amorphous solid).

Methyl (3,4,6-tri-*O*-acetyl-2-deoxy-2-formamido-β-D-glucopyranosyl)-(1→6)-2,3,4-tri-*O*-benzyl-α-D-glucopyranoside **48**



48³⁴ was obtained under microwave conditions using donor **39** (48 mg, 0.129 mmol, 2 eq.), TTBP (32 mg, 0.129 mmol, 2 eq.), Fe(OTf)₃·6.2DMSO (10 mg, 0.010 mmol, 15 mol-%) and acceptor **3** (30 mg, 0.065 mmol, 1 eq.) in dry CH₂Cl₂ (1 mL), according to general procedure A (CEM Discover instrument, 80 °C, 45 min) (25 mg, 50 %, white amorphous solid).

Methyl (3,4,6-tri-*O*-acetyl-2-deoxy-2-(4-methylbenzamido)-β-D-glucopyranosyl)-(1→6)-2,3,4-tri-*O*-benzyl-α-D-glucopyranoside **49**



49⁴ was obtained under microwave conditions using donor **40** (60 mg, 0.129 mmol, 2 eq.), TTBP (32 mg, 0.129 mmol, 2 eq.), Fe(OTf)₃·6.2DMSO (10 mg, 0.010 mmol, 15 mol-%) and acceptor **32** (30 mg, 0.065 mmol, 1 eq.) in dry CH₂Cl₂ (1 mL), according to general procedure A (CEM Discover instrument, 80 °C, 45 min) (46 mg, 82 %, white amorphous solid).

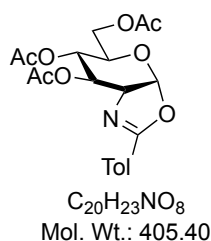
³² D. Macmillan, A. M. Daines, M. Bayrhuber, S. L. Flitsch, *Org. Lett.* **2002**, 4, 1467-1470.

³³ M. Suihko, M. Ahlgrén, P. Aulaskari, J. Rouvinen, *Carbohydr. Res.* **2001**, 334, 337-341.

³⁴ M. Trumtel, P. Tavecchia, A. Veyrières, P. Sinaÿ, *Carbohydr. Res.* **1989**, 191, 29-52.

2-(4-Methylphenyl) oxazoline **55**

(3,4,6-tri-*O*-acetyl-1,2-di-deoxy- α -D-glucopyrano)-[2,1-d]-2-



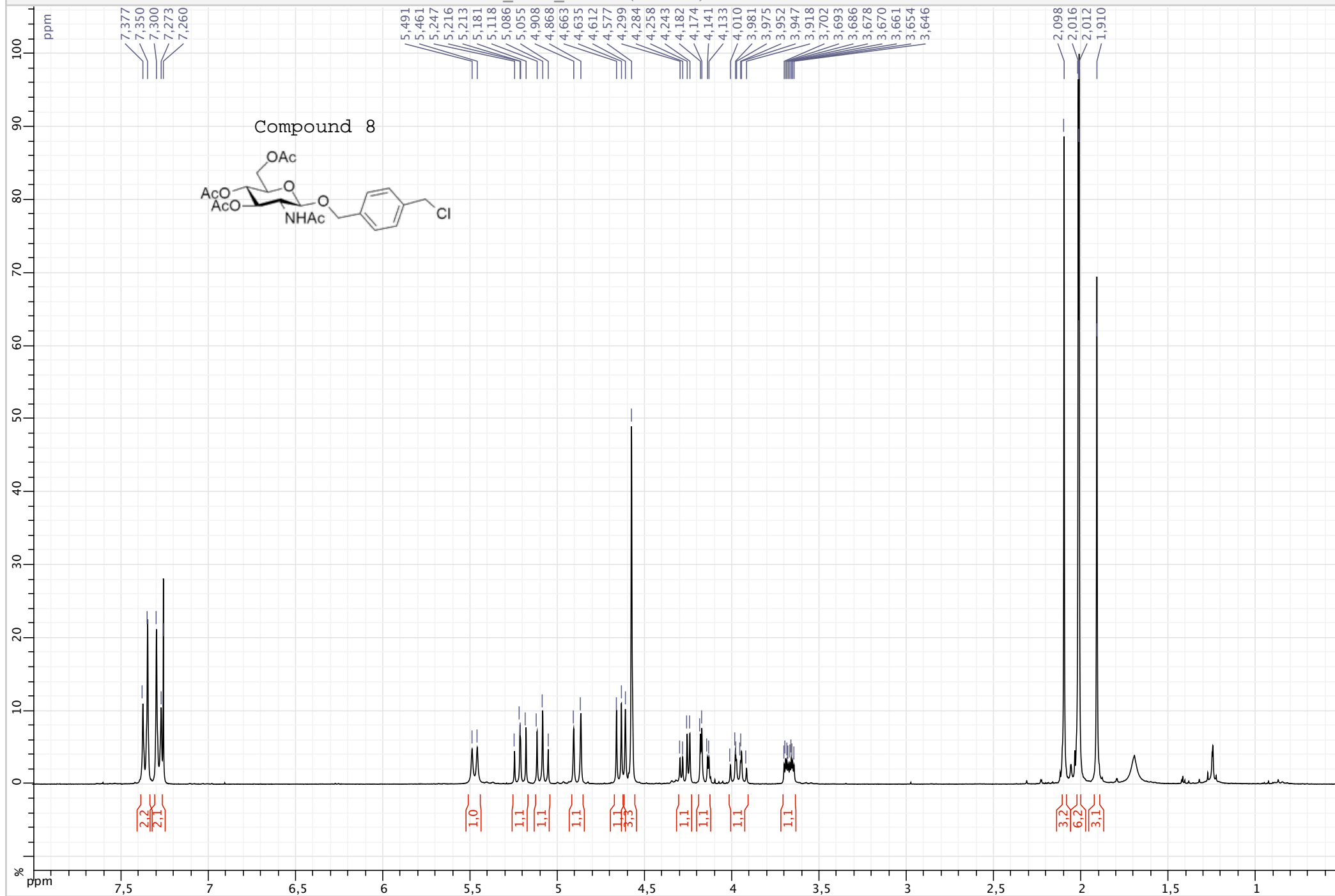
TTBP (53 mg, 0.215 mmol, 2 eq.) and Fe(OTf)₃·6.2DMSO (16 mg, 0.016 mmol, 15 mol%) are added donor **40** (50 mg, 0.107 mmol, 1 eq.), in an oven-dried, argon-purged microwave vial equipped with a magnetic stirring bar. Everything is flushed under argon and dry CH₂Cl₂ is added (1 mL). After sealing the vial, the reaction mixture is heated to 80 °C under microwave irradiation for 45 minutes (CEM Discover instrument). Then, the reaction mixture is diluted in CH₂Cl₂ (20 mL) and washed with a saturated aqueous solution of NaHCO₃ (20 mL). The aqueous layer is extracted with CH₂Cl₂ (4×20 mL) and the combined organic layers are washed with brine (20 mL), dried over Na₂SO₄, filtered and concentrated under reduced pressure. The crude product is purified by chromatography on silica gel (heptane/EtOAc 8:2 to 4:6) to give the oxazoline **554** (27 mg, 62 %, white amorphous solid).

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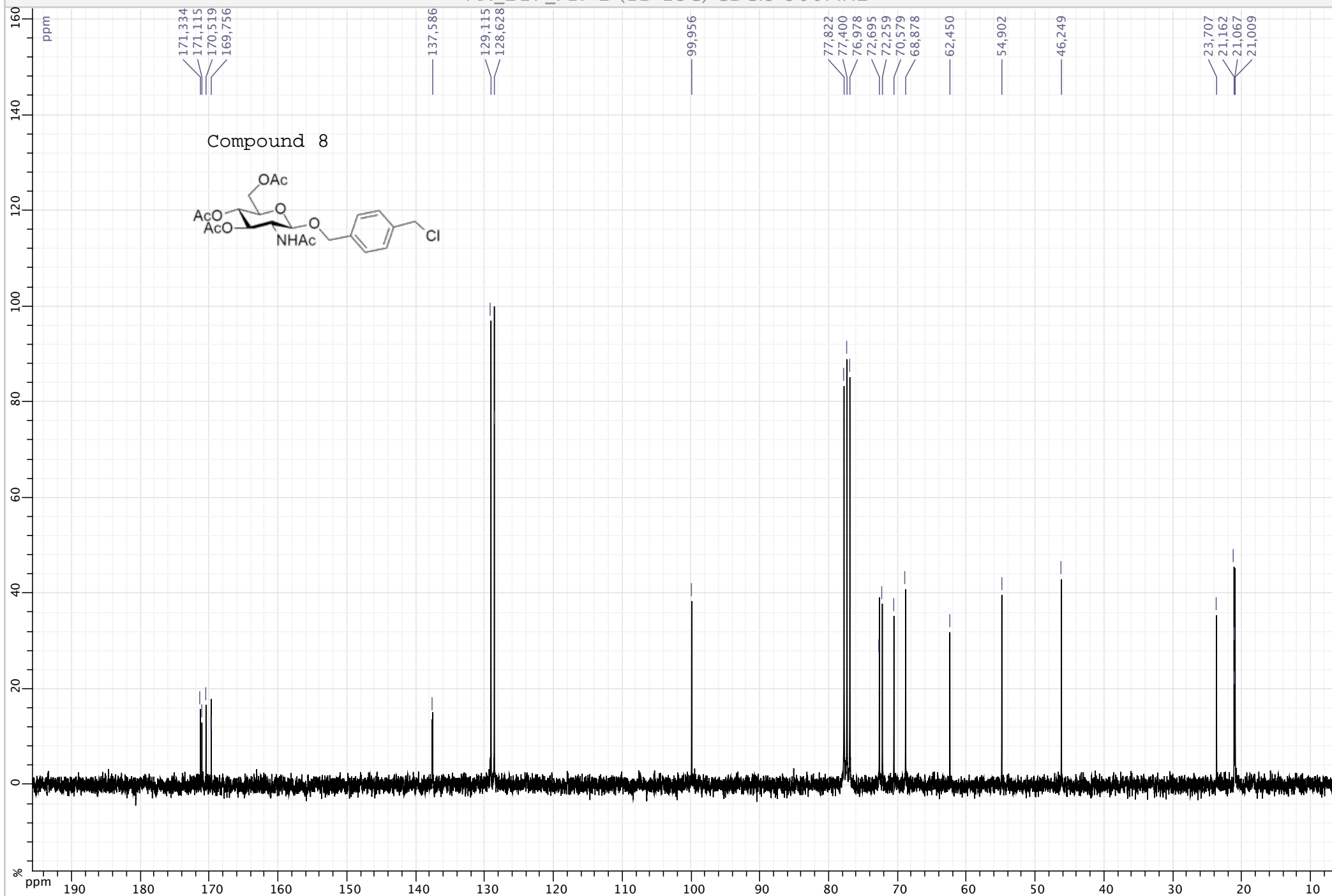
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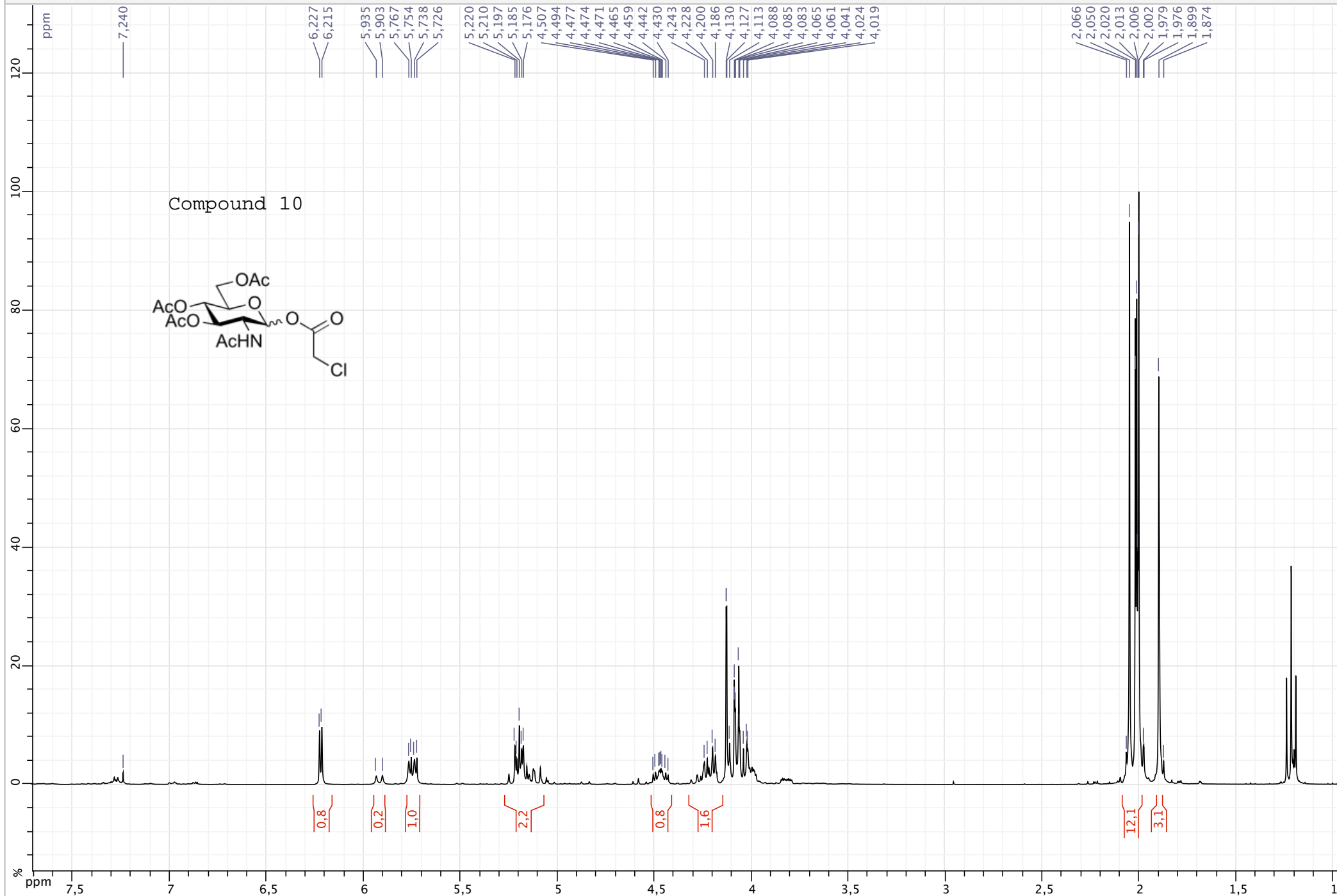
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AX_217_F1P 2 (1D 13C) CDCl3 300MHz

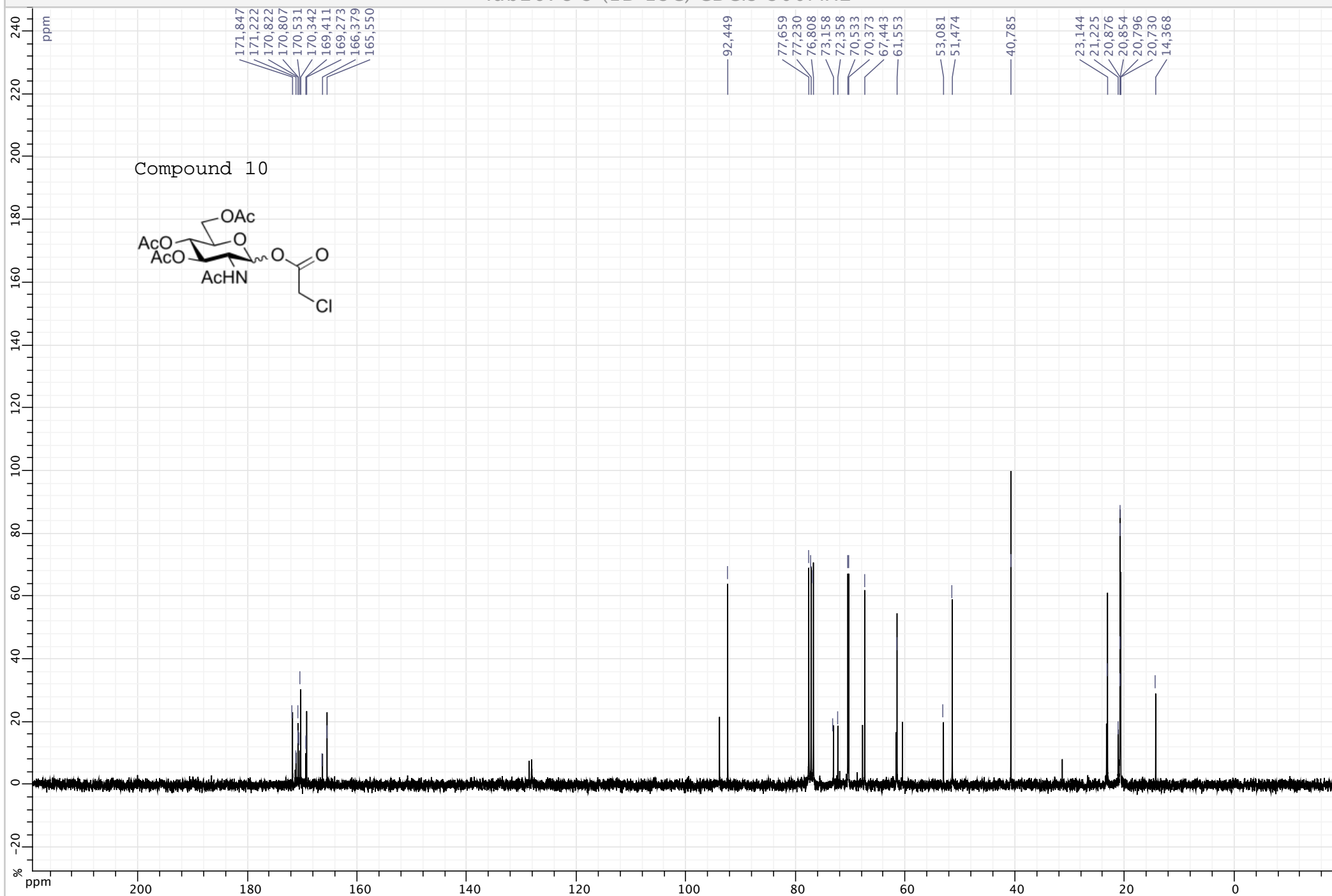
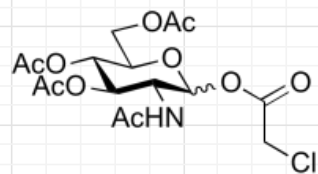


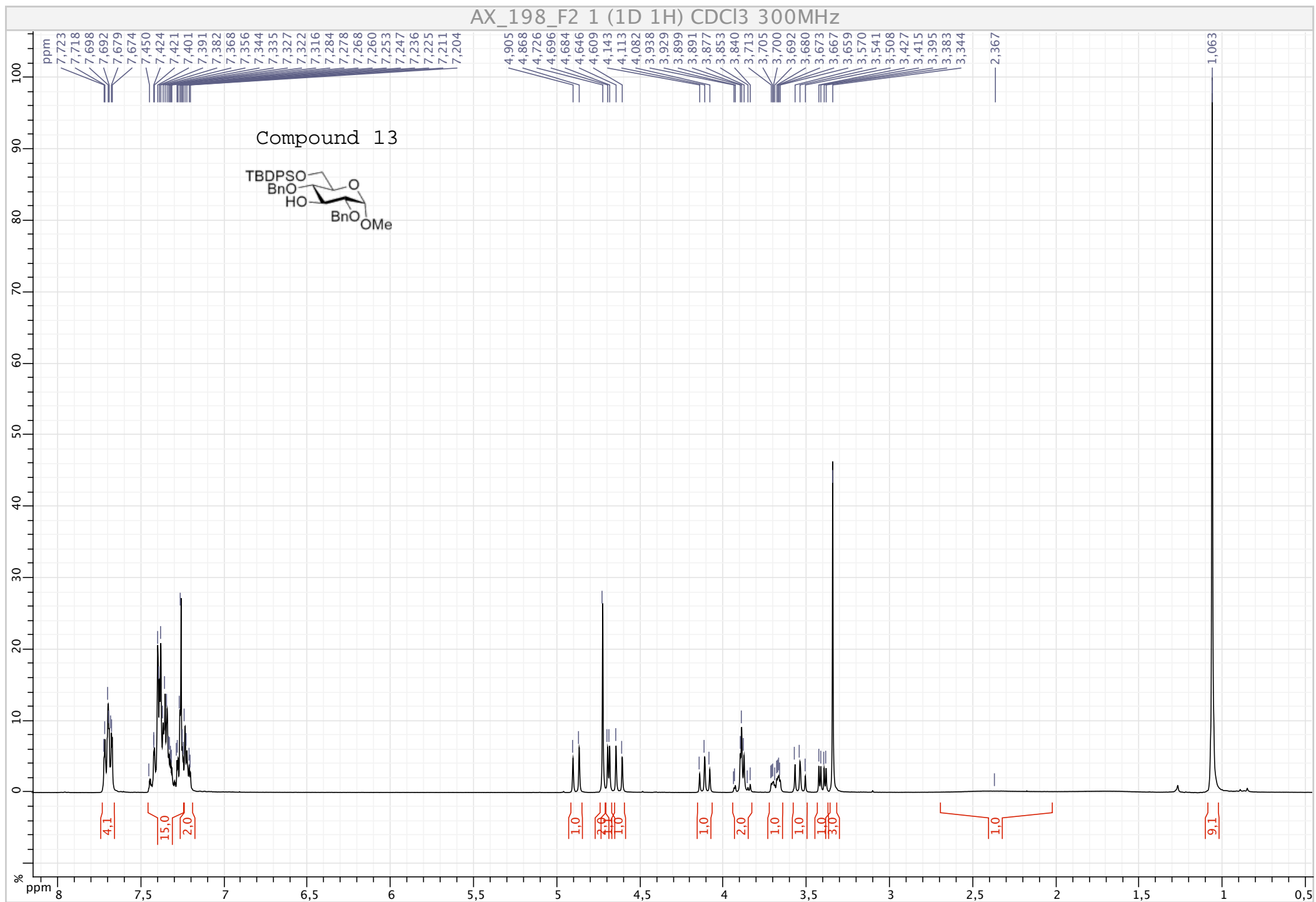
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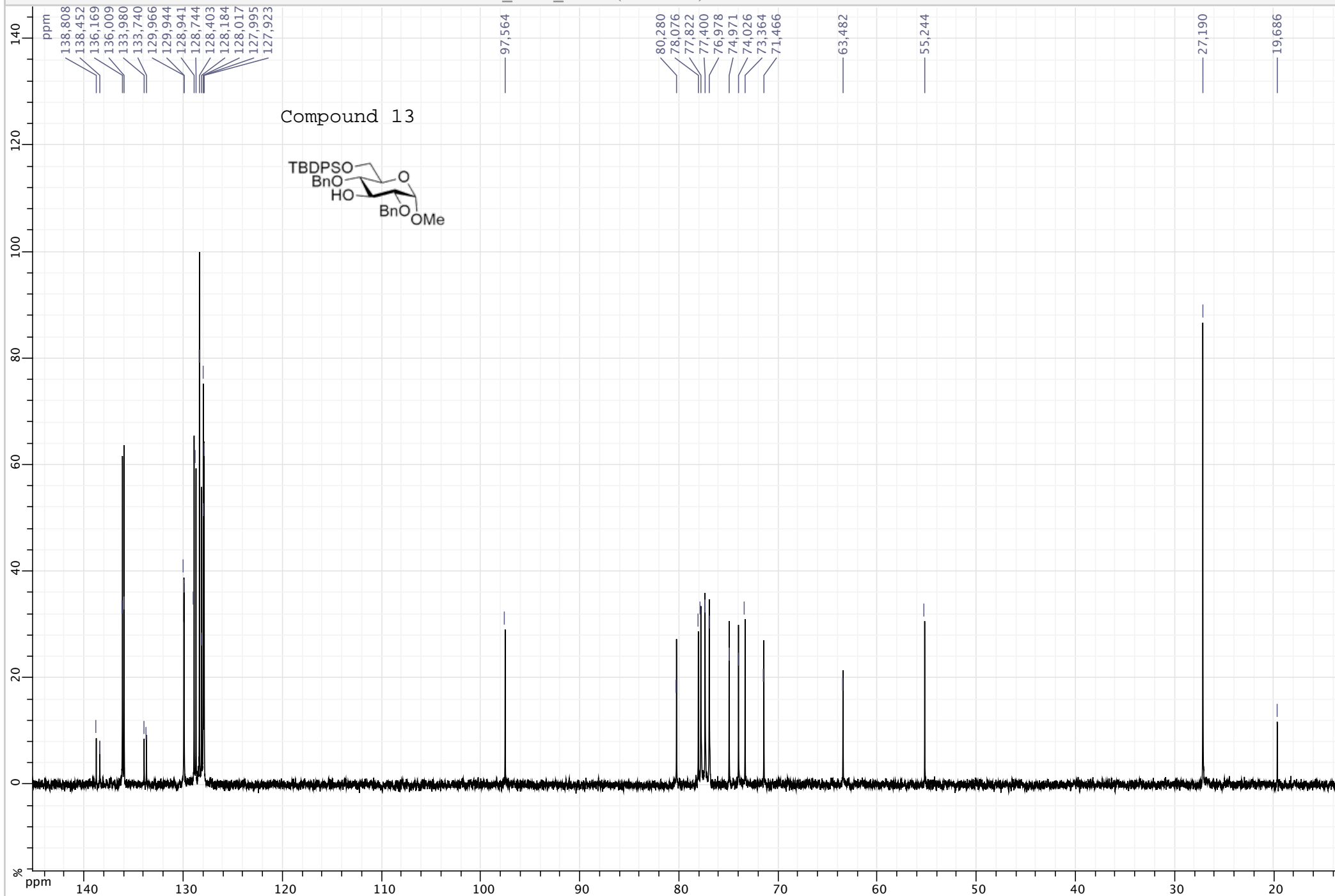
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Compound 10



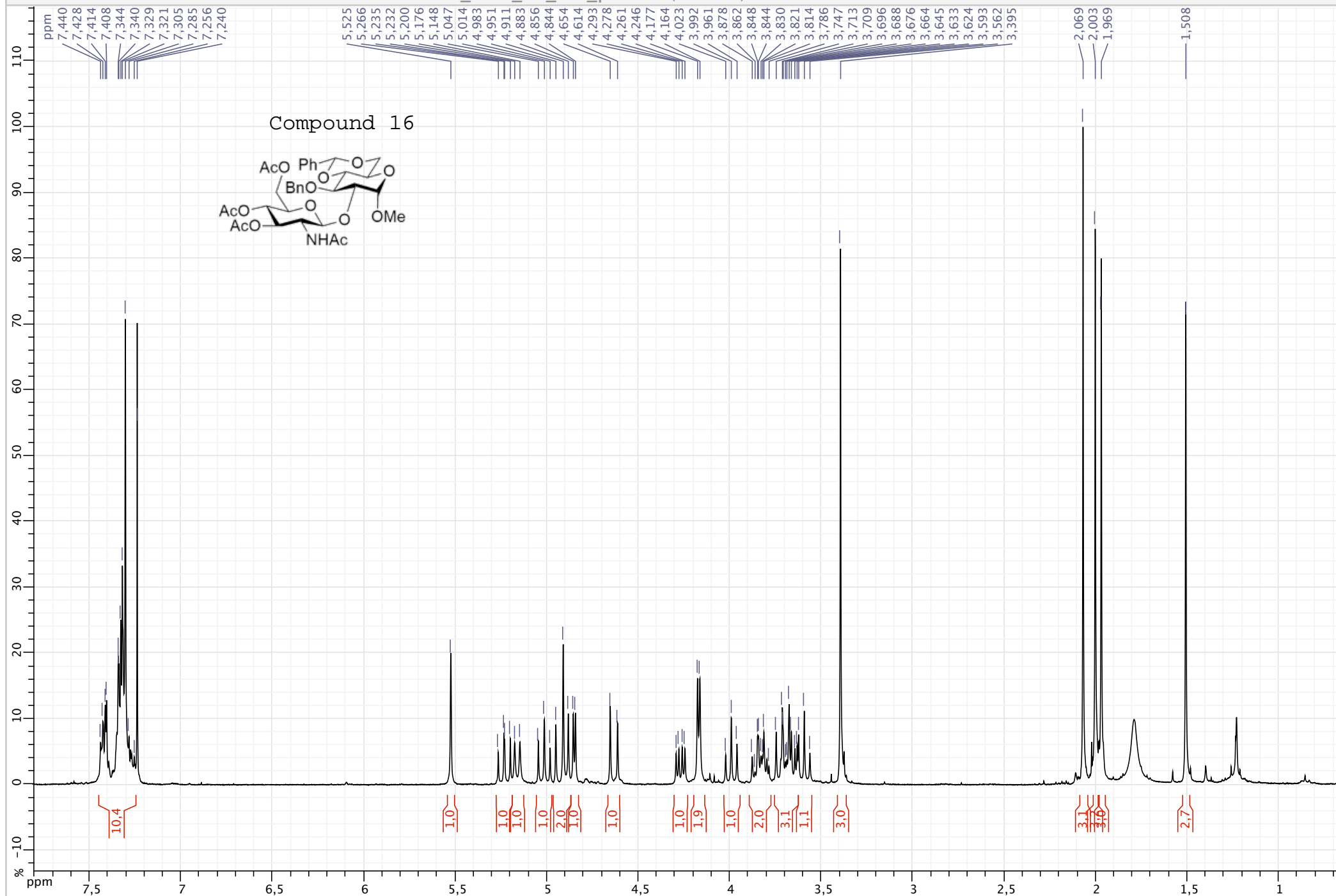
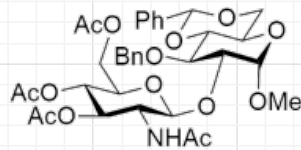


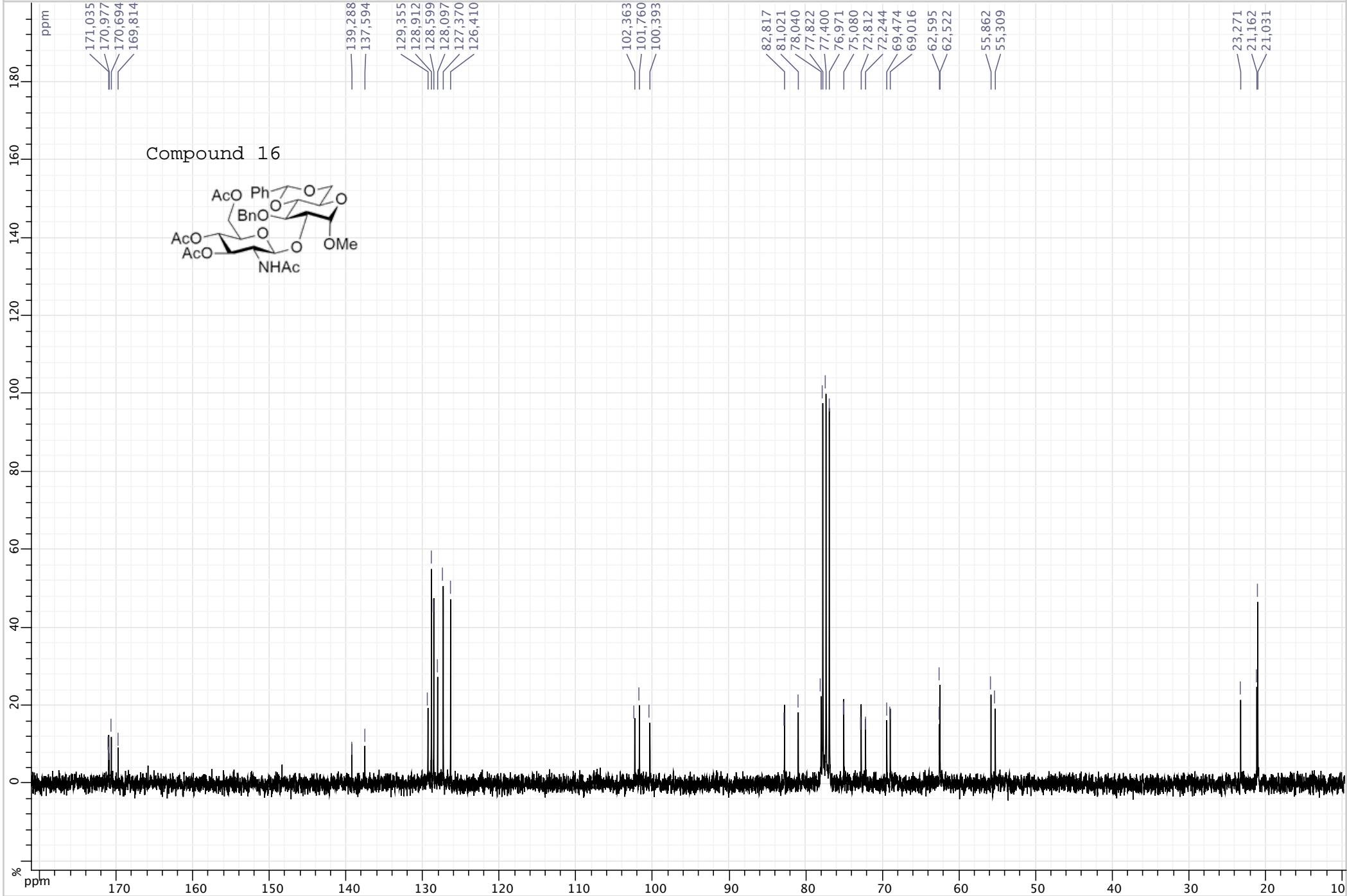
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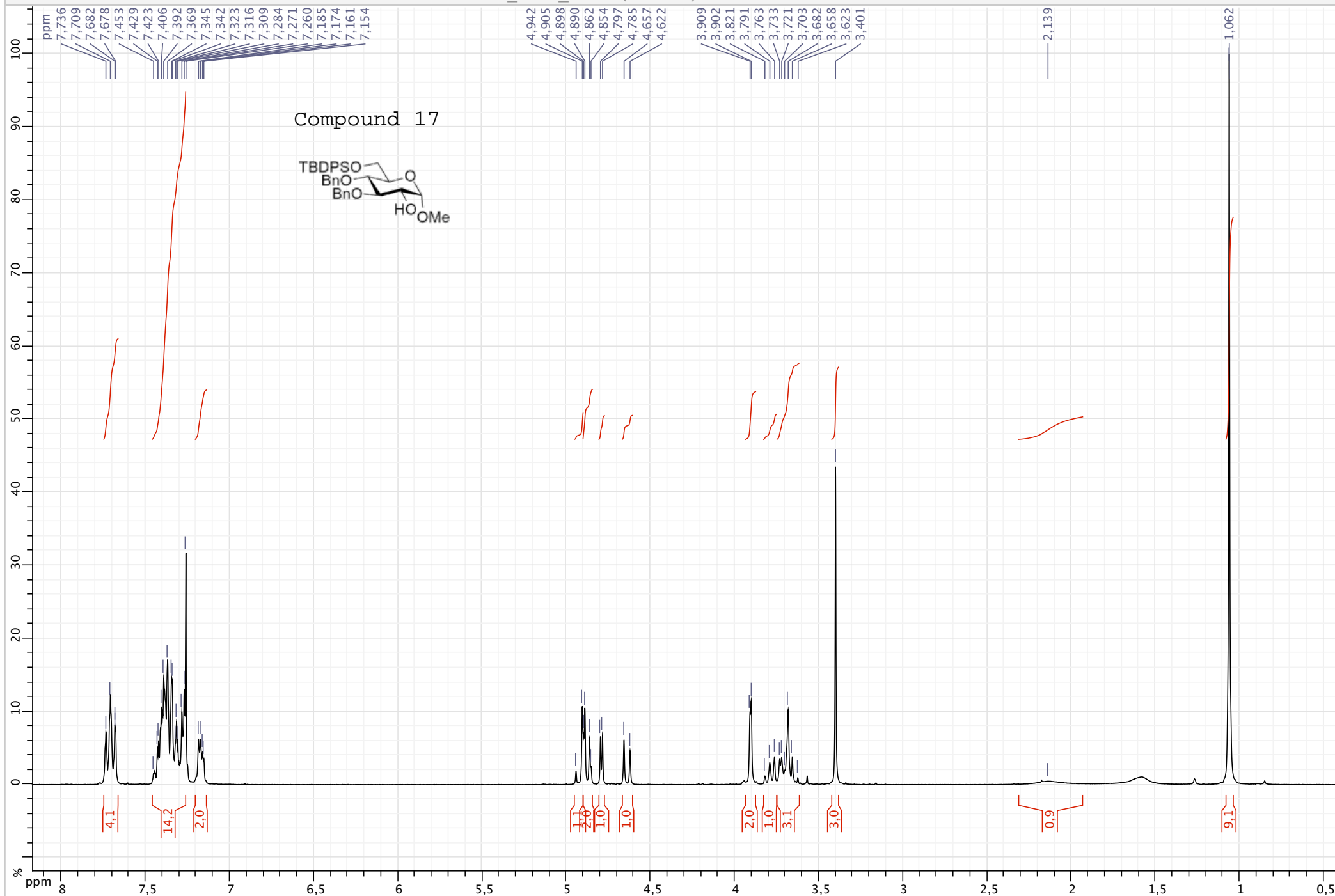
AX_213_F1_ext_pent 1 (1D 1H) CDCl3 300MHz

Compound 16

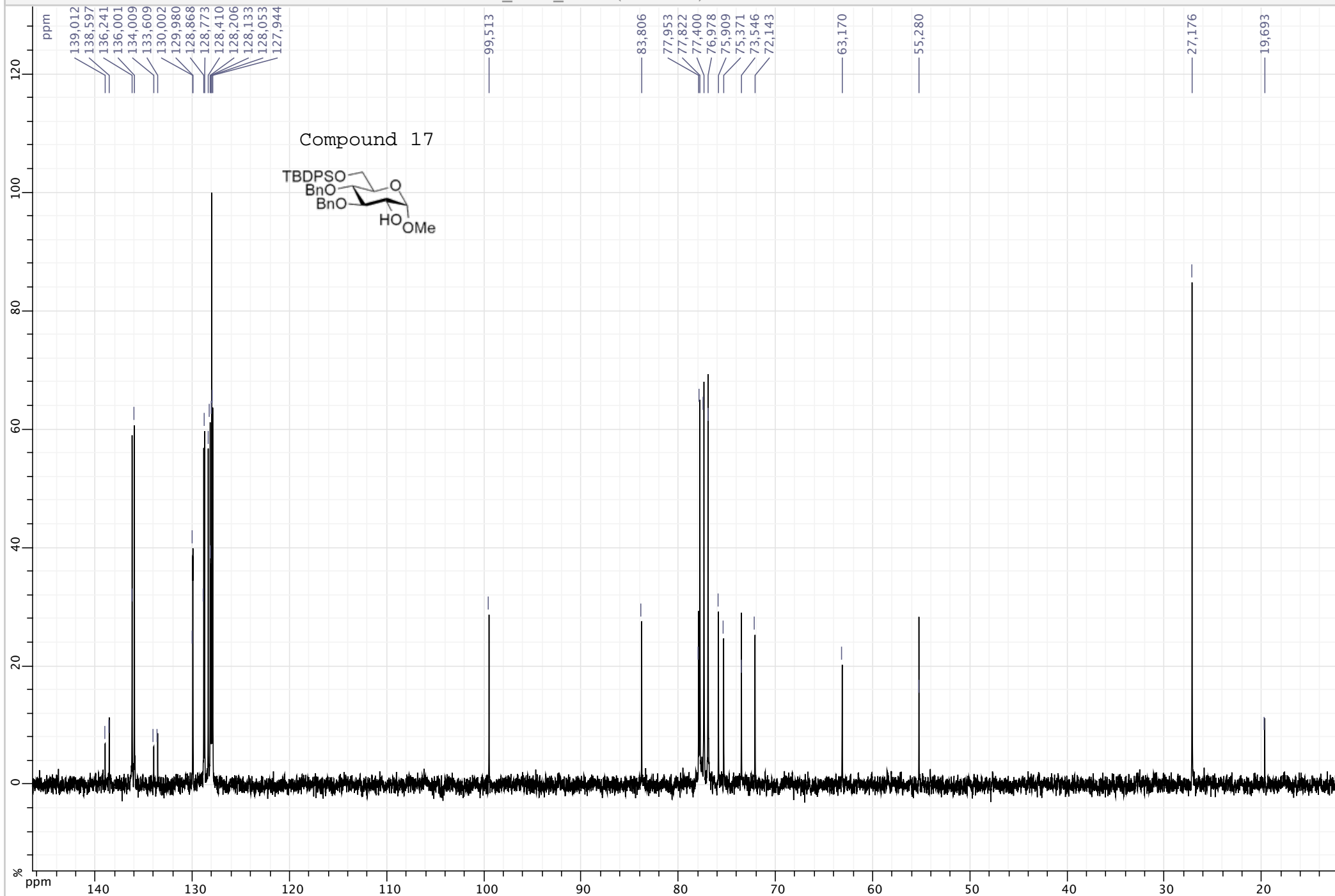




AX_199_F2 1 (1D 1H) CDCl3 300MHz



AX_199_F2 3 (1D 13C) CDCl3 300MHz



AX_202_F2_1 (1D 1H) CDCl3 300MHz

Compound 18

Chemical structure of Compound 18 is shown, featuring a bicyclic core with various substituents including AcO, BnO, NHAc, OMe, and OTBDPS.

The 1H NMR spectrum (CDCl3, 300 MHz) displays peaks from 0.5 to 10.0 ppm. Key peaks are labeled with their chemical shifts (ppm) and integration values (red numbers below the baseline):

- 7.728, 7.723, 7.702, 7.697, 7.672, 7.668, 7.428, 7.416, 7.410, 7.405, 7.391, 7.365, 7.354, 7.339, 7.311, 7.306, 7.303, 7.291, 7.274, 7.260, 7.217, 7.211, 7.200, 7.195, 7.062, 7.055, 7.043, 7.030 (Aromatic region, integration: 4.2, 12.1, 2.5, 2.0)
- 5.379, 5.348, 5.344, 5.313, 5.207, 5.178, 5.065, 5.033, 5.000, 4.997, 4.969, 4.917, 4.905, 4.834, 4.777, 4.741, 4.593, 4.558, 4.227, 4.217, 4.210, 4.000, 3.971, 3.968, 3.939, 3.884, 3.876, 3.842, 3.836, 3.814, 3.808, 3.798, 3.792, 3.779, 3.764, 3.759, 3.747, 3.731, 3.723, 3.690, 3.681, 3.673, 3.656, 3.645, 3.624, 3.612, 3.370, 2.089, 2.034, 1.993, 1.445 (Aliphatic region, integration: 1.0, 1.0, 1.1, 1.0, 2.0, 1.0, 1.0, 1.9, 1.1, 2.0, 5.3, 3.0)
- 1.066 (Methyl singlet, integration: 9.3)

The spectrum shows a complex pattern of peaks, particularly in the aromatic and aliphatic regions, consistent with the structure of Compound 18.

AX_202_F2_1 (1D 1H) CDCl3 300MHz

Compound 18

Chemical structure of Compound 18 is shown, featuring a bicyclic system with various substituents including AcO, BnO, NHAc, OMe, and OTBDPS.

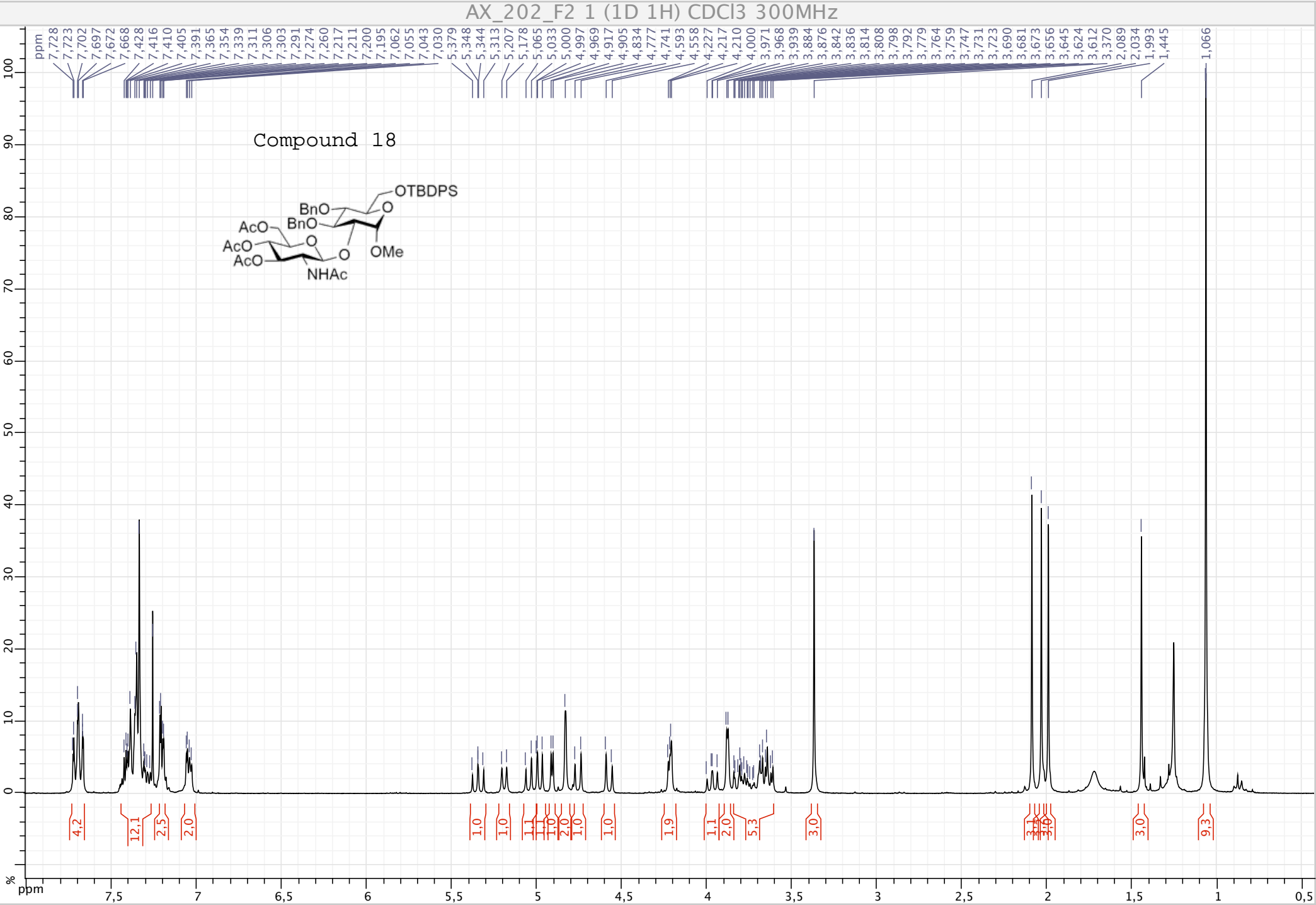
The 1H NMR spectrum (CDCl3, 300 MHz) displays peaks in the aromatic region (7.0-7.8 ppm) and the aliphatic region (1.0-5.5 ppm). Integration values are provided below the peaks.

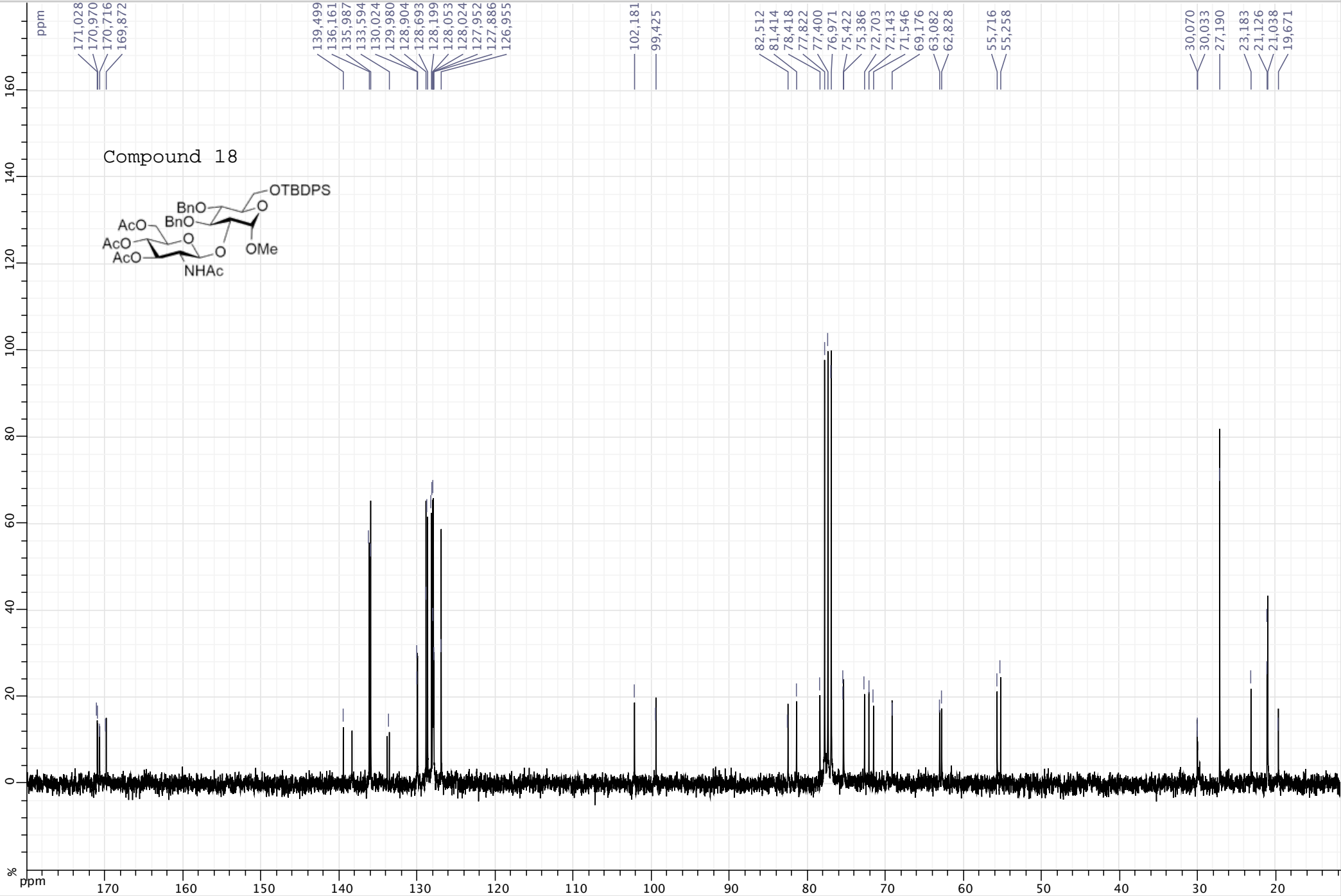
Peak list (ppm):

- 7.728, 7.723, 7.702, 7.697, 7.672, 7.668, 7.428, 7.416, 7.410, 7.405, 7.391, 7.365, 7.354, 7.339, 7.311, 7.306, 7.303, 7.291, 7.274, 7.260, 7.217, 7.211, 7.200, 7.195, 7.062, 7.055, 7.043, 7.030, 5.379, 5.348, 5.344, 5.313, 5.207, 5.178, 5.065, 5.033, 5.000, 4.997, 4.969, 4.917, 4.905, 4.834, 4.777, 4.741, 4.593, 4.558, 4.227, 4.217, 4.210, 4.000, 3.971, 3.968, 3.939, 3.884, 3.876, 3.842, 3.836, 3.814, 3.808, 3.798, 3.792, 3.779, 3.764, 3.759, 3.747, 3.731, 3.723, 3.690, 3.681, 3.673, 3.656, 3.645, 3.624, 3.612, 3.370, 2.089, 2.034, 1.993, 1.445, 1.066

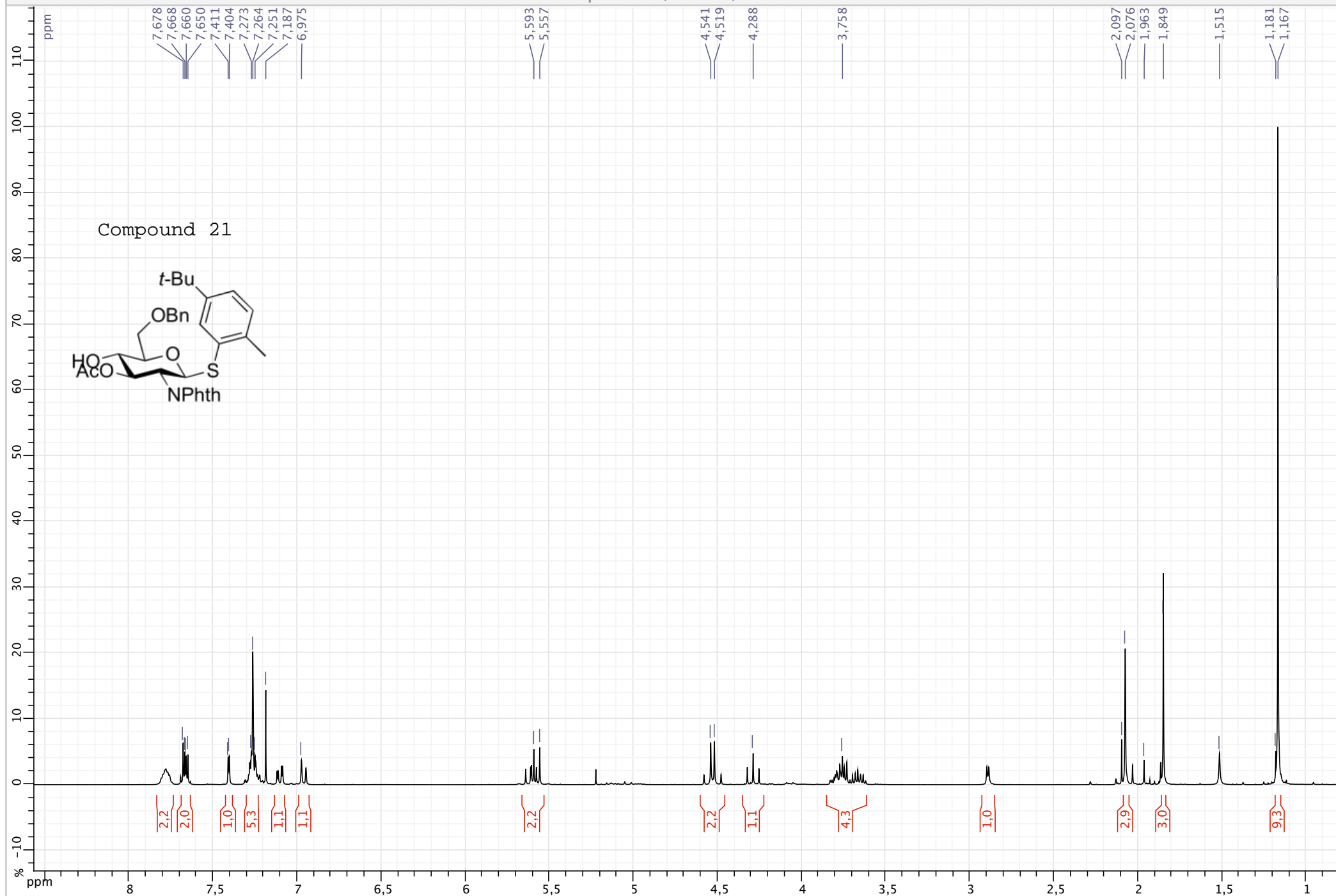
Integration values (from left to right):

- 4.2, 12.1, 2.5, 2.0, 1.0, 1.0, 1.1, 1.0, 2.0, 1.0, 1.0, 1.9, 1.1, 2.0, 5.3, 3.0, 3.1, 3.0, 3.0, 3.0, 9.3

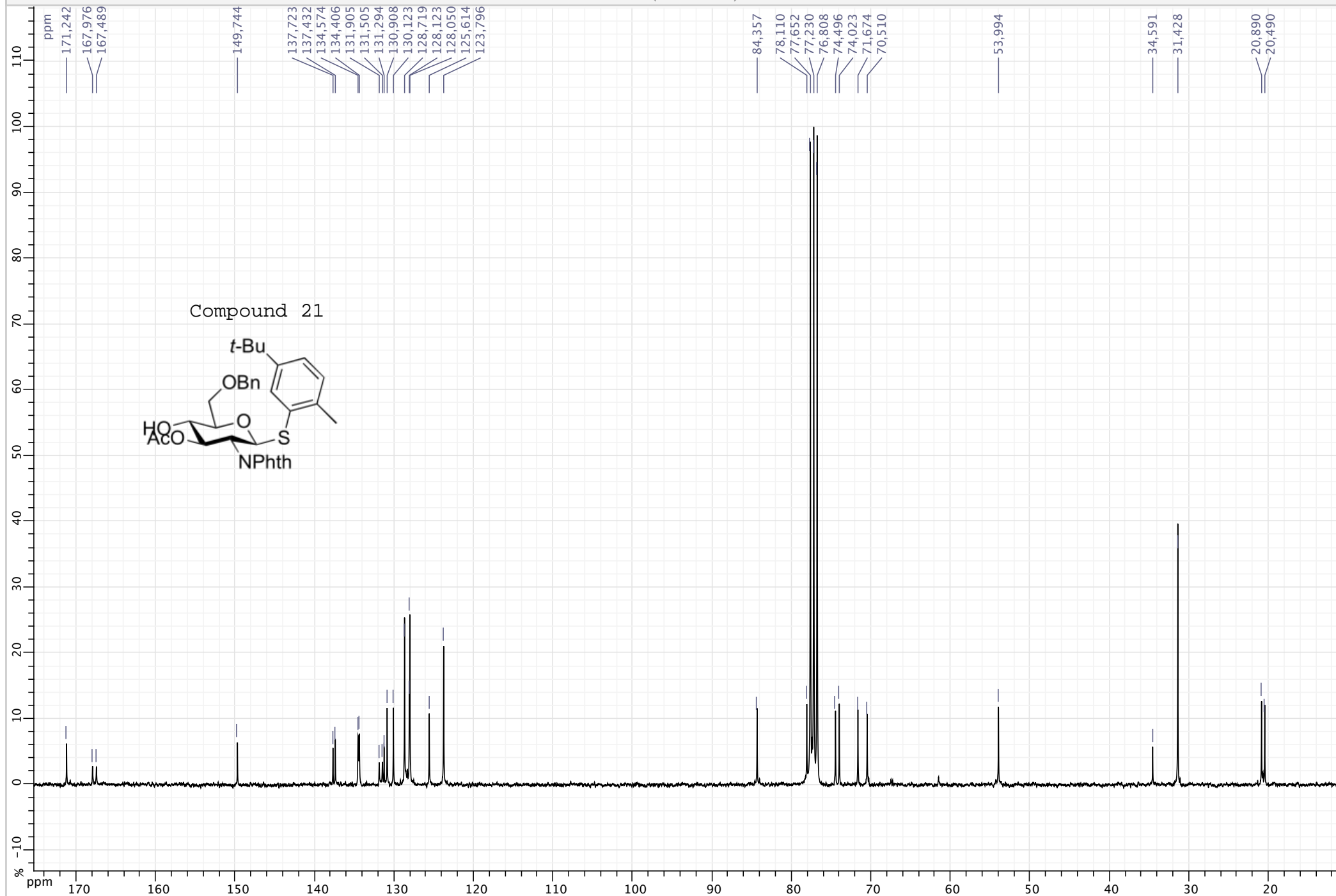




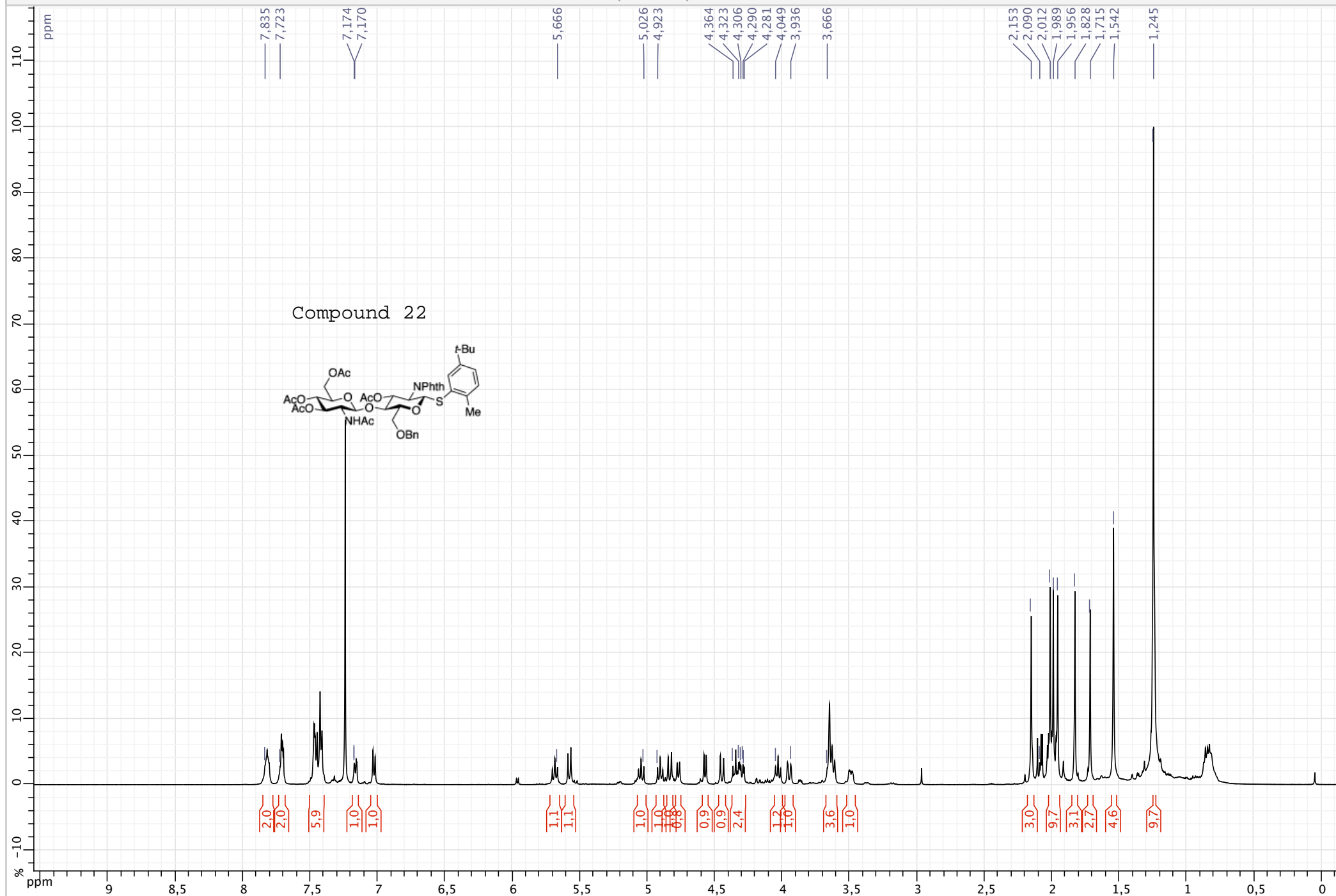
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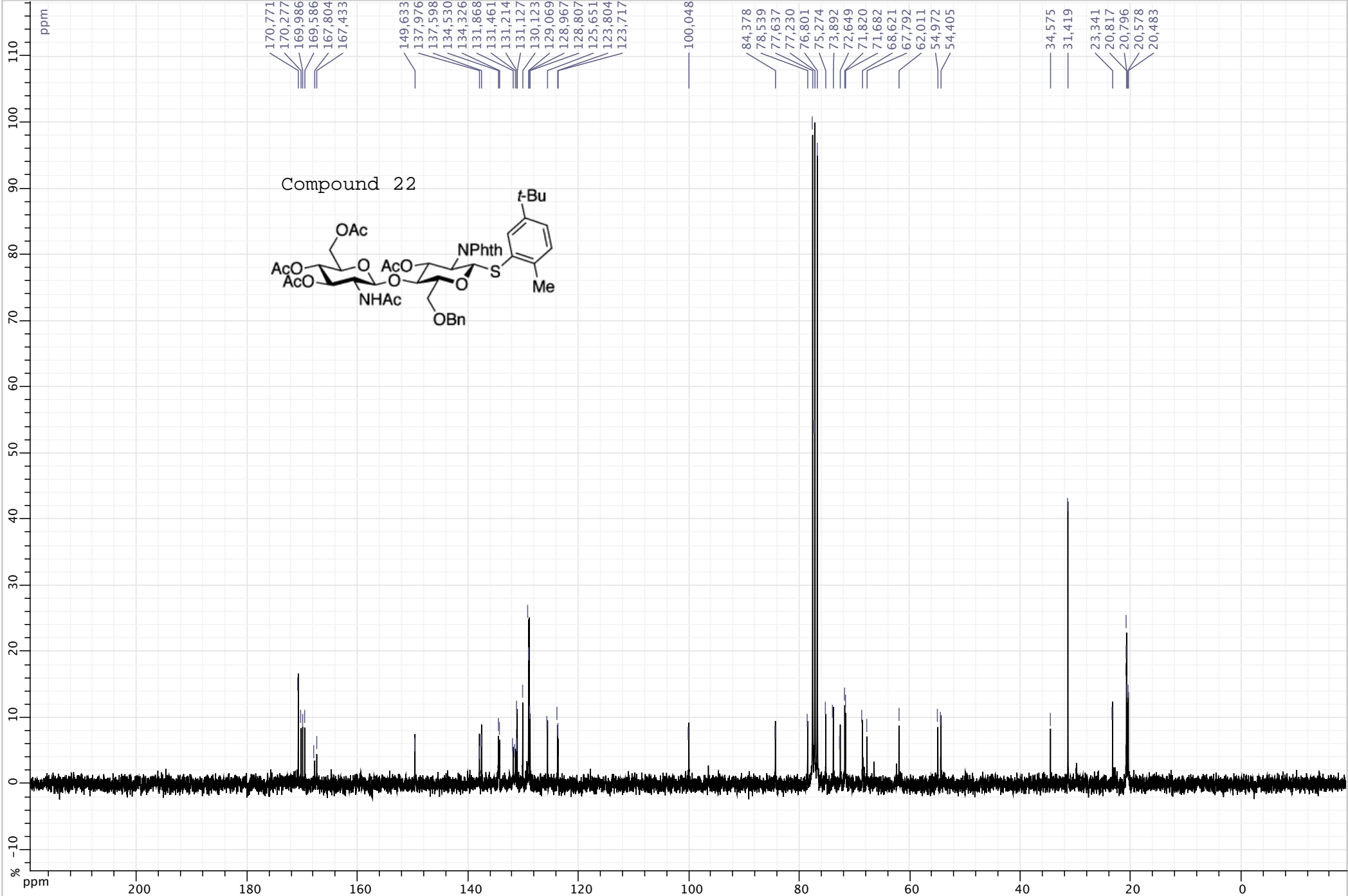
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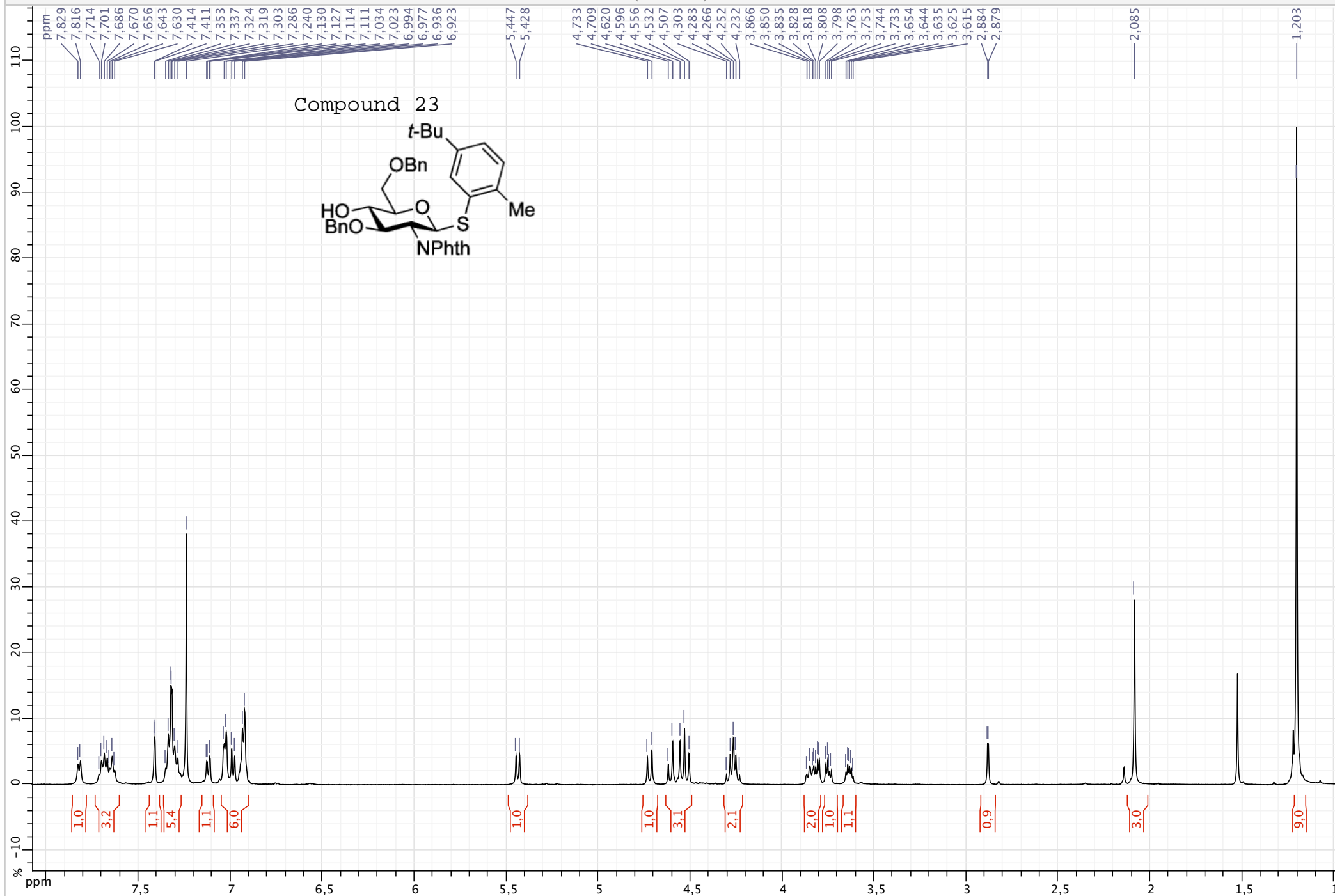
41ast548 1 (1D 1H) CDCl3 500MHz



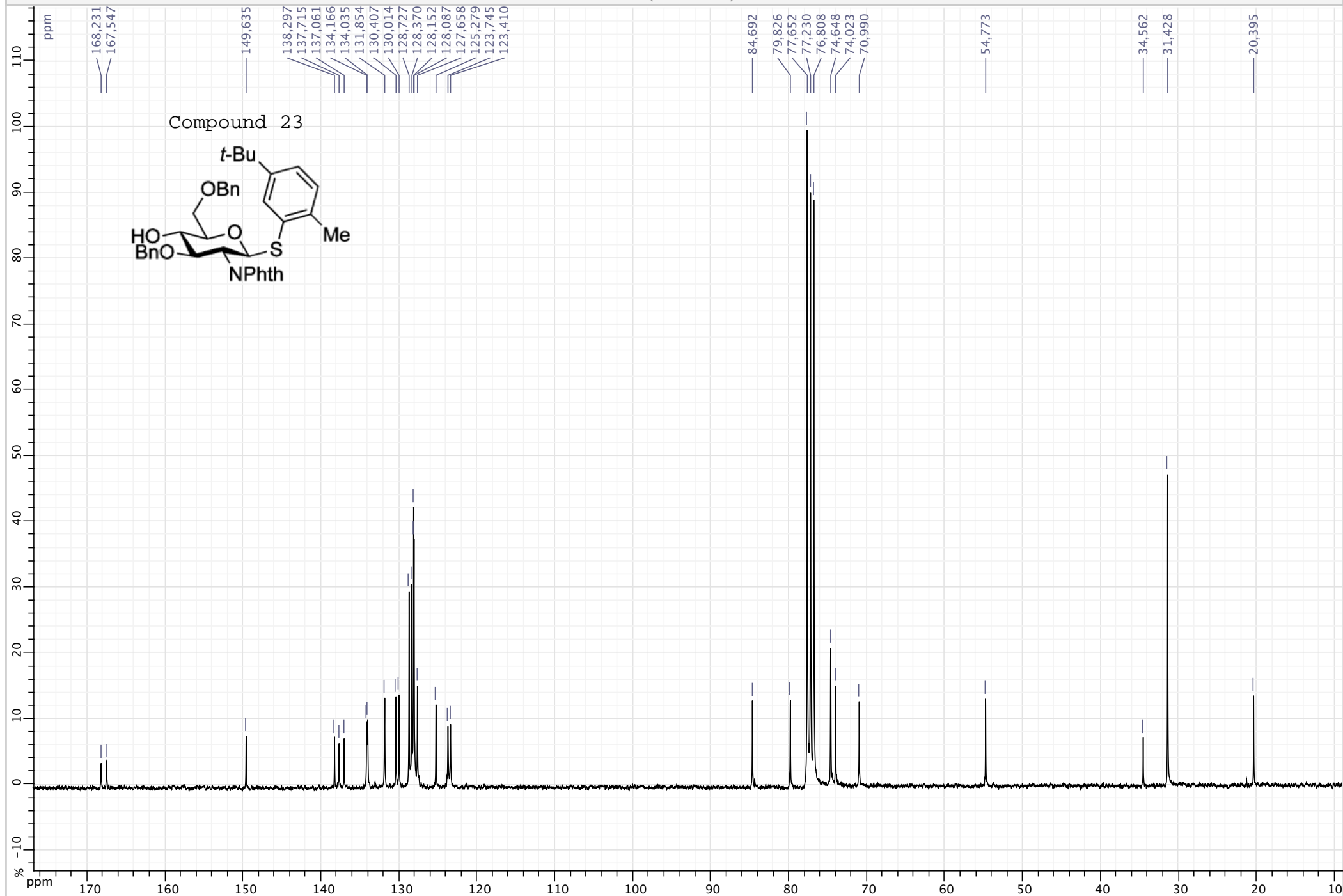
41ast-548-pass2 3 (1D 13C) CDCl₃ 300MHz



41astasarobn 1 (1D 1H) CDCl3 500MHz

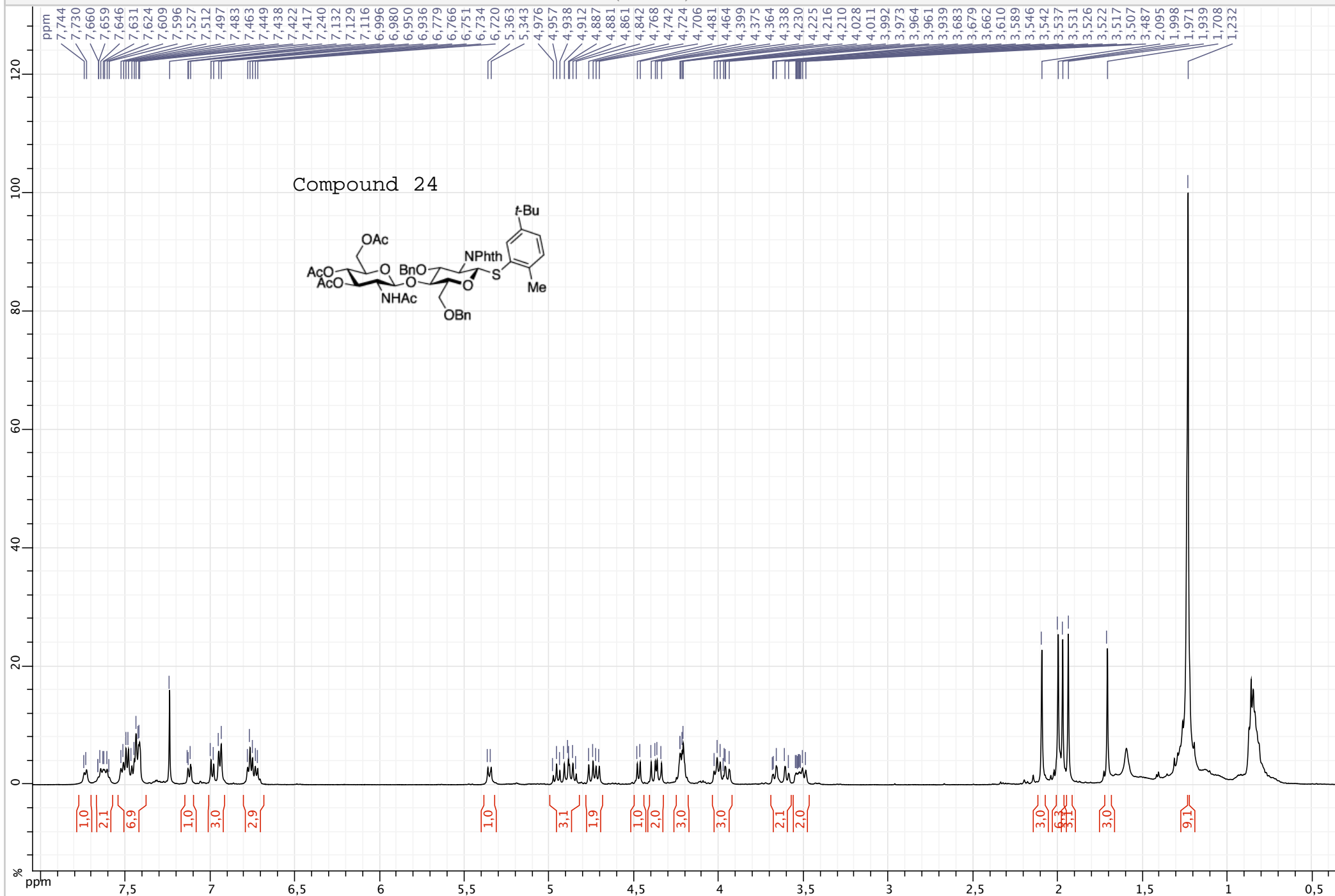
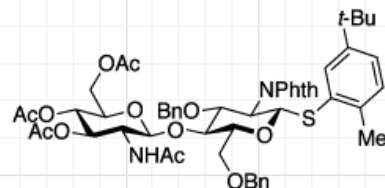


41astasarobnc13 1 (1D 13C) CDCl3 300MHz

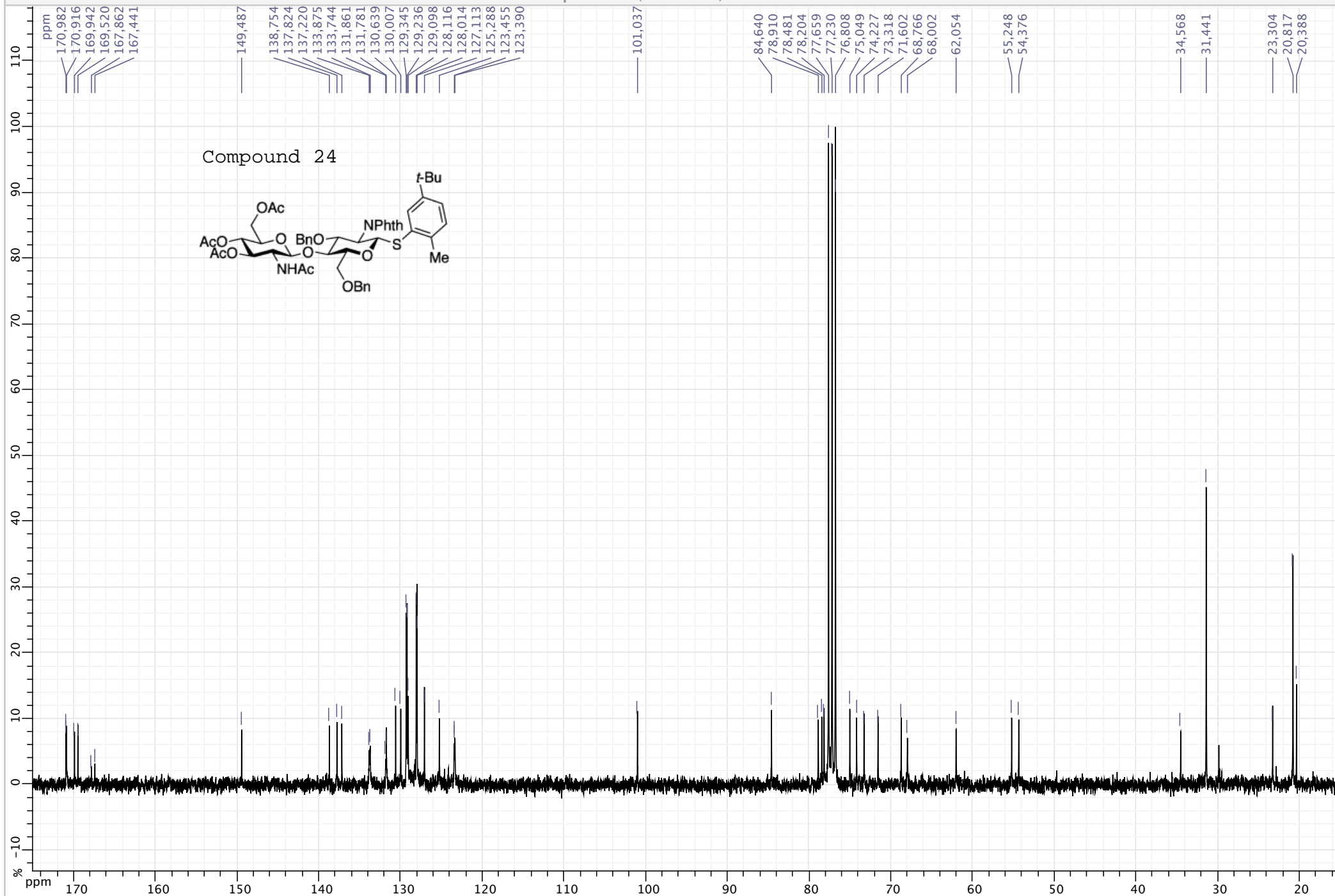


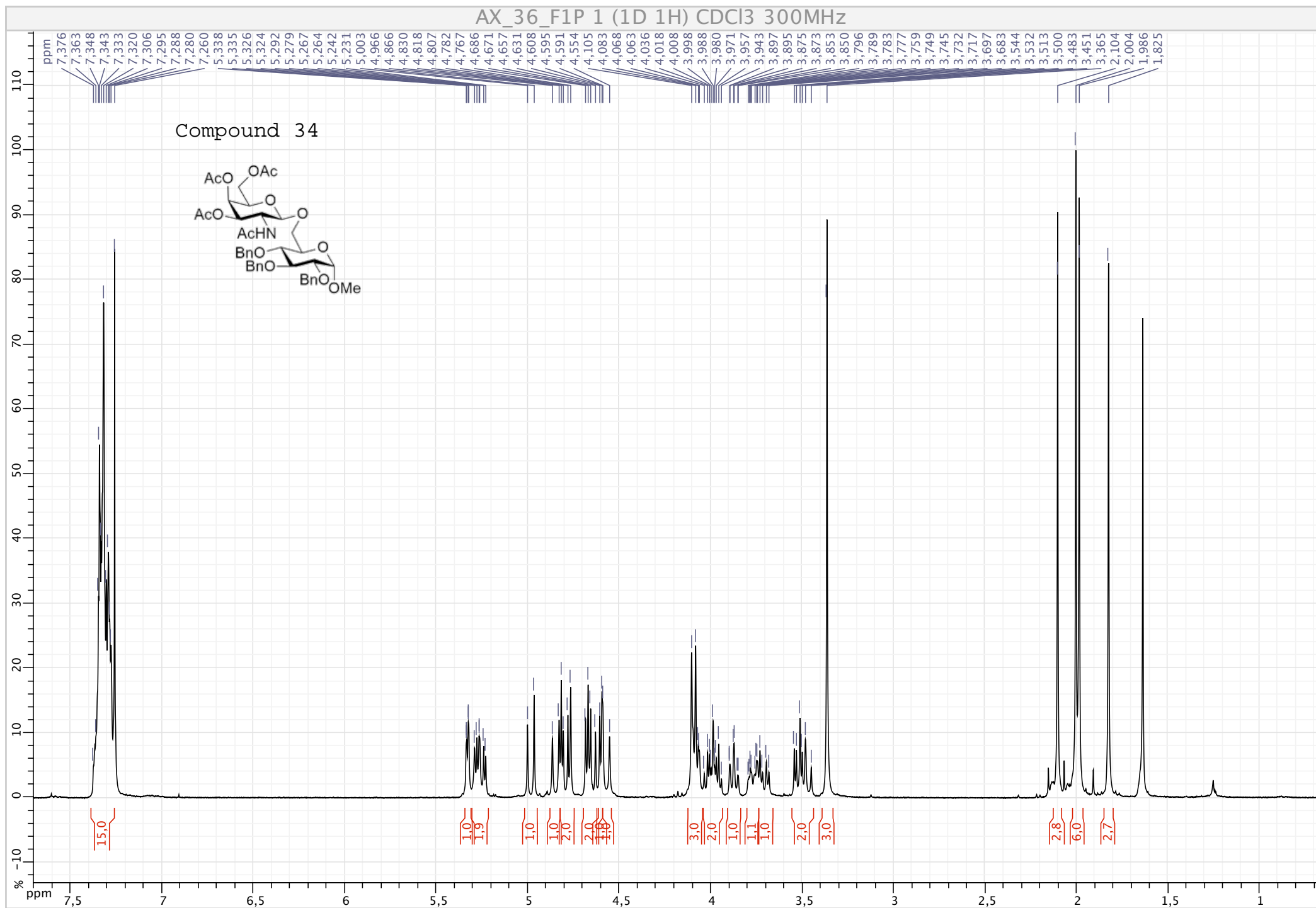
41ast283 1 (1D 1H) CDCl3 500MHz

Compound 24

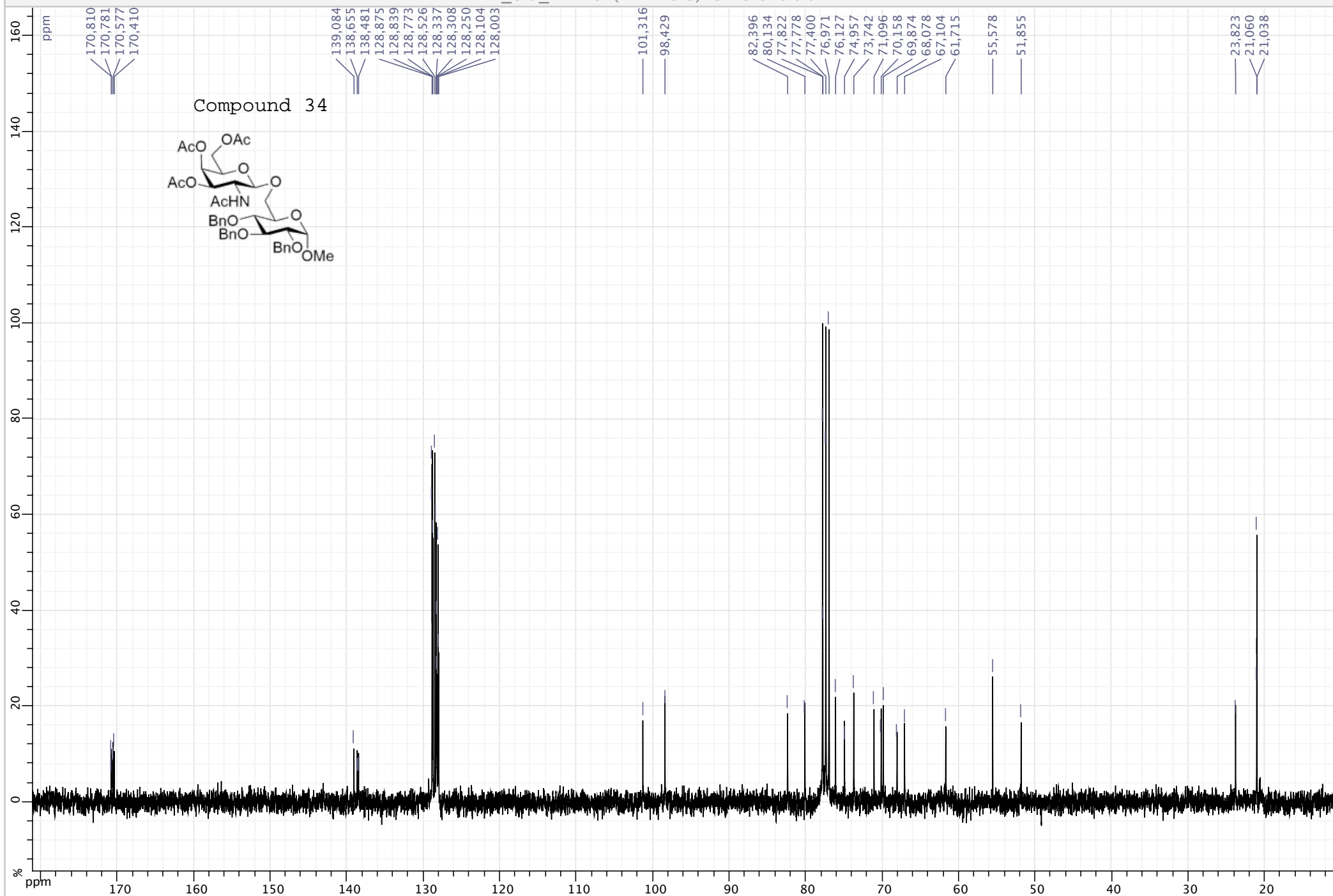


41ast550-pass 3 (1D 13C) CDCl3 300MHz

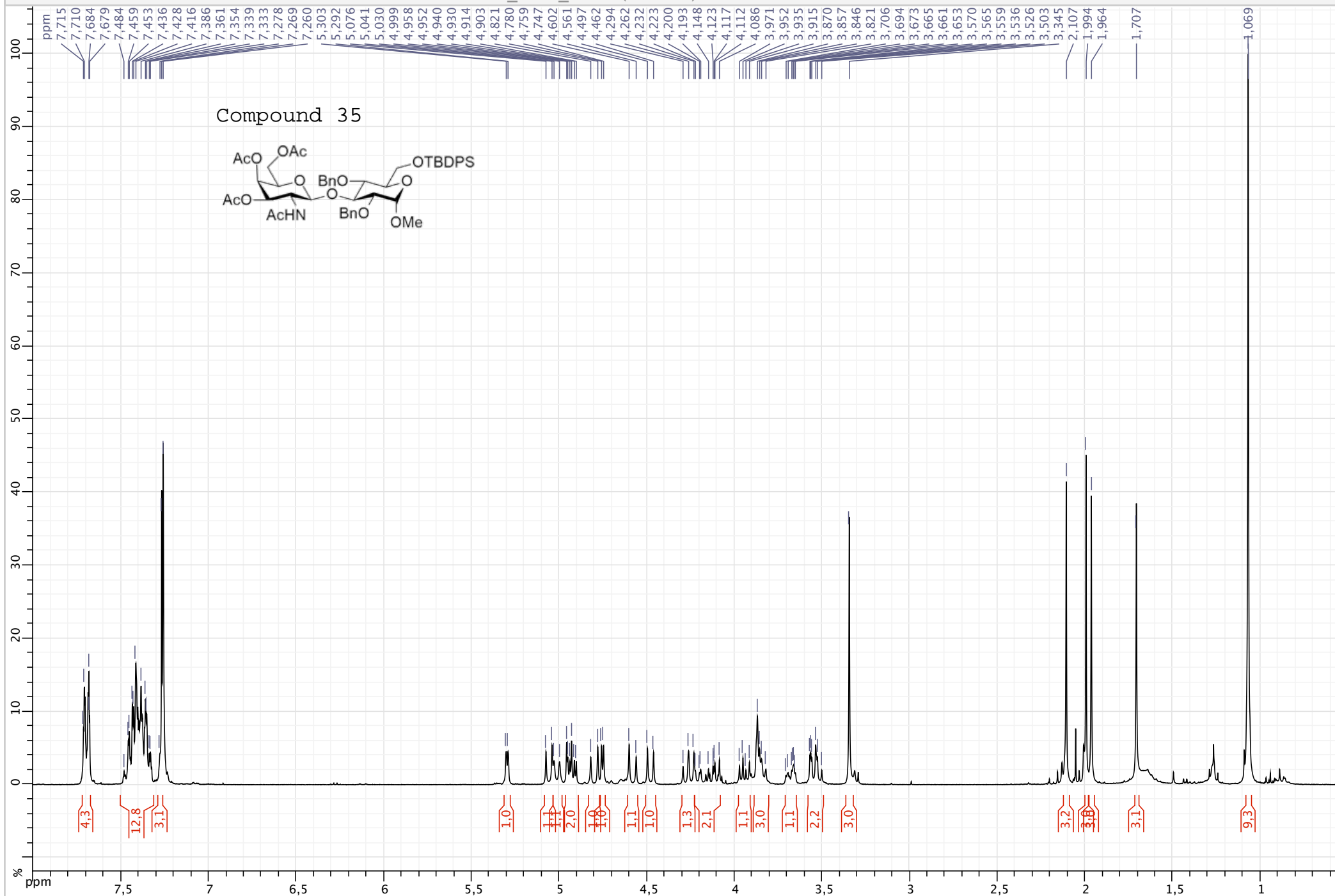




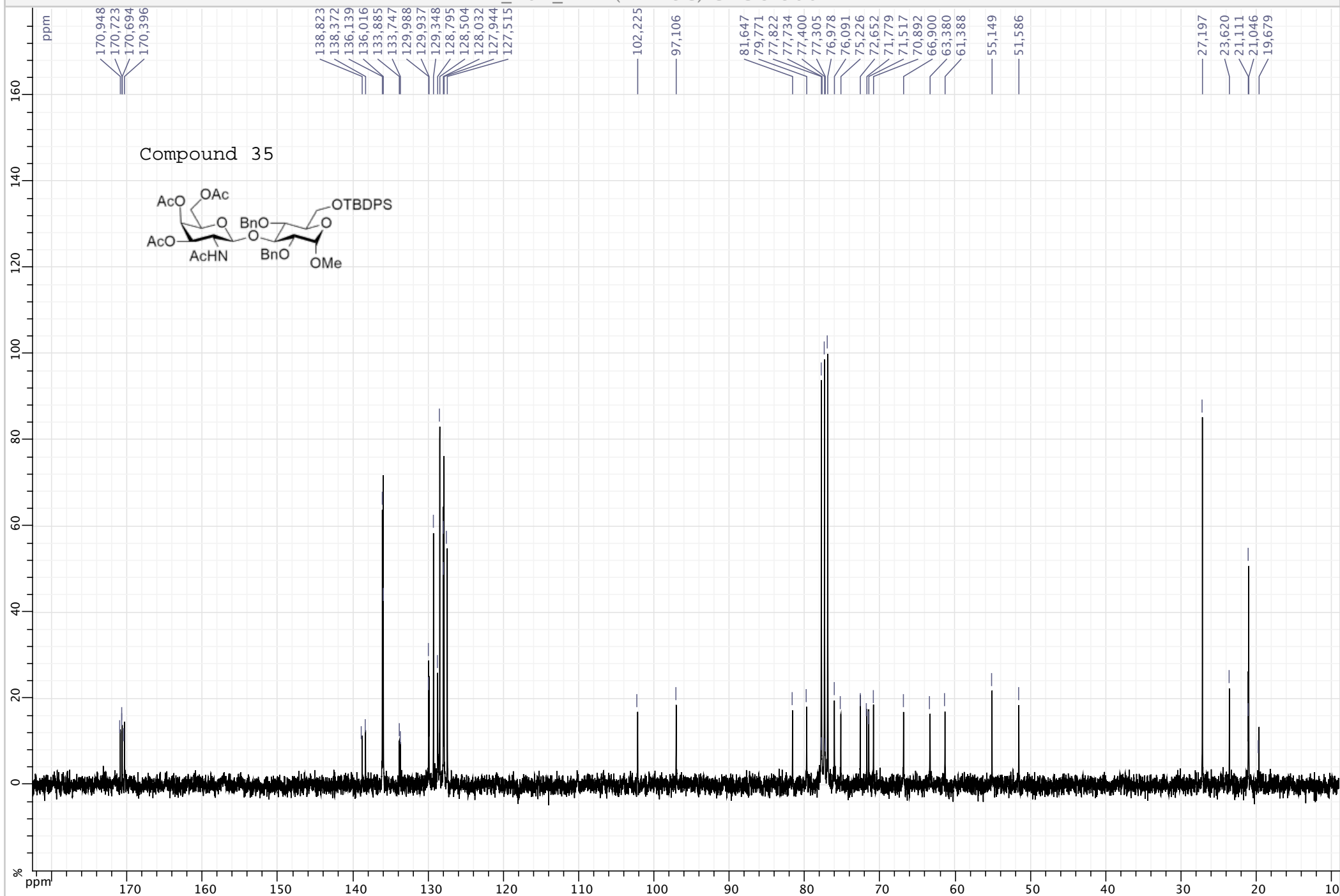
AX_36_F1P 3 (1D 13C) CDCl3 300MHz



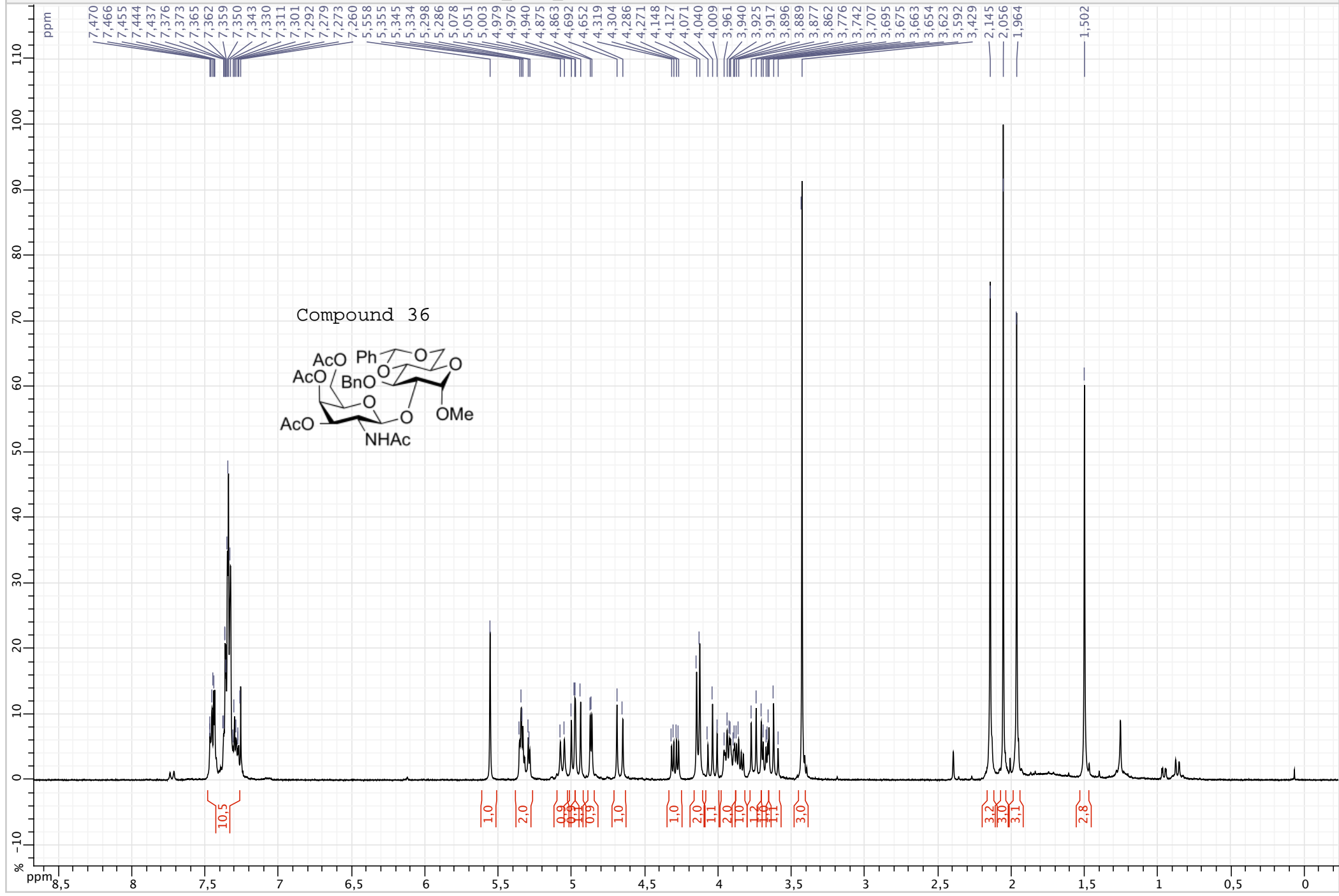
AX_201_F2 1 (1D 1H) CDCl3 300MHz



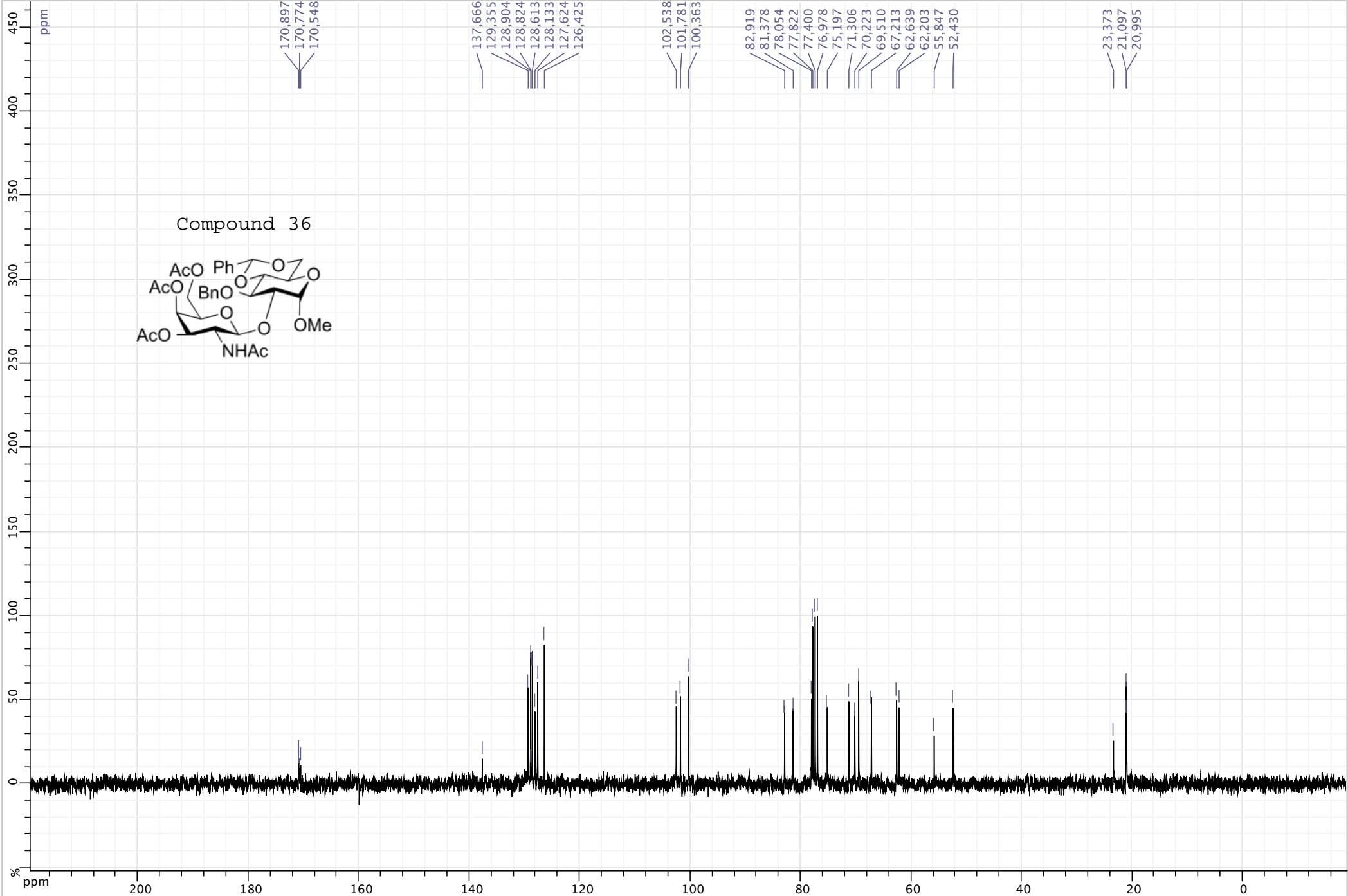
AX_201_F2 2 (1D 13C) CDCl3 300MHz



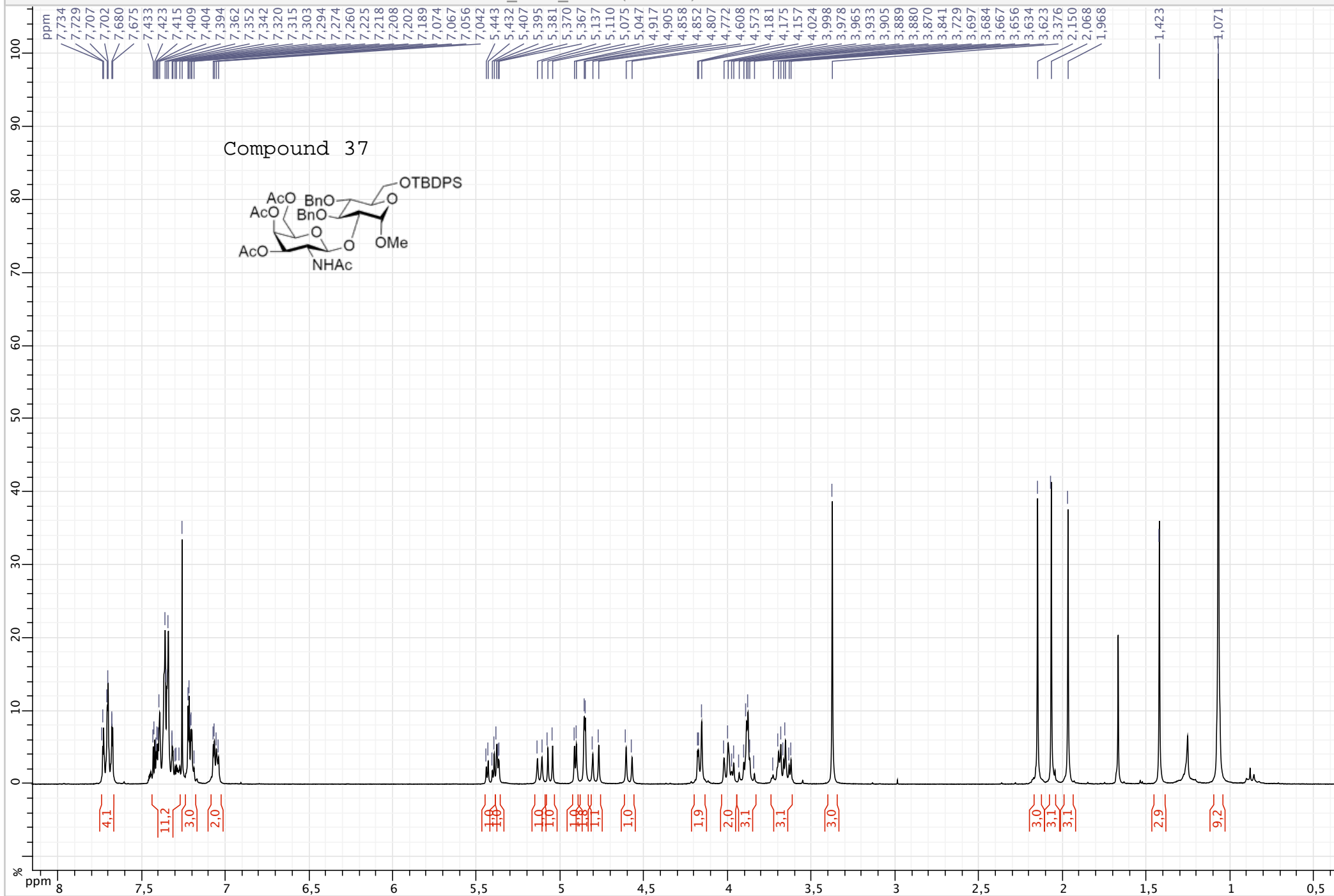
AX_304_F2P 1 (1D 1H) CDCl3 300MHz



AX_304_F2P 2 (13C) CDCI3 300MHz

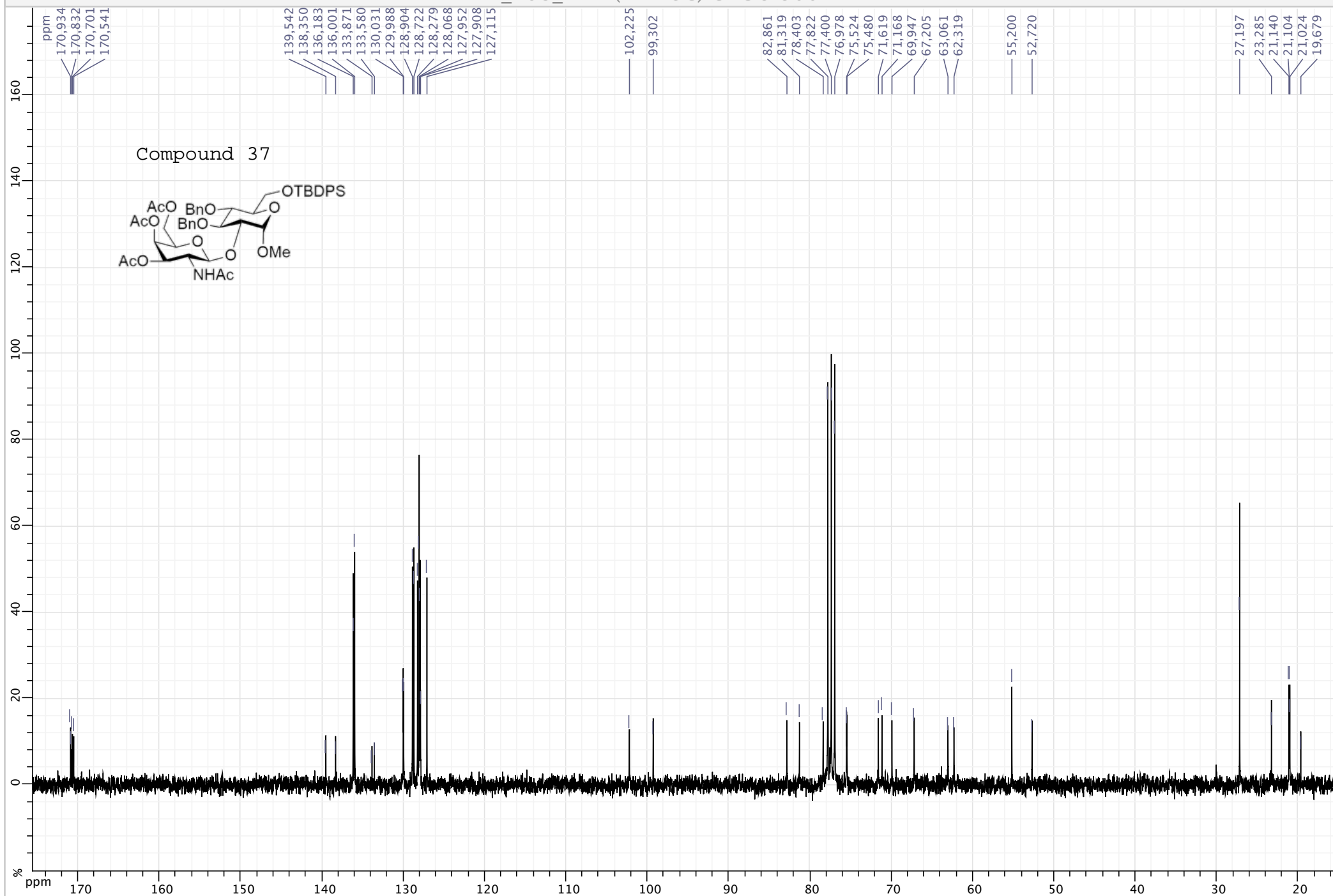
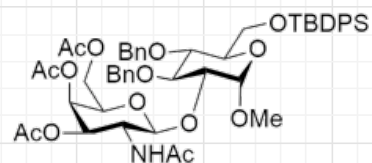


AX_268_F2 1 (1D 1H) CDCl3 300MHz

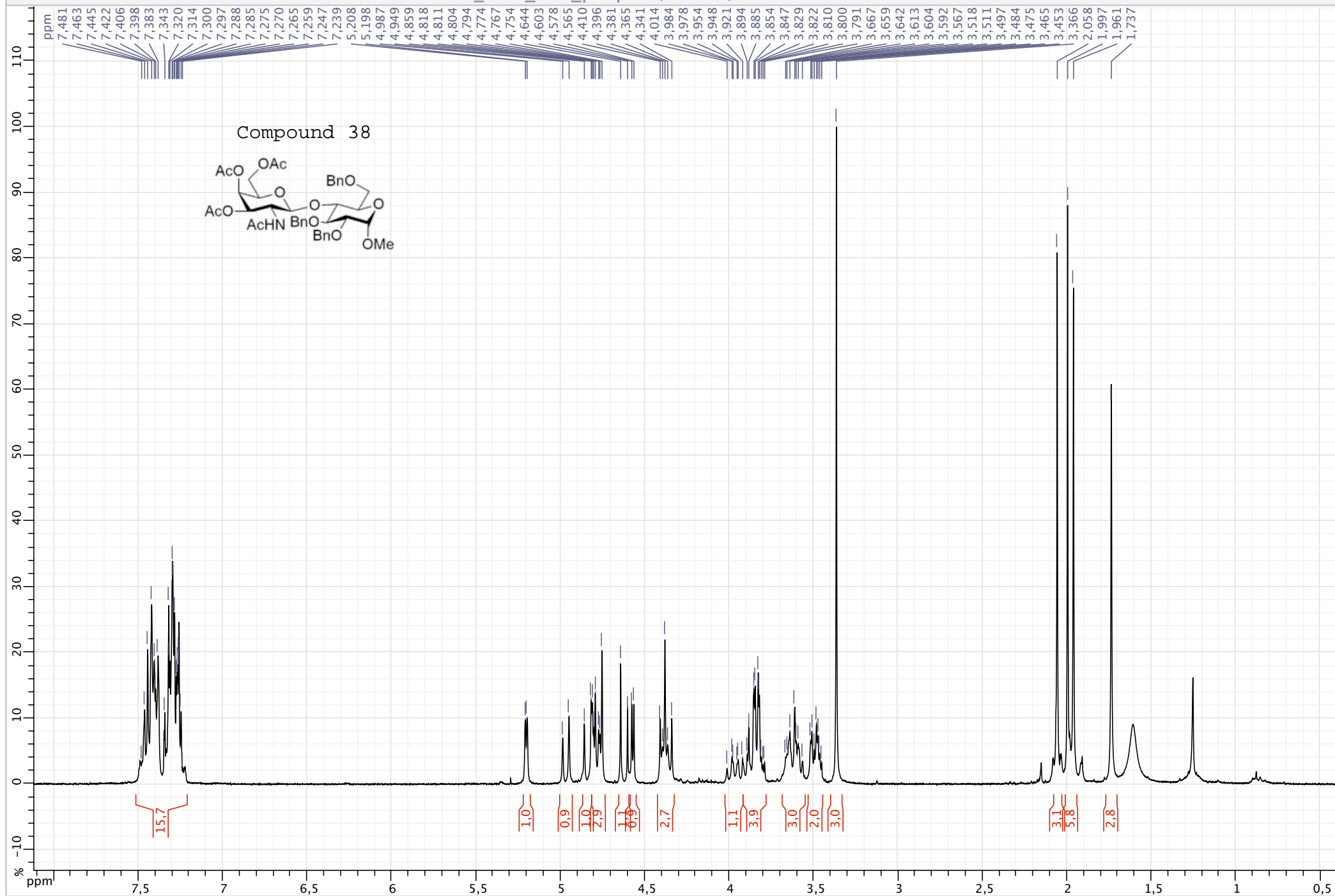


AX_268_F2 2 (1D 13C) CDCl3 300MHz

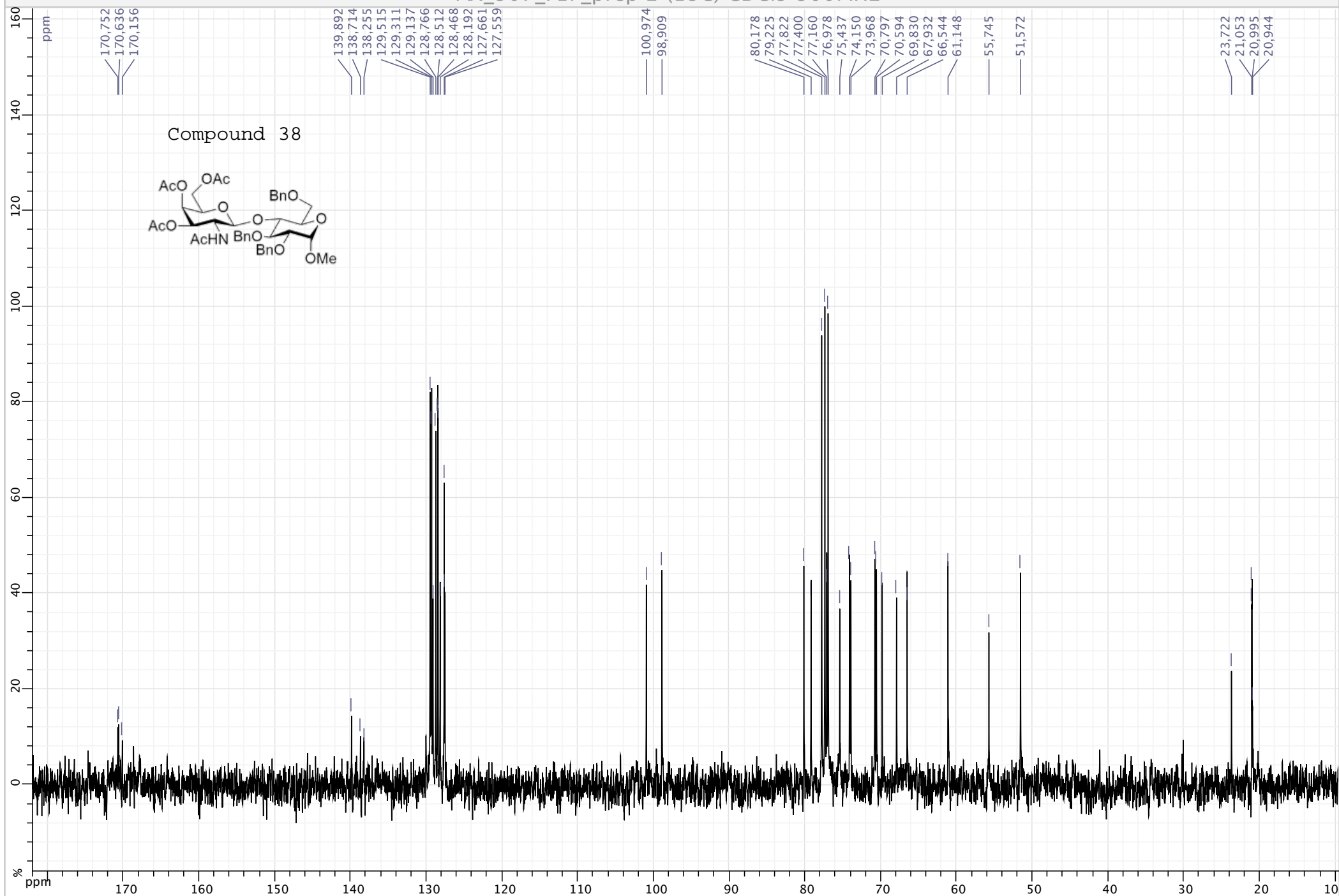
Compound 37



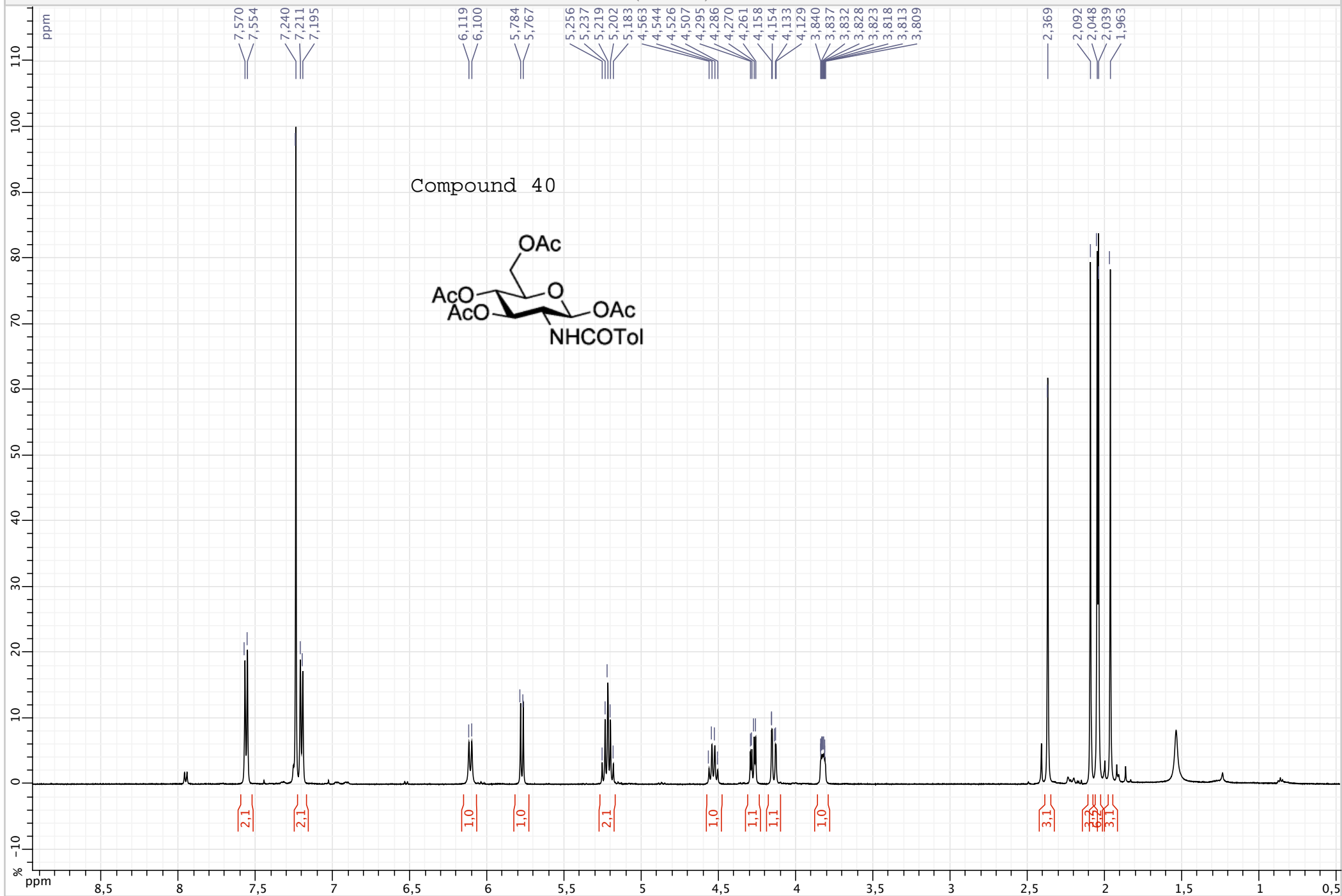
AX_307_F1P_prep 1 (1D 1H) CDCl3 300MHz



AX_307_F1P_prep 2 (13C) CDCl3 300MHz



41astdnhcotol 1 (1D 1H) CDCl3 500MHz



41astdnhcitol 3 (1D 13C) CDCl3 300MHz

