Supporting information

Facile Functionalization of FK506 for Biological Studies by the

Thiol-Ene 'Click' Reaction

Zhi-Fo Guo, Roushu Zhang and Fu-Sen Liang

Table of content

Contents	Page
Supplementary Figure 1	2
Supplementary Figure 2	3
Experiment section	
Chemicals and instrument	4
Synthesis of compounds	4-7
Plasmid construction	7
Cell Culture and transfection	7
Luciferase assay	7-8
Compound Characterization	
NMR of compound 17	9
NMR of compound 18	10
NMR and HRMS of compound 19	11-12
NMR and HRMS of compound 20	12-13
NMR and HRMS of compound 21	14-15
NMR and HRMS of compound 9	15-16
¹ H NMR and HRMS of compound 3	17
¹ H NMR and HRMS of compound 4	18
¹ H NMR and HRMS of compound 6	19
¹ H NMR and HRMS of compound 11	20
¹ H NMR and HRMS of compound 12	21
¹ H NMR and HRMS of compound 13	22
¹ H NMR and HRMS of compound 15	23



Fig. S1. TEC reaction setup. Reaction was carried out using handheld UV lamp at room temperature.



Fig. S2. TEC reaction time profile. The conversion was based on FK506 consumption followed under HPLC.

Experiment section

Chemicals and instruments:

Bulk solvents were obtained from EMD. Cysteamine, 3-thiopropanoic acid, Cysteine, Dithiothreitol, 5-hexynoic acid, N-Ethyl-N'-(3-dimethylaminopropyl)carbodiimide hydrochloride (EDCI), Et₃N, 1-Hydroxybenzotriazole hydrate (HOBt), 3-Aminopropanol, di-tert-butyl dicarbonate $(Boc_2O),$ 4-(Dimethylamino)pyridine (DMAP), p-Toluenesulfonyl chloride (TsCl), triethylsilane, NaN_3 and 2.2-Dimethoxy-2-phenylacetophenone (DPAP) were obtained from Sigma-Aldrich and Alfa-Aesar and were used directly without further purification. Other chemicals are commercially available. Boc-cysteamine was synthesized from Cysteamine following the general procedure. 3-(tritylthio)propinoic acid was synthesized following the reported literature (Langmuir, 2008, 24, 13581). NMR spectra were recorded on a Bruker instrument (300 MHz). Mass and NMR spectra for new compounds were recorded at the Mass Spectrometry and NMR Facilities, Department of Chemistry and Chemical Biology, University of New Mexico.

Synthesis of compounds:



Scheme S1. a) (Boc)₂O, DIPEA, MeOH (99%); b) TsCl, DMAP, Et₃N, DCM (96%); c) NaN₃, DMF, rt, 13 h (89%); d) TFA/DCM, rt, 1 h (99%); e) HATU, DIPEA, DCM, rt, overnight (92%); f) TFA/DCM(v/v= 2: 8), Et₃SiH, rt, 1h (94%).

Prepare of compound 17

3-Aminopropanol (7.0 g, 92 mmol) dissolved in methanol (100 mL) was successively treatd with di-tert-butyl dicarbonate (22.0 g, 101.2 mmol) and di-iso-propylethylamine (32 mL, 184 mmol) stir overnight at room temperature. After removal of the solvent under reduced pressure, the product was dissolved in DCM (100 mL). The organic layer was washed with 10 % citric acid (X 2). The aqueous layers were extracted with DCM. The combined organic layers were dried over sodium sulfate. Removal of the solvent under vacuum gave compound **17** as colorless viscous oil (17.5 g, yield: 99 %). ¹H NMR (CDCl₃, 300 MHz): 4.78 (bs, 1H), 3.67-3.63 (t, J = 11.4 Hz, 2H), 3.29-3.25 (t, J = 12.3 Hz, 2H), 1.69-1.63 (m, 2H), 1.44 (s, 9H). ¹³C NMR (CDCl₃, 300 MHz): 157.50, 79.64, 59.26, 36.97, 32.89, 28.36.

Prepare of compound 18

Compound 17 (3.50 g, 20 mmol), Et₃N (2.02 g, 20 mmol) and DMAP (0.244g, 2 mmol) were stirred in DCM about 5 min, then add TsCl (4.00 g, 21 mmol) and stirred further 2 h at room temperature. Colorless viscous solid was

obtained after purification by silica gel column chromatography using hexane/ethyl acetate (v/v= 2: 1) as an eluting solvent (R_f = 0.72). Yield: 96 %. ¹H NMR (CDCl₃, 300 MHz): 7.79-7.77 (d, *J* = 8.4 Hz, 2H), 7.36-7.33 (d, *J* = 8.1 Hz, 2H), 4.61 (bs, 1H), 4.09-4.05 (t, *J* = 12.0 Hz, 2H), 3.15 (s, 2H), 2.44 (s, 3H), 1.87- 1.79 (m, 2H), 1.41 (s, 9H). ¹³C NMR (CDCl₃, 300 MHz): 155.87, 144.91, 129.92, 127.88, 79.36, 68.00, 36.82, 29.26, 28.35, 21.64.

Prepare of compound 19

Compound **18** (3.29 g, 10 mmol) and NaN₃ (3.30 g, 30 mmol) were stirred in THF/H₂O (v/v= 5: 1) for 2 h. The mixture was extracted with ethyl acetate; the organic layers were combined, washed three times with NaHCO₃ and brine, and subsequently dried with anhydrous Na₂SO₄. Colorless viscous solid was obtained after the solvent was removed under reduced pressure. Yield: 89%. ¹H NMR (CDCl₃, 300 MHz): 4.66 (bs, 1H), 3.37-3.33 (t, J = 13.2 Hz, 2H), 3.22-3.18 (t, J = 12.9 Hz, 2H), 1.80- 1.71 (m, 2H), 1.43 (s, 9H). ¹³C NMR (CDCl₃, 300 MHz): 155.94, 79.42, 49.11, 38.10, 29.29, 28.37. TOF-HRMS (m/z) found (calcd.) for C₈H₁₆N₄OS (M): [M+Na]⁺, 223.1171 (223.1171).

Prepare of compound 20

Compound **19** (1g, 5 mmol) was stirred in DCM/TFA (v/v= 5: 1) for 1 h, white solid was obtained after the solvent was removed under reduced pressure. Yield: 99%. ¹H NMR (CDCl₃, 300 MHz): 8.30 (bs, 2H), 3.57 (s, 2H), 3.18 (s, 2H), 2.09 (s, 2H). ¹³C NMR (CDCl₃, 300 MHz): 48.51, 37.81, 26.80. TOF-HRMS (m/z) found (calcd.) for $C_3H_8N_4$ (M): [M+H]⁺, 101.0814 (101.0827).

Prepare of compound 21

3-(tritylthio)propanoic acid (697 mg, 2.0 mmol), HATU (836 mg, 2.2 mmol), DIPEA (516 mg, 4.0 mmol), and compound **20** (220 mg, 2.2 mmol) were stirred at room temperature overnight in DCM. White solid was obtained after purification by silica gel column chromatography using hexane/ethyl acetate (v/v= 1: 1) as an eluting solvent (R_f = 0.65). Yield: 96 %. ¹H NMR (Acetone-d₆, 300 MHz): 7.41-7.39 (m, 6H), 7.34-7.28 (m, 6H), 7.26-7.20 (m, 3H), 7.14 (bs, 1H), 3.39-3.35 (t, *J*= 13.5 Hz, 2H), 3.27-3.20 (q, *J*= 19.2 Hz, 2H), 2.44-2.39 (t, *J*= 14.7 Hz, 2H), 2.22-2.17 (t, *J*= 14.7 Hz, 2H), 1.77- 1.68 (m, 2H). ¹³C NMR (Acetone-d₆, 300 MHz): 171.11, 145.83, 130.34, 128.70, 127.47, 67.17, 49.66, 37.10, 35.40, 28.53. TOF-HRMS (m/z) found (calcd.) for C₂₅H₂₆N₄OS (M): [M+Na]⁺, 453.1717 (453.1725), [2M+Na]⁺, 883.3400 (883.3552).

Prepare of compound 9

To a solution of compound **21** (215 mg, 0.5 mmol) in trifluoroacetic acid (TFA, 1mL) and CH₂Cl₂ (2 mL) was added triethylsilane (174 mg, 1.5 mmol). The resulting mixture was stirred for a half hour at room temperature. After evaporating the reaction solvent, CH₂Cl₂ (20 mL) was added to the resulting residues. The organic layer was extracted with H₂O (10 mL). The aqueous layer was then evaporated to give the transparent oil product. Yield: 94 %. ¹H NMR (CDCl₃, 300 MHz): 5.93 (s, 1H), 3.41-3.36 (m, 4H), 2.84-2.80 (q, *J*= 12.9 Hz, 2H), 2.52-2.49 (t, *J*= 7.8 Hz, 2H), 1.84-1.79 (m, 2H), 1.62-1.59 (t, *J*= 9.9 Hz, 1H). ¹³C NMR (CDCl₃, 300 MHz): 171.58, 49.45, 40.46, 37.48, 28.79, 20.54. TOF-HRMS (m/z) found (calcd.) for C₆H₁₂N₄OS (M): [M+Na]⁺, 211.0628 (211.0630), [2M+Na-2H]⁺, 397.1207 (397.1207).

Synthesis of compound 3

FK506 (201.0 mg, 0.25 mmol), Boc-cysteamine (46.0 mg, 0.26 mmol), DPAP (3.2 mg, 12.5 nmol) and 0.4 mL dichloromethane were put in a vials, and stirred 15 min under UV light. White solid (246 mg, 98% yield) was

obtained after purification by silica gel column chromatography using ethyl acetate as an eluting solvent (R_f = 0.5). ¹H NMR (300 MHz, CDCl₃): 5.33-5.21(d, *J*= 36.8 Hz, 1H), 5.12-5.08 (m, 2H), 4.88-4.40 (m, 1H), 3.94-3.57 (m, 3H), 3.41-3.29 (m, 9H), 3.05-2.95 (m, 2H), 2.78-2.50 (m, 6H), 2.38-1.26 (m, 45H), 1.07-0.82(m, 13H). TOF-HRMS (m/z) found (calcd.) for C₅₁H₈₄N₂O₁₄S (M): [M+Na]⁺, 1003.5583 (1003.5541).

Synthesis of compound 4

Method A: compound 3 disovled in DCM/TFA and stir 1 h at room temprature.

Method B: FK506 (201.0 mg, 0.25 mmol), cysteamine (19.3 mg, 0.26 mmol), DPAP (3.2 mg, 12.5 nmol) and 0.4 mL methanol were put in a vials, and stirred 15 min under UV light. White solid (211 mg, 96% yield) was obtained after purification by silica gel column chromatography using ethyl DCM/methanol (v/v= 5: 1) as an eluting solvent (R_f = 0.56). ¹H NMR (300 MHz, Acetone-d₆): 6.93-6.80 (m, 1H), 6.45-6.29 (m, 1H), 5.30-4.98 (m, 3H), 4.66-4.34 (m, 1 H), 4.13-4.02 (m, 2H), 3.80-3.32 (m, 11H), 3.02-3.00 (m, 2H), 2.68-2.50 (m, 4H), 2.49-1.58 (m, 36 H), 1.21-0.88 (m, 13H). TOF-HRMS (m/z) found (calcd.) for C₄₆H₇₆N₂O₁₂S (M): [M+H]⁺, 881.5215 (881.5197).

Synthesis of compound 6

5-hexynoic acid (28.0 mg, 0.25 mmol), EDCI (52.7 mg, 0.275 mmol), HOBt (37.1 mg, 0.275 mmol), and Et₃N (50.5 mg, 0.5 mmol), stir about 1 h in DCM, then add compound **4** (242 mg, 0.275 mmol) and stir overnight. White solid (190 mg, 78% yield) was obtained after purification by silica gel column chromatography using ethyl acetate as an eluting solvent (R_f = 0.42). ¹H NMR (300 MHz, CDCl₃): 6.80-6.69 (m, 1H), 6.34-6.10 (m, 1H), 5.97 (s, 1H), 5.26-4.85 (m, 2H), 4.75-4.72 (m, 1H), 4.49-4.16 (m, 1H), 3.92-3.72 (m, 1H), 3.59-3.26 (m, 11H), 3.06-2.92 (m, 2H), 2.63-2.61 (m, 2H), 2.49-2.23 (m, 6H), 2.12-1.23 (m, 39H), 1.80-0.83 (m, 13H). TOF-HRMS (m/z) found (calcd.) for $C_{52}H_{82}N_2O_{13}S$ (M): [M+H]⁺, 979.5360 (976.2860).

Synthesis of compound 11

FK506 (201.0 mg, 0.25 mmol), 3-thiopropanoic acid (28.0 mg, 0.26 mmol), DPAP (3.2 mg, 12.5 nmol) and 0.4 mL dichloromethane were put in a vials, and stirred 15 min under UV light. White solid (222 mg, 98% yield) was obtained after purification by silica gel column chromatography using ethyl acetate/ acetone (v/v= 1: 1) as an eluting solvent (R_f = 0.51). ¹H NMR (300 MHz, CDCl₃): 5.32-5.20(d, *J*= 36.4 Hz, 1H), 5.10-5.00 (m, 2H), 4.75-4.30 (m, 1H), 3.94-3.54 (m, 3H), 3.40-3.29 (m, 11H), 3.05-2.95 (m, 2H), 2.75-2.51 (m, 6H), 2.38-1.25 (m, 36H), 1.06-0.82(m, 13H). TOF-HRMS (m/z) found (calcd.) for C₄₇H₇₅NO₁₄S (M): [M-H]⁺, 908.4821 (908.4830).

Synthesis of compound 12

FK506 (201.0 mg, 0.25 mmol), compound **9** (48.9 mg, 0.26 mmol), DPAP (3.2 mg, 12.5 nmol) and 0.4 mL DCM were put in a vials, and stirred 15 min under UV light. Pale yellow solid (235 mg, 95% yield) was obtained after purification by silica gel column chromatography using ethyl acetate/ acetone (v/v= 4: 1) as an eluting solvent (R_f = 0.56). ¹H NMR (300 MHz, CDCl₃): 6.50 (s, 1H), 5.32-4.95 (m, 3H), 4.74-4.22 (m, 2H), 3.91-3.53 (m, 2H), 3.38-3.27 (m, 17H), 2.98-2.95 (m, 4H), 2.80-2.64 (m, 4H), 2.58-2.39 (m, 4H), 2.15-1.34 (m, 34H), 0.98-0.80 (m, 13H). TOF-HRMS (m/z) found (calcd.) for C₅₀H₈₁N₅O₁₃S (M): [M+Na]⁺, 1014.5485 (1014.5449).

Synthesis of compound 13

FK506 (201.0 mg, 0.25 mmol), Cysteine (31.5 mg, 0.26 mmol), DPAP (3.2 mg, 12.5 nmol) and 0.5 mL methanol/water (1: 1) were put in a vials, and stirred 15 min under UV light. White solid (220 mg, 95% yield) was

obtained after purification by silica gel column chromatography using acetone/methanol (v/v= 1: 1) as an eluting solvent (R_f = 0.46). ¹H NMR (300 MHz, CDCl₃): 5.27-4.95 (m, 3H), 4.63-4.33 (m, 1H), 4.08-3.97 (m, 2H), 3.70-3.41 (m, 3H), 3.38-2.75 (m, 13H), 2.09-1.20 (m, 34H), 0.96-0.71(m, 13H). TOF-HRMS (m/z) found (calcd.) for C₄₇H₇₆N₂O₁₄S (M): [M+H]⁺, 925.5076 (925.5096).

Synthesis of compound 15

FK506 (201.0 mg, 0.25 mmol), dithiothreitol (20.2 mg, 0.13 mmol), DPAP (3.2 mg, 12.5 nmol) and 0.4 mL dichloromethane were put in a vials, and stirred 15 min under UV light. White solid (220 mg, 99% yield) was obtained after purification by silica gel column chromatography using ethyl acetate/acetone (v/v=1:1) as an eluting solvent (R_f = 0.56). ¹H NMR (300 MHz, CDCl₃): 5.33-5.20 (d, *J*= 36.5 Hz, 2H), 5.10-5.06 (m, 4H), 4.74-4.28 (m, 4H), 3.92-3.57 (m, 8H), 3.41-3.30 (m, 22H), 3.06-2.97 (m, 4H), 2.74-2.54 (m, 14H), 2.30-1.25 (m, 64H), 1.08-0.83 (m, 26H). TOF-HRMS (m/z) found (calcd.) for C₉₂H₁₄₈N₂O₂₆S₂ (M): [M+Na]⁺, 1783.9602 (1783.9659).

Plasmid construction

All DNA fragments were amplified by PCR (Polymerase chain reaction) from other intermediate constructs. PCR was carried out with Phusion DNA Polymerase (New England Biolabs), PfuUltra II Fusion HotStart DNA Polymerase (Agilent Technologies) under S1000 thermal cycler with Dual 48/48 Fast Reaction Module (Bio-Rad).

- For SV-ires-GalDBD-3FKBP12, a DNA construct SV-VP16-Frb-ires-GalDBD-FKBP12x3 (*Sci. Signal.*, 2011, 4, rs2) was firstly digested by EcoRI and BamHI. The sticky ends of the vector were blunted by DNA Polymerase I Lg (Klenow) Fragment (New England Biolabs) under the present of dNTPs. Finally, the blunt ends were ligated by T4 DNA Ligase (New England Biolabs).
- 2) For SV-VP16-FKBP12x2, the DNA construct was first made as SV-VP16-FKBP12. SV-VP16-Frb-ires-GalDBD-FKBP12x3 was firstly digested by AscI and NotI. The PCR product of FKBP12 was inserted into the vector by recombination using the In-Fusion HD Enzyme Premix (ClonTech). The second copy of FKBP12 was inserted via the AcsI site using T4 DNA ligase.

Cell Culture and transfection

CHO cells were cultured in Dulbecco's modified Eagle's medium (DMEM, Gibco) with 10% Fetal Bovine Serum (FBS, Atlanta Biologicals), 1×glutamate (100x from Gibco) and 1×penicillin/streptomycin (Pen/Strep, 100X from Gibco). Cells were plated with the starting concentration of 50,000 cells per well in a 24-well plate (Greiner Bio-one) the day before transfection. An amount of 0.2µg of each DNA construct was mixed with Opti-MEM (Gibco) and PEI. After incubation at room temperature for 15min, the mixture was added to the cells and cultured for 24h. Then FK506, FK1012-DT and FK1012-ZE dissolved in DMSO were added into the cell culture with the final concentration of 200nM, 100nM and 100nM respectively. Each type of the experiments, including the one with transfected DNA but without drugs, was carried out as triplets. After the incubation of 10 hours, the cells were harvested and washed by PBS buffer (Gibco) for 3 times.

Luciferase assay

Cells in 24-well plates were frozen under the temperature of -80° C at first and then lysed with 100 µL of 1×Passive Lysis Buffer (Promega Corporation) at room temperature for 10min on a shaker. Cell lysates were then collected and centrifuged in tubes and 10µL of supernatant was added separately into a 96 well plate for Luciferase assay. 90µL

Luciferase substrate solution (5mg of D-luciferin and 7mg of coenzyme A in 33mL of Luciferase reading buffer, which includes 20mM tricine, 1.07mM (MgCO₃)₄Mg(OH)₂·5H₂O, 2.67mM MgSO₄, 0.1mM EDTA, 33.3mM dithiothreitol and 0.53mM ATP in water) was added into each well with cell lysates. The signal was read with a 3s delay and 1s integration with Clomax Multi Detection System (Promega). Obtained data were analyzed by KaleidaGraph. The shown results are from 3 experimental repeats.



¹³C NMR of compound **17**



¹³C NMR of compound **18**



¹³C NMR of compound **19**



¹H NMR of compound **20**





HRMS of compound ${\bf 20}$



¹³C NMR of compound **21**



HRMS of compound 21







HRMS of compound 9



HRMS of compound 3





HRMS of compound 6



HRMS of compound 11



HRMS of compound 12



HRMS of compound 13



Elemental Composition Report

Single Mass Analysis

Tolerance = 20.0 PPM / DBE: min = -1.5, max = 50.0 Isotope cluster parameters: Separation = 1.0 Abundance = 1.0%

Monoisotopic Mass, Odd and Even Electron lons 3 formula(e) evaluated with 3 results within limits (up to 50 closest results for each mass)

Zhifo Guo / Lia 130424_167b 100-) / Liang 20130423A 67b 364 (8.343) Cm (364:435) 1784.9646 ↓ │ 1785.9642						UNM MS Facility Inst.KD116 1: TOF MS ES+ 1.14e4			
%- 0- 1750.0	1756.9945 1760.0	<u>1769.9293</u> 1770.0	1778.9663	33.9095	1786.9602	<u>1800.920</u> 1800.0	<u>1803.0487</u> 1810.0	1815.9733 1818.9415 1827.8989 1820.0 1830.0		
Minimum: Maximum:		200.0	20.0	-1.5 50.0						
Mass	Calc. Mass	mDa	PPM	DBE	Score	Formula				
1783.9602	1783.9659 1783.9320 1783.9296	-5.7 28.2 30.6	-3.2 15.8 17.2	19.5 23.5 20.5	2 3 1	C92 H14 C93 H14 C91 H14	43 N2 027	Na 3252 3252 Na 3252		

HRMS of compound 15