

Synthesis of interesting β -nitrohydrazides through thiourea organocatalysed aza-Michael addition

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ELECTRONIC SUPPLEMENTARY INFORMATION (ESI)

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General experimental methods. Purification of reaction products was carried out by flash chromatography using silical-gel (0.063-0.200 mm). Analytical thin layer chromatography was performed on 0.25 mm silical gel 60-F plates. ESI ionization method and mass analyser type MicroTof-Q were used for the HRMS measurements. ¹H-NMR spectra were recorded at 300 MHz and 400 MHz; ¹³C-NMR spectra were recorded at 75 MHz and 100 MHz; CD₃CN, CDCl₃, CD₂Cl₂, THF-d₈ and CD₃COCD₃ as the solvent. Chemical shifts were reported in the δ scale relative to residual CH₃CN (1.94 ppm), CHCl₃ (7.26 ppm), CH₂Cl₂ (5.32 ppm), THF (1.73 ppm) and CH₃COCH₃ (2.05 ppm) for ¹H-NMR and to the central line of CDCl₃ (77 ppm), CD₂Cl₂ (53.8 ppm), THF-d₈ (25.3 ppm) and CD₃COCD₃ (30.8 ppm) for ¹³C-NMR.

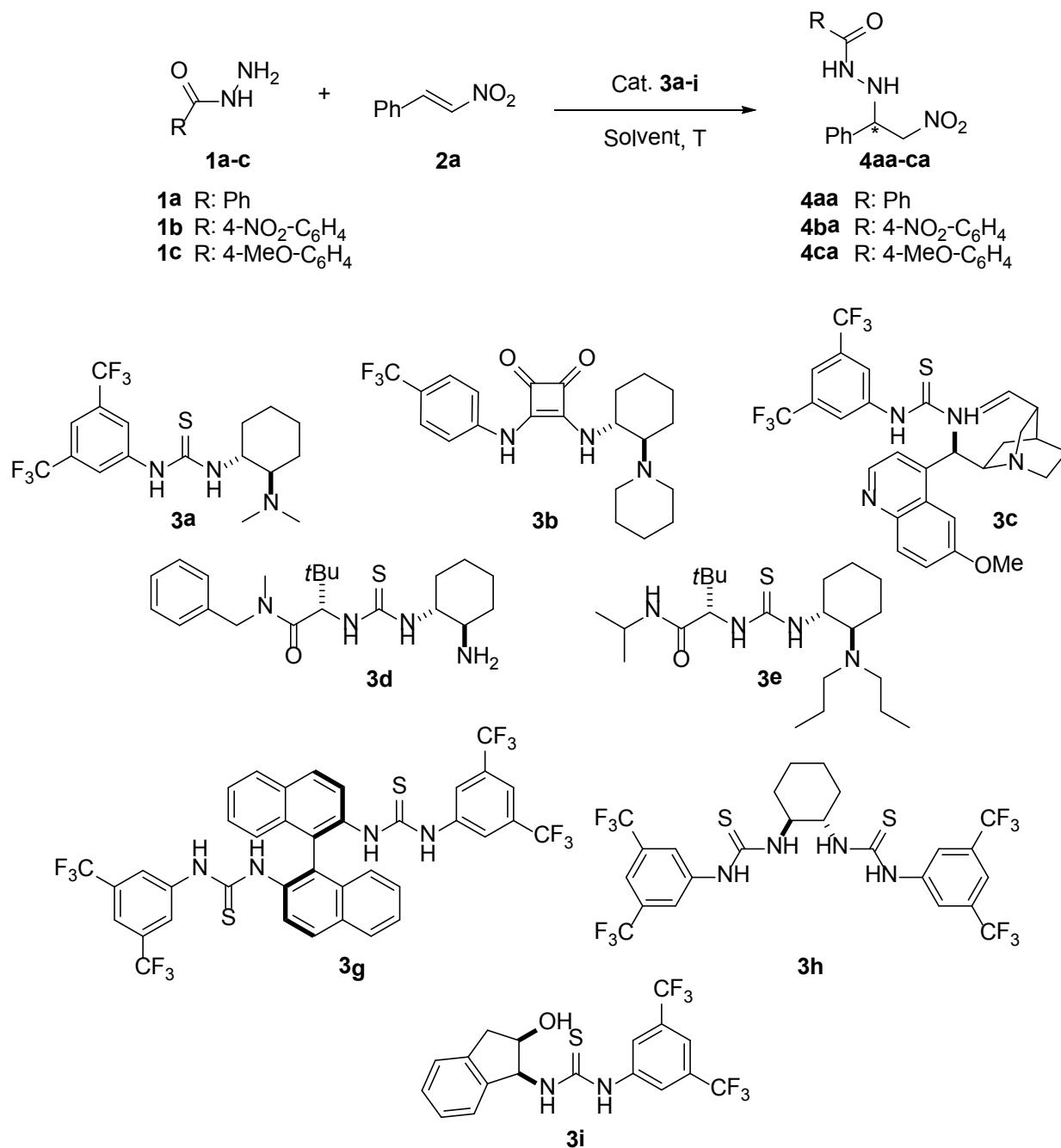
Materials. All commercially available solvents and reagents were used as received. Nitroalkenes **2k**[1] and **2l**[1] have been synthesised following the described method in the literature. The NMR spectra (¹H-NMR and APT) for nitroalkenes **2k**[1] and **2l**[2] are consistent with values previously reported in the literature.

[1] S. E. Denmark and L. R. Marcin, *J. Org. Chem.*, 1993, **58**, 3850.

[2] E. C. Taylor and B. Liu., *J. Org. Chem.*, 2003, **68**, 9938.

Screening of Catalysts and Study of the Reaction Conditions

Scheme S1. Organocatalytic enantioselective aza-Michael addition reaction of hydrazine **1** to nitrostyrene **2a**.



In an initial screening of solvents at room temperature (Table S1, entries 1-7), CH₃CN (entry 3) and EtOAc (entry 7) were identified as the solvents of choice for further variations, since they led to the highest values of enantiomeric ratio. Although we also continued with EtOAc in the ensuing screenings, we finally decided to proceed with CH₃CN because slightly better results were rendered for similar conditions. It was found that lowering the temperature to 5 or -27 °C improved enantiomeric ratios were achieved, although longer reaction

times were required (entries 8 and 11). Decreasing the catalyst loading until 10 or 5 mol% (entries 13 and 14), and increasing the concentration of the reaction mixture had a positive effect on the enantioselectivity without altering the final good yield. Unfortunately, experiments made lowering further the temperature to $-35\text{ }^{\circ}\text{C}$ afforded poorer enantioselectivities and yields (entries 15 and 16). Finally, the combination of 10 mol% of catalyst **3a** cooling down the reaction mixture to $-27\text{ }^{\circ}\text{C}$ in 0.25 mL of solvent, led to a slight increase in the asymmetric induction as well as in the yield, providing the best reaction conditions (entry 13).

Table S1. aza-Michael addition reaction of hydrazine **1** to nitrostyrene **2a** catalysed by thiourea **3a**.^[a]

| Entry | Solvent (mL) | 3a (mol%) | T ($^{\circ}\text{C}$) | Time (h) | Yield (%) ^[b] | e.r. ^[c] |
|-------|---------------------------------------|------------------|--------------------------|----------|--------------------------|---------------------|
| 1 | Toluene (0.5) | 20 | r.t. | 72 | 12 | 56:44 |
| 2 | Xylene (0.5) | 20 | r.t. | 48 | 24 | 55:45 |
| 3 | CH ₃ CN (0.5) | 20 | r.t. | 48 | >95 | 75:25 |
| 4 | CHCl ₃ (0.5) | 20 | r.t. | 72 | 33 | 52:48 |
| 5 | CH ₂ Cl ₂ (0.5) | 20 | r.t. | 48 | 70 | 55:45 |
| 6 | THF (0.5) | 20 | r.t. | 78 | 91 | 55:45 |
| 7 | EtOAc (0.5) | 20 | r.t. | 20 | 79 | 65:35 |
| 8 | CH ₃ CN (0.5) | 20 | 5 | 84 | 87 | 77:23 |
| 9 | CH ₃ CN (0.5) | 10 | 5 | 40 | >95 | 67:33 |
| 10 | CH ₃ CN (0.25) | 10 | 5 | 42 | >95 | 80:20 |
| 11 | CH ₃ CN (0.5) | 20 | -27 | 93 | 91 | 82:18 |
| 12 | CH ₃ CN (0.5) | 10 | -27 | 72 | 88 | 82:18 |
| 13 | CH ₃ CN (0.25) | 10 | -27 | 84 | >95 | 88:12 |
| 14 | CH ₃ CN (0.25) | 5 | -27 | 72 | 79 | 86:14 |
| 15 | CH ₃ CN (0.5) | 20 | -35 | 48 | 12 | 70:30 |
| 16 | CH ₃ CN (0.25) | 10 | -35 | 120 | 27 | 75:25 |

[a] Experimental conditions: To a solution of thiourea **3a** and nitrostyrene **2a** (0.1 mmol) in solvent, hydrazide **1b** (0.1 mmol) was added at the indicated temperature. After a reasonable reaction time, product **4ba** was isolated by flash chromatography (Hex:EtOAc, 7:3). [b] Isolated yield. [c] Determined by chiral HPLC analysis (Chiraldak IC, 80:20 Hex:EtOAc, 1 mL/min).

We have also explored different conditions with thioureas **3b-e** (Table S2) and **3g-i** (Table S3) before to discard these catalysts as suitable ones for developing this reaction.

Table S2. aza-Michael addition reaction of hydrazine **1** to nitrostyrene **2a** catalysed by thiourea **3a-e**.^[a]

| Entry | Cat. (mol%) | Hydraz. | Solvent (mL) | T (°C) | Time (h) | Yield (%) ^[b] | e.r. ^[c] |
|------------------|-----------------|-----------|--|--------|----------|--------------------------|---------------------|
| 1 ^[d] | 3b (20) | 1b | CH ₃ CN (0.25) | r.t. | 18 | 60 | 52:48 |
| 2 ^[d] | 3c (20) | 1b | CH ₃ CN (0.25) | r.t. | 18 | 60 | 65:35 |
| 3 | 3d (20) | 1b | CH ₃ CN (0.5) | r.t. | 48 | 76 | 52:48 |
| 4 | 3e (20) | 1b | CH ₃ CN (0.5) | r.t. | 48 | 74 | 53:47 |
| 5 | 3a (20) | 1a | CH ₃ CN (0.5) | r.t. | 20 | 63 | 52:48 |
| 6 | 3a (20) | 1c | CH ₃ CN (0.5) | r.t. | 20 | 86 | 54:46 |
| 7 | 3a (20) | 1b | EtOAc (0.5) | 5 | 91 | 67 | 69:31 |
| 8 | 3a (30) | 1b | EtOAc (0.5) | 5 | 85 | 76 | 74:26 |
| 9 | 3a (20) | 1b | EtOAc (0.5) | -27 | 113 | 33 | 81:19 |
| 10 | 3a (30) | 1b | EtOAc (0.5) | -27 | 109 | 52 | 80:20 |
| 11 | 3a (30) | 1b | CH ₃ CN (0.5) | 5 | 84 | 79 | 64:36 |
| 12 | 3a (30) | 1b | CH ₃ CN (0.5) | -27 | 92 | 97 | 82:18 |
| 13 | 3a (2.5) | 1b | CH ₃ CN (0.25) | -27 | 72 | 70 | 82:18 |
| 14 | 3a (10) | 1b | CH ₃ CN (0.25)/EtOAc (0.25) | -27 | 72 | 61 | 81:19 |
| 15 | 3a (20) | 1b | CH ₃ CN (0.25)/EtOAc (0.25) | -27 | 73 | 80 | 82:18 |
| 16 | 3a (10) | 1b | CH ₃ CN (0.225)/EtOAc (0.025) | -27 | 76 | <95 | 86:14 |
| 17 | 3a (20) | 1b | CH ₃ CN (0.450)/EtOAc (0.050) | -27 | 76 | 48 | 86:14 |

[a] Experimental conditions: To a solution of thiourea **3a-e** and nitrostyrene **2a** (0.1 mmol) in the corresponding solvent, hydrazide **1a-c** (0.1 mmol) was added at the corresponding temperature. After a reasonable reaction time, final product was isolated by flash chromatography (Hex:EtOAc, 7:3). [b] Isolated yield. [c] Determined by chiral HPLC analysis. [d] Reaction performed using 0.05 mmol of nitroalkenes.

Table S3. aza-Michael addition reaction of hydrazine **1** to nitrostyrene **2a** catalysed by thiourea **3g-i**.^[a]

| Entry | Cat. | Hydraz. | Solvent | Time (h) | Yield (%) ^[b] | e.r. ^[c] |
|-------|-----------|-----------|--------------------|----------|--------------------------|---------------------|
| 1 | 3g | 1a | Toluene | 24 | 62 | 56:44 |
| 2 | 3g | 1a | CHCl ₃ | 24 | 68 | 56:44 |
| 3 | 3g | 1a | THF | 24 | 47 | 52:48 |
| 4 | 3g | 1a | CH ₃ CN | 48 | 46 | 56:44 |
| 5 | 3g | 1a | Xylene | 48 | 49 | 57:43 |
| 6 | 3g | 1b | CH ₃ CN | 40 | 15 | 62:38 |
| 7 | 3g | 1b | Toluene | 72 | 22 | 52:48 |
| 8 | 3g | 1b | CHCl ₃ | 72 | 36 | 56:44 |
| 9 | 3g | 1b | THF | 72 | 42 | 56:44 |
| 10 | 3g | 1b | CH ₃ CN | 72 | 45 | 61:39 |
| 11 | 3g | 1b | Xylene | 72 | 32 | 56:44 |
| 12 | 3g | 1c | Toluene | 24 | 63 | 57:43 |
| 13 | 3g | 1c | CHCl ₃ | 24 | 66 | 56:44 |
| 14 | 3g | 1c | THF | 72 | 64 | 52:48 |
| 15 | 3g | 1c | CH ₃ CN | 72 | 63 | 55:45 |
| 16 | 3g | 1c | Xylene | 72 | 42 | 58:42 |
| 17 | 3h | 1b | CH ₃ CN | 40 | 39 | 53:47 |
| 18 | 3i | 1b | CH ₃ CN | 42 | 27 | 57:43 |

[a] Experimental conditions: To a solution of thiourea **3g-i** (20 mol%) and nitrostyrene **2a** (0.1 mmol) in the corresponding solvent (0.5 mL), hydrazide **1a-c** (0.1 mmol) was added at room temperature. After a reasonable reaction time, final products were isolated by flash chromatography (Hex:EtOAc, 7:3). [b] Isolated yield. [c] Determined by chiral HPLC analysis.

The background reaction at room temperature was found to be very high for the three hydrazides (Table S4, entries 1,2 and 4). However, at the temperature for best reaction conditions, the background is clearly minor (entry 5). Moreover, since the catalysed reaction is faster than the background one (compare entry 6 with 5), no competition between both routes should be expected at -27 °C.

Table S4. Background reaction for the aza-Michael addition reaction of hydrazines **1** to nitrostyrene **2a**.^[a]

| Entry | Hydraz. | Solvent (mL) | T (°C) | Time (days) | Yield (%) ^[b] | er (%) ^[c] |
|--------------------|-----------|---------------------------------------|--------|-------------|--------------------------|-----------------------|
| 1 | 1a | CH ₂ Cl ₂ (0.5) | r.t. | 8 | 94 | — |
| 2 | 1b | CH ₂ Cl ₂ (0.5) | r.t. | 8 | 68 | — |
| 3 ^[d] | 1b | CH ₂ Cl ₂ (0.5) | r.t. | 48 h | 70 | 55:45 |
| 4 | 1c | CH ₂ Cl ₂ (0.5) | r.t. | 8 | 91 | — |
| 5 ^[e] | 1b | CH ₃ CN (0.25) | -27 | 7 | 18 | — |
| 6 ^[e,f] | 1b | CH ₃ CN (0.25) | -27 | 84 h | >95 | 88:12 |

[a] Experimental conditions: To a solution of nitrostyrene **2a** (0.1 mmol) in the corresponding solvent, hydrazide **1a-c** (0.15 mmol) was added at the indicated temperature. After a reasonable reaction time, final products were isolated by flash chromatography (Hex:EtOAc, 7:3). Reactions performed in absence of catalyst. [b] Isolated yield. [c] Determined by chiral HPLC analysis. [d] Reaction performed in the presence of 20 mol% of catalyst **3a**. [e] 1 equiv. of hydrazide. [f] Reaction performed in the presence of 10 mol% of catalyst **3a**.

¹H-RMN EXPERIMENTS

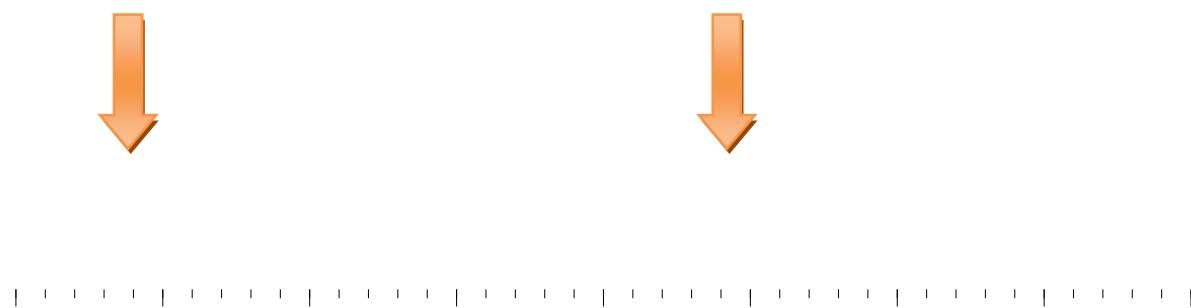
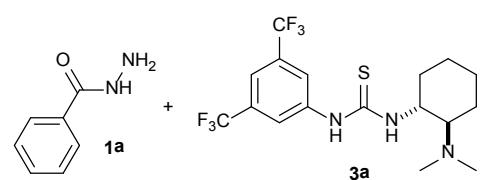


Figure S1. ¹H-NMR experiment performed in CD_3CN with thiourea **3a** and hydrazide **1a** (400 MHz).

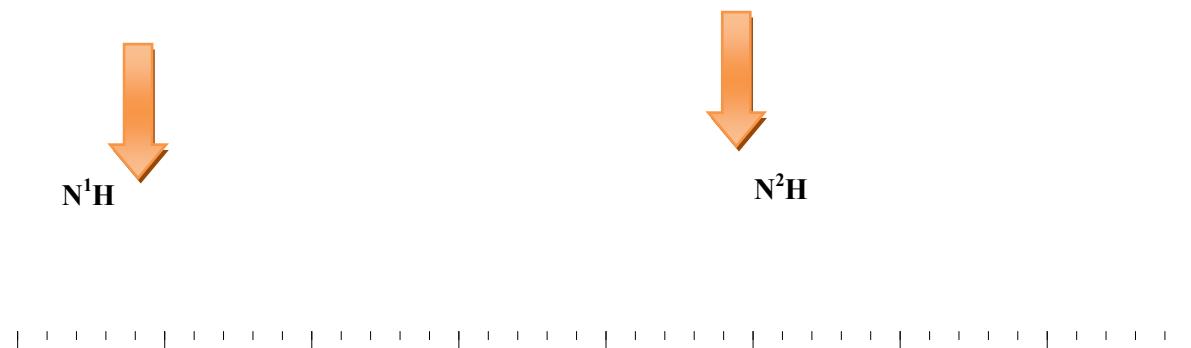
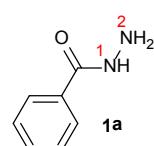


Figure S2. ¹H-NMR experiment performed in CD_3CN with hydrazide **1a** (400 MHz).

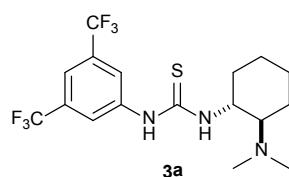
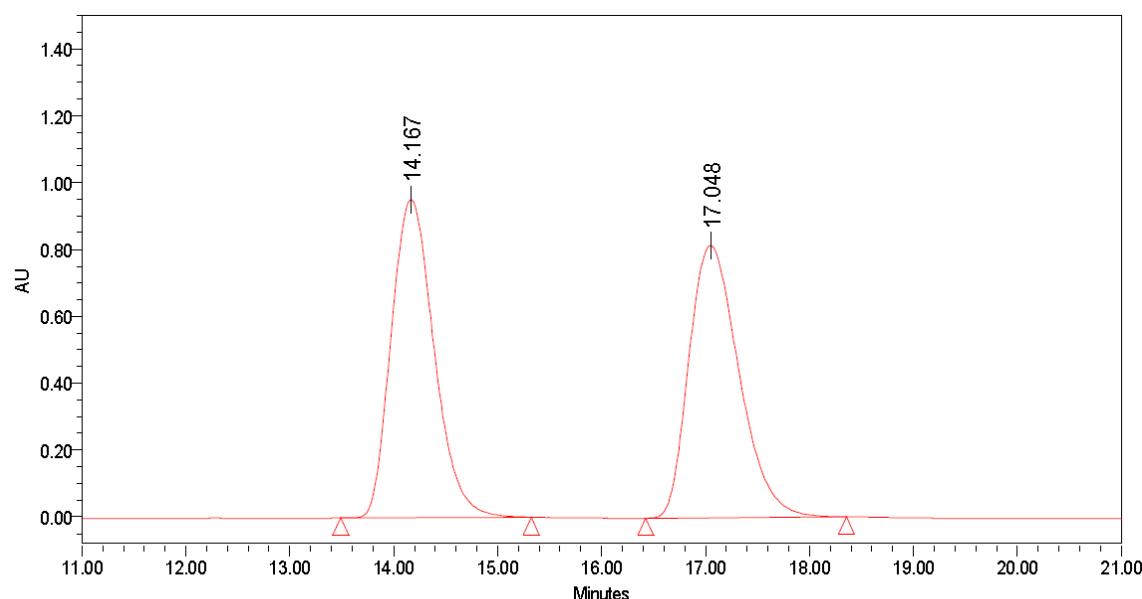


Figure S3. ^1H -NMR experiment performed in CD_3CN with thiourea **3a** (400 MHz).

In order, to support if a deprotonation occurs between the hydrazide **1b** and the catalyst **3a**, we performed the same ^1H -NMR experiments (Figures S1-3) for the less acidic hydrazide **1a** (compared with hydrazide **1b**). First, catalyst **3a** (0.01 mmol), and hydrazide **1a** (0.01 mmol) in CD_3CN (0.5 mL) were analysed separately in two NMR tubes at room temperature (Figures S2 and S3). Subsequently, a mixture of both, catalyst **3a** and hydrazide **1a** in the same concentration in CD_3CN , was analysed by ^1H -NMR (Figure S1). Interestingly, the signal belonging to the N^1H disappears becoming a very broad singlet in the base line of the spectrum and N^2H of the hydrazide does not disappear but it becomes a broader singlet. This fact will support a possible interaction between the NH's of the hydrazide and the catalyst more than a deprotonation, since in this case the deprotonation using hydrazide **1a** is clearly less favourable but the signals do not remain unaltered.

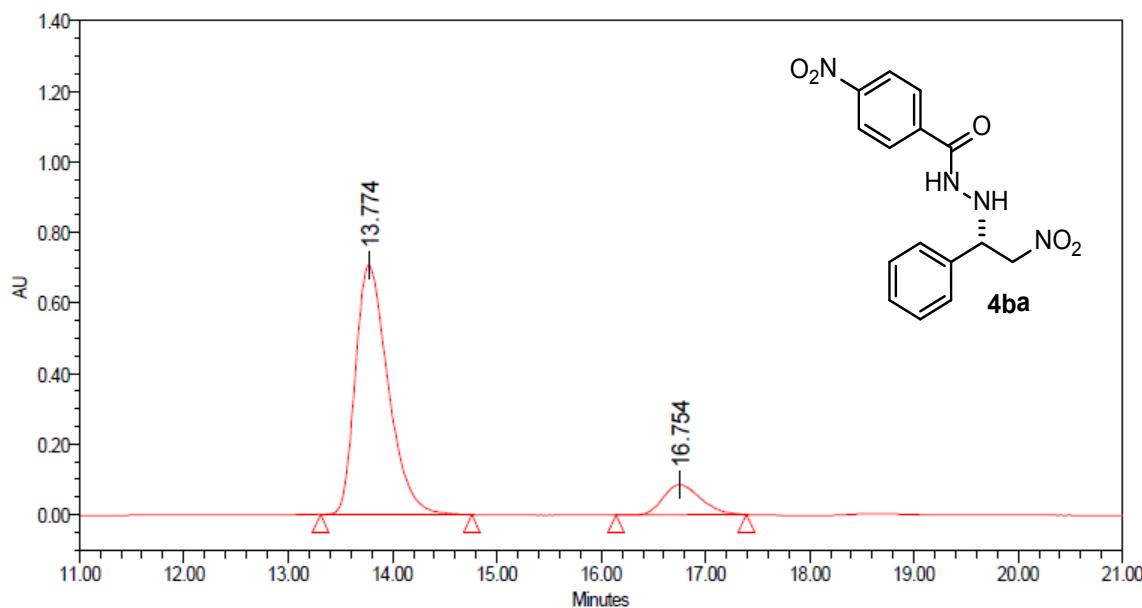
HPLC CROMATOGRAMS OF COMPOUNDS 4ba-bl



Processed Channel: PDA 258.8 nm

| | Processed Channel | Retention Time (min) | Area | % Area | Height |
|---|-------------------|----------------------|----------|--------|--------|
| 1 | PDA 258.8 nm | 14.167 | 27281313 | 50.03 | 952005 |
| 2 | PDA 258.8 nm | 17.048 | 27247520 | 49.97 | 814501 |

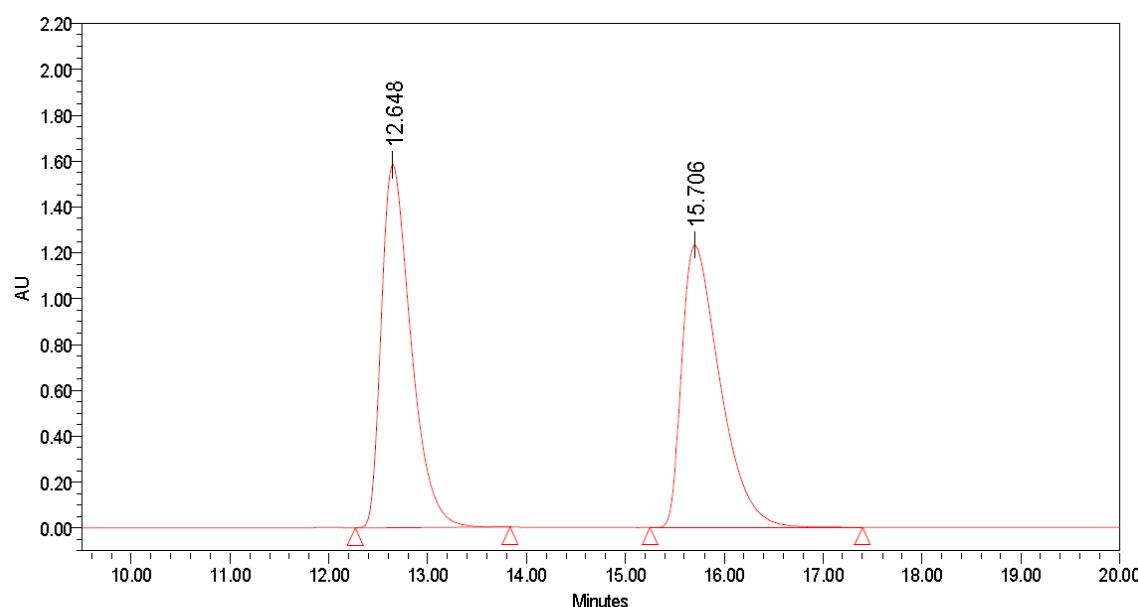
Figure S4. Racemic mixture of **4ba**. Daicel Chiraldex IC column (*n*-hexane/EtOAc = 80:20, flow rate 1 mL/min).



Processed Channel: PDA 258.8 nm

| | Processed Channel | Retention Time (min) | Area | % Area | Height |
|---|-------------------|----------------------|----------|--------|--------|
| 1 | PDA 258.8 nm | 13.774 | 15429883 | 87.78 | 706798 |
| 2 | PDA 258.8 nm | 16.754 | 2147592 | 12.22 | 85677 |

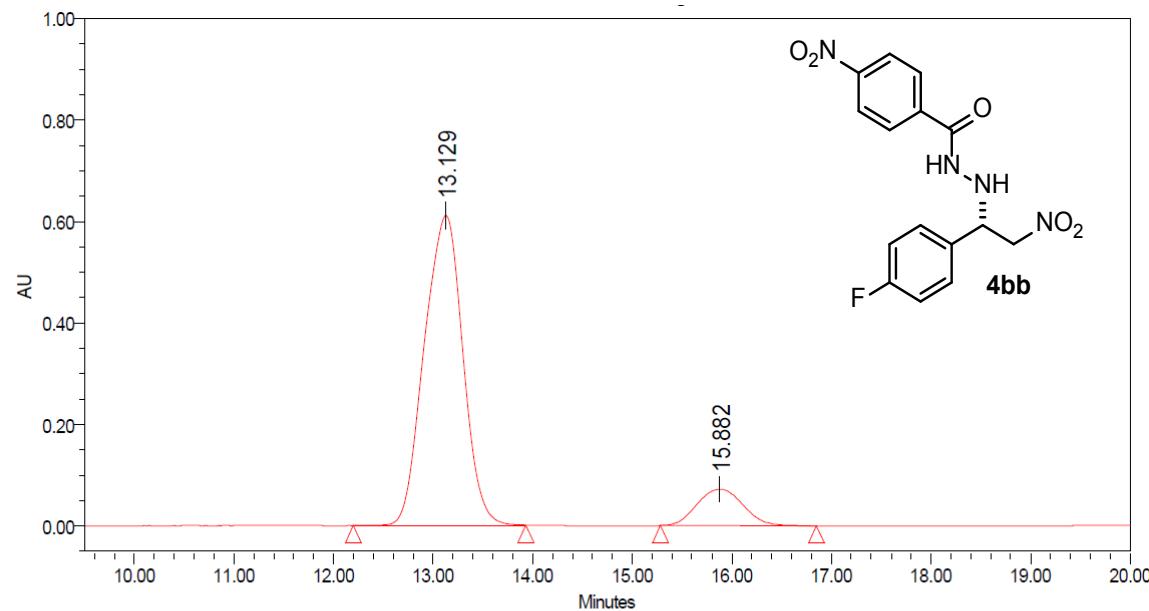
Figure S5. Chiral sample of compound (*S*)-**4ba**.



Processed Channel: PDA 260.0 nm

| | Processed Channel | Retention Time (min) | Area | % Area | Height |
|---|-------------------|----------------------|----------|--------|---------|
| 1 | PDA 260.0 nm | 12.648 | 33095830 | 49.94 | 1583528 |
| 2 | PDA 260.0 nm | 15.706 | 33181412 | 50.06 | 1232497 |

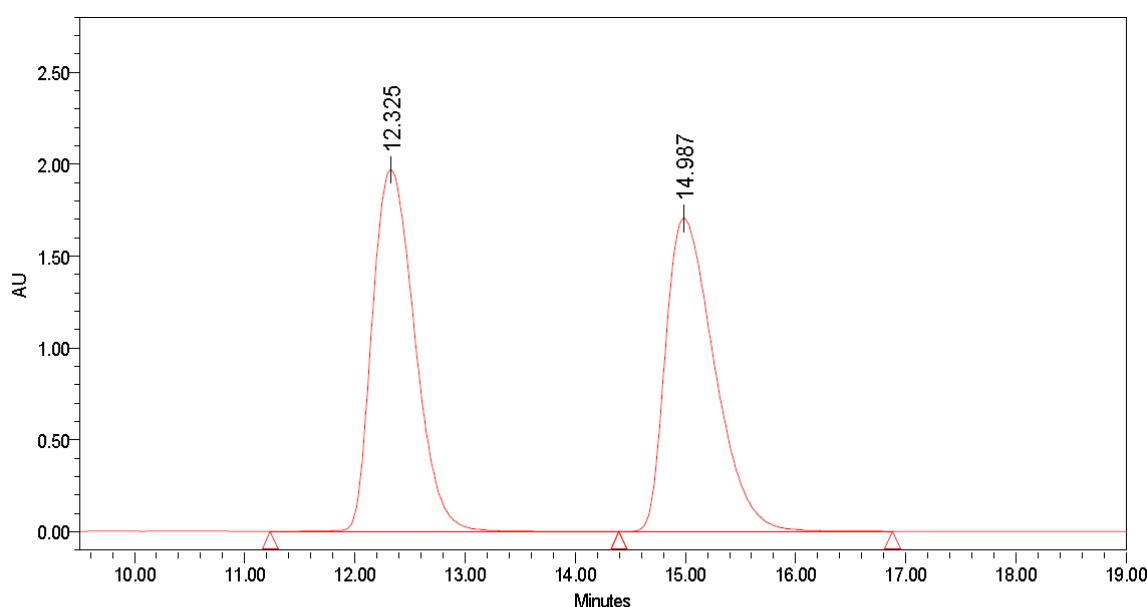
Figure S6. Racemic mixture of **4bb**. Daicel Chiralpak IC column (*n*-hexane/EtOAc = 80:20, flow rate 1 mL/min).



Processed Channel: PDA 260.0 nm

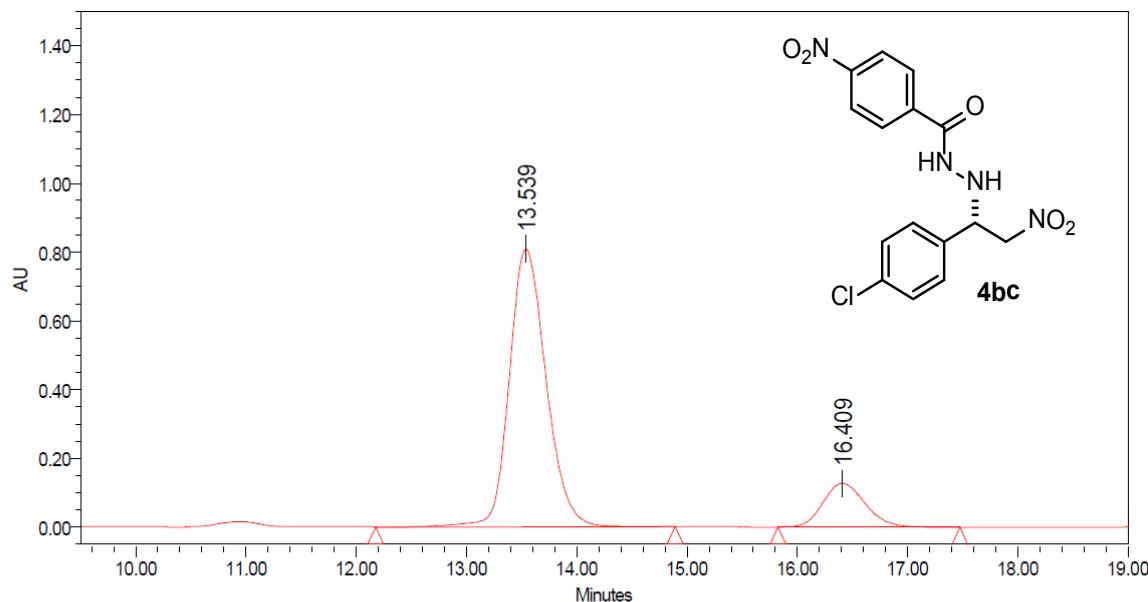
| | Processed Channel | Retention Time (min) | Area | % Area | Height |
|---|-------------------|----------------------|----------|--------|--------|
| 1 | PDA 260.0 nm | 13.129 | 16481791 | 87.85 | 610743 |
| 2 | PDA 260.0 nm | 15.882 | 2279291 | 12.15 | 71775 |

Figure S7. Chiral sample of compound (*S*)-**4bb**.



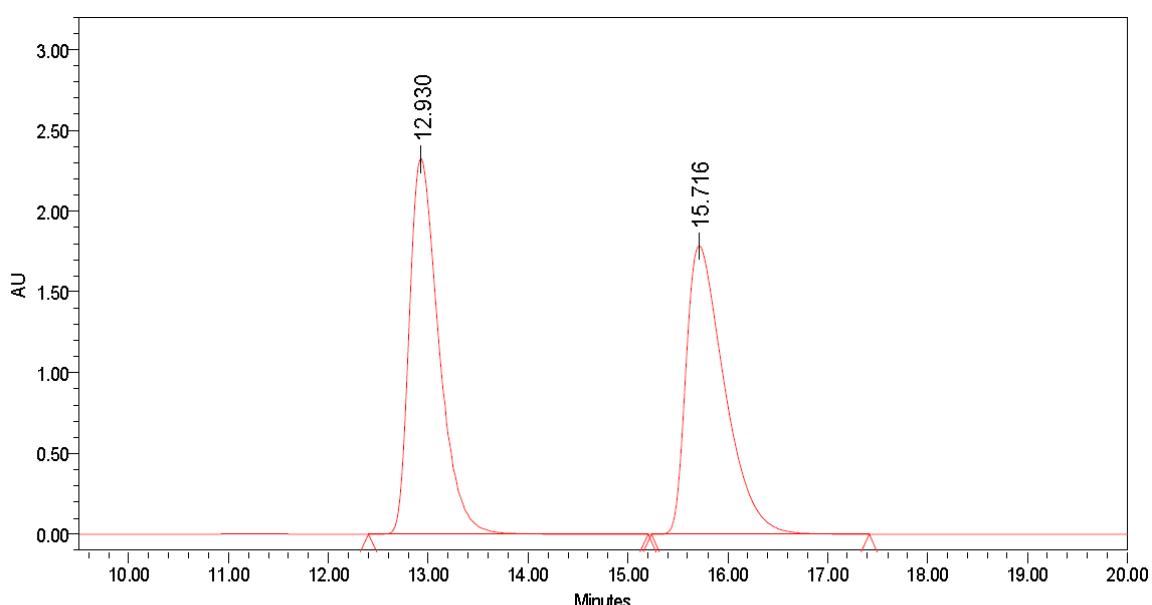
| | Processed Channel | Retention Time (min) | Area | % Area | Height |
|---|-------------------|----------------------|----------|--------|---------|
| 1 | PDA 258.8 nm | 12.325 | 52585626 | 50.08 | 1969868 |
| 2 | PDA 258.8 nm | 14.987 | 52409891 | 49.92 | 1705808 |

Figure S8. Racemic mixture of **4bc**. Daicel Chiraldak IC column (*n*-hexane/EtOAc = 80:20, flow rate 1 mL/min).



| | Processed Channel | Retention Time (min) | Area | % Area | Height |
|---|-------------------|----------------------|----------|--------|--------|
| 1 | PDA 258.8 nm | 13.539 | 18732559 | 84.86 | 808404 |
| 2 | PDA 258.8 nm | 16.409 | 3341180 | 15.14 | 126918 |

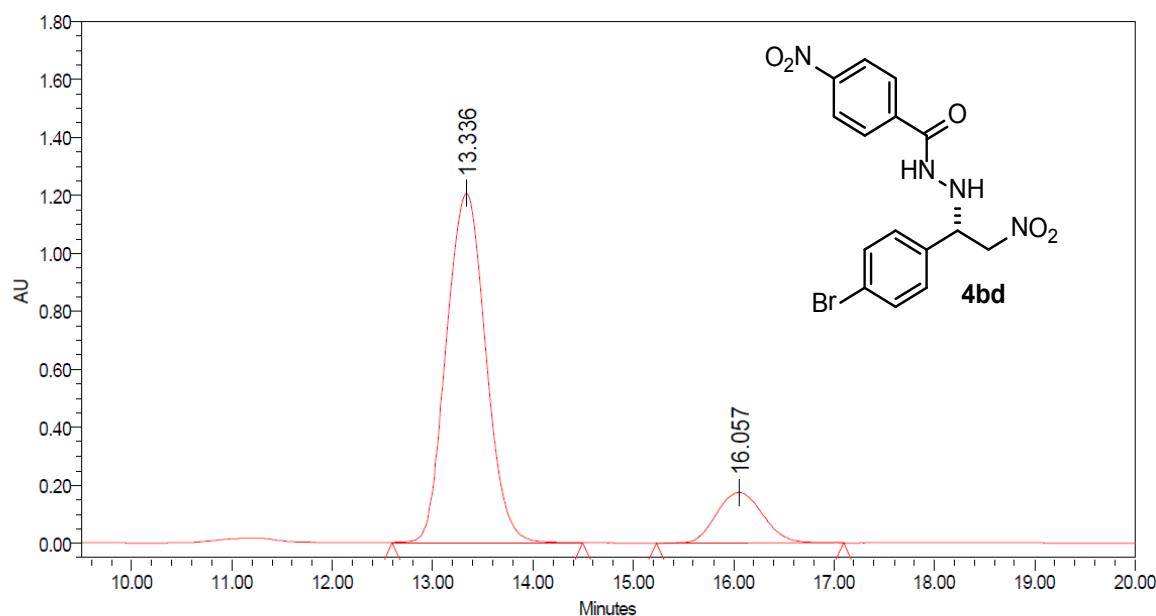
Figure S9. Chiral sample of compound (*S*)-**4bc**.



Processed Channel: PDA 258.8 nm

| | Processed Channel | Retention Time (min) | Area | % Area | Height |
|---|-------------------|----------------------|----------|--------|---------|
| 1 | PDA 258.8 nm | 12.930 | 47902806 | 50.03 | 2326800 |
| 2 | PDA 258.8 nm | 15.716 | 47853605 | 49.97 | 1787406 |

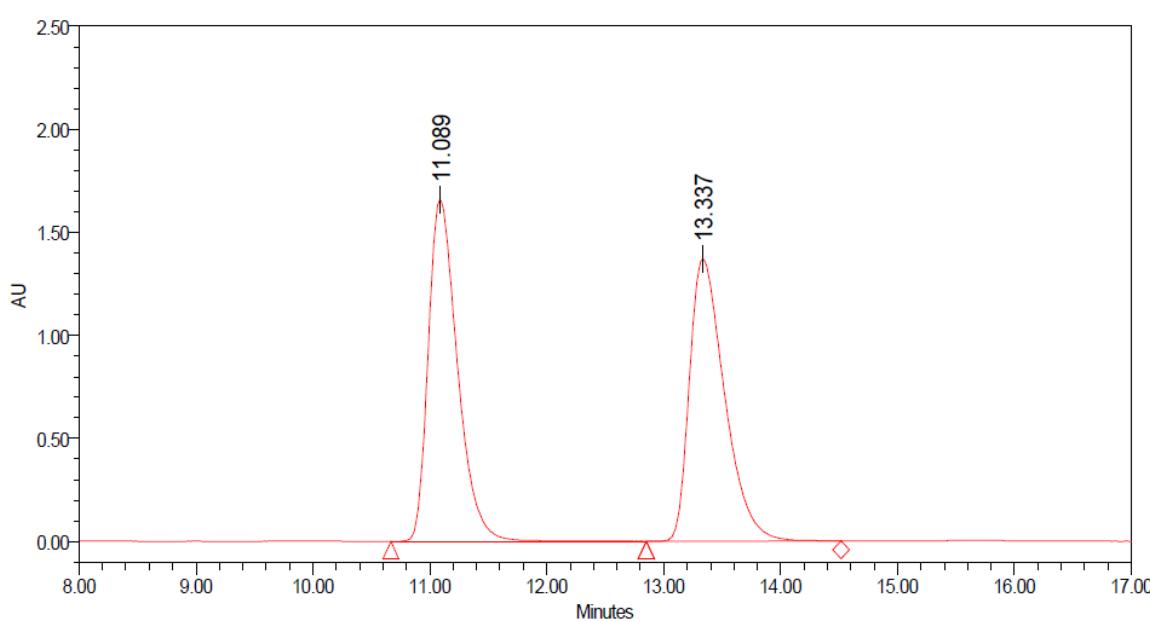
Figure S10. Racemic mixture of **4bd**. Daicel Chiraldpak IC column (*n*-hexane/EtOAc = 80:20, flow rate 1 mL/min).



Processed Channel: PDA 258.8 nm

| | Processed Channel | Retention Time (min) | Area | % Area | Height |
|---|-------------------|----------------------|----------|--------|---------|
| 1 | PDA 258.8 nm | 13.336 | 32888243 | 85.52 | 1205292 |
| 2 | PDA 258.8 nm | 16.057 | 5566538 | 14.48 | 174056 |

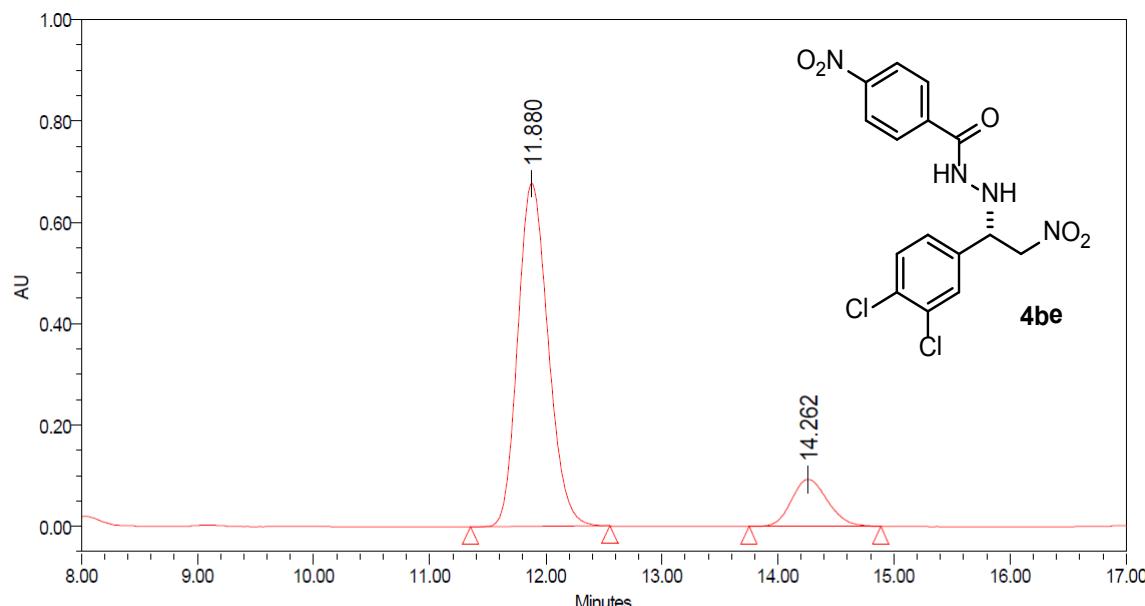
Figure S11. Chiral sample of compound (*S*)-4bd.



Processed Channel: PDA 258.8 nm

| | Processed Channel | Retention Time (min) | Area | % Area | Height |
|---|-------------------|----------------------|----------|--------|---------|
| 1 | PDA 258.8 nm | 11.089 | 28782672 | 50.08 | 1661031 |
| 2 | PDA 258.8 nm | 13.337 | 28690952 | 49.92 | 1371165 |

Figure S12. Racemic mixture of **4be**. Daicel Chiraldpak IC column (*n*-hexane/EtOAc = 80:20, flow rate 1 mL/min).



Processed Channel: PDA 258.8 nm

| | Processed Channel | Retention Time (min) | Area | % Area | Height |
|---|-------------------|----------------------|----------|--------|--------|
| 1 | PDA 258.8 nm | 11.880 | 12240193 | 85.98 | 677085 |
| 2 | PDA 258.8 nm | 14.262 | 1996160 | 14.02 | 92786 |

Figure S13. Chiral sample of compound (*S*)-**4be**.

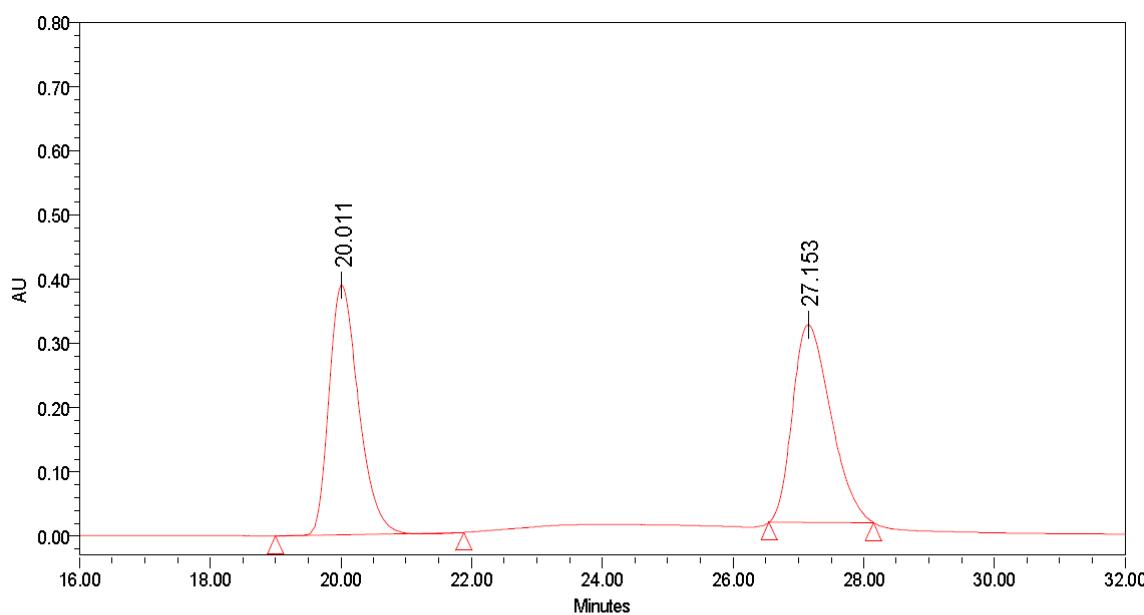


Figure S14. Racemic mixture of **4bf**. Daicel Chiraldak IC column (*n*-hexane/EtOAc = 80:20, flow rate 1 mL/min).

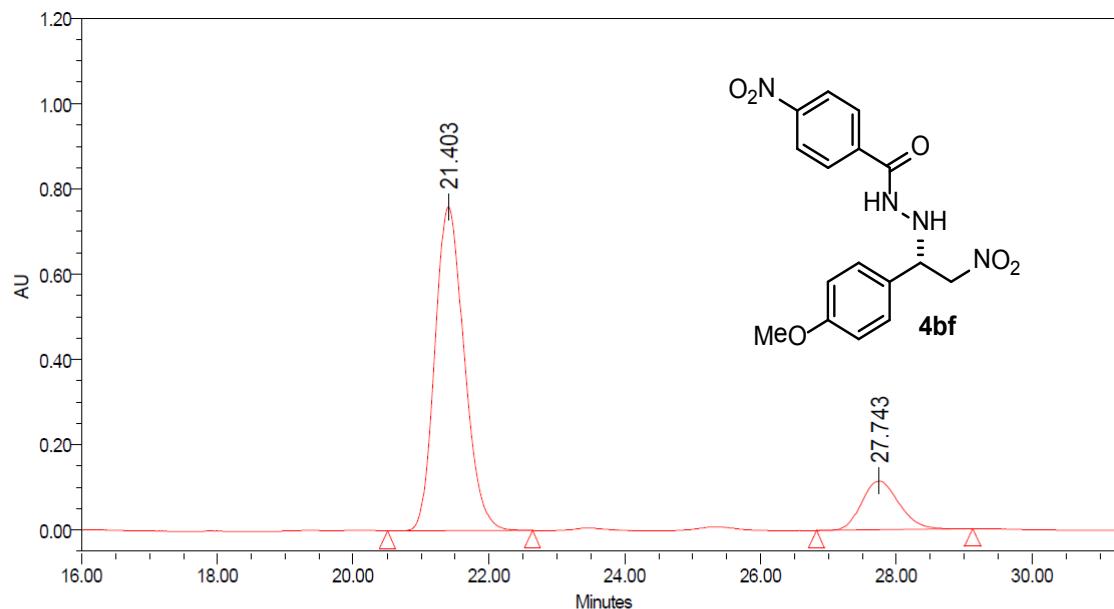
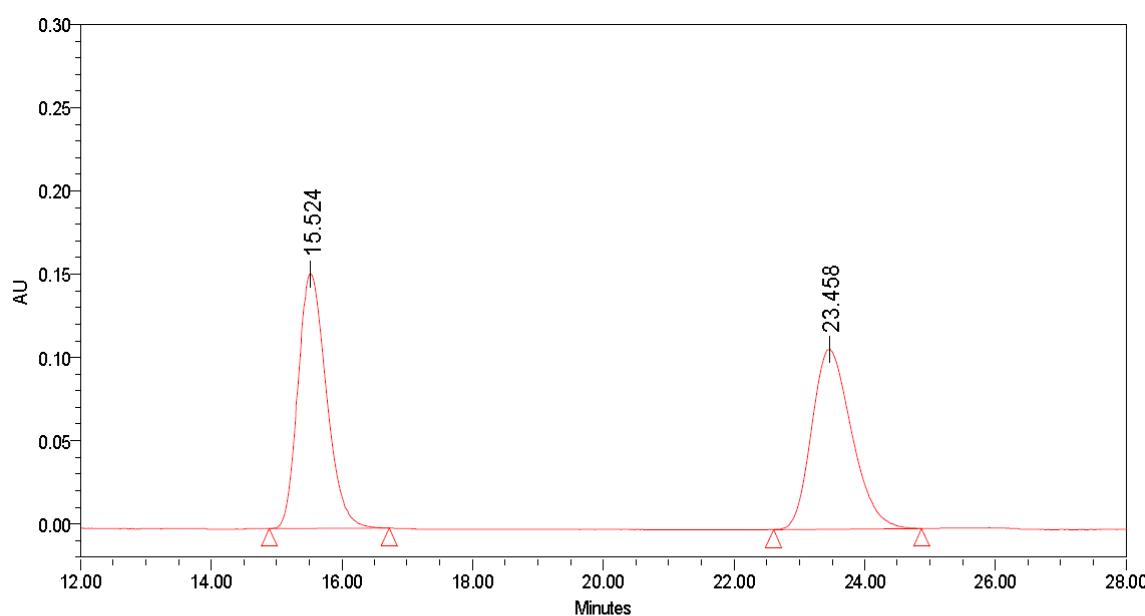


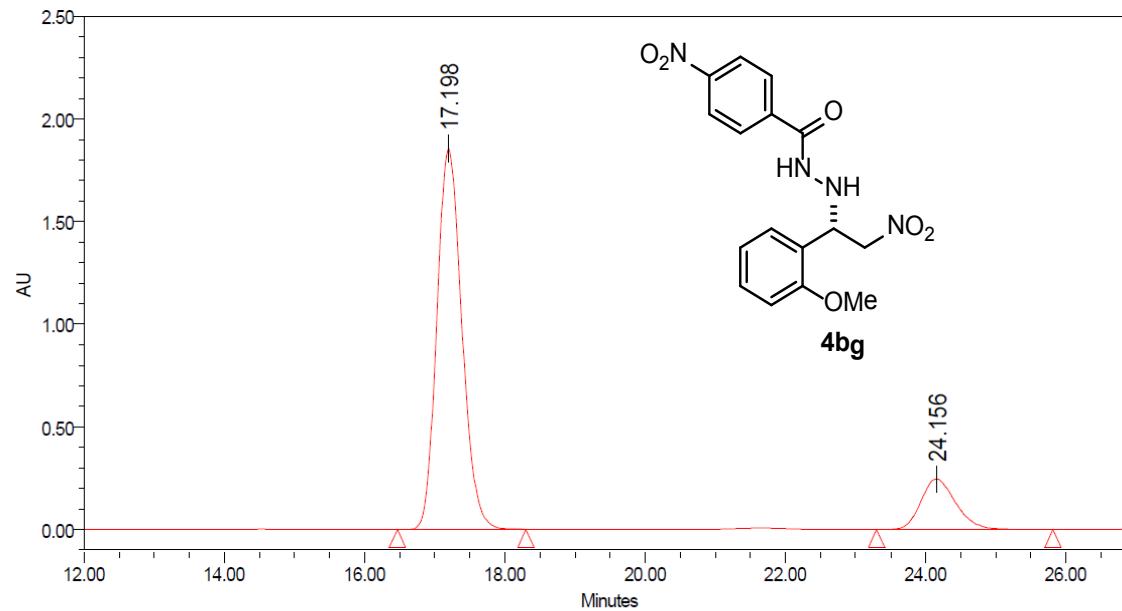
Figure S15. Chiral sample of compound (*S*)-**4bf**.



Processed Channel: PDA 262.4 nm

| | Processed Channel | Retention Time (min) | Area | % Area | Height |
|---|-------------------|----------------------|---------|--------|--------|
| 1 | PDA 262.4 nm | 15.524 | 4643265 | 50.20 | 153044 |
| 2 | PDA 262.4 nm | 23.458 | 4607086 | 49.80 | 108001 |

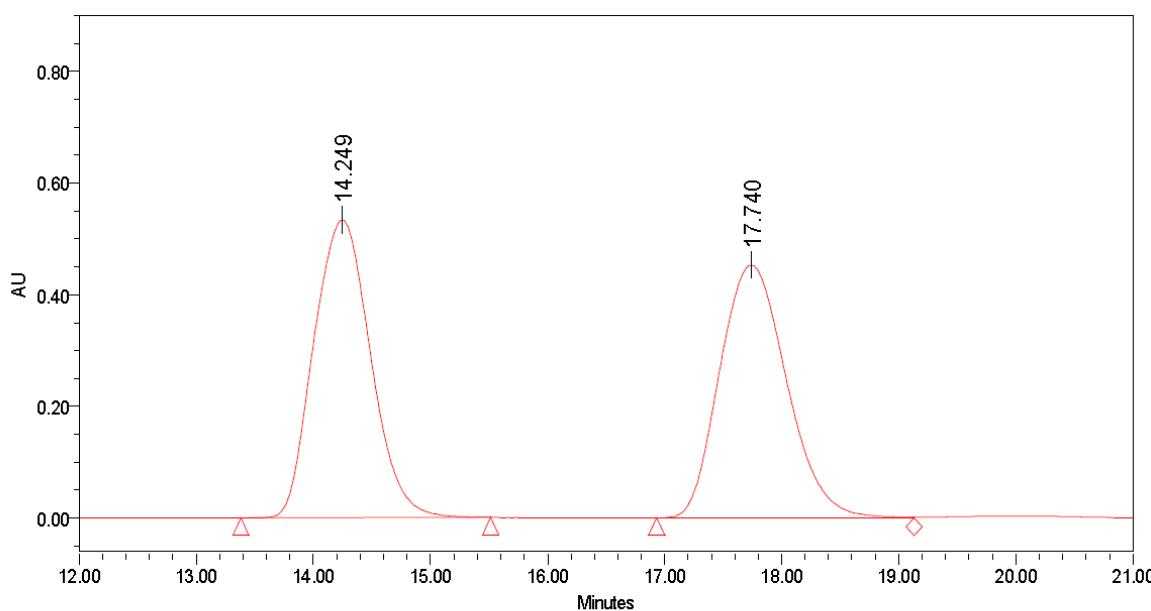
Figure S16. Racemic mixture of **4bg**. Daicel Chiraldpak IC column (*n*-hexane/EtOAc = 80:20, flow rate 1 mL/min).



Processed Channel: PDA 262.4 nm

| | Processed Channel | Retention Time (min) | Area | % Area | Height |
|---|-------------------|----------------------|----------|--------|---------|
| 1 | PDA 262.4 nm | 17.198 | 45955714 | 84.45 | 1855358 |
| 2 | PDA 262.4 nm | 24.156 | 8464751 | 15.55 | 246044 |

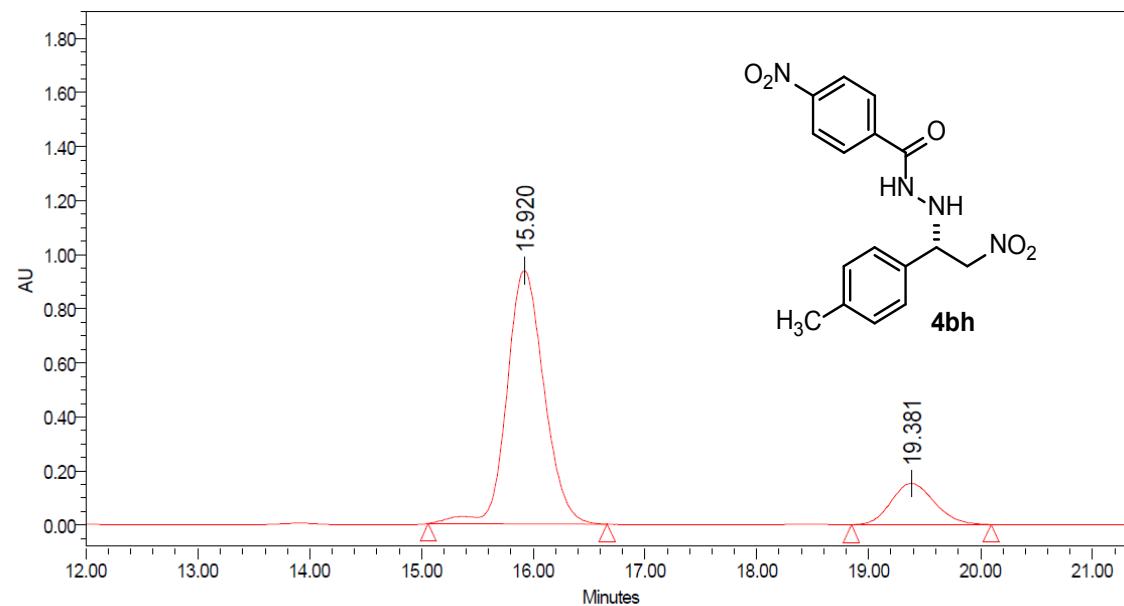
Figure S17. Chiral sample of compound (*S*)-**4bg**.



Processed Channel: PDA 260.0 nm

| | Processed Channel | Retention Time (min) | Area | % Area | Height |
|---|-------------------|----------------------|----------|--------|--------|
| 1 | PDA 260.0 nm | 14.249 | 17979659 | 50.08 | 533398 |
| 2 | PDA 260.0 nm | 17.740 | 17924412 | 49.92 | 452351 |

Figure S18. Racemic mixture of **4bh**. Daicel Chiralpak IC column (*n*-hexane/EtOAc = 80:20, flow rate 1 mL/min).



Processed Channel: PDA 260.0 nm

| | Processed Channel | Retention Time (min) | Area | % Area | Height |
|---|-------------------|----------------------|----------|--------|--------|
| 1 | PDA 260.0 nm | 15.920 | 21672667 | 84.16 | 938517 |
| 2 | PDA 260.0 nm | 19.381 | 4077907 | 15.84 | 152398 |

Figure S19. Chiral sample of compound (S)-4bh.

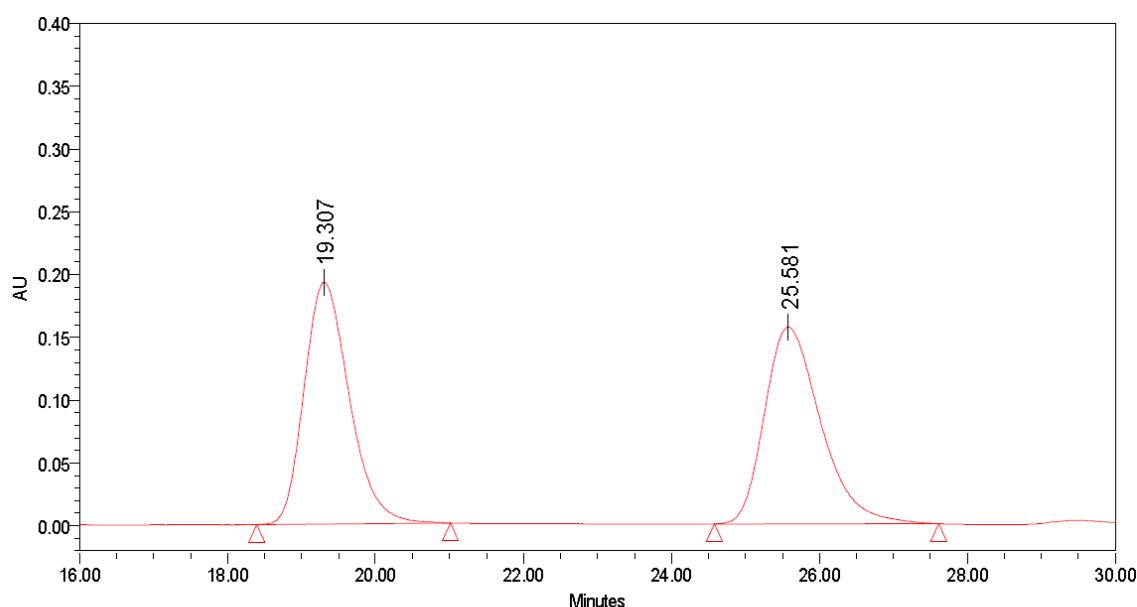


Figure S20. Racemic mixture of **4bi**. Daicel Chiraldpak IC column (*n*-hexane/EtOAc = 80:20, flow rate 1 mL/min).

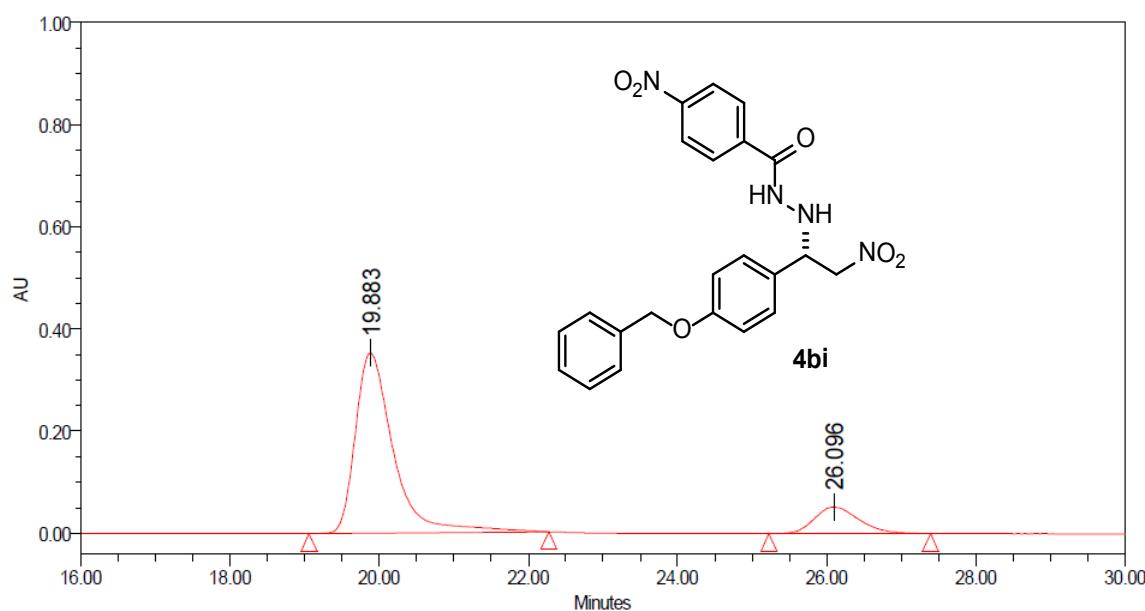
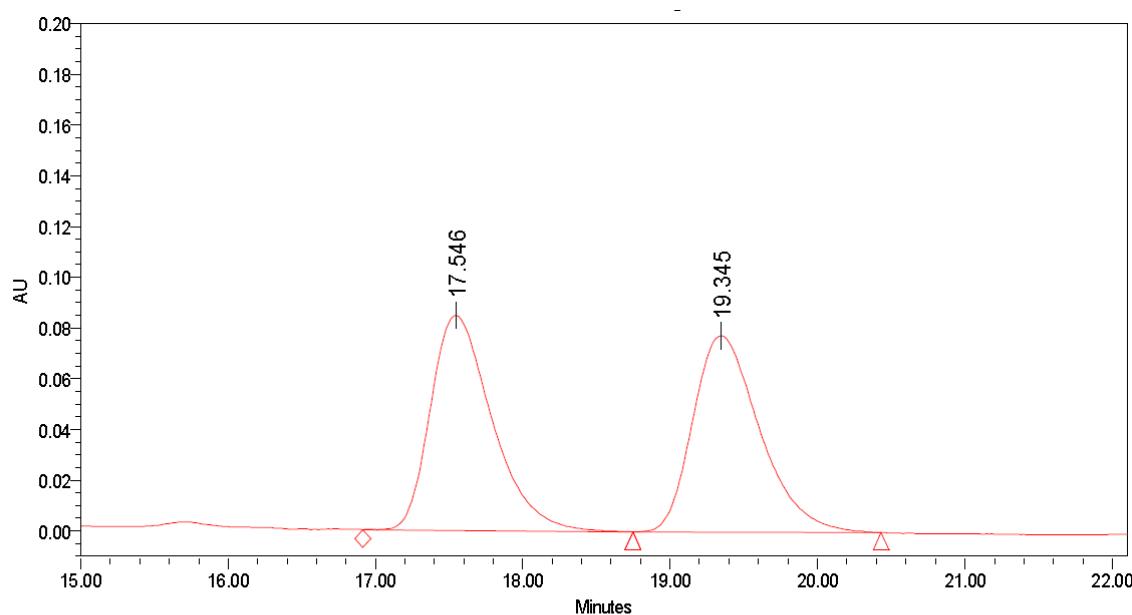


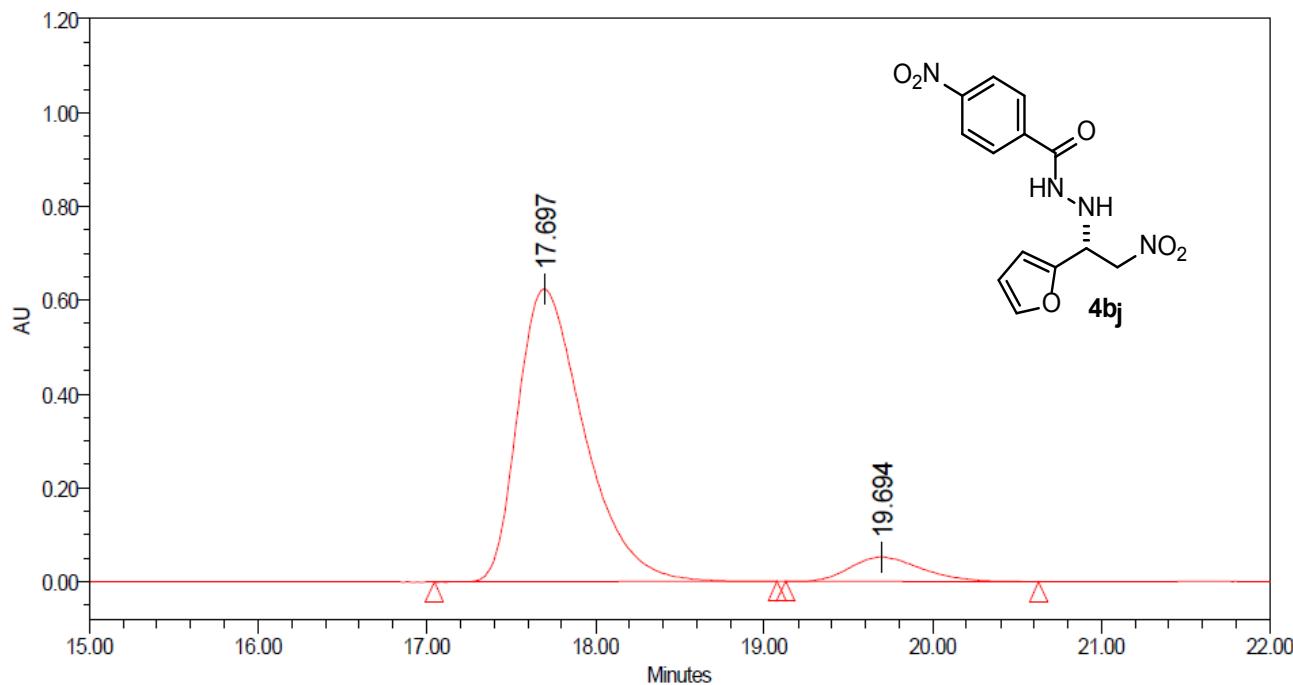
Figure S21. Chiral sample of compound (*S*)-**4bi**.



Processed Channel: PDA 258.8 nm

| | Processed Channel | Retention Time (min) | Area | % Area | Height |
|---|-------------------|----------------------|---------|--------|--------|
| 1 | PDA 258.8 nm | 17.546 | 2471201 | 50.32 | 84916 |
| 2 | PDA 258.8 nm | 19.345 | 2439618 | 49.68 | 77441 |

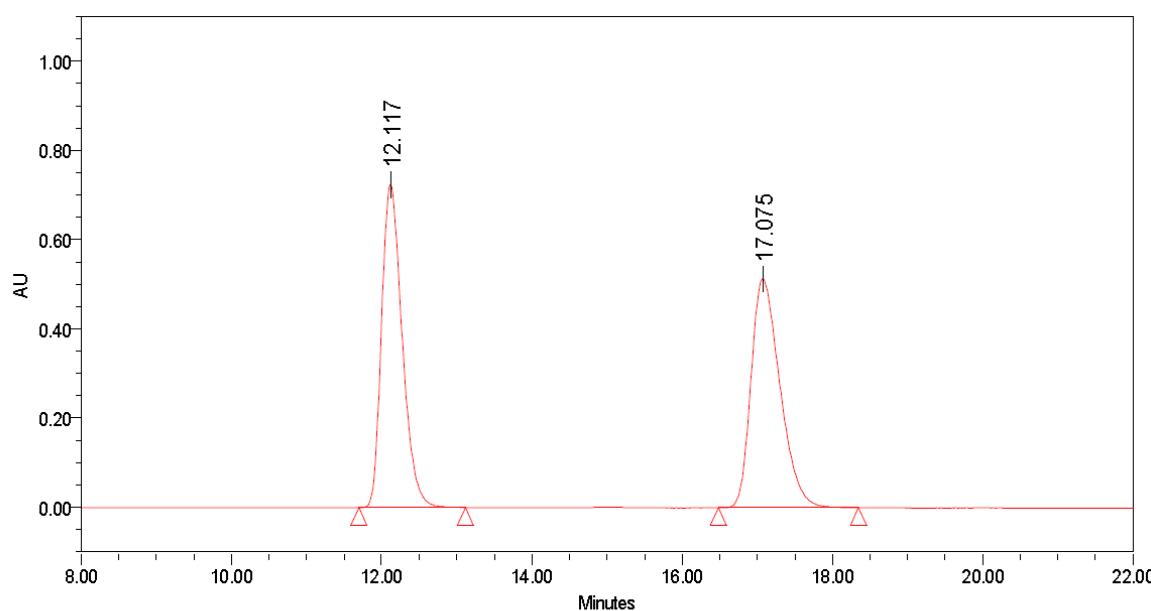
Figure S22. Racemic mixture of **4bj**. Daicel Chiralpak IC column (*n*-hexane/EtOAc = 80:20, flow rate 1 mL/min).



Processed Channel: PDA 258.8 nm

| | Processed Channel | Retention Time (min) | Area | % Area | Height |
|---|-------------------|----------------------|----------|--------|--------|
| 1 | PDA 258.8 nm | 17.697 | 17159347 | 91.70 | 625003 |
| 2 | PDA 258.8 nm | 19.694 | 1552558 | 8.30 | 52253 |

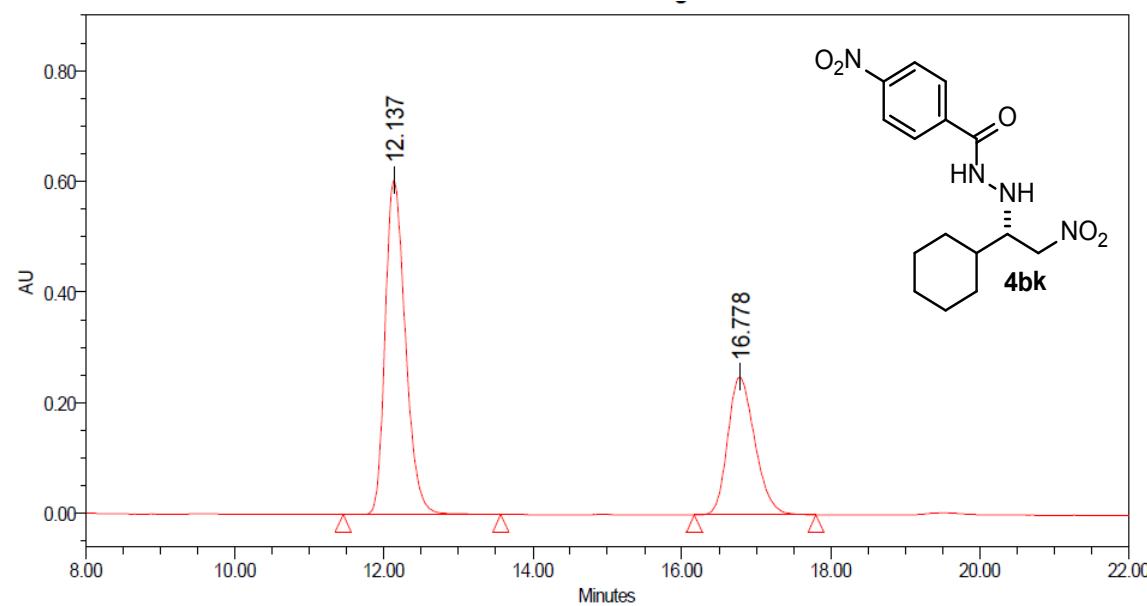
Figure S23. Chiral sample of compound (*S*)-**4bj**.



Processed Channel: PDA 258.8 nm

| | Processed Channel | Retention Time (min) | Area | % Area | Height |
|---|-------------------|----------------------|----------|--------|--------|
| 1 | PDA 258.8 nm | 12.117 | 13751895 | 50.05 | 724094 |
| 2 | PDA 258.8 nm | 17.075 | 13726466 | 49.95 | 512952 |

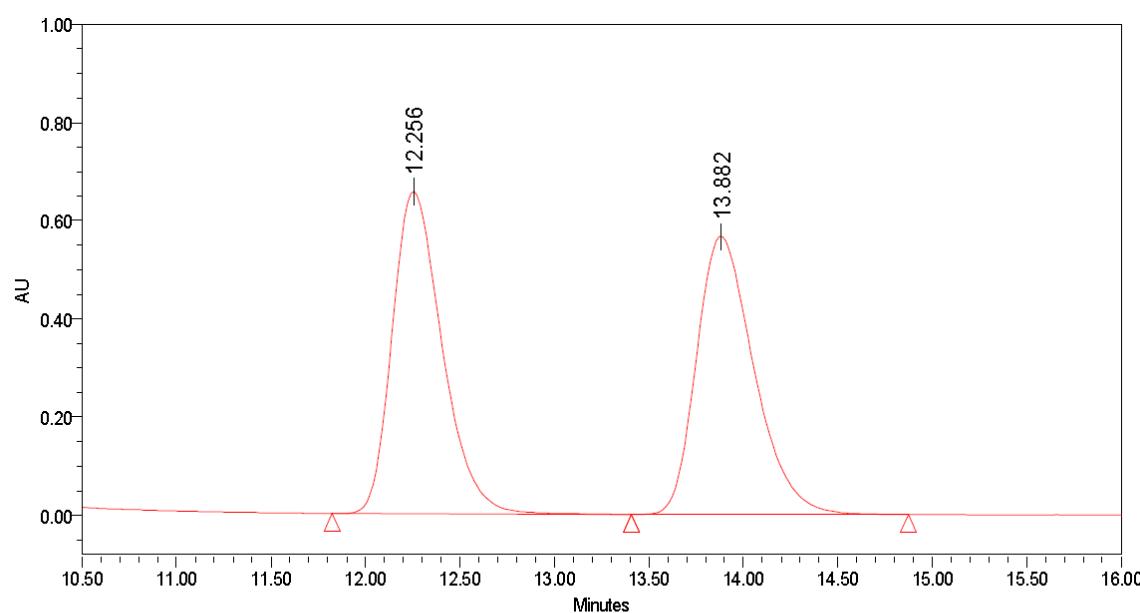
Figure S24. Racemic mixture of **4bk**. Daicel Chiralpak IC column (*n*-hexane/EtOAc = 80:20, flow rate 1 mL/min).



Processed Channel: PDA 258.8 nm

| | Processed Channel | Retention Time (min) | Area | % Area | Height |
|---|-------------------|----------------------|----------|--------|--------|
| 1 | PDA 258.8 nm | 12.137 | 11386921 | 64.37 | 604942 |
| 2 | PDA 258.8 nm | 16.778 | 6302012 | 35.63 | 249410 |

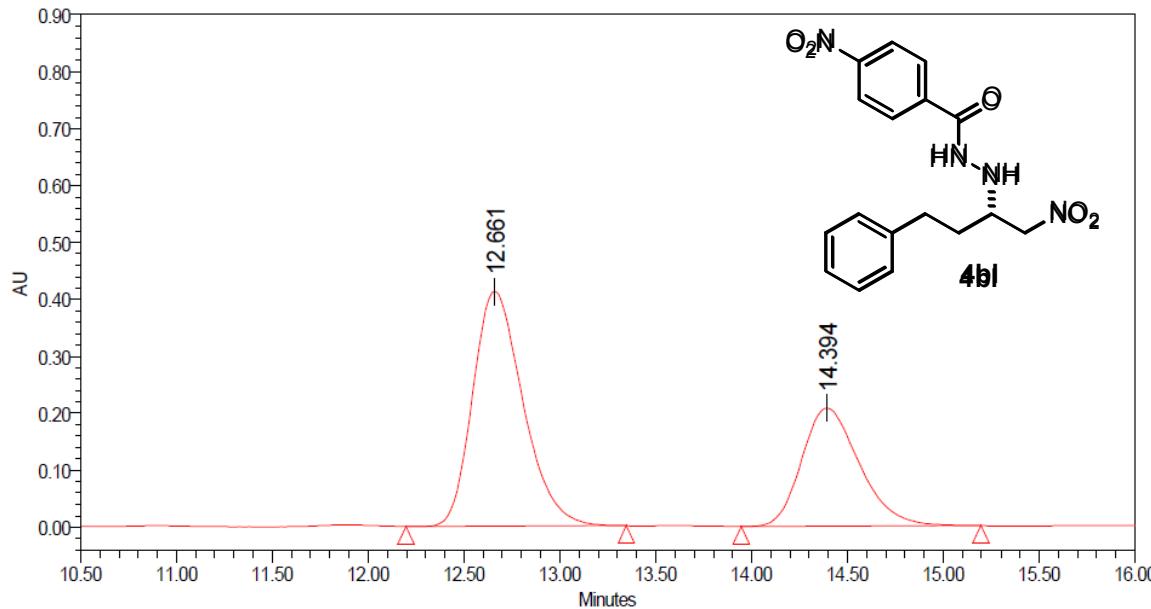
Figure S25. Chiral sample of compound (*S*)-4bk.



Processed Channel: PDA 258.8 nm

| | Processed Channel | Retention Time (min) | Area | % Area | Height |
|---|-------------------|----------------------|----------|--------|--------|
| 1 | PDA 258.8 nm | 12.256 | 12056055 | 50.16 | 656886 |
| 2 | PDA 258.8 nm | 13.882 | 11977637 | 49.84 | 566832 |

Figure S26. Racemic mixture of **4bl**. Daicel Chiralpak IC column (*n*-hexane/EtOAc = 80:20, flow rate 1 mL/min).



Processed Channel: PDA 258.8 nm

| | Processed Channel | Retention Time (min) | Area | % Area | Height |
|---|-------------------|----------------------|---------|--------|--------|
| 1 | PDA 258.8 nm | 12.661 | 7517318 | 63.36 | 412648 |
| 2 | PDA 258.8 nm | 14.394 | 4347250 | 36.64 | 207456 |

Figure S27. Chiral sample of compound (*S*)-**4bl**.

NMR SPECTRA OF COMPOUNDS 4ba-bl

(*S*)-4-Nitro-*N'*-(2-nitro-1-phenylethyl)benzohydrazide (4ba)



(S)-N'-(1-(4-Fluorophenyl)-2-nitroethyl)-4-nitrobenzohydrazide (4bb)



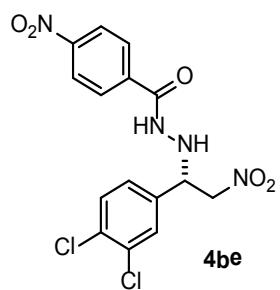
(S)-N'-(1-(4-Chlorophenyl)-2-nitroethyl)-4-nitrobenzohydrazide (4bc)



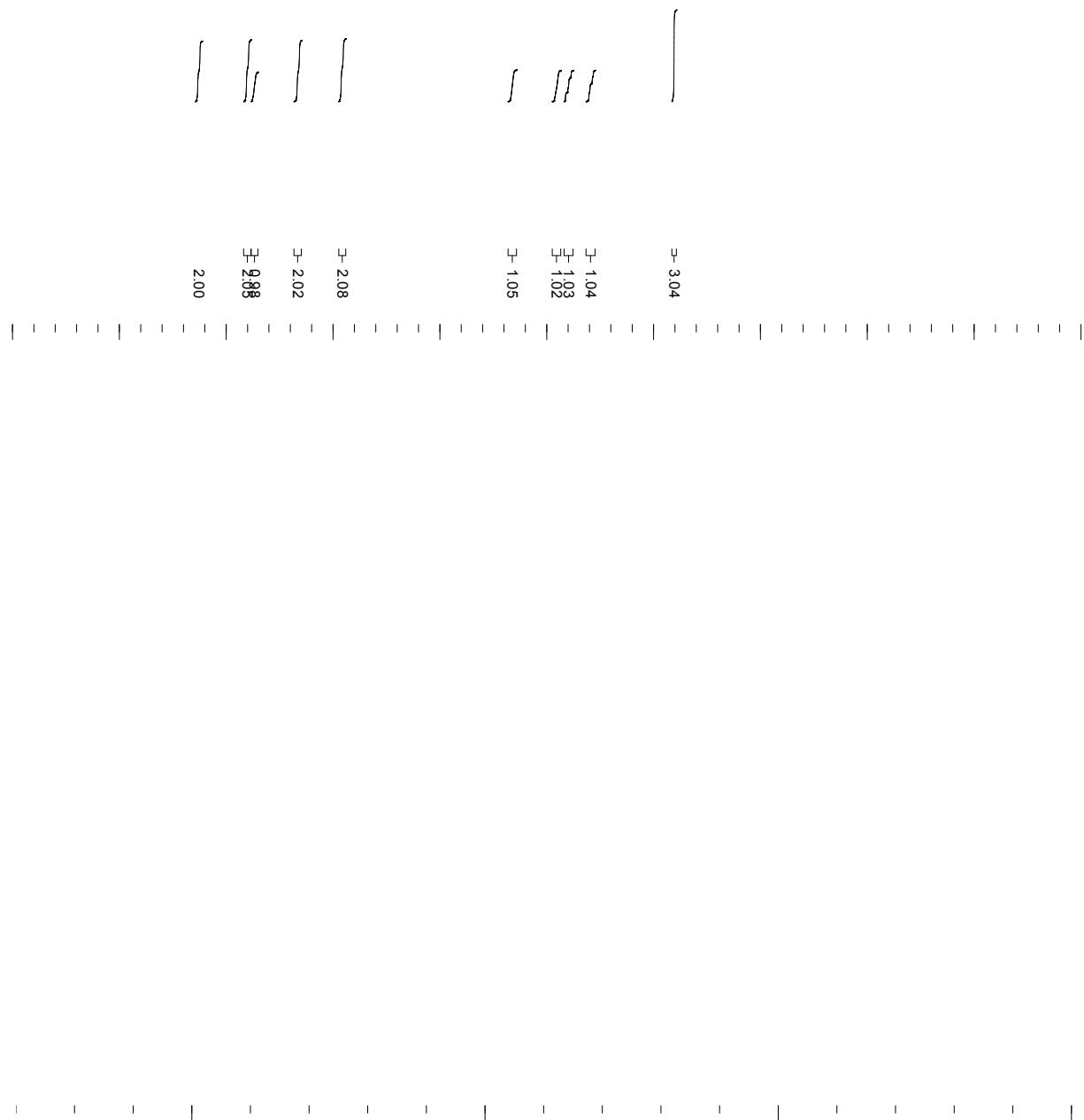
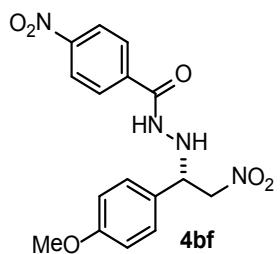
(*S*)-*N'*-(1-(4-Bromophenyl)-2-nitroethyl)4-nitrobenzohydrazide (**4bd**)



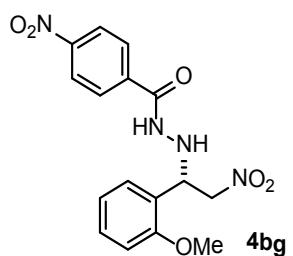
(*S*)-*N'*-(1-(3,4-Dichlorophenyl)-2-nitroethyl)-4-nitrobenzohydrazide (**4be**)



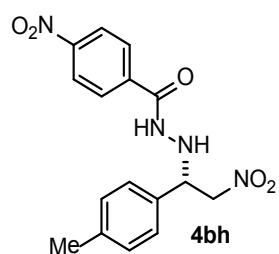
(*S*)-*N'*-(1-(4-Methoxyphenyl)-2-nitroethyl)-4-nitrobenzohydrazide (**4bf**)



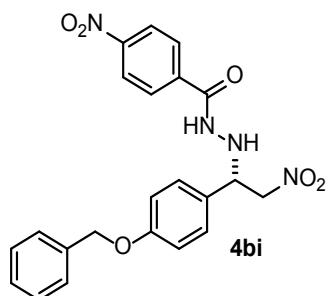
(*S*)-*N'*-(1-(2-Methoxyphenyl)-2-nitroethyl)-4-nitrobenzohydrazide (**4bg**)



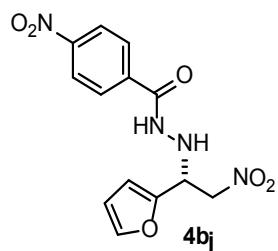
(*S*)-4-Nitro-*N'*-(2-nitro-1-p-tolylethyl)benzohydrazide (**4bh**)



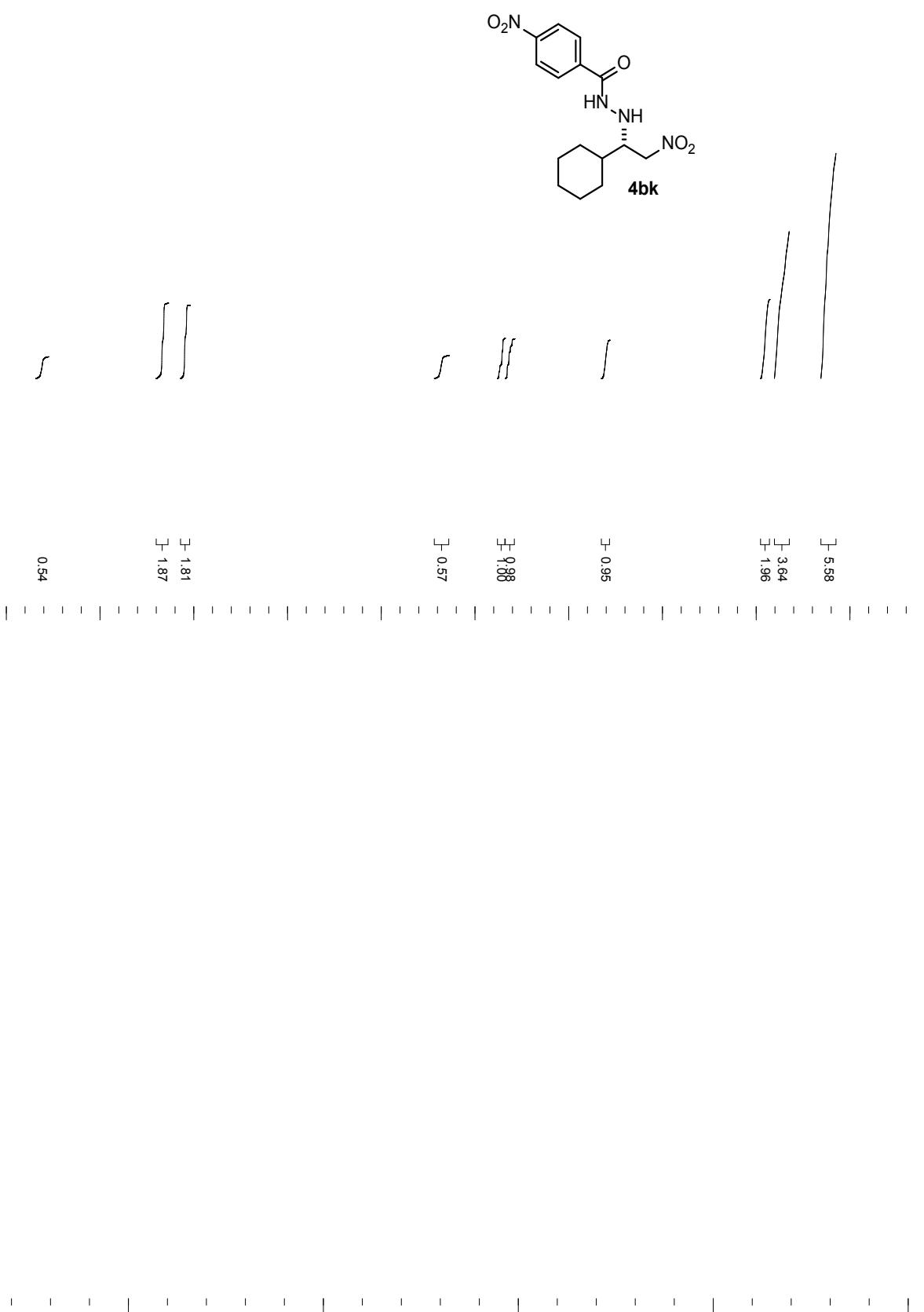
(S)-N'-(1-(4-(Benzyl)phenyl)-2-nitroethyl)-4-nitrobenzohydrazide (4bi)



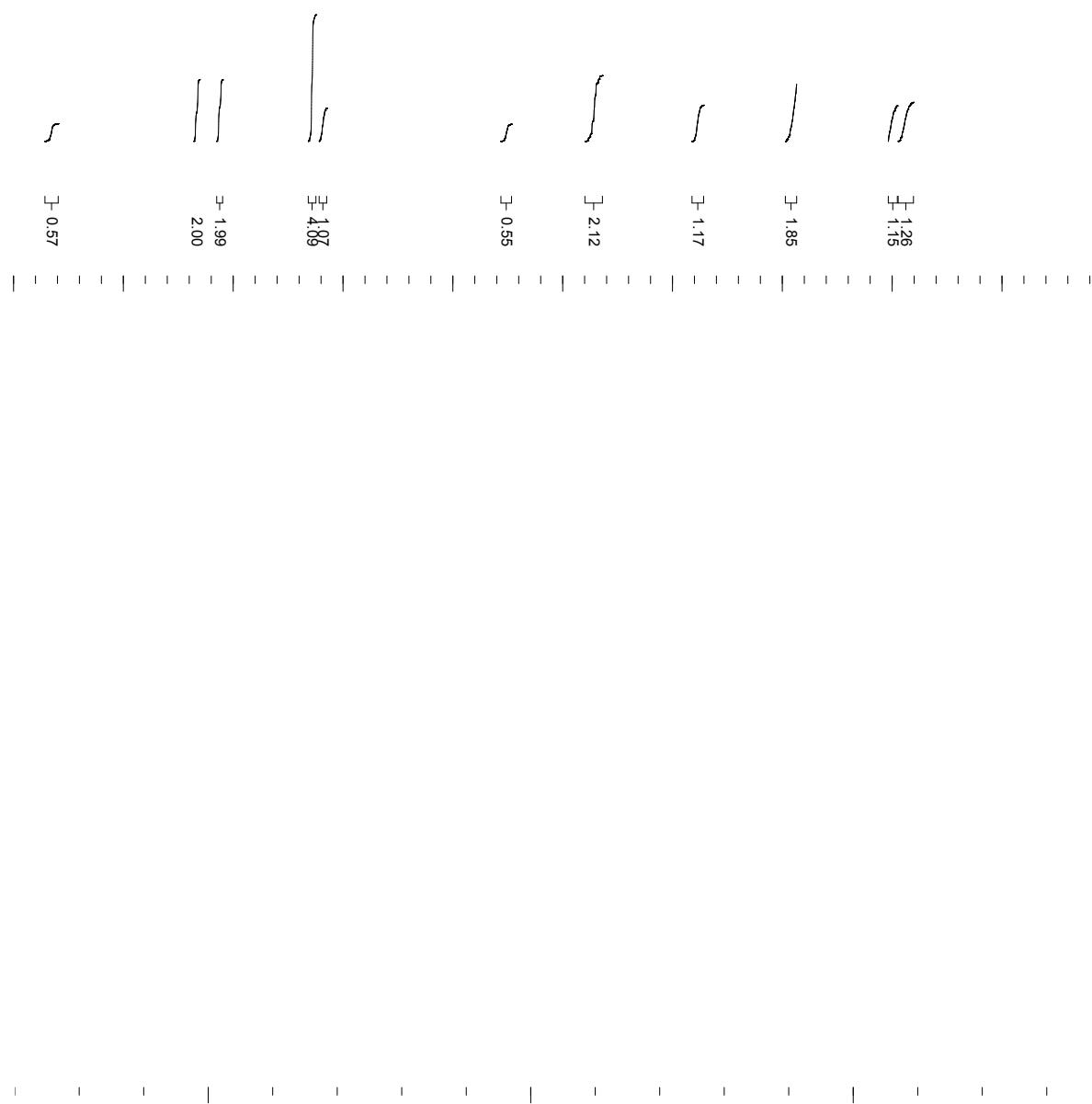
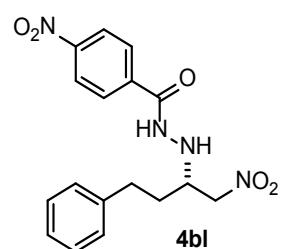
(S)-N'-(1-(Furan-2-yl)-2-nitroethyl)-4-nitrobenzohydrazide (4bj)



(S)-N'-(1-Cyclohexyl-2-nitroethyl)-4-nitrobenzohydrazide (4bk)

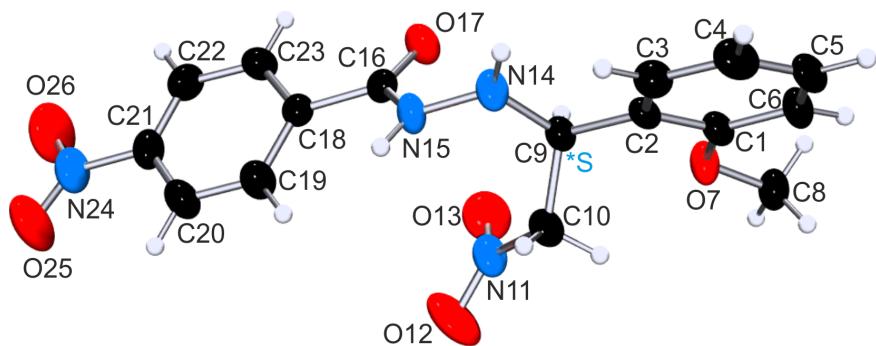


(S)-4-Nitro-N'-(1-nitro-4-phenylbutan-2-yl)benzohydrazide (4bl)

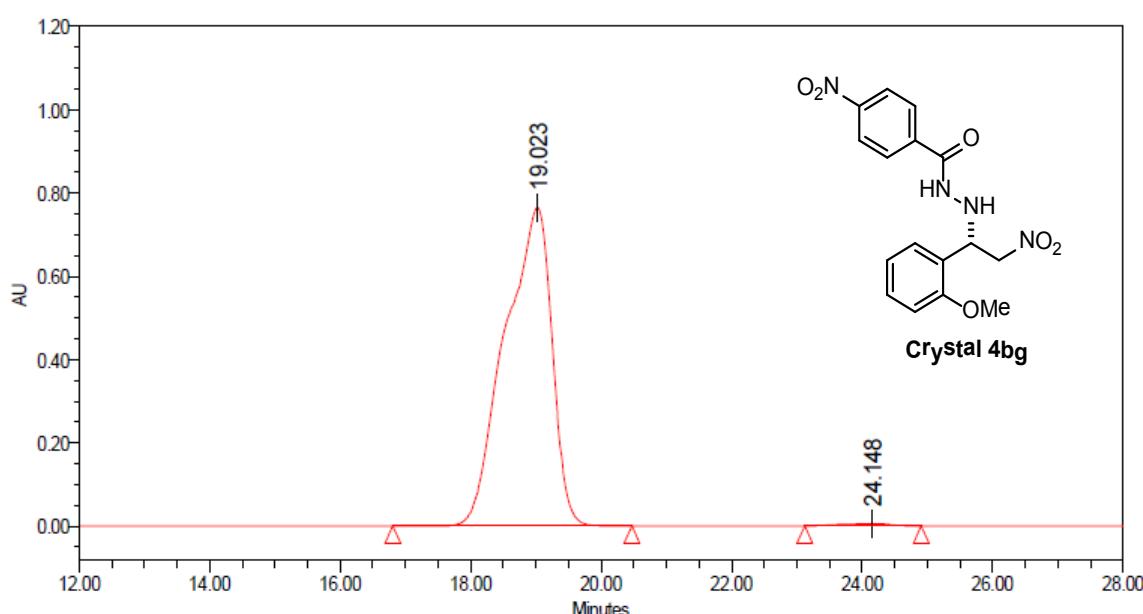


X-RAY CRYSTALLOGRAPHY

The crystals for compound **4bg** were obtained by slow evaporation of a cool AcOEt solution maintained in the fridge during two weeks. The purity of the crystals was determined by HPLC using a Daicel Chiralpak IC column (*n*-hexane/EtOAc = 80:20, flow rate 1 mL/min, λ = 262.4 nm): $t_{\text{major}} = 19.0$ min; $t_{\text{minor}} = 24.1$ min. $[\alpha]_D^{25} = +23.4$ ($c = 0.10$, acetone, 99:1 er). The e.r. of the crystals was found to be >99:1. M.p. for crystallized compound **4bg** is 140–144 °C.



Crystal data of (*S*)-**4bg**: $[\text{C}_{16}\text{H}_{16}\text{N}_4\text{O}_6]$, monoclinic, $P21$, $a = 5.0171(2)$ Å, $b = 16.1781(6)$ Å, $c = 10.3154(4)$ Å, $\beta = 102.463(2)^\circ$, $Z = 2$, $M_r = 360.33$ g mol⁻¹, $V = 817.54(5)$ Å³, $D_{\text{calcd}} = 1.464$ g cm⁻³, $\lambda(\text{Cu K}\alpha) = 1.54178$ Å, $T = 295$ K, $\mu = 0.970$ mm⁻¹, 2471 reflections collected, 2342 unique, $R1(F_o) = 0.0598$ [$I > 2\sigma(I)$], $wR2 (F_o^2) = 0.1503$ (all data), GOF = 1.038. CCDC-959956.



Processed Channel: PDA 262.4 nm

| | Processed Channel | Retention Time (min) | Area | % Area | Height |
|---|-------------------|----------------------|----------|--------|--------|
| 1 | PDA 262.4 nm | 19.023 | 38209576 | 99.15 | 764560 |
| 2 | PDA 262.4 nm | 24.148 | 329183 | 0.85 | 6118 |

Figure S28. Crystal of compound (*S*)-4bg. Daicel Chiraldpak IC column (*n*-hexane/EtOAc = 80:20, flow rate 1 mL/min).