

Electronic Supplementary Information (ESI)

Learn from T-Hg-T and C-Ag-T: Four-input Dual-core Molecular Logic Gate and Its

New Application in Cryptography

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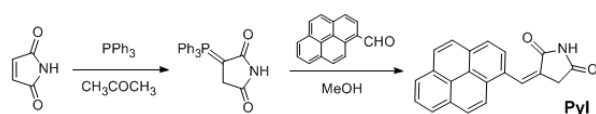
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Material & Instrumentation

Maleimide was purchased from Energy-Chemical (China). PPh_3 and Benzaldehyde were purchased from GuangFu (China). 1-Formylpyrene was purchased from TCI. Quinine sulphate was purchased from Alfa Aesar. All other common reagents and solvent were received from commercial sources and used without further purification. ^1H and ^{13}C NMR spectra were measured in $\text{DMSO-}d_6$ on Bruker-Avance (400 MHz for ^1H and 101 MHz for ^{13}C) with TMS as an internal reference. HPLC-HRMS were measured with Bruker MicrOTOF QII. UV-vis absorption spectra were obtained using a Varian-Cary50. All the fluorescence measurements were acquired on a Varian-FLR006 and Edinburgh Instrument FLS920.

Synthesis of Pyl



A mixture of maleimide (116 mg, 1.2 mmol), PPh_3 (315 mg, 1.2 mmol), and acetone (3 ml) was heated under reflux for 2 h. After cooling, the precipitate was filtered and washed with acetone (20 ml), then dried in vacuo and got a colorless solid (308 mg, 0.86 mmol, yield 72%). It was used for the next step without further purification and mixed with 1-Formylpyrene (198 mg, 0.86 mmol) and methanol (5 ml). The mixture was heated under reflux for 3 h, then cooled at room temperature. The resulting precipitate was filtered, washed with methanol (20 mL) and dried in vacuo to give a yellow power **PyI**: 204 mg, 76% yield. Metamorphosed into brown over 260 °C. ^1H NMR (400 MHz, DMSO) δ 11.56 (s, 1H), 8.52 – 8.19 (m, 9H), 8.13 (t, $J = 7.6$ Hz, 1H), 3.78 (s, 2H). ^{13}C NMR (101 MHz, DMSO) δ 175.85, 171.80, 131.57, 130.78, 130.17, 129.50, 129.27, 128.62, 128.53, 128.09, 127.81, 127.29, 126.63, 126.27, 126.10, 125.91, 125.03, 123.95, 123.66, 122.64, 34.76. HRMS ($\text{M}+\text{H}^+$): Cald. For $\text{C}_{21}\text{H}_{14}\text{NO}_2$ 312.1019; Found: 312.1014.

Synthesis of PhI

This imide derivative was synthesized according to the literature method.¹

Synthesis of PhI-Ag-PhI

PhI (187 mg, 1.0 mmol) was dissolved in 5 mL methanol / DMF. Triethylamine (TEA) (1 equiv.) and AgNO_3 solution (85 mg, 0.5 mmol, in DMF) were then added. The resulting precipitate was filtered, washed with methanol / H_2O and dried in vacuo to give a white power **PhI-Ag-PhI**: 52 mg, yield 22%.

Method for making Pyl fluorescent paper

The filter papers were soaked in the ethanol solution of 10 mM **PyI**. After 0.5 h, they were removed out of solution and dried at room temperature.

(1) (a) Y. Luo, L. Ma, H. Zheng, L. Chen, R. Li, C. He, S. Yang, X. Ye, Z. Chen, Z. Li, Y. Gao, J. Han, G. He, L. Yang and Y. Wei, *J. Med. Chem.*, 2010, **53**, 273; (b) H. M. Albers, L. J. Hendrickx, R. J. van Tol, J. Hausmann, A. Perrakis and H. Ovaa, *J. Med. Chem.*, 2011, **54**, 4619.

Emission spectra of PyI in EtOH / H₂O mixture

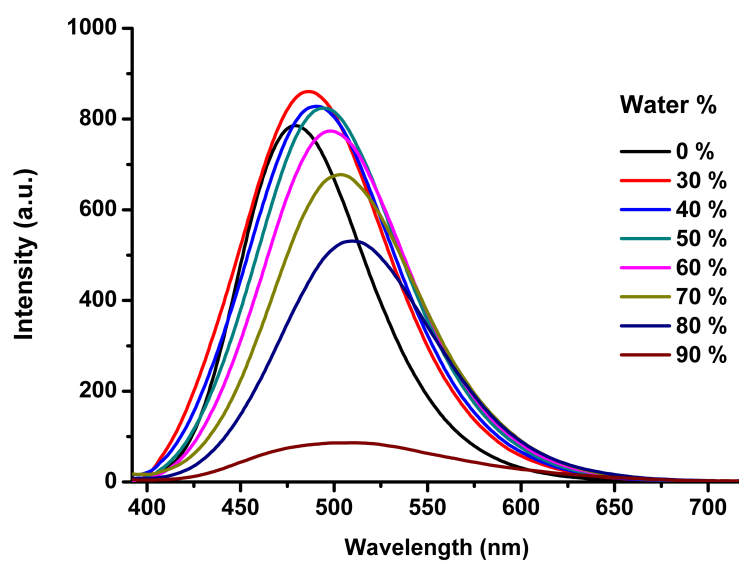


Fig. S1 Emission spectra of the **PyI** (5 μ M) in EtOH and 30~90 % water in EtOH.

Photo of **PyI** in the solid and EtOH solution under UV lamp

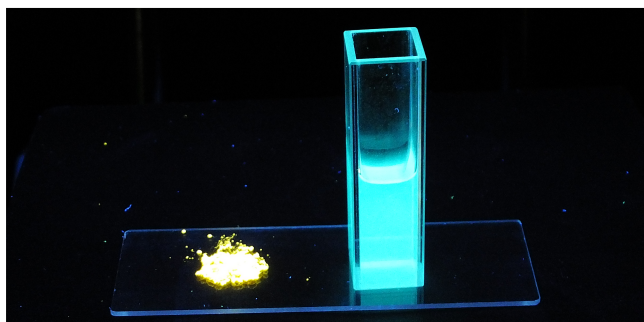


Fig. S2 Fluorescence of **PyI** under UV lamp (365 nm) (left) in the solid (right) 5 μ M in EtOH solution.

Quenching effect by Hg^{2+} and Ag^+ in different pH

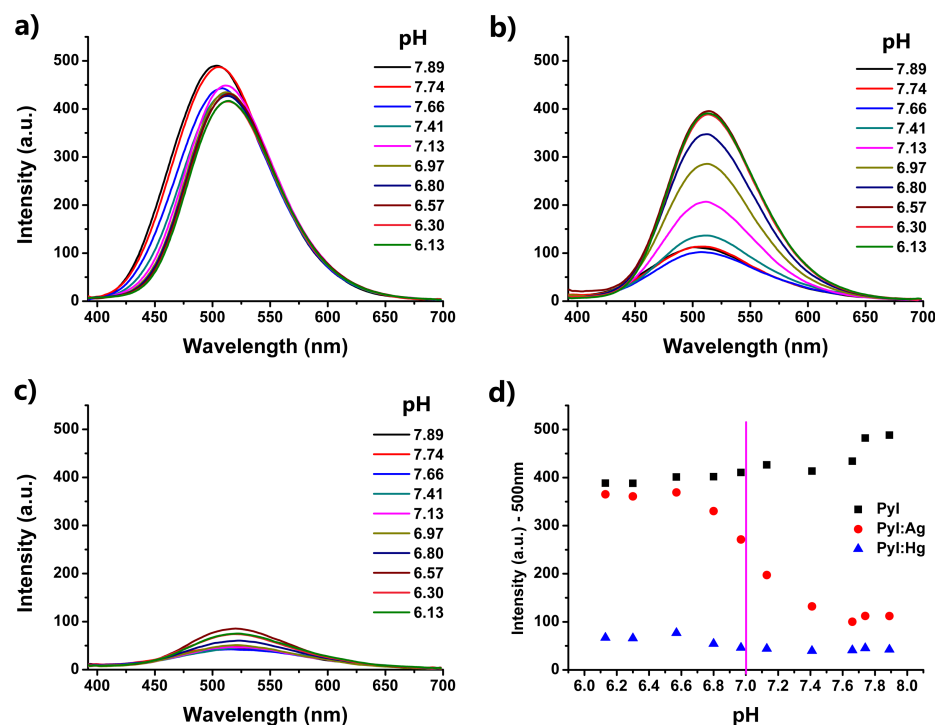


Fig. S3 Emission spectra of **PyI** (5 μM) in EtOH-PBS buffer (5 mM) (1:4, v/v) (a) in pH 6.13~7.89, without Ag^+ and Hg^{2+} ; (b) adding 5 μM Ag^+ or Hg^{2+} in pH 6.13~7.89; (c) adding 5 μM Hg^{2+} in pH 6.13~7.89; (d) The fluorescence intensity at 500 nm without and with 5 μM Ag^+ or Hg^{2+} vs pH, ($\lambda_{\text{ex}} = 375$ nm).

Fluorescence quantum yield (Φ) and % fluorescence quenching of **PyI**.

pH	Compound	Fluorescence quantum yield ^a Φ	% Fluorescence quenching ^{b,c}
6	PyI	0.39	-
6	PyI : Hg^{2+}	0.08	77 (92)
8	PyI	0.45	-
8	PyI : Ag^+	0.10	85 (90)
8	PyI : Hg^{2+}	0.09	87 (93)

Table. S1 ^aQuinine sulphate was used as standard whose quantum yield is 0.63 in 0.1N H_2SO_4 at $\lambda_{\text{ex}} = 375$ nm; ^bcalculated at 0.5 equiv. of metal ions; ^cvalues in parentheses correspond to 1.0 equiv. of metal ions. (EtOH-PBS buffer (5 mM) (1:4, v/v))

Emission spectra of PyI versus Hg^{2+} or Ag^+ concentration

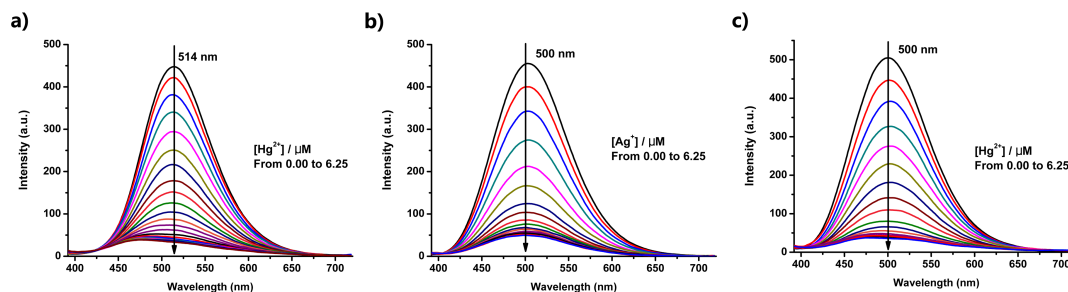


Fig. S4 Emission spectra of **PyI** (5 μM) in EtOH-PBS buffer (5 mM) (1:4, v/v) (a) in pH 6.0, adding different concentrations of Hg^{2+} ; (b) in pH 8.0, adding different concentrations of Ag^+ ; (c) in pH 8.0, adding different concentrations of Hg^{2+} , ($\lambda_{\text{ex}} = 375 \text{ nm}$).

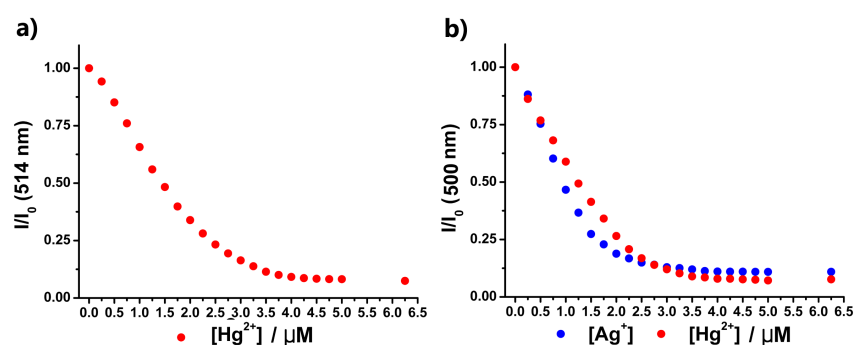


Fig. S5 Plots of fluorescence intensity **PyI** (5.0 μM) at (a) 514 nm versus Hg^{2+} concentration in EtOH-PBS buffer (5 mM, pH = 6.0) (1:4, v/v); (b) 500 nm versus Hg^{2+} and Ag^+ concentration EtOH-PBS buffer (5 mM, pH = 8.0) (1:4, v/v). ($\lambda_{\text{ex}} = 375 \text{ nm}$).

Job plot of Hg^{2+} and Ag^+ complex formation

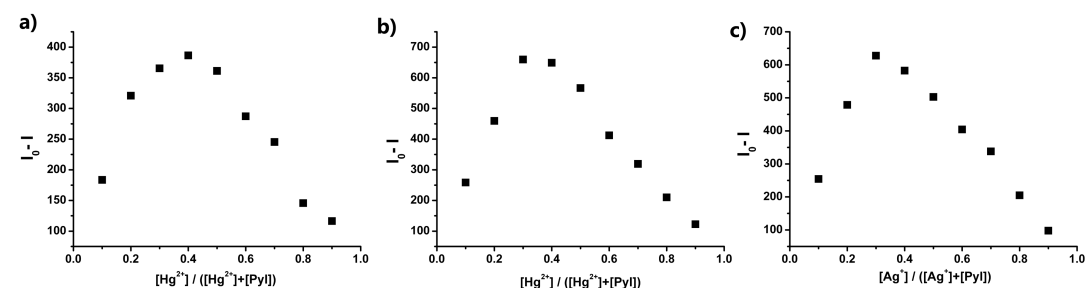


Fig. S6 Job plot of Hg^{2+} and Ag^+ complex formation. I_0 is the fluorescence intensity ((a) at 500 nm in pH 6.0; (b, c) at 514 nm in pH 8.0) when $[\text{Hg}^{2+}]$ or $[\text{Ag}^{2+}] = 0$. $[\text{Hg}^{2+}] + [\text{PyI}] = 0.01 \text{ mM}$ or $[\text{Ag}^+] + [\text{PyI}] = 0.01 \text{ mM}$.

Emission spectra of logic gate Pyl with different inputs

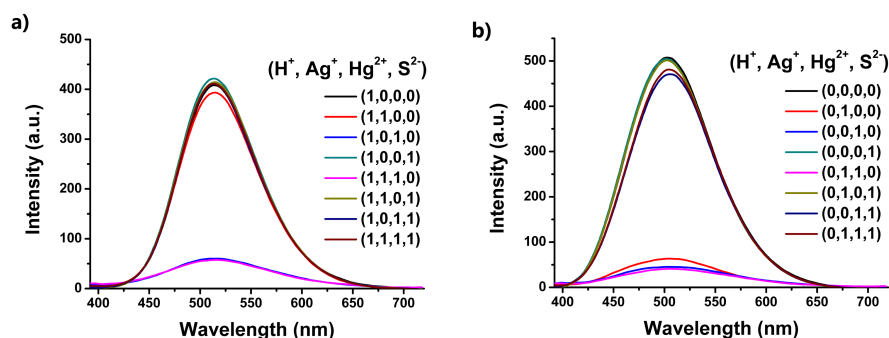


Fig. S7 Emission spectra of Pyl (5 μM) in EtOH-PBS buffer (5 mM) (1:4, v/v) with different inputs (5 μM Hg(NO₃)₂, 5 μM AgNO₃ and 15 μM Na₂S) (a) in pH 6.0; (b) in pH 8.0, (λ_{ex} = 375 nm).

Photos of the reversibility of logic gate Pyl

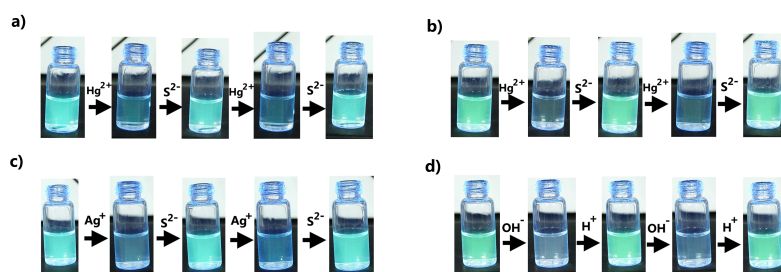


Fig. S8 Fluorescence of Pyl (5 μM) under UV lamp (365 nm) in EtOH-PBS buffer (5 mM) (1:4, v/v) by alternating addition of (a) Hg²⁺ and S²⁻ in pH 6.0; (b) Hg²⁺ and S²⁻ in pH 8.0; (c) Ag⁺ and S²⁻ in pH 8.0; (d) a little dilute HNO₃ and NaOH in the presence of 5 μM Ag⁺.

UV-Vis spectra of Pyl for Hg²⁺ and Ag⁺ in different pH

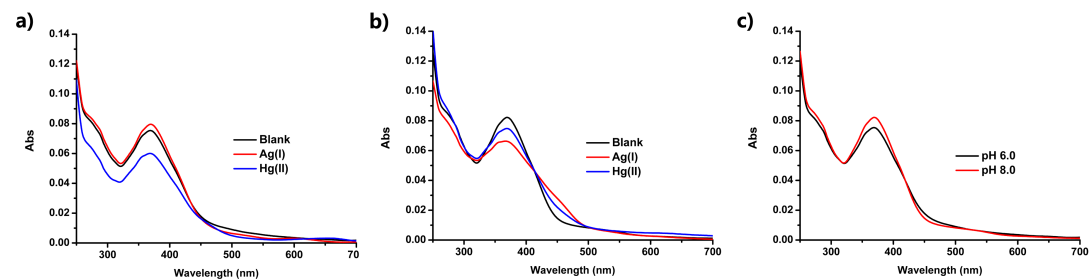


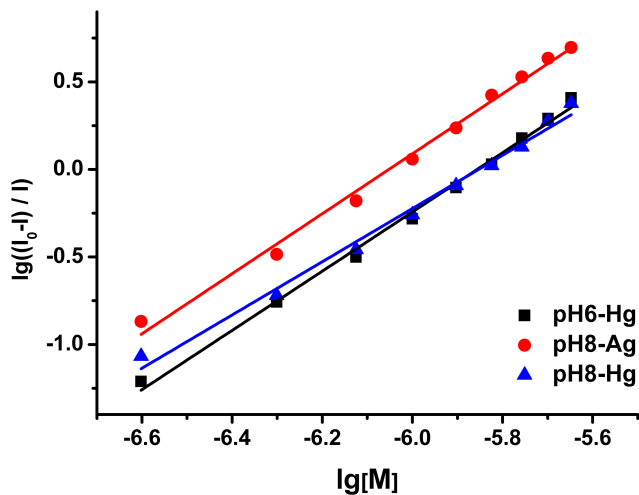
Fig. S9 UV-Vis spectra of **PyI** (0.02 mM) in EtOH-PBS buffer (20 mM) (1:4, v/v) (a) in pH 6.0, adding 0.02 mM Ag⁺ or Hg²⁺; (b) in pH 8.0, adding 0.02 mM Ag⁺ or Hg²⁺; (c) in pH 6.0 or pH 8.0, without Ag⁺ or Hg²⁺.

Hill equation²

Hill equation describes the quantitative relationship between the ligand with receptor. Because the quenching efficiency of Ag⁺ or Hg²⁺ to fluorescent probe **PyI** are high, the **Hill equation** can be described as the following:

$$\lg \frac{I_0 - I}{I} = n \lg [M] + \lg K_{app}$$

where I_0 and I are fluorescence intensities of **PyI** in the absence and presence of Ag⁺ or Hg²⁺ and $[M]$ is the concentration of Ag⁺ or Hg²⁺ respectively. By fitting the fluorescence quenching data, the apparent association constant K_{app} and Hill coefficient n (the binding of **PyI** to Ag⁺ or Hg²⁺) can then be obtained. ($n > 1$, positive cooperativity; $n = 1$, noncooperativity and $n < 1$, negative cooperativity.)



pH	Compound	$\lg K_{app}$	n	R^2
6	PyI :Hg ²⁺	9.91	1.7	0.994
8	PyI :Ag ⁺	10.4	1.7	0.992
8	PyI :Hg ²⁺	8.90	1.5	0.990

Fig. S10 / Table S2. Fluorescence quenching data of **PyI** by Hg²⁺ and Ag⁺ fitted with **Hill equation** (solid line).

(2) J. Gao, Y. Lai, C. Wu and Y. Zhao, *Nanoscale*, 2013, **5**, 8242.

PyI in different pH

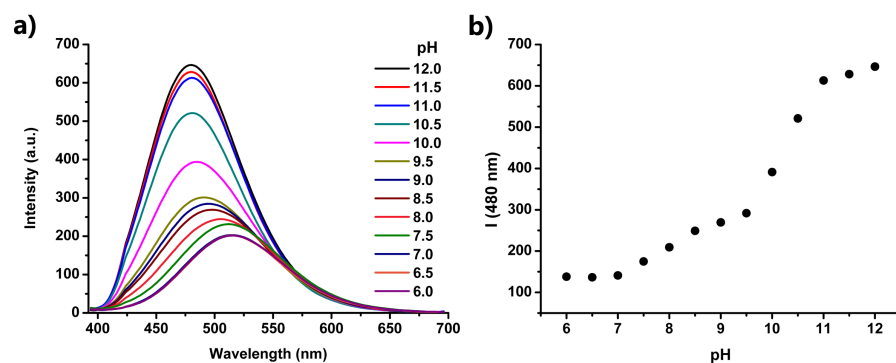
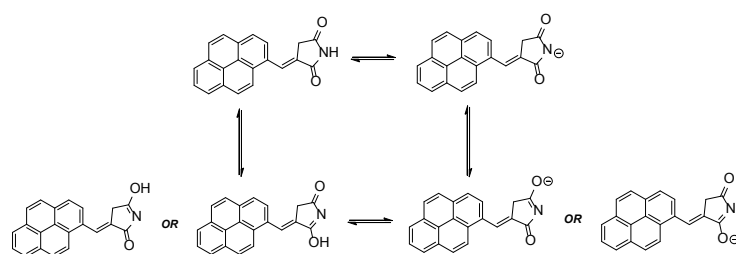


Fig. S11 (a) Emission spectra of **PyI** (5 μM) in EtOH-PBS buffer (5 mM) (1:4, v/v) in pH 6.0–12.0; (b) The fluorescence intensity at 480 nm vs pH, ($\lambda_{\text{ex}} = 375$ nm).



Scheme S1 The conversion of **PyI** (5 μM) in the base environment.

Fluorescence lifetime spectra of PyI

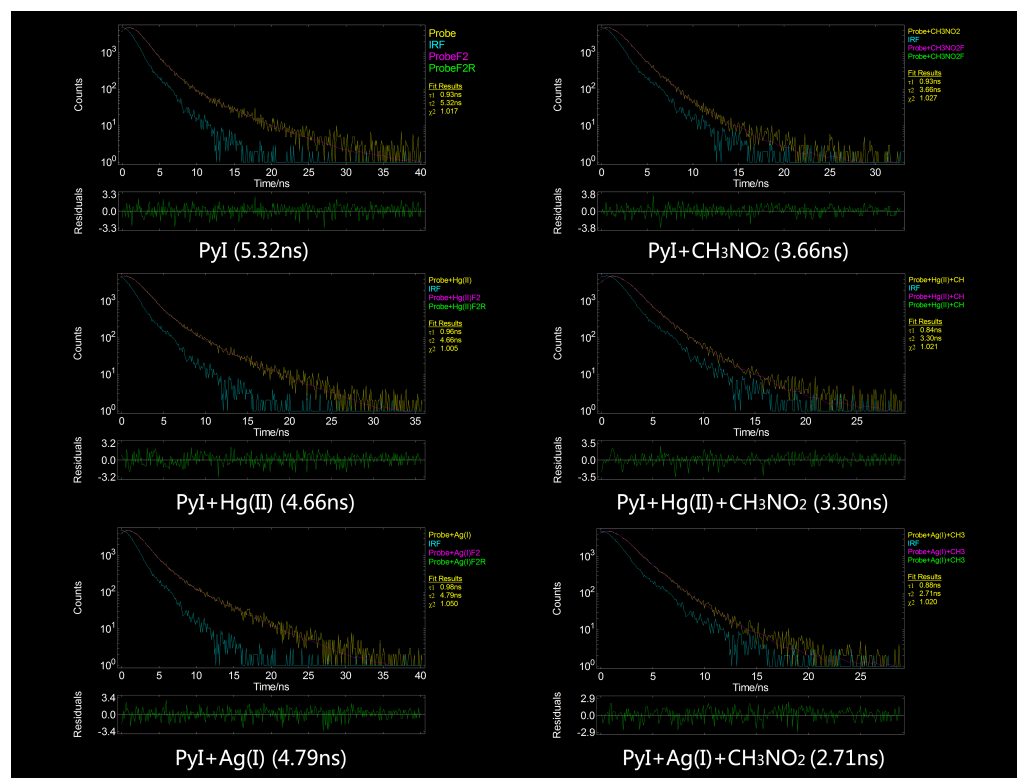


Fig. S12 Fluorescence lifetime spectra of **PyI**, **PyI** + CH₃NO₂, **PyI** + Hg²⁺, **PyI** + Hg²⁺ + CH₃NO₂, **PyI** + Ag⁺, **PyI** + Ag⁺ + CH₃NO₂ in EtOH-PBS buffer (1:4, v/v) in pH 8.0.

The function of the logic gate Pyl

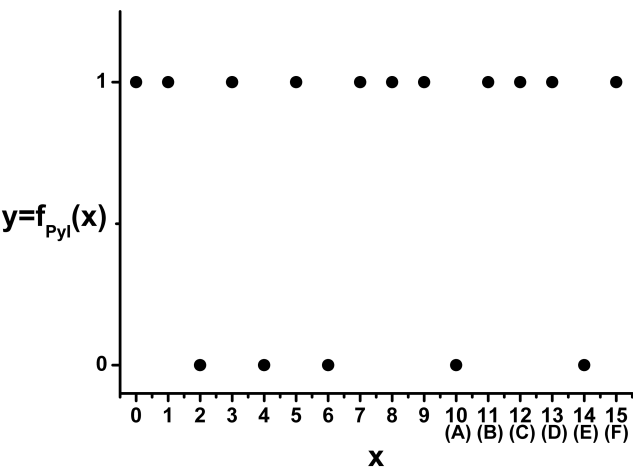


Fig. S13 The encryption function of the logic gate Pyl.

Transcoded by the logic gate Pyl

Table S3. Detailed steps.

Steps	Details
1	ABCD 2346 EAF (Secret letter)
2	A→1010 B→1011 C→1100 D→1101 2→0010 3→0011 4→0100 6→0110 E→1110 A→1010 F→1111 (Hexadecimal→Binary)
3	1010→0 1011→1 1100→1 1101→1 0010→0 0011→1 0100→0 0110→0 1110→0 1010→0 1111→1 (Outputs by Pyl)
4 ^a	0111→.--- 0100→.-. 001→..-
5	.---→J .-.→L ..-→U (Transcoded by Morse alphabet)

[a] “0” = “.” / “1” = “-”

The security and reliability of the encryption by Pyl

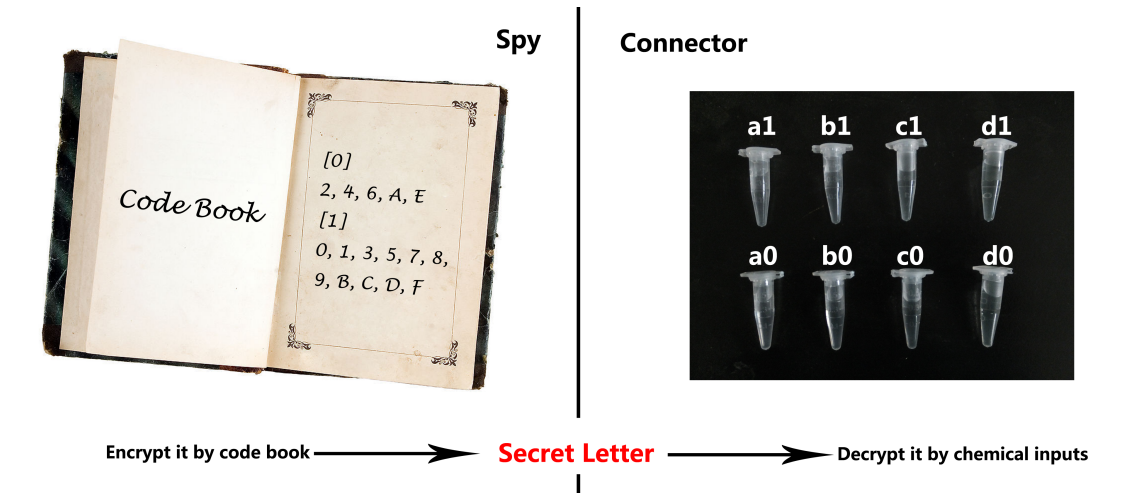


Fig. S14 The code book (spy) and the chemical inputs (connector).

Table S4. Chemical input tubes.

a1	b1	c1	d1	a0	b0	c0	d0
PBS pH=6.0	Ag ⁺ 5 μM	Hg ²⁺ 5 μM	S ²⁻ 15 μM	PBS pH=8.0	H ₂ O -	H ₂ O -	H ₂ O -

Example:

(Encryption) When the the spy wanted to sent the message “0”, he could write “A” in the fluorescent paper.

(Decryption) After getting the secret letter, the connector added (a1, b0, c1, d0) into the fluorescent paper to get the message “0”.

Others: By changing the chemical input tubes as following, we could get 384 kinds of code systems by **PyI**.

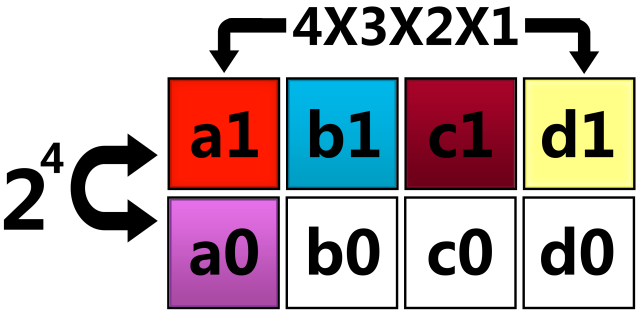


Fig. S15 The changing of the chemical input tubes.

International Morse Code alphabet³

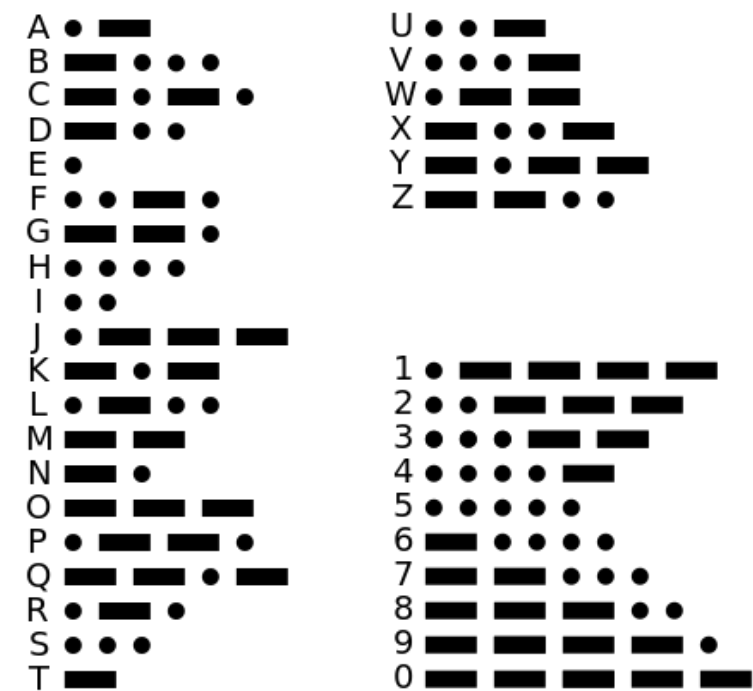


Fig. S16 International Morse Code alphabet

(3) http://en.wikipedia.org/wiki/Morse_code

Electronic engineering symbols for some double-input logic gates⁴

In ₁ In ₂		Out						
0	0	0	0	1	0	1	0	1
0	1	0	1	0	1	0	1	0
1	0	0	1	0	1	0	0	1
1	1	1	1	0	0	1	0	1
		AND	OR	NOR	XOR	XNOR	INH	IMP
								

Fig. S17 Names, truth tables, and electronic engineering symbols for some double-input logic gates which will be built-up into larger arrays in subsequent sections. INH = INHIBIT. IMP = IMPLICATION.

(4) A. P. de Silva, *Chem. Asian J.*, 2011, **6**, 750.

^1H NMR, ^{13}C NMR spectra of PyI

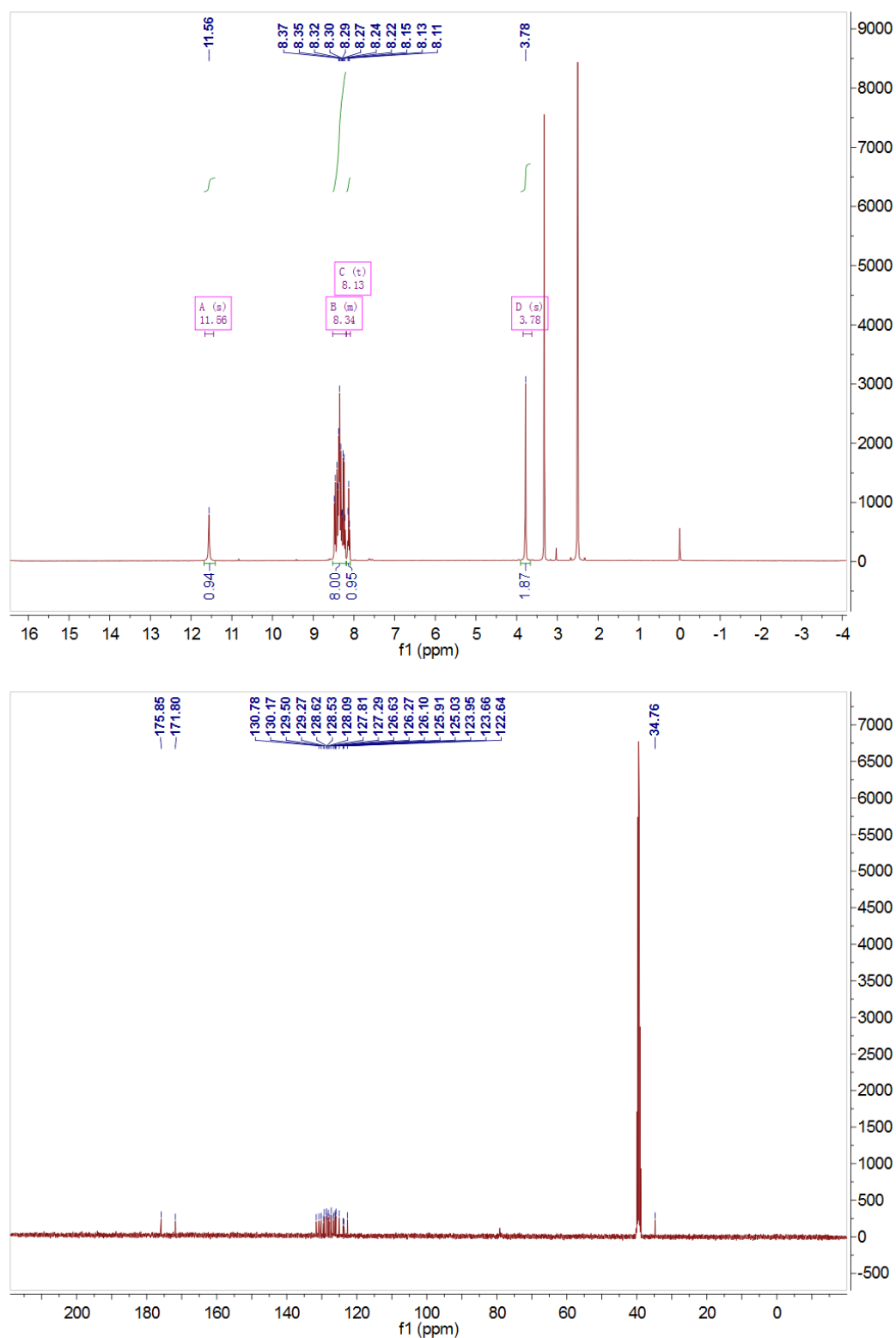


Fig. S18 ^1H NMR, ^{13}C NMR spectra of PyI.

HRMS data of Pyl

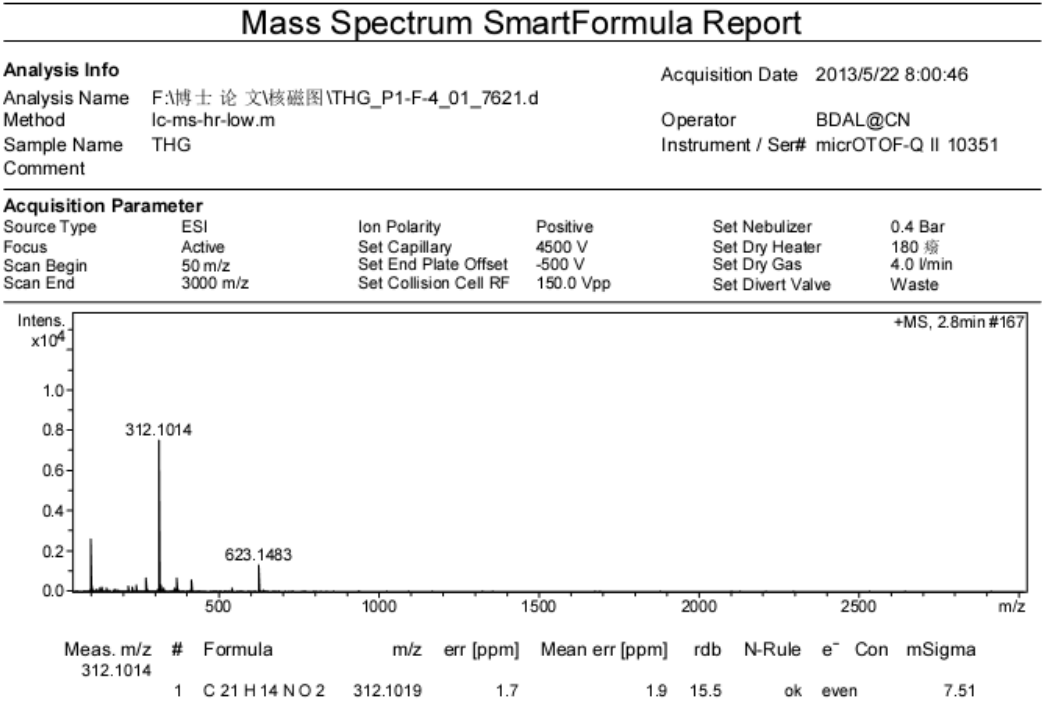


Fig. S19 HRMS data of Pyl

HRMS data of PhI-Ag-PhI

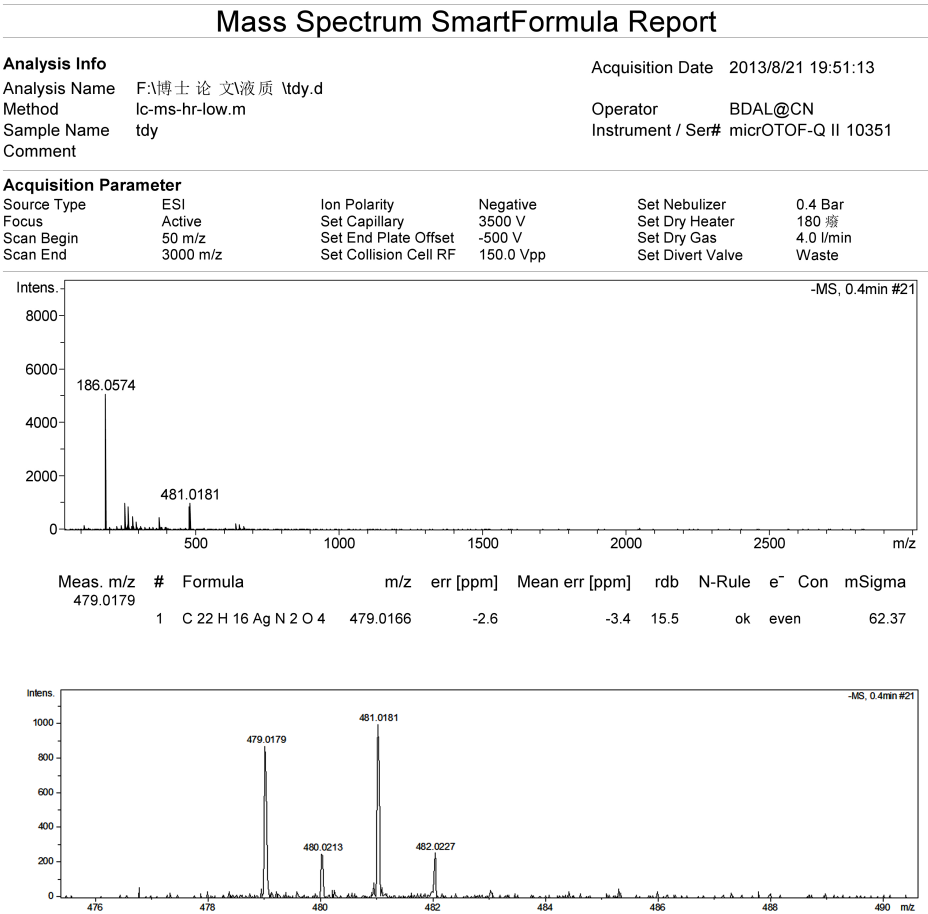


Fig. S20 HRMS data of PhI-Ag-PhI.

¹H NMR spectra of PhI and PhI-Ag-PhI

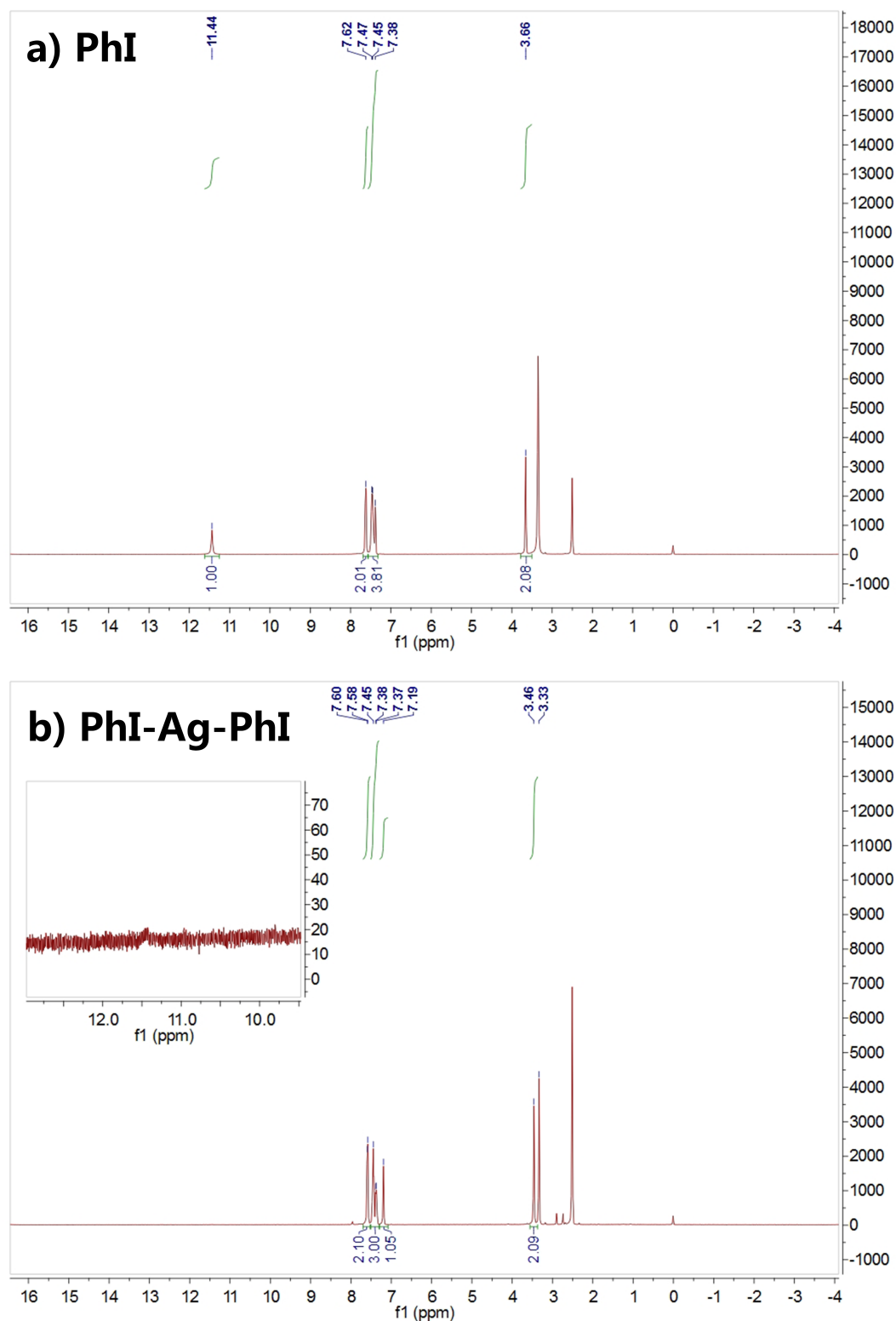


Fig. S21 ¹H NMR spectra of **PhI** and **PhI-Ag-PhI**.