#### **Supplementary Information**

# Phase-transfer-catalyzed asymmetric desymmetrizations of cyclopentanones

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# **General Information**

<sup>1</sup>H NMR spectra were measured on JEOL JNM-FX 400 NMR instrument (400 MHz for <sup>1</sup>H NMR). <sup>13</sup>C NMR spectra were measured on JEOL JNM-FX 400 NMR instrument (100 MHz for <sup>13</sup>C NMR). Tetramethylsilane (TMS) served as the internal standard (0 ppm) for <sup>1</sup>H NMR, and CDCl<sub>3</sub> served as the internal standard (77.0 ppm) for  ${}^{13}$ C NMR. The following abbreviations were used to express the multiplicities: s = singlet; d = doublet; t = triplet; q = quartet; m = multiplet; br = broad. Highperformance liquid chromatography (HPLC) was performed on Shimadzu 10A instruments using Daicel Chiralpak AD-H or IC (4.6 mm × 250 mm) columns. High-resolution mass spectra (HRMS) were performed on BRUKER microTOF focus-KR. Optical rotations were measured on a JASCO DIP-1000 digital polarimeter. All reactions were monitored by thin-layer chromatography carried out on Merck precoated TLC plates (silica gel 60GF-254, 0.25 mm), visualization by using UV (254 nm), or dyes such as KMnO<sub>4</sub>. The products were purified by flash column chromatography on silica gel 60N [Kanto Chemical Co., Inc. (spherical, neutral)] or Merck preparative thin layer chromatography on silica gel (PLC 60 F254, 0.5 mm). All simple chemicals were purchased and used as received.

## **Experimental Section**

#### Synthesis of substrates 1 and 6.

Substrates 1 and 6 were prepared according to the literature.<sup>1</sup>

### Synthesis of chiral phase-transfer catalysts.

Catalysts (S)-**3**, (S)-**4**, (S)-**5**<sup>4</sup> were prepared according to the literature.

# General procedure for the catalytic asymmetric desymmetrization of epoxy ketones 1 (Table 2).

To a solution of epoxy ketone **1** (0.050 mmol) and chiral phase-transfer catalyst (*S*)-**4b** (1 mol %, 0.00050 mmol) in toluene (0.50 mL) was added 5% aqueous  $K_2CO_3$  (0.35 mL) at 0 °C. The reaction mixture was vigorously stirred for 24 h at 0 °C. The resulting mixture was quenched with saturated aqueous NH<sub>4</sub>Cl and diluted with ethyl acetate. The organic phase was separated and the aqueous phase was extracted with ethyl acetate. The combined extracts were dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated. The residue was purified by column chromatography on silica gel (hexane/ethyl acetate as eluent) to give product **2**. The enantiomeric excess of the product **2** was determined by chiral HPLC analysis.

HO W Ph

2a:<sup>1a</sup>  $[\alpha]^{23}{}_D = -142.4$  (c = 0.40, CHCl<sub>3</sub>), HPLC analysis (92% ee): Daicel Chiralpak AD-H, haxane/2-propanol = 9:1, flow rate = 1.0 mL/min, 254 nm; retention time: 8.7 min (major) and 9.7 min (minor). <sup>1</sup>H NMR (400

MHz, CDCl<sub>3</sub>)  $\delta$  7.43–7.56 (m, 4H), 7.26–7.39 (m, 6H), 6.71 (s, 1H), 3.02 (d, J = 18.4 Hz, 1H), 2.91 (d, J = 18.4 Hz, 1H), 2.59 (br, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  204.7, 173.9, 144.1, 131.4, 130.8, 129.1, 128.9, 128.8, 127.5, 124.2, 81.7, 56.6; IR (neat) 3392, 1681, 1592, 1569, 1494, 1447, 1249, 1208, 1051, 977, 766, 723, 701, 689 cm<sup>-1</sup>; HRMS (ESI-TOF) calcd for C<sub>17</sub>H<sub>14</sub>NaO<sub>2</sub><sup>+</sup>: 273.0886 ([M+Na]<sup>+</sup>), found 273.0880.



**2b**:  $[\alpha]_{D}^{25} = -83.3$  (*c* = 0.36, CHCl<sub>3</sub>), HPLC analysis (90% ee): Daicel Chiralpak AD-H, haxane/2-propanol = 9:1, flow rate = 1.0 mL/min, 254 nm; retention time: 9.2 min (major) and 12.2 min (minor). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.43 (d, *J* = 8.0 Hz, 2H), 7.32 (d, *J* = 8.0 Hz, 2H), 7.14 (d, *J* = 8.0 Hz, 2H), 7.10 (d, *J* = 8.0 Hz, 2H), 7.14 (d, *J* = 8.0 Hz, 2H), 7.10 (d, *J* = 8.0 Hz, 2H), 7.14 (d, *J* = 8.0 Hz, 2H), 7.10 (d, *J* = 8.0 Hz, 2H), 7.14 (d, *J* = 8.0 Hz, 2H), 7.10 (d, *J* = 8.0 Hz, 2H), 7.14 (d, *J* = 8.0 Hz, 2H), 7.10 (d, *J* = 8.0 Hz, 2H), 7.14 (d, *J* = 8.0 Hz, 2H), 7.10 (d, *J* = 8.0 Hz, 2H), 7.14 (d, *J* = 8.0 Hz, 2H), 7.10 (d, *J* = 8.0 Hz, 2H), 7.14 (d, *J* = 8.0 Hz, 2H), 7.10 (d, *J* = 8.0 Hz, 10 Hz), 7.10 (d, *J* = 8.0 Hz), 7.10 (d, *J* = 8.0 Hz), 8.0 Hz (d, J) = 8.0 Hz (d, J) = 8.0 Hz), 8.0 Hz (d, J) = 8.0

2H), 6.65 (s, 1H), 2.98 (d, J = 18.4 Hz, 1H), 2.85 (d, J = 18.4 Hz, 1H), 2.66 (br, 1H), 2.32 (s, 3H), 2.31 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  204.8, 173.9, 141.4, 141.3, 137.1, 129.5, 129.2, 129.1, 128.6, 128.2, 124.1, 81.6, 56.7, 21.4, 21.0; IR (neat) 3390, 1679, 1610, 1590, 1511, 1323, 1301, 1275, 1248, 1220, 1203, 819, 730 cm<sup>-1</sup>; HRMS (ESI-TOF) calcd for C<sub>19</sub>H<sub>18</sub>NaO<sub>2</sub><sup>+</sup>: 301.1199 ([M+Na]<sup>+</sup>), found 301.1194.



**2c**:  $[\alpha]^{26}{}_{D} = -100.7$  (*c* = 0.38, CHCl<sub>3</sub>), HPLC analysis (91% ee): Daicel Chiralpak AD-H, haxane/2-propanol = 9:1, flow rate = 1.0 mL/min, 254 nm; retention time: 7.3 min (major) and 8.1 min (minor). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.35 (s, 1H), 7.22–7.27 (m, 4H), 7.14–7.21 (m, 2H), 7.05–7.12 (m, 1H), 6.67 (s,

1H), 2.99 (d, J = 18.8 Hz, 1H), 2.88 (d, J = 18.8 Hz, 1H), 2.55 (br, 1H), 2.33 (s, 3H), 2.29 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  204.7, 174.1, 144.2, 138.6, 138.5, 131.7, 131.4, 129.7, 129.1, 128.7, 128.2, 126.2, 124.8, 121.2, 116.7, 81.7, 56.6, 21.6, 21.4; IR (neat) 3392, 1680, 1591, 1576, 1487, 1304, 1283, 1252, 1210, 1178, 1058, 788, 722, 703 cm<sup>-1</sup>; HRMS (ESI-TOF) calcd for C<sub>19</sub>H<sub>18</sub>NaO<sub>2</sub><sup>+</sup>: 301.1199 ([M+Na]<sup>+</sup>), found 301.1207.



**2d**:  $[\alpha]_{D}^{23} = -210.8$  (*c* = 0.78, CHCl<sub>3</sub>), HPLC analysis (93% ee): Daicel Chiralpak AD-H, haxane/2-propanol = 9:1, flow rate = 1.0 mL/min, 254 nm; retention time: 6.9 min (major) and 8.5 min (minor). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.53–7.61 (m, 2H), 7.36–7.44 (m, 2H), 6.96–7.06 (m, 4H), 6.64 (s, 1H), 2.99 (d, *J* = 18.8 Hz, 1H), 2.88 (d, J = 18.8 Hz, 1H), 2.88 (d, J = 18.8 Hz, 1H), 2.88 (d, J = 18.8 Hz, 1H)

1H), 2.72 (br, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  204.0, 172.4, 164.1 (d,  $J_{C-F} = 255.1$  Hz), 162.1 (d,  $J_{C-F} = 247.8$  Hz), 139.5 (d,  $J_{C-F} = 3.3$  Hz), 131.3 (d,  $J_{C-F} = 9.0$  Hz), 128.8, 127.4 (d,  $J_{C-F} = 3.3$  Hz), 126.1 (d,  $J_{C-F} = 8.3$  Hz), 116.0 (d,  $J_{C-F} = 22.2$  Hz), 115.8 (d,  $J_{C-F} = 21.4$  Hz), 81.3, 56.6; IR (neat) 3381, 1684, 1601, 1579, 1508, 1414, 1279, 1238, 1208, 1163, 1052, 980, 837 cm<sup>-1</sup>; HRMS (ESI-TOF) calcd for C<sub>17</sub>H<sub>12</sub>F<sub>2</sub>NaO<sub>2</sub><sup>+</sup>:

309.0698 ([M+Na]<sup>+</sup>), found 309.0686.



**2e**:  $[\alpha]^{26}{}_{D} = -145.9$  (*c* = 0.36, CHCl<sub>3</sub>), HPLC analysis (90% ee): Daicel Chiralpak AD-H, haxane/2-propanol = 9:1, flow rate = 1.0 mL/min, 254 nm; retention time: 6.9 min (major) and 8.8 min (minor). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.49 (d, *J* = 8.8 Hz, 2H), 7.35 (d, *J* = 8.8 Hz, 2H), 7.30 (d, *J* = 8.8 Hz, 2H), 7.27 (d, *J* = 8.8 Hz, 2H), 6.63 (s,

1H), 2.98 (d, J = 18.8 Hz, 1H), 2.96 (br, 1H), 2.87 (d, J = 18.8 Hz, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  204.1, 172.3, 142.2, 137.3, 133.6, 130.4, 129.6, 129.2, 129.13, 129.07, 125.7, 81.2, 56.5; IR (neat) 3390, 1680, 1589, 1490, 1319, 1210, 1093, 1013, 975, 827, 749, 730 cm<sup>-1</sup>; HRMS (ESI-TOF) calcd for C<sub>17</sub>H<sub>12</sub>Cl<sub>2</sub>NaO<sub>2</sub><sup>+</sup>: 341.0107 ([M+Na]<sup>+</sup>), found 341.0108.



**2f**:  $[\alpha]^{25}_{D}$  = +8.6 (*c* = 0.78, CHCl<sub>3</sub>), HPLC analysis (96% ee): Daicel Chiralpak IC, haxane/2-propanol = 9:1, flow rate = 0.75 mL/min, 254 nm; retention time: 28.5 min (minor) and 34.0 min (major). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.63–7.69 (m, 2H), 7.51–7.62 (m, 10H), 7.39–7.46 (m, 4H), 7.31–7.38 (m, 2H), 6.77 (s, 1H), 3.07 (d, *J* = 18.8

Hz, 1H), 2.96 (d, J = 18.8 Hz, 1H), 2.77 (br, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  204.5, 173.3, 143.6, 143.1, 140.4, 139.7, 130.2, 129.7, 128.9, 128.8, 128.1, 127.5, 127.44, 127.37, 127.0, 124.7, 81.6, 56.7; IR (neat) 3367, 1678, 1588, 1487, 1408, 1298, 1208, 1007, 978, 908, 841, 768, 734, 696 cm<sup>-1</sup>; HRMS (ESI-TOF) calcd for C<sub>29</sub>H<sub>22</sub>NaO<sub>2</sub><sup>+</sup>: 425.1512 ([M+Na]<sup>+</sup>), found 425.1501.



**2g**:  $[\alpha]^{22}{}_{D} = -126.1$  (*c* = 1.57, CHCl<sub>3</sub>), HPLC analysis (90% ee): Daicel Chiralpak AD-H, haxane/2-propanol = 9:1, flow rate = 1.0 mL/min, 254 nm; retention time: 13.2 min (major) and 15.7 min (minor). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.10 (d, *J* = 1.6 Hz, 1H), 8.07 (d, *J* = 1.6 Hz, 1H), 7.78–7.86 (m, 3H), 7.71–7.75 (m,

2H), 7.61–7.66 (m, 2H), 7.43–7.52 (m, 4H), 7.39 (dd, *J* = 7.6, 7.6 Hz, 1H), 6.88 (s, 1H), 3.12 (d, *J* = 18.8 Hz, 1H), 3.01 (d, *J* = 18.8 Hz, 1H), 2.96 (br, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 204.6, 173.6, 141.5, 134.1, 133.3, 132.8, 132.6, 130.2, 129.5, 129.1, 129.0, 128.7, 128.5, 128.2, 127.8, 127.6, 127.5, 126.6, 126.5, 126.2, 125.4, 123.1, 122.4, 82.0,

56.7; IR (neat) 3368, 1679, 1600, 1584, 1567, 1299, 1272, 1250, 1197, 907, 857, 817, 749, 731 cm<sup>-1</sup>; HRMS (ESI-TOF) calcd for  $C_{25}H_{18}NaO_2^+$ : 373.1199 ([M+Na]<sup>+</sup>), found 373.1200.

#### Procedure for the catalytic asymmetric isomerization of ketone 6 (Scheme 2).

To a solution of ketone **6** (0.050 mmol) and chiral phase-transfer catalyst (*S*)-**4b** (3 mol %, 0.0015 mmol) in trifluoromethylbenzene (0.50 mL) was added 50% aqueous  $Cs_2CO_3$  (0.50 mL) at -20 °C. The reaction mixture was vigorously stirred for 48 h at -20 °C. The resulting mixture was quenched with saturated aqueous NH<sub>4</sub>Cl and diluted with ethyl acetate. The organic phase was separated and the aqueous phase was extracted with ethyl acetate. The combined extracts were dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated. The residue was purified by column chromatography on silica gel (hexane/ethyl acetate 10:1–3:1 as eluent) to give product 7. The enantiomeric excess of the product 7 was determined by chiral HPLC analysis.

7:  $[\alpha]^{26}_{D} = -315.1$  (c = 0.75, CHCl<sub>3</sub>), HPLC analysis (85% ee): Daicel Chiralpak AD-H, haxane/2-propanol = 9:1, flow rate = 1.0 mL/min, 254 nm; retention time: 8.8 min (major) and 11.6 min (minor). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.53 (dd, J = 1.6, 8.0 Hz, 2H), 7.22–7.36 (m, 5H), 7.11–7.21 (m, 3H), 6.75 (d, J = 1.2 Hz, 1H), 4.64 (d, J = 7.2 Hz, 1H), 3.12 (dd, J = 7.2, 19.2 Hz, 1H), 2.46 (dd, J = 2.0, 19.2 Hz, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  207.9, 175.1, 142.5, 133.3, 130.8, 129.2, 129.1, 128.7, 127.9, 127.1, 127.0, 46.9, 46.7; IR (neat) 1708, 1683,

1594, 1570, 1494, 1447, 1326, 1307, 1268, 1186, 943, 788, 760, 701, 690 cm<sup>-1</sup>; HRMS (ESI-TOF) calcd for  $C_{17}H_{14}NaO^+$ : 257.0937 ([M+Na]<sup>+</sup>), found 257.0938.

#### Determination of absolute configuration of products 2.

The absolute configuration of product 2a was determined by comparison of the optical rotation with the literature value.<sup>1a</sup>

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S-23



S-24





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