

Supporting Information

Substrate and catalytic promiscuity of secondary metabolite enzymes: O-prenylation of hydroxyxanthenes with different prenyl donors by a bisindolyl benzoquinone C- and N-prenyltransferase

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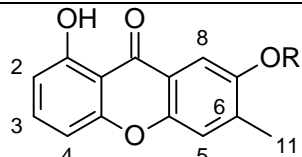
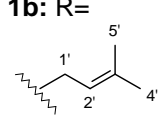
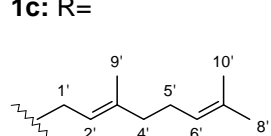
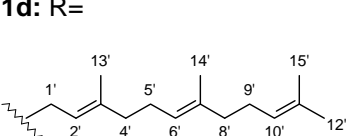
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Table S1. Sequence identities of AstPT and homologues.

Enzyme	TdiB	AstPT	XptB	FgaPT2	FtmPT1	7-DMATS	AnaPT	CdpNPT
CdpNPT	25	24	24	31	28	25	32	100
AnaPT	26	23	26	31	30	30	100	
7-DMATS	23	26	27	31	26	100		
FtmPT1	20	23	26	36	100			
FgaPT2	26	25	27	100				
XptB	25	23	100					
AstPT	45	100						
TdiB	100							

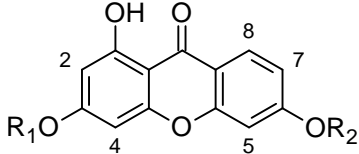
Sequence identities are given in percentage. TdiB (ABU51603.1) and XptB (BN001302.1) are from *Aspergillus nidulans*, AstPT (EAU29429.1) from *Aspergillus terreus*, FgaPT2 (AAX08549.1), FtmPT1 (AAX56314.1), 7-DMATS (ABS89001.1) and CdpNPT (ABR14712.1) are from *Aspergillus fumigatus*, AnaPT (EAW16181.1) is from *Neosartorya fischeri*.

Table S2. ¹H NMR data of **1a** and enzyme products **1b-1d**, (CD₃)₂CO, 500 MHz.

Compound				
	1a: R= H	1b: R= 	1c: R= 	1d: R= 
Proton	δ_H , mult., J	δ_H , mult., J	δ_H , mult., J	δ_H , mult., J
OH-1	12.83, s	12.82, s	12.82, s	12.82, s
2	6.74, dd, 8.3, 0.9	6.77, dd, 8.3, 0.9	6.77, dd, 8.3, 0.8	6.77, dd, 8.3, 0.9
3	7.67, t, 8.3	7.69, t, 8.3	7.69, t, 8.3	7.69, t, 8.3
4	6.98, dd, 8.3, 0.9	7.01, dd, 8.3, 0.9	7.01, dd, 8.3, 0.8	7.01, dd, 8.3, 0.9
5	7.40, q, 0.9	7.46, q, 0.8	7.46, br s	7.46, br s
OH-7	12.83, s	-	-	-
8	7.58, s	7.57, s	7.57, s	7.56, s
11	2.37, d, 0.9	2.39, d, 0.8	2.39, s	2.39, d, 0.8
1'	-	4.75, d, 6.6	4.78, d, 6.7	4.79, d, 6.5
2'	-	5.54, t, 6.6	5.54, td, 6.5, 1.2	5.53, td, 6.4, 1.2
4'	-	1.83, br s ^a	2.14, m	2.15, m
5'	-	1.81, br s ^a	2.14, m	2.15, m
6'	-	-	5.11, t, 6.6	5.13, t, 6.6
8'	-	-	1.62, br s ^b	2.02, m
9'	-	-	1.85, s	2.02, m
10'	-	-	1.59, s ^b	5.03, t, 7.0
12'	-	-	-	1.62, br s ^c
13'	-	-	-	1.86, s
14'	-	-	-	1.60, s
15'	-	-	-	1.55, s ^c

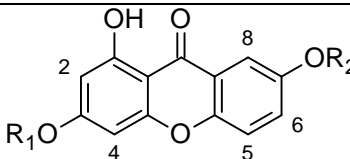
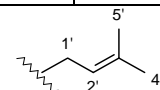
J values are given in Hz. ^{a-c}: Signals with same letters are interchangeable.

Table S3. ¹H NMR data of **2a** and enzyme products **2b1-2d2**, (CD₃)₂CO, 500 MHz.

Compound							
	2a R ₁ = R ₂ = H	2b1 R ₁ =	2b2 R ₂ =	2c1 R ₁ =	2c2 R ₂ =	2d1 R ₁ =	2d2 R ₂ =
Proton	δ _H , mult., J	δ _H , mult., J	δ _H , mult., J	δ _H , mult., J	δ _H , mult., J	δ _H , mult., J	δ _H , mult., J
OH-1	13.12, s	-	13.06, s	13.03, s	-	13.05, s	-
2	6.23, d, 2.2	6.29, d, 2.3	6.24, d, 2.1	6,10, d, 2,0	6.21, d, 2.2	6.19, d, 2.1	6.25, d, 2.3
OH-3	13.12, s	-	13.06, s	-	-	-	-
4	6.39, d, 2.2	6.49, d, 2.3	6.41, d, 2.1	6.25, d, 2.0	6.42, d, 2.2	6.35, d, 2.1	6.45, d, 2.3
5	6.88, d, 2.3	6.87, d, 2.2	7.01, s	6.94, s	6.60, d ^d	6.97, d, 2.3	6.76, d, 2.1
OH-6	13.12, s	-	-	13.03, s	-	13,05, s	-
7	6.96, dd, 8.7, 2.3	6.95, dd, 8.8, 2.2	7.01, dd, 8.6, 2.4	6.93, dd, 7.9, 2.0	6.74, dd ^d	6.97, dd, 9.5, 2.3	6.87, dd, 8.6, 2.1
8	8.05, d, 8.7	8.03, d, 8.8	8.07, dd, 8.6, 0.5	8.02, m	7.90, d, 8.7	8.05, d	7.98, d, 9.1
1'	-	4.70, d, 6.5	4.76, d, 6.7	4.76, d, 6.5	4.69, d, 6.8	4.78, d, 6.3	4.72, d, 6.1
2'	-	5.50, t, 6.5	5.53, t, 6.7	5.52, t, 6.5	- ^e	5.53, t, 6.3	- ^f
4'	-	1.78, s	1.80, s	2.14, m	- ^e	2.14, m	- ^f
5'	-	1.78, s	1.80, s	2.14, m	- ^e	2.14, m	- ^f
6'	-	-	-	5.11, t, 6.9	- ^e	5.13, t, 6.8	- ^f
8'	-	-	-	1.63, br s ^a	1.57, s ^b	1.94, m	- ^f
9'	-	-	-	1.80, br s	1.79, s	1.94, m	- ^f
10'	-	-	-	1.59, br s ^a	1.56, s ^b	5.05, t, 7.0	- ^f
12'	-	-	-	-	-	1.63, br s ^c	1.65, s
13'	-	-	-	-	-	1.81, br s	1.80, d, 1.2
14'	-	-	-	-	-	1.60, br s	1.59, s
15'	-	-	-	-	-	1.56, br s ^c	1.56, s ^f

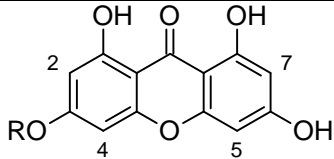
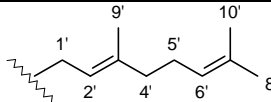
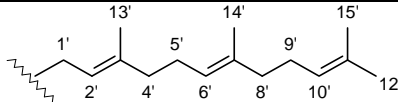
J values are given in Hz. ^{a-c}: Signals with same letters are interchangeable. ^d: Due to low product amount coupling constants could not be determined. ^e: Signals overlap with those of **2c1**. ^f: Signals overlap with those of **2d1**.

Table S4. ^1H NMR data of **3a** and enzyme products **3b1** and **3b2**, $(\text{CD}_3)_2\text{CO}$, 500 MHz.

Compound			
	3a $\text{R}_1 = \text{R}_2 = \text{H}$	3b1 $\text{R}_1 =$	3b2 $\text{R}_2 =$
			
Proton	δ_{H} , mult., J	δ_{H} , mult., J	δ_{H} , mult., J
OH-1	12.99, s	-	12.96, s
2	6.25, d, 2.1	6.33, d, 2.3	6.28, d, 2.1
OH-3	12.99, s	-	12.96, s
4	6.41, d, 2.1	6.53, d, 2.3	6.45, d, 2.1
5	7.45, d, 9.0	7.47, dd, 9.1, 0.4	7.50, dd, 9.1, 0.4
6	7.36, dd, 9.0, 3.0	7.38, dd, 9.1, 3.0	7.42, dd, 9.1, 3.1
OH-7	12.99 s	-	-
8	7.57, d, 3.0	7.58, d, 3.0	7.60, d, 3.1
1'	-	4.72, d, 6.9	4.70, d, 6.6
2'	-	5.35, t, 5.2	5.50, t, 6.6
4'	-	1.81 ^a	1.81, s ^a
5'	-	1.79 ^a	1.79, s ^a

J values are given in Hz. ^a: Signals are interchangeable.

Table S5. ^1H NMR data of **4a** and enzyme products **4c** and **4d**, $(\text{CD}_3)_2\text{CO}$, 500 MHz.

Compound			
	4a R = H	4c R = 	4d R = 
Proton	δ_{H} , mult., J	δ_{H} , mult., J	δ_{H} , mult., J
OH-1	11.99, s	-	12.16, s
2	6.25, d, 2.2	6.33, d, 2.3 ^a	6.28, d, 2.3 ^d
OH-3	11.99, s	-	-
4	6.38, d, 2.2	6.51, d, 2.3 ^b	6.46, d, 2.3 ^e
5	6.38, d, 2.2	6.41, d, 2.1 ^b	6.32, d, 1.9 ^e
OH-6	11.99, s	-	11.91, s
7	6.25, d, 2.2	6.26, d, 2.1 ^a	6.18, d, 1.9 ^d
OH-8	11.99, s	-	12.16, s
1'	-	4.75, d, 6.6	4.74, d, 6.6
2'	-	5.50, td, 6.6, 1.4	5.49, td, 6.6, 1.3
4'	-	2.13, m	2.15, m
5'	-	2.13, m	2.15, m
6'	-	5.12, t, 6.9	5.13, t, 7.0
8'	-	1.64, br s ^c	2.05, m
9'	-	1.80, br s	2.05, m
10'	-	1.60, br s ^c	5.06, t, 7.1
12'	-	-	1.61, br s ^f
13'	-	-	1.80, br s
14'	-	-	1.63, br s
15	-	-	1.56, br s ^f

J values are given in Hz.^{a-f}: Signals with same letters are interchangeable.

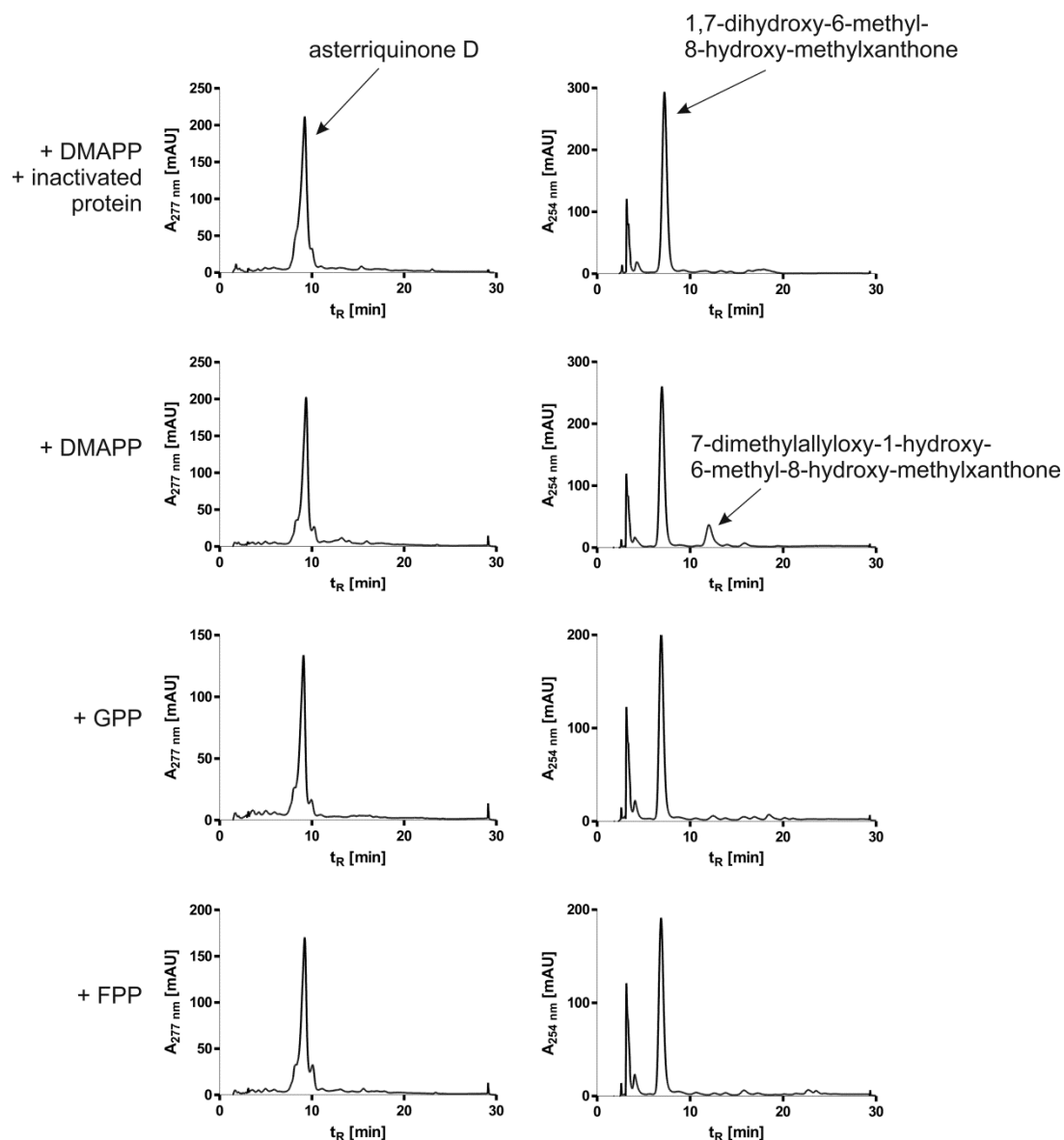


Figure S1. HPLC chromatograms of incubation mixtures of XptB with AQ D and 1,7-dihydroxy-6-methyl-8-hydroxymethylxanthone.

Assays with XptB were incubated for 16 h at 37 °C and subsequently extracted three times with ethyl acetate. Chromatograms of incubations with inactivated protein, with DMAPP, GPP and FPP as prenyl donors are shown. Incubation of XptB with 1,7-dihydroxy-6-methyl-8-hydroxymethylxanthone in the presence of DMAPP serves as positive control.

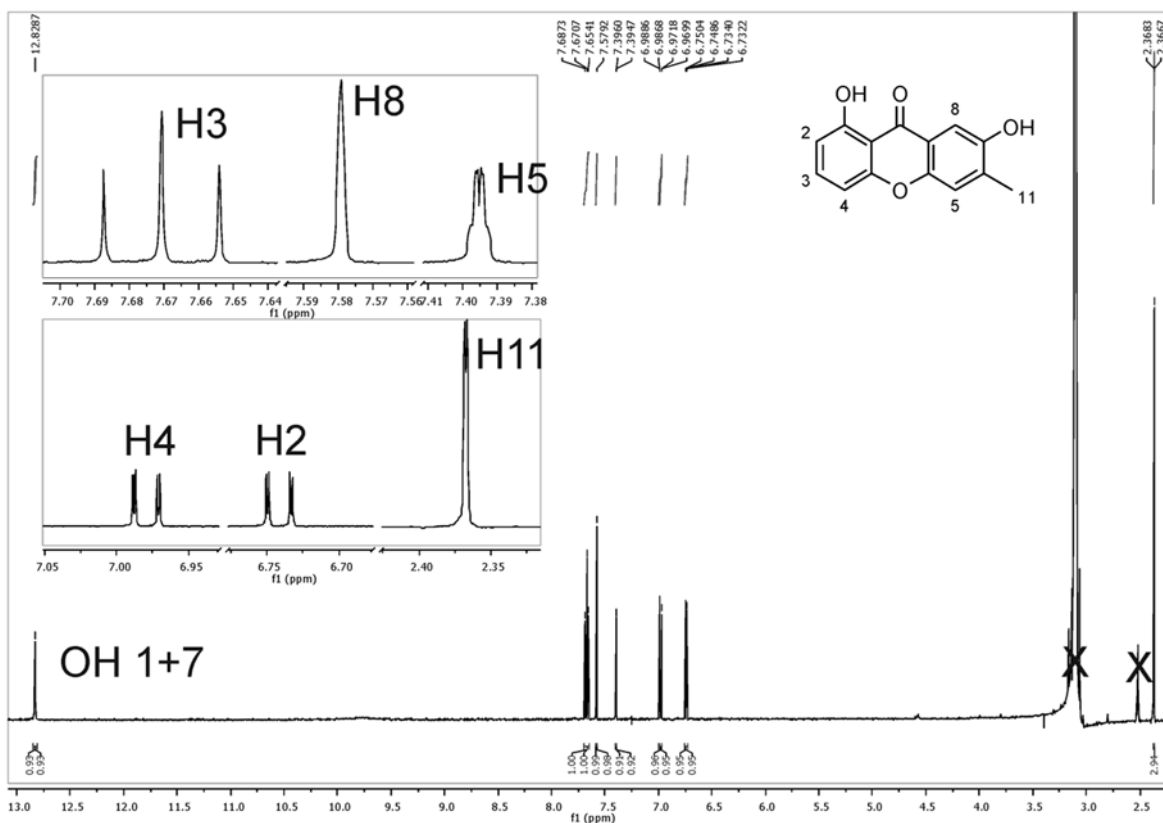


Figure S2. ¹H NMR spectrum of **1a** in (CD₃)₂CO (500 MHz).

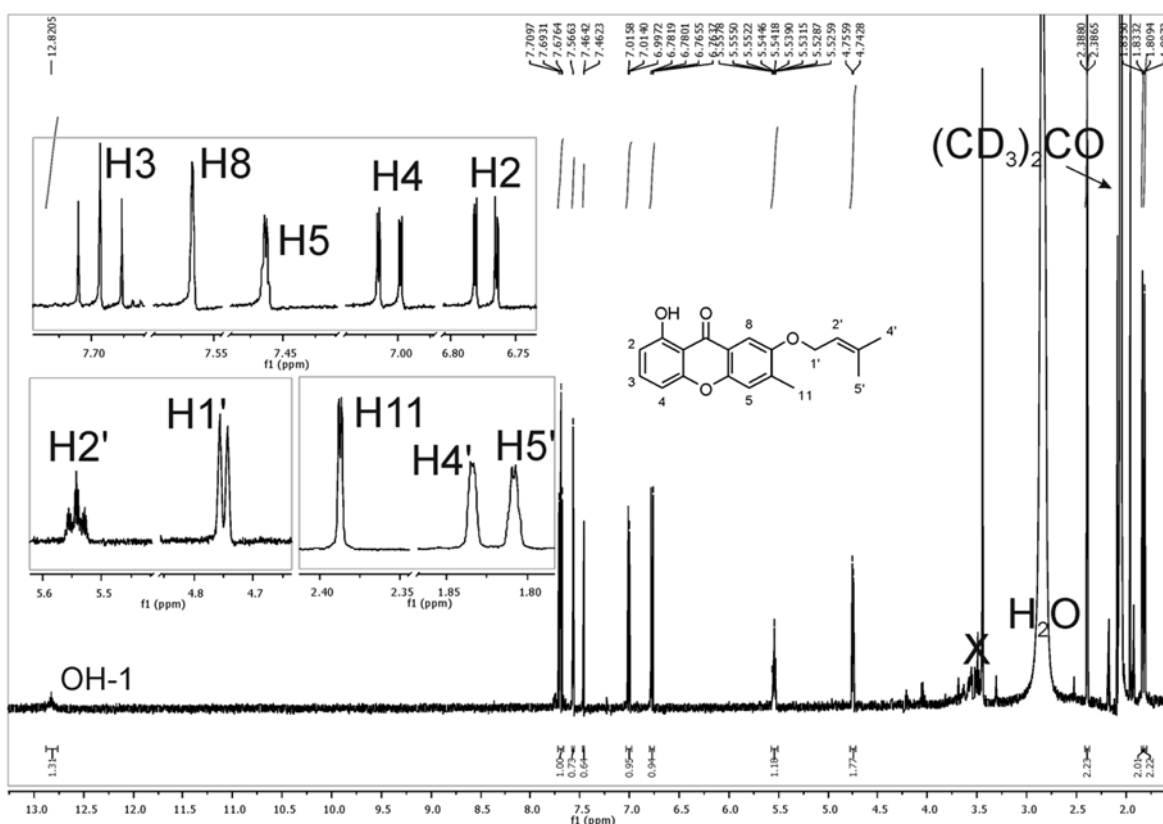


Figure S3. ¹H NMR spectrum of **1b** in (CD₃)₂CO (500 MHz).

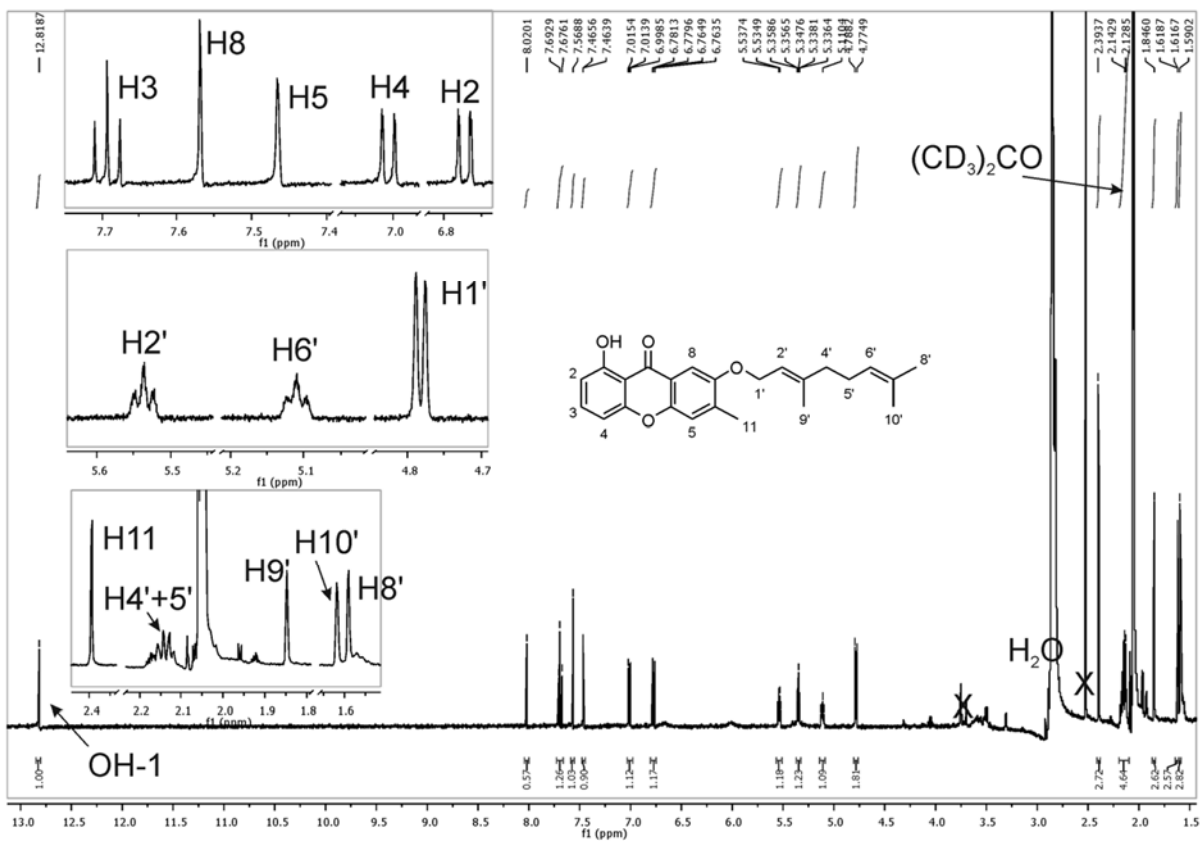


Figure S4. ¹H NMR spectrum of **1c** in (CD₃)₂CO (500 MHz).

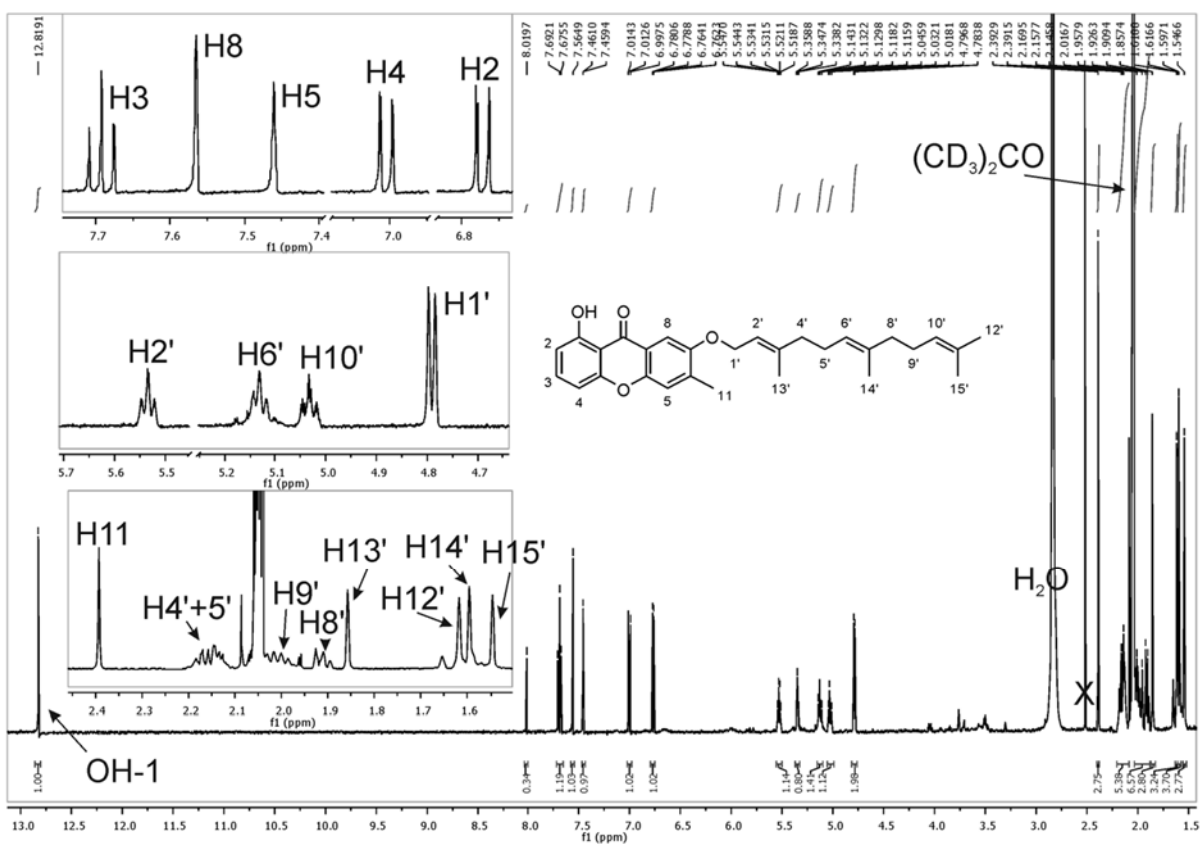
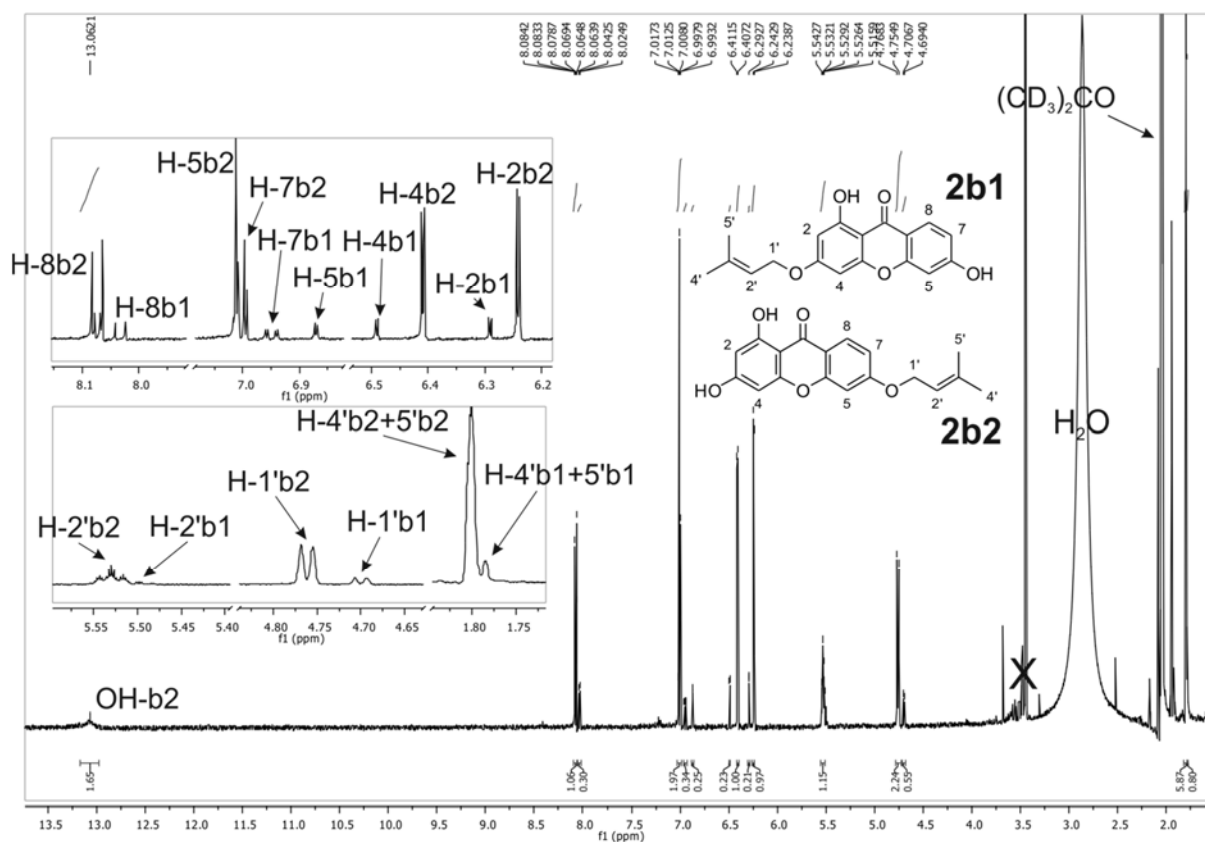
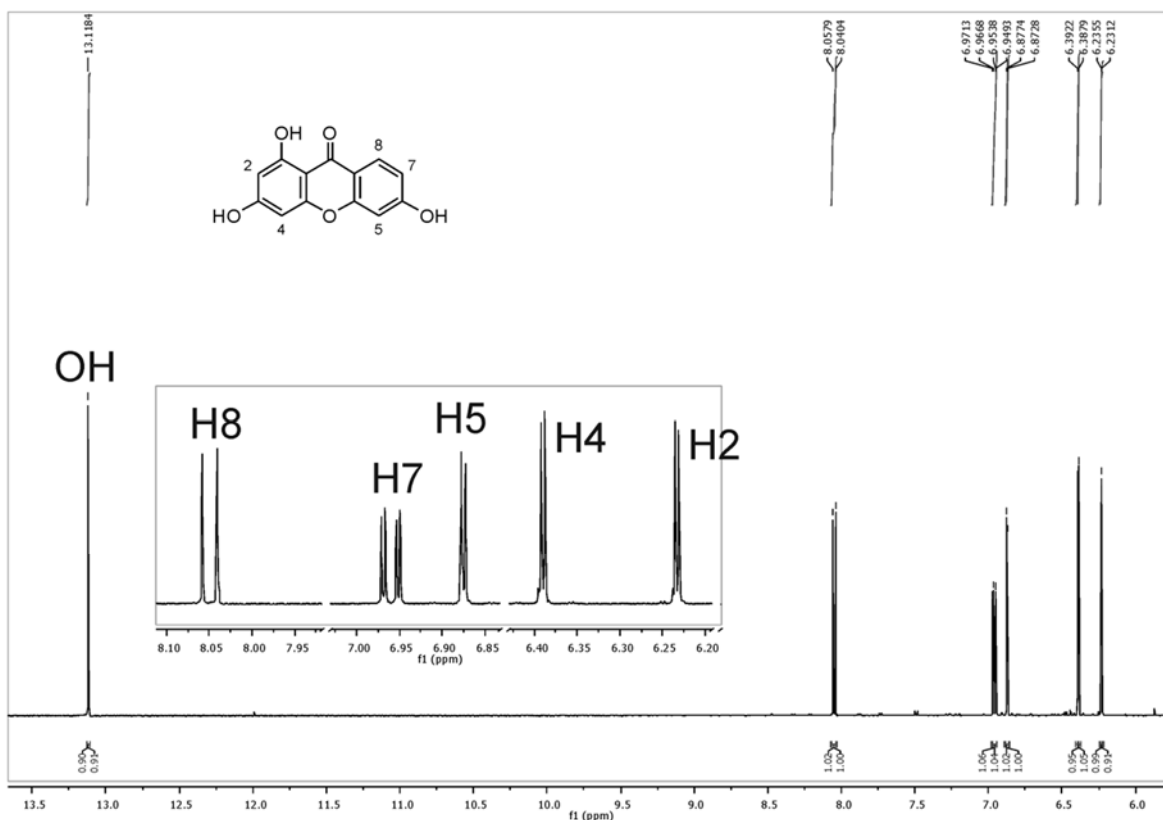


Figure S5. ¹H NMR spectrum of **1d** in (CD₃)₂CO (500 MHz).



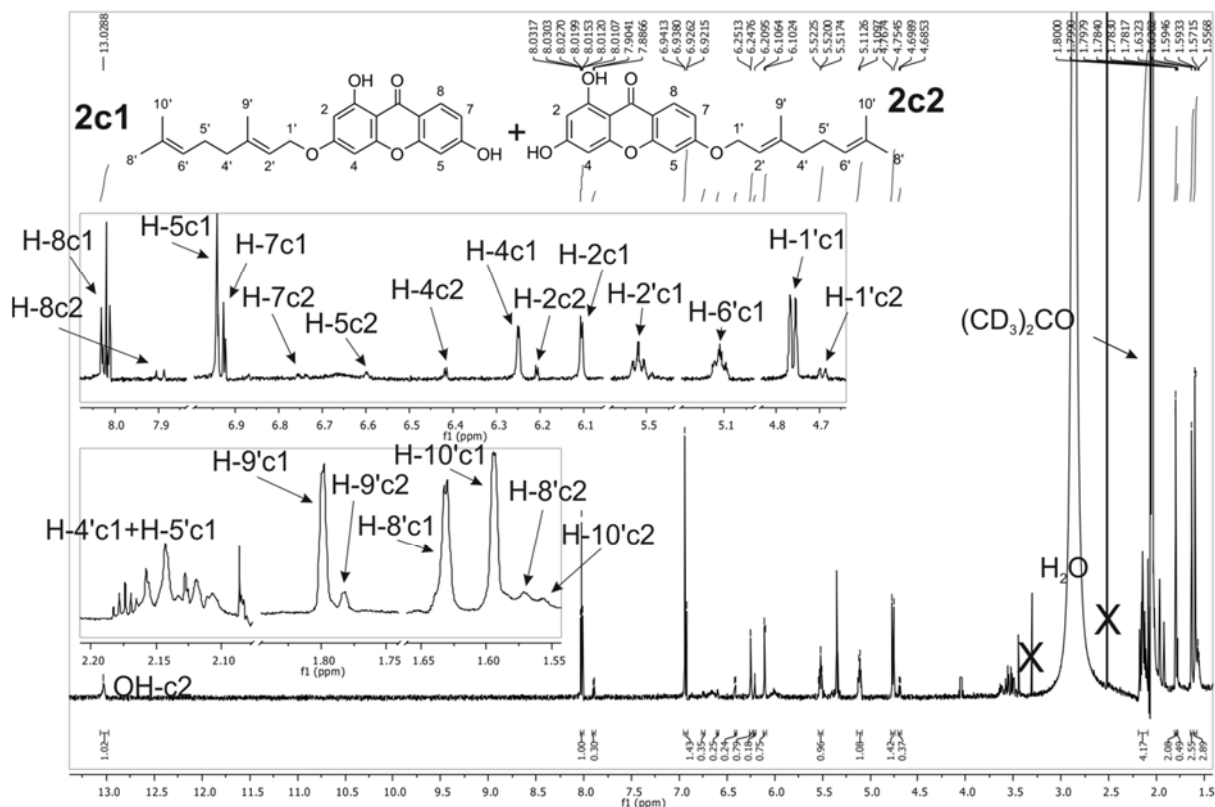


Figure S8. ¹H NMR spectrum of **2c1** and **2c2** in (CD₃)₂CO (500 MHz).

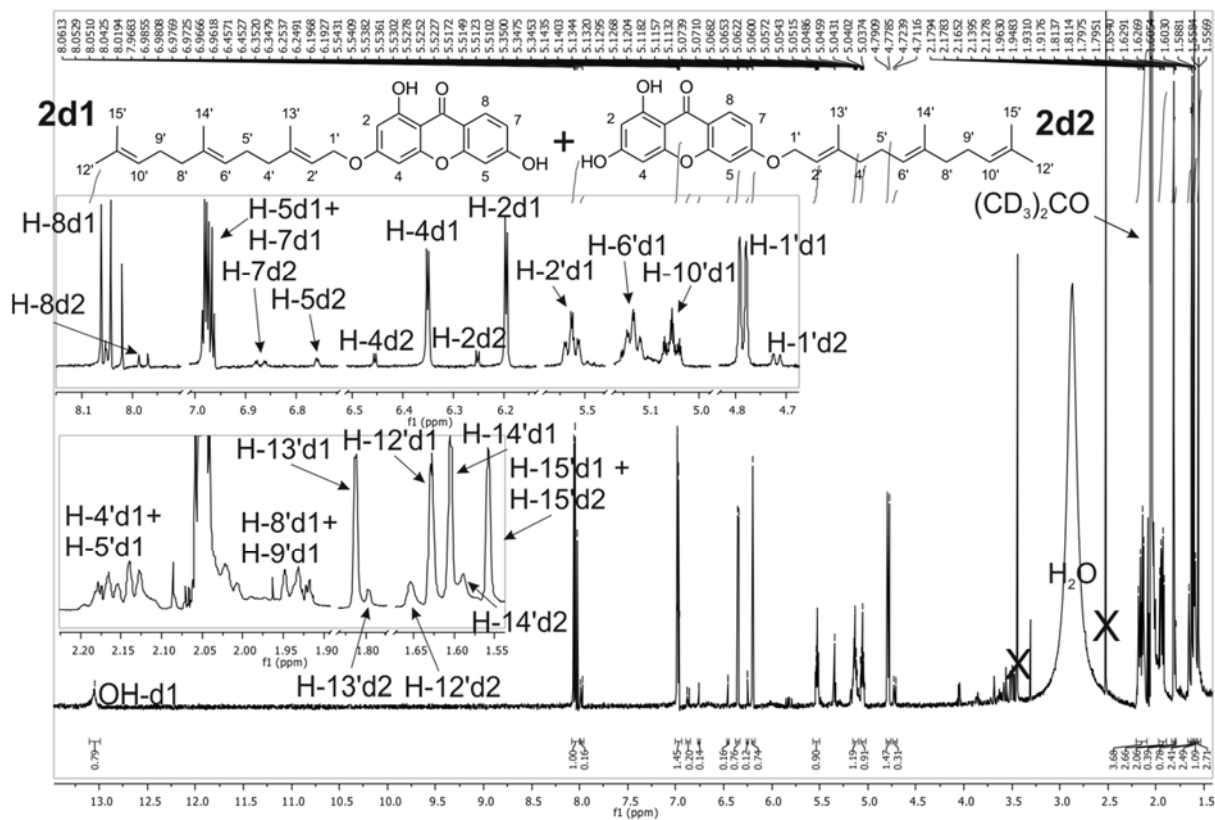


Figure S9. ¹H NMR spectrum of **2d1** and **2d2** in (CD₃)₂CO (500 MHz).

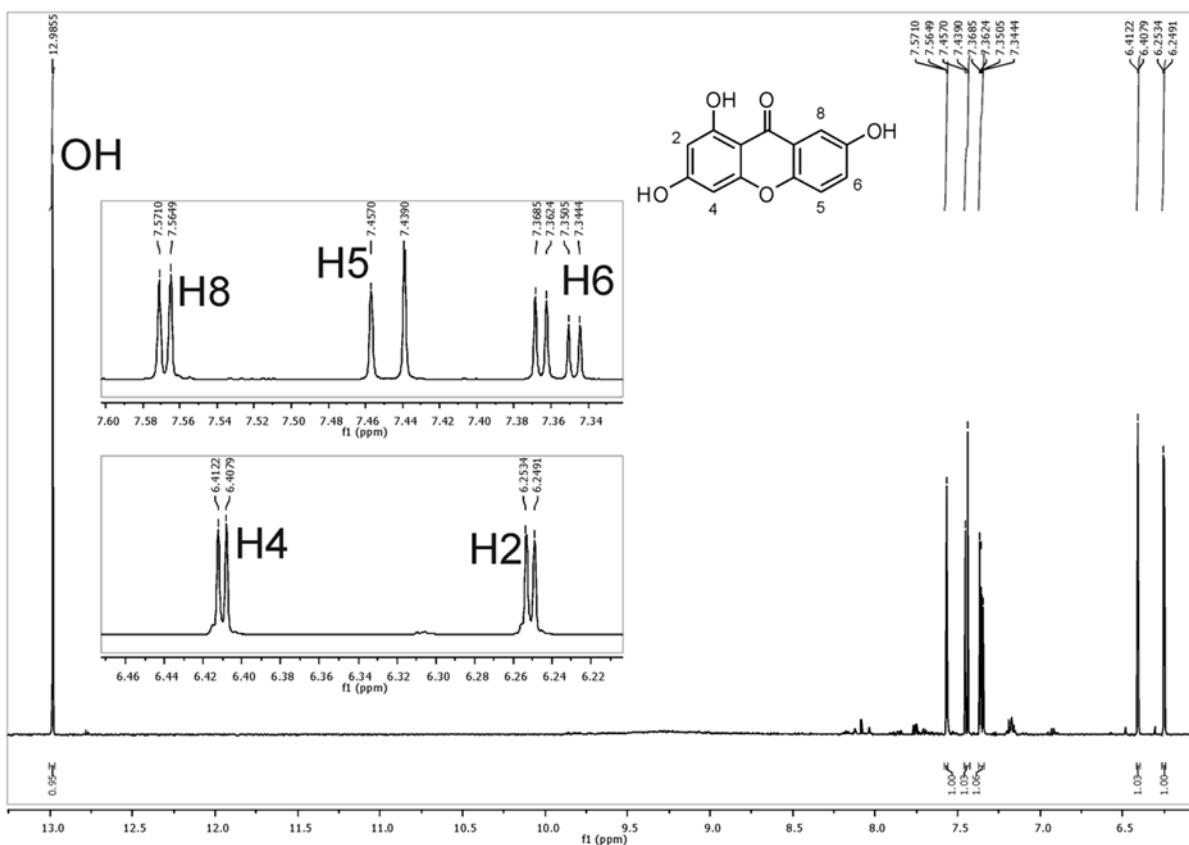


Figure S10. ^1H NMR spectrum of **3a** in $(\text{CD}_3)_2\text{CO}$ (500 MHz).

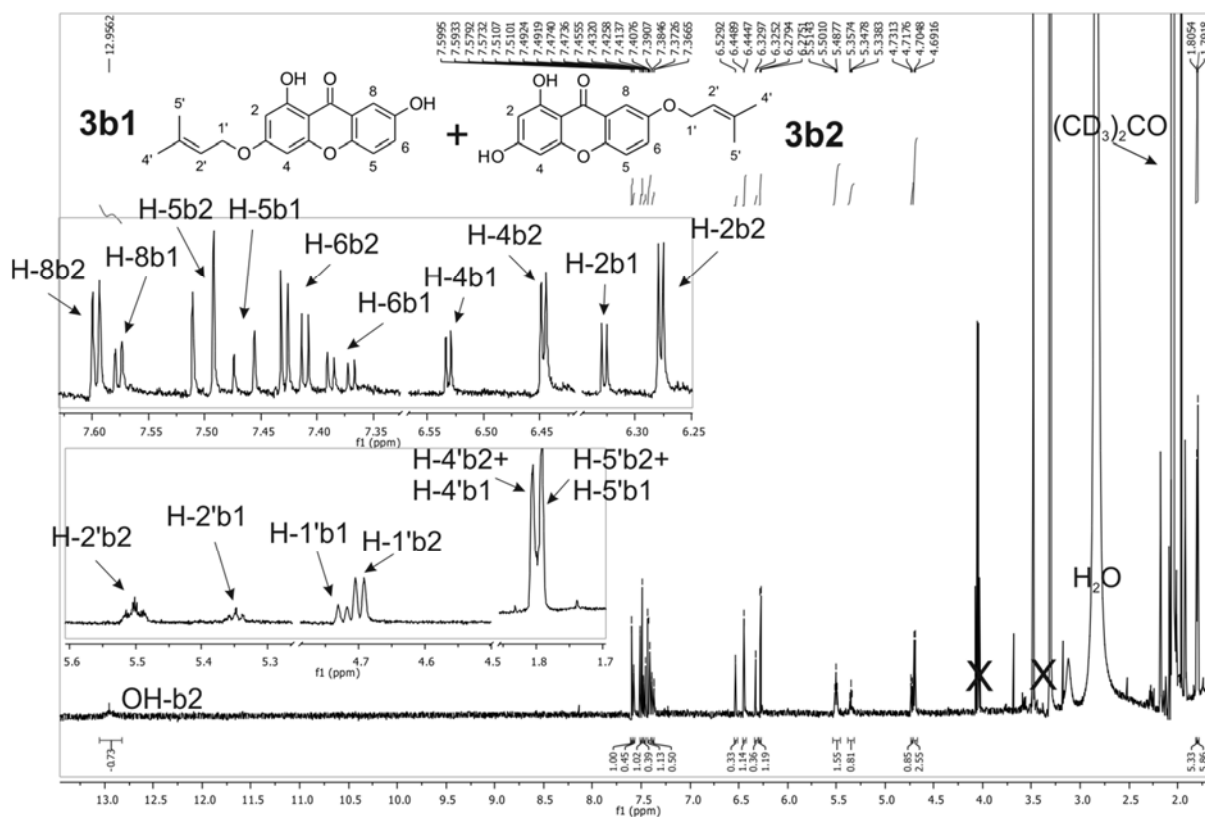


Figure S11. ^1H NMR spectrum of **3b1** and **3b2** in $(\text{CD}_3)_2\text{CO}$ (500 MHz).

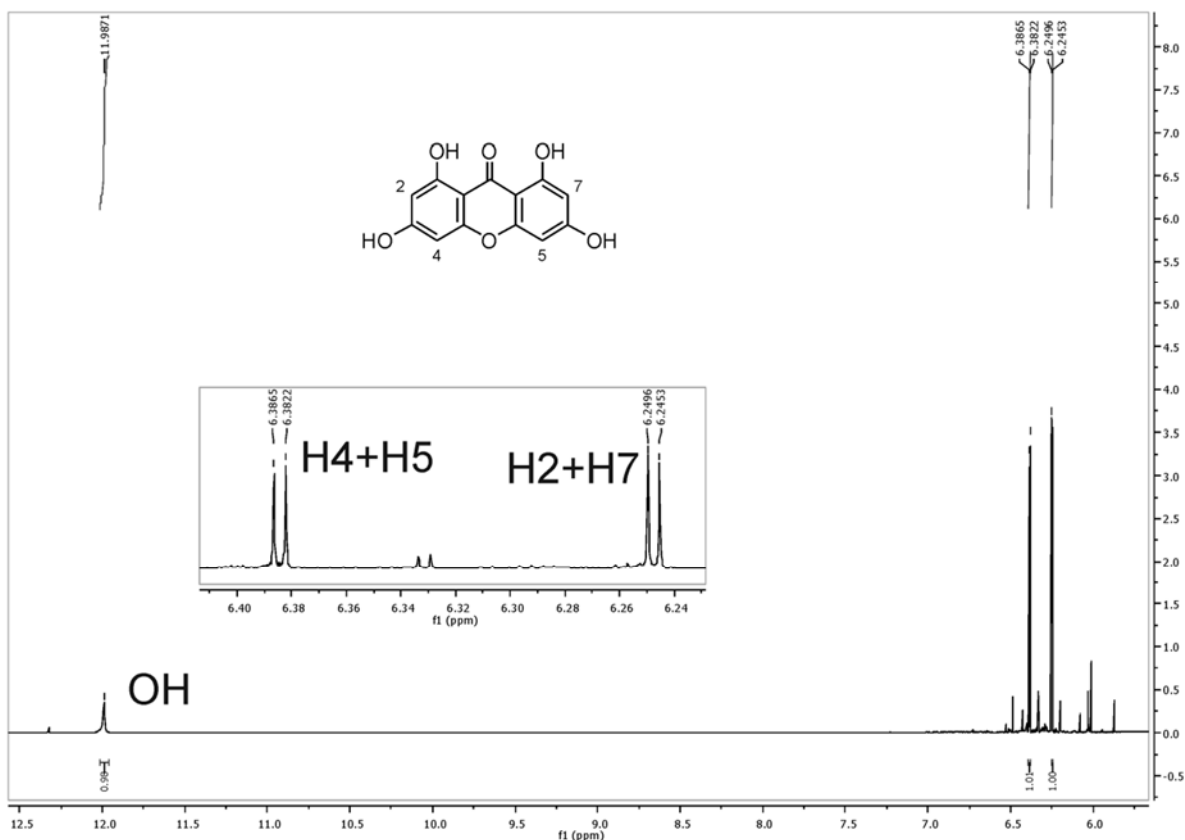


Figure S12. ^1H NMR spectrum of **4a** in $(\text{CD}_3)_2\text{CO}$ (500 MHz).

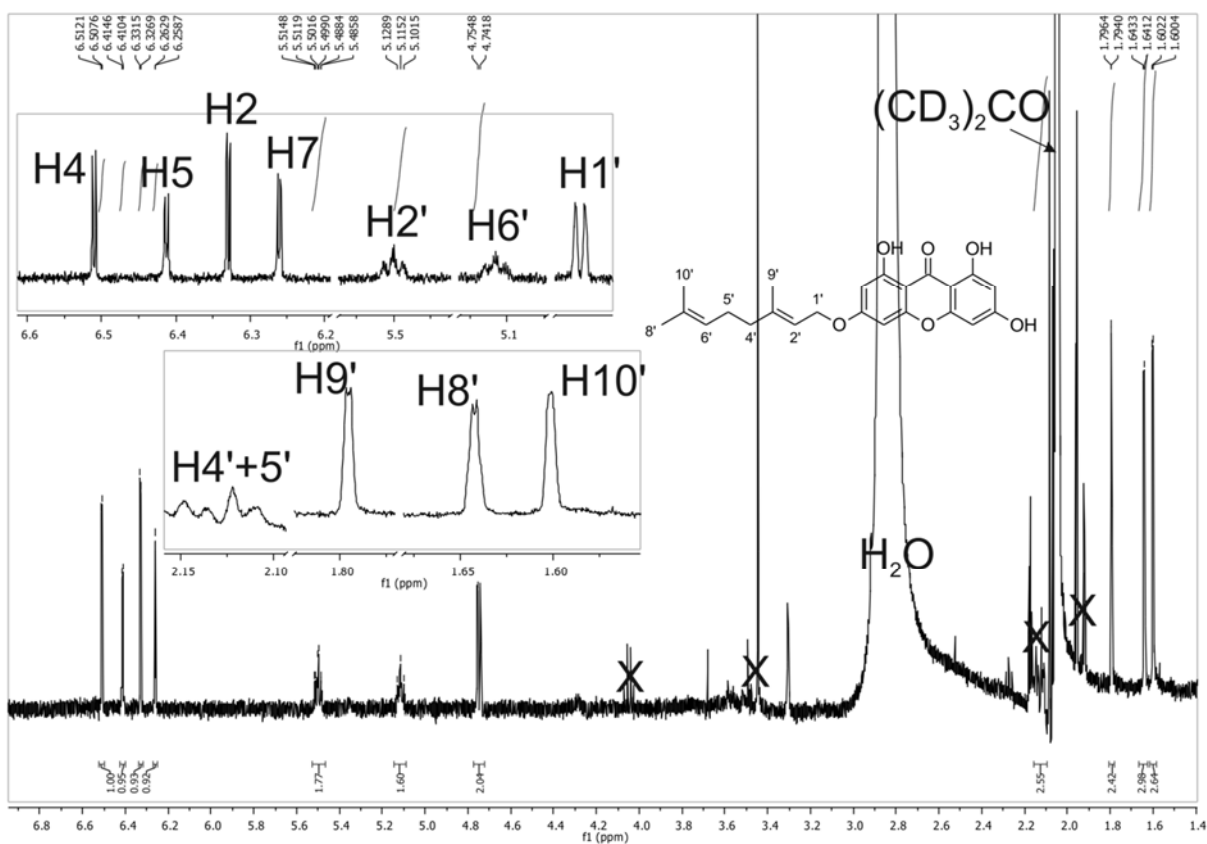


Figure S13. ^1H NMR spectrum of **4c** in $(\text{CD}_3)_2\text{CO}$ (500 MHz).

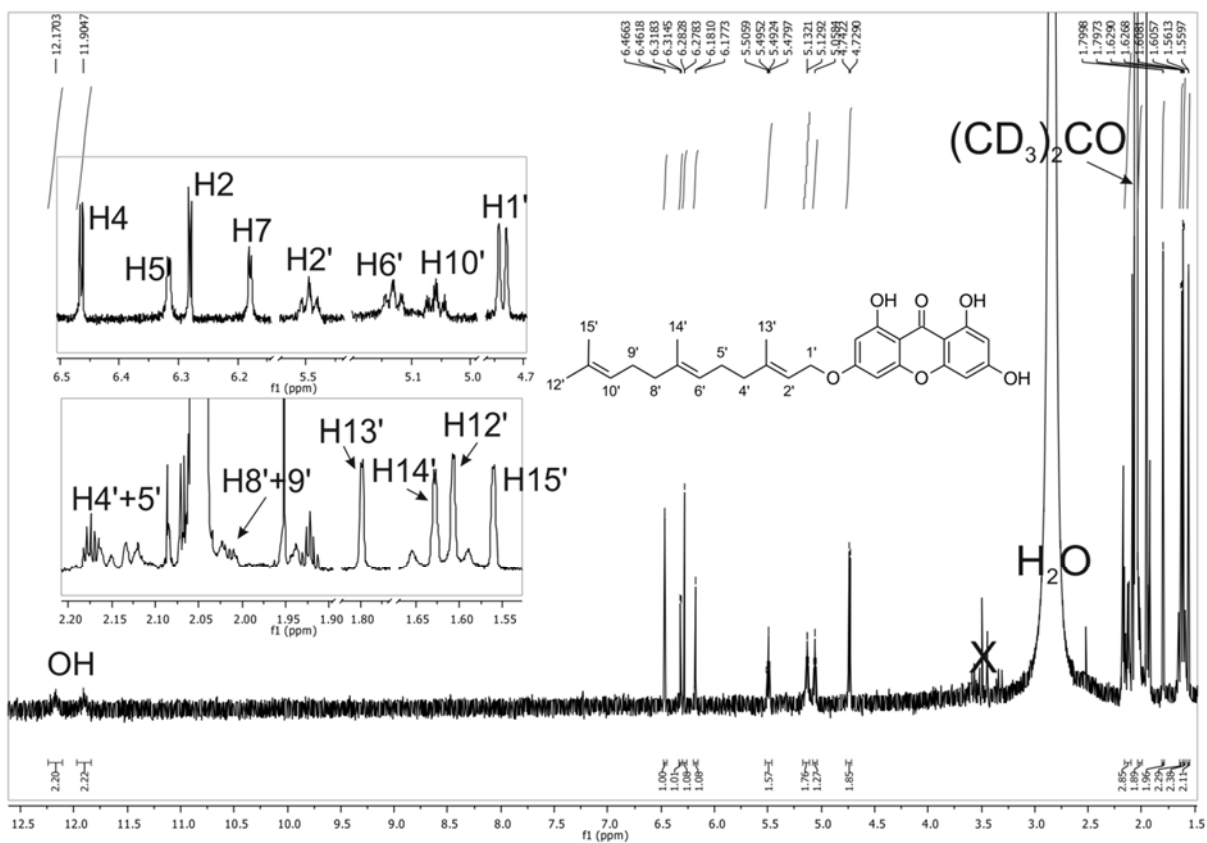


Figure S14. ^1H NMR spectrum of **4d** in $(\text{CD}_3)_2\text{CO}$ (500 MHz).

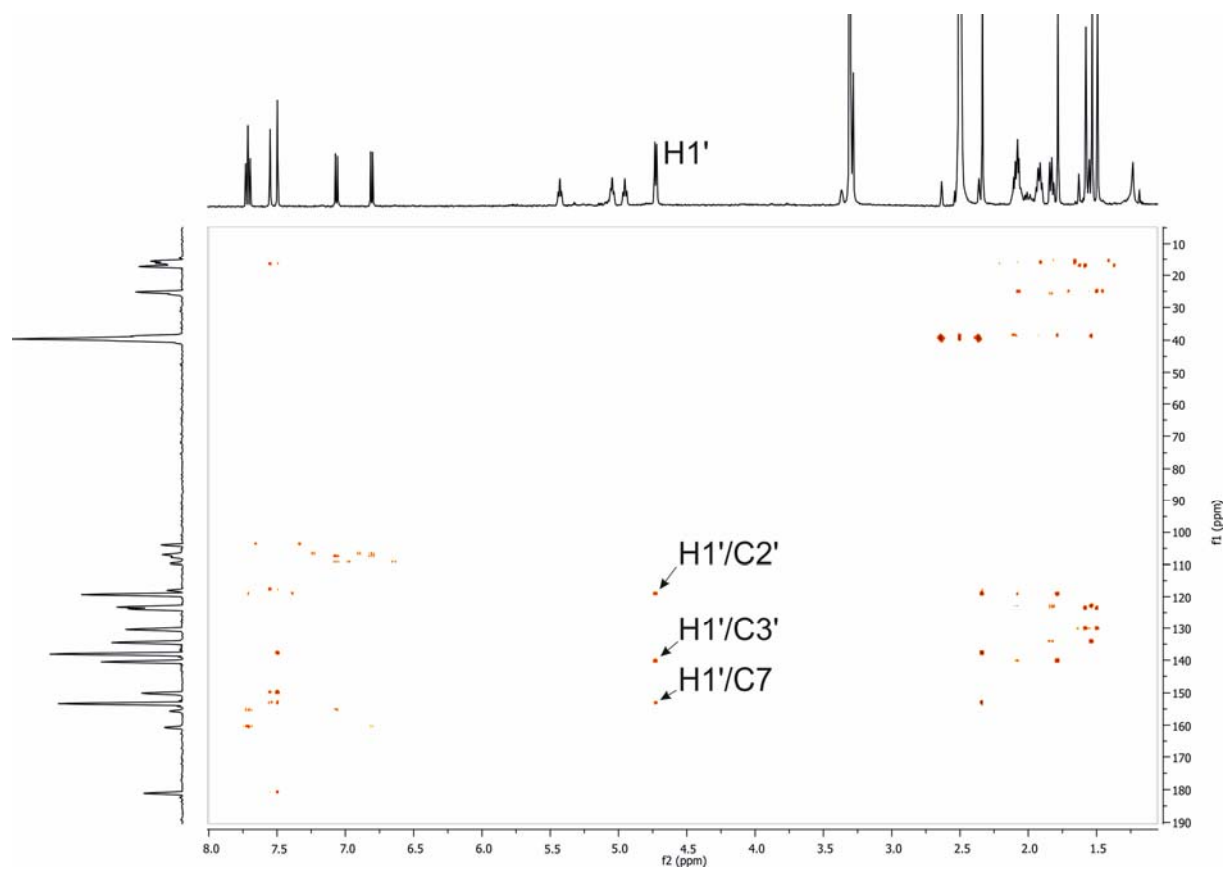


Figure S15. HMBC spectrum of **1d** in $(\text{CD}_3)_2\text{SO}$.

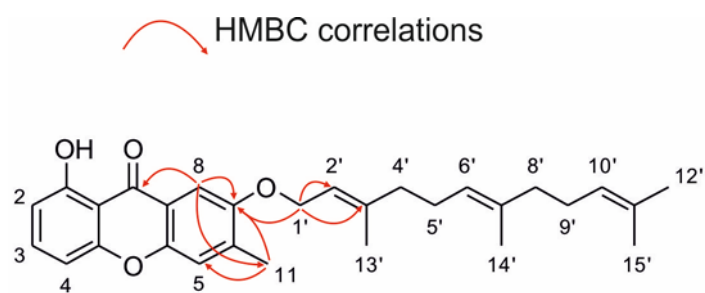


Figure S16. HMBC correlations of **1d**.

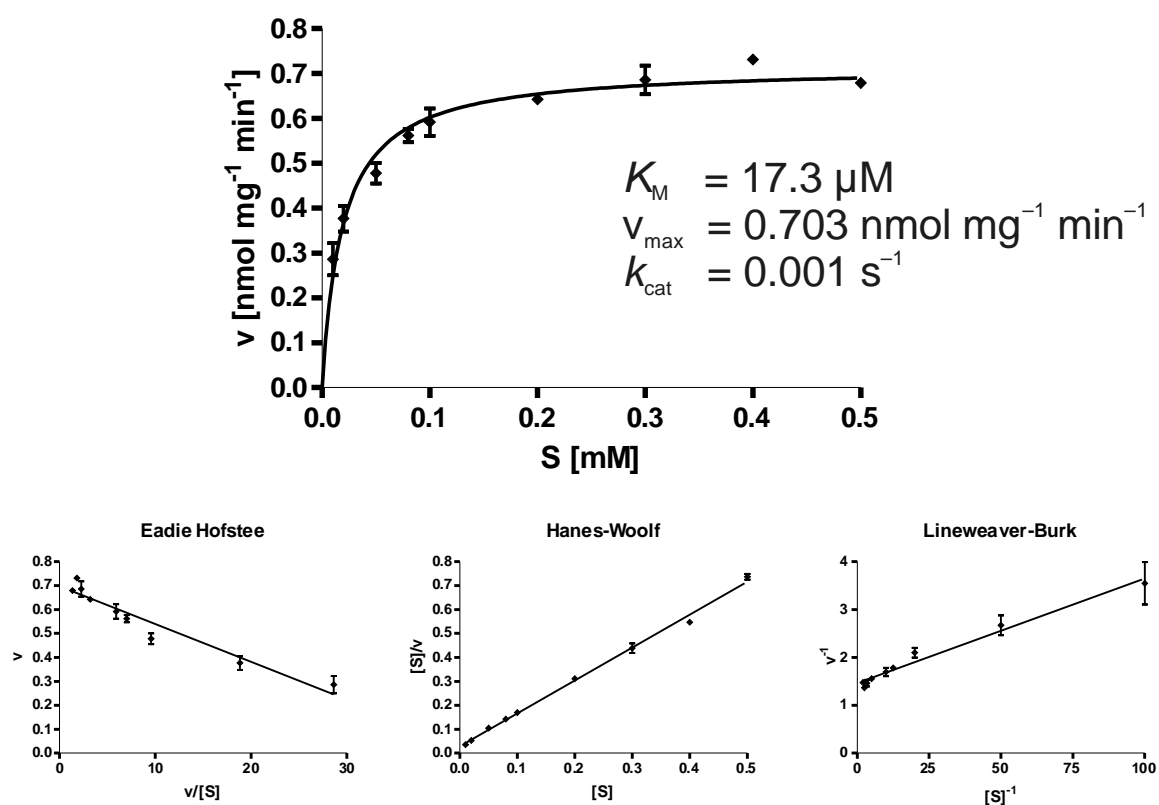


Figure S17. Determination of kinetic parameters for **1a**.

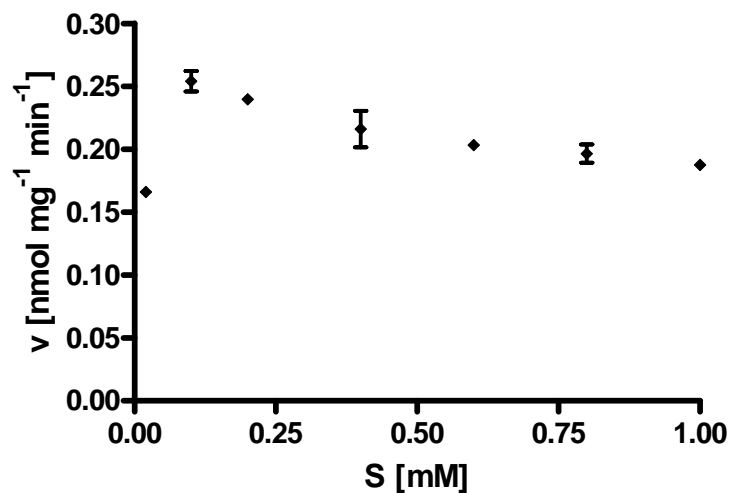


Figure S18. Determination of kinetic parameters for **2a**.

Inhibition occurred and maximal velocity was not reached under our assay conditions.

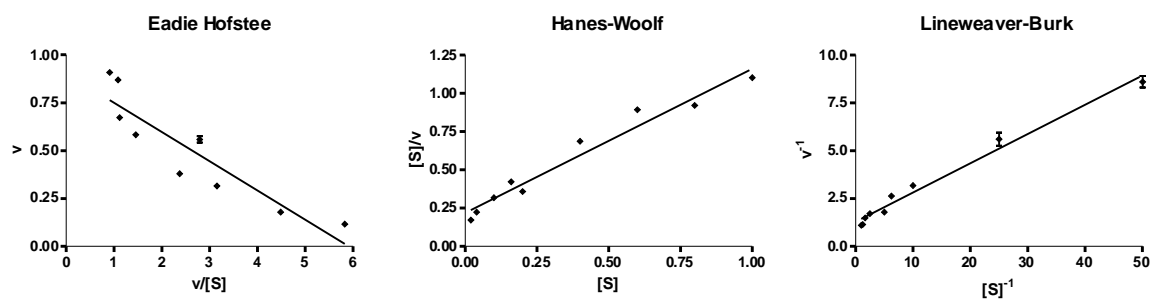
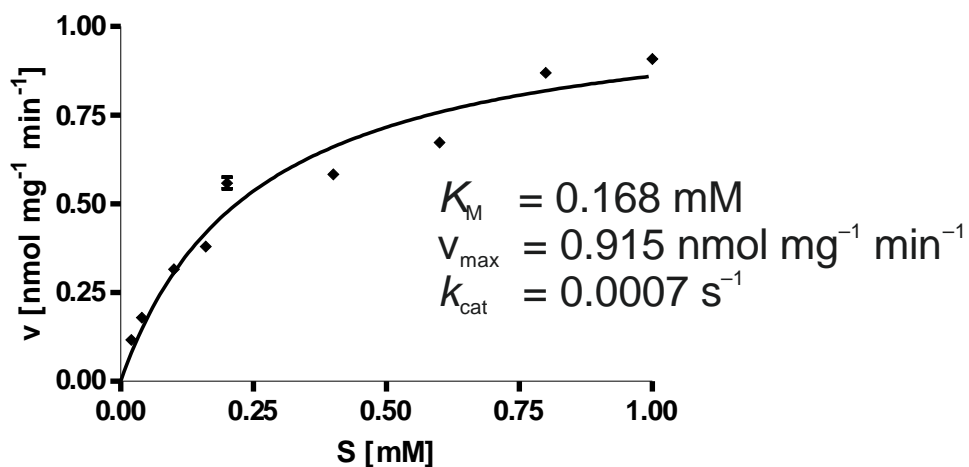


Figure S19. Determination of kinetic parameters for **3a**.

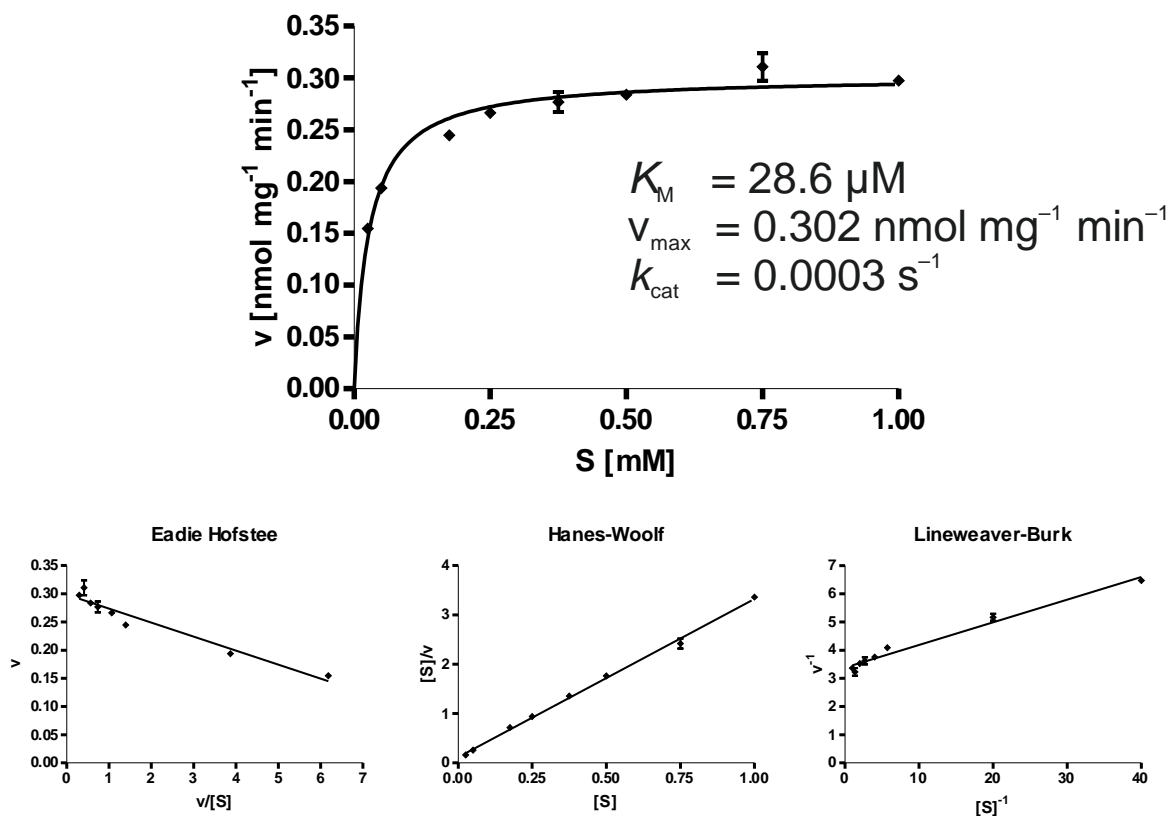


Figure S20. Determination of kinetic parameters for **4a**.

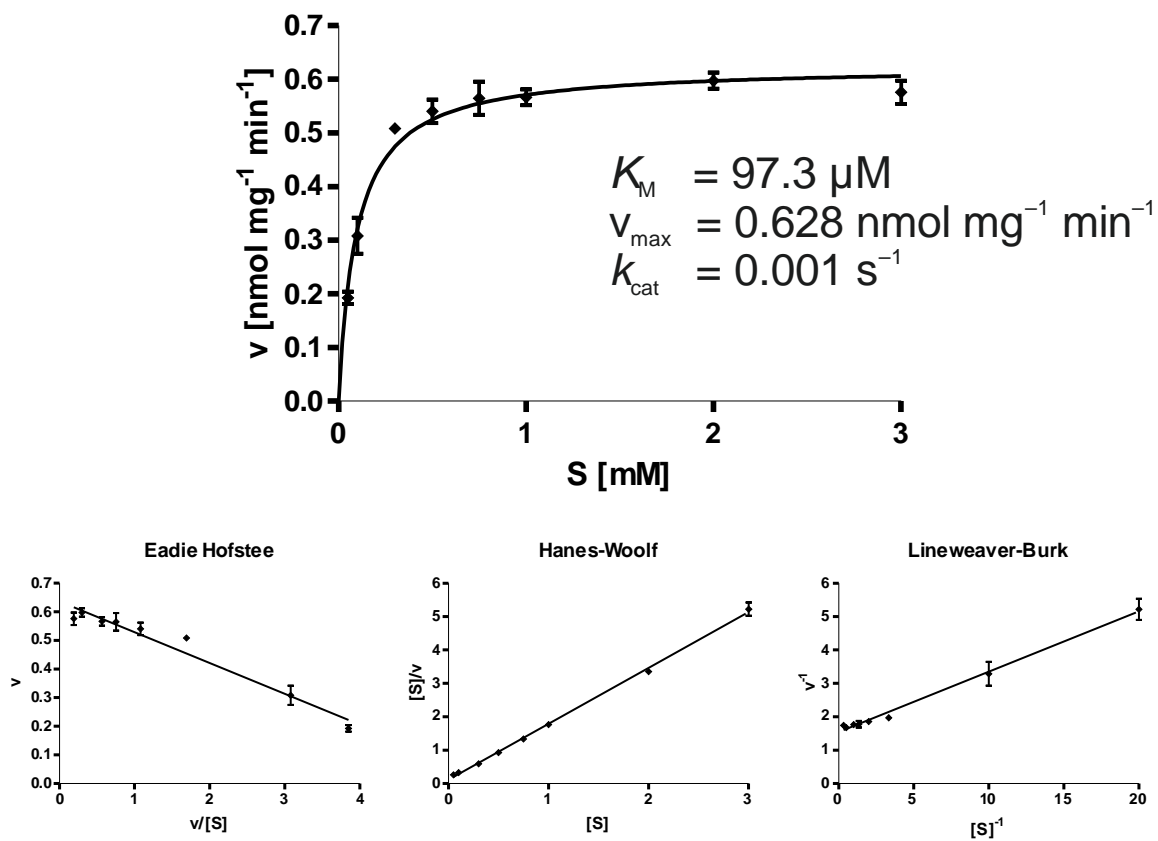


Figure S21. Determination of kinetic parameters for **DMAPP**.

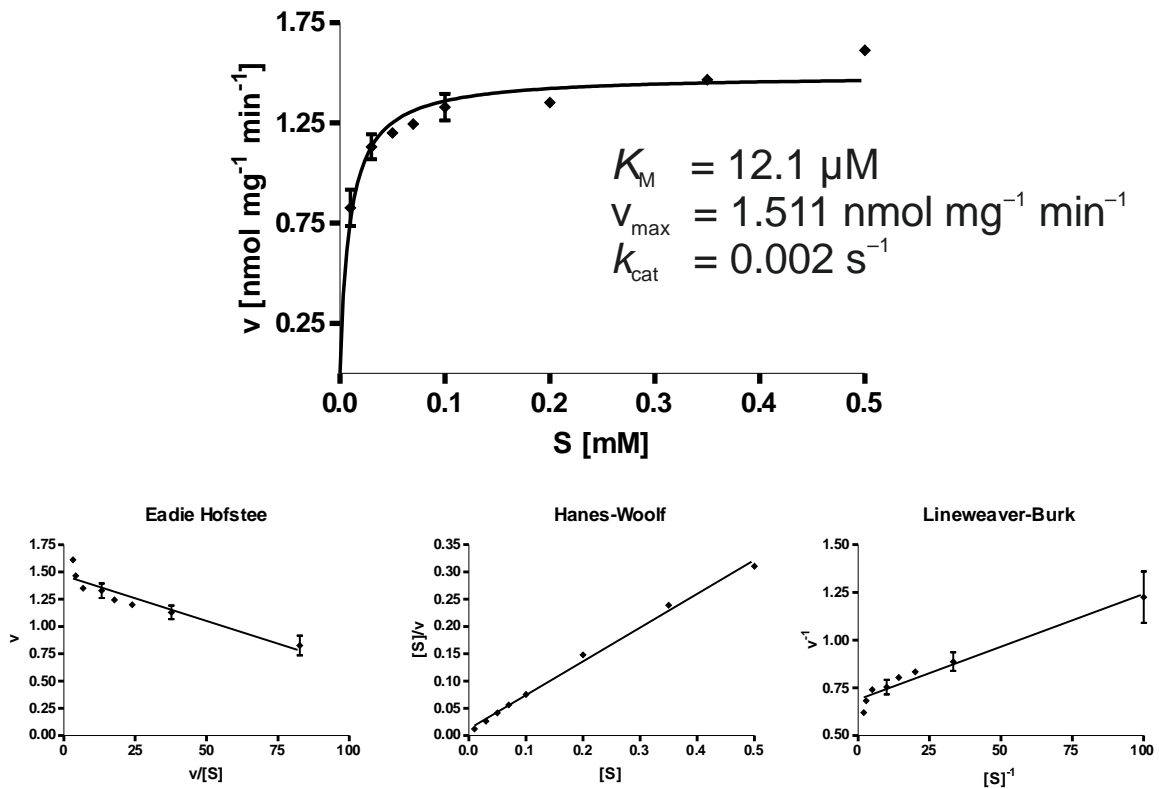


Figure S22. Determination of kinetic parameters for GPP.

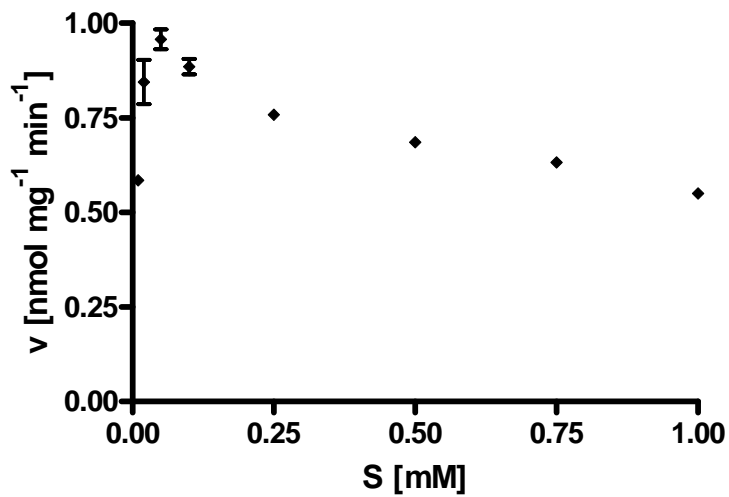


Figure S23. Determination of kinetic parameters for FPP.

Inhibition occurred and maximal velocity was not reached under our assay conditions.

FgaPT2	-----MKA-----NASSAEAYRVLS	16
7-DMATS	MSIGAEIDSLVPAPQGLNGTAAGYPAKTQKE-----LSNGDFDAHDGLSLAQLTPYDVLVLT	55
AnaPT	-----MSPLSMQTTDSVQGTAEEN-KSLETNGTSDNQQLPWKVLG	37
FtmPT1	-----MPPAPPDQKPC-----HQLQPAPYRALS	23
CdpNPT	-----MDGEMTASPPDISACDTSVAVDEQTTQSGSQQAPIPKDIAIYHTLTL	44
XptB	-----MVL-HKRSLSE---GGTSTQAWKVLVS	22
TdiB	-----	0
AstPT	-----MTIH	4
FgaPT2	RAFR-FDNE--DQKLWWHSTAPMFAKML-ETANYTTPCQYQYLITYKECVIPSLGCYPTN	72
7-DMATS	AALP-LPAPASSTGFWWRETGPVMSKLL-AKANYPLYTHYKYLMLYHTHILPLLGP RPPL	113
AnaPT	KSLG-LPTI--EQEQYWLNTAPYFNNLL-IQCGYDVHQYQYLAFAFYHRHVLVPLGPFIRS	93
FtmPT1	ESIL-FGSV--DEERWWHSTAPILSRLL-ISSNYDVVDVQYKYLSLYRHLVLPALGPYPQR	79
CdpNPT	KALL-FPDI--DQYQHWHHVAPMLAKML-VDGKYSIHQQYELCLFAQLVAVPLGYPYSP	100
XptB	QTLF-SRGP--DVDAWQLTGRHLAVLL-DAAAYPIEKQYECLELHYHYAAPYLGPAPE	78
TdiB	-----MATEYWSRHLRSVLAPLFAAAGTYSPEQESHAFIDEHIAPNLGLPLWE	50
AstPT	TDLPPPEAKASDTEFWSEHIRSVIGPLMKATGYSYSGTAQEANRFLDNYIAPALGPHPTV	64
	: : : * : *	
FgaPT2	S-----APRWLSILTRYGTPFELSLNC-----SNSIVRYTFEPINQHTGT-DKDPFN	118
7-DMATS	ENSTHPSPSNAPWRSFLTDDFTPLPESWNVNGNSEAQSTIRLGIPIGFEAGA-AADPFN	172
AnaPT	SAEAN-----YISGSAEG-YPMELSVNY---QASKATVRLGCEPVGEFAGT-SQDPMN	142
FtmPT1	DPETG--IIATQWRSGMVLTLGLPIEFSSNNV-----ARALIRIGVDPVTADSGT-AQDPFN	131
CdpNPT	GRDVI-----RCTLGGN-MTVELSQNF---QRSGSTTRIAFEPVRYQASV-GHDRFN	147
XptB	GAS-----PPTWKSMLQLDGTPEFESWKWNW-PGGEPEVRFGLPEIGPMAGT-SLDPLN	130
TdiB	PHGPY-----STPSSLVGSPPDPSINIV--SSGKAKVRFDFDVLSPDRT-GDDPFA	99
AstPT	AHPTY-----VAPCTIVGTLFNPSISLS--AKGKPTVRFDYDLPLPLDRASSDDPWG	114
	: * . . * :	
FgaPT2	THAIWESLQHLPLEKSIDLEWFRHFHKLTLNSEEASAFLAHN---DRLVGGTIRTQNKL	175
7-DMATS	QAAVTQFMHSYEATEVGTATLTLFEHFRNDMFVGPETYAALR-----AKIPEGEHTTQSFL	227
AnaPT	QFMTREVLGRLSRLDPTFDLRLFDYFDSQFSLTTSE-ANLAASKL-----IKQRRQSKVI	196
FtmPT1	TTRPKVYLETAARLLPGVDLTRFYEFETELVITKAEAEVLQAN---PDLFRSPWKSQILT	188
CdpNPT	RTSVNAFFSLOQLLVKSVNIELHLLSEHLTLTAKDERNLNEEQLTKYLTNFQVKTYVV	207
XptB	HLAMREILYKLVSSAVPGSDLTWTHHFLATLFDHDY--AKYTQK---AATMGSSIGTSLVY	185
TdiB	EGSAREILHRLADL-VGADTQWMGYLMDALYLTPAEAEVAK-----TKLPPGVAIPSSV	153
AstPT	EKGARTLFRRLAAA-LGADTQWLEYFMARFLSPAETEALR-----SKIPADLVIPSAMV	168
	: : :	
FgaPT2	ALDL-KDGRFALKTYIYPALKAVVTGKTIHELVFSGVRRLAVREPRILPPLNMLEEYIRS	234
7-DMATS	AFDL-DAGRVTTKAYFFPILMSLKTQSTTKVVSDSLHLALKSGVWGVTIAAMSVMEA	286
AnaPT	AFDL-KDGALIPKAYFFLKGKSLASGIPVQDVAFNAIESIAPKQIESPLRVL--RTFVTK	253
FtmPT1	AMDQKSGTVLVKAYFPQPKSAVTGRSTEDLLVNAIRKVD-REGRFETQLANLQRYIER	247
CdpNPT	ALDL-RKTGIVAKYFFPQKCAATGQTGSNACFGAIRAVDKDG---HLDSL--CQLIEA	261
XptB	SLEF-QRKSTGLKTYFHPKLDQQA-----FLDIP-----S-----WEA	218
TdiB	GFDF-DGPERTLKFIIPSVRKALATQDVSSELMKTLRGLQPLGSELVPA---MDLIAS	208
AstPT	GVAF-DDAQPRLLKAVPTMRRRAILEGRSSNEIAVEVLRGLSPLGSEITPA---IDMVEA	223
	.. : * : :	
FgaPT2	RGS-----KSTASPRLVSCDLTSPAK---SRIKIYVLEEQ	265
7-DMATS	WIG---SY-----GGAAKTEMISVDCVNEAD---SRIKIYVRMP	319
AnaPT	LFS---KP-----TVTSDVFLAVDCIVPEK---SRIKIYVADS	286
FtmPT1	RRRGLHVPVGTADKPPATAA---DKAFDACSFFPHFLSTDLVPEPK---SRVVFYASER	300
CdpNPT	HFQ---QS-----KI--DDAFLCCDLVDPAH---TRFKVIADP	292
XptB	SFRGLHPN-----SPSRTAVHEFLSTNPEGKLLKPFCLSVNDCSPAK---ARIKWFNSP	270
TdiB	YLST---R-----TNDAMLPLVGDIDCLDPRTHKNARVKCYLHTS	244
AstPT	WIAA---S-----PHDPRMLVGMDCGEPD---SRIKIYLVTT	256
	: * : * * *	

Figure S24. Multiple sequence alignment of AstPT and homologues by Clustal Omega. Identical amino acids in all proteins are marked with an asteriks. Identical amino acids which stabilise the substrates in the active site are highlighted by black boxes. TdiB (ABU51603.1) and XptB (BN001302.1) are from *Aspergillus nidulans*, AstPT (EAU29429.1) from *Aspergillus terreus*, FgaPT2 (AAX08549.1), FtmPT1 (AAX56314.1), 7-DMATS (ABS89001.1) and CdpNPT (ABR14712.1) are from *Aspergillus fumigatus*, AnaPT (EAW16181.1) is from *Neosartorya fischeri*.

FgaPT2	MVSLEAMEDLWTLGGRRRDASTLEGLSLVRELWDLIQLSPGLKS-YP----APYLP LG--	318
7-DMATS	HTSLRKVKEAYCLGGRLTDENTKEGLKLLDELWRTVFGIDEDA-----ELP--	366
AnaPT	QLSLATLRFWTLGGSVTDSATMKGLEIAEELWRILQYDDAVCS-----H----	331
FtmPT1	HVNLQMVEDIWTFGGRLRRDPDALRGLLELLRHFWADIQMGEGYIT-MP----RGFCELG--	353
CdpNPT	LVTTLARAEHHTLGGRLTDEDAAVGLEIIRGLWSELGI IQGPLE-----PSA--	339
XptB	HTNFRAIREIMTLGGRIADTETRTRK--QFSELFNLLKTVTEEHADFPETSEFPYVPNMGD	328
TdiB	SNSFAVVRDVLTLGGRLSDDTSLKRVETLKSVMPLLINELEGPQSDAATMDESWSKPE--	302
AstPT	KNSWATVQDVMTLGGRLDDEPTRKQLALLRK IWPYLINEPDN---TRIADENWCKPE--	310
	. : : ** * :	
FgaPT2	-VIPD-----E-RLPLMANFTLH-QNDPVPEPQVYFTTFGMND--MAVADALTTFFERRG	368
7-DMATS	-Q-NS-----HRTAGTIFNFE LR-PGKWFPEPKVYLPVRHYCESDMQIASRLQTFFGRLG	418
AnaPT	---SN-----MDQLPLVVNYELS-SGSATPKPQLYMLPLHGRND--EAMANALTKFWDYLG	380
FtmPT1	-KSSA-----GFEAPMMFHFHLDGSSQSPFPDPQMYVCVFGMNS--RKLVEGLTTFYRRVG	405
CdpNPT	-M-ME-----KGLLPIMLNEMK-AGQRLPKPKLYMPLTGIPE--TKIARIMTAFQQRHD	389
XptB	SIIPNFADAPMLKGCVYFFDIA-PGRNLPKIKVYFPVRNHCNRNDLAVTQNLNRWLESRG	387
TdiB	-RLNR-----TGYSGIQYTIETIT-PGQAIPTKIYVPLFQYTDSSVAERNFESALKKLG	355
AstPT	-RMPR-----VGFFGLMYSLEIK-PGRPTPEVKLYVPLFQYAESWAI AENNMETVLKLLD	363
	: * : : *	
FgaPT2	--WSEMARTYETTLKSYYPHADHDKLNYLHAMISFSYR-DRTPYLSVYLQSFETG-----	420
7-DMATS	--WHNMEKDYCKHLEDLFPHHPLSSSTGHTHTFLSFSYKKQKGVYMTMYNLRVYS-----	471
AnaPT	--WKGAAQYKLDLYANNPCRNLAETTTVQRWVAFSYTESGGAYLTVYFHAVGGM-----	433
FtmPT1	--WEEMASHYQANFLANYPDEDFEKA AHLCAVVSFAYK-NGGAYVTLNHSFNPNV-----	457
CdpNPT	--MPEQAEVFMENLQAYYEGKNLEEA TRYQAWLSFAYTKEKGPYLSIYYFWPE-----	440
XptB	--RGQYGAAFGRALETIADYRRLLEDSSGLLSFLSCQFMEDGELDLTSYFNPQAFH-----	440
TdiB	NEWGLSG-KYRSVMQEIFKD---VENYQTYASFYSYTEGKGVYTTSYVAMP IKDEGGGS	410
AstPT	IDWGHSG-KYRQAMEMTFGK---GNSYQGIFVAYSYSERTGGYINSYVSMVPKDVPAHA	418
	: : : : *	
FgaPT2	---DWAVANLSESKVKCQDAACQPTALPPDL SKTGVYYSGLH	459
7-DMATS	---T-----	472
AnaPT	---KG---NL-----	437
FtmPT1	---GDVSFPN-----	464
CdpNPT	-----	440
XptB	---SGRLTHRRA-TRRRGDDRW-----	458
TdiB	LAGDFGFRN-----	419
AstPT	FTGDYV-----	424

Figure S24. Continued.