

Electronic Supporting Information (ESI):

Multifunctional BODIPY derivatives to image cancer cells and sense copper(II) ions in living cells.

Zan Li, Qiu-Yun Chen*, Pei-Dong Wang, Yi Wu

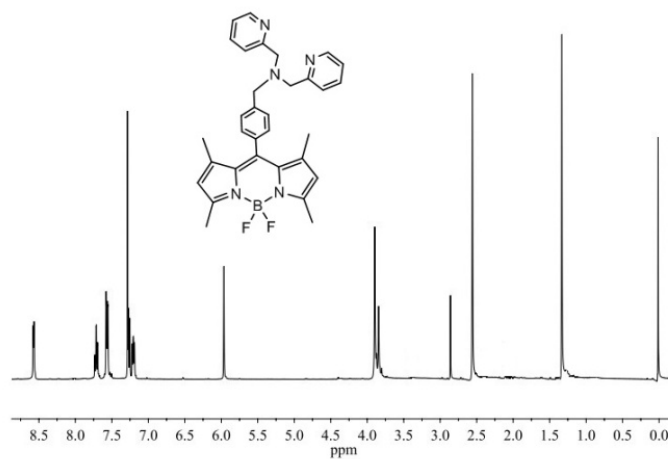


Fig. S1(a) ¹H NMR spectrum of 8-[di(2-picoly)amine-4-benzyl]-4,4-difluoro-1,3,5,7-tetramethyl-4-bora-3a,4a-diaza-s-indacene (**L1**) in chloroform (CDCl₃).

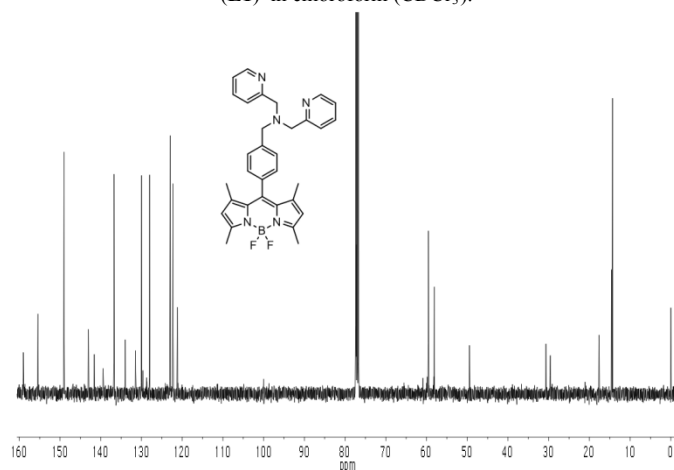


Fig. S1(b) ¹³C NMR spectrum of 8-[di(2-picoly)amine-4-benzyl]-4,4-difluoro-1,3,5,7-tetramethyl-4-bora-3a,4a-diaza-s-indacene (**L1**) in chloroform (CDCl₃).

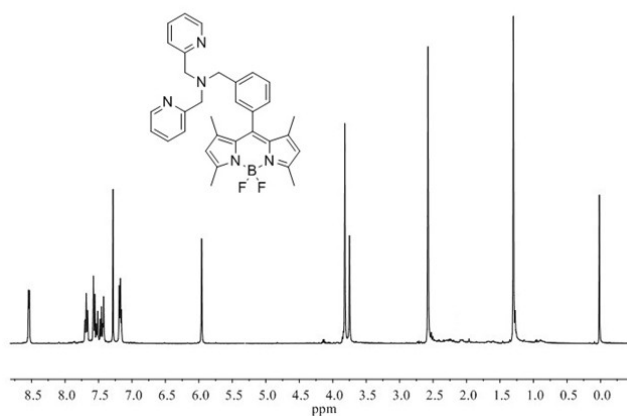


Fig.S2(a) ^1H NMR spectrum of 8-[di(2-picolyl)amine-3-benzyl]-4,4-difluoro-1,3,5,7-tetramethyl-4-bora-3a,4a-diaza-s-indacene (**L2**) in chloroform (CDCl_3).

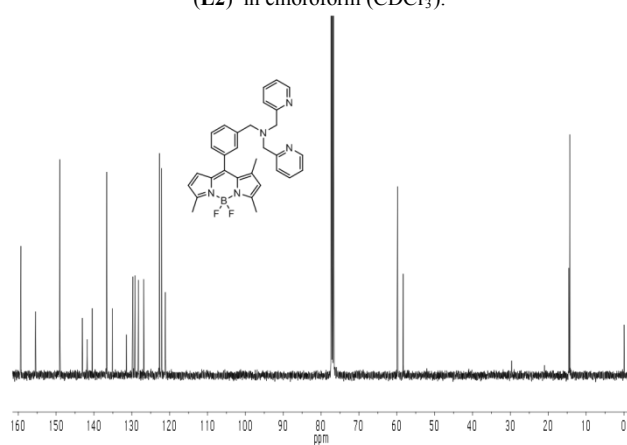


Fig.S2(b) ^{13}C NMR spectrum of 8-[di(2-picolyl)amine-3-benzyl]-4,4-difluoro-1,3,5,7-tetramethyl-4-bora-3a,4a-diaza-s-indacene (**L2**) in chloroform (CDCl_3).

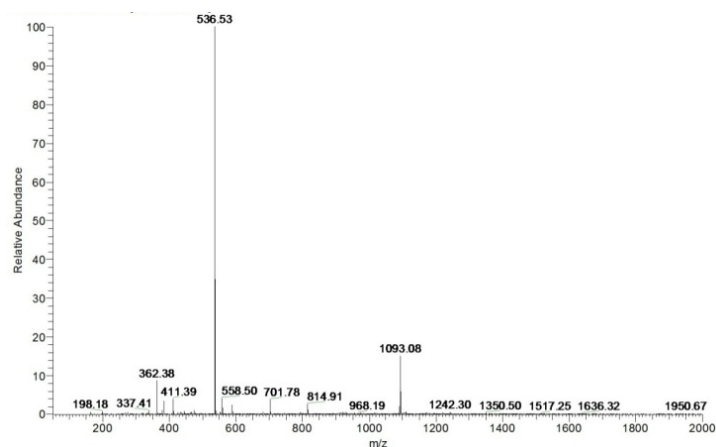


Fig. S3(a) ESI-MS spectrum of $[\text{L1}+\text{H}^+]$, $m/z=536.53$.

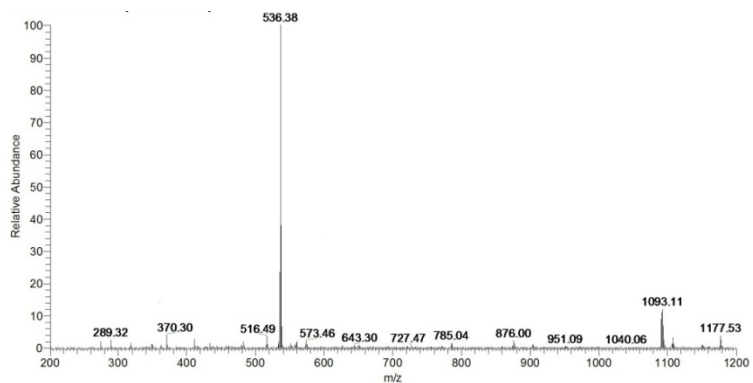


Fig. S3 (b) ESI-MS spectrum of $[L2+H^+]$, $m/z=536.38$.

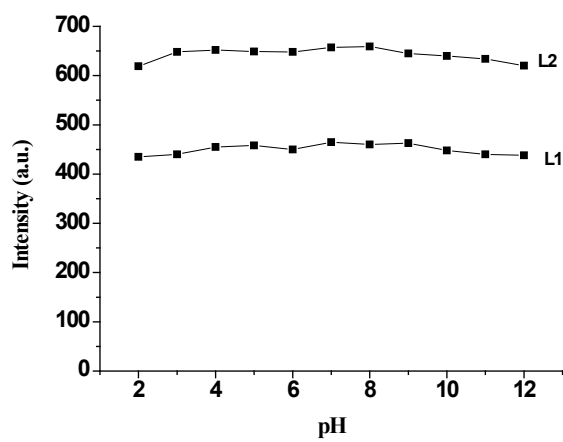


Fig. S4 Fluorescence intensities at 510 nm of L1 and L2 at various pH values. Tris-DMSO (pH 7.4, 9:1, v/v). $\lambda_{ex}=460$ nm. Slit width was 5 nm. All measurements were taken at 25 °C.

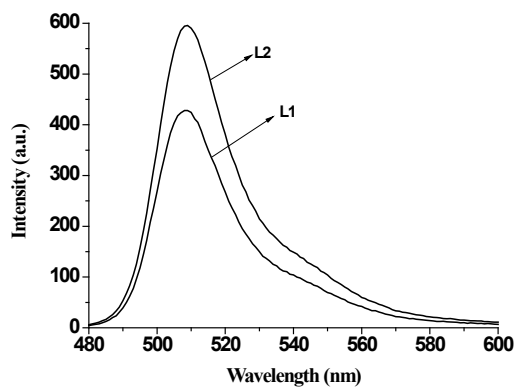


Fig. S5 Fluorescence spectra of probe L1, L2 (2 μ M) measured in Tris-DMSO (9:1, v/v) solution, $\lambda_{ex}=460$ nm. Measurements were taken at 25°C, Slit width was 5 nm.

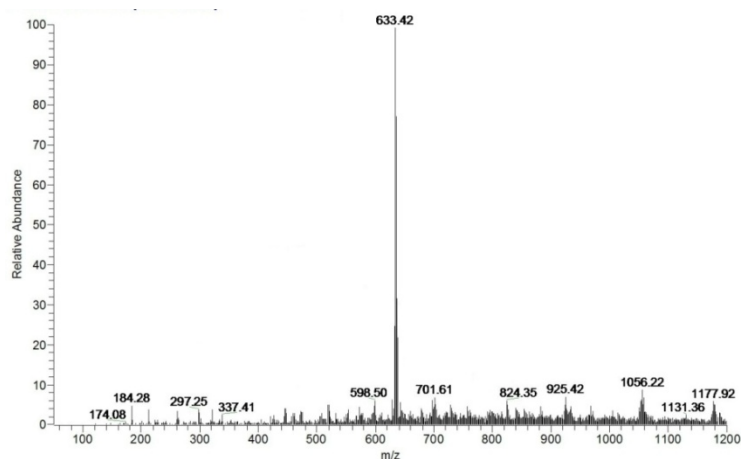


Fig. S6(a) ESI-MS spectrum of [L1+Cu²⁺+Cl⁻], m/z=633.42.

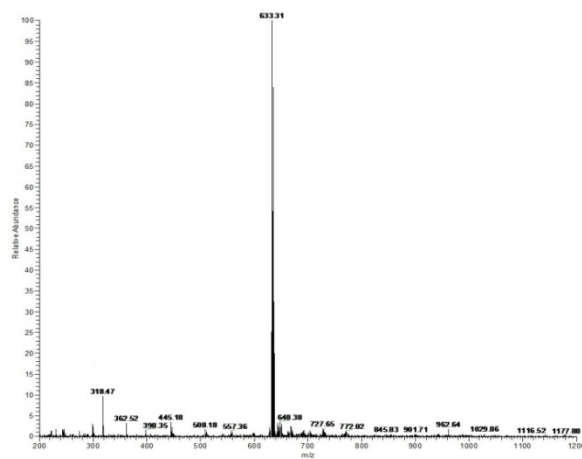


Fig. S6(b) ESI-MS spectrum of [L2+Cu²⁺+Cl⁻], m/z=633.31.

Determination of quantum yields [1]

Quantum yields were determined at 25°C, Fluorescein ($\Phi = 0.90$) in 0.1 M NaOH was used as a standard. The absorption of standard was adjusted to the same value ($abs < 0.1$, 491nm) as that of probe L. Excitation was chosen at 460 nm; the emission spectra were corrected and integrated from 480 to 600 nm. The quantum yields were calculated with the following equation:

$$U_{\text{sample}} = U_{\text{standard}} \times \frac{\int_{\text{emission sample}}}{\int_{\text{emission standard}}}$$

Association constant[2]

The association constant was calculated from the emission intensity-titration curves according to the equation:

$$I_F^0 / (I_F - I_F^0) = (1/f) [(1/K_s[M]) + 1]$$

Where I_F^0 is the emission intensity of probe L1 and L2 at 510 nm, I_F is the emission intensity of probe L at 510 nm upon the addition of different amount of Cu^{2+} , f is the fraction of the initial fluorescence which is accessible to the sensor, $[M]$ is the concentration of Cu^{2+} . The association constant values K_S is given by the ratio intercept/slope.

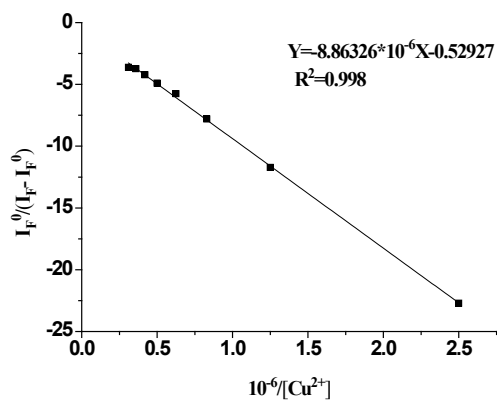


Fig. S7 Fitting of Fluorescence titration curve of L1 in Tris-DMSO (9:1, v/v). The association constant is $K_S = 5.97 \times 10^4 \text{ M}^{-1}$.

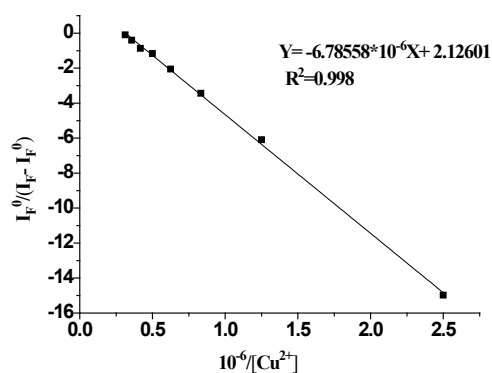


Fig. S8 Fitting of Fluorescence titration curve of L2 in Tris-DMSO (9:1, v/v). The association constant is $K_S = 3.13 \times 10^5 \text{ M}^{-1}$.

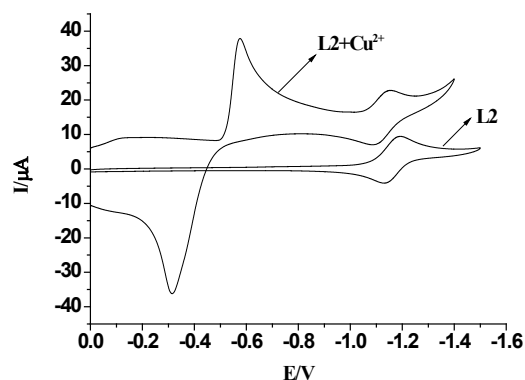


Fig.S9 Cyclic voltammogram of complex $[\text{L2}+\text{Cu}^{2+}]$ in MeCN + 0.1 M Bu_4NClO_4 . Scan rate: 100 mV S^{-1} .

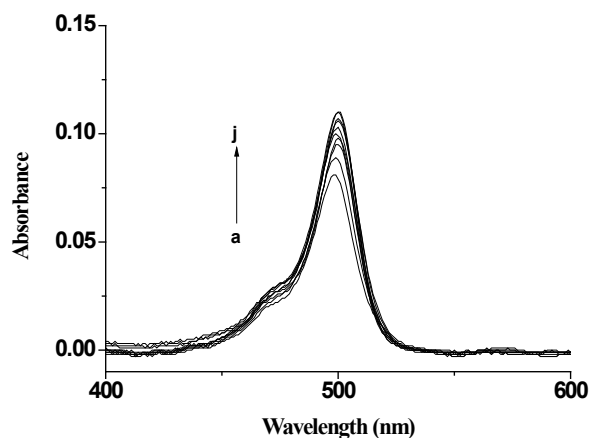


Fig. S10 UV-vis absorption spectra of probe **L2** (2 μM) in Tris-DMSO (9:1, v/v) solution with titration of Cu²⁺ (a-j: 0, 0.4, 0.8, 1.2, 1.6, 2.0, 2.4, 2.8, 3.2, 4.0 μM). All measurements were taken at 25°C, The data were recorded 1 min after Cu²⁺ was added.

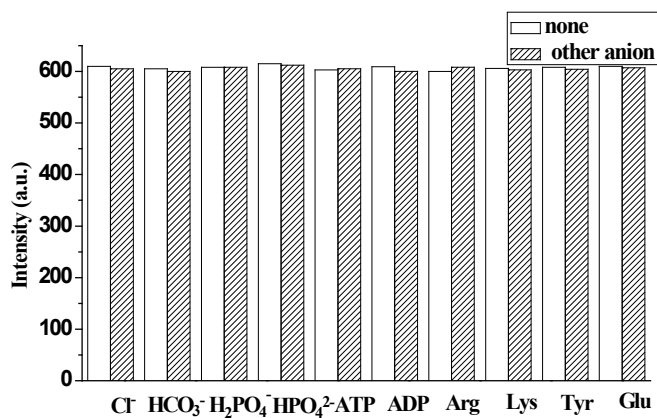


Fig. S11 Fluorescence of the chemosensor **L2** (2 μM) in the presence of important anions and molecules (2 μM) within cancer cells. Measured in Tris-DMSO (9:1, v/v) solution at 25°C, $\lambda_{\text{ex}}=460$ nm, Slit width was 5 nm.

Notes and references

1. J. N. Demas and G. A. Crosby, *J. Phys. Chem.*, 1971, **75**, 991-1024.
2. F.-P. Hou, J. Cheng, P.-X. Xi, F.-J. Chen, L. Huang, G.-Q. Xie, Y.-J. Shi, *Dalton Trans.*, 2012, **41**, 5799 -5804.