Electronic Supporting Information

A highly selective G-quadruplex-based luminescent switch-on probe for the detection of nanomolar strontium(II) ion in sea water

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Experimental section

Materials. Reagents were purchased from Sigma Aldrich and used as received. Iridium chloride hydrate (IrCl₃.xH₂O) was purchased from Precious Metals Online. All oligonucleotides were synthesized by Techdragon Inc. (Hong Kong, China)

DNA sequences:

	Sequence
T2	$5' - G_4 T_2 G_4 T_2 G_4 T_2 G_4 - 3'$
T2m1	5'-GCAGT ₂ GCAGT ₂ GCAGT ₂ GCAG-3'
T2m2	5'-GAAGT ₂ GAAGT ₂ GAAGT ₂ GAAG-3'

General experimental. Mass spectrometry was performed at the Mass Spectroscopy Unit at the Department of Chemistry, Hong Kong Baptist University, Hong Kong (China). Melting points were determined using a Gallenkamp melting apparatus and are uncorrected. Deuterated solvents for NMR purposes were obtained from Armar and used as received.

¹H and ¹³C NMR were recorded on a Bruker Avance 400 spectrometer operating at 400 MHz (¹H) and 100 MHz (¹³C). ¹H and ¹³C chemical shifts were referenced internally to solvent shift (CD₃CN: ¹H, δ 1.94, ¹³C δ 118.7; d₆-DMSO: ¹H δ 2.50, ¹³C δ 39.5). Chemical shifts (δ) are quoted in ppm, the downfield direction being defined as positive. Uncertainties in chemical shifts are typically ±0.01 ppm for ¹H and ±0.05 for ¹³C. Coupling constants are typically ± 0.1 Hz for ¹H-¹H and ±0.5 Hz for ¹H-¹³C couplings. The following abbreviations are used for convenience in reporting the multiplicity of NMR resonances: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet; br, broad. All NMR data was acquired and processed using standard Bruker software (Topspin). Emission spectra were recorded on a PTI QM4 spectrometer.

Synthesis of [Ir(ppy)₂(biq)]PF₆

The following complexes were prepared according to the reported literature method, the precursor complex $[Ir_2(ppy)_4Cl_2]$, and $[Ir(ppy)_2(biq)]PF_6(1)$, and characterized by ¹H-NMR, ¹³C-NMR and HRMS.

1 as molecular probe for oligonucleotide based Sr^{2+} detection

5 μ M of oligonucleotides (T2 or mutants) and indicated concentrations of Sr²⁺ ion were added into Tris-HCl buffer (20 mM Tris, pH 7.0). The solutions were heated to 95 °C for 10 min, and were cool to room temperature at 0.1 °C/s.

The pre-annealed solution was added into 400 μ L Tris-HCl buffer followed by addition of complex **1** at a final concentration of 2 μ M. Emission spectra were recorded in the 550–750 nm range, after equilibration at 25.0 °C for 10 min. Excitation wavelength = 390 nm.

Assay selectivity

5 μ M of oligonucleotides (T2) and 10 μ M of each ion (total 12 ions classified in three groups) were added into Tris-HCl buffer (20 mM Tris, pH 7.0) to make up a total volume of 100 μ L. The solutions were heated to 95 °C for 10 min, and were cool to room temperature at 0.1 °C/s.

The pre-annealed solution was added into 400 μ L Tris-HCl buffer followed by the addition of complex **1** at a final concentration of 2 μ M. Emission spectra were recorded in the 550–750 nm range, after equilibration at 25.0 °C for 10 min. Excitation wavelength = 390 nm.

Real sample analysis

Sea water samples collected from Lamma Island, Hong Kong were pretreated using a syringe filter to remove particulate impurities. Indicated concentrations of Sr^{2+} were then spiked into the water samples (100 µL). 5 µM of T2 was added into the sea water samples. The resulting solutions were heated to 95 °C for 10 min, and were cool to room temperature at 0.1 °C/s.

The pre-annealed water sample was added into 400 μ L Tris-HCl buffer followed by the addition of complex **1** at the final concentration of 2 μ M. Emission spectra were recorded in the 550–750 nm range, after equilibration at 25.0 °C for 10 min. Excitation wavelength = 390 nm.

References

- 1. M. S. Lowry, W. R. Hudson, R. A. Jr. Pascal and S. Bernhard, *J. Am. Chem. Soc.*, **2004**, *126*, 14129–14135.
- 2. H.-C. Su, H.-F. Chen, F.-C. Fang, C.-C. Liu, C.-C. Wu, K.-T. Wong, Y.-H. Liu and S.-M. Peng, J. Am. Chem. Soc., 2008, 130, 3413–3419.

Fig. S1 Photograph image of **1** and T2 (5 μ M) in Tris buffer (20 mM, pH 7.0) in the absence (left) or presence (right) of 10 μ M Sr²⁺ ions.



Fig. S2 Intensity of complex 1 (2 μ M) and T2 5 μ M) in the presence of Sr²⁺ ions (5 μ M) or 100-fold excess of common ions in sea water (K⁺, Ca²⁺, Na⁺ and Mg²⁺). Error bars represent the standard deviations of the results from three independent experiments.



Fig. S3 Relative luminescence intensity of complex **1** (2 μ M) in the presence of Sr²⁺ ions (0.1, 1, 5 and 10 μ M) and T2 or T2 mutants (5 μ M). Error bars represent the standard deviations of the results from three independent experiments.



Fig. S4 Sr^{2+} ion determination in sea water sample collected from Lamma Island, Hong Kong. Sr^{2+} ions (0.01, 0.1, 1, 3, 7, 10 and 20 μ M) were spiked into sea water samples, and the emission spectra were recorded with complex **1** (2 μ M) and T2 (5 μ M). Inset: Change in intensity at $\lambda = 638$ nm versus Sr^{2+} concentration in sea water samples. Error bars represent the standard deviations of the results from three independent experiments.



Fig. S5 Time course of luminescence response of the system which contains various concentrations of Sr^{2+} (0–10 μ M) and T2 (5 μ M) in diluted sea water sample upon addition of **1** (2 μ M) at 25 °C.



Table S1. Comparison of detection limit and range for some recently reported analytical techniques for Sr^{2+} ions. References refer to those in the main text.

Method	Detection limit	Range	Analyte	Ref.
Functionalized gold nanoparticles	7 μΜ	7–20 μM	$Ca^{2+}, Ba^{2+},$	8
			Sr^{2+}	
Potentiometric membrane sensor	240 nM	240 nM-100 μM	Sr^{2+}	10b

PVC-membrane sensor	550 nM	550 nM-100 μM	$\mathrm{Sr}^{2+},\mathrm{Cd}^{2+},\mathrm{Cu}^{2}$	10c
			+	
Ion-selective electrode	75 nM	75 nM–100 μM	Sr ²⁺	7b
G-quadruplex-based fluorescence assay	10 nM	10 nM-100 μM	Sr ²⁺	16
Ion-imprinted Au NPs composites and	20 fM	20 fM-100 pM	Alkaline-earth	9
SPR spectroscopy			ions	
Method in this study	13 nM	13 nM–10 μM	Sr^{2+}	