Electronic Supplementary Information

New hybrid materials with porphyrin-ferrocene and porphyrinpyrene covalently linked to single-walled carbon nanotubes.

Solon P. Economopoulos,^a Angeliki Skondra,^b Kalliopi Ladomenou,^b Nikolaos Karousis,^a Georgios Charalambidis,^b Athanassios G. Coutsolelos^{*b} and Nikos Tagmatarchis^{*a}

 ^a Theoretical and Physical Chemistry Institute National Hellenic Research Foundation 48 Vass. Constantinou Avenue, 11635 Athens, Hellas
^b Department of Chemistry, University of Crete, P O Box 1470, 71409 Heraklion Crete, Hellas.

Table of contents

| Experimental Details: | page 3 |
|--|---------|
| Synthesis of 5,15-bis(4-methoxyphenyl)-10,20-bis(2,4,6-trimethylphenyl)porphyrin 2 : | page 4 |
| Synthesis of 5,15-bis(4-hydroxyphenyl)-10,20-bis(2,4,6-trimethylphenyl)porphyrin 3 : | page 4 |
| Synthesis of mono 5-(4-hydroxyphenyl)-15-(4-(ferrocene-2-carbonyloxy)phenyl)- | |
| 10,20-bis(2,4,6-trimethylphenyl)porphyrin 4 : | page 5 |
| Synthesis of 5-(4-(2-(benzyloxy)-2-oxoethoxy)phenyl)-15-(4-(ferrocene-2-carbonyloxy) | |
| phenyl)-10,20-bis(2,4,6-trimethylphenyl)porphyrin 6: | page 5 |
| Synthesis of 5-(4-hydroxyphenyl)-15-(4-(pyrene-2-carbonyloxy)phenyl)-10,20- | |
| bis(2,4,6-trimethylphenyl)porphyrin 5 : | page 6 |
| Synthesis of 5-(4-(2-(benzyloxy)-2-oxoethoxy)phenyl)-15-(4-(pyrene-2-carbonyloxy) | |
| phenyl)-10,20-bis(2,4,6-trimethylphenyl)porphyrin 7: | page 7 |
| Fig. S1. ¹ H NMR spectrum of 2 in CDCl ₃ .: | page 8 |
| Fig. S2. ¹³ C NMR spectrum of 2 in CDCl ₃ : | page 9 |
| Fig. S3. ¹ H NMR spectrum of 3 in THF- d_8 : | page 10 |
| Fig. S4. ¹³ C NMR spectrum of 3 in THF- d_8 : | page 11 |
| Fig. S5. ¹ H NMR spectrum of 4 in CDCl ₃ : | page 12 |
| Fig. S6. ¹³ C NMR spectrum of 4 in CDCl ₃ : | page 13 |
| Fig. S7. ¹ H NMR spectrum of 6 in CDCl ₃ : | page 14 |
| Fig. S8. ¹³ C NMR spectrum of 6 in CDCl ₃ : | page 15 |
| Fig. S9. ¹ H NMR spectrum of H ₂ P-Fc 8 in CDCl ₃ : | page 16 |
| Fig. S10. ¹³ C NMR spectrum of 8 in CDCl ₃ : | page 17 |
| Fig. S11. ¹ H NMR spectrum of 5 in CDCl ₃ : | page 18 |
| Fig. S12. ¹³ C NMR spectrum of 5 in CDCl ₃ : | page 19 |
| Fig. S13. ¹ H NMR spectrum of 7 in CDCl ₃ : | page 20 |
| Fig. S14. ¹³ C NMR spectrum of 7 in CDCl ₃ : | page 21 |
| Fig. S15. ¹ H NMR spectrum of 9 in CDCl ₃ : | page 22 |
| Fig. S16. ¹³ C NMR spectrum of 9 in CDCl ₃ : | page 23 |
| Fig. S17. ATR-IR spectra of SWCNT-(H ₂ P-Fc) 11 (red) and SWCNT-(H ₂ P-pyr) 12 | |
| (blue): | page 24 |
| Fig. S18. Raman spectra of pristine SWCNTs (black), Boc-functionalized SWCNTs (gray | y), |
| SWCNT-(H ₂ P-Fc) 11 (red) and SWCNT-(H ₂ P-pyr) 12 (blue): | page 25 |

Experimental Details

All solvents and reagents were purchased from Aldrich and used without further purification unless otherwise stated. Organic extracts were dried over magnesium sulphate unless indicated otherwise. Evaporation of the solvents was accomplished on a rotary evaporator. Thin layer chromatography was performed on silica gel 60 F₂₅₄ plates. Chromatography refers to flash chromatography and was carried out on SiO₂ (silica gel 60, SDS, 70-230 mesh ASTM). ¹H and ¹³C NMR spectra were recorded unless otherwise specified, as deuteriochloroform solutions using the solvent peak as internal standard on a Bruker AMX-500 MHz and on a Bruker DPX-300 MHz spectrometers. High resolution mass spectra were performed on a Bruker ultraflextreme MALDI-TOF/TOF spectrometer. Steady-state UV-Vis electronic absorption spectra were recorded on a Perkin Elmer (Lambda 19) UV-VIS-NIR spectrophotometer or on a Shimadzu Multispec-1501 instrument. Mid-infrared spectra in the region 550-4000 cm⁻¹ were obtained on a Fourier Transform IR spectrometer (Equinox 55 from Bruker Optics) equipped with a single reflection diamond ATR accessory (DuraSamp1IR II by SensIR Technologies). Micro-Raman scattering measurements were performed at room temperature in the backscattering geometry using a RENISHAW inVia Raman microscope equipped with a CCD camera and a Leica microscope. A 2400 lines mm⁻¹ grating was used for all measurements, providing a spectral resolution of ± 1 cm⁻¹. As an excitation source the Ar⁺ laser (514 nm with less than 0.5 mW laser power) was used. Measurements were taken with 60 seconds of exposure times at varying numbers of accumulations. The laser spot was focused on the sample surface using a long working distance 50x objective. Raman spectra were collected on numerous spots on the sample and recorded with Peltier cooled CCD camera. The intensity ratio I_D/I_G was obtained by taking the peak intensities following any baseline corrections. The data were collected and analyzed with Renishaw Wire and Origin software. Thermogravimetric analysis was performed using a TGA Q500 V20.2 Build 3 instrument by TA in an inert atmosphere of nitrogen. In a typical experiment, 1 mg of the material was placed in the sample pan and the temperature was equilibrated at 60 °C. Subsequently, the temperature was increased to 800 °C at a rate of 10 °C min⁻¹ and the weight changes were recorded as a function of temperature. Fluorescence spectra were taken on an Aminco Bowman spectrofluorimetre (Spectronocs Co., USA) and а Fluorolog-3 Jobin Yvon-Spex spectrofluorometer (model GL3-21). Picosecond time-resolved fluorescence spectra were measured by the time-correlated single-photon counting (TCSPC) method on a NanoLog spectrofluorometer (Horiba Jobin Yvon), using a laser diode as an excitation source (NanoLED, 441 nm, 200 ps pulse width). Lifetimes were evaluated with the DAS6 Fluorescence-Decay

Analysis Software. All measurements were performed at room temperature. Electrochemistry was conducted on a Princeton Applied Research potentiostat/galvanostat model 2273 connected to a personal computer running Power Suite. The measurements were conducted using in solution using 0.1M tetrabutylammonium hexafluorophosphate 0.1M in DMF as supporting electrolyte, on a standard 3 electrode cell, with Pt disk (1.6mm diameter) as a working electrode, Pt gauze 52 mesh, 99.9% and a Ag/AgNO₃ reference electrode (Ag wire in 0.1M AgNO₃ in dry acetonitrile). Experiments were conducted in inert atmosphere using N₂ and data calibrated with ferrocene. Specifically, for the differential pulse voltammograms of materials **8** and **10** the following parameters were applied: a) reduction run: Pulse Height 0.05V, Pulse Width 50ms, Step Height 5mV and Step Time 100 ms, and b) oxidation run: Pulse Height 0.05V, Pulse Width 50ms, Step Height 5mV and Step Time 100ms, Step Height 5mV and Step Time 100ms.

Synthesis of 5,15-bis(4-methoxyphenyl)-10,20-bis(2,4,6-trimethylphenyl)porphyrin 2.

of 4-methoxy-benzaldehyde (0.32 mL, 2.6 mmol) А solution and 2,2'-(2,4,6trimethylphenylmethylene)bis(1H-pyrrole) 1 (0.7 g, 2.6 mmol) in 275 mL of dichloromethane was purged with nitrogen for 5 min. Then trifluoroacetic acid (0.37 mL, 4.9 mmol) was added and the solution stirred at room temperature for 45 min. 2,3-Dichloro-5,6-dicyano-1,4-benzoquinone (DDQ, 0.9 g, 3.9 mmol) was added and the solution stirred for a further 1.5 h. The mixture was filtered through an alumina (Al₂O₃) column (CH₂Cl₂). The product was eluted with dichloromethane to give 2 as a purple solid (177 mg, 18 %). ¹H NMR (500 MHz, CDCl₃): δ 8.85 (d, J = 5.0 Hz, 4H), 8.71 (d, J = 4.5 Hz, 4H), 8.16 (d, J = 7.5 Hz, 4H), 7.30 (m, 8H), 4.10 (s, 6H),2.65 (s, 6H), 1.87 (s, 12H), -2.56 (s, 2H); ¹³C NMR (125 MHz, CDCl₃): δ 159.8, 139.8, 139.0, 138.1, 135.9, 134.8, 131.8, 130.3, 128.2, 119.5, 118.6, 112.6, 56.0, 22.1, 21.9; UV/vis (CH₂Cl₂) λ_{max} , nm (ε , mM⁻¹ cm⁻¹) 420 (473.4), 516 (18.8), 552 (9.0), 592 (5.7), 649 (5.2); HRMS (MALDI-TOF) calcd for $C_{52}H_{47}N_4O_2 [M+H]^+$ 759.3699, found 759.3691.

Synthesis of 5,15-bis(4-hydroxyphenyl)-10,20-bis(2,4,6-trimethylphenyl)porphyrin 3

To a solution of porphyrin **2** (100 mg, 0.13 mmol) in dry CH_2Cl_2 (100 mL) was added boron tribromide (6.0 mL, 60.0 mmol) and the solution was stirred at room temperature under nitrogen for 2 days. Then, CH_2Cl_2 (100 mL) was added and the mixture was cooled down to 0 ^{0}C for 30 min. After that time MeOH (70 mL) and water (140 mL) were added and left stirring fo further 30

min. The resulting solution was washed with saturated NaHCO₃ (3 x 50 mL) and then with H₂O (2 x 50 mL). The organic layer dried, filtered, concentrated and the residue was chromatographed on a silica gel column (CH₂Cl₂/THF). The product was eluted with THF to obtain **3** as a purple solid (85 mg, 89 %). ¹H NMR (300 MHz, THF-*d*₈): δ 8.85 (d, *J* = 4.5, 4H), 8.79 (s, 2H), 8.62 (d, *J* = 4.8 Hz, 4H), 8.00 (d, *J* = 8.4 Hz, 4H), 7.30 (s, 4H), 7.15 (d, *J* = 8.4, 4H), 2.60 (s, 6H), 1.84 (s, 12H); ¹³C NMR (75 MHz, THF-*d*₈): δ 159.0, 140.3, 140.0, 138.8, 136.6, 134.0, 132.4, 130.5, 128.9, 121.0, 118.8, 114.8, 22.0, 21.8; UV/vis (toluene) λ_{max} , nm (ε , mM⁻¹ cm⁻¹) 420 (199.5), 515 (10.9), 550 (6.9), 592 (5.7), 650 (5.1); HRMS (MALDI-TOF) calcd for C₅₀H₄₃N₄O₂ [M+H]⁺ 731.3386, found 731.3391.

Synthesis of 5-(4-hydroxyphenyl)-15-(4-(ferrocene-2-carbonyloxy)phenyl)-10,20-bis(2,4,6-trimethylphenyl)porphyrin 4

To a solution of ferrocene carboxylic acid (12.6 mg, 0.05 mmol) in dry CH₂Cl₂ (1 mL) oxalyl chloride (7 µl, 0.08 mmol) was added and the solution was stirred under nitogen at 40 0 C for 1h. Then the solvent and excess oxalyl chloride were removed and the mixture was dried under vacum for 1 h, to yield the ferrocene carboxylic acid chloride, which was dissolved in dry THF (4 mL). Porphyrin **3** (40 mg, 0.05 mmol) and triethylamine (0.13 mL, 0.9 mmol) were added and the solution was stirred overnight at 70 0 C. After removal of the solvent the residue was obtained by chromatography (CH₂Cl₂). First the bis-ferrocene porphyrin was eluted and then the desired product **4** obtained as a purple solid (19 mg, 40 %). ¹H NMR (500 MHz, CDCl₃): δ 8.89 (d, *J* = 4.5 Hz, 2H), 8.84 (d, *J* = 4.5 Hz, 2H), 8.74 (d, *J* = 4.5 Hz, 2H), 8.71 (d, *J* = 4.5 Hz, 2H), 8.30 (d, *J* = 8.0 Hz, 2H), 8.08 (d, *J* = 8.0 Hz, 2H), 7.61 (d, *J* = 8.5 Hz, 2H), 7.30 (s, 4H), 7.16 (d, *J* = 8.5 Hz, 2H), 5.15 (s, 2H), 4.61 (s, 2H), 4.44 (s, 5H), 2.65 (s, 6H), 1.87 (s, 12H), -2.57(s, 2H); ¹³C NMR (125 MHz, CDCl₃): δ 171.2, 155.9, 151.2, 139.8, 138.9, 138.2, 136.1, 135.8, 134.9, 131.8, 130.6, 128.2, 120.4, 119.6, 118.8, 118.6, 114.1, 72.6, 71.2, 70.5, 22.1, 21.9; UV/vis (CH₂Cl₂) λ_{max} , nm (ε , mM⁻¹ cm⁻¹) 419 (446.1), 447 (24.2), 515 (17.8), 550 (7.5), 591 (5.4), 647 (4.0); HRMS (MALDI-TOF) calcd for C₆₁H₅₁FeN₄O₃ [M+H]⁺ 943.3311, found 943.3305.

Synthesis of 5-(4-(2-(benzyloxy)-2-oxoethoxy)phenyl)-15-(4-(ferrocene-2-carbonyloxy) phenyl)-10,20-bis(2,4,6-trimethylphenyl)porphyrin 6.

To a solution of porphyrin **4** (36 mg, 0.04 mmol) in dry DMF (3 mL) K_2CO_3 (7 mg, 0.05 mmol) and benzylacetylbromide (9 µl, 0.05 mmol) were added and the mixture was stirred under nitrogen at 80 0 C overnight. Then CH₂Cl₂ (50 mL) was added and extracted with water (4 x 50

mL), dried, filtered and concentrated under vacuum. The resulting crude product was purified by column chromatography (CH₂Cl₂). The product was eluted with dichloromethane to obtain **6** as a purple solid (33 mg, 76 %). ¹H NMR (500 MHz, CDCl₃): δ 8.85 (d, *J* = 4.5 Hz, 2H), 8.79 (d, *J* = 4.5 Hz, 2H), 8.72 (t, *J* = 4.5 Hz, 4H), 8.26 (d, *J* = 8.1 Hz, 2H), 8.12 (d, *J* = 8.4 Hz, 2H), 7.58 (d, *J* = 8.1 Hz, 2H), 7.44 (m, 5H), 7.29 (s, 6H), 5.39 (s, 2H), 5.14 (s, 2H), 4.98 (s, 2H), 4.62 (s, 2H), 4.45 (s, 5H), 2.67 (s, 6H), 1.89 (s, 12H), -2.57 (s, 2H); ¹³C NMR (75 MHz, CDCl₃): δ 170.6, 168.9, 157.6, 150.8, 139.4, 139.3, 138.4, 137.7, 135.5, 135.3, 135.2, 131.4, 130.0, 128.7, 128.6, 128.5, 128.4, 127.8, 120.0, 118.8, 118.3, 118.2, 117.1, 112.9, 72.1, 71.6, 70.1, 70.0, 67.2, 65.7, 21.6, 21.5; UV/vis (CH₂Cl₂) λ_{max} , nm (ε , mM⁻¹ cm⁻¹) 419 (440.3), 447 (22.1), 515 (14.9), 550 (6.5), 591 (4.6), 647 (3.2); HRMS (MALDI-TOF) calcd for C₇₀H₅₉FeN₄O₅ [M+H]⁺ 1091.3835, found 1091.3825.

Synthesis of 5-(4-hydroxyphenyl)-15-(4-(pyrene-1-carbonyloxy)phenyl)-10,20-bis(2,4,6trimethylphenyl)porphyrin 5

To a solution of pyrene carboxylic acid (33 mg, 0.13 mmol) in dry CH₂Cl₂ (2.0 mL) oxalyl chloride (18 µl, 0.21 mmol) was added and the solution was stirred under nitogen at 40 °C for 1h. Then the solvent and excess oxalvl chloride were removed and the mixture was dried under vacum for 1 h, to yield the pyrene carboxylic acid chloride, which was dissolved in THF (3.5 mL). Porphyrin 3 (100 mg, 0.13 mmol) and triethylamine (0.13 mL, 0.9 mmol) were added and the solution was stirred overnight at 70 °C. After removal of the solvent the residue was obtained by chromatography (CH₂Cl₂/Hexane, 7:3). First the bis-pyrene porphyrin was eluted and then the desired product 5 obtained as a purple solid (66 mg, 53 %). ¹H NMR (500 MHz, CDCl₃): δ 9.62 (d, J = 9.5 Hz, 1H), 9.13 (d, J = 8.0 Hz, 1H), 8.98 (d, J = 4.5 Hz, 2H), 8.87 (d, J = 4.5 Hz, 2H),8.79 (d, J = 4.5 Hz, 2H), 8.75 (d, J = 4.5 Hz, 2H), 8.38 (m, 6H), 8.28 (d, J = 9.0 Hz, 1H), 8.20 (d, J = 9.0 Hz, 1H), 8.14 (t, J = 7.5 Hz, 1H), 8.09 (d, J = 8.0 Hz, 2H), 7.82 (d, J = 8.0 Hz, 2H), 7.33 (s, 4H), 7.15 (d, J = 8.5 Hz, 2H), 2.67 (s, 6H), 1.90 (s, 12H), -2.52 (s br, 2H); ¹³C NMR (125 MHz. CDCl₃): δ 167.0, 155.8, 151.5, 140.2, 139.9, 138.9, 138.2, 136.1, 135.9, 135.6, 134.9, 132.6, 131.5, 130.9, 130.7, 130.6, 129.7, 128.2, 127.7, 127.2, 127.1, 127.0, 128.6, 125.3, 124.8, 124.6, 122.6, 120.7, 119.7, 118.8, 118.6, 114.1, 22.1, 21.9; UV/vis (CH₂Cl₂) λ_{max} , nm (ε , mM⁻¹ cm⁻¹) 284 (32.4), 357 (33.8), 419 (356.1), 515 (15.1), 550 (6.5), 591 (4.7), 647 (3.7); HRMS (MALDI-TOF) calcd for $C_{67}H_{51}N_4O_3$ [M+H]⁺ 959.3961, found 959.3957.

Synthesis of 5-(4-(2-(benzyloxy)-2-oxoethoxy)phenyl)-15-(4-(pyrene-1-carbonyloxy) phenyl)-10,20-bis(2,4,6-trimethylphenyl)porphyrin 7

To a solution of porphyrin 5 (40 mg, 0.04 mmol) in dry DMF (3 mL), K₂CO₃ (8 mg, 0.06 mmol) and benzylacetylbromide (10 µl, 0.06 mmol) were added and the mixture was stirred under nitrogen at 80 °C overnight. Then CH₂Cl₂ (50 mL) was added and extracted with water (4 x 50 mL), dried, filtered and concentrated under vacuum. The resulting crude product was purified by column chromatography (CH₂Cl₂:Hex 8/2). The product 7 was eluted as a purple solid (43 mg, 97 %). ¹H NMR (500 MHz, CDCl₃): δ 9.60 (d, J= 9.5 Hz, 1H), 9.12 (d, J= 8.0 Hz, 1H), 8.95 (d, J= 4.5 Hz, 2H), 8.83 (d, J = 4.5 Hz, 2H), 8.76 (d, J = 4.5 Hz, 2H), 8.73 (d, J = 5.0 Hz, 2H), 8.37 (m, 6H), 8.29 (d, J = 9.0 Hz, 1H), 8.20 (d, J = 9.0 Hz, 1H), 8.15 (m, 3H), 7.80 (d, J = 8.5 Hz, 2H), 7.48 (d, J = 7.5 Hz, 2H), 7.43 (t, J = 7.5 Hz, 2H), 7.38 (d, J = 7.5 Hz, 1H), 7.32 (s, 4H), 7.29 (d, J = 8.0Hz, 2H), 5.40 (s, 2H), 4.98 (s, 2H), 2.67 (s, 6H), 1.89 (s, 12H), -2.56 (s br. 2H); ¹³C NMR (125 MHz, CDCl₃): δ 169.4, 166.9, 158.1, 151.5, 140.1, 139.8, 138.9, 138.2, 136.0, 135.9, 135.6, 135.6, 132.6, 131.5, 130.9, 130.7, 130.6, 129.6, 129.2, 129.1, 129.0, 128.2, 127.7, 127.1, 127.0, 127.0, 125.5, 125.3, 124.8, 124.6, 122.6, 120.7, 119.4, 118.8, 118.6, 113.4, 67.6, 66.2, 22.1, 21.9; UV/vis $(CH_2Cl_2) \lambda_{max}$, nm (ϵ , mM⁻¹ cm⁻¹) 284 (30.2), 357 (31.1), 419 (346.1), 515 (13.9), 550 (5.1), 591 (3.2), 647 (3.0); HRMS (MALDI-TOF) calcd for C₇₆H₅₉N₄O₅ [M+H]⁺ 1107.4485, found 1107.4492.



Fig. S1. ¹H NMR spectrum of 2 in CDCl₃.



Fig. S2. ¹³C NMR spectrum of 2 in CDCl₃.



Fig. S3. ¹H NMR spectrum of **3** in THF- d_8 .



Fig. S4. ¹³C NMR spectrum of **3** in THF- d_8 .



Fig. S5. ¹H NMR spectrum of 4 in CDCl₃.



Fig. S6. ¹³C NMR spectrum of 4 in CDCl₃.



Fig. S7. ¹H NMR spectrum of 6 in CDCl₃.



Fig. S8. ¹³C NMR spectrum of 6 in CDCl₃.



Fig. S9. ¹H NMR spectrum of H₂P-Fc 8 in CDCl₃.



Fig. S10. ¹³C NMR spectrum of H₂P-Fc 8 in CDCl₃.



Fig. S11. ¹H NMR spectrum of 5 in CDCl₃.



Fig. S12. ¹³C NMR spectrum of 5 in CDCl₃.



Fig. S13. ¹H NMR spectrum of 7 in CDCl₃.



Fig. S14. ¹³C NMR spectrum of 7 in CDCl₃.



Fig. S15. ¹H NMR spectrum of H₂P-Fc **9** in CDCl₃.



Fig. S16. ¹³C NMR spectrum of H₂P-pyr 9 in CDCl₃.



Fig. S17. ATR-IR spectra of SWCNT-(H₂P-Fc) 11 (red) and SWCNT-(H₂P-pyr) 12 (blue).



Fig. S18. Raman spectra of pristine SWCNTs (black), Boc-functionalized SWCNTs (gray), SWCNT-(H₂P-Fc) **11** (red) and SWCNT-(H₂P-pyr) **12** (blue), using 514 nm laser excitation. All intensities are normalized versus the G-band (1591 cm⁻¹) of the pristine SWCNTs.