

Supporting Information

Controlled biosilification using self-assembled short peptides A₆K and V₆K

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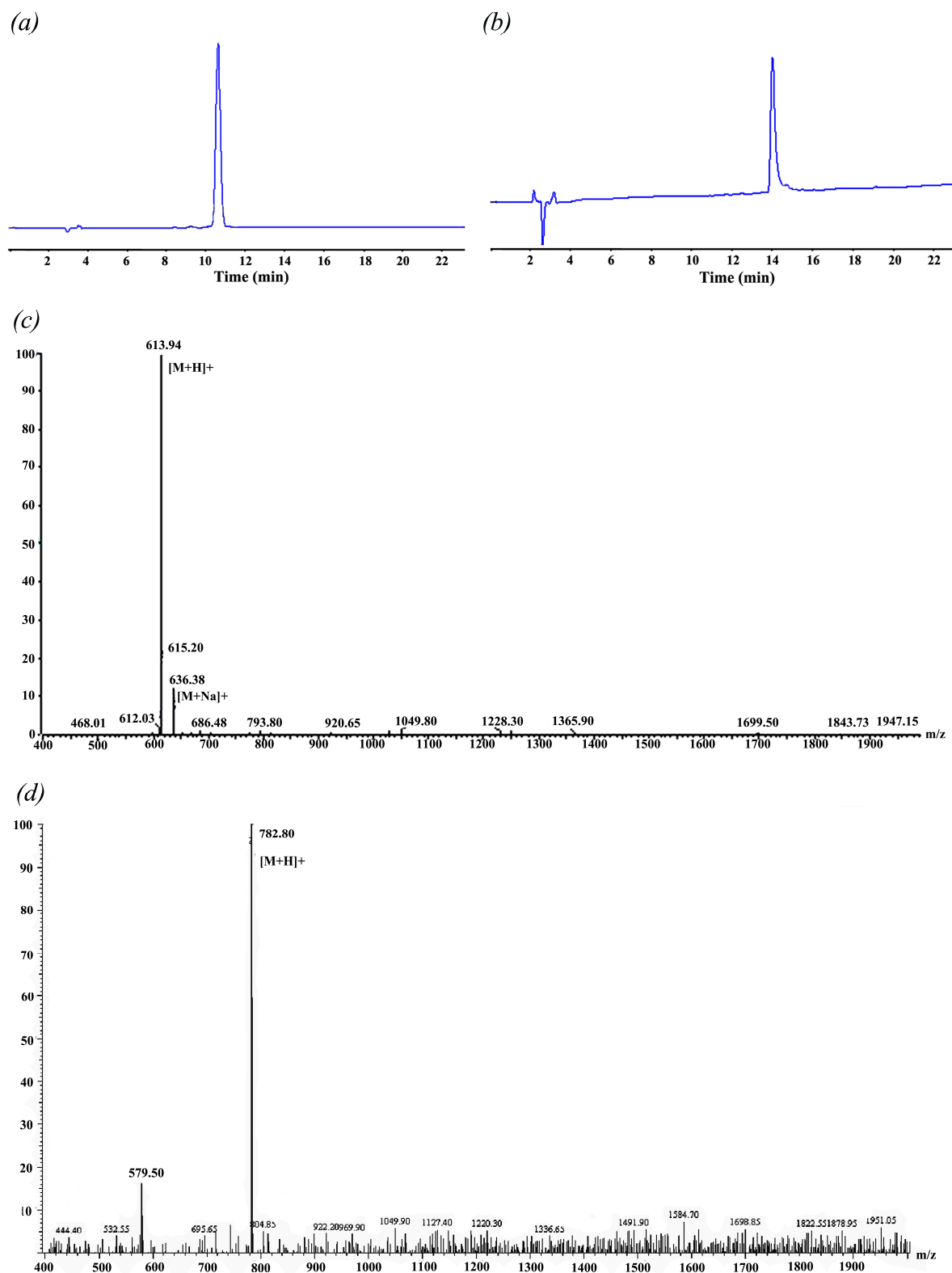


Fig. S1 HPLC spectra of (a) A₆K and (b) V₆K. ESI-MS spectra of (c) A₆K and (d) V₆K. The conditions for HPLC analysis of the two peptides are as follows: eluent A, 0.1% trifluoroacetic acid in acetonitrile, eluent B, 0.1% trifluoroacetic acid in water, 0→2 min, 95% (A%), 3→22 min, 95→40% (A%); UV detector, 214 nm; flow rate, 1.0 mL/min; column, VYDAC-C18, 4.6 mm × 250 mm.

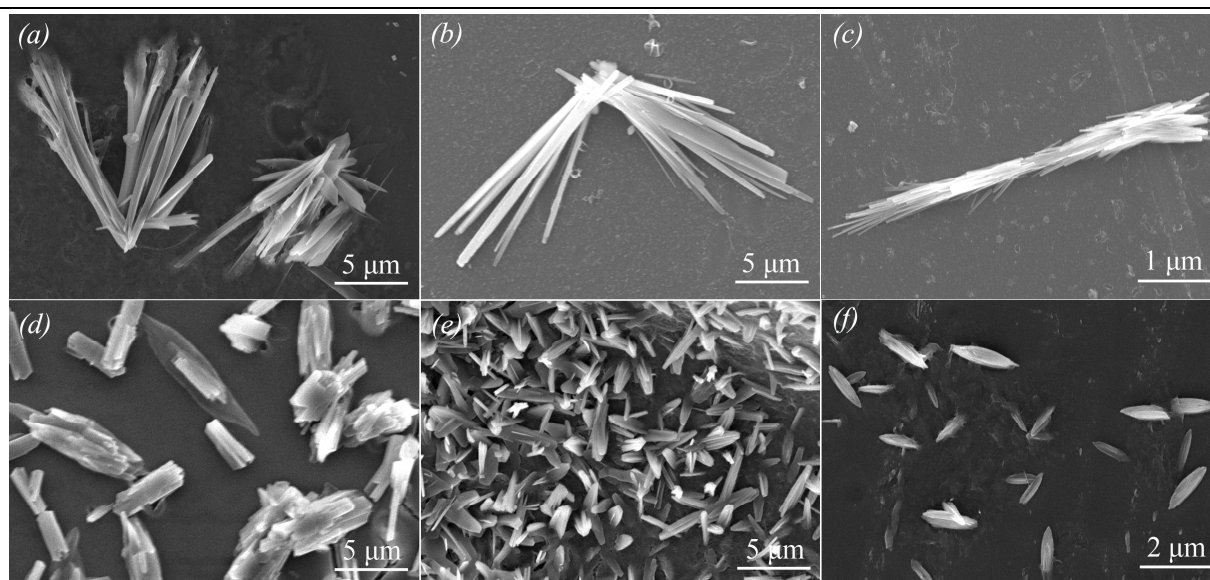


Fig. S2 SEM images of biosilica produced by A_6K using different TEOS concentration of (a) 100 mM, (b) 20 mM, and (c) 5 mM. SEM images of biosilica produced by V_6K using different TEOS concentration of (a) 100 mM, (b) 20 mM, and (c) 5 mM. The additive anion is phosphate.

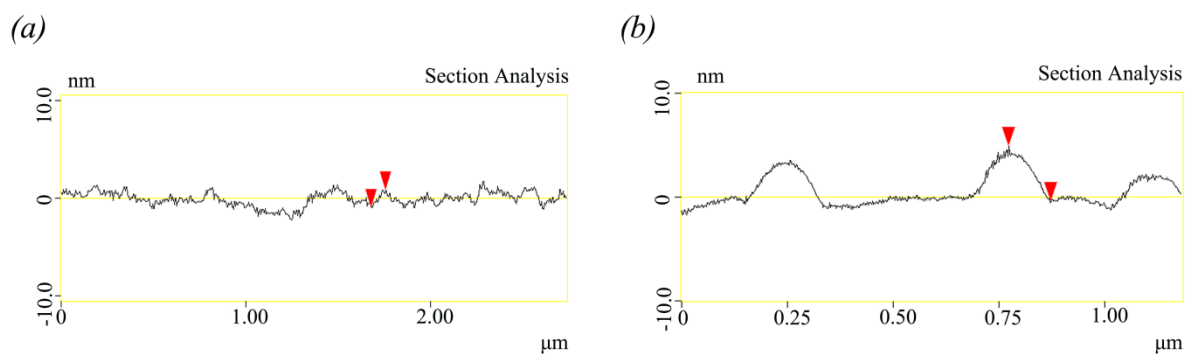


Fig. S3 Typical height sections of (a) A₆K nanofibers and (b) V₆K nanostacks, respectively.

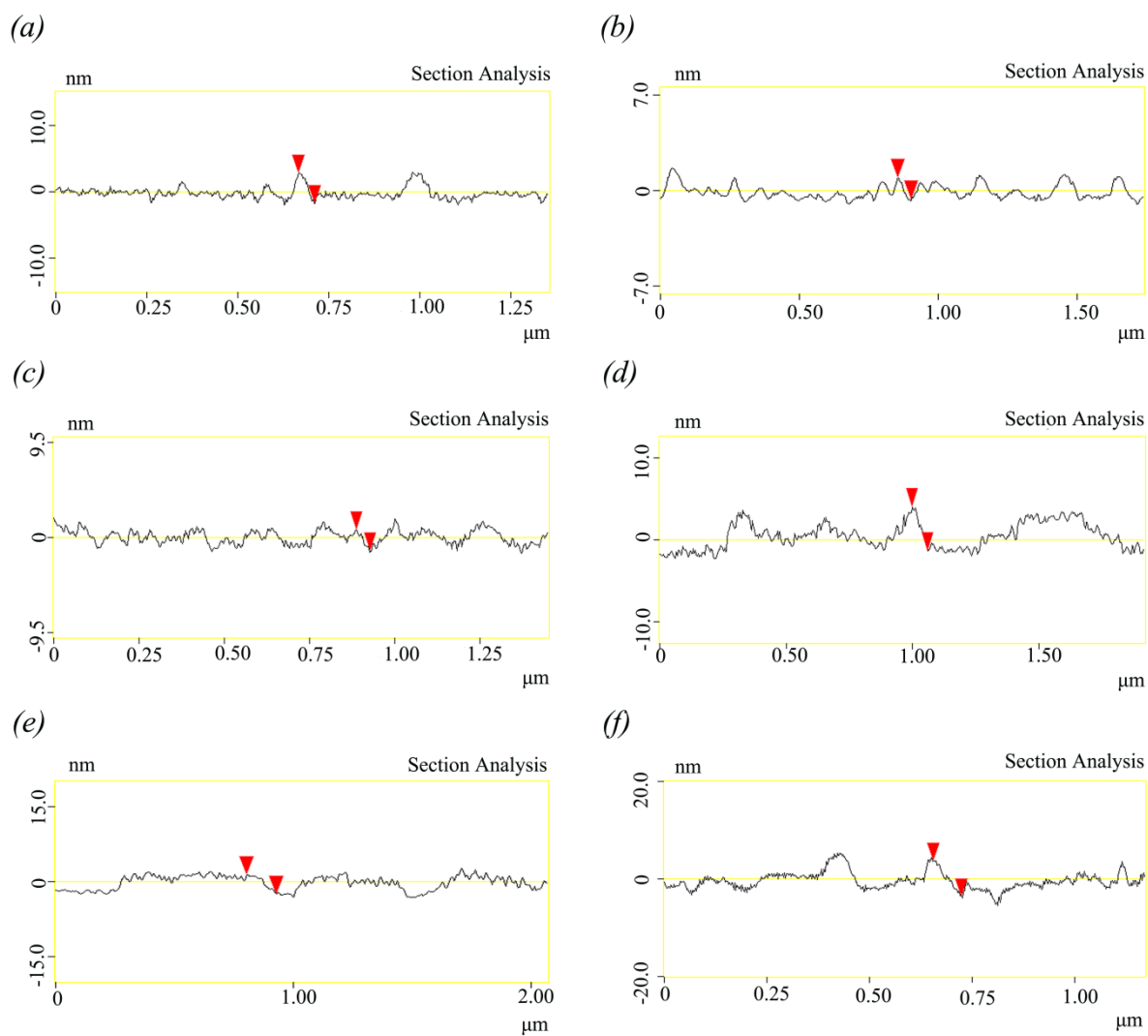


Fig. S4 Typical height sections of A_6K in the presence of (a) sodium carbonate, (b) sodium sulfate, and (c) sodium phosphate. Typical height sections of V_6K in the presence of (d) sodium carbonate, (e) sodium sulfate, and (f) sodium phosphate.

Table 1. Shifts in the negative CD peak from A₆K/V₆K solutions with different counterions with reference to the A₆K/V₆K peak without any added counterions

	A ₆ K/carbonate	A ₆ K/sulfate	A ₆ K/phosphate	V ₆ K/carbonate	V ₆ K/sulfate	V ₆ K/phosphate
peak shift (nm)	-7	-8	-10	1	1	2

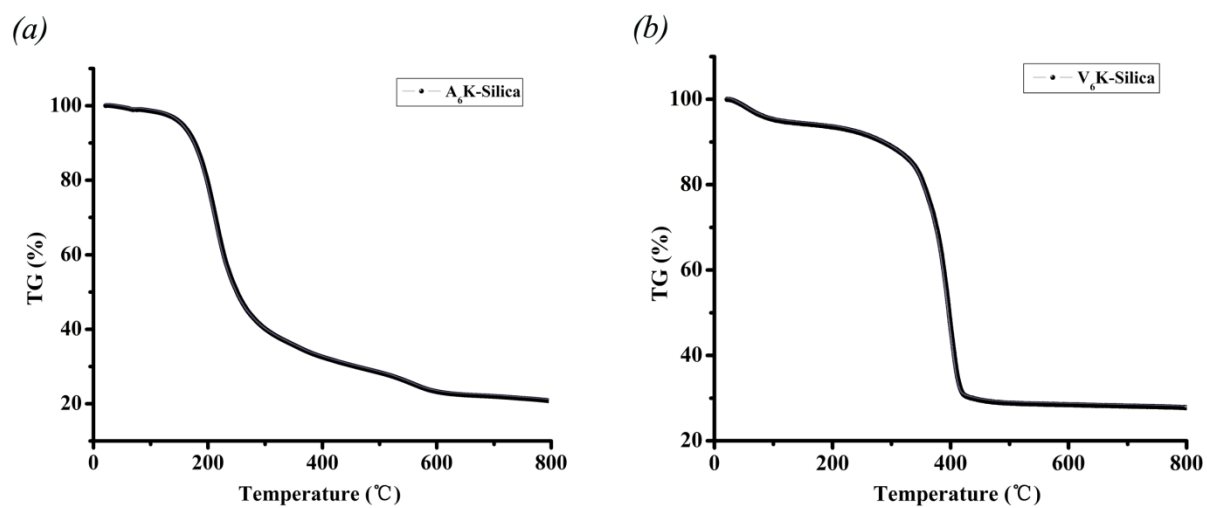


Fig. S5 Thermogravimetric analysis (TGA) spectrum of (a) A₆K-silica and (b) V₆K-silica composites, respectively.