

## SUPPORTING INFORMATION

### Palladium nanoparticles in glycerol: clear-cut catalyst for one-pot multi-step processes applied in the synthesis of heterocyclic compounds

Faouzi Chahdoura,<sup>a</sup> Sonia Mallet-Ladeira<sup>b</sup> and Montserrat Gómez<sup>\*a</sup>

#### Experimental part

**General.** All preparations and manipulations were performed using standard Schlenk techniques under argon atmosphere. Unless stated otherwise, commercially compounds were used without further purification. Glycerol was treated under vacuum at 80 °C overnight prior to use. NMR spectra were recorded on a Bruker Avance 300 spectrometer at 293 K (299.7 MHz for <sup>1</sup>H NMR and 75.5 MHz for <sup>13</sup>C NMR). GC analyses were carried out on an Agilent GC6890 with a flame ionization detector, using a SGE BPX5 column composed by 5% of phenylmethylsiloxane. IR spectra were recorded in the range of 4000-400 cm<sup>-1</sup> on a Perkin Elmer Spectrum One FT-IR. Mass chromatograms were carried out by the “Service commun de spectrométrie de masse” of the “Institute de Chimie de Toulouse de l’Université Paul Sabatier”. Electronic impact (EI) and Chemical ionization (CI using methane or ammonia as reactant gas), on a TSQ 7000 Thermo Electron apparatus and electrospray (ES) on a API-365 MS/MS Spectrometer (Perkin Elmer Sciex). Gas chromatography analyses were performed with a Perkin–Elmer Clarus 500 chromatograph fitted with a FID and MS-detector, using dodecane as internal standard. The column employed was SGE BPX5 (30 m x 0.32 mm x 0.25 mm) phase composed by 5% of phenyl methylsiloxane. The injector temperature was 250 °C and the flow 2 mL/min. Temperature programme: 40 °C for 2 min; 10 °C/min to 300 °C hold for 5 min. SFC analyses (CO<sub>2</sub>/methanol, 4 mL/min) were carried out at 35 °C using a Chiralpak OJ-H 5 µm column and a UV PDA detector. TEM images of particles dispersed in glycerol and in solid state were obtained from transmission electron microscopes JEOL JEM 1400 running at 120 kV and JEOL JEM 2100F running at 200 kV equipped with X PGT analyzer (detection of light elements, resolution 135 eV), at the “Service Commun de Microscopie Electronique de l’Université Paul Sabatier, TEMSCAN”. A drop of solution was deposited on a holey carbon grid and the excess of glycerol was removed in order to obtain a film as thin as possible. The nanoparticles size distribution and average diameter were directly determined from TEM images by Image-J software associated to a Microsoft Excel macro developed by Christian Pradel.

#### Crystallographic data collection and structure determination

The data were collected at low temperature (193 K) on a Bruker-AXS APEX II QUAZAR diffractometer equipped with a 30W air-cooled microfocus source (**a1**, **a30**, **s40u**, **r33** and **g8**) and on a Bruker-AXS SMART APEX II diffractometer (**n35** and **a17u**), using MoK $\alpha$  radiation ( $\lambda = 0.71073\text{Å}$ ). Phi- and omega- scans were used. The data were integrated with SAINT, and an empirical absorption correction with SADABS was applied.<sup>[1]</sup> The structures were solved by direct methods (SHELXS-97)<sup>[2]</sup> and refined using the least-squares method on  $F^2$ <sup>[2]</sup>. All non-H atoms were refined with anisotropic displacement parameters. The H atoms were refined isotropically at calculated positions using a riding model.

CCDC-1020806 (**a1**), CCDC-1020807 (**a30**), CCDC-1020808 (**s40u**), CCDC-1020809 (**r33**), CCDC-1020810 (**n35**), CCDC-1020811 (**a17u**) and CCDC-1020812 (**g8**) contain the

supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via [www.ccdc.cam.ac.uk/data\\_request/cif](http://www.ccdc.cam.ac.uk/data_request/cif).

#### References

- [1] SAINT and SADABS. Bruker AXS Inc., Madison, Wisconsin, USA.  
[2] G. M. Sheldrick, *Acta Cryst.*, 2008, **A64**, 112-122.

#### Synthesis of Palladium Nanoparticles in glycerol

Pd(OAc)<sub>2</sub> ( $5.10^{-2}$  mmol, 11.2 mg) both in the presence of m-TPPTS ( $5.10^{-2}$  mmol, 28.4 mg), were dissolved in 5 mL of glycerol and stirred at room temperature under argon in a Fischer-Porter bottle until complete dissolution. The system was then pressurised with 3bar of dihydrogen and stirred at 60 °C overnight, leading to a black solution. After releasing the residual gas, the solution was washed with pentane (2x5 mL) under argon atmosphere and dried under reduced pressure for 1 h.

#### General Procedure for Synthesis of N-Substituted Phthalimides catalyzed by PdNPs in glycerol.

o-Dihaloarene/o-iododerivative (0.4 mmol), amine (0.6 mmol), and DABCO (1 mmol) were consecutively added to 1 mL of a solution of preformed nanoparticles in glycerol. The resulting mixture was stirred at room temperature under argon in a Fisher-Porter bottle. The system was then pressurised with carbon monoxide (0.5 bar) and stirred at 120 °C for 30 min. The mixture was then cooled to room temperature. The catalytic mixture was extracted with dichloromethane (3 x 10 mL) and the combined organic phases were dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and the solvent evaporated under reduced pressure. The product was purified by short column chromatography on silica gel. All the products were previously reported, and identified by comparison of their <sup>1</sup>H and <sup>13</sup>C NMR spectra and GC-MS data with those of authentic samples.

#### Pd-catalysed domino Sonogashira reaction/Cycloisomerization process. Synthesis of Benzo[b]furan in glycerol

Aryl halide (0.4 mmol), the corresponding terminal alkyne (0.6 mmol), t-BuOK (1 mmol) were consecutively added to 1 mL of a solution of preformed nanoparticles in glycerol. The resulting mixture was heated at 100 °C during 2 h and then cooled to room temperature. The organic products were extracted from the catalytic mixture with dichloromethane (3 x 10 mL) and the combined organic phases were dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and the solvent evaporated under reduced pressure. The product was purified by short-column chromatography on silica gel. All the products were previously reported, and identified by comparison of their <sup>1</sup>H and <sup>13</sup>C NMR spectra and GC-MS data with those of authentic samples.

#### Pd-catalysed sequential processes: carbonylative cyclization followed by domino reaction Sonogashira/Cycloisomerization

1,2-Diiodobenzene (0.4 mmol, 131.96 mg), 4-pentyn-1-amine (0.6 mmol, 49.87 mg), and DABCO (1 mmol, 112.17 mg) were consecutively added to 1 mL of a solution of preformed nanoparticles in glycerol (0.01 mmol Pd, 2.5 mol%). The resulting mixture was stirred at room temperature under argon in a Fisher-Porter bottle. The system was then pressurised with carbon monoxide (0.5 bar) and stirred at 120 °C for 30 min and then cooled to room temperature. The system were added 2-Iodophenol (0.4 mmol, 88.01 mg). The resulting mixture was heated at 100 °C for 2 h. The mixture was then cooled to room temperature. The organic products were extracted from the catalytic mixture with dichloromethane or dichloromethane (3x10 mL), and the combined organic phases were dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and the solvent was

evaporated under reduced pressure. The product was purified by short-column chromatography on silica gel. Yield: for xx, 111.10 mg (91%)

**Pd-catalysed sequential process: carbonylative cyclization followed by hydrogenation**

o-dihaloarene/o-iododerivative (0.4 mmol), amine (0.6 mmol), and DABCO (1 mmol) were consecutively added to 1 mL of a solution of preformed nanoparticles in glycerol (0.01 mmol Pd, 2.5 mol%). The resulting mixture was stirred at room temperature under argon in a Fisher-Porter bottle. The system was then pressurised with carbon monoxide (0.5 bar) and stirred at 120 °C for 30 min and then cooled to room temperature. The system was then pressurized with dihydrogen (3 bar) and stirred at 100 °C for 2 h. The mixture was then cooled to room temperature. The catalytic mixture was extracted with dichloromethane (3 x 10 mL) and the combined organic phases were dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and the solvent evaporated under reduced pressure. The product was purified by short-column chromatography on silica gel. The corresponding product was identified by comparison of its <sup>1</sup>H and <sup>13</sup>C NMR spectra and GC–MS data with those of an authentic sample.

**Pd-catalysed domino process: Sonogashira coupling/Cycloisomerization process. Synthesis of isoindolinone in glycerol**

2-Iodo-benzamide (0.4 mmol), terminal alkyne (0.6 mmol), and t-BuOK (1 mmol) were consecutively added to 1 mL of a solution of preformed nanoparticles in glycerol (0.01 mmol Pd, 2.5 mol%). The resulting mixture was heated at 100 °C during 2 h and then cooled to room temperature. The organic products were extracted from the catalytic mixture with dichloromethane (3 x 10 mL) and the combined organic phases were dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and the solvent evaporated under reduced pressure. The product was purified by short-column chromatography on silica gel. All the products were previously reported, and identified by comparison of their <sup>1</sup>H and <sup>13</sup>C NMR spectra and GC-MS data with those of authentic samples.

**Pd-catalysed sequential process: domino Sonogashira/Cycloisomerization process followed by hydrogenation**

2-Iodo-benzamide (0.4 mmol), terminal alkyne (0.6 mmol), and t-BuOK (1 mmol) were consecutively added to 1 mL of a solution of preformed nanoparticles in glycerol (0.01 mmol Pd, 2.5 mol%). The resulting mixture was heated at 100 °C during 2 h and then cooled to room temperature. The system was then pressurized with dihydrogen (3 bar) and stirred at 100 °C for 2 h. The mixture was then cooled to room temperature. The catalytic mixture was extracted with dichloromethane (3 x 10 mL) and the combined organic phases were dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and the solvent evaporated under reduced pressure. The product was purified by short-column chromatography on silica gel. The corresponding product was identified by comparison of its <sup>1</sup>H and <sup>13</sup>C NMR spectra and GC–MS data with those of an authentic sample.

**Pd/Cu-catalysed sequential process: domino carbonylative cyclization followed by Azide–Alkyne cycloaddition**

1,2-Diodobenzene (0.4 mmol, 131.96 mg), 4-pentyn-1-amine (0.6 mmol, 49.87 mg), and DABCO (1 mmol, 112.17 mg) were consecutively added to 1 mL of a solution of preformed nanoparticles in glycerol (0.01 mmol Pd, 2.5 mol%). The resulting mixture was stirred at room temperature under argon in a Fisher-Porter bottle. The system was then pressurised with carbon monoxide (0.5 bar) and stirred at 120 °C for 30 min and then cooled to room temperature. The system were added consecutively to a solution of preformed Cu<sub>2</sub>ONP in glycerol (1 mL, 0.01 mmol of Cu) and benzyl azide (0.4 mmol). The resulting mixture was heated at 100 °C for 2 h. The mixture was then cooled to room temperature. The organic products were extracted from

the catalytic mixture with dichloromethane or dichloromethane (3×10 mL), and the combined organic phases were dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and the solvent was evaporated under reduced pressure. The product was purified by short-column chromatography on silica gel. Yield: for xx, 128.70 mg (93%)

### **Pd/Cu-catalysed sequential process: domino Sonogashira/Cycloisomerization followed by Azide–Alkyne cycloaddition**

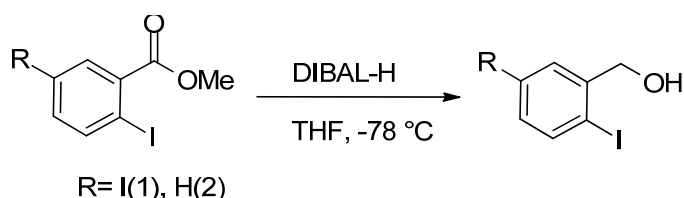
2-Iodophenol (0.4 mmol, 88.01 mg), 1,7-Octadiyne (0.6 mmol, 63.70 mg), t-BuOK (1 mmol, 112.21 mg) were consecutively added to 1 mL of a solution of preformed nanoparticles in glycerol (0.01 mmol Pd, 2.5 mol%). The resulting mixture was heated at 100 °C during 2 h and then cooled to room temperature. The system were added consecutively to a solution of preformed Cu<sub>2</sub>ONP in glycerol (1 mL, 0.01 mmol of Cu) and benzyl azide (0.4 mmol). The resulting mixture was heated at 100 °C for 2 h. The mixture was then cooled to room temperature. The organic products were extracted from the catalytic mixture with dichloromethane or dichloromethane (3×10 mL), and the combined organic phases were dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and the solvent was evaporated under reduced pressure. The product was purified by short-column chromatography on silica gel. Yield: for xx, 125.70 mg (95%)

### **General Procedure for Recycling the Catalytic Phase**

A typical experimental procedure to recycle the catalytic phase is described. The glycerol phase from the previous run was maintained for 30 min under dynamic vacuum while stirring at 100 °C. Then, the corresponding reagents were added to the glycerol phase. The catalytic mixture was the treated under the corresponding conditions applied for the first run, the final product was extracted with dichloromethane and analyzed by GC and NMR.

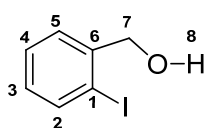
### **Synthesis of starting materials**

a) Synthesis of benzylic alcohols (*Org. Lett.*, 2012, **14**, 3264)



To solution of 1 or 2 (3.79 mmol) in THF (25mL) was added DIBAL-H ( 9.5 mL, 9.47 mmol) at -78 °C. After completion, the mixture was added 1M NaOH (2mL) and MgSO<sub>4</sub>. The reaction mixture was filtered and concentrated under reduce pressure. The residue was purified by flash chromatography on silica (30% EtOAc/hexane) to give a white solid.

**(2-iodophenyl)methanol.** Yield 95%. The spectroscopic data was in good agreement with that reported previously.



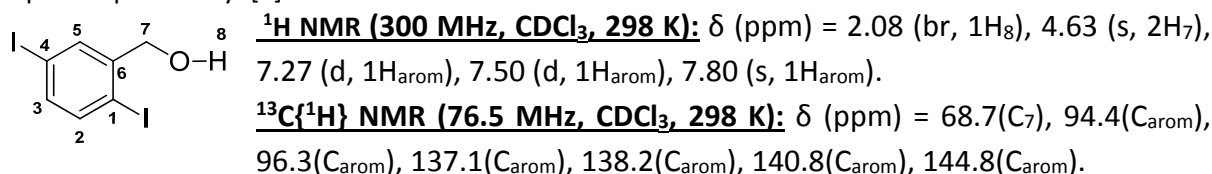
**<sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, 298 K):** δ (ppm) = 2.02 (br, 1H<sub>8</sub>), 4.68 (s, 2H<sub>7</sub>), 6.97 (m, 1H<sub>arom</sub>), 7.34 (m, 1H<sub>arom</sub>), 7.45 (m, 1H<sub>arom</sub>), 7.84 (m, 1H<sub>arom</sub>).

**<sup>13</sup>C{<sup>1</sup>H} NMR (76.5 MHz, CDCl<sub>3</sub>, 298 K):** δ (ppm) = 69.4(C<sub>7</sub>), 97.6(C<sub>arom</sub>), 128.5(C<sub>arom</sub>), 128.6(C<sub>arom</sub>), 129.4(C<sub>arom</sub>), 139.3(C<sub>arom</sub>), 142.6(C<sub>arom</sub>).

**M/S (EI):** t<sub>r</sub>= 9.67 min, calc. for C<sub>7</sub>H<sub>7</sub>IO: 233.95. Found: (m/z) = 233.71 (M<sup>+</sup>, 100).



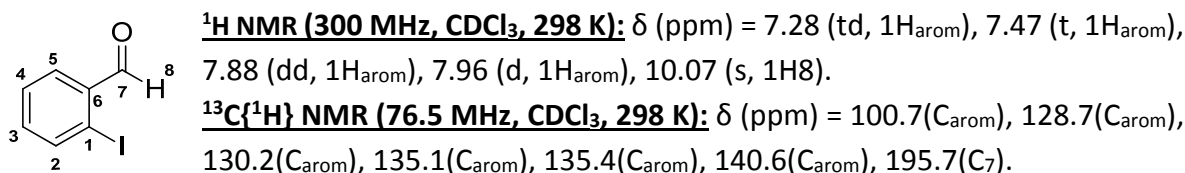
**2,5-diiodophenyl)methanol.** Yield 95%. The spectroscopic data was in good agreement with that reported previously. [1]



**M/S (EI):**  $t_r$  = 12.77 min, calc. for  $\text{C}_7\text{H}_6\text{I}_2\text{O}$ : 359.8. Found: ( $m/z$ ) = 359.9 ( $\text{M}^+$ , 100).

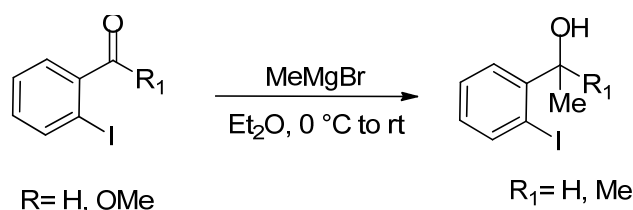
a) Synthesis of 2-iodobenzylalcohol (J. Tummatorn and G. Dudley, *Org. Lett.*, 2011, **13**, 1572)

To a solution of 2-iodobenzylalcohol (1.31 g, 5.60 mmol) in DCM was added PCC (1.33 g, 6.16 mmol). after being stirred at room temperature for 2 h, the reaction solvent was removed in vacuum and the residue was dissolved with 30% EtOAc/hexane and filtered through a silica gel plug. Evaporation gave 1.26 g (97%) of 2-iodobenzaldehyde as a white solid.



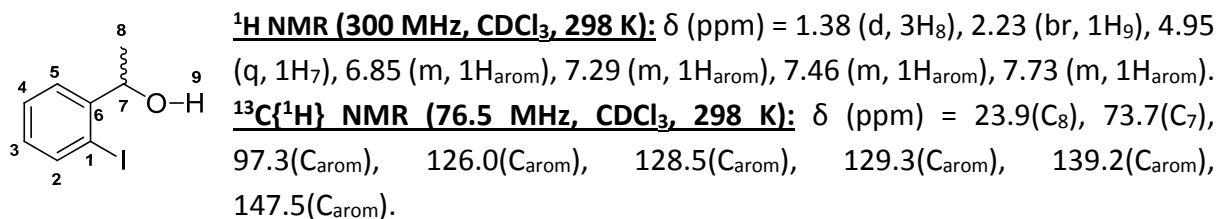
**M/S (EI):**  $t_r$  = 9.07 min, calc. for  $\text{C}_7\text{H}_5\text{IO}$ : 231.9. Found: ( $m/z$ ) = 231.1 ( $\text{M}^+$ , 100).

c) Synthesis of 2-iodophenylalcohols



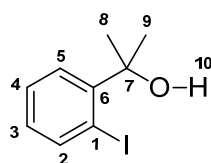
A solution of methyl Grignard reagent in Et<sub>2</sub>O (4.5 mmol) was added to a stirred solution of *o*-iodobenzaldehyde or methyl-*o*-iodobenzoate (3.0 mmol) in Et<sub>2</sub>O (15 mL) at 0 °C. The mixture was warmed to room temperature over 1 h, monitoring through TLC (30% EtOAc in hexanes). The reaction was quenched through the addition of saturated aqueous NH<sub>4</sub>Cl (15 mL), the aqueous phase was extracted with Et<sub>2</sub>O (3\*20 mL). The combined organic phases were dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated in vacuum. The residue was purified through FCC (SiO<sub>2</sub>: 30% EtOAc in hexanes) to yield a white solid.

**1-(2-iodophenyl)ethanol.** Yield 95%. The spectroscopic data was in good agreement with that reported previously (D. Sälinger and R. Brückner, *Chem. Eur. J.*, 2009, **15**, 6688)



**M/S (EI):**  $t_r$  = 9.53 min, calc. for  $\text{C}_8\text{H}_9\text{IO}$ : 247.71. Found: ( $m/z$ ) = 247.54 ( $\text{M}^+$ , 100).

**2-(2-iodophenyl)propan-2-ol.** Yield 91%. The spectroscopic data was in good agreement with that reported previously (R. Moss, K. Jespersen and J. Westbrook, *J. Am. Chem. Soc.*, 1989, **111**, 250)

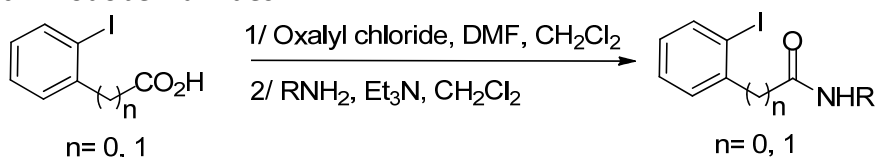


**$^1\text{H NMR}$  (300 MHz,  $\text{CDCl}_3$ , 298 K):**  $\delta$  (ppm) = 1.65 (s, 6 $\text{H}_{8,9}$ ), 2.58 (br, 1 $\text{H}_{10}$ ), 6.78 (m, 1 $\text{H}_{\text{arom}}$ ), 7.22 (m, 1 $\text{H}_{\text{arom}}$ ), 7.55 (m, 1 $\text{H}_{\text{arom}}$ ), 7.85 (m, 1 $\text{H}_{\text{arom}}$ ).

**$^{13}\text{C}\{^1\text{H}\}$  NMR (76.5 MHz,  $\text{CDCl}_3$ , 298 K):**  $\delta$  (ppm) = 29.7( $\text{C}_{8,9}$ ), 72.1( $\text{C}_7$ ), 93.3( $\text{C}_{\text{arom}}$ ), 126.8( $\text{C}_{\text{arom}}$ ), 128.1( $\text{C}_{\text{arom}}$ ), 128.6( $\text{C}_{\text{arom}}$ ), 142.8( $\text{C}_{\text{arom}}$ ), 148.6( $\text{C}_{\text{arom}}$ ).

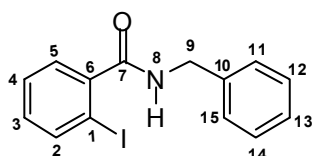
**M/S (EI):**  $t_r$  = 9.94 min, calc. for  $\text{C}_9\text{H}_{11}\text{IO}$ : 261.9. Found: ( $m/z$ ) = 261.2 ( $\text{M}^+$ , 100).

c) Synthesis of 2-iodobenzamides



To a solution of the corresponding carboxylic acid (10 mmol) in  $\text{CH}_2\text{Cl}_2$  (20ml) was added oxalyl chloride (1.29 ml, 15 mmol) and DMF (73 $\mu\text{L}$ , 0.95 mmol). Stirring was continued till the solid disappeared. The volatile was removed under reduced pressure. Then  $\text{CH}_2\text{Cl}_2$  (20ml) was added followed by addition of the corresponding amine (10 mmol) and triethylamine (1.39 ml, 10 mmol). After being stirred at room temperature for another one hour, the mixture was quenched with water and the aqueous phase was extracted with  $\text{CH}_2\text{Cl}_2$ . The combined organic phase was washed with 10% HCl (aq.), Water, saturated  $\text{NaHCO}_3$  (aq.) and brine, successively, and dried over  $\text{Na}_2\text{SO}_4$ . Evaporation of the solvent afforded the corresponding amide used without further purification.

**N-benzyl-2-iodobenzamide.** Yield 99%. The spectroscopic data was in good agreement with that reported previously (B. Yao, C. Jaccoud, Q. Wang and J. Zhu, *Chem. Eur. J.*, 2012, **18**, 5864)

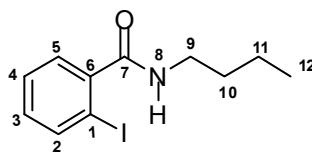


**$^1\text{H NMR}$  (300 MHz,  $\text{CDCl}_3$ , 298 K):**  $\delta$  (ppm) = 4.64 (s, 2 $\text{H}_9$ ), 6.16 (br, 1 $\text{H}_8$ ), 7.07 (m, 1 $\text{H}_{\text{arom}}$ ), 7.30 (m, 7 $\text{H}_{\text{arom}}$ ), 7.84 (d, 1 $\text{H}_{\text{arom}}$ ).

**$^{13}\text{C}\{^1\text{H}\}$  NMR (76.5 MHz,  $\text{CDCl}_3$ , 298 K):**  $\delta$  (ppm) = 44.3( $\text{C}_9$ ), 92.5( $\text{C}_{\text{arom}}$ ), 127.8( $\text{C}_{\text{arom}}$ ), 128.2( $\text{C}_{\text{arom}}$ ), 128.4( $\text{C}_{\text{arom}}$ ), 128.8( $\text{C}_{\text{arom}}$ ), 131.2( $\text{C}_{\text{arom}}$ ), 137.7( $\text{C}_{\text{arom}}$ ), 139.9( $\text{C}_{\text{arom}}$ ), 142.1( $\text{C}_{\text{arom}}$ ), 169.4( $\text{C}_7$ ).

**M/S (EI):**  $t_r$  = 15.32 min, calc. for  $\text{C}_{14}\text{H}_{12}\text{INO}$ : 337.16. Found: ( $m/z$ ) = 337.18 ( $\text{M}^+$ , 100).

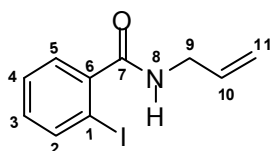
**N-butyl-2-iodobenzamide.** Yield 97%. The NMR spectral data was in good agreement with that reported previously (N. Schröder, J. Wencel-Delord, and F. Glorius, *J. Am. Chem. Soc.*, 2012, **134**, 8298)



**$^1\text{H NMR}$  (300 MHz,  $\text{CDCl}_3$ , 298 K):**  $\delta$  (ppm) = 0.80 (t, 3 $\text{H}_{12}$ ), 1.22 (m, 2 $\text{H}_{11}$ ), 1.44 (q, 2 $\text{H}_{10}$ ), 3.22 (q, 2 $\text{H}_9$ ), 6.82 (br, 1 $\text{H}_8$ ), 6.91 (m, 1 $\text{H}_{\text{arom}}$ ), 7.13 (m, 2 $\text{H}_{\text{arom}}$ ), 7.67 (d, 1 $\text{H}_{\text{arom}}$ ).

**$^{13}\text{C}\{^1\text{H}\}$  NMR (76.5 MHz,  $\text{CDCl}_3$ , 298 K):**  $\delta$  (ppm) = 13.4( $\text{C}_{12}$ ), 19.9( $\text{C}_{11}$ ), 30.8( $\text{C}_{10}$ ), 39.4( $\text{C}_9$ ), 92.4( $\text{C}_{\text{arom}}$ ), 127.5( $\text{C}_{\text{arom}}$ ), 127.7( $\text{C}_{\text{arom}}$ ), 130.3( $\text{C}_{\text{arom}}$ ), 139.1( $\text{C}_{\text{arom}}$ ), 142.1( $\text{C}_{\text{arom}}$ ), 159.4( $\text{C}_{\text{arom}}$ ), 169.0( $\text{C}_7$ ).

**M/S (EI):**  $t_r$  = 12.93 min, calc. for  $\text{C}_{11}\text{H}_{14}\text{INO}$ : 303.01. Found: ( $m/z$ ) = 303.07 ( $\text{M}^+$ , 100).

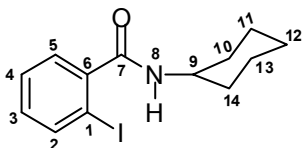


Yield 97%. The NMR spectral data was in good agreement with that reported previously (S. Couty, B. Liegault, C. Meyer and J. Cossy, *Tetrahedron*, 2006, **62**, 3882)

**$^1\text{H NMR}$  (300 MHz,  $\text{CDCl}_3$ , 298 K):**  $\delta$  (ppm) = 4.02 (d, 2H<sub>9</sub>), 5.24 (d, 2H<sub>11</sub>), 5.90 (m, 1H<sub>10</sub>), 6.09 (br, 1H<sub>8</sub>), 7.03 (m, 1H<sub>arom</sub>), 7.33 (m, 1H<sub>arom</sub>), 7.83 (d, 2H<sub>arom</sub>).

**$^{13}\text{C}\{^1\text{H}\}$  NMR (76.5 MHz,  $\text{CDCl}_3$ , 298 K):**  $\delta$  (ppm) = 42.2(C<sub>9</sub>), 92.5(C<sub>arom</sub>), 117.1(C<sub>11</sub>), 128.1(C<sub>arom</sub>), 128.4(C<sub>arom</sub>), 131.1(C<sub>arom</sub>), 133.7(C<sub>10</sub>), 139.9(C<sub>arom</sub>), 142.1(C<sub>arom</sub>), 169.2(C<sub>7</sub>).

**$\text{M/S (EI)}$**  :  $t_r$  = 12.63 min, calc. for C<sub>10</sub>H<sub>10</sub>INO: 287.21. Found: ( $m/z$ ) = 287.04 (M<sup>+</sup>, 100).

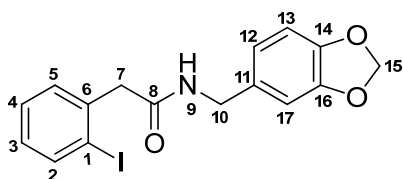


Yield 97%. The NMR spectral data was in good agreement with that reported previously.

**$^1\text{H NMR}$  (300 MHz,  $\text{CDCl}_3$ , 298 K):**  $\delta$  (ppm) = 1.12 (m, 4H<sub>11,13</sub>), 1.60 (m, 2H<sub>12</sub>), 1.80 (m, 4H<sub>10,14</sub>), 2.52 (m, 1H<sub>9</sub>), 7.62 (m, 1H<sub>arom</sub>), 7.80 (m, 1H<sub>arom</sub>), 7.97 (m, 2H<sub>arom</sub>), 8.03 (br, 1H<sub>8</sub>).

**$^{13}\text{C}\{^1\text{H}\}$  NMR (76.5 MHz,  $\text{CDCl}_3$ , 298 K):**  $\delta$  (ppm) = 24.8(C<sub>12</sub>), 24.9(C<sub>13</sub>), 25.7(C<sub>11</sub>), 32.3(C<sub>14</sub>), 32.4(C<sub>10</sub>), 51.6(C<sub>9</sub>), 92.8(C<sub>arom</sub>), 127.7(C<sub>arom</sub>), 130.7(C<sub>arom</sub>), 131.2(C<sub>arom</sub>), 141.5(C<sub>arom</sub>), 142.4(C<sub>arom</sub>), 167.2(C<sub>7</sub>).

**$\text{M/S (EI)}$**  :  $t_r$  = 15.04 min, calc. for C<sub>13</sub>H<sub>16</sub>INO: 329.03. Found: ( $m/z$ ) = 329.14 (M<sup>+</sup>, 100).

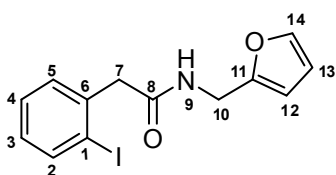


Yield 91%. The NMR spectral data was in good agreement with that reported previously.

**$^1\text{H NMR}$  (300 MHz,  $\text{CDCl}_3$ , 298 K):**  $\delta$  (ppm) = 3.85 (s, 2H<sub>10</sub>), 4.24 (s, 2H<sub>7</sub>), 6.07 (s, 2H<sub>15</sub>), 6.76 (m, 2H<sub>arom</sub>), 7.03 (m, 3H<sub>arom</sub>), 7.32 (m, 1H<sub>arom</sub>), 7.83 (m, 1H<sub>arom</sub>), 8.03 (br, 1H<sub>9</sub>).

**$^{13}\text{C}\{^1\text{H}\}$  NMR (76.5 MHz,  $\text{CDCl}_3$ , 298 K):**  $\delta$  (ppm) = 39.0(C<sub>7</sub>), 43.9(C<sub>10</sub>), 98.8(C<sub>arom</sub>), 101.2(C<sub>15</sub>), 107.7(C<sub>arom</sub>), 109.2(C<sub>arom</sub>), 128.1(C<sub>arom</sub>), 129.2(C<sub>arom</sub>), 130.6(C<sub>arom</sub>), 131.2(C<sub>arom</sub>), 145.9(C<sub>arom</sub>), 146.8(C<sub>arom</sub>), 144.8(C<sub>arom</sub>), 171.9(C<sub>8</sub>).

**$\text{M/S (EI)}$**  :  $t_r$  = 16.57 min, calc. for C<sub>16</sub>H<sub>14</sub>INO<sub>3</sub>: 395.01. Found: ( $m/z$ ) = 395.19 (M<sup>+</sup>, 100).



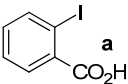
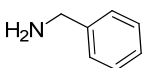
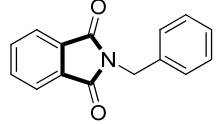
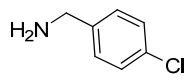
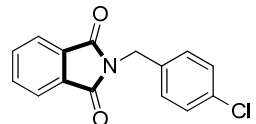
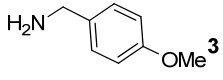
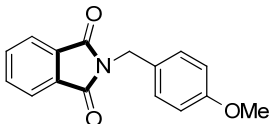
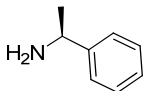
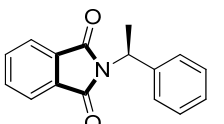
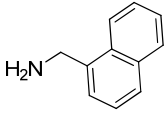
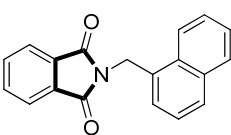
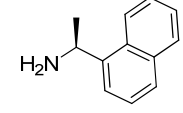
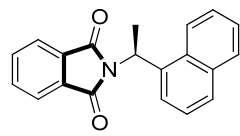
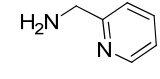
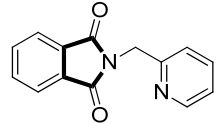
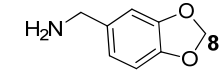
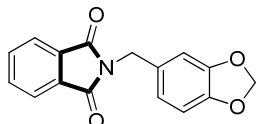
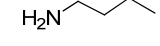
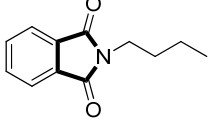
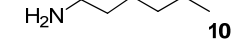
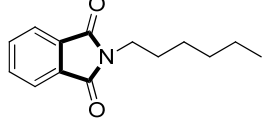
Yield 91%. The NMR spectral data was in good agreement with that reported previously.


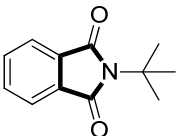
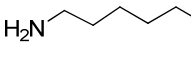
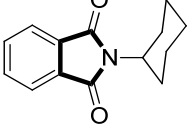
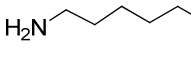
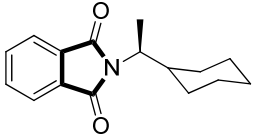
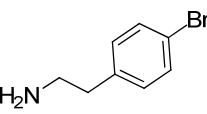
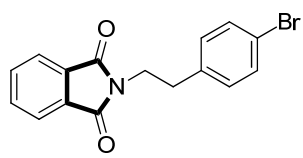
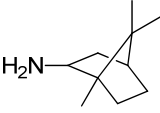
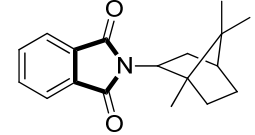
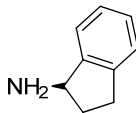
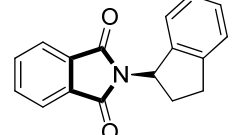
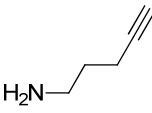
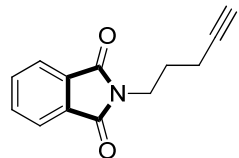
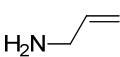
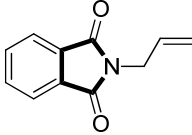
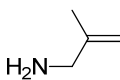
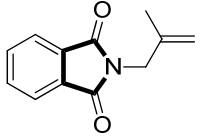
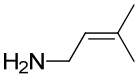
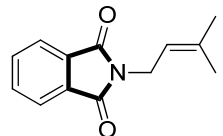
**$^1\text{H NMR}$  (300 MHz,  $\text{CDCl}_3$ , 298 K):**  $\delta$  (ppm) = 3.74 (s, 2H<sub>7</sub>), 4.43 (d, 2H<sub>10</sub>), 5.78 (br, 1H<sub>9</sub>), 6.30 (d, 2H<sub>12,13</sub>), 6.98 (m, 1H<sub>14</sub>), 7.34 (m, 3H<sub>arom</sub>), 7.87 (d, 1H<sub>arom</sub>).

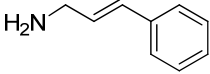
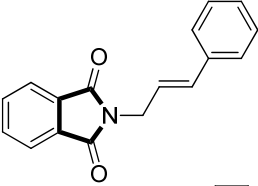
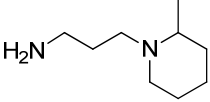
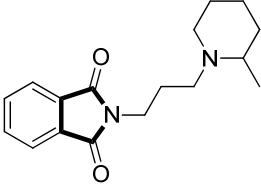
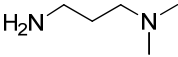
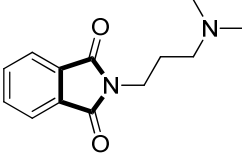
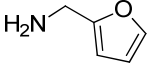
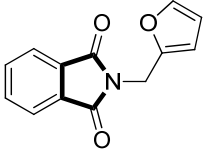
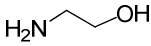
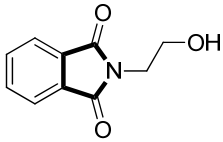
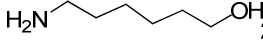
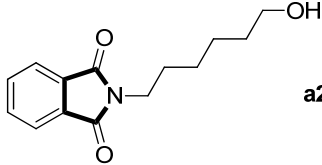
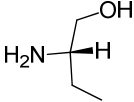
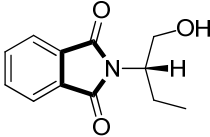
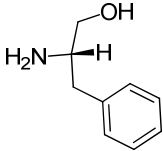
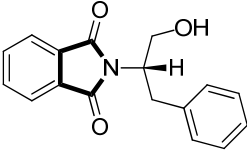
**$^{13}\text{C}\{^1\text{H}\}$  NMR (76.5 MHz,  $\text{CDCl}_3$ , 298 K):**  $\delta$  (ppm) = 36.8(C<sub>10</sub>), 48.6(C<sub>7</sub>), 101.5(C<sub>12</sub>), 107.4(C<sub>13</sub>), 110.5(C<sub>arom</sub>), 128.9(C<sub>arom</sub>), 129.2(C<sub>arom</sub>), 131.0(C<sub>arom</sub>), 138.1(C<sub>arom</sub>), 140.2(C<sub>arom</sub>), 142.3(C<sub>14</sub>), 151.4(C<sub>11</sub>), 169.6(C<sub>8</sub>).

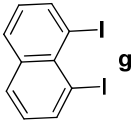
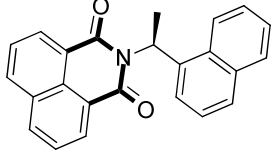
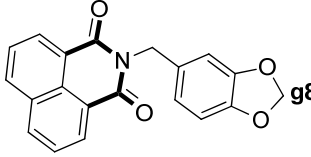
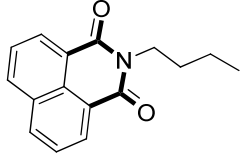
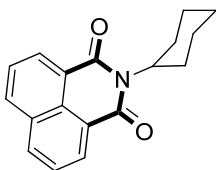
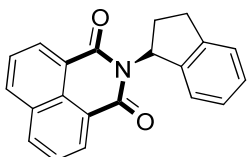
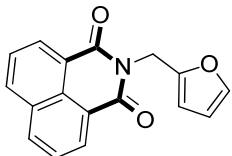
**$\text{M/S (EI)}$**  :  $t_r$  = 14.94 min, calc. for C<sub>13</sub>H<sub>12</sub>INO<sub>2</sub>: 341.19. Found : ( $m/z$ ) = 341.20 (M<sup>+</sup>, 100).

**Table S1.** Pd-catalysed three-component carbonylative cyclisations for the synthesis of N-substituted isoindol-1,3-diones (**a1-a28**) and isoquinolin-1,3-diones (**g derivatives**).<sup>[a]</sup>

Entry	Substrat	Amine	Product	Yield (%) <sup>[b]</sup>
1	 <b>a</b>	 <b>1</b>	 <b>a1</b>	98
2	<b>a</b>	 <b>2</b>	 <b>a2</b>	98
3	<b>a</b>	 <b>3</b>	 <b>a3</b>	95
4	<b>a</b>	 <b>4</b>	 <b>a4</b>	90
5	<b>a</b>	 <b>5</b>	 <b>a5</b>	90
6	<b>a</b>	 <b>6</b>	 <b>a6</b>	90
7	<b>a</b>	 <b>7</b>	 <b>a7</b>	96
8	<b>a</b>	 <b>8</b>	 <b>a8</b>	95
9	<b>a</b>	 <b>9</b>	 <b>a9</b>	98
10	<b>a</b>	 <b>10</b>	 <b>a10</b>	96

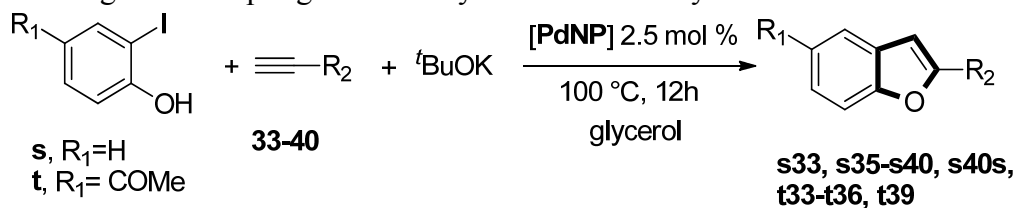
11	a		11		a11	92
12	a		12		a12	98
13	a		13		a13	96
14	a		14		a14	95
15	a		15		a15	92
16	a		16		a16	90
17	a		17		a17	90
18	a		18		a18	91
19	a		19		a19	96
20	a		20		a20	91

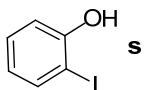
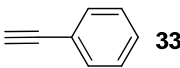
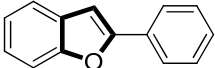
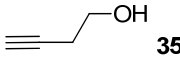
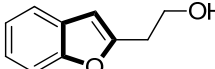
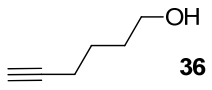
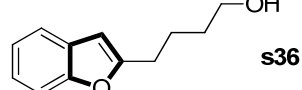
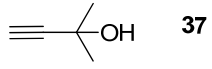
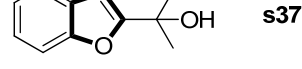
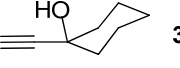
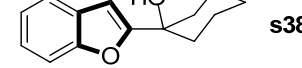
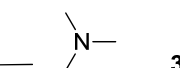
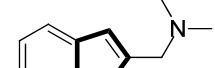
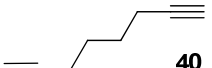
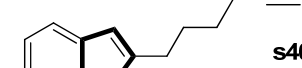
21	a		21		a21	94
22	a		22		a22	91
23	a		23		a23	95
24	a		24		a24	91
25	a		25		a25	92
26	a		26		a26	92
27	a		27		a27	96
28	a		28		a28	90

29		<b>g</b>	6		<b>g6</b>	93
30	<b>g</b>	<b>g</b>	(8)		<b>g8</b>	95
31	<b>g</b>	<b>g</b>	(9)		<b>g9</b>	98
32	<b>g</b>	<b>g</b>	12		<b>g12</b>	91
33	<b>g</b>	<b>g</b>	16		<b>g16</b>	91
34	<b>g</b>	<b>g</b>	24		<b>g24</b>	97

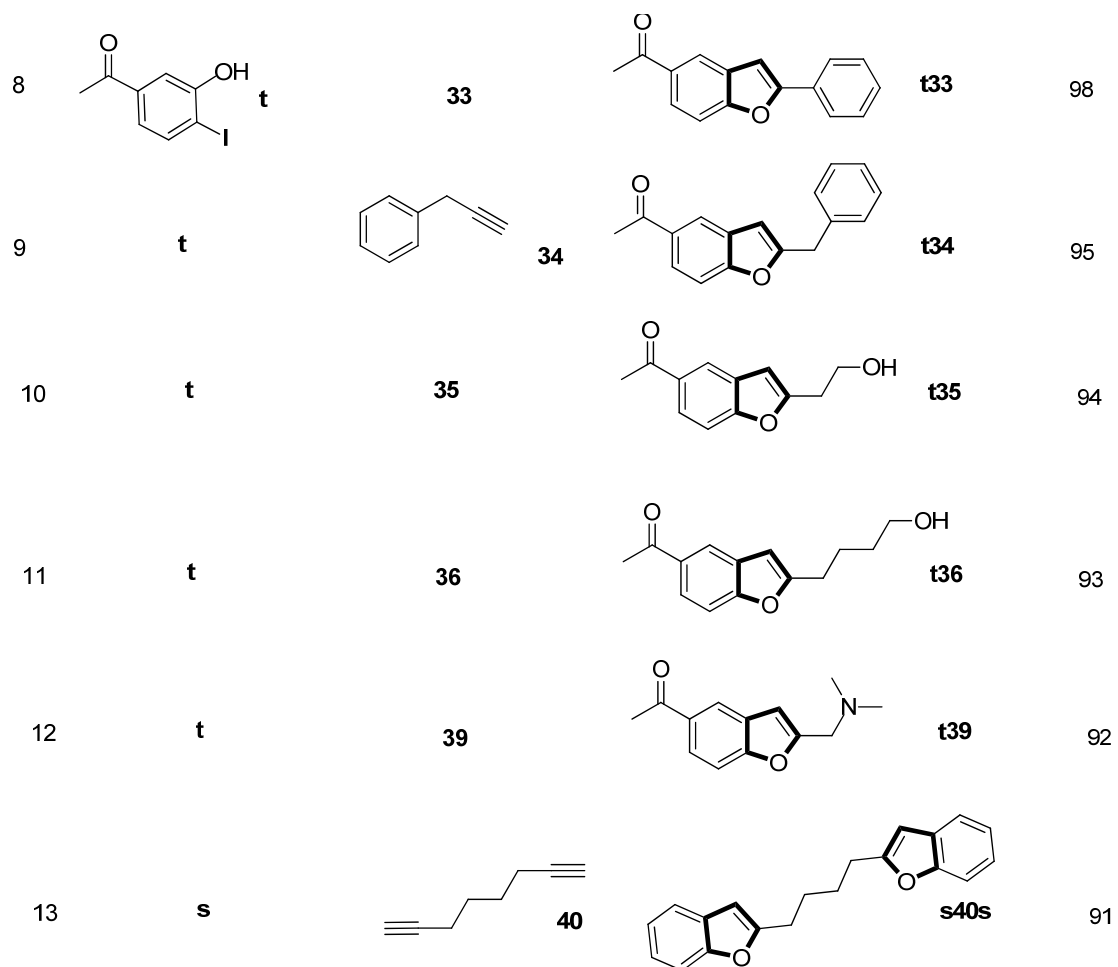
<sup>[a]</sup> Results from duplicate experiments. Reaction conditions: 0.4 mmol of 2-iodobenzoic acid or 1,8-diiodonaphthalene, 0.4 mmol of amine in glycerol (1 mL, 13.6 mmol) at 120 °C, 0.5 bar CO for 0.5h, using 2.5 mol% **PdNP**. <sup>[b]</sup> Isolated yields after column chromatography; compounds identified by <sup>1</sup>H and <sup>13</sup>C NMR and GC-MS.

**Table S2.** Synthesis of 2-substituted benzofurans (s33, s35-s40, t33-t36 and t39-t40) by Pd-catalysed Sonogashira coupling followed by intramolecular cyclisation.<sup>[a]</sup>



Entry	Substrat	$\equiv\text{-R}_2$	Product	Yield (%) <sup>[b]</sup>
1	 <b>s</b>	 <b>33</b>	 <b>s33</b>	91
2	<b>s</b>	 <b>35</b>	 <b>s35</b>	93
3	<b>s</b>	 <b>36</b>	 <b>s36</b>	98
4	<b>s</b>	 <b>37</b>	 <b>s37</b>	91
5	<b>s</b>	 <b>38</b>	 <b>s38</b>	97
6	<b>s</b>	 <b>39</b>	 <b>s39</b>	97
7	<b>s</b>	 <b>40</b>	 <b>s40</b>	96



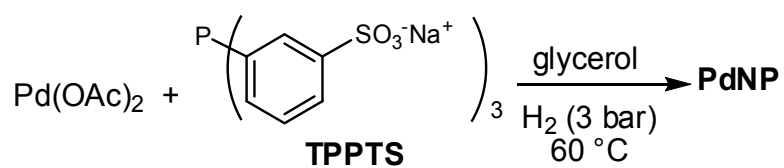


<sup>[a]</sup> Results from duplicate experiments. Reaction conditions: 0.4 mmol of iodophenol or 4'-hydroxy-3'-iodoacetophenone and 0.6 mmol of alkyne in glycerol (1 mL, 13.6 mmol) at 100 °C for 12 h, using 2.5 mol% **PdNP**.

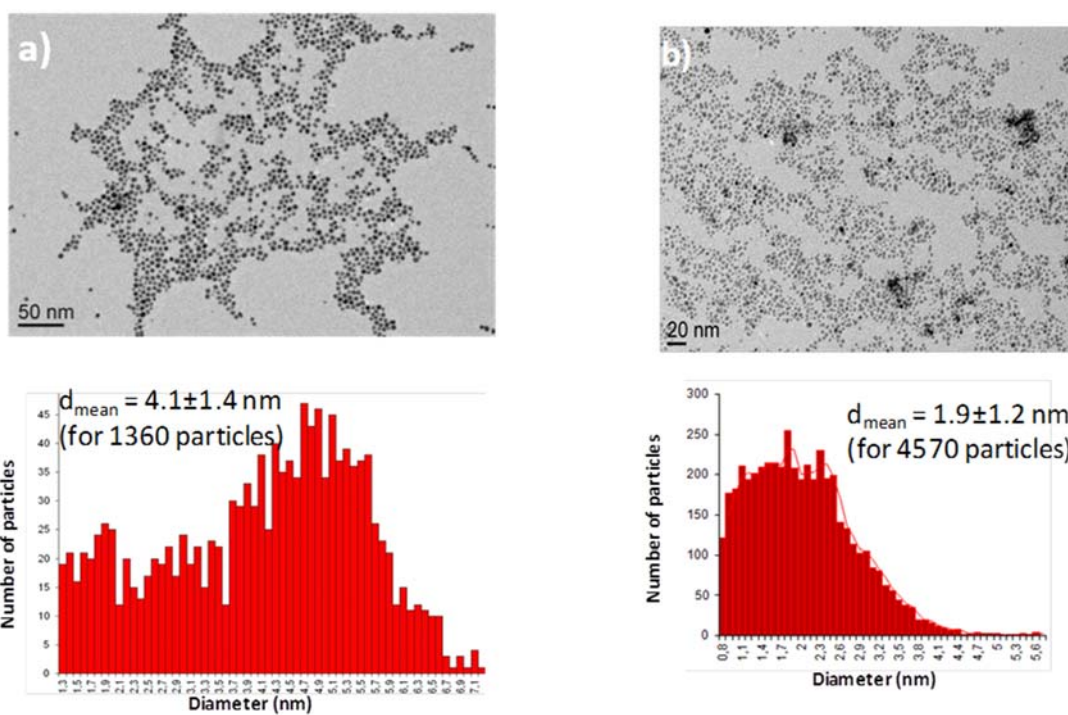
<sup>[b]</sup> Isolated yields after column chromatography; compounds identified by <sup>1</sup>H and <sup>13</sup>C NMR and GC-MS.

**Table S3.** Crystallographic data for **a1**, **a30**, **s40u**, **r33**, **n35**, **a17u** and **g8**.

	<b>a1</b>	<b>a30</b>	<b>s40u</b>	<b>r33</b>	<b>n35</b>	<b>a17u</b>	<b>g8</b>
chemical formula	C <sub>15</sub> H <sub>11</sub> NO <sub>2</sub>	C <sub>22</sub> H <sub>20</sub> N <sub>2</sub> O <sub>4</sub>	C <sub>21</sub> H <sub>21</sub> N <sub>3</sub> O	C <sub>23</sub> H <sub>16</sub> O	C <sub>22</sub> H <sub>23</sub> NO <sub>2</sub>	C <sub>20</sub> H <sub>18</sub> N <sub>4</sub> O <sub>2</sub>	C <sub>20</sub> H <sub>13</sub> NO <sub>4</sub>
<i>Mr</i>	237.25	376.4	331.41	308.36	333.41	346.38	331.31
crystal system	triclinic	monoclinic	monoclinic	monoclinic	triclinic	triclinic	triclinic
space group	<i>P</i> $\bar{1}$	<i>P</i> 2 <sub>1</sub> /n	<i>P</i> c	<i>P</i> 2 <sub>1</sub>	<i>P</i> $\bar{1}$	<i>P</i> $\bar{1}$	<i>P</i> $\bar{1}$
<i>a</i> [Å]	7.1432(4)	7.0179(8)	18.2723(16)	11.008(3)	9.1264(5)	5.6257(7)	7.1728(2)
<i>b</i> [Å]	8.4932(4)	4.6365(4)	5.5654(4)	7.5883(19)	9.7169(6)	19.7469(19)	8.0196(2)
<i>c</i> [Å]	10.1874(5)	29.074(3)	8.7266(7)	19.571(5)	10.2731(7)	23.768(2)	12.8455(4)
$\alpha$ [°]	99.388(2)	90	90	90	95.504(4)	79.737(6)	84.142(2)
$\beta$ [°]	97.577(2)	92.757(3)	103.633(4)	90.988(10)	91.157(4)	88.280(7)	89.687(2)
$\gamma$ [°]	106.771(2)	90	90	90	101.221(4)	82.746(7)	87.901(2)
<i>V</i> [Å <sup>3</sup> ]	573.27(5)	944.93(17)	862.43(12)	1634.6(7)	888.78(10)	2577.2(5)	734.56(4)
<i>Z</i>	2	2	2	4	2	6	2
Reflections collected	8247	7457	11512	11659	13707	44315	12101
Independent reflections	2080	1947	3137	5736	3588	8707	2691
	[R(int) = 0.0205]	[R(int) = 0.0402]	[R(int) = 0.0256]	[R(int) = 0.08]	[R(int) = 0.0591]	[R(int) = 0.1375]	[R(int) = 0.0279]
$\rho_{calc}$ [g cm <sup>-3</sup> ]	1.374	1.323	1.276	1.253	1.246	1.339	1.498
$\mu$ (MoK $\alpha$ ) [mm <sup>-1</sup> ]	0.092	0.092	0.080	0.075	0.079	0.090	0.106
crystal size (mm <sup>3</sup> )	0.10 x 0.08 x 0.08	0.18 x 0.16 x 0.04	0.24 x 0.24 x 0.20	0.16x 0.08 x 0.02	0.20 x 0.12 x 0.08	0.40 x 0.08 x 0.03	0.12 x 0.10 x 0.06
GOF on F <sup>2</sup>	1.048	1.048	1.039	0.972	1.079	1.075	1.028
<i>R</i> ( <i>I</i> > 2 $\sigma$ ( <i>I</i> ))	0.0340	0.0445	0.0412	0.0704	0.0587	0.0759	0.0379
wR <sup>2</sup> (all data)	0.0899	0.1093	0.1114	0.1720	0.2170	0.1814	0.1044
Largest difference peak and hole [e Å <sup>-3</sup> ]	0.230 and -0.190	0.159 and -0.170	0.312 and -0.227	0.243 and -0.197	0.326 and -0.319	0.219 and -0.271	0.161 and -0.253



**Scheme S1.** Synthesis of PdNPs in neat glycerol.



**Figure S1** TEM images corresponding to the PdNP in neat glycerol (a) and after the fifth recycling run (b).

## **SPECTRA OF SYNTHESISED HETEROCYCLES**

**following the order of appearance in the main text**

501

fc-501-2 Sm (Mn, 1x3)

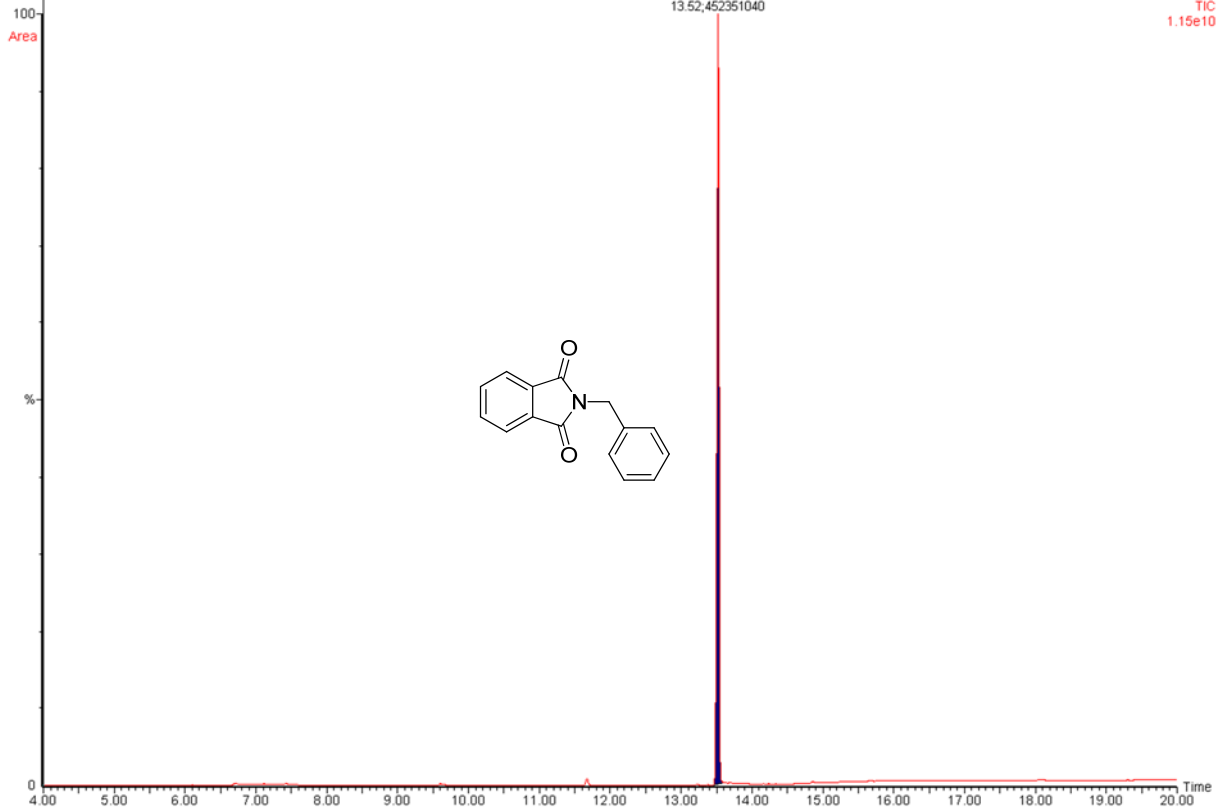
, 17-Jan-2012 + 14:55:56

Scan E1+

TIC

1.15e10

Area



501

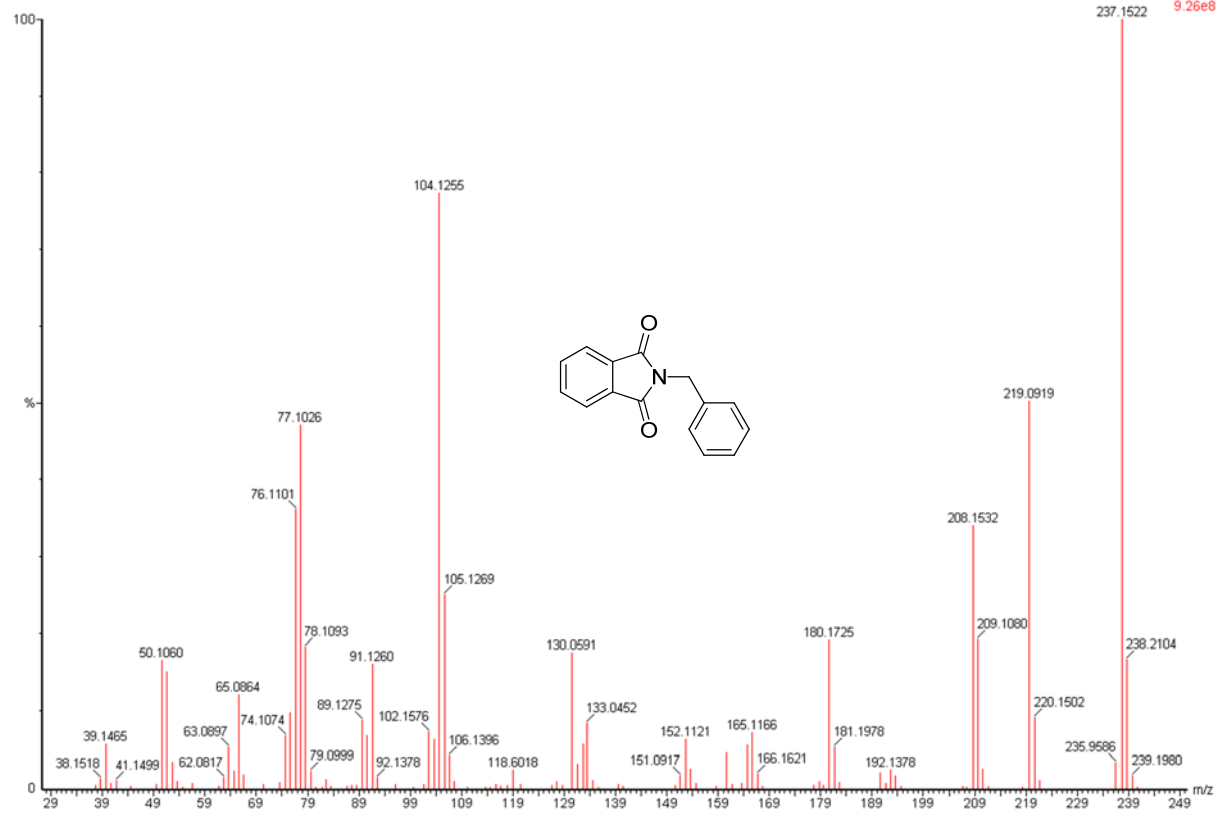
fc-501-2 1906 (13.532) Cm (1896:1909)

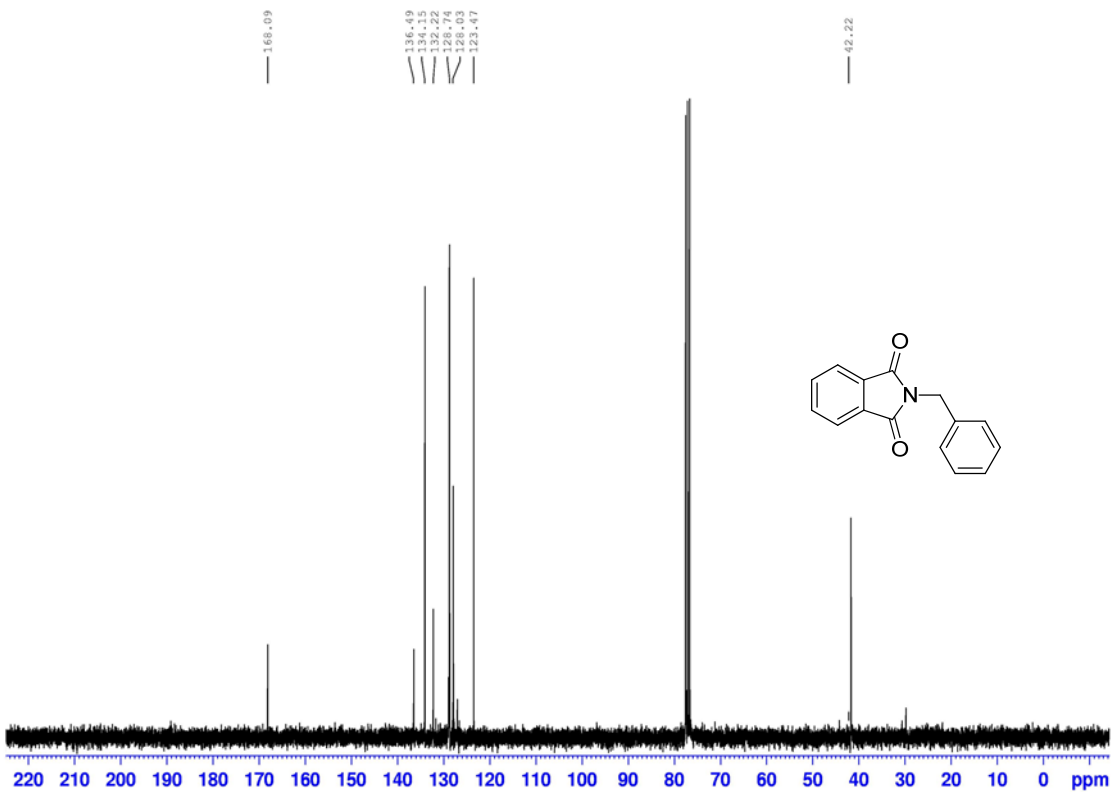
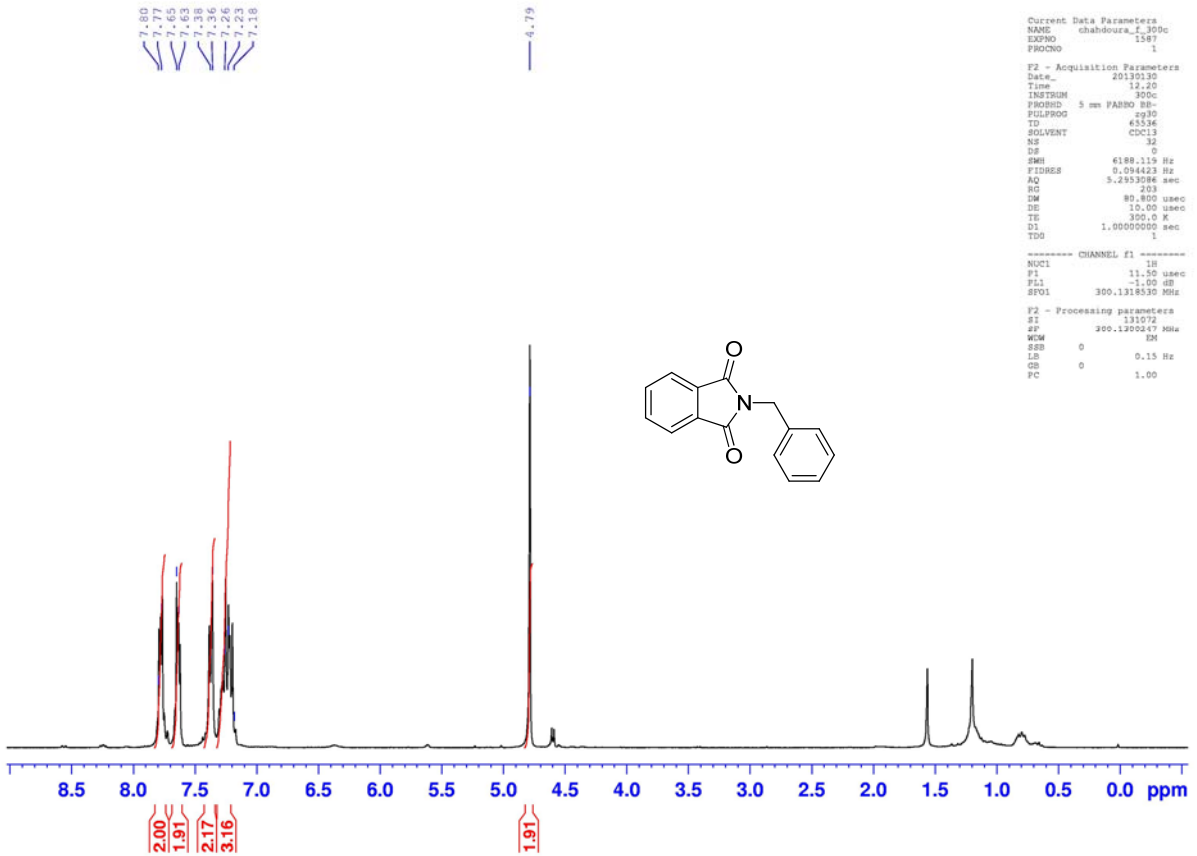
, 17-Jan-2012 + 14:55:56

Scan E1+

9.26e8

%



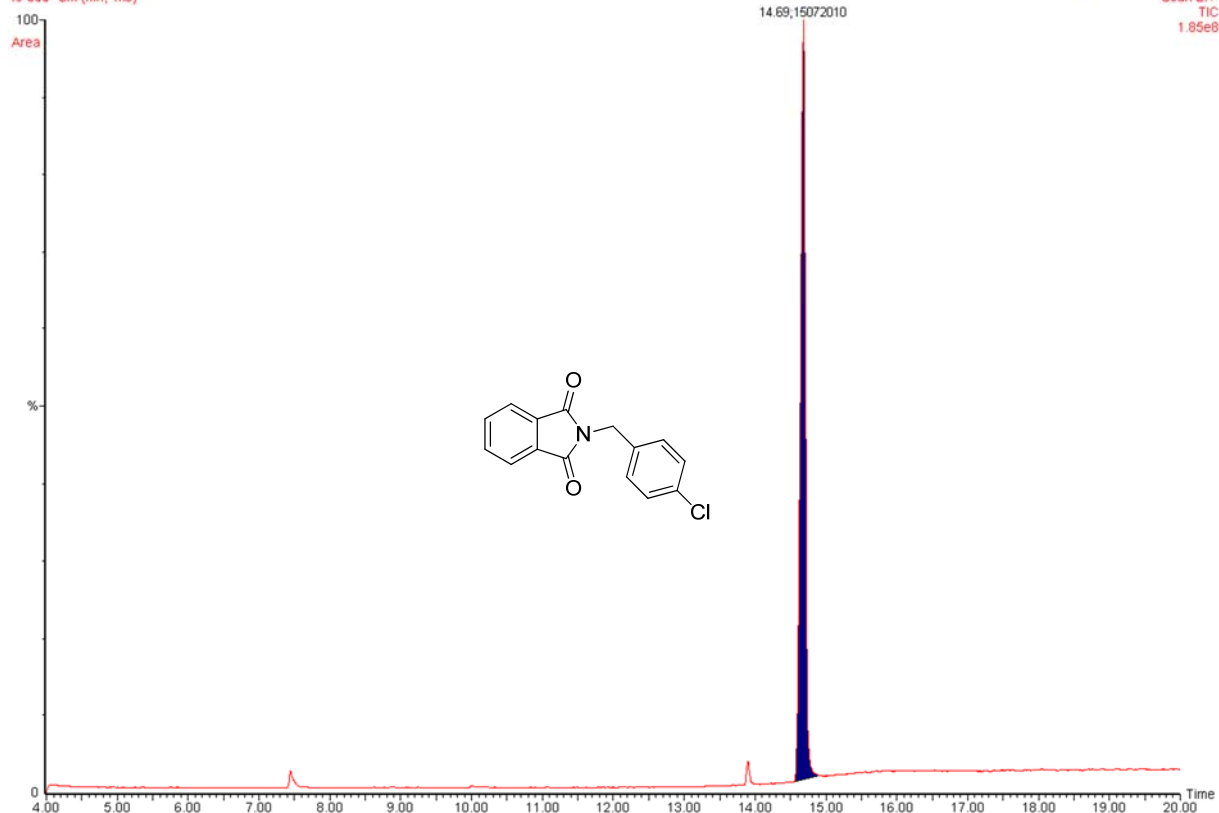


GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **a1**

855  
tc-855- Sm (Mn, 1x3)

, 18-Sep-2012 + 10:09:14

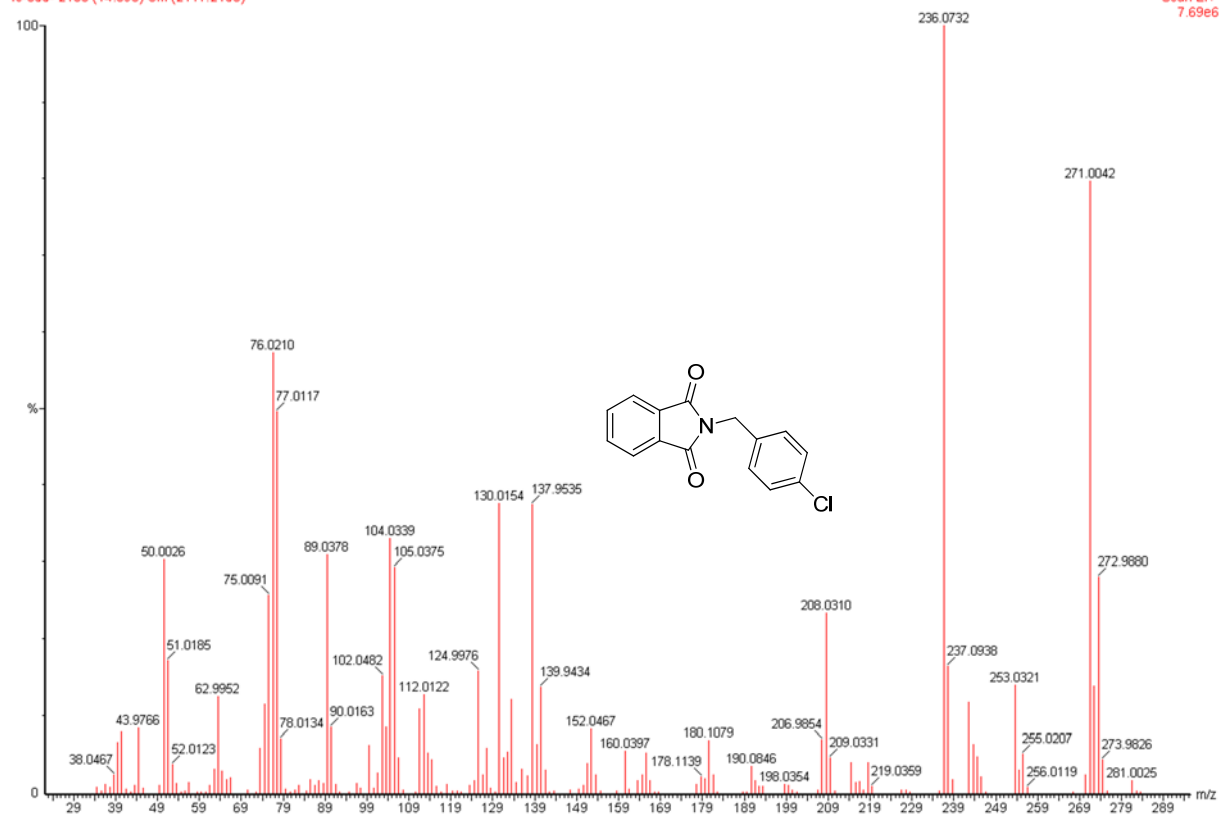
Scan EI+  
TIC  
1.85e8

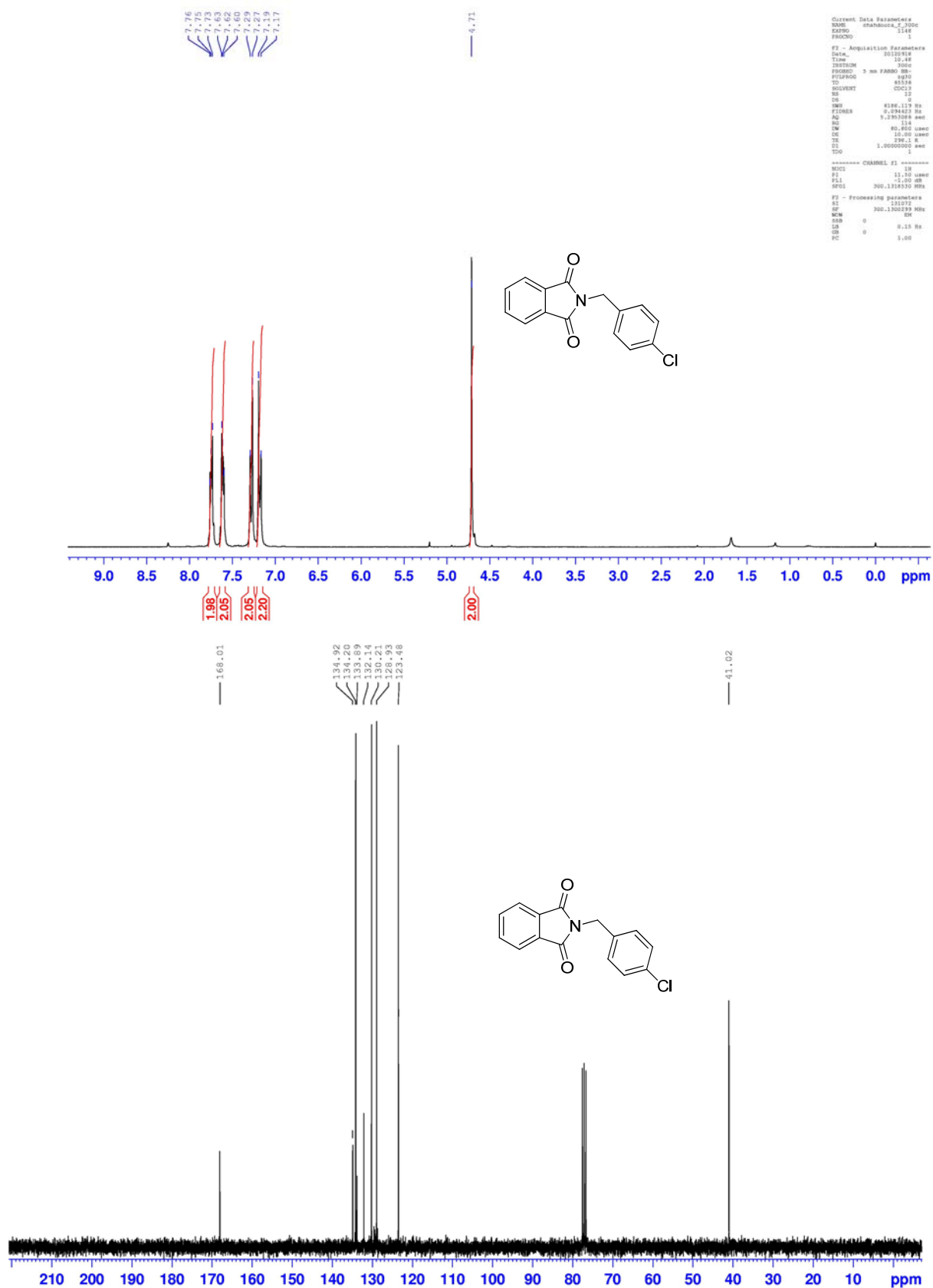


855  
tc-855- 2138 (14.693) Cm (2117:2156)

, 18-Sep-2012 + 10:09:14

Scan EI+  
7.69e6

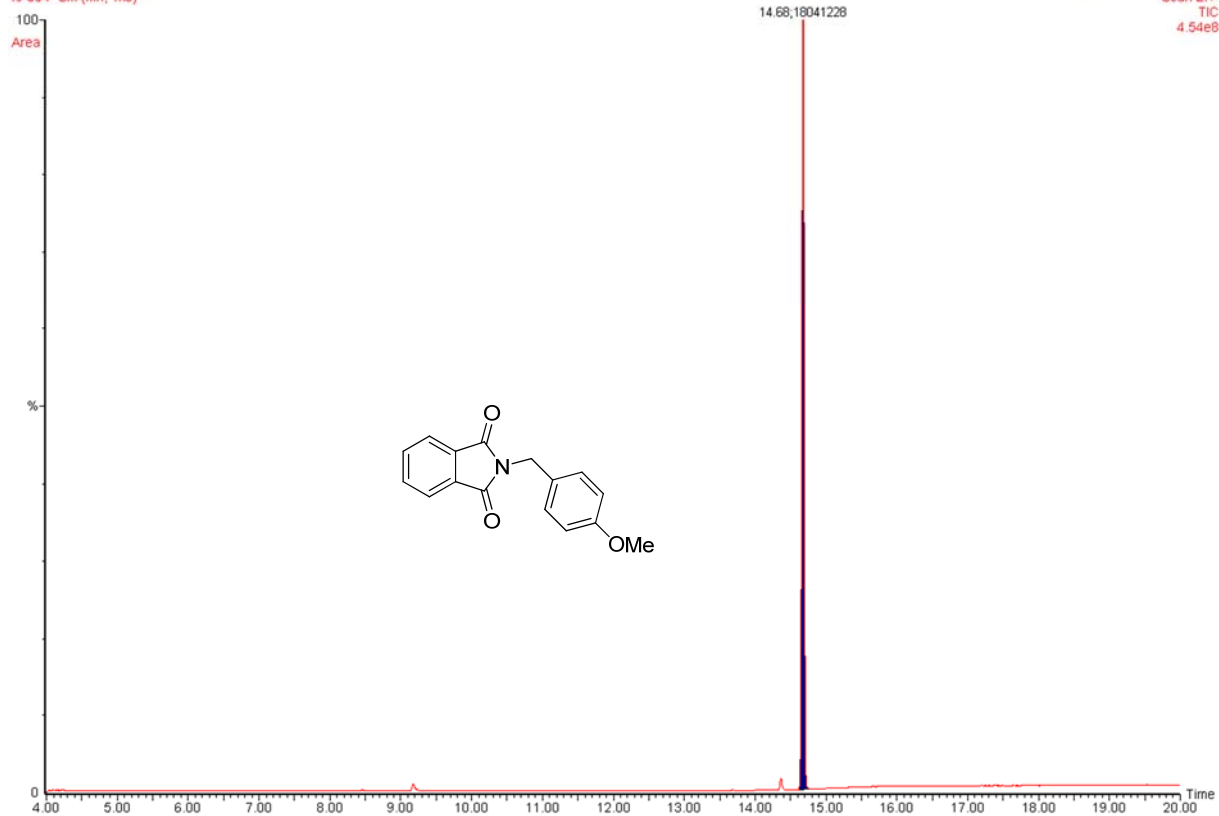






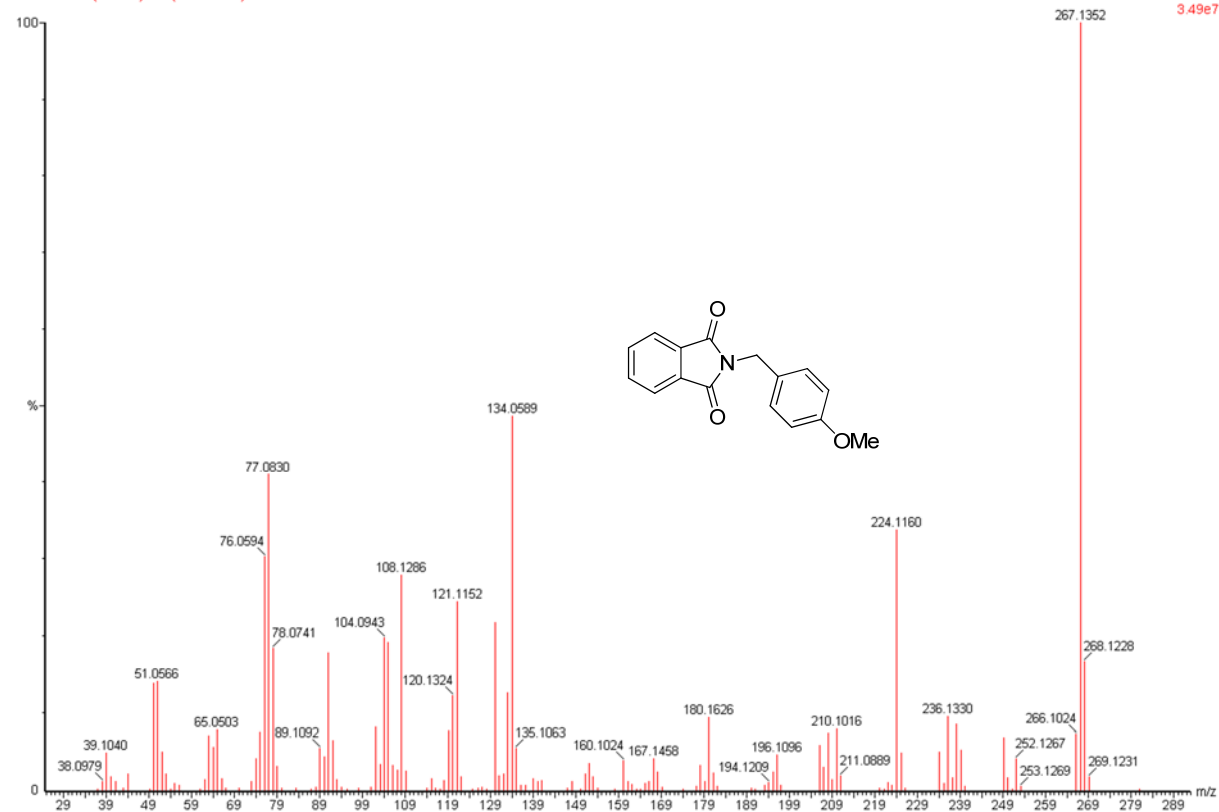
854  
fc-854- Sm (Mn, 1x3)

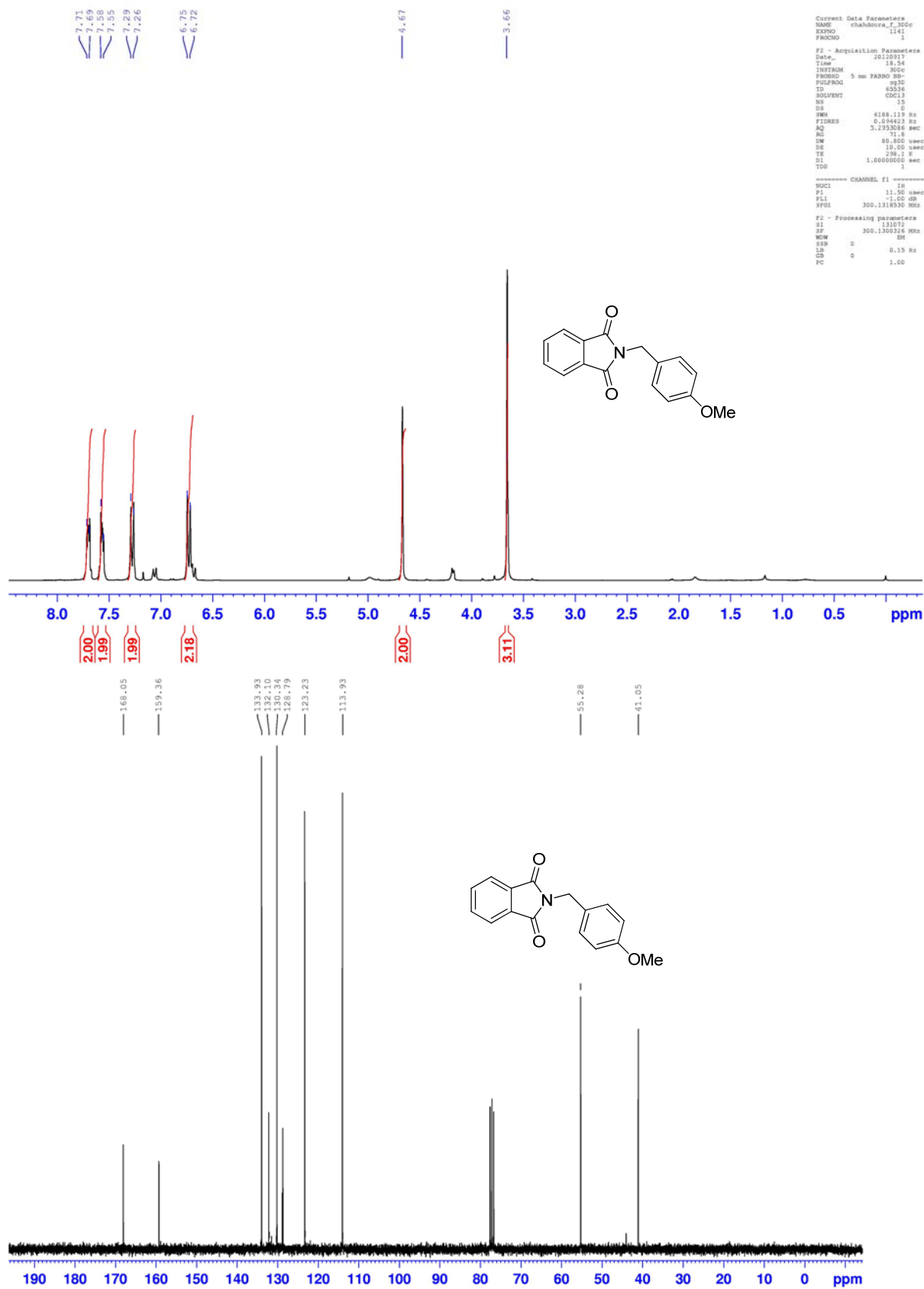
, 17-Sep-2012 + 18:33:41  
Scan EI+  
TIC  
4.54e8



854  
fc-854- 2137 (14.688) Cm (2128:2141)

, 17-Sep-2012 + 18:33:41  
Scan EI+  
3.49e7

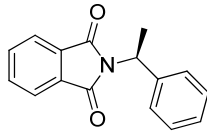
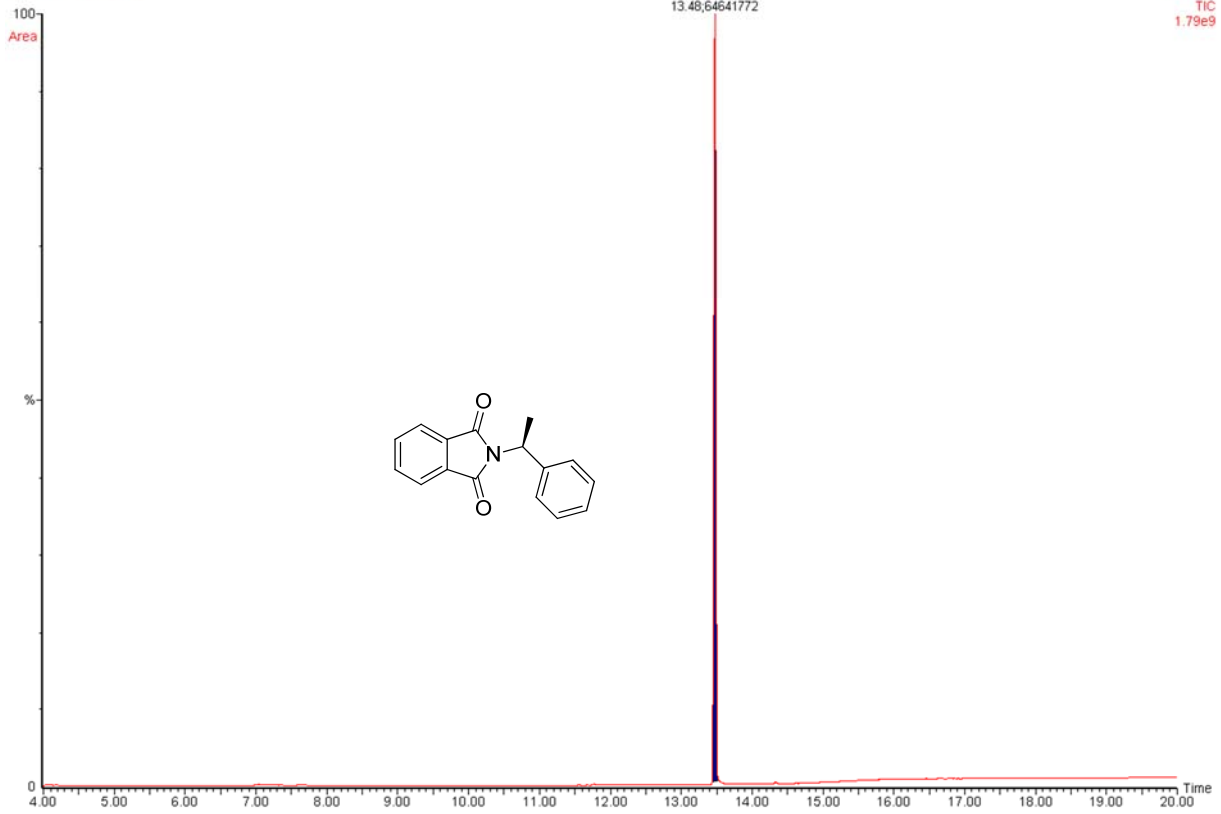




GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **a3**

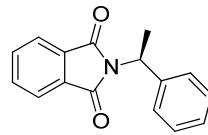
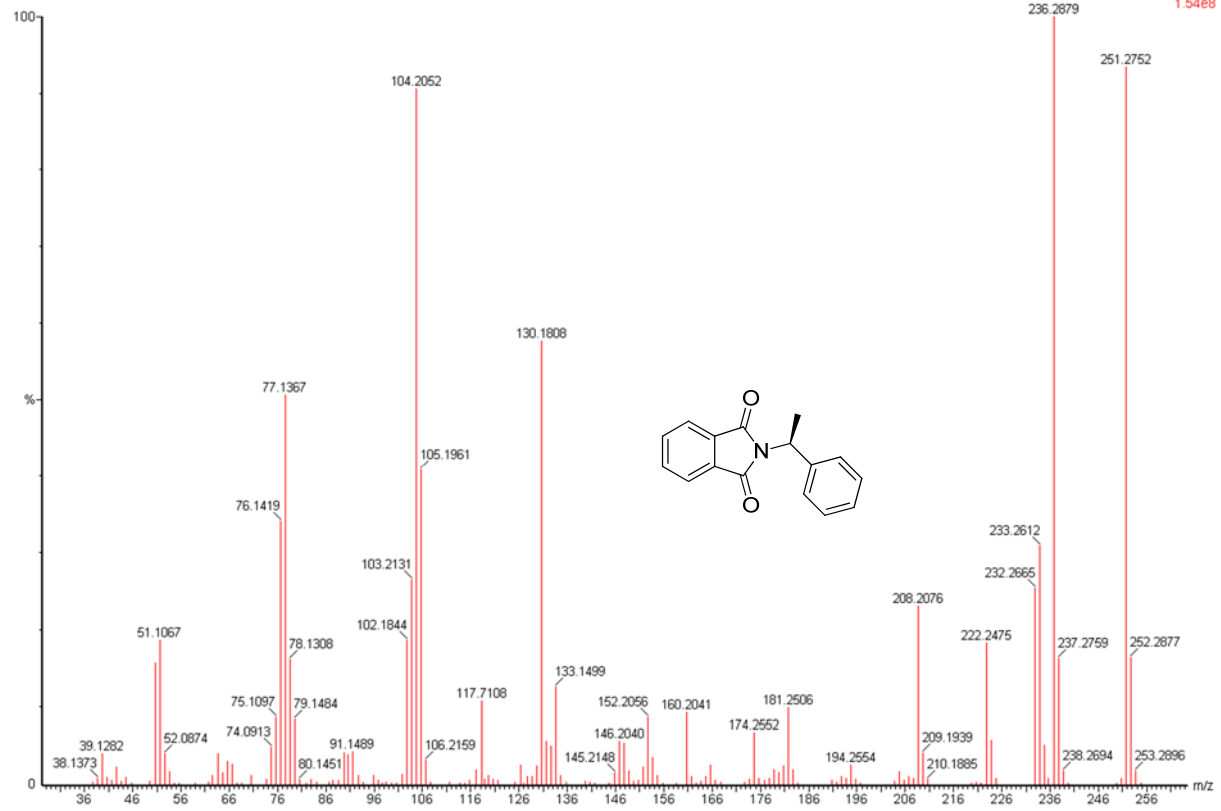
784  
fc-784-2 Sm (Mn, 1x3)

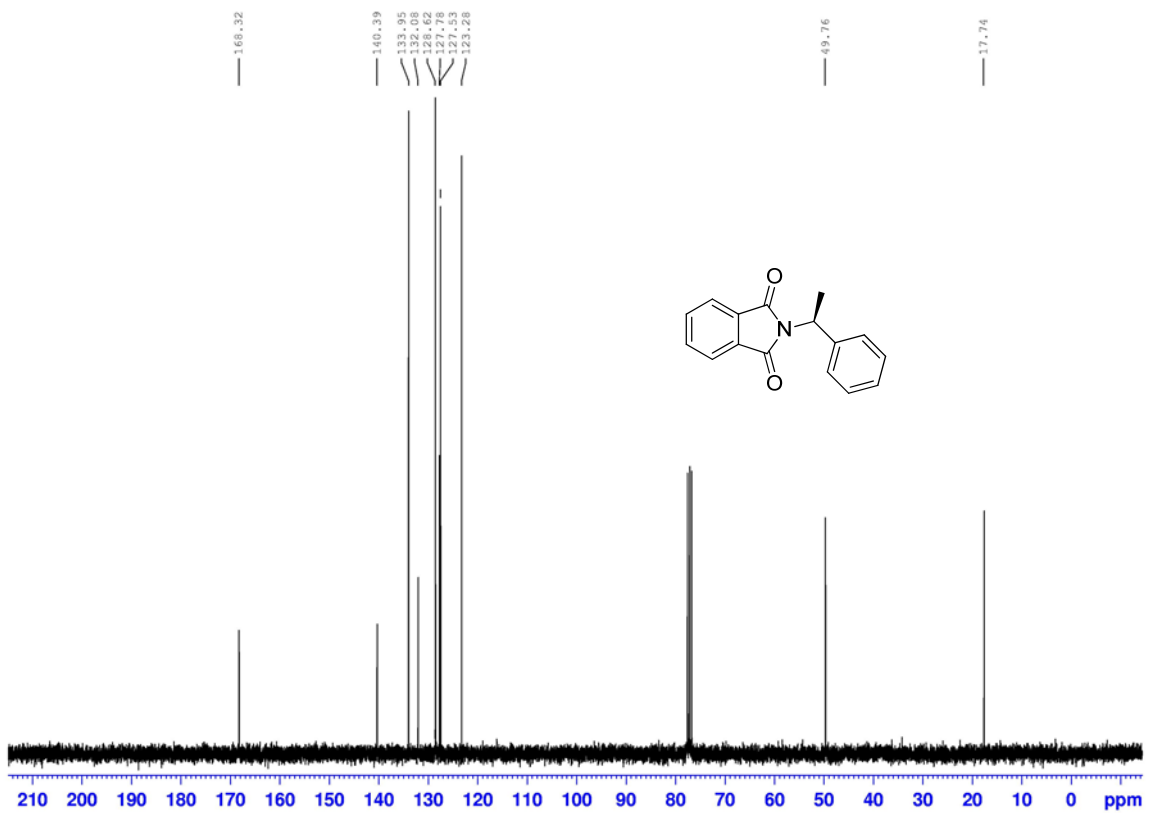
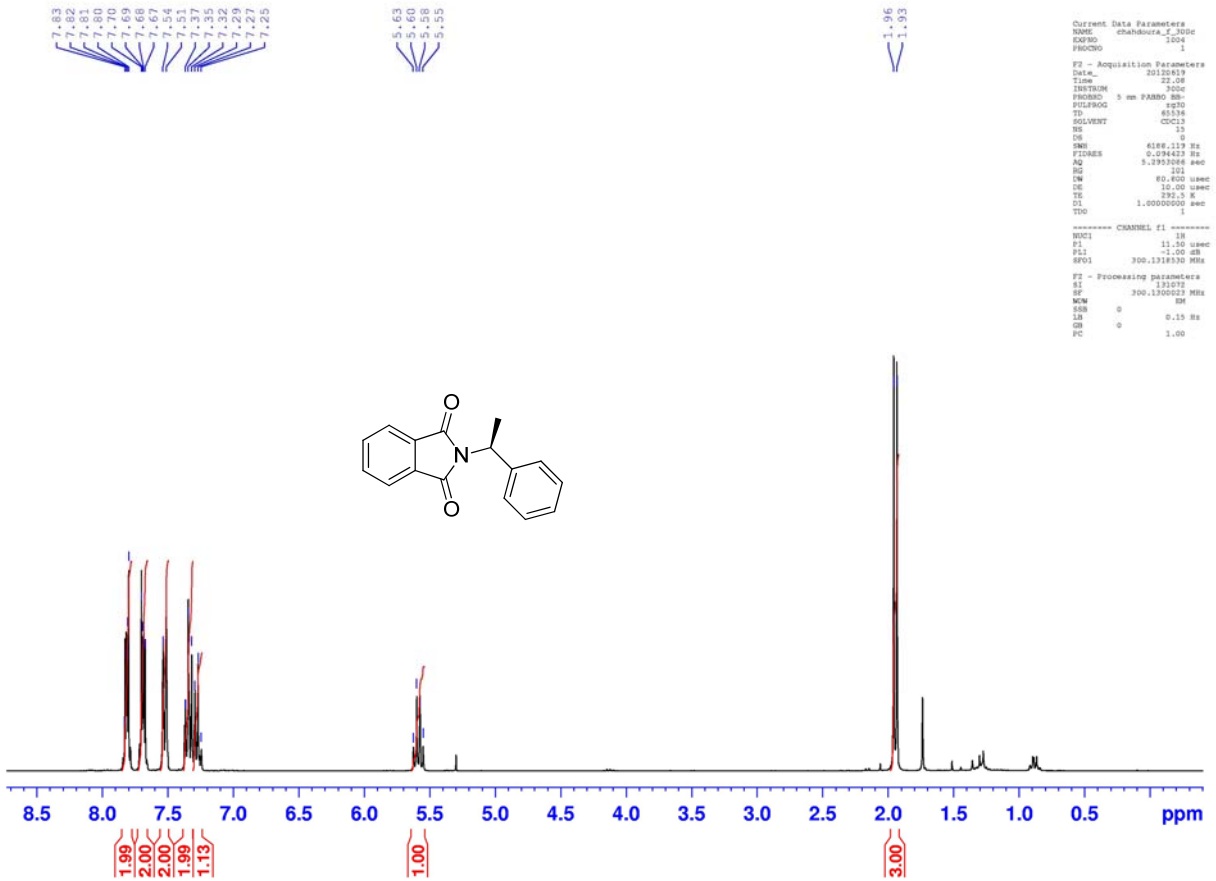
, 19-Jun-2012 + 18:30:06  
Scan EI+  
TIC  
1.79e9



784  
fc-784-2 1896 (13.482) Cm (1890:1898)

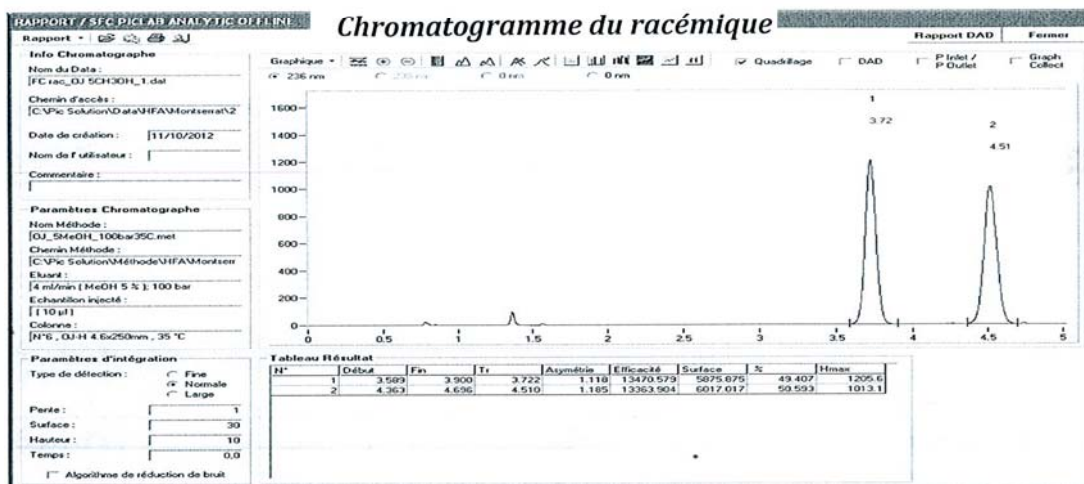
, 19-Jun-2012 + 18:30:06  
Scan EI+  
1.54e8



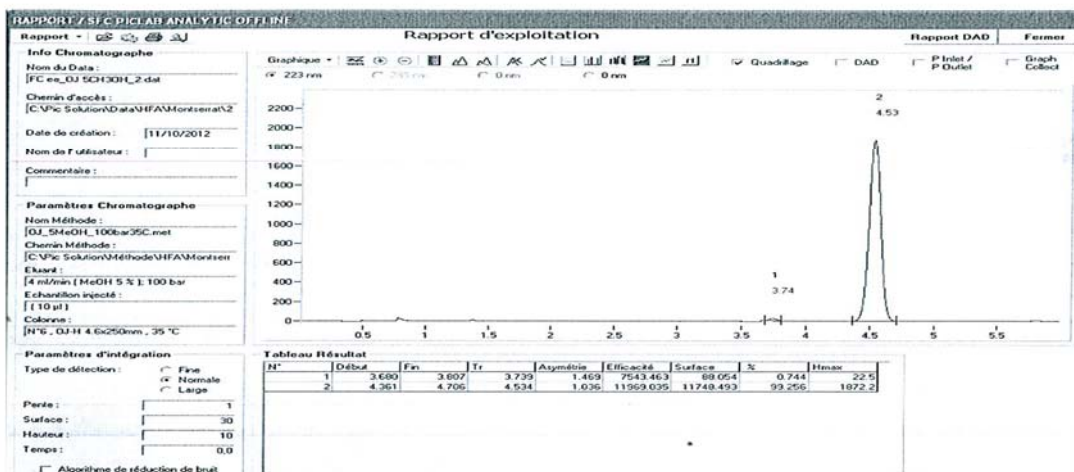


### SFC analyses:

Colonne	Chiralpak OJ-H 5 $\mu$ m (4.6 x 250) mm
Débit total CO <sub>2</sub> + co-solvant (mL/min)	4
Co-solvant	CH <sub>3</sub> OH
% Co-solvant	5
Température (°C)	35
Pin (bar)	138
Pout (bar)	100
$\lambda$ (nm)	236 ( $\lambda_{max}$ = 223, 285)
$t_R$ (min)	3.72 et 4.51
Durée du run (min)	6



### Chromatogramme de l'excès énantiomérique



GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) spectra in CDCl<sub>3</sub>, and SFC analysis (bottom) for **a4**

866

tc-866 Sm (Mn, 1x3)

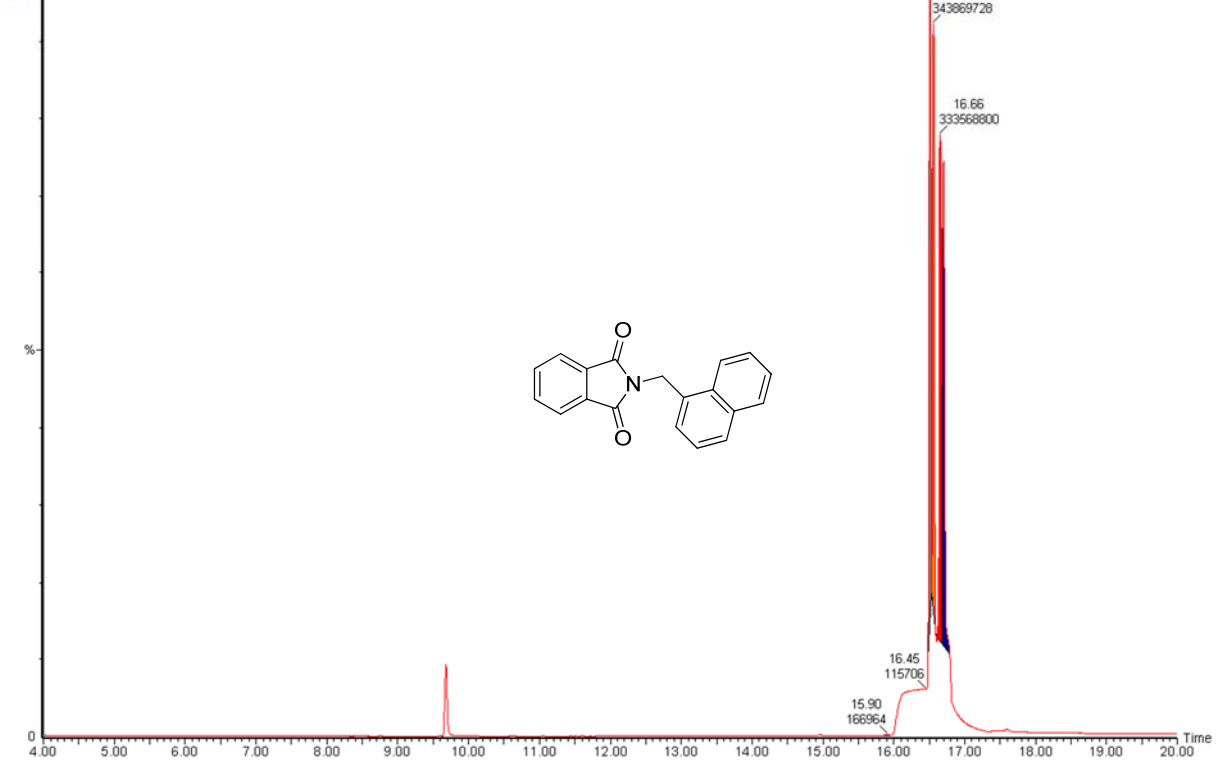
, 05-Nov-2012 + 22:26:48

Scan EI+

TIC

1.31e10

Area



866

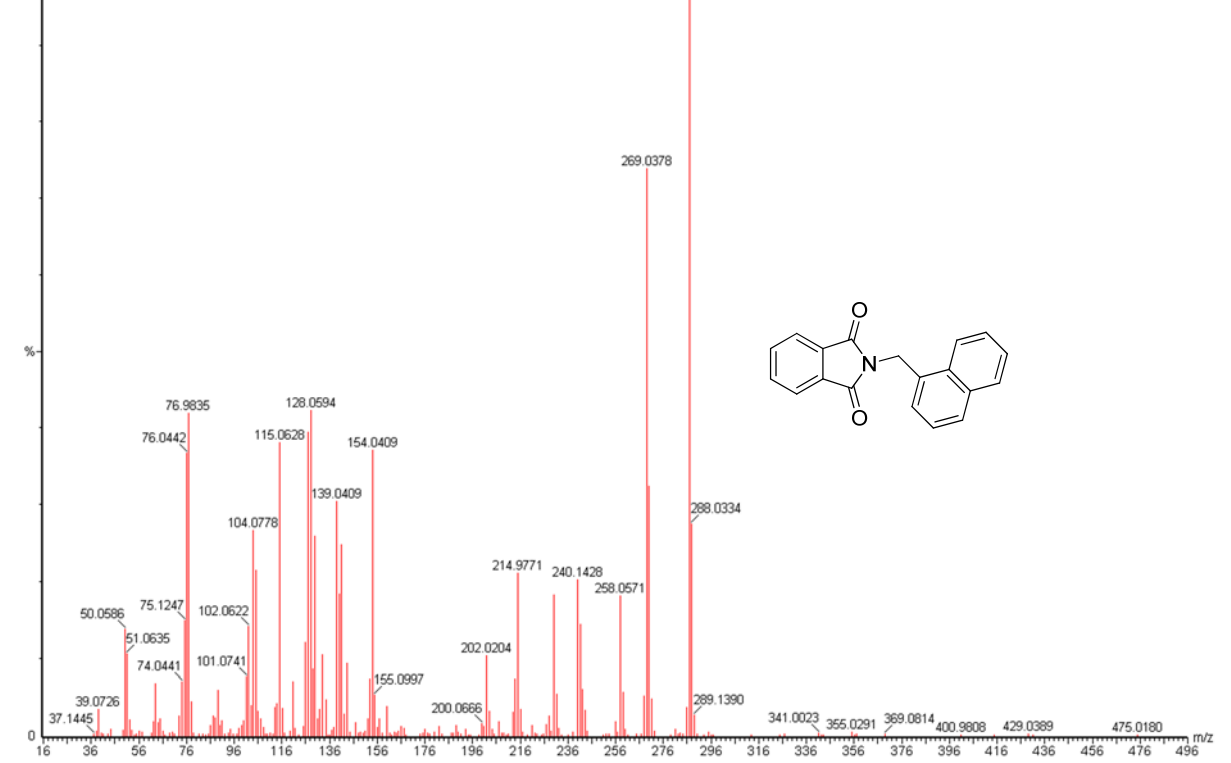
tc-866- 2503 (16.518) Cm (2499:2514)

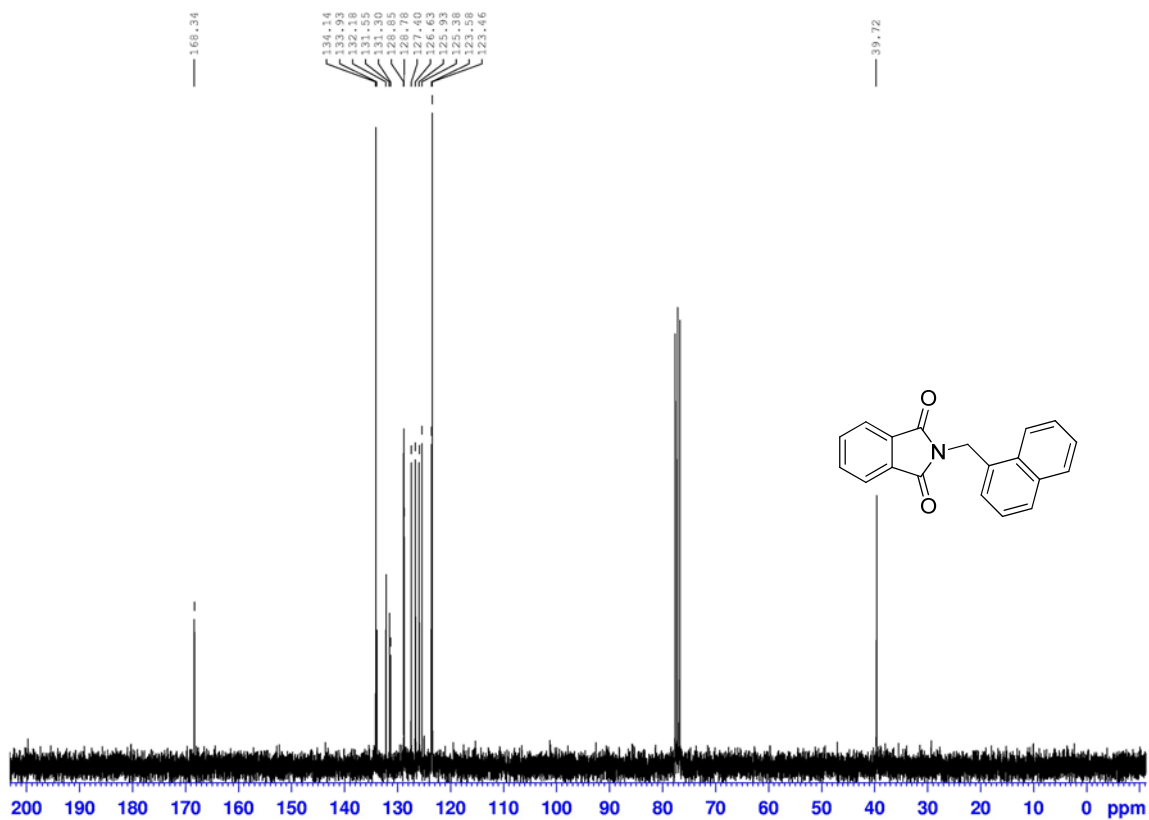
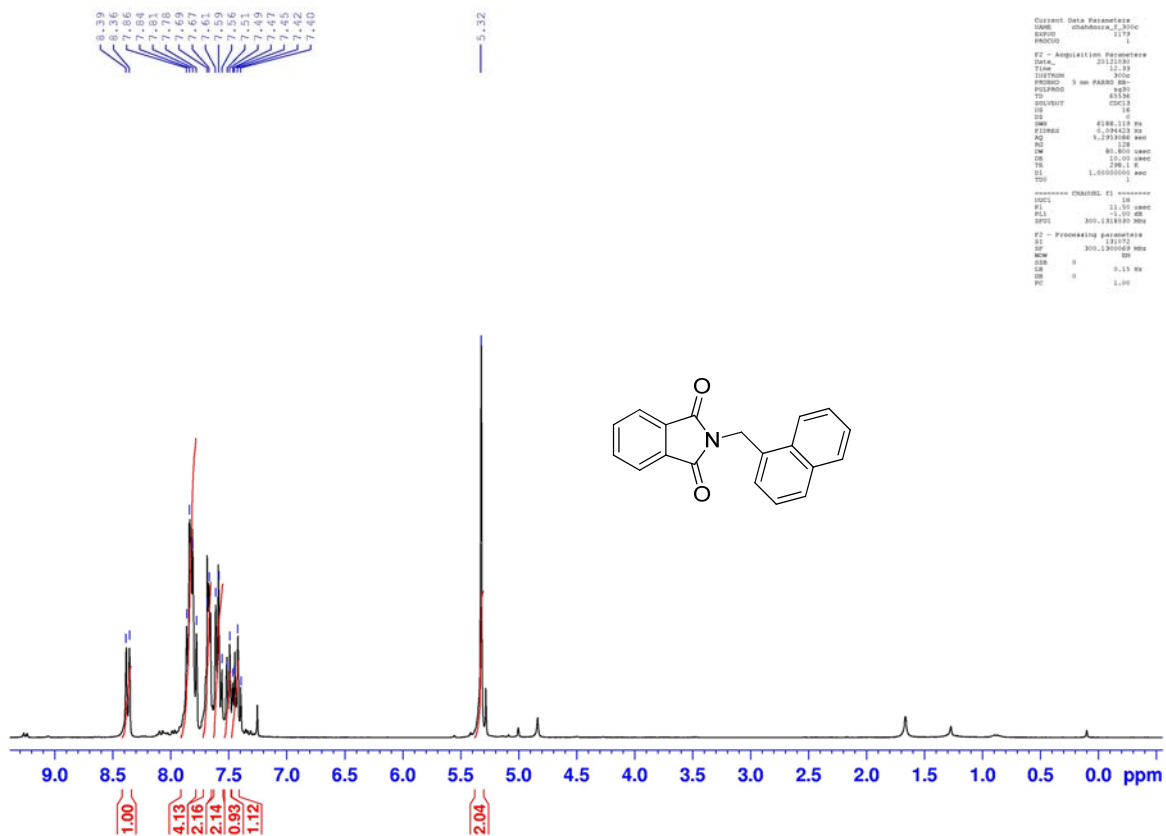
, 05-Nov-2012 + 22:26:48

Scan EI+

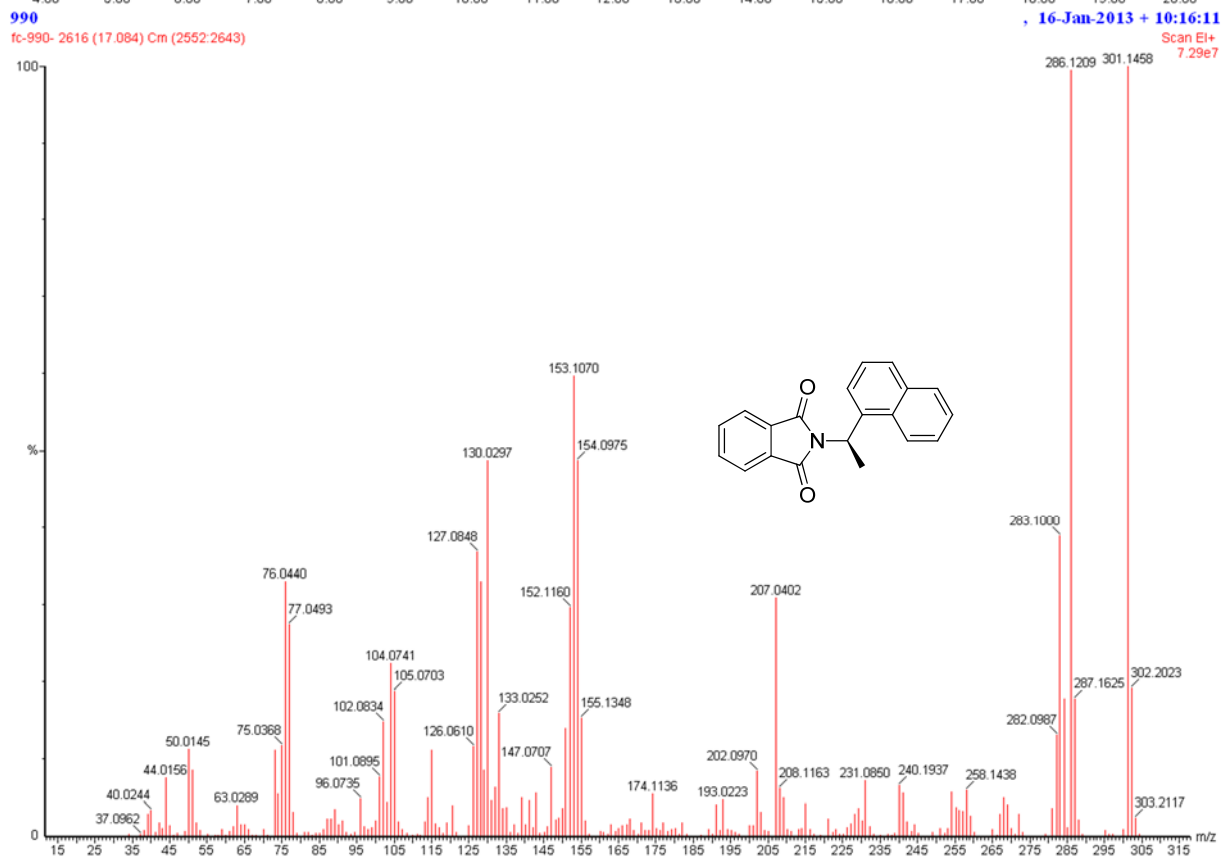
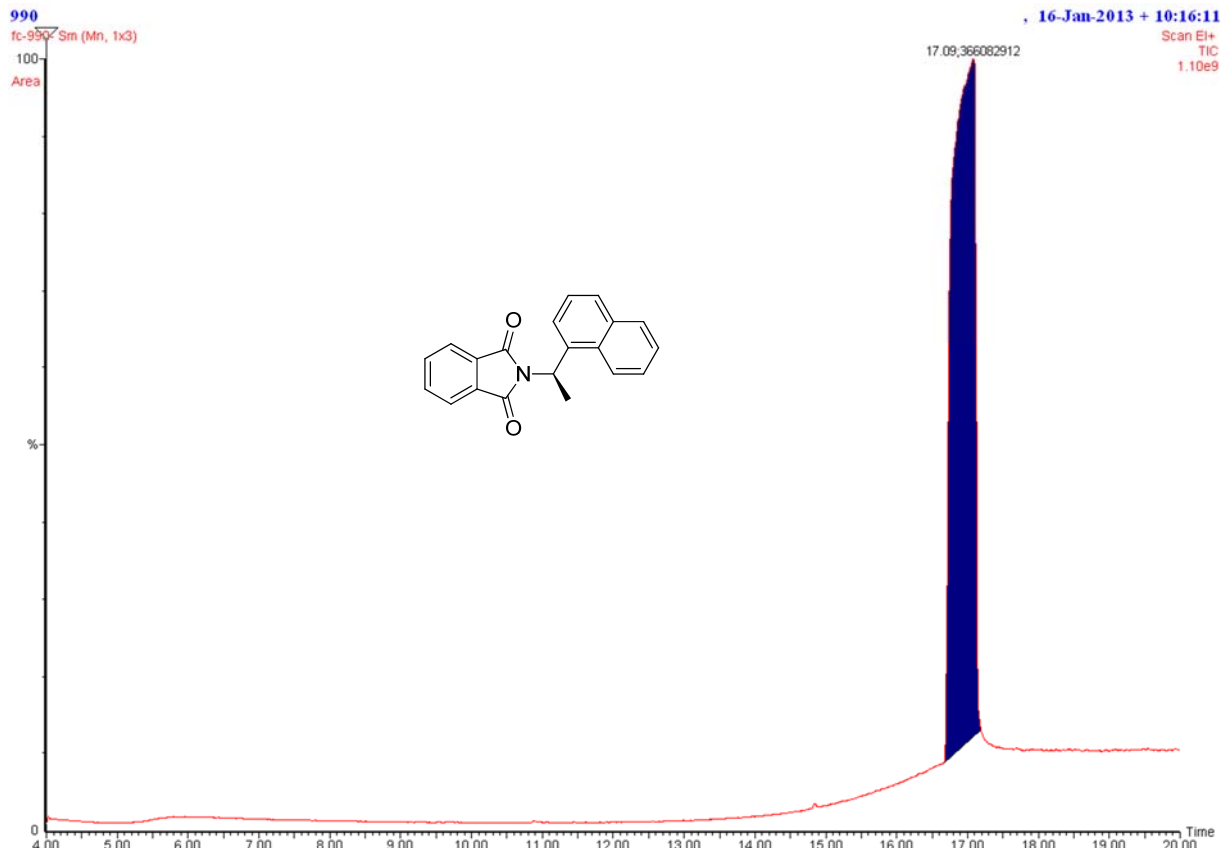
1.06e9

%

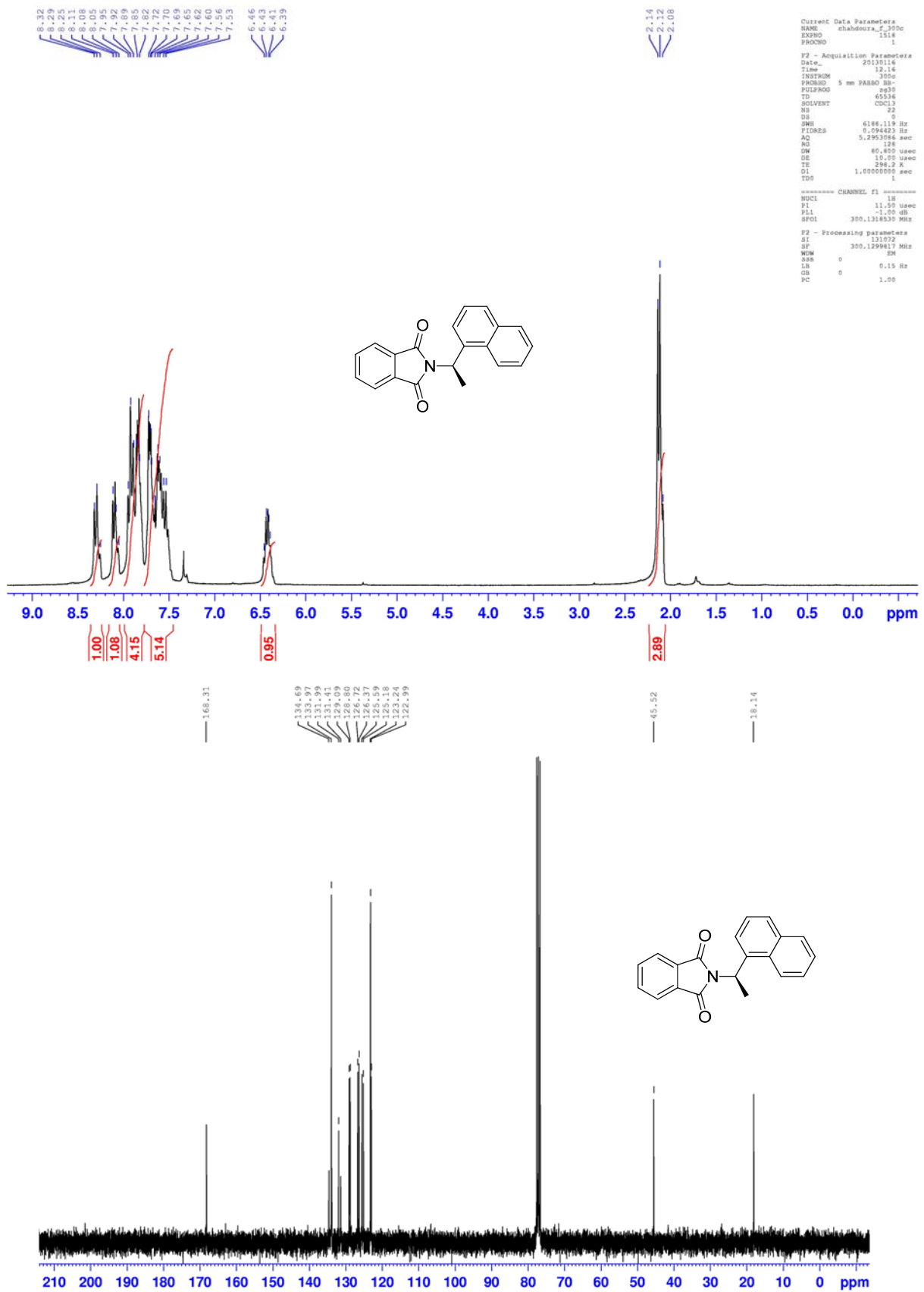




GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **a5**



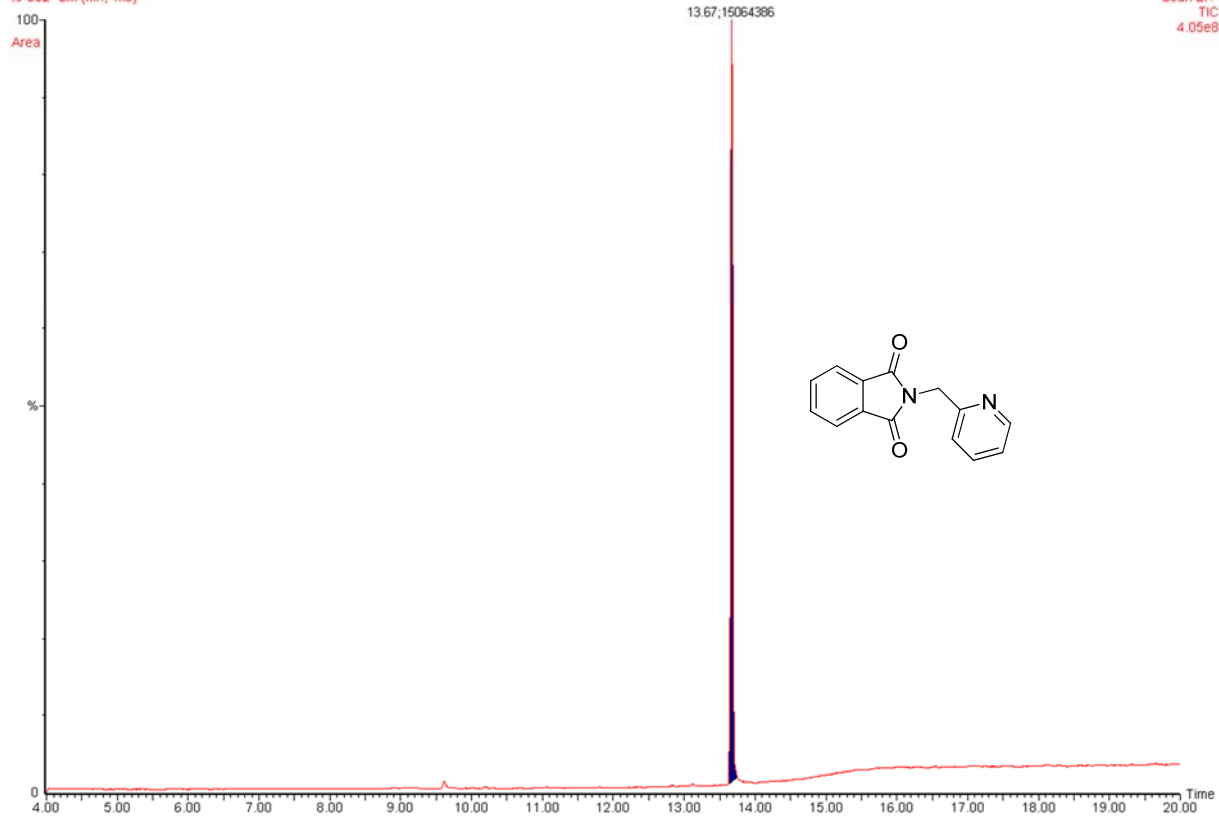




GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **a6**

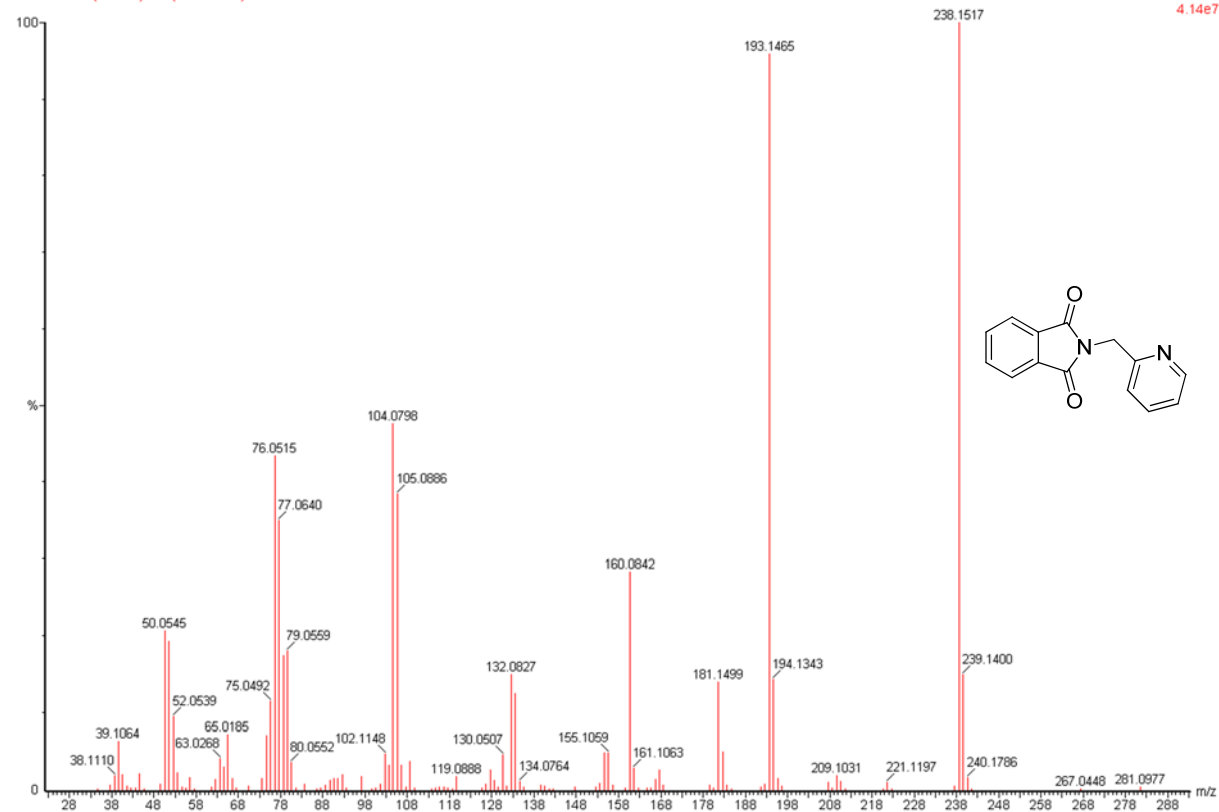
852  
fc-852- Sm (Mn, 1x3)

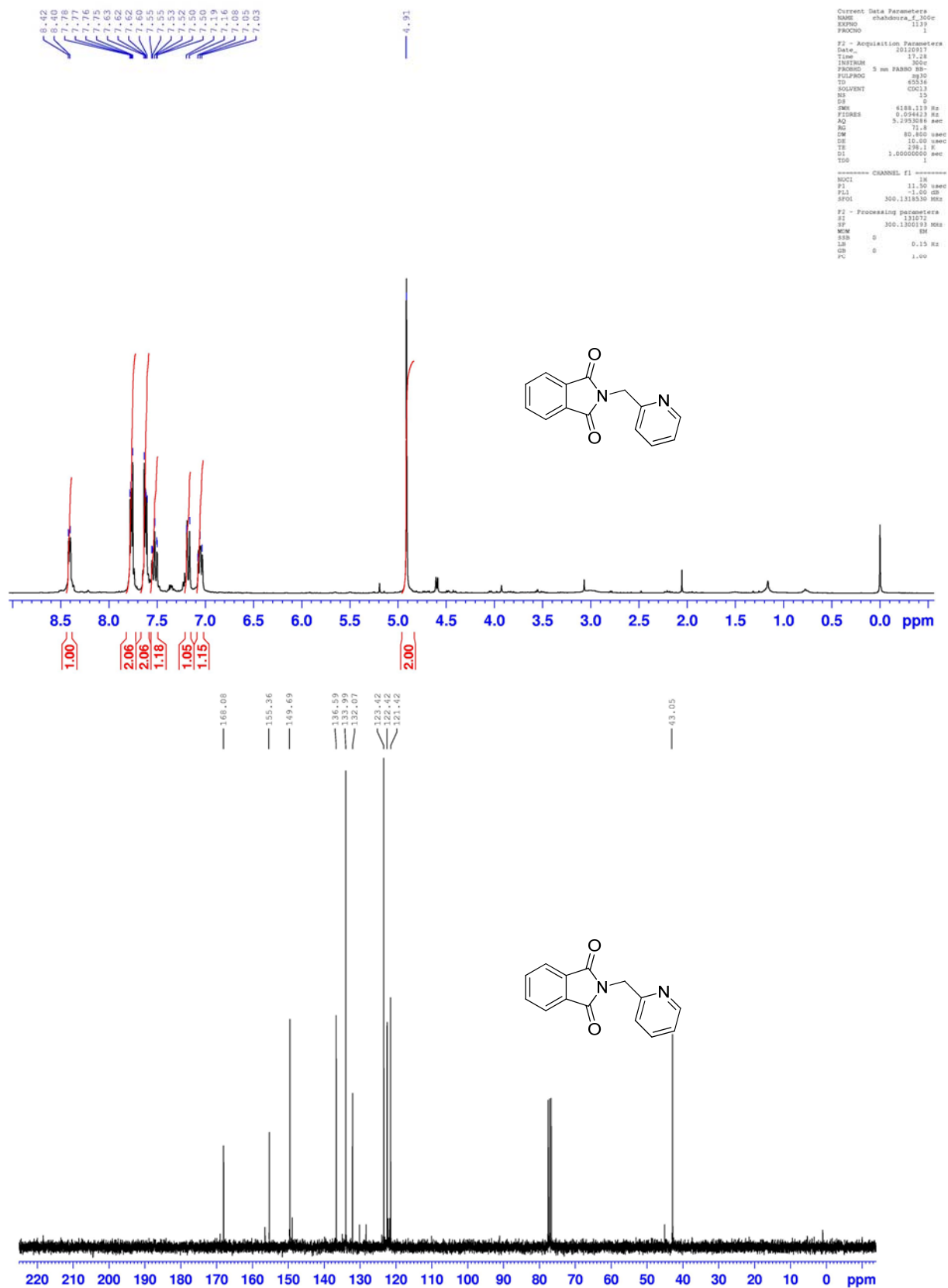
, 17-Sep-2012 + 16:06:30  
Scan EI+  
TIC  
4.05e8



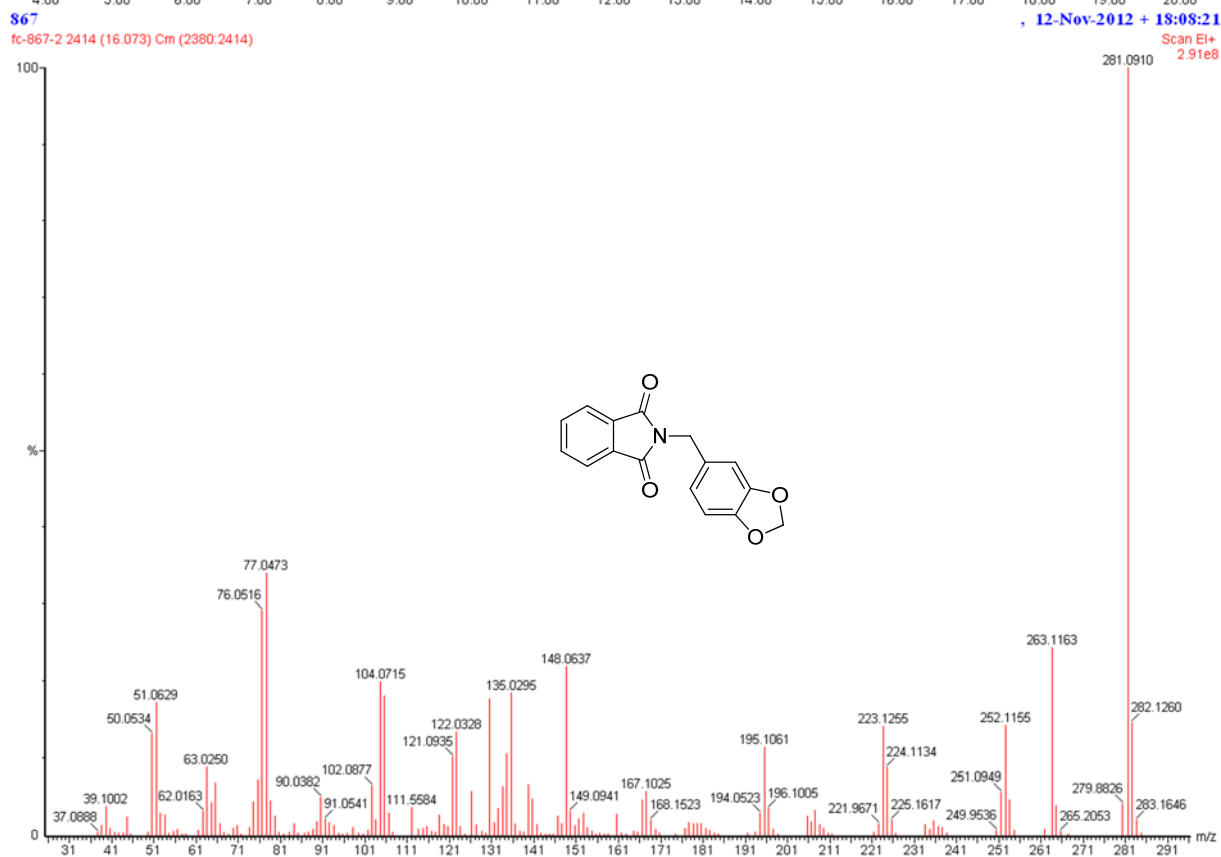
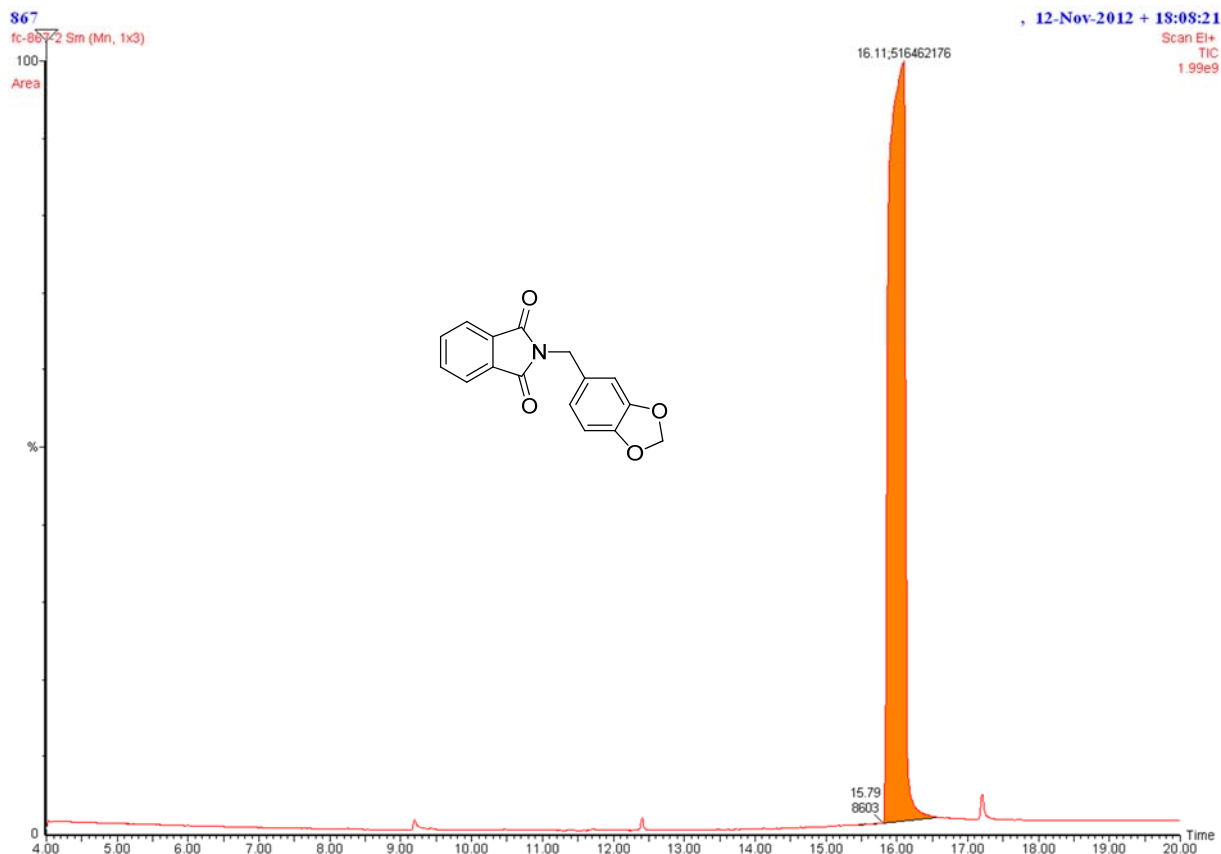
852  
fc-852- 1935 (13.677) Cm (1932:1941)

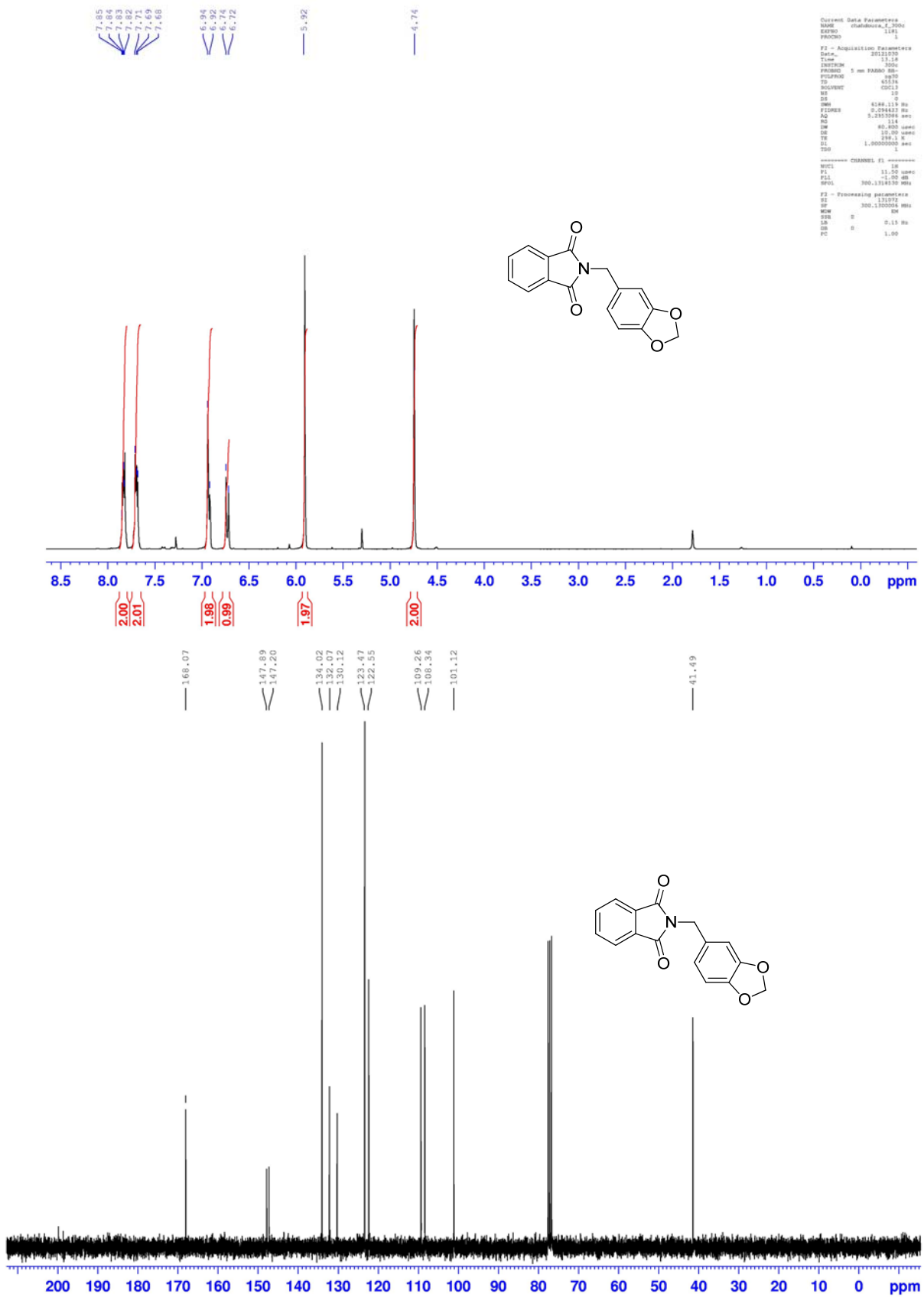
, 17-Sep-2012 + 16:06:30  
Scan EI+  
4.14e7



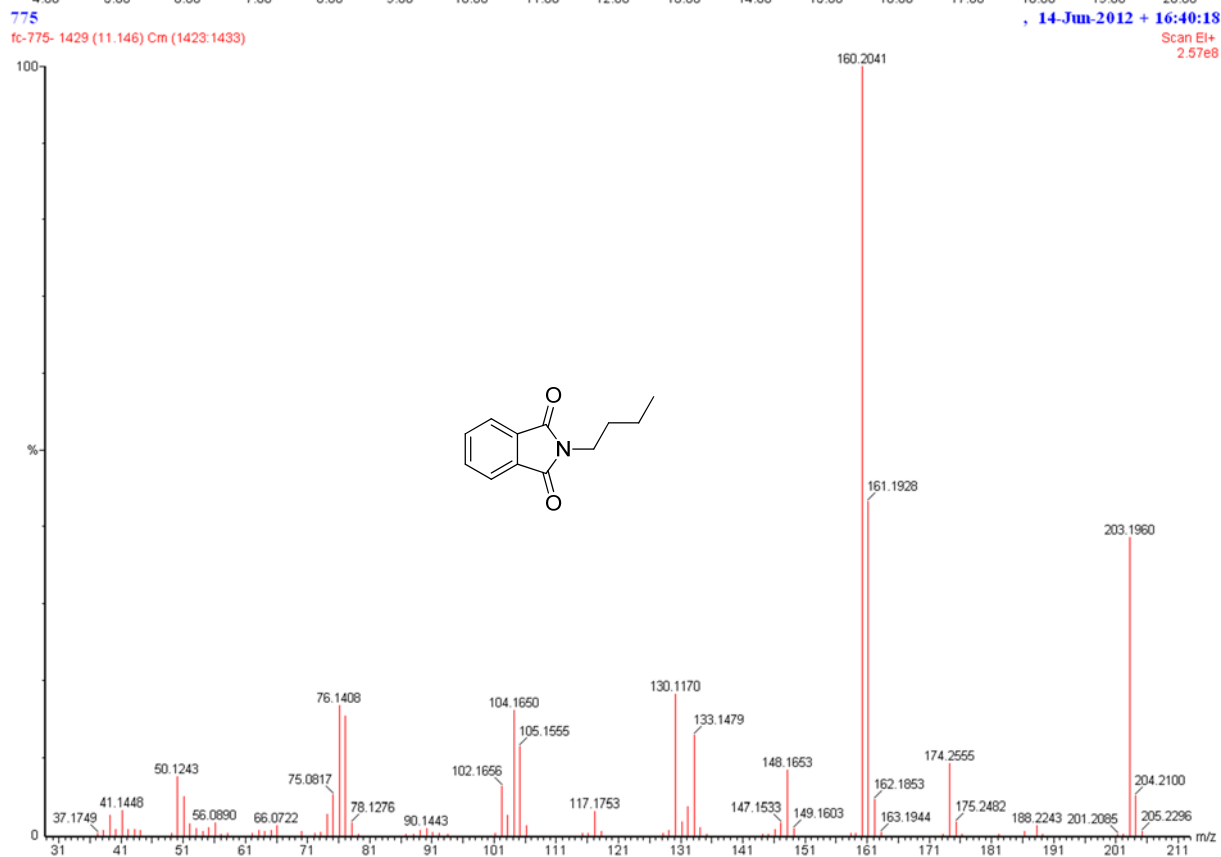
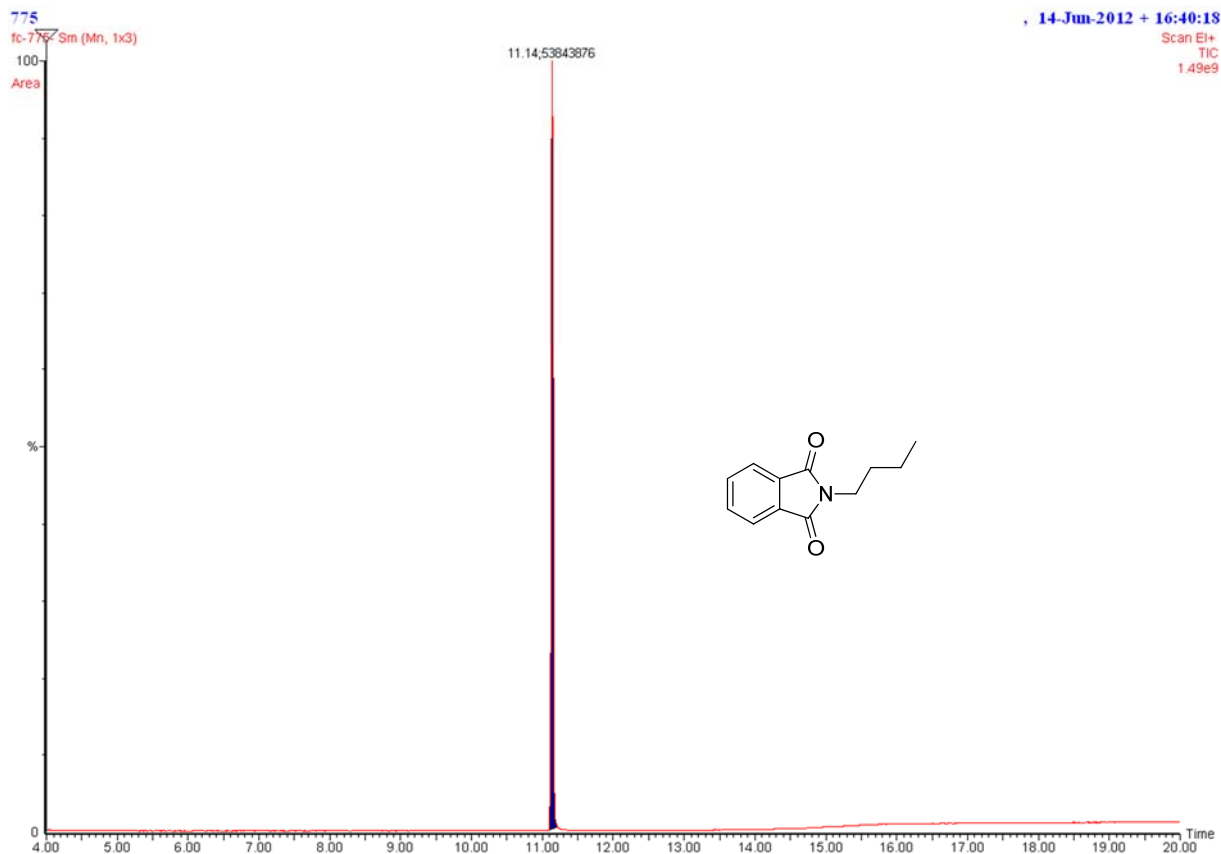


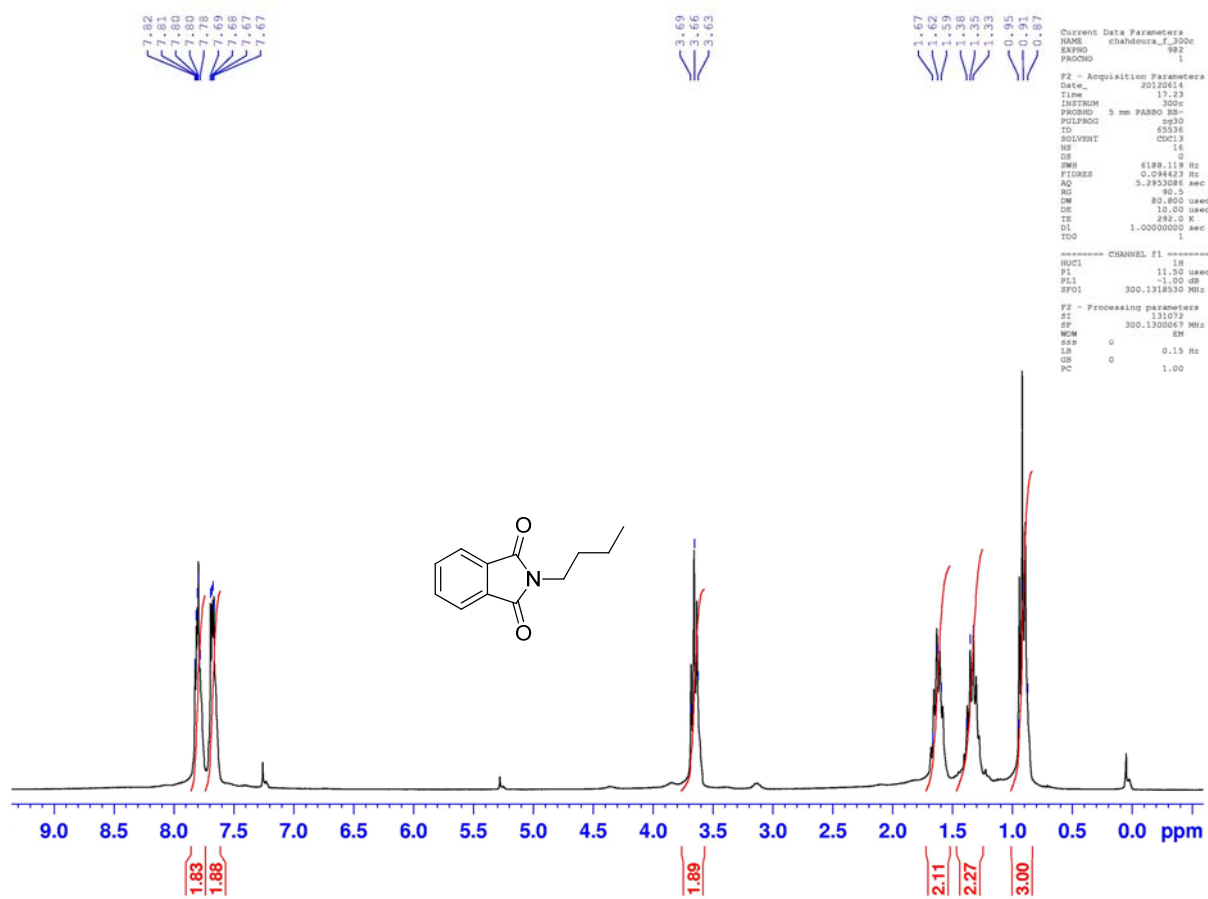
GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **a7**





GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **a8**





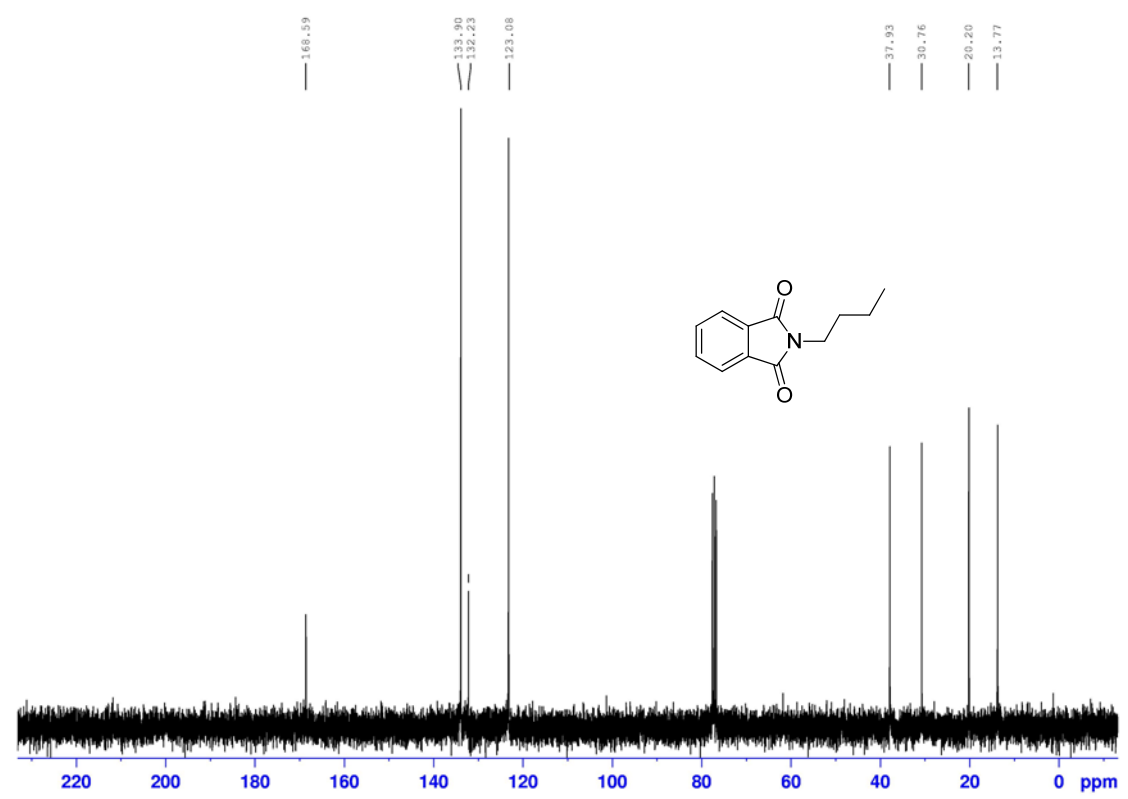
```

Current Data Parameters
NAME chahdoura_f_300c
EXPNO 322
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120614
Time 17.23
INSTRUM 300c
PROBHD 3 mm F4BBO 88-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 0
SWH 4188.119 Hz
FIDRES 0.094423 Hz
AQ 5.2932086 sec
RG 95.5
DM 80.800 usec
DE 10.00 usec
TE 292.0 K
D1 1.00000000 sec
TDO 1

===== CHANNEL f1 =====
NUC1 1H
P1 11.50 usec
PL1 -1.00 dB
SFO1 300.1318330 MHz

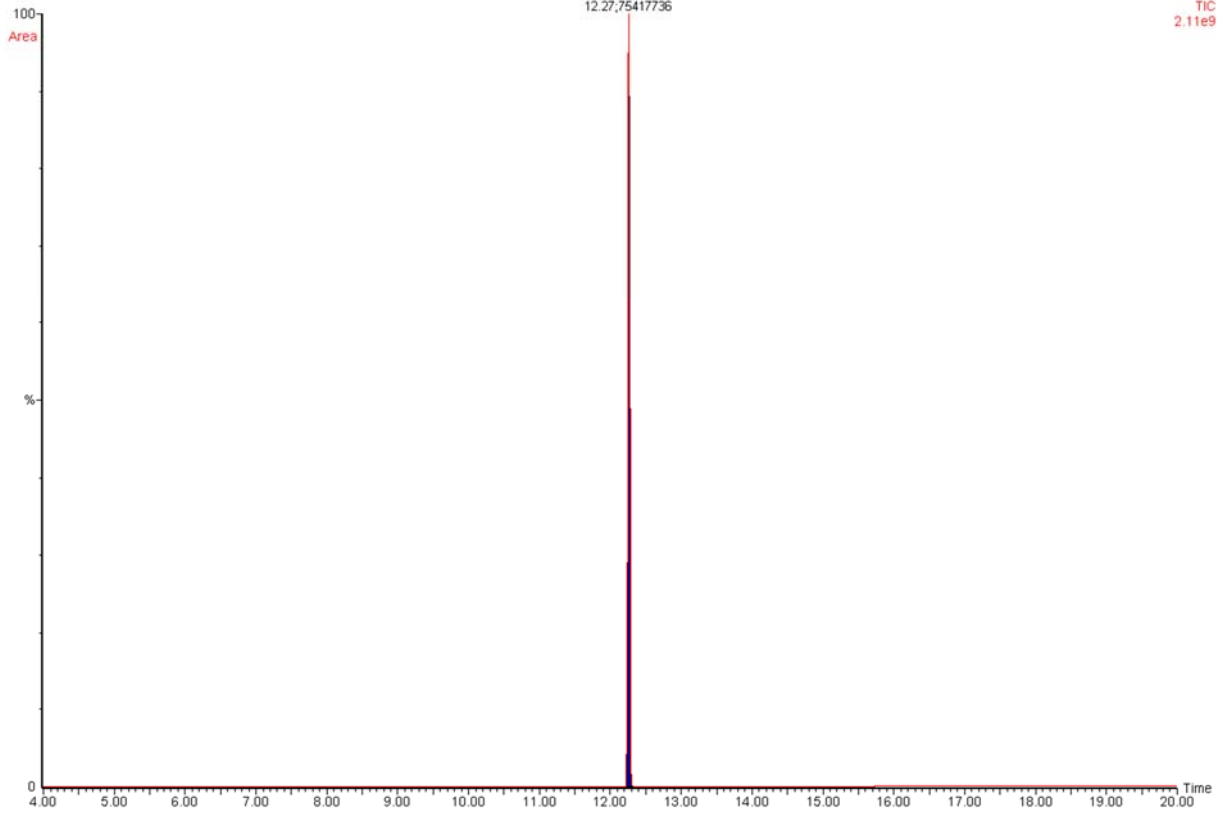
F2 - Processing parameters
SI 131072
SF 300.1300000 MHz
WDW EM
SSB 0
LB 0.15 Hz
GB 0
PC 1.00
  
```



GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **a9**

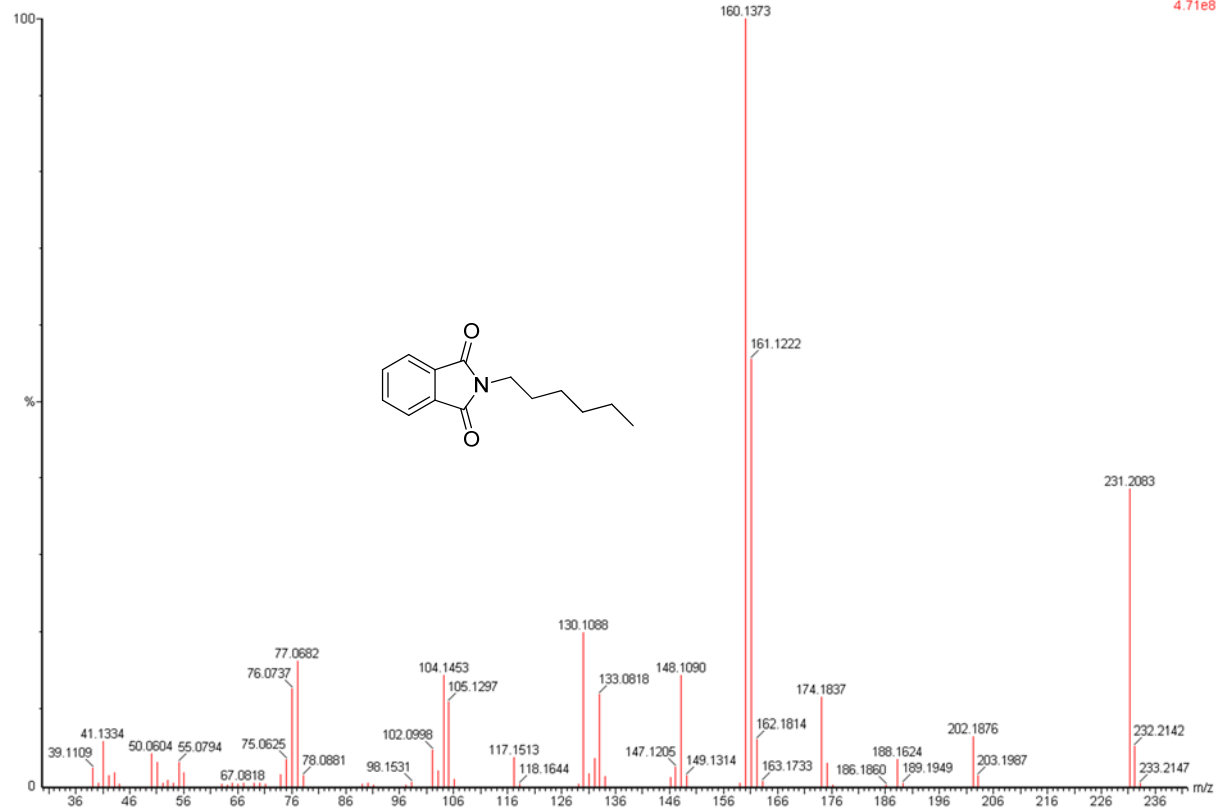
773  
fc-773- Sm (Mn, 1x3)

, 13-Jun-2012 + 19:28:31  
Scan EI+  
TIC  
2.11e9

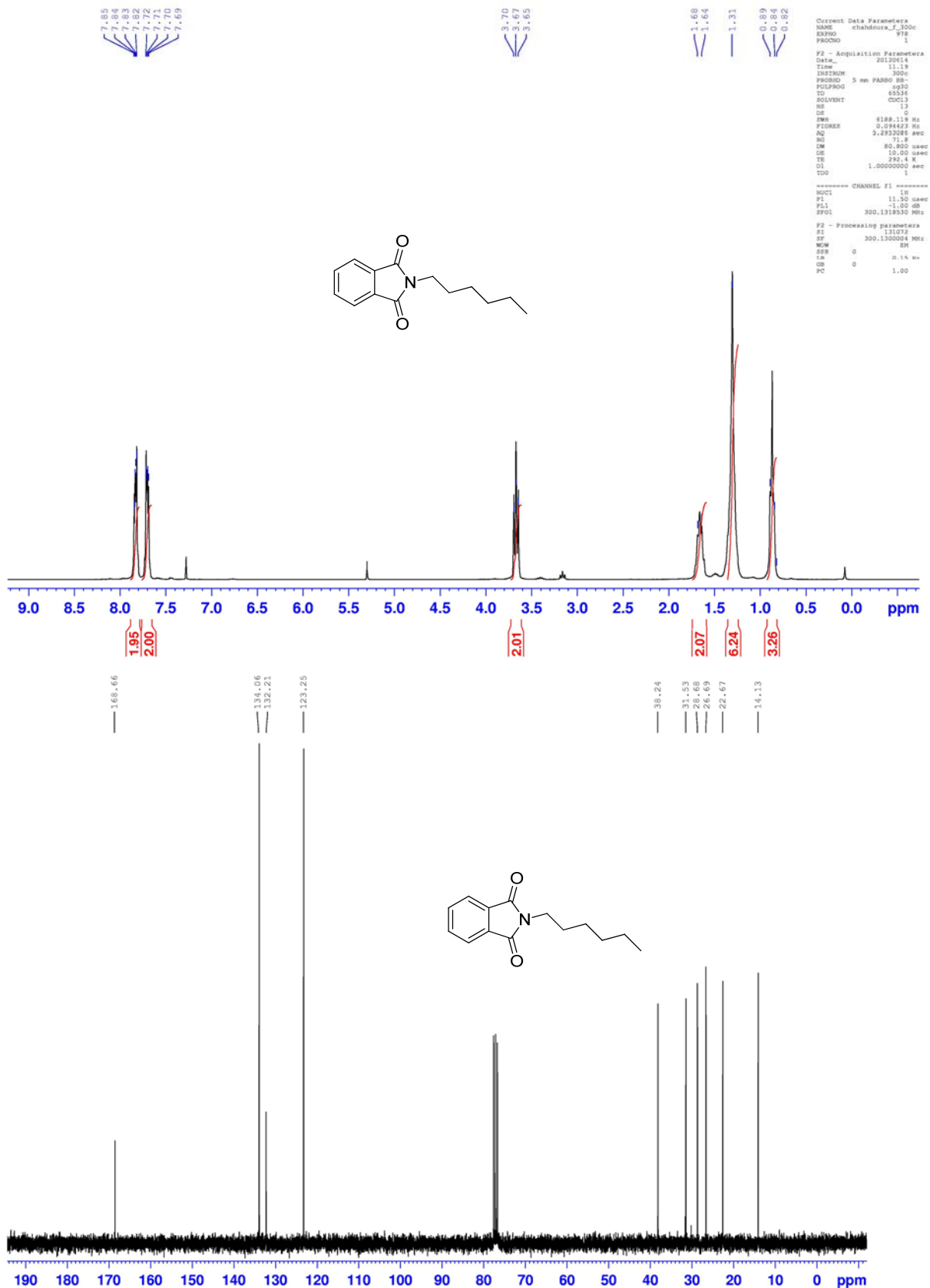


773  
fc-773- 1653 (12.267) Cm (1650:1657)

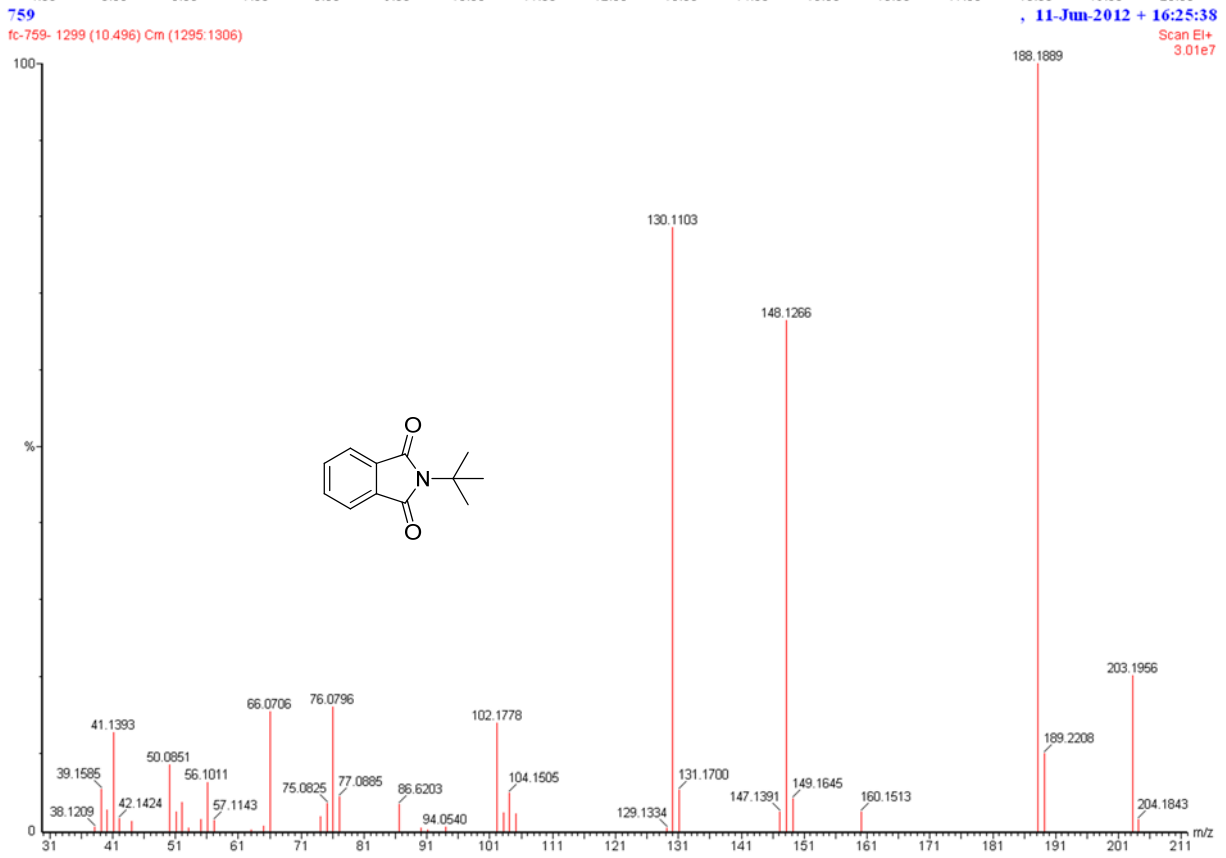
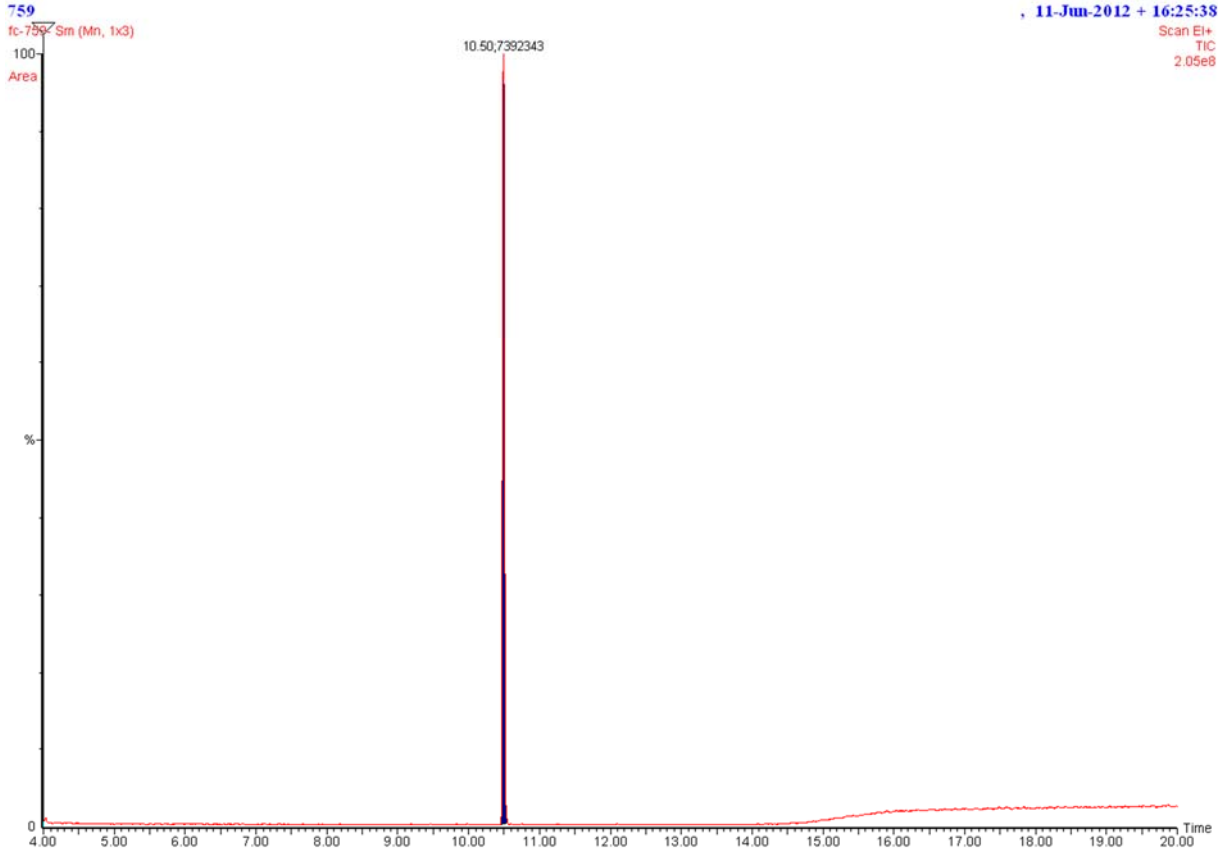
, 13-Jun-2012 + 19:28:31  
Scan EI+  
4.71e8

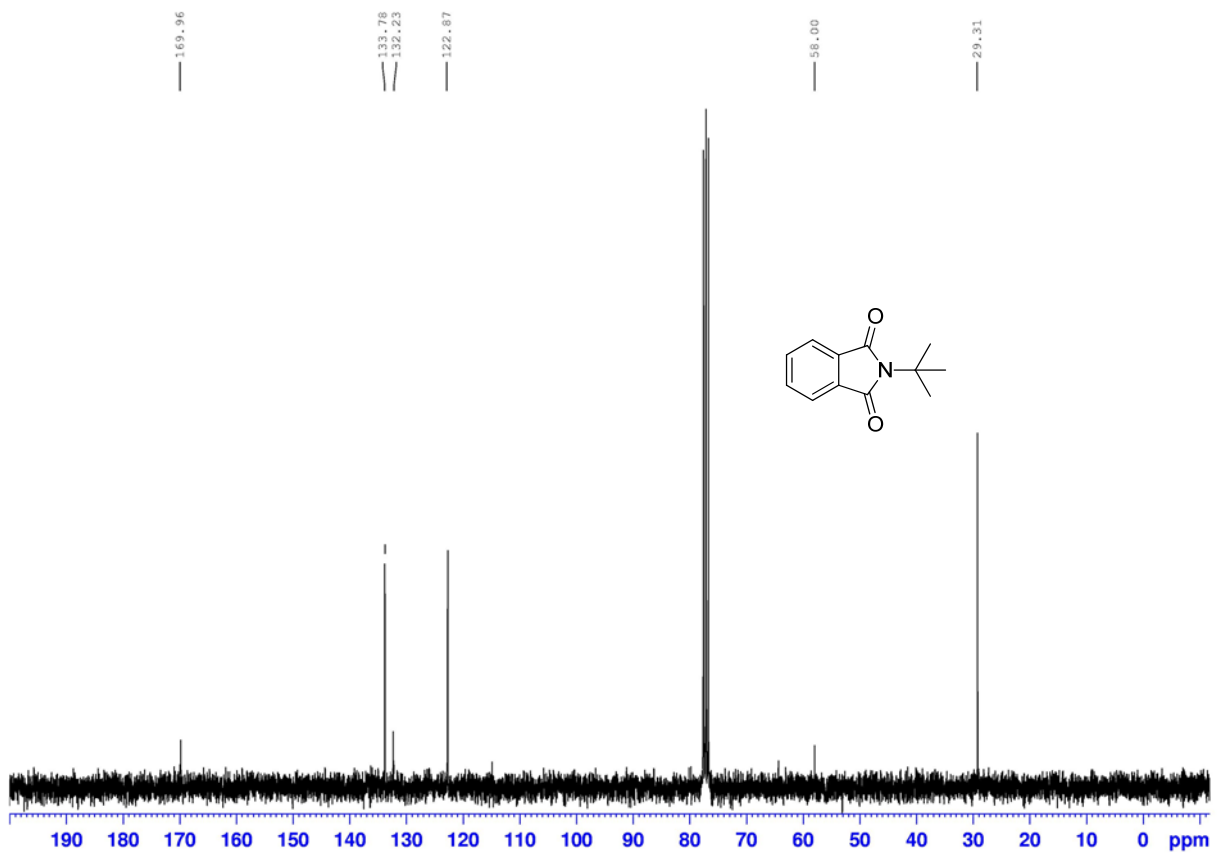
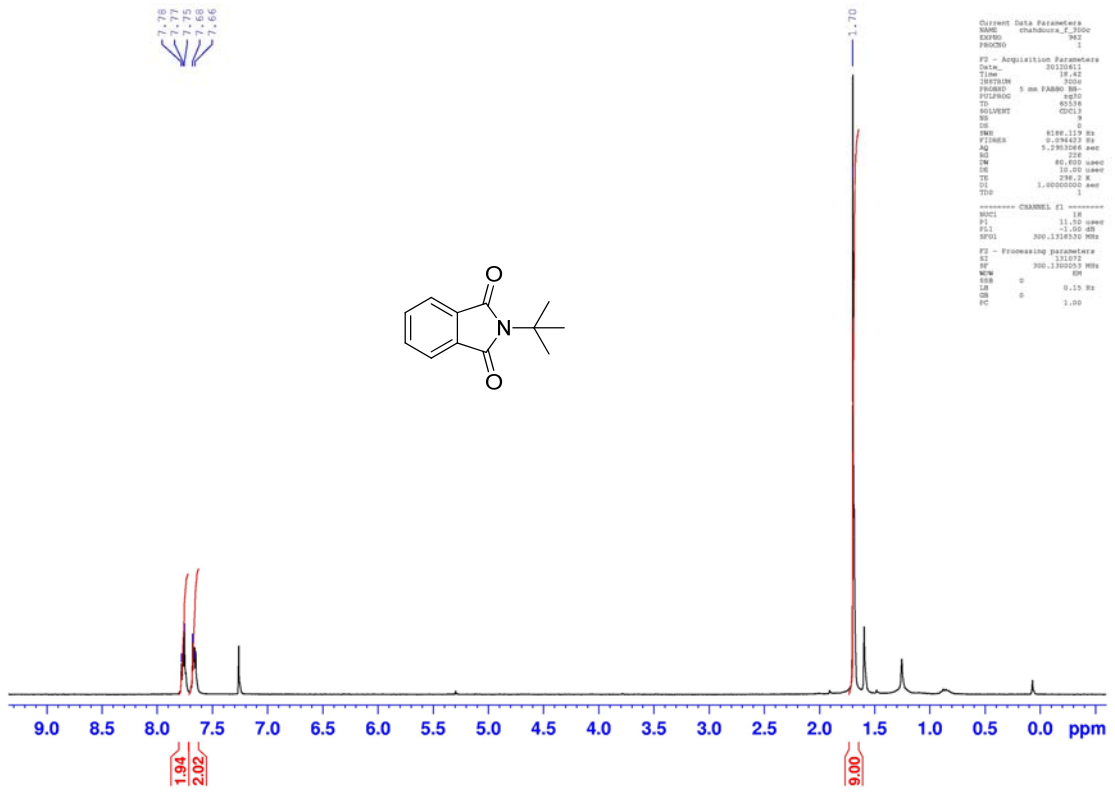






GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **a10**

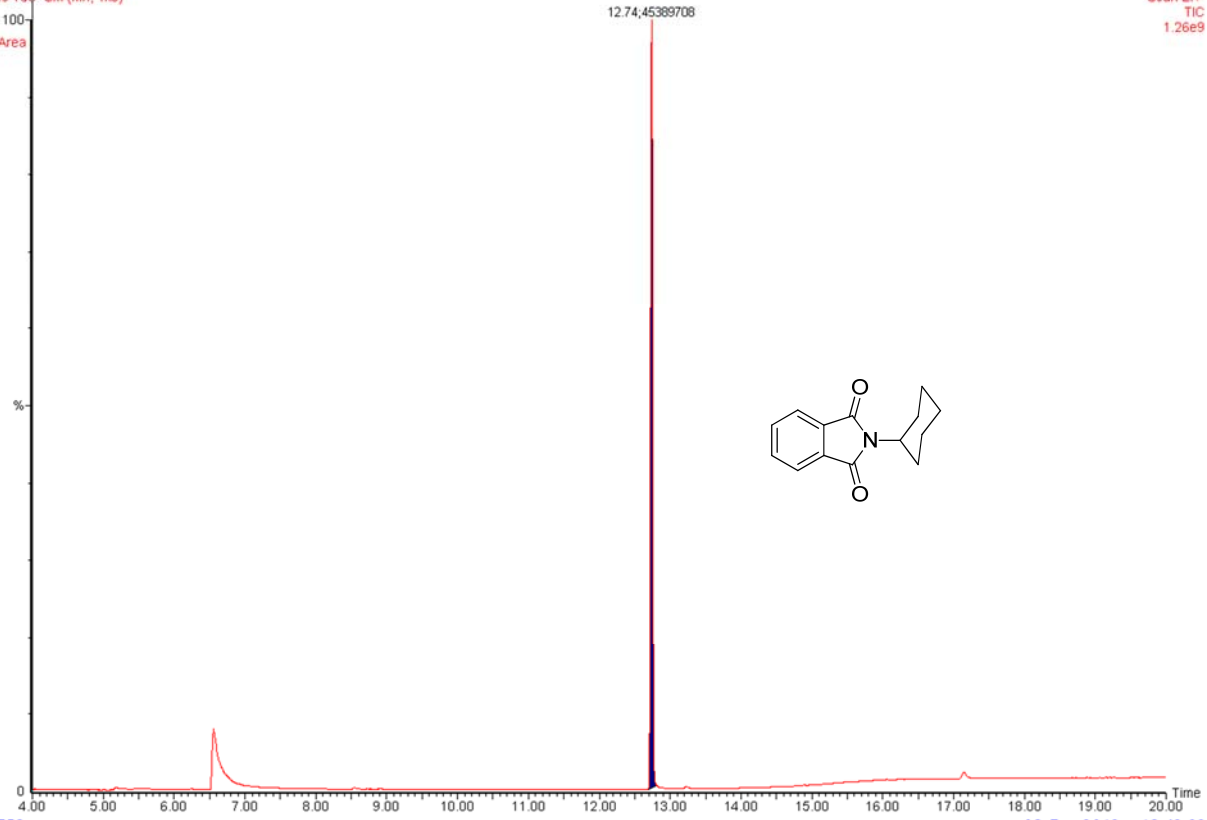




GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **a11**

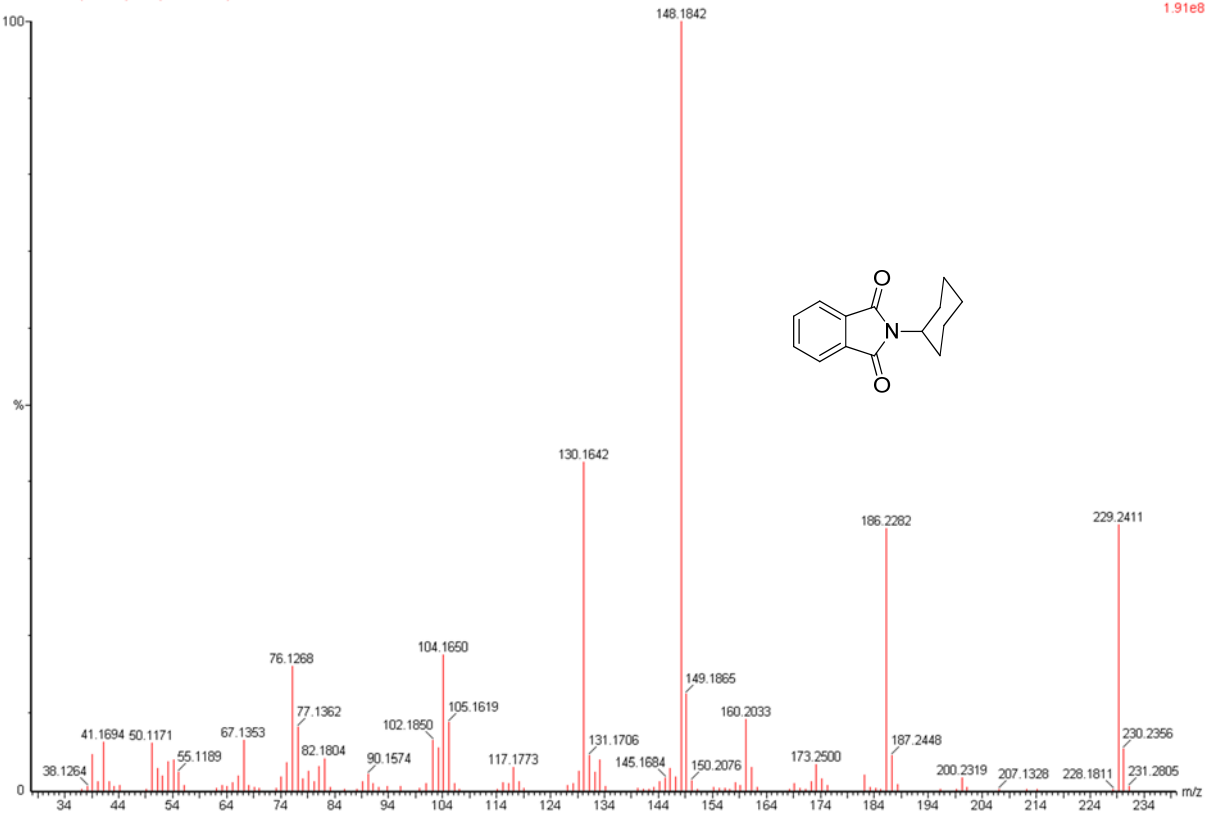
758  
tc-758 Sm (Mn, 1x3)  
Area

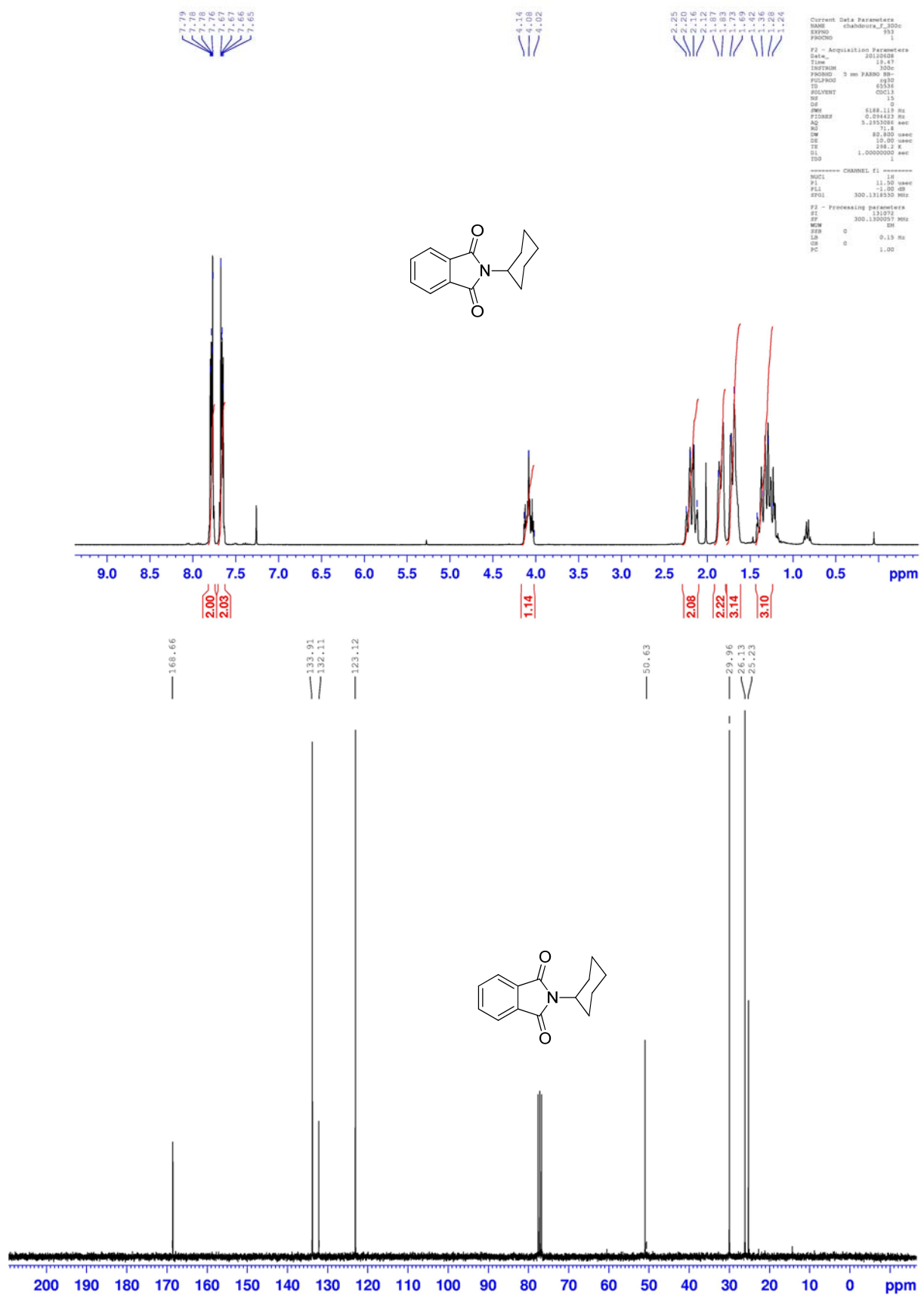
, 08-Jun-2012 + 18:42:32  
Scan EI+  
TIC  
1.26e9



758  
tc-758- 1749 (12.747) Cm (1744:1754)

, 08-Jun-2012 + 18:42:32  
Scan EI+  
1.91e8

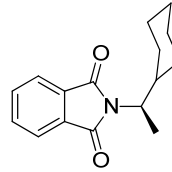
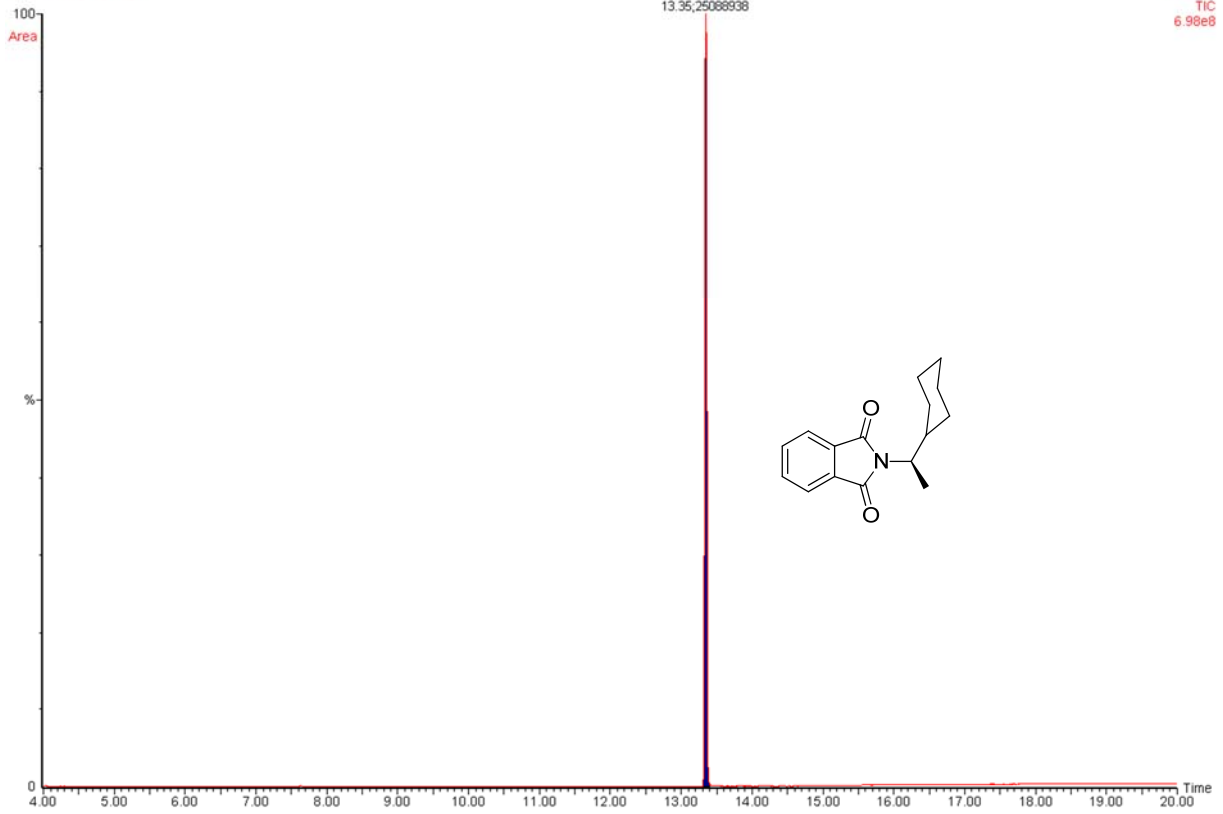




GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **a12**

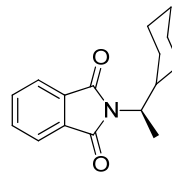
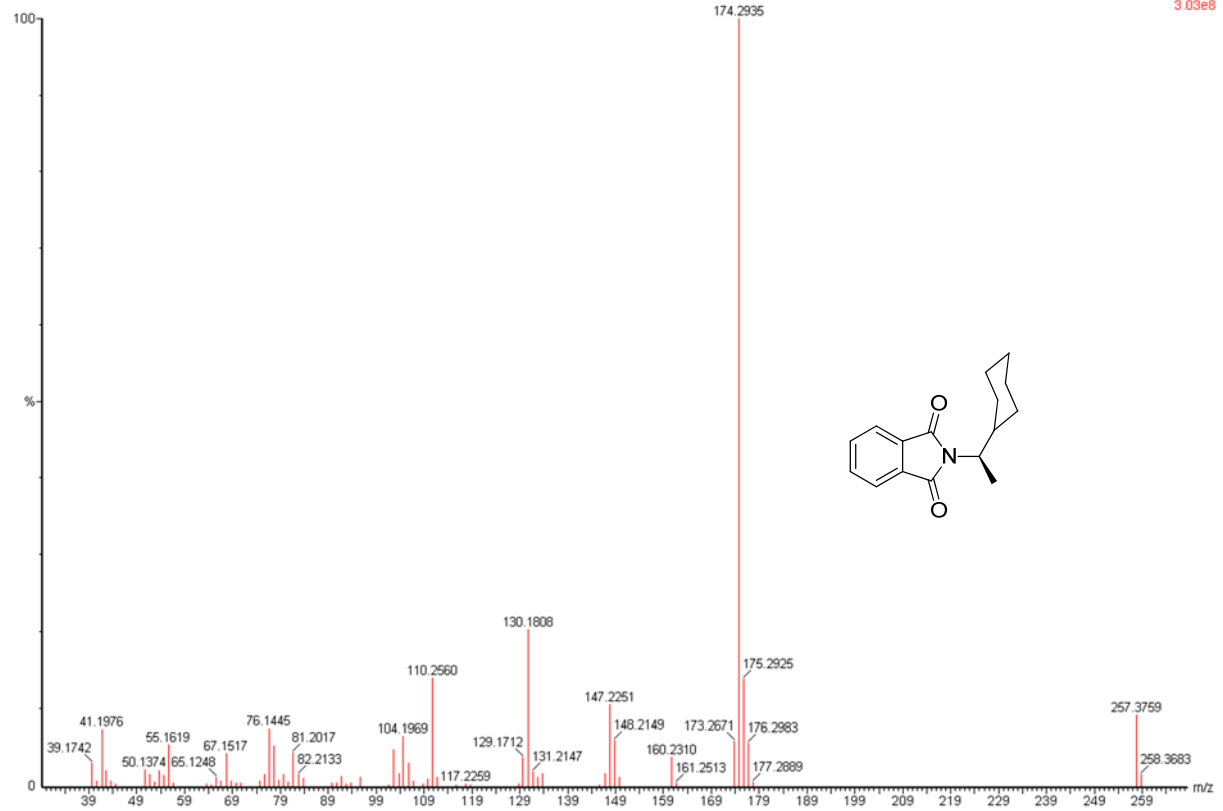
794  
fc-794- Sm (Mn, 1x3)

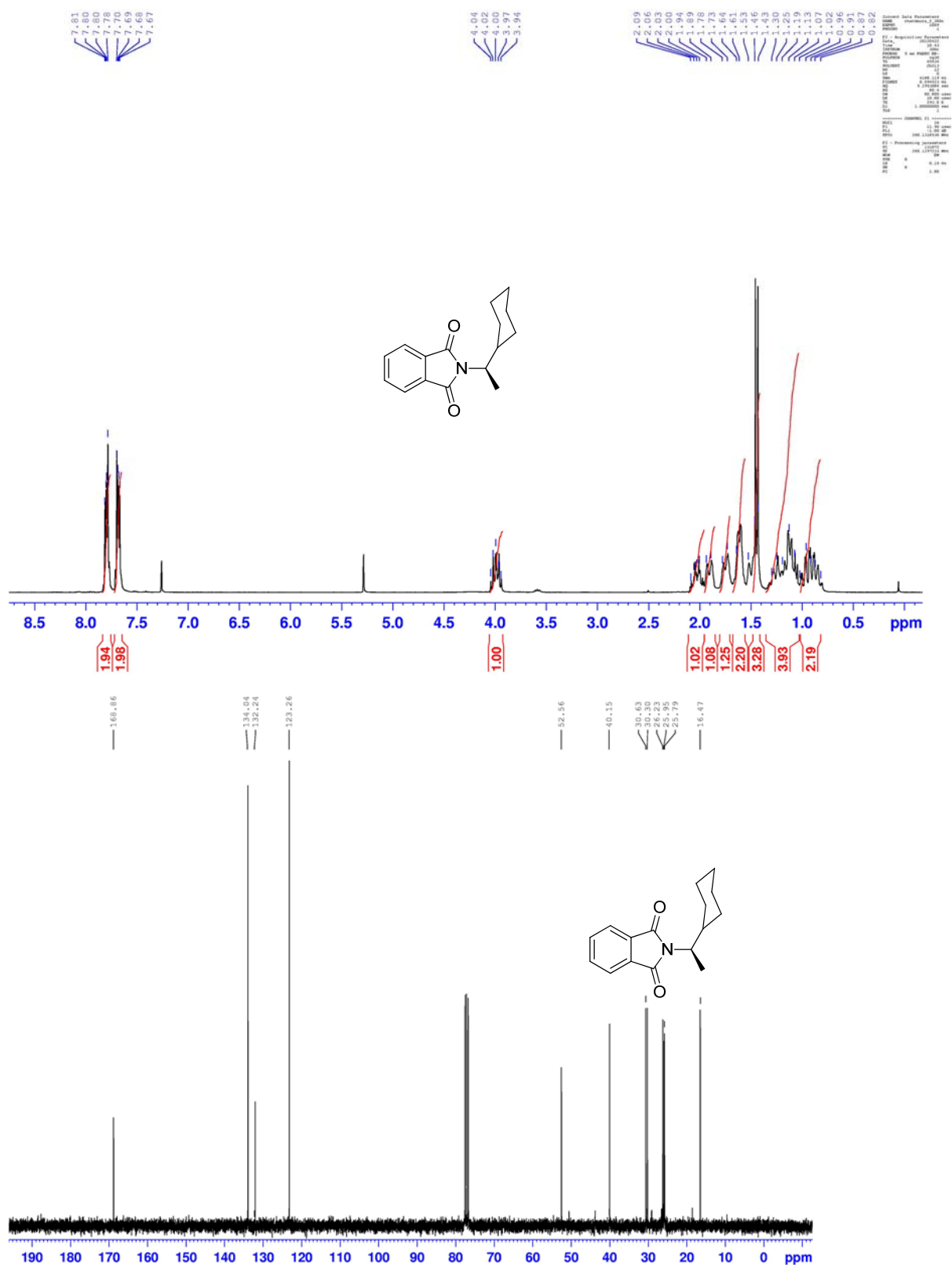
, 22-Jun-2012 + 09:43:19  
Scan EI+  
TIC  
6.98e8



794  
fc-794- 1870 (13.352) Cm (1867:1871)

, 22-Jun-2012 + 09:43:19  
Scan EI+  
3.03e8

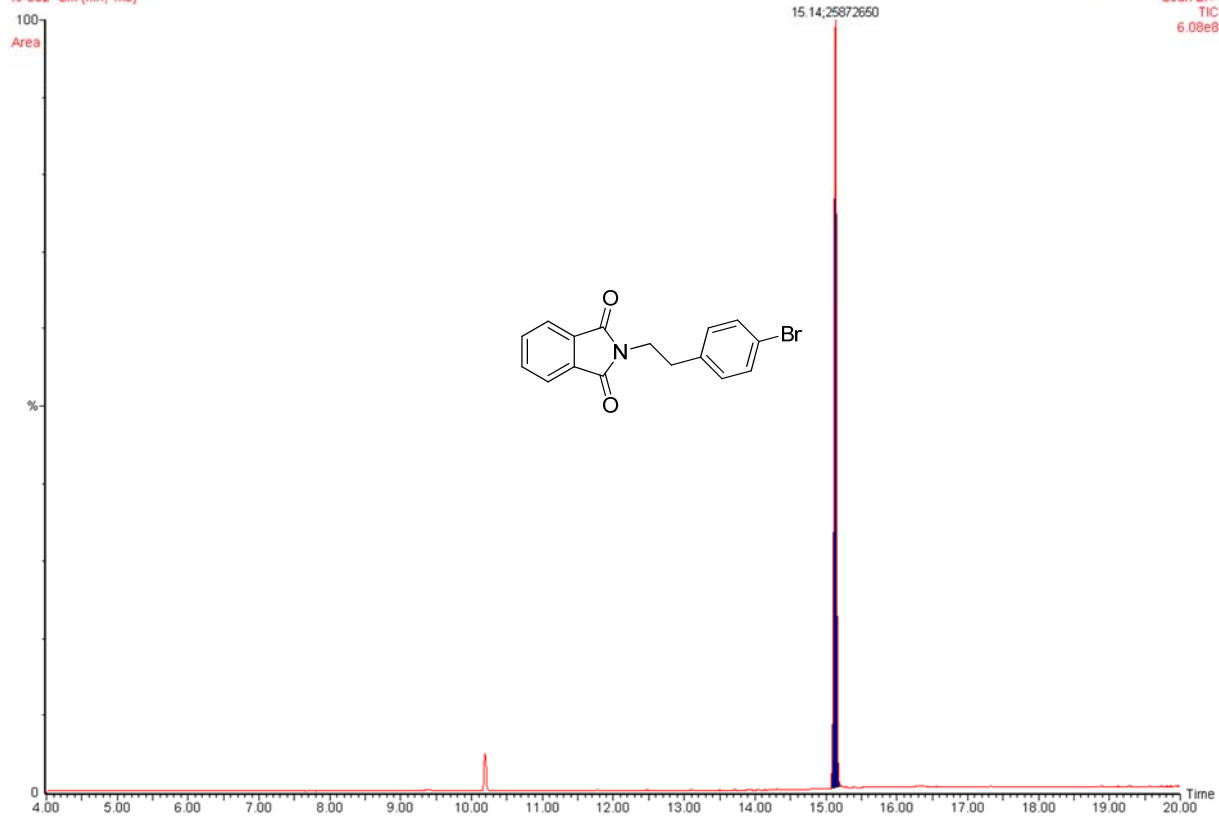




GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$  { $^1\text{H}$ } NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **a13**

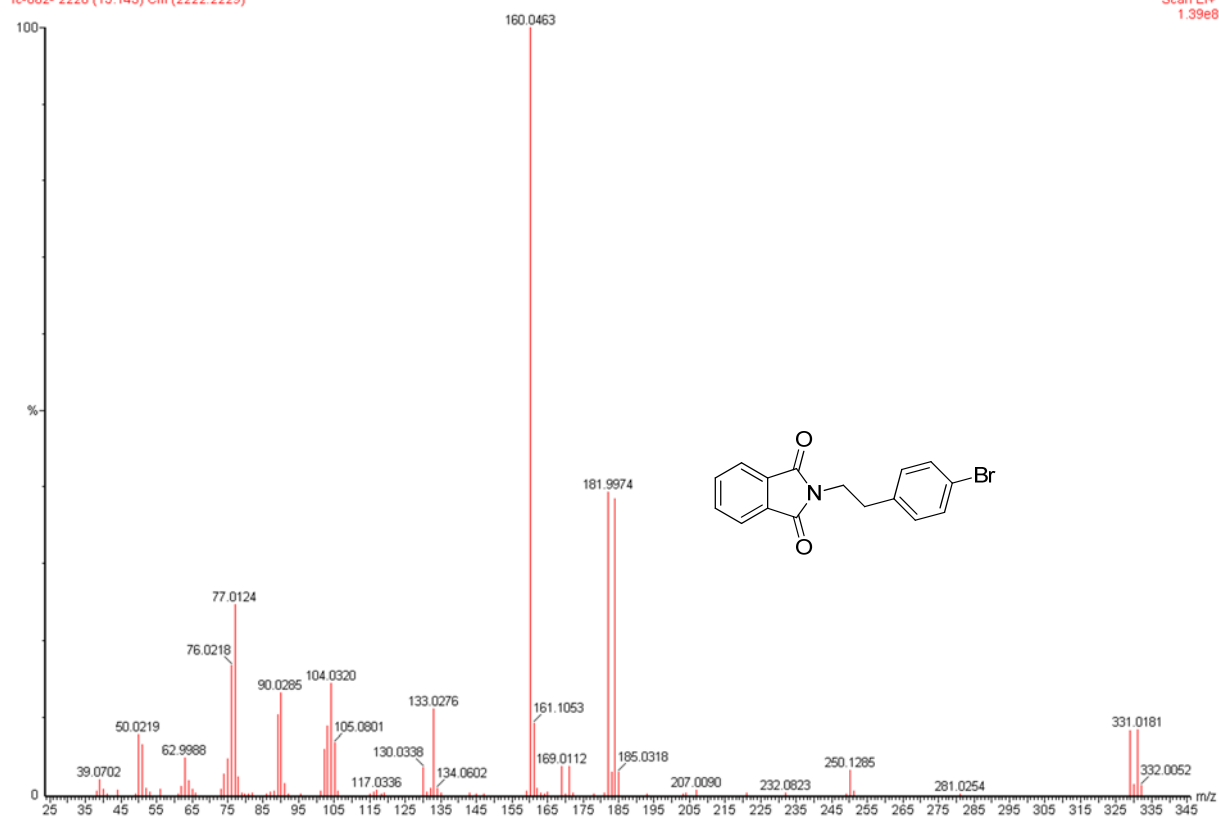
S62  
fc-862- Sm (Mn, 1x3)

, 24-Sep-2012 + 17:04:53

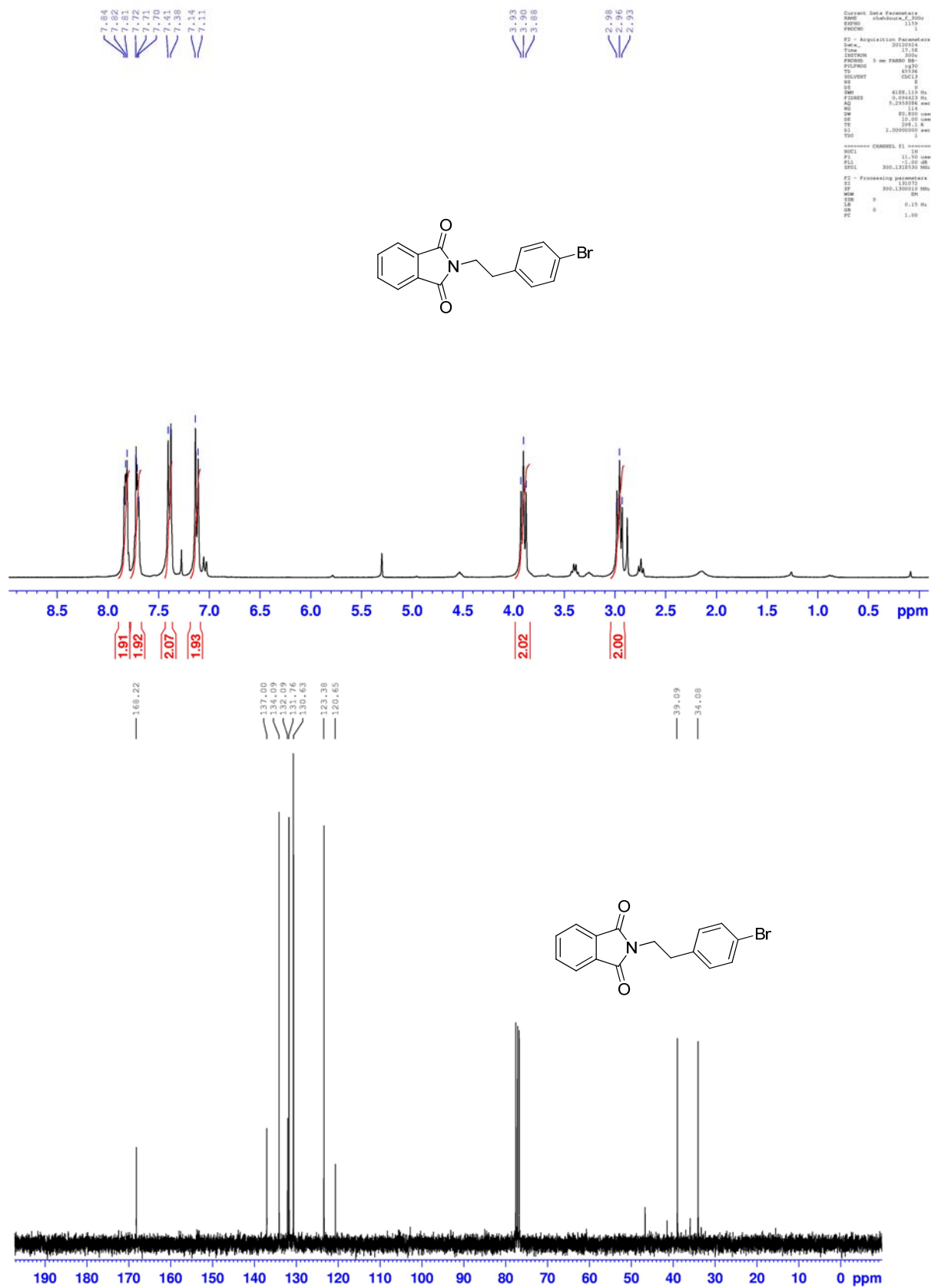


S62  
fc-862- 2228 (15.143) Cm (2222.2229)

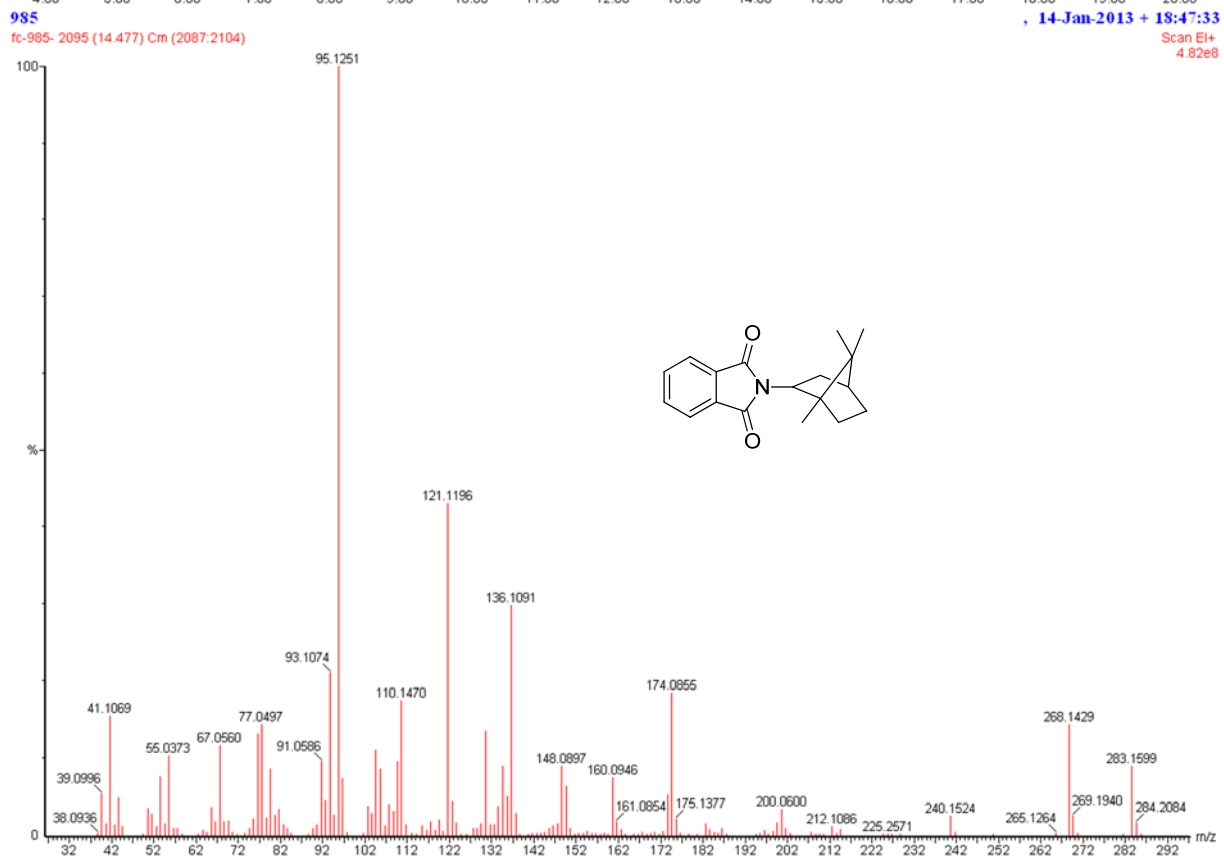
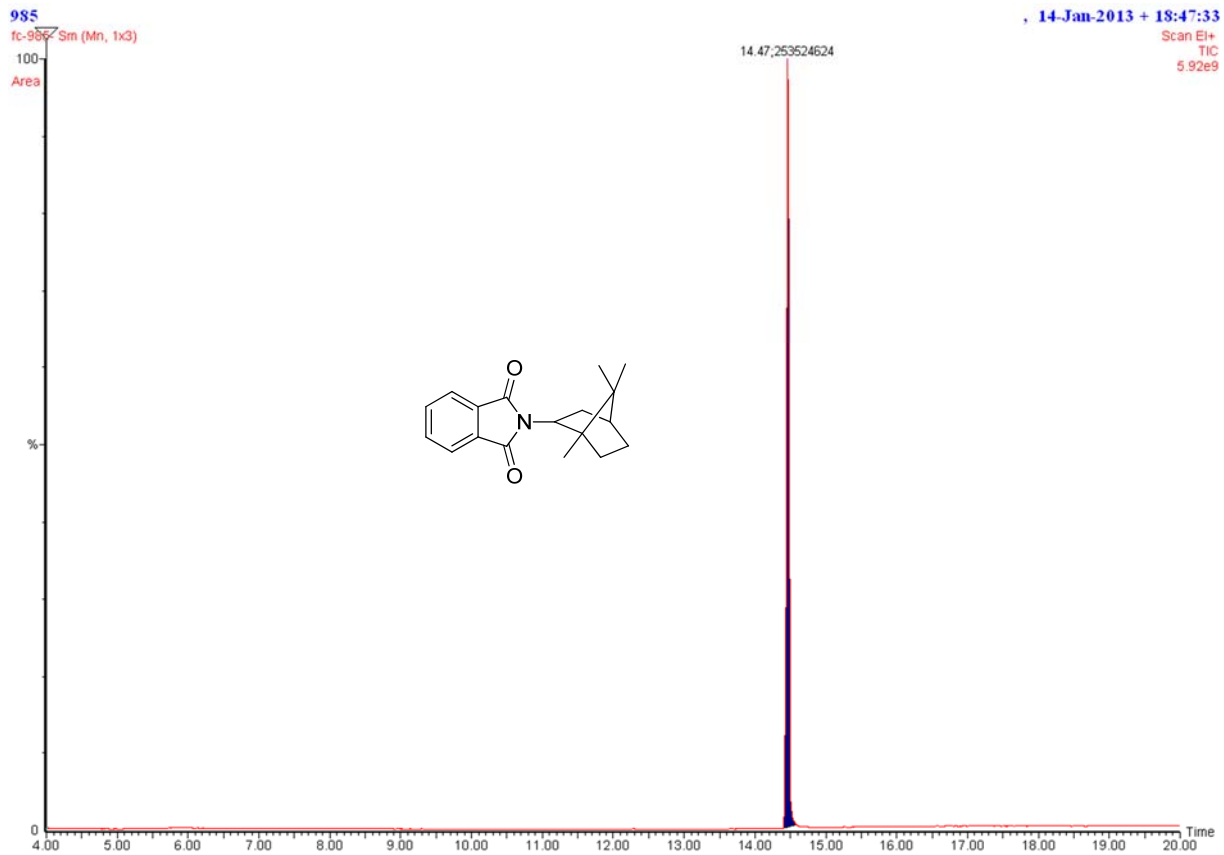
, 24-Sep-2012 + 17:04:53

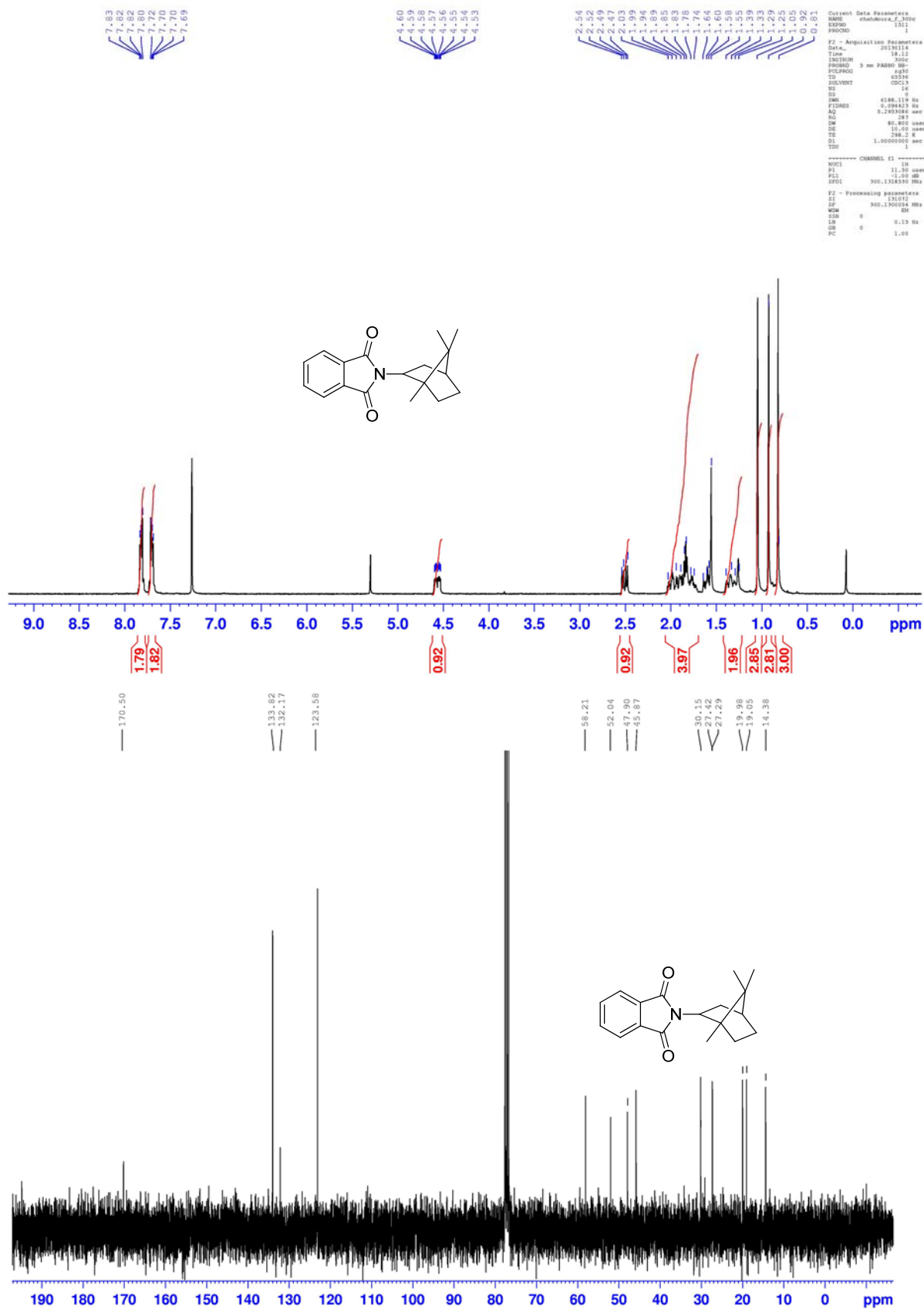






GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **a14**

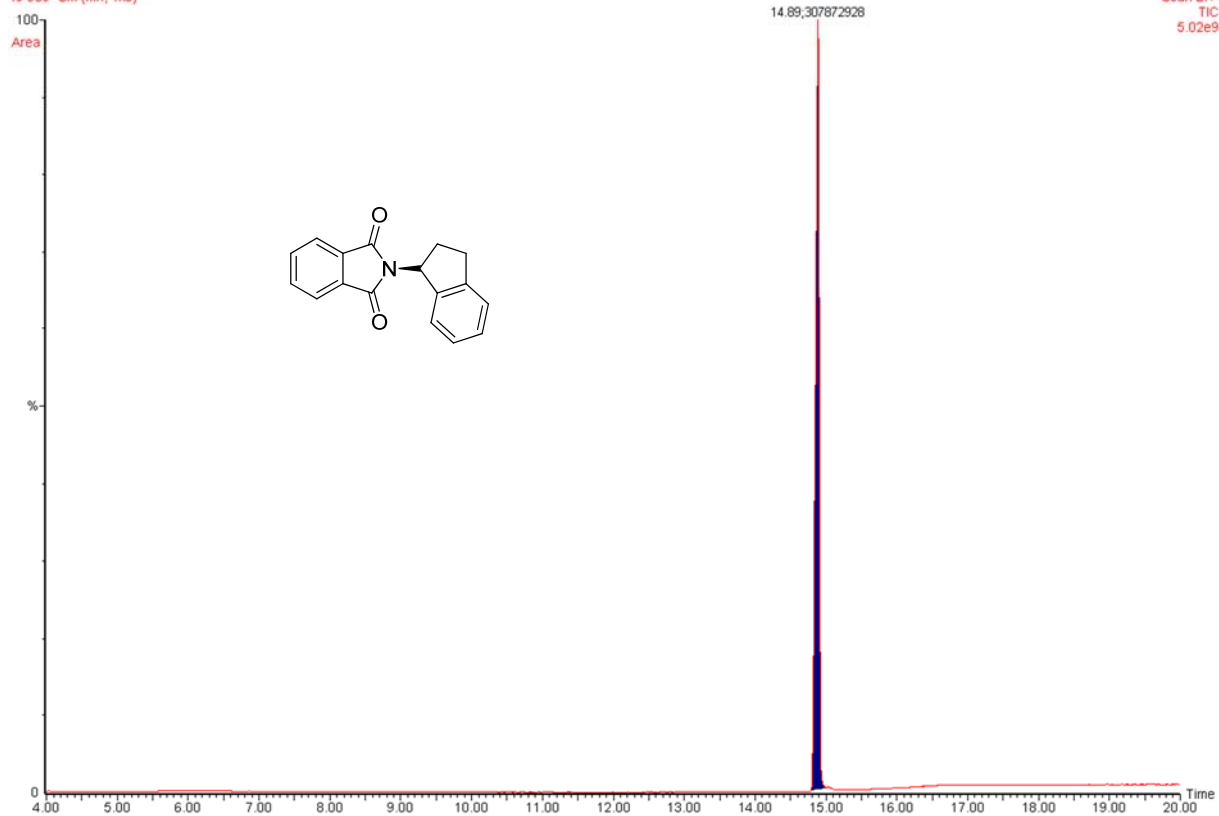




GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **a15**

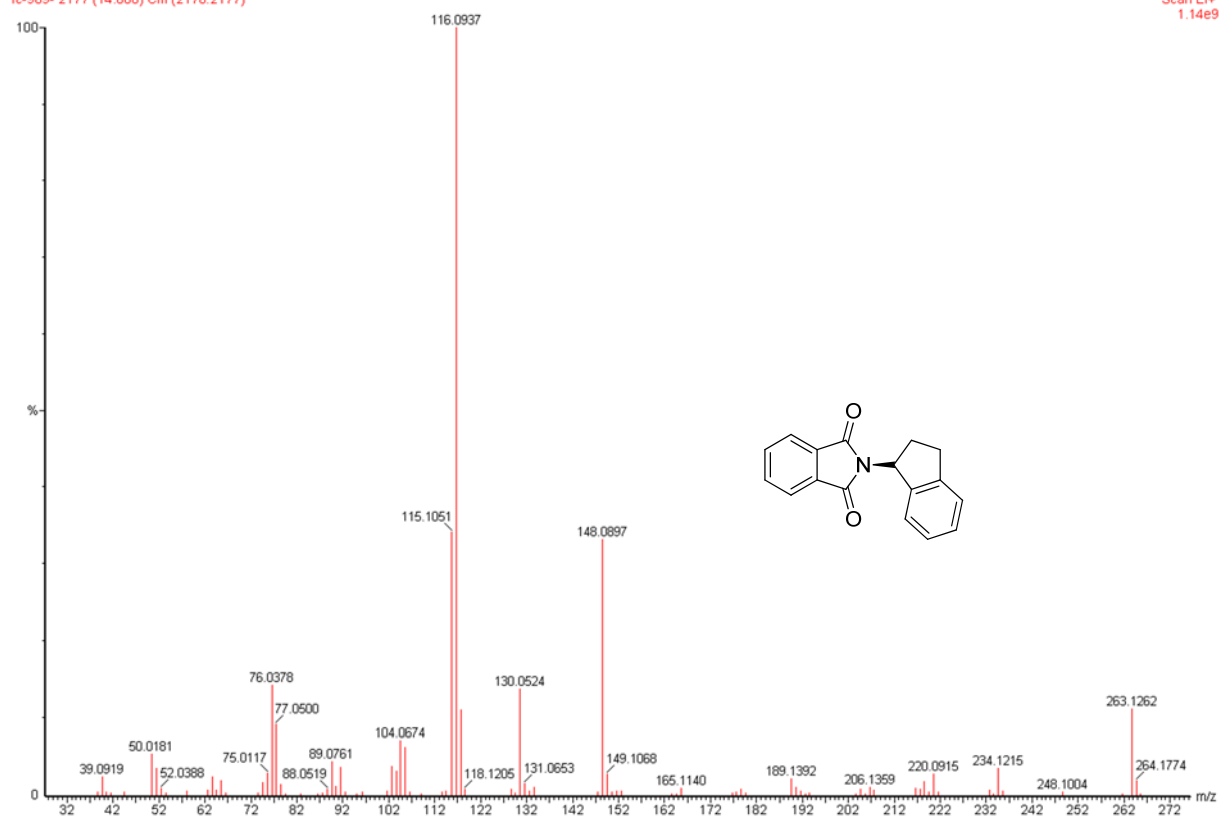
989  
fc-989- Sm (Mn, 1x3)

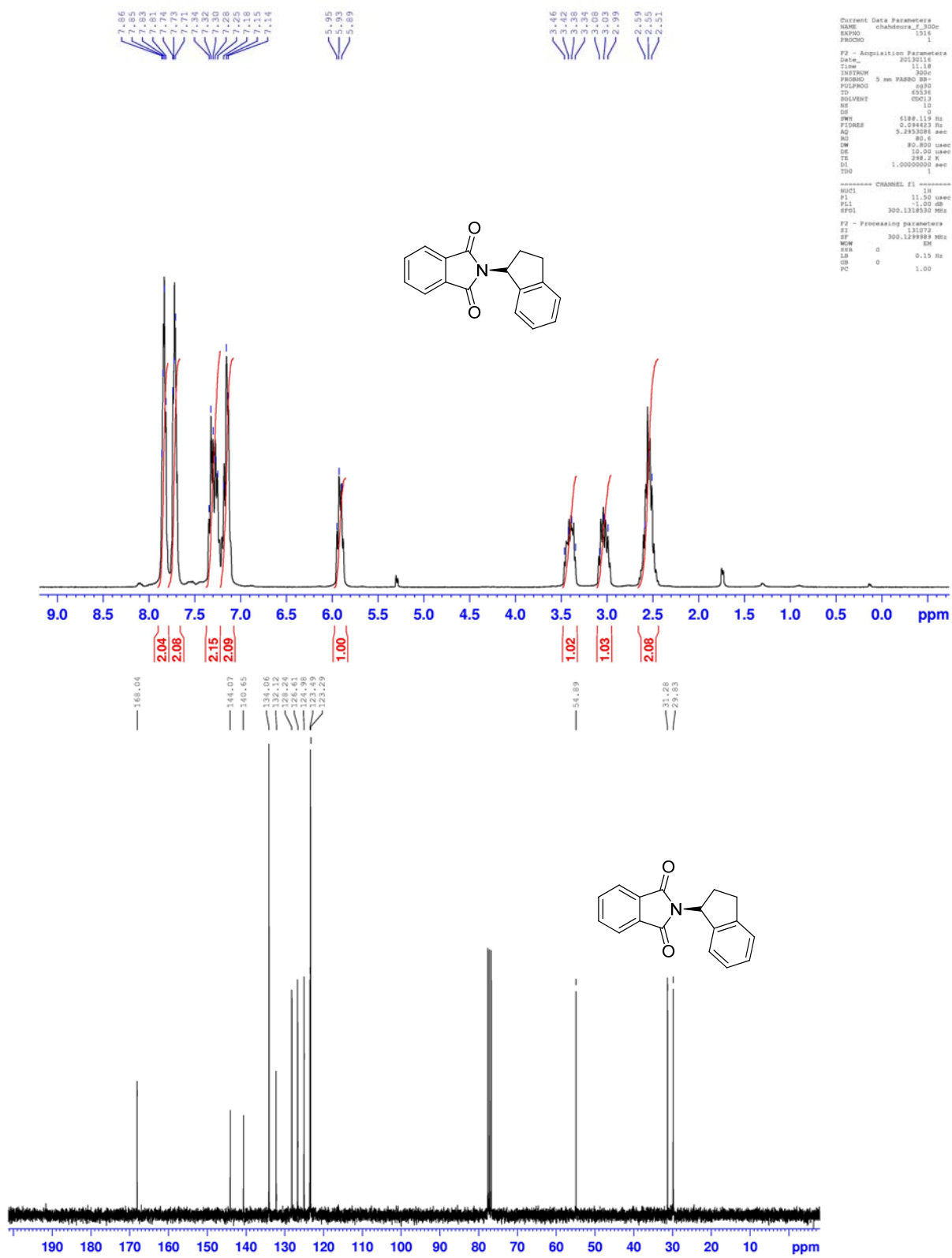
, 16-Jan-2013 + 09:41:55



989  
fc-989- 2177 (14.888) Cm (2170:2177)

, 16-Jan-2013 + 09:41:55



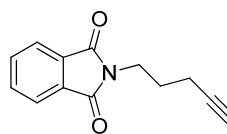
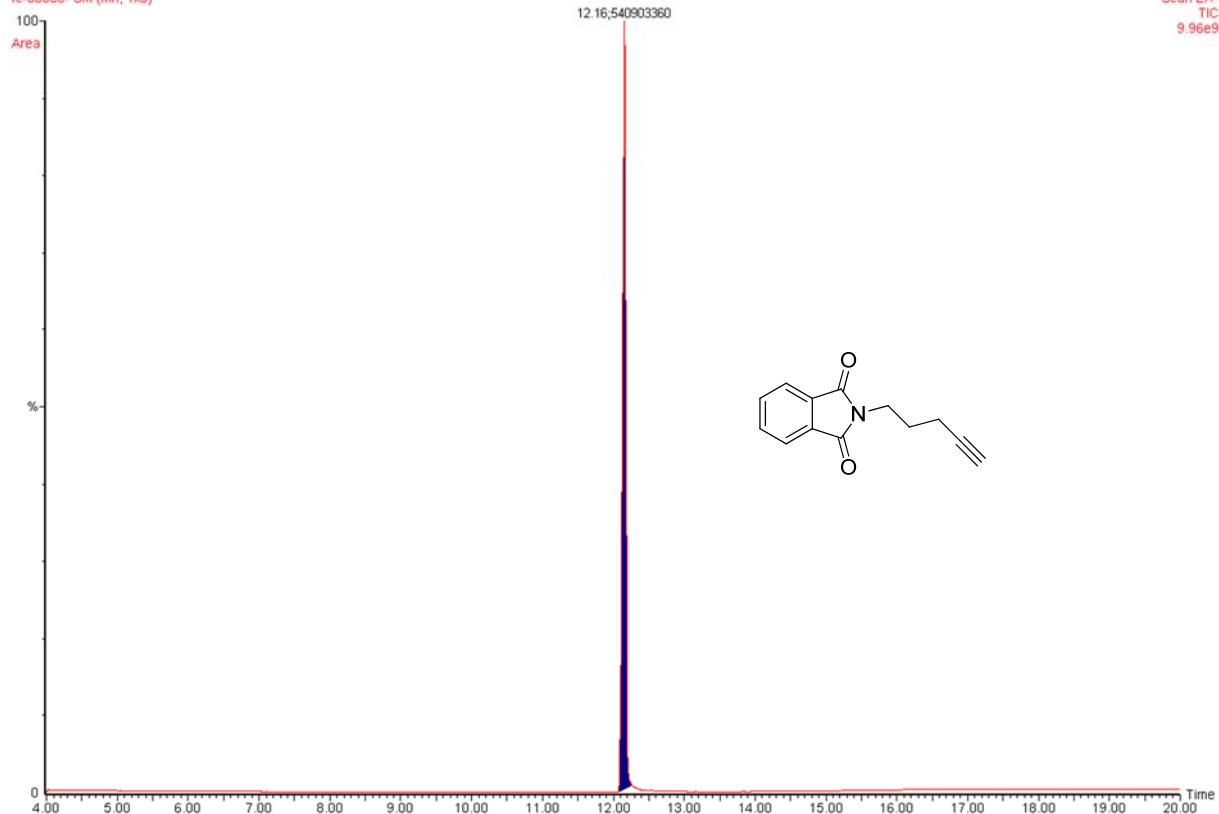


GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **a16**

0  
fc-00000- Sm (Mn, 1x3)

, 23-Nov-2012 + 13:16:58

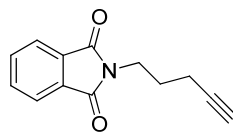
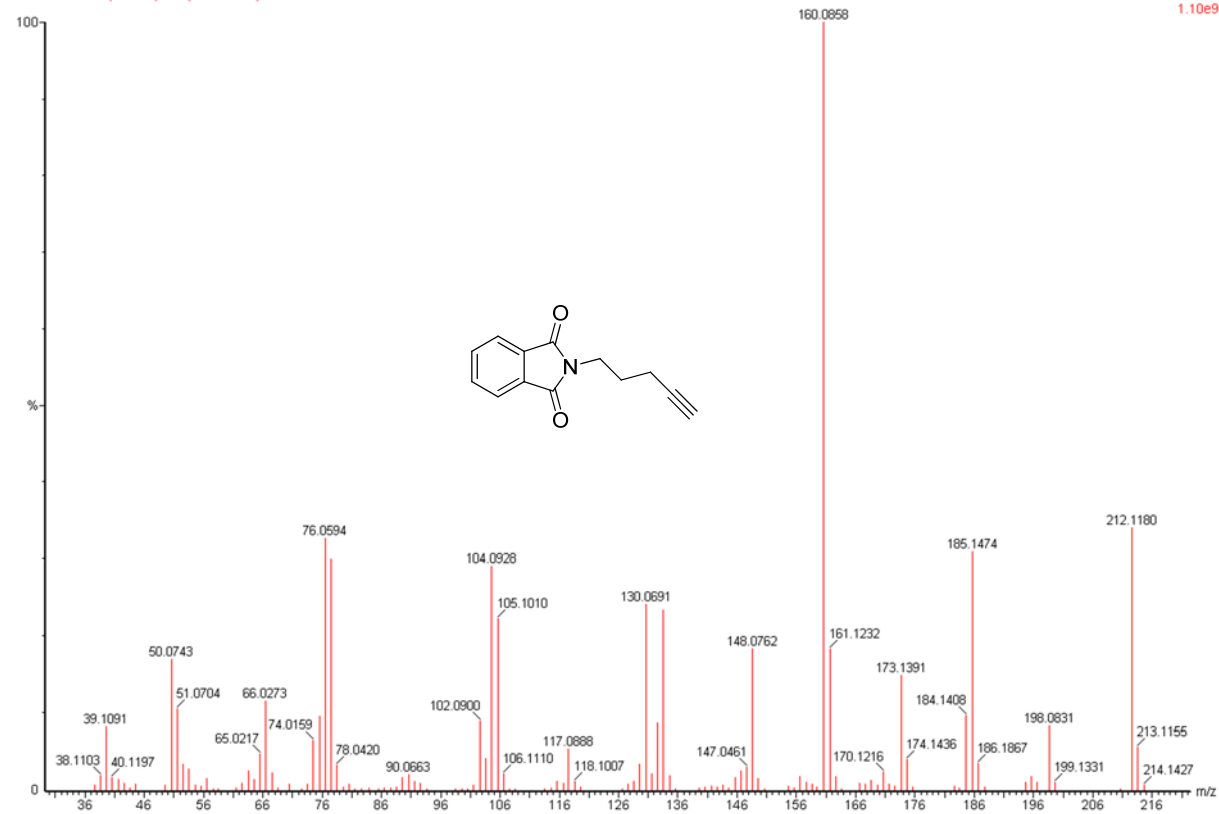
Scan E1+  
TIC  
9.96e9

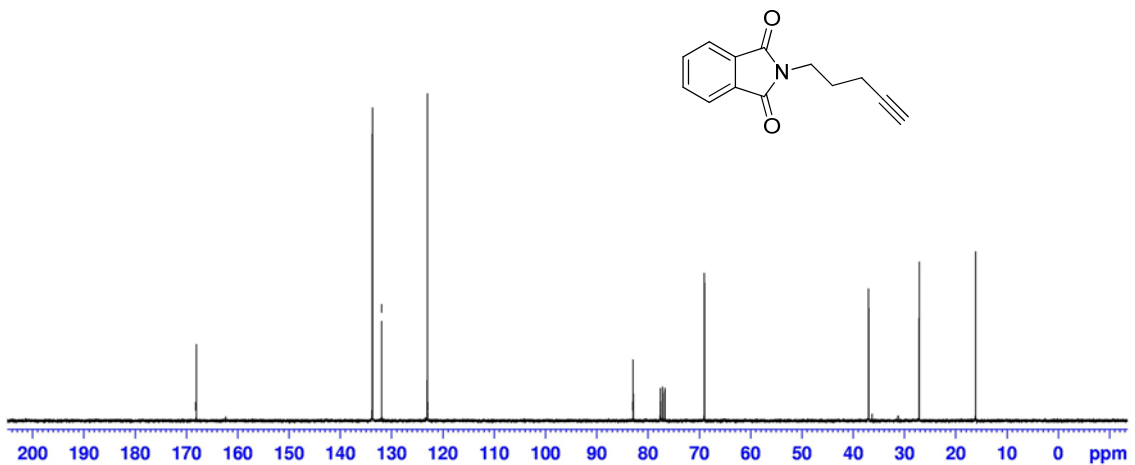
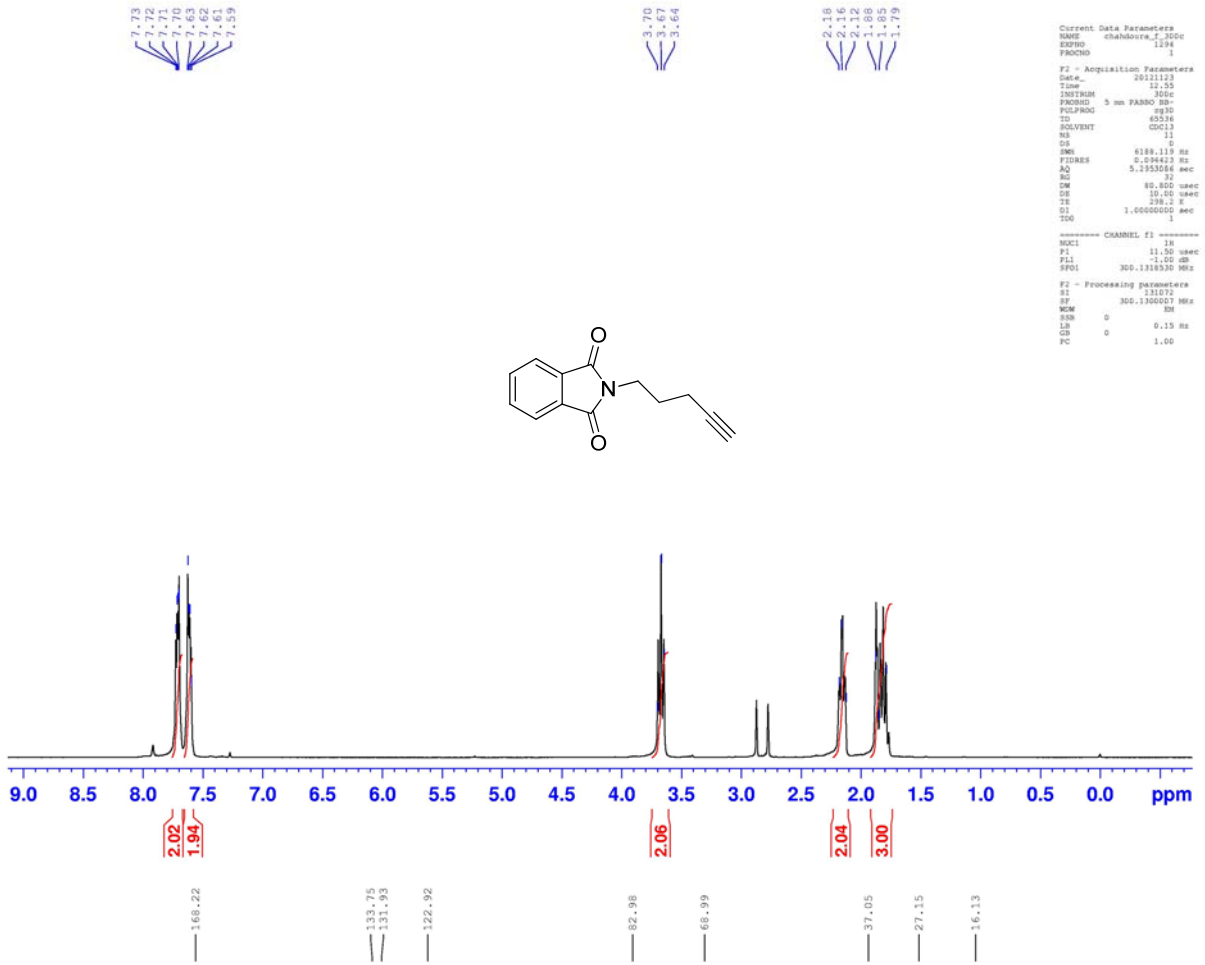


0  
fc-00000- 1634 (12.172) Cm (1623:1638)

, 23-Nov-2012 + 13:16:58

Scan E1+  
1.10e9

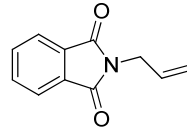
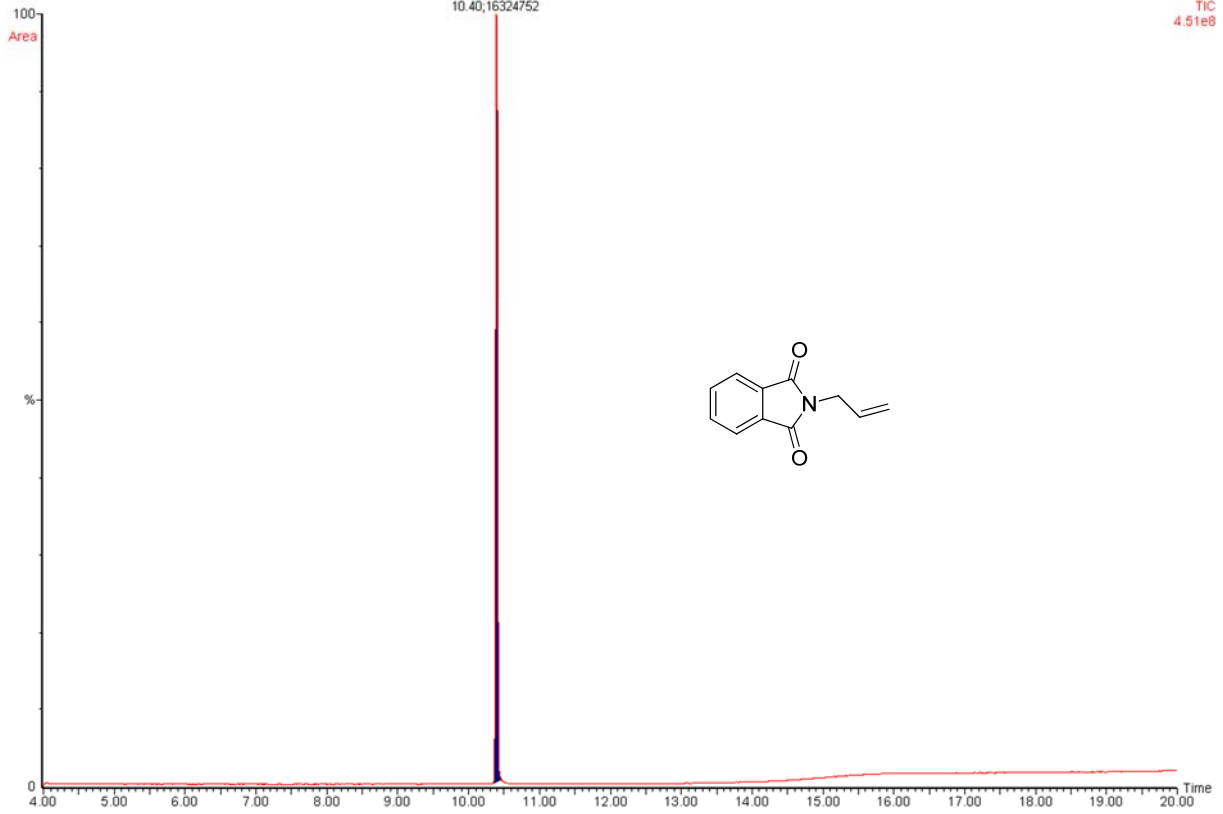




GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **a17**

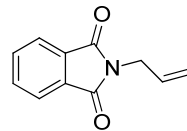
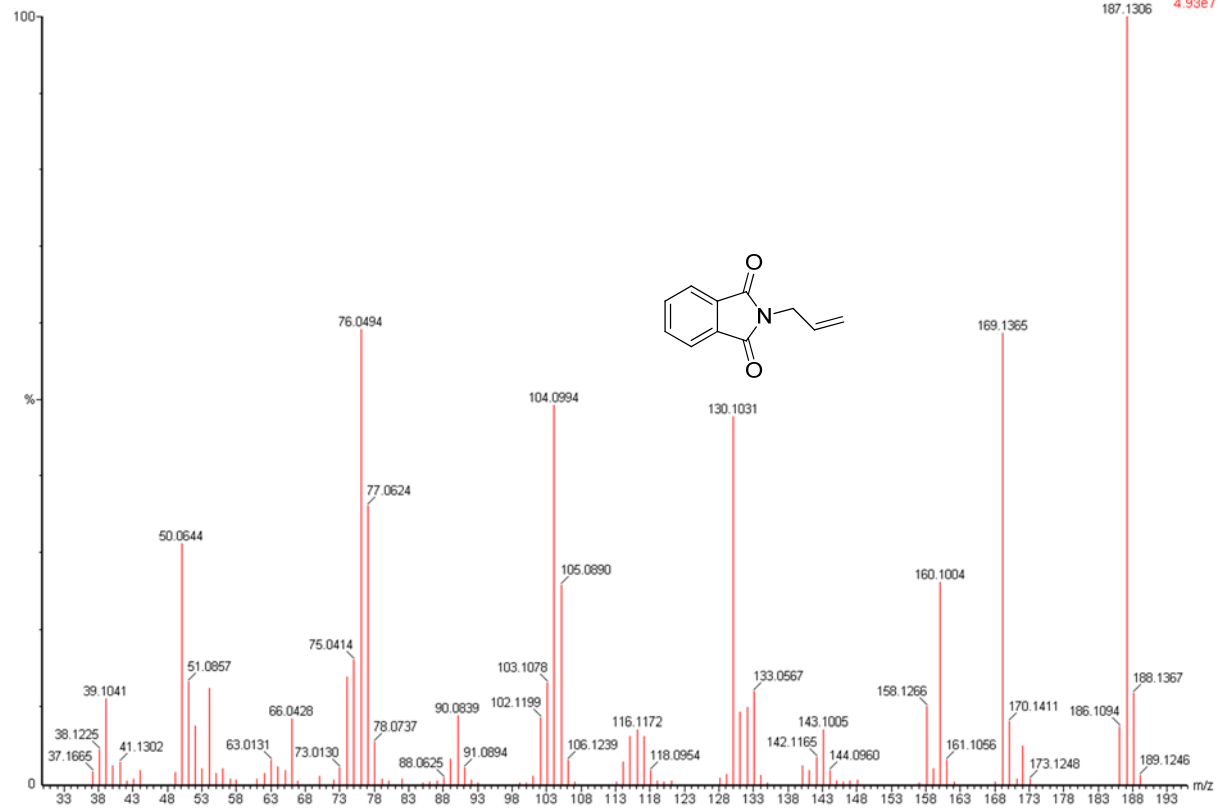
853  
fc-853-2 Sm (Mn, 1x3)

, 17-Sep-2012 + 20:51:12  
Scan EI+  
TIC  
4.51e8

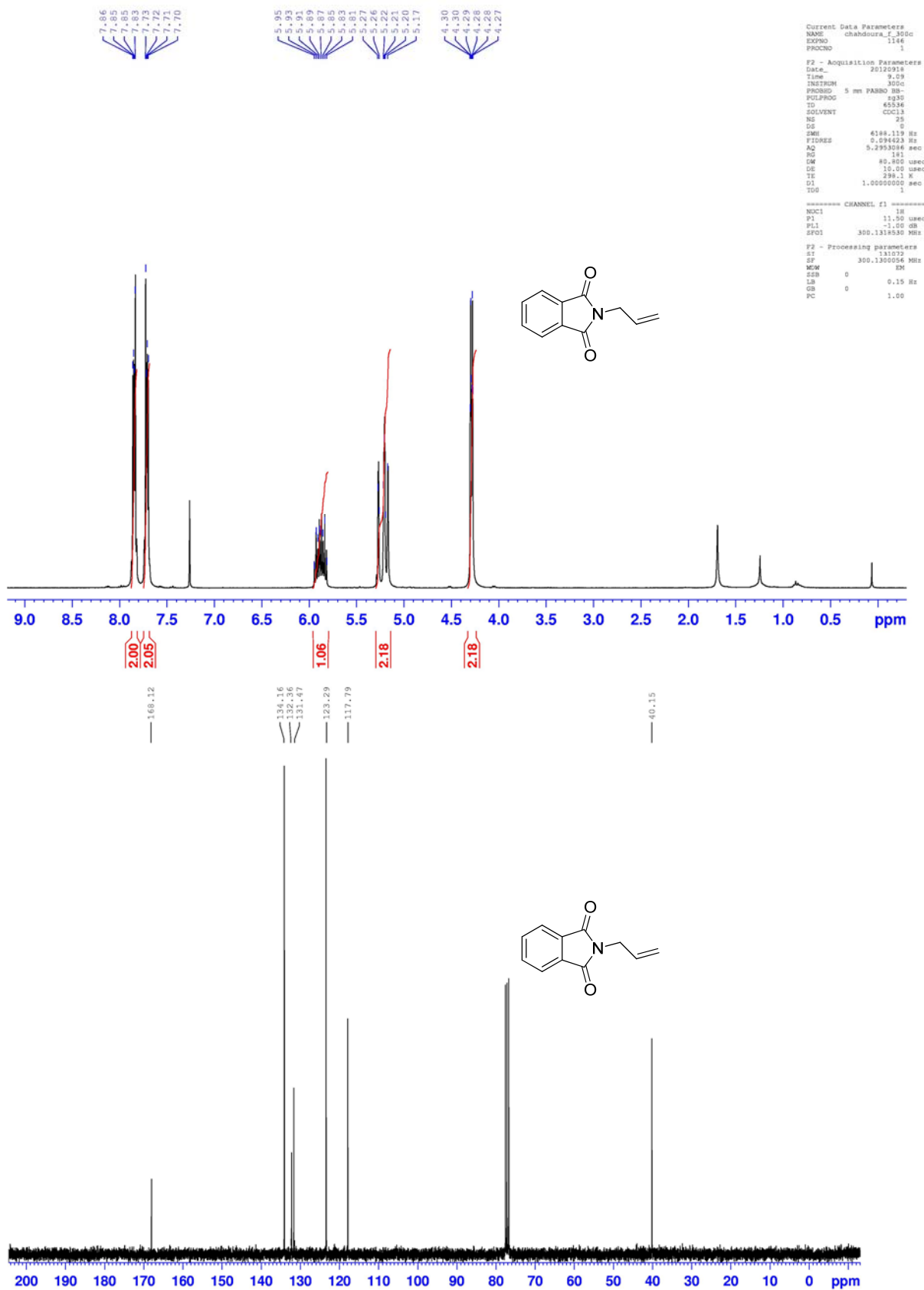


853  
fc-853-2 1280 (10.401) Cm (1275:1283)

, 17-Sep-2012 + 20:51:12  
Scan EI+  
4.93e7



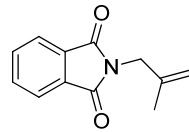
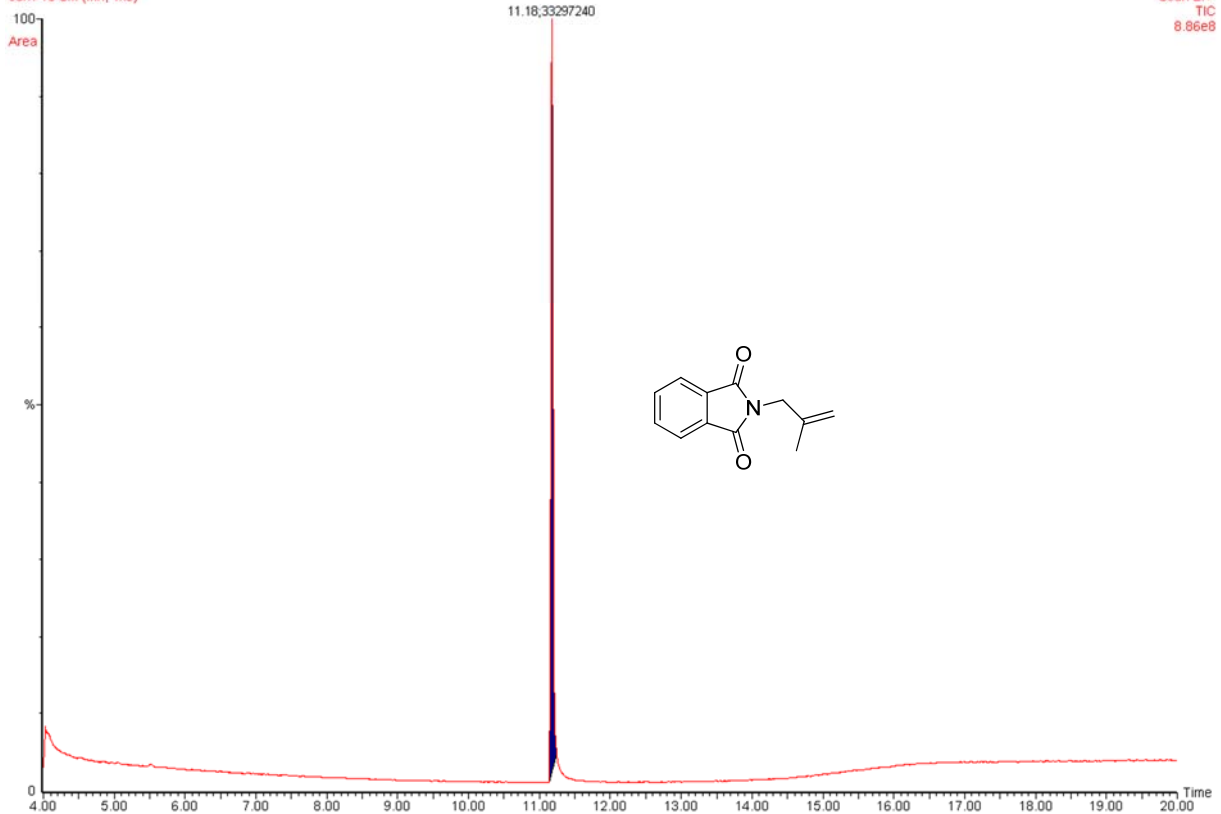




GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **a18**

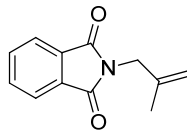
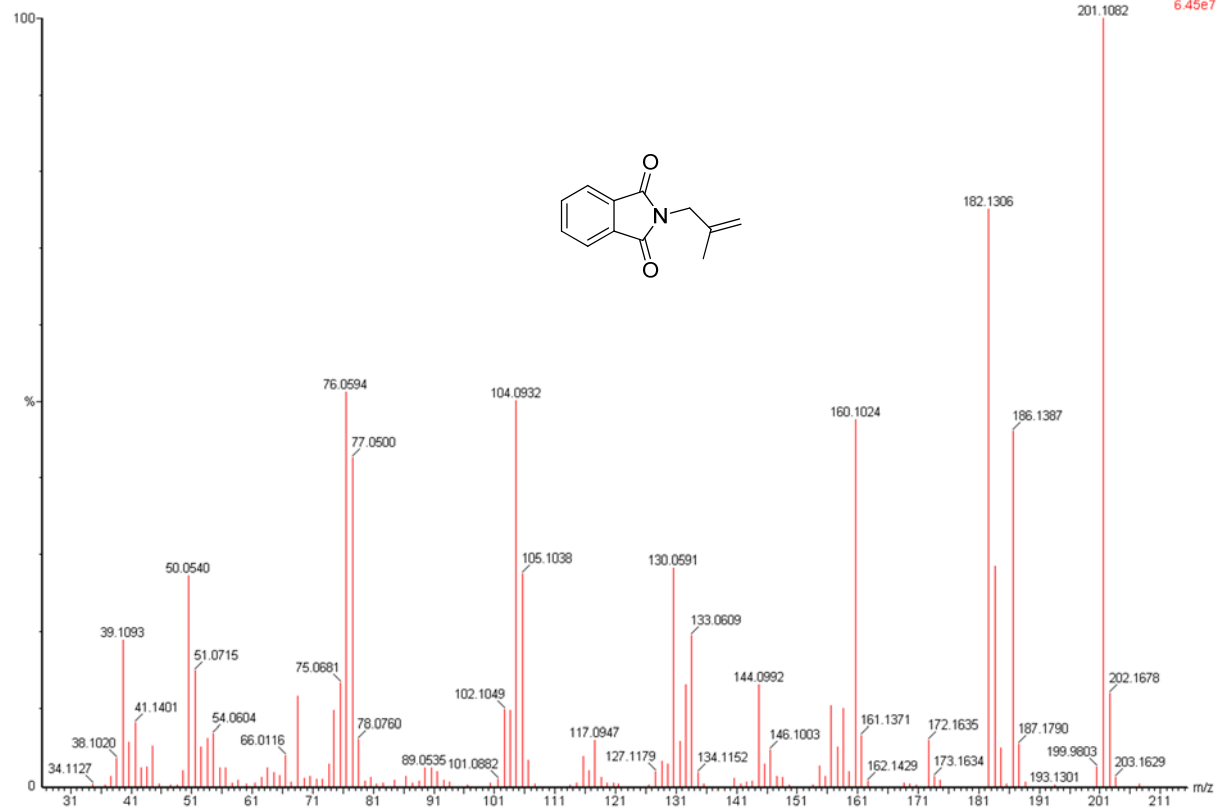
16  
suivi-16 Sm (Mn, 1x3)

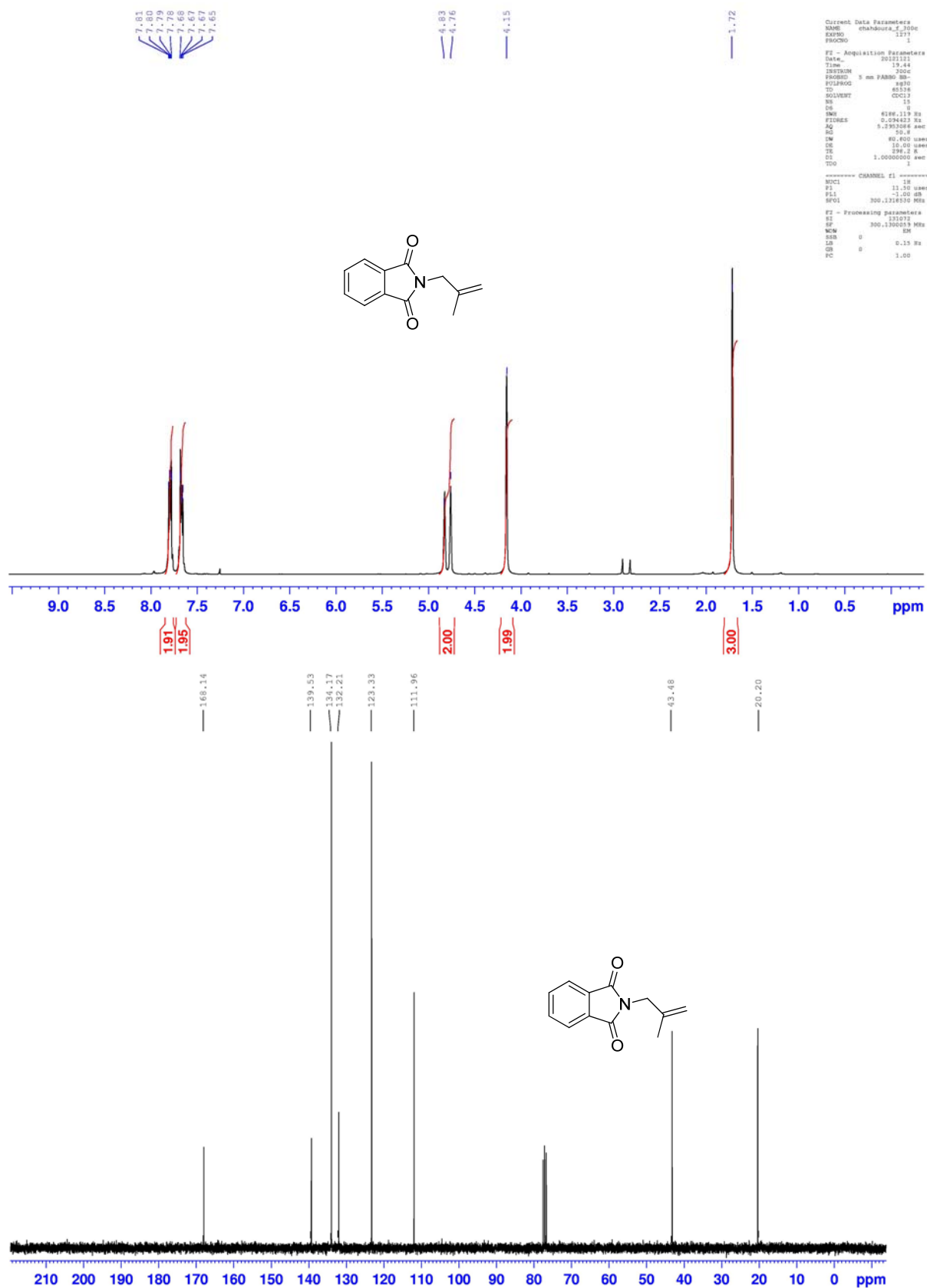
, 21-Nov-2012 + 17:34:05  
Scan E1+  
TIC  
8.86e8



16  
suivi-16 1436 (11.181) Cm (1433:1444)

, 21-Nov-2012 + 17:34:05  
Scan E1+  
6.45e7



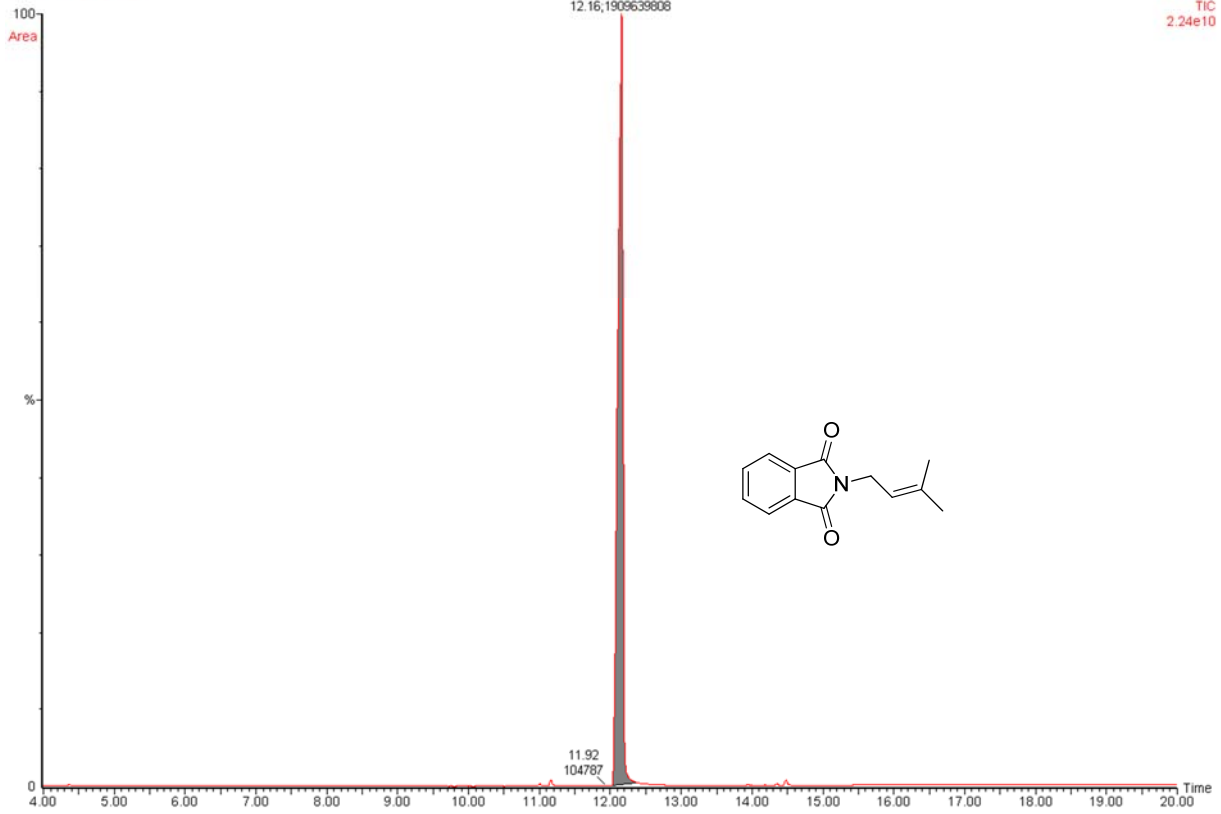


GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **a19**

1  
fc-0000 Sm (Mn, 1x3)

, 22-Nov-2012 + 11:52:40

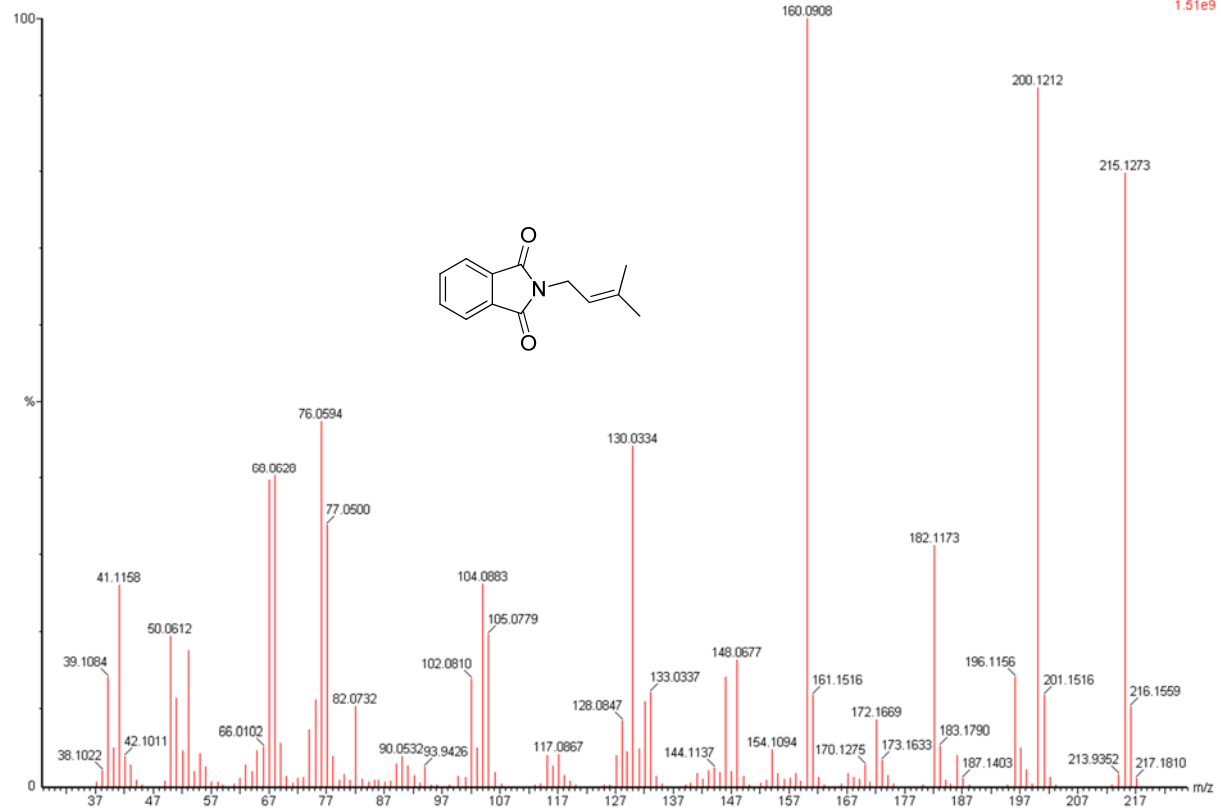
Scan EI+  
TIC  
2.24e10

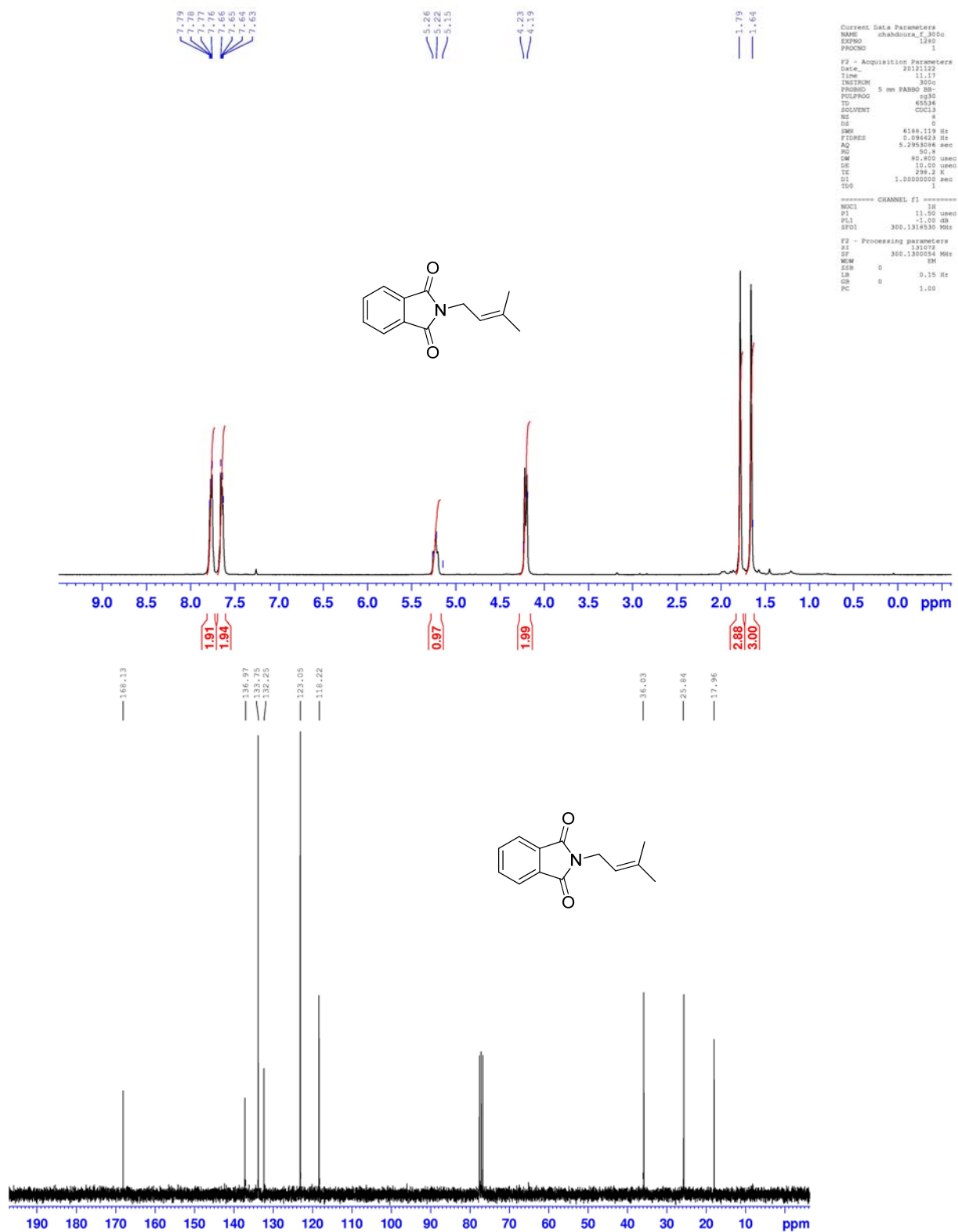


1  
fc-0000 1632 (12.162) Cm (1615:1632)

, 22-Nov-2012 + 11:52:40

Scan EI+  
1.51e9

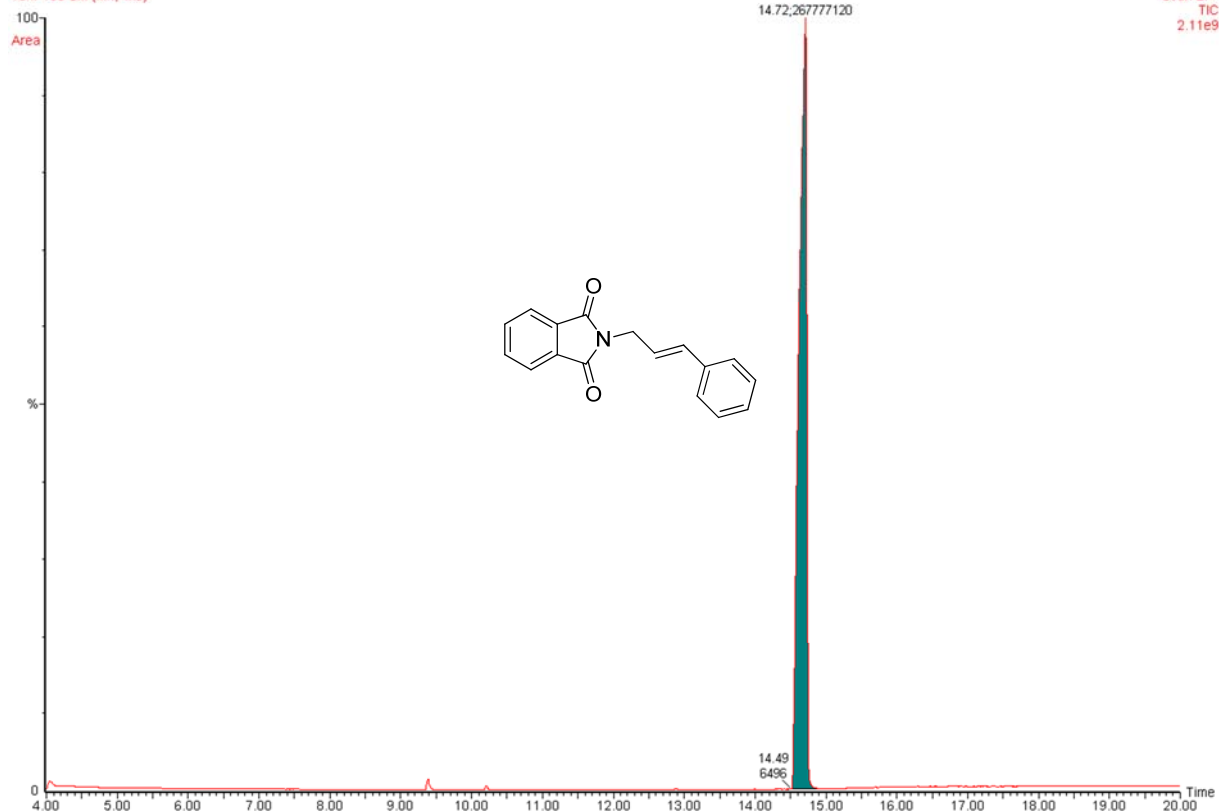




GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **a20**

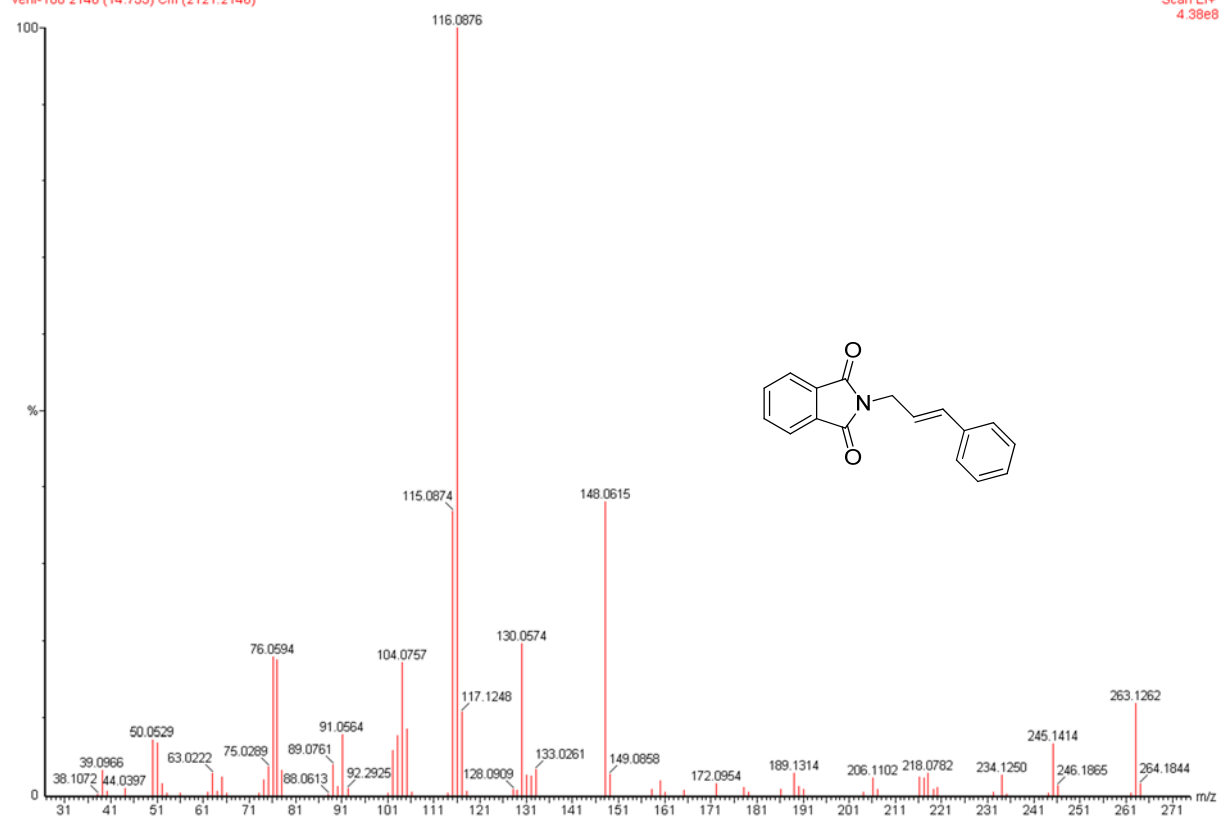
V  
verif-100 Sm (Mn, 1x3)

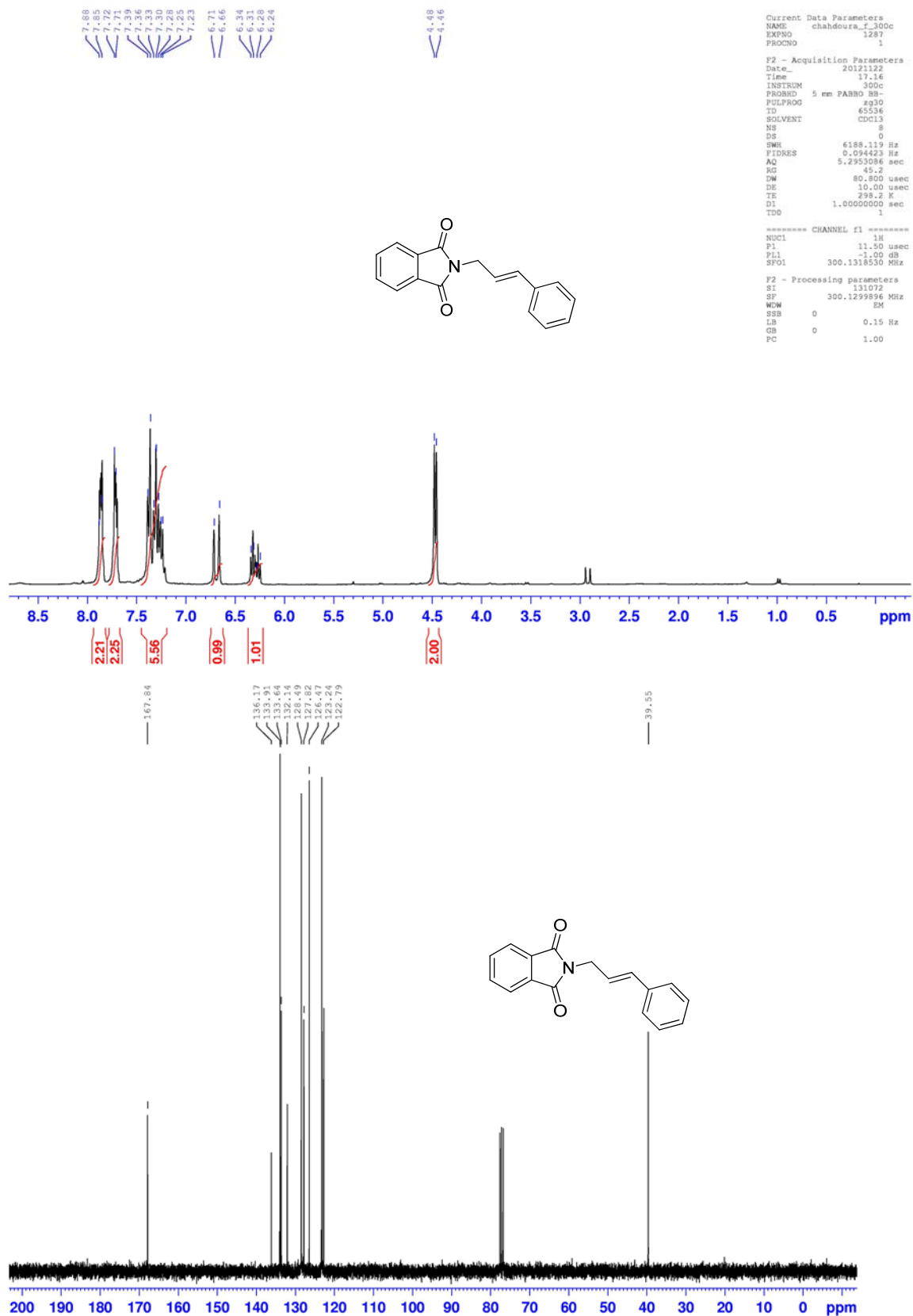
, 23-Nov-2012 + 17:48:47



V  
verif-100 2146 (14.733) Cm (2121:2146)

, 23-Nov-2012 + 17:48:47

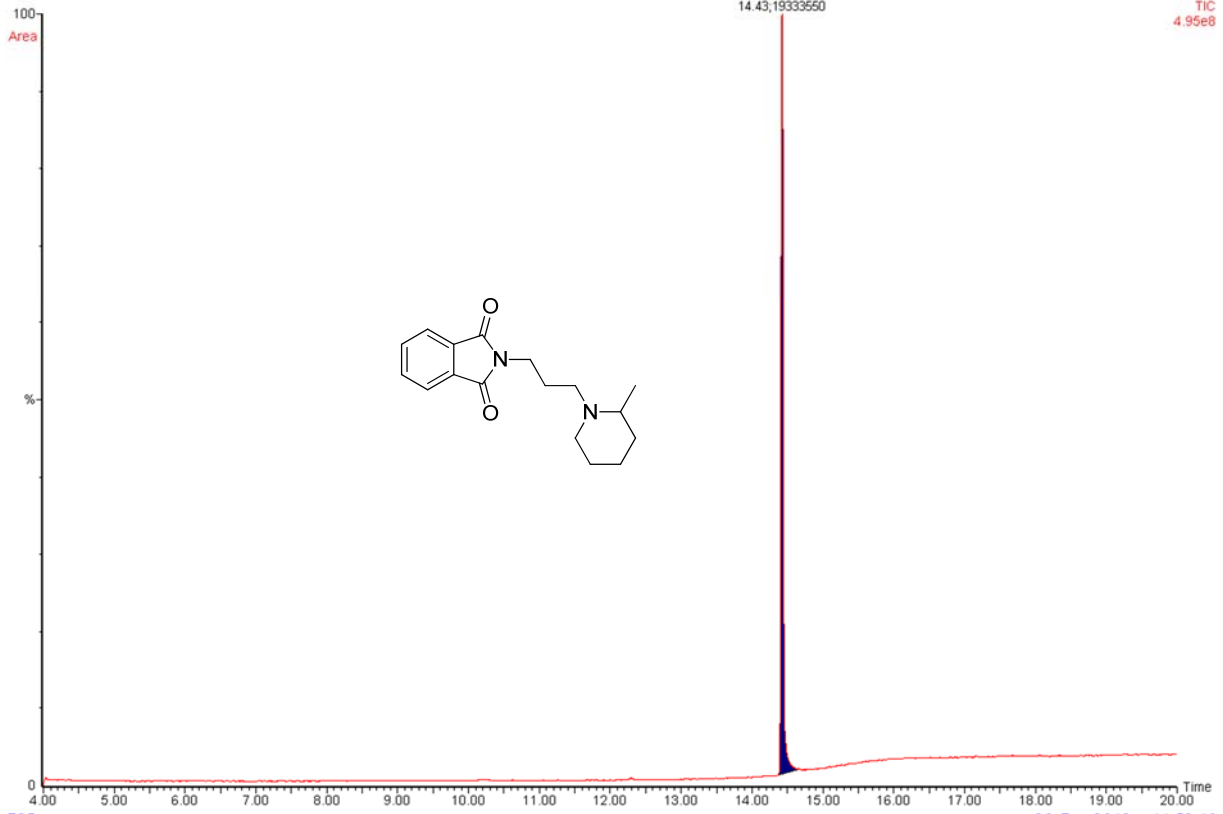




GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **a21**

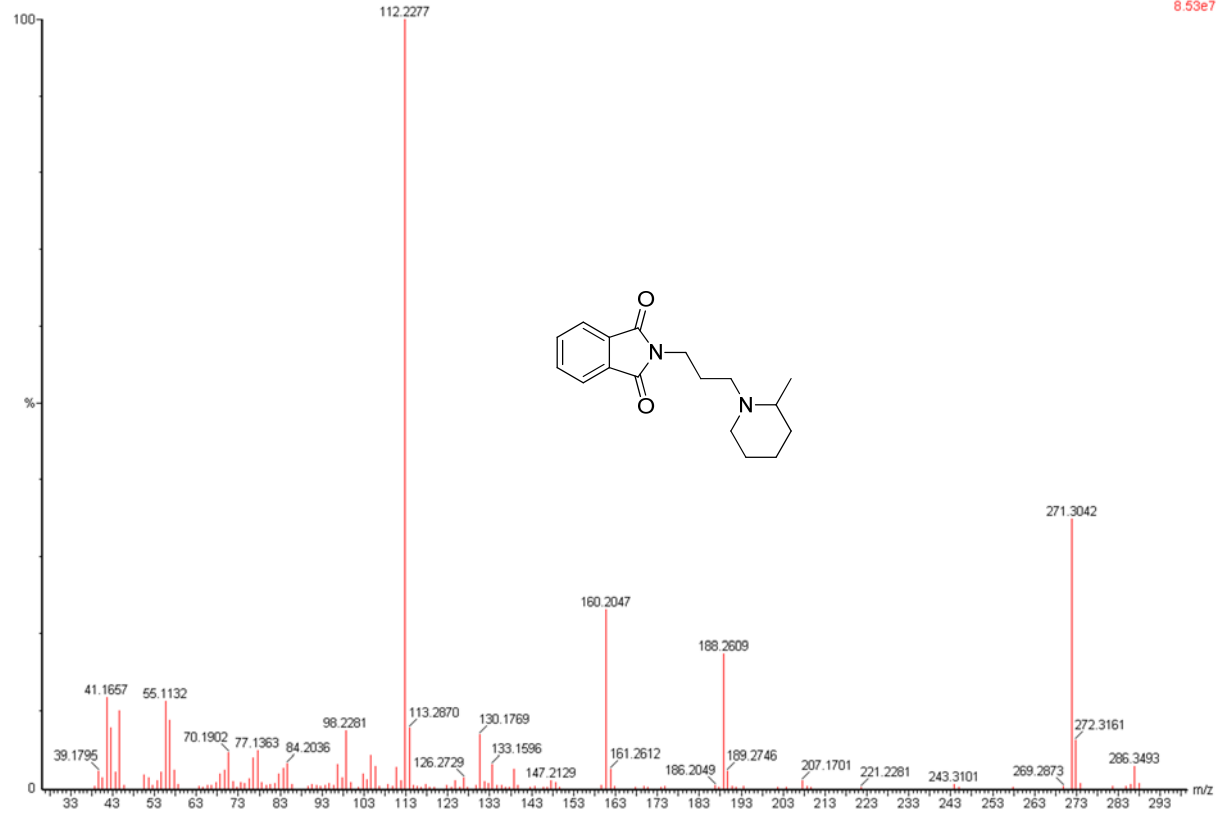
785  
fc-785-1 Sm (Mn, 1x3)

, 20-Jun-2012 + 11:52:19  
Scan EI+  
TIC  
4.95e8

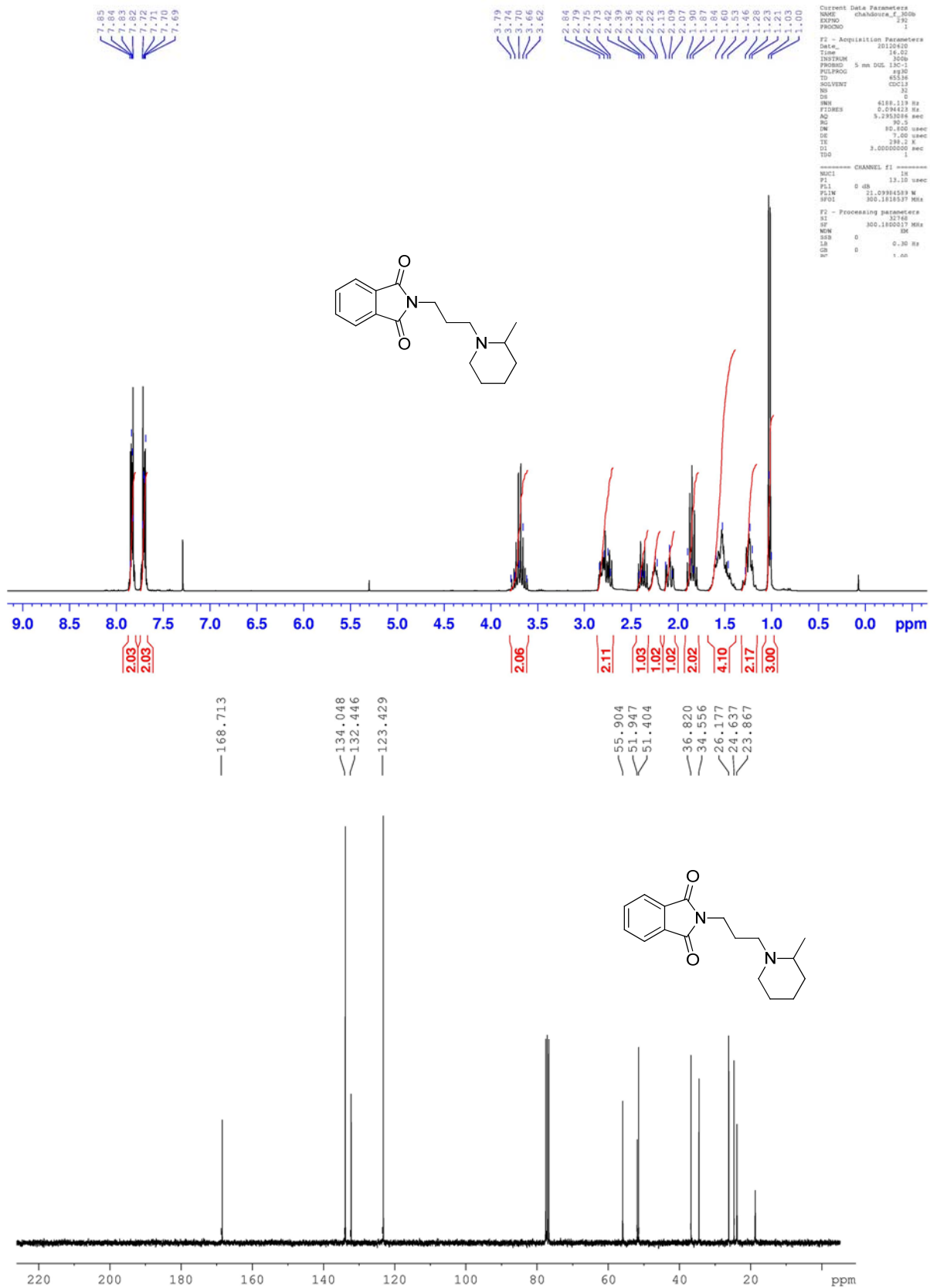


785  
fc-785-1 2085 (14.428) Cm (2083:2094)

, 20-Jun-2012 + 11:52:19  
Scan EI+  
8.53e7



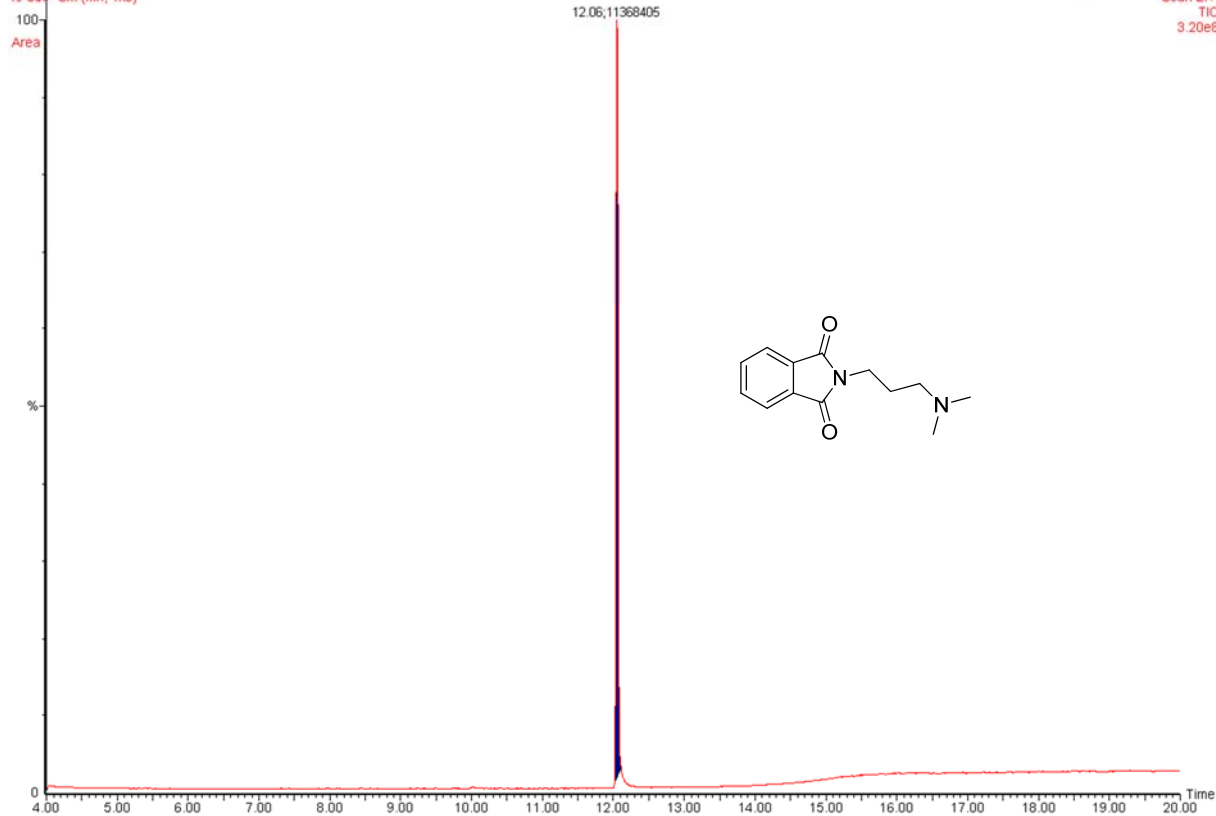




GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **a22**

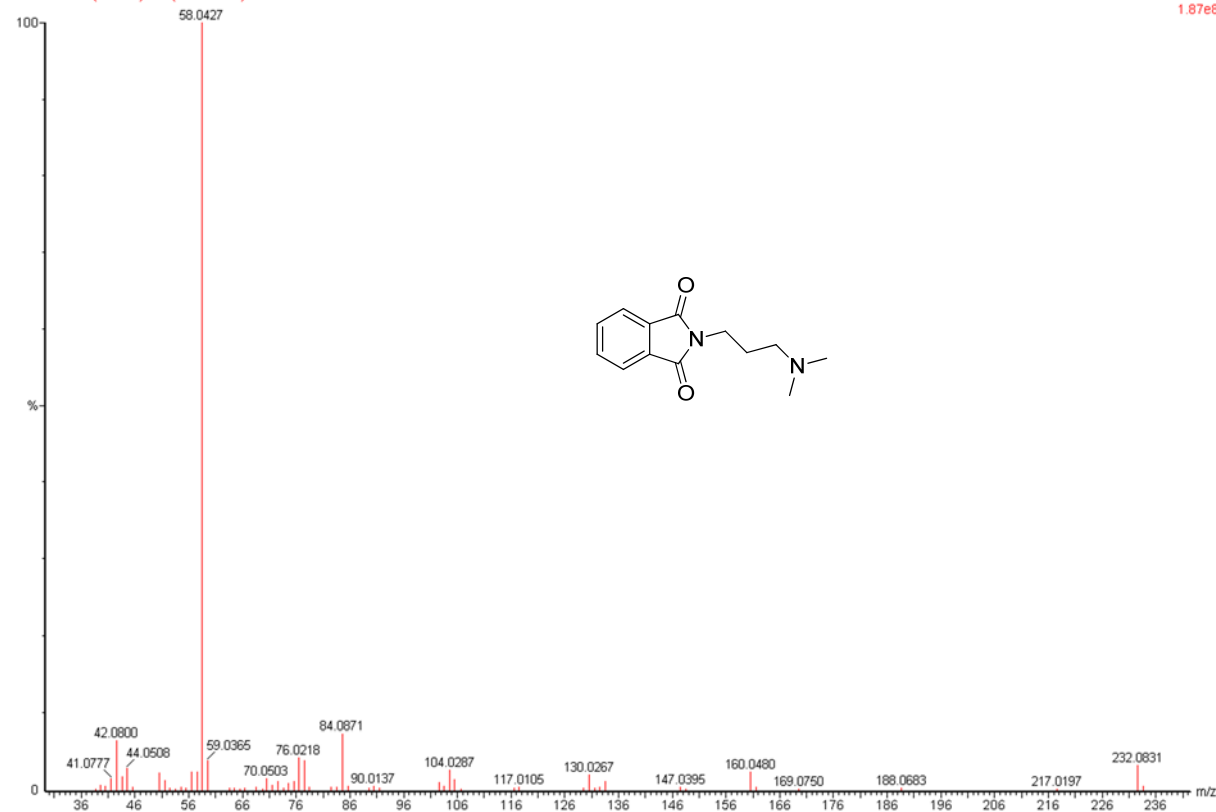
859  
tc-859 Sm (Mn, 1x3)  
Area

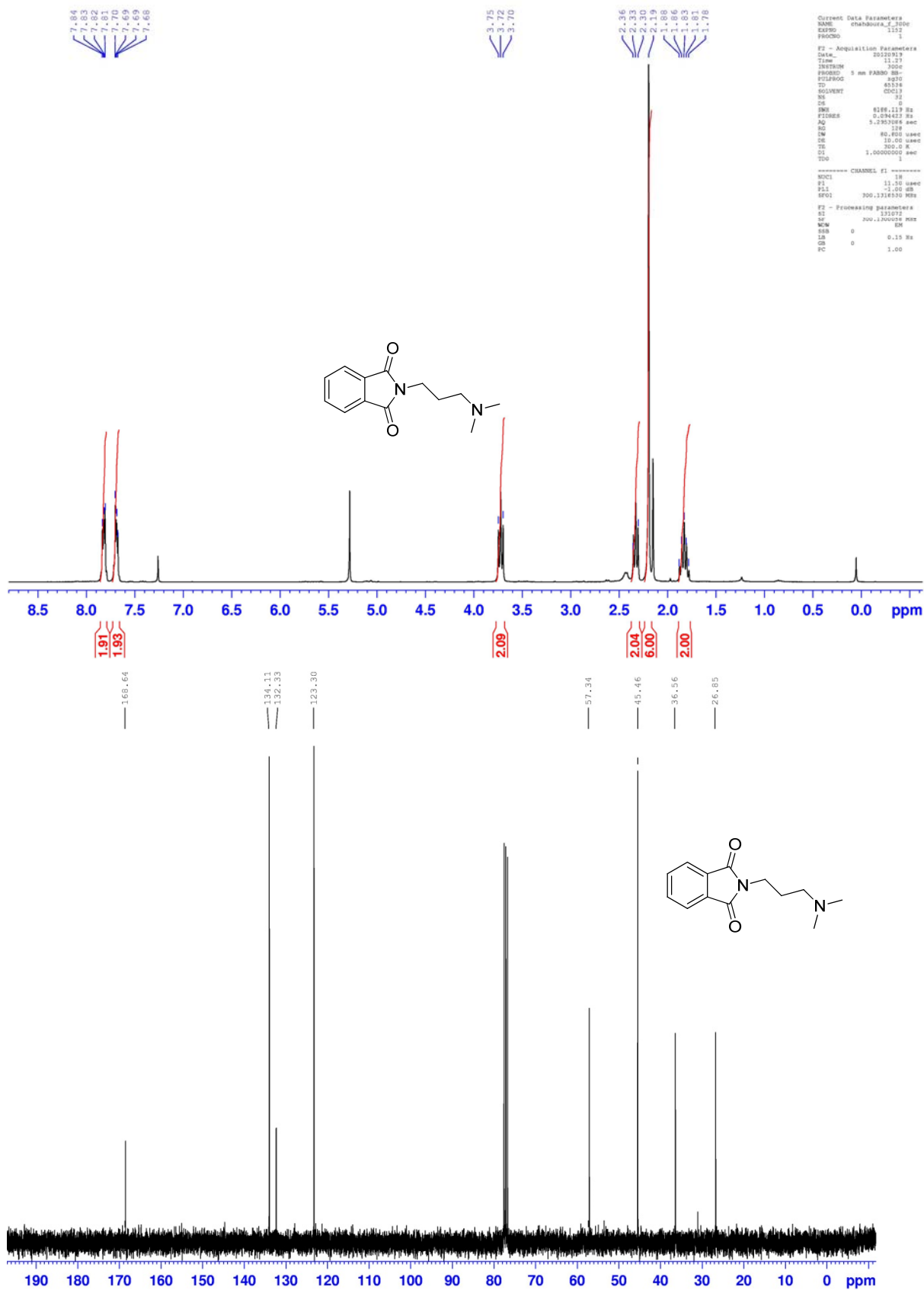
, 19-Sep-2012 + 10:38:22  
Scan EI+  
TIC  
3.20e8



859  
tc-859- 1611 (12.057) Cm (1610:1615)

, 19-Sep-2012 + 10:38:22  
Scan EI+  
1.87e8

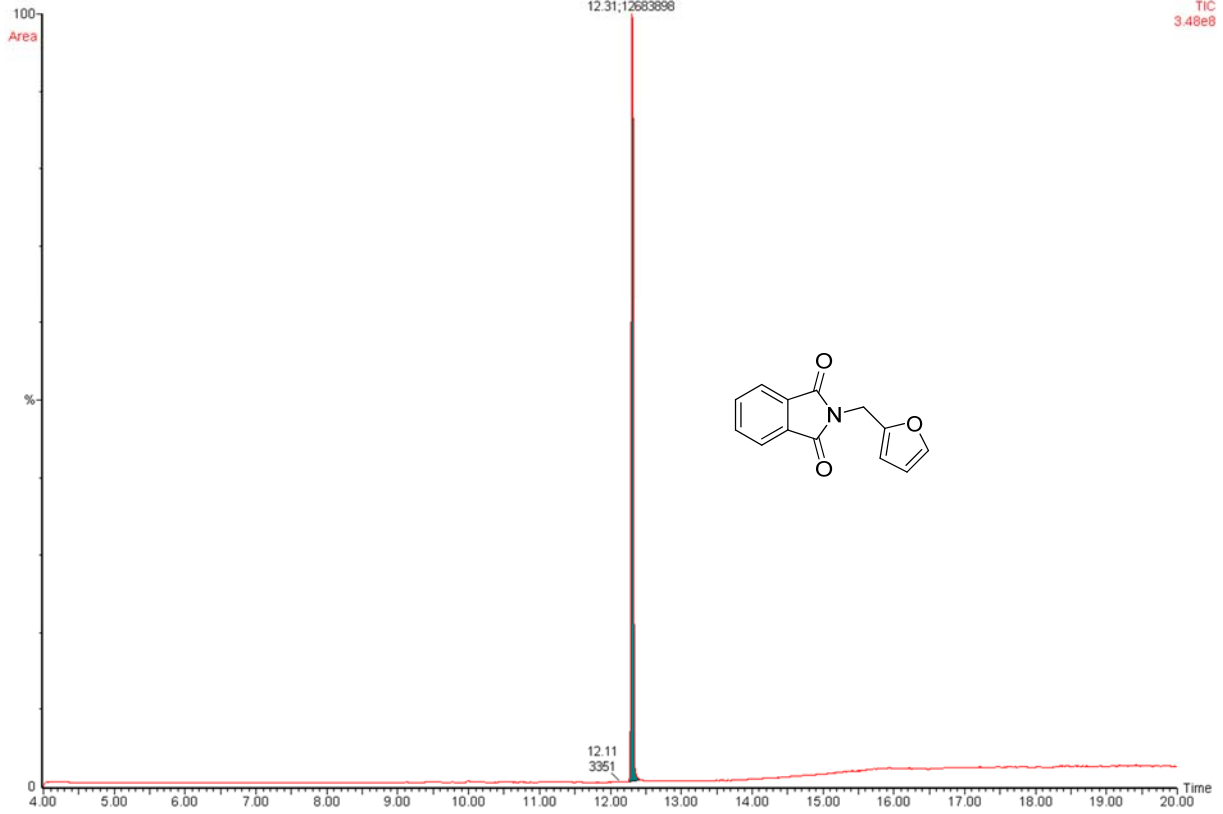




GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **a23**

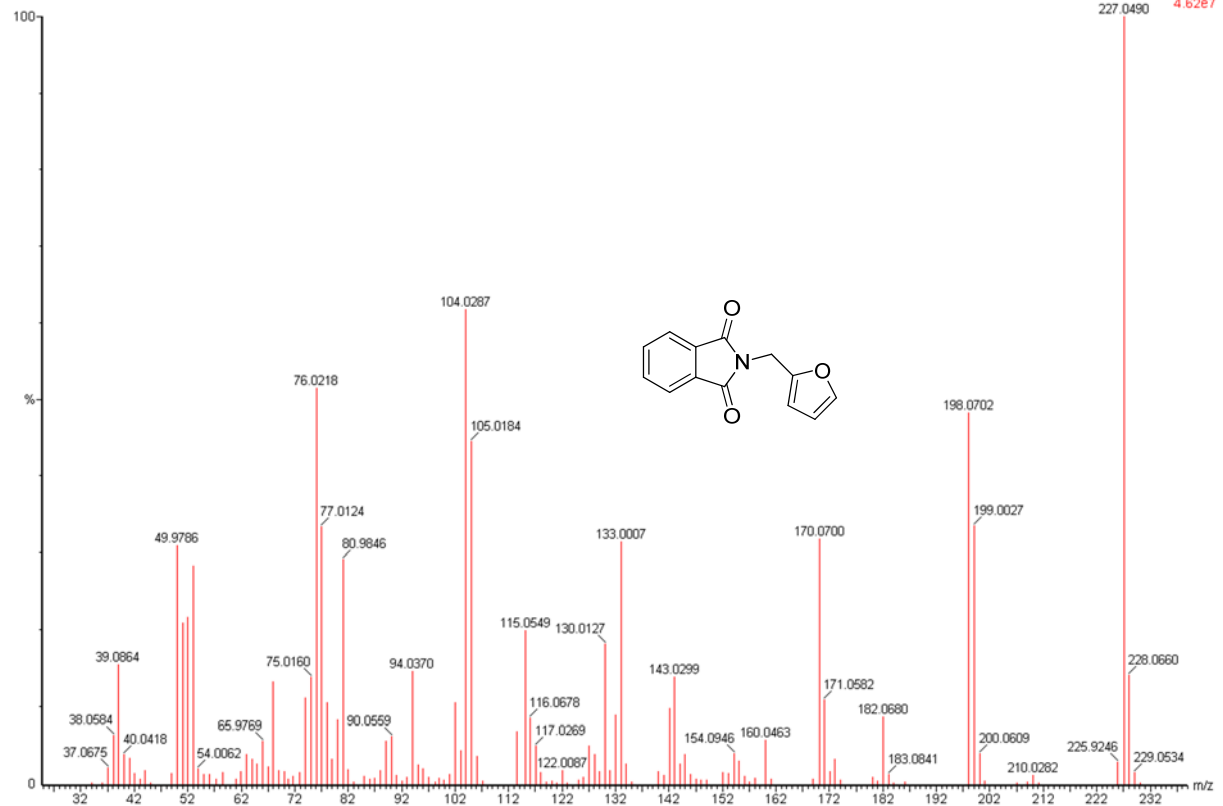
864  
tc-864- Sm (Mn, 1x3)

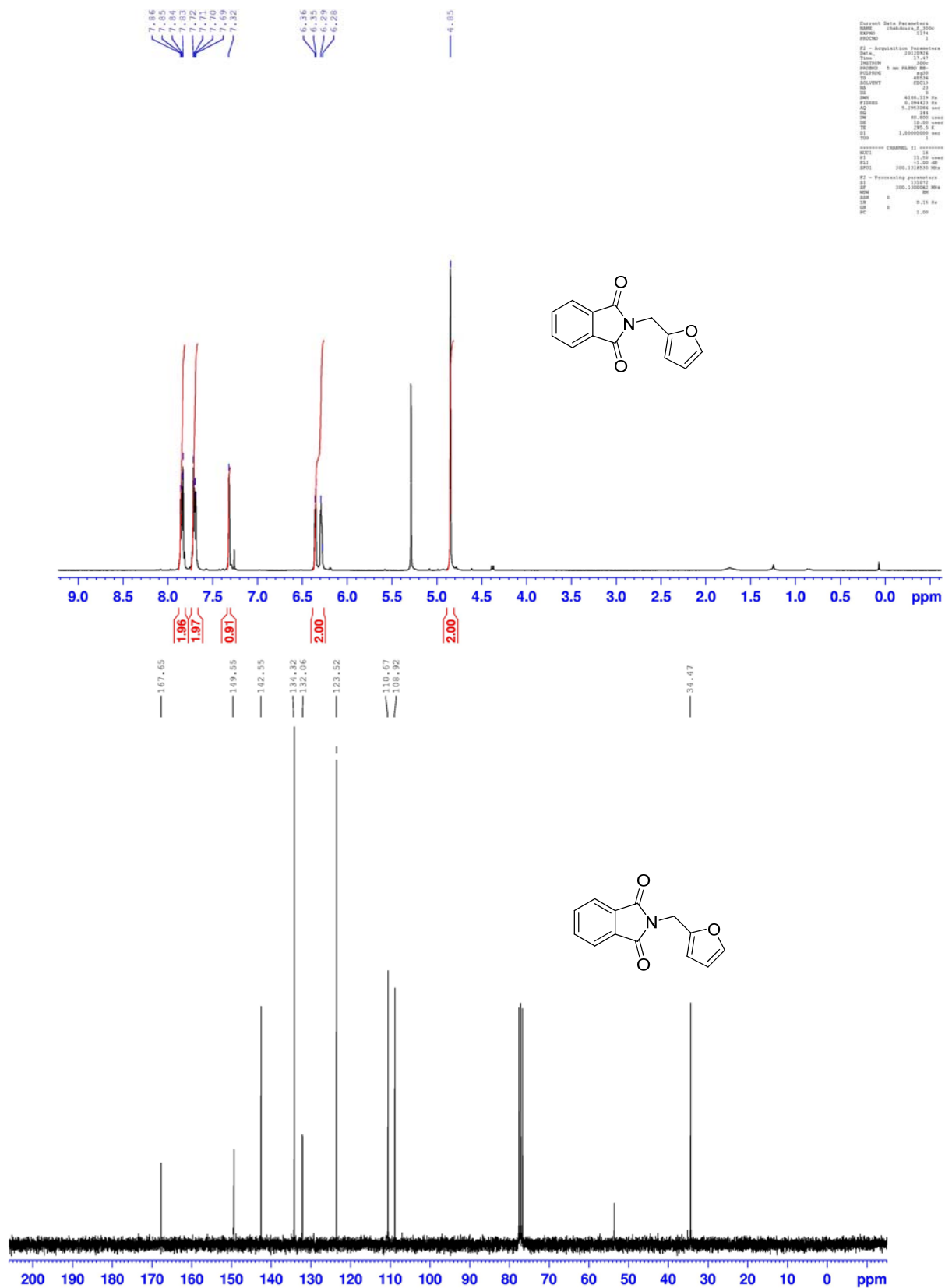
, 26-Sep-2012 + 17:16:26  
Scan EI+  
TIC  
3.48e8



864  
tc-864- 1663 (12.317) Cm (1660:1664)

, 26-Sep-2012 + 17:16:26  
Scan EI+  
4.62e7

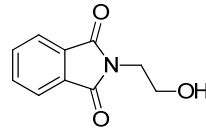
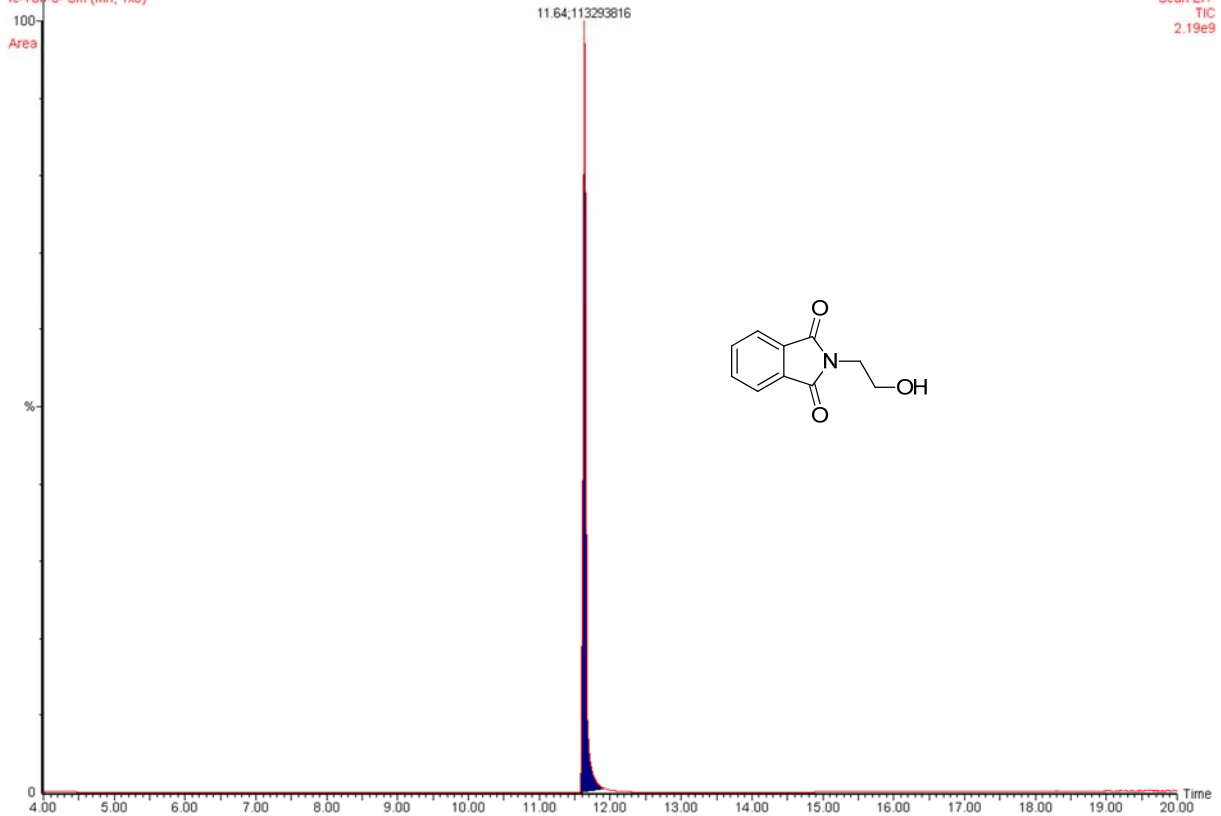




GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **a24**

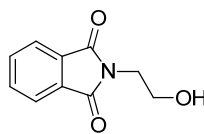
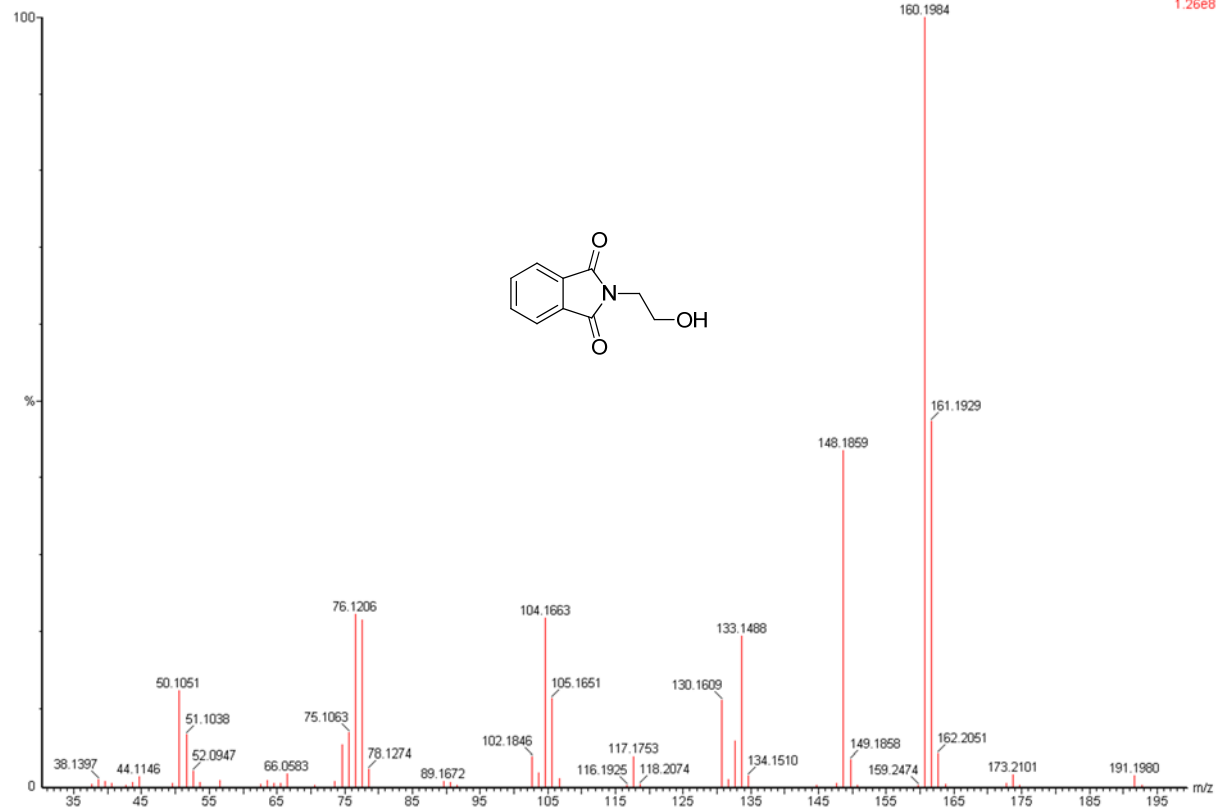
760  
fc-760-3- Sm (Mn, 1x3)  
Area

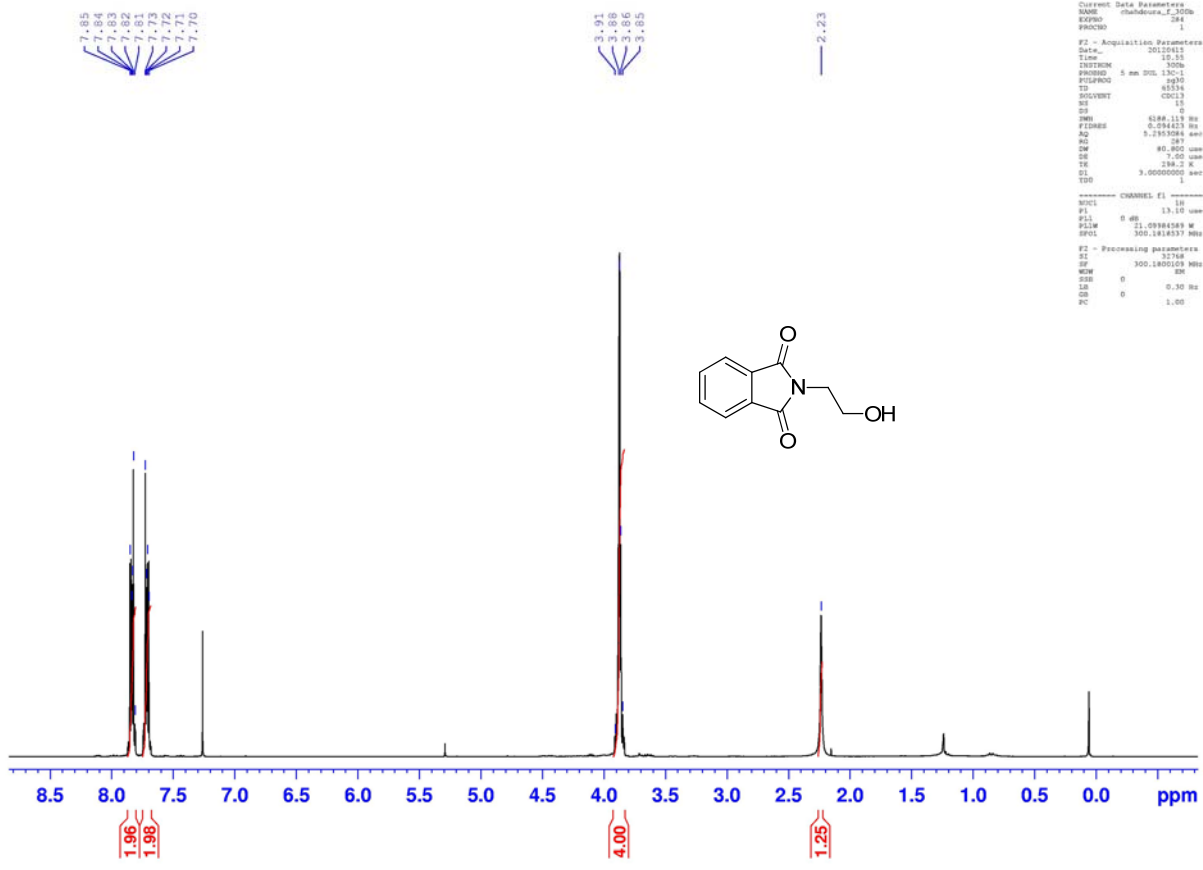
, 15-Jun-2012 + 18:02:53  
Scan E1+  
TIC  
2.19e9



760  
fc-760-3- 1529 (11.646) Cm (1521:1568)

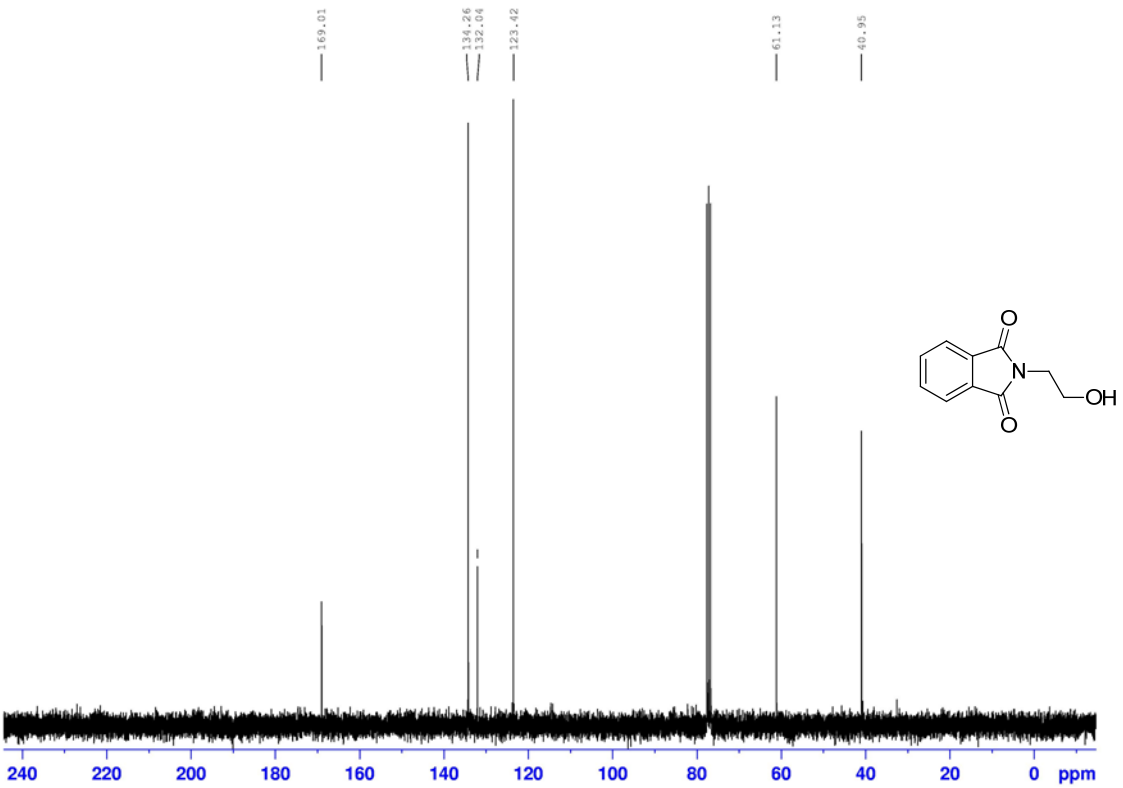
, 15-Jun-2012 + 18:02:53  
Scan E1+  
1.26e8





```

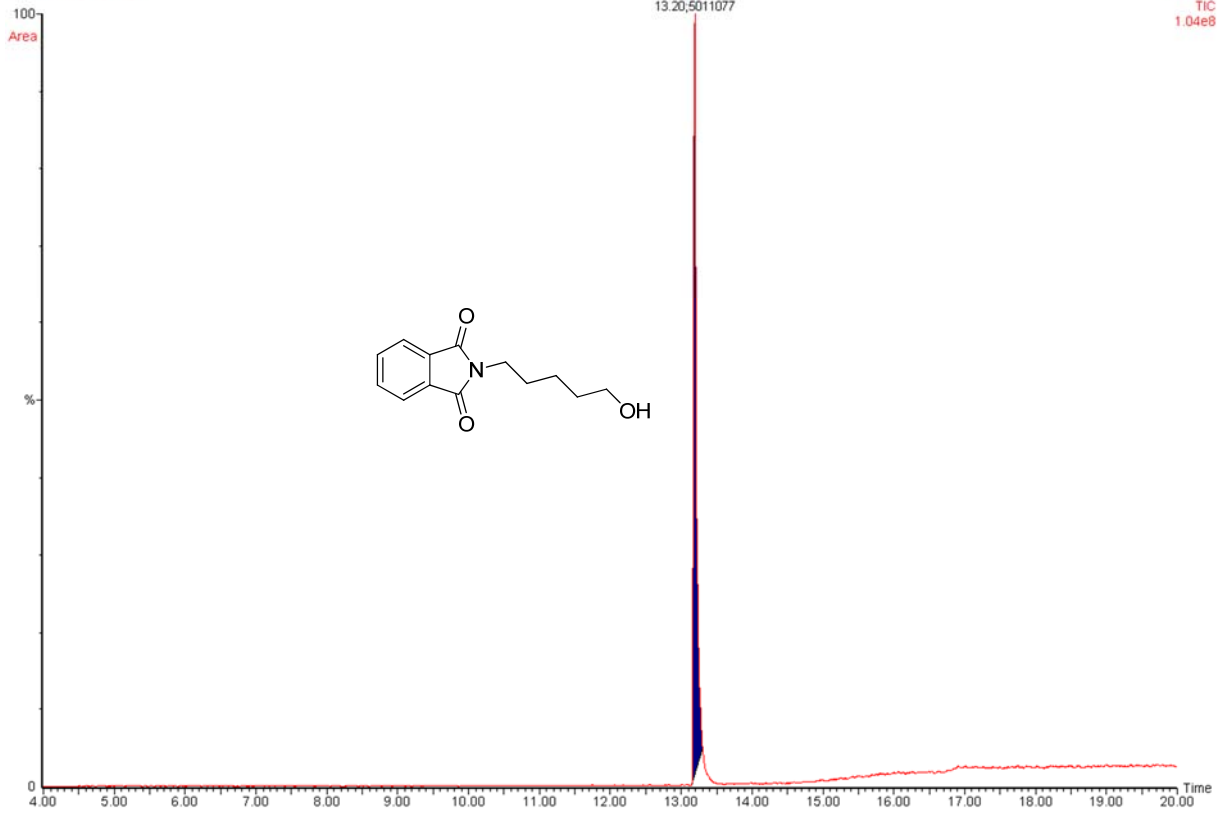
Current Data Parameters
NAME: chmdeura_f_300
EXPNO: 284
PROCNO: 1
F2 - Acquisition Parameters
Date_: 20120415
Time: 10:55
INSTRUM: spect
PROBHD: 5 mm BBO 13C-1
PULPROG: zgpg30
TD: 65536
SOLVENT: CDCl3
SI: 32
SF: 300.1360950 MHz
AQ: 0.0942338 sec
RG: 327.500
AQ: 5.2502084 sec
RG: 327.500
SFO: 300.1360950 MHz
WDW: EM
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00
  
```



GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **a25**

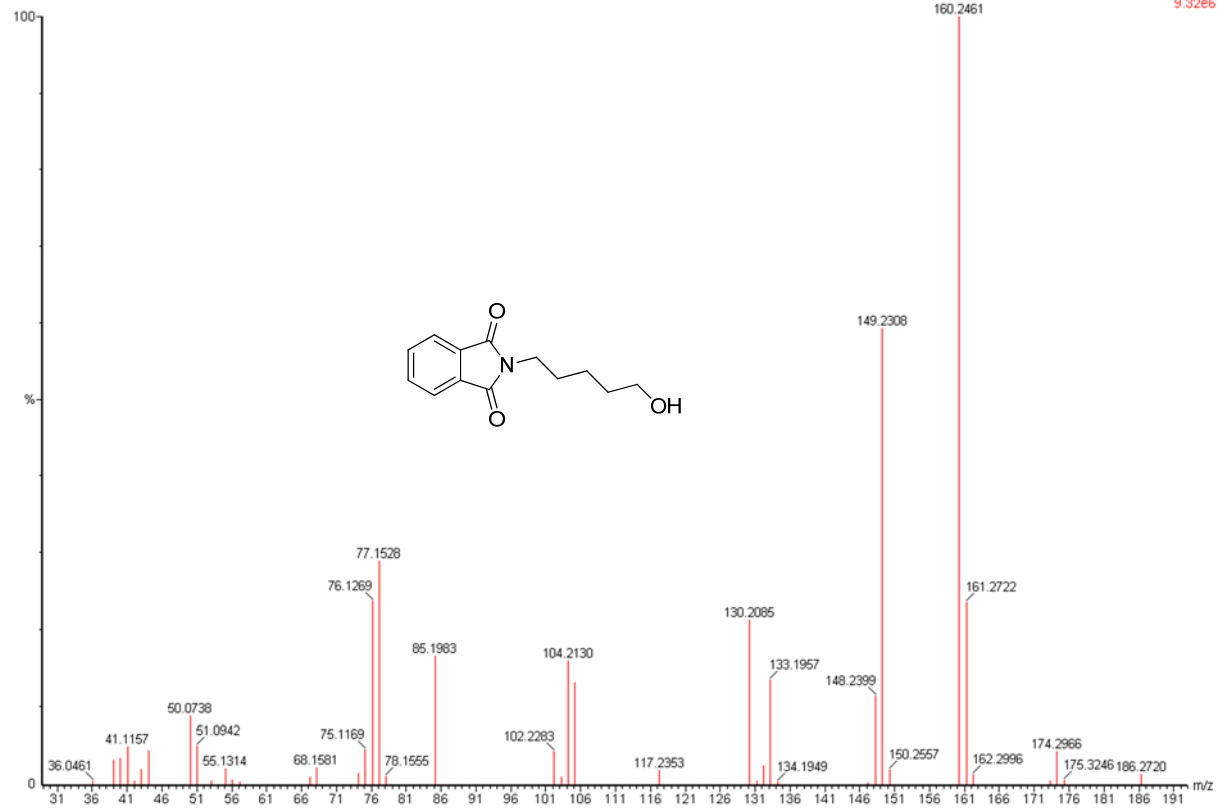
829  
fc-829- Sm (Mn, 1x3)

, 17-Jul-2012 + 17:28:14  
Scan EI+  
TIC  
1.04e8

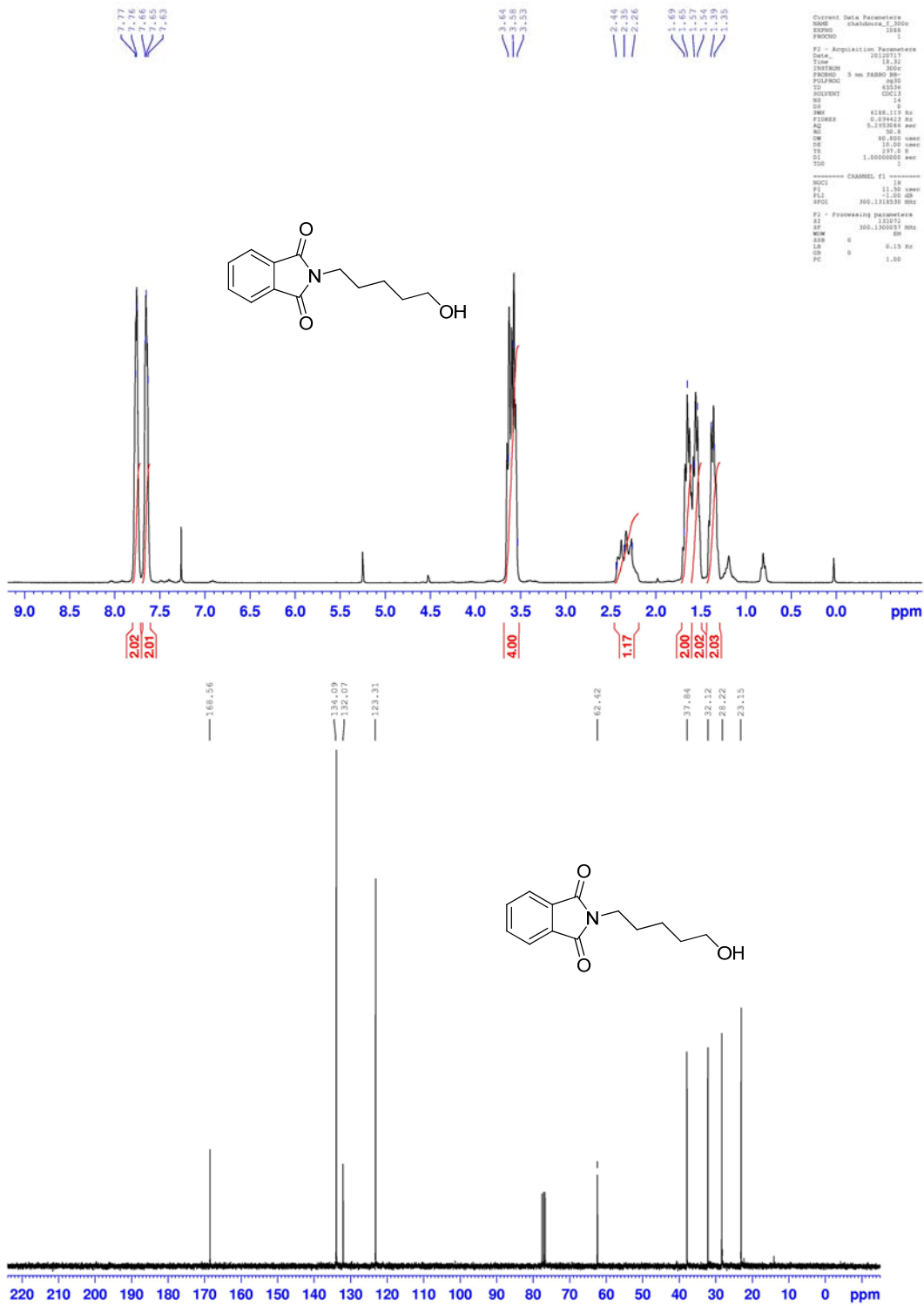


829  
fc-829- 1838 (13.192) Cm (1835:1864)

, 17-Jul-2012 + 17:28:14  
Scan EI+  
9.32e6



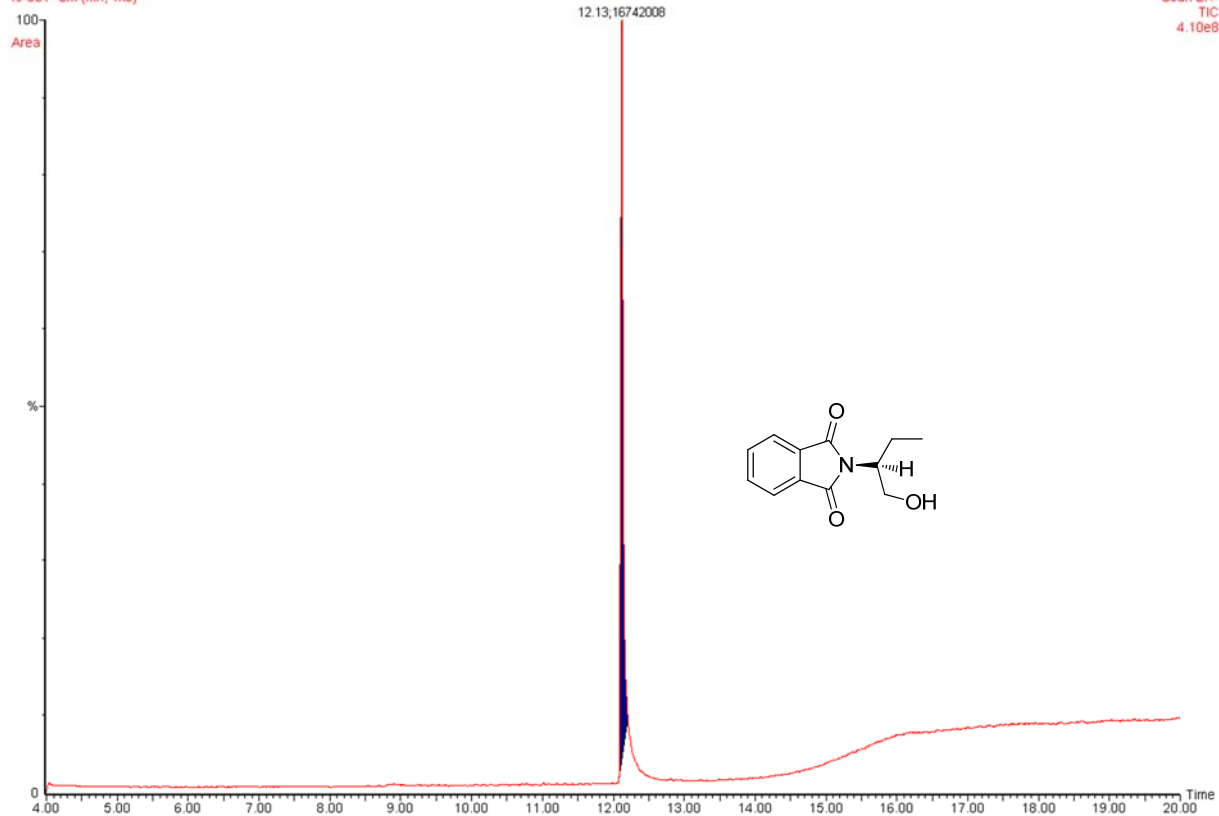




GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **a26**

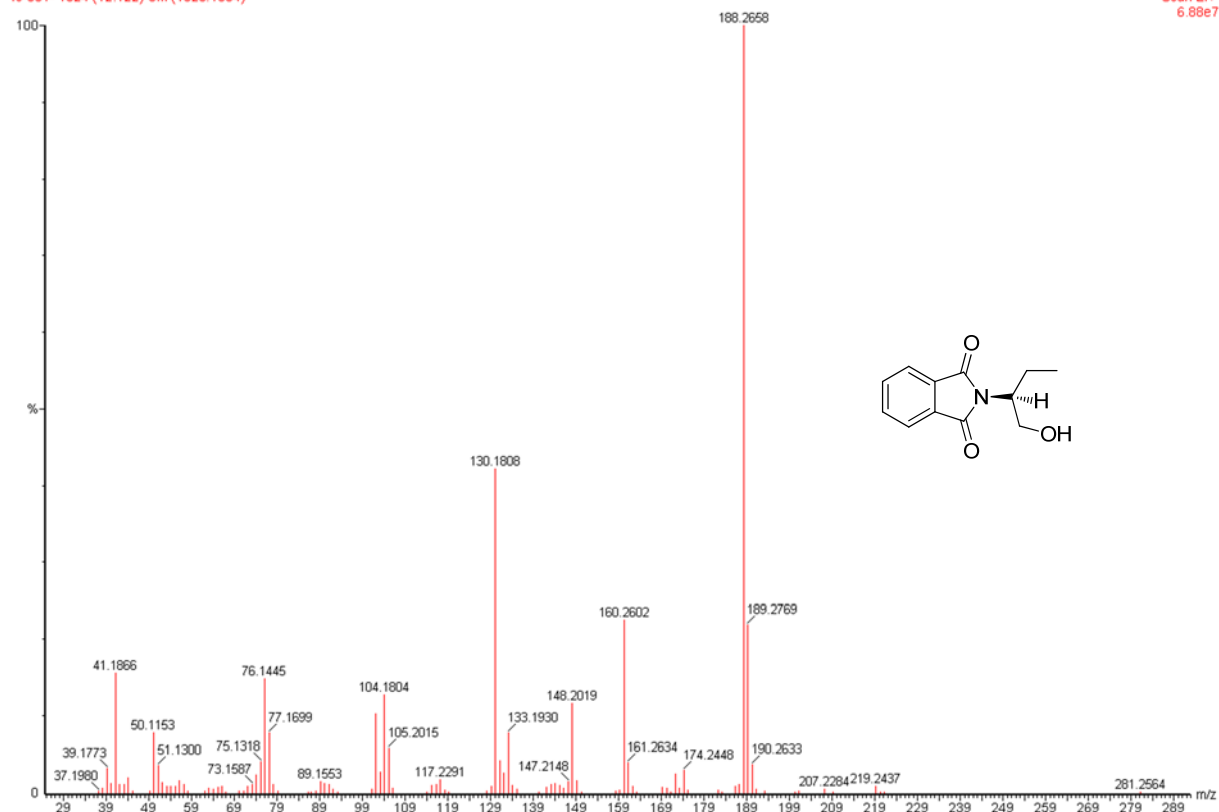
801  
fc-801- Sm (Mn, 1x3)

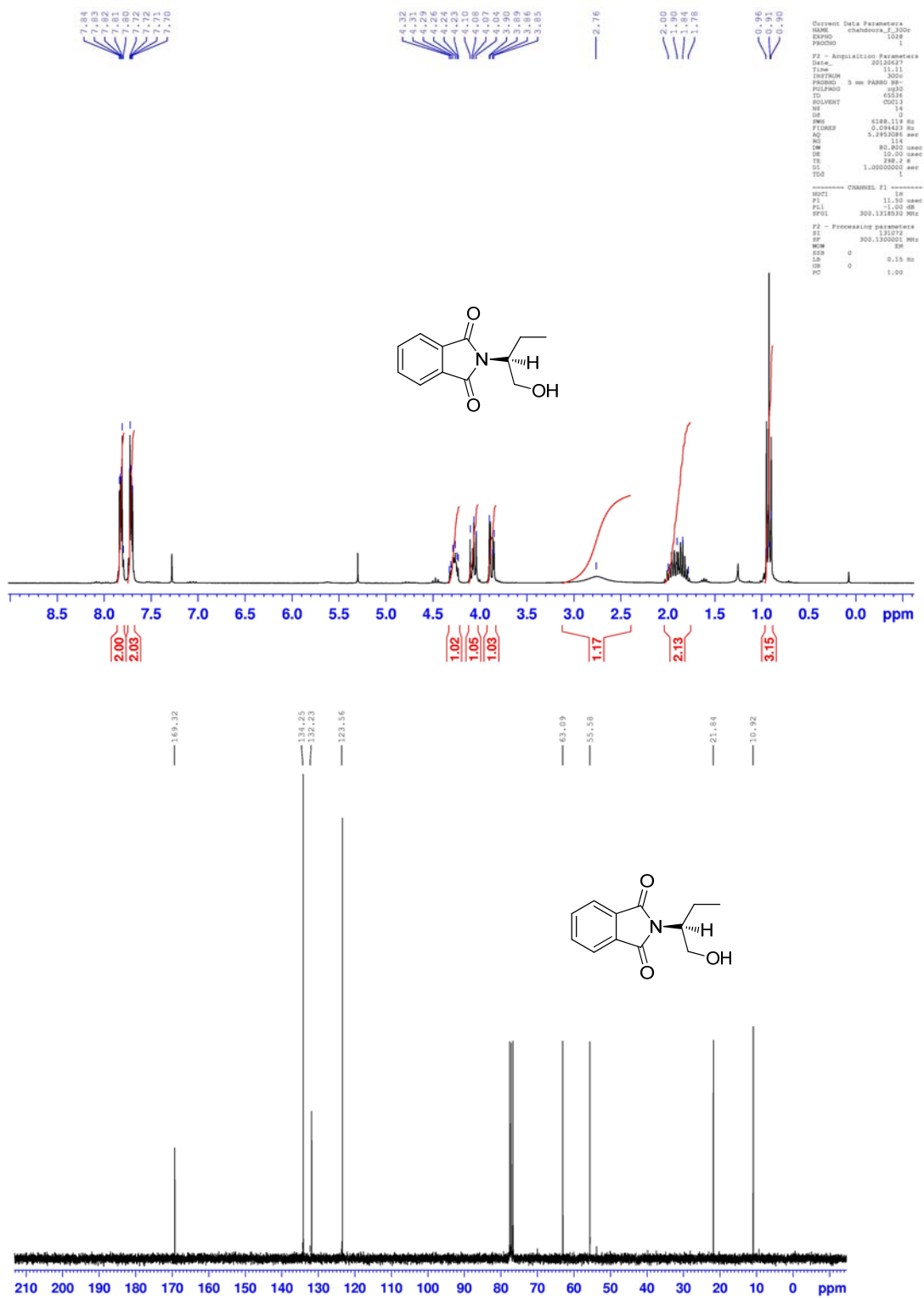
, 27-Jun-2012 + 10:25:25



801  
fc-801- 1624 (12.122) Cm (1623:1634)

, 27-Jun-2012 + 10:25:25

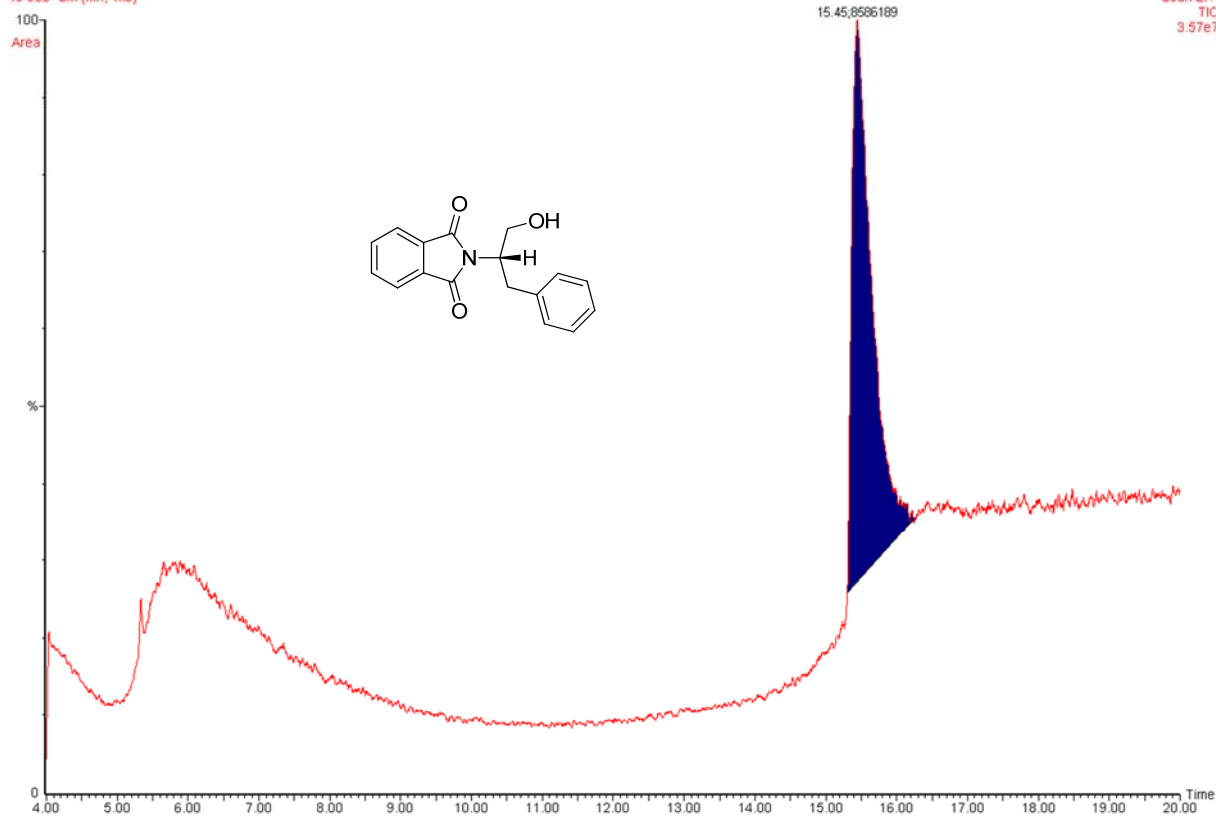




GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **a27**

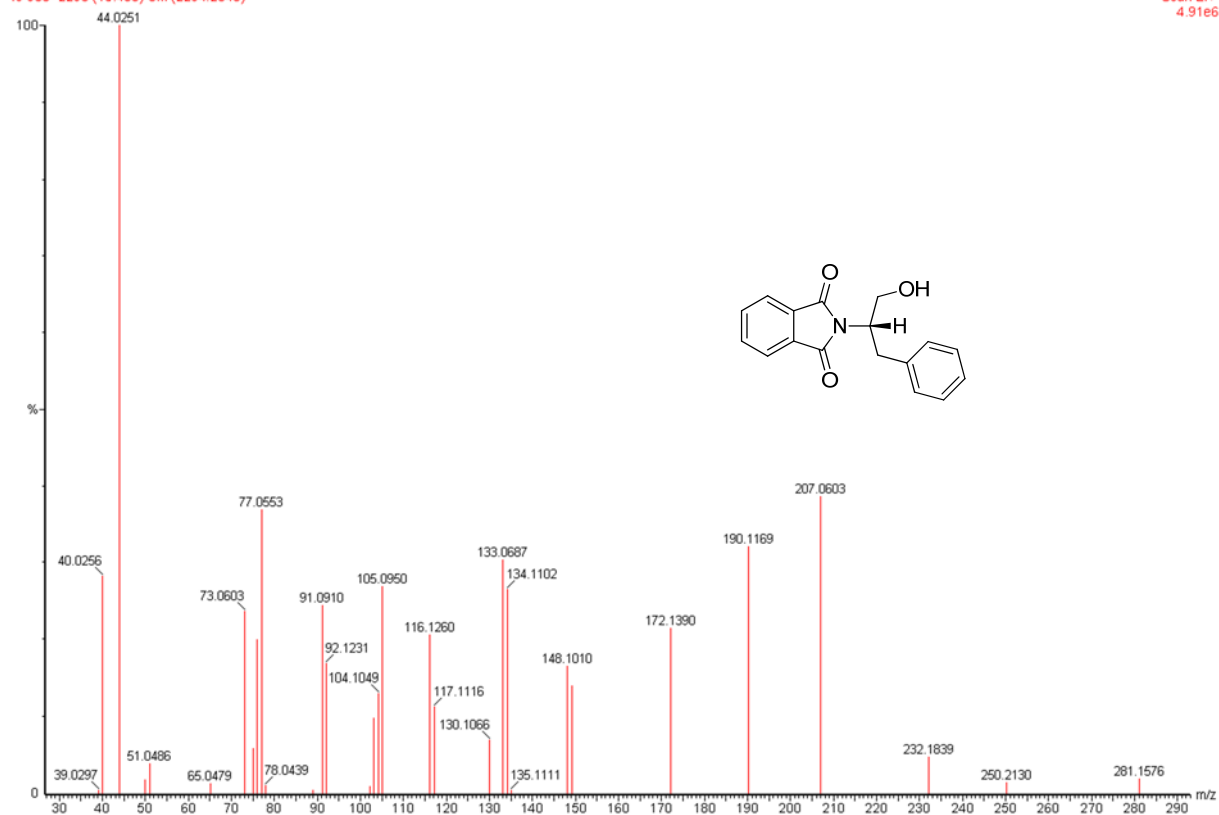
986  
fc-986- Sm (Mn, 1x3)

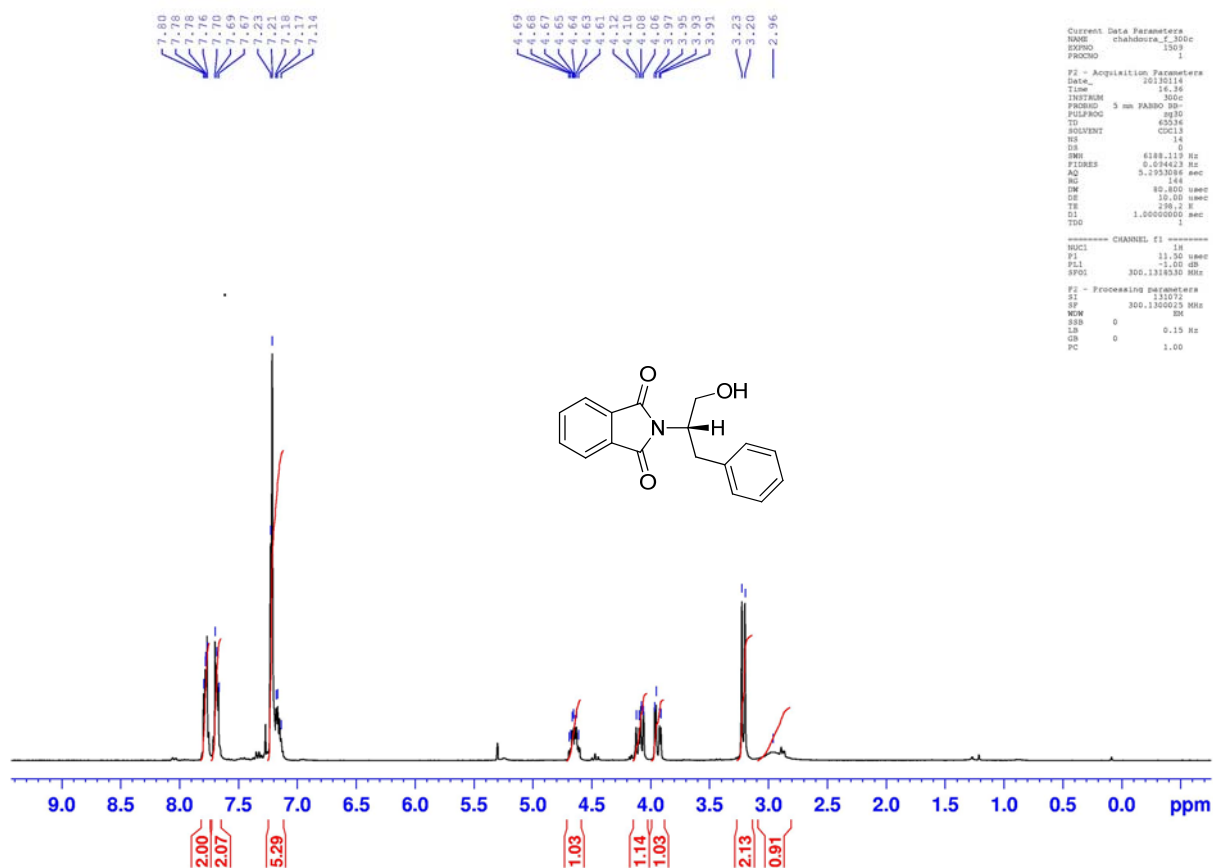
, 14-Jan-2013 + 15:57:38



986  
fc-986- 2296 (15.483) Cm (2294:2346)

, 14-Jan-2013 + 15:57:38





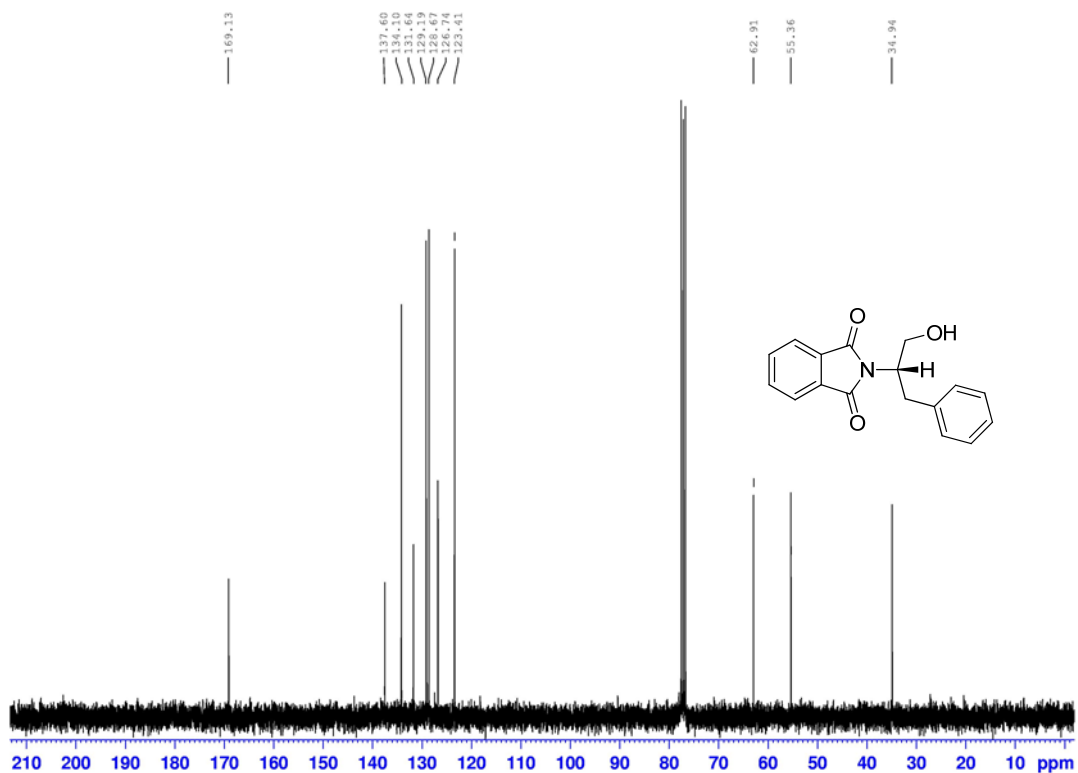
```

Current Data Parameters
NAME chahdora_f_300c
EXPNO 1
PROCNO 1

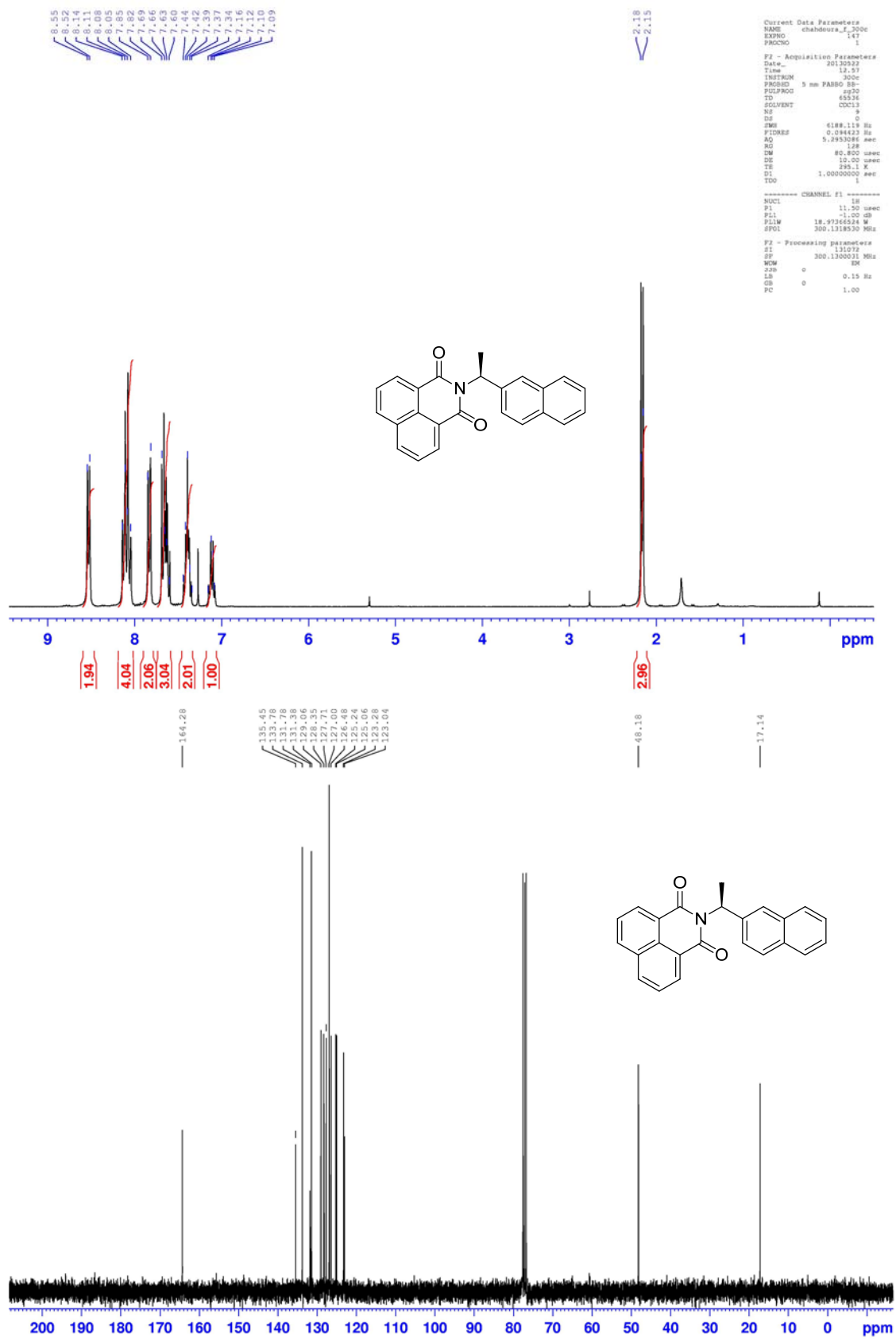
F2 - Acquisition Parameters
Date_ 20191114
Time 14.26
INSTRUM spect
PROBHD 5 mm PABBO DQ-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 14
DS 4
SWH 6188.119 Hz
FIDRES 0.094423 Hz
AQ 5.2953038 sec
RG 144
RM 80.800 usec
DE 16.00 usec
TE 298.2 K
SI 1.0000000 sec
TDC 5

===== CHANNEL f1 =====
NUC1 13C
P1 11.00 usec
PL1 -1.00 dB
SFO1 300.131535 MHz

F2 - Processing parameters
SI 33072
SF 300.1300025 MHz
WDW EM
SSB 0
LB 0.15 Hz
GB 0
PC 1.00
  
```



GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **a28**

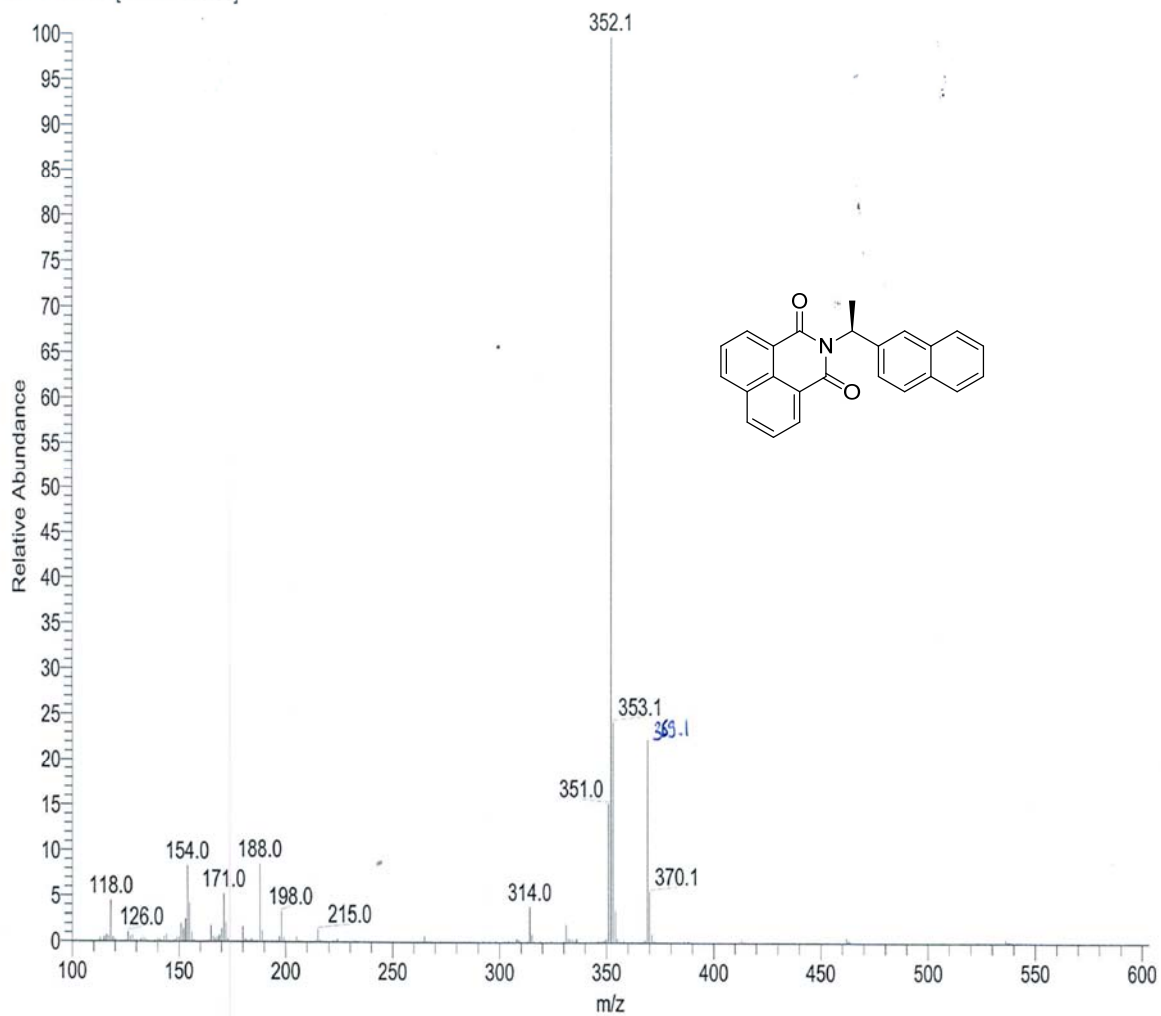


C:\Xcalibur\data\2013\May 2013\FC1124  
DCI/NH3

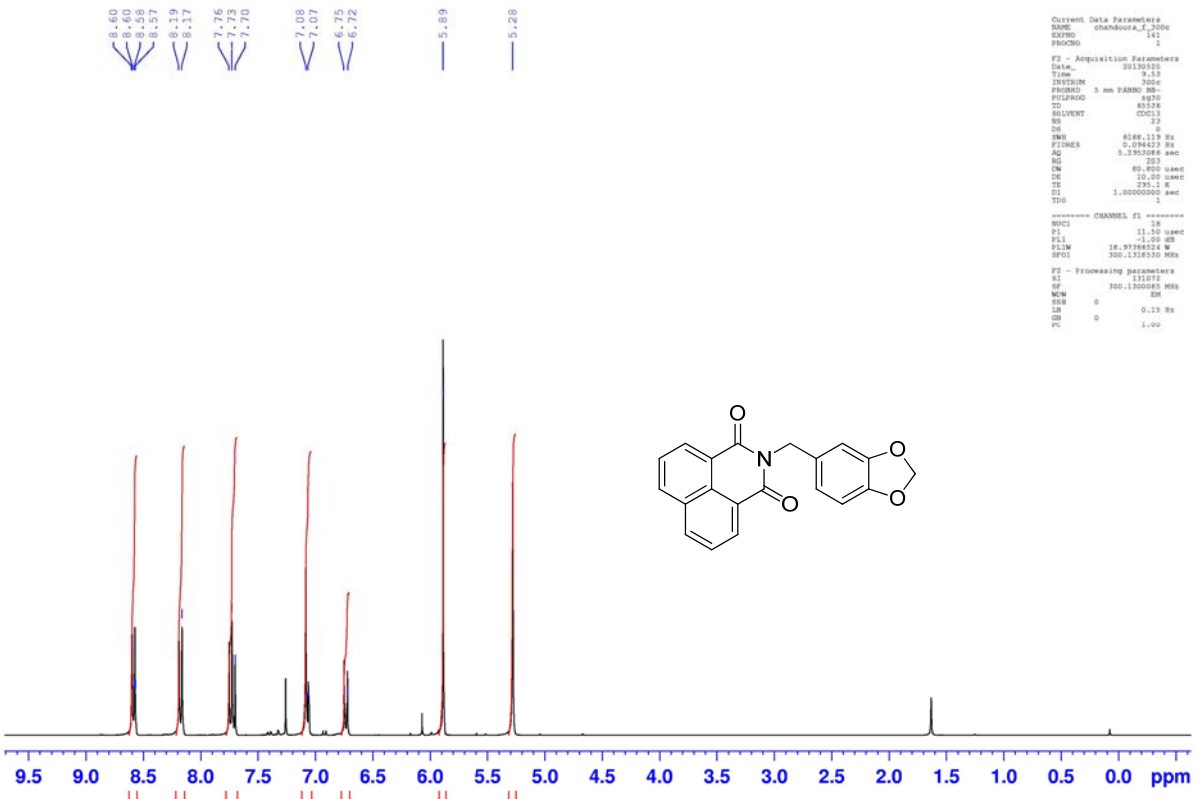
5/24/2013 9:50:17 AM

FC1124 #12-13 RT: 0.27-0.30 AV: 2 NL: 8.25E6

T: + c Full ms [100.00-800.00]

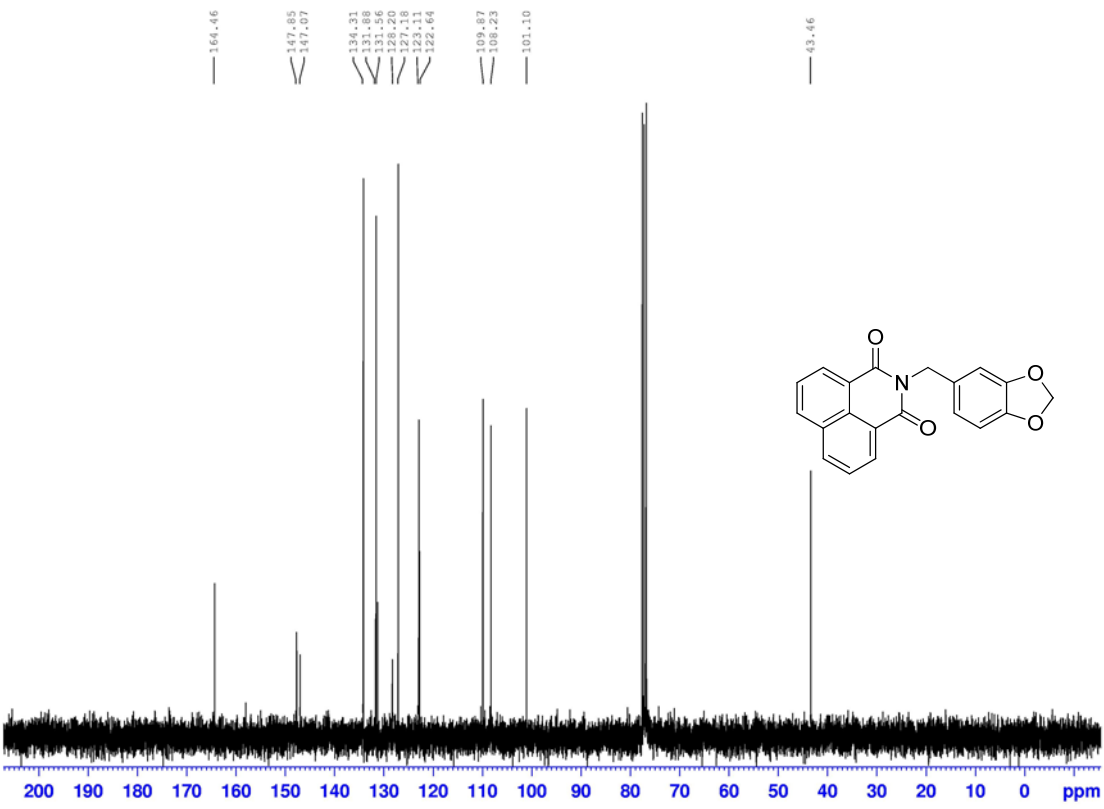


$^1\text{H}$  NMR (300 MHz) and  $^{13}\text{C}$  { $^1\text{H}$ } NMR (75 MHz) (top) spectra in  $\text{CDCl}_3$ , and DCI( $\text{NH}_3$ )-MS (bottom) for **g6**

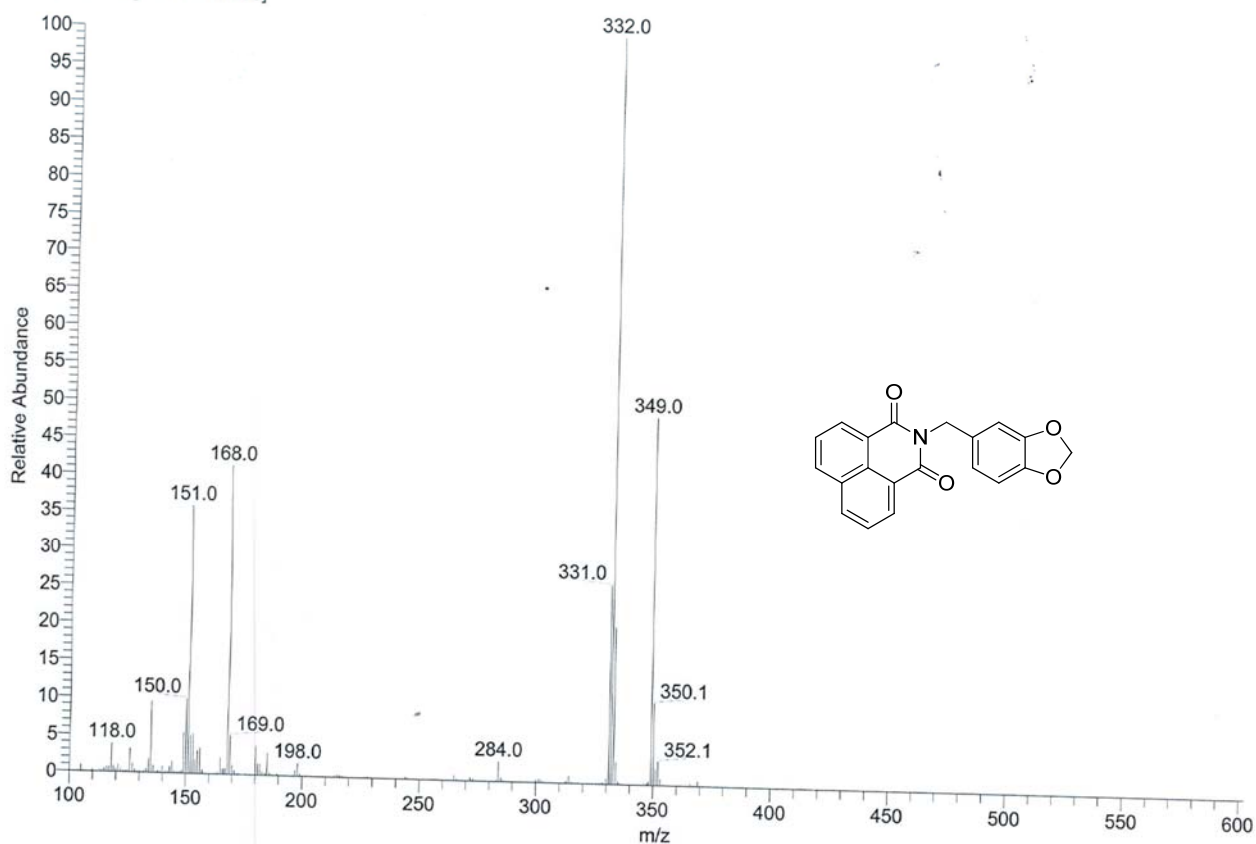


```

Current Data Parameters
NAME: chamsoua_f_2000
EXPNO: 141
PROCNO: 1
F2 - Acquisition Parameters
Date_: 20130510
Time: 9:53
INSTRUM: spect
PROBHD: 5 mm F400 BBO
PULPROG: zgpg30
TD: 65536
SOLVENT: CDCl3
NS: 2023
DS: 4
SWH: 8166.119 Hz
FIDRES: 0.096427 Hz
AQ: 5.2353066 sec
RG: 257
AQ: 80.000 usec
DE: 10.00 usec
TE: 299.15 K
C1: 1.00000000 sec
TD0: 1
----- CHANNEL f1 -----
NUC1: 13
P1: 11.00 usec
PL1: -1.00 dB
PULSE: 18.9758054 Hz
SFO1: 300.1318530 MHz
F2 - Processing parameters
SI: 13182
SF: 300.1300885 MHz
WDW: EM
SSB: 0
GB: 0
PC: 1.00
  
```



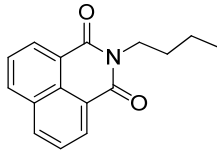
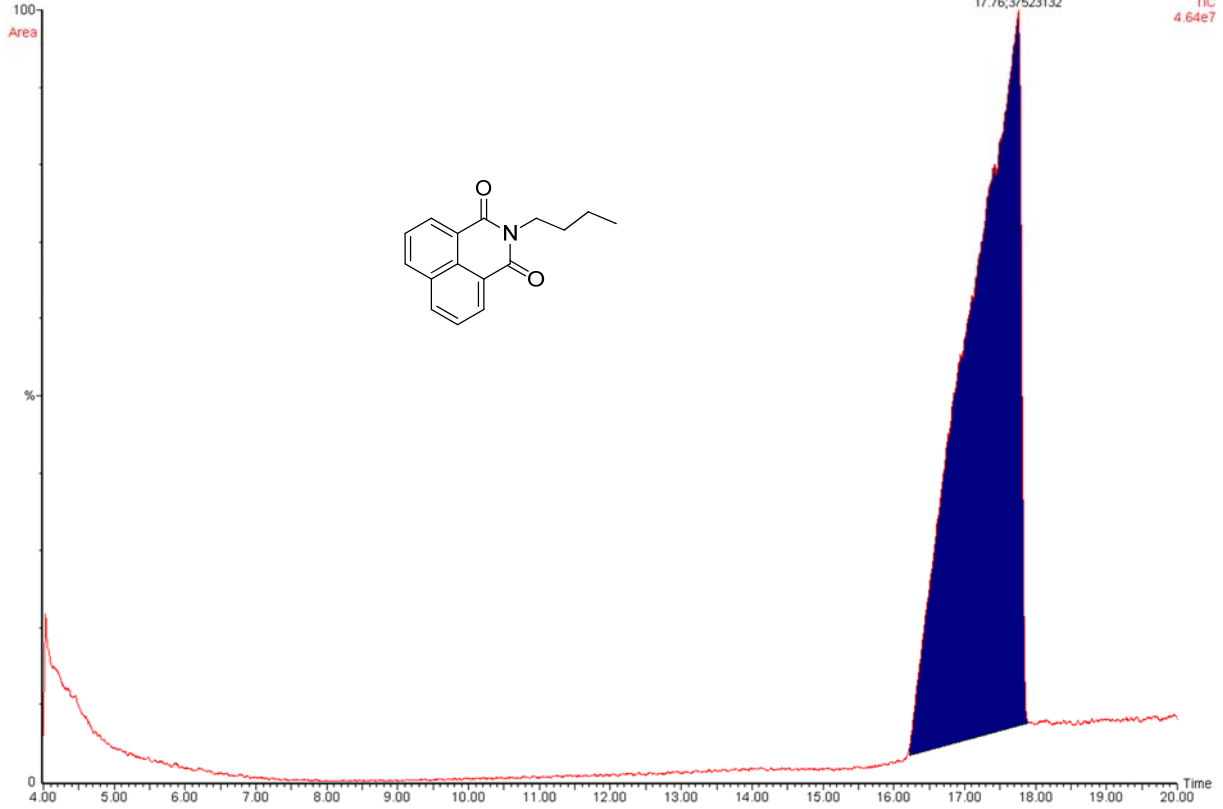




$^1\text{H}$  NMR (300 MHz) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (top) spectra in  $\text{CDCl}_3$ , and DCI( $\text{NH}_3$ )-MS (bottom) for **g8**

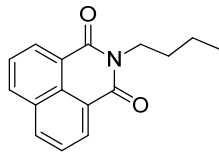
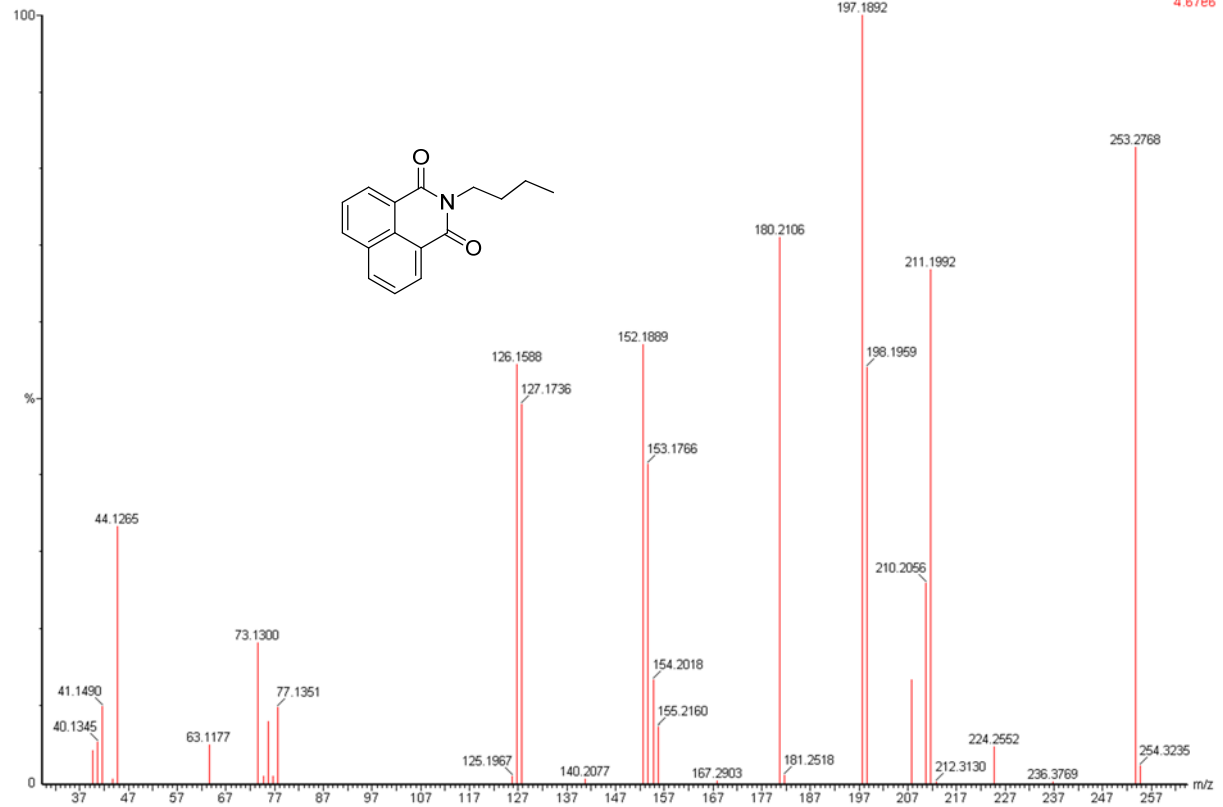
1108  
fc-1108- Sm (Mn, 1x3)

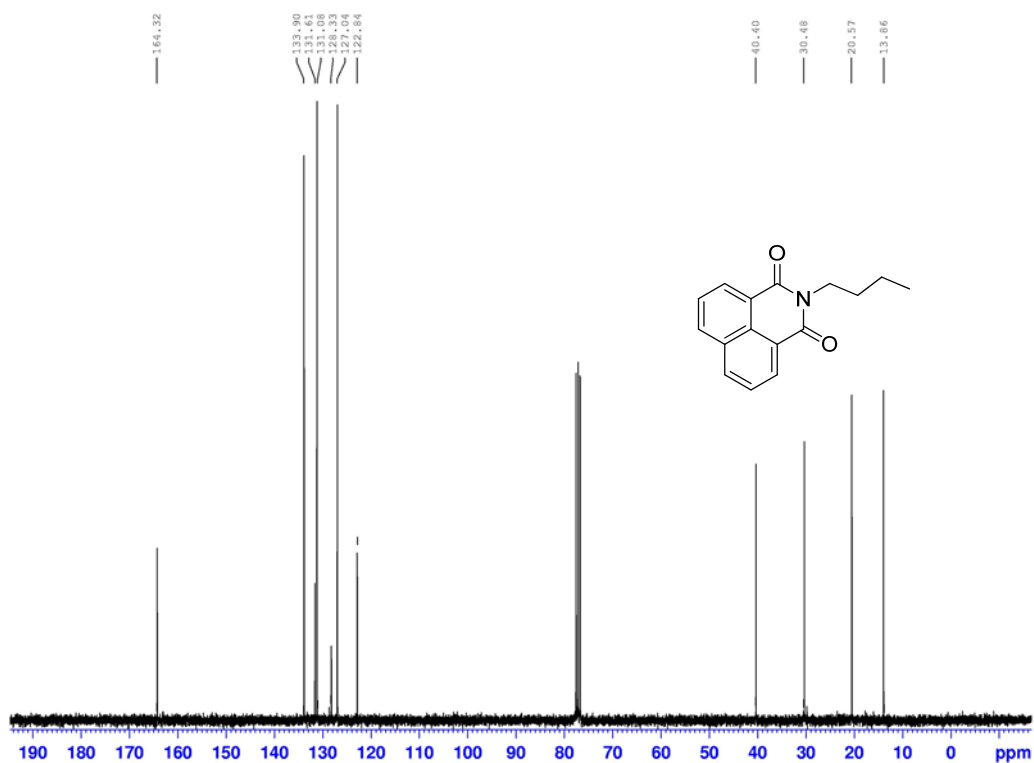
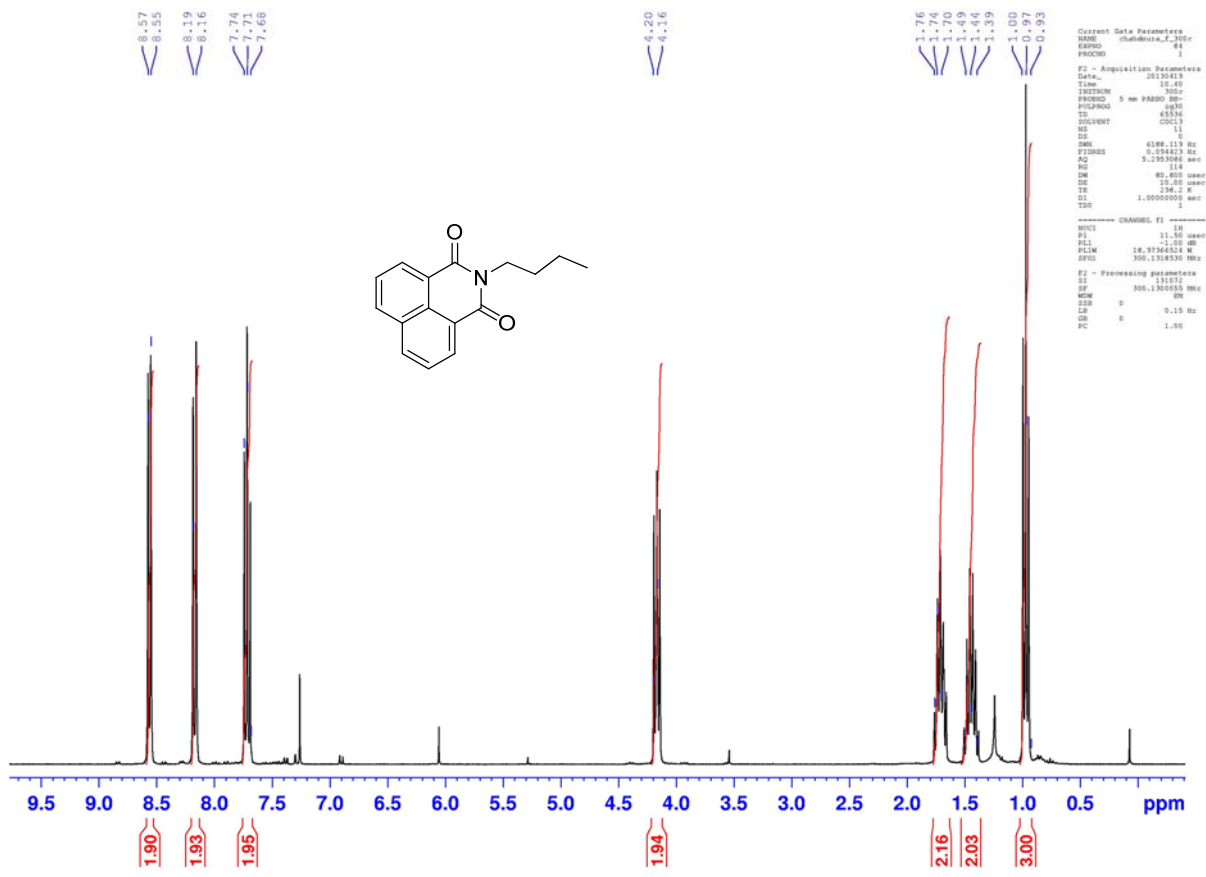
, 19-Apr-2013 + 11:04:07  
Scan E1+  
TIC  
4.64e7



1108  
fc-1108- 2682 (17.414) Cm (2640:2689)

, 19-Apr-2013 + 11:04:07  
Scan E1+  
4.67e6



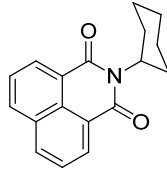
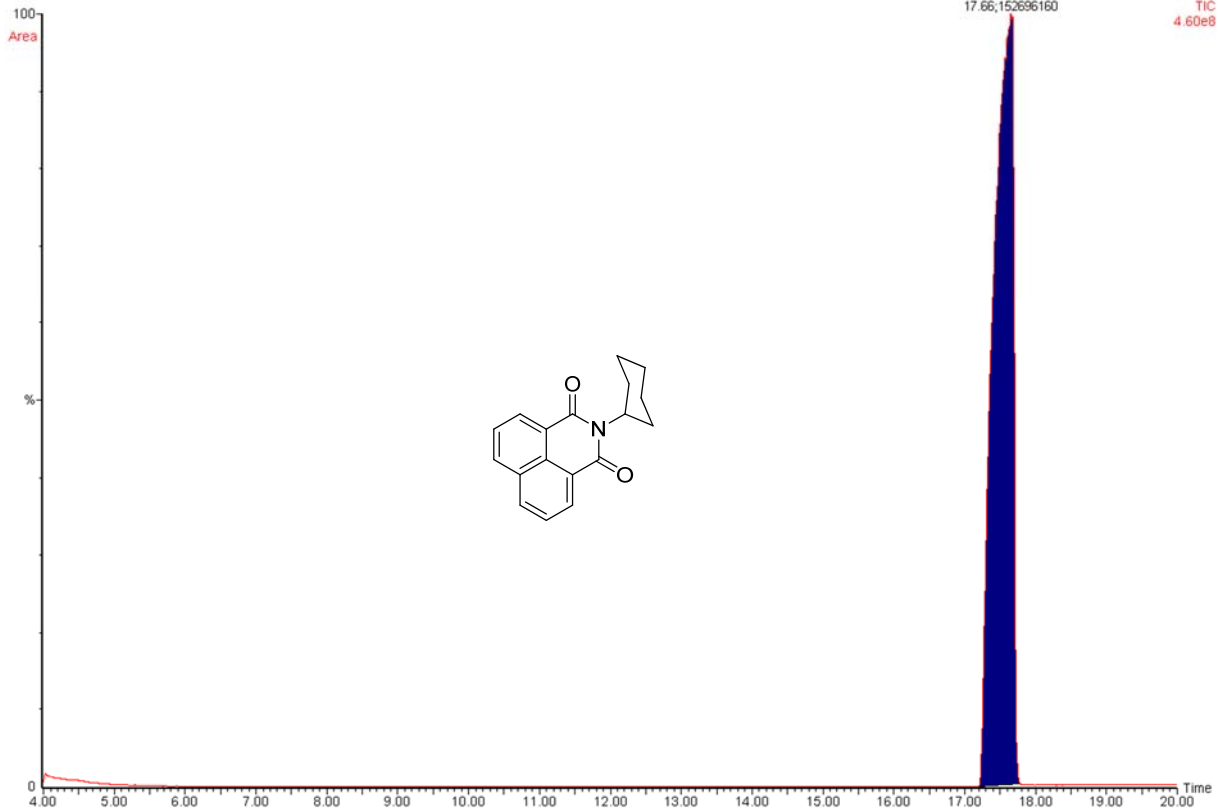


GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **g9**

1115  
fc-1115- Sm (Mn, 1x3)

, 29-Apr-2013 + 14:34:00

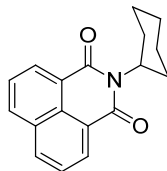
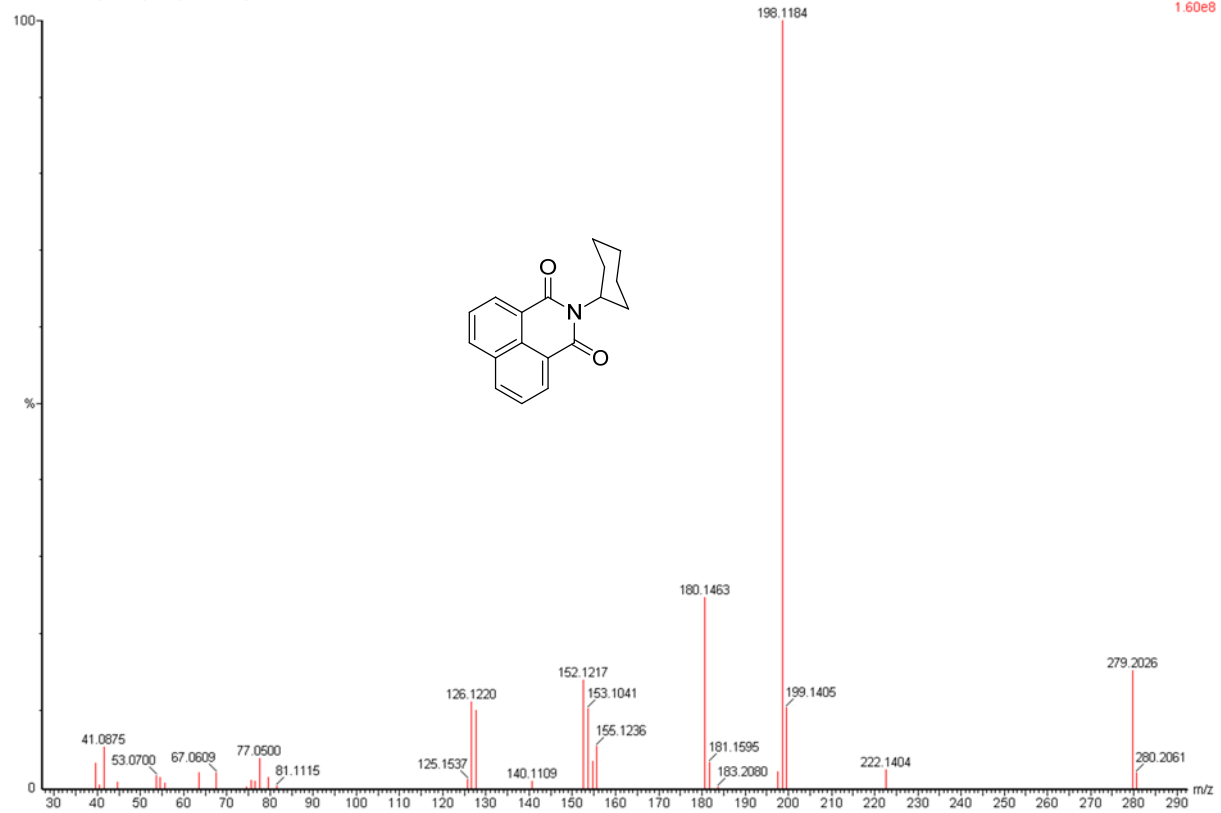
Scan EI+  
TIC  
4.60e8

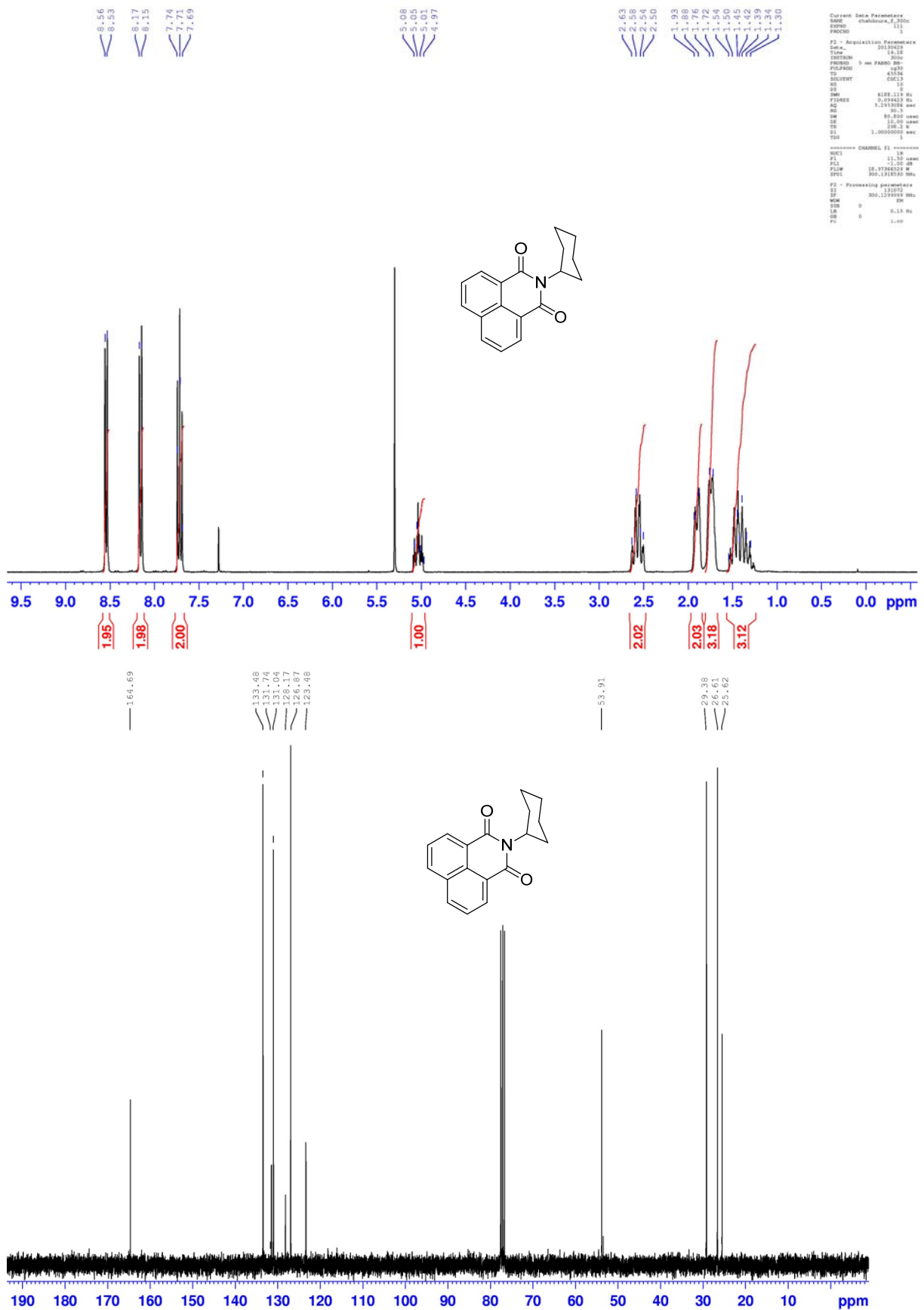


1115  
fc-1115- 2716 (17.584) Cm (2682:2719)

, 29-Apr-2013 + 14:34:00

Scan EI+  
1.60e8





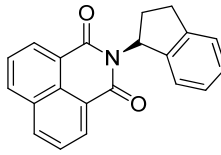
GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **g12**

8.59  
8.56  
8.22  
8.19  
7.77  
7.75  
7.71  
7.37  
7.34  
7.28  
7.24  
7.12  
7.09  
7.09  
6.88  
6.86  
6.82

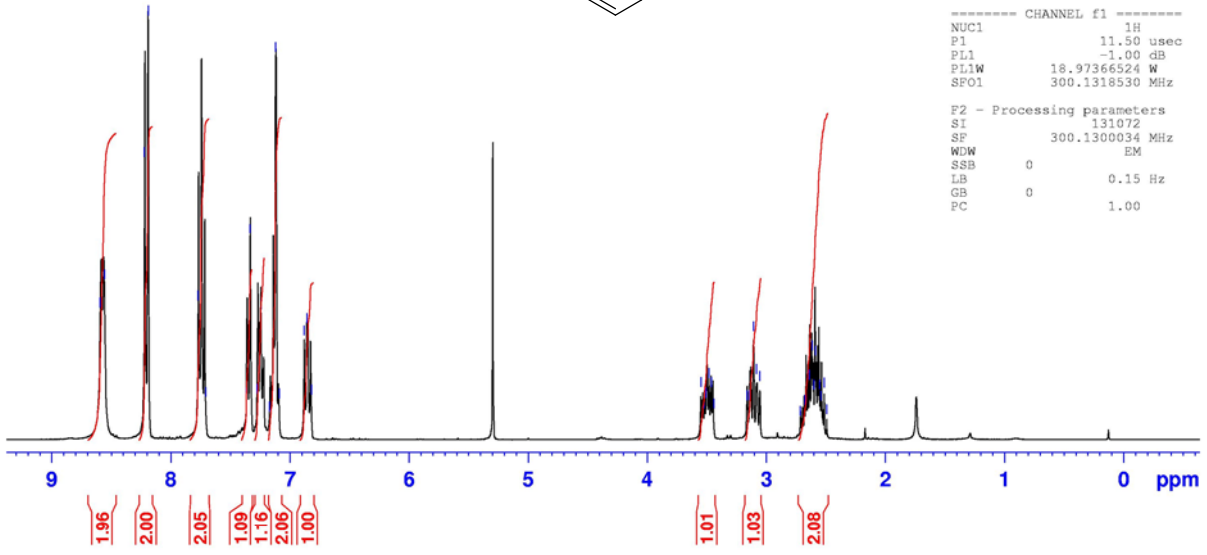
3.55  
3.51  
3.51  
3.48  
3.46  
3.44  
3.17  
3.14  
3.11  
3.08  
3.05  
2.71  
2.68  
2.61  
2.60  
2.59  
2.55  
2.54  
2.52  
2.49

Current Data Parameters  
NAME chahdoura\_f\_300c  
EXPNO 149  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20130523  
Time 9.44  
INSTRUM 300c  
PROBHD 5 mm PABBO BB-  
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 12  
DS 0  
SWH 6188.119 Hz  
FIDRES 0.094423 Hz  
AQ 5.2953086 sec  
RG 90.5  
DW 80.800 usec  
DE 10.00 usec  
TE 295.1 K  
D1 1.0000000 sec  
TDO 1



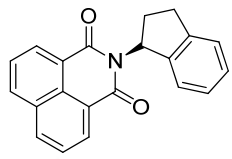
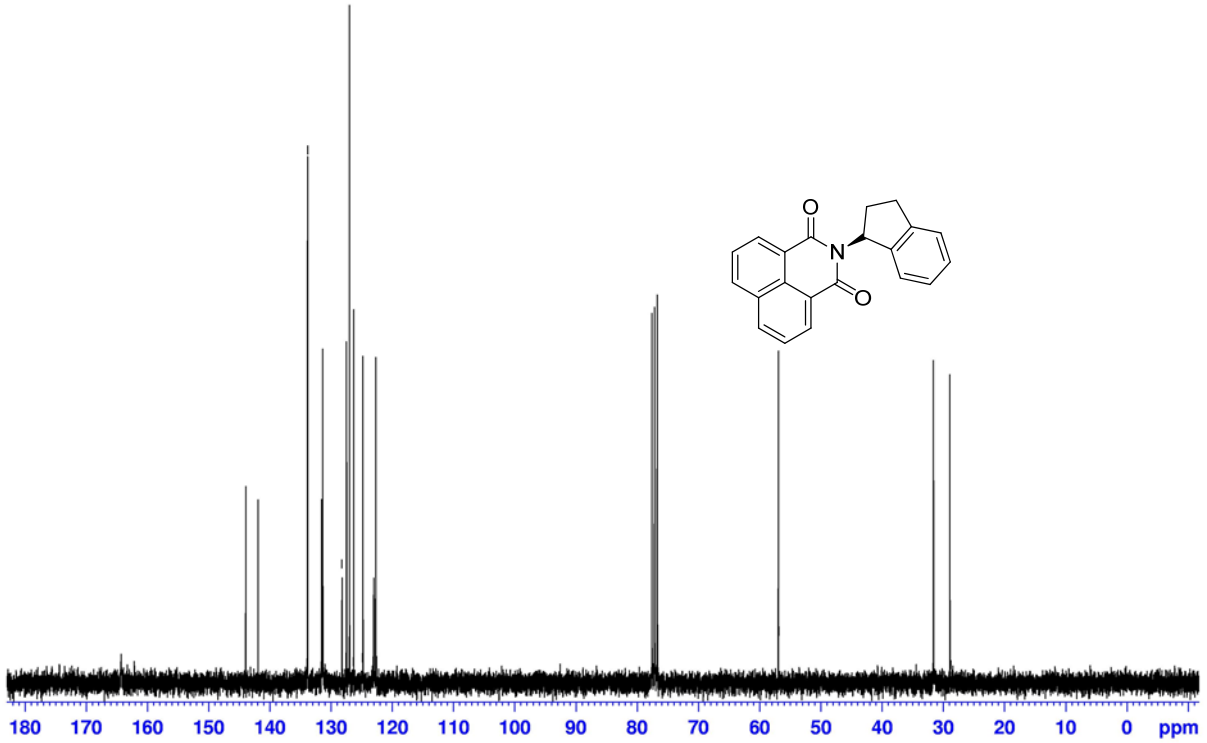
----- CHANNEL f1 -----  
NUC1 1H  
P1 11.50 usec  
PL1 -1.00 dB  
PLW 18.97366524 W  
SFO1 300.1318530 MHz  
  
F2 - Processing parameters  
SI 131072  
SF 300.1300034 MHz  
WDW EM  
SSB 0  
LB 0.15 Hz  
GB 0  
PC 1.00



1.96  
2.00  
2.05  
1.09  
1.16  
2.06  
1.00  
1.01  
1.03  
2.08

143.98  
141.93  
133.64  
133.66  
131.22  
127.60  
126.95  
126.35  
124.86  
123.15  
122.19

56.88  
31.65  
28.76

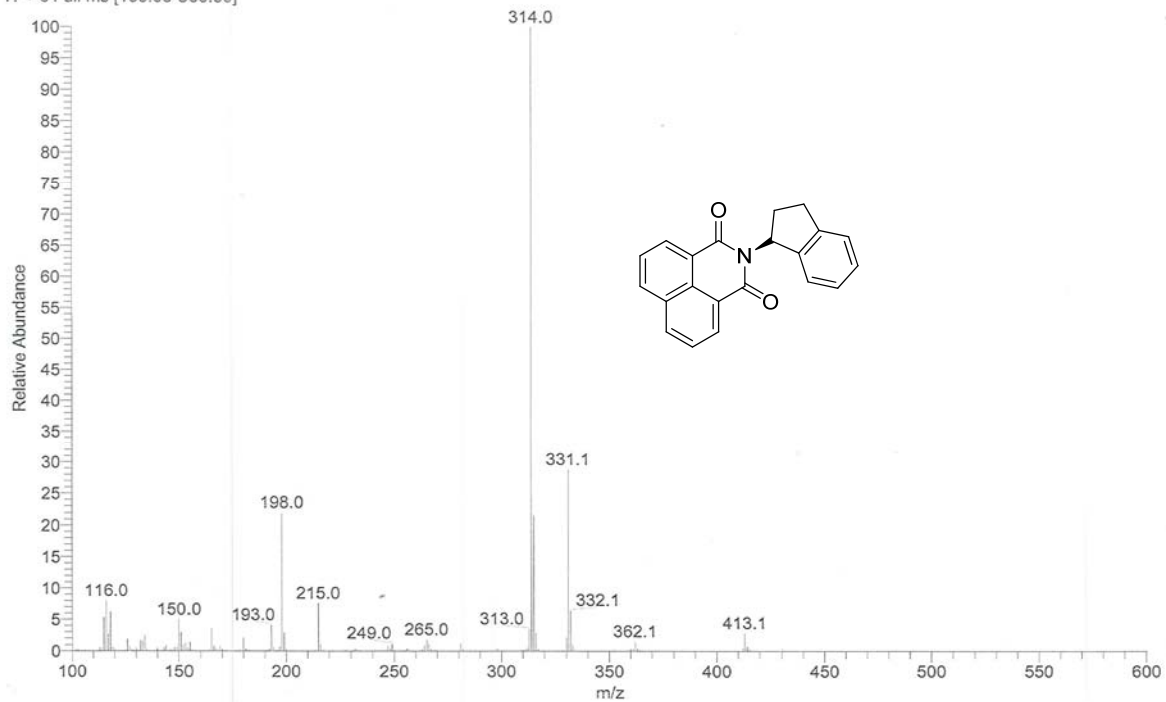


180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 ppm

C:\Xcalibur\data\2013\Mai 2013\FC1125  
DCI/NH3

5/24/2013 9:47:22 AM

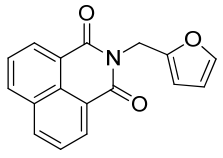
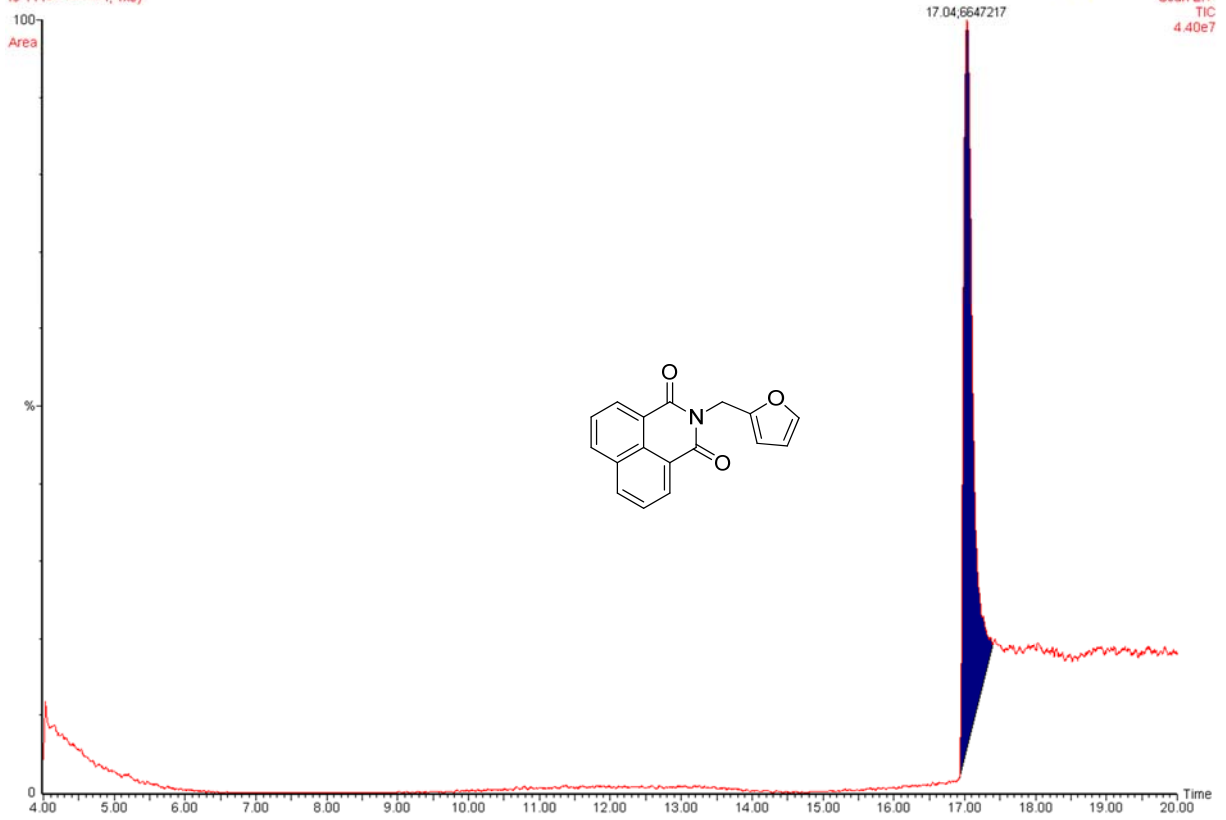
FC1125 #11-13 RT: 0.25-0.30 AV: 3 NL: 7.80E6  
T: + c Full ms [100.00-800.00]



$^1\text{H}$  NMR (300 MHz) and  $^{13}\text{C}$  { $^1\text{H}$ } NMR (75 MHz) (top) spectra in  $\text{CDCl}_3$ , and DCI( $\text{NH}_3$ )-MS (bottom) for **g16**

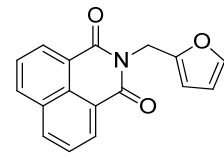
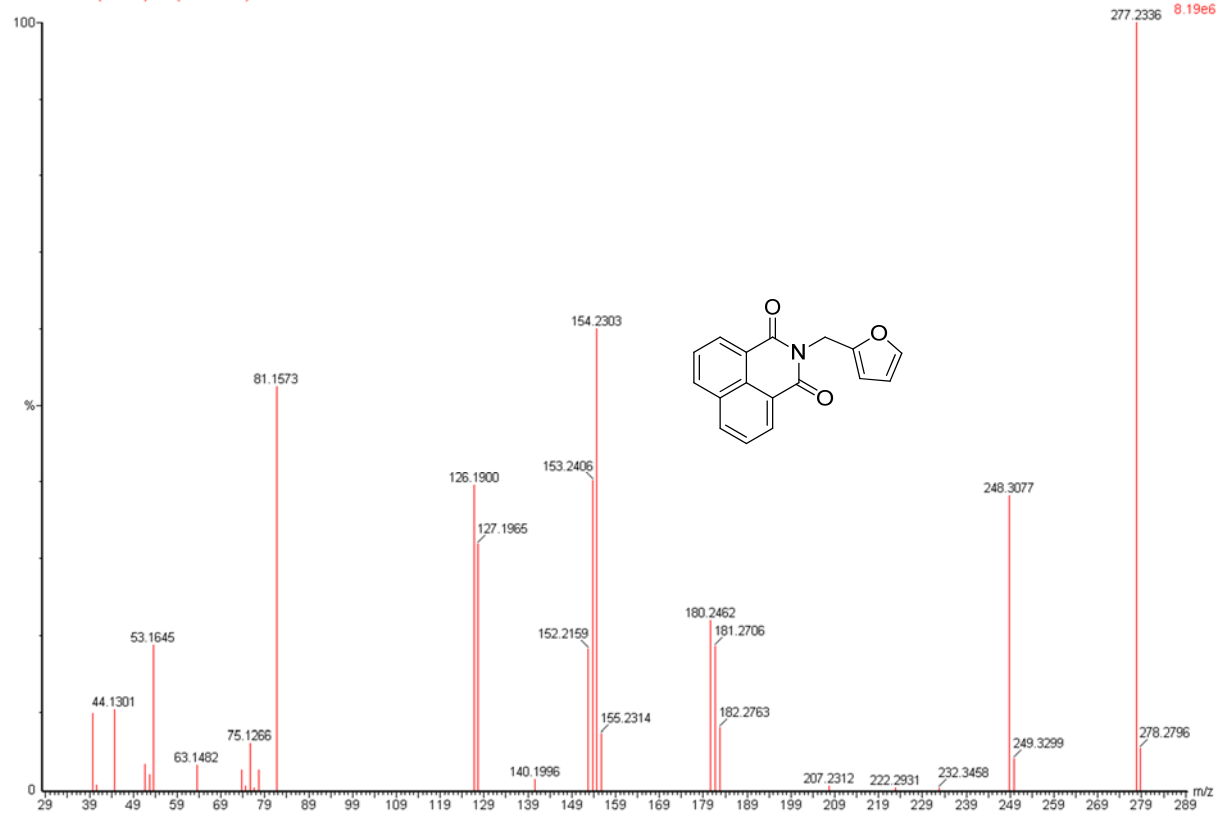
1114  
fc-1114- Sm (Mn, 1x3)

, 26-Apr-2013 + 15:58:54

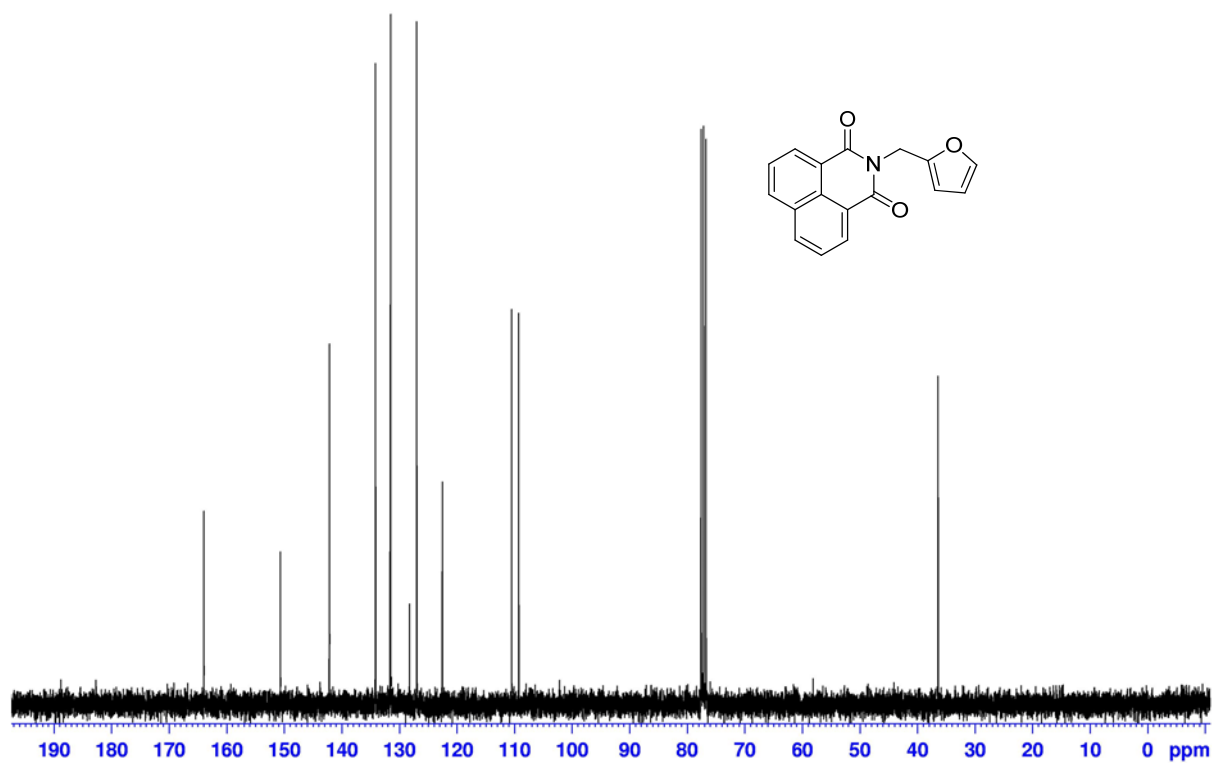
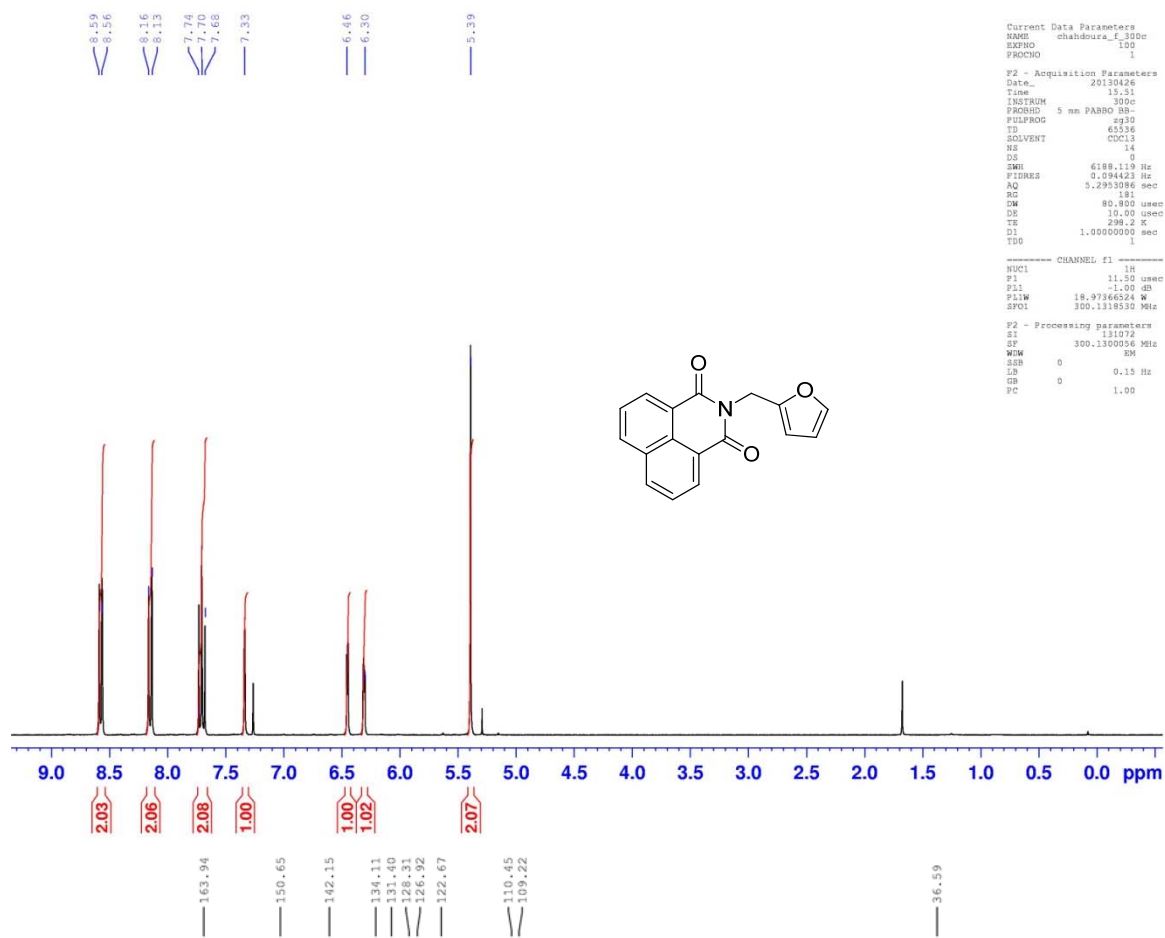


1114  
fc-1114- 2606 (17.033) Cm (2602:2617)

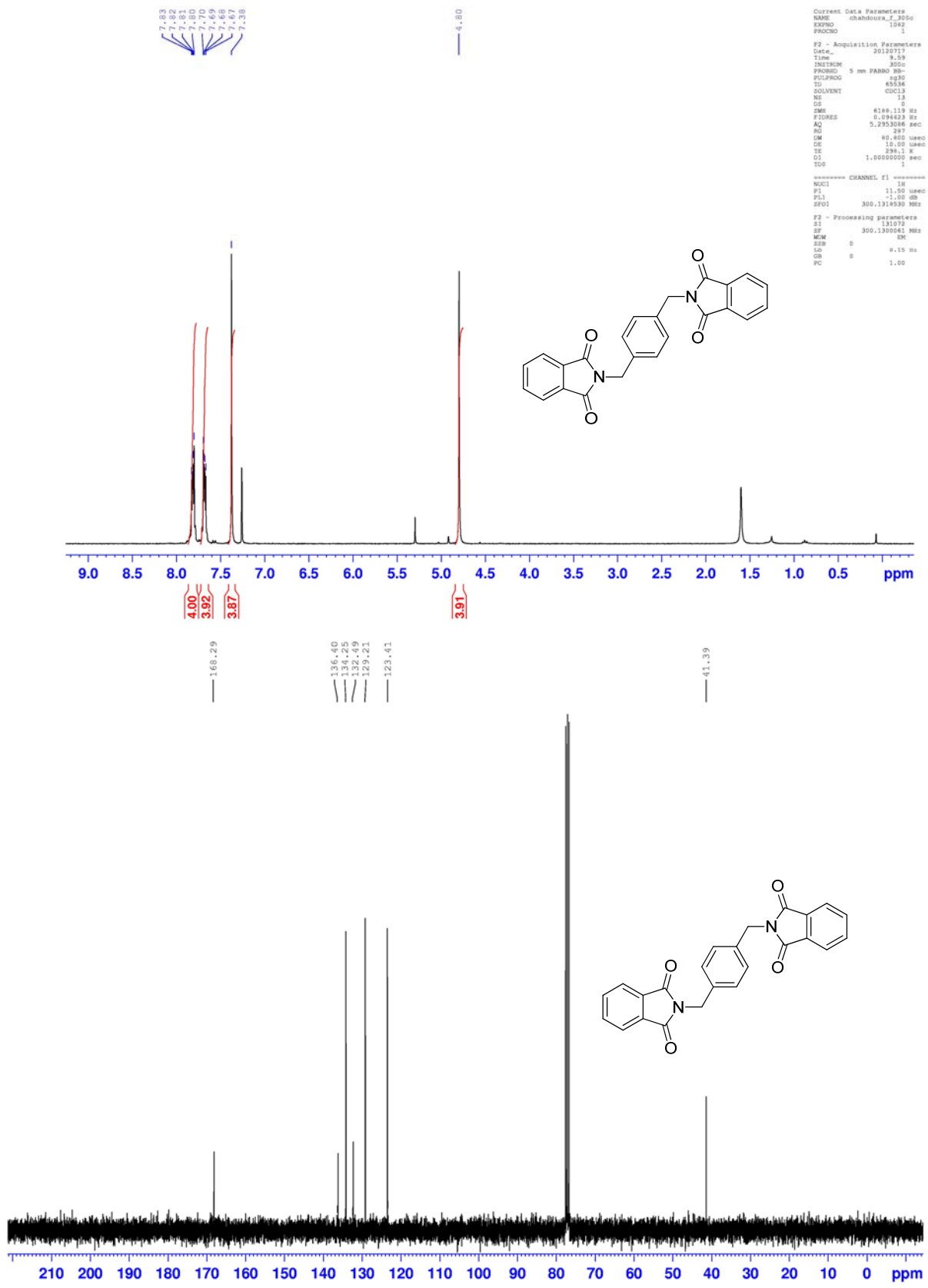
, 26-Apr-2013 + 15:58:54



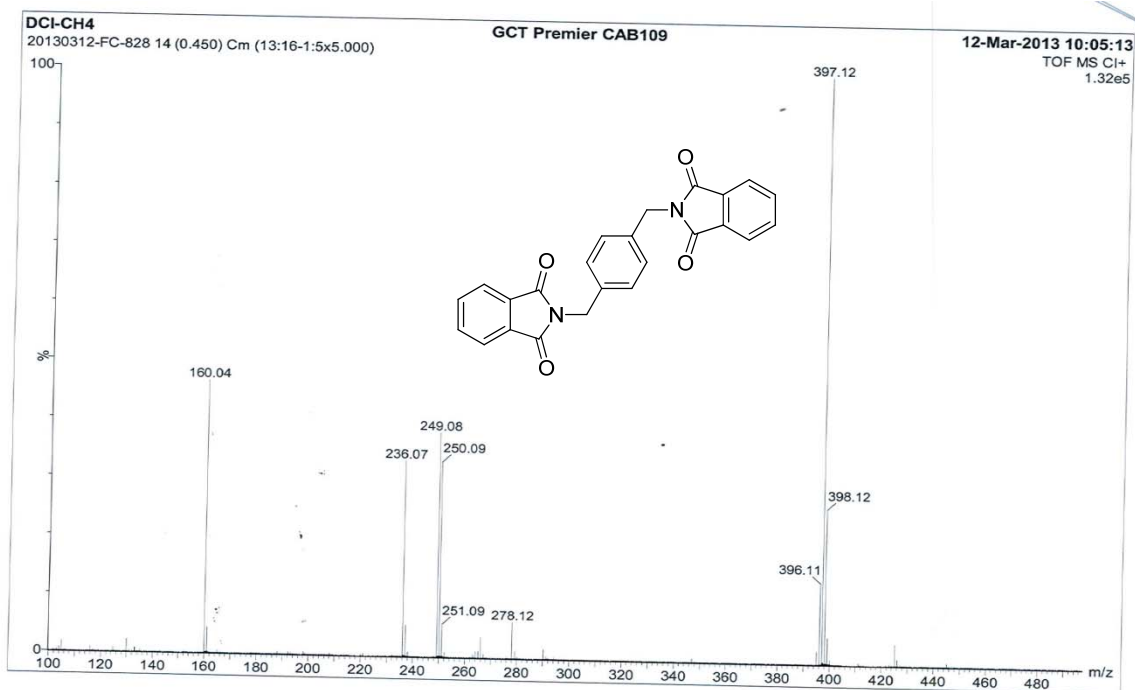




GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C} \{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **g24**



Current Data Parameters  
 NAME cshahdura\_2\_305c  
 EXPNO 1082  
 PROCNO 1  
 F2 - Acquisition Parameters  
 Date\_ 20120717  
 Time 9.59  
 INSTRUM spect  
 PPROBHD 5 mm PABBO BB-  
 PULPROG zgpg30  
 TD 32768  
 SFO1 300.1314530 MHz  
 SOLVENT CDCl3  
 NS 13  
 DS 0  
 SWH 6188.119 Hz  
 FIDRES 0.094423 Hz  
 AQ 5.2953086 sec  
 RG 287  
 PC 80.405 usec  
 DE 18.00 usec  
 TE 298.1 K  
 D1 1.0000000 sec  
 TSD  
 ===== CHANNEL f1 =====  
 NUC1 13  
 P1 11.00 usec  
 PL1 -1.00 dB  
 SFO1 300.1314530 MHz  
 F2 - Processing parameters  
 SI 131074  
 SF 300.1300041 MHz  
 MCM 64  
 ZSB 0  
 LB 0  
 GB 0  
 PC 1.00



#### Single Mass Analysis

Tolerance = 3.0 PPM / DBE: min = -1.5, max = 50.0

Element prediction: Off

Monoisotopic Mass, Odd and Even Electron Ions

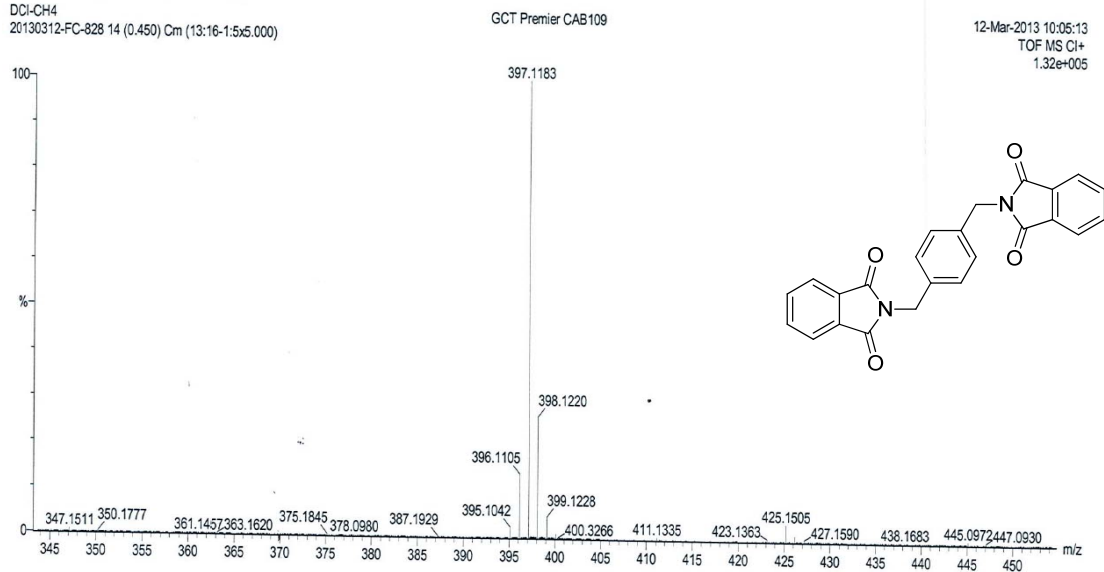
619 formula(e) evaluated with 3 results within limits (all results (up to 1000) for each mass)

Elements Used:

C: 0-60 H: 0-60 N: 0-15 O: 0-8

DCI-CH4

20130312-FC-828 14 (0.450) Cm (13:16-1:5x5.000)



Minimum:

Maximum: 1.3 3.0 -1.5

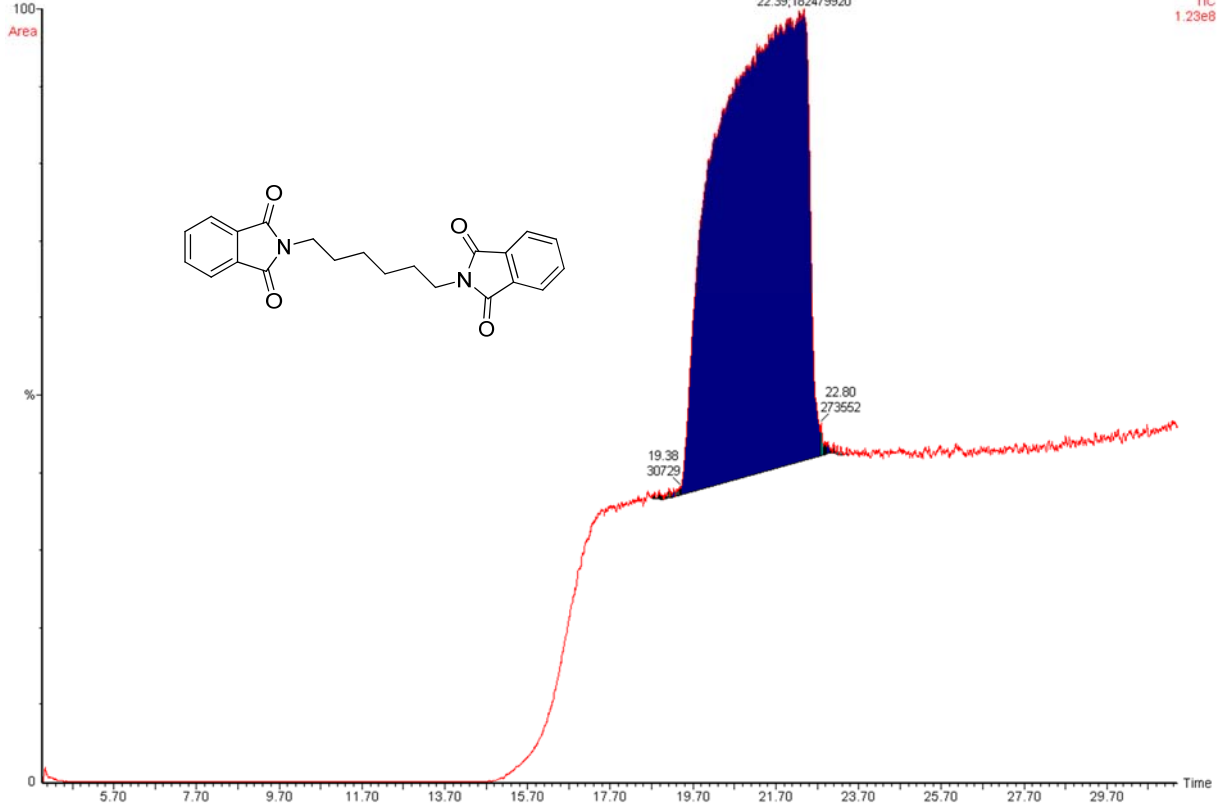
Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	Formula
397.1183	397.1175	0.8	2.0	18.0	41.6	C22 H15 N5 O3
	397.1188	-0.5	-1.3	17.5	45.1	C24 H17 N2 O4
	397.1193	-1.0	-2.5	10.5	3396.0	C9 H13 N14 O5

$^1\text{H}$  NMR (300 MHz) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (top) spectra in  $\text{CDCl}_3$ , and low and high resolution DCI( $\text{CH}_4$ )-MS (bottom) for **a29**

S10  
fc-810-3 Sm (Mn, 1x3)

, 03-Jul-2012 + 16:32:30

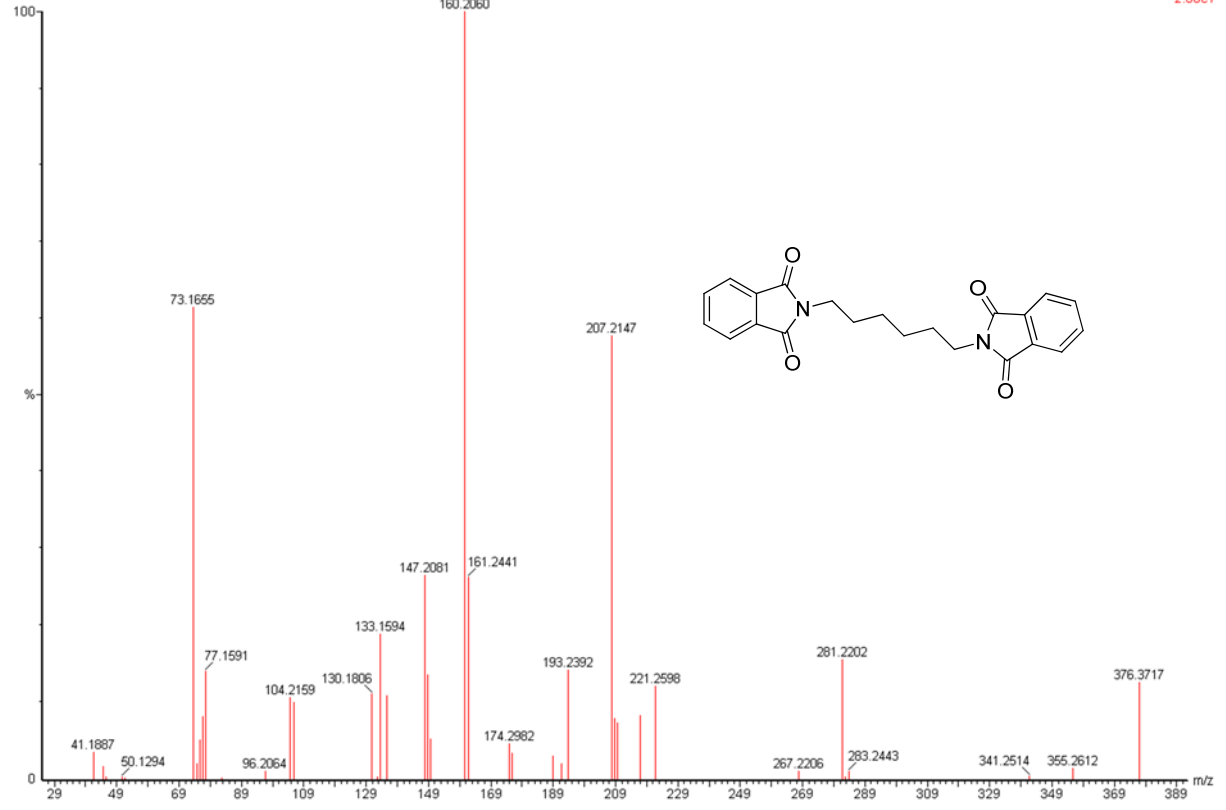
Scan EI+  
TIC  
1.23e8

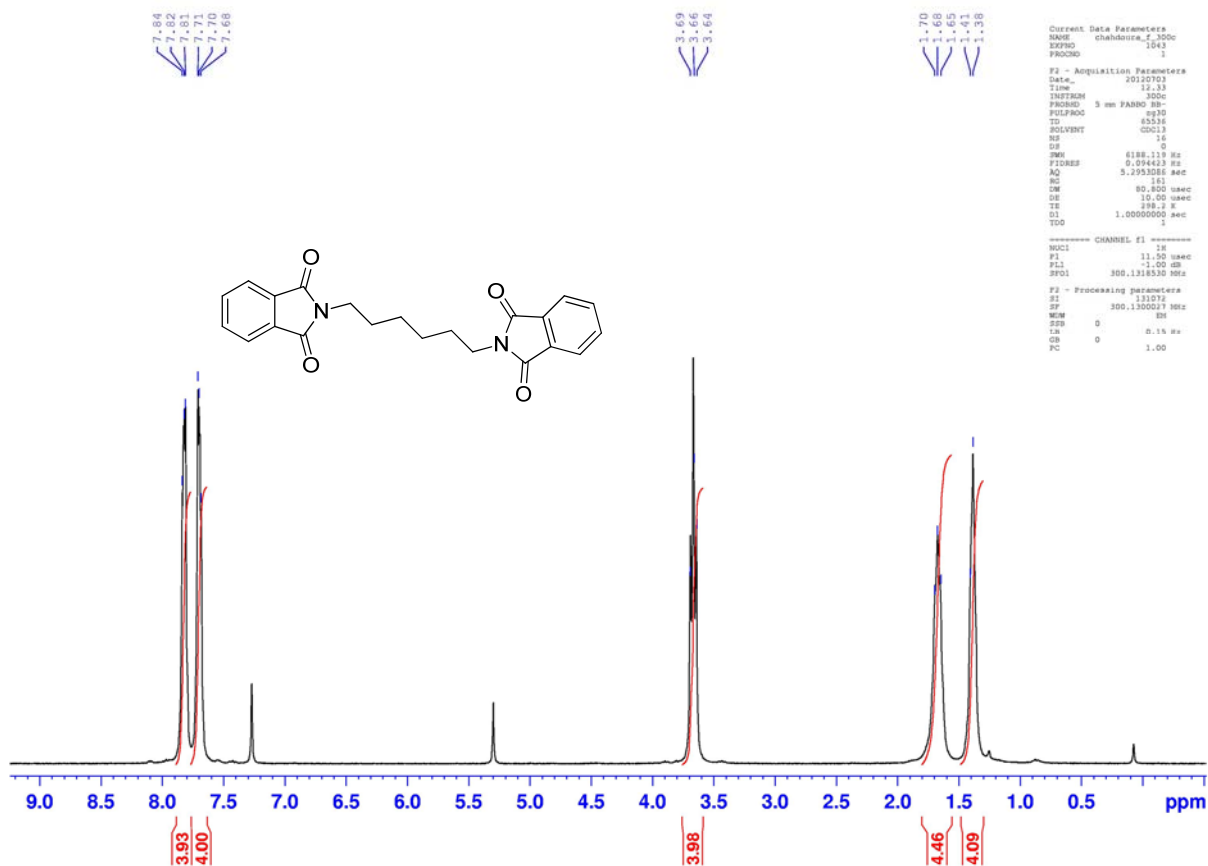


S10  
fc-810-3 3541 (21.710) Cm (3242:3604)

, 03-Jul-2012 + 16:32:30

Scan EI+  
2.36e7

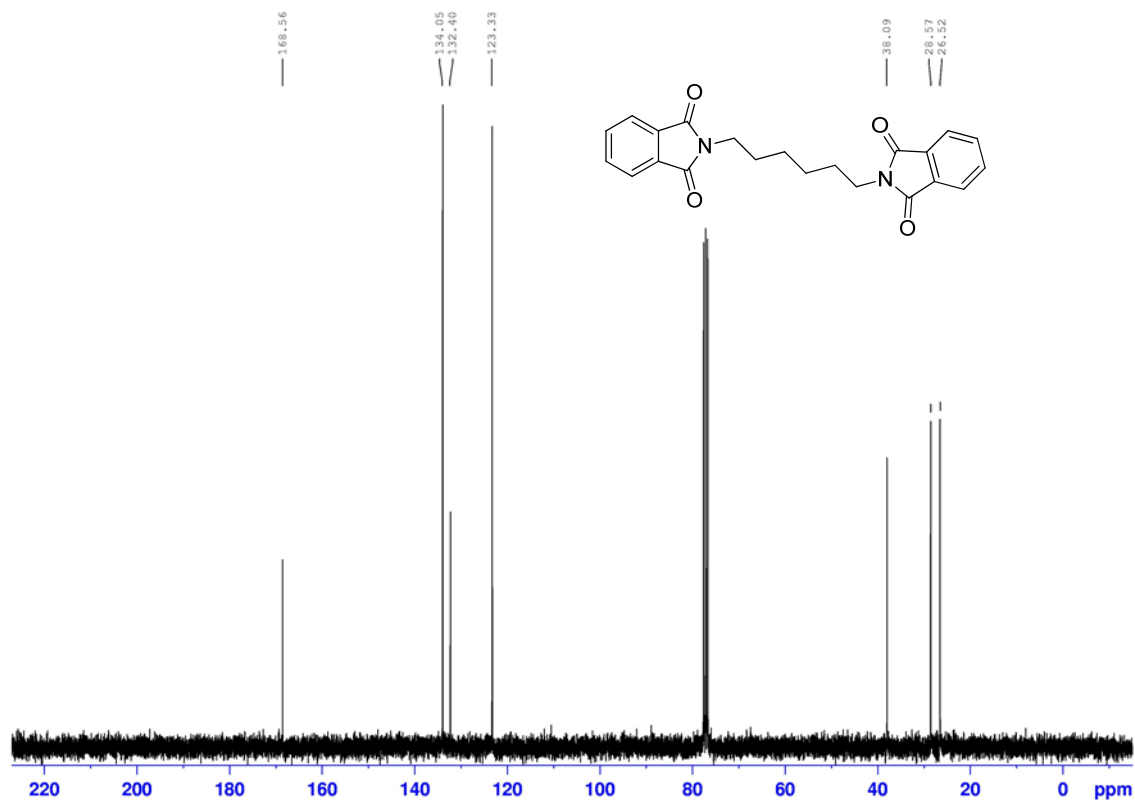




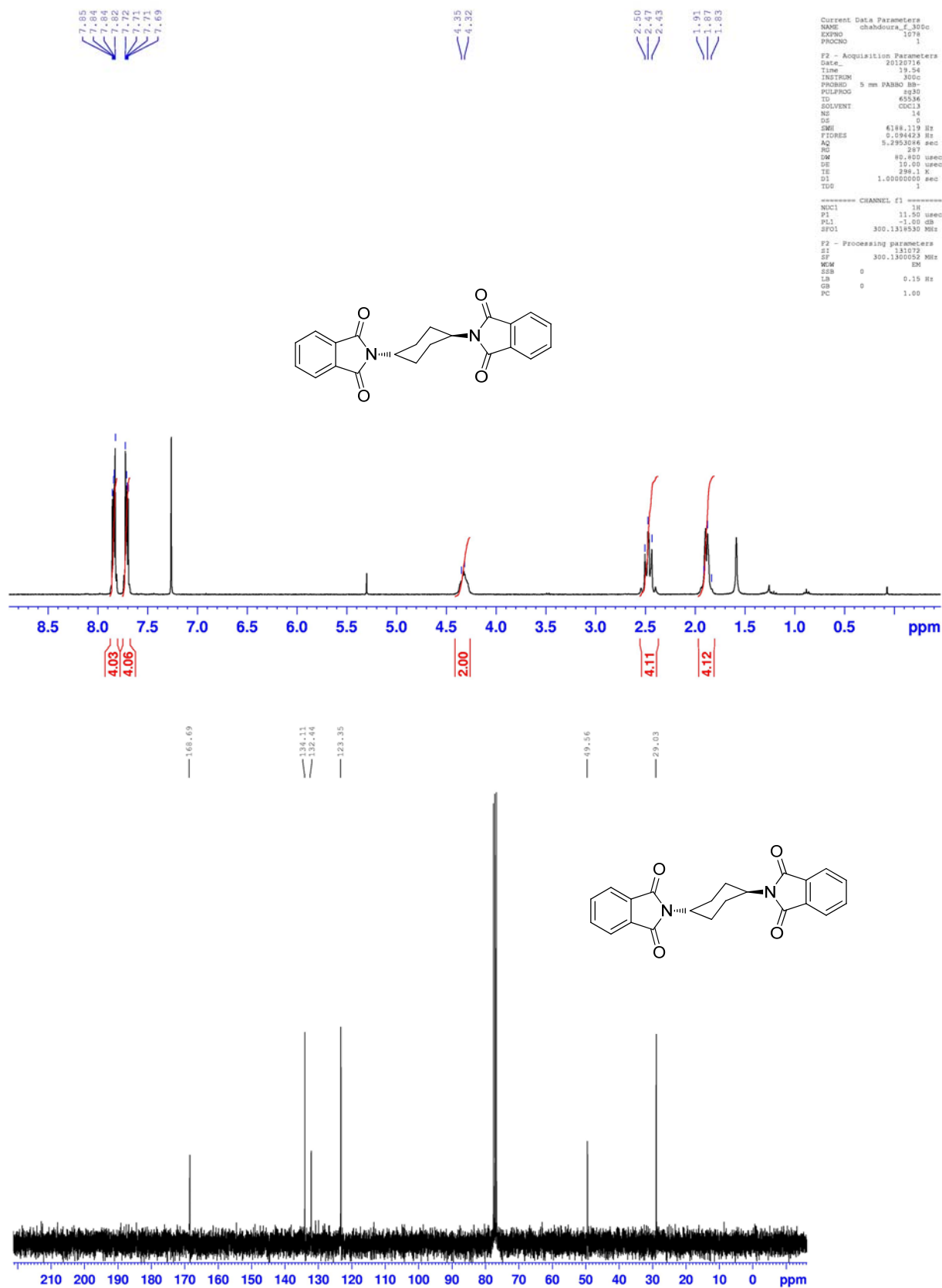
Current Data Parameters  
 NAME chahdosa\_f\_300c  
 EXPNO 1543  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 2012013  
 Time 12.33  
 INSTRUM 300c  
 PROBRD 5 mm PABBO BB-  
 PULPROG zg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 16  
 DS 8  
 SWH 6188.119 Hz  
 FIDRES 0.094423 Hz  
 AQ 3.2935868 sec  
 RG 181  
 CW 80.800 usec  
 DE 10.00 usec  
 TE 298.2 K  
 DQ 1.0000000 sec  
 TDO 0

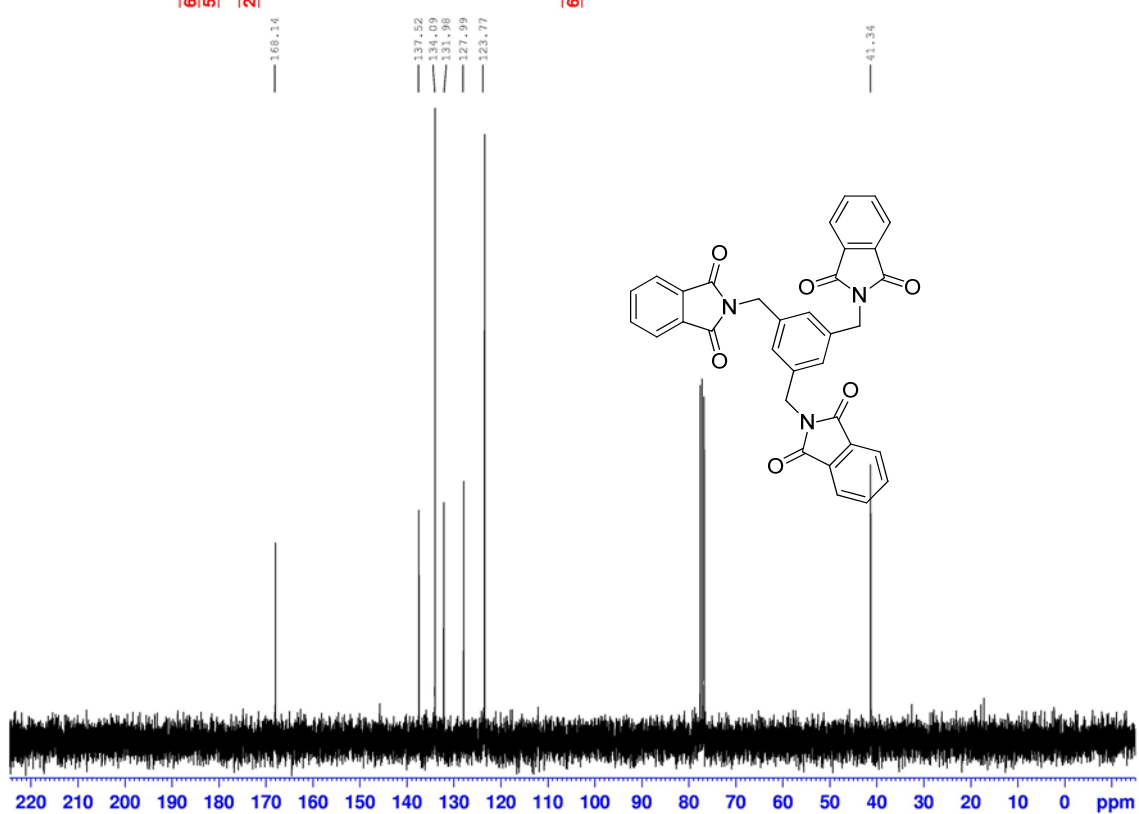
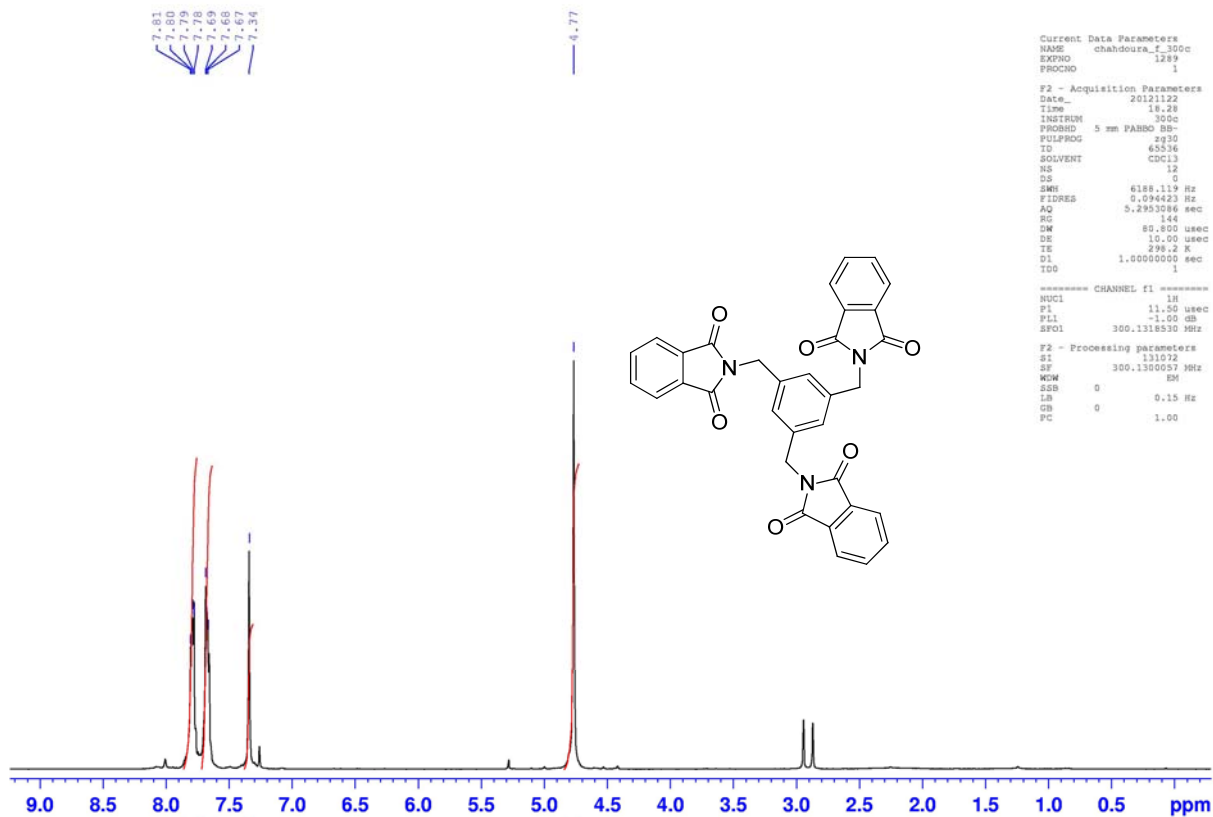
----- CHANNEL f1 -----  
 NUC1 13C  
 P1 11.00 usec  
 PL1 -1.00 dB  
 SFO1 300.1318330 MHz  
 F2 - Processing parameters  
 SI 131072  
 SF 300.1300277 MHz  
 WF 30  
 ZF 0  
 GB 0 0.1% Hz  
 CB 0  
 PC 1.00

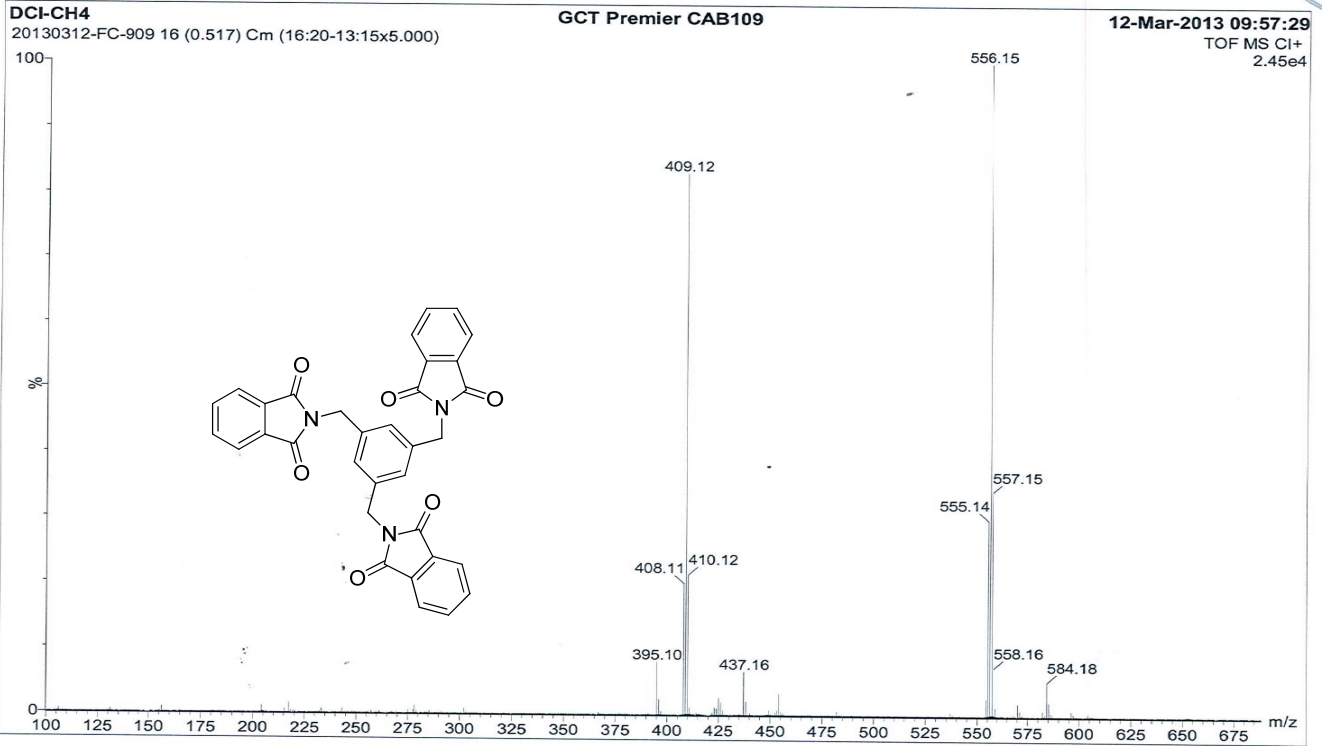


GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **a30**



$^1\text{H}$  NMR (300 MHz) (top)) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **a31**





**Elemental Composition Report**

**Single Mass Analysis**

Tolerance = 3.0 PPM / DBE: min = -1.5, max = 50.0  
Element prediction: Off

Monoisotopic Mass, Odd and Even Electron Ions

744 formula(e) evaluated with 6 results within limits (all results (up to 1000) for each mass)

Elements Used:

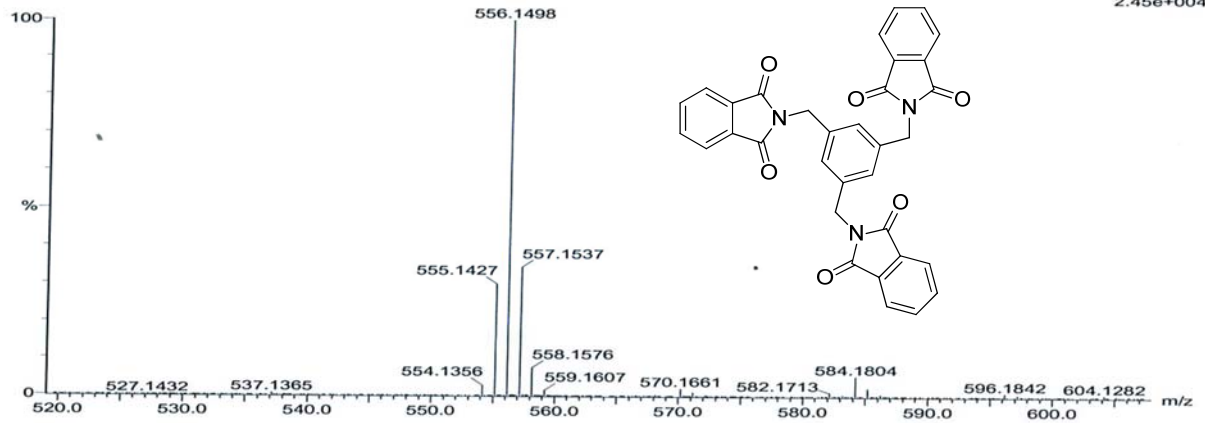
C: 0-60 H: 0-60 N: 0-15 O: 0-8

DCI-CH4

20130312-FC-909 16 (0.517) Cm (16:20-13:15x5.000)

GCT Premier CAB109

12-Mar-2013 09:57:29  
TOF MS CI+  
2.45e+004



Minimum:  
Maximum:

1.3      3.0      -1.5  
25.5      23.2      45.7      52.2      30.0      70.3      293.7

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	Formula
556.1498	556.1482	1.6	2.9	25.5	11.1	C29 H18 N9 O4
	556.1495	0.3	0.5	25.0	23.2	C31 H20 N6 O5
	556.1495	0.3	0.5	30.5	45.7	C30 H14 N13
	556.1509	-1.1	-2.0	24.5	52.2	C33 H22 N3 O6
	556.1509	-1.1	-2.0	30.0	70.3	C32 H16 N10 O
	556.1514	-1.6	-2.9	17.5	293.7	C18 H18 N15 O7

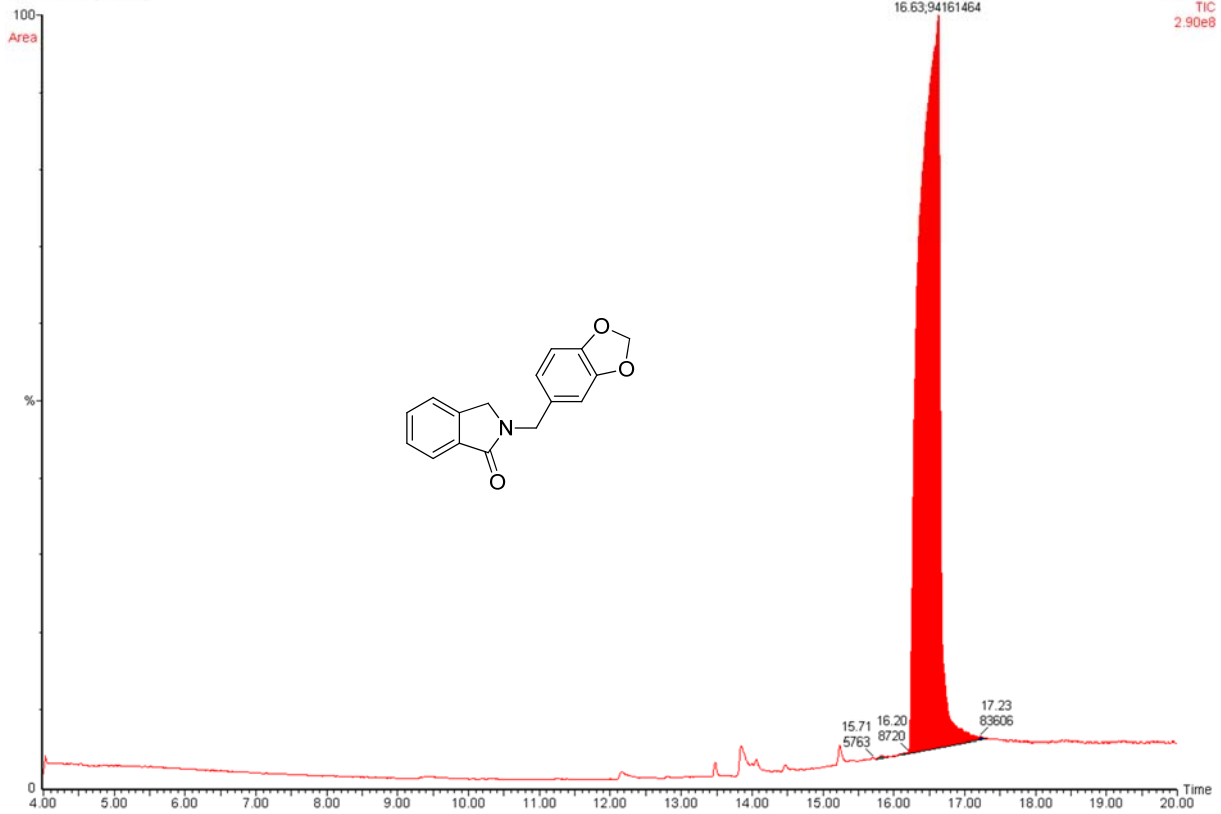
<sup>1</sup>H NMR (300 MHz) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (top) spectra in CDCl<sub>3</sub>, and low and high resolution DCI(CH<sub>4</sub>)-MS (bottom) for **a32**



1081-  
fc-1081-3 Sm (Mn, 1x3)

, 22-Mar-2013 + 13:00:36

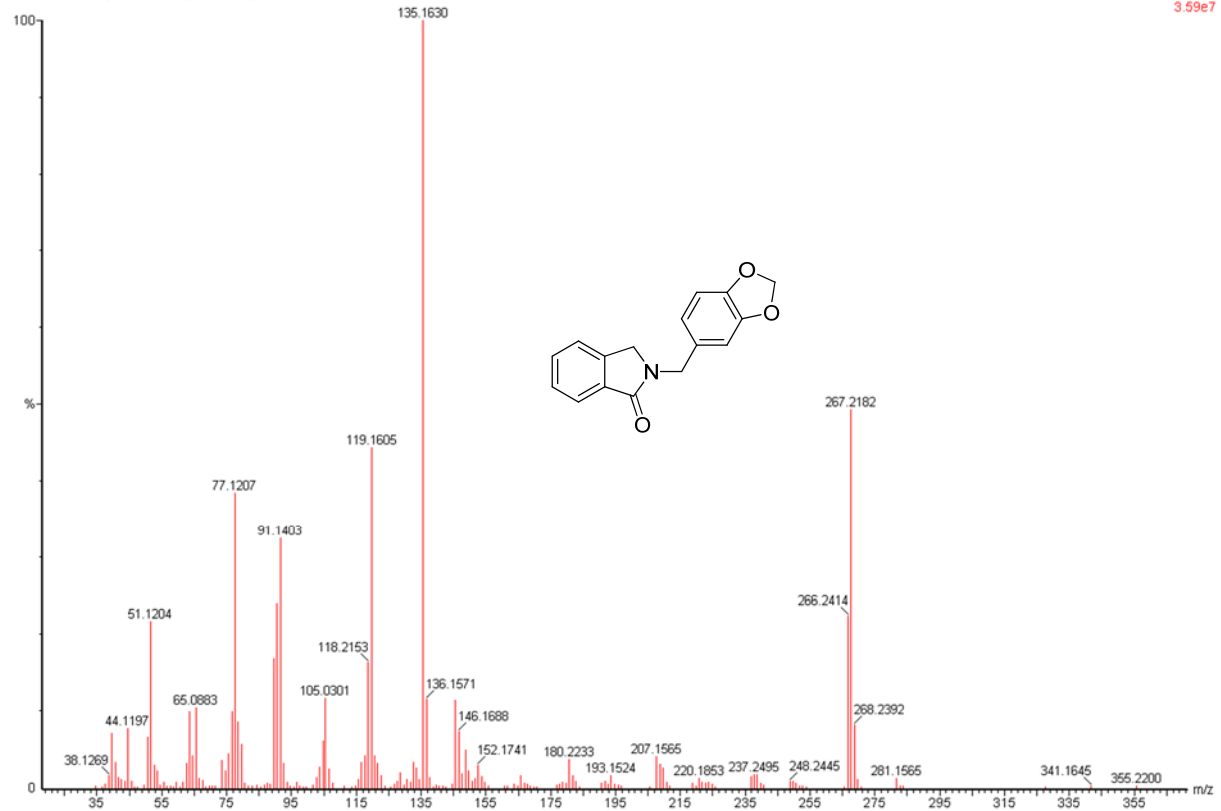
Scan EI+  
TIC  
2.90e8

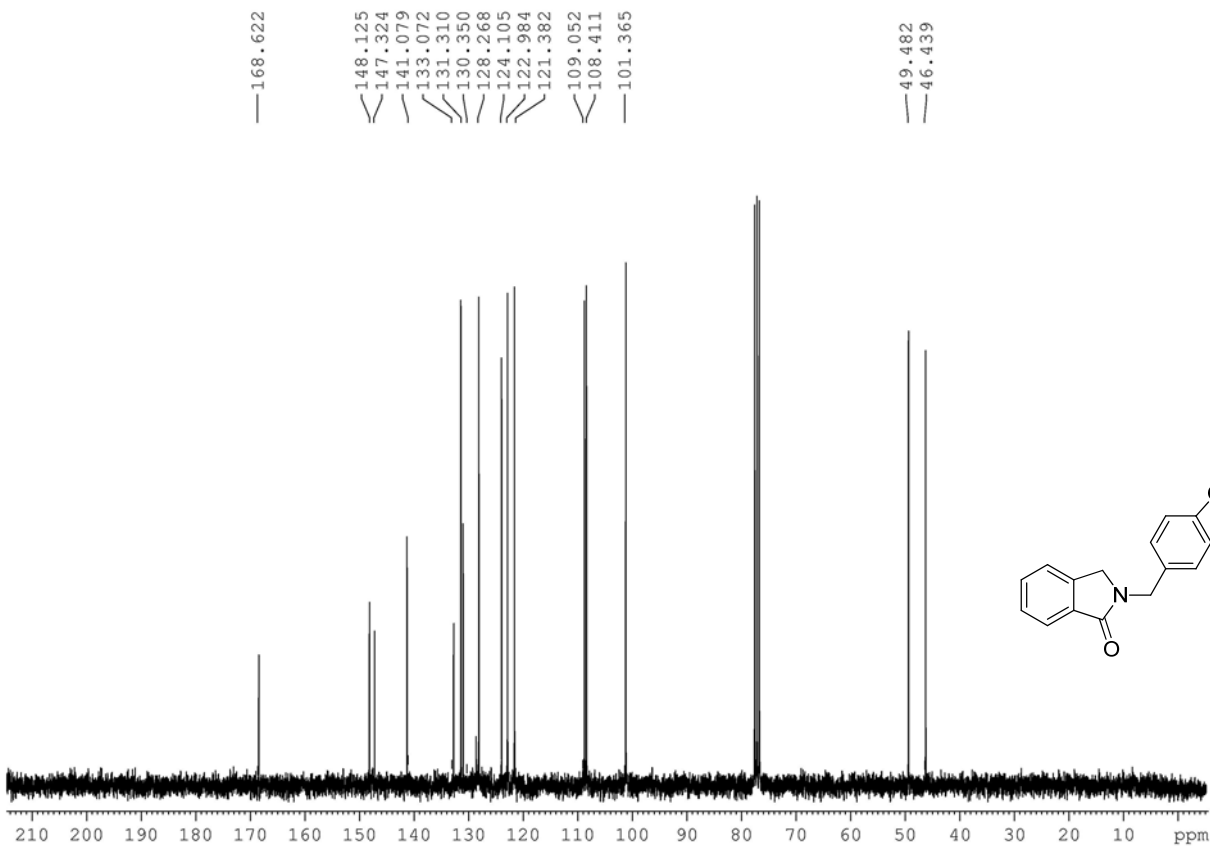
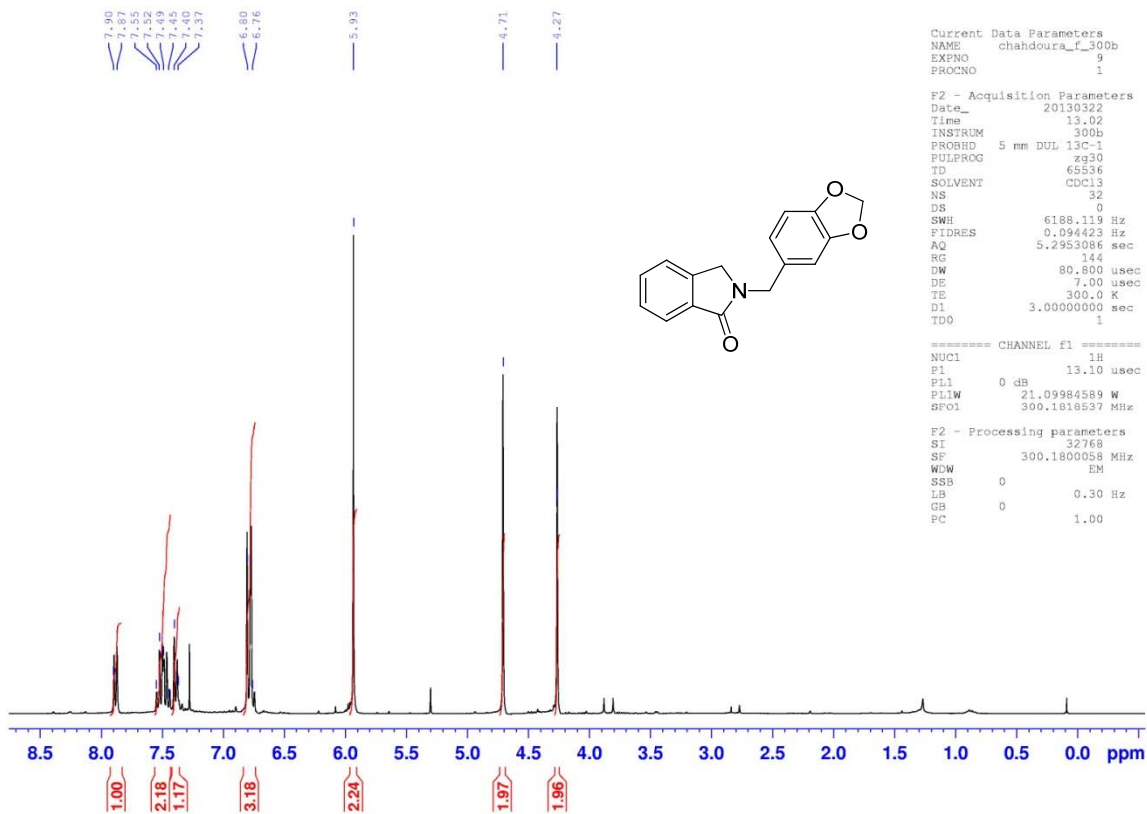


1081-  
fc-1081-3 2511 (16.558) Cm (2459:2511)

, 22-Mar-2013 + 13:00:36

Scan EI+  
3.59e7





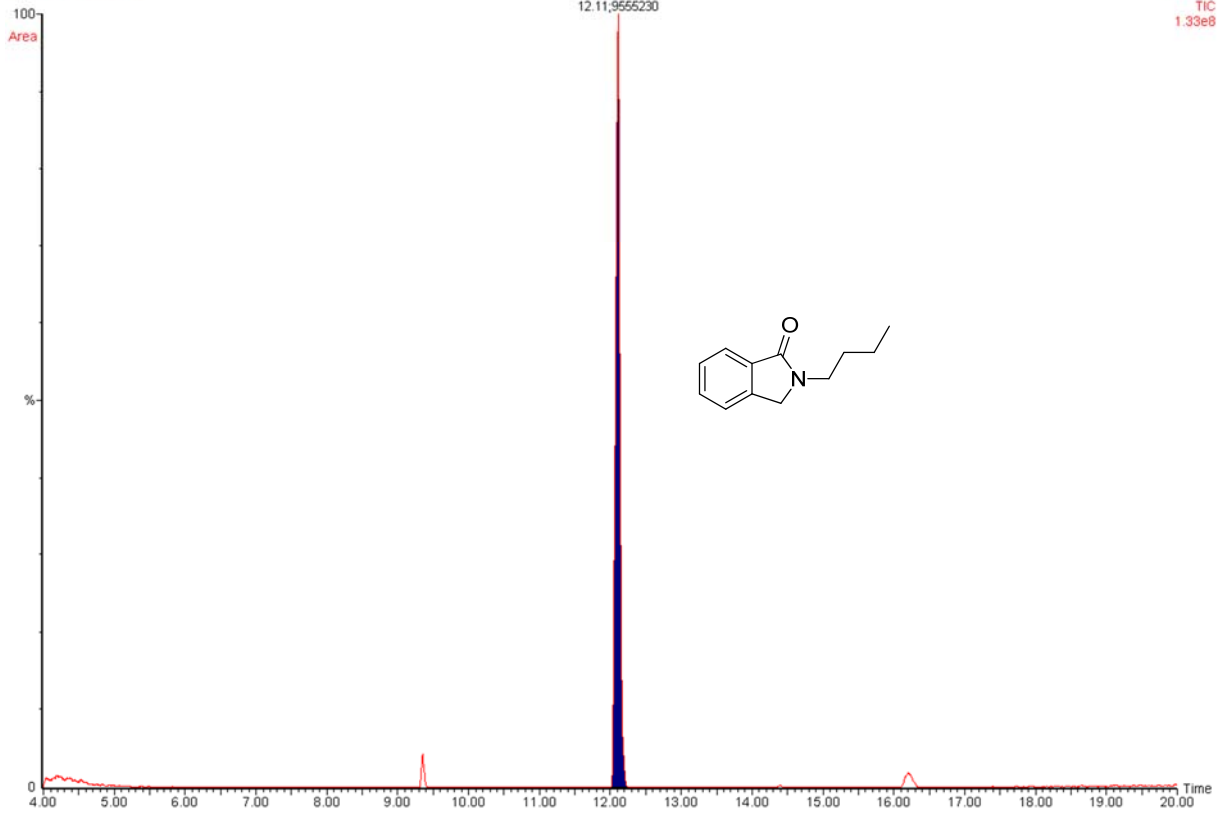
GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **h8**

1072

fc-1072- Sm (Mn, 1x3)

, 08-Mar-2013 + 10:13:53

Scan EI+  
TIC  
1.33e8

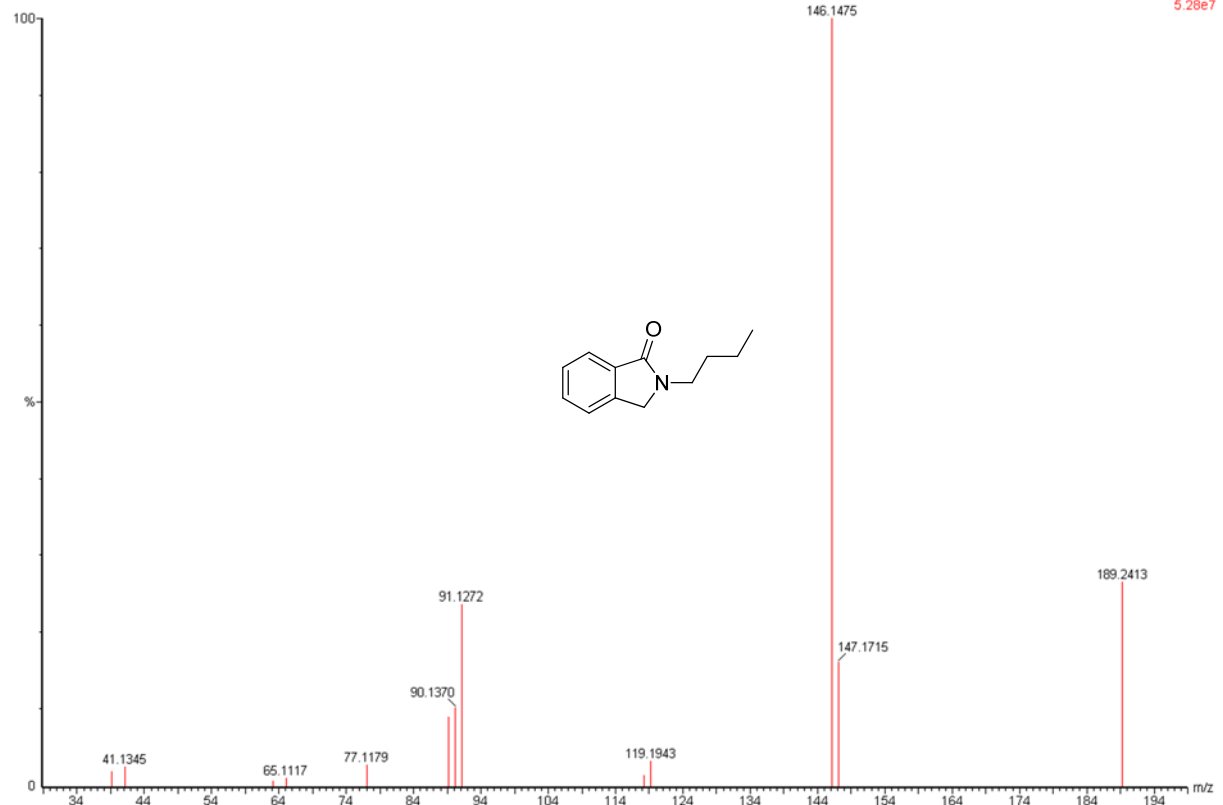


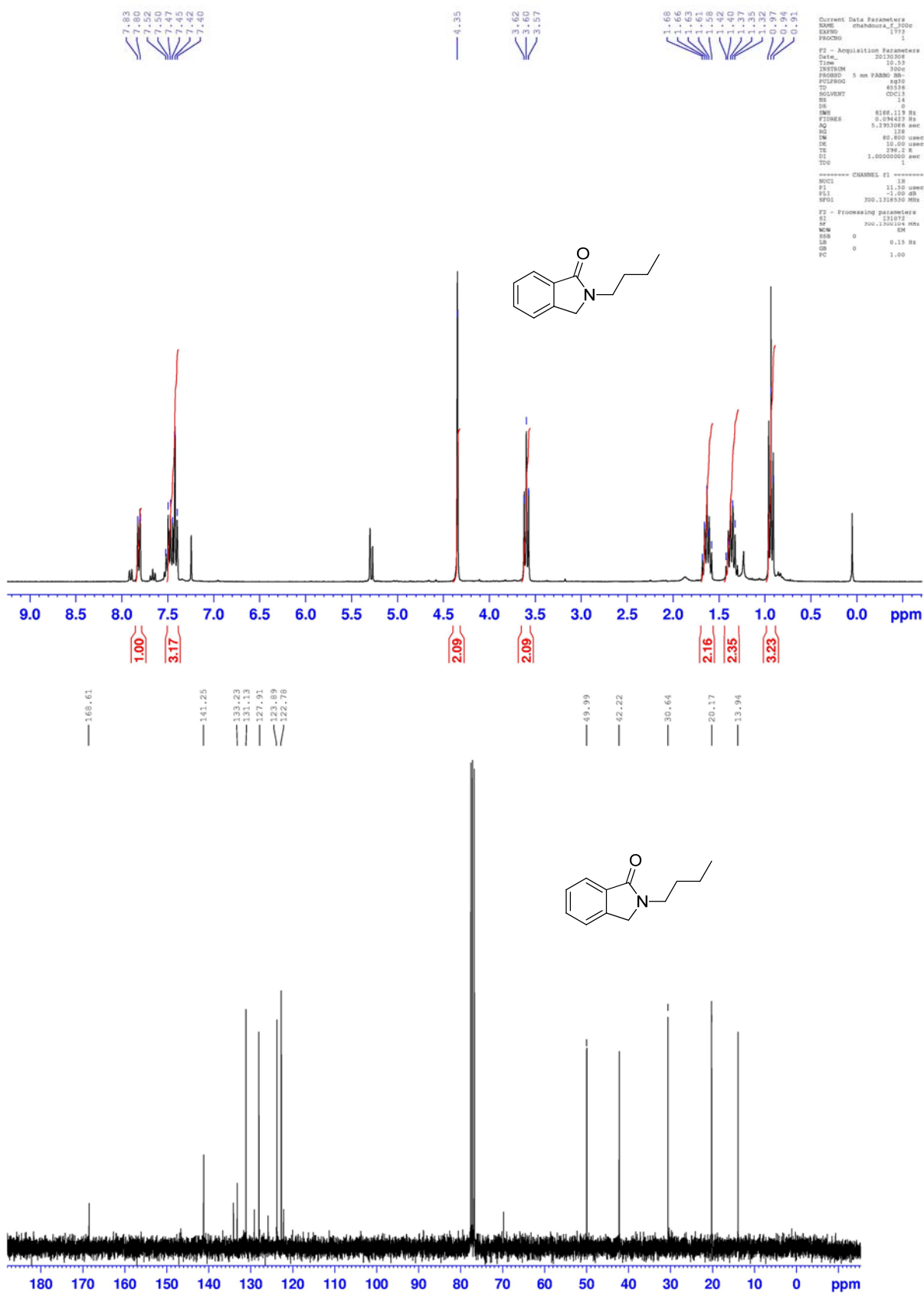
1072

fc-1072- 1624 (12.122) Cm (1616:1629)

, 08-Mar-2013 + 10:13:53

Scan EI+  
5.28e7

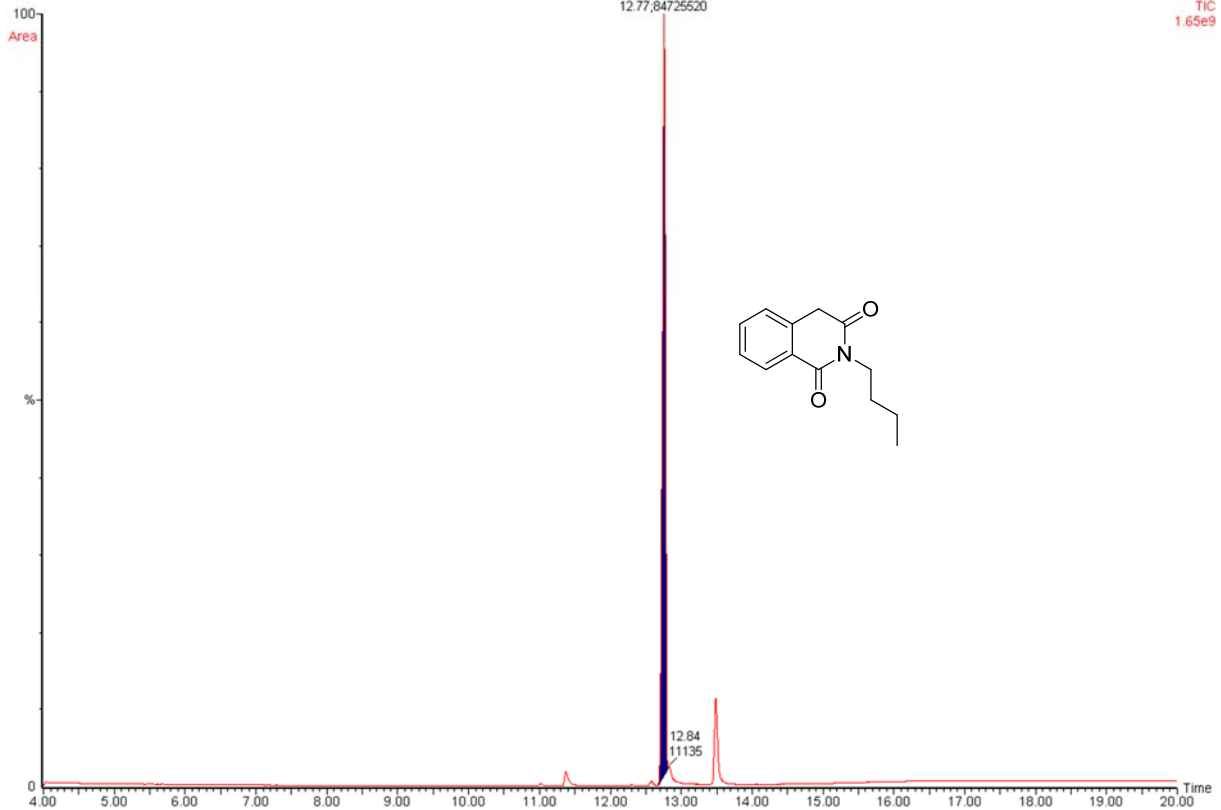




GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **i9**

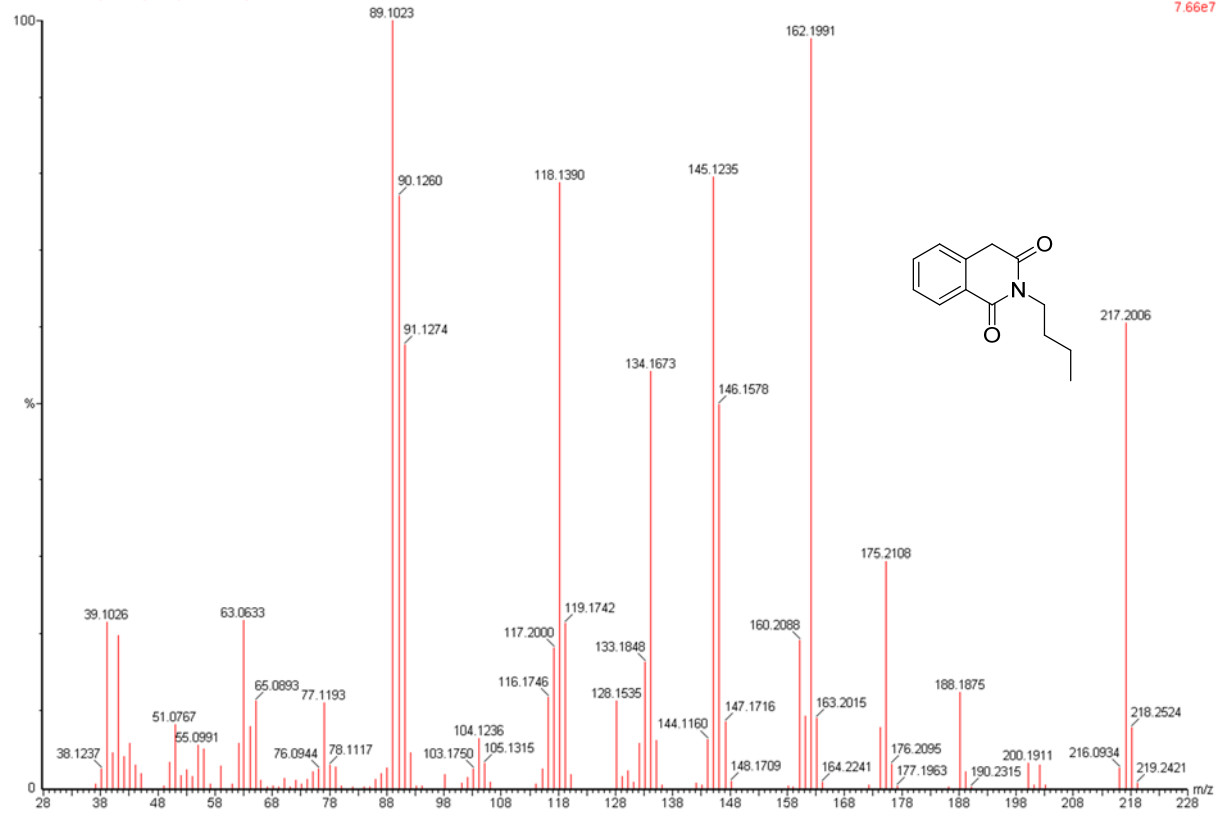
1095  
fc-1095- Sm (Mn, 1x3)

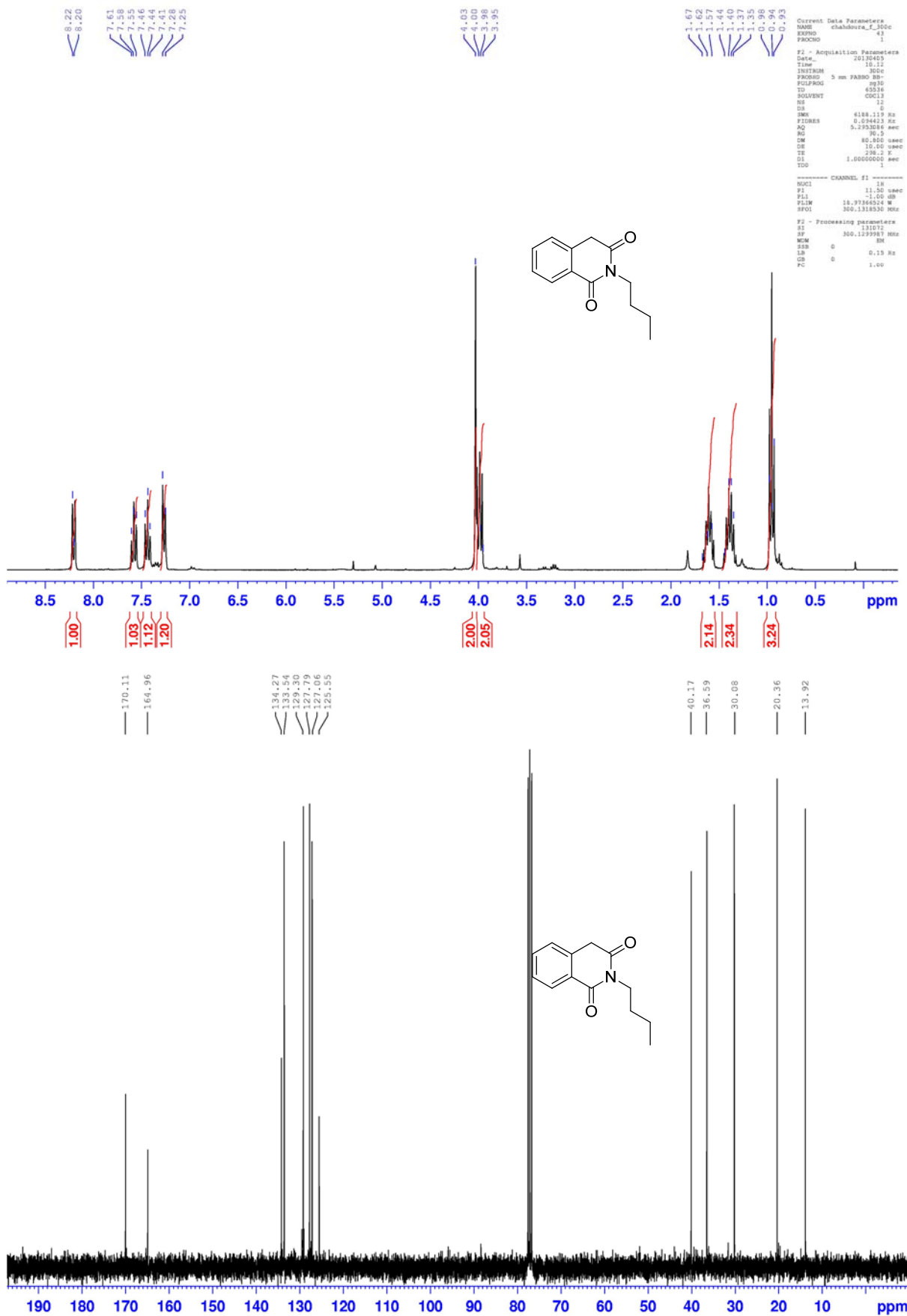
, 05-Apr-2013 + 10:33:01  
Scan EI+  
TIC  
1.65e9



1095  
fc-1095- 1756 (12.782) Cm (1740:1760)

, 05-Apr-2013 + 10:33:01  
Scan EI+  
7.66e7





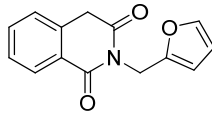
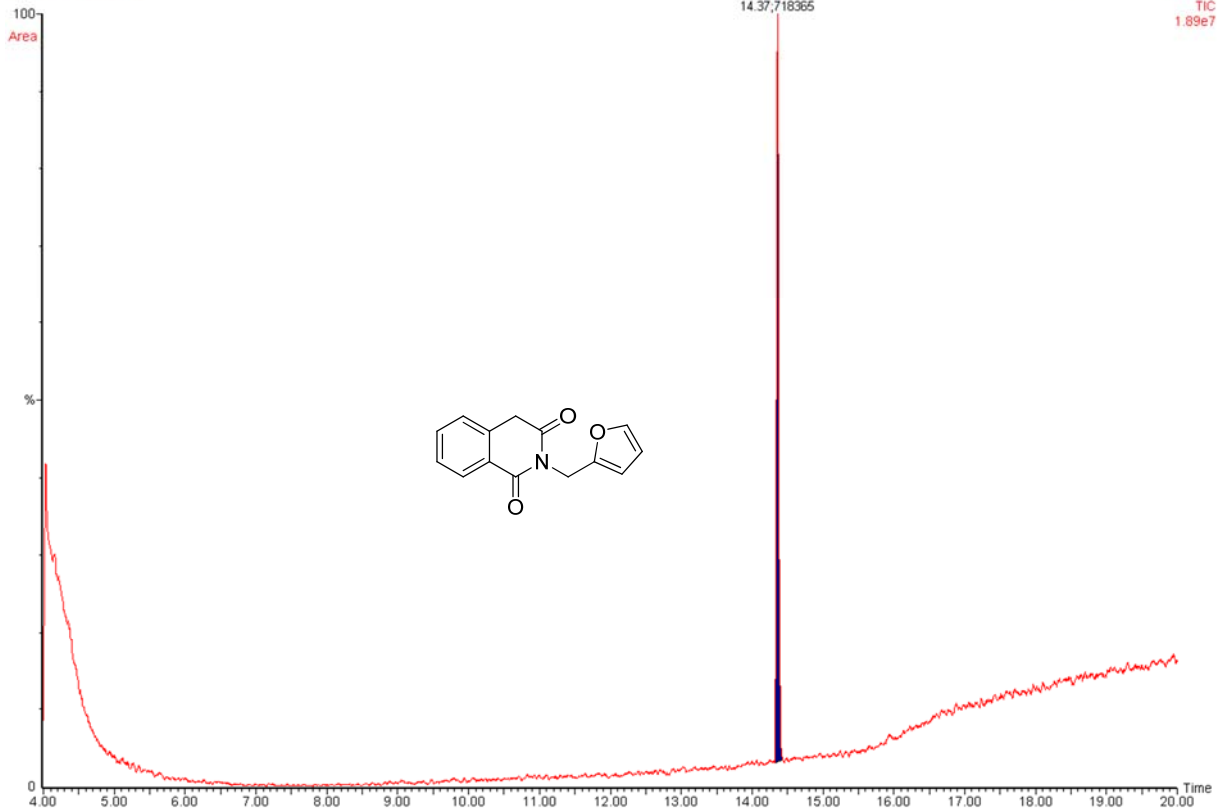
GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **j9**

1105

fc-1105- Sm (Mn, 1x3)

, 16-Apr-2013 + 10:41:16

Scan EI+  
TIC  
1.89e7

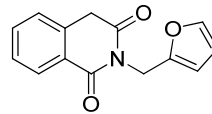
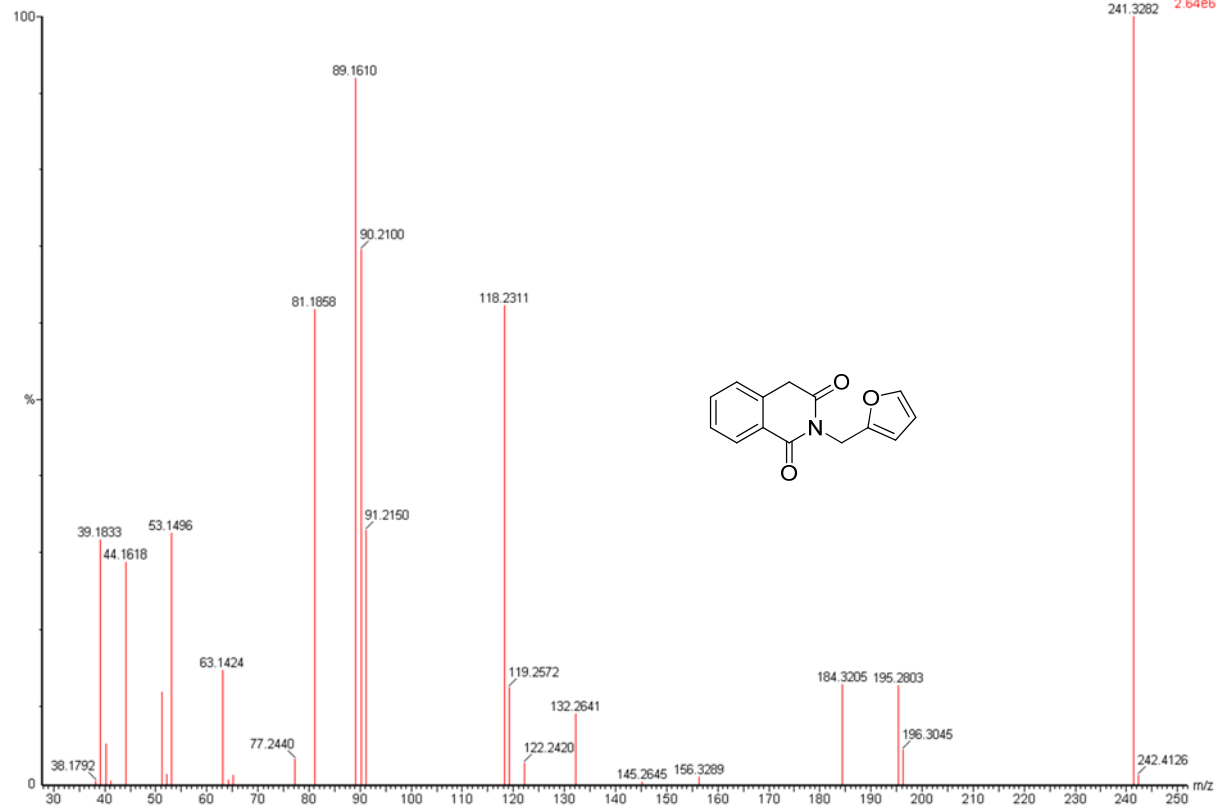


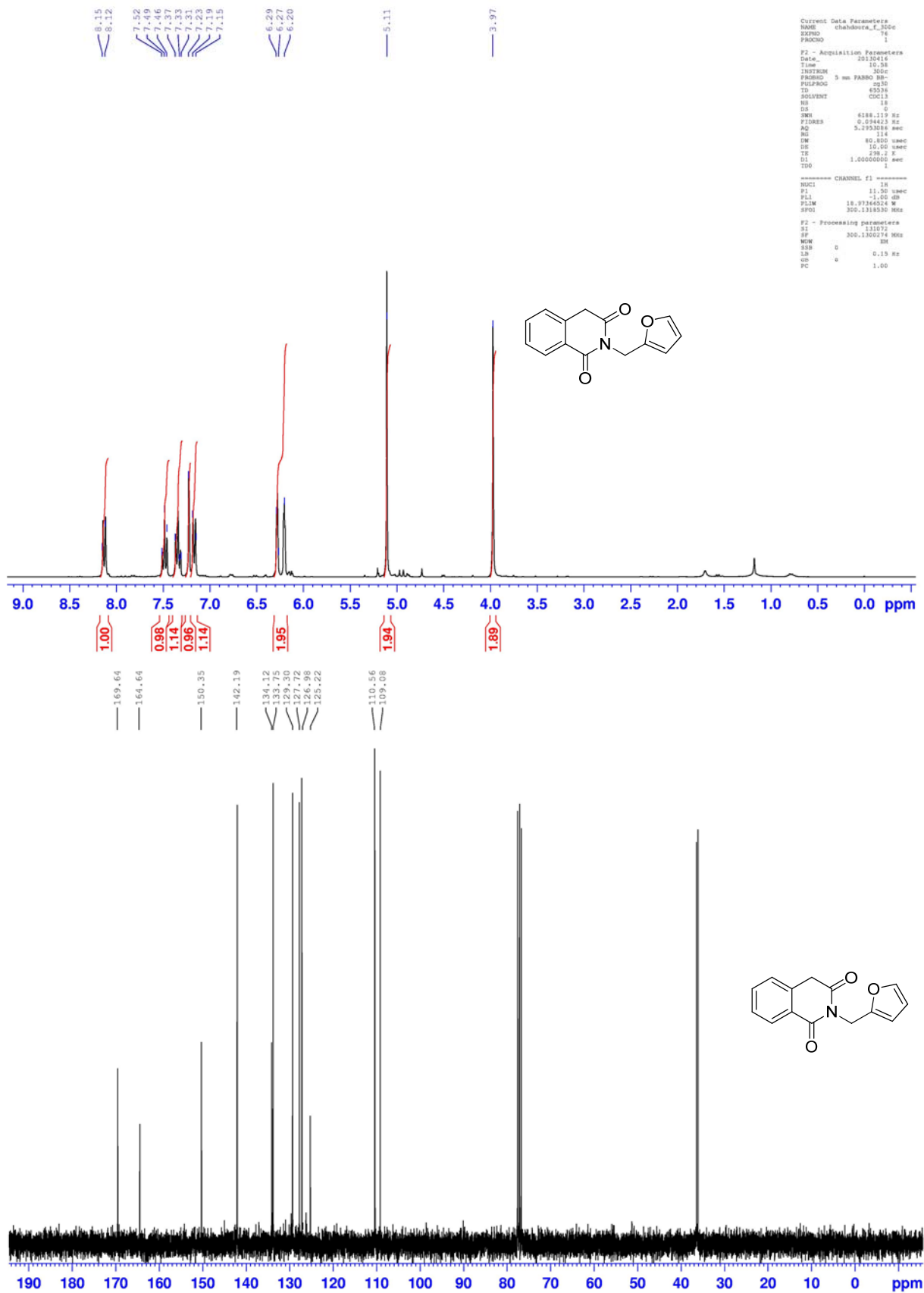
1105

fc-1105- 2072 (14.363) Cm (2069:2077)

, 16-Apr-2013 + 10:41:16

Scan EI+  
2.64e6





GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **k24**



1018

fc-1018-3 Sm (Mn, 1x3)

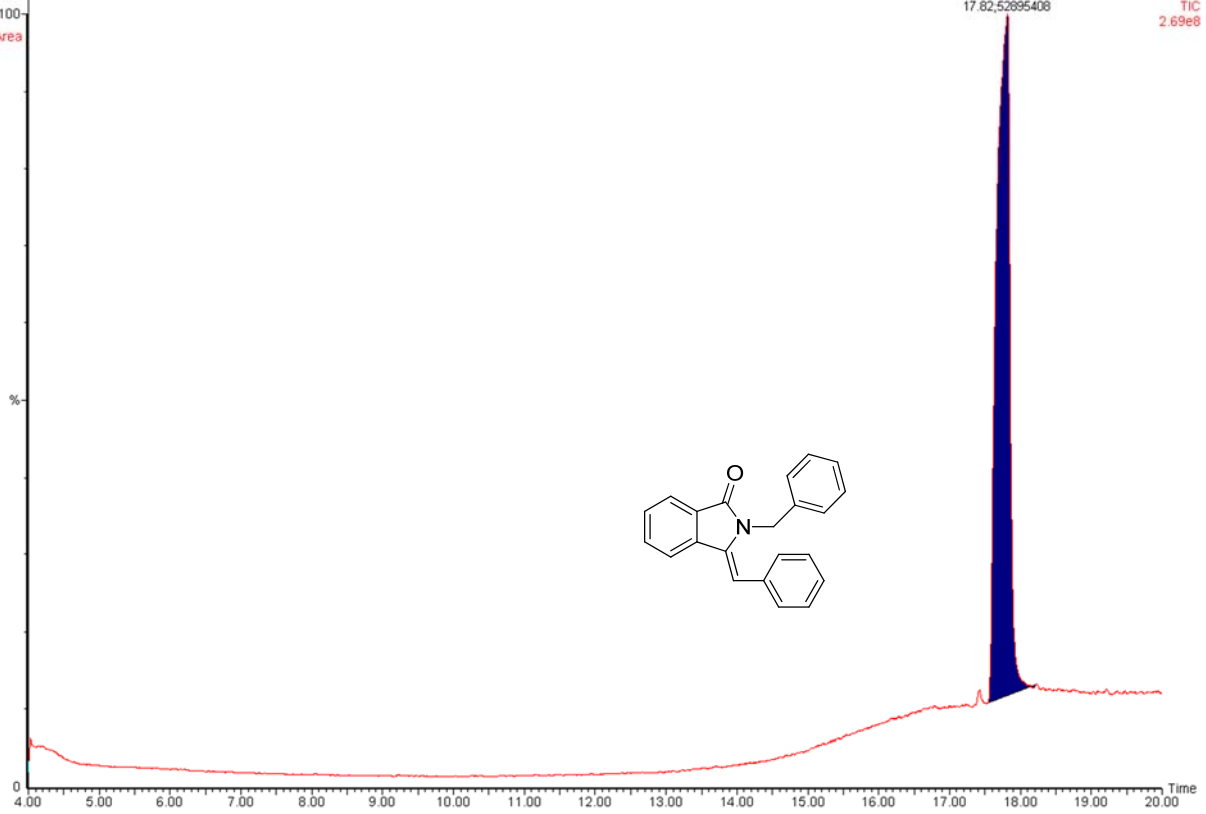
, 30-Jan-2013 + 15:41:38

Scan EI+

Area

17.82,52895408

TIC  
2.69e8



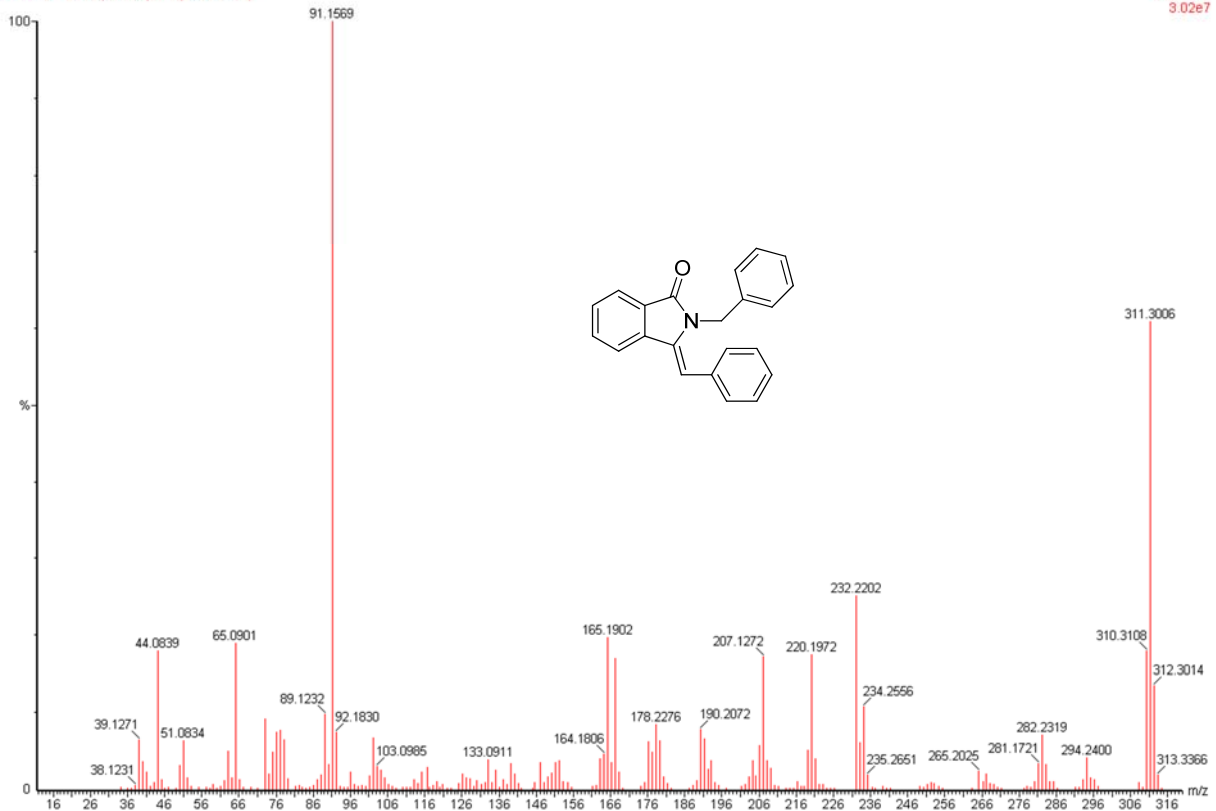
1018

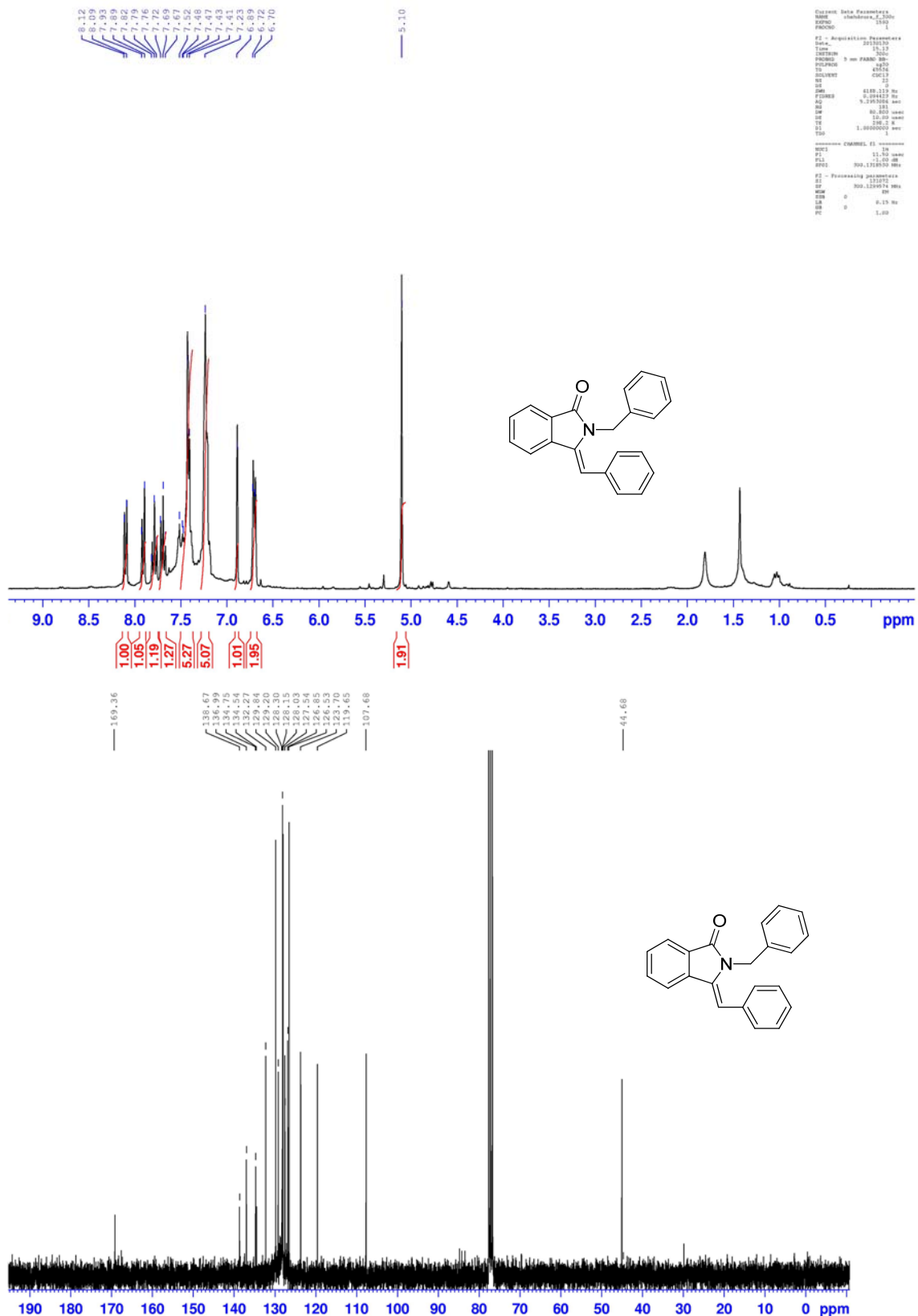
fc-1018-3 2764 (17.824) Cm (2723:2780)

, 30-Jan-2013 + 15:41:38

Scan EI+

3.02e7





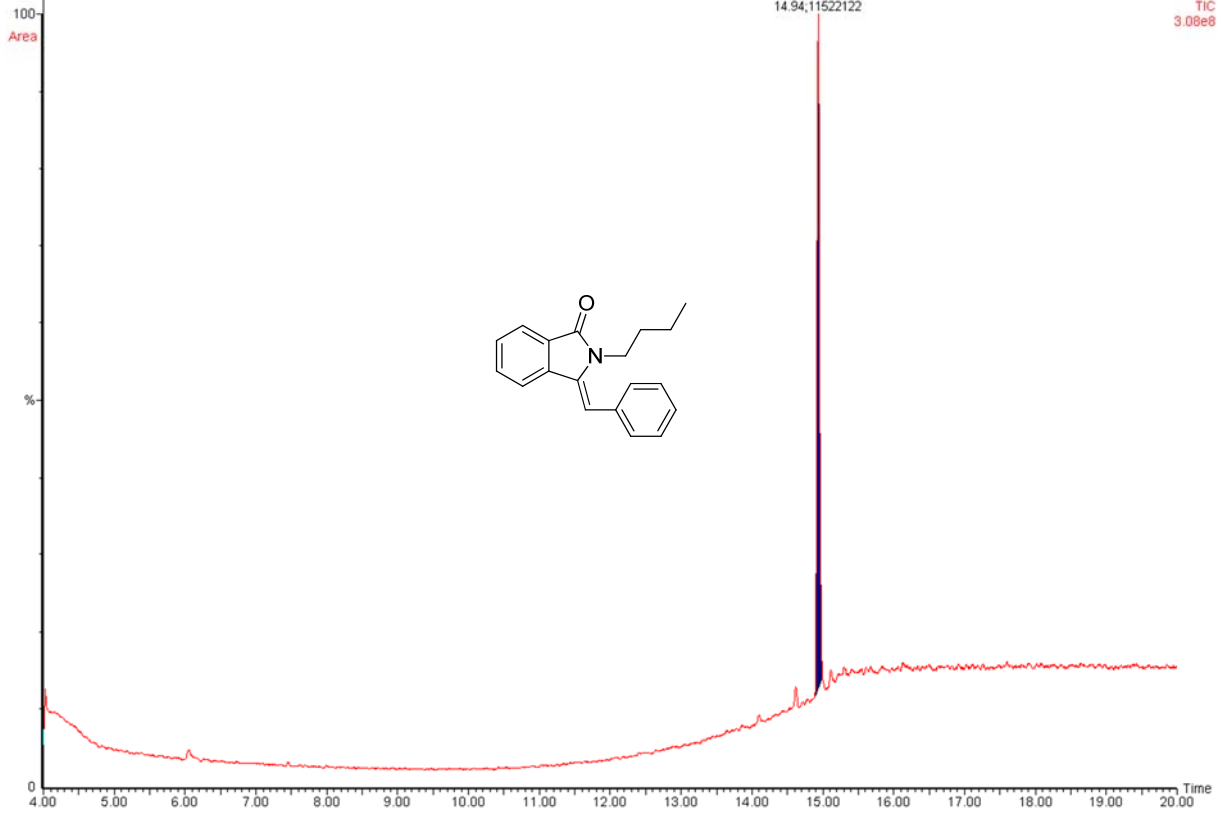
GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **b33**

1022

fc-1022-2 Sm (Mn, 1x3)

, 06-Feb-2013 + 11:47:37

Scan EI+  
TIC  
3.08e8

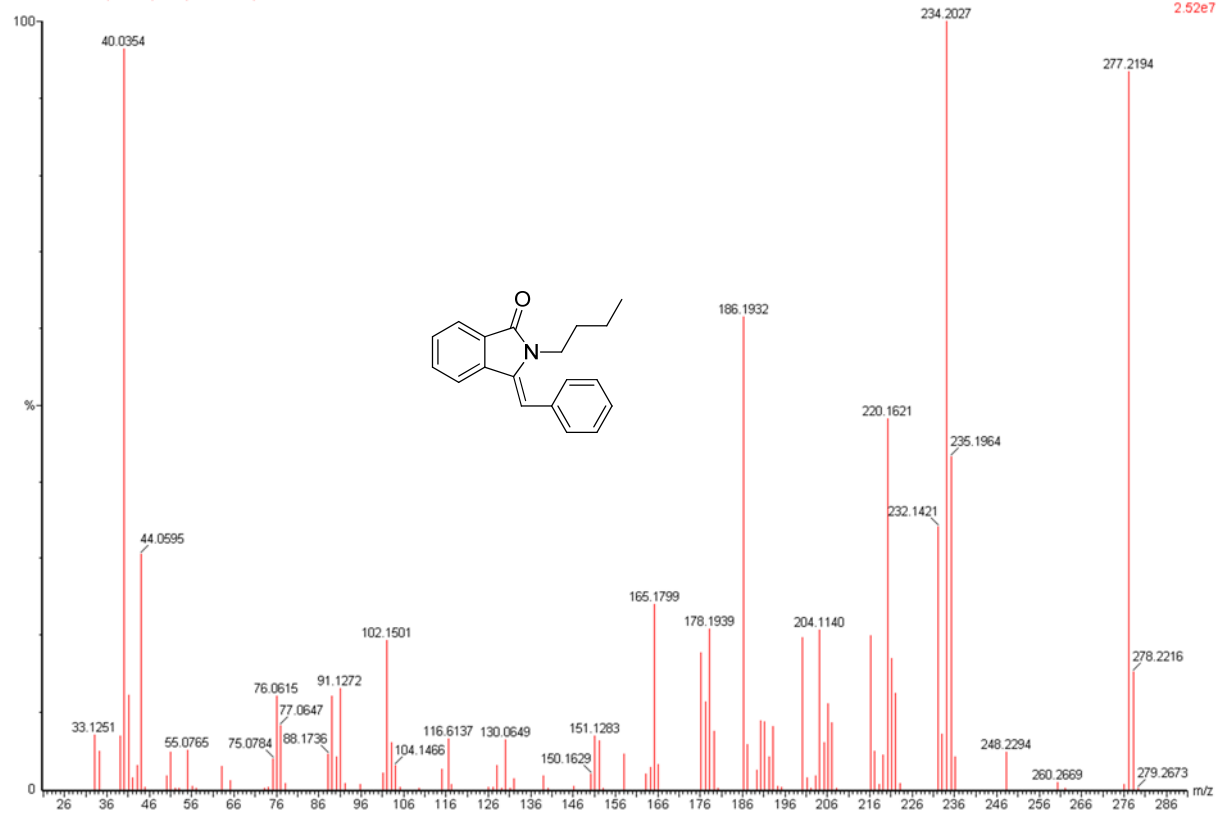


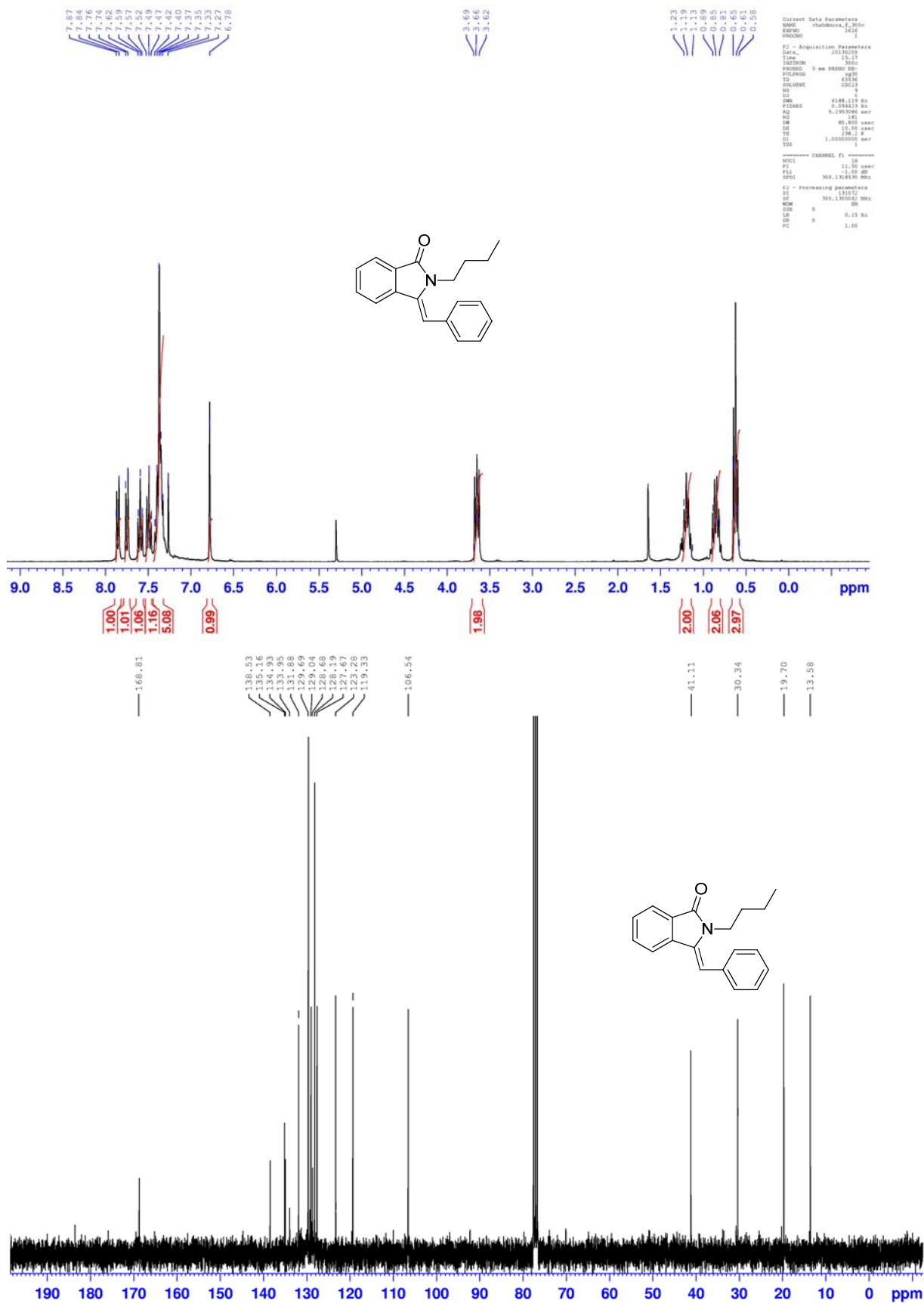
1022

fc-1022-2 2188 (14.943) Cm (2182:2191)

, 06-Feb-2013 + 11:47:37

Scan EI+  
2.52e7





GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **133**

1026

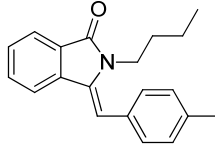
fc-1026-2 Sm (Mn, 1x3)

, 07-Feb-2013 + 17:31:59

Scan EI+

TIC

1.66e9



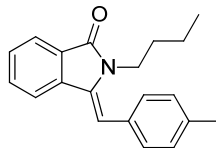
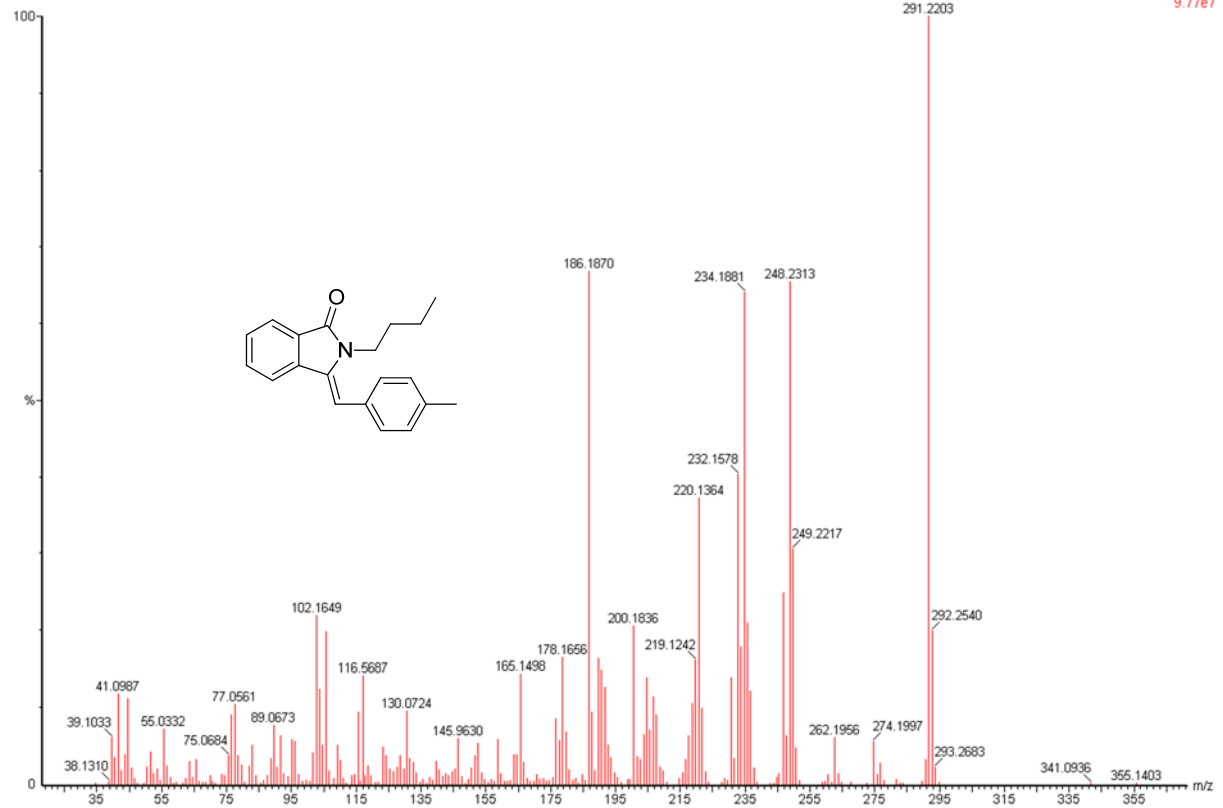
1026

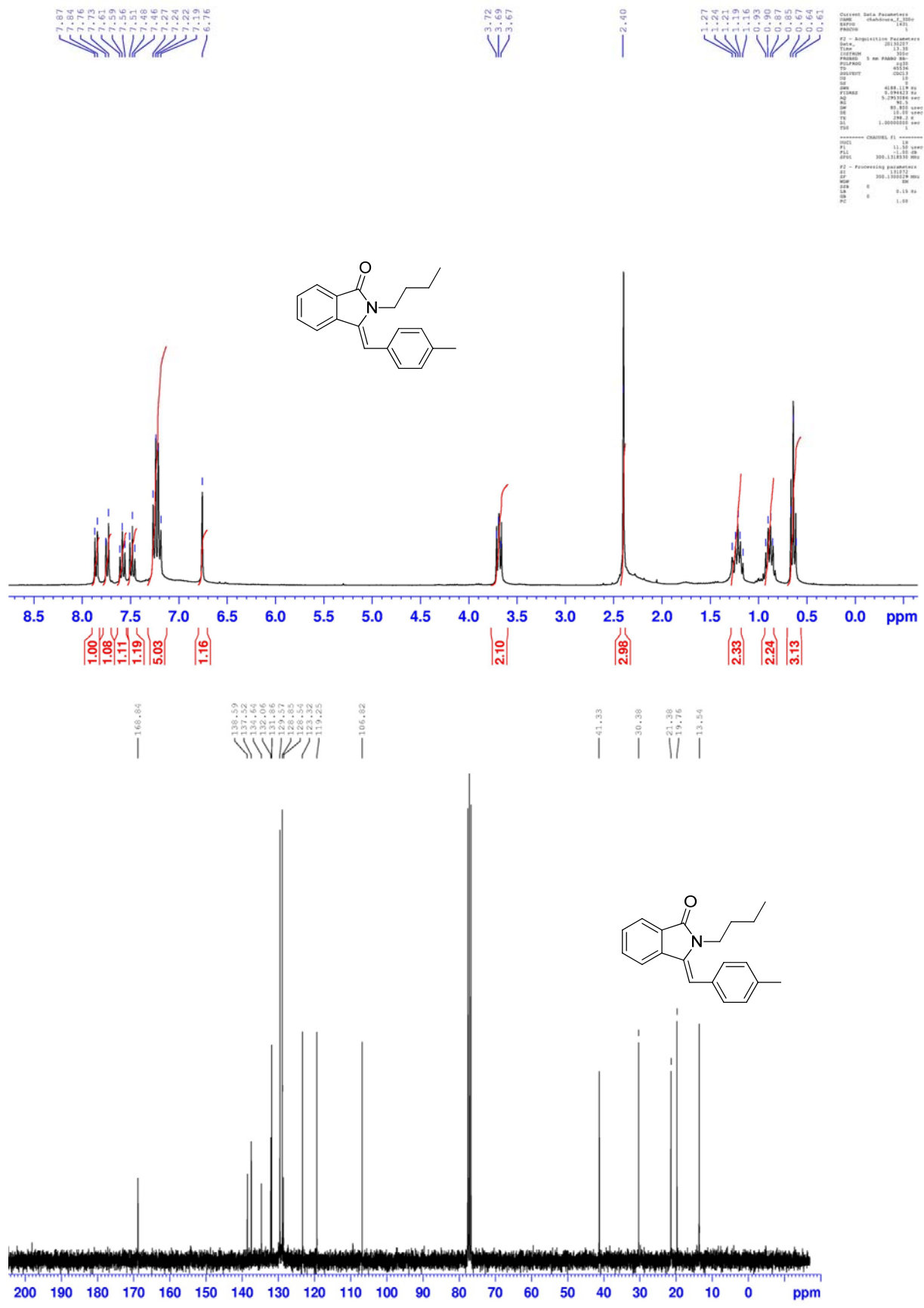
fc-1026-2 2383 (15.918) Cm (2362:2389)

, 07-Feb-2013 + 17:31:59

Scan EI+

9.77e7





GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **13a**

1027

fc-1027- Sm (Mn, 1x3)

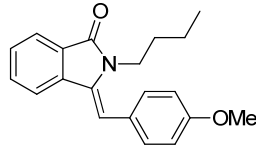
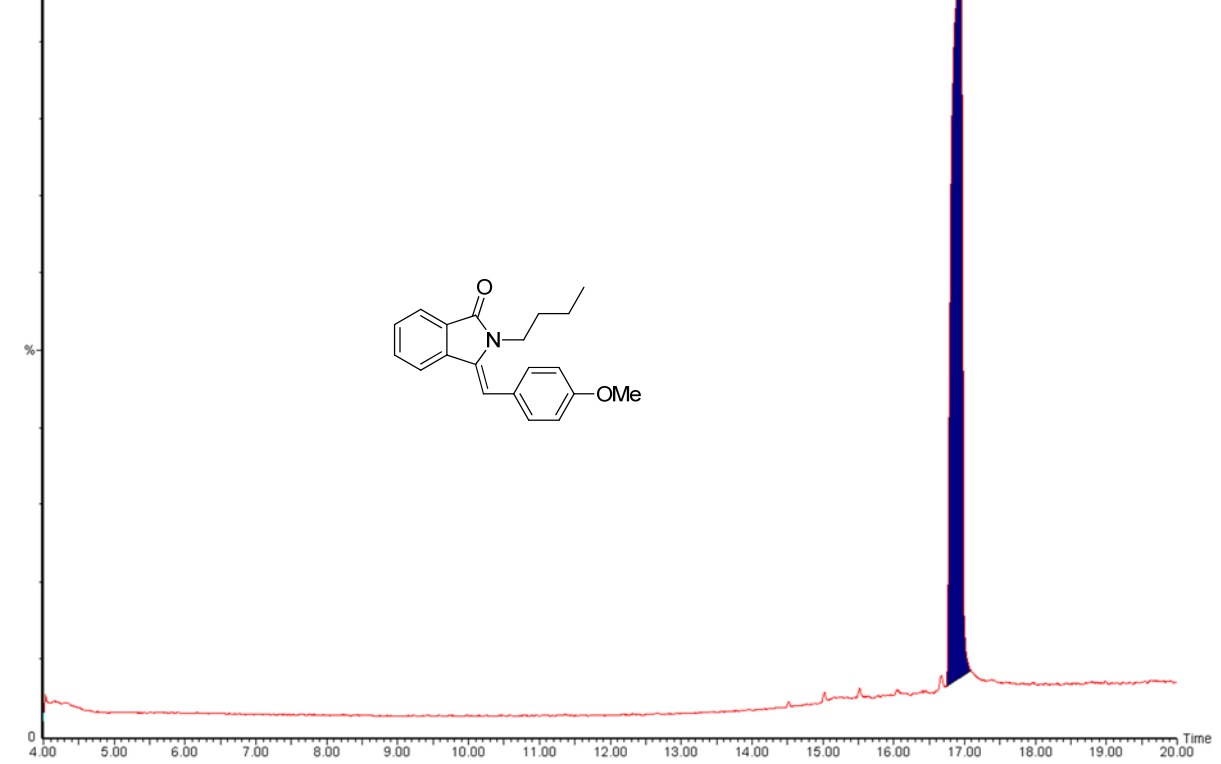
, 07-Feb-2013 + 18:07:11

Scan EI+

TIC

4.63e8

Area



1027

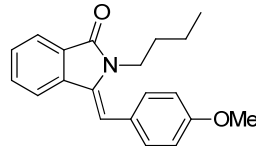
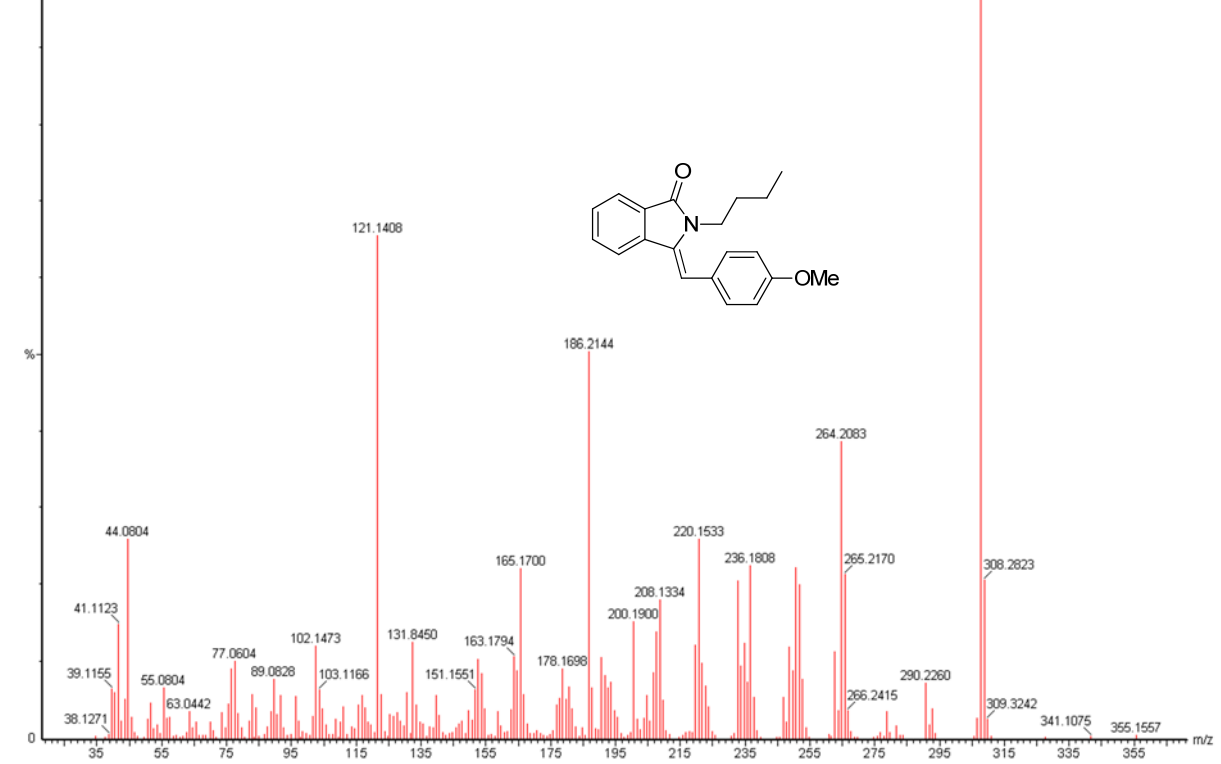
fc-1027- 2590 (16.953) Cm (2558:2593)

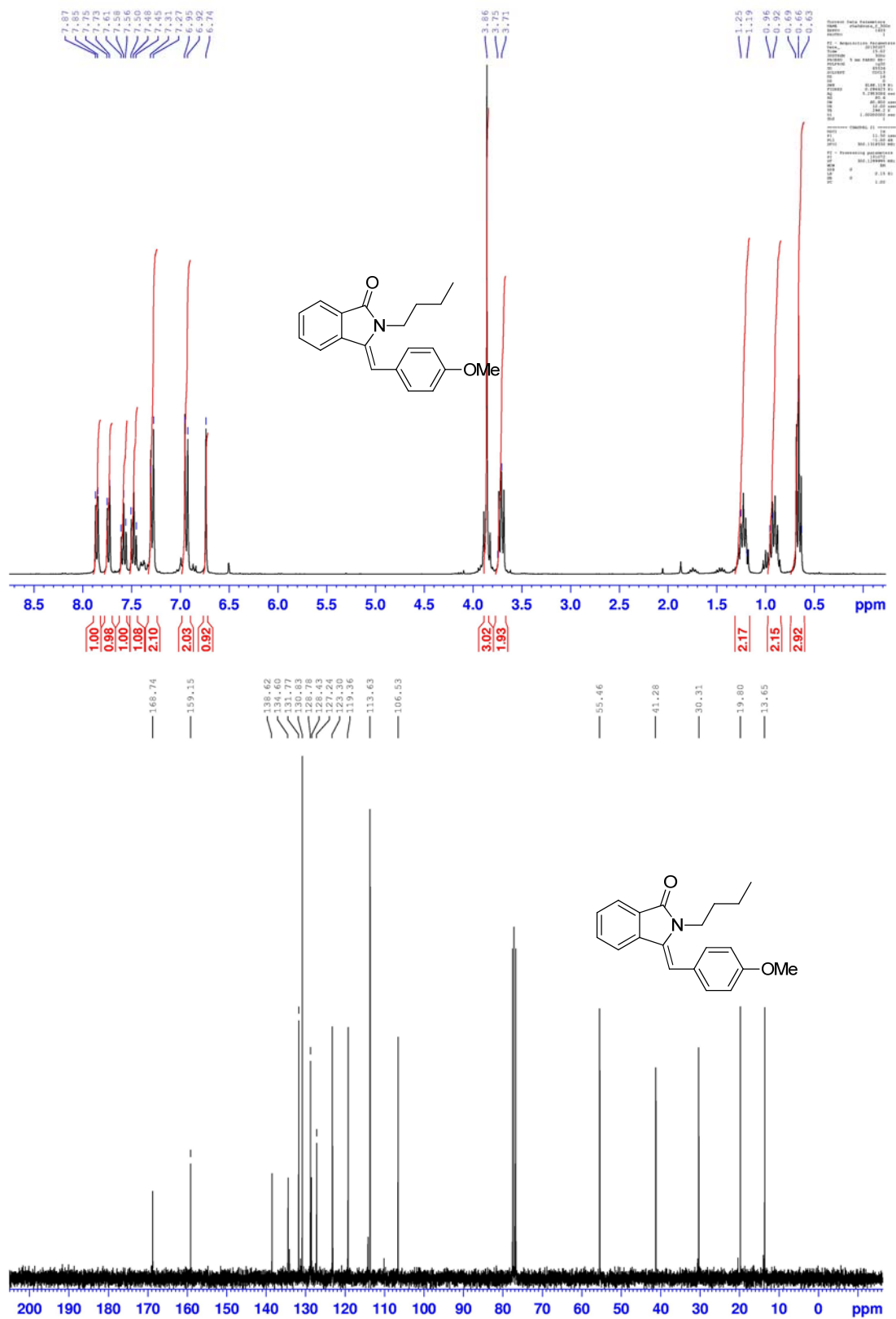
, 07-Feb-2013 + 18:07:11

Scan EI+

3.68e7

%



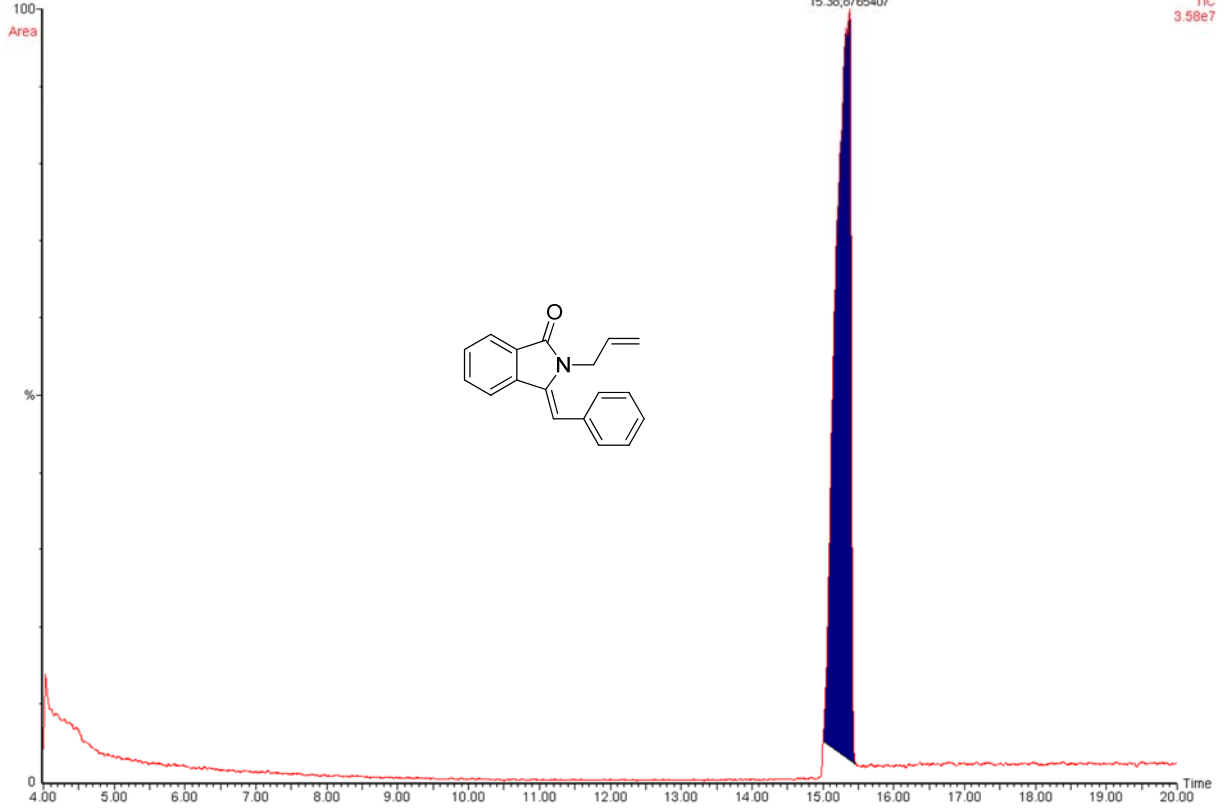


GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **135**



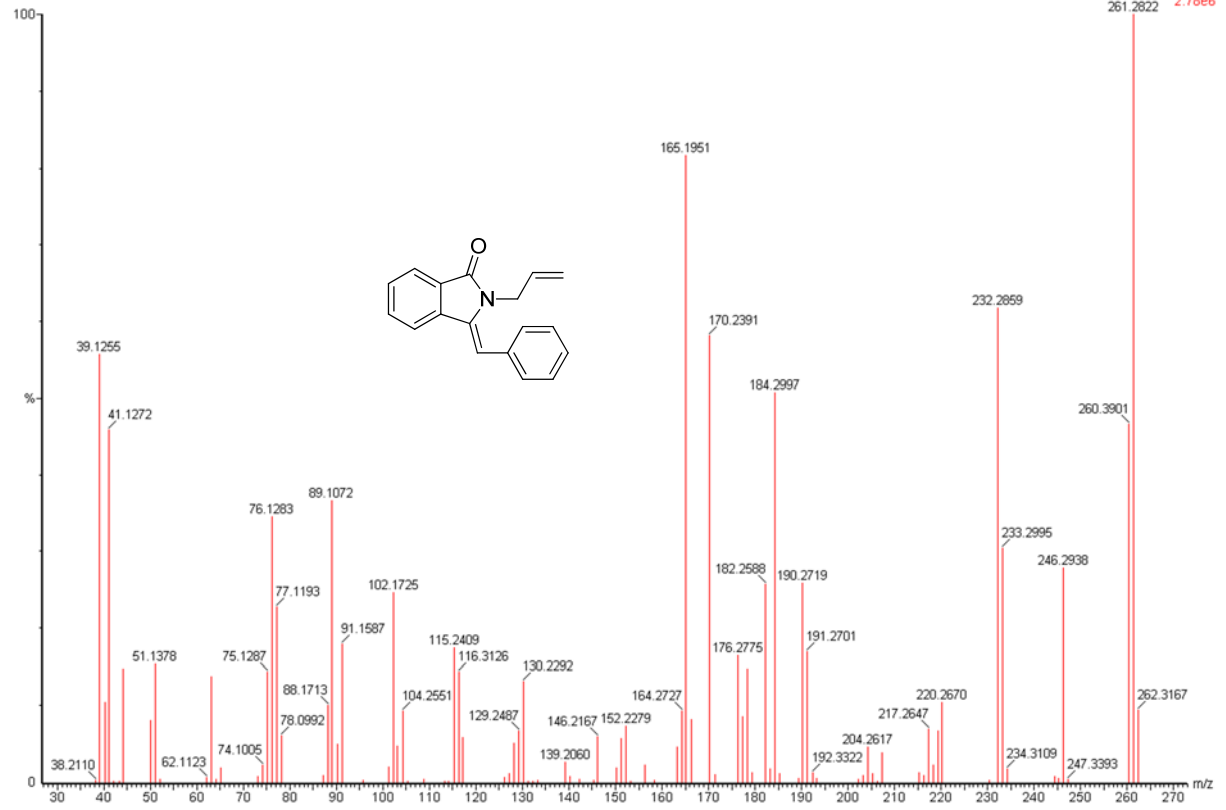
1067  
fc-1067- Sm (Mn, 1x3)

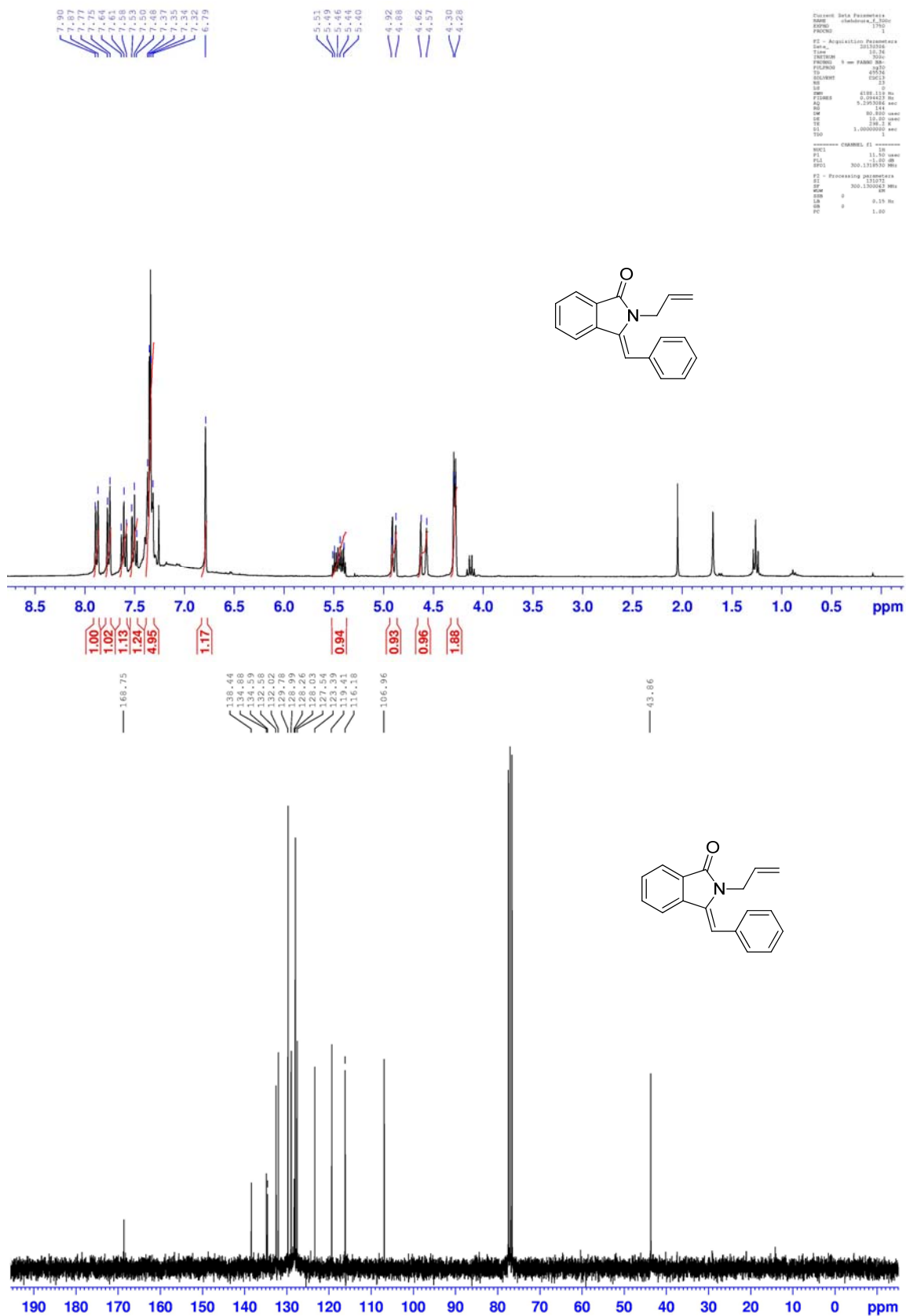
, 06-Mar-2013 + 10:58:26  
Scan EI+  
TIC  
3.58e7



1067  
fc-1067- 2264 (15.323) Cm (2237:2270)

, 06-Mar-2013 + 10:58:26  
Scan EI+  
2.78e6





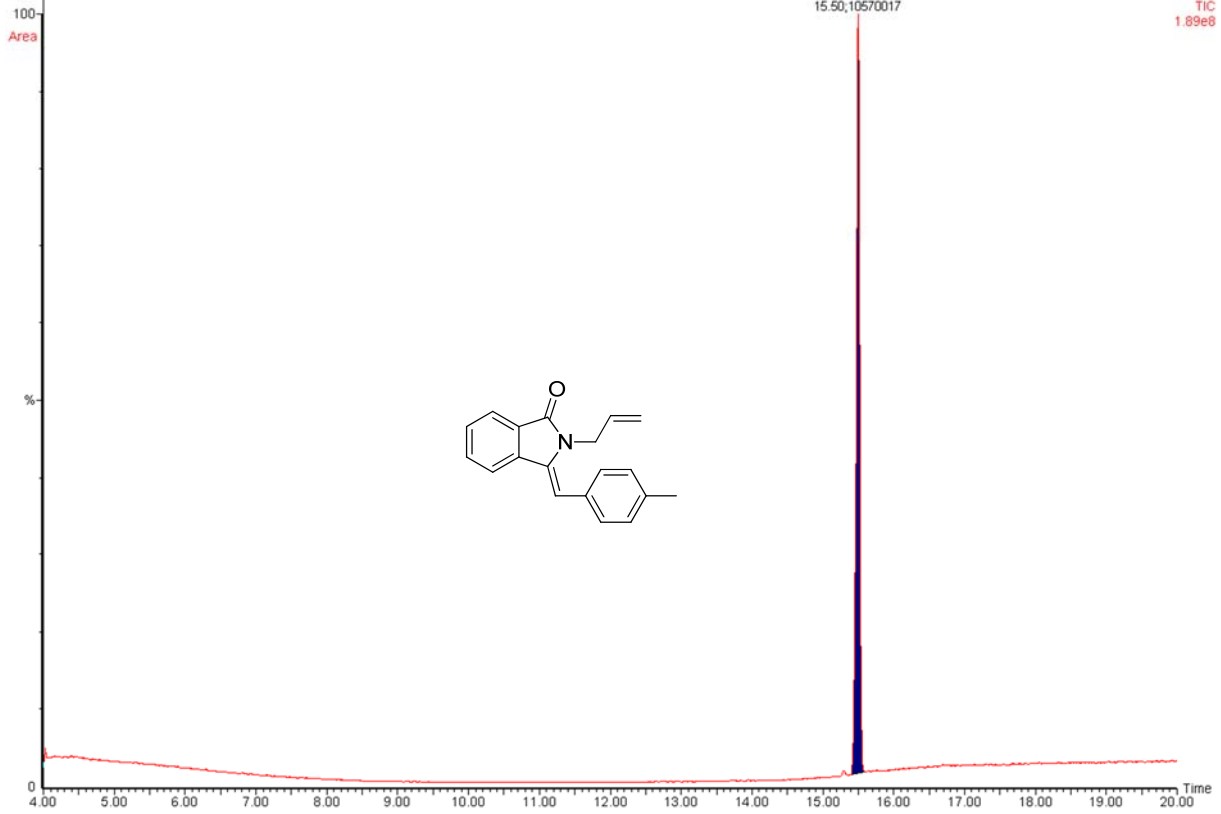
GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **m33**

1068

fc-1068-bis-1 Sm (Mn, 1x3)

, 07-Mar-2013 + 12:37:54

Scan EI+  
TIC  
1.89e8

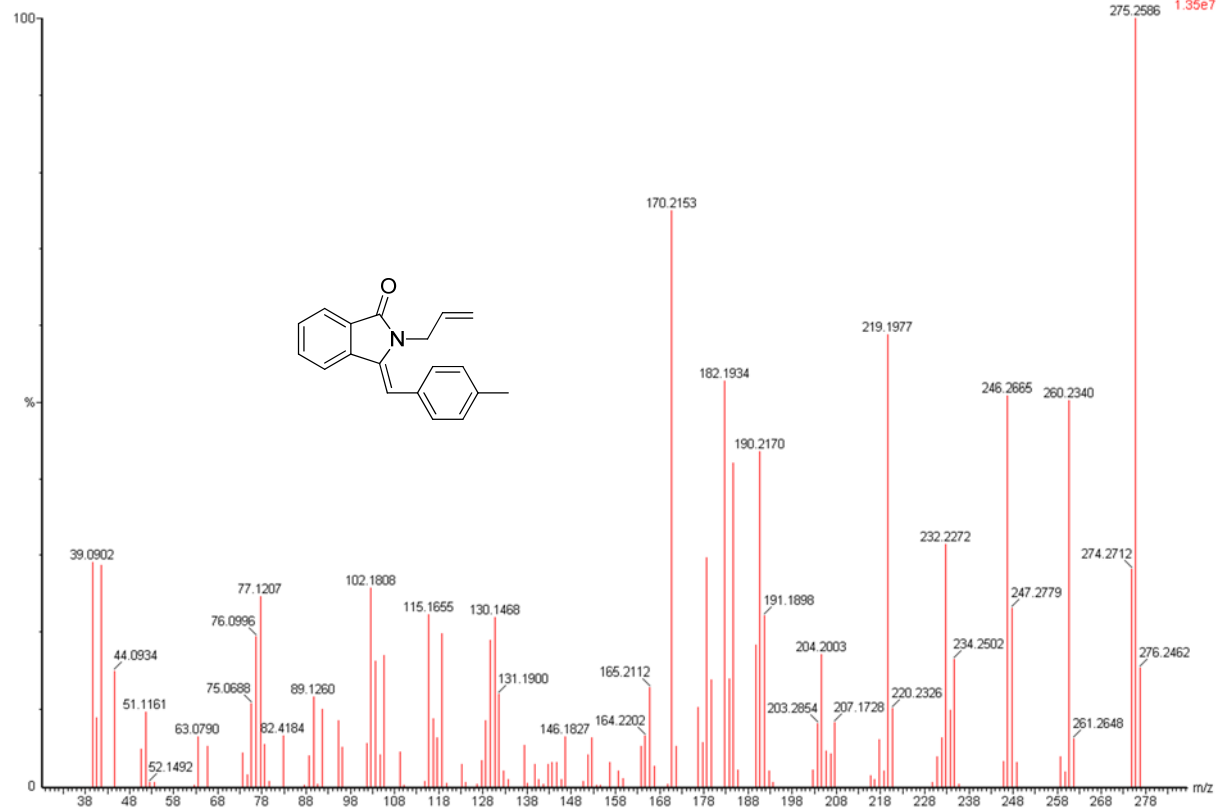


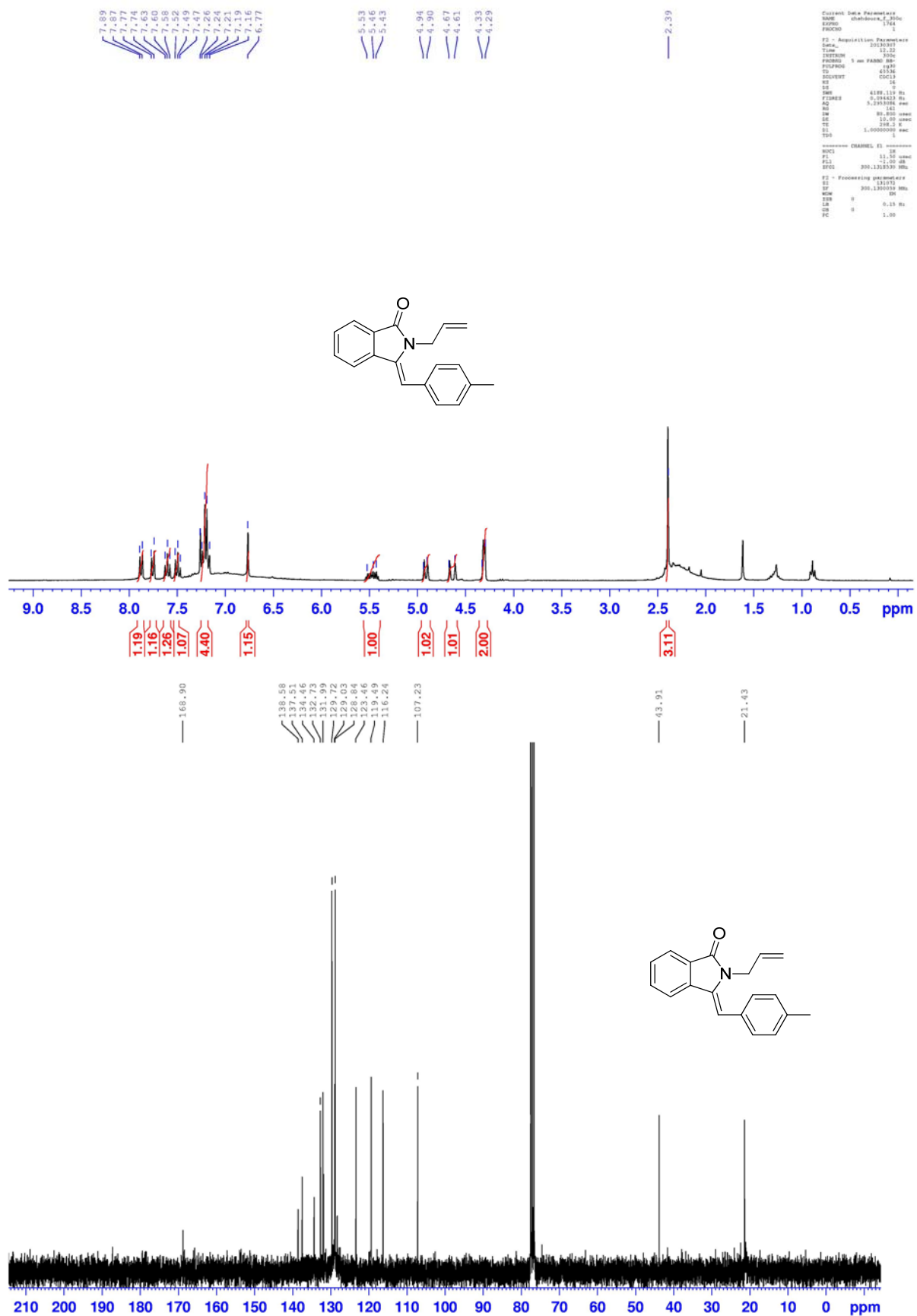
1068

fc-1068-bis-1 2300 (15.503) Cm (2296:2301)

, 07-Mar-2013 + 12:37:54

Scan EI+  
1.35e7





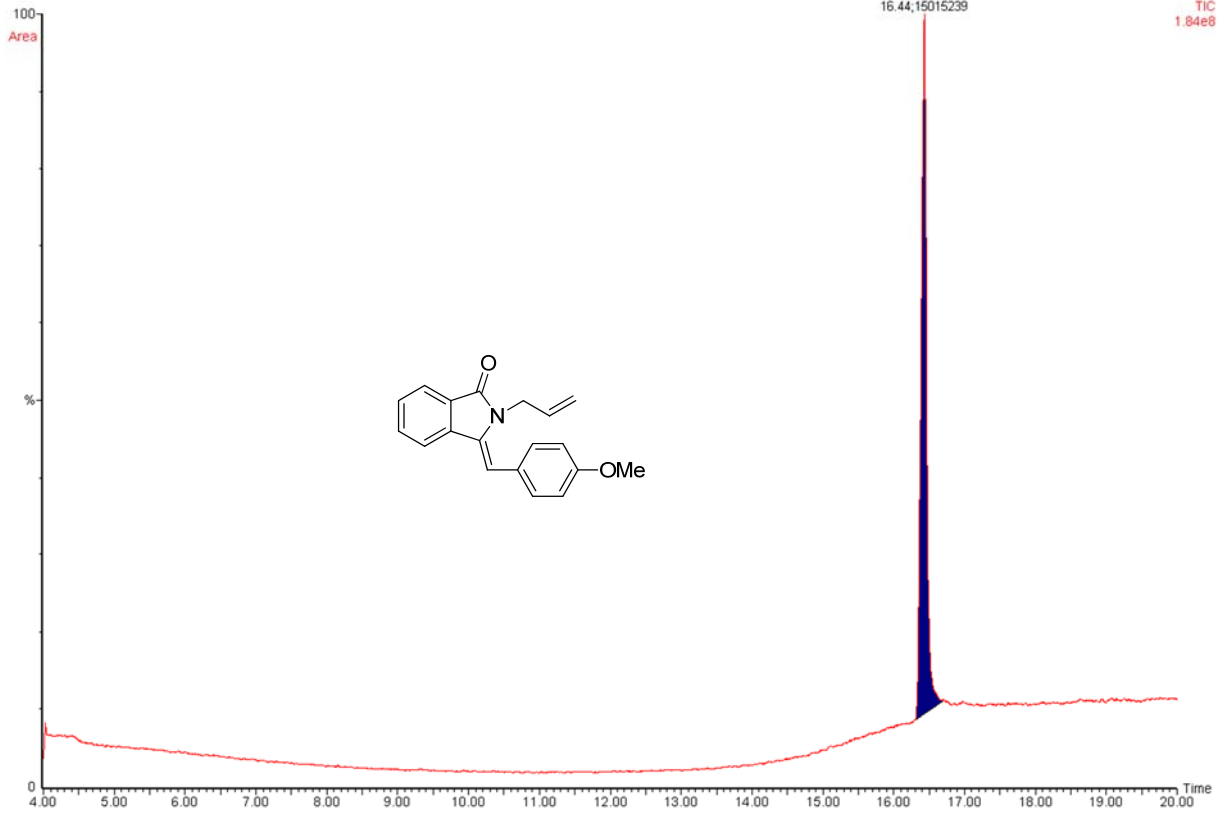
GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **m34**

1069

fc-1069-1 Sm (Mn, 1x3)

, 06-Mar-2013 + 13:59:26

Scan EI+  
TIC  
1.84e8

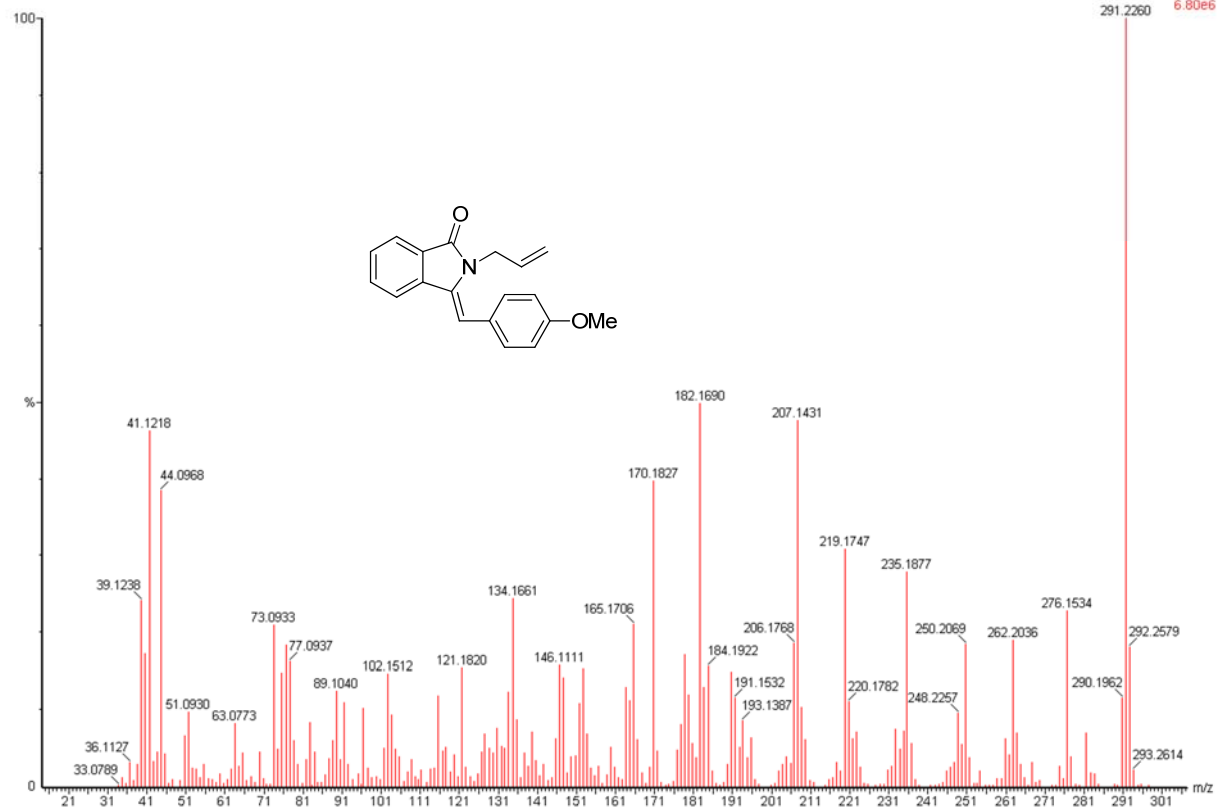


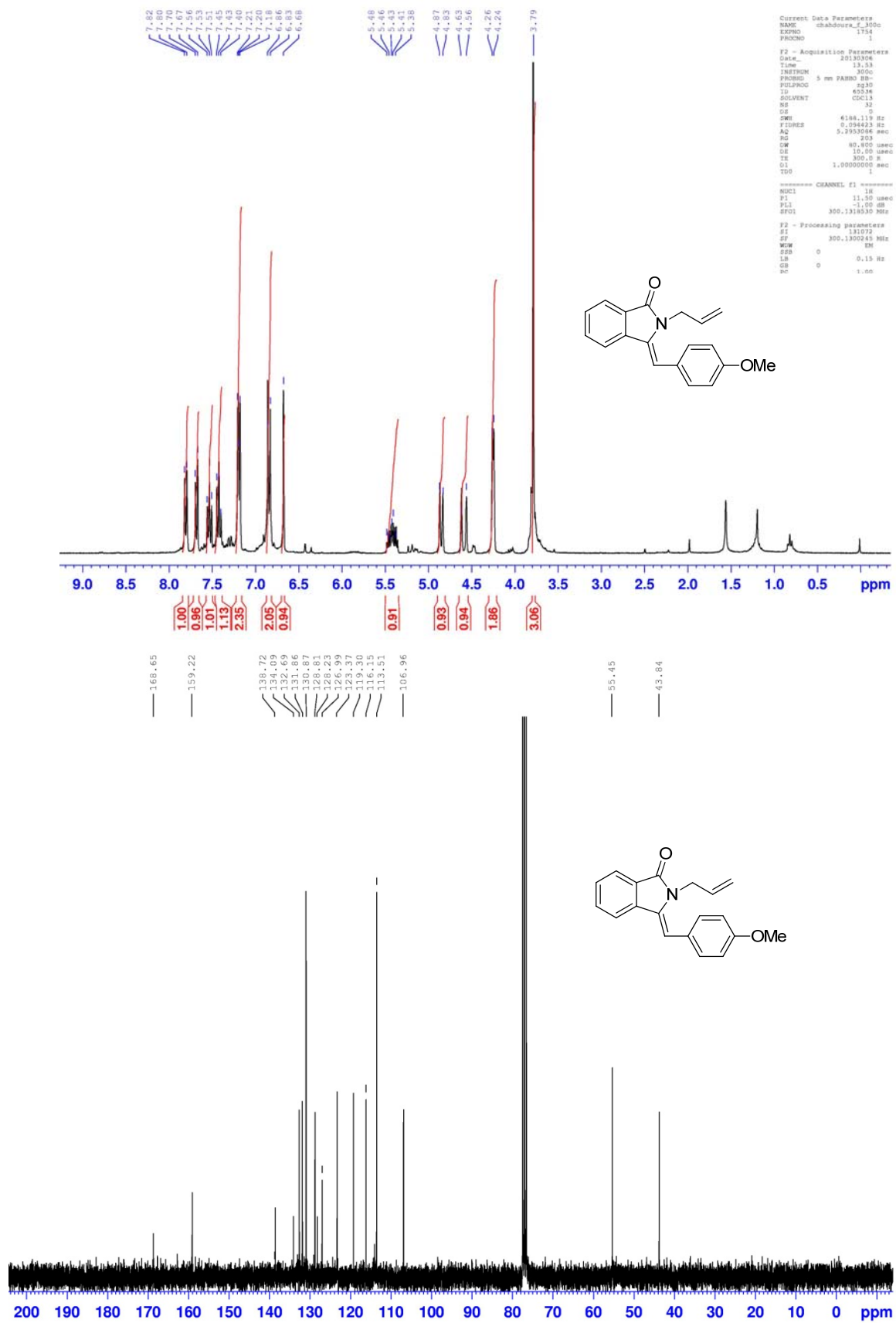
1069

fc-1069-1 2488 (16.443) Cm (2474:2507)

, 06-Mar-2013 + 13:59:26

Scan EI+  
6.80e6





GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **m35**

1077

fc-1077-bis Sm (Mn, 1x3)

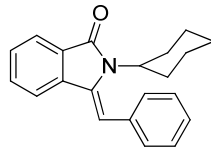
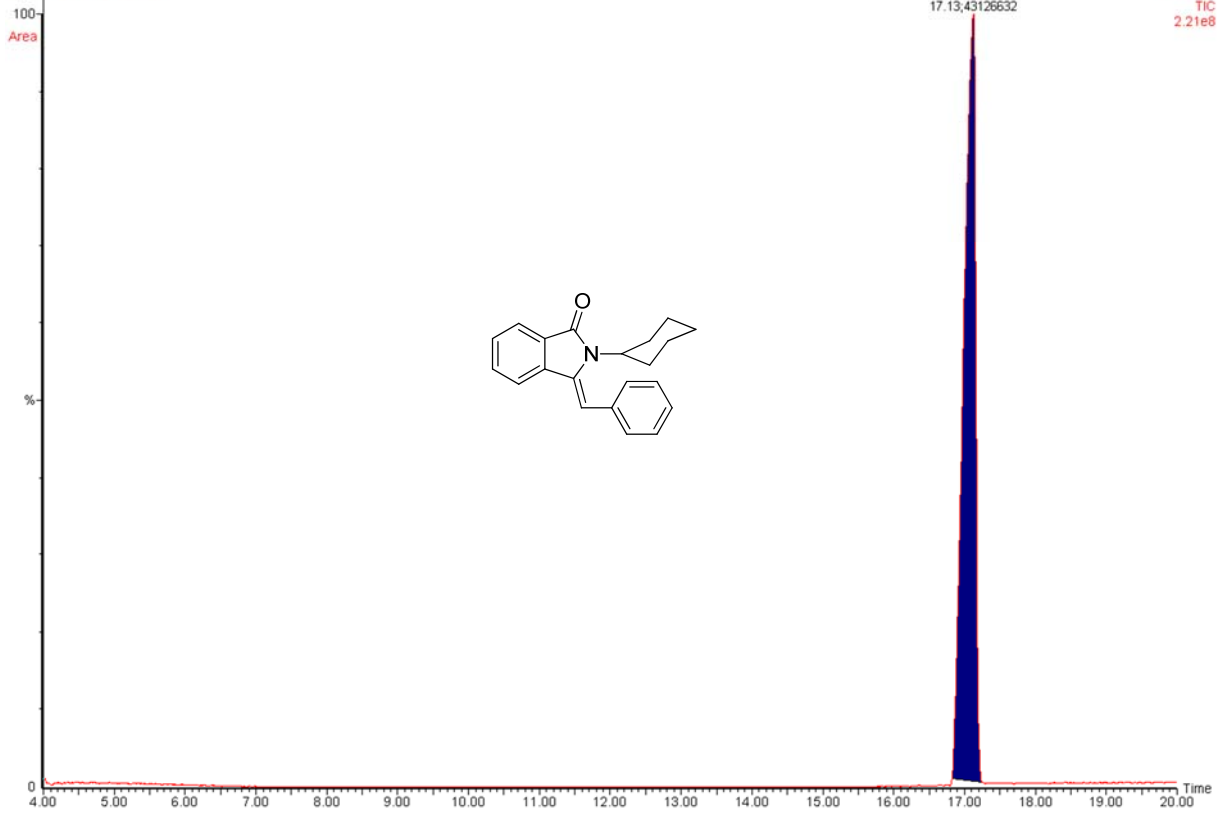
, 19-Mar-2013 + 19:51:10

Scan E1+

TIC

2.21e8

Area



1077

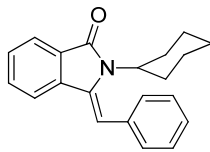
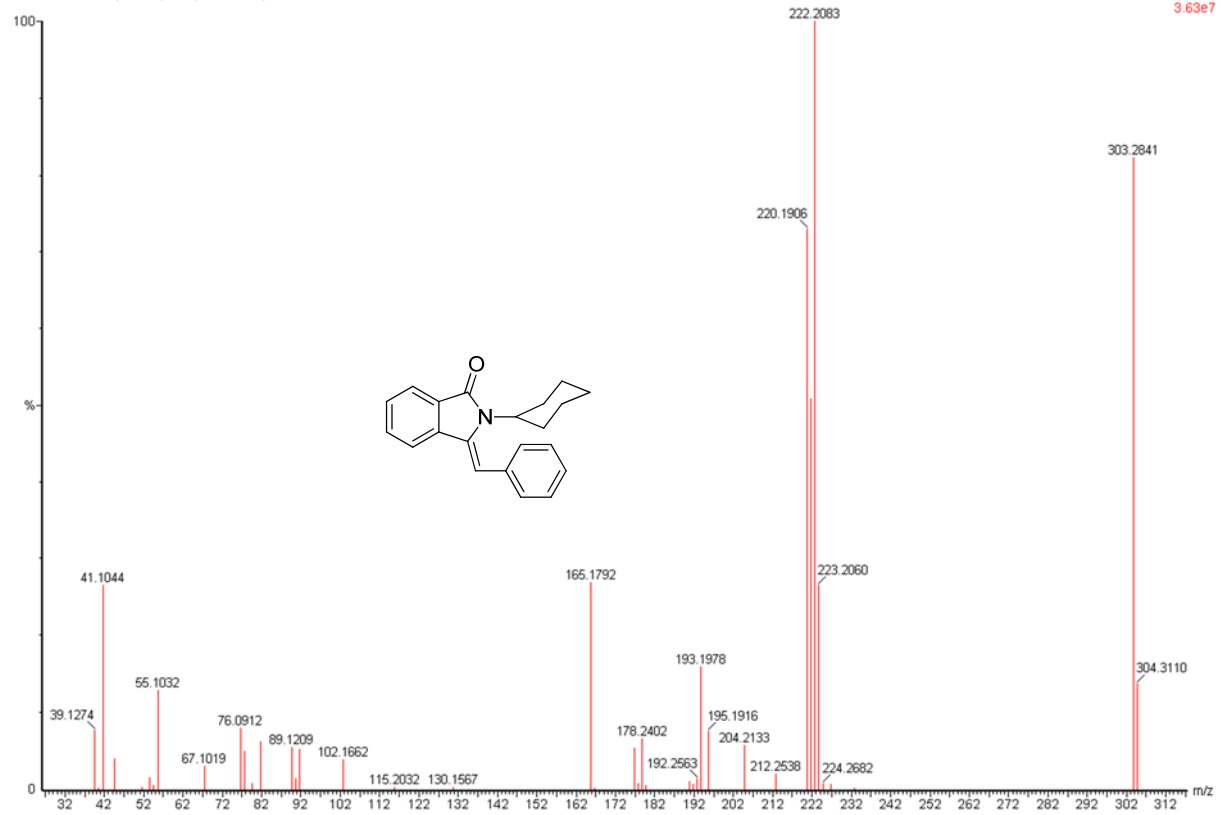
fc-1077-bis 2624 (17.124) Cm (2599:2625)

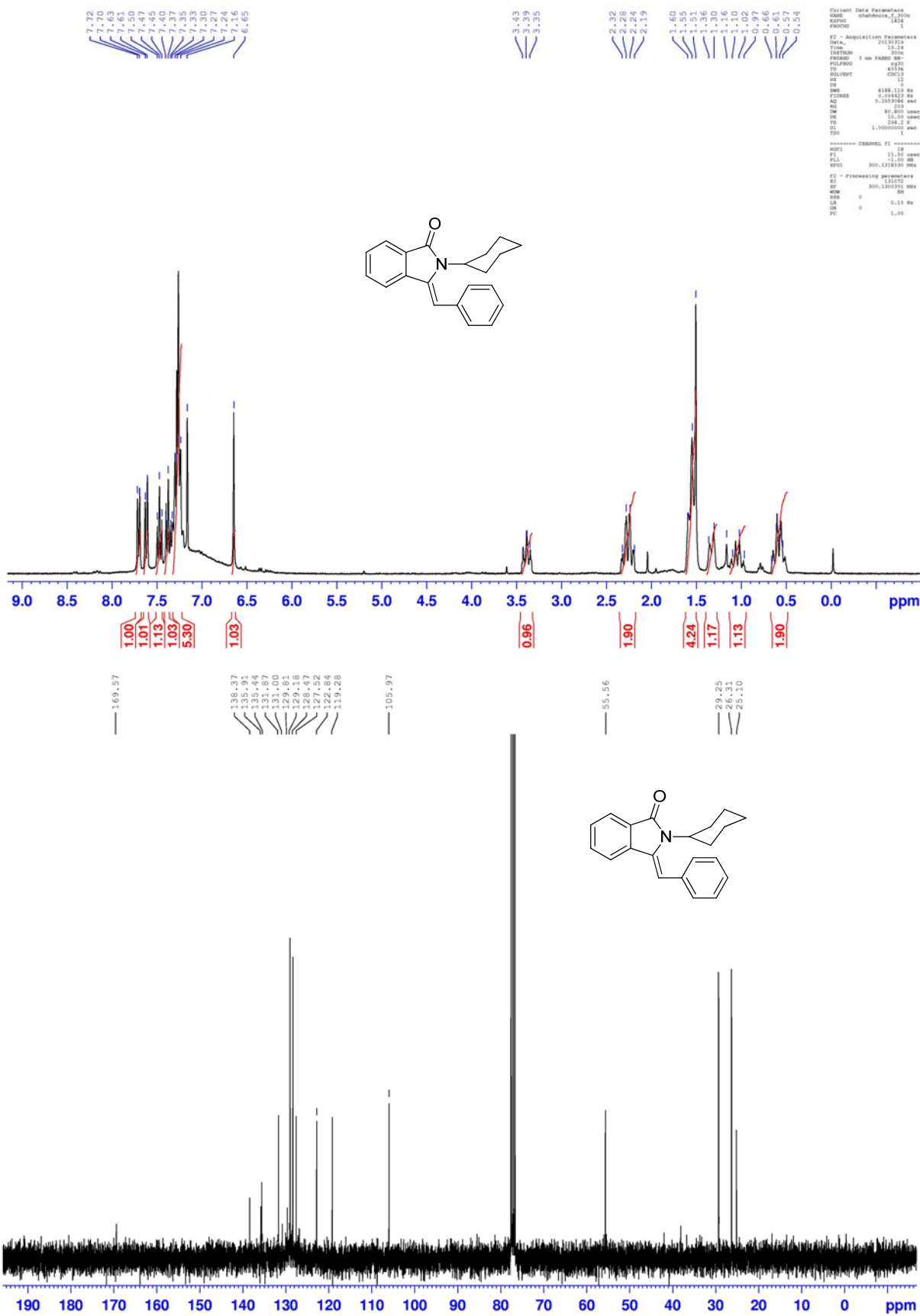
, 19-Mar-2013 + 19:51:10

Scan E1+

3.63e7

%



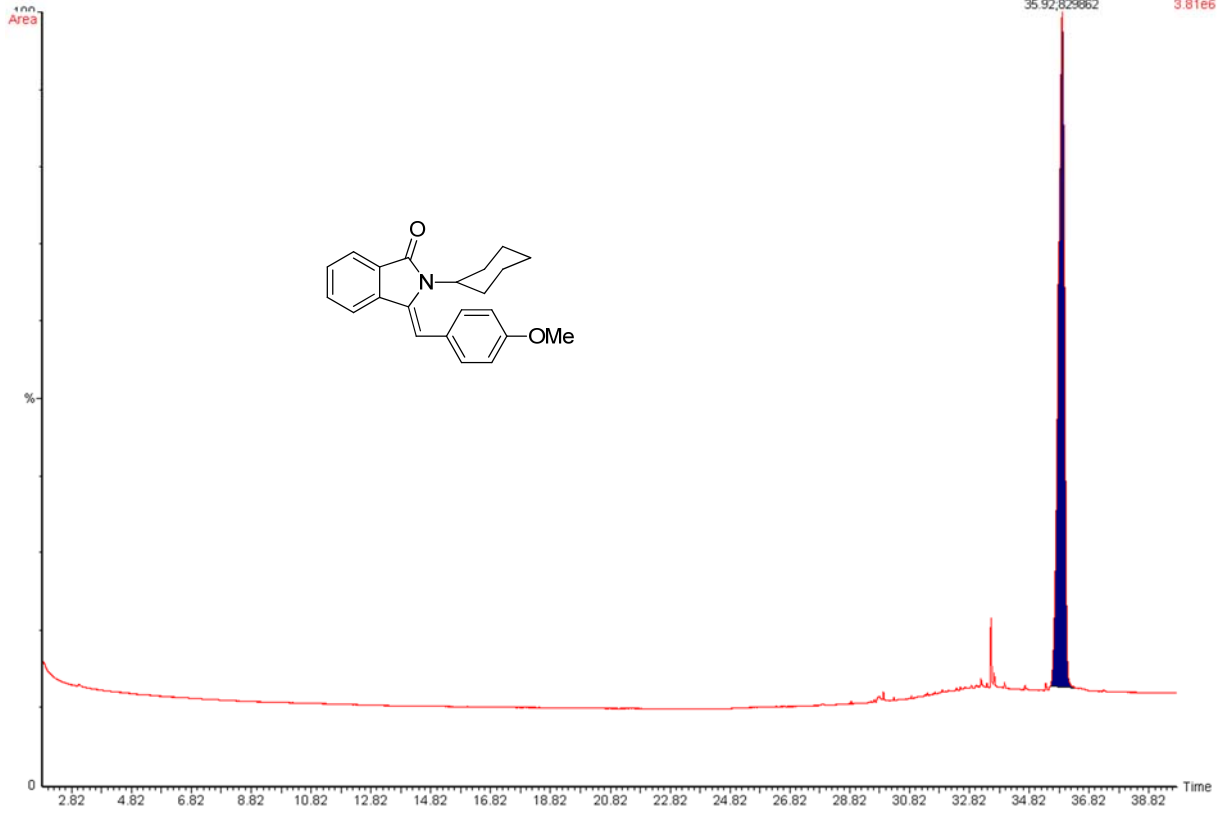


GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **n33**



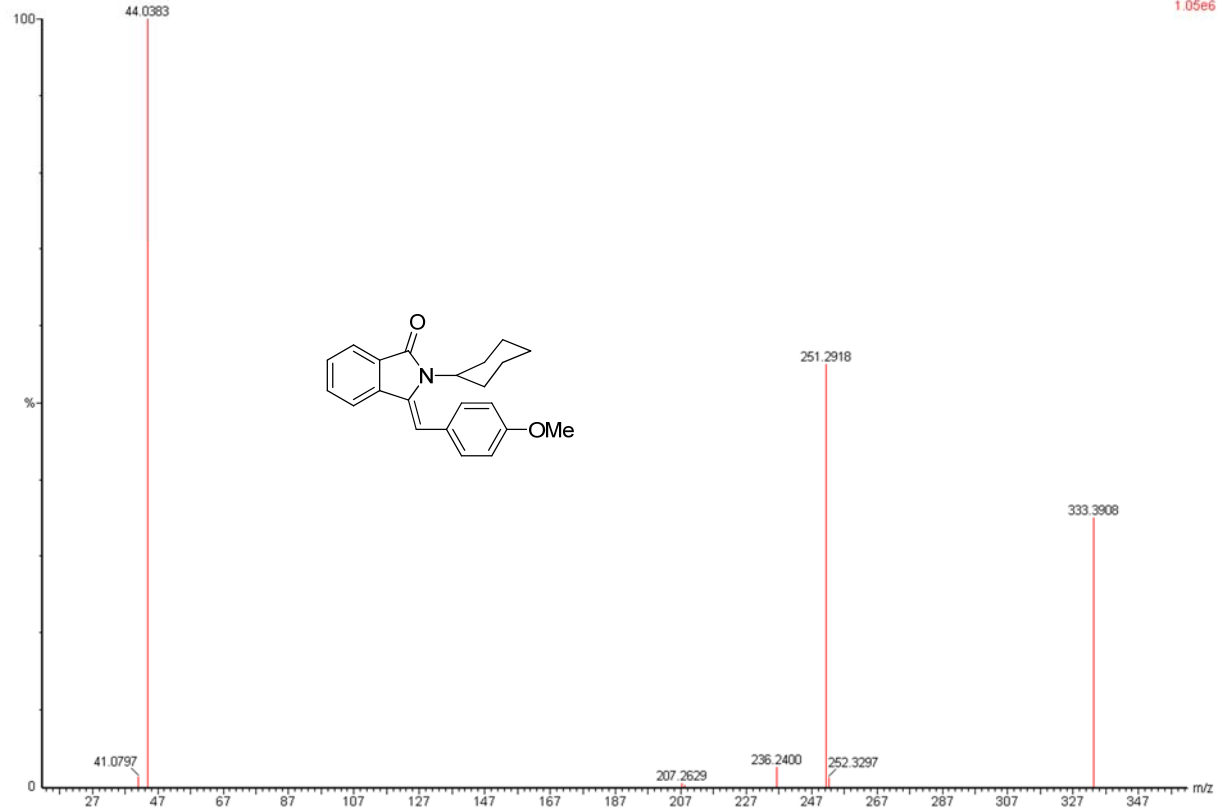
1078  
fc-1078- Sm (Mn, 1x3)

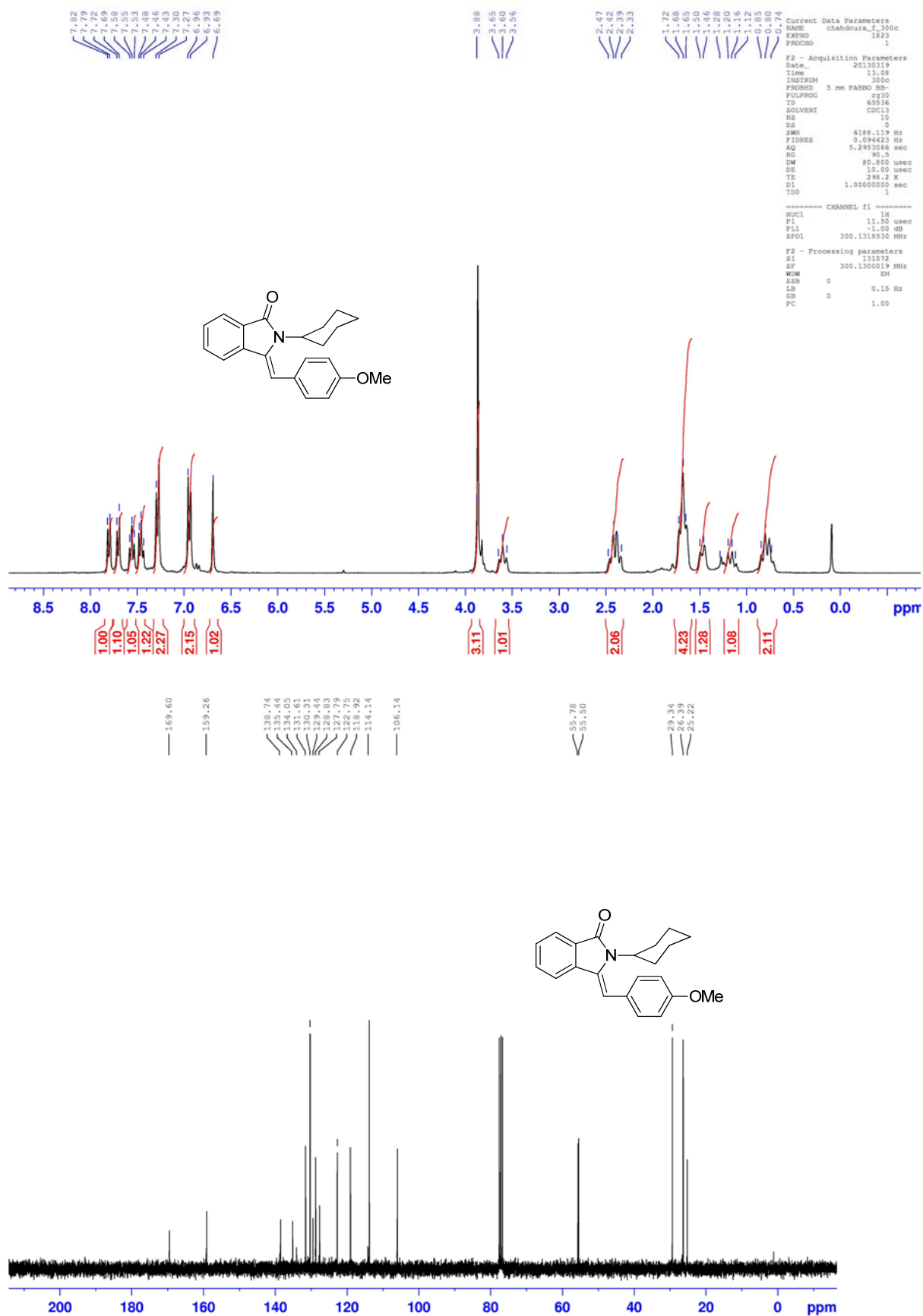
, 26-Mar-2013 + 19:48:16  
A: FID  
3.81e6



1078  
fc-1078- 7095 (38.487) Cm (6798.7095)

, 26-Mar-2013 + 19:48:16  
Scan EI+  
1.05e6





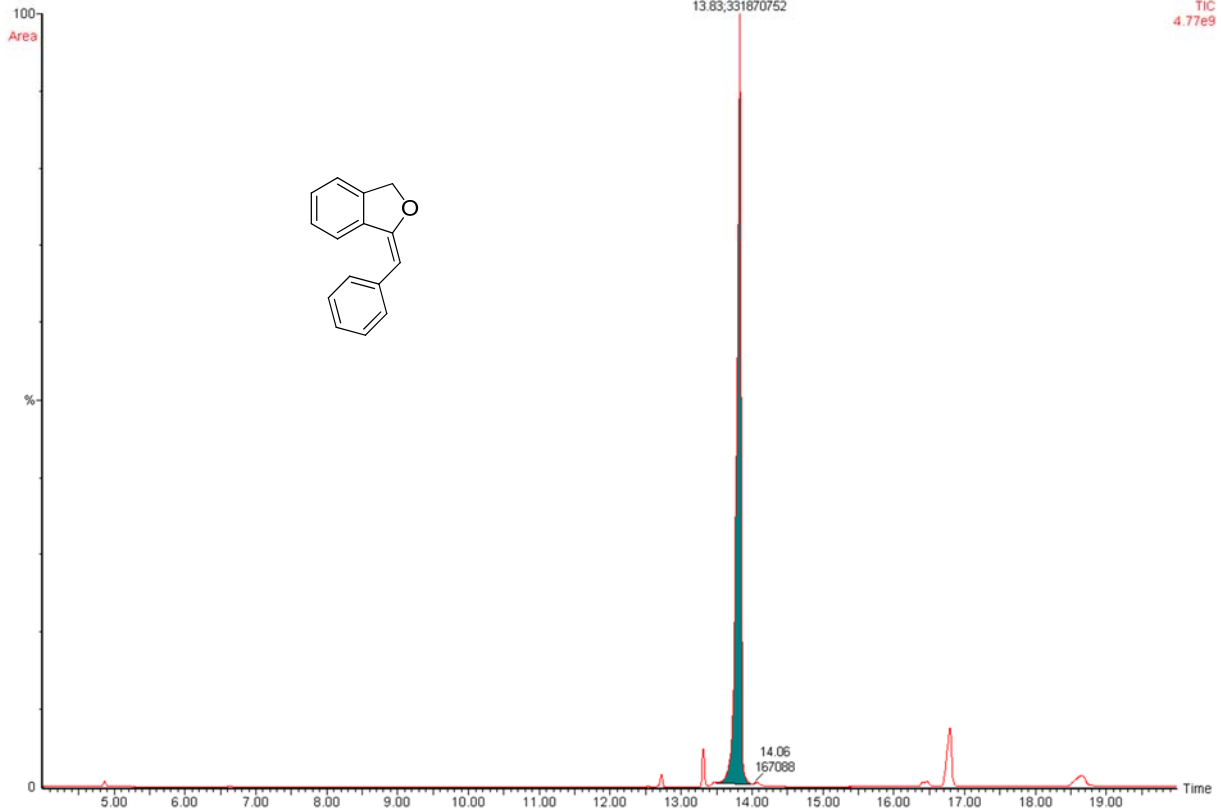
GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **n35**

1063

tc-1063- Sm (Mn, 1x3)

, 04-Mar-2013 + 15:10:38

Scan EI+  
TIC  
4.77e9

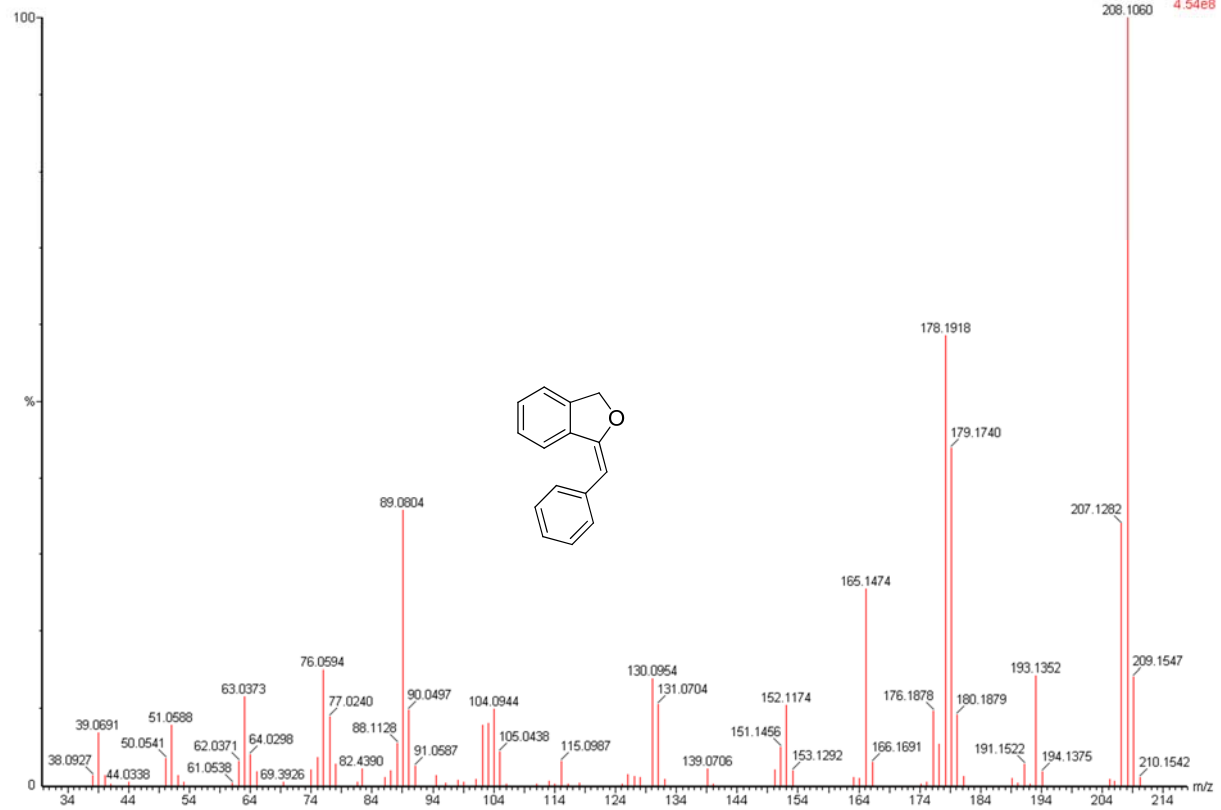


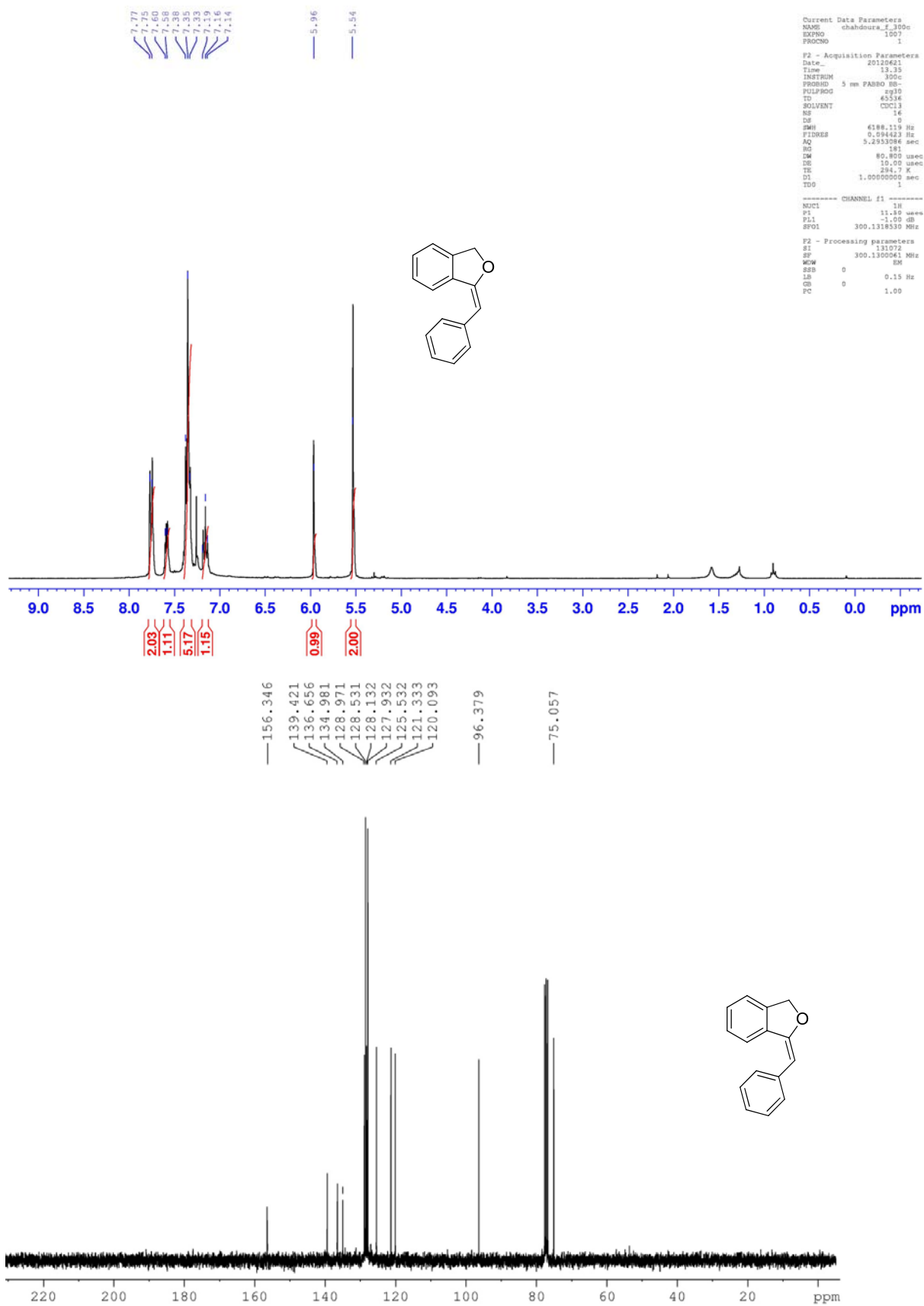
1063

tc-1063- 1968 (13.842) Cm (1945:1968)

, 04-Mar-2013 + 15:10:38

Scan EI+  
4.54e8



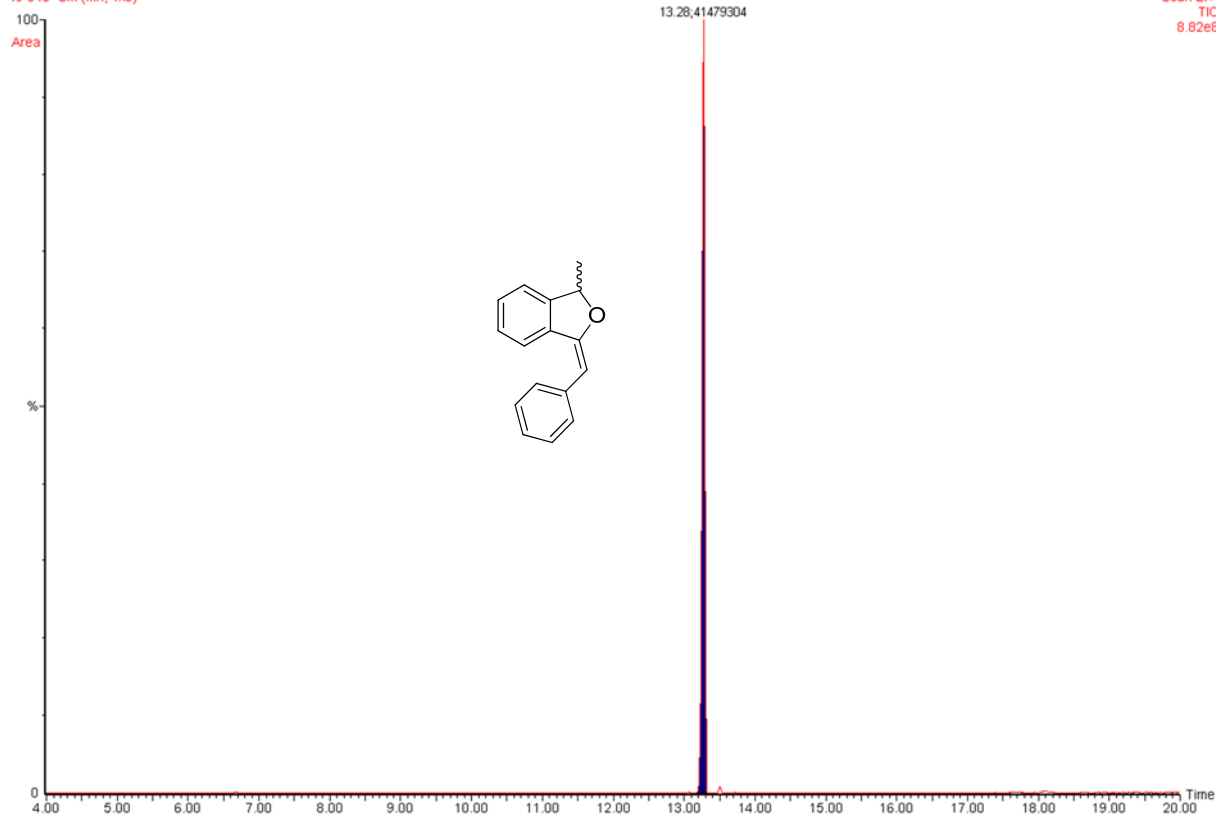


GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C} \{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **o33**

840  
fc-840- Sm (Mn, 1x3)

, 27-Jul-2012 + 09:07:23

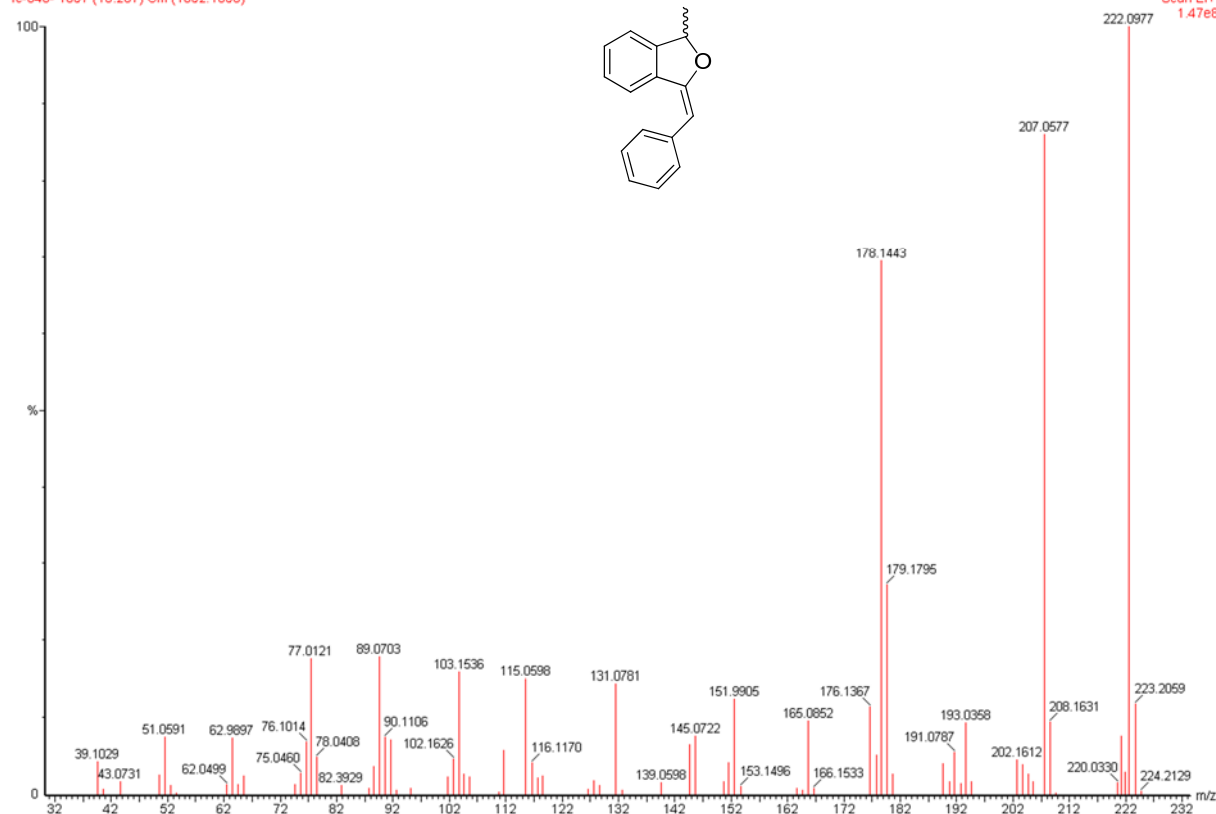
Scan EI+  
TIC  
8.82e8

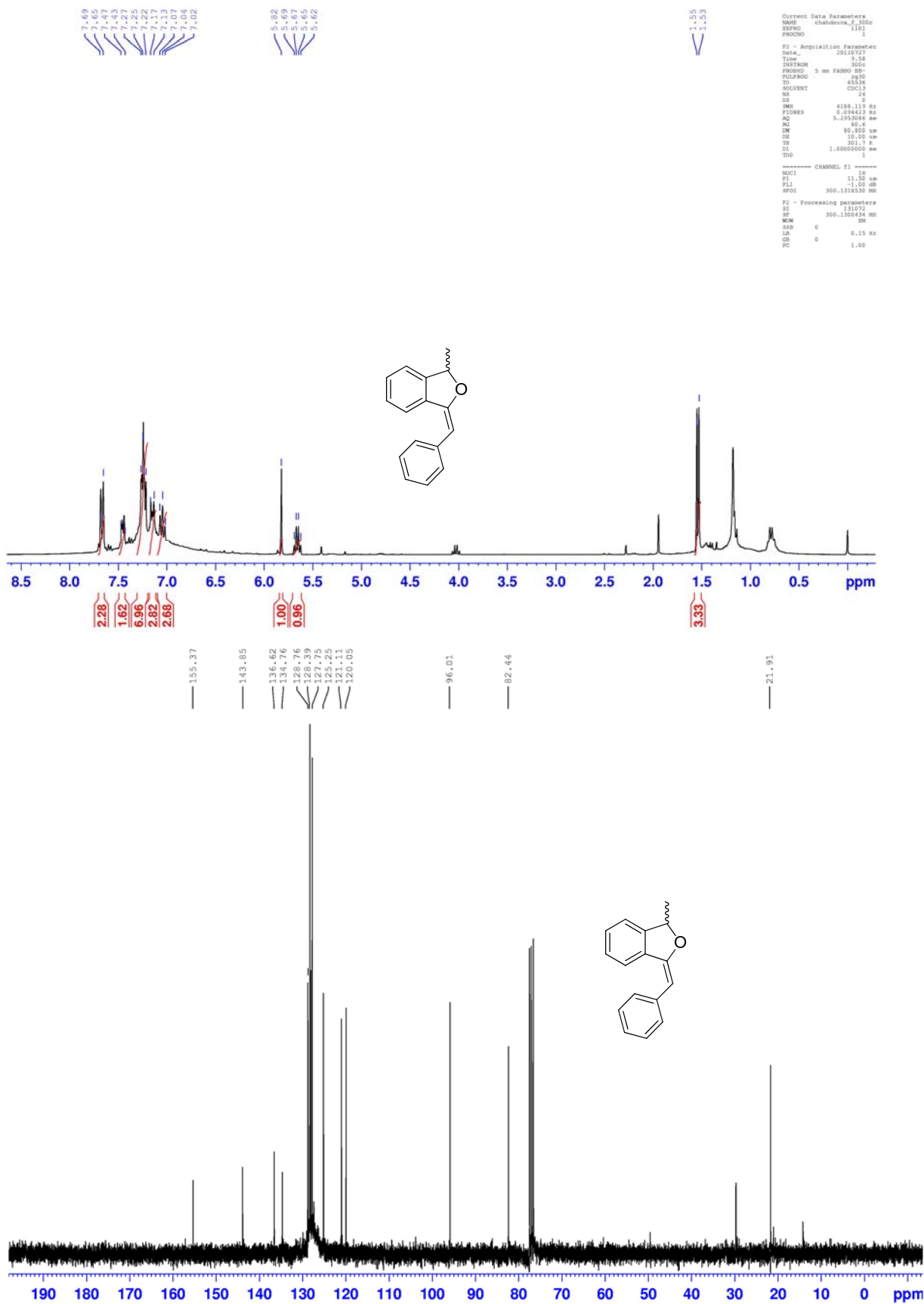


840  
fc-840- 1857 (13.287) Cm (1852:1858)

, 27-Jul-2012 + 09:07:23

Scan EI+  
1.47e8

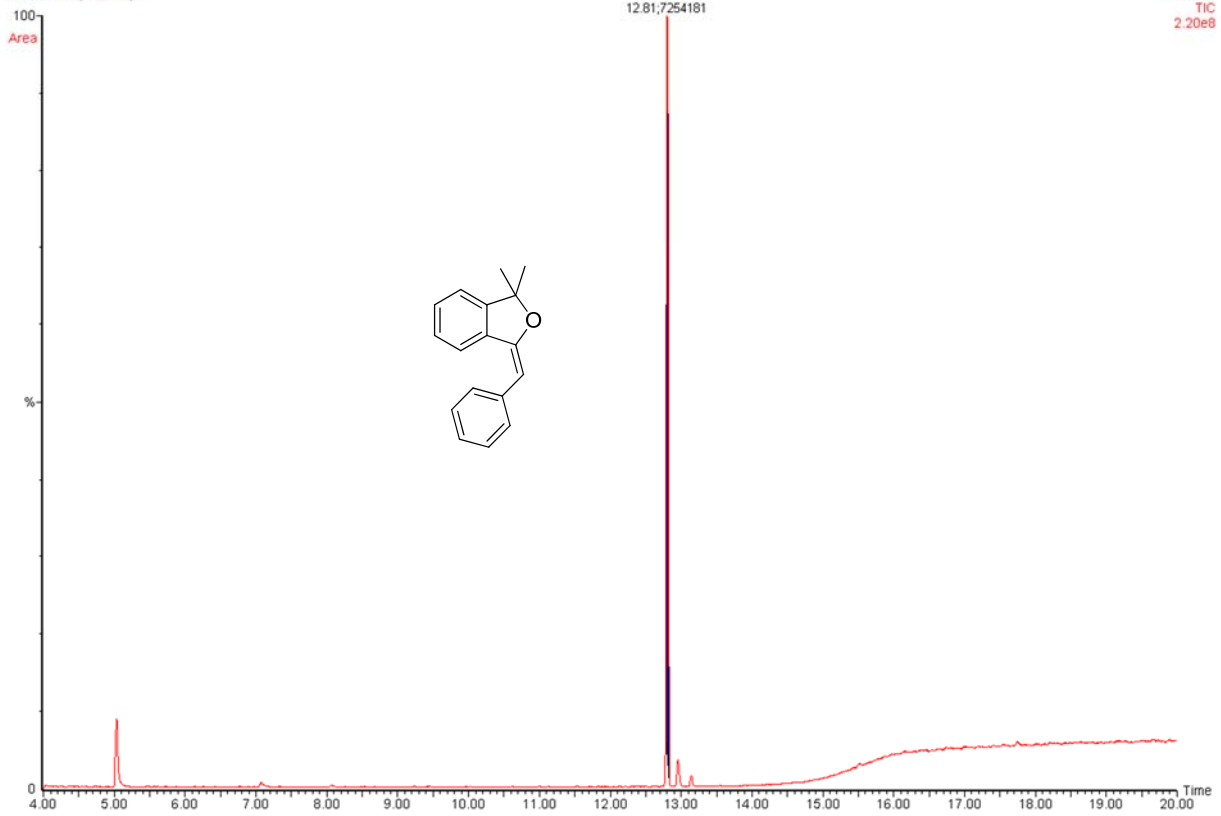




GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **p33**

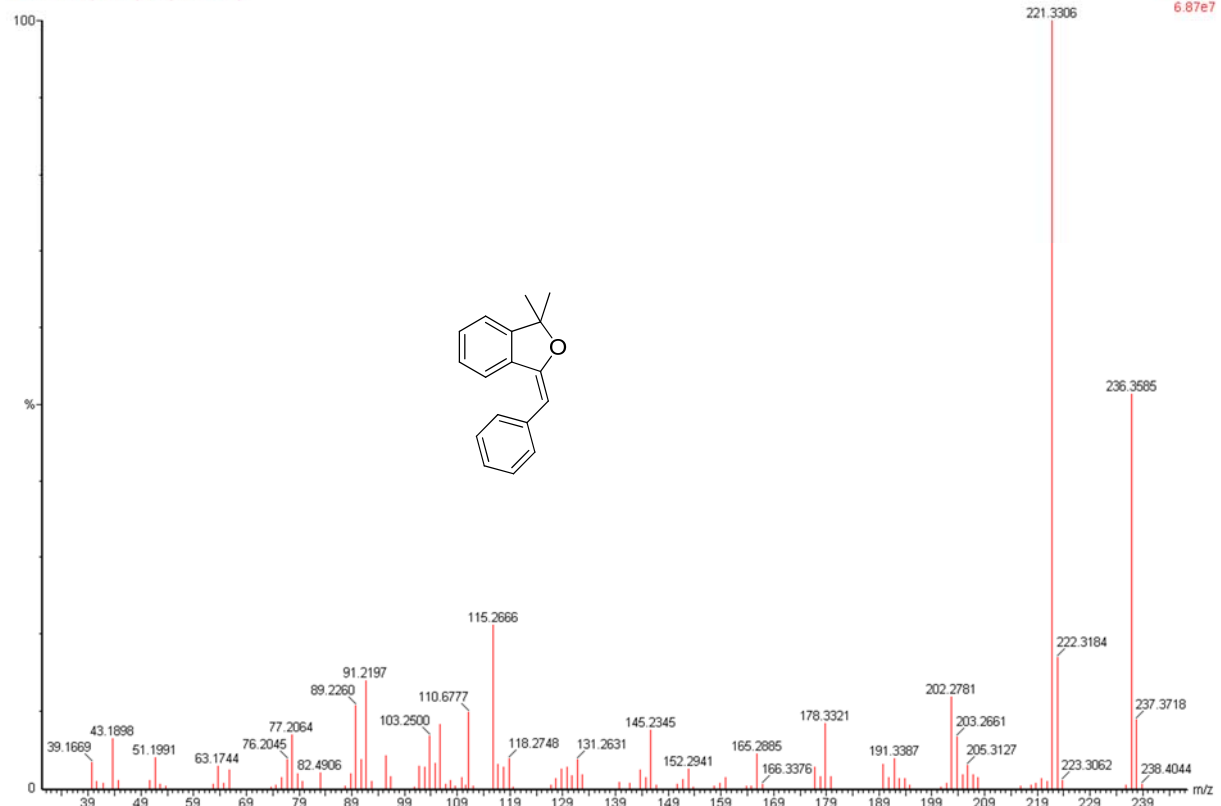
799  
tc-799- Sm (Mn, 1x3)

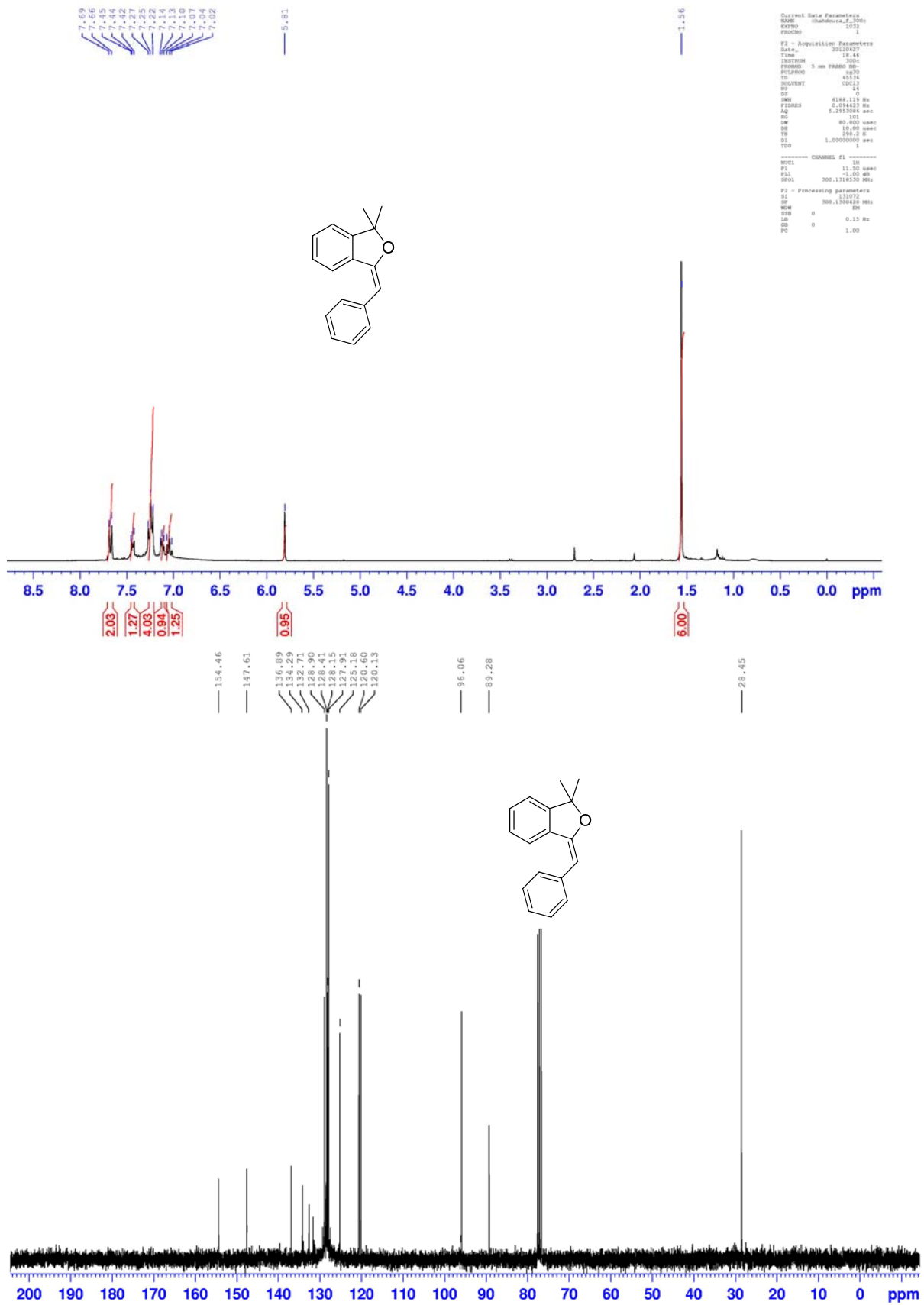
, 26-Jun-2012 + 19:47:23  
Scan EI+  
TIC  
2.20e8



799  
tc-799- 1762 (12.812) Cm (1760:1764)

, 26-Jun-2012 + 19:47:23  
Scan EI+  
6.87e7



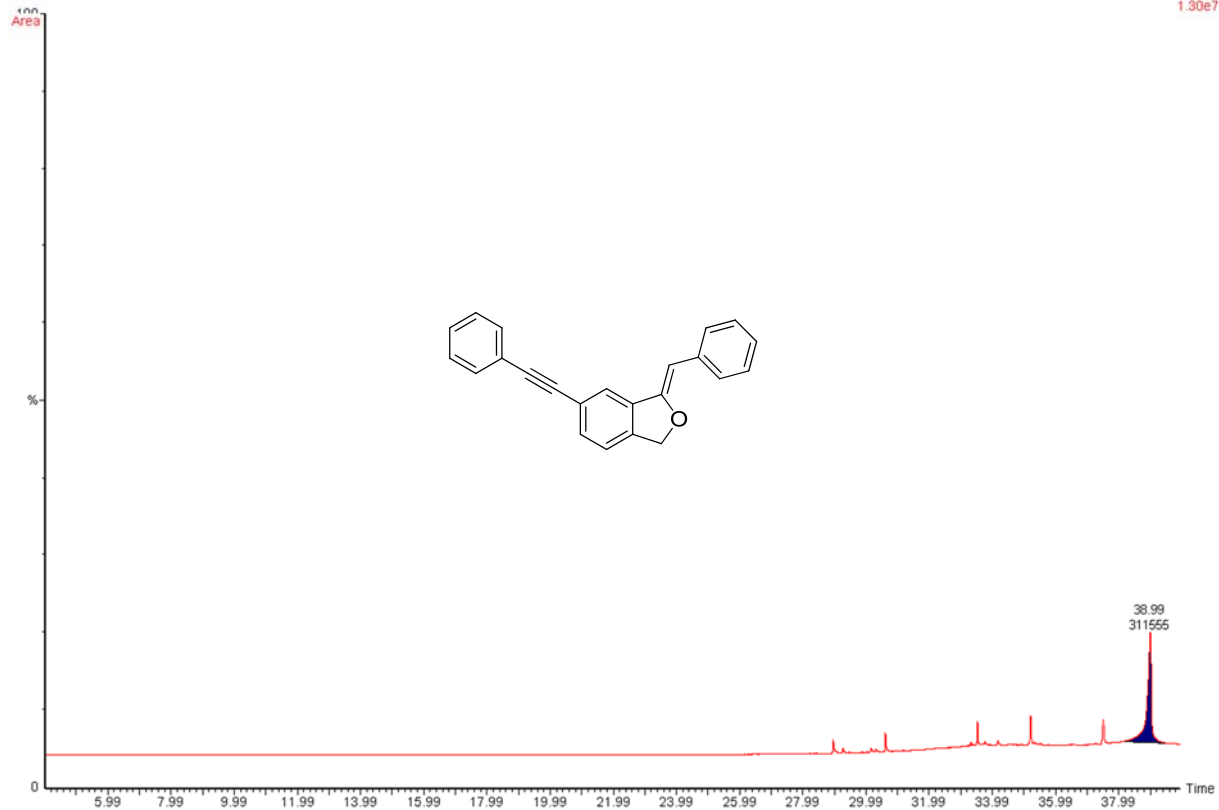


GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **q33**



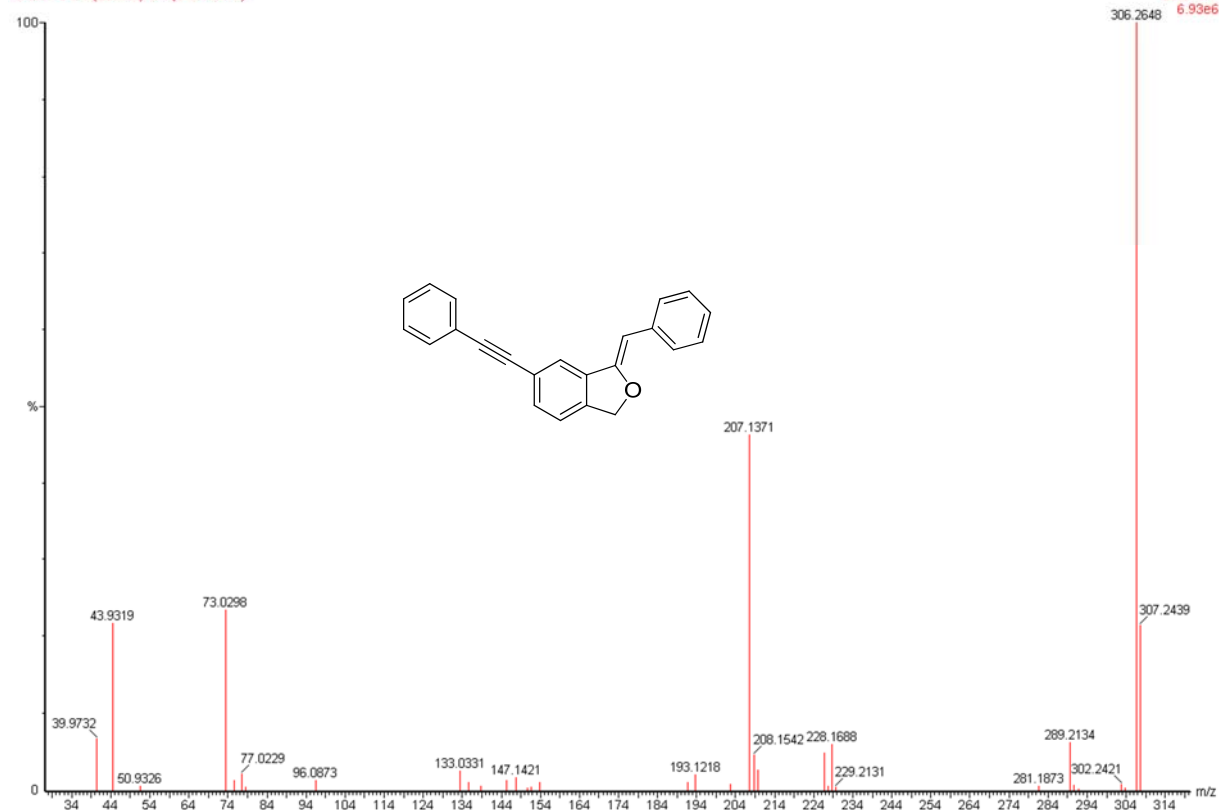
997  
fc-997- Sm (Mn, 1x3)

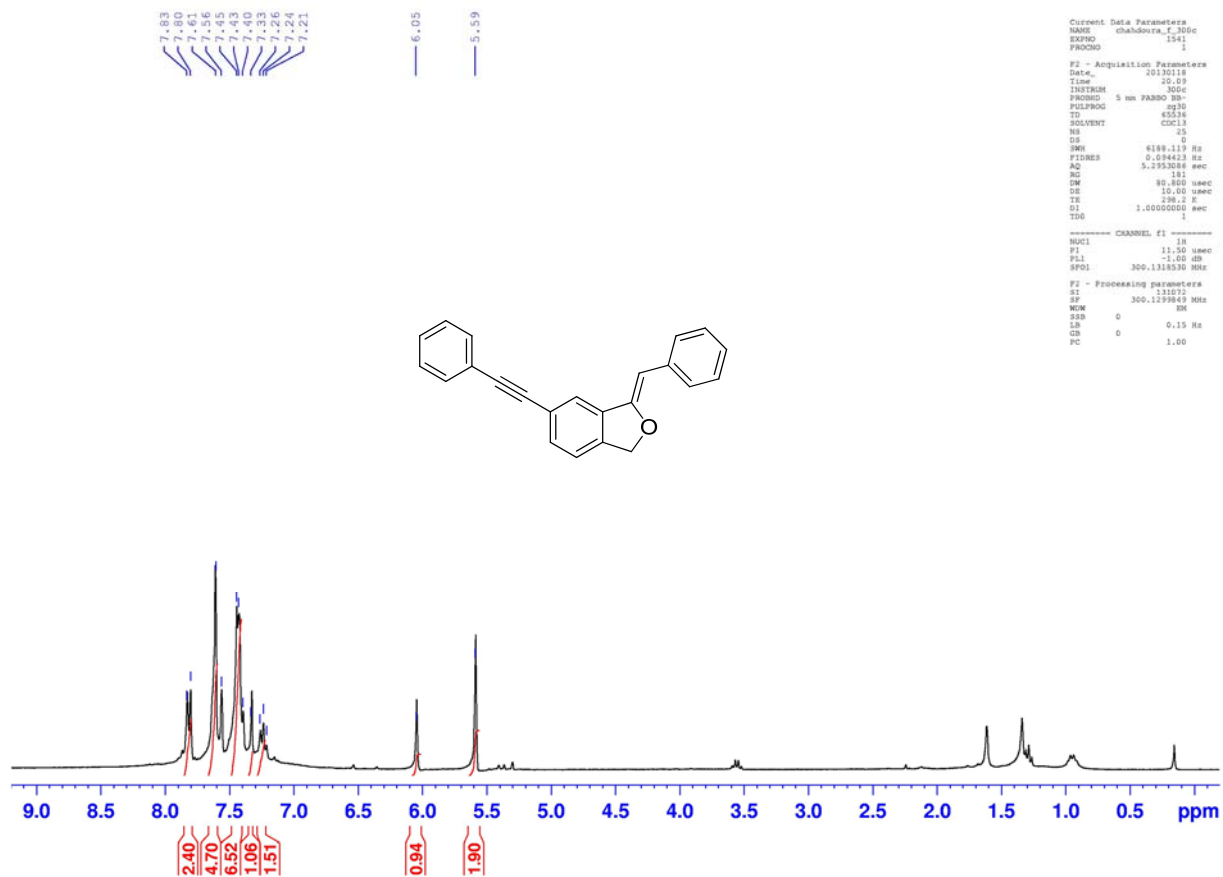
, 18-Jan-2013 + 15:36:44  
A: FID  
1.30e7



997  
fc-997- 6471 (35.366) Cm (6467.6476)

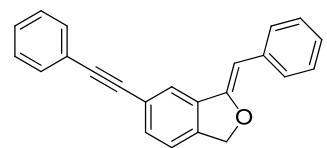
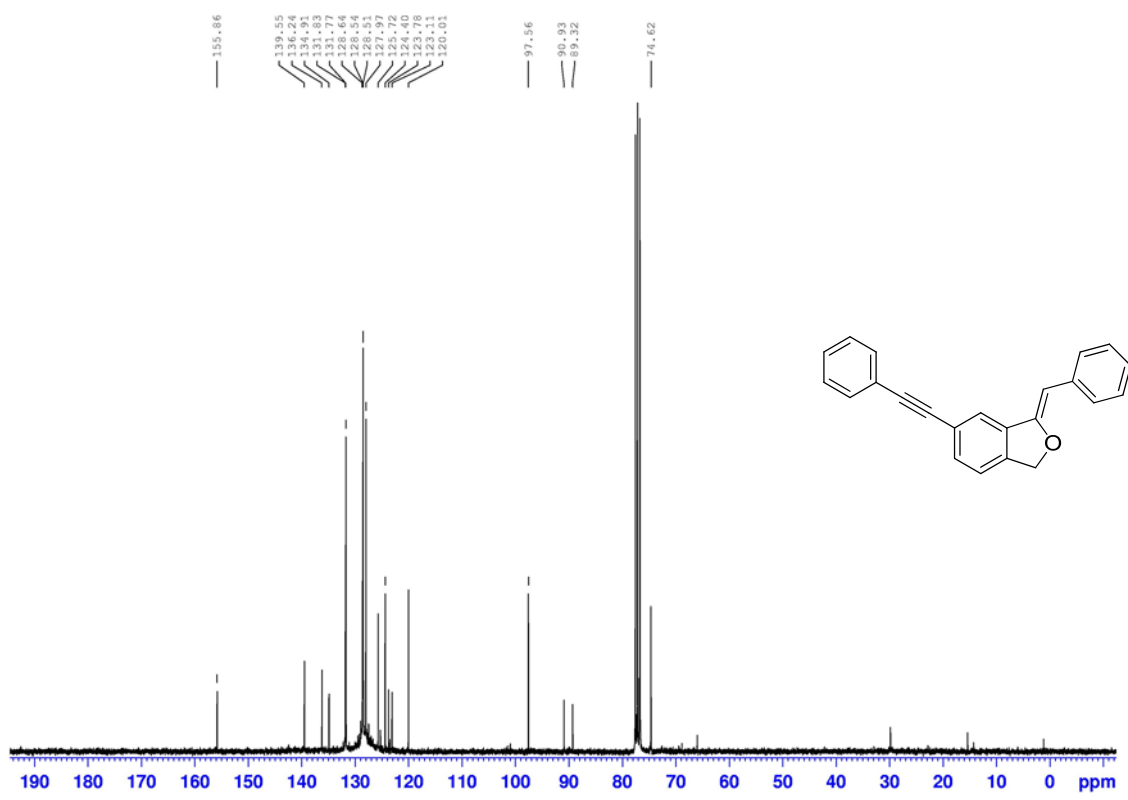
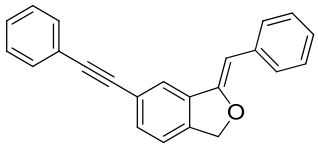
, 18-Jan-2013 + 15:36:44  
Scan EI+  
6.93e6





```

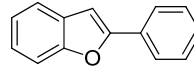
Current Data Parameters
NAME: Chahdova_r_330c
EXPNO: 1541
PROCNO: 3
F2 - Acquisition Parameters
Date_: 20130118
Time: 22:03
INSTRUM: zgpg
PROBHD: 5 mm PABBO MM-
PULPROG: zg30
TD: 65536
SOLVENT: CDCl3
NS: 15
DS: 4
SFR: 4188.115 Hz
FIDRES: 0.094423 Hz
AQ: 5.2953068 sec
RG: 181
DM: 80.800 usec
DE: 15.00 usec
TE: 300.2 K
D1: 1.0000000 sec
TDO: 3
----- CHANNEL f1 -----
NUC1: 1H
P1: 11.50 usec
PL1: -1.00 dB
SFO1: 300.1318330 MHz
F2 - Processing parameters
SI: 11672
SF: 300.1298649 MHz
WDW: RM
SSB: 0
CB: 0
PC: 1.00
  
```



GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **r33**

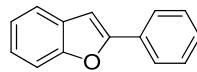
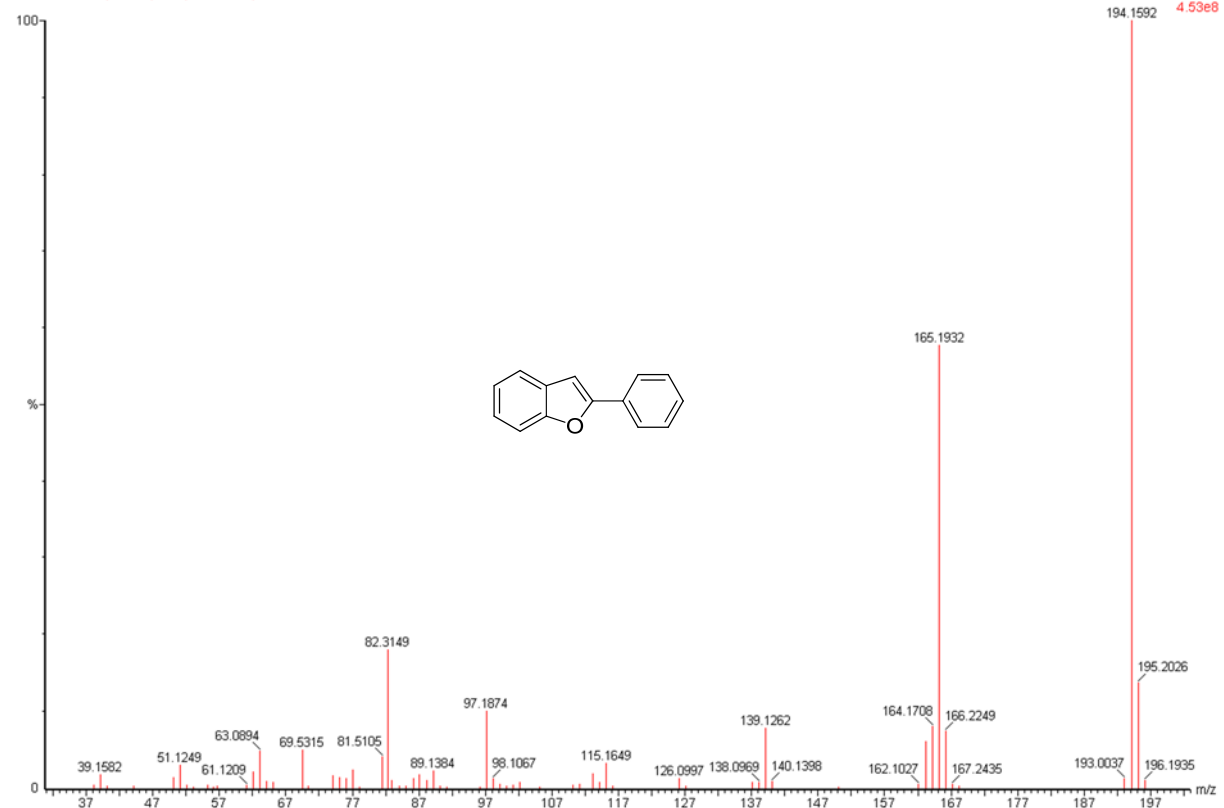
FC-246-F1  
fc-246-f1 Sm (Mn, 1x3)

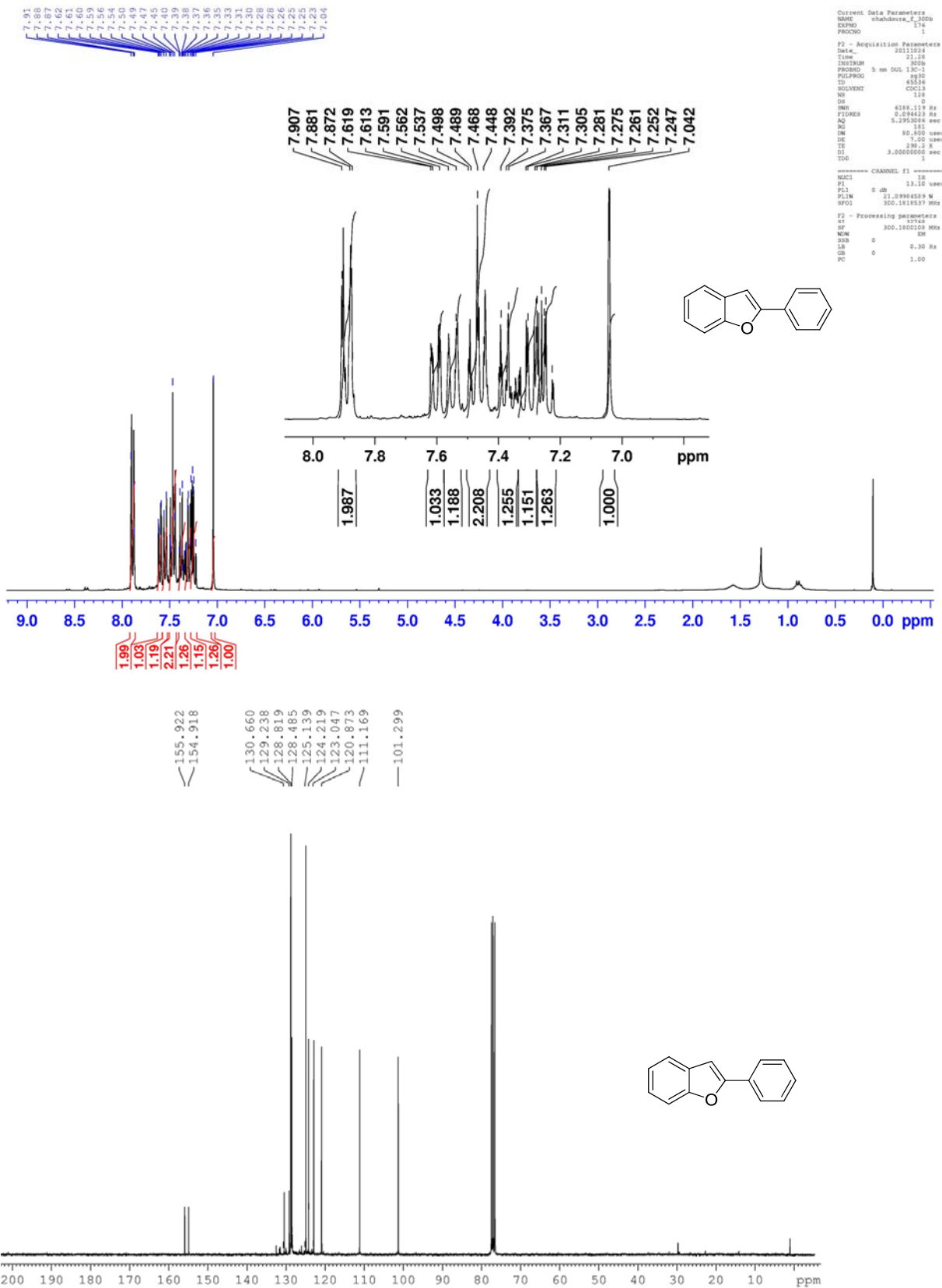
, 15-Jun-2011 + 14:15:30  
Scan E1+  
TIC  
1.48e9



FC-246-F1  
fc-246-f1 1616 (12.082) Cm (1612:1619)

, 15-Jun-2011 + 14:15:30  
Scan E1+  
4.53e8



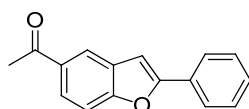
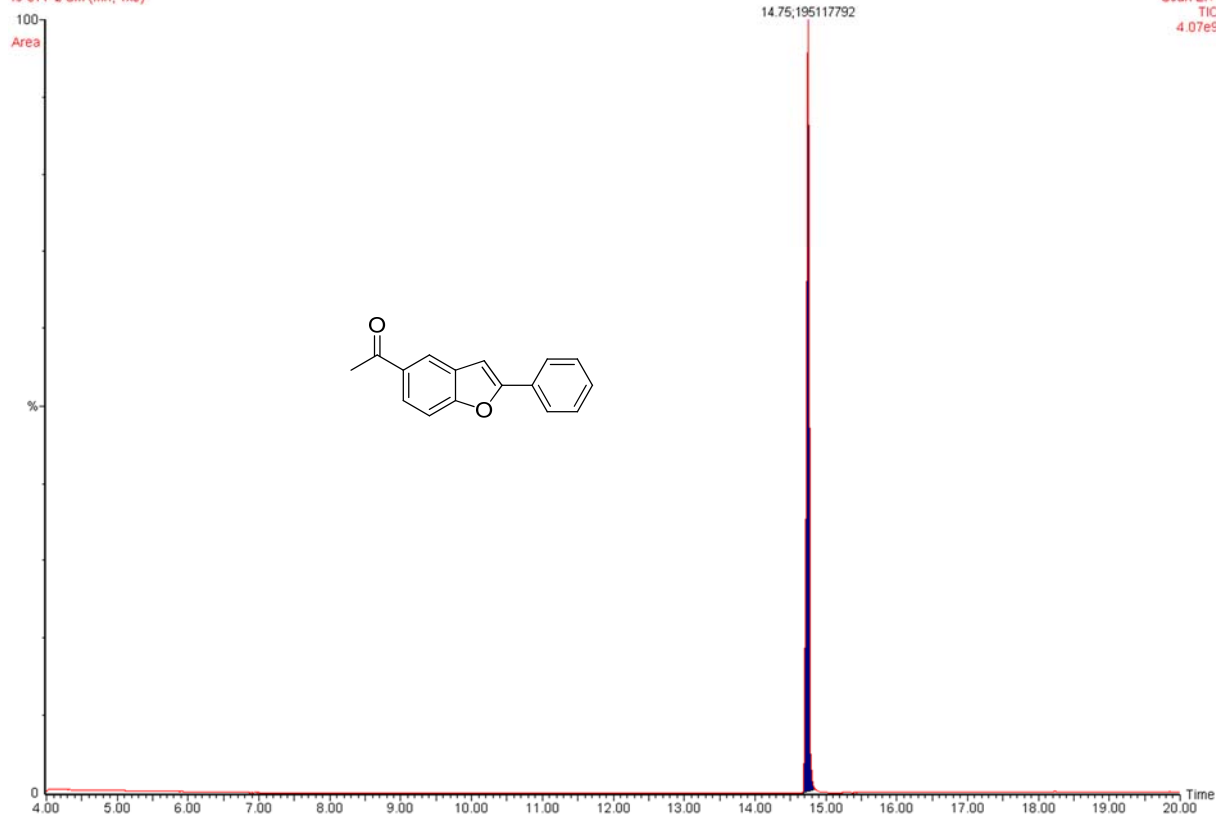


GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **s33**

877  
fc-877-2 Sm (Mn, 1x3)

, 20-Nov-2012 + 21:08:48

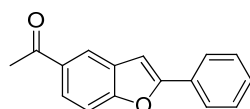
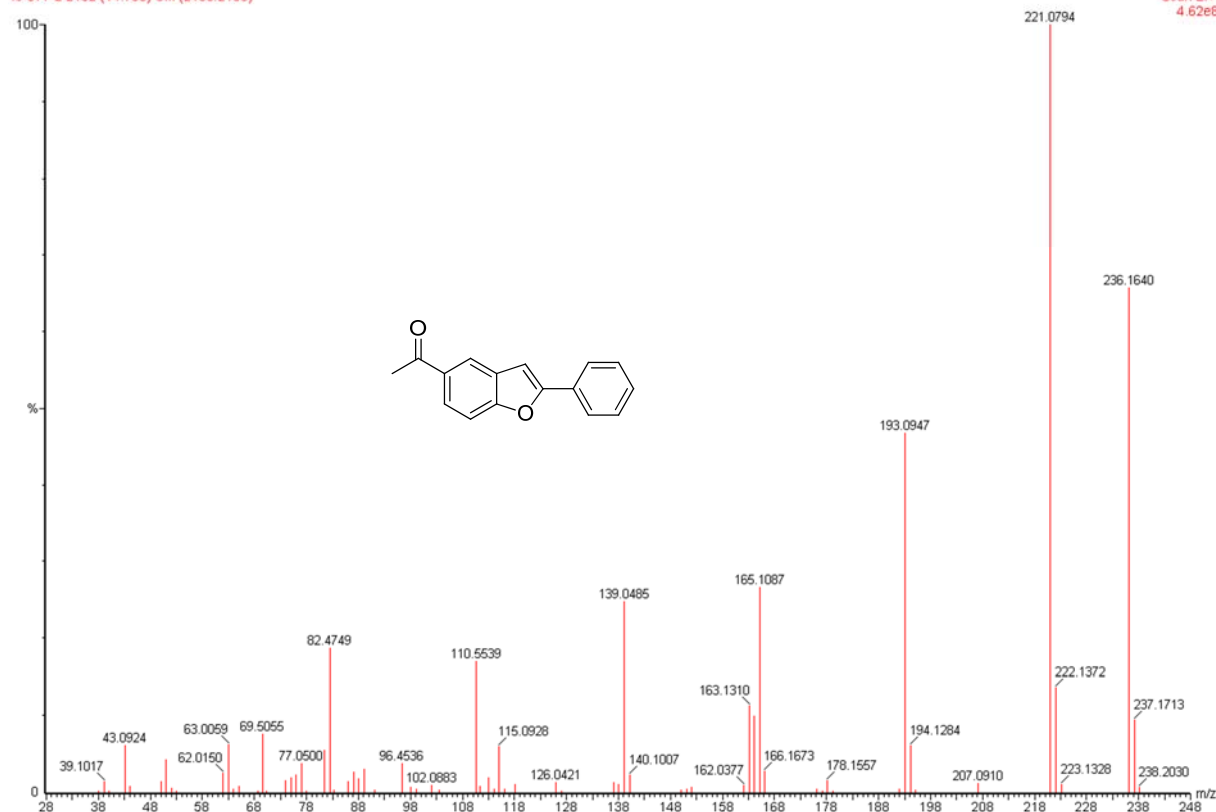
Scan EI+  
TIC  
4.07e9

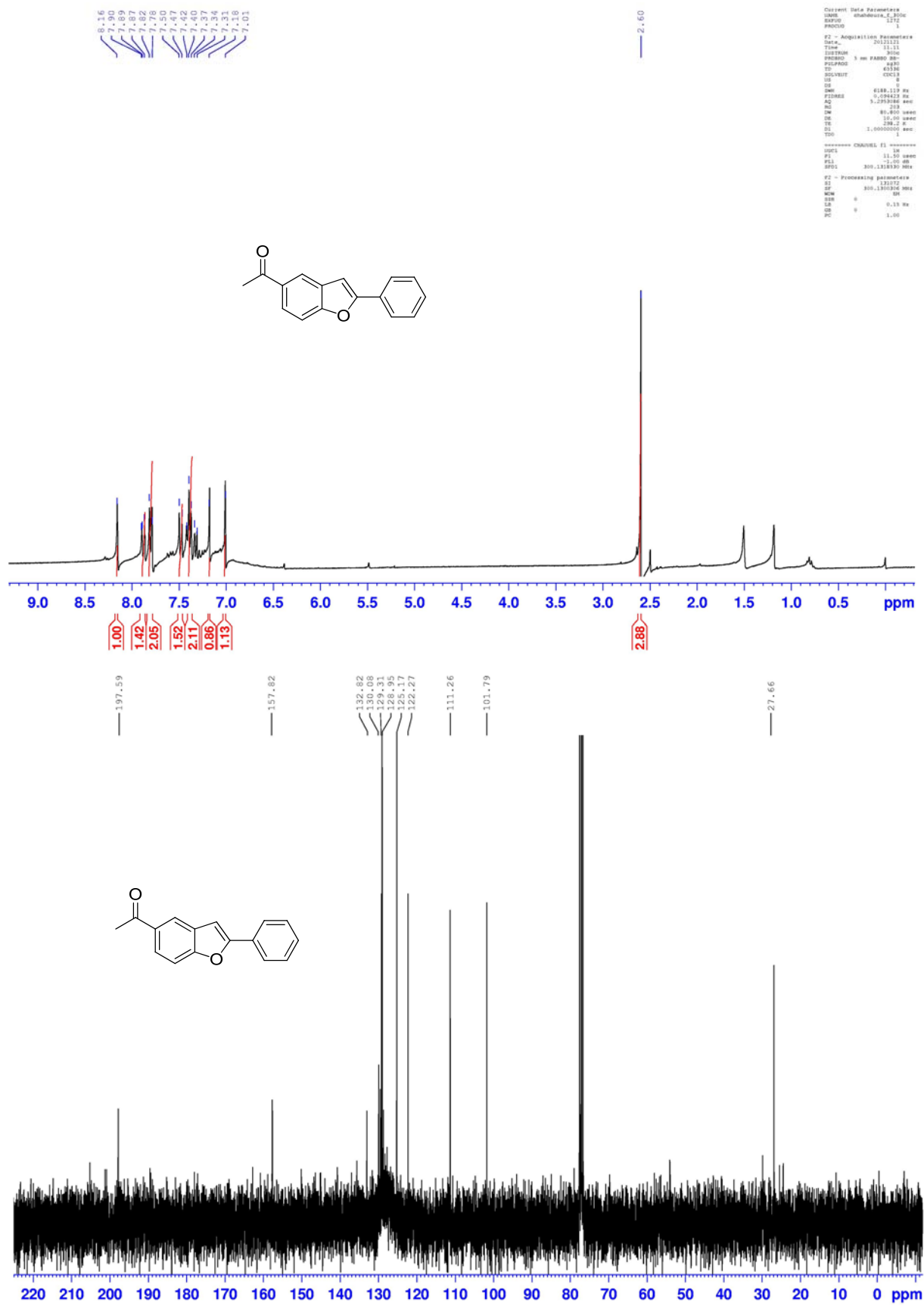


877  
fc-877-2 2152 (14.763) Cm (2138:2156)

, 20-Nov-2012 + 21:08:48

Scan EI+  
4.62e8

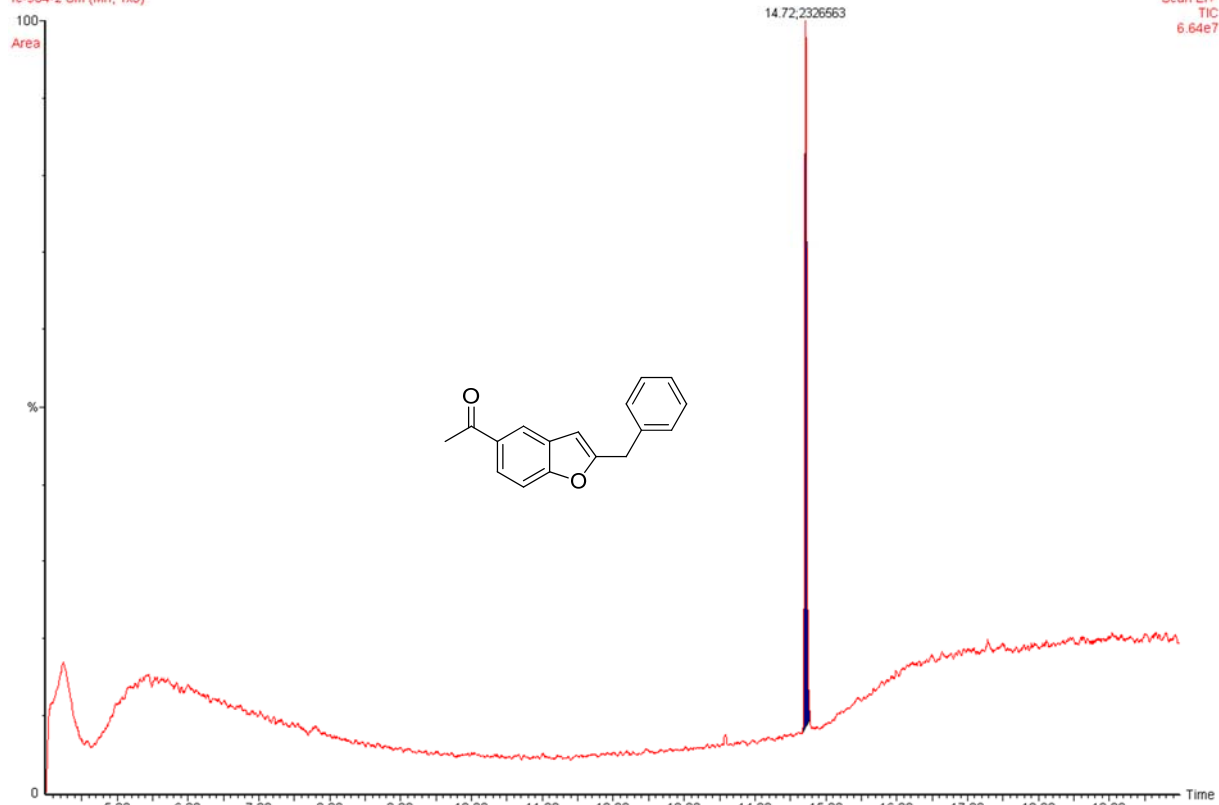




GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **t33**

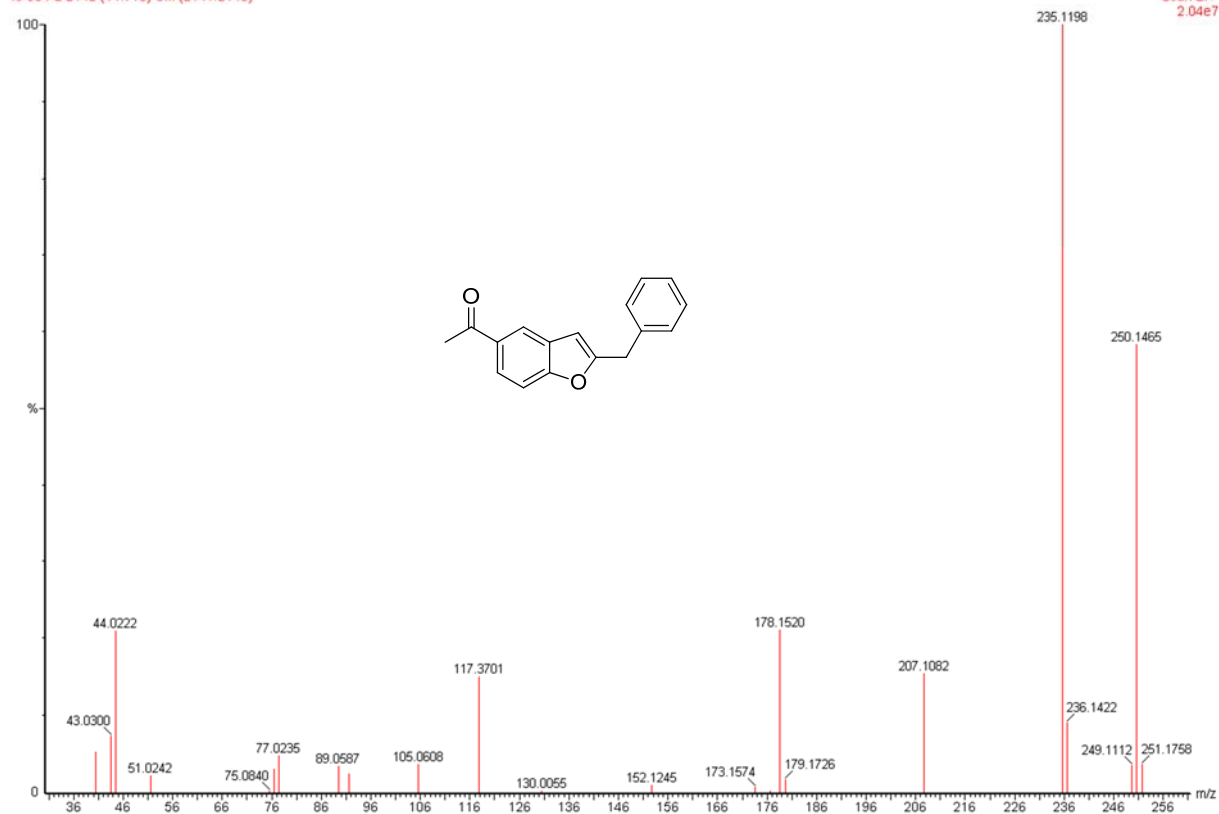
934  
fc-934-2 Sm (Mn, 1x3)

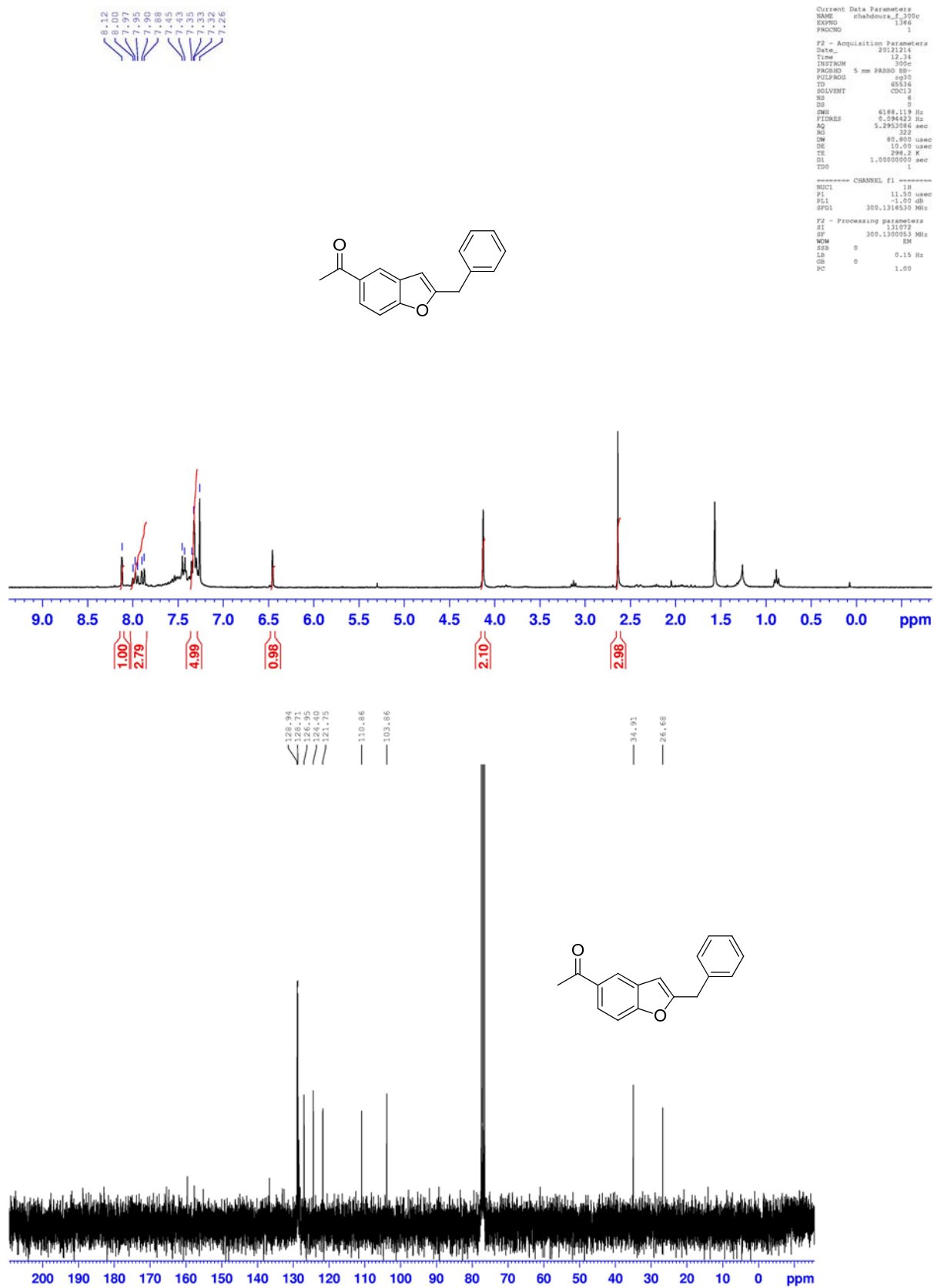
, 14-Dec-2012 + 10:30:50  
Scan EI+  
TIC  
6.64e7



934  
fc-934-2 2142 (14.713) Cm (2141:2148)

, 14-Dec-2012 + 10:30:50  
Scan EI+  
2.04e7



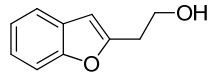
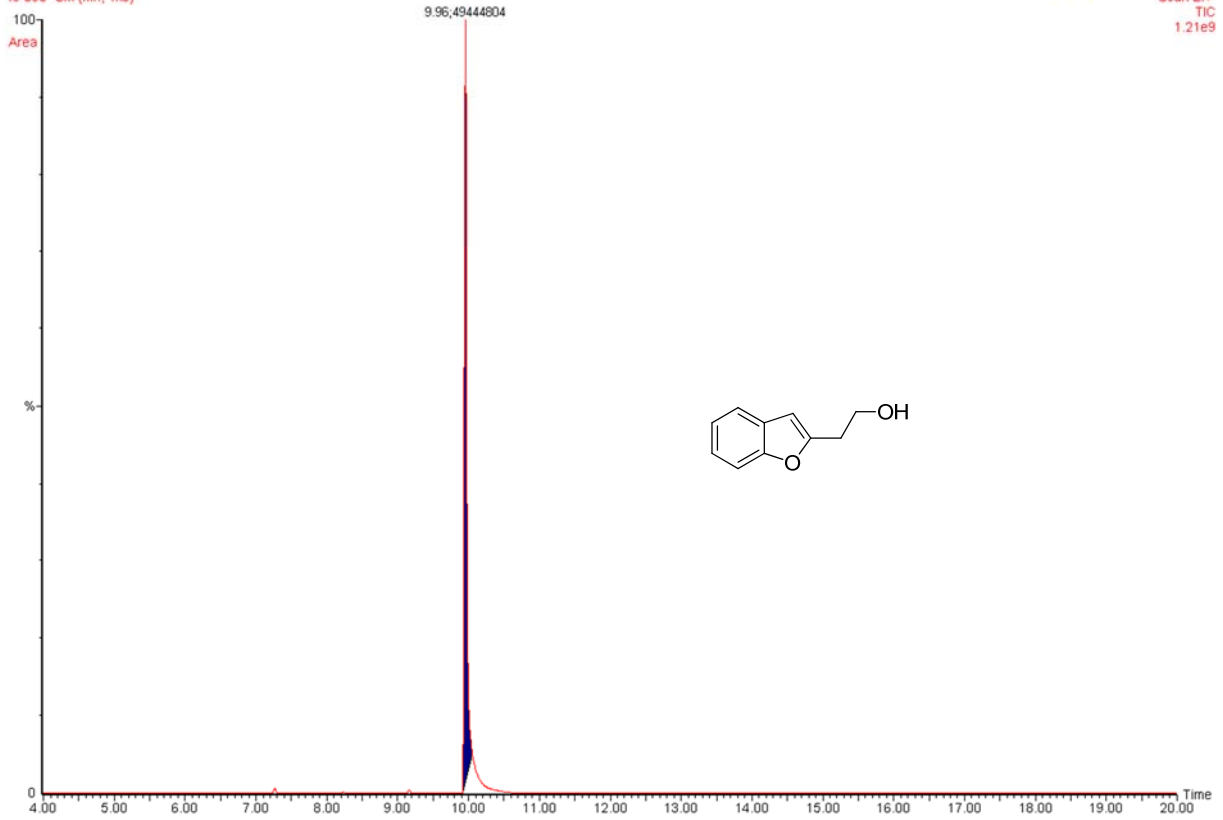


GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **t34**



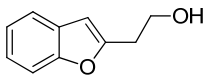
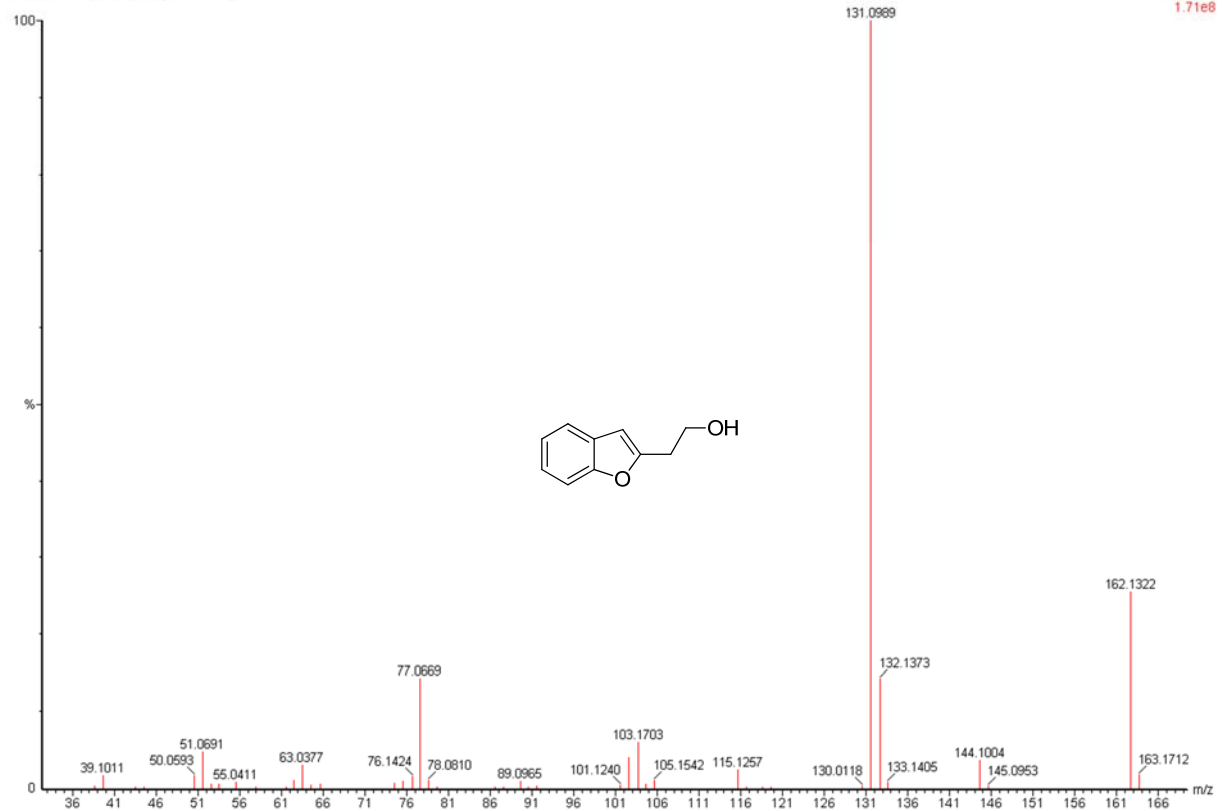
693  
tc-693- Sm (Mn, 1x3)

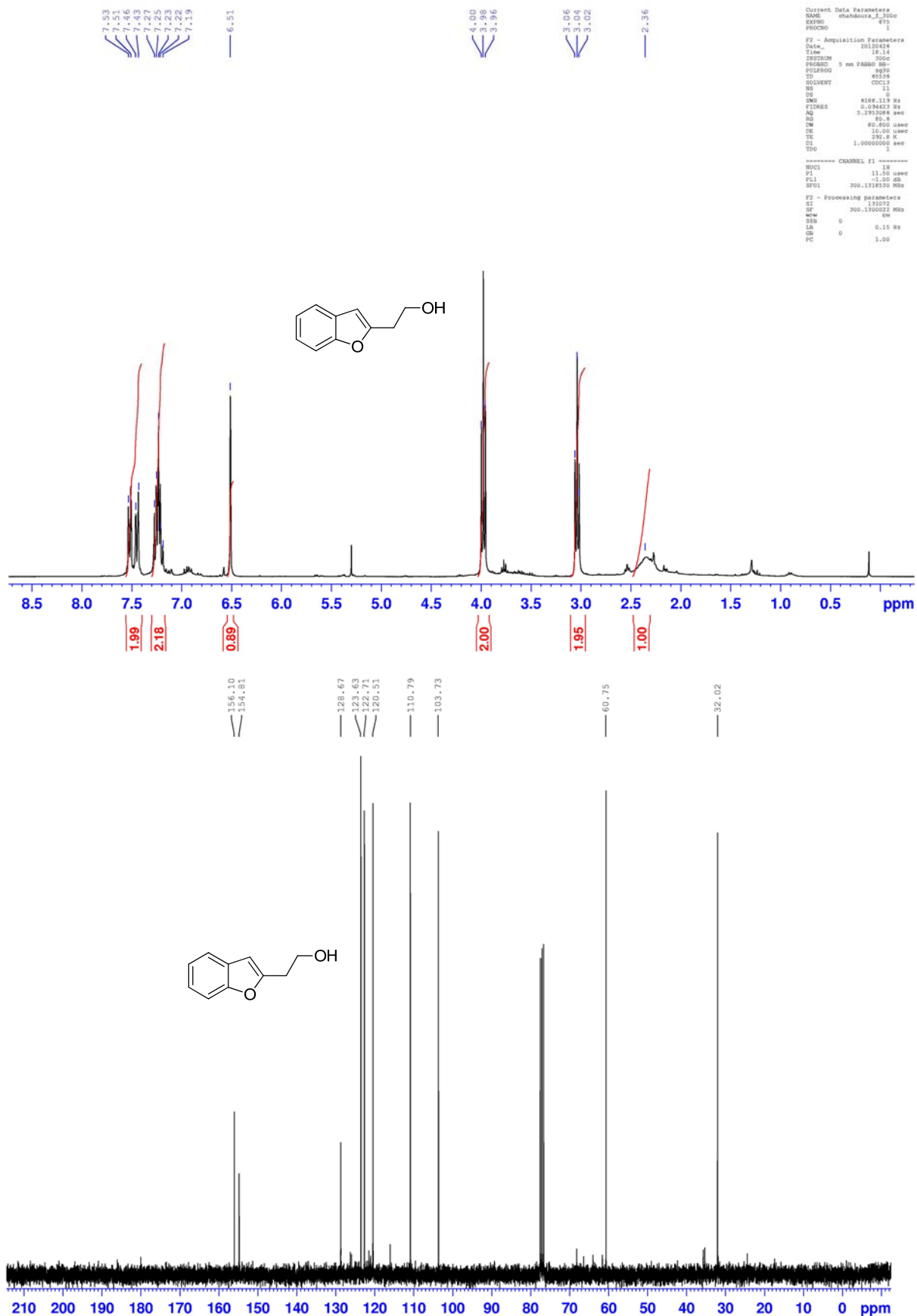
, 26-Apr-2012 + 17:05:14  
Scan EI+  
TIC  
1.21e9



693  
tc-693- 1192 (9.961) Cm (1191:1217)

, 26-Apr-2012 + 17:05:14  
Scan EI+  
1.71e8

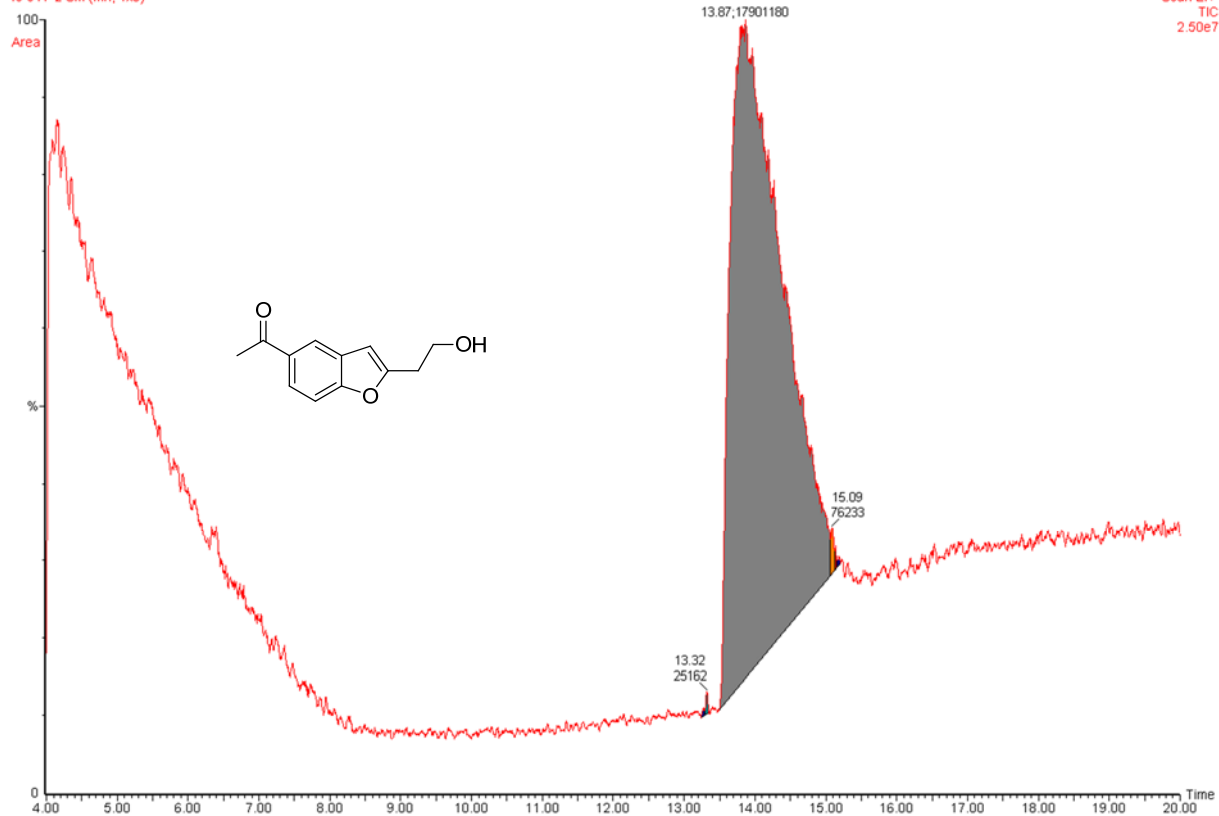




GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for s35

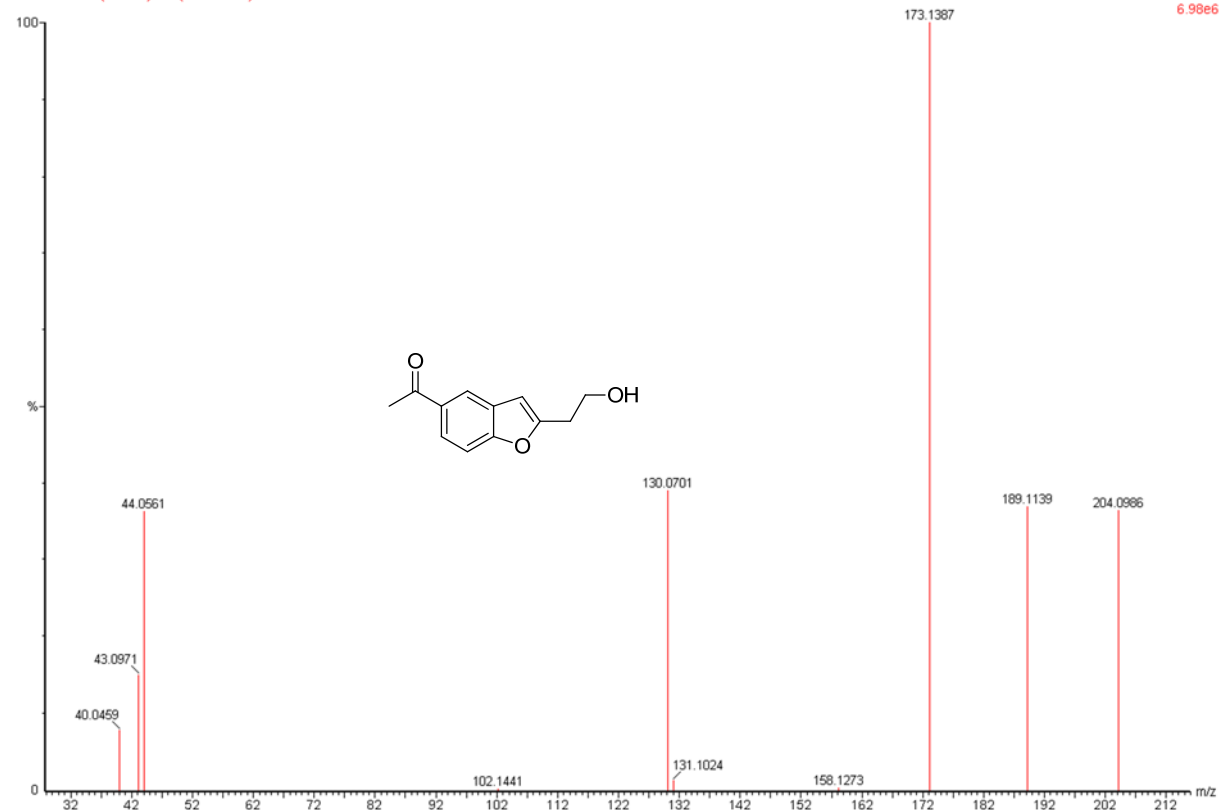
917  
fc-917-2 Sm (Mn, 1x3)

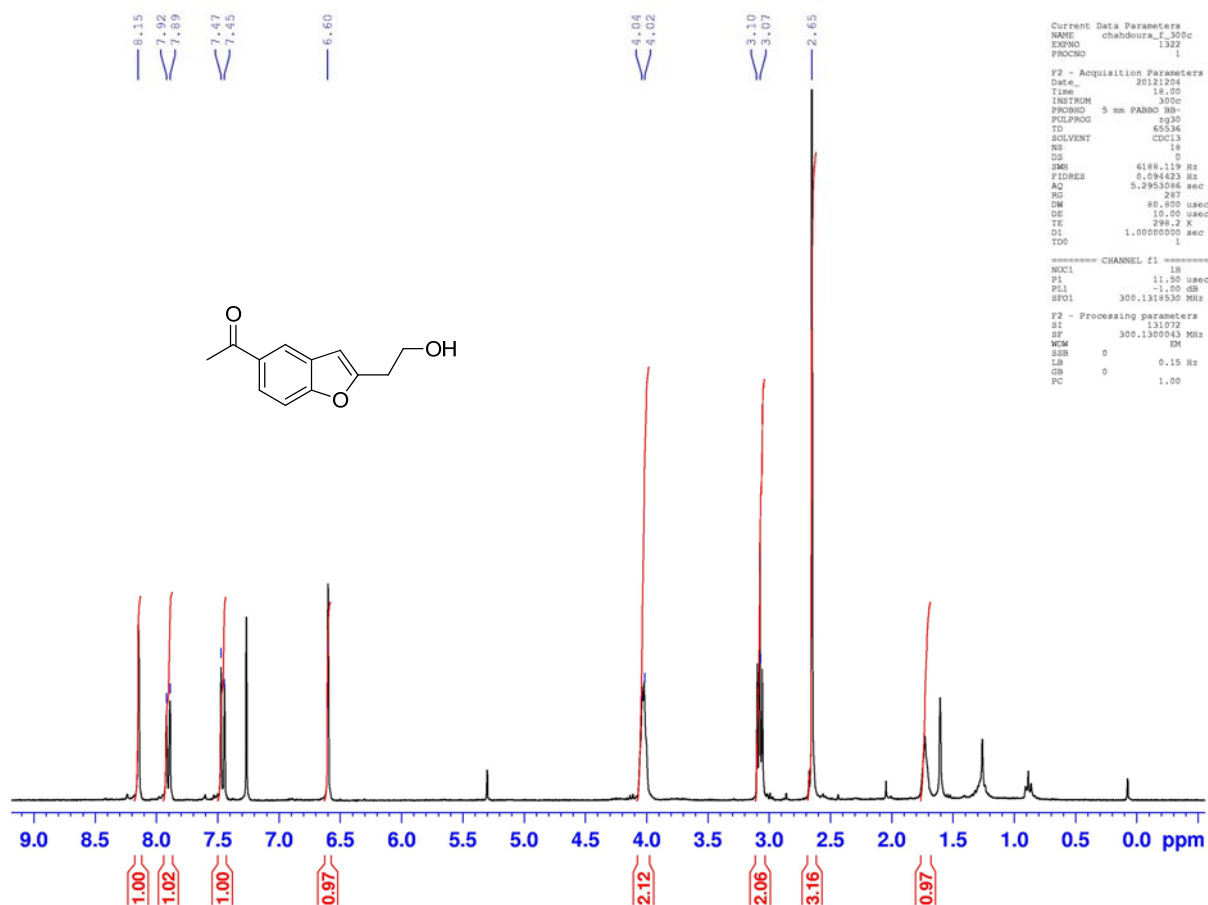
, 04-Dec-2012 + 18:35:47  
Scan EI+  
TIC  
2.50e7



917  
fc-917-2 1995 (13.977) Cm (1977:2121)

, 04-Dec-2012 + 18:35:47  
Scan EI+  
6.98e6





```

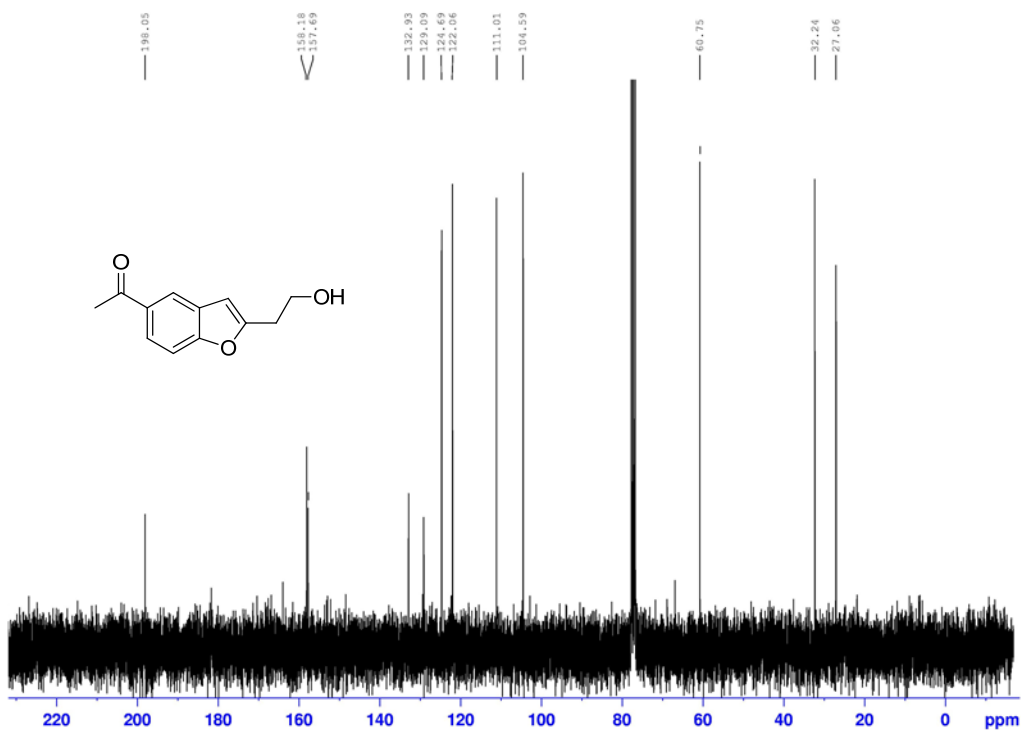
Current Data Parameters
NAME chahdoura_f_303c
EXPNO 1322
PROCNO 1

F2 - Acquisition Parameters
Date_ 20121204
Time 18.00
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 18
DS 0
SFO 618.119 Hz
FIDRES 0.094423 Hz
AQ 5.2953286 sec
RG 287
DM 80.800 usec
DE 10.00 usec
TE 298.2 K
D1 1.0000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
PI 11.50 usec
P1 1.00 dB
SFO1 300.1318530 MHz

F2 - Processing parameters
SI 131072
SF 300.1300043 MHz
WDW EM
SSB 0
LB 0.15 Hz
GB 0
PC 1.00

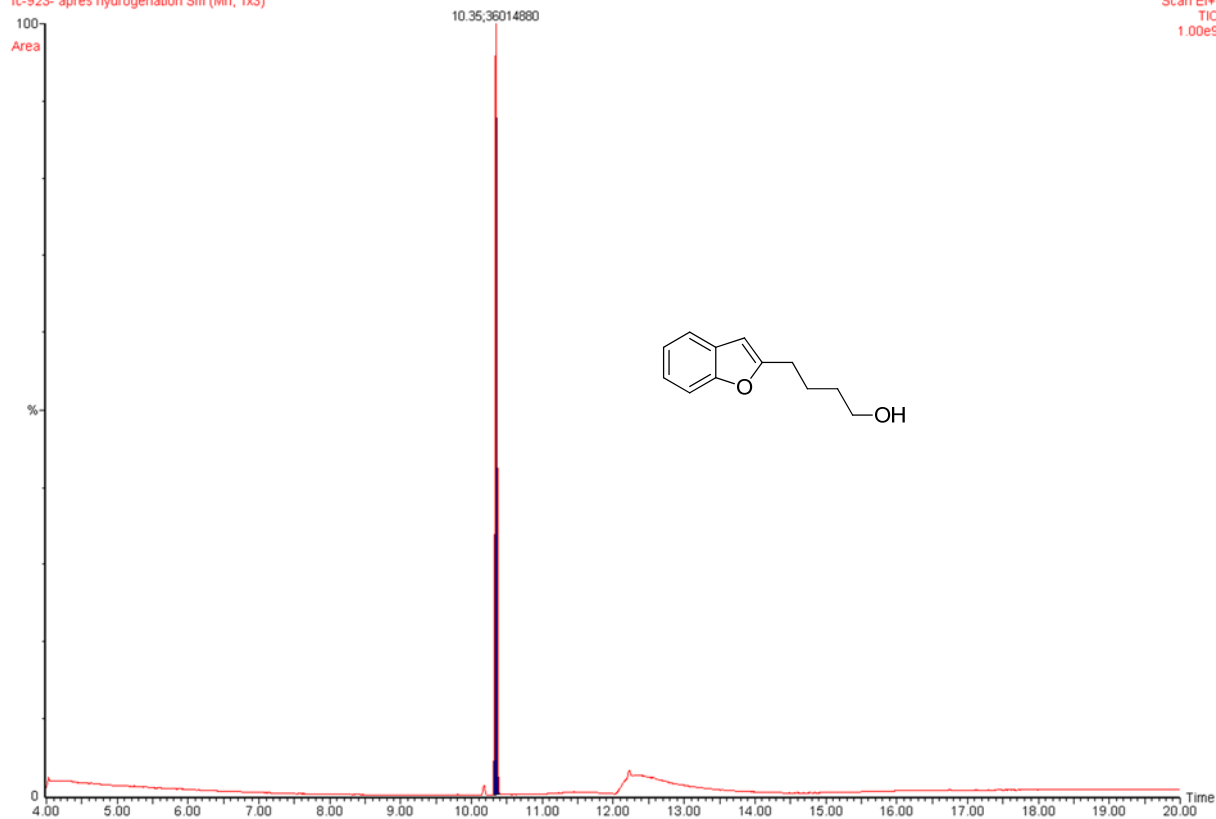
```



GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **t35**

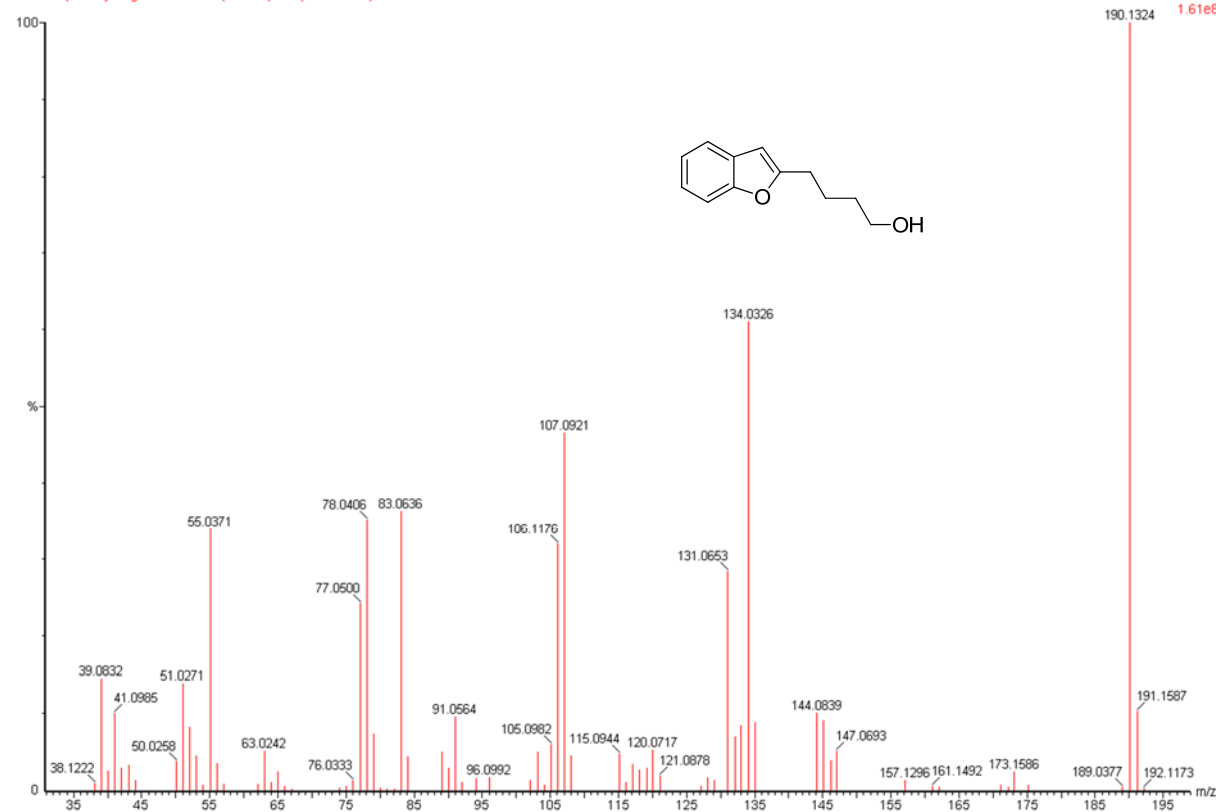
923  
fc-923- apres hydrogenation Sm (Mn, 1x3)

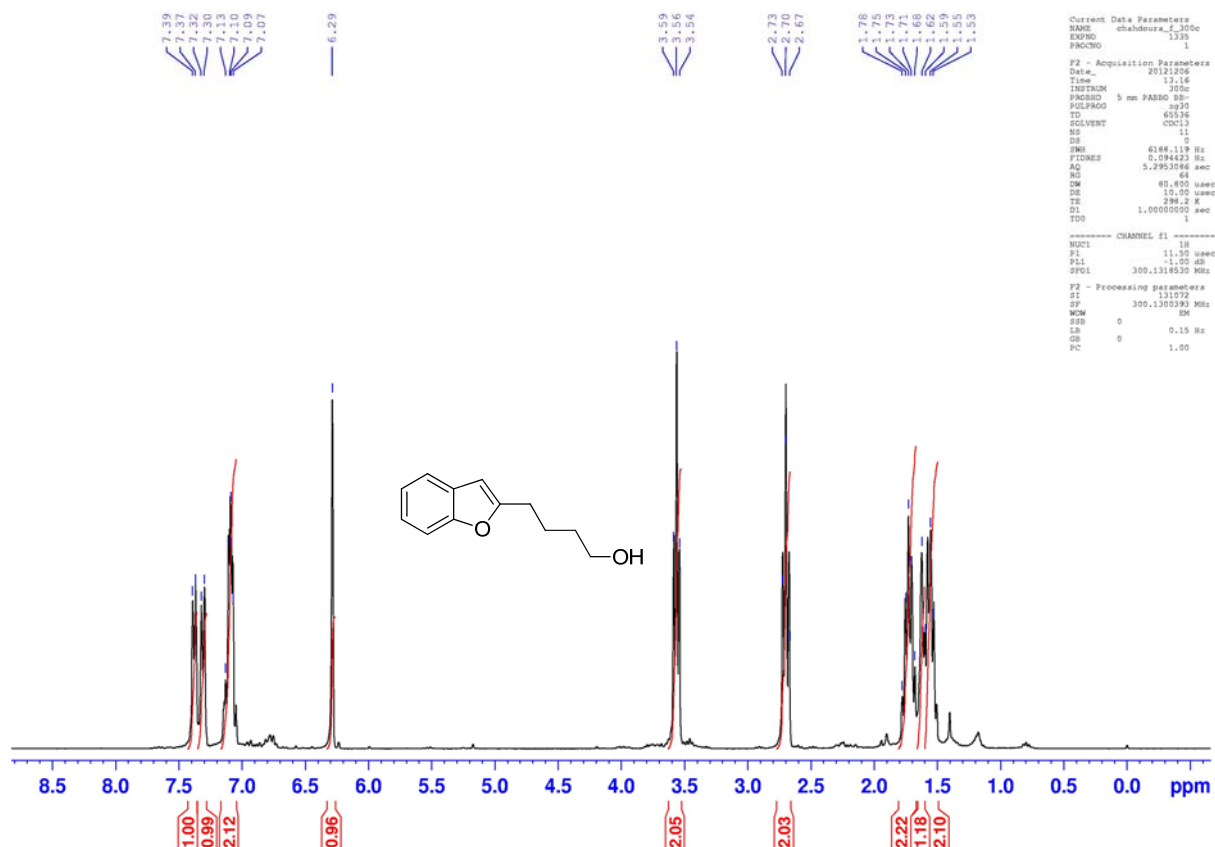
, 07-Dec-2012 + 14:47:57  
Scan E1+  
TIC  
1.00e9



923  
fc-923- apres hydrogenation 1270 (10.351) Cm (1267:1273)

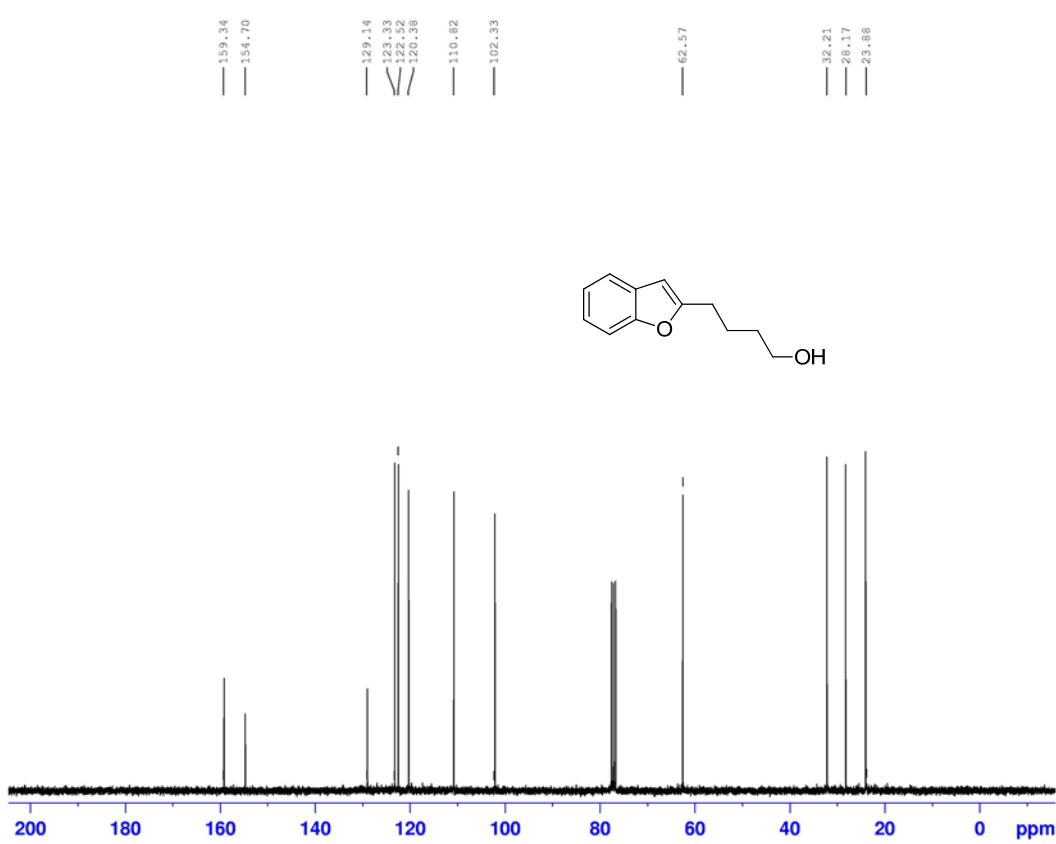
, 07-Dec-2012 + 14:47:57  
Scan E1+  
1.61e8





```

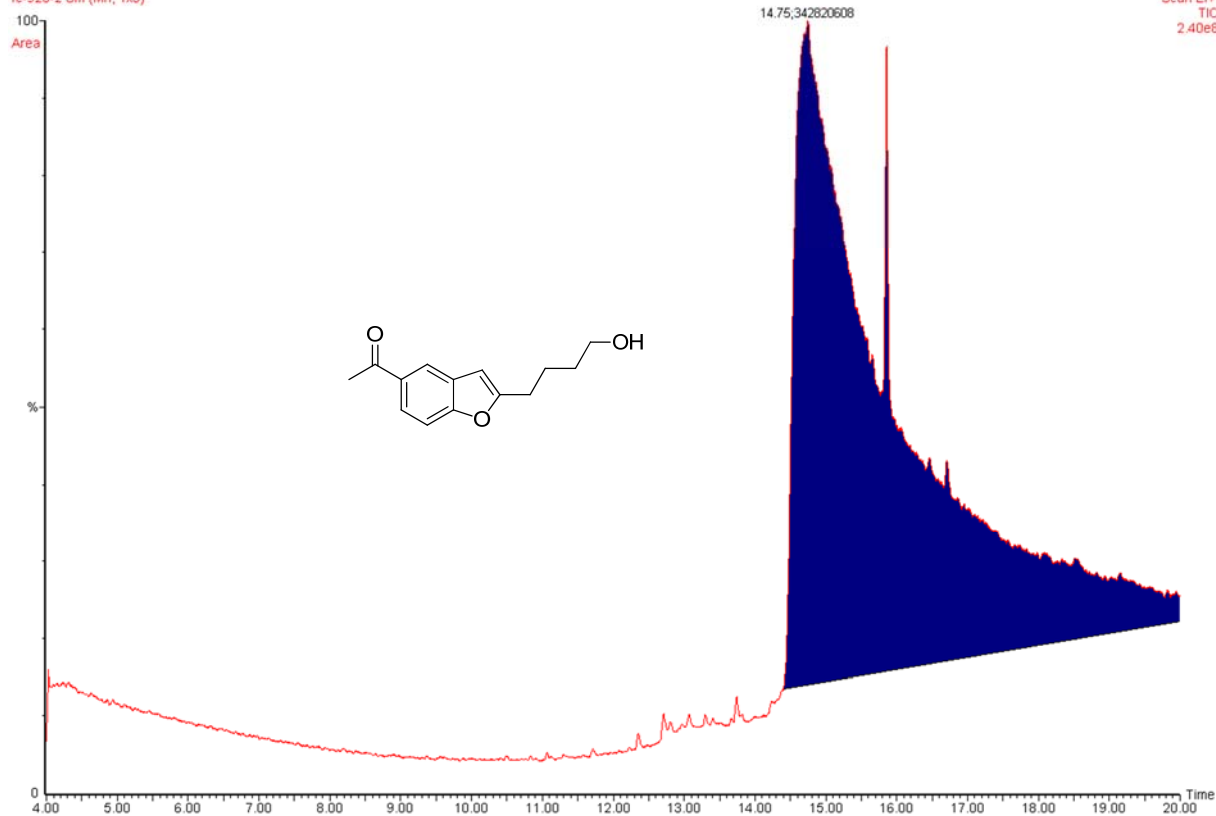
Current Data Parameters
NAME chahdura_s36
EXPNO 133
PROCNO 1
F2 - Acquisition Parameters
Date_ 201206
Time 11:16
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 11
DS 0
SWH 6188.119 Hz
FIDRES 0.094423 Hz
AQ 5.2953066 sec
RG 64
AQ 5.2953066 sec
DE 80.800 usec
TE 298.2 K
D1 1.00000000 sec
TSD 1
----- CHANNEL f1 -----
NUC1 1H
P1 11.00 usec
PL1 -1.00 dB
SFO1 300.1318530 MHz
F2 - Processing parameters
SI 131072
SF 300.1305390 MHz
WDW EM
SSB 0
GB 0
PC 1.00
  
```



GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for s36

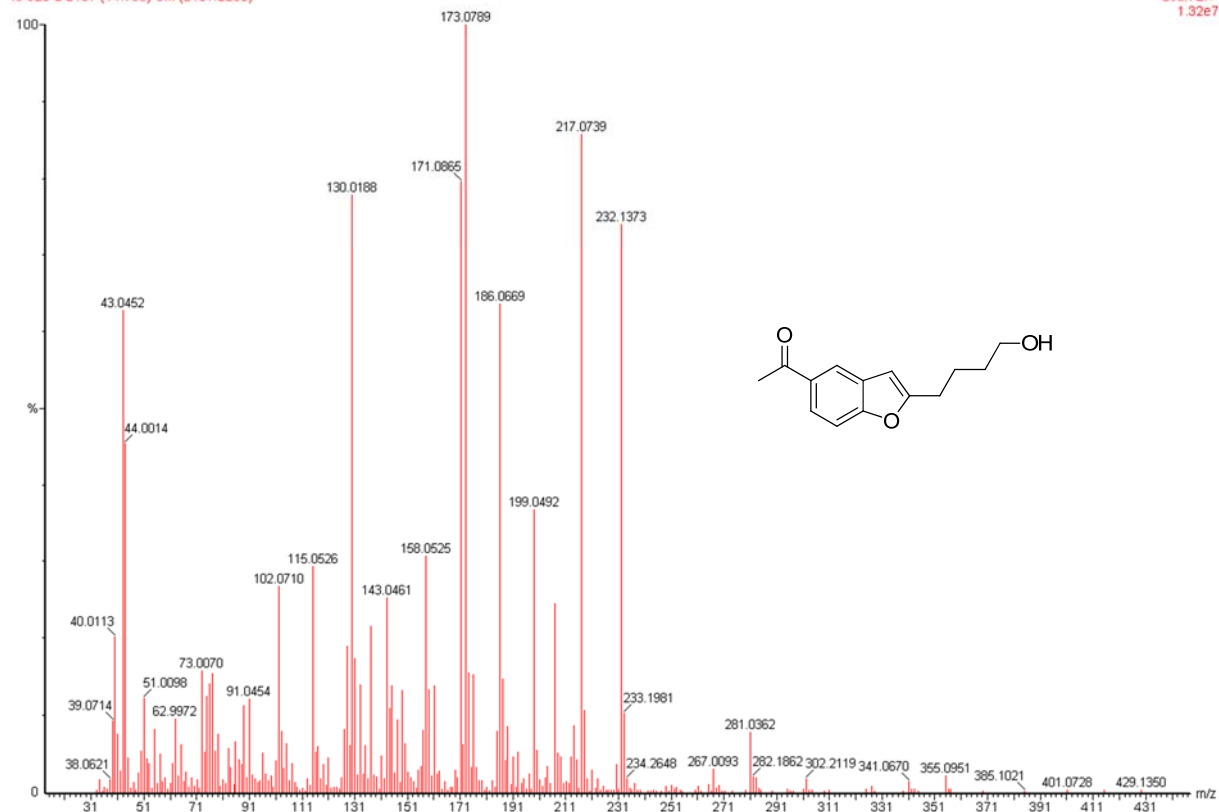
926  
fc-926-2 Sm (Mn, 1x3)

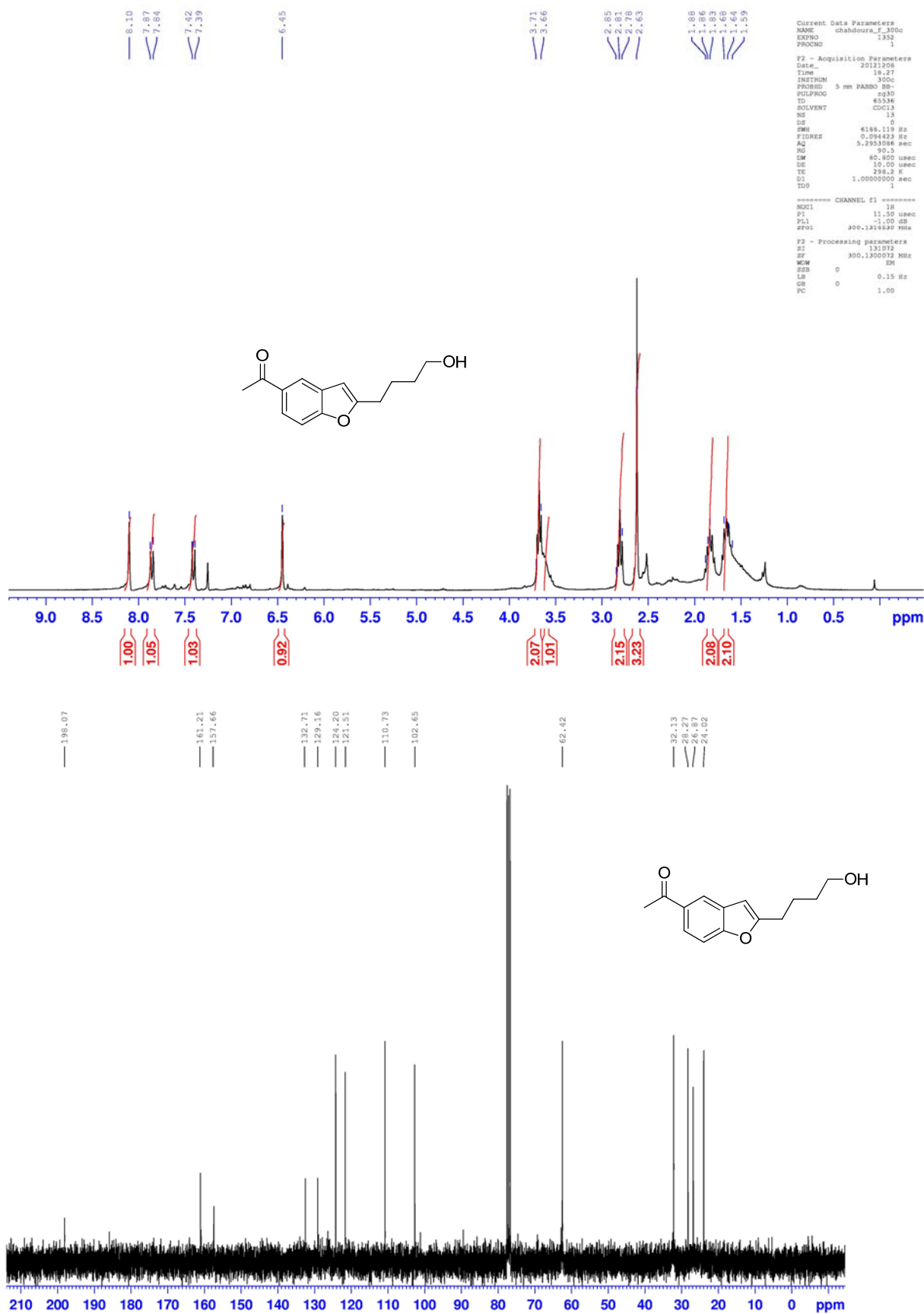
, 09-Dec-2012 + 11:04:31  
Scan E1+  
TIC  
2.40e8



926  
fc-926-2 2157 (14.788) Cm (2157:2236)

, 09-Dec-2012 + 11:04:31  
Scan E1+  
1.32e7



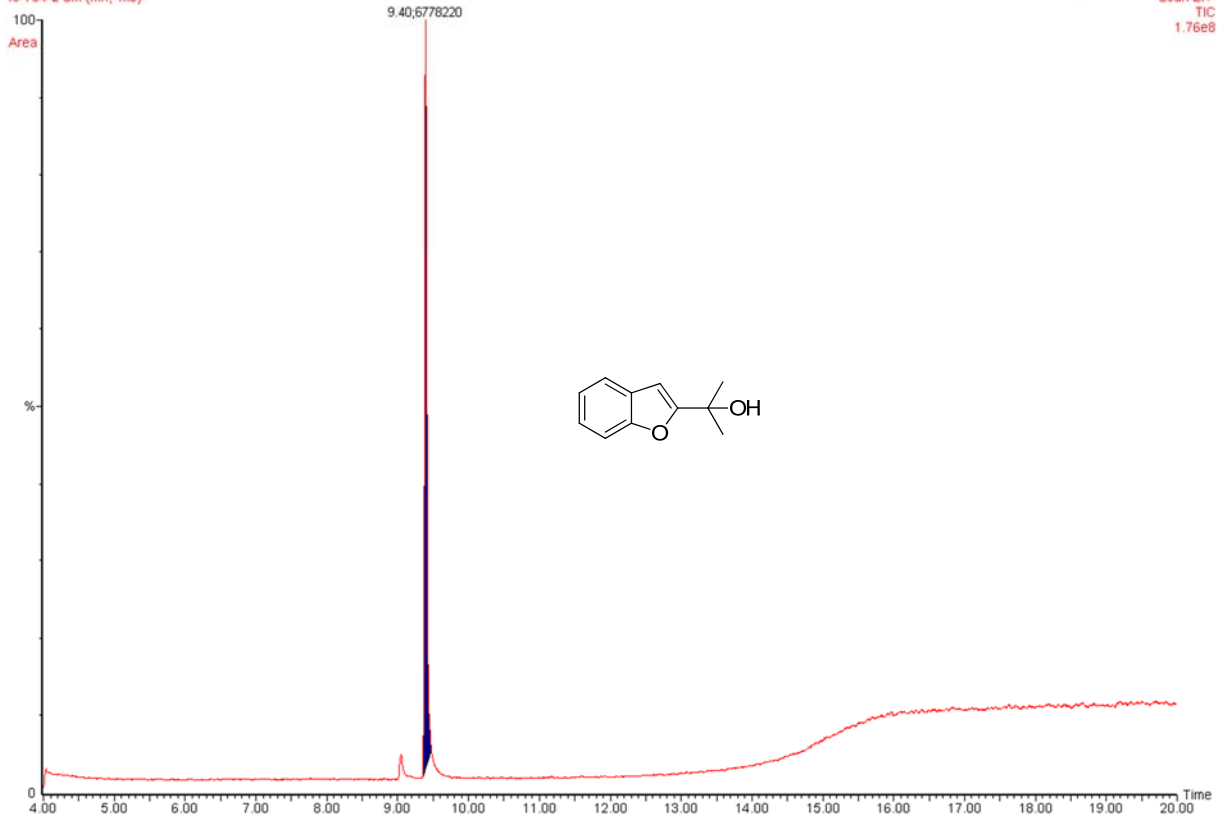


GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **t36**



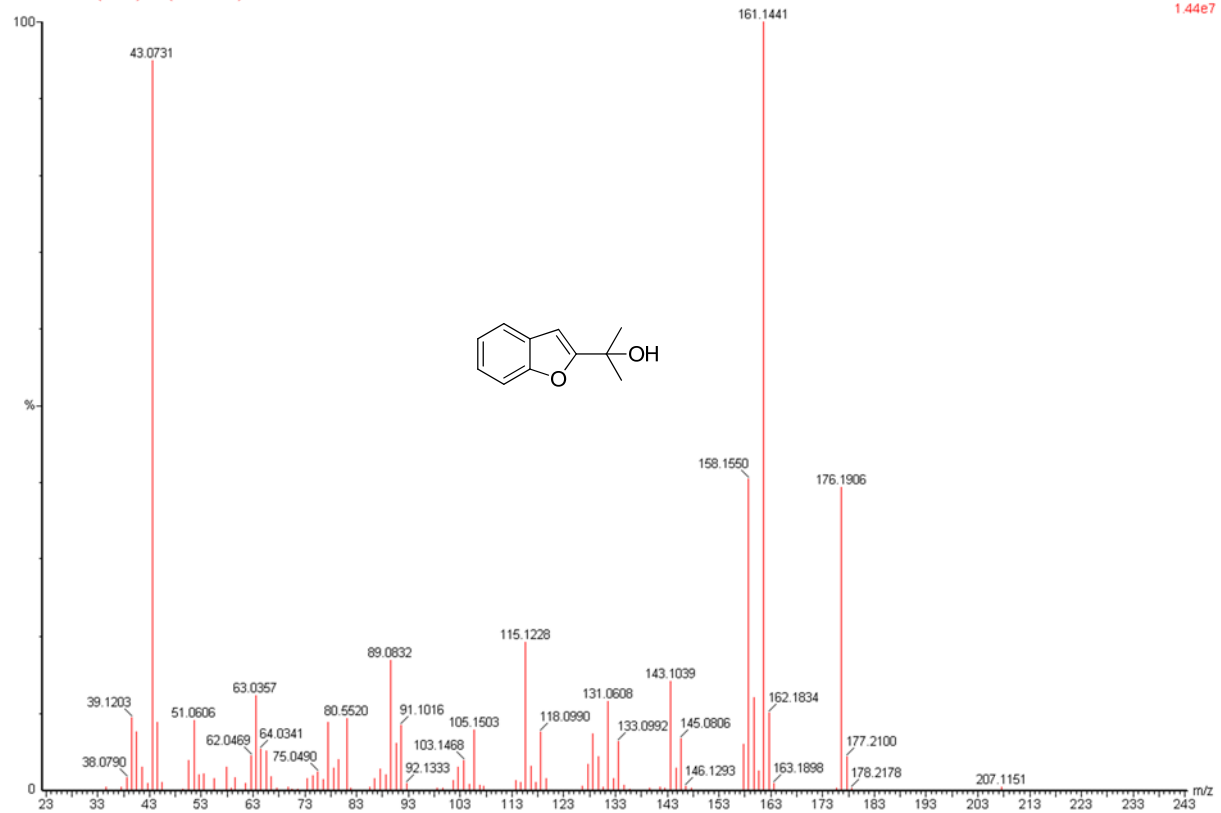
704  
fc-704-2 Sm (Mn, 1x3)

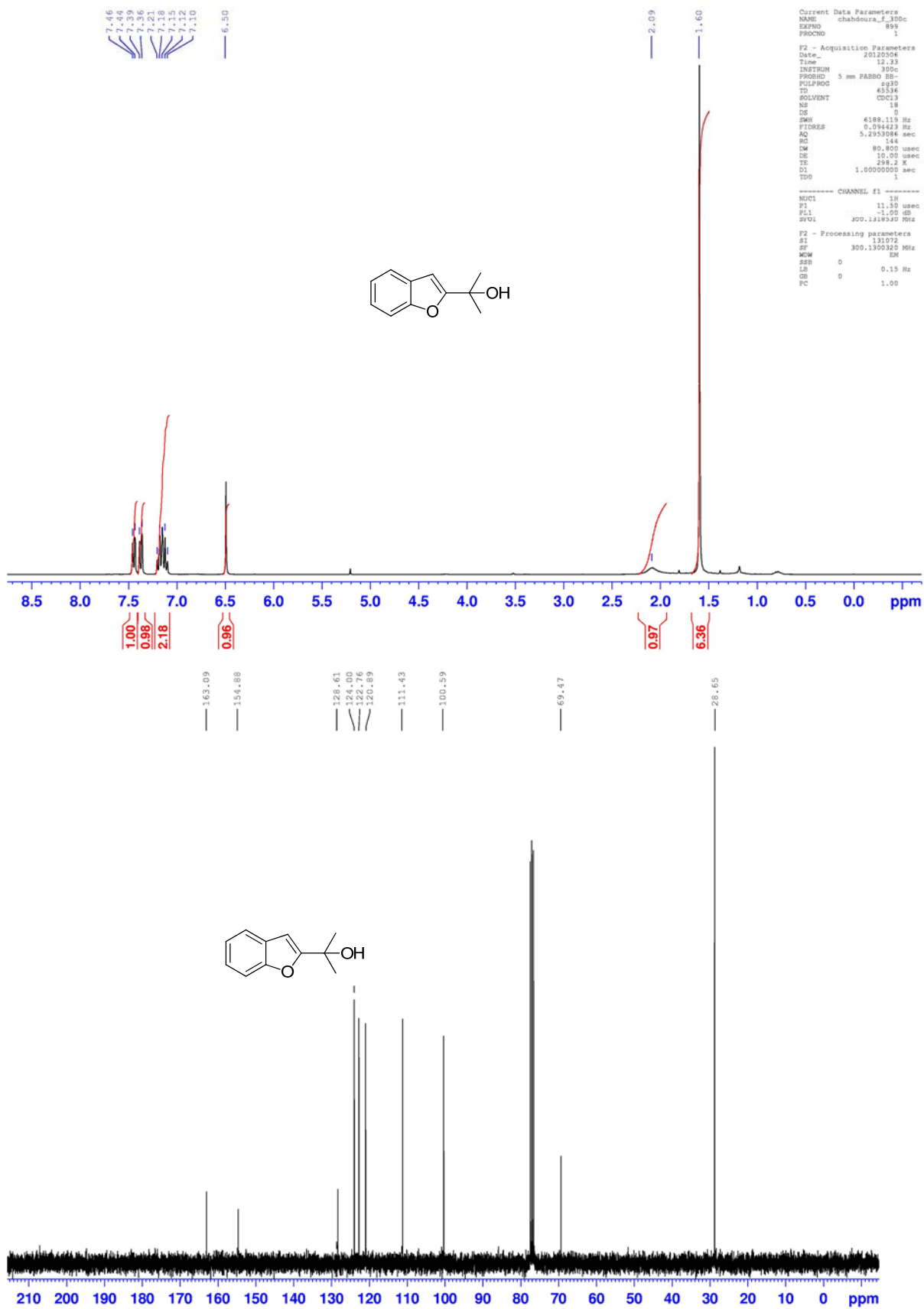
, 06-May-2012 + 12:42:04  
Scan EI+  
TIC  
1.76e8



704  
fc-704-2 1079 (9.396) Cm (1076:1092)

, 06-May-2012 + 12:42:04  
Scan EI+  
1.44e7



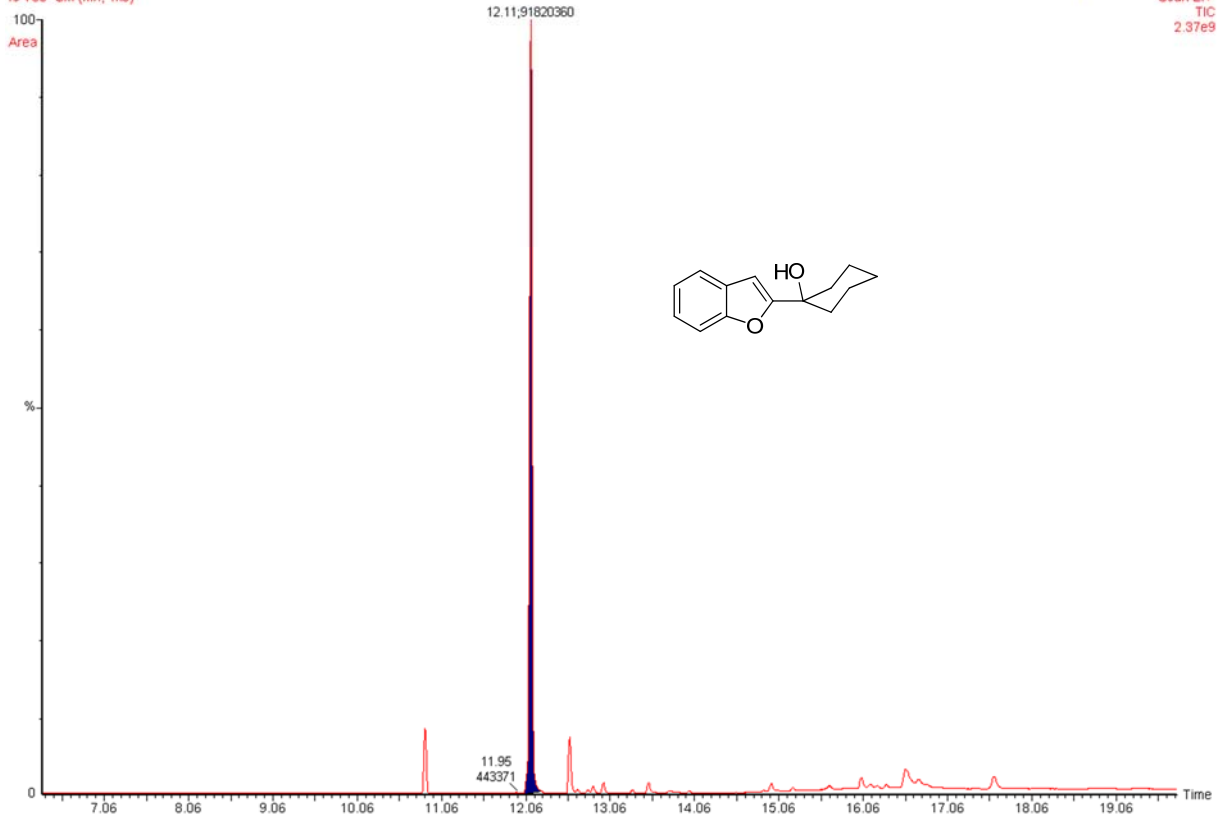


GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **s37**

700  
tc-700- Sm (Mn, 1x3)

, 04-May-2012 + 11:41:06

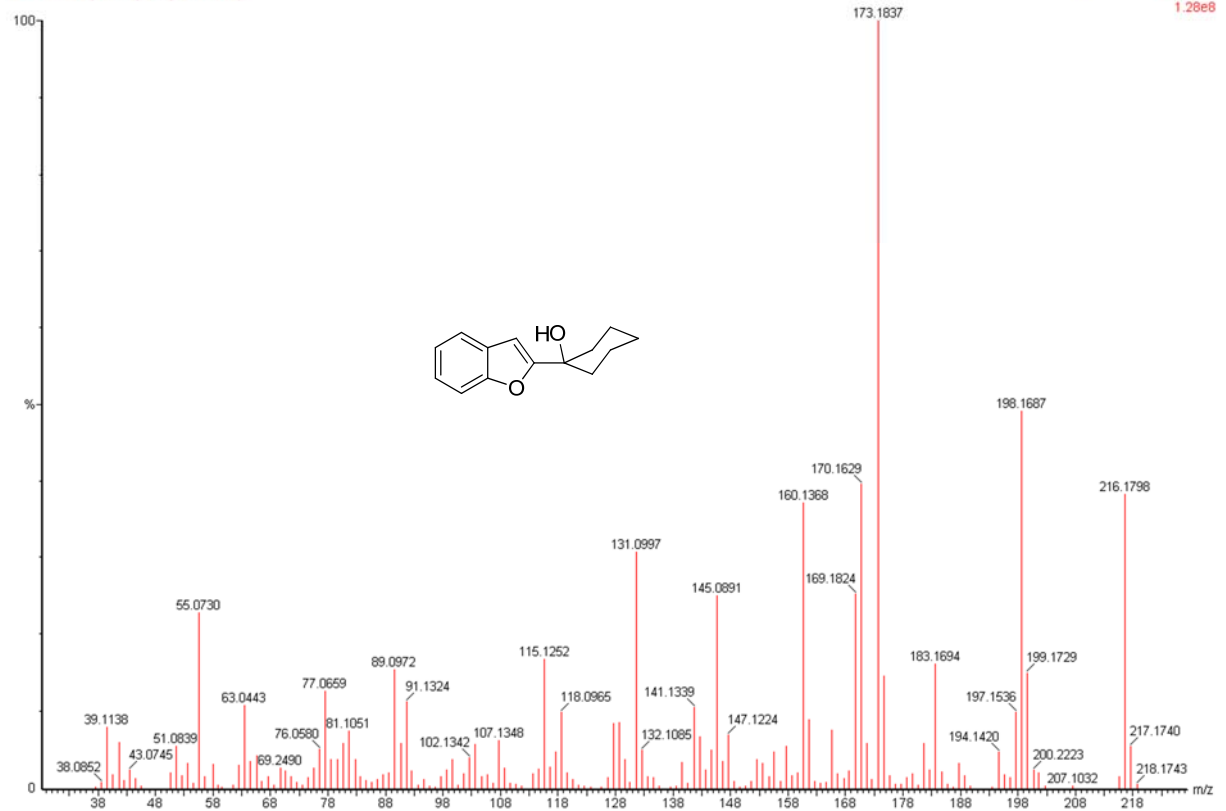
Scan EI+  
TIC  
2.37e9

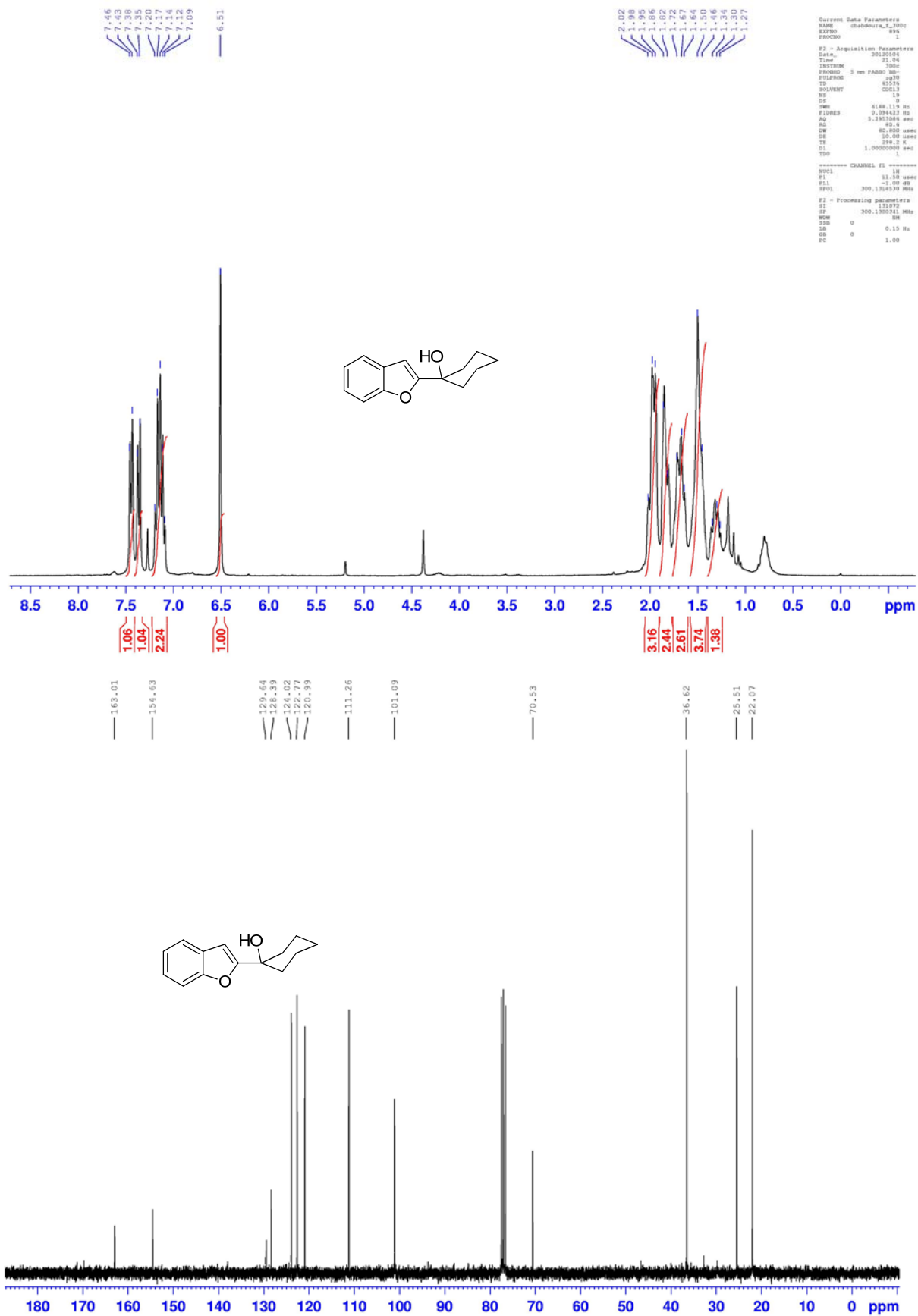


700  
tc-700- 1623 (12.117) Cm (1612:1628)

, 04-May-2012 + 11:41:06

Scan EI+  
1.28e8

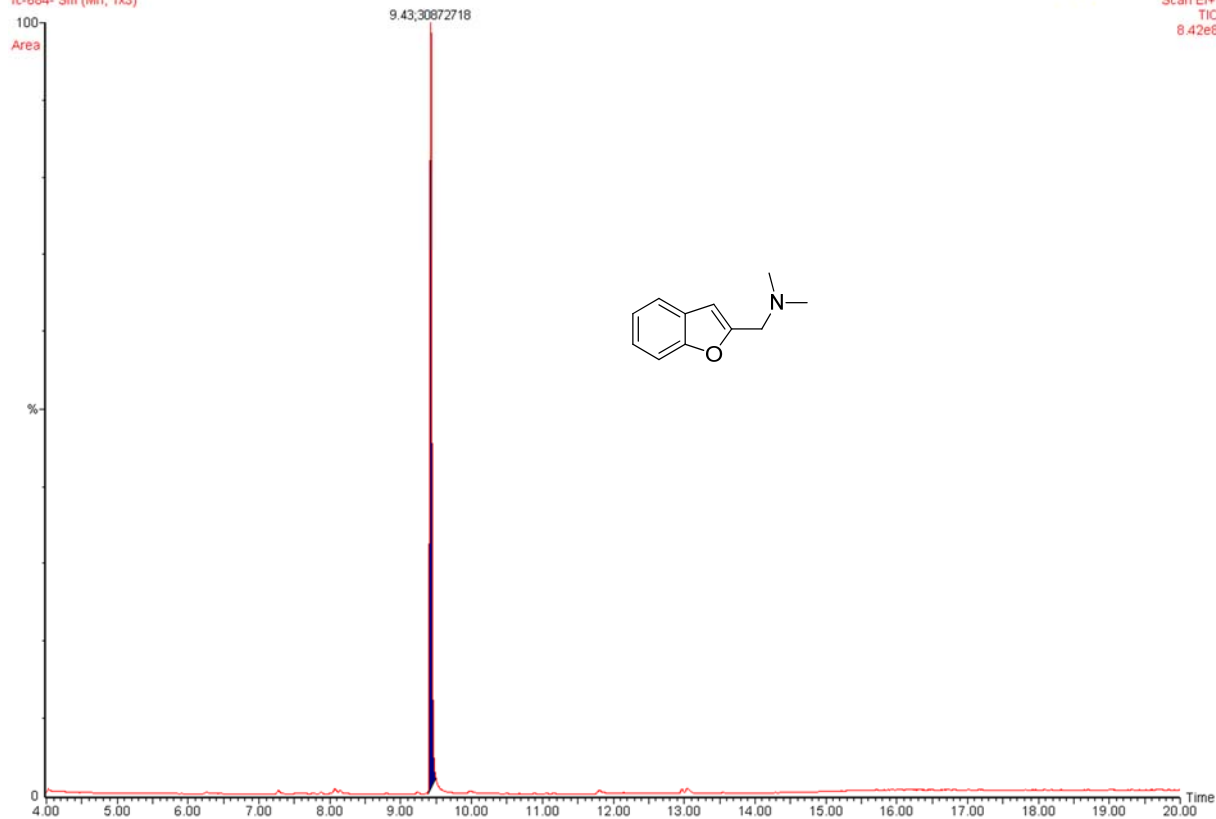




GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **s38**

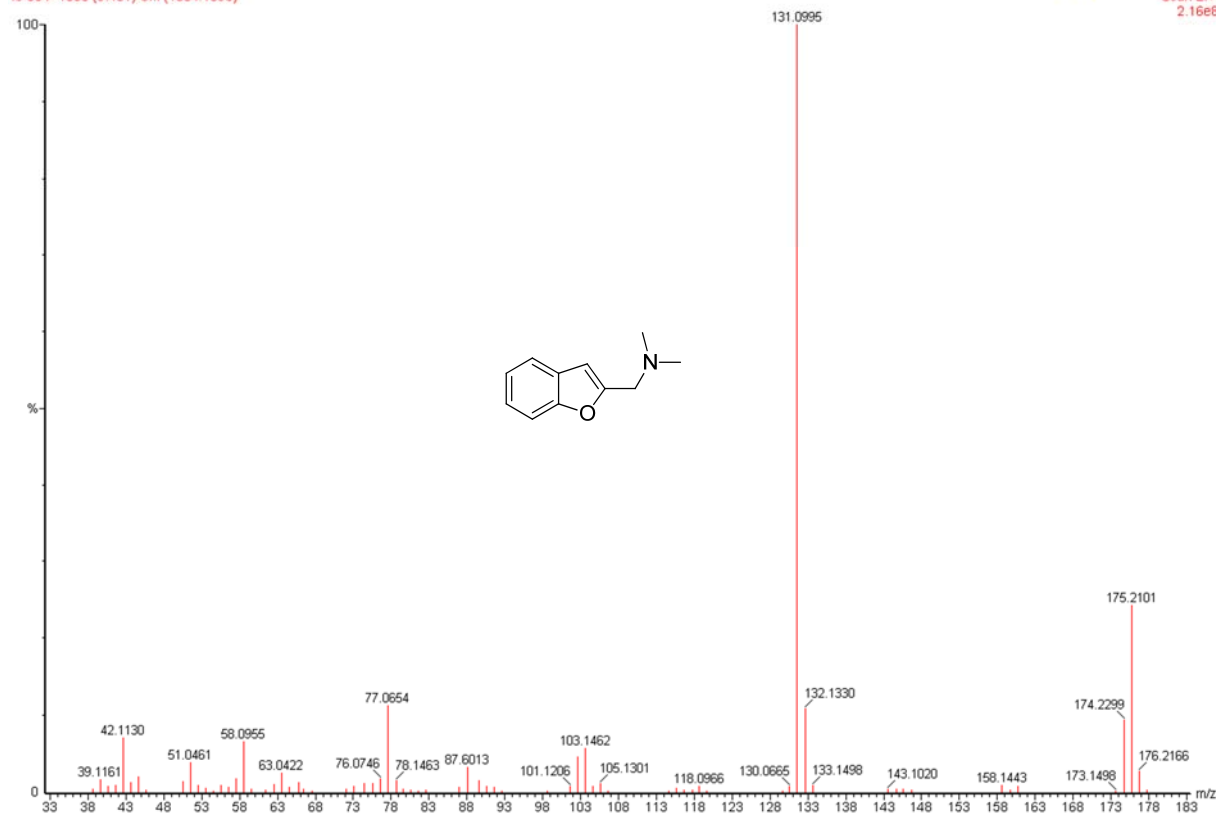
684  
tc-684- Sm (Mn, 1x3)

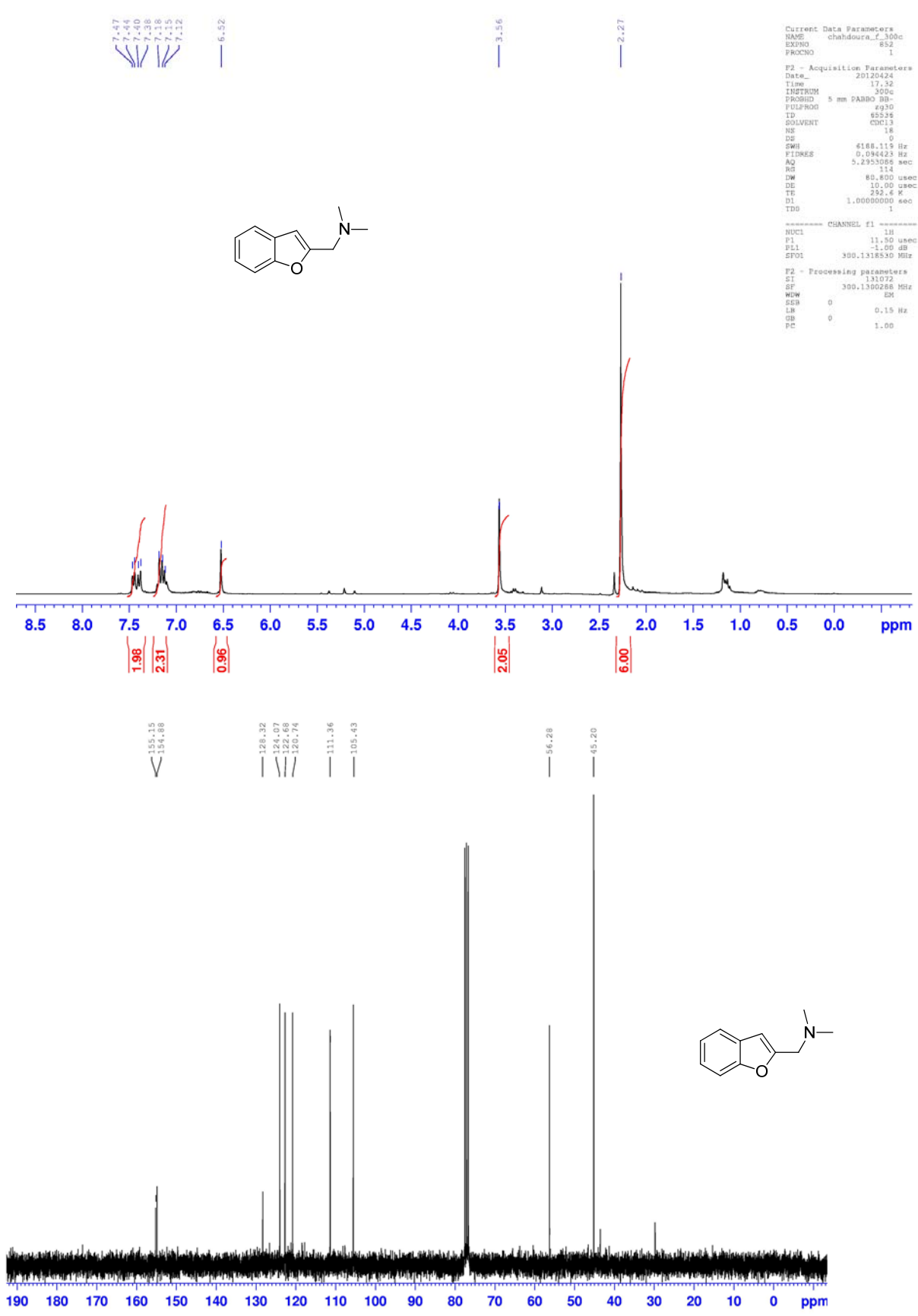
, 24-Apr-2012 + 14:33:17  
Scan EI+  
TIC  
8.42e8



684  
tc-684- 1086 (9.431) Cm (1084:1095)

, 24-Apr-2012 + 14:33:17  
Scan EI+  
2.16e8

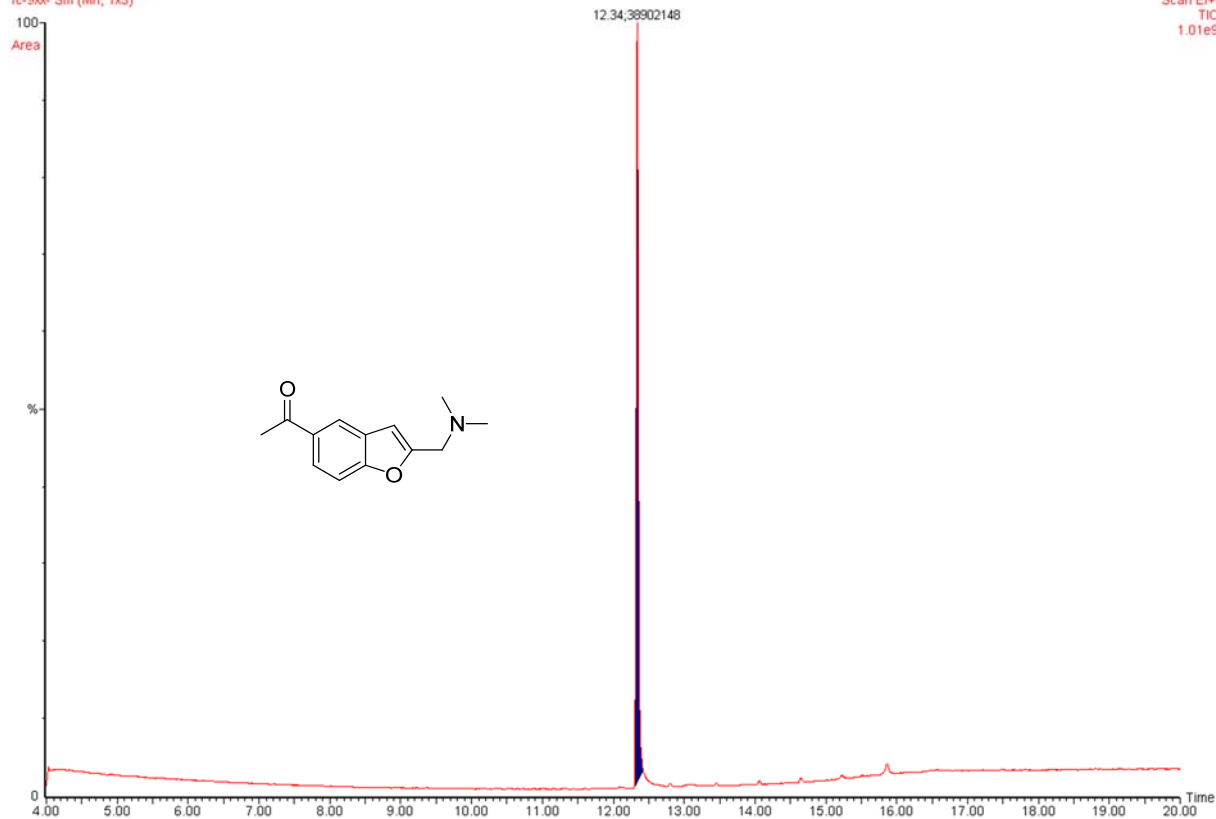




GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **s39**

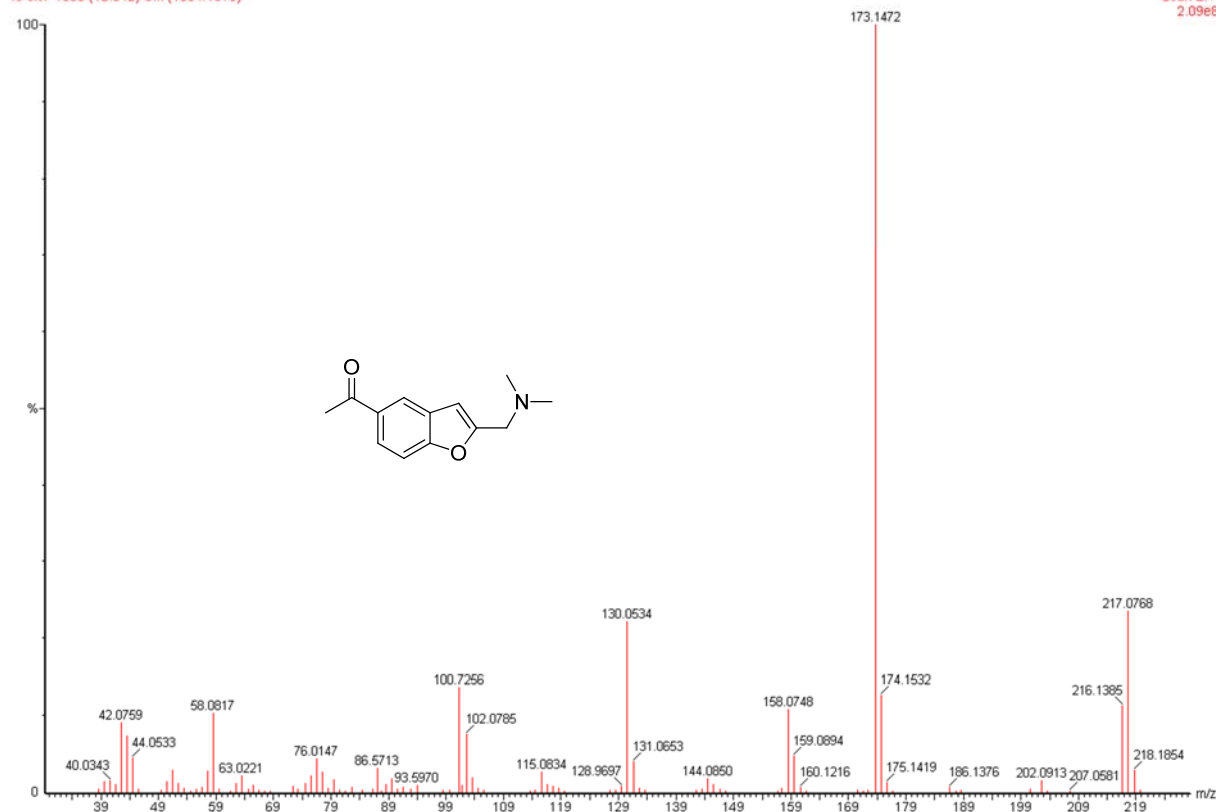
99XX  
fc-9xx- Sm (Mn, 1x3)

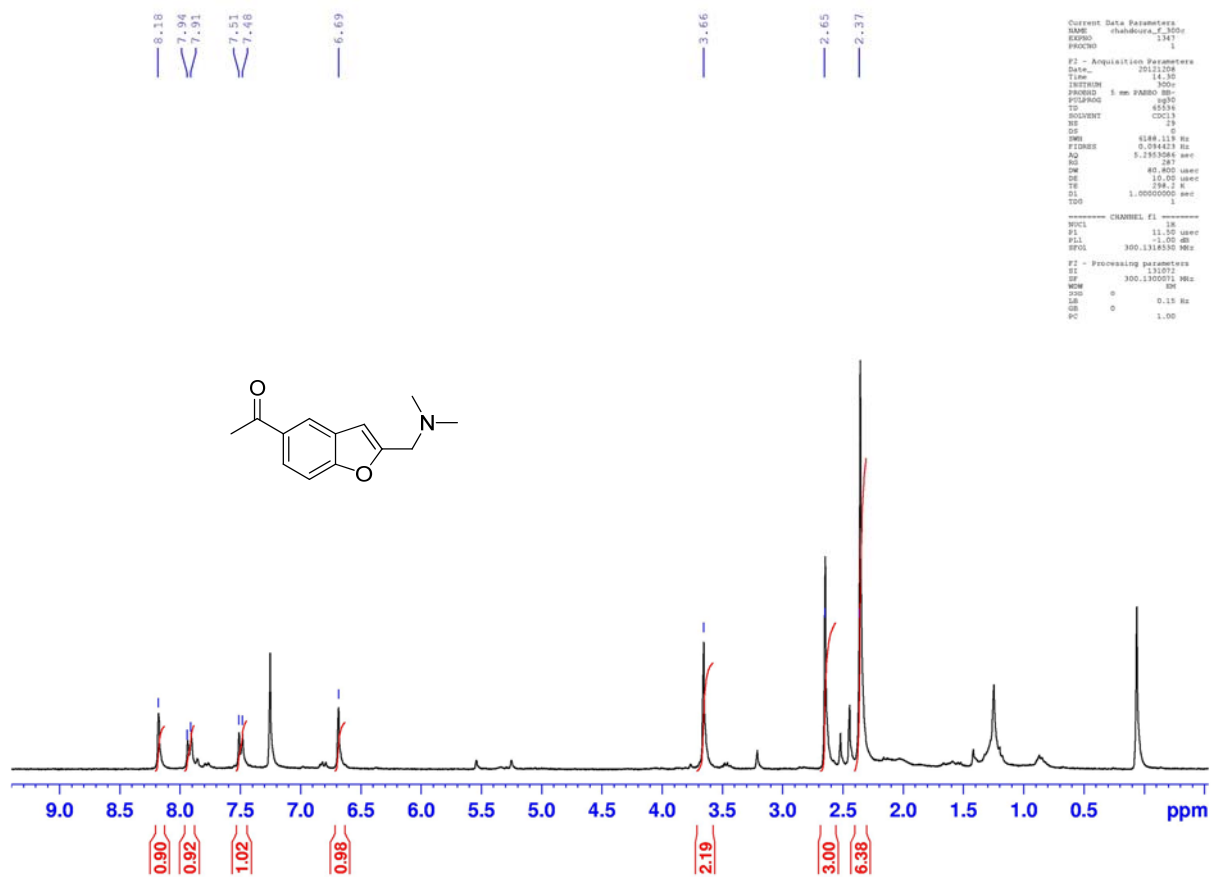
, 08-Dec-2012 + 15:25:52  
Scan EI+  
TIC  
1.01e9



99XX  
fc-9xx- 1668 (12.342) Cm (1664:1675)

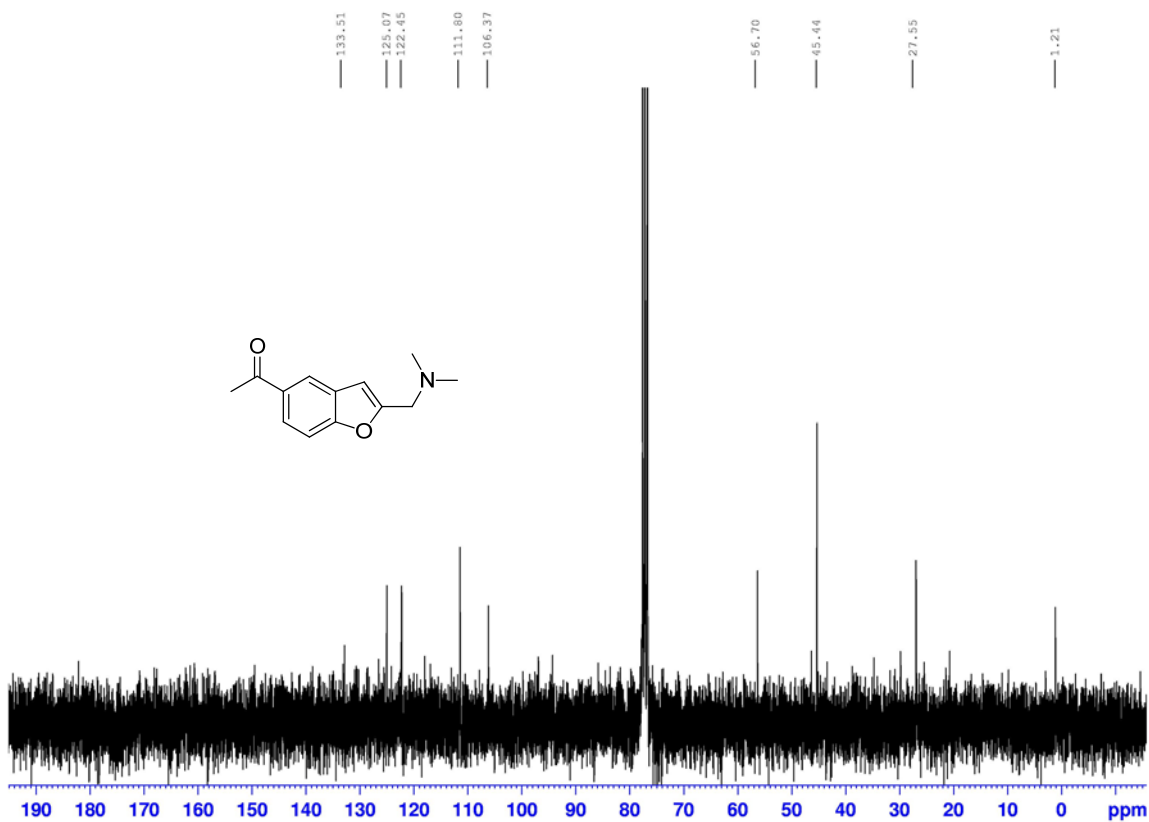
, 08-Dec-2012 + 15:25:52  
Scan EI+  
2.09e8





```

Current Data Parameters
NAME          chabouca_7_30a
EXPNO        1
PROCNO       1
F2 - Acquisition Parameters
Date_       201109
Time        14:30
INSTRUM     zgpg30
PROBHD      5 mm PABBO BB-
PULPROG     zgpg30
TD          65536
SOLVENT     CDCl3
NS          19
DS          4
SWH         6188.119 Hz
FIDRES     0.094243 Hz
AQ         5.2953284 sec
RG          287
DM         60.800 usec
DE         10.00 usec
TE         298.2 K
D1         1.00000000 sec
TDS        1
----- CHANNEL f1 -----
NUC1        13
P1          11.00 usec
PL         -1.00 dB
RF1         300.1318530 MHz
F2 - Processing parameters
SI          32768
SF          300.1300711 MHz
WDW         EM
SSB         0
LB          0.15 Hz
GB          0
PC          1.00
  
```



GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **t39**



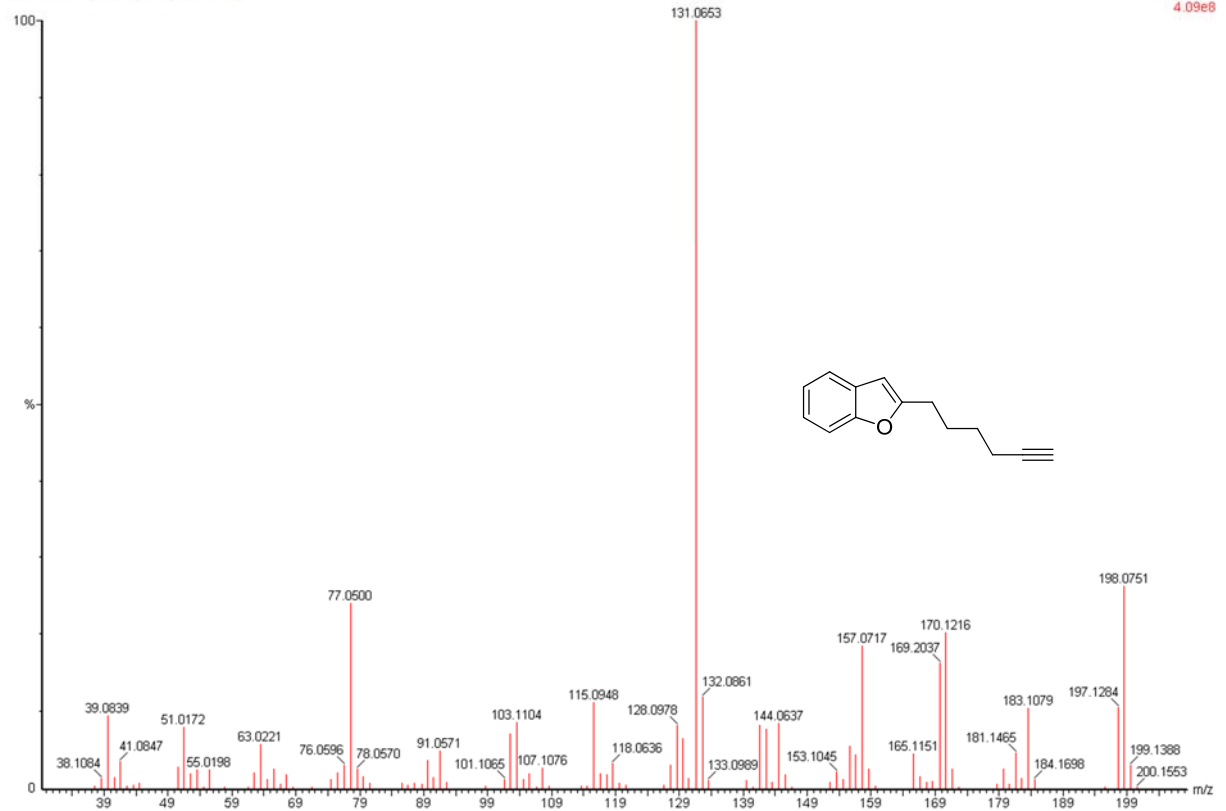
924  
fc-924- Sm (Mn, 1x3)

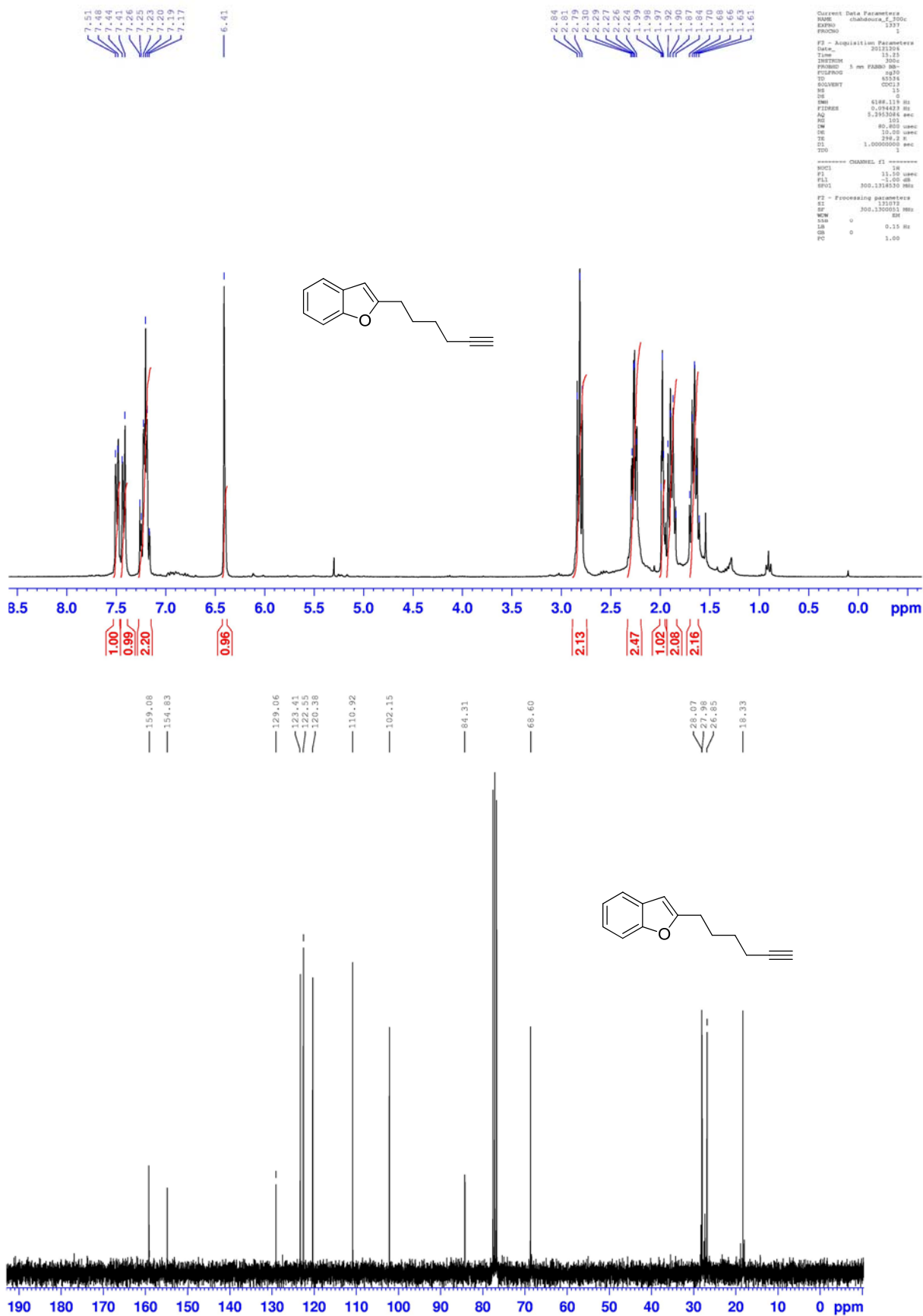
06-Dec-2012 + 14:19:34  
Scan EI+  
TIC  
2.42e9



924  
fc-924- 1414 (11.071) Cm (1412:1420)

06-Dec-2012 + 14:19:34  
Scan EI+  
4.09e8





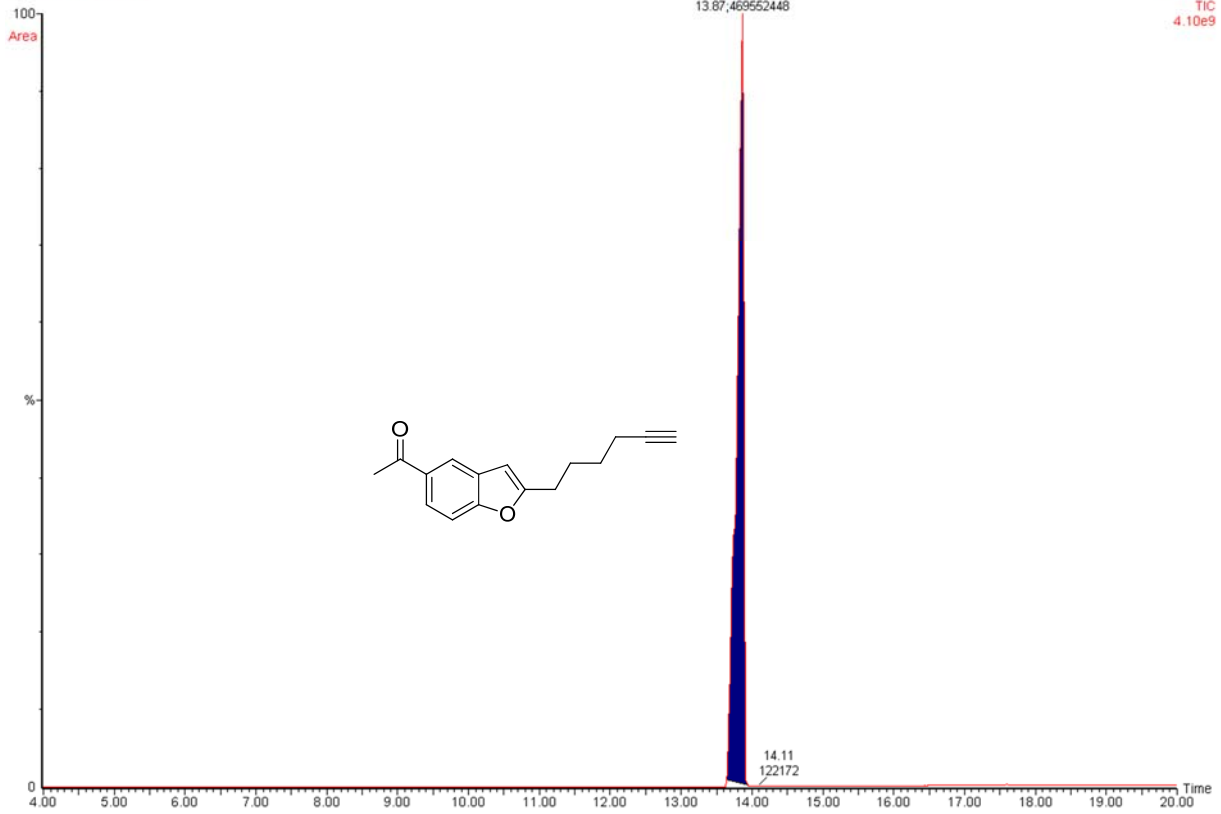
GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **s40**

1009

fc-1009-2 Sm (Mn, 1x3)

, 26-Jan-2013 + 13:50:14

Scan EI+  
TIC  
4.10e9

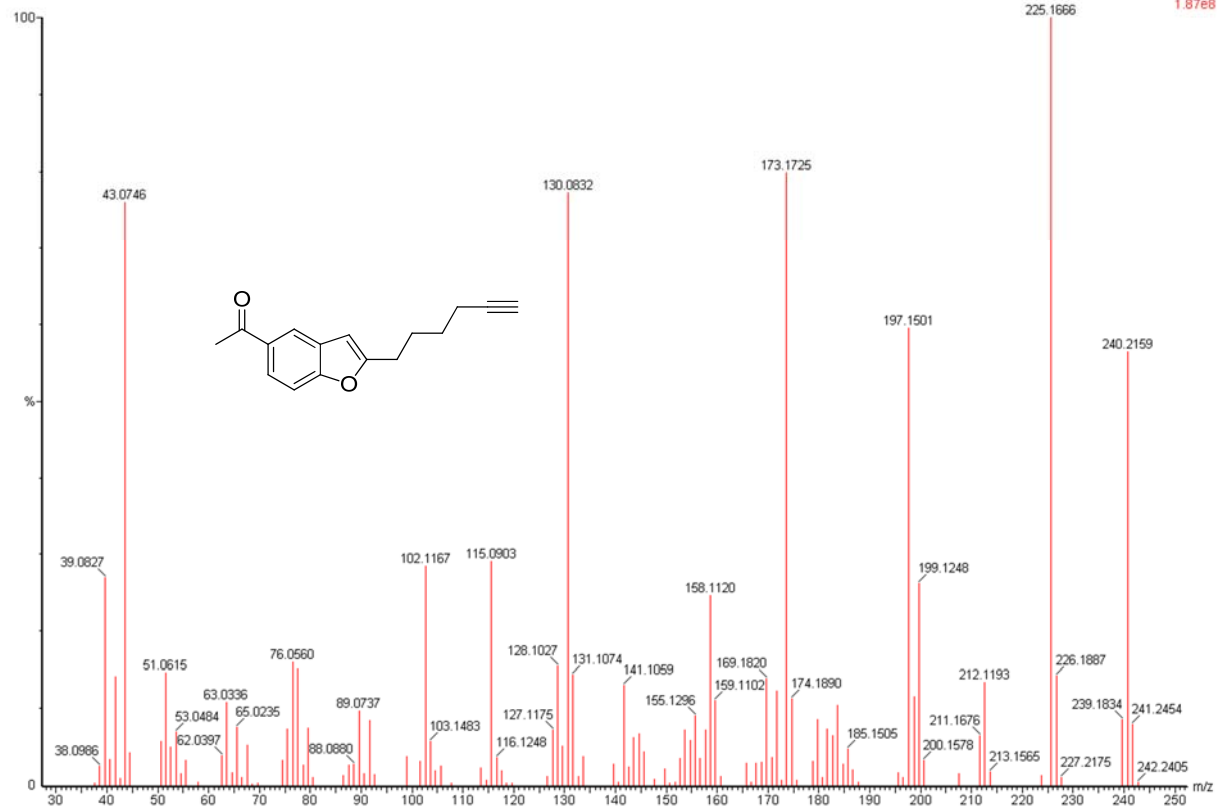


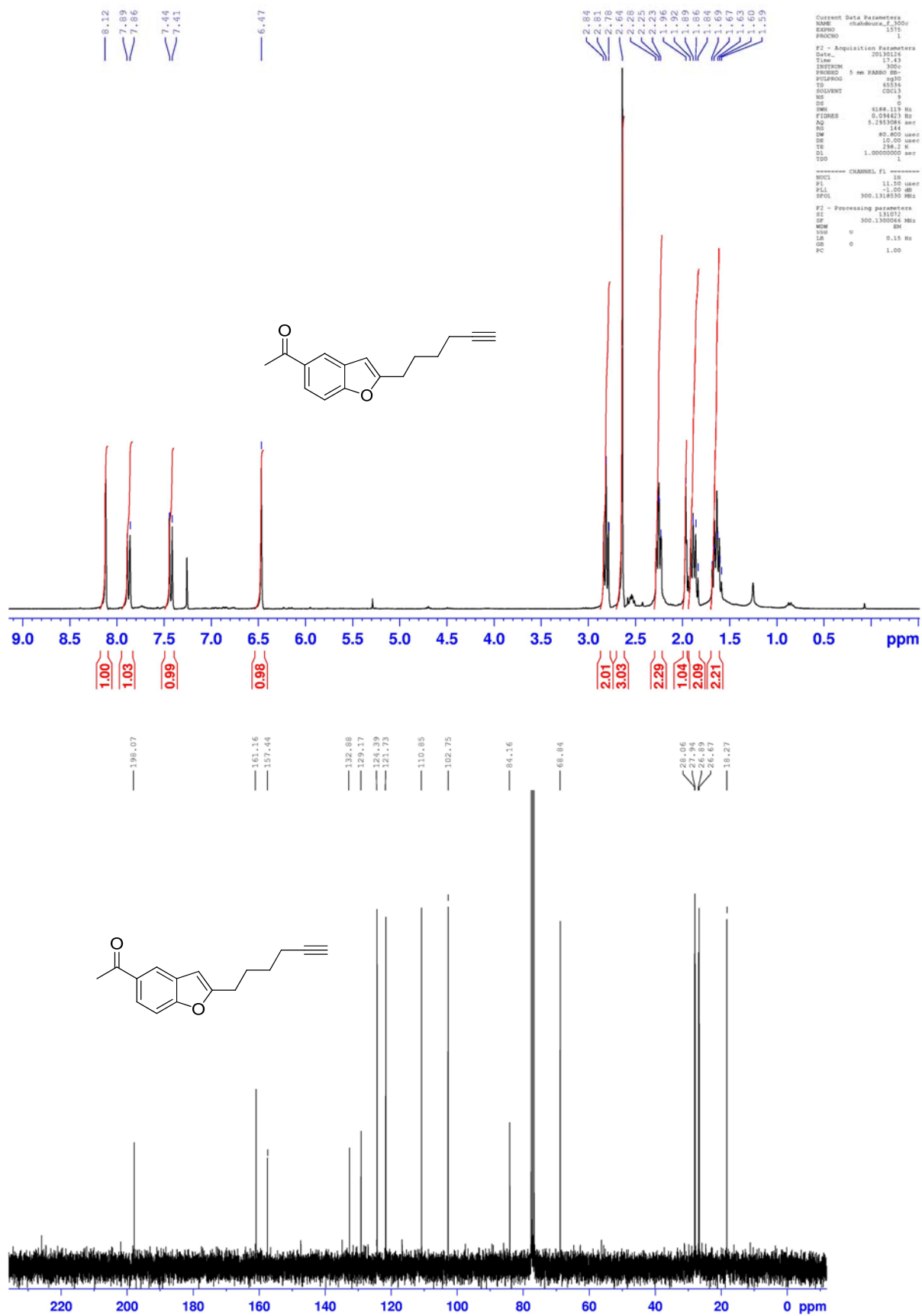
1009

fc-1009-2 1971 (13.857) Cm (1941:1971)

, 26-Jan-2013 + 13:50:14

Scan EI+  
1.87e8

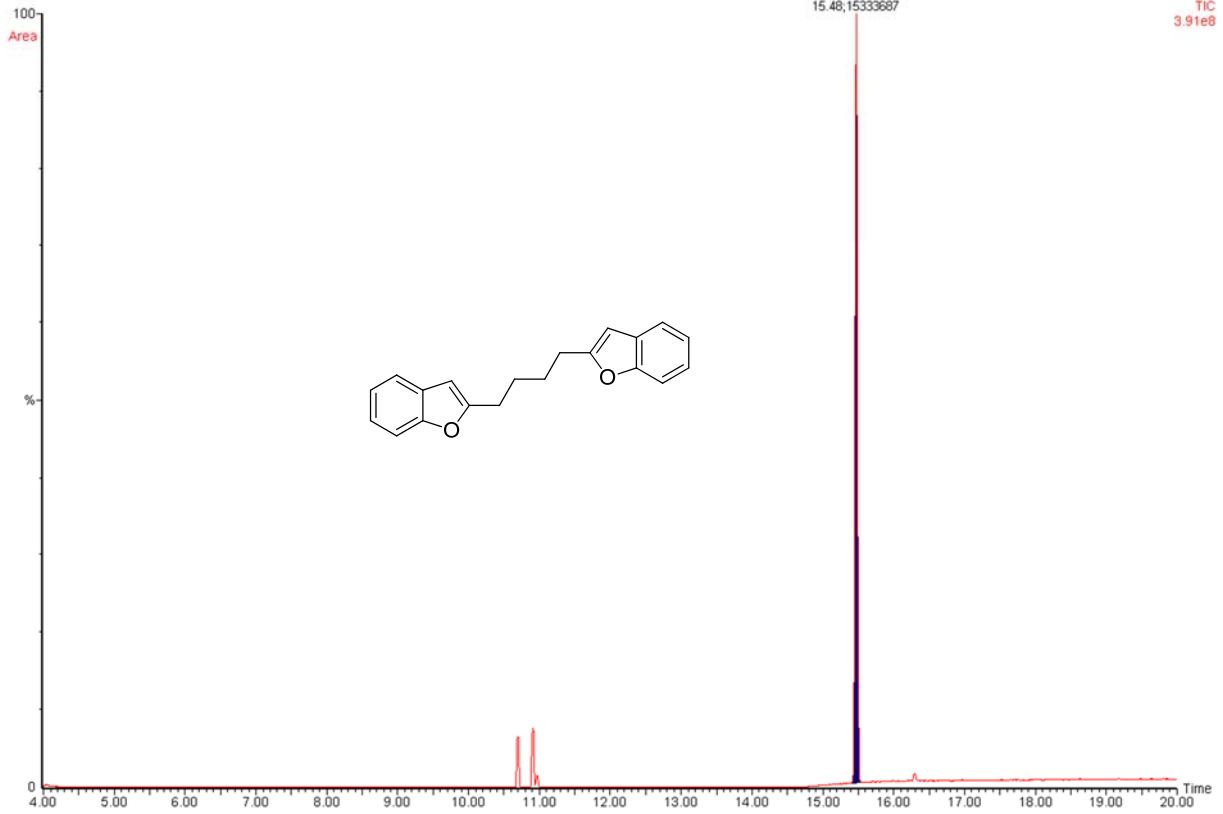




GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **t40**

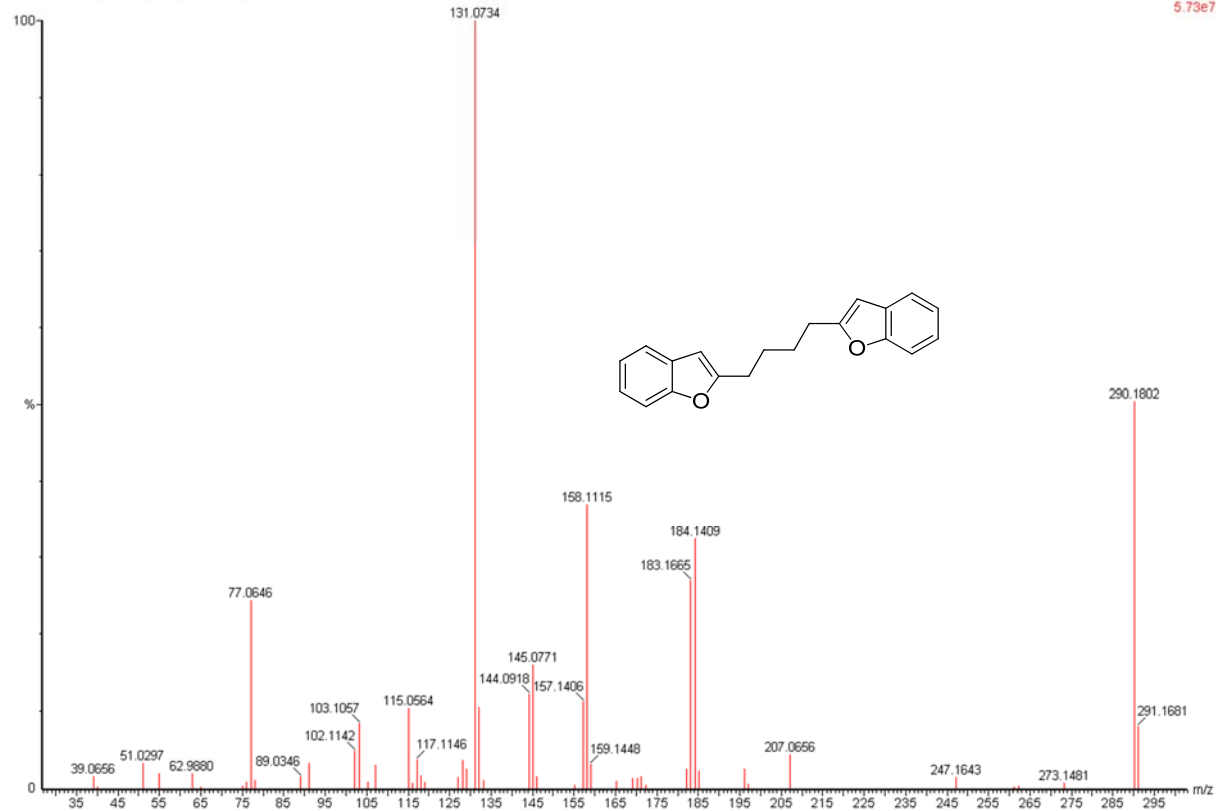
712  
fc-712-3 Sm (Mn, 1x3)

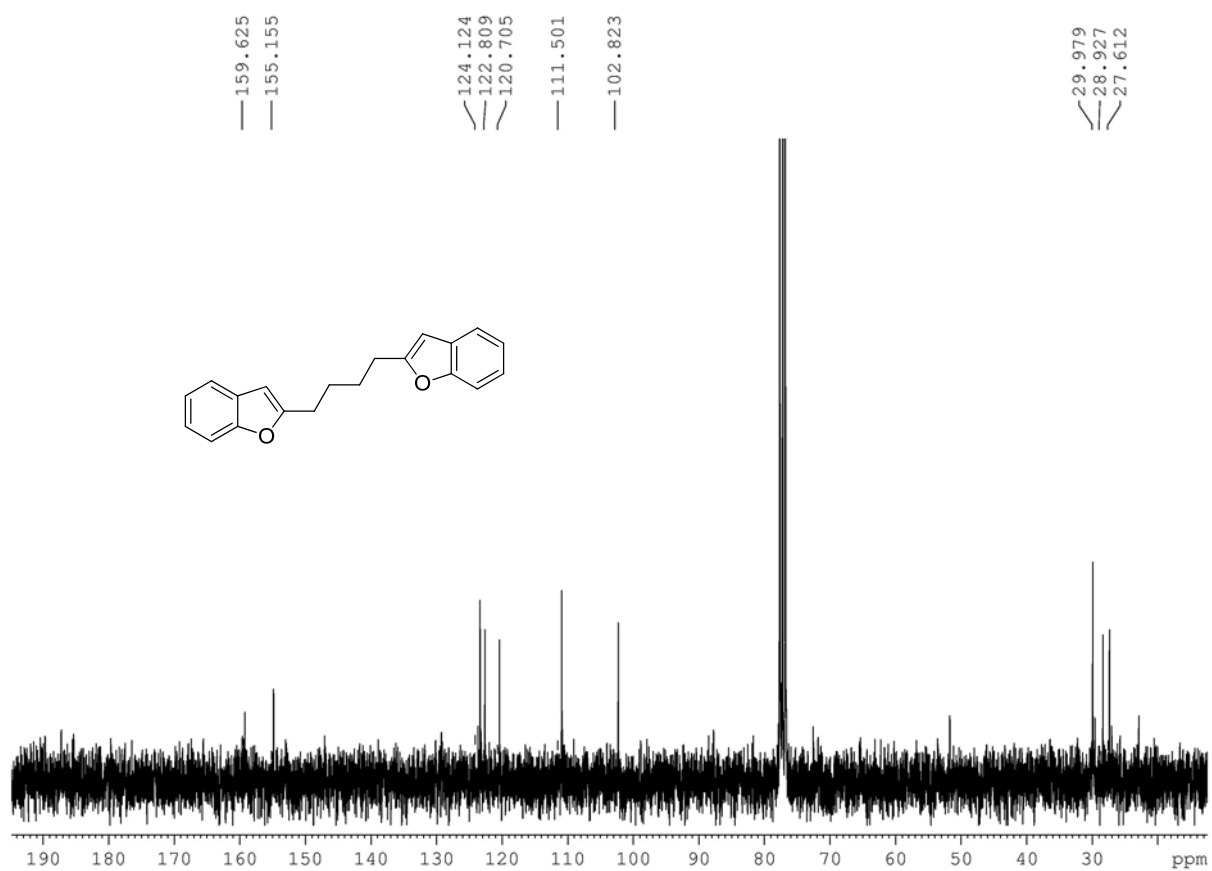
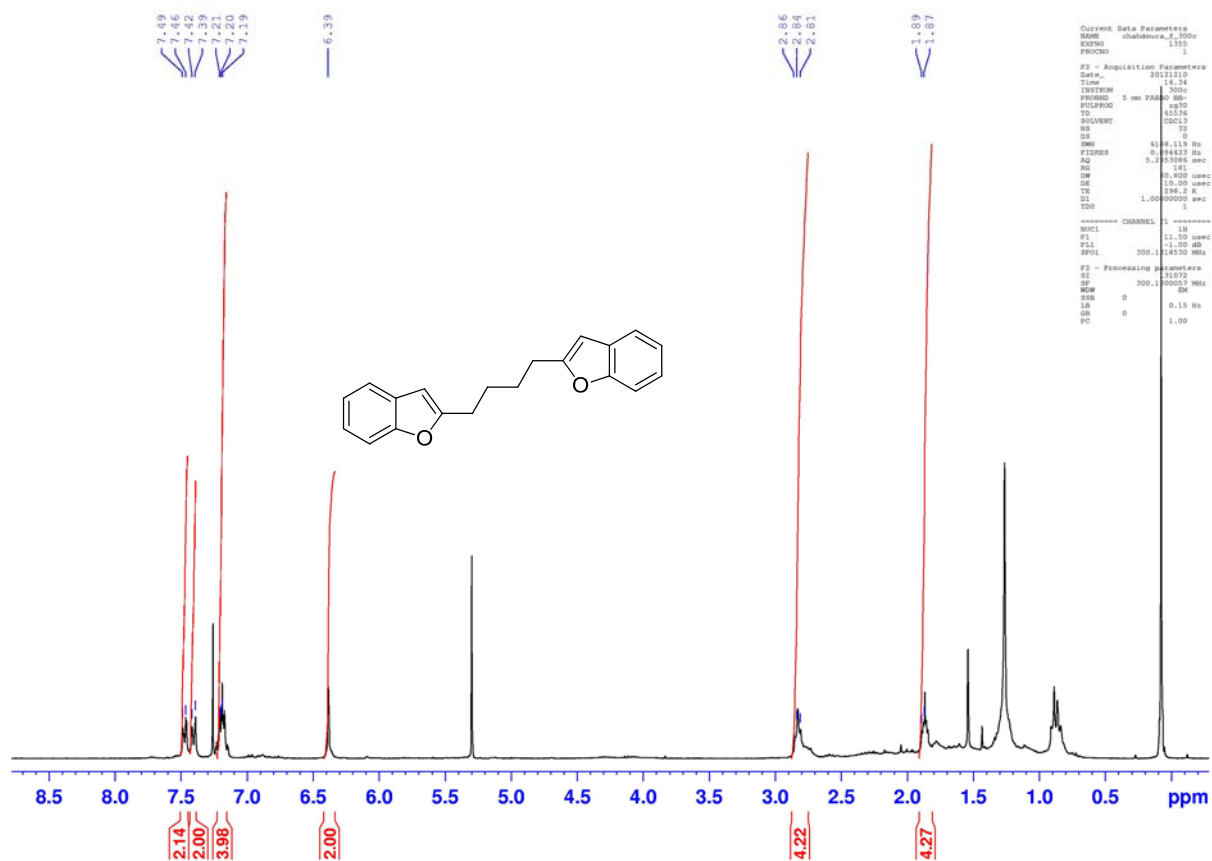
, 21-May-2012 + 18:14:21  
Scan EI+  
TIC  
3.91e8



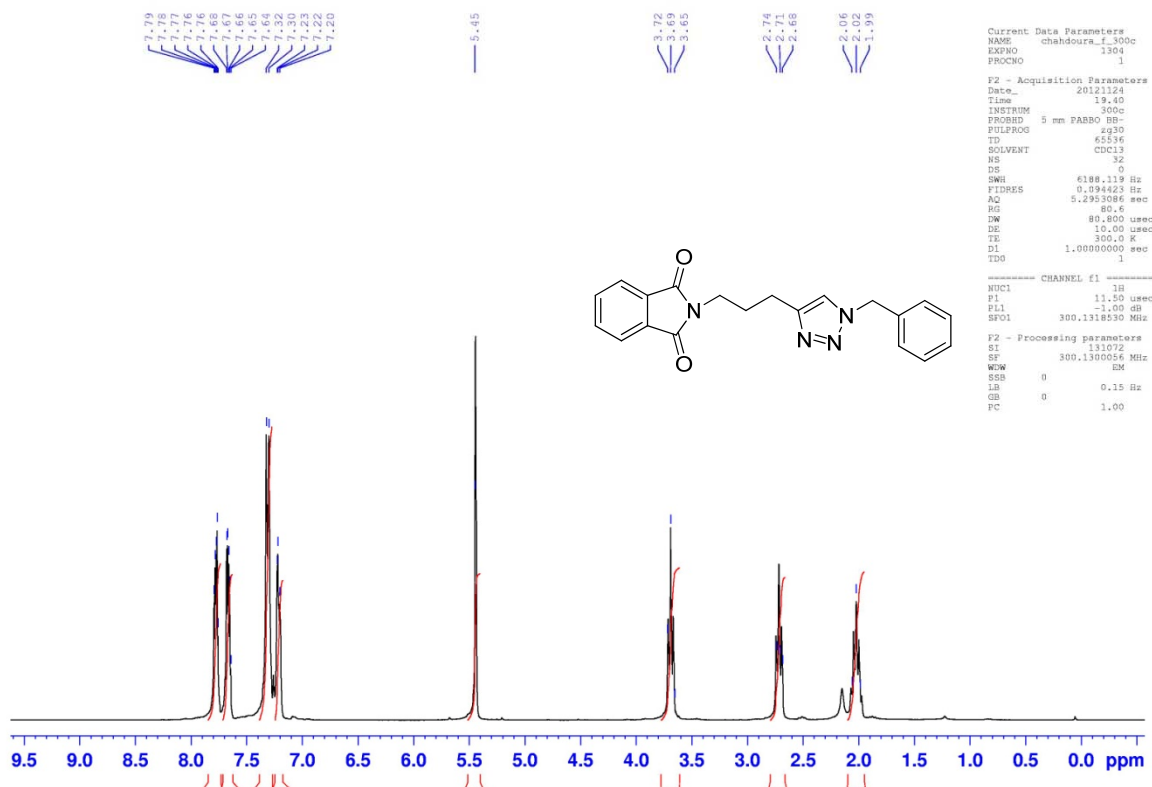
712  
fc-712-3 2295 (15.478) Cm (2289:2301)

, 21-May-2012 + 18:14:21  
Scan EI+  
5.73e7





GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **s40s**

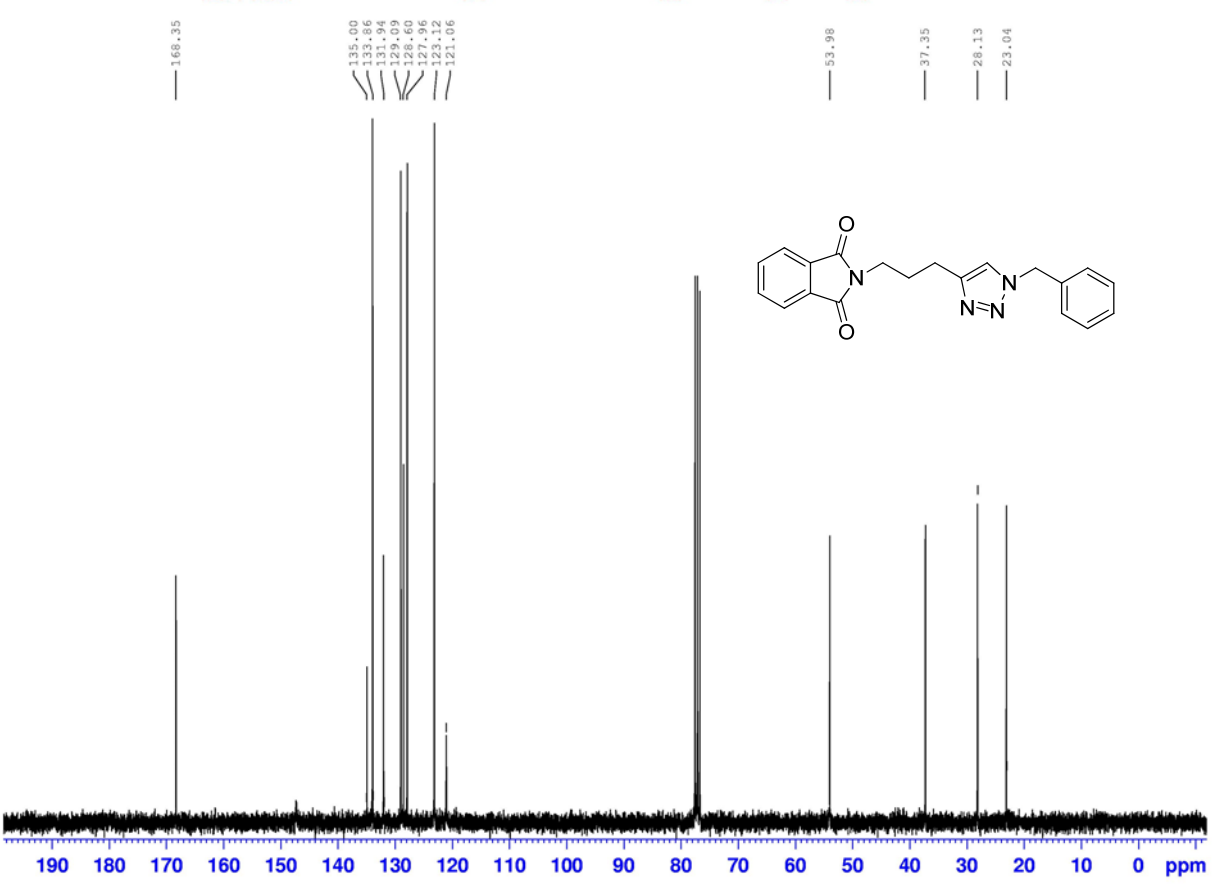


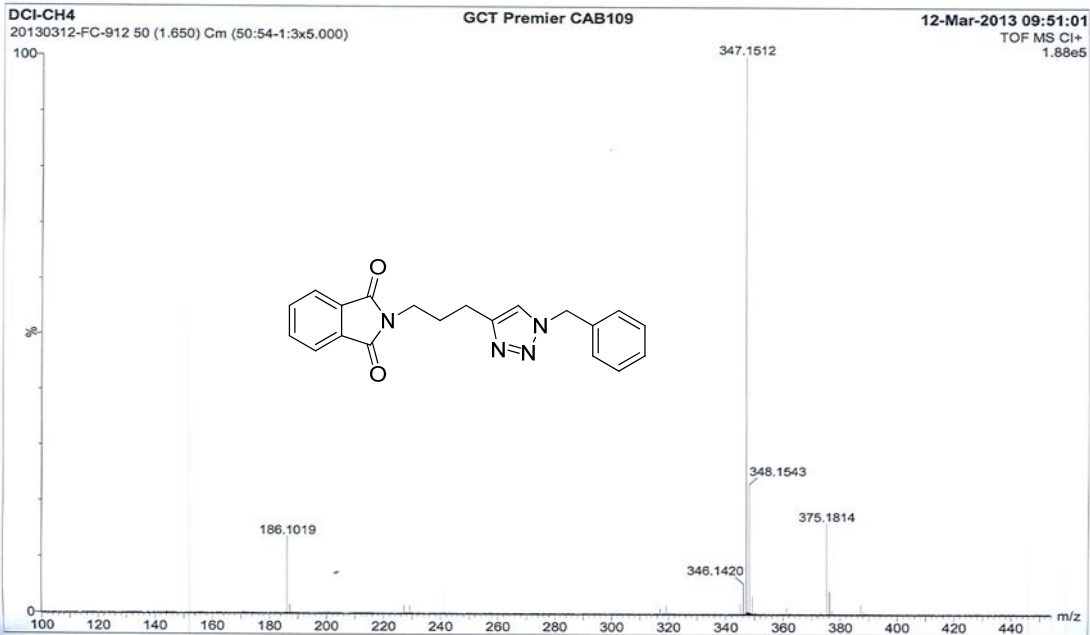
Current Data Parameters  
 NAME chahdoura\_f\_300c  
 EXPNO 1304  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20121124  
 Time 19.40  
 INSTRUM 300c  
 PROBED 5 mm PABBO BB-  
 PULPROG zg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 32  
 DS 0  
 SWH 6188.119 Hz  
 FIDRES 0.094423 Hz  
 AQ 5.2953086 sec  
 RG 80.6  
 DW 80.800 usec  
 DE 10.00 usec  
 TE 300.0 K  
 D1 1.0000000 sec  
 TDO 1

CHANNEL f1  
 NUCl 1H  
 P1 11.50 usec  
 PL1 -1.00 dB  
 SFO1 300.1318030 MHz

F2 - Processing parameters  
 SI 131072  
 SF 300.1300056 MHz  
 WDW EM  
 SSB 0  
 LB 0.15 Hz  
 GB 0  
 PC 1.00





### Elemental Composition Report

Page 1

#### Single Mass Analysis

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0  
Element prediction: Off

Monoisotopic Mass, Odd and Even Electron Ions  
453 formula(e) evaluated with 4 results within limits (all results (up to 1000) for each mass)  
Elements Used:

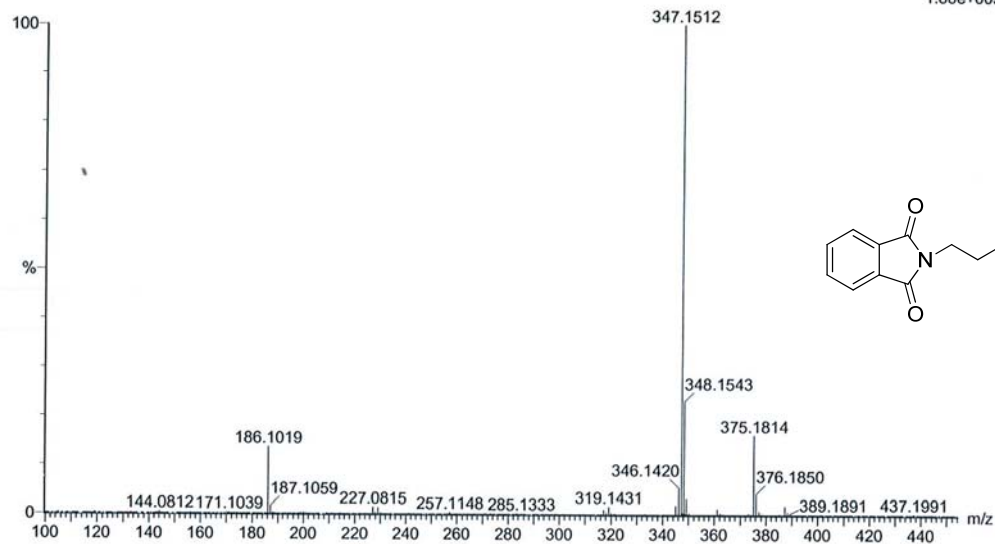
C: 0-100 H: 0-100 N: 0-10 O: 0-10

DCI-CH4

GCT Premier CAB109

12-Mar-2013 09:51:01  
TOF MS Cl+  
1.88e+005

20130312-FC-912 50 (1.650) Cm (50:54-1:3x5.000)

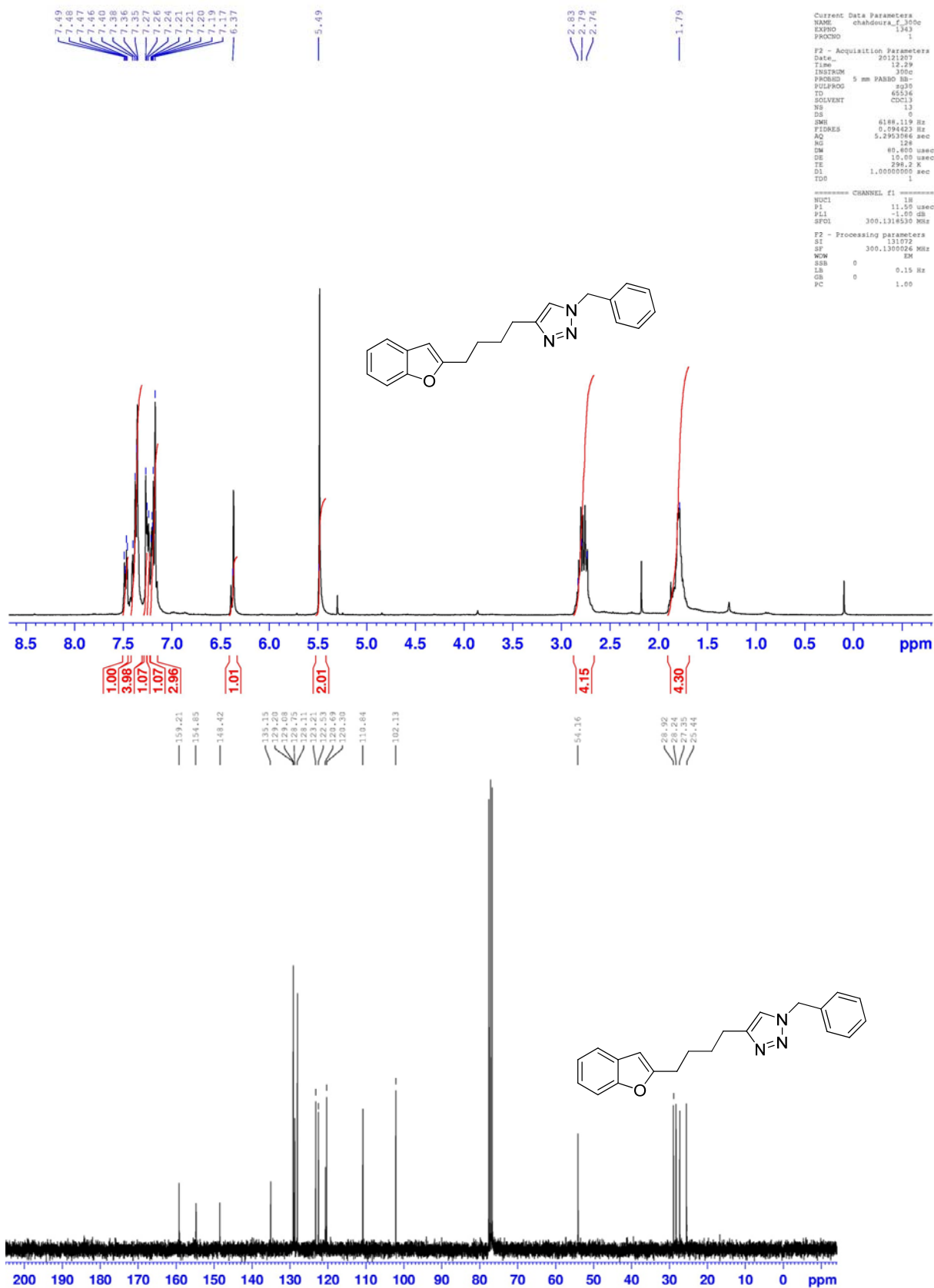


Minimum:  
Maximum:

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	Formula
347.1512	347.1508	0.4	1.2	13.5	32.4	C20 H19 N4 O2
	347.1521	-0.9	-2.6	13.0	215.9	C22 H21 N O3
	347.1527	-1.5	-4.3	0.5	4995.1	C8 H23 N6 O9
	347.1513	-0.1	-0.3	1.0	6289.8	C6 H21 N9 O8

<sup>1</sup>H NMR (300 MHz) (top) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (middle) spectra in CDCl<sub>3</sub>, low and high resolution DCI (CH<sub>4</sub>) spectre and for **s17u**

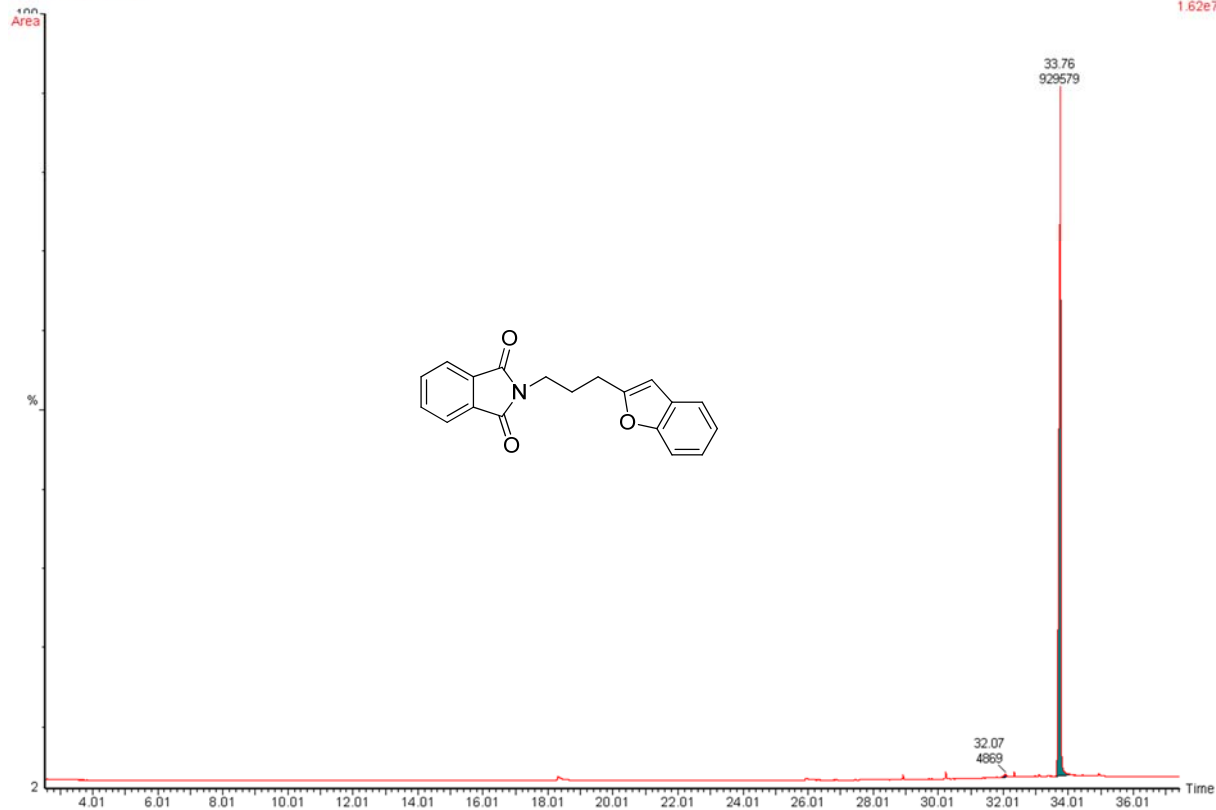




$^1\text{H}$  NMR (300 MHz) (top) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **s40u**

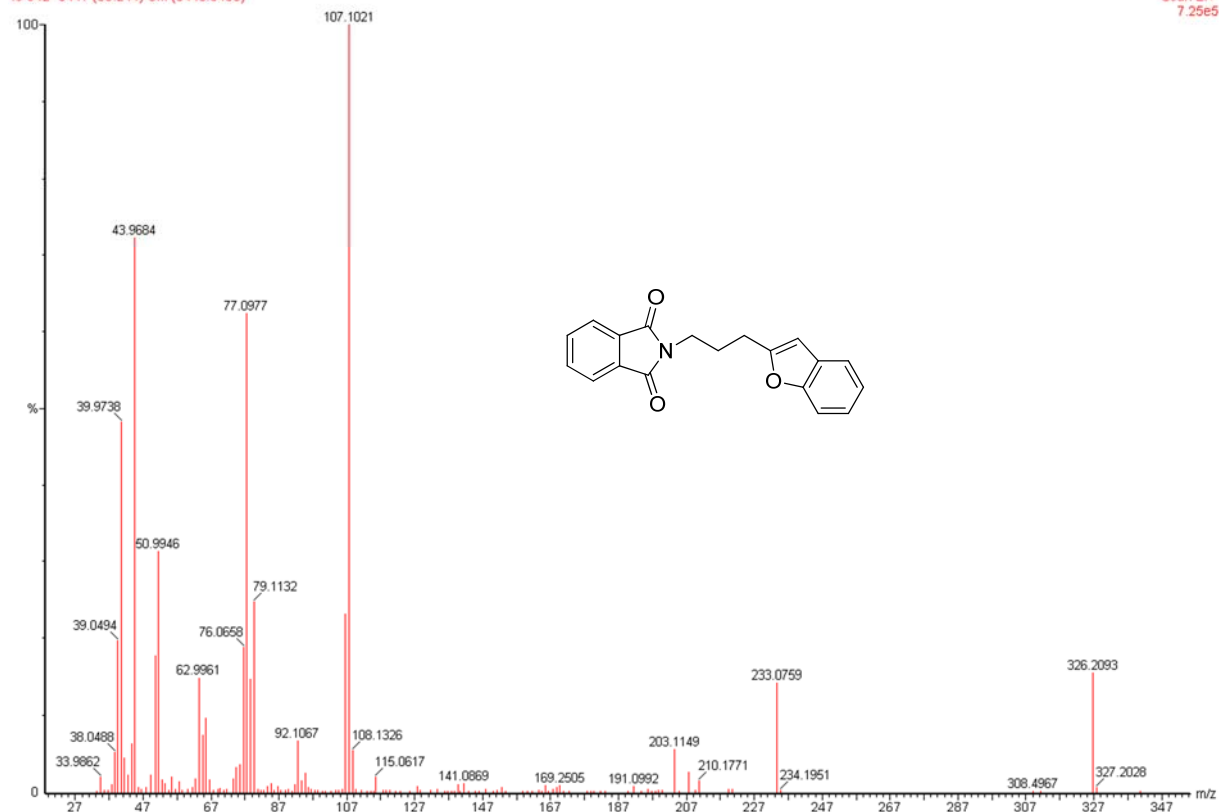
942  
fc-942- Sm (Mn, 1x3)

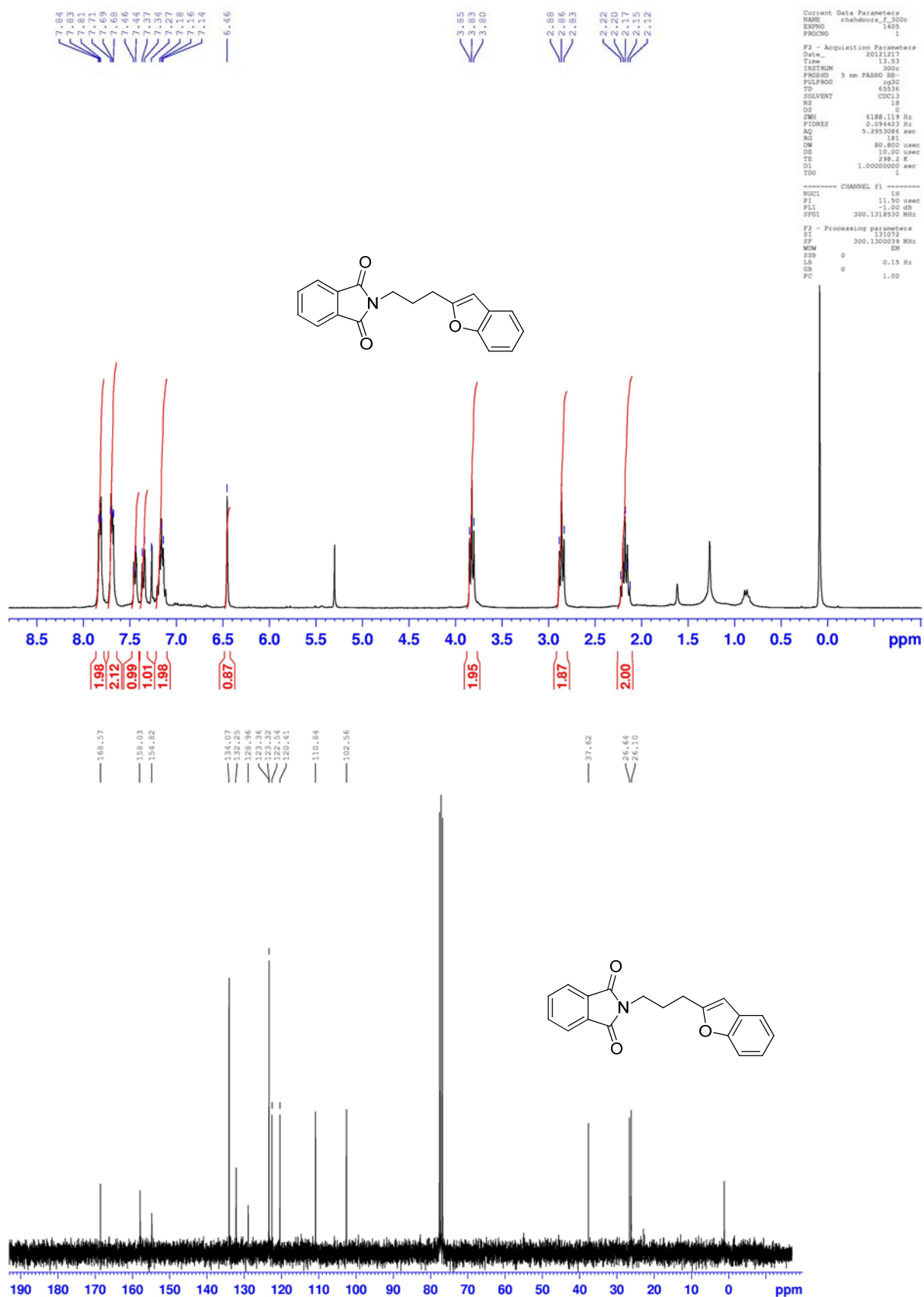
, 17-Dec-2012 + 14:35:09  
A: FID  
1.62e7



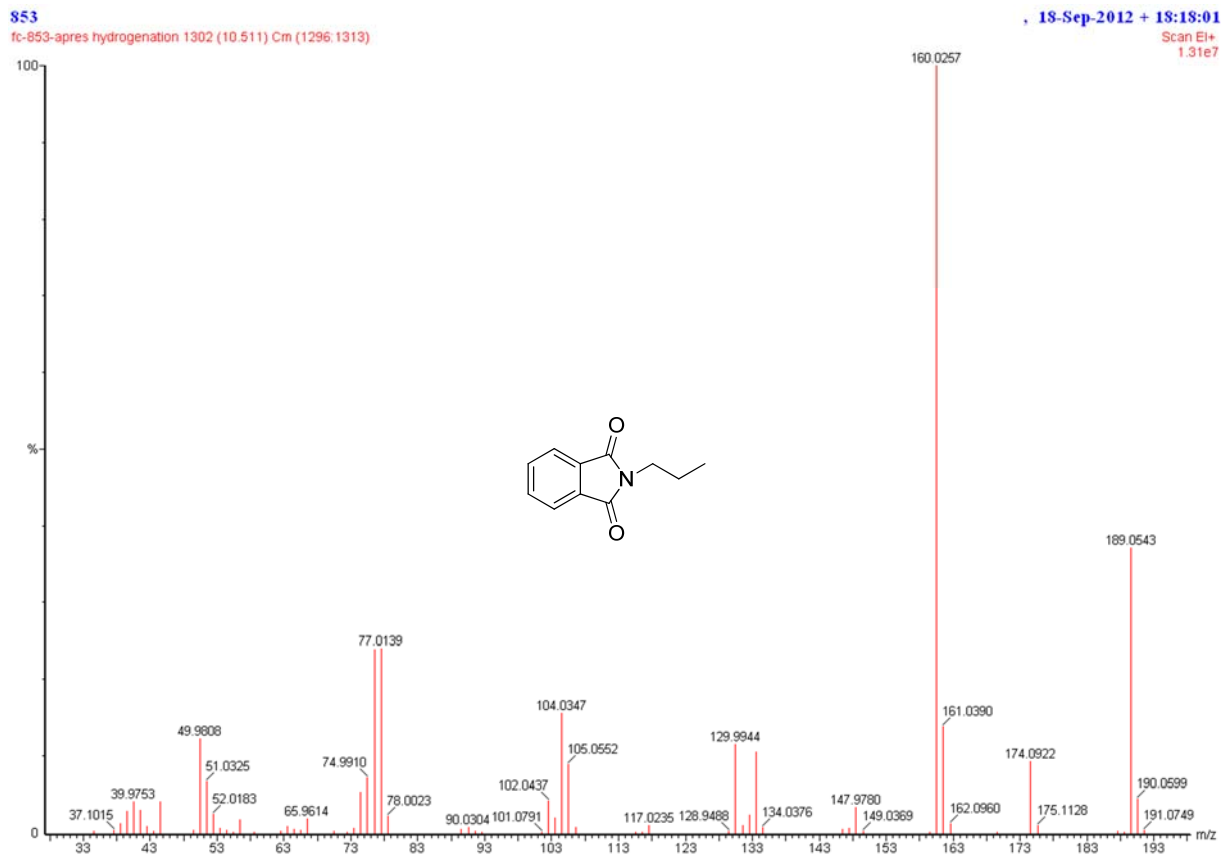
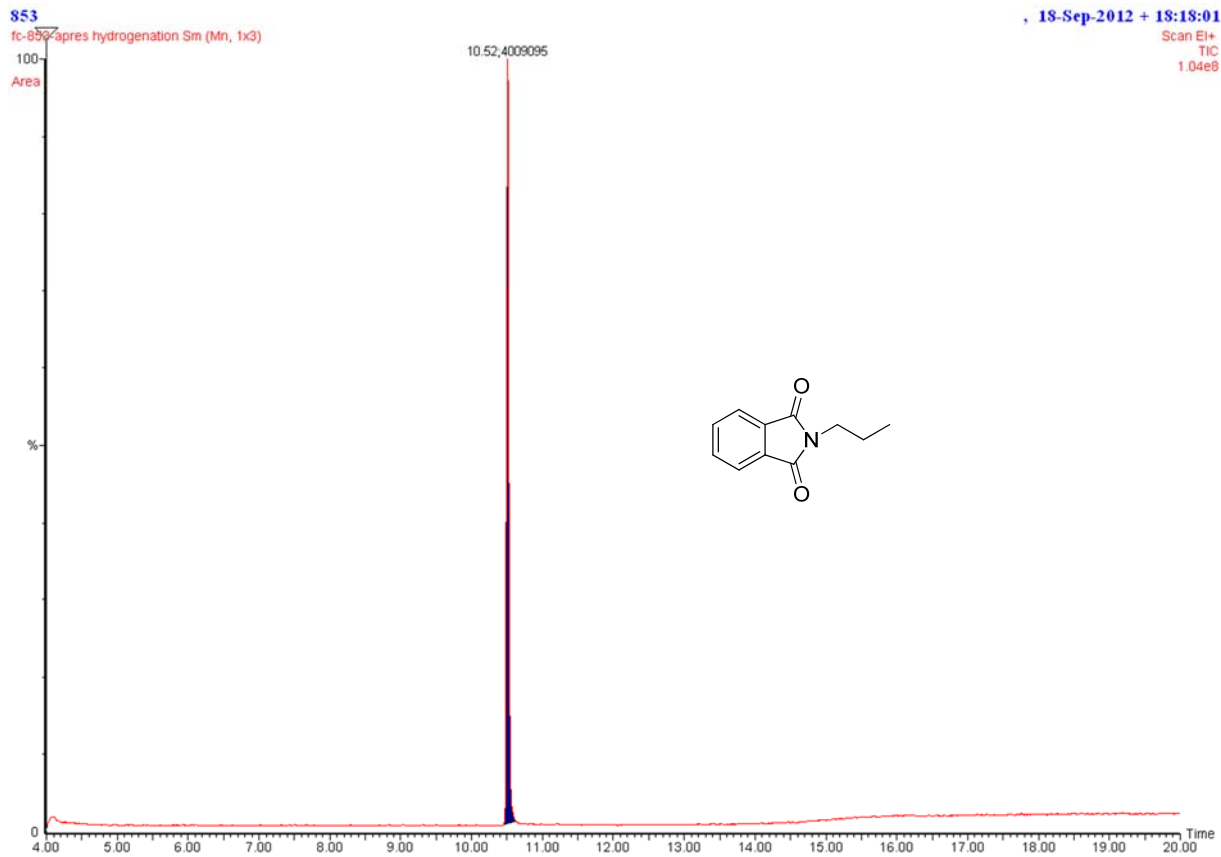
942  
fc-942- 5447 (30.244) Cm (5443:5453)

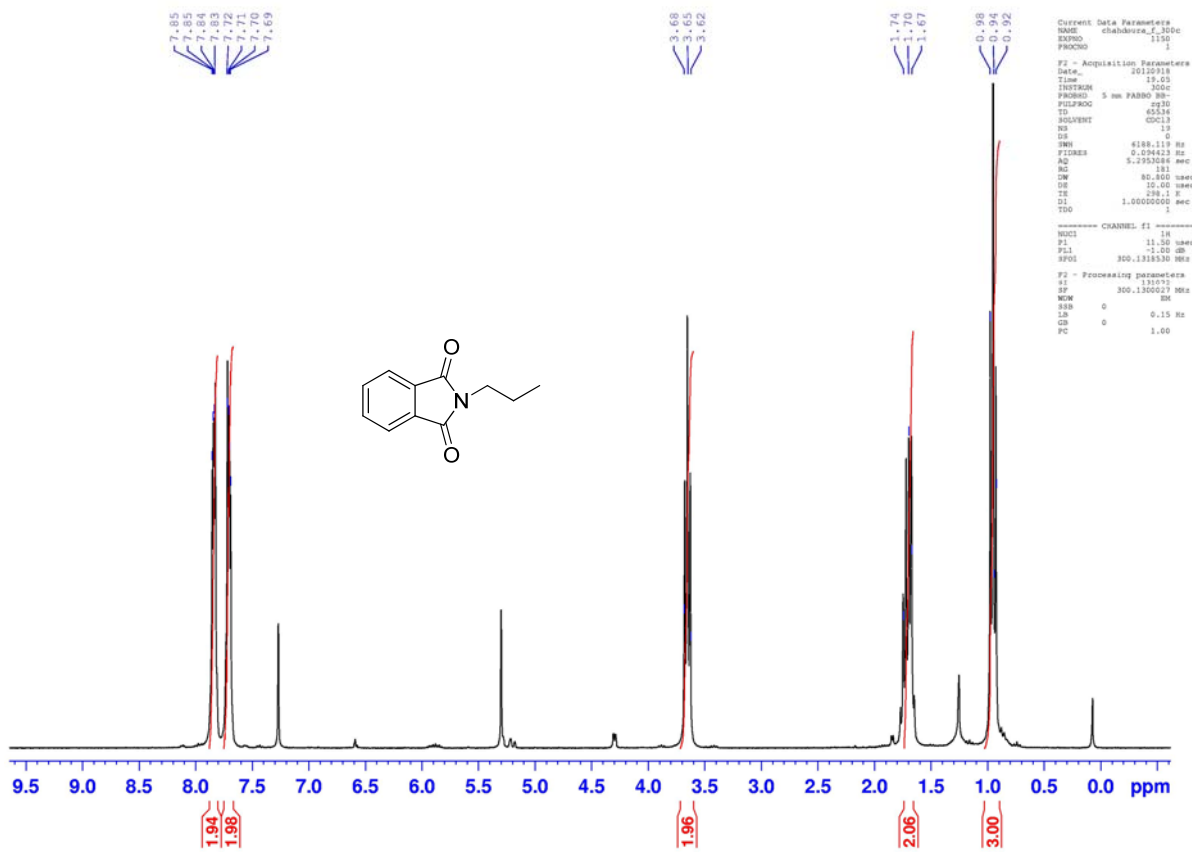
, 17-Dec-2012 + 14:35:09  
Scan EI+  
7.25e5





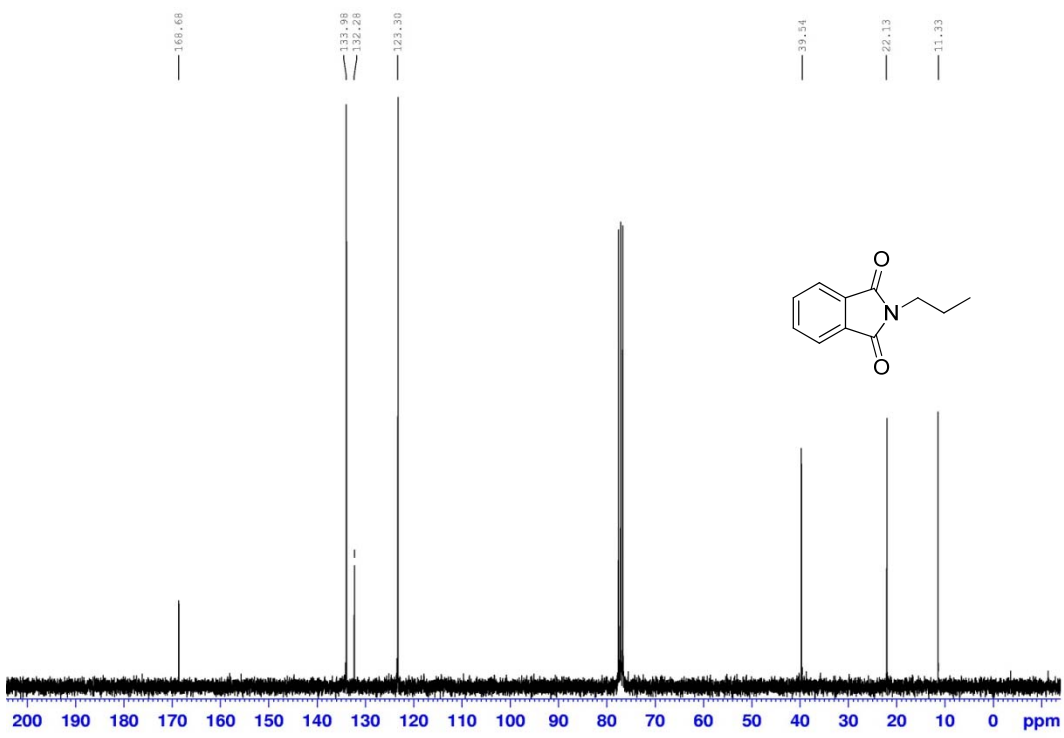
GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **a17s**





```

Current Data Parameters
NAME: 21abakura_1_30dc
EXPNO: 1350
PROCNO: 5
F2 - Acquisition Parameters
Date_: 20120318
Time: 12.03
INSTRUM: zgpg
PROBHD: 5 mm PABBO MM-
PULPROG: zg30
TD: 65536
SOLVENT: CDCl3
NS: 12
DS: 0
SWH: 6188.111 Hz
FIDRES: 0.094423 Hz
AQ: 5.295026 sec
RG: 181
DM: 80.000 usec
DE: 10.00 usec
TE: 29.2 K
D1: 1.0000000 sec
TDC:
===== CHANNEL f1 =====
NUC1: 1H
P1: 11.00 usec
PL1: -1.00 dB
RF01: 300.131530 MHz
F2 - Processing parameters
F1: 130.013
SF: 300.1300212 MHz
WDW: EM
SSB: 0
LB: 0.15 Hz
GB: 0
PC: 1.00
  
```



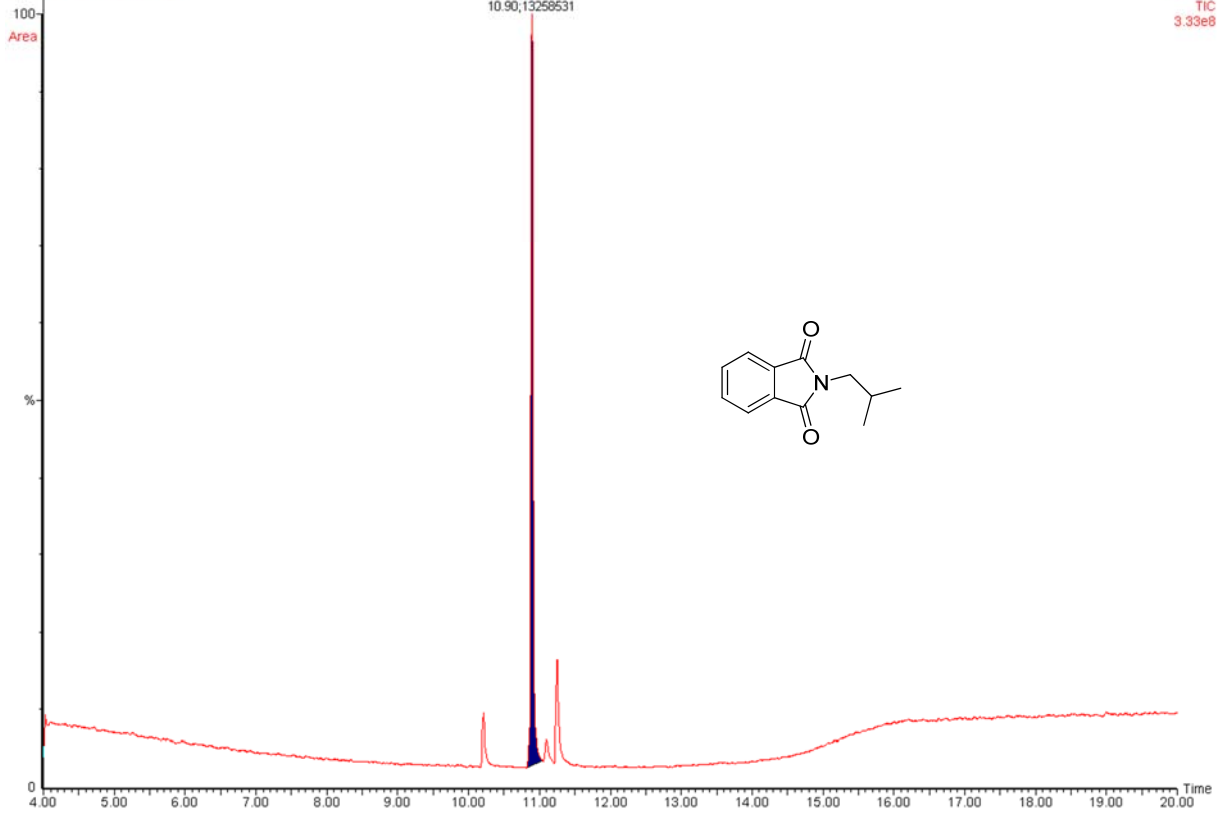
GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **a18H**

12

reduction-13- Sm (Mn, 1x3)

, 22-Nov-2012 + 19:44:23

Scan EI+  
TIC  
3.33e8

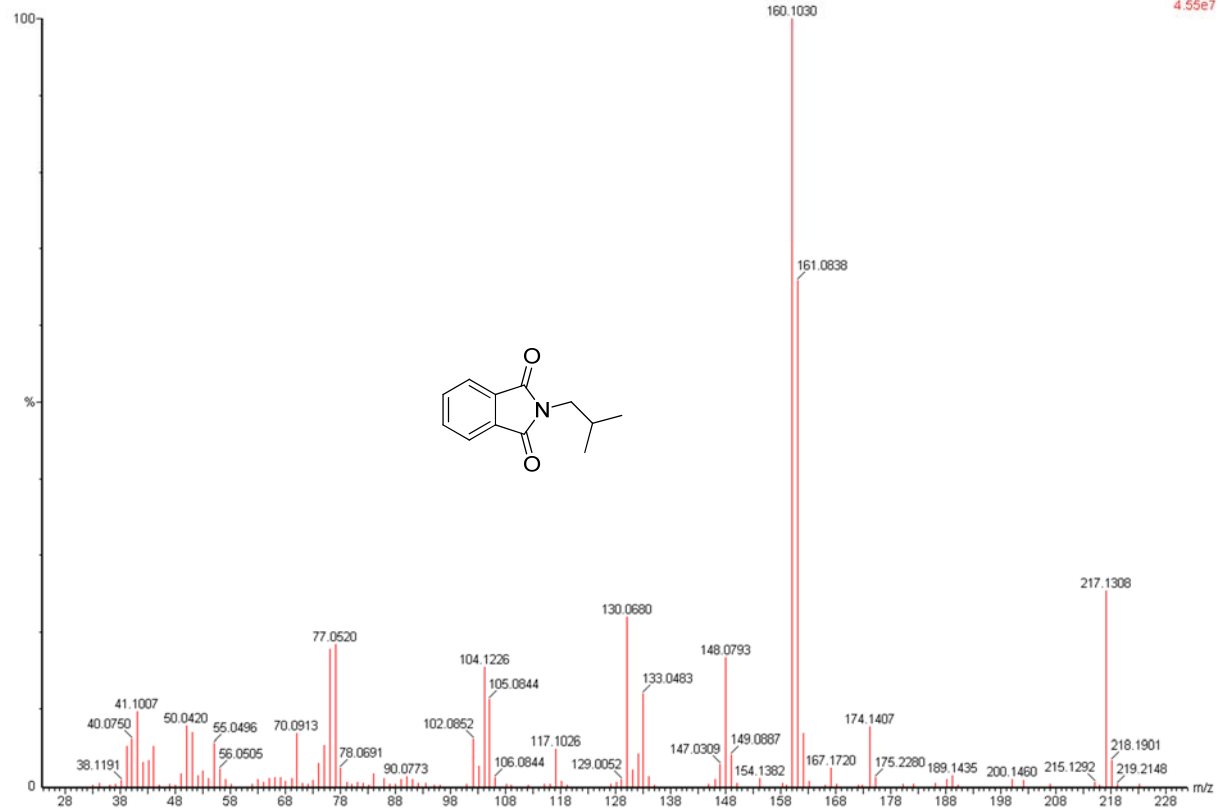


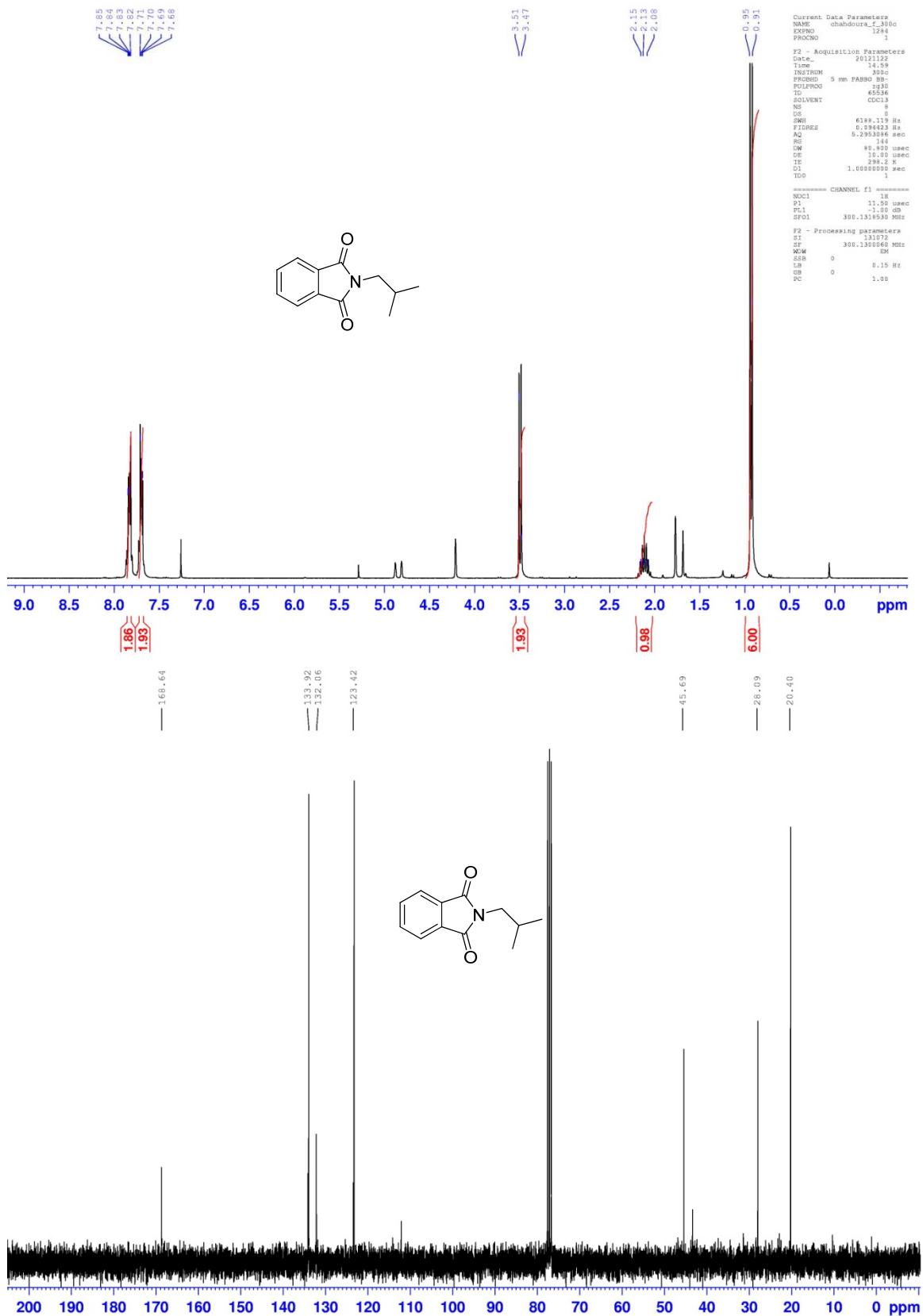
12

reduction-13- 1379 (10.896) Cm (1376:1387)

, 22-Nov-2012 + 19:44:23

Scan EI+  
4.55e7

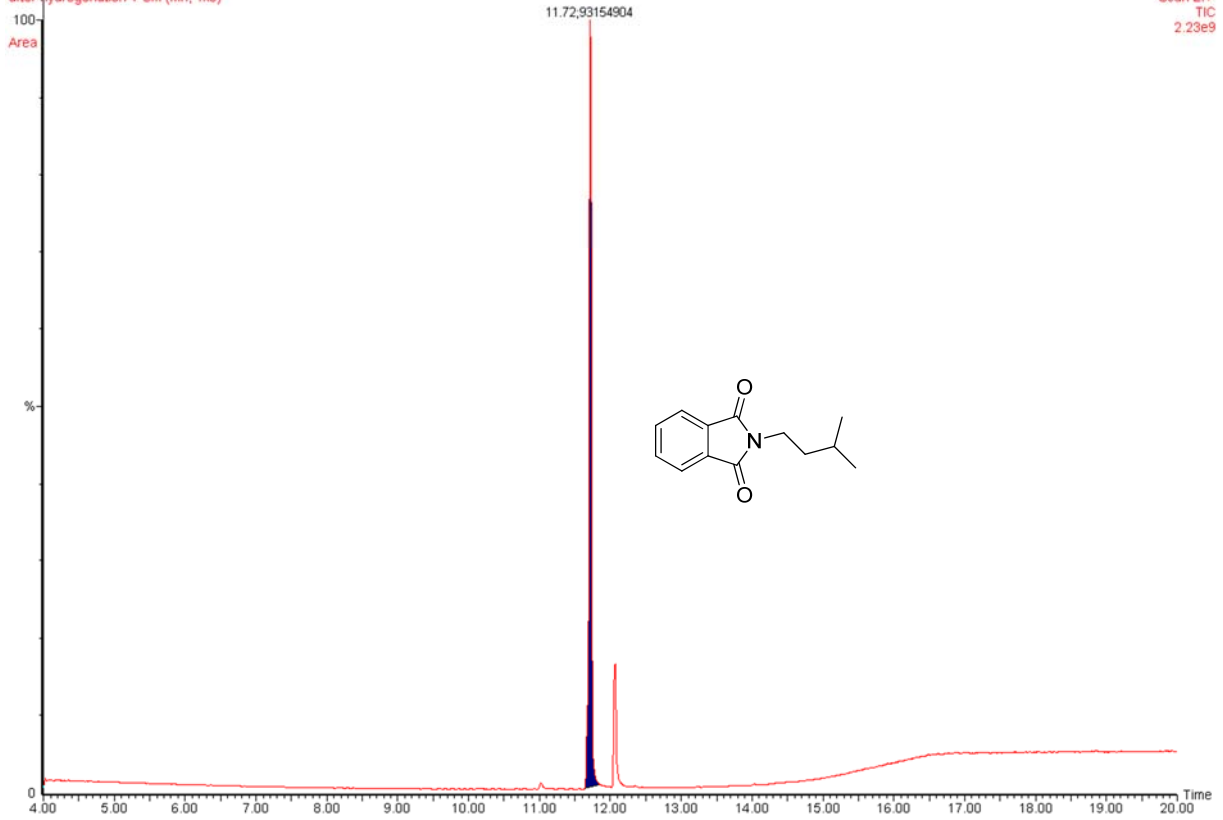




GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **a19H**

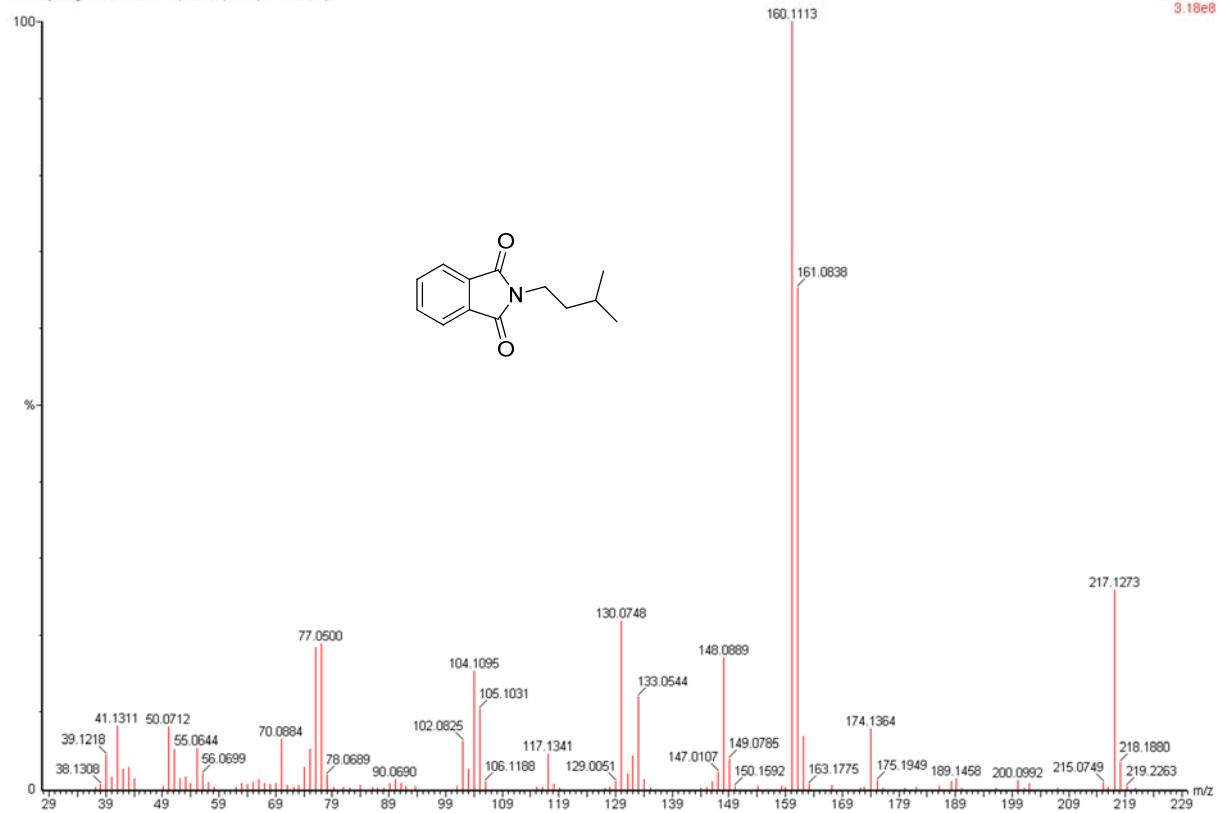
7  
after hydrogenation-7 Sm (Mn, 1x3)

, 23-Nov-2012 + 19:00:33

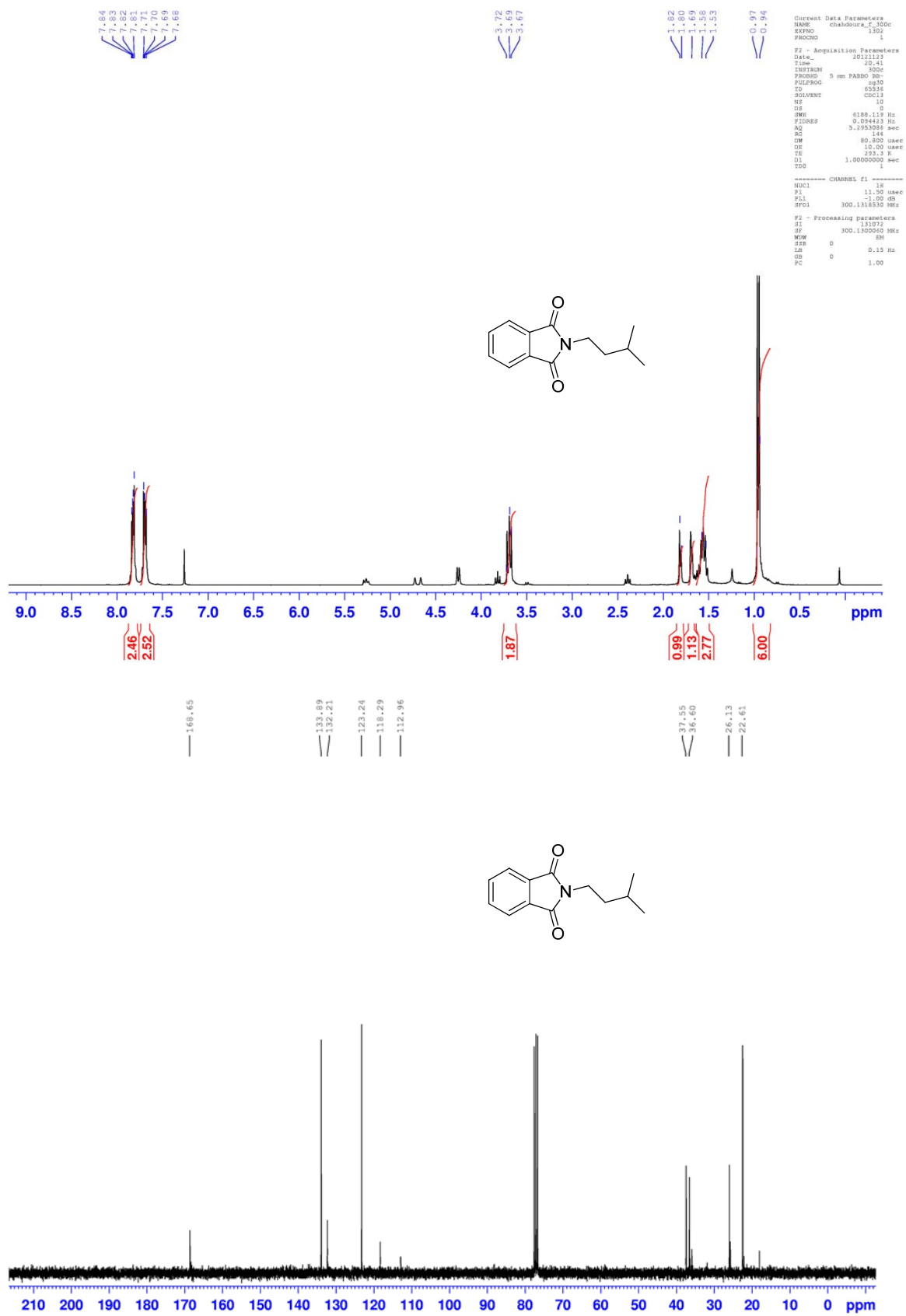


7  
after hydrogenation-7 1544 (11.722) Cm (1539:1550)

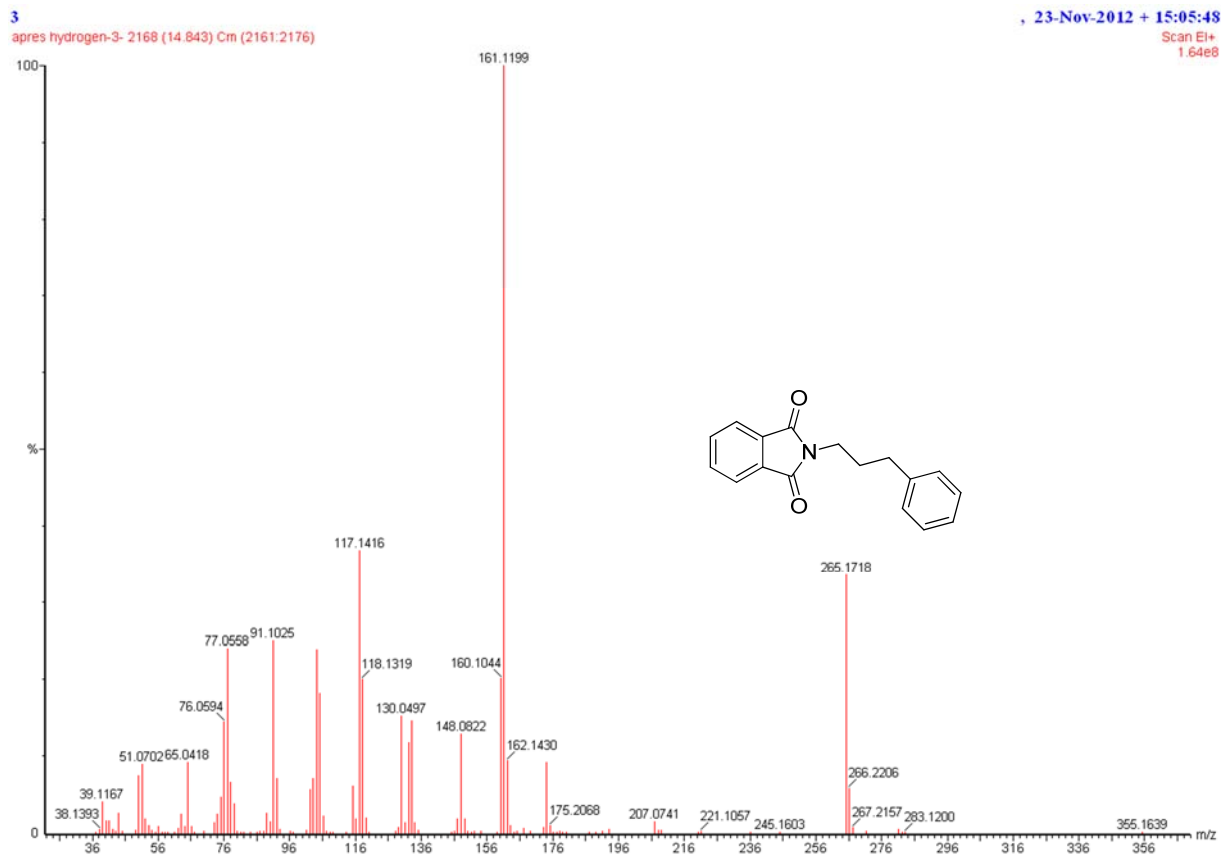
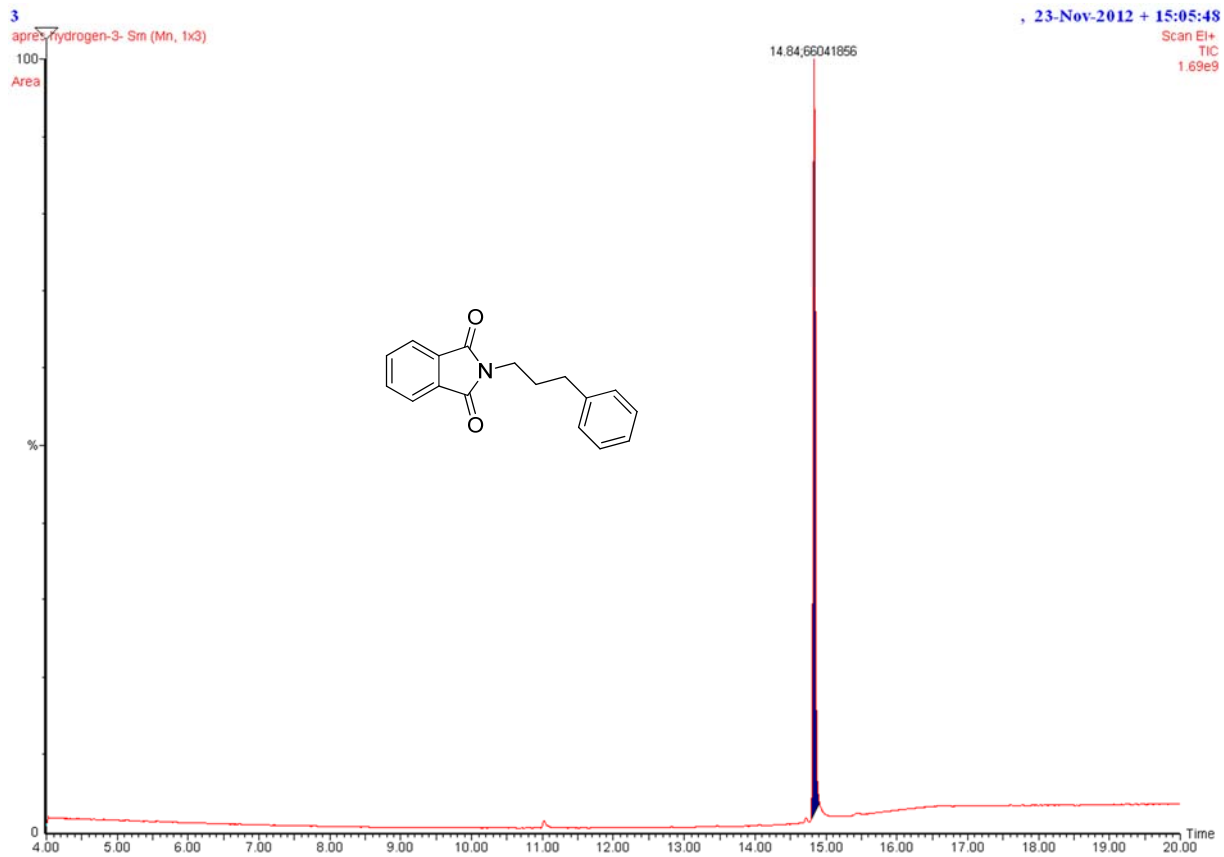
, 23-Nov-2012 + 19:00:33

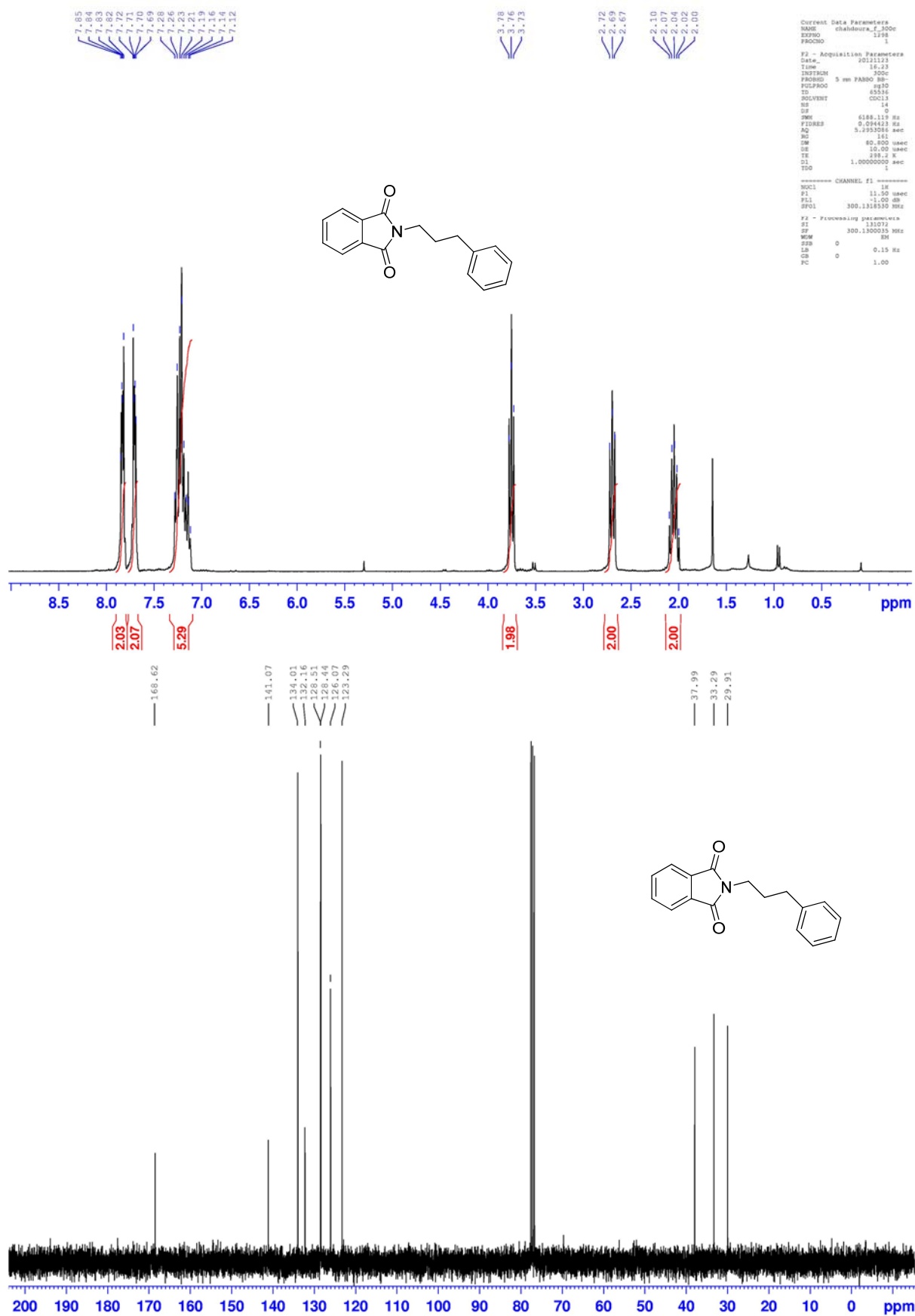






GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **a20H**





GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **a21H**

1062

fc-1062-apres hydrogenation Sm (Mn, 1x3)

, 05-Mar-2013 + 10:31:38

Scan EI+  
TIC  
1.47e9

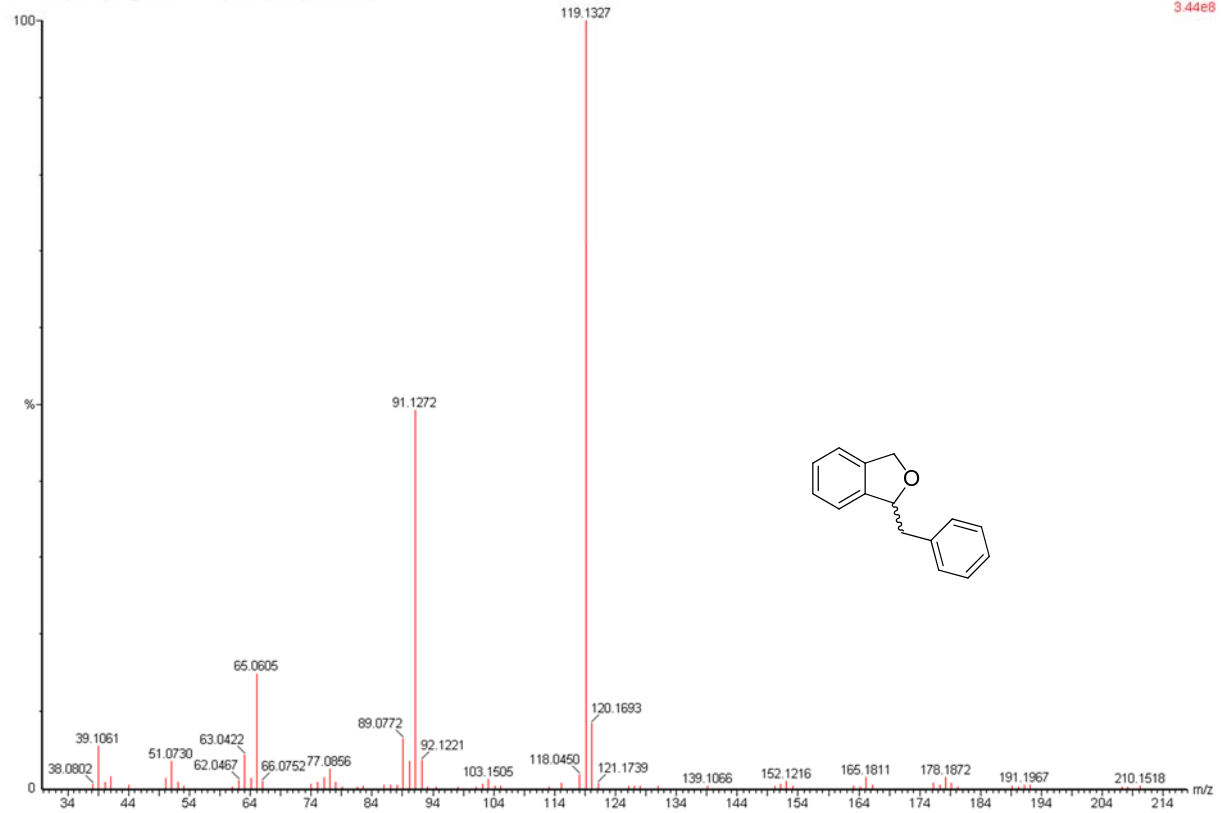


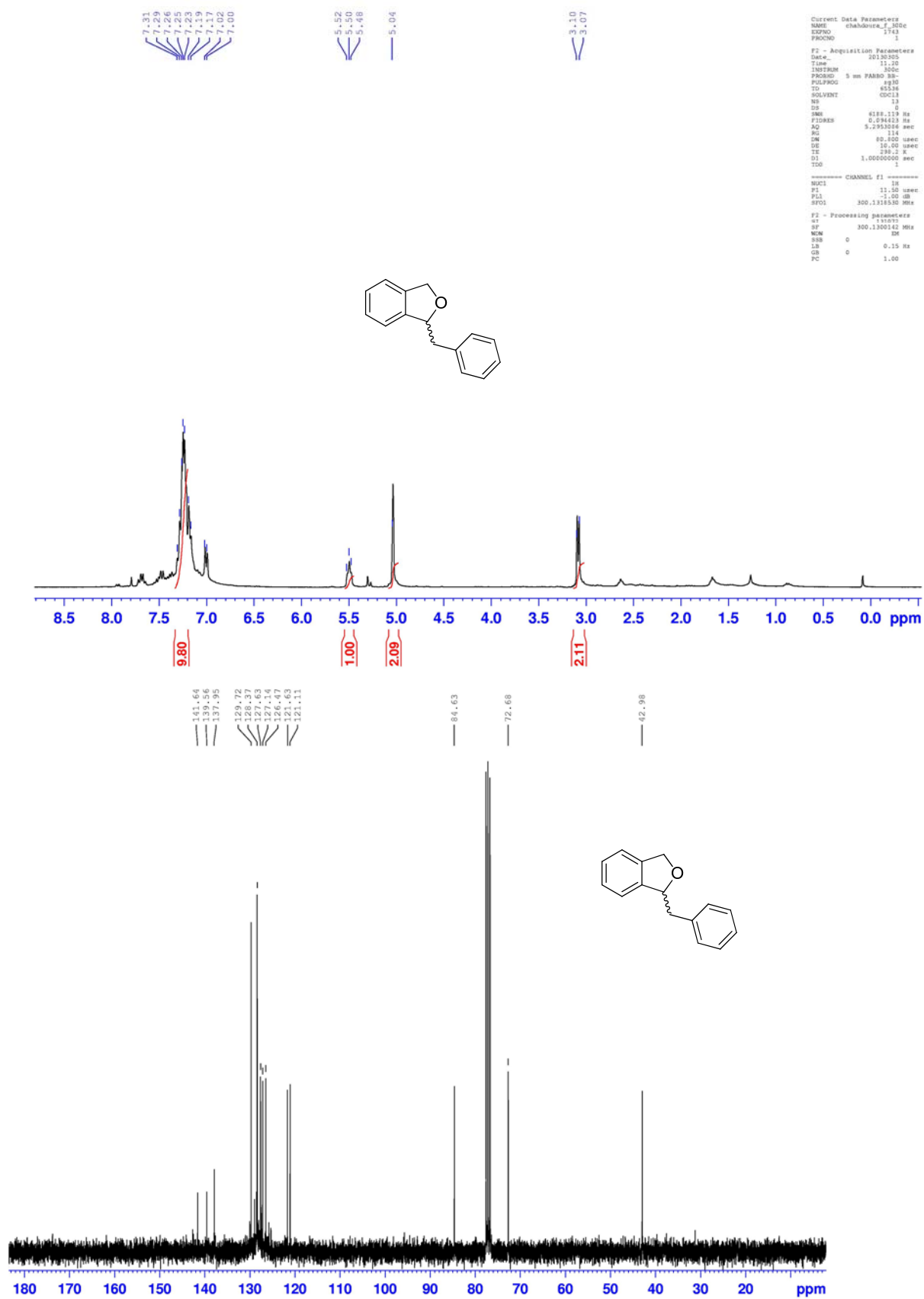
1062

fc-1062-apres hydrogenation 1584 (11.922) Cm (1576.1589)

, 05-Mar-2013 + 10:31:38

Scan EI+  
3.44e8





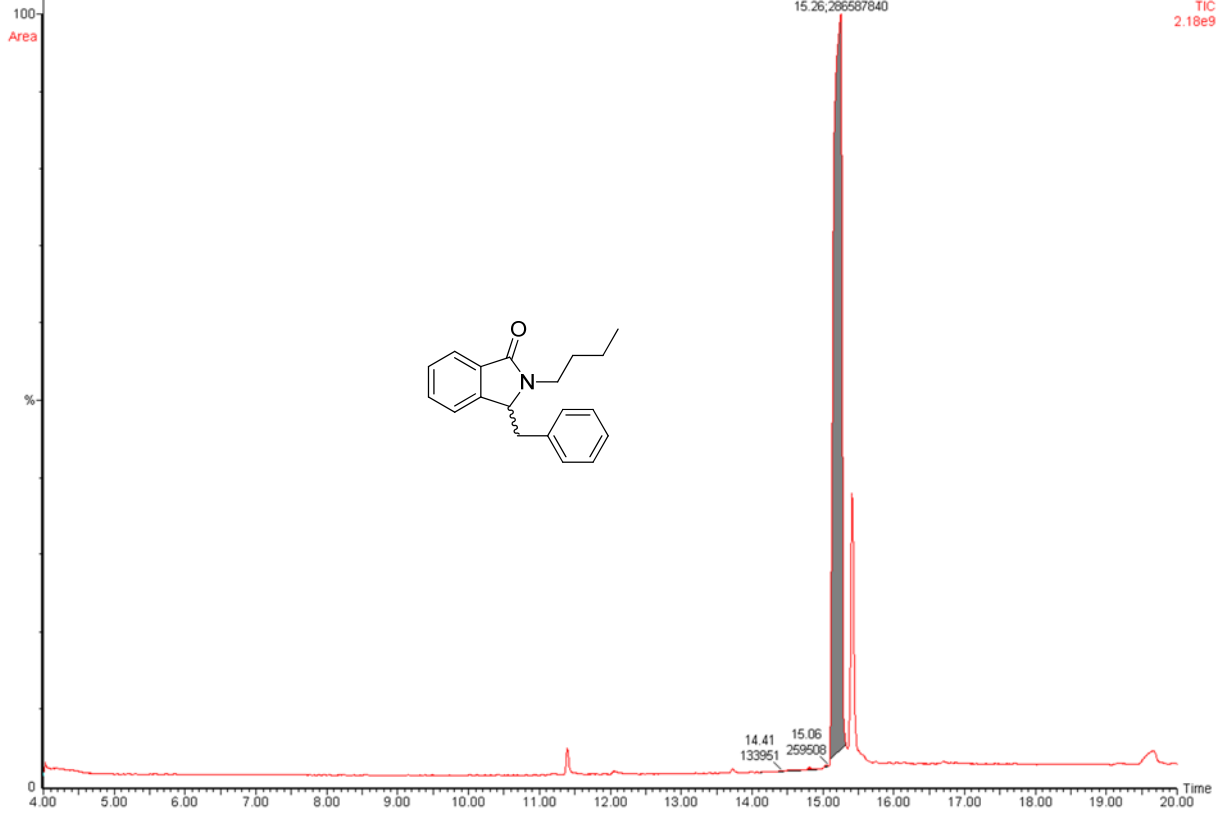
GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **o33H**

1022

fc-1022-after hydrogen-2ation Sm (Mn, 1x3)

, 07-Feb-2013 + 16:57:02

Scan EI+  
TIC  
2.18e9

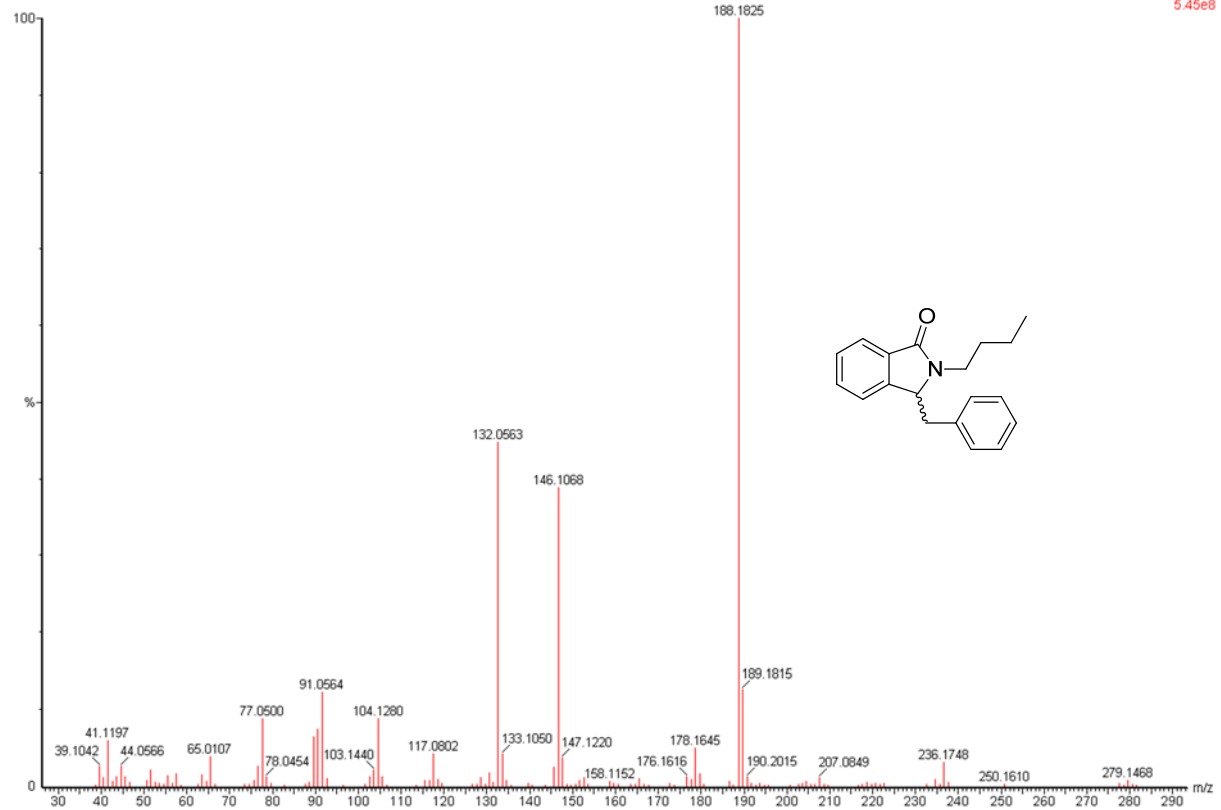


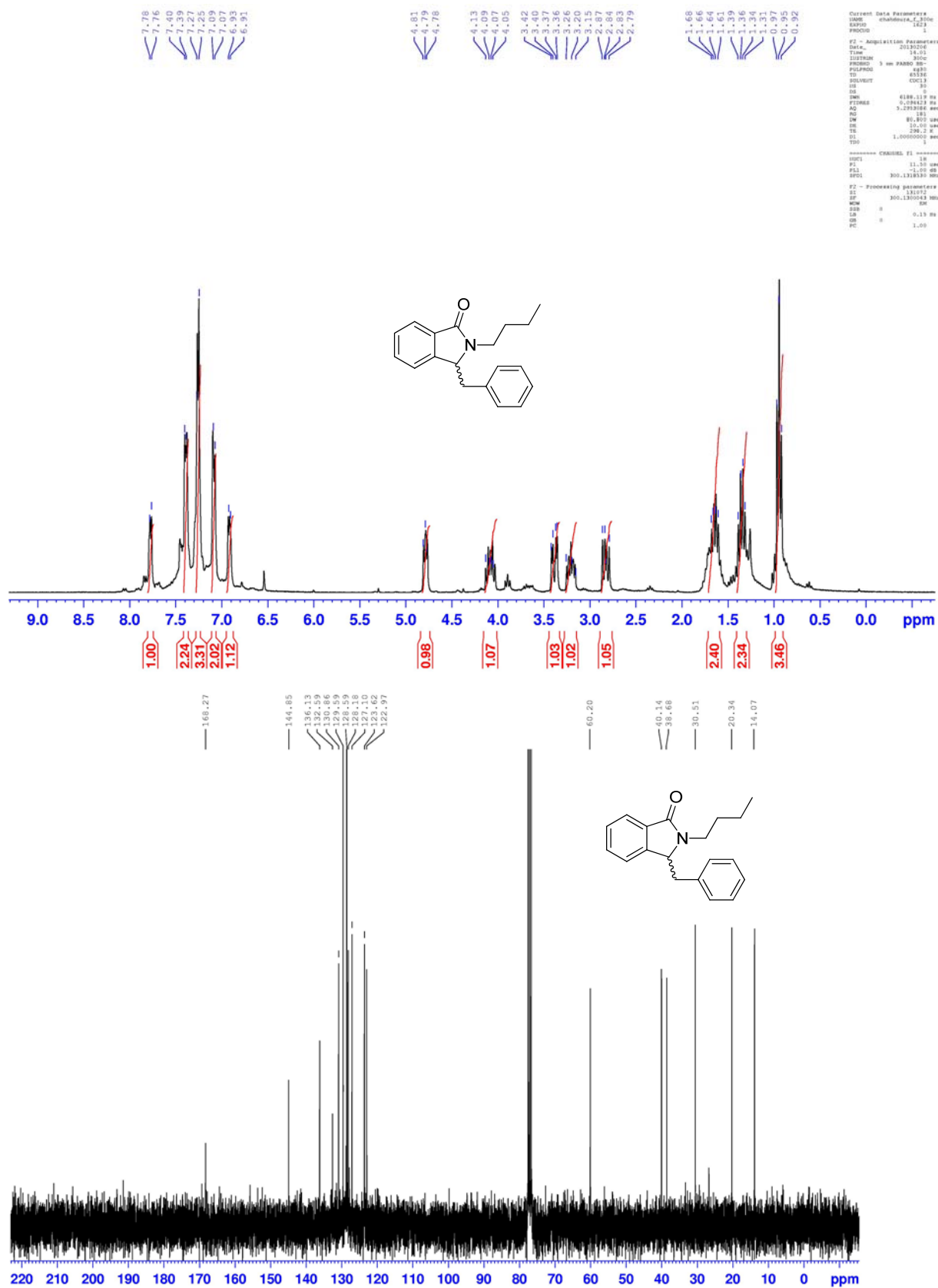
1022

fc-1022-after hydrogen-2ation 2252 (15.263) Cm (2224:2254)

, 07-Feb-2013 + 16:57:02

Scan EI+  
5.45e8





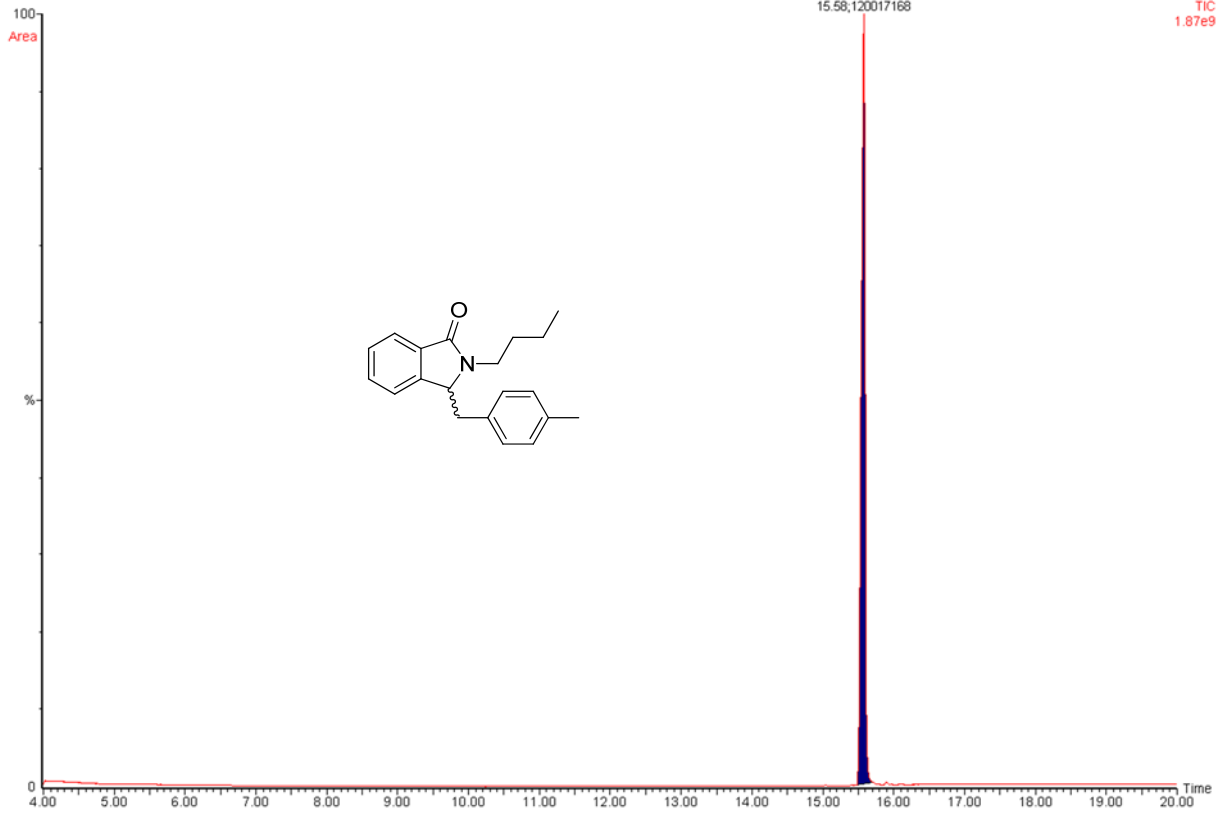
GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **133H**

1026

fc-1026-after hydrogenation Sm (Mn, 1x3)

, 08-Feb-2013 + 17:03:56

Scan EI+  
TIC  
1.87e9

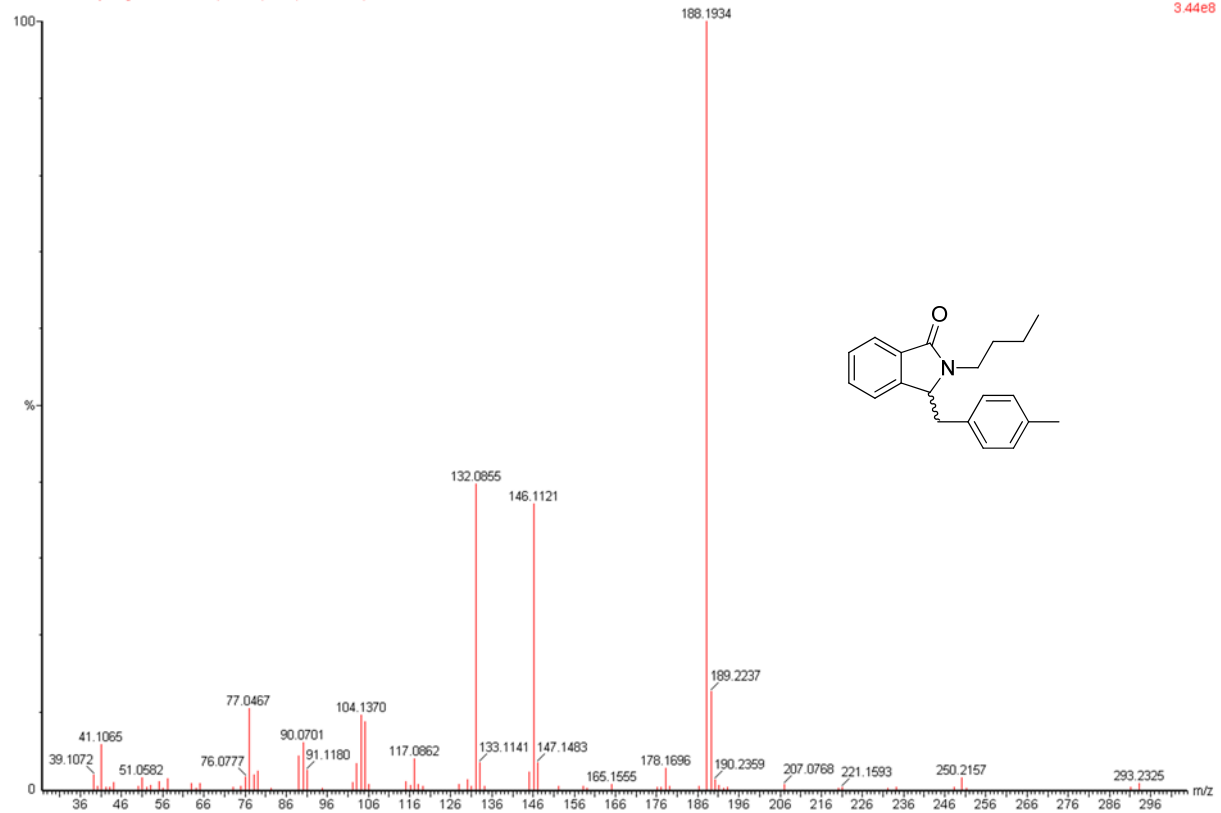


1026

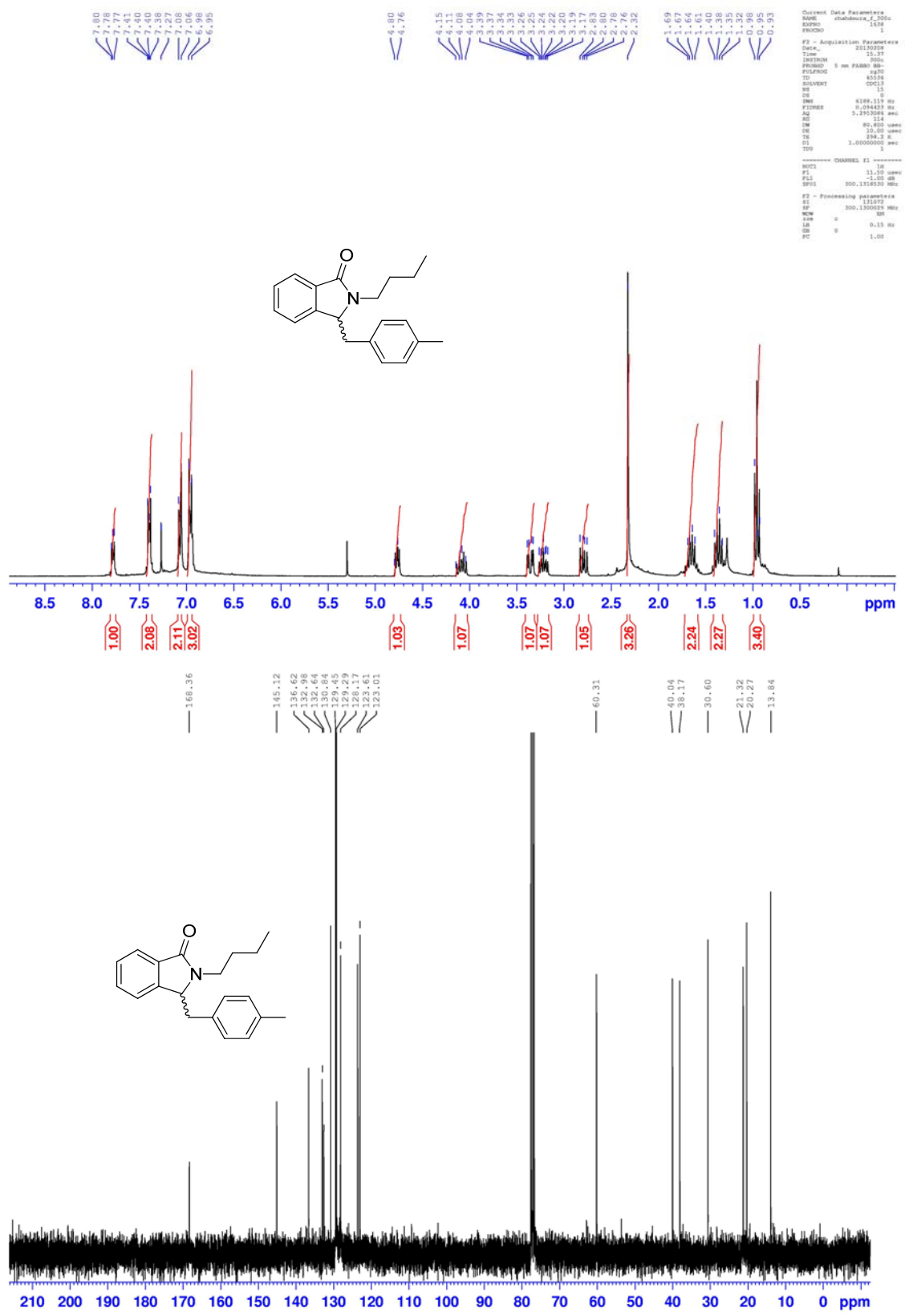
fc-1026-after hydrogenation 2319 (15.598) Cm (2301:2324)

, 08-Feb-2013 + 17:03:56

Scan EI+  
3.44e8







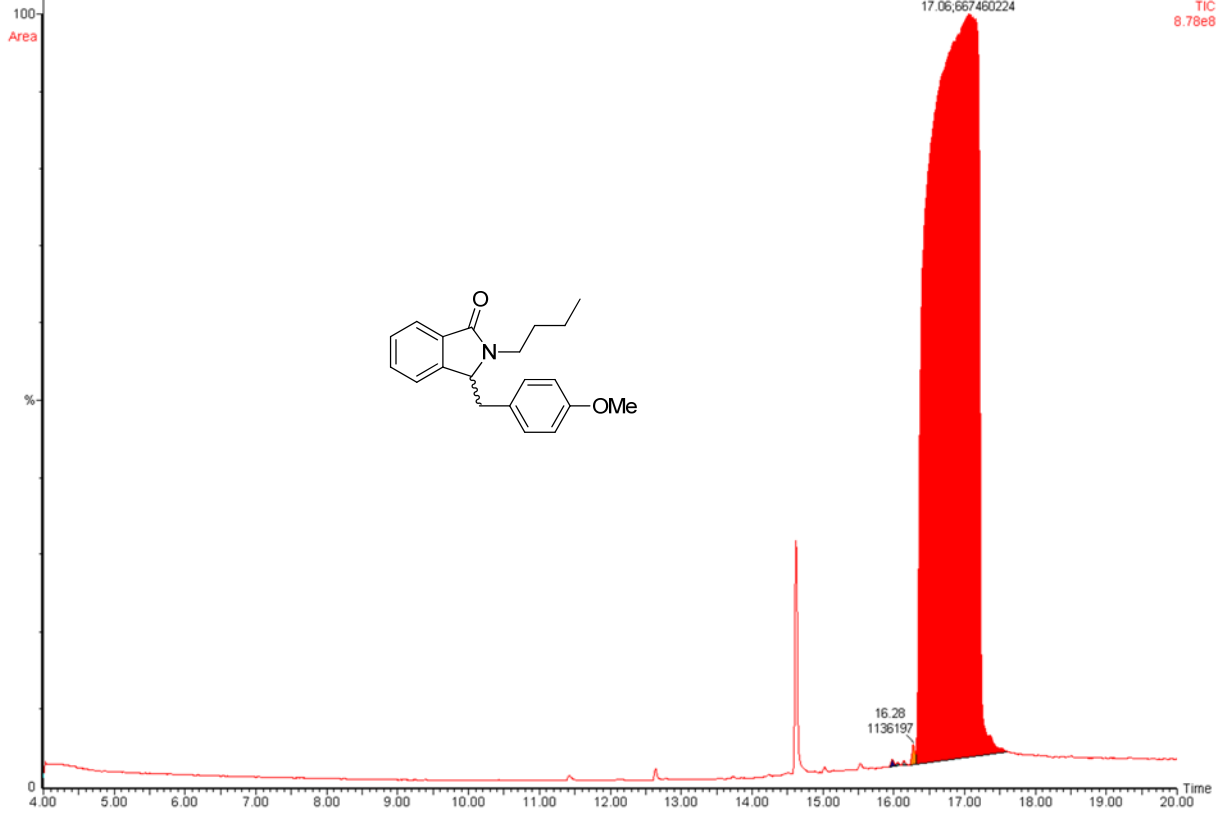
GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C} \{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **134H**

1027

fc-1027-after hydrogenation Sm (Mn, 1x3)

, 08-Feb-2013 + 17:39:00

Scan EI+  
TIC  
8.78e8

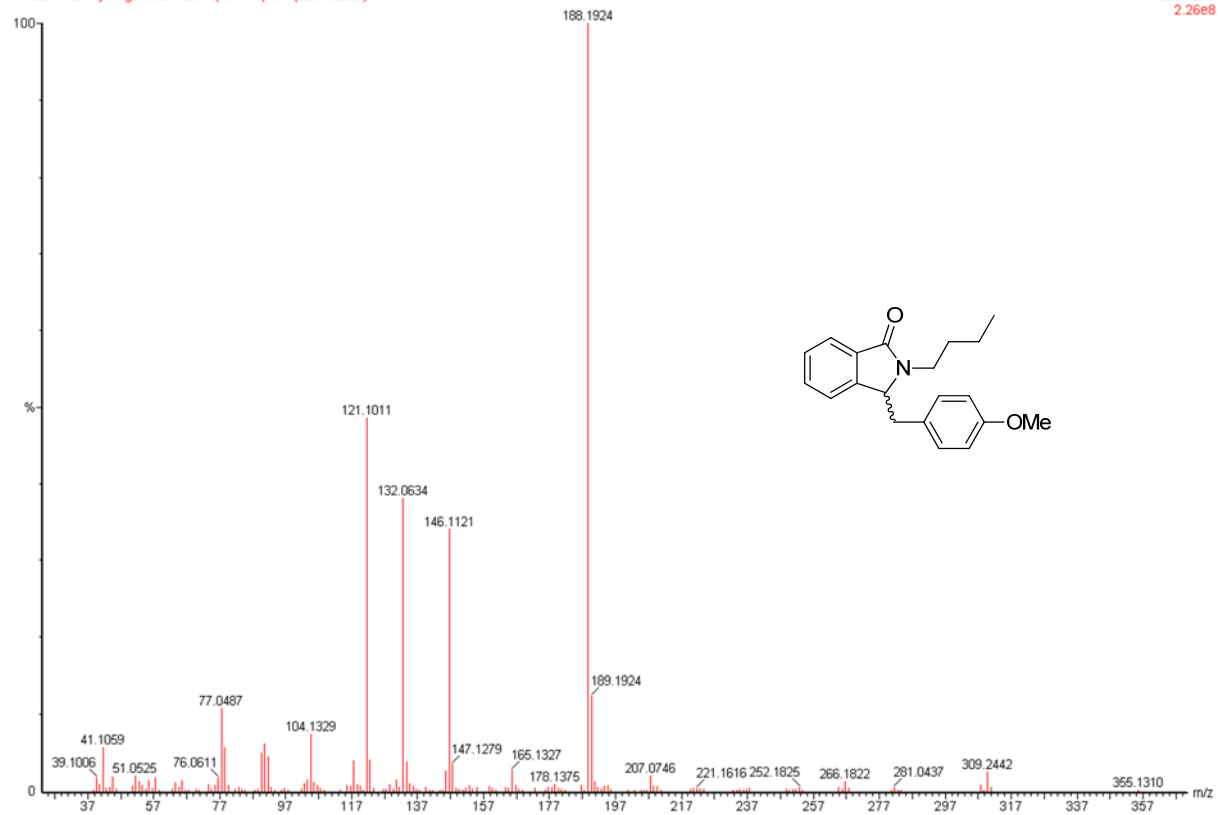


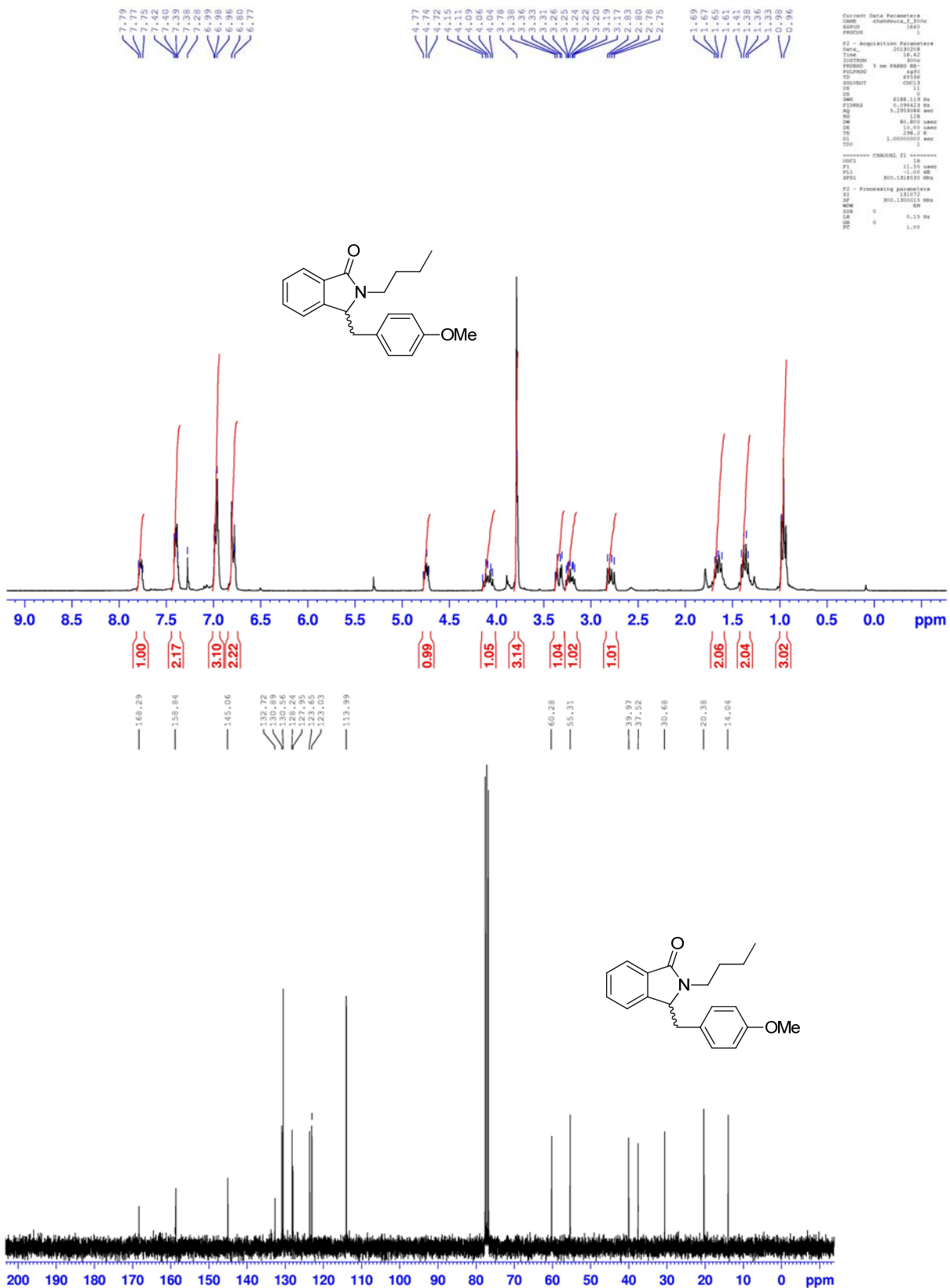
1027

fc-1027-after hydrogenation 2594 (16.974) Cm (2531:2600)

, 08-Feb-2013 + 17:39:00

Scan EI+  
2.26e8





GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **135H**

1078

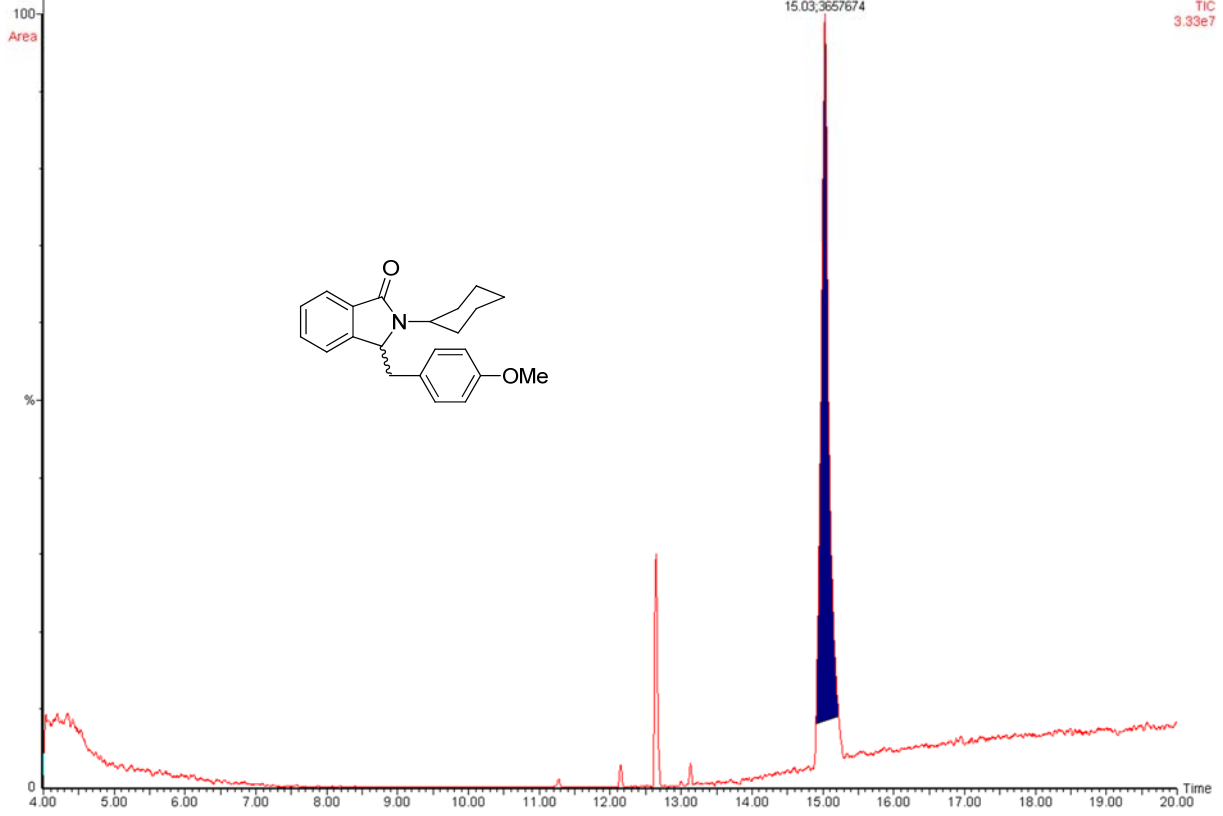
fc-1078-after hydrogenation Sm (Mn, 1x3)

, 28-Mar-2013 + 10:00:20

Scan EI+

TIC

3.33e7



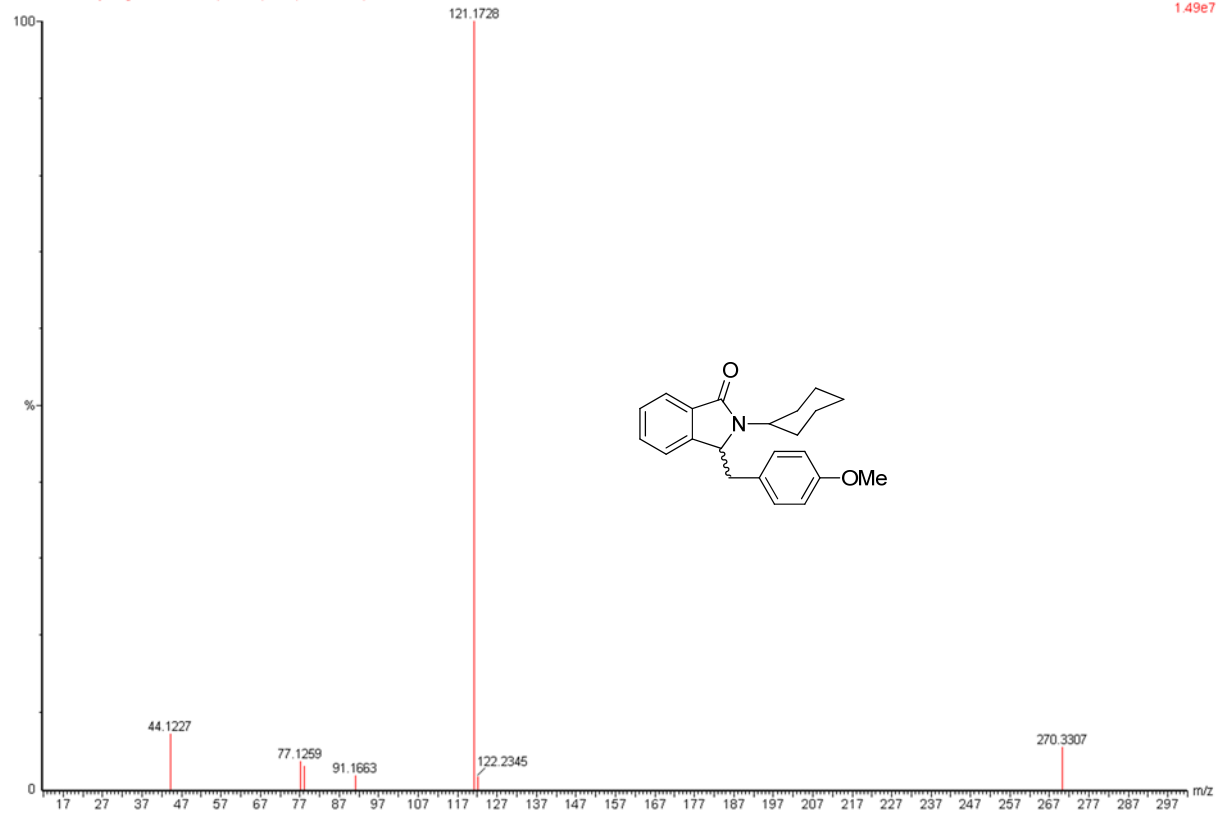
1078

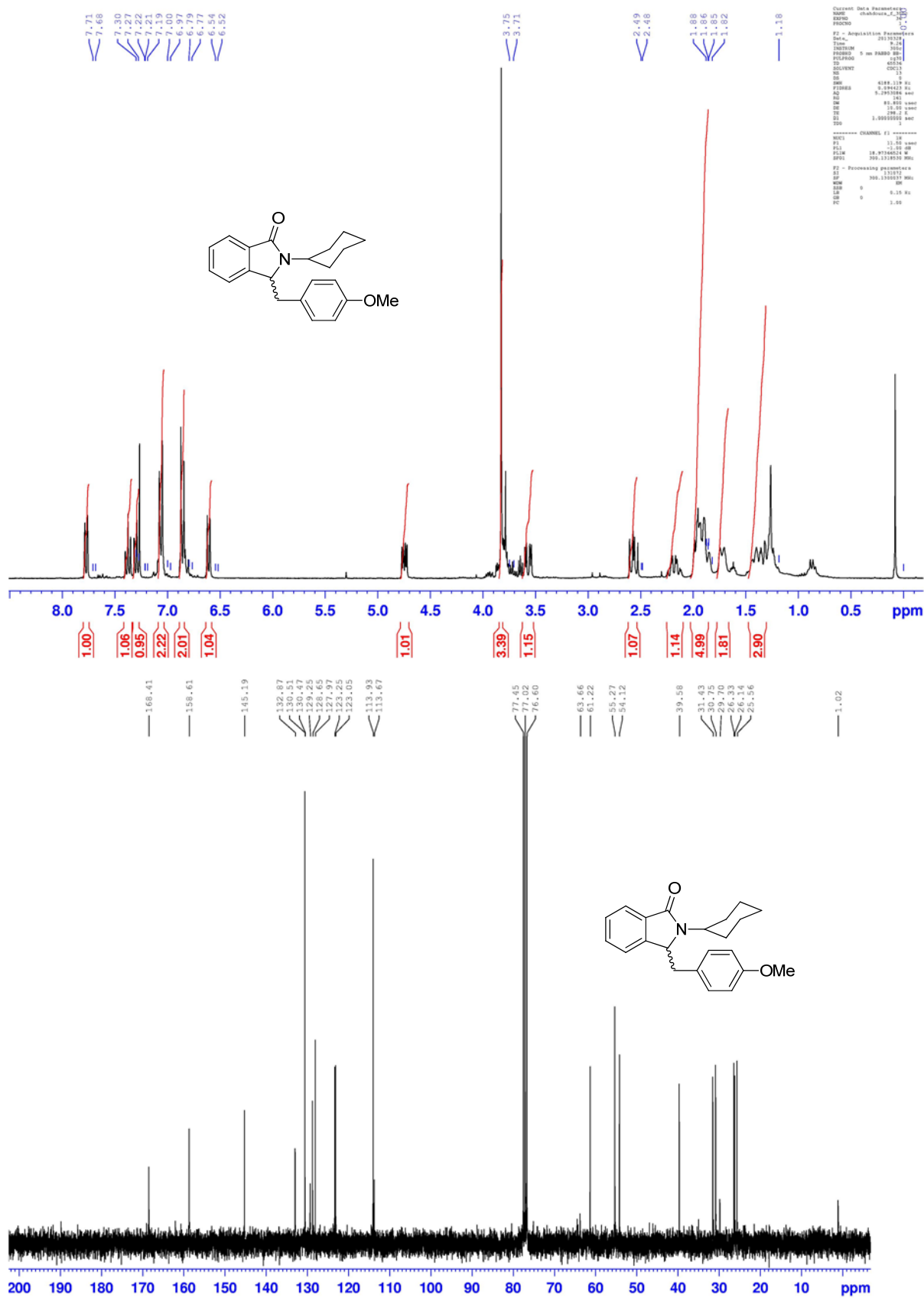
fc-1078-after hydrogenation 2206 (15.033) Cm (2184:2228)

, 28-Mar-2013 + 10:00:20

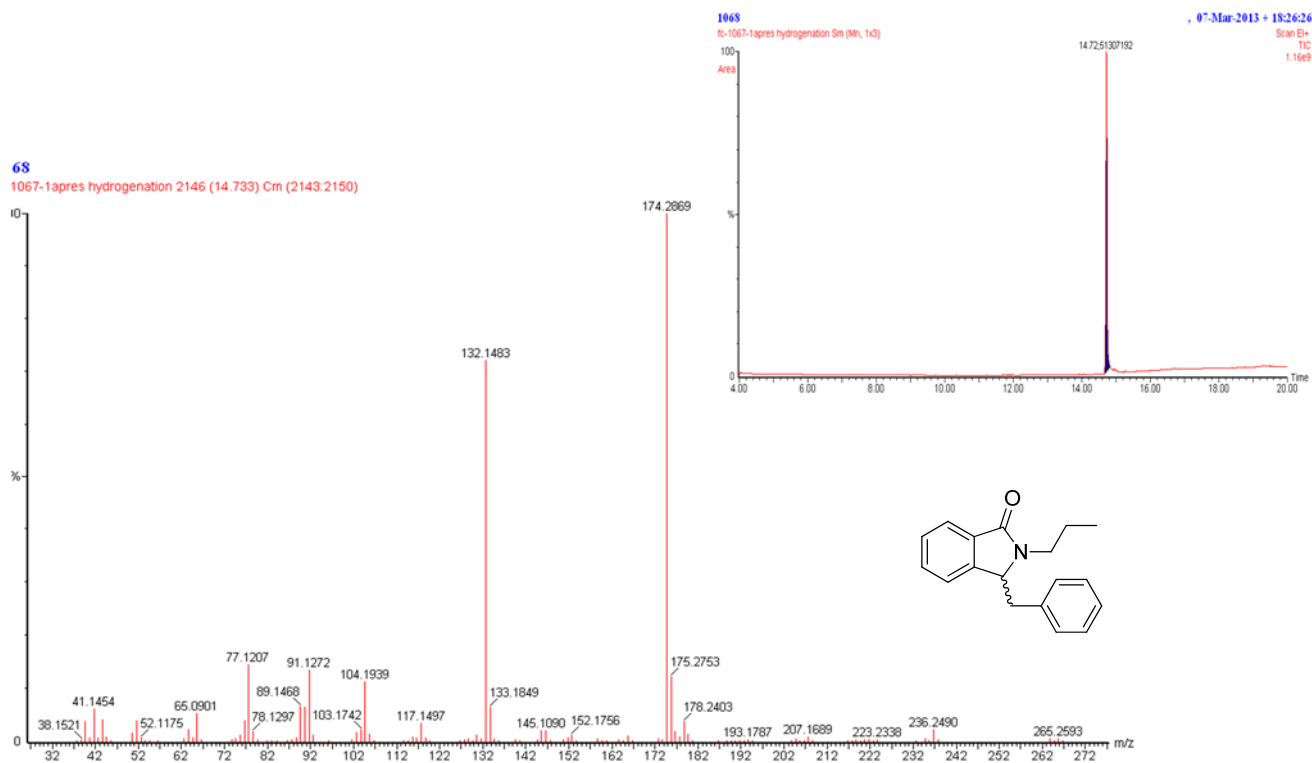
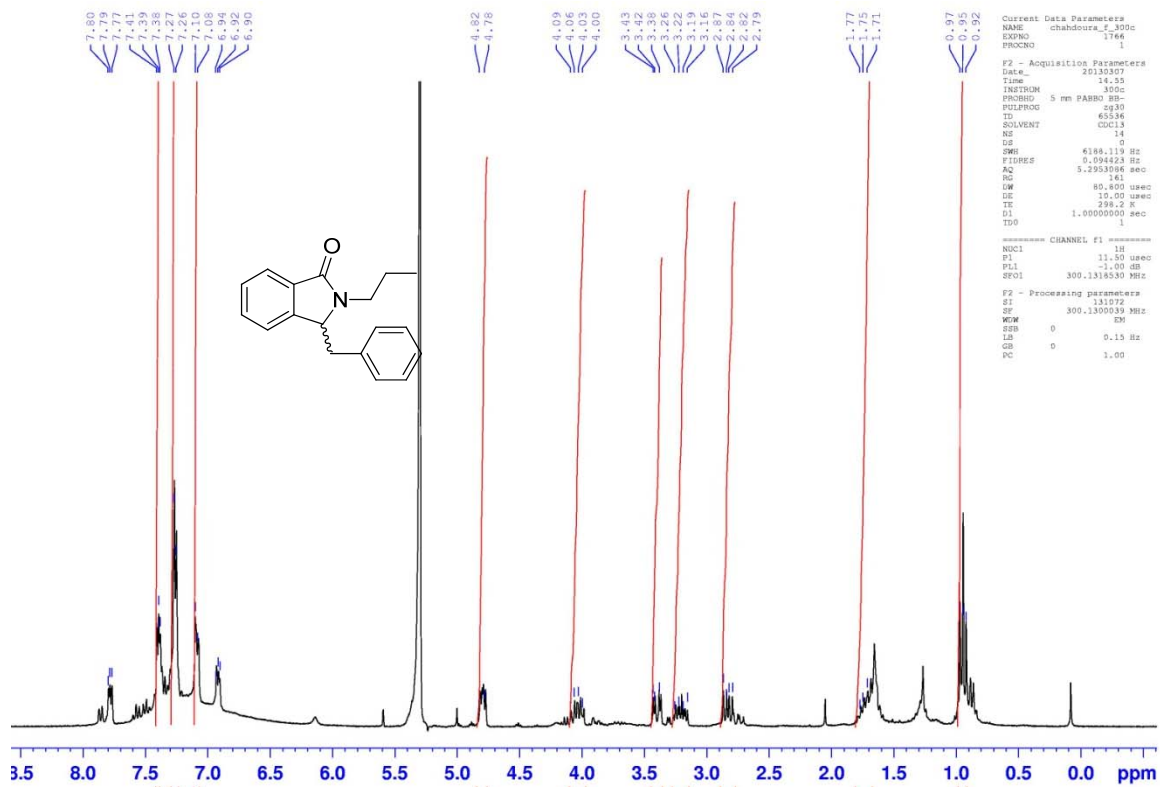
Scan EI+

1.49e7





GC (FID and MS) spectra (top),  $^1\text{H}$  NMR (300 MHz) (middle) and  $^{13}\text{C}$   $\{^1\text{H}\}$  NMR (75 MHz) (bottom) spectra in  $\text{CDCl}_3$  for **n35H**



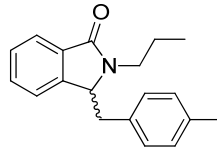
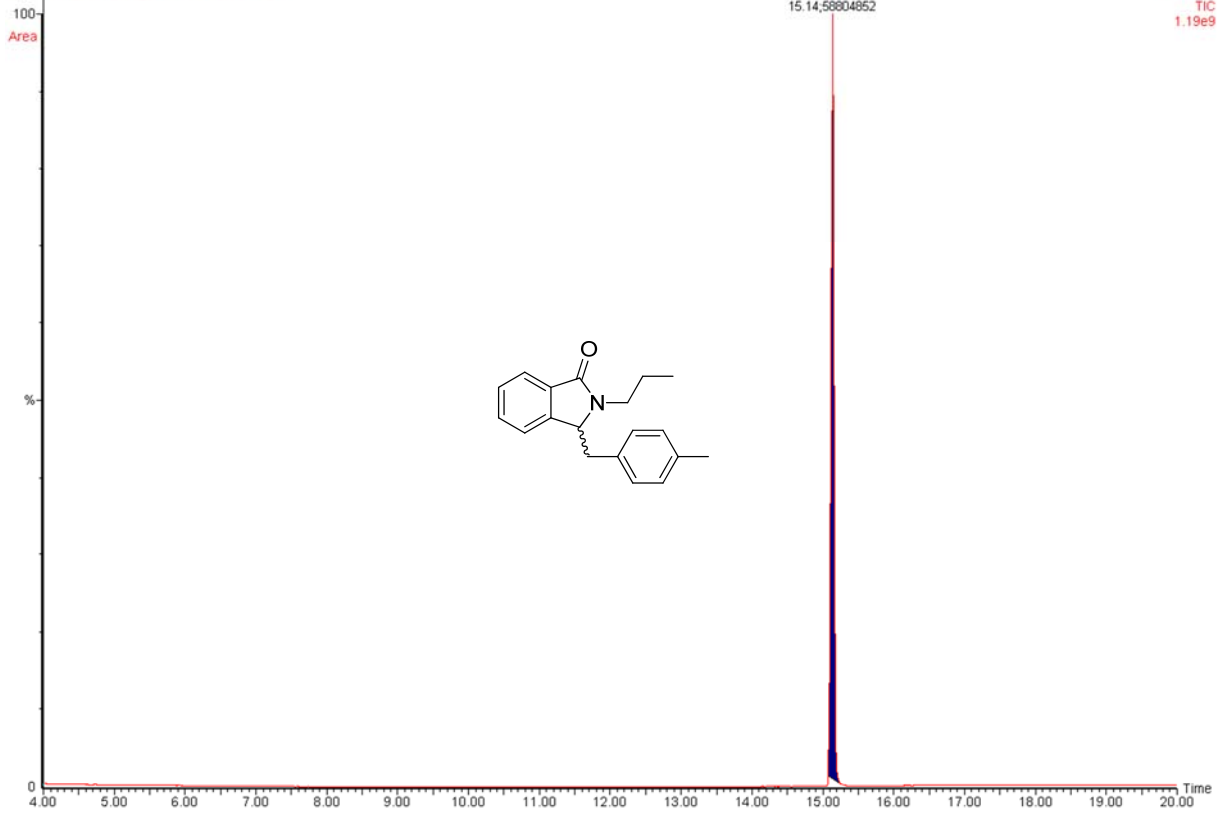
<sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) (top) and GC (FID and MS) spectra (bottom) for **m33H**

1068

fc-1068-2apres hydrogenation Sm (Mn, 1x3)

, 08-Mar-2013 + 13:21:04

Scan EI+  
TIC  
1.19e9

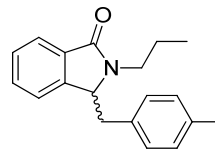
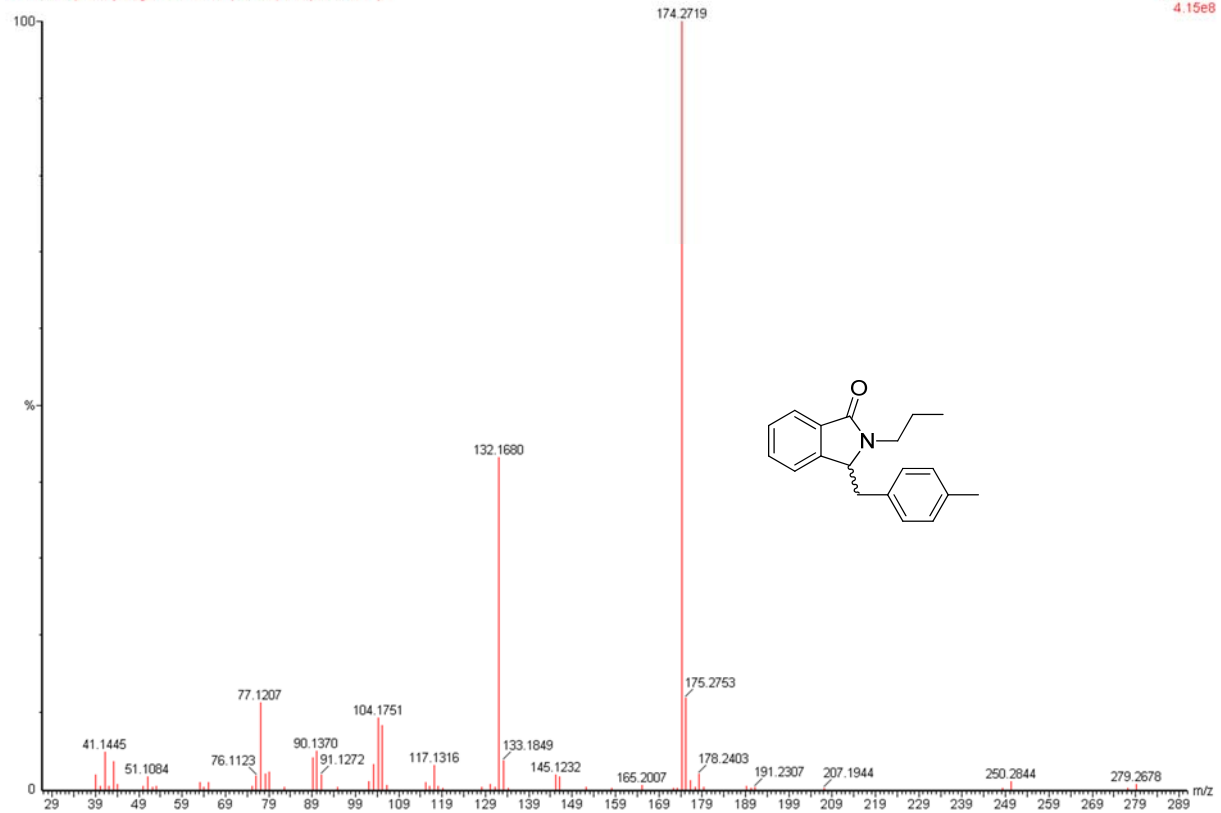


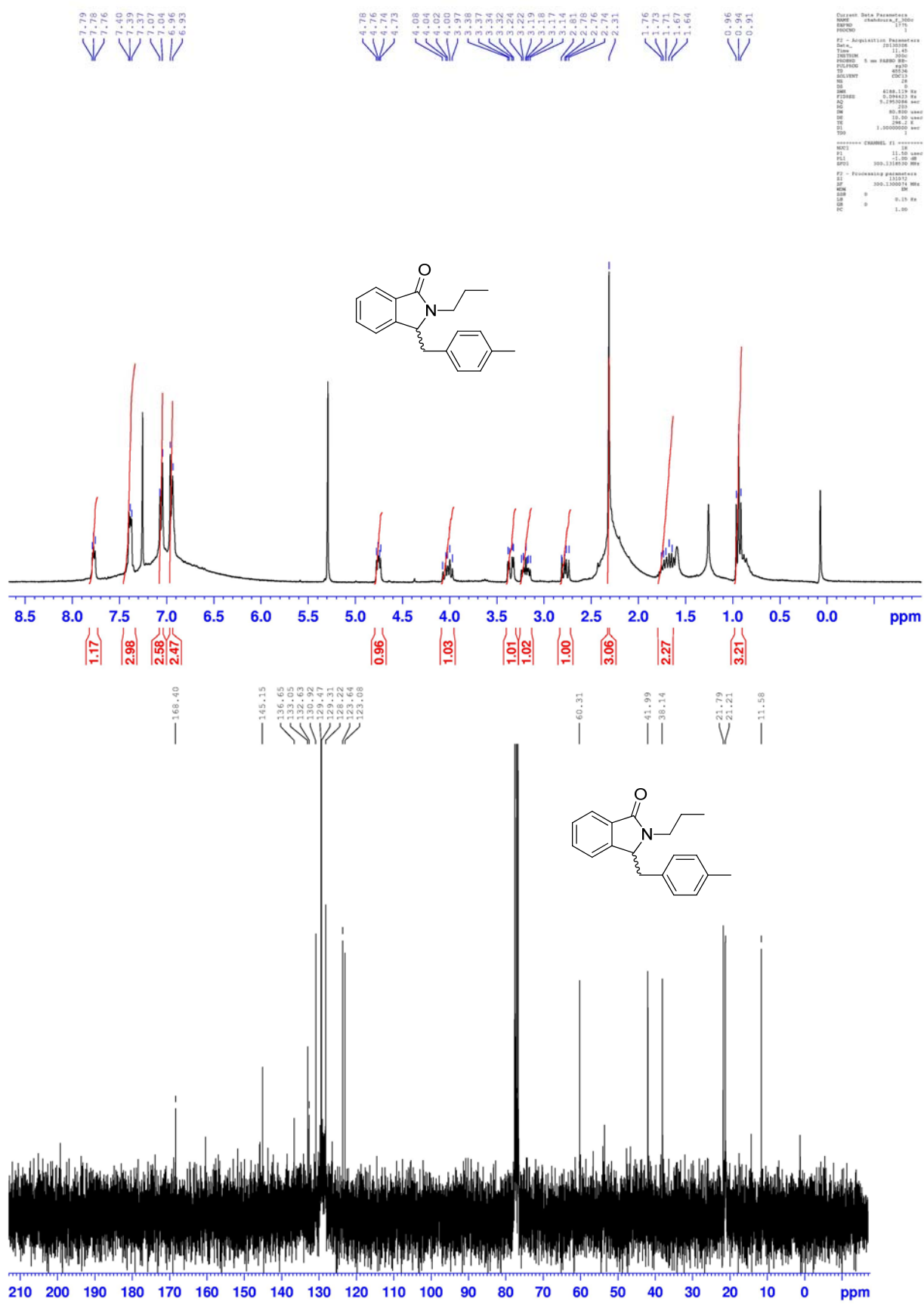
1068

fc-1068-2apres hydrogenation 2230 (15.153) Cm (2222:2231)

, 08-Mar-2013 + 13:21:04

Scan EI+  
4.15e8





GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **m34H**

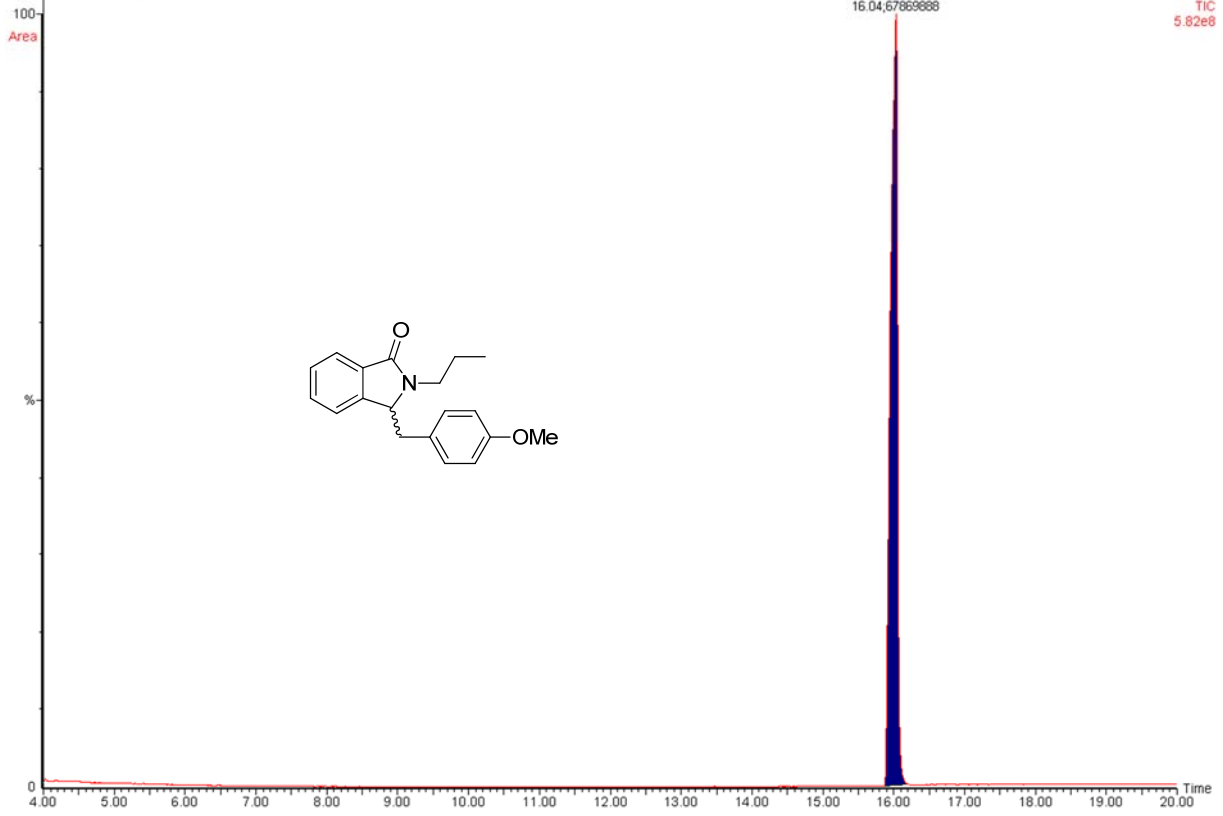


1069

fc-1069-1apres hydrogenation Sm (Mn, 1x3)

, 07-Mar-2013 + 19:24:19

Scan EI+  
TIC  
5.82e8

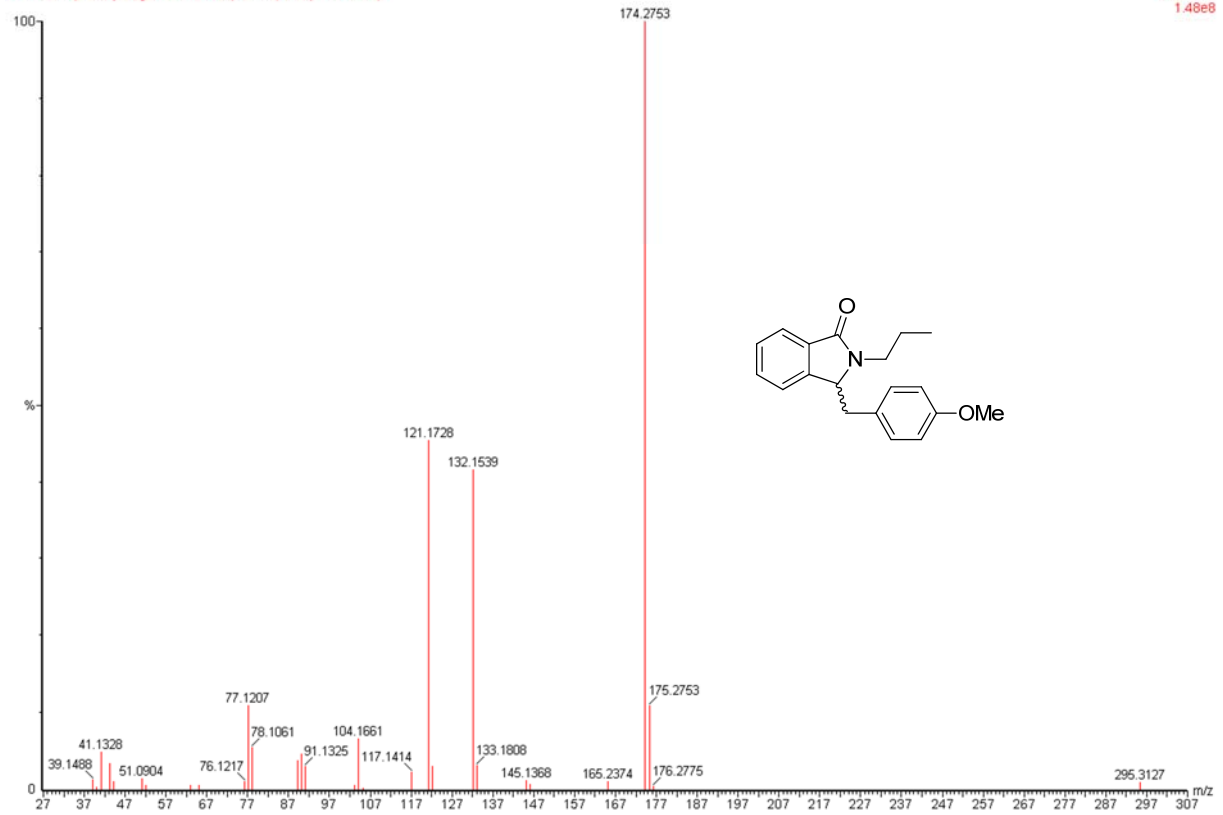


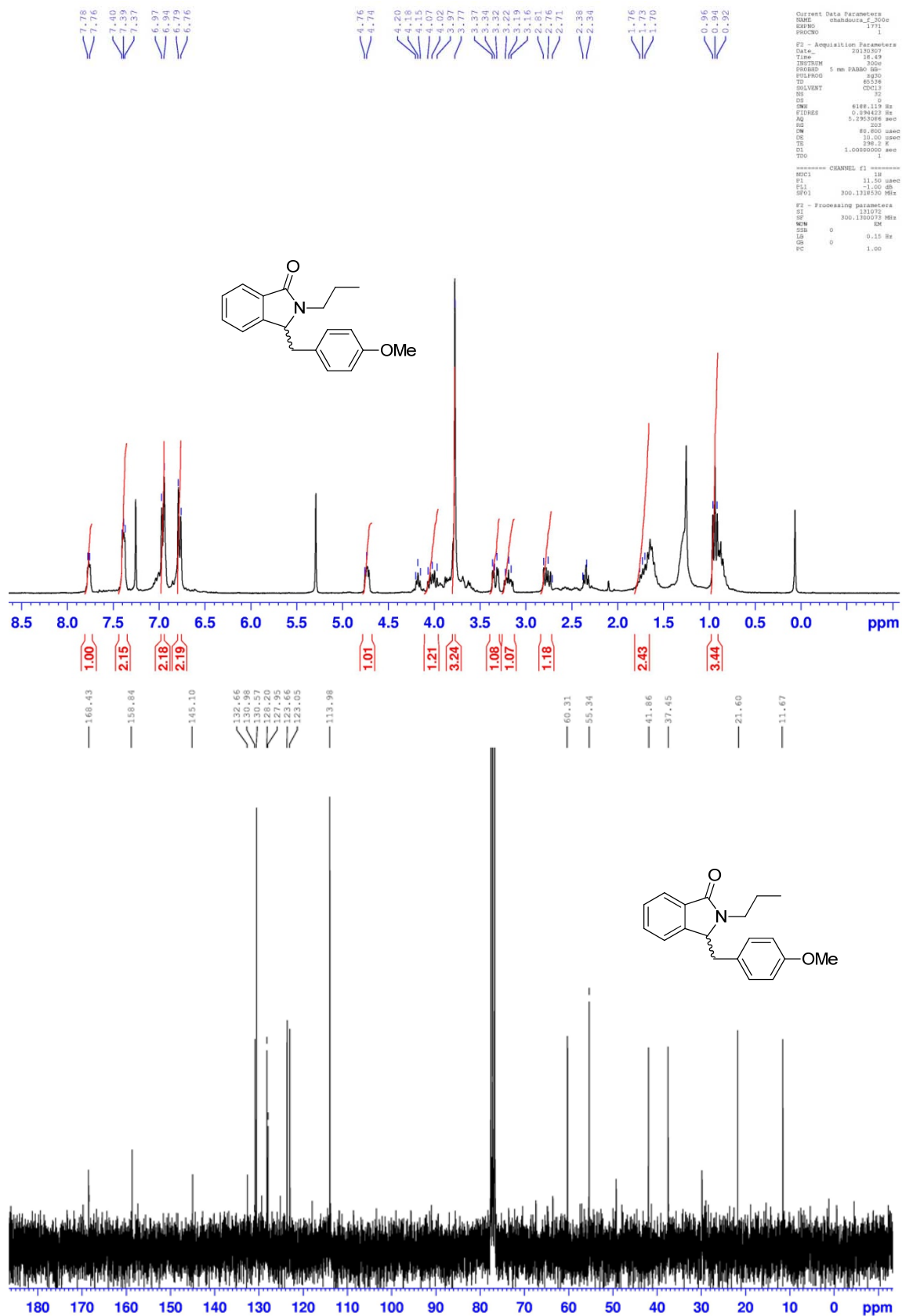
1069

fc-1069-1apres hydrogenation 2406 (16.033) Cm (2381:2406)

, 07-Mar-2013 + 19:24:19

Scan EI+  
1.48e8





GC (FID and MS) spectra (top), <sup>1</sup>H NMR (300 MHz) (middle) and <sup>13</sup>C {<sup>1</sup>H} NMR (75 MHz) (bottom) spectra in CDCl<sub>3</sub> for **m35H**