

Supporting Information for

**Asymmetric hydrogenation of 3-substituted 2*H*-1,4-benzoxazines
with chiral cationic Ru-MsDPEN catalysts: remarkable counteranion
effect**

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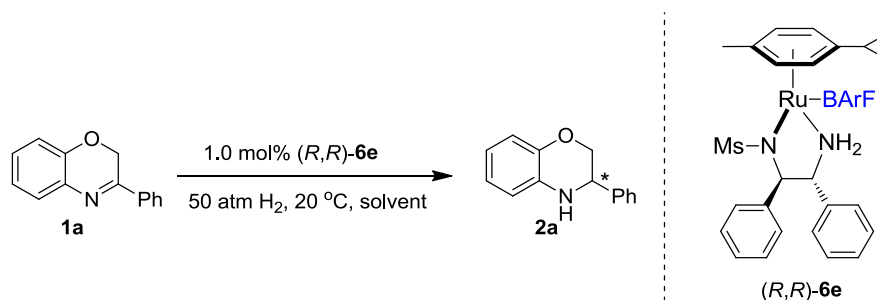
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1. General information

Unless otherwise noted, all experiments were carried out under an atmosphere of nitrogen using standard Schlenk techniques or in a nitrogen-filled glovebox. ^1H NMR and ^{13}C NMR spectra were recorded on a Bruker Model Avance DMX 300 Spectrometer (^1H 300 MHz and ^{13}C 75 MHz, respectively). Chemical shifts (δ) were given in ppm and were referenced to residual solvent or TMS peaks. Optical rotations were measured with PerkinElmer 341 polarimeter. High resolution MS (P-ESI HRMS) were obtained on Bruker Apex IV FTMS spectrometer. HPLC analyses were performed on a Varian Prostar 210 liquid chromatograph. All organic solvents were dried using standard, published methods and were distilled before use. All other chemicals were used as received from Aldrich or Acros without further purification. All Ru-catalysts were prepared according to the published method.¹ 3-Substituted-2*H*-1,4-benzoxazines **1** and **3** were synthesized according to modified literature method.²

2. Optimization of conditions for asymmetric hydrogenation

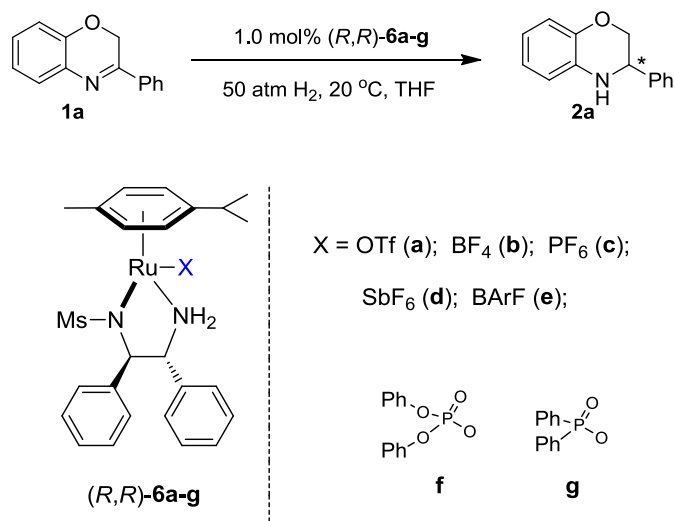
Table S1 Screening of the solvents for asymmetric hydrogenation of **1a** with (*R,R*)-**6e**^a



Entry	Solvent	Time (h)	Conv. (%) ^b	Ee (%) ^{c, d}
1	ClCH ₂ CH ₂ Cl	12	>95	19 (S)
2	CH ₂ Cl ₂	12	>95	15 (S)
3	toluene	12	>95	9 (S)
4	MeOH	12	>95	55 (R)
5	EtOH	12	>95	57 (R)
6	<i>i</i> -PrOH	12	>95	67 (R)
7	THF	12	>95	65 (R)
8	MeOH/CH ₂ Cl ₂ (1/1, v/v)	12	>95	45 (R)

^a Reaction conditions: **1a** (0.1 mmol) in solvent (1 mL), (*R,R*)-**6e** (1.0 mol%), H₂ (50 atm), stirred at 20 °C for 12 h. ^b The conversions were determined by ¹H NMR spectroscopy of the crude reaction mixture. ^c The enantiomeric excesses were determined by chiral HPLC with a chiral OD-H column. ^d The absolute configurations of the products were assigned by comparison of the optical rotations with those in the published literature.

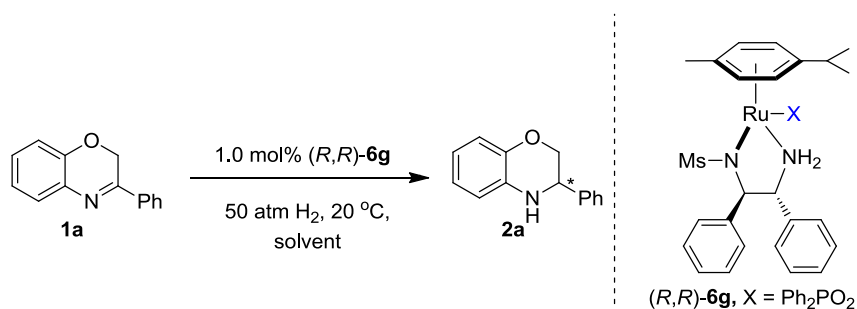
Table S2 Screening of the catalysts for asymmetric hydrogenation of **1a** in THF^a



Entry	Catalyst	X	Conv. (%) ^b	Ee (%) ^{c, d}
1	(R,R)-6a	OTf	>95	8 (<i>S</i>)
2	(R,R)-6b	BF ₄	>95	13 (<i>R</i>)
3	(R,R)-6c	PF ₆	>95	47 (<i>R</i>)
4	(R,R)-6d	SbF ₆	>95	55 (<i>R</i>)
5	(R,R)-6e	BArF	>95	65 (<i>R</i>)
6	(R,R)-6f	(PhO) ₂ PO ₂	92	83 (<i>R</i>)
7	(R,R)-6g	Ph₂PO₂	19	92 (<i>R</i>)

^a Reaction conditions: **1a** (0.1 mmol) in THF (1 mL), catalyst (1.0 mol%), H₂ (50 atm), stirred at 20 °C for 12 h. ^b The conversions were determined by ¹H NMR spectroscopy of the crude reaction mixture. ^c The enantiomeric excesses were determined by chiral HPLC with a chiral OD-H column. ^d The absolute configurations of the products were assigned by comparison of the optical rotations with those in the published literature.

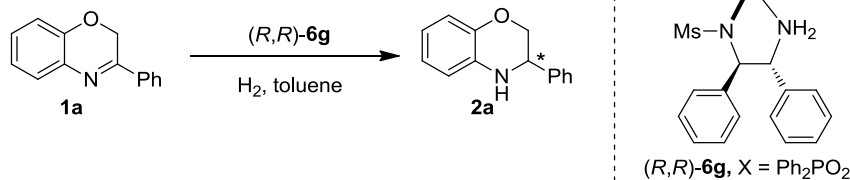
Table S3 Screening of the solvents for asymmetric hydrogenation of **1a** with (*R,R*)-**6g**^a



Entry	Solvent	Conv. (%) ^b	Ee (%) ^c
1	THF	19	92
2	CF ₃ CH ₂ OH	>95	35
3	CH ₂ Cl ₂	32	82
4	ClCH ₂ CH ₂ Cl	57	88
5	toluene	>95	94

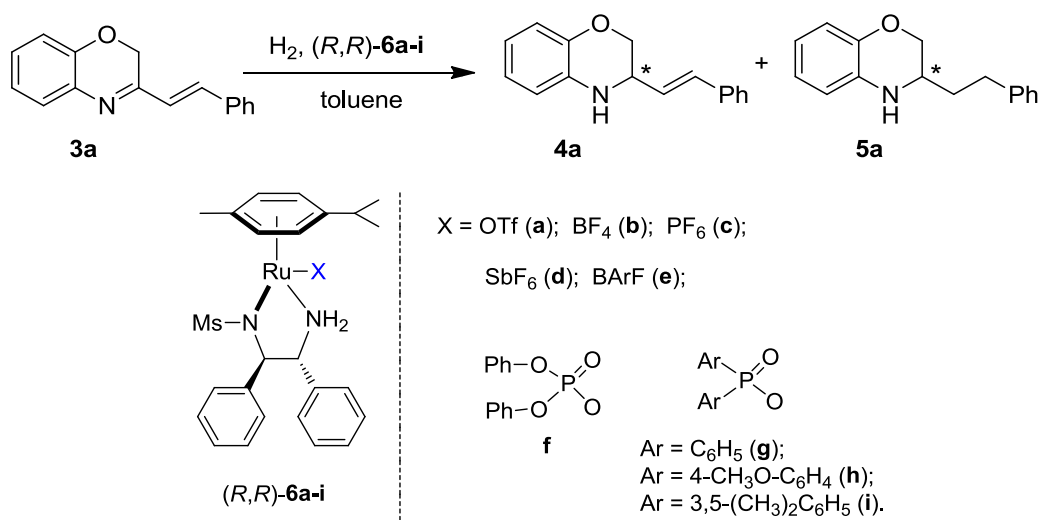
^a Reaction conditions: **1a** (0.1 mmol) in solvent (1 mL), (*R,R*)-**6g** (1.0 mol%), H₂ (50 atm), stirred at 20 °C for 12 h. ^b The conversions were determined by ¹H NMR spectroscopy of the crude reaction mixture. ^c The enantiomeric excesses were determined by chiral HPLC with a chiral OD-H column.

Table S4 Optimization of conditions for asymmetric hydrogenation of **1a** with *(R,R)*-**6g**^a



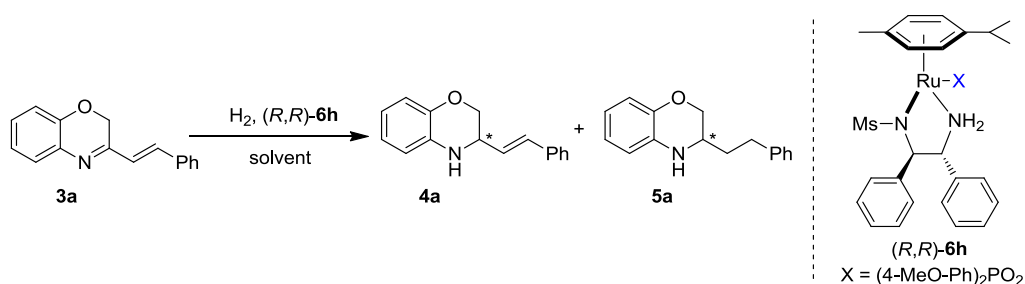
Entry	S/C	H ₂ (atm)	Temp. (°C)	Time (h)	Conv. (%) ^b	Ee (%) ^c
1	100	10	20	12	74	94
2	100	50	20	12	>95	94
3	100	80	20	12	>95	94
4	100	50	40	12	>95	94
5	100	50	0	12	79	94
6	100	50	40	6	>95	94
7	200	50	40	6	>95	94
8	500	50	40	24	>95	94
9	1000	50	40	24	40	93
10	1000	50	40	72	73	93

^a Reaction conditions: **1a** (0.1 mmol) in solvent (1 mL), *(R,R)*-**6g**. ^b The conversions were determined by ¹H NMR spectroscopy of the crude reaction mixture. ^c The enantiomeric excesses were determined by chiral HPLC with a chiral OD-H column.

Table S5: Screening of the catalysts for asymmetric hydrogenation of **3a** in toluene^a

Entry	Catalyst	X	Conv. (%) ^b	4a:5a ^c	Ee (%) ^d 4a/5a
1	(R,R)-6e	BArF	>95	33:67	75/92
2	(R,R)-6c	PF ₆	>95	70:30	8/14
3	(R,R)-6a	OTf	>95	89:11	47/ 0
4	(R,R)-6f	(PhO) ₂ PO ₂	>95	91:9	75/nd
5	(R,R)-6g	Ph ₂ PO ₂	>95	95:5	98/nd
6	(R,R)-6h	(4-MeO-Ph) ₂ PO ₂	>95	97:3	99/nd
7	(R,R)-6i	(3,5-Me ₂ -Ph) ₂ PO ₂	>95	96:4	98/nd
8^e	(R,R)-6h	(4-MeO-Ph)₂PO₂	>95	97:3	99/nd
9 ^f	(R,R)-6h	(4-MeO-Ph) ₂ PO ₂	>95	97:3	98/nd

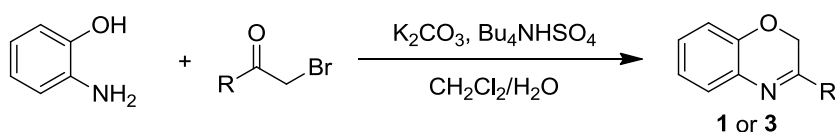
^a Reaction conditions: **3a** (0.1 mmol) in toluene (1 mL), catalyst (1.0 mol%), H₂ (50 atm), stirred at 20 °C for 24 h. ^b The conversions were determined by ¹H NMR spectroscopy of the crude reaction mixture. ^c The product ratios of **4a/5a** were determined by ¹H NMR spectroscopy of the crude reaction mixture. ^d The enantiomeric excesses were determined by chiral HPLC. ^e 0.5 mol% catalyst was used. ^f 0.2 mol% catalyst was used.

Table S6: Optimization of conditions for asymmetric hydrogenation of **3a**^a

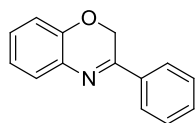
Entry	Solvent	H ₂ (atm)	Temp. (°C)	T (h)	Conv. (%) ^b	4a/5a ^c	Ee (%) ^d 4a/5a
1	MeOH	50	20	12	>95	53:47	28/50
2	CH ₂ Cl ₂	50	20	12	>95	97:3	95/nd
3	DCE	50	20	12	>95	97:3	94/nd
4	toluene	50	20	12	>95	98:2	99/nd
5	toluene	50	20	1.5	68	98:2	99/nd
6	toluene	10	20	1.5	30	96:4	99/nd
8	toluene	80	20	1.5	>95	98:2	99/nd
9	toluene	50	40	1.5	>95	97:3	99/nd

^a Reaction conditions: **3a** (0.1 mmol) in toluene (1 mL), **(R,R)-6h** (0.5 mol %). ^b The conversions were determined by ¹H NMR spectroscopy of the crude reaction mixture. ^c The product ratios of **4a/5a** were determined by ¹H NMR spectroscopy of the crude reaction mixture. ^d The enantiomeric excesses were determined by chiral HPLC.

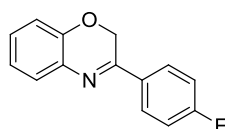
3. Synthesis of 3-substituted-2H-1,4-benzoxazines



General procedure:³ To a solution of 2-aminophenol (1.09 g, 10 mmol) in dichloromethane (60 mL) was added 20% aqueous K₂CO₃ solution (40 mL) and *n*-Bu₄NHSO₄ (10 mg). Then a solution of substituted 2-bromoacetophenone or (*E*)-1-bromo-4-phenylbut-3-en-2-one (10 mmol) in dichloromethane (25 mL) was added dropwise to the reaction mixture over a period of 15 min. The reaction mixture was stirred at room temperature and monitored by TLC. After the consumption of the starting materials, the reaction mixture was washed by water (50 mL) and brine (50 mL), then the organic layer was separated and dried by anhydrous Na₂SO₄. The solvent was removed on vacuum and the crude product was purified by column chromatography using dichloromethane as eluent to obtain the corresponding 3-substituted-2H-1,4-benzoxazines.

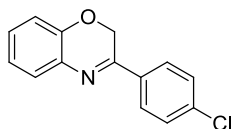


3-phenyl-2H-1,4-benzoxazine (1a): Yellow solid; 70% yield; ¹H NMR (300 MHz, CDCl₃): δ (ppm) 7.95-7.91 (m, 2H), 7.50-7.44 (m, 4H), 7.19-7.14 (m, 1H), 7.07-7.01 (m, 1H), 6.95-6.92 (m, 1H), 5.07 (s, 2H); ¹³C NMR (75 MHz, CDCl₃): δ (ppm) 158.8, 146.5, 135.6, 133.9, 131.3, 128.9, 128.8, 127.9, 126.6, 122.5, 115.7, 63.0.



3-(4-fluorophenyl)-2H-1,4-benzoxazine (1b): Yellow solid; 75% yield; ¹H NMR (300 MHz, CDCl₃): δ (ppm) 7.95-7.89 (m, 2H),

7.44-7.41 (m, 1H), 7.19-7.12 (m, 3H), 7.06-7.00 (m, 1H), 6.93-6.90 (m, 1H), 5.03 (s, 2H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 166.4, 163.0, 157.5, 146.3, 133.9, 131.8, 131.8, 128.8, 128.8, 128.7, 127.9, 122.5, 116.1, 115.9, 115.7, 62.8.



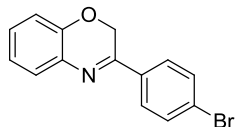
3-(4-chlorophenyl)-2H-1,4-benzoxazine (1c): Yellow solid; 68%

yield; ^1H NMR (300 MHz, CDCl_3): δ (ppm) 7.88-7.85 (m, 2H),

7.46-7.41 (m, 3H), 7.19-7.14 (m, 1H), 7.06-7.01 (m, 1H),

6.93-6.90 (m, 1H), 5.03 (s, 2H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 157.4, 146.3,

137.4, 133.9, 133.7, 129.2, 129.0, 128.0, 127.9, 122.6, 115.7, 62.8.



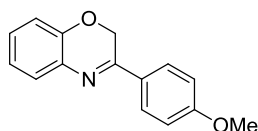
3-(4-bromophenyl)-2H-1,4-benzoxazine (1d): Yellow solid; 72%

yield; ^1H NMR (300 MHz, CDCl_3): δ (ppm) 7.81-7.76 (m, 2H),

7.62-7.58 (m, 2H), 7.44-7.41 (m, 1H), 7.19-7.13 (m, 1H), 7.06-7.00 (m, 1H),

6.93-6.90 (m, 1H), 5.02 (s, 2H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 157.5, 146.4,

134.4, 133.7, 132.1, 129.1, 128.1, 128.0, 125.9, 122.6, 115.7, 62.7.



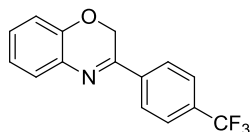
3-(4-methoxyphenyl)-2H-1,4-benzoxazine (1e): Yellow solid;

70% yield; ^1H NMR (300 MHz, CDCl_3): δ (ppm) 7.91-7.88 (m,

2H), 7.44-7.41 (m, 1H), 7.16-7.10 (m, 1H), 7.05-6.90 (m, 4H), 5.02 (s, 2H), 3.86 (s,

3H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 162.2, 158.3, 146.4, 134.0, 128.4, 128.2,

128.1, 127.6, 122.4, 115.6, 114.3, 62.8, 55.5.

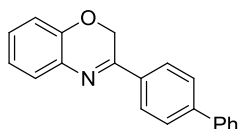


3-(4-(trifluoromethyl)phenyl)-2H-1,4-benzoxazine (1f): Yellow

solid; 75% yield; ^1H NMR (300 MHz, CDCl_3): δ (ppm)

8.04-8.01 (m, 2H), 7.74-7.71 (m, 2H), 7.47-7.44 (m, 1H), 7.22-7.16 (m, 1H),

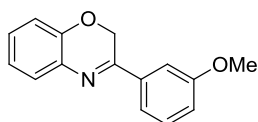
7.07-7.02 (m, 1H), 6.95-6.92 (m, 1H), 5.07 (s, 2H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 157.2, 146.4, 138.7, 133.7, 132.9, 132.5, 129.5, 128.3, 126.9, 125.9, 125.8, 122.7, 115.8, 62.9.



3-([1,1'-biphenyl]-4-yl)-2H-1,4-benzoxazine (1g): Yellow solid;

71% yield; ^1H NMR (300 MHz, CDCl_3): δ (ppm) 8.02-8.00 (m, 2H), 7.73-7.70 (m, 2H), 7.67-7.64 (m, 2H), 7.51-7.46 (m, 3H),

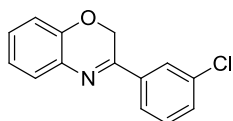
7.43-7.38 (m, 1H), 7.20-7.14 (m, 1H), 6.96-6.93 (m, 1H), 5.10 (s, 2H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 158.4, 146.5, 144.0, 140.2, 134.4, 134.0, 129.1, 128.8, 128.1, 128.0, 127.5, 127.3, 127.1, 122.5, 115.7, 63.0.



3-(3-methoxyphenyl)-2H-1,4-benzoxazine (1h): Yellow solid;

71% yield; ^1H NMR (300 MHz, CDCl_3): δ (ppm) 7.57-7.56 (m,

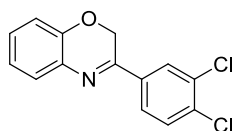
1H), 7.47-7.35 (m, 3H), 7.19-7.13 (m, 1H), 7.07-7.00 (m, 2H), 6.94-6.91 (m, 1H), 5.05 (s, 2H), 3.89 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 160.1, 158.6, 146.5, 137.0, 133.9, 129.8, 128.8, 128.0, 122.5, 119.0, 117.6, 115.7, 111.3, 63.1, 55.6.



3-(3-chlorophenyl)-2H-1,4-benzoxazine (1i): Yellow solid; 71%

yield; ^1H NMR (300 MHz, CDCl_3): δ (ppm) 7.96-7.95 (m, 1H),

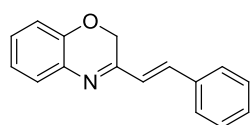
7.76-7.73 (m, 1H), 7.48-7.38 (m, 3H), 7.20-7.14 (m, 1H), 7.06-7.01 (m, 1H), 6.94-6.90 (m, 1H), 5.03 (s, 2H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 157.3, 146.4, 137.3, 135.2, 133.7, 131.2, 130.1, 129.2, 128.1, 126.7, 124.6, 122.6, 115.8, 62.9.



3-(3,4-dichlorophenyl)-2H-1,4-benzoxazine (1j): Yellow solid;

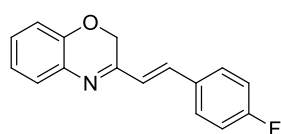
71% yield; ^1H NMR (300 MHz, CDCl_3): δ (ppm) 8.04-8.03 (m,

1H), 7.72-7.68 (m, 1H), 7.54-7.51 (m, 1H), 7.43-7.40 (m, 1H), 7.20-7.14 (m, 1H), 7.06-7.00 (m, 1H), 6.93-6.89 (m, 1H), 4.99 (s, 2H); ¹³C NMR (75 MHz, CDCl₃): δ (ppm) 156.1, 146.3, 135.4, 135.3, 133.5, 133.5, 130.8, 129.4, 128.5, 128.1, 125.6, 122.7, 115.8, 62.6.



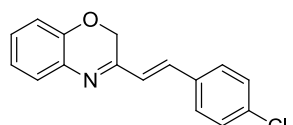
(E)-3-styryl-2H-1,4-benzoxazine (3a): Yellow solid; 71% yield;

¹H NMR (300 MHz, CDCl₃): δ (ppm) 7.56-7.53 (m, 2H), 7.43-7.33 (m, 4H), 7.17-7.11 (m, 1H), 7.06-6.99 (m, 3H), 6.92-6.89 (dd, *J* = 8.0, 1.2 Hz, 1H), 4.92 (s, 2H); ¹³C NMR (75 MHz, CDCl₃): δ (ppm) 159.0, 146.7, 137.4, 135.6, 134.2, 129.8, 129.1, 128.9, 127.6, 126.9, 122.6, 115.8, 62.2.



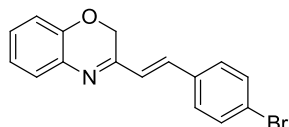
(E)-3-(4-fluorostyryl)-2H-1,4-benzoxazine (3b): Yellow

solid; 75% yield; ¹H NMR (300 MHz, CDCl₃): δ (ppm) 7.54-7.49 (m, 2H), 7.36-7.33 (m, 1H), 7.16-6.95 (m, 6H), 6.91-6.88 (m, 1H), 4.89 (s, 2H); ¹³C NMR (75 MHz, CDCl₃): δ (ppm) 165.2, 161.9, 158.8, 146.7, 135.9, 134.2, 131.9, 129.4, 129.3, 128.9, 127.7, 126.8, 122.6, 116.4, 116.1, 115.8, 62.2.



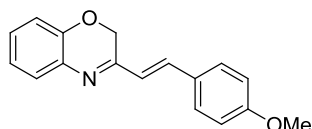
(E)-3-(4-chlorostyryl)-2H-1,4-benzoxazine (3c): Yellow

solid; 80% yield; ¹H NMR (300 MHz, CDCl₃): δ (ppm) 7.47-7.44 (m, 2H), 7.37-7.33 (m, 3H), 7.16-7.11 (m, 1H), 7.05-6.99 (m, 3H), 6.93-6.88 (m, 1H), 4.89 (s, 2H); ¹³C NMR (75 MHz, CDCl₃): δ(ppm) 158.7, 146.7, 135.8, 135.5, 134.2, 134.1, 129.4, 129.0, 128.7, 127.8, 127.6, 122.6, 115.8, 62.2.



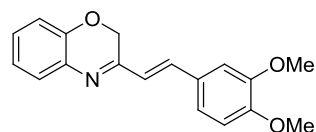
(E)-3-(4-bromostyryl)-2H-1,4-benzoxazine (3d): Yellow

solid; 75% yield; ^1H NMR (300 MHz, CDCl_3): δ (ppm) 7.54-7.50 (m, 2H), 7.41-7.34 (m, 3H), 7.17-7.11 (m, 1H), 7.04-6.98 (m, 3H), 6.91-6.88 (m, 1H), 4.89 (s, 2H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 158.7, 146.7, 135.8, 134.6, 134.2, 132.3, 129.0, 128.9, 127.8, 127.7, 123.8, 122.7, 115.8, 62.2. HRMS-ESI exact mass calcd. for $\text{C}_{16}\text{H}_{13}\text{BrNO}^+$ ($[\text{M}+\text{H}]^+$) requires m/z 314.01797, found m/z 314.01750.



(E)-3-(4-methoxystyryl)-2H-1,4-benzoxazine (3e): Yellow

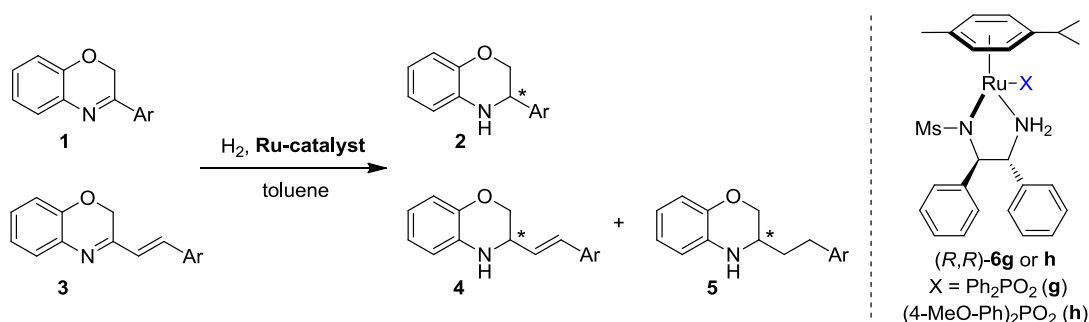
solid; 78% yield; ^1H NMR (300 MHz, CDCl_3): δ (ppm) 7.50-7.47 (m, 2H), 7.36-7.33 (m, 1H), 7.14-7.08 (m, 1H), 7.05-6.97 (m, 2H), 6.93-6.86 (m, 4H), 4.88 (s, 2H), 3.83 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 161.0, 159.2, 146.7, 137.0, 134.4, 129.1, 128.5, 128.3, 127.5, 124.8, 122.5, 115.7, 114.6, 62.2, 55.5. HRMS-ESI exact mass calcd. for $\text{C}_{17}\text{H}_{16}\text{NO}_2^+$ ($[\text{M}+\text{H}]^+$) requires m/z 266.11775, found m/z 266.11756.



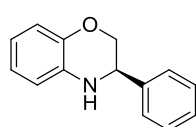
(E)-3-(3,4-dimethoxystyryl)-2H-1,4-benzoxazine (3f):

Yellow solid; 73% yield; ^1H NMR (300 MHz, CDCl_3): δ (ppm) 7.35-7.31 (m, 1H), 7.14-6.97 (m, 5H), 6.92-6.85 (m, 3H), 4.89 (s, 2H), 3.91 (s, 3H), 3.90 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 159.1, 150.8, 149.5, 146.7, 137.2, 134.4, 128.6, 128.5, 127.5, 125.0, 122.6, 122.1, 115.7, 111.2, 108.8, 62.1, 56.1, 56.0. HRMS-ESI exact mass calcd. for $\text{C}_{18}\text{H}_{18}\text{NO}_3^+$ ($[\text{M}+\text{H}]^+$) requires m/z 296.12850, found m/z 296.12812.

4. Asymmetric hydrogenation of 3-substituted-2*H*-1,4-benzoxazines



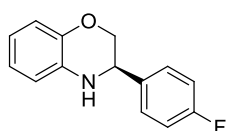
A 50 mL glass-lined stainless-steel reactor equipped with a magnetic stirrer bar was charged with Ru-catalyst (0.001 mmol, 0.5 mol%) and the corresponding 3-substituted-2*H*-1,4-benzoxazines (0.2 mmol) in toluene (2 mL) under nitrogen atmosphere in a glove box. The autoclave was closed, and the final pressure of the hydrogen gas was adjusted to 50 atm after purging the autoclave with hydrogen gas several times. The reaction mixture was stirred at 40 °C for 12 h. Then the hydrogen gas was carefully released and the conversion was determined by ^1H NMR spectroscopy after the solvent was evaporated under reduced pressure. The reaction mixture was filtered through a short pad of silica eluted with DCM to give the pure products. The enantiomeric excess of the product was determined by HPLC with a chiral OD-H column.



(*R*)-3-phenyl-3,4-dihydro-2*H*-1,4-benzoxazine (2a): known

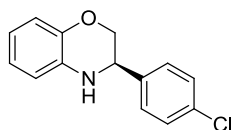
compound,³ Yellow oil; 95% yield, 94% ee, $[\alpha]_{\text{D}}^{20} = -134.5$ (c 1.0, CHCl_3), {Lit.^[2] $[\alpha]_{\text{D}}^{20} = -118.1$ (c 1.0, CHCl_3) for 98% ee of (*R*)-enantiomer}; ^1H NMR (300 MHz, CDCl_3): δ (ppm) 7.44-7.34 (m, 5H), 6.91-6.683 (m, 2H), 6.77-6.69 (m, 2H), 4.54-4.50 (dd, $J_1 = 2.8$ Hz, $J_2 = 8.6$ Hz, 1H), 4.34-4.29 (dd, $J_1 = 2.9$ Hz, $J_2 =$

10.7 Hz, 1H), 4.06-4.00 (m, 2H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 143.6, 139.2, 133.9, 128.9, 128.4, 127.3, 121.6, 119.1, 116.7, 115.6, 71.0, 54.3; HPLC (OD-H, elute: Hexane / *i*-PrOH = 80 / 20, detector: 254 nm, flow rate: 1 mL / min), (*R*) t_1 = 9.9 min (major), (*S*) t_2 = 13.7 min (minor).



(*R*)-3-(4-fluorophenyl)-3,4-dihydro-2H-1,4-benzoxazine (2b):

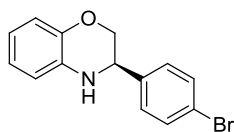
Yellow oil; 98% yield, 95% ee, $[\alpha]_{\text{D}}^{20}$ = -139.8 (*c* 1.0, CHCl_3), {Lit.³ $[\alpha]_{\text{D}}^{20}$ = +74.8 (*c* 1.2, CHCl_3) for 88% ee of (*S*)-enantiomer}; ^1H NMR (300 MHz, CDCl_3): δ (ppm) 7.41-7.36 (m, 2H), 7.12-7.06 (m, 2H), 6.89-6.81 (m, 2H), 6.76-6.67 (m, 2H), 4.51-4.47 (dd, J_1 = 2.9 Hz, J_2 = 8.5 Hz, 1H), 4.29-4.24 (dd, J_1 = 3.0 Hz, J_2 = 10.6 Hz, 1H), 4.04-3.94 (m, 2H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 164.4, 161.1, 143.6, 135.1, 135.0, 133.8, 129.0, 128.9, 121.7, 119.2, 116.7, 116.0, 115.7, 115.6, 71.0, 53.6; HPLC (OD-H, elute: Hexane / *i*-PrOH = 80 / 20, detector: 254 nm, flow rate: 1 mL / min), (*R*) t_1 = 9.9 min (major), (*S*) t_2 = 15.4 min (minor).



(*R*)-3-(4-chlorophenyl)-3,4-dihydro-2H-1,4-benzoxazine (2c):

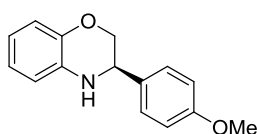
Yellow oil; 97% yield, 96% ee, $[\alpha]_{\text{D}}^{20}$ = -131.3 (*c* 1.0, CHCl_3), {Lit.^[3] $[\alpha]_{\text{D}}^{20}$ = +59.1 (*c* 0.7, CHCl_3) for 90% ee of (*S*)-enantiomer}; ^1H NMR (300 MHz, CDCl_3): δ (ppm) 7.39-7.32 (m, 4H), 6.87-6.80 (m, 2H), 6.74-6.67 (m, 2H), 4.52-4.48 (dd, J_1 = 2.8 Hz, J_2 = 8.3 Hz, 1H), 4.28-4.24 (dd, J_1 = 2.9 Hz, J_2 = 10.7 Hz, 1H), 4.00-3.93 (m, 2H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 143.6, 137.9, 134.2, 133.7, 129.2, 128.7, 121.7, 119.3, 116.8, 115.6, 70.9, 53.8; HPLC (OD-H, elute: Hexane / *i*-PrOH = 80 / 20, detector: 254 nm, flow rate: 1 mL / min), (*R*) t_1 = 11.6 min

(major), (*S*) t_2 = 20.4 min (minor).



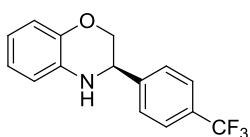
(*R*)-3-(4-bromophenyl)-3,4-dihydro-2*H*-1,4-benzoxazine (2d):

Yellow oil; 95% yield, 96% ee, $[\alpha]_D^{20} = -111.5$ (c 1.0, CHCl_3), {Lit.³ $[\alpha]_D^{20} = +89.5$ (c 1.03, CHCl_3) for 89% ee of (*S*)-enantiomer}; ^1H NMR (300 MHz, CDCl_3): δ (ppm) 7.55-7.50 (m, 2H), 7.30-7.27 (m, 2H), 6.89-6.81 (m, 2H), 6.76-6.67 (m, 2H), 4.48-4.45 (dd, $J_1 = 2.9$ Hz, $J_2 = 8.3$ Hz, 1H), 4.28-4.23 (dd, $J_1 = 2.9$ Hz, $J_2 = 10.7$ Hz, 1H), 4.00-3.93 (m, 2H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 143.6, 138.3, 133.6, 132.0, 129.0, 122.3, 121.7, 119.3, 117.0, 115.6, 70.7, 53.7; HPLC (OD-H, elute: Hexane / *i*-PrOH = 80 / 20, detector: 254 nm, flow rate: 1 mL / min), (*R*) $t_1 = 12.6$ min (major), (*S*) $t_2 = 22.5$ min (minor).



(*R*)-3-(4-methoxyphenyl)-3,4-dihydro-2*H*-1,4-benzoxazine (2e):

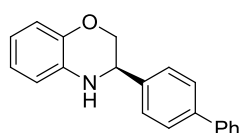
Yellow oil; 96% yield, 98% ee, $[\alpha]_D^{20} = -126.5$ (c 1.0, CHCl_3), {Lit.^[2] $[\alpha]_D^{20} = -55.5$ (c 1.0, CHCl_3) for 98% ee of (*R*)-enantiomer}; ^1H NMR (300 MHz, CDCl_3): δ (ppm) 7.35-7.31 (m, 2H), 6.96-6.92 (m, 2H), 6.89-6.80 (m, 2H), 6.75-6.66 (m, 2H), 4.47-4.43 (dd, $J_1 = 2.8$ Hz, $J_2 = 8.6$ Hz, 1H), 4.28-4.24 (dd, $J_1 = 2.9$ Hz, $J_2 = 10.7$ Hz, 1H), 4.01-3.95 (m, 2H), 3.83 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 159.7, 143.6, 134.1, 131.2, 128.4, 121.5, 119.0, 116.4, 115.5, 114.3, 71.2, 55.4, 53.7; HPLC (OD-H, elute: Hexane / *i*-PrOH = 80 / 20, detector: 254 nm, flow rate: 1 mL / min), (*R*) $t_1 = 10.5$ min (major), (*S*) $t_2 = 15.9$ min (minor).



(*R*)-3-(4-(trifluoromethyl)phenyl)-3,4-dihydro-2*H*-1,4-benzoxazine (2f):

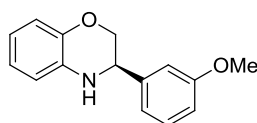
Yellow oil; 97% yield, 92% ee, $[\alpha]_D^{20} = -122.8$ (c

1.0, CHCl₃), {Lit.^[3] $[\alpha]_D^{20} = +63.1$ (*c* 1.48, CHCl₃) for 89% ee of (*S*)-enantiomer}; ¹H NMR (300 MHz, CDCl₃): δ (ppm) 7.68-7.66 (m, 2H), 7.55-7.53 (m, 2H), 6.90-6.83 (m, 2H), 6.78-6.70 (m, 2H), 4.60-4.57 (dd, $J_1 = 2.9$ Hz, $J_2 = 8.1$ Hz, 1H), 4.32-4.28 (dd, $J_1 = 2.9$ Hz, $J_2 = 10.7$ Hz, 1H), 4.04-3.98 (m, 2H); ¹³C NMR (75 MHz, CDCl₃): δ (ppm) 143.6, 143.5, 133.5, 130.9, 130.4, 127.7, 125.9, 125.9, 125.8, 122.3, 121.9, 119.4, 116.9, 115.7, 70.6, 54.0; HPLC (OD-H, elute: Hexane / *i*-PrOH = 80 / 20, detector: 254 nm, flow rate: 1 mL / min), (*R*) $t_1 = 11.3$ min (major), (*S*) $t_2 = 16.4$ min (minor).



(*R*)-3-([1,1'-biphenyl]-4-yl)-3,4-dihydro-2H-1,4-benzoxazine

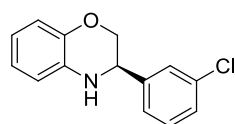
(2g): Yellow oil; 95% yield, 97% ee, $[\alpha]_D^{20} = -129.2$ (*c* 1.0, CHCl₃), {Lit.^[2] $[\alpha]_D^{20} = -53.4$ (*c* 1.0, CHCl₃) for 98% ee of (*R*)-enantiomer}; ¹H NMR (300 MHz, CDCl₃): δ (ppm) 7.67-7.63 (m, 4H), 7.53-7.48 (m, 4H), 7.44-7.39 (m, 1H), 6.95-6.86 (m, 2H), 6.80-6.71 (m, 2H), 4.59-4.55 (dd, $J_1 = 2.9$ Hz, $J_2 = 8.5$ Hz, 1H), 4.38-4.34 (dd, $J_1 = 2.9$ Hz, $J_2 = 10.7$ Hz, 1H), 4.11-4.05 (m, 2H); ¹³C NMR (75 MHz, CDCl₃): δ (ppm) 143.6, 141.4, 140.7, 138.3, 133.9, 129.0, 127.7, 127.6, 127.2, 121.6, 119.1, 116.7, 115.5, 71.0, 54.0; HPLC (OD-H, elute: Hexane / *i*-PrOH = 80 / 20, detector: 254 nm, flow rate: 1 mL / min), (*S*) $t_1 = 17.7$ min (major), (*R*) $t_2 = 25.5$ min (minor).



(*R*)-3-(3-methoxyphenyl)-3,4-dihydro-2H-1,4-benzoxazine

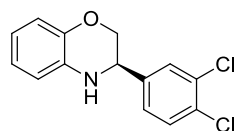
(2h): Yellow oil; 95% yield, 90% ee, $[\alpha]_D^{20} = -129.8$ (*c* 1.0, CHCl₃), {Lit.^[2] $[\alpha]_D^{20} = +84.6$ (*c* 0.87, CHCl₃) for 87% ee of (*S*)-enantiomer}; ¹H

NMR (300 MHz, CDCl₃): δ (ppm) 7.35-7.30 (m, 1H), 7.02-6.99 (m, 2H), 6.92-6.82 (m, 3H), 6.76-6.68 (m, 2H), 4.50-4.46 (dd, $J_1 = 2.9$ Hz, $J_2 = 8.6$ Hz, 1H), 4.33-4.28 (dd, $J_1 = 2.9$ Hz, $J_2 = 10.7$ Hz, 1H), 4.04-3.98 (m, 2H), 3.84 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ (ppm) 160.1, 143.6, 140.9, 133.9, 129.9, 121.6, 119.6, 119.0, 116.7, 115.5, 113.8, 112.8, 71.0, 55.4, 54.2; HPLC (OD-H, elute: Hexane / *i*-PrOH = 80 / 20, detector: 254 nm, flow rate: 1 mL / min), (*R*) $t_1 = 17.3$ min (major), (*S*) $t_2 = 19.8$ min (minor).



(*R*)-3-(3-chlorophenyl)-3,4-dihydro-2*H*-1,4-benzoxazine (2i):

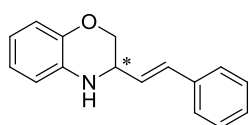
Yellow oil; 94% yield, 82% ee, $[\alpha]_D^{20} = -123.2$ (*c* 1.0, CHCl₃), {Lit.^[2] $[\alpha]_D^{20} = +71.0$ (*c* 1.38, CHCl₃) for 88% ee of (*S*)-enantiomer}; ¹H NMR (300 MHz, CDCl₃): δ (ppm) 7.42 (s, 1H), 7.34-7.29 (m, 3H), 6.89-6.82 (m, 2H), 6.76-6.68 (m, 2H), 4.50-4.46 (dd, $J_1 = 2.9$ Hz, $J_2 = 8.3$ Hz, 1H), 4.30-4.26 (dd, $J_1 = 2.9$, $J_2 = 10.7$ Hz, 1H), 4.01-3.94 (m, 2H); ¹³C NMR (75 MHz, CDCl₃): δ (ppm) 143.6, 141.4, 134.9, 133.6, 130.2, 128.6, 127.4, 125.5, 121.7, 119.3, 116.8, 115.6, 70.7, 53.9. HPLC (OD-H, elute: Hexane / *i*-PrOH = 80 / 20, detector: 254 nm, flow rate: 1 mL / min), (*R*) $t_1 = 11.0$ min (major), (*S*) $t_2 = 15.3$ min (minor).



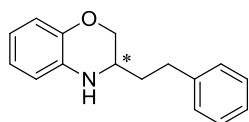
(*R*)-3-(3,4-dichlorophenyl)-3,4-dihydro-2*H*-1,4-benzoxazine

(2j): Yellow oil; 98% yield, 86% ee, $[\alpha]_D^{20} = -117.8$ (*c* 1.0, CHCl₃), {Lit.^[2] $[\alpha]_D^{20} = +52.9$ (*c* 1.12, CHCl₃) for 92% ee of (*S*)-enantiomer}; ¹H NMR (300 MHz, CDCl₃): δ (ppm) 7.48-7.47 (m, 1H), 7.44-7.41 (m, 1H), 7.23-7.19 (m, 1H), 6.85-6.78 (m, 2H), 6.73-6.65 (m, 2H), 4.45-4.41 (dd, $J_1 = 2.9$ Hz, $J_2 = 8.2$ Hz,

1H), 4.24-4.20 (dd, $J_1 = 2.9$ Hz, $J_2 = 10.7$ Hz, 1H), 3.95-3.88 (m, 2H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 143.5, 139.7, 133.3, 133.1, 132.4, 130.9, 129.2, 126.6, 121.8, 119.5, 116.8, 115.7, 70.5, 53.3; HPLC (OD-H, elute: Hexane / *i*-PrOH = 80 / 20, detector: 254 nm, flow rate: 1 mL / min), (*R*) $t_1 = 14.0$ min (major), (*S*) $t_2 = 23.8$ min (minor).

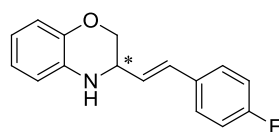


(-)-(E)-3-styryl-3,4-dihydro-2H-1,4-benzoxazine (4a): Yellow oil; 95% yield, 99% ee, $[\alpha]_{\text{D}}^{20} = -34.5$ (*c* 1.0, CHCl_3); ^1H NMR (300 MHz, CDCl_3): δ (ppm) 7.41-7.24 (m, 5H), 6.84-6.64 (m, 5H), 6.23-6.15 (m, 1H), 4.32-4.28 (m, 1H), 4.17-4.12 (m, 1H), 4.02-3.96 (m, 1H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 143.7, 136.3, 133.1, 133.0, 128.8, 128.1, 126.6, 126.5, 121.7, 119.0, 116.7, 115.5, 69.1, 52.4; HPLC (AD-H, elute: Hexane / *i*-PrOH = 90 / 10, detector: 254 nm, flow rate: 1 mL / min), $t_1 = 9.8$ min (minor), $t_2 = 11.1$ min (major). HRMS-ESI exact mass calcd. for $\text{C}_{16}\text{H}_{16}\text{NO}^+$ ($[\text{M}+\text{H}]^+$) requires m/z 238.12297, found m/z 238.12264.



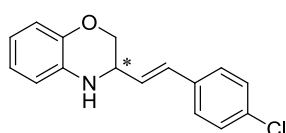
(S)-3-phenethyl-3,4-dihydro-2H-1,4-benzoxazine (5a): Yellow solid; m.p. 89-90 °C; yield 91%, 89% ee, $[\alpha]_{\text{D}}^{20} = +82.5$ (*c* 1.0, CHCl_3); ^1H NMR (300 MHz, CDCl_3): δ (ppm) 7.31-7.26 (m, 2H), 7.21-7.17 (m, 3H), 6.79-6.71 (m, 2H), 6.66-6.60 (m, 1H), 6.54-6.51 (m, 1H), 4.20-4.16 (m, 1H), 3.89-3.83 (m, 1H), 3.65 (b, 1H), 3.41-3.33 (m, 1H), 2.75-2.70 (m, 2H), 1.83-1.75 (m, 2H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 143.9, 141.2, 133.3, 128.7, 128.4, 126.3, 121.5, 118.8, 116.6, 115.6, 69.3, 49.3, 33.9, 32.0; HPLC (OD-H, elute: Hexane / *i*-PrOH = 80 / 20, detector: 254 nm, flow rate: 1 mL / min), (*S*) $t_1 = 14.4$ min (major),

(*R*) t_2 = 17.9 min (minor).



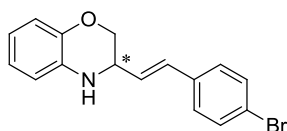
(-)-(E)-3-(4-fluorostyryl)-3,4-dihydro-2H-1,4-benzoxazine

(3b): Yellow oil; 96% yield, 91% ee, $[\alpha]_D^{20} = -28.6$ (c 1.0, CHCl_3); ^1H NMR (300 MHz, CDCl_3): δ (ppm) 7.38-7.34 (m, 2H), 7.05-6.99 (m, 2H), 6.85-6.78 (m, 2H), 6.72-6.64 (m, 3H), 6.14-6.06 (m, 1H), 4.31-4.27 (m, 1H), 4.15-4.10 (m, 1H), 4.02-3.96 (m, 1H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 164.3, 161.0, 143.7, 133.1, 132.5, 132.5, 131.8, 128.2, 128.1, 126.3, 126.3, 121.7, 119.1, 116.7, 115.8, 115.6, 69.1, 52.3; HPLC (AD-H, elute: Hexane / *i*-PrOH = 90 / 10, detector: 254 nm, flow rate: 1 mL / min), t_1 = 10.1 min (minor), t_2 = 13.5 min (major). HRMS-ESI exact mass calcd. for $\text{C}_{16}\text{H}_{15}\text{FNO}^+$ ($[\text{M}+\text{H}]^+$) requires m/z 256.11359, found m/z 256.11322.



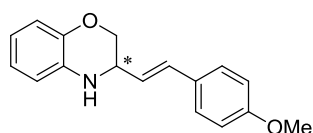
(-)-(E)-3-(4-chlorostyryl)-3,4-dihydro-2H-1,4-benzoxazine

(3c): Yellow oil; 94% yield, 98% ee, $[\alpha]_D^{20} = -35.2$ (c 1.0, CHCl_3); ^1H NMR (300 MHz, CDCl_3): δ (ppm) 7.33-7.28 (m, 4H), 6.85-6.78 (m, 2H), 6.73-6.64 (m, 3H), 6.21-6.13 (m, 1H), 4.31-4.27 (m, 1H), 4.16-4.10 (m, 1H), 4.02-3.96 (m, 1H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 143.7, 134.8, 133.8, 133.0, 131.7, 128.9, 127.8, 127.3, 121.7, 119.1, 116.7, 115.6, 69.0, 52.3; HPLC (AD-H, elute: Hexane / *i*-PrOH = 90 / 10, detector: 254 nm, flow rate: 1 mL / min), t_1 = 11.2 min (minor), t_2 = 15.4 min (major). HRMS-ESI exact mass calcd. for $\text{C}_{16}\text{H}_{15}\text{ClNO}^+$ ($[\text{M}+\text{H}]^+$) requires m/z 272.08405, found m/z 272.08367.



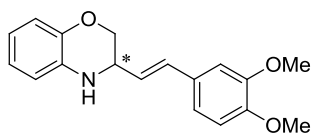
(-)-(E)-3-(4-bromostyryl)-3,4-dihydro-2H-1,4-benzoxazine

e (3d): Yellow oil; 93% yield, 98% ee, $[\alpha]_D^{20} = -32.9$ (c 1.0, CHCl_3); ^1H NMR (300 MHz, CDCl_3): δ (ppm) 7.24-7.14 (m, 4H), 6.83-6.75 (m, 2H), 6.70-6.56 (m, 3H), 6.17-6.09 (m, 1H), 4.27-4.22 (m, 1H), 4.10-4.04 (m, 1H), 3.98-3.92 (m, 1H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 143.7, 134.8, 133.8, 133.0, 131.7, 128.9, 127.8, 127.3, 121.7, 119.1, 116.7, 115.6, 69.0, 52.3; HPLC (AD-H, elute: Hexane / *i*-PrOH = 90 / 10, detector: 254 nm, flow rate: 1 mL / min), $t_1 = 12.2$ min (minor), $t_2 = 16.9$ min (major). HRMS-ESI exact mass calcd. for $\text{C}_{16}\text{H}_{15}\text{BrNO}^+$ ($[\text{M}+\text{H}]^+$) requires m/z 316.03343, found m/z 316.03315.



(-)-(E)-3-(4-methoxystyryl)-3,4-dihydro-2H-1,4-benzoxazine (3e):

Yellow oil; 95% yield, 97% ee, $[\alpha]_D^{20} = -38.9$ (c 1.0, CHCl_3); ^1H NMR (300 MHz, CDCl_3): δ (ppm) 7.24-7.14 (m, 4H), 6.83-6.75 (m, 2H), 6.70-6.56 (m, 3H), 6.17-6.09 (m, 1H), 4.27-4.22 (m, 1H), 4.10-4.04 (m, 1H), 3.98-3.92 (m, 1H), 3.77 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 143.7, 134.8, 133.8, 133.0, 131.7, 128.9, 127.8, 127.3, 121.7, 119.1, 116.7, 115.6, 69.0, 55.9, 52.3; HPLC (AD-H, elute: Hexane / *i*-PrOH = 90 / 10, detector: 254 nm, flow rate: 1 mL / min), $t_1 = 14.1$ min (minor), $t_2 = 17.6$ min (major). HRMS-ESI exact mass calcd. for $\text{C}_{17}\text{H}_{18}\text{NO}_2^+$ ($[\text{M}+\text{H}]^+$) requires m/z 268.13377, found m/z 268.13321.



(-)-(E)-3-(3,4-dimethoxystyryl)-3,4-dihydro-2H-1,4-benzoxazine (3f):

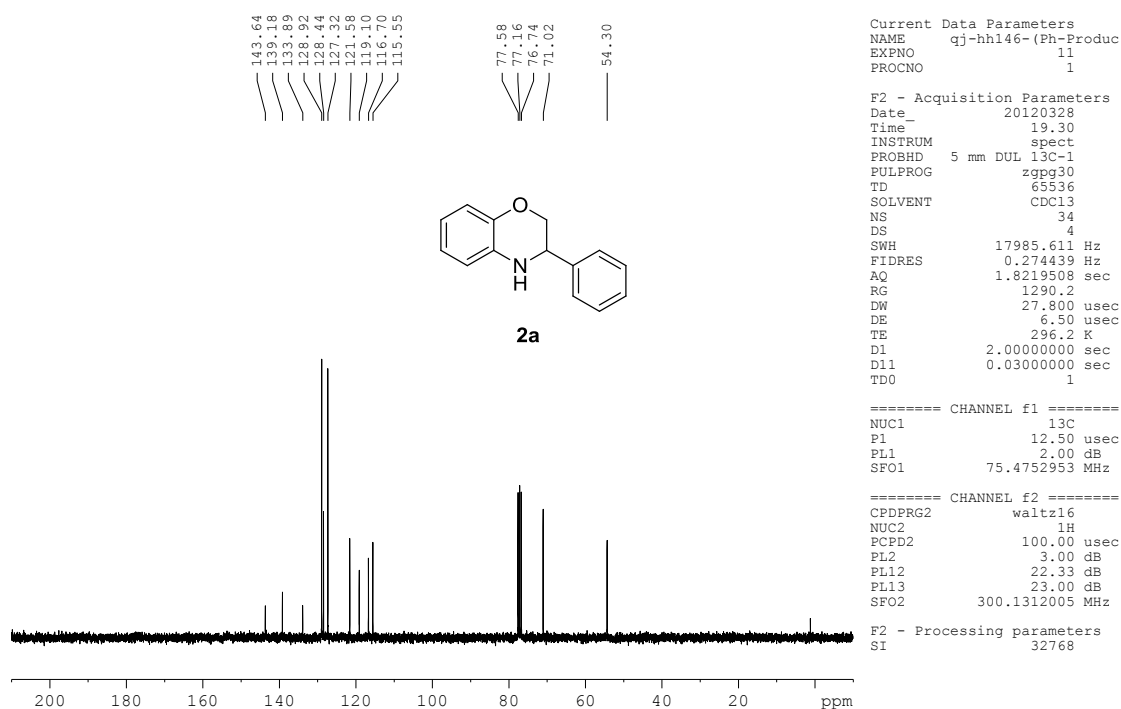
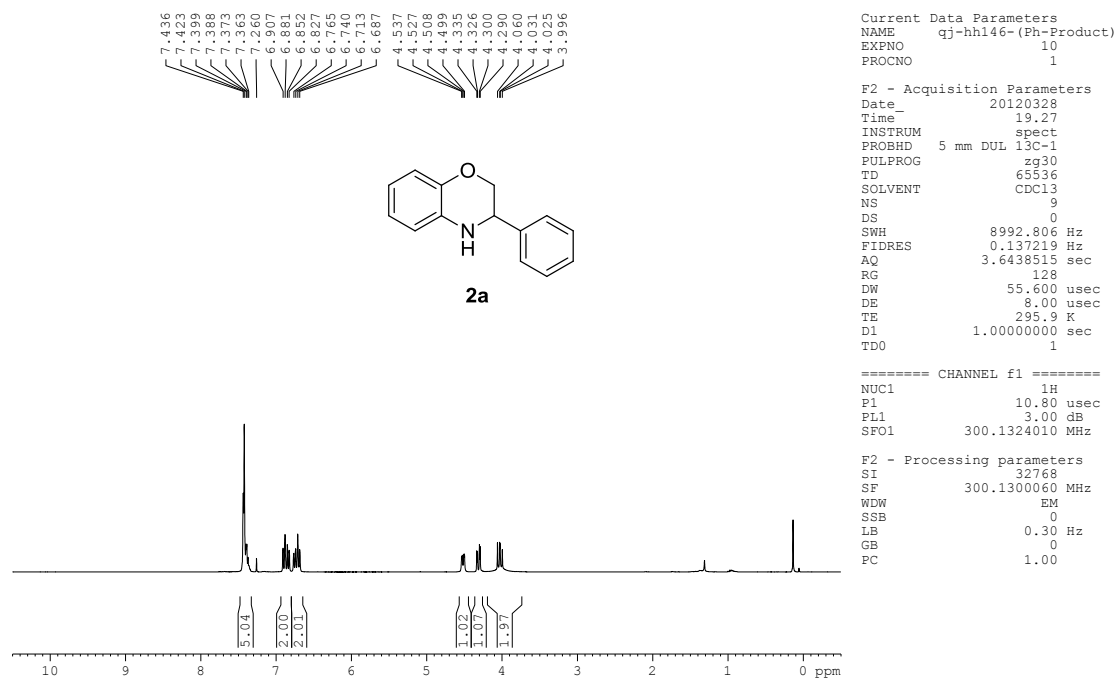
Yellow oil; 97% yield, 98% ee, $[\alpha]_D^{20} = -40.5$ (c 1.0, CHCl_3); ^1H NMR (300 MHz, CDCl_3): δ (ppm) 6.95-6.92 (m, 2H), 6.85-6.78 (m, 3H), 6.71-6.60 (m, 3H), 6.07-5.99 (m, 1H), 4.30-4.26 (m, 1H), 4.12-4.06 (m, 1H),

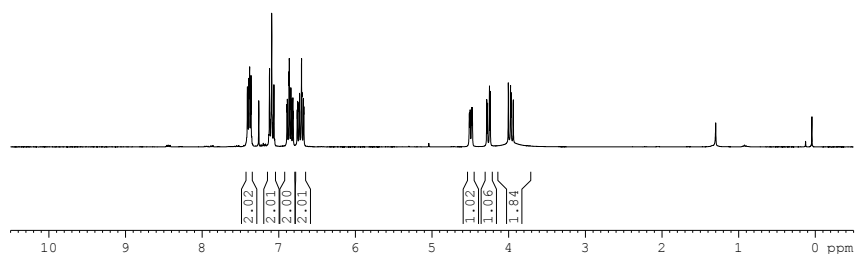
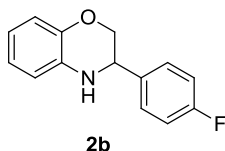
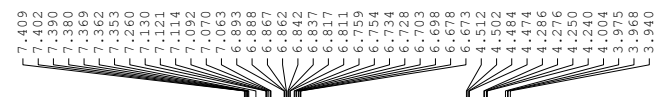
4.01-3.95 (m, 1H), 3.90 (s, 3H), 3.88 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ (ppm) 149.2, 149.1, 143.6, 133.2, 132.7, 129.3, 124.4, 121.6, 119.9, 118.9, 116.6, 115.5, 111.2, 108.8, 69.2, 55.9, 55.9, 52.4; HPLC (OD-H, elute: Hexane / *i*-PrOH = 80 / 20, detector: 254 nm, flow rate: 1 mL / min), t_1 = 28.2 min (major), t_2 = 37.1 min (minor). HRMS-ESI exact mass calcd. for $\text{C}_{18}\text{H}_{20}\text{NO}_3^+$ ($[\text{M}+\text{H}]^+$) requires m/z 298.14425, found m/z 298.14377.

5. Reference

- [1] G. Sabitha, A. V. S. Rao, *Synth. Commun.* **1987**, 17, 341.
- [2] M. Rueping, A. P. Antonchick, T. Theissmann, *Angew. Chem. Int. Ed.* **2006**, 45, 6751.
- [3] K. Gao, C-B. Yu, D-S. Wang, Y-G. Zhou, *Adv. Synth. Catal.* **2012**, 354, 483.

6. Copy of NMR spectra of reduced products



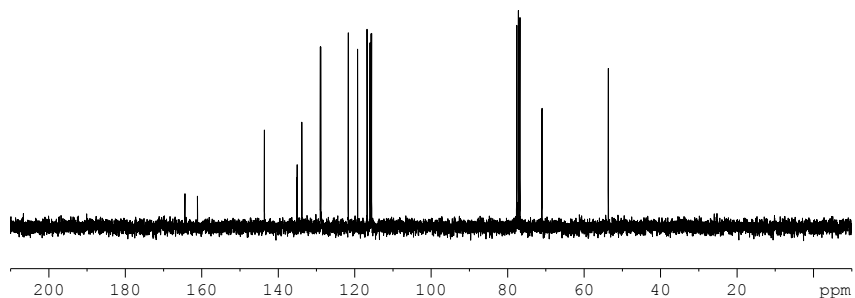
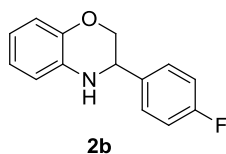
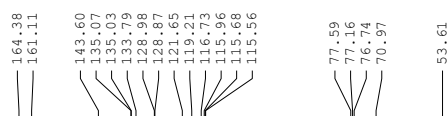


Current Data Parameters
NAME qj-hh149
EXPNO 101
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120411
Time 15.08
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 6
DS 0
SWH 8992.806 Hz
FIDRES 0.137219 Hz
AQ 3.6438515 sec
RG 128
DW 55.600 usec
DE 8.00 usec
TE 297.4 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 10.80 usec
PL1 3.00 dB
SFO1 300.1324010 MHz

F2 - Processing parameters
SI 32768
SF 300.1300057 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



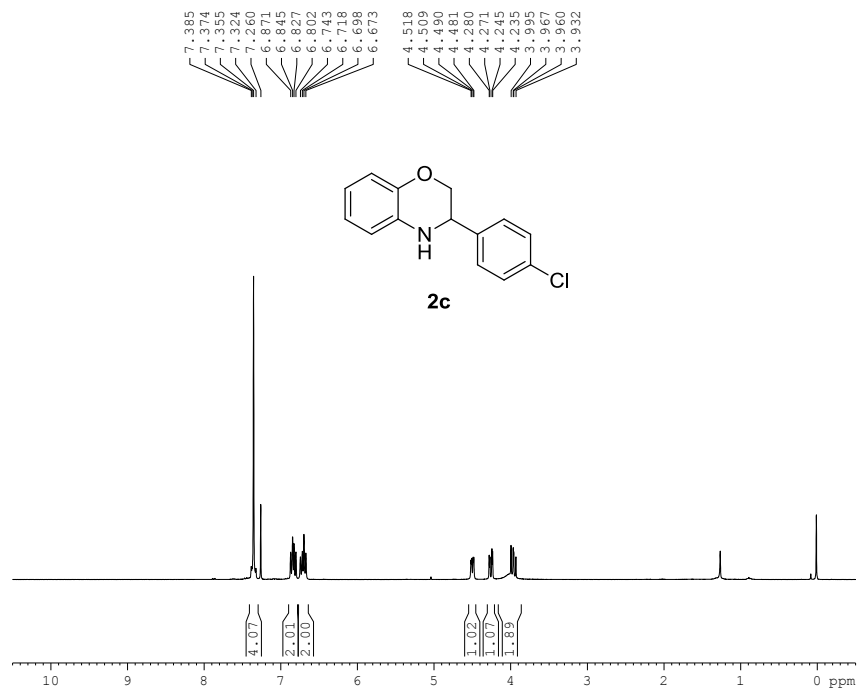
Current Data Parameters
NAME qj-hh149
EXPNO 102
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120411
Time 15.10
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 24
DS 4
SWH 17985.611 Hz
FIDRES 0.274439 Hz
AQ 1.8219508 sec
RG 2298.8
DW 27.800 usec
DE 6.50 usec
TE 297.6 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 12.50 usec
PL1 2.00 dB
SFO1 75.4752953 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 3.00 dB
PL12 22.33 dB
PL13 23.00 dB
SFO2 300.1312005 MHz

F2 - Processing parameters
SI 32768

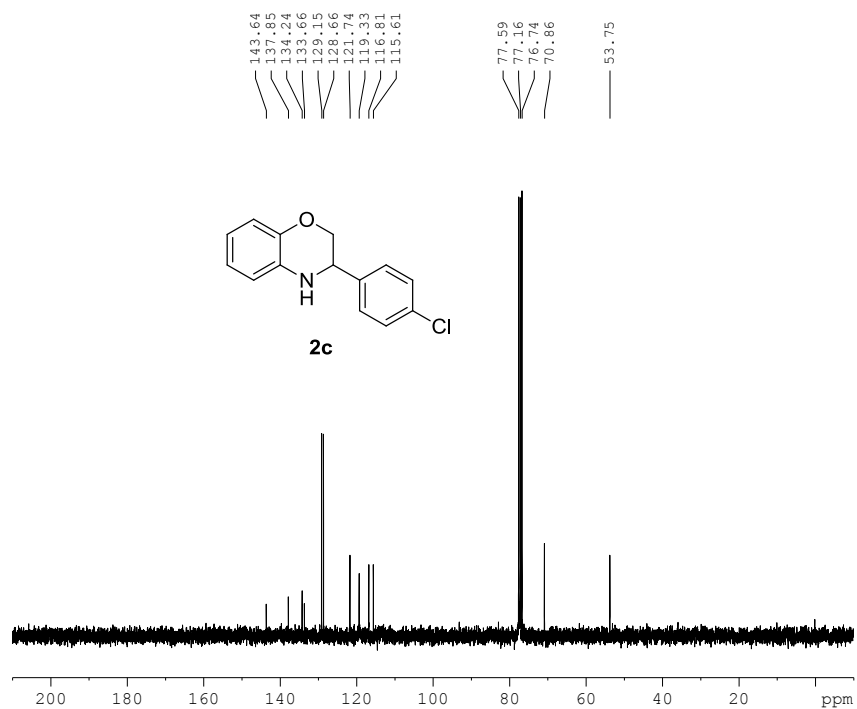


Current Data Parameters
 NAME qj-hh149
 EXPNO 21
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20120411
 Time_ 19.17
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 8992.806 Hz
 FIDRES 0.137219 Hz
 AQ 3.6438515 sec
 RG 256
 DW 55.600 usec
 DE 8.00 usec
 TE 297.4 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 10.80 usec
 PL1 3.00 dB
 SFO1 300.1324010 MHz

F2 - Processing parameters
 SI 32768
 SF 300.1300057 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



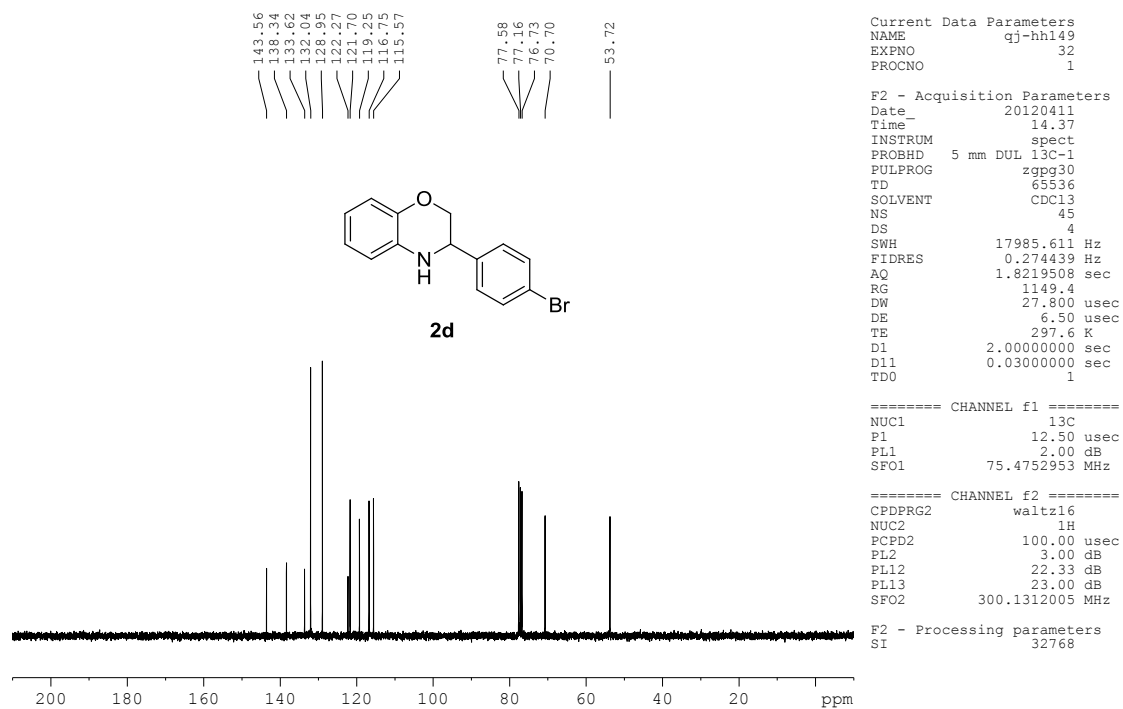
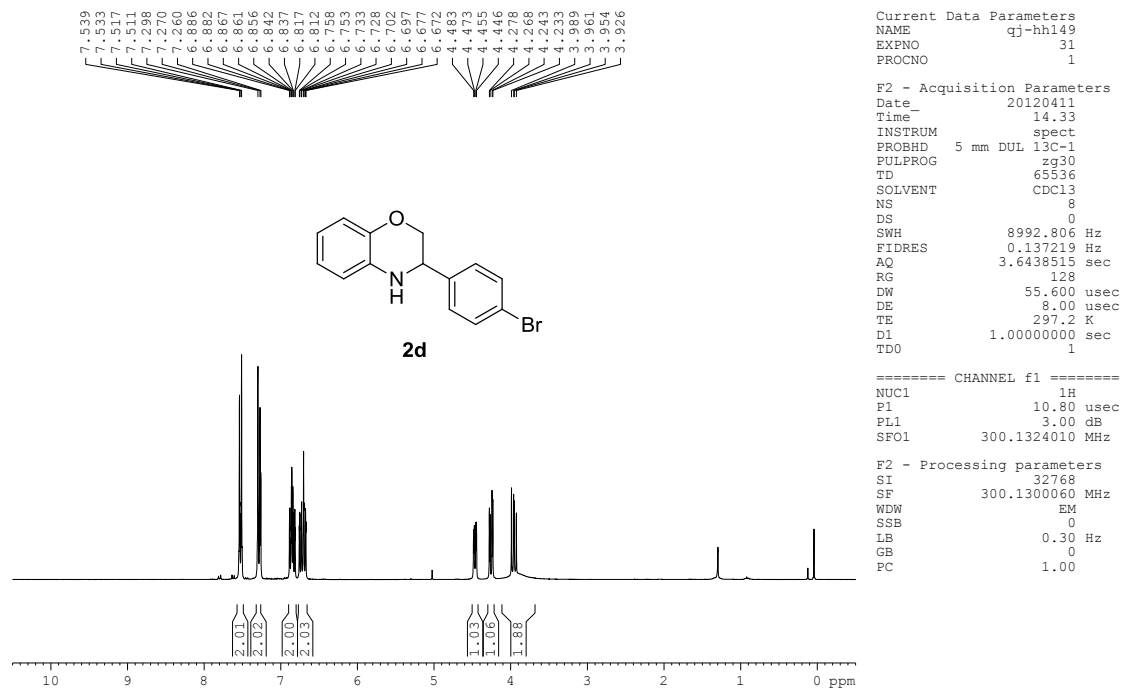
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 NAME qj-hh149
 EXPNO 22
 PROCNO 1

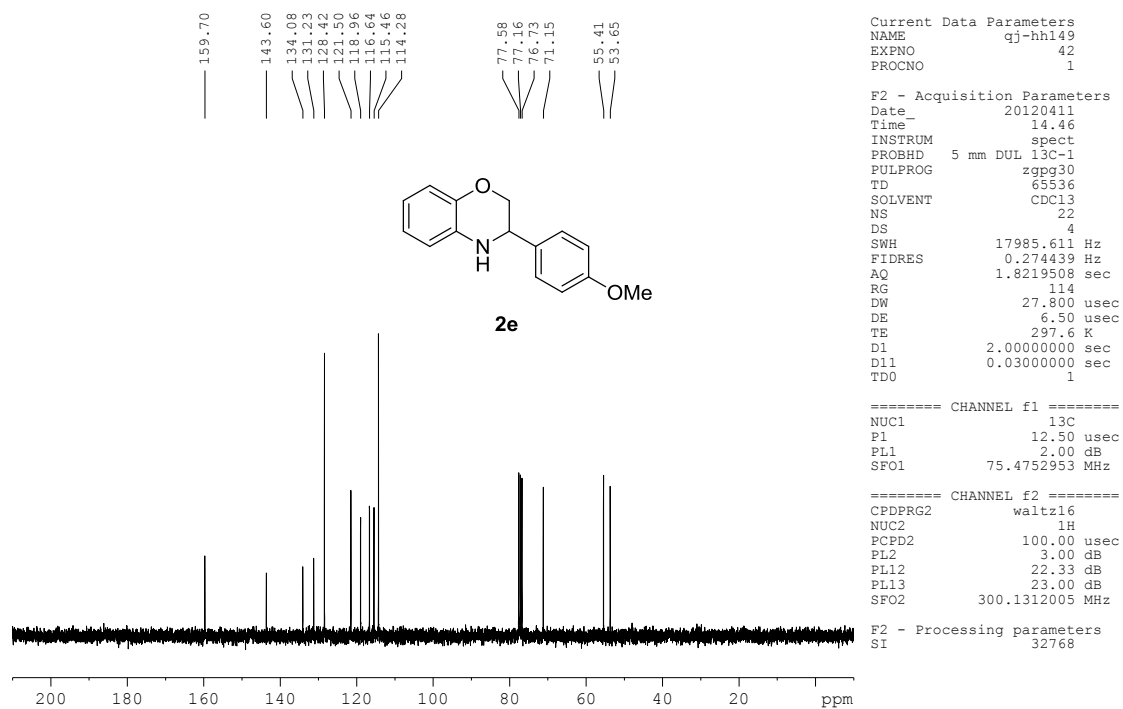
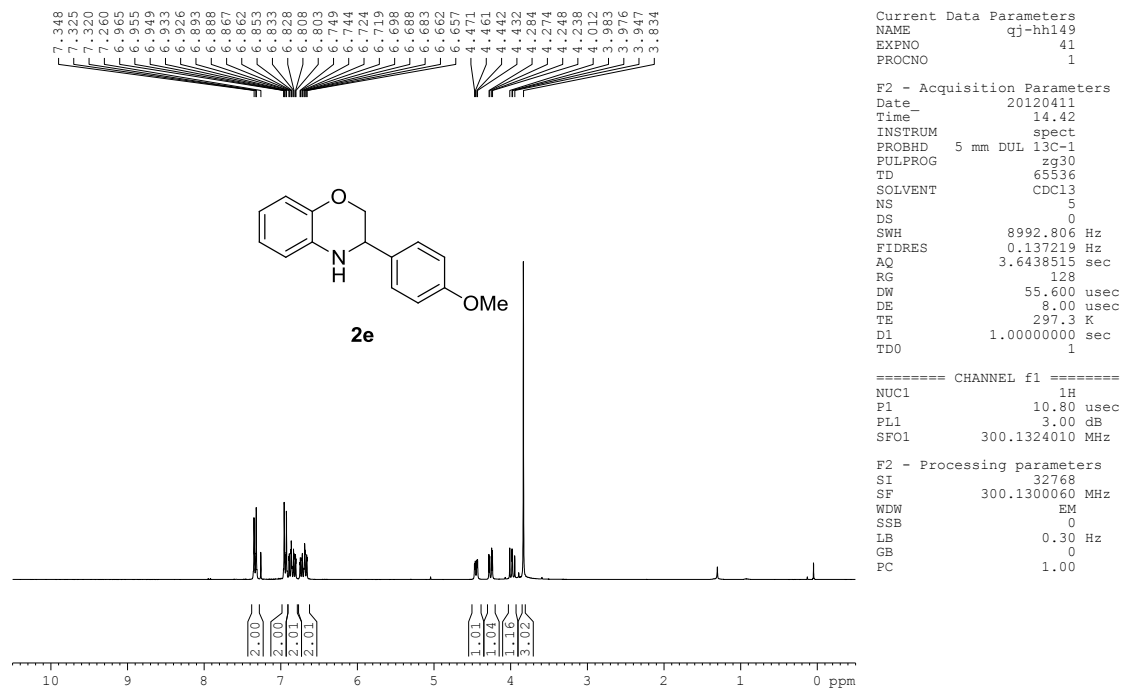
F2 - Acquisition Parameters
 Date_ 20120411
 Time_ 19.20
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 85
 DS 4
 SWH 17985.611 Hz
 FIDRES 0.274439 Hz
 AQ 1.8219508 sec
 RG 1625.5
 DW 27.800 usec
 DE 6.50 usec
 TE 297.6 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

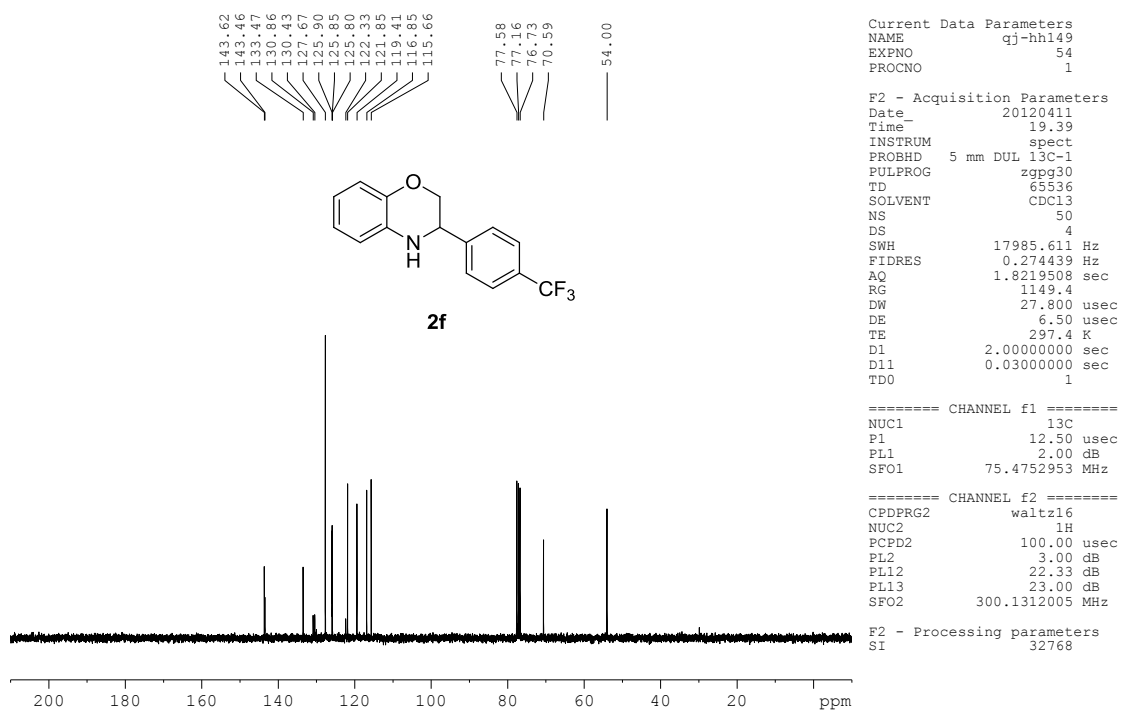
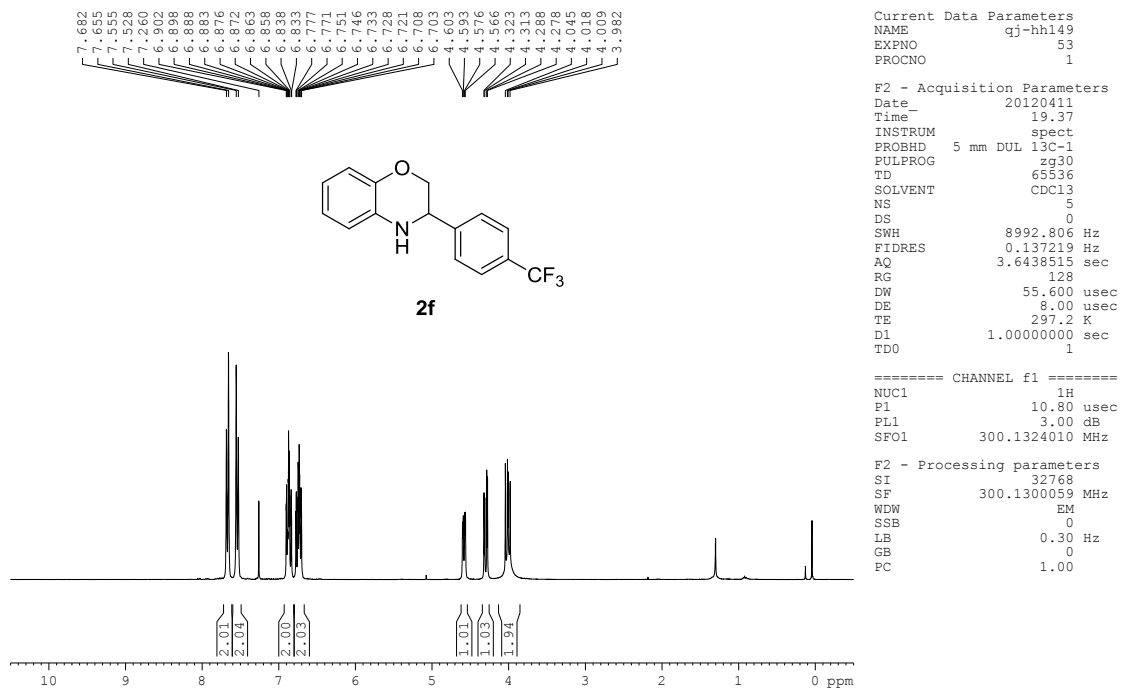
===== CHANNEL f1 =====
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 P1 12.50 usec
 PL1 2.00 dB
 SFO1 75.4752953 MHz

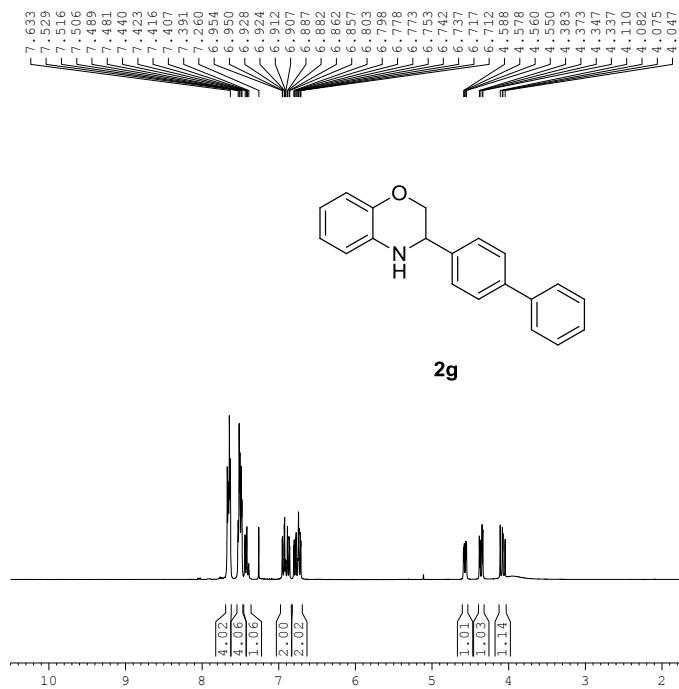
===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 3.00 dB
 PL12 22.33 dB
 PL13 23.00 dB
 SFO2 300.1312005 MHz

F2 - Processing parameters
 SI 32768







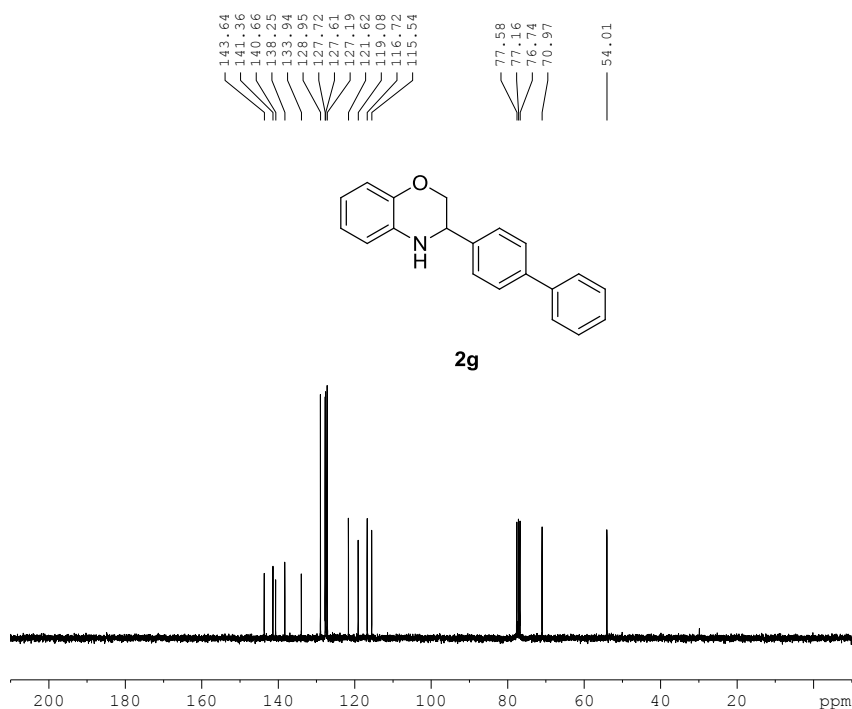


Current Data Parameters
 NAME qj-hh149
 EXPNO 91
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20120411
 Time 14.59
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 6
 DS 0
 SWH 8992.806 Hz
 FIDRES 0.137219 Hz
 AQ 3.6438515 sec
 RG 64
 DW 55.600 usec
 DE 8.00 usec
 TE 297.4 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 10.80 usec
 PL1 3.00 dB
 SFO1 300.1324010 MHz

F2 - Processing parameters
 SI 32768
 SF 300.1300057 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



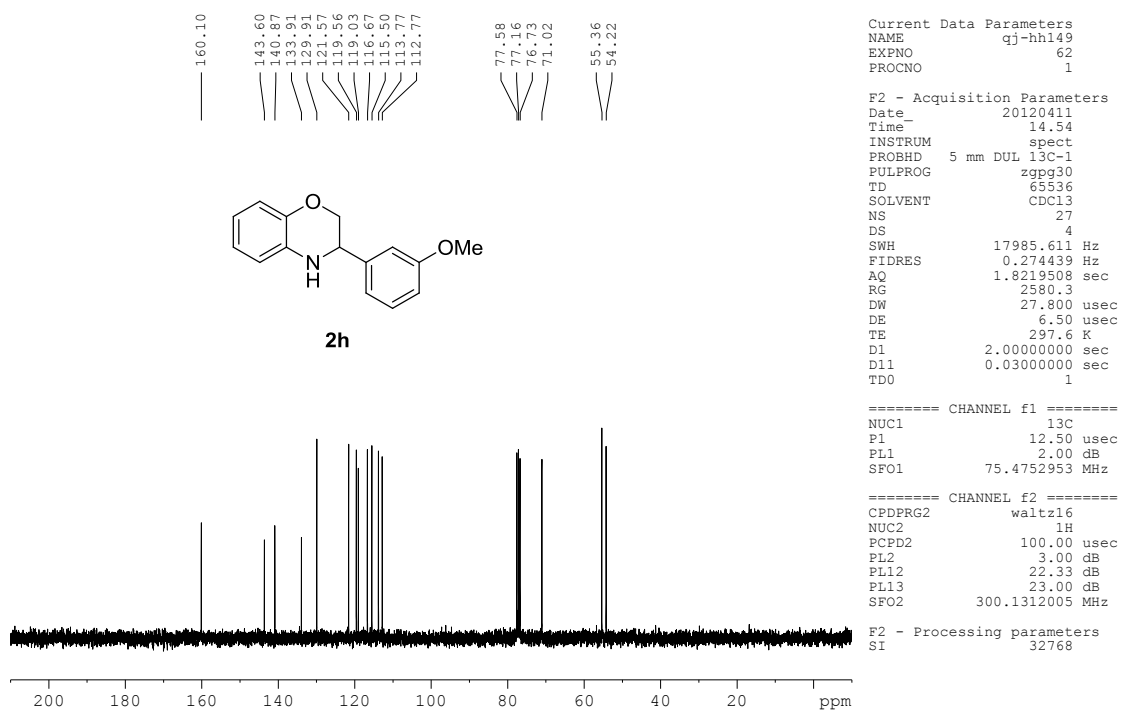
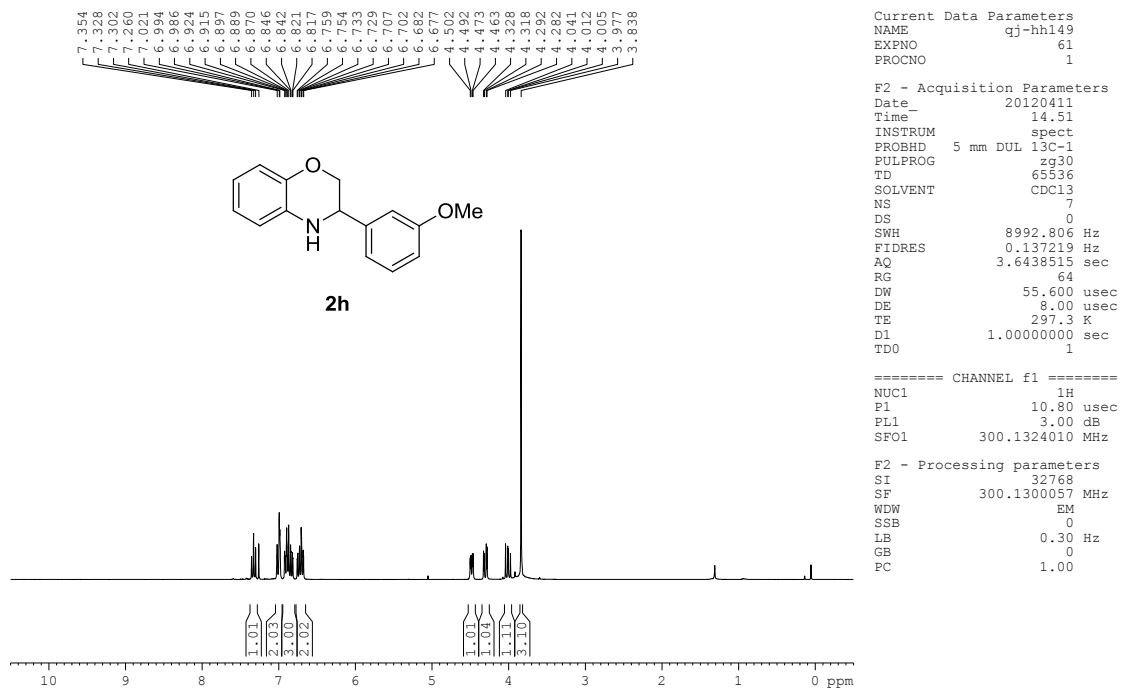
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 NAME qj-hh149
 EXPNO 92
 PROCNO 1

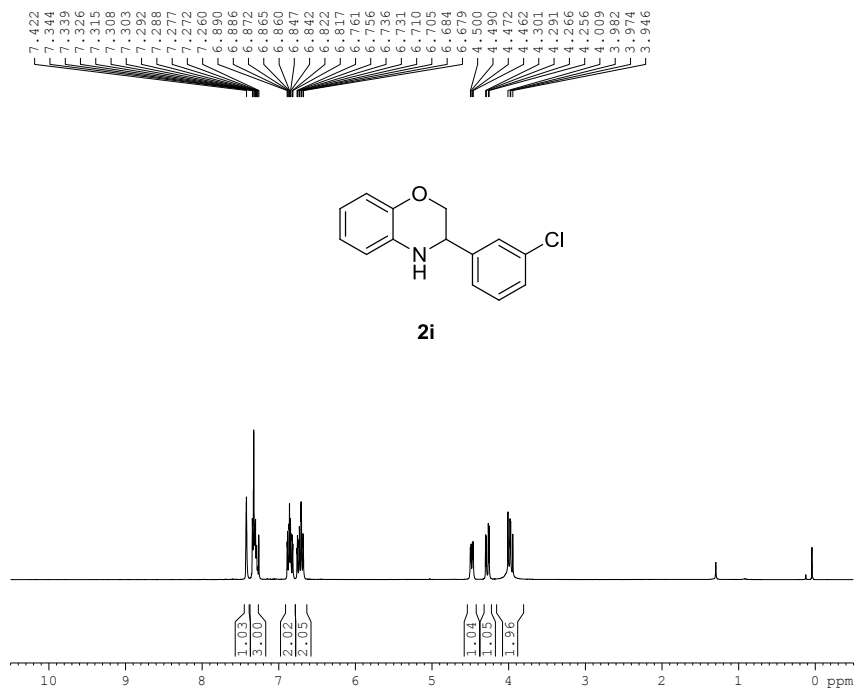
F2 - Acquisition Parameters
 Date_ 20120411
 Time 15.01
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 54
 DS 4
 SWH 17985.611 Hz
 FIDRES 0.274439 Hz
 AQ 1.8219508 sec
 RG 1149.4
 DW 27.800 usec
 DE 6.50 usec
 TE 297.6 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 12.50 usec
 PL1 2.00 dB
 SFO1 75.4752953 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 3.00 dB
 PL12 22.33 dB
 PL13 23.00 dB
 SFO2 300.1312005 MHz

F2 - Processing parameters
 SI 32768



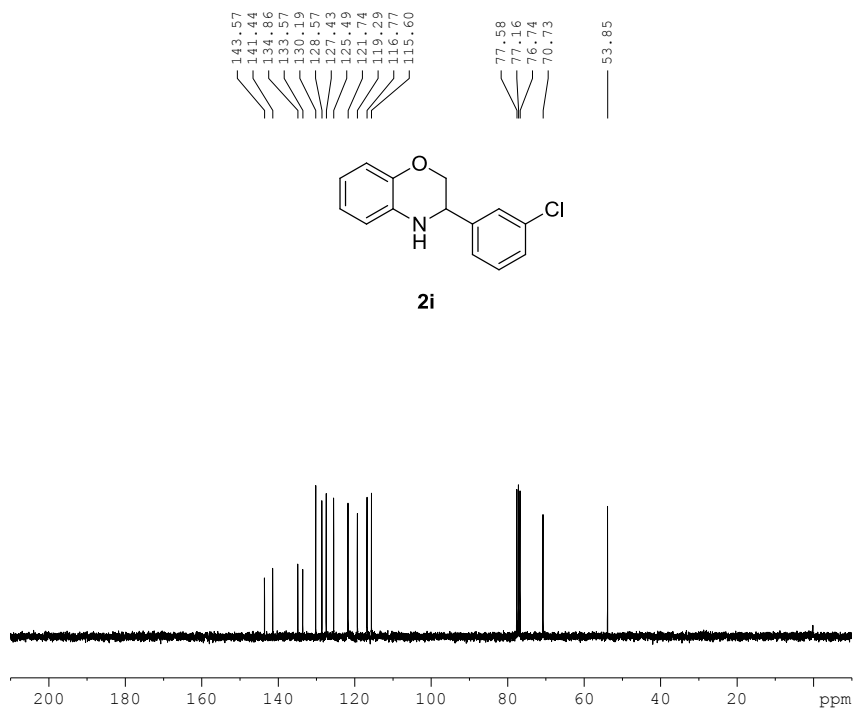


Current Data Parameters
NAME qj-hh151
EXPNO 21
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120428
Time 21.01
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 8
DS 0
SWH 8992.806 Hz
FIDRES 0.137219 Hz
AQ 3.6438515 sec
RG 90.5
DW 55.600 usec
DE 8.00 usec
TE 298.3 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 10.80 usec
PL1 3.00 dB
SFO1 300.1324010 MHz

F2 - Processing parameters
SI 32768
SF 300.1300060 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



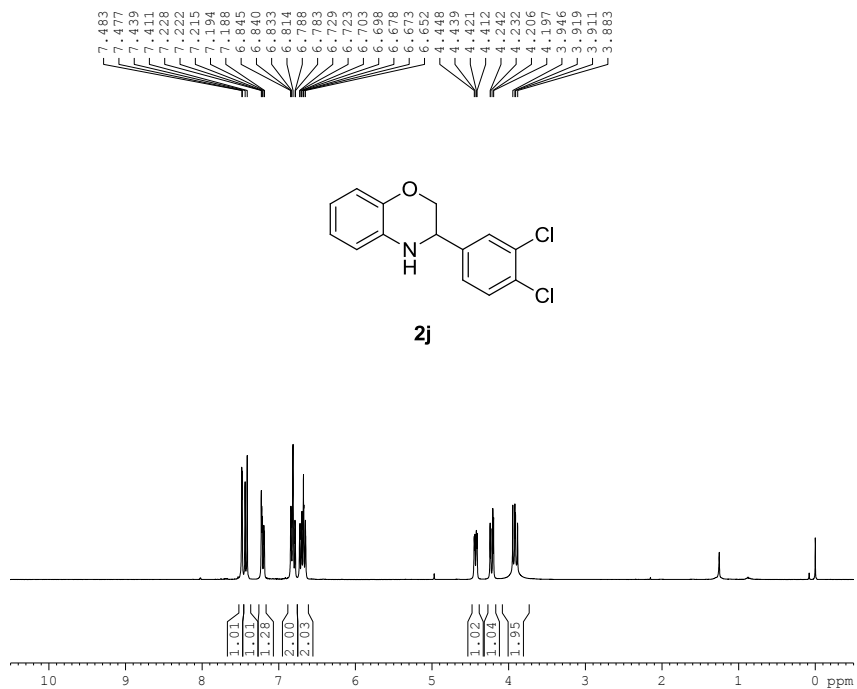
Current Data Parameters
NAME qj-hh151
EXPNO 31
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120428
Time 21.03
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zgig30
TD 32768
SOLVENT CDCl3
NS 74
DS 0
SWH 18832.393 Hz
FIDRES 0.574719 Hz
AQ 0.8700404 sec
RG 11585.2
DW 26.550 usec
DE 8.00 usec
TE 298.5 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 12.50 usec
PL1 2.00 dB
SFO1 75.4752953 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 3.00 dB
PL12 22.33 dB
SFO2 300.1312005 MHz

F2 - Processing parameters
SI 32768
SF 75.4677438 MHz

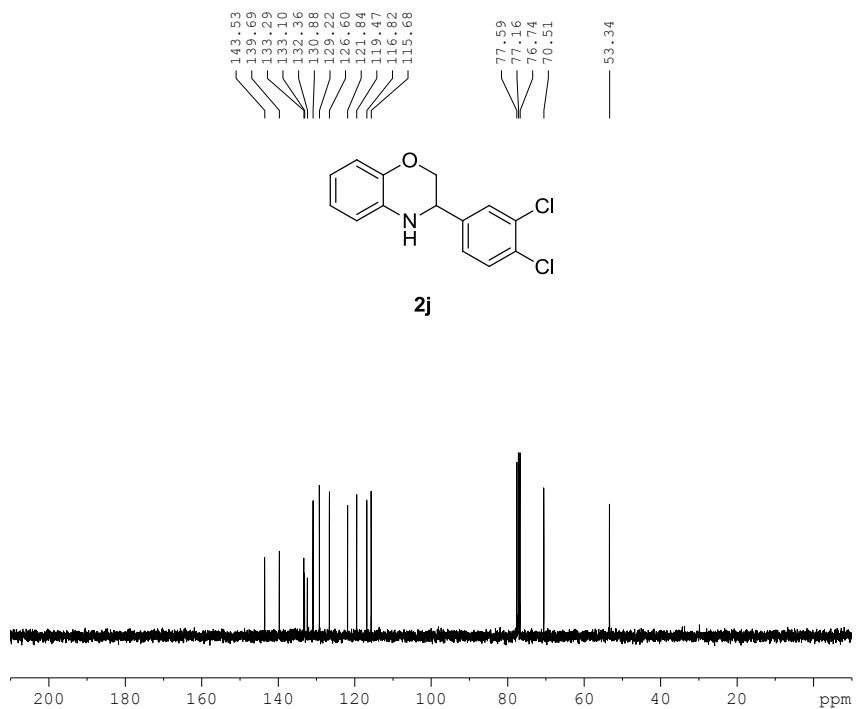


Current Data Parameters
NAME qj-hh149
EXPNO 82
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120411
Time 19.56
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 11
DS 0
SWH 8992.806 Hz
FIDRES 0.137219 Hz
AQ 3.6438515 sec
RG 128
DW 55.600 usec
DE 8.00 usec
TE 297.3 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 10.80 usec
PL1 3.00 dB
SFO1 300.1324010 MHz

F2 - Processing parameters
SI 32768
SF 300.1300155 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



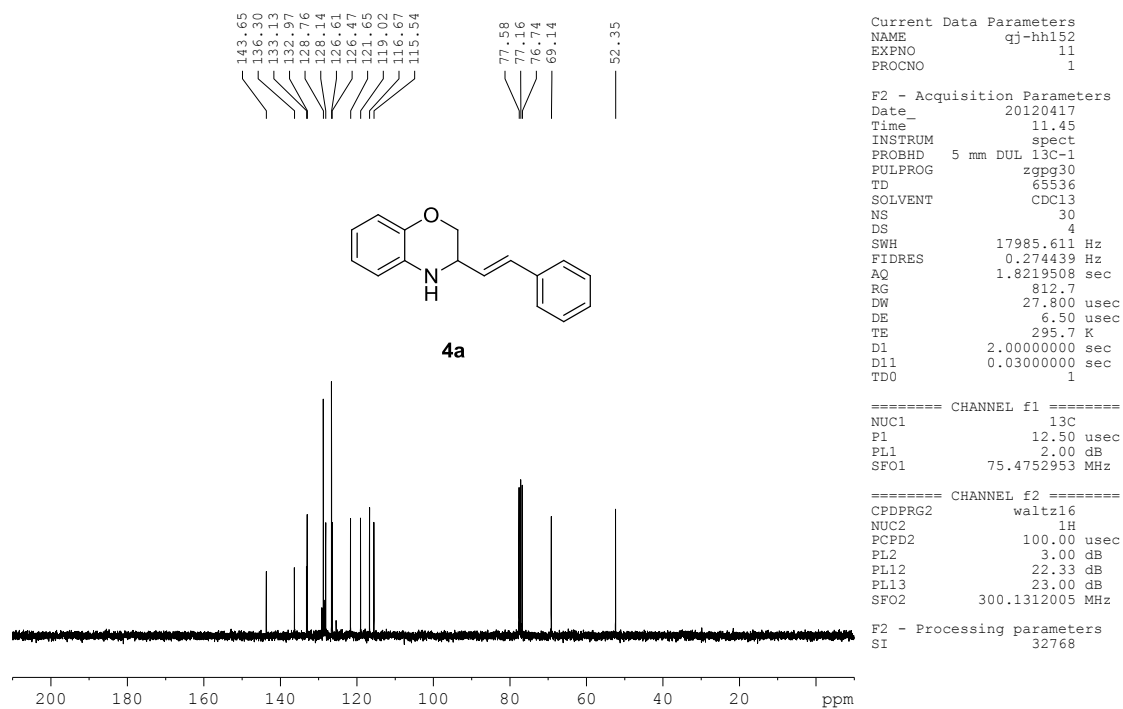
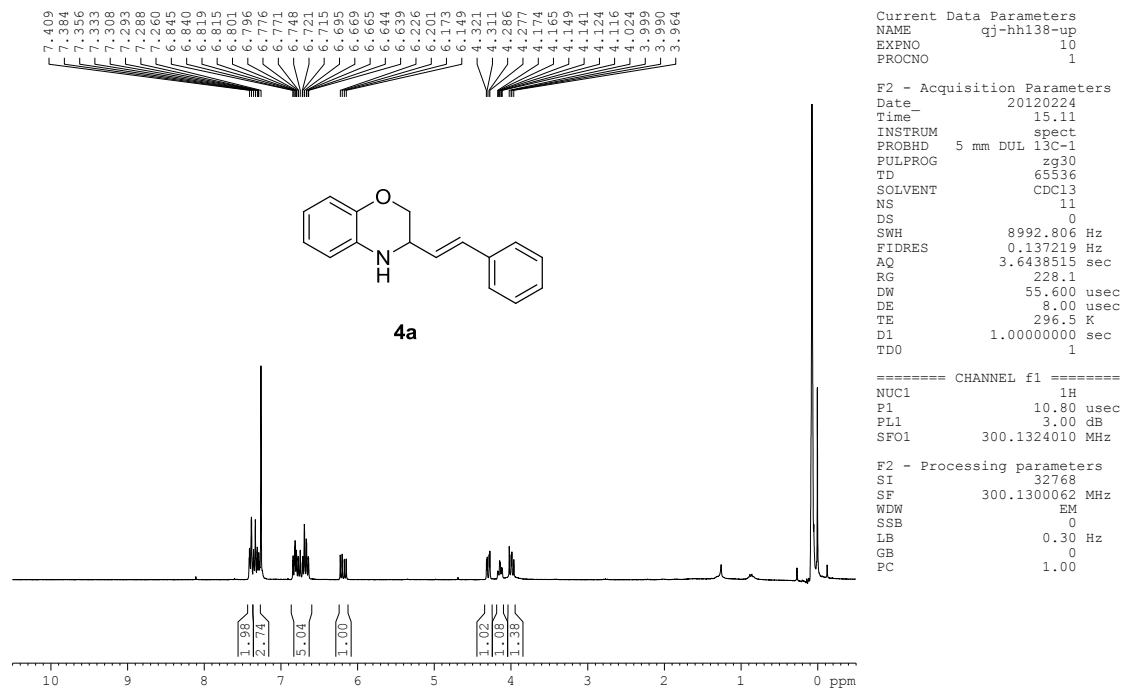
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NAME qj-hh149
EXPNO 83
PROCNO 1

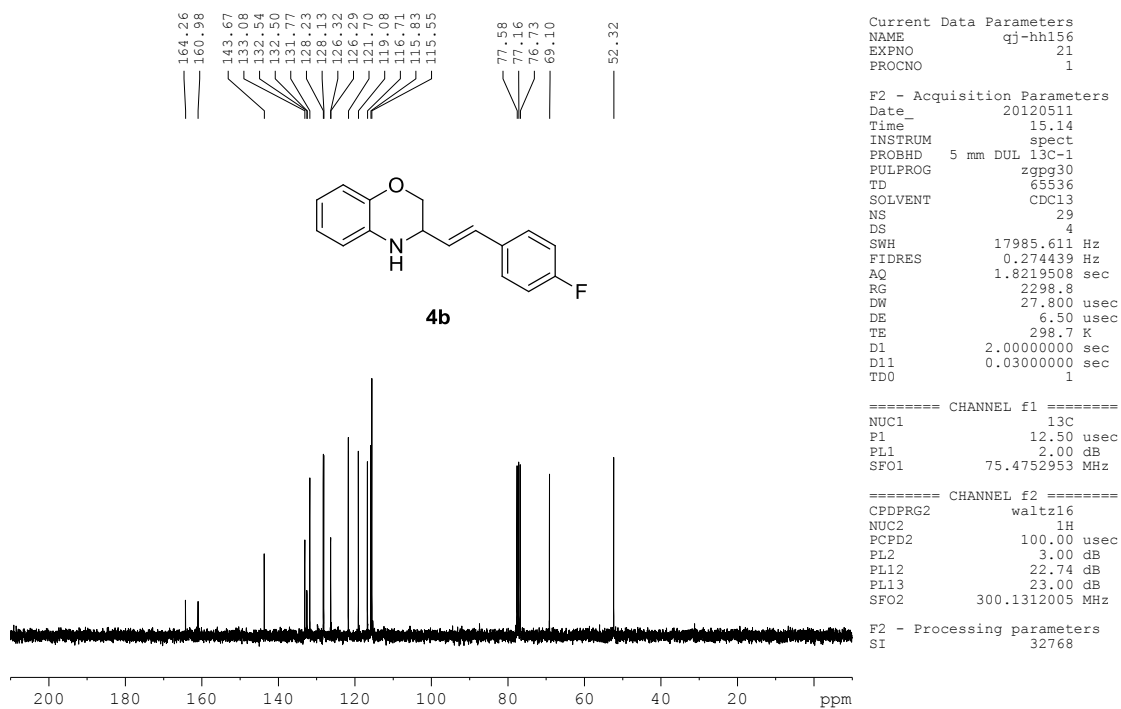
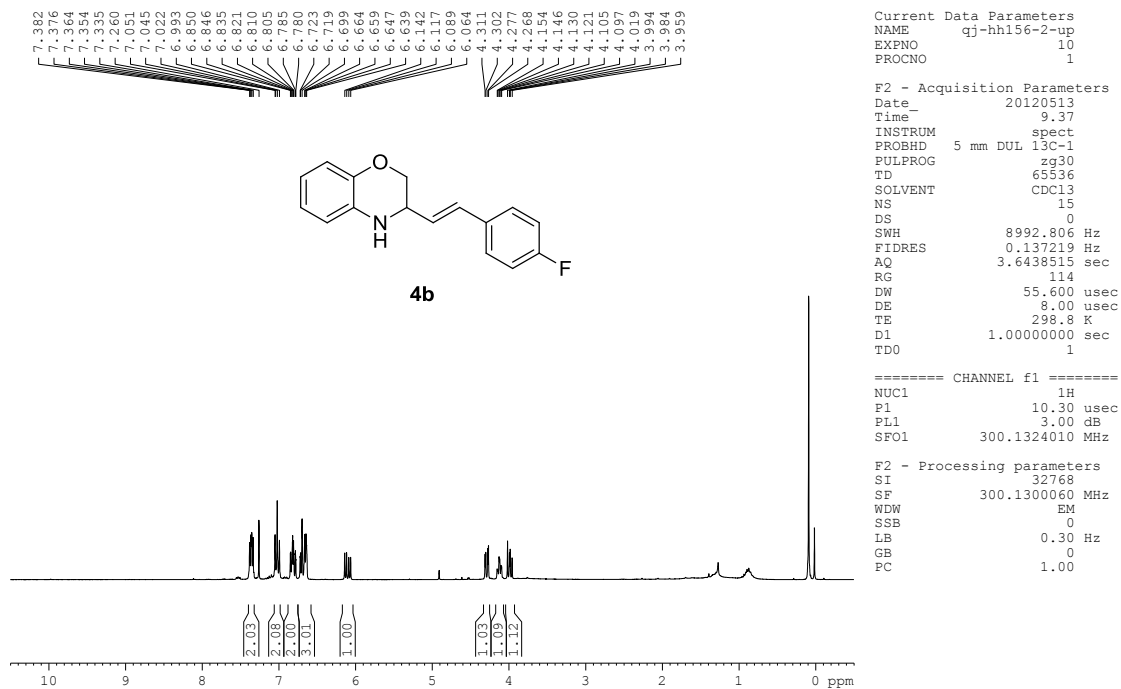
F2 - Acquisition Parameters
Date_ 20120411
Time 19.59
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 32
DS 4
SWH 17985.611 Hz
FIDRES 0.274439 Hz
AQ 1.8219508 sec
RG 1290.2
DW 27.800 usec
DE 6.50 usec
TE 297.5 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

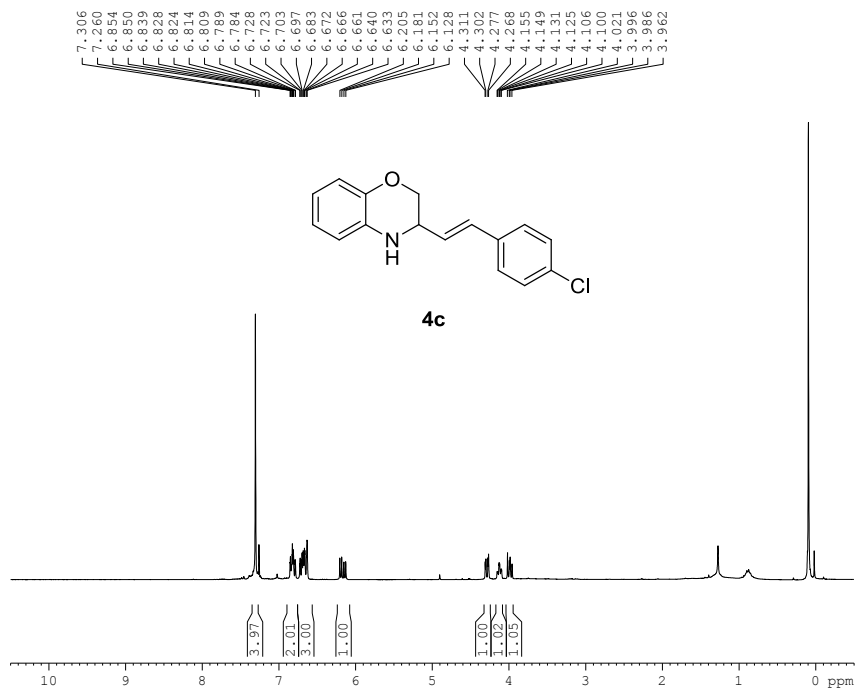
===== CHANNEL f1 =====
NUC1 13C
P1 12.50 usec
PL1 2.00 dB
SFO1 75.4752953 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 3.00 dB
PL12 22.33 dB
PL13 23.00 dB
SFO2 300.1312005 MHz

F2 - Processing parameters
SI 32768





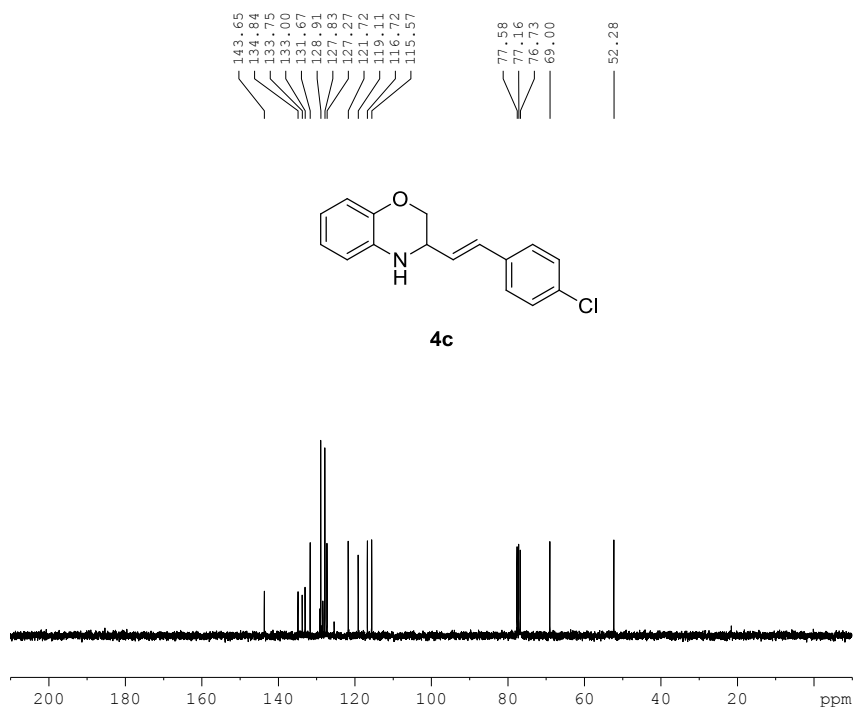


Current Data Parameters
NAME qj-hhl56-3-up
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120513
Time 9.44
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 12
DS 0
SWH 8992.806 Hz
FIDRES 0.137219 Hz
AQ 3.6438515 sec
RG 71.8
DW 55.600 usec
DE 8.00 usec
TE 298.8 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 10.30 usec
PL1 3.00 dB
SFO1 300.1324010 MHz

F2 - Processing parameters
SI 32768
SF 300.1300062 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



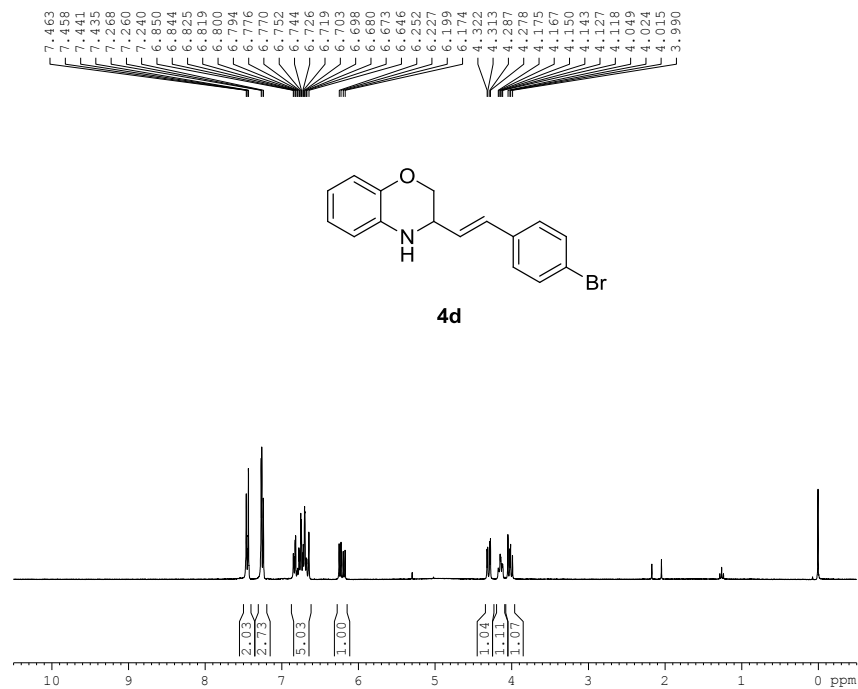
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NAME qj-hhl56
EXPNO 31
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120511
Time 15.22
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 18
DS 4
SWH 17985.611 Hz
FIDRES 0.274439 Hz
AQ 1.8219508 sec
RG 2580.3
DW 27.800 usec
DE 6.50 usec
TE 298.5 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 12.50 usec
PL1 2.00 dB
SFO1 75.4752953 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 3.00 dB
PL12 22.74 dB
PL13 23.00 dB
SFO2 300.1312005 MHz

F2 - Processing parameters
SI 32768

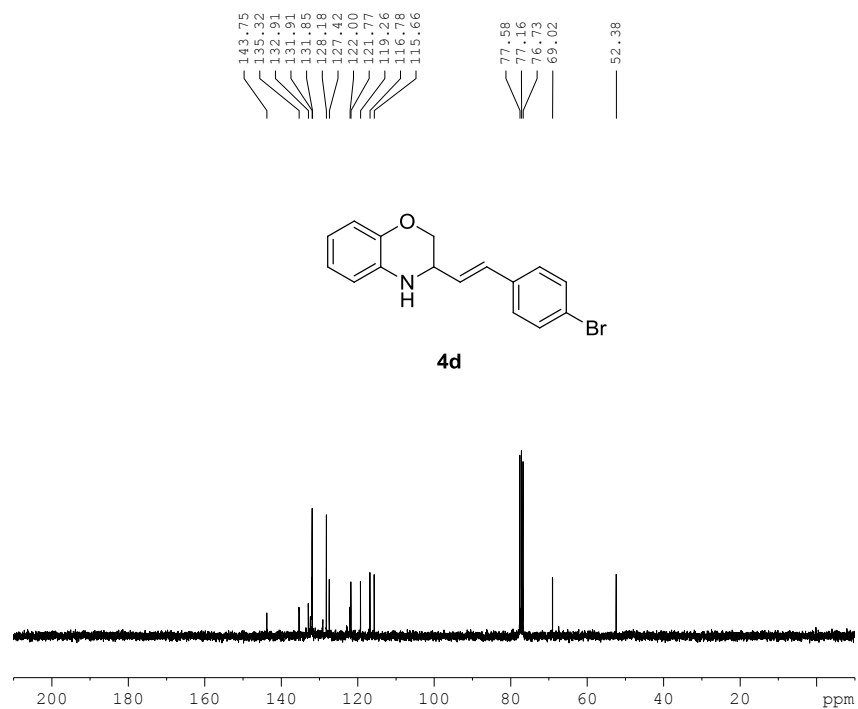


Current Data Parameters
NAME qj-hh-br
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120618
Time 11.16
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 7
DS 0
SWH 8992.806 Hz
FIDRES 0.137219 Hz
AQ 3.6438515 sec
RG 512
DW 55.600 usec
DE 8.00 usec
TE 300.3 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 10.30 usec
PL1 3.00 dB
SFO1 300.1324010 MHz

F2 - Processing parameters
SI 32768
SF 300.1300060 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



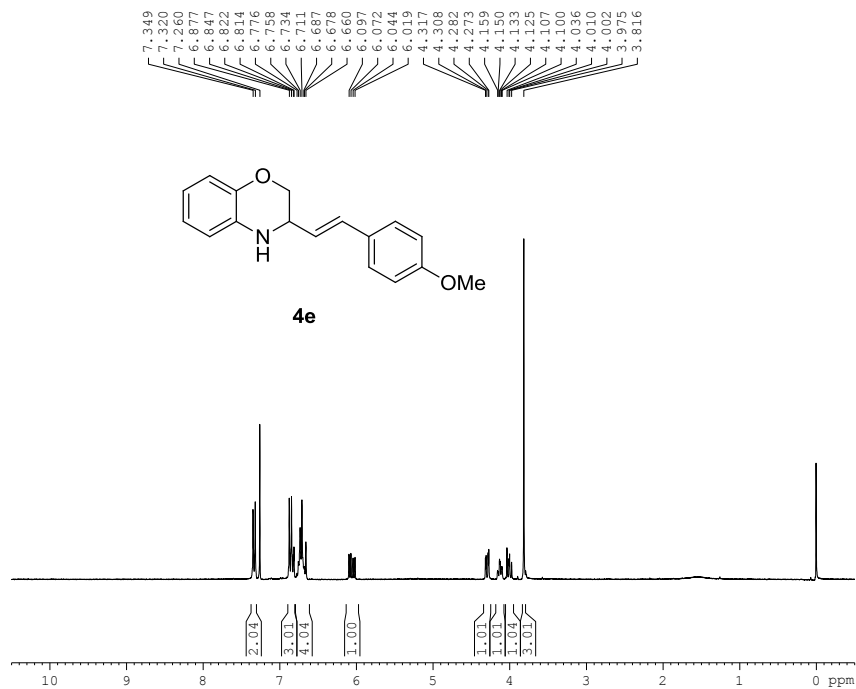
Current Data Parameters
NAME qj-hh-br
EXPNO 21
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120619
Time 11.53
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 152
DS 4
SWH 17985.611 Hz
FIDRES 0.274439 Hz
AQ 1.8219508 sec
RG 512
DW 27.800 usec
DE 6.50 usec
TE 300.3 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 12.50 usec
PL1 2.00 dB
SFO1 75.4752953 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 3.00 dB
PL12 22.74 dB
PL13 23.00 dB
SFO2 300.1312005 MHz

F2 - Processing parameters
SI 32768

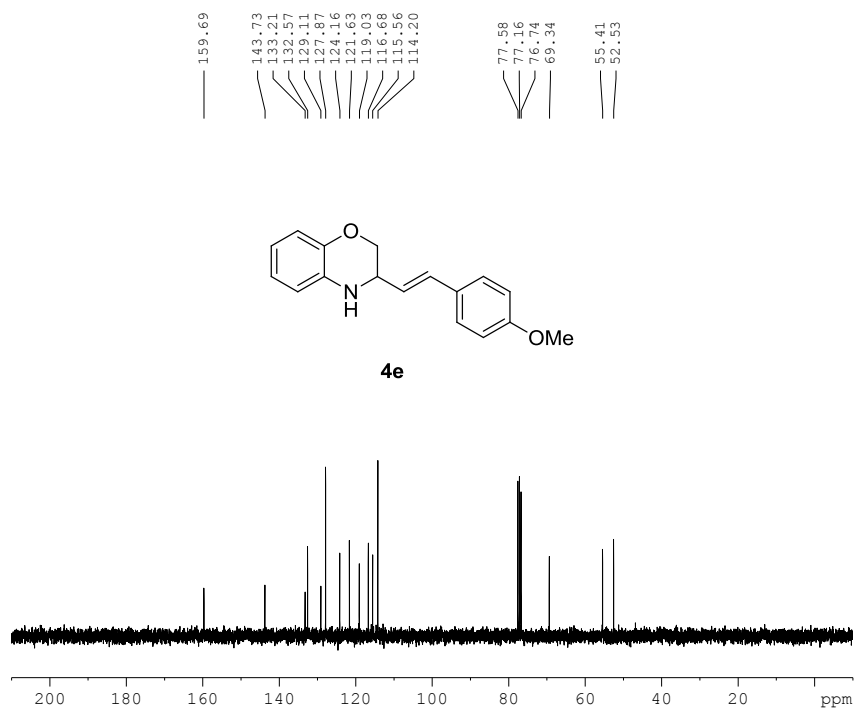


Current Data Parameters
NAME gj-hh-meo
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120618
Time 11.21
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 7
DS 0
SWH 8992.806 Hz
FIDRES 0.137219 Hz
AQ 3.6438515 sec
RG 574.7
DW 55.600 usec
DE 8.00 usec
TE 300.3 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 10.30 usec
PL1 3.00 dB
SFO1 300.1324010 MHz

F2 - Processing parameters
SI 32768
SF 300.1300060 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



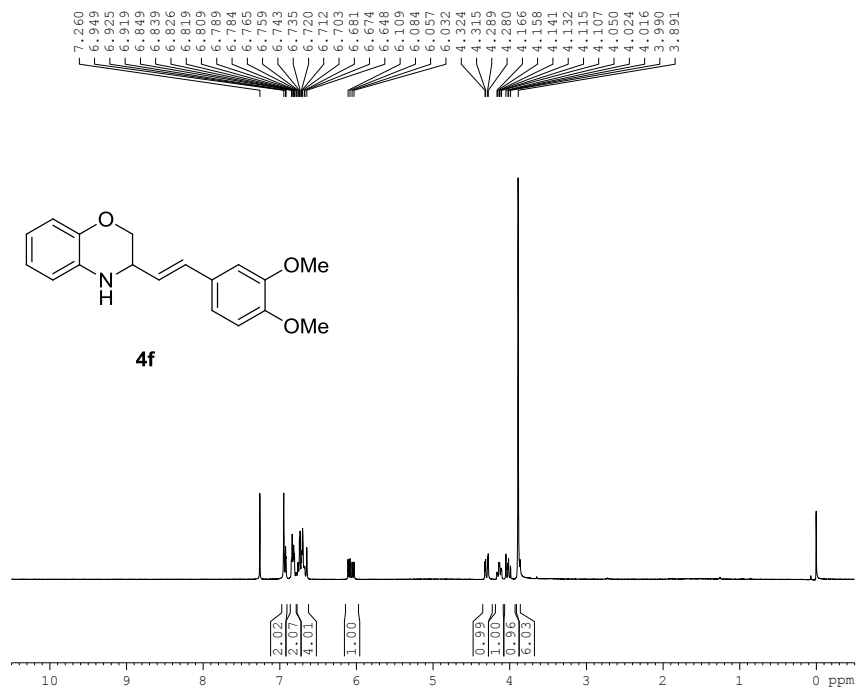
Current Data Parameters
NAME gj-hh-meo
EXPNO 20
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120619
Time 12.07
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 19
DS 4
SWH 17985.611 Hz
FIDRES 0.274439 Hz
AQ 1.8219508 sec
RG 574.7
DW 27.800 usec
DE 6.50 usec
TE 300.5 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 12.50 usec
PL1 2.00 dB
SFO1 75.4752953 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 3.00 dB
PL12 22.74 dB
PL13 23.00 dB
SFO2 300.1312005 MHz

F2 - Processing parameters
SI 32768

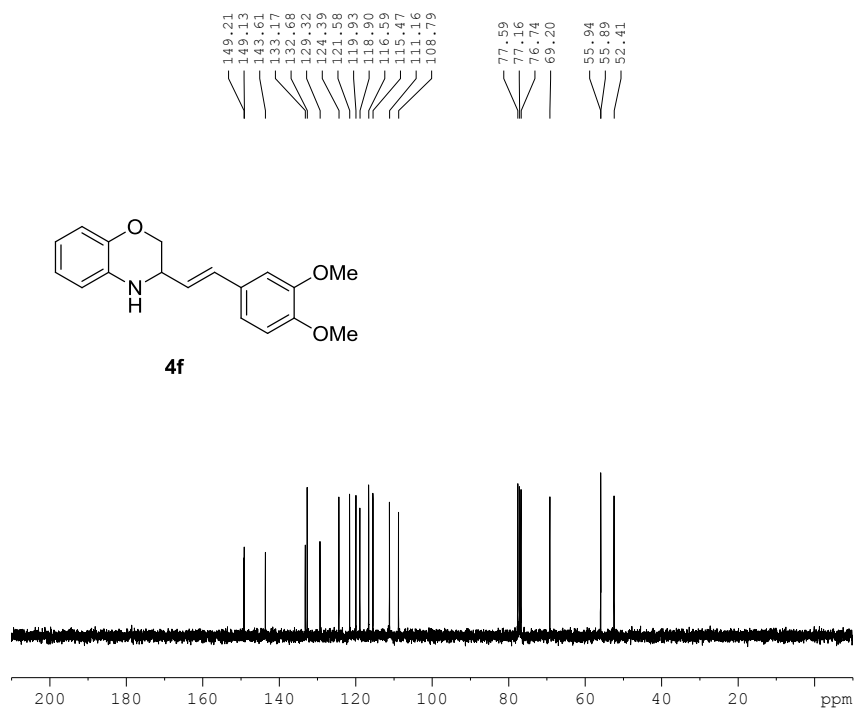


Current Data Parameters
 NAME qj-hh-dimeo
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20120618
 Time 11.25
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 9
 DS 0
 SWH 8992.806 Hz
 FIDRES 0.137219 Hz
 AQ 3.6438515 sec
 RG 456.1
 DW 55.600 usec
 DE 8.00 usec
 TE 300.3 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 10.30 usec
 PL1 3.00 dB
 SFO1 300.1324010 MHz

F2 - Processing parameters
 SI 32768
 SF 300.1300062 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



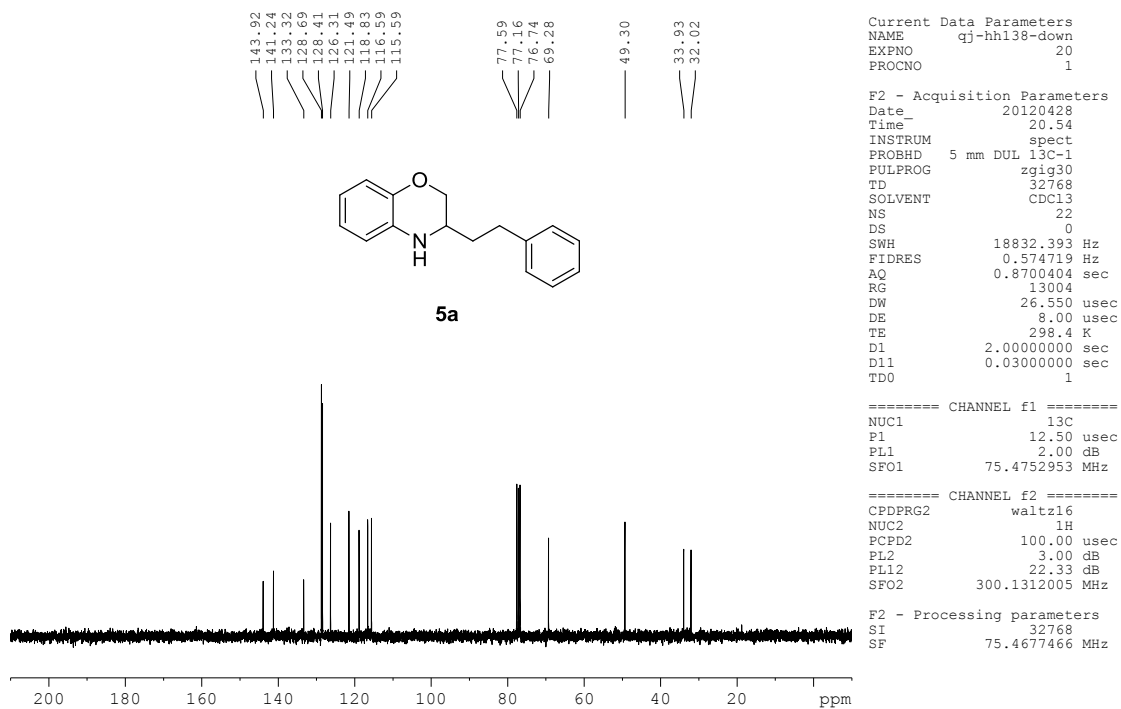
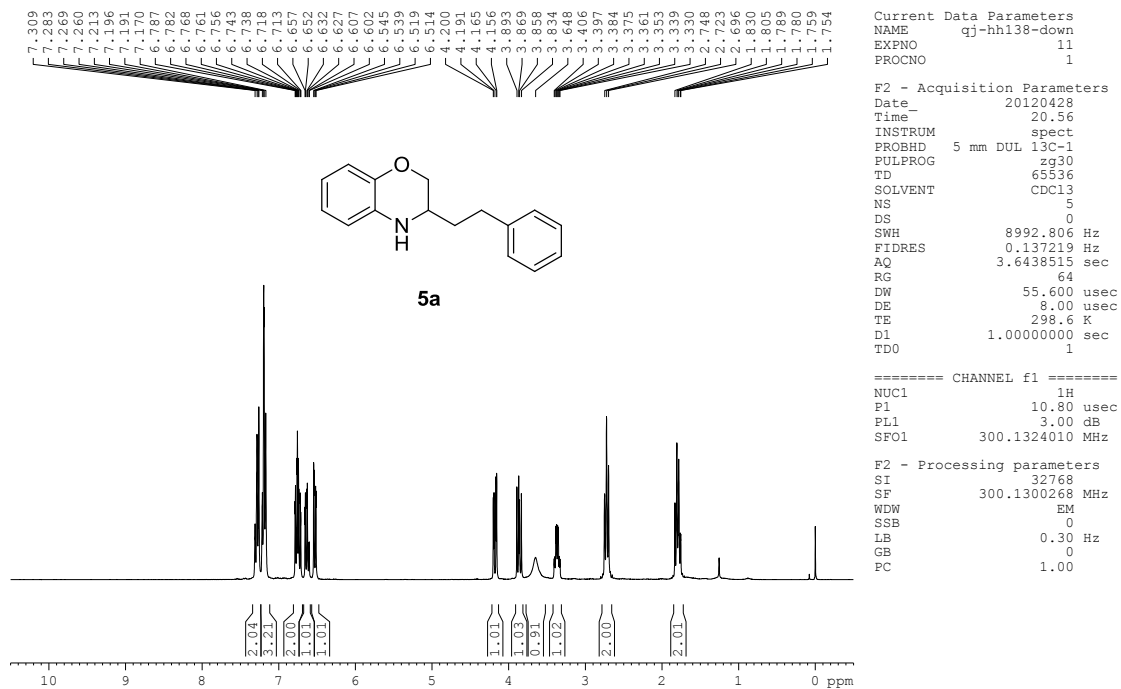
Current Data Parameters
 NAME qj-hh156
 EXPNO 41
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20120511
 Time 15.06
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 22
 DS 4
 SWH 17985.611 Hz
 FIDRES 0.274439 Hz
 AQ 1.8219508 sec
 RG 2580.3
 DW 27.800 usec
 DE 6.50 usec
 TE 298.6 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

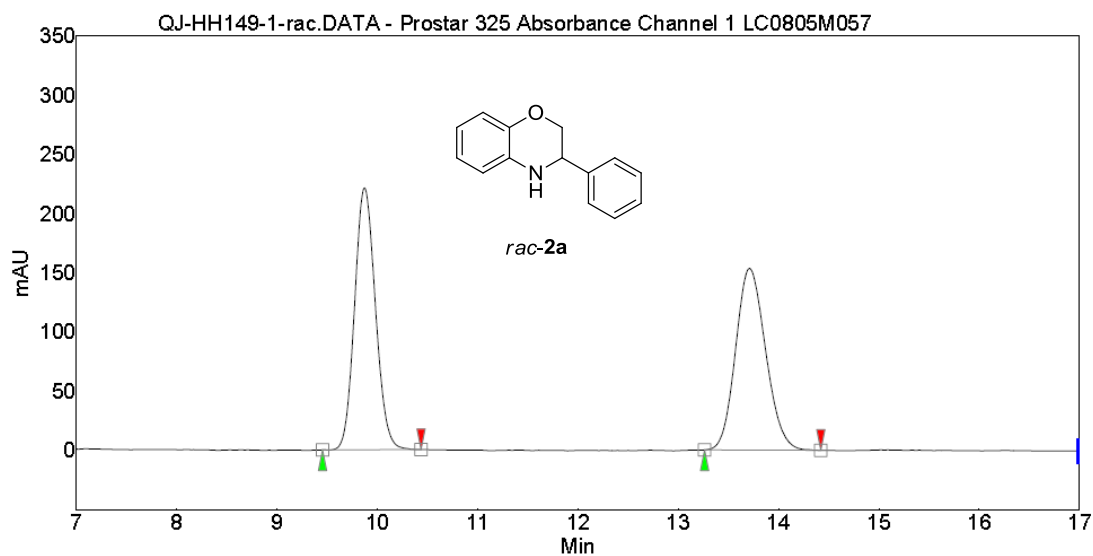
===== CHANNEL f1 =====
 NUC1 13C
 P1 12.50 usec
 PL1 2.00 dB
 SFO1 75.4752953 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 3.00 dB
 PL12 22.74 dB
 PL13 23.00 dB
 SFO2 300.1312005 MHz

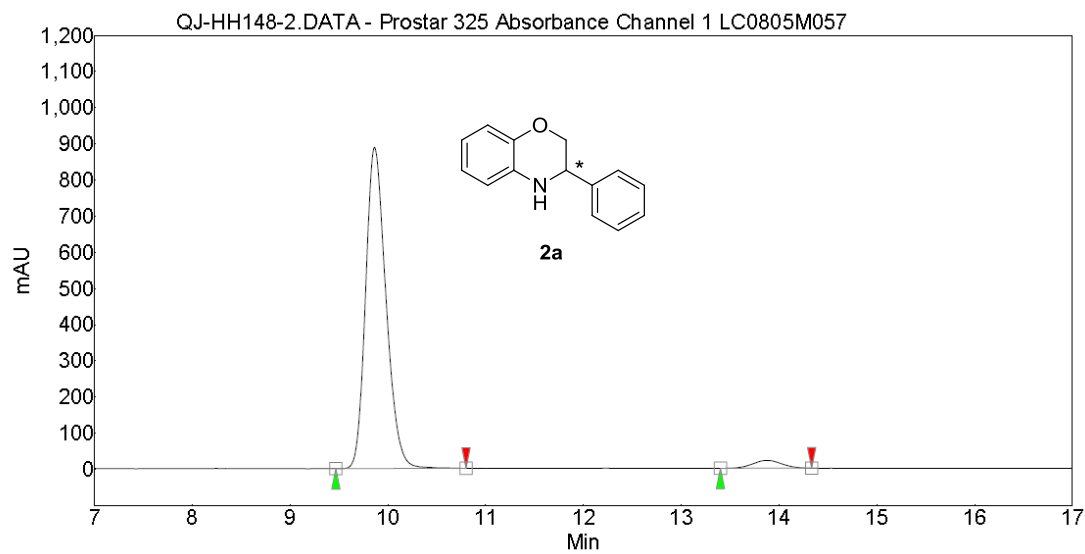
F2 - Processing parameters
 SI 32768



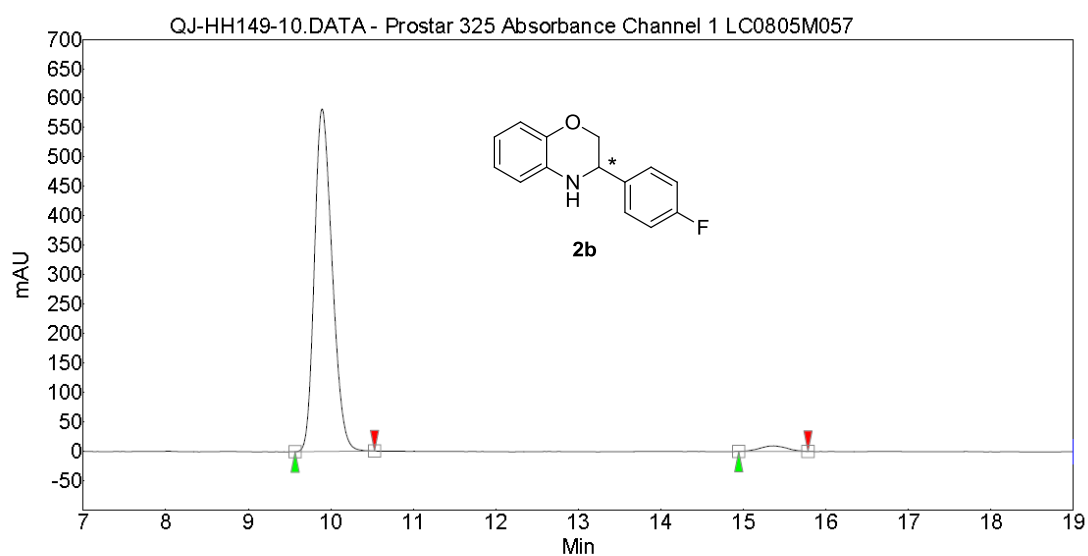
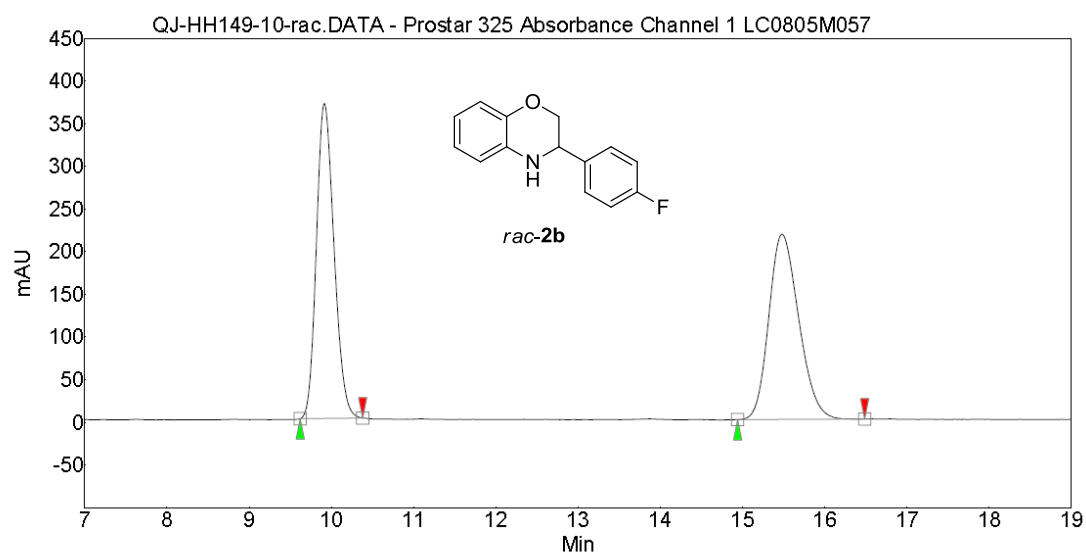
7. HPLC spectra of the reduced products

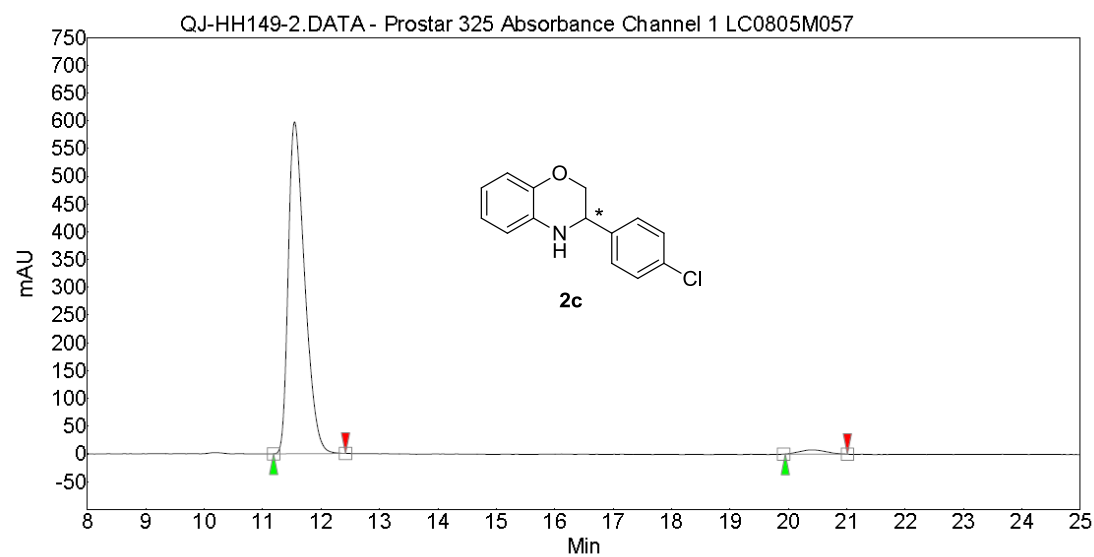
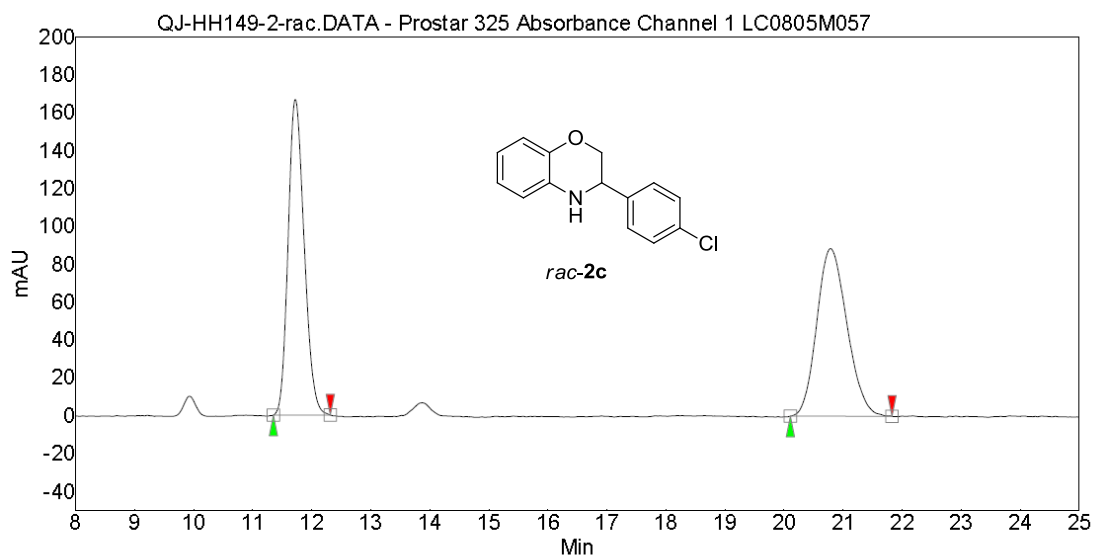


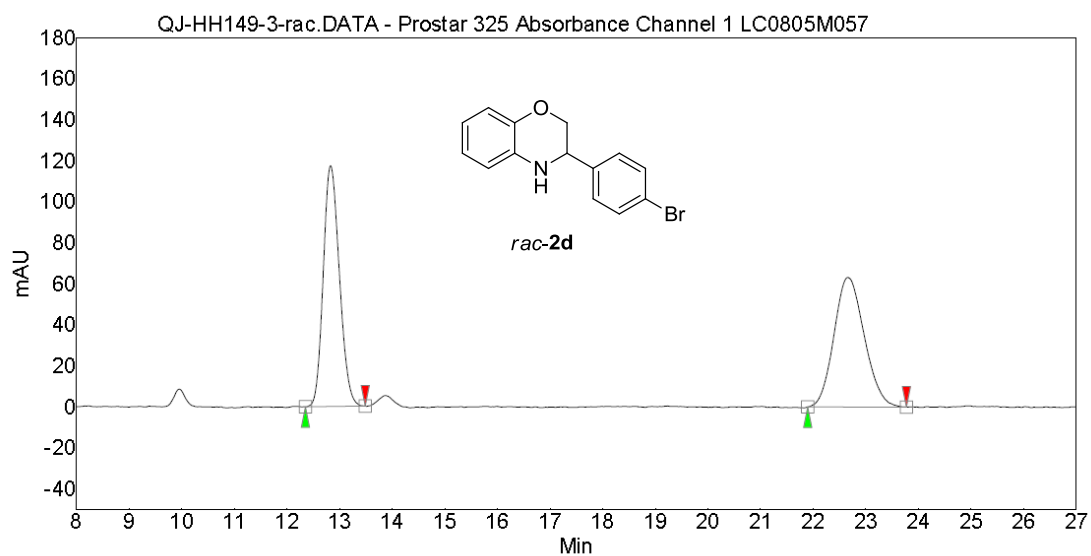
Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	9.87	50.03	221.3	53.3	50.031
2	未知	13.71	49.97	153.5	53.3	49.969
Total			100.00	374.8	106.6	100.000



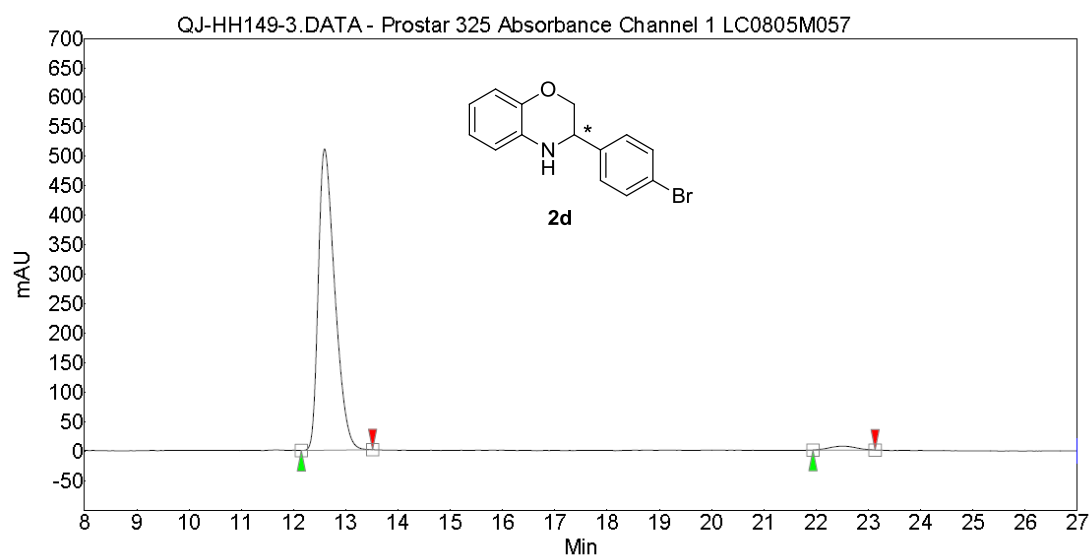
Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
2	未知	9.86	96.60	888.4	214.4	96.603
1	未知	13.87	3.40	21.7	7.5	3.397
Total			100.00	910.1	222.0	100.000



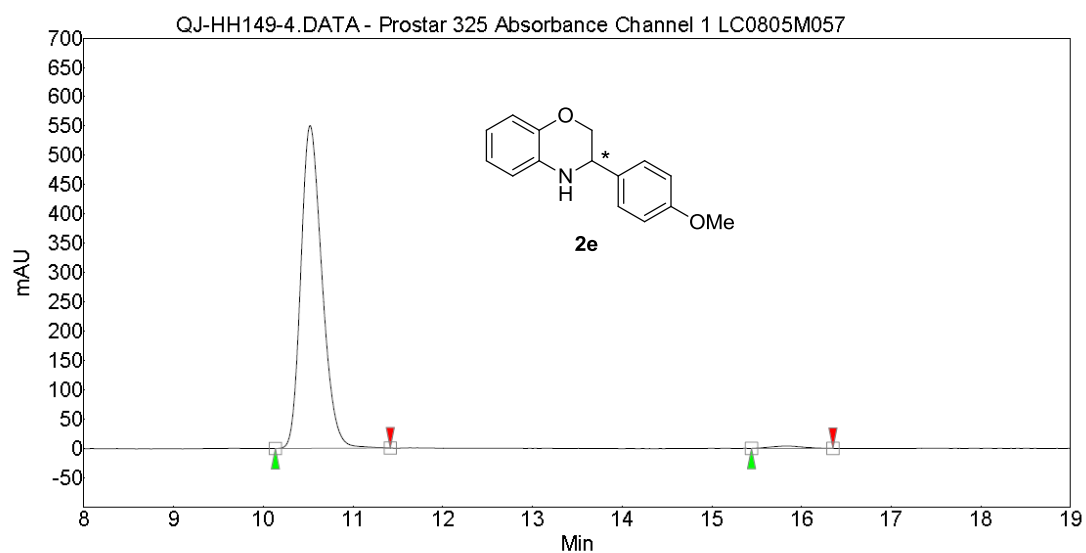
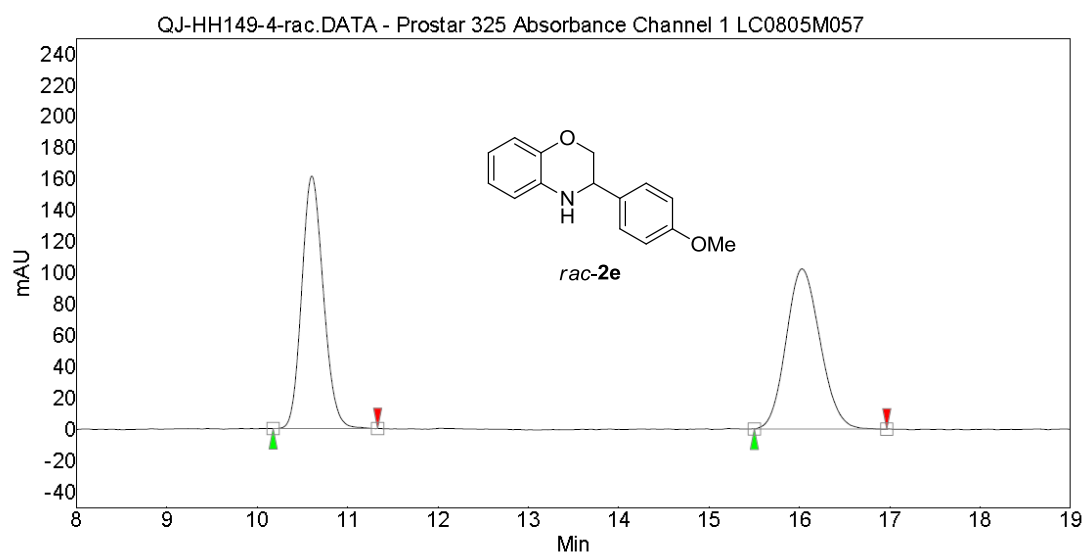


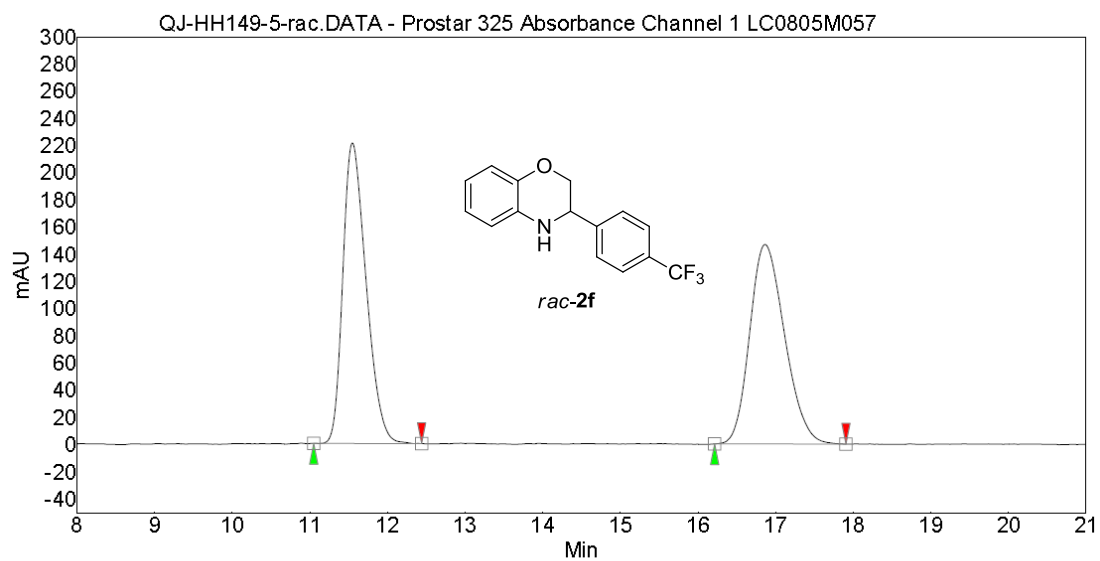


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	12.83	49.75	117.4	41.7	49.749
2	未知	22.65	50.25	63.2	42.2	50.251
Total			100.00	180.6	83.9	100.000

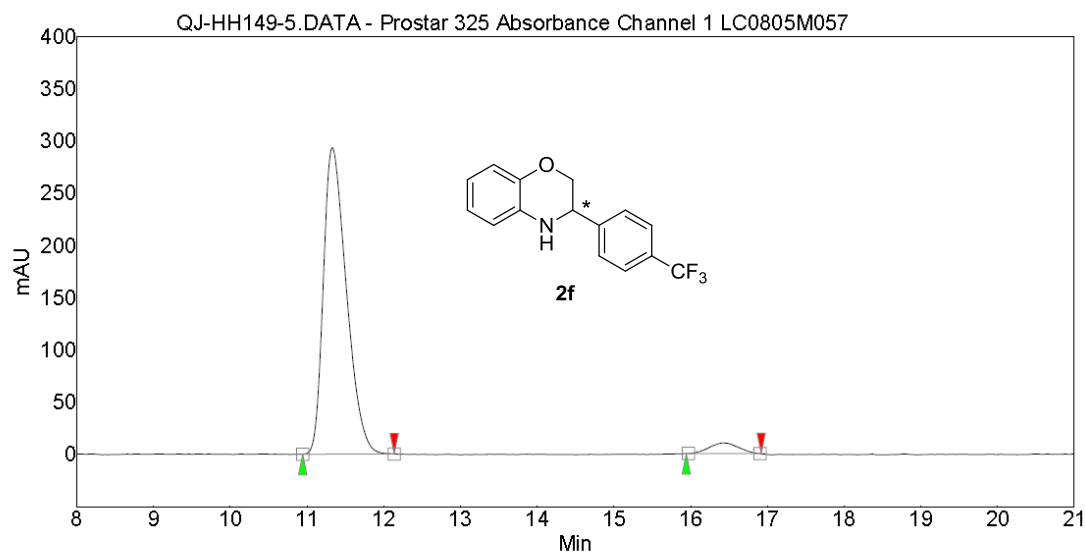


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	12.59	97.91	511.0	187.3	97.912
2	未知	22.54	2.09	6.7	4.0	2.088
Total			100.00	517.7	191.3	100.000

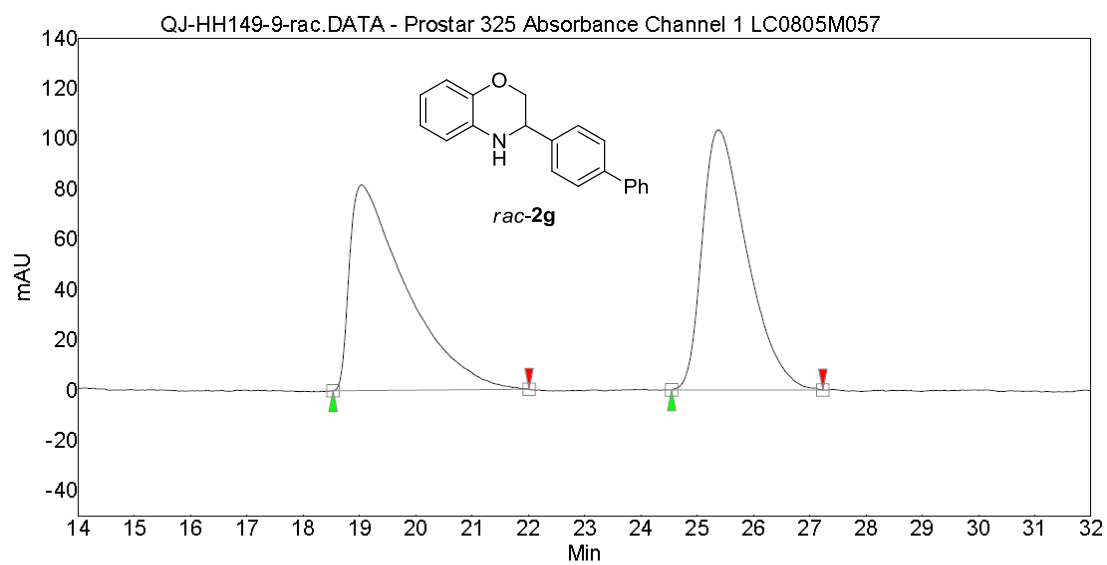




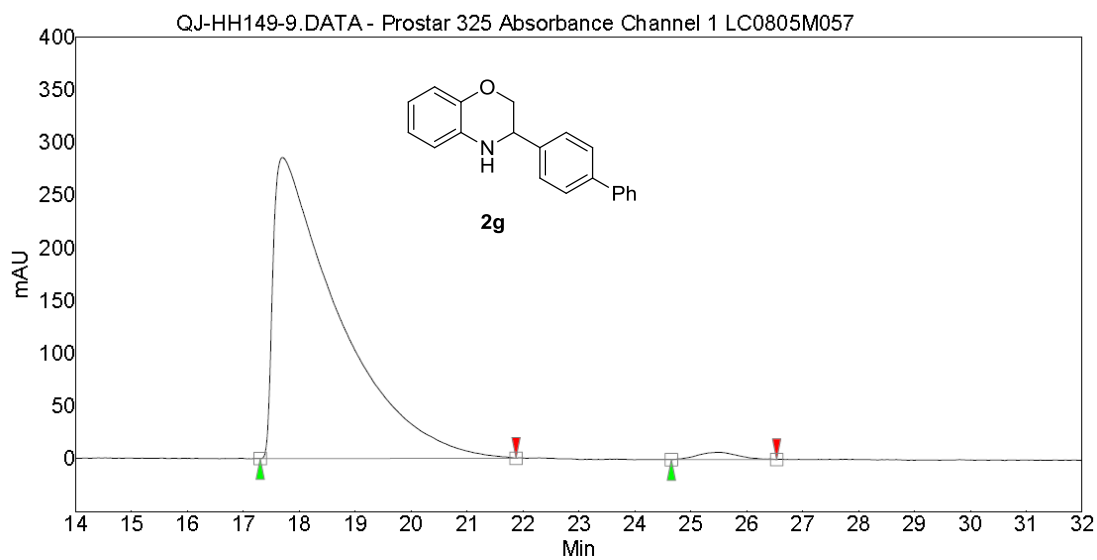
Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	11.55	50.06	221.0	76.7	50.060
2	未知	16.87	49.94	146.7	76.5	49.940
Total			100.00	367.6	153.2	100.000



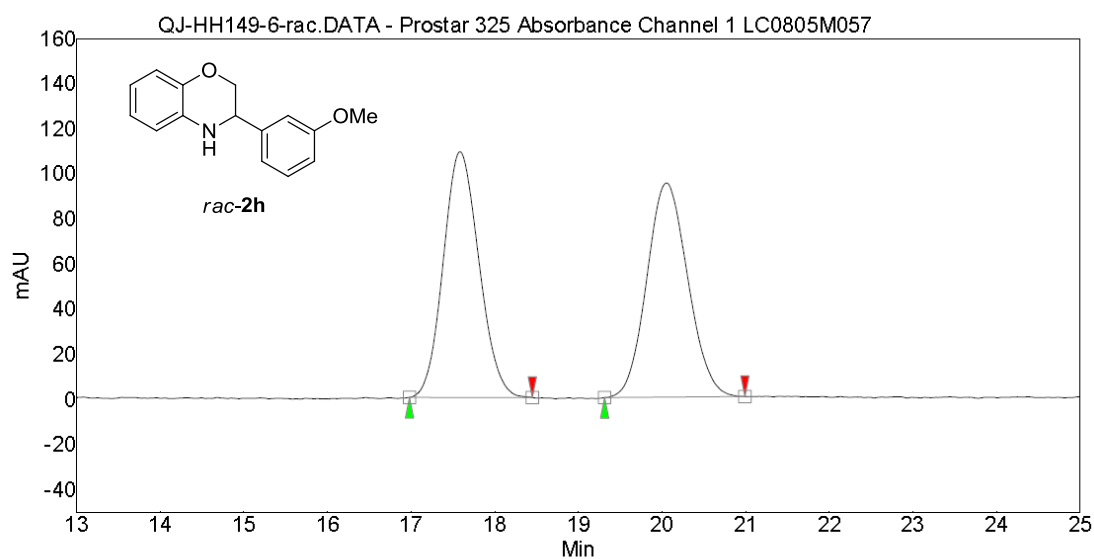
Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	11.33	95.82	293.1	102.1	95.816
2	未知	16.42	4.18	10.1	4.5	4.184
Total			100.00	303.2	106.6	100.000



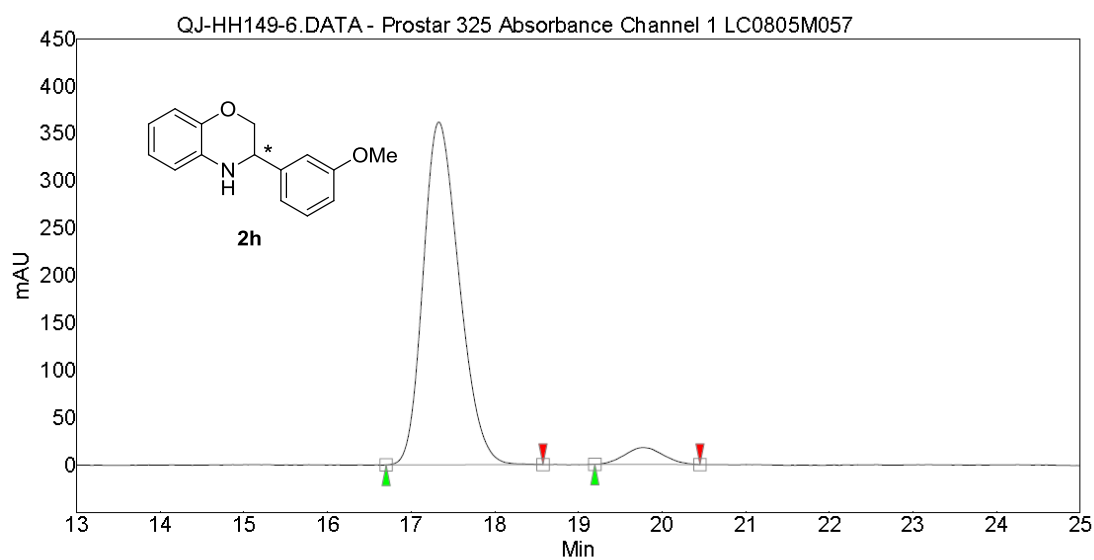
Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
2	未知	19.03	50.03	81.7	95.4	50.027
1	未知	25.37	49.97	103.4	95.3	49.973
Total			100.00	185.1	190.7	100.000



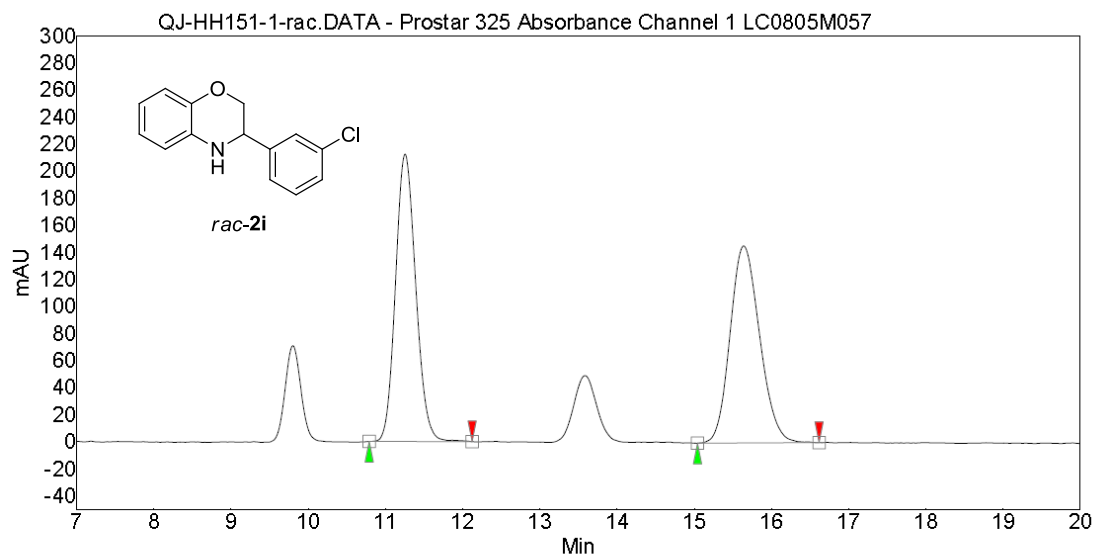
Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
2	未知	17.70	98.50	285.3	385.1	98.504
1	未知	25.49	1.50	6.9	5.8	1.496
Total			100.00	292.3	391.0	100.000



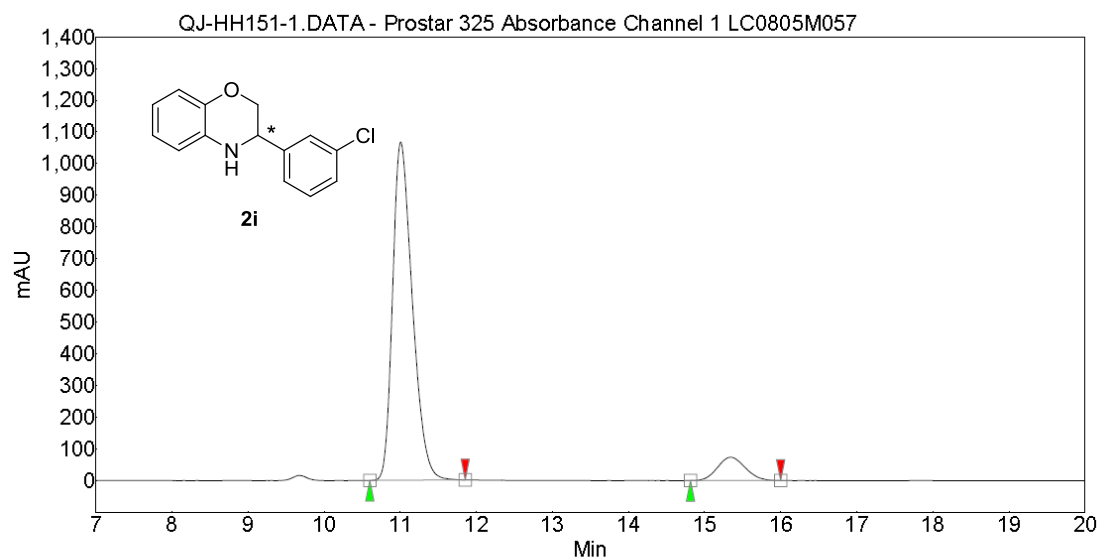
Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
2	未知	17.59	49.96	108.9	53.9	49.963
1	未知	20.05	50.04	94.9	54.0	50.037
Total			100.00	203.9	107.9	100.000



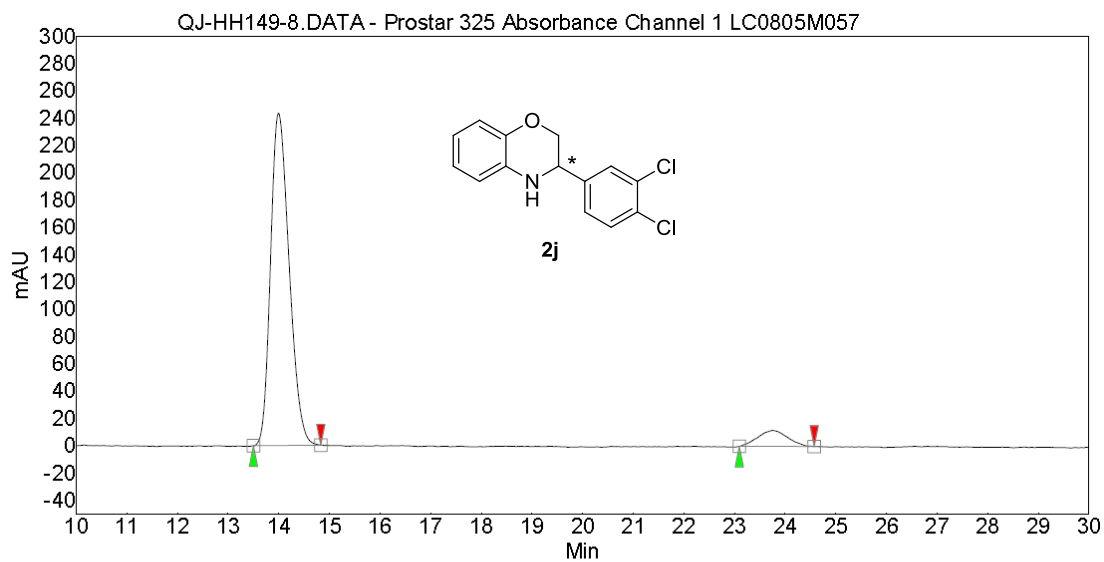
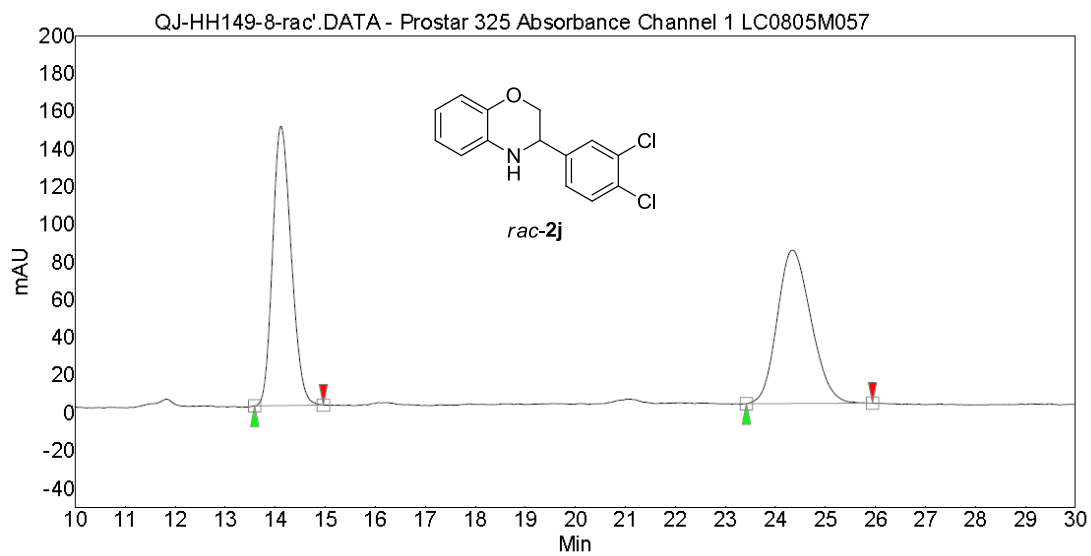
Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
2	未知	17.33	94.99	361.7	180.5	94.988
1	未知	19.77	5.01	17.8	9.5	5.012
Total			100.00	379.5	190.0	100.000

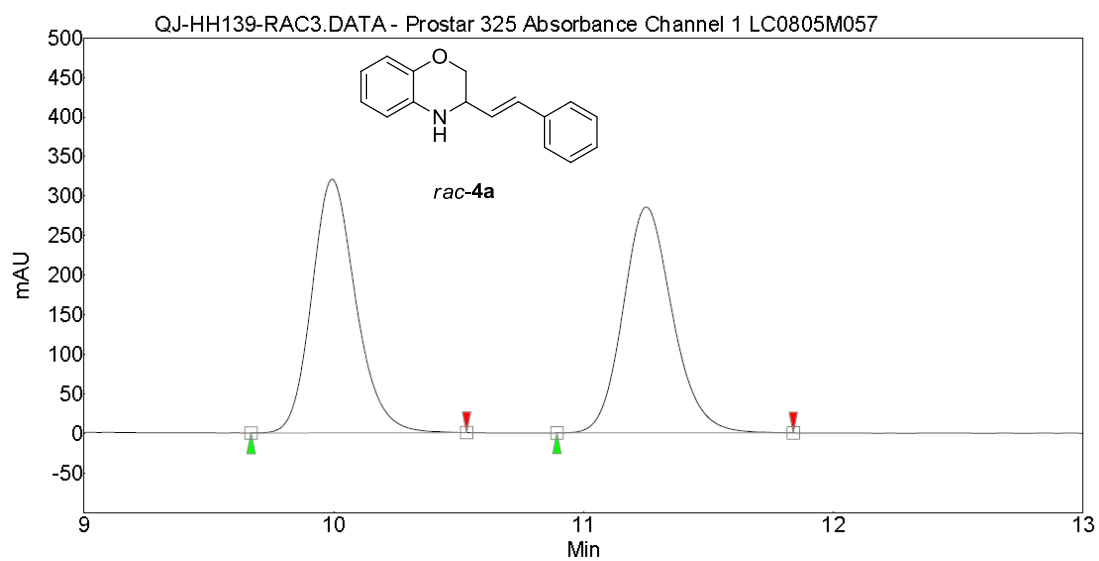


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
2	未知	11.25	49.99	212.3	64.6	49.993
1	未知	15.64	50.01	145.6	64.6	50.007
Total			100.00	357.9	129.2	100.000

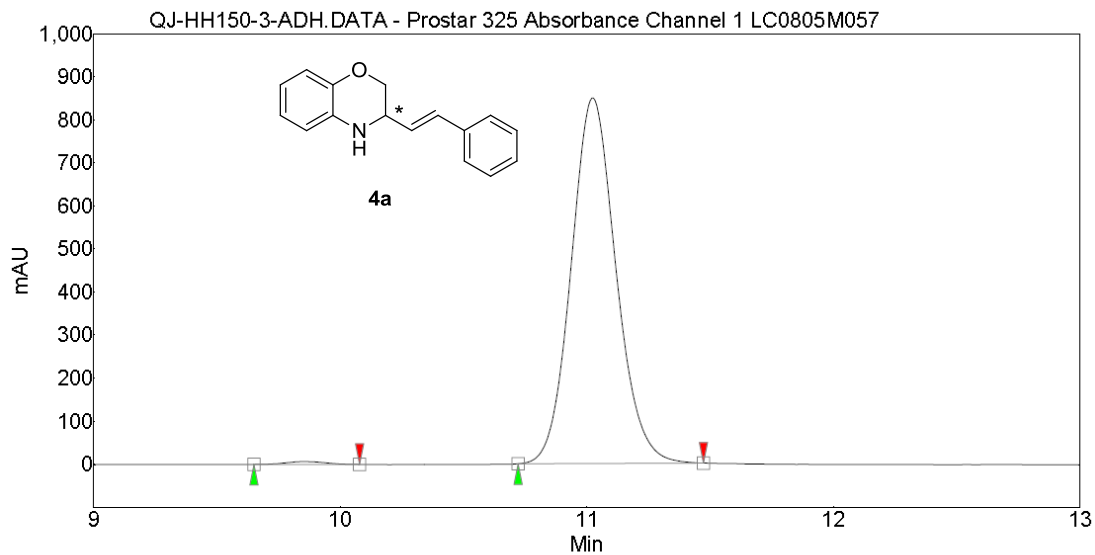


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	11.01	91.21	1067.5	320.7	91.210
2	未知	15.34	8.79	73.4	30.9	8.790
Total			100.00	1140.9	351.6	100.000

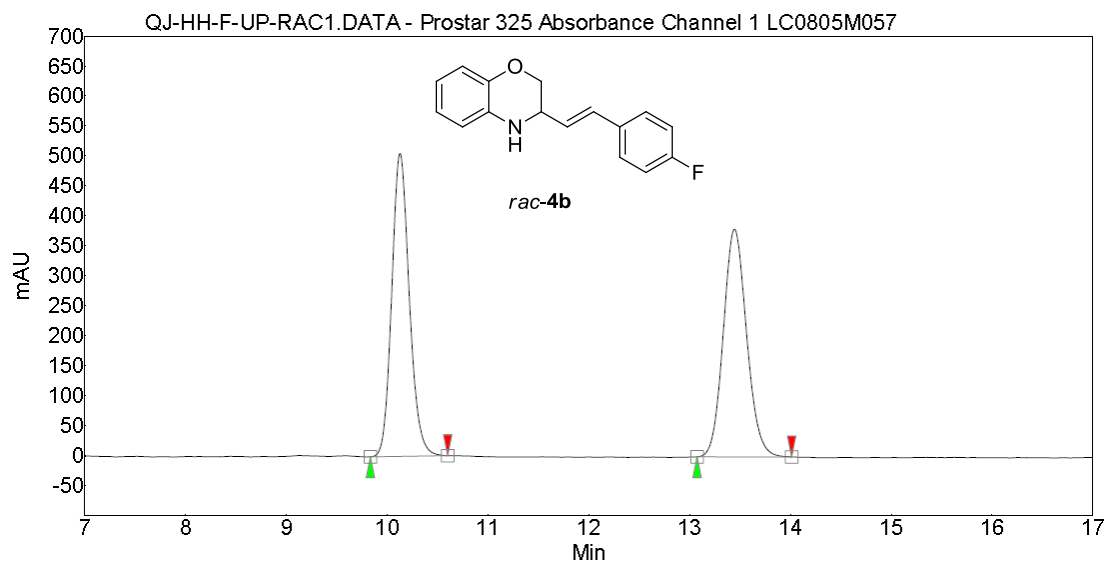




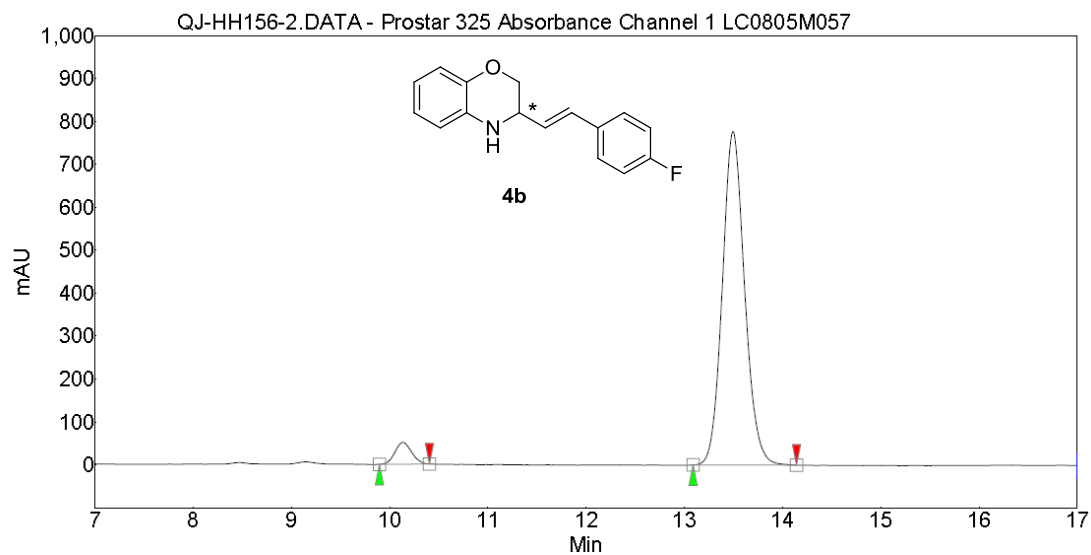
Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	9.99	49.99	320.6	65.7	49.987
2	未知	11.25	50.01	285.5	65.7	50.013
Total			100.00	606.2	131.4	100.000



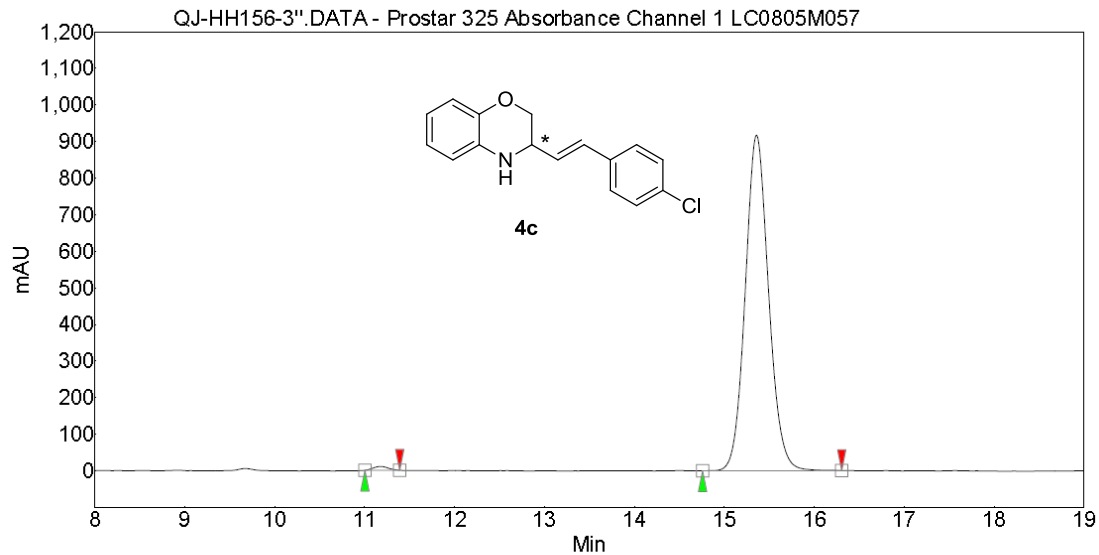
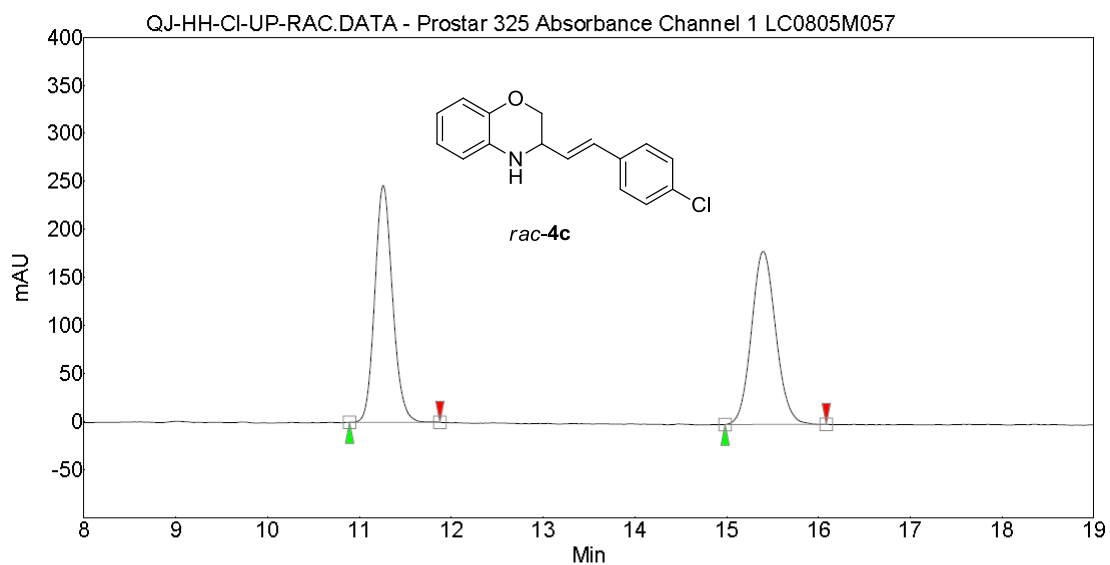
Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	9.85	0.67	6.6	1.2	0.666
2	未知	11.02	99.33	849.0	178.5	99.334
Total			100.00	855.6	179.7	100.000

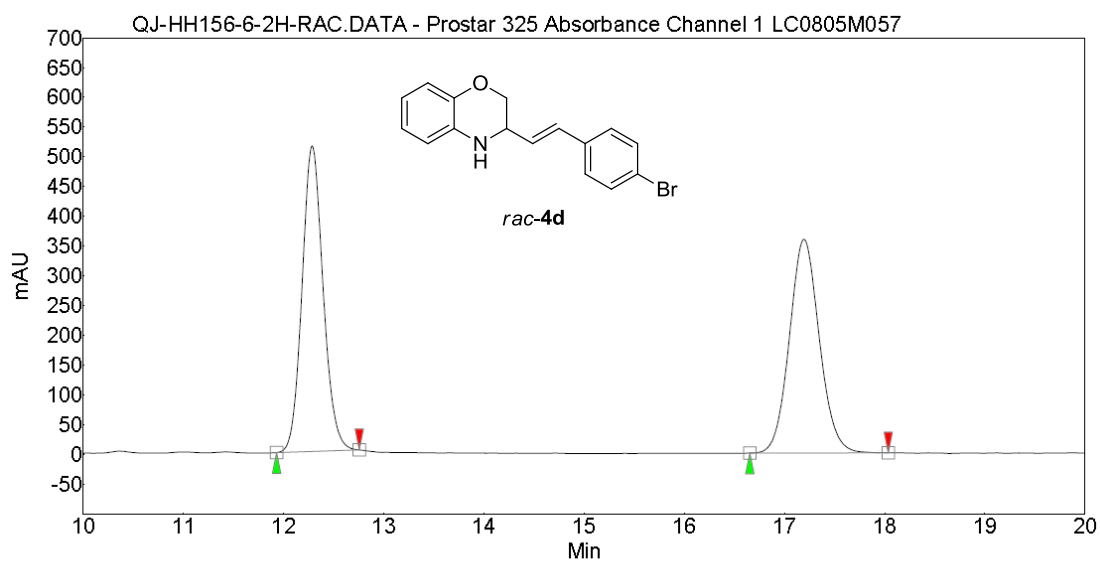


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	10.13	50.02	505.9	102.1	50.023
2	未知	13.45	49.98	380.2	102.0	49.977
Total			100.00	886.2	204.2	100.000

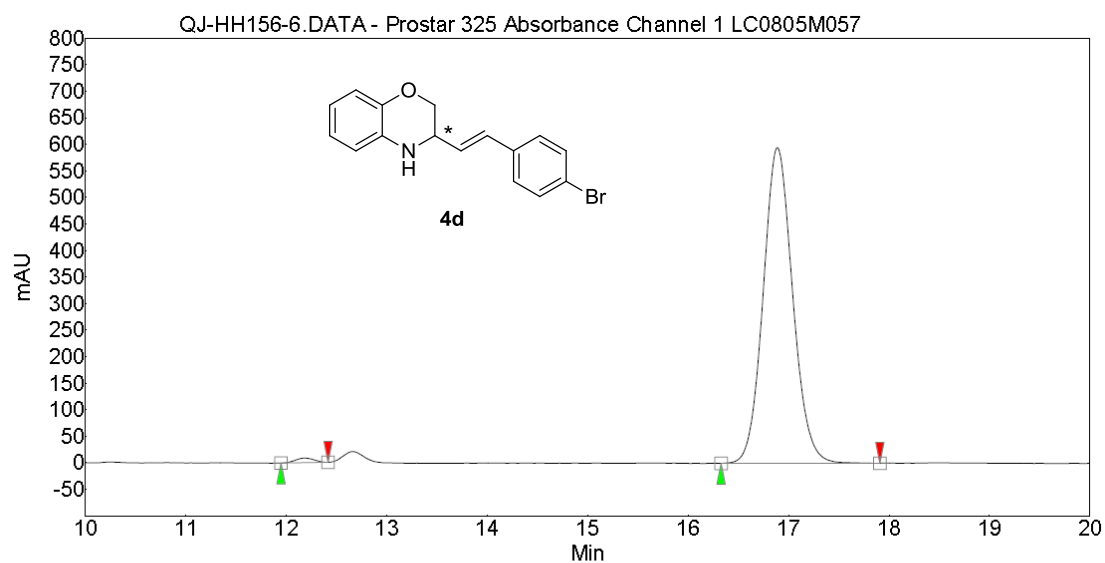


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	10.13	4.68	50.8	10.0	4.680
2	未知	13.49	95.32	776.9	203.9	95.320
Total			100.00	827.6	213.9	100.000

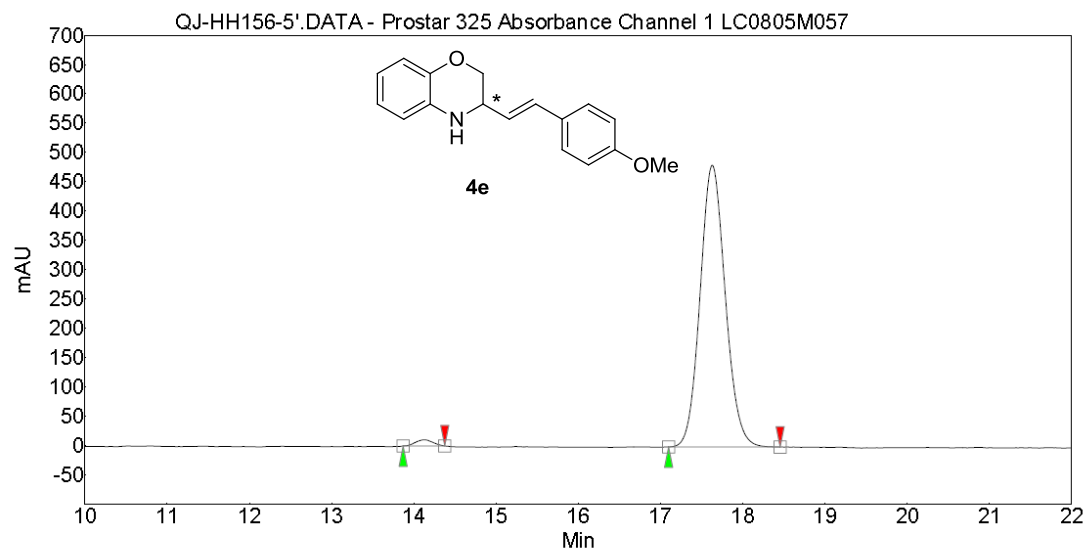
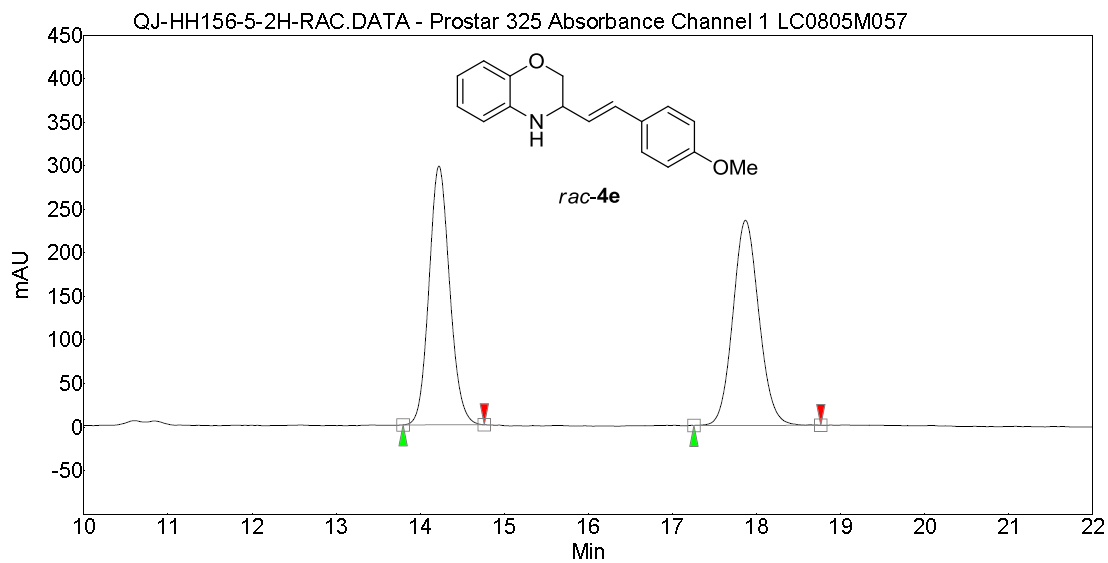


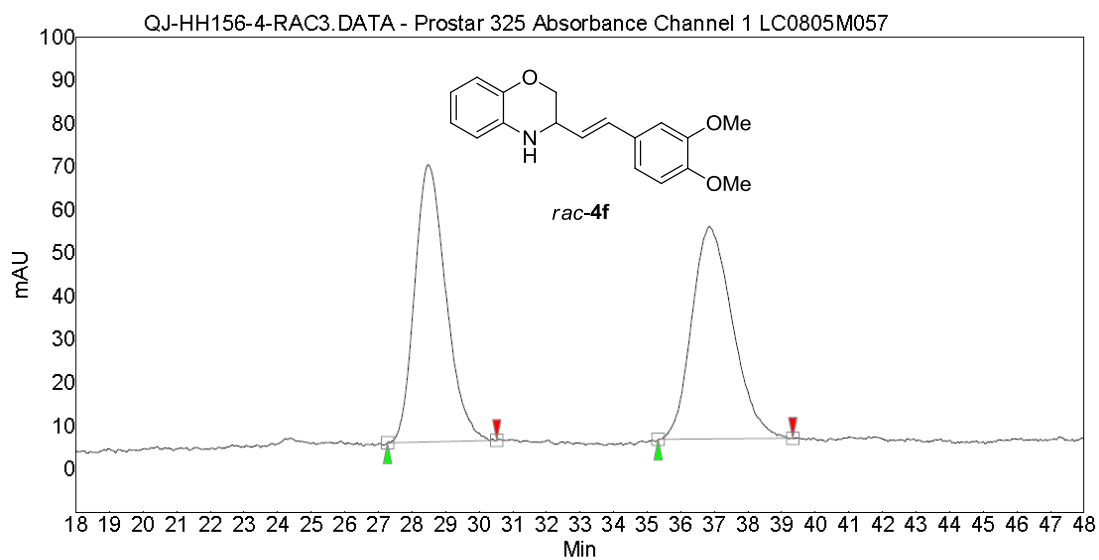


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
2	未知	12.29	50.15	513.3	125.0	50.153
1	未知	17.19	49.85	359.2	124.2	49.847
Total			100.00	872.5	249.2	100.000

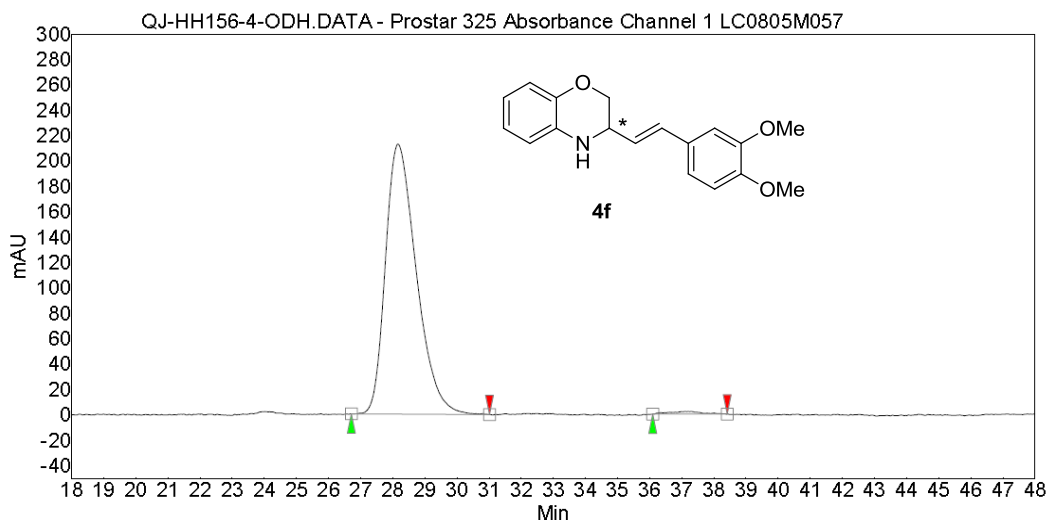


Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
2	未知	12.18	0.90	8.6	1.8	0.904
1	未知	16.89	99.10	594.7	202.2	99.096
Total			100.00	603.3	204.0	100.000





Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	28.49	49.97	64.2	69.2	49.972
2	未知	36.85	50.03	49.2	69.3	50.028
Total			100.00	113.4	138.5	100.000



Index	文件名	时间 [Min]	数量 [% 面积]	高度 [mAU]	Area [mAU.Min]	Area % [%]
1	未知	28.16	98.93	212.7	236.6	98.931
2	未知	37.10	1.07	2.2	2.6	1.069
Total			100.00	214.9	239.2	100.000