

Supplementary Table 1 Environmental safety evaluation of ENMs after acute exposure in *C. elegans*

ENMs (size)	Dose	Duration	Adverse effects	Ref
CeO ₂ -NPs (8.5 nm)	1-100 nM	L1-larvae to adult	Decreased lifespan, increased intestinal lipofuscin accumulation, increased oxidative damage.	[1]
CeO ₂ -NPs (15, 30, 45 nm)		24-hr	Altered expression of <i>hsp-16.1</i> , <i>hsp-16.2</i> , <i>gst-4</i> , and <i>cyp35a2</i> , decreased egg number.	[2]
CeO ₂ -NPs (15 and 45 nm)		24-hr	Increased expression of <i>cyp35a2</i> , and decrease in fertility, and survival.	[3]
ZnO-NPs (1.5 nm)	325-1625 mg/L	4, 24, 72-hr	Increased lethality and <i>Pmtl-2::gfp</i> expression, and decreased brood size and movement.	[4]
ZnO-NPs (20 nm)	0.4-8.1 mg/L	24-hr, 5-day	Increased lethality, suppressed reproduction and growth.	[5]
ZnO-NPs (25 and 100 nm)			LC ₅₀ was 0.32 mg/L or 2 mg/L (25 nm or 100 nm)	[6]
TiO ₂ -NPs (7 and 20 nm)		24-hr	Increased expression of <i>cyp35a2</i> , and decrease in fertility, and survival.	[3]
TiO ₂ -NPs (50 nm)	24-239.6 mg/L	24-hr, 5-day	Increased lethality, suppressed reproduction and growth.	[5]
TiO ₂ -NPs (4, 10, 60, 90 nm)	0.001-10 µg/L	L1-larvae to day-1 adult	Decreased survival, growth, reproduction, locomotion behavior, and metabolism, and increased intestinal autofluorescence and ROS production.	[7]
TiO ₂ -NPs (25 and 100 nm)			LC ₅₀ was 77 mg/L (25 nm)	[6]
Al ₂ O ₃ -NPs (60 nm)	10.2-407.8 mg/L	24-hr, 5-day	Increased lethality, suppressed reproduction and growth.	[5]
Al ₂ O ₃ -NPs (60 nm)	6.3-203.9 mg/L	24-hr, L1 larvae-adult	Increases in lethality, stress response, and intestinal lipofuscin autofluorescence.	[8]
Al ₂ O ₃ -NPs (60 nm)	0.1-50000 µg/L	6 and 12-hr, L1-larvae to adult	Decreased locomotion behavior.	[9]
Ag-NPs (14-20 nm)	0.05-0.5 mg/L	24, 72-hr	Altered gene expression profile, and suppressed reproduction.	[10]

CIT-Ag-NPs (7 nm), PVP-Ag-NPs (21 nm), PVP-Ag-NPs (75 nm)	0.5-50 mg/L	24-hr	Growth inhibition.	[11]
Ag-NPs (20 nm)	0.05-0.5 mg/L	4, 24-hr	Increase in ROS formation, altered gene expression, and reduced reproduction.	[12]
Ag-NPs (1 nm), PVP-coated Ag-NPs (28 nm)	0.5-10 mg/L or 0.6-3 mg/L	24, 48, 72-hr	Increased lethality.	[13]
Citrate coated Ag-NPs (50.6 nm)	1-1000 mg/L	24, 48-hr	Decreased survival and reproduction.	[14]
CuO-NPs (<50 nm)	1-50 µg/mL	24-hr	Altered gene expression patterns.	[15]
DMSA-coated Fe ₂ O ₃ -NPs (9 nm)	0.001-100 mg/L	24-hr, L1-larvae to day-1 adult	Decreased survival, growth, reproduction, locomotion behavior, and metabolism, and increased intestinal autofluorescence and ROS production.	[16]
hydroxylated fullerene (4.7 and 40.1 nm)	1-100 µg/mL		Decreased survival ratio, shortened lifespan, and reduced reproduction rate and body size.	[17]
NaYF ₄ :Yb,Er	1-5 mg/mL	3, 24-hr	Reduced survival.	[18]
MSA-capped CdSe/ZnS	0.01-1 µM	3 days	Increased embryo mortality, defect in egg-laying, and reduced lifespan.	[19]
Citrate-coated Au-NPs	5.9 mg/L	4 nm	Different expression of 797 genes.	[20]

Supplementary Table 2 Environmental safety evaluation of ENMs after chronic exposure in *C. elegans*

ENMs (size)	Dose	Duration	Adverse effects	Ref
Silica-NPs (60 nm)	0.25-50 mg.mL	The full generation from day-1 adult	Reproductive senescence.	[11]
Al ₂ O ₃ -NPs (60 nm)	8.1-30.6 mg/L	10-days	Increase of intestinal autofluorescence, and formation of stress response and oxidative damage.	[21]
Al ₂ O ₃ -NPs (60 nm)	0.005-23.1 mg/L	10 days	Decreased locomotion behavior, and increased stress response and oxidative stress.	[22]
DMSA coated Fe ₂ O ₃ -NPs (9 nm)	1-5000 µg/L	L1-larvae to day-8 adult	Decreased survival, growth, reproduction, locomotion behavior, and metabolism, and increased intestinal autofluorescence and ROS production.	[16]
MPA-CdSe QDs (5-6 nm)	20 nM	16 days	Difficulty in egg laying, damaged egg left in the vulva and egg without an intact egg shell, and decreased growth, brood size and lifespan.	[23]

Supplementary Table 3 Comparative analysis of *C. elegans* based toxicities of TiO₂-NPs with those in other *in vitro* and *in vivo* assay systems

Organism	Size	Toxicity array used	Dose	Duration	Adverse effects	Ref.
Human pulmonary epithelial, cervical cancer, hepatocarcinoma, and monocytic leukemia cells	25 nm	Cytotoxicity and gene expression	75 mg/L	6 hr	Cell viability and induce expression of stress-related genes	[24]
Human skin keratinocytes	25 and 100 nm	Lipid and protein peroxidation	50 mg/L,	4 hr	Photocytotoxicity, cell membrane damage	[25]
Human embryonic kidney cell	25 nm	Oxidative stress and apoptosis	50 mg/L	24, 48 and 72 hr	Oxidative stress and apoptosis	[26]
Human hepatocarcinoma, lung epithelial, and monocytic leukemia cells	21 nm	Metabolic activation and cell death	10 mg/L	24, 48, 72 hr	Decreased metabolic activation, and increased cell death	[27]
Human hepatoblastoma cells	7 and 10 nm	Total glutathione, DCFH-DA, DNA strand breaks	10 mg/L	4 hr	DNA damage	[28]
Human keratinocytes cells	20 nm	Micronucleus, mitochondrial DNA damage, ROS	5 mg/L	24, 48, 72 hr	Cytotoxic and genotoxic	[29]
Murine fibroblast and pre-osteoblast cells	25 nm	Morphology, LDH assay, IL-6 and TNF-β assay, DNA fragmented	10 mg/L	2, 24, 48 hr	Cellular toxicity and inflammation	[30]
Murine pre-osteoblast s cells	5 and 32 nm	Cell viability, LDH assay, apoptosis, mitochondrial membrane permeability, gene expression	5 mg/L	24, 48, and 72 hr	Cytotoxicity	[31]
Murine microglia cells		Morphology, chromosome condensation, cell cycle	16 mg/L	24 hr	Apoptosis	[32]
Human	3.2 nm	Live viability, LDH assay,	100	50 hr	Cytotoxicity and	[33]

dermal fibroblast and lung epithelial cells		mitochondrial activity, IL-8 expression	mg/L		inflammatory response	
Human lymphocyte	100 nm	Comet assay, DNA laddering assay	0.25 mM	3 hr	Mitochondrial damage, genotoxic	[34]
Rat adrenal medulla pheochromocytoma	21 nm	Viability, ROS	10 mg/L	6, 12, 24 and 48 hr	Apoptosis	[35]
Non-small cell lung cancer cells	11-150 nm	Cell viability, necrosis and apoptosis, LDH assay	50 mg/L	1 hr	Cytotoxicity	[36]
Human monoblastoid cells	100 nm	Membrane permeability, DNA quantification	0.005 g/L	24 and 48 hr	Apoptotic and necrotic modifications	[37]
Human keratinocytes cells	25 nm	Lipid peroxidation, cell proliferation	10 mg/L	4 hr	Perturbs the cytoskeleton architecture, and inhibit cell growth	[38]
Bottlenose dolphin leukocytes	25 nm	Comet assay, cytotoxicity	50 mg/L	4, 24,48 hr	Genotoxic	[39]
Human keratinocytes cells	25 and 100 nm	ROS, cell toxicity	200 mg/L	4 hr	Mitochondria disruption	[40]
Mouse fibroblast cells	100 nm	ROS, SOD, LDH, GSH, survival, cell morphology	60 mg/L	24 and 48 hr	Cell morphology changed, apoptosis	[41]
Mice	100 nm	Pathological examination, monoamine neurotransmitter levels	500 µg/mice	30 day	Lesions on murine brain	[42]
Mice	33 nm	Comet assay, karyological assay	40 mg/kg	7 day	Genotoxicity	[43]
Mice	8 and 10.4 nm	Cells and total protein, TNF-α and IL-6 expression	5 µg/mice	24 hr and 30 day	Inflammation	[44]
Apolipoprotein E knockout (ApoE-/-) mice	12 and 21.6 nm	Plaque progression in aorta, vasodilatory function, gene expression levels	0.5 mg/kg	4 weeks	Increase in plaque progression in aorta	[45]
Mice	25 nm	Cytoplasmic aconitase, L-lactate dehydrogenase A chain, carbonic anhydrase 1, pyruvate kinase isoform M2	2.5 mg/mice	7 day	Induced toxicities evaluated at the protein level	[46]

		and peroxiredoxin 6, expression of heat shock protein, moesin and apolipoprotein A-1 precursor				
Mice	5-6 nm	Nephric inflammation, cell necrosis and dysfunction, expression of nucleic factor- κ B, tumor necrosis factor- α , macrophage migration inhibitory factor, interleukin-2, interleukin-4, interleukin-6, interleukin-8, interleukin-10, interleukin 18, interleukin-1 β , cross-reaction protein, transforming growth factor- β , interferon- γ and CYP1A1, and heat shock protein 70	2.5 mg/kg	90 day	Nephric injury	[47]
Mice	6.9 nm	Histopathological and biochemical analysis including ROS, lipid peroxidation, antioxidant enzyme activity, glutathione, and ascorbic acid	5 mg/kg	14 day	Nephrotoxicity, oxidative stress	[48]
Mice	21 nm	Maternal lung inflammation, gestational and litter parameters, offspring neurobehavioral alterations, and fertility	42 mg/m ³	8-18 day	Lung inflammation, offspring neurobehavioral alterations	[49]
Mice	5 nm	Expressions of inflammatory cytokines, histopathological changes, and hepatocytes apoptosis	5-150 mg/kg	14 day	Inflammatory responses, and liver injury	[50]
Mice	5 nm	Organs and serum biochemical parameters	5 mg/kg	14 day	Toxicity formed in liver	[51]
Mice	80-110 nm	Histopathology, passive behavior, loss of appetite, tremor, and lethargy	324 mg/kg	24 and 48 hr, 7 and 14 days	Hhepatocellular necrosis and apoptosis, hepatic fibrosis, renal glomerulus swelling and interstitial pneumonia	[52]
Mice	15, 50 and 100 nm	Expression of LPS, IL-1 β , MCP-1, and KC	8mg/kg	24 hr	Lung inflammation	[53]
Mice	5 nm	Norepinephrine, dopamine,	5	60 day	Impaired spatial	[54]

		dihydroxyphenylacetic acid, 5-hydroxytryptamine, acetylcholine, glutamate, and NO	mg/kg		recognition memory ability	
Rat	21 nm	LC50, microvascular oxidative stress, NO	5 µg/rat	24 hr	Microvascular dysfunction	[55]
Rat	13. and 42 nm	GSH, SOD, H ₂ O ₂ , TNF-α, and IL-1β	0.2 mg/kg	7 day	Oxidative damage, synovium hypotrophy and lymphocytes and plasma cells infiltration	[56]
Rat	5 nm	LDH, 8-isoprostanate	2 mg/m ³	6 hr	Inflammatory response	[57]
Rat	25 nm	Input/output (I/O) functions, pairedpulse reaction (PPR), field excitatory postsynaptic potential, population spike amplitude	100 mg/kg	2 and 21 days	Neurotoxicity	[58]
Larval zebrafish	25-70 nm	Hatch ability, survival, malformation, moving velocity and activity	0.1 mg/L	5 day	Behavioral toxicity	[59]
Zebrafish	30 nm	SOD, CAT, GSH, MDA, .OH	50 mg/L	7 day	Oxidative damage	[60]
Trout erythrocyte	10-150 nm	Hemolysis rate, NADH, comet assay	1.6 mg/L	10 and 60 min	DNA damage,	[61]
<i>D. magna</i>	21 nm	Survival	0.25 mM,	96 hr, 21 day	Mortality	[62]
<i>D. magna</i>	100 nm	Molting	0.73 mg/L	24, 48, 72, 96 hr	Molting inhibition	[63]
<i>E. fetida</i>	14-16 nm	Expression of metallothionein and superoxide dismutase, induction of apoptotic activity, phagocytosis	0.1 mg/L	24 hr	Altered antioxidant system	[64]
<i>E. fetida</i>	10-20 nm	Antioxidant enzymes, DNA damage, cellulase activity, mitochondria damage	1.0 g/kg dry soil	7 day	Mitochondrial damages, DNA damage	[65]
<i>Porcellio scaber</i>	15 nm	CAT and GST	0.5 µg/g food	3 days	Decreased antioxidant enzymes	[66]
<i>Lumbricus terrestris</i>	100-300 nm	Intestinal epithelium and chloragogenous	10 mg/kg soil	7 days	Apoptosis	[67]
<i>C. elegans</i>	30 nm	lethality, growth,	0.05-5	L1-larvae	decreased survival,	[68]

		reproduction, locomotion behavior, ROS production	0 µg/L	to day-1 adult	growth, reproduction, locomotion behavior, and increased ROS production	
<i>C. elegans</i>				24-hr	Increased expression of <i>cyp35a2</i> , and decrease in fertility, and survival.	[3]
<i>C. elegans</i>	50 nm	lethality, reproduction and growth	24-23 9.6 mg/L	24-hr, 5-day	Increased lethality, suppressed reproduction and growth.	[5]
<i>C. elegans</i>	4, 10, 60, 90 nm	survival, growth, reproduction, locomotion behavior, metabolism, intestinal autofluorescence, and ROS production.	0.001- 10 µg/L	L1-larvae to day-1 adult	Decreased survival, growth, reproduction, locomotion behavior, and metabolism, and increased intestinal autofluorescence and ROS production.	[7]
<i>C. elegans</i>	25 and 100 nm	survival			LC ₅₀ was 77 mg/L (25 nm)	[76]

References

- 1 H. Zhang, X. He, Z. Zhang, P. Zhang, Y. Liu, Y. Ma, Y. Kuang, Y. Zhao, Z. Chai, *Environ. Sci. Technol.*, 2011, **45**, 3725.
- 2 J. Roh, Y. Park, J. Choi, *J. Environ. Toxicol.*, 2008, **23**, 87.
- 3 J. Roh, Y. Park, K. Park, J. Choi, *Environ. Toxicol. Pharmacol.*, 2010, **29**, 167.
- 4 H. Ma, P. M. Bertsch, T. C. Glenn, N. J. Kabengi, P. L. Williams, *Environ. Toxicol. Chem.*, 2009, **28**, 1324.
- 5 H. Wang, R. L. Wick, B. Xing, *Environ. Pollut.*, 2009, **157**, 1171.
- 6 P. Khare, M. Sonane, R. Pandey, S. Ali, K. C. Gupta, A. Satish, *J. Biomed. Nanotechnol.*, 2011, **7**, 116.
- 7 Y. -X. Li, W. Wang, Q. -L. Wu, Y. -P. Li, M. Tang, B. -P. Ye, D. -Y. Wang, *PLoS ONE*, 2012, **7**, e44688.
- 8 S. Wu, J. -H. Lu, Q. Rui, S. -H. Yu, T. Cai, D. -Y. Wang, *Environ. Toxicol. Pharmacol.*, 2011, **31**, 179.
- 9 Y. -X. Li, S. -H. Yu, Q. -L. Wu, M. Tang, D. -Y. Wang, *Nanotoxicology*, 2012, doi: 10.3109/17435390.2012.689884.

- 10 J. Roh, S. J. Sim, J. Yi, K. Park, K. H. Chung, D. Ryu, J. Choi, *Environ. Sci. Technol.*, 2009, **43**, 3933.
- 11 J. N. Meyer, C. A. Lord, X. Y. Yang, E. A. Turner, A. R. Badireddy, S. M. Marinakos, A. Chilkoti, M. R. Wiesner, M. Auffan, *Aquat. Toxicol.*, 2010, **100**, 140.
- 12 D. Lim, J. Roh, H. Eom, J. Choi, J. Hyun, J. Choi, *Environ. Toxicol. Chem.*, 2012, **31**, 585.
- 13 L. Ellegaard-Jensen, K. A. Jensen, A. Johansen, *Ecotoxicol. Environ. Safety*, 2012, **80**, 216.
- 14 S. W. Kim, S. H. Nam, Y. J. An, *Ecotoxicol. Environ. Safety*, 2012, **77**, 64.
- 15 Y. Zhang, D. Chen, M. A. Smith, B. Zhang, X. Pan, *PLoS ONE*, 2012, **7**, e31849.
- 16 Q. -L. Wu, Y. -P. Li, M. Tang, D. -Y. Wang, *PLoS ONE*, 2012, **7**, e43729.
- 17 Y. J. Cha, J. Lee, S. S. Choi, *Chemosphere*, 2012, **87**, 49.
- 18 J. Chen, C. Guo, M. Wang, L. Huang, L. Wang, C. Mi, J. Li, X. Fang, C. Mao, S. Xu, *J. Mater. Chem.*, 2011, **21**, 2632.
- 19 P. L. Hsu, M. O'Callaghan, N. Al-Salim, M. R. H. Hurst, *Environ. Toxicol. Chem.*, 2012, **31**, 2366.
- 20 O. V. Tsyusko, J. M. Unrine, D. Spurgeon, E. Blaslock, D. Stames, M. Tseng, G. Joice, P. M. Bertsch, *Environ. Sci. Technol.*, 2012, **46**, 4115.
- 21 S. -H. Yu, Q. Rui, T. Cai, Q. -L. Wu, Y. -X. Li, D. -Y. Wang, *Environ. Toxicol. Pharmacol.*, 2011, **32**, 233.
- 22 Y. -X. Li, S. -H. Yu, Q. -L. Wu, M. Tang, Y. -P. Pu, D. -W. Wang, *J. Hazard. Mater.*, 2012, **219-220**, 221.
- 23 Y. Qu, W. Li, Y. Zhou, X. Liu, L. Zhang, Y. Li, A. Iida, Z. Tang, Y. Zhao, Z. Chai, C. Chen, *Nano Lett.*, 2011, **11**, 3174.
- 24 S.S. Mano, K. Kanehira, S. Sonezaki, A. Taniguchi, *Int. J. Mol. Sci.*, 2012, **13**, 3703.
- 25 J.J. Yin, J. Liu, M. Ehrenshaft, J.E. Roberts, P.P. Fu, R.P. Mason, B. Zhao, *Toxicol. Appl. Pharmacol.*, 2012, **263**, 81.
- 26 R. Meena, M. Rani, R. Pal, P. Rajamani, *Appl. Biochem. Biotechnol.*, 2012, **167**, 791.
- 27 A. Lankoff, W.J. Sandberg, A. Wegierek-Ciuk, H. Lisowska, M. Refsnes, B. Sartowska, P.E. Schwarze, S. Meczynska-Wielgosz, M. Wojewodzka, M. Kruszewski, *Toxicol. Lett.*, 2012, **208**, 197.
- 28 A. Kermanizadeh, B.K. Gaiser, G.R. Hutchison, V. Stone, *Part. Fibre Toxicol.*, 2012, **9**, 28.

- 29 A. Jaeger, D.G. Weiss, L. Jonas, R. Kriehuber, *Toxicology*, 2012, **296**, 27.
- 30 M.C. Bernier, K. El Kirat, M. Besse, S. Morandat, M. Vayssade, *Colloids Surf. B Biointerfaces*, 2012, **90**, 68.
- 31 Y. Zhang, W. Yu, X. Jiang, K. Lv, S. Sun, F. Zhang, *J. Mater. Sci. Mater. Med.*, 2011, **22**, 1933.
- 32 X. Li, S. Xu, Z. Zhang, H.J. Schluesener, *Chin. Sci. Bull.*, 2009, **54**, 3830.
- 33 C.M. Sayes, R. Wahi, P.A. Kurian, Y. Liu, J.L. West, K.D. Ausman, D.B. Warheit, V.L. Colvin, *Toxicol. Sci.*, 2006, **92**, 174.
- 34 M. Ghosh, M. Bandyopadhyay, A. Mukherjee, *Chemosphere*, 2010, **81**, 1253.
- 35 S. Liu, L. Xu, T. Zhang, G. Ren, Z. Yang, *Toxicology*, 2010, **267**, 172.
- 36 Y.S. Lee, S. Yoon, H.J. Yoon, K. Lee, H.K. Yoon, J.H. Lee, C.S. Song, *Toxicol. Lett.*, 2009, **189**, 191.
- 37 C.I. Vamanu, M.R. Cimpan, P.J. Høl, S. Sørnes, S.A. Lie, N.R. Gjerdet, *Toxicol. in Vitro*, 2008, **22**, 1689.
- 38 J. Chan, T. Ying, Y.F. Guang, L.X. Lin, T. Kai, Z.Y. Fang, Y.X. Ting, L.F. Xing, Y.Y. Ji, *Biol. Trace Elem. Res.*, 2011, **144**, 183.
- 39 M. Bernardeschi, P. Guidi, V. Scarcelli, G. Frenzilli, M. Nigro, *Anal. Bioanal. Chem.*, 2010, **396**, 619.
- 40 C. Jin, Y. Tang, F.G. Yang, X.L. Li, S. Xu, X.Y. Fan, Y.Y. Huang, Y.J. Yang, *Biol. Trace Elem. Res.*, 2011, **141**, 3.
- 41 C. Jin, B. Zhu, X. Wang, Q. Lu, *Chem. Res. Toxicol.*, 2008, **21**, 1871.
- 42 L. Zhang, R. Bai, B. Li, C. Ge, J. Du, Y. Liu, L. Le Guyader, Y. Zhao, Y. Wu, S. He, Y. Ma, C. Chen, *Toxicol. Lett.*, 2011, **207**, 73.
- 43 L.P. Sycheva, V.S. Zhurkov, V.V. Iurchenko, N.O. Daugel-Dauge, M.A. Kovalenko, E.K. Krivtsova, A.D. Durnev, *Mutat. Res.*, 2011, **726**, 8.
- 44 M. Roursgaard, K.A. Jensen, S.S. Poulsen, N.E. Jensen, L.K. Poulsen, M. Hammer, G.D. Nielsen, S.T. Larsen, *Sci. World J.*, 2011, **11**, 801.
- 45 L. Mikkelsen, M. Sheykhzade, K.A. Jensen, A.T. Saber, N.R. Jacobsen, U. Vogel, H. Wallin, S. Loft, P. Moller, *Part. Fibre Toxicol.*, 2011, **8**, 32.
- 46 Y.-M. Jeon, S.-K. Park, W.-J. Kim, J.-H. Ham, M.-Y. Lee, *Mol. Cell. Toxicol.*, 2011, **7**, 283.

- 47 S. Gui, Z. Zhang, L. Zheng, Y. Cui, X. Liu, N. Li, X. Sang, Q. Sun, G. Gao, Z. Cheng, J. Cheng, L. Wang, M. Tang, F. Hong, *J. Hazard. Mater.*, 2011, **195**, 365.
- 48 J. Zhao, N. Li, S. Wang, X. Zhao, J. Wang, J. Yan, J. Ruan, H. Wang, F. Hong, *J. Exp. Nanosci.*, 2010, **5**, 447.
- 49 K.S. Hougaard, P. Jackson, K.A. Jensen, J.J. Sloth, K. Loschner, E.H. Larsen, R.K. Birkedal, A. Vibenholt, A.M. Boisen, H. Wallin, U. Vogel, *Part. Fibre Toxicol.*, 2010, **7**, 16.
- 50 L. Ma, J. Zhao, J. Wang, J. Liu, Y. Duan, H. Liu, N. Li, J. Yan, J. Ruan, H. Wang, F. Hong, *Nanoscale Res. Lett.*, 2009, **4**, 1275.
- 51 H. Liu, L. Ma, J. Zhao, J. Liu, J. Yan, J. Ruan, F. Hong, *Biol. Trace Elem. Res.*, 2009, **129**, 170.
- 52 J. Chen, X. Dong, J. Zhao, G. Tang, *J. Appl. Toxicol.*, 2009, **29**, 330.
- 53 K. Inoue, H. Takano, M. Ohnuki, R. Yanagisawa, M. Sakurai, A. Shimada, K. Mizushima, T. Yoshikawa, *Int. J. Immunopathol. Pharmacol.*, 2008, **21**, 197.
- 54 R. Hu, X. Gong, Y. Duan, N. Li, Y. Che, Y. Cui, M. Zhou, C. Liu, H. Wang, F. Hong, *Biomaterials*, 2010, **31**, 8043.
- 55 T.R. Nurkiewicz, D.W. Porter, A.F. Hubbs, S. Stone, B.T. Chen, D.G. Frazer, M.A. Boegehold, V. *Toxicol. Sci.*, 2009, **110**, 191.
- 56 J.X. Wang, Y.-B. Fang, Y. Gao, Q.H. Hu, T.C. Wang, *Biomaterials*, 2009, **30**, 4590.
- 57 A. Noël, K. Maghni, Y. Cloutier, C. Dion, K.J. Wilkinson, S. Hallé, R. Tardif, G. Truchon, *Toxicol. Lett.*, 2012, **214**, 109.
- 58 X. Gao, S. Yin, M. Tang, J. Chen, Z. Yang, W. Zhang, L. Chen, B. Yang, Z. Li, Y. Zha, D. Ruan, M. Wang, *Biol. Trace Elem. Res.*, 2011, **143**, 1616.
- 59 T. Chena, C.-Y. Lin, M.C. Tseng, *Marine Pollut. Bull.*, 2011, **63**, 303.
- 60 D. Xiong, T. Fang, L. Yu, X. Sima, W. Zhu, *Sci. Total Environ.*, 2011, **409**, 1444.
- 61 D. Sekar, M.L. Falcioni, G. Barucca, G. Falcioni, *Environ. Toxicol.*, 2011, doi: 10.1002/tox.20778
- 62 S.H. Bang, T.H. Le, S.K. Lee, P. Kim, J.S. Kim, J. Min, *Environ. Health Toxicol.*, 2011, **26**, e2011002.
- 63 D. Andre, L. Duester, C. Prasse, F. Seitz, R. Rosenfeld, C. Schilde, G.E. Schaumann, R.

- Schulz, *PLoS ONE*, 2011, **6**, e20112.
- 64 E. Bigorgne, L. Foucaud, E. Lapiède, J. Labille, C. Botta, C. Sirguey, J. Falla, J. Rose, E.J. Joner, F. Rodius, J. Nahmani, *Environ. Pollut.*, 2011, **159**, 2698.
- 65 C.W. Hu, M. Li, Y.B. Cui, D.S. Li, J. Chen, L.Y. Yang, *Soil Biol. Biochem.*, 2010, **42**, 586.
- 66 A. Jemec, D. Drobne, M. Remskar, K. Sepčić, T. Tisler, *Environ. Toxicol. Chem.*, 2008, **27**, 1904.
- 67 E. Lapiède, J.Y. Nahmani, E. Moudilou, P. Chaurand, J. Labille, J. Rose, J. Exbrayat, D.H. Oughton, E.J. Joner, *Environ. Int.*, 2011, **37**, 1105.
- 68 Q.-L. Wu, A. Nouara, Y.-P. Li, M. Zhang, W. Wang, M. Tang, B.-P. Ye, J.-D. Ding, D.-Y. Wang, *Chemosphere*, 2013, **90**, 1123.