

Electronic Supplementary

Structural Characterization, Solution Stability, and Potential Health and Environmental Effects of the Nano-TiO₂ Bioencapsulation Matrix and the Model Product of its Biodegradation TiBALDH

Nicole Groenke,^{‡,§} Gulaim A. Seisenbaeva,[†] Vitaliy Kaminsky,[‡] Boris Zhivotovsky,[‡]
Benedikt Kost,[§] Vadim G. Kessler^{‡*}

[‡] Department of Chemistry, SLU BioCenter, Box 7015, 75007 Uppsala, Sweden;

[§] Department of Plant Biology and Forest Pathology, SLU BioCenter, Box 7082, 75007
Uppsala, Sweden; [‡] Department of Toxicology, Institute of Environmental Medicine,
Karolinska Institutet, Box 210, 17177 Stockholm, Sweden

Figure FS1 TGA of Captigel derived xerogel

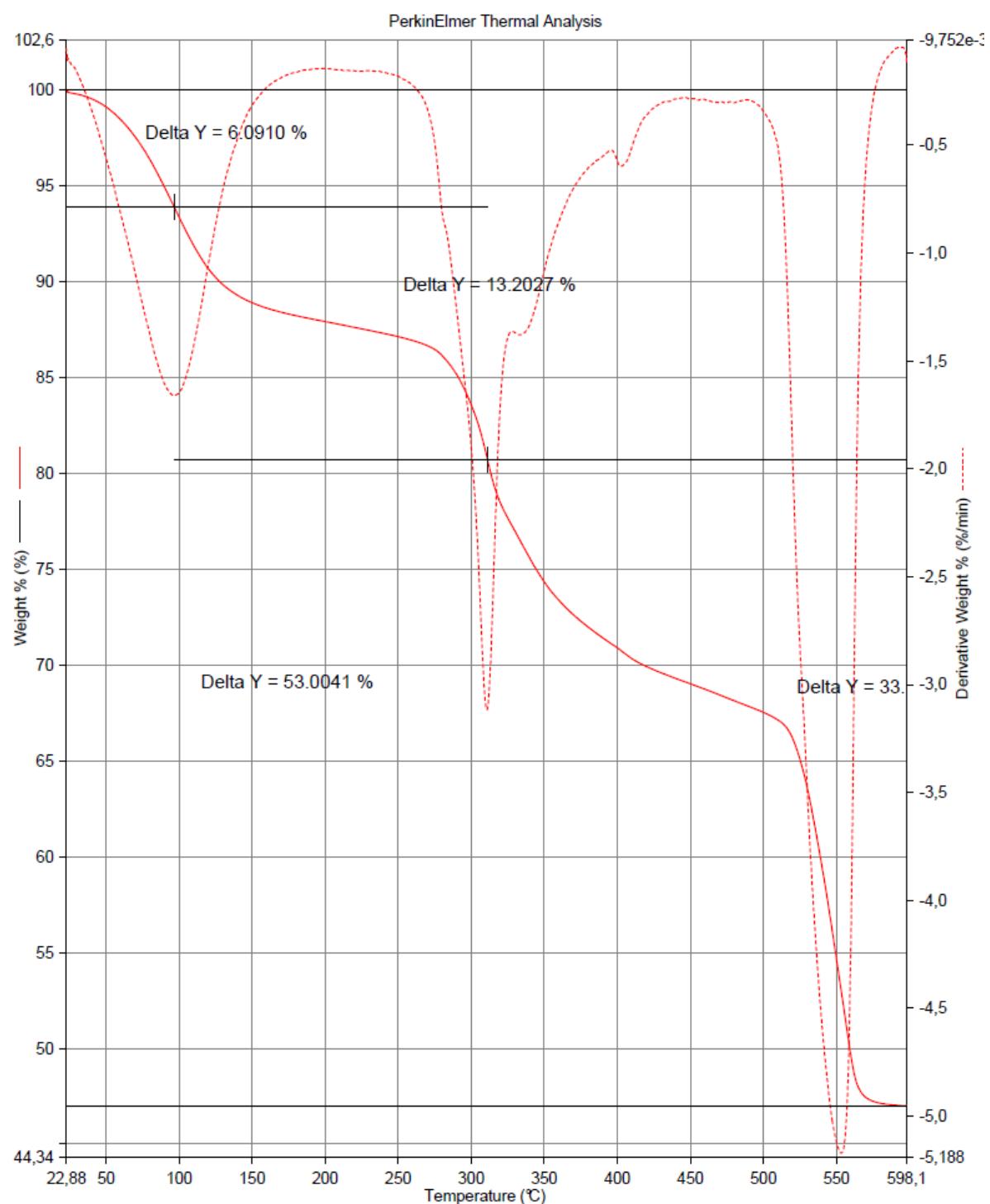


Figure FS2 X-ray powder pattern of the TiO₂ powder produced by heat treatment of CaptiGel at 500°C

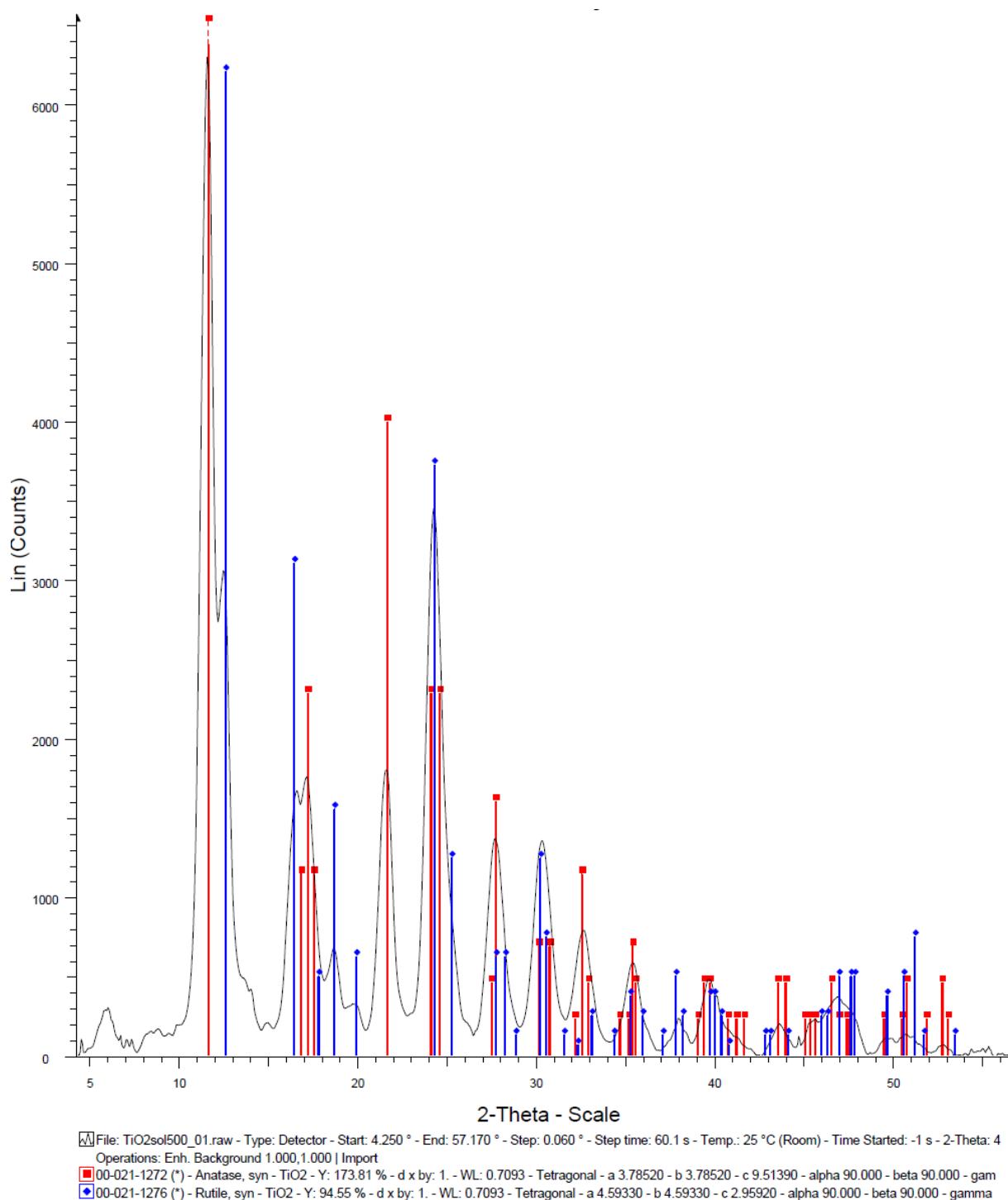


Figure FS3 Phase-corrected Fourier transformed EXAFS spectra of CaptiGel (a, this work) and of crystalline anatase (b, reprinted from R. Bouchet, A. Weibel, P. Knauth, G. Mountjoy, A.V. Chadwick, Chem. Mater. 2003, 15, 4996-5002), in nanopowder (solid line) and bulk sample (dotted line).

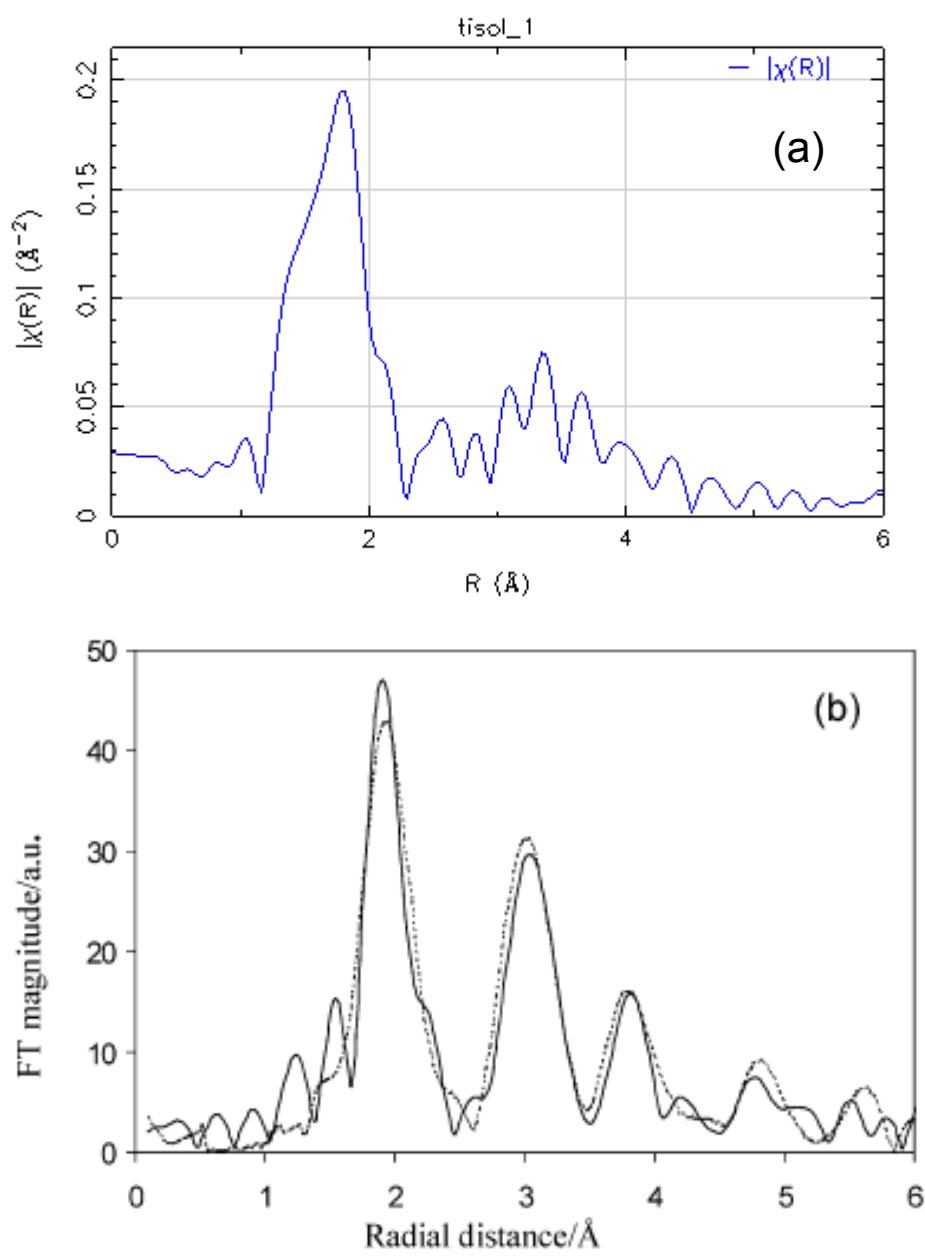


Table TS1 Size-measurement of TiO₂, non-dispersed and ultra sonificated

Direct Run								
Non-dispersed				Dispersed				
	Z-Ave (dia.nm)	Peak 1 (dia.nm)	%	Peak 2 (dia.nm)	%	Peak 3 (dia.nm)	%	Z-Ave (dia.nm)
1	59,98	151,4	80,7	6,605	13,3	24,41	4,9	26,71
2	61,36	142,6	77,3	5,519	9,8	15,49	9,7	24,24
3	61,27	134,3	77,9	4,011	13,2	10,94	5,1	23,41

48 h delay								
Non-dispersed				Dispersed				
	Z-Ave (dia.nm)	Peak 1 (dia.nm)	%	Peak 2 (dia.nm)	%	Peak 3 (dia.nm)	%	Z-Ave (dia.nm)
1	73,42	125,6	68	6,067	19,2	22,06	10,8	29,39
2	48,53	154,1	66	5,04	15,5	24,92	15,4	63,08
3	86,25	140,9	71	4,364	13,8	15,52	12,7	30,65

Figure FS4 TGA of air-dried crystalline TiBALDH sample

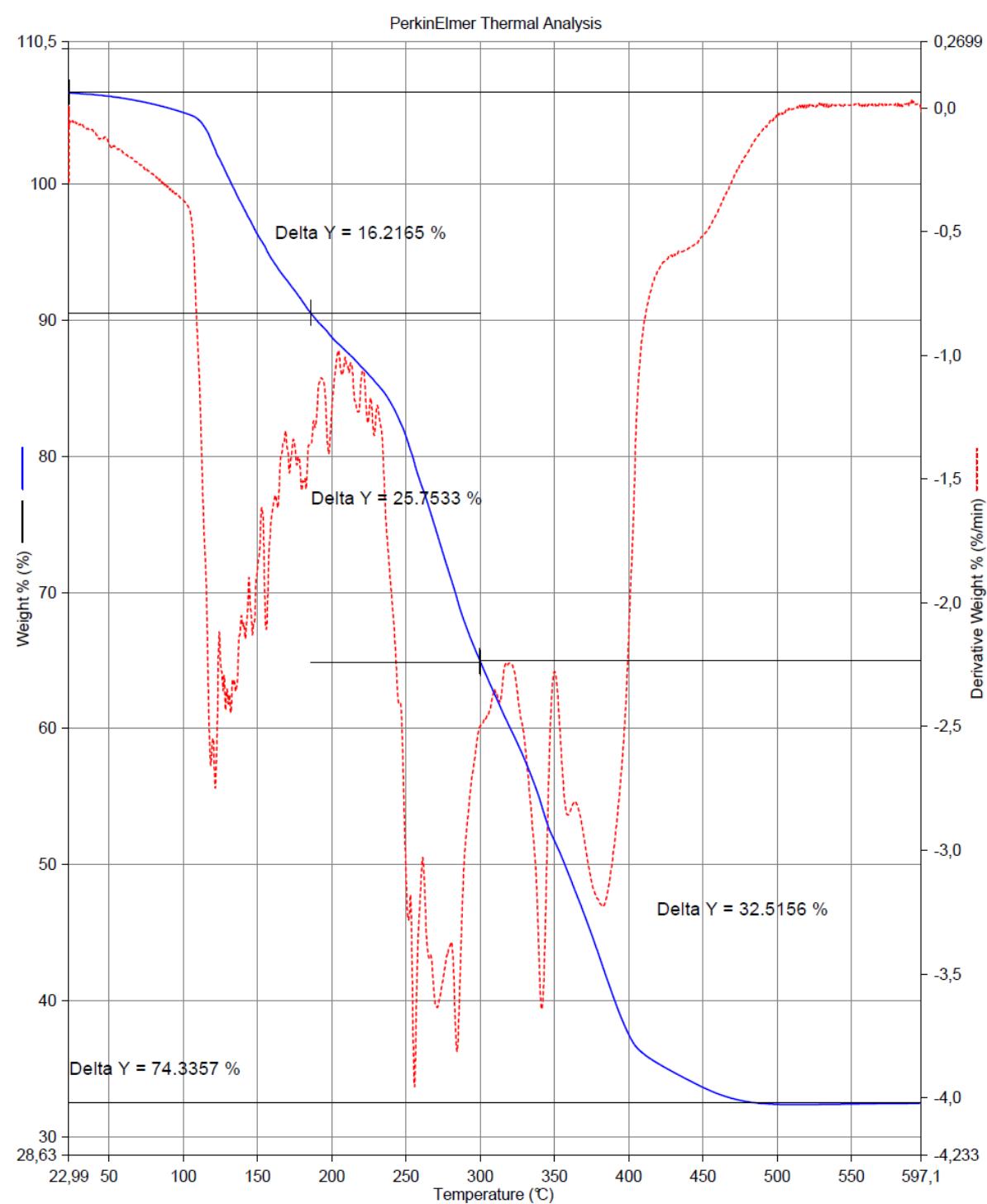
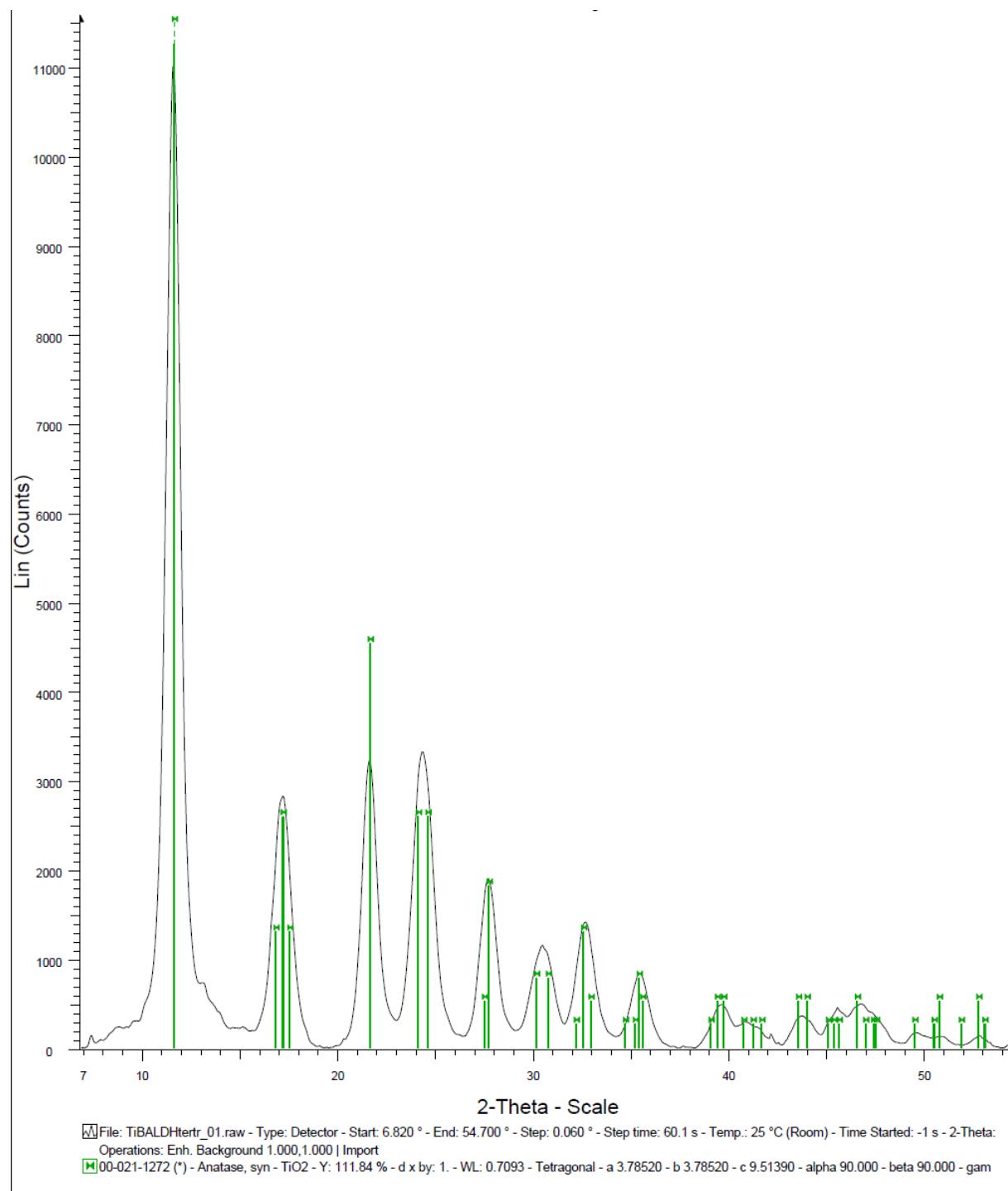


Figure FS5 X-ray powder pattern of the TiO₂ powder produced by heat treatment of TiBALDH at 500°C



Preparation of pollen suspension

A pollen suspension was prepared by collecting stamens of tobacco plants and putting them into a liquid medium. The pollen suspension was filtered through a colander to remove stamens and vacuum filtrated on a filter paper circles. Then filter papers were transferred onto a medium solidified with 0.5% phytagel in small Petri dishes. After removal of the filter paper circles, pollen remains on the surface of the solid medium. Pollen bombardment was performed immediately after plating.

Figure FS6 Working scheme for sample preparation in the shooting experiment:



Figure FS7 The shooting a quantified portion of nanoparticles suspension using Bio-rad instrument

