ESI for

Bis-triazolium containing macrocycles, pseudorotaxanes and interlocked structures for anion recognition

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Details of instrumentation

Routine NMR spectra were recorded on a Varian Mercury 300 spectrometer with ¹H NMR operating at 300 MHz, ¹³C at 75.5 MHz, ¹⁹F at 283 MHz, and ³¹P at 122 MHz. Some compounds were too poorly-soluble, or not enough compound was synthesized to allow ¹³C NMR spectra to be recorded on the 300 MHz spectrometer. In these cases, the spectra were collected on a Bruker AVII 500 spectrometer with a 5 mm ¹³C(¹H) dual cryoprobe with ¹³C operating at 126 MHz and ¹H operating at 500 MHz.

High resolution ESI mass spectra were recorded on a Bruker μ TOF spectrometer. High resolution EI mass spectra were recorded on a Waters GCT Classic spectrometer. Low resolution ESI mass spectra were recorded on a Walters LCT premier spectrometer.

NMR Spectra of new compounds

NMR Spectra of polyether bis-azide 1





Figure S2. ¹³C NMR spectrum of 1 (CDCl₃, 293 K, 76 MHz).

NMR Spectra of phenyl bis-triazole macrocycle 3



Figure S3. ¹H NMR spectrum of 3 (d₆-acetone, 293 K, 500 MHz).



Figure S4. ¹³C NMR spectrum of 3 (CDCl₃, 293 K, 76 MHz).

NMR Spectra of propyl bis-triazole macrocycle 4



Figure S5. ¹H NMR spectrum of 4 (d₆-acetone, 293 K, 500 MHz).



Figure S6. ¹³C NMR spectrum of 4 (CDCl₃, 293 K, 76 MHz).

NMR Spectra of bis-triazolium macrocycle 5·(BF₄)₂



Figure S7. ¹H NMR spectrum of 5·(BF₄)₂ (CD₃CN, 293 K, 300 MHz).



Figure S8. ¹³C NMR spectrum of 5·(BF₄)₂ (CD₃CN, 293 K, 76 MHz).

NMR Spectra of phenyl bis-triazole thread 6



Figure S9. ¹H NMR spectrum of 6 (CDCl₃, 293 K, 300 MHz).



Figure S10. ¹³C NMR spectrum of 6 (CDCl₃, 293 K, 76 MHz).

NMR Spectra of propyl bis-triazole thread 7





Figure S12. ¹³C NMR spectrum of 7 (CDCl₃, 293 K, 76 MHz).

NMR Spectra of phenyl bis-triazolium thread $8 \cdot (BF_4)_2$



Figure S13. ¹H NMR spectrum of 8·(BF₄)₂ (d₆-acetone, 293 K, 500 MHz).



Figure S14. ¹³C NMR spectrum of 8·(BF₄)₂ (CDCl₃, 293 K, 76 MHz).





Figure S15. ¹H NMR spectrum of 9·(BF₄)₂ (d₆-acetone, 293 K, 500 MHz).



Figure S16. ¹³C NMR spectrum of 9·(BF₄)₂ (CDCl₃, 293 K, 76 MHz).

NMR Spectra of phenyl bis-triazole axle 11



(ppm)

Figure S18. ¹³C NMR spectrum of 11 (CDCl₃, 293 K, 76 MHz).

NMR Spectra of propyl bis-triazole axle 12



Figure S19. ¹H NMR spectrum of 12 (CDCl₃, 293 K, 300 MHz).



Figure S20. ¹³C NMR spectrum of **12** (CDCl₃, 293 K, 76 MHz).





Figure S21. ¹H NMR spectrum of 14·(BF₄)₂ (d₆-DMSO, 293 K, 300 MHz).



Figure S22. ¹³C NMR spectrum of 14·(BF₄)₂ (d₆-DMSO, 293 K, 76 MHz).



Figure S23. ¹H NMR spectrum of 15·(BF₄)₂ (d₆-DMSO, 293 K, 300 MHz).



Figure S24. ¹³C NMR spectrum of 15·(BF₄)₂ (d₆-DMSO, 293 K, 76 MHz).

NMR Spectra of bis(triazole xylyl azide) 19



Figure S25. ¹H NMR spectrum of 19 (CDCl₃, 293 K, 300 MHz).



Figure S26. ¹³C NMR spectrum of 19 (CDCl₃, 293 K, 76 MHz).

NMR Spectra of bis(triazolium xylyl azide) 20·(BF₄)₂



Figure S27. ¹H NMR spectrum of 20·(BF₄)₂ (d₆-acetone, 293 K, 300 MHz).



Figure S28. ¹³C NMR spectrum of 20·(BF₄)₂ (d₆-acetone, 293 K, 76 MHz).

NMR Spectra of bis-triazolium rotaxane $22 \cdot (PF_6)_2$



Figure S30. ¹³C NMR spectrum of 22·(PF₆)₂ (1:1 CDCl₃:CD₃OD, 293 K, 126 MHz).

NMR Spectra of bis-triazole bis-azide 24



Figure S31. ¹H NMR spectrum of 24 (CDCl₃, 293 K, 300 MHz).



Figure S32. ¹³C NMR spectrum of 24 (CDCl₃, 293 K, 76 MHz).

NMR Spectra of bis-triazolium bis-azide $25 \cdot (BF_4)_2$



Figure S33. ¹H NMR spectrum of 25·(BF₄)₂ (CD₃CN, 293 K, 300 MHz).



Figure S34. ¹³C NMR spectrum of 25·(BF₄)₂ (CD₃CN, 293 K, 76 MHz).

NMR Spectra of *hetero*-catenane 26 · (BPh₄)₄



Figure S35. ¹H NMR spectrum of 26 · (BPh₄)₄ (d₆-acetone, 293 K, 500 MHz).



Figure S36. Truncated and labelled ¹H NMR spectrum of $26 \cdot (BPh_4)_4$ (d₆-acetone, 293 K, 500 MHz). Peaks that could belong to either macrocyclic component are labelled purple. In one case, the two *h* environments appear to be inequivalent, these have been labelled h^{*}.

Due to the similar chemical environments of a number of signals, it is not possible to distinguish between them (e.g. it is not possible to say which of the two triazolium resonances e belongs to which macrocycle). Despite this, and despite the slight impurities present, all proton resonances can be reasonably assigned, which along with the high resolution ESI mass spectrometry data confirm the identity of the tetra-triazolium catenane.



Scheme S1. Synthesis of bis-triazole macrocycles S1 and S2.



Scheme S2. Synthesis of bis(triazolium octyl azide) S4·(BF₄)₂.



Scheme S3. Attempted synthesis of octyl-containing bis-triazolium rotaxanes (rotaxanes appeared to be formed in $\sim 25\%$ yield, but could not be separated from by-products.



Scheme S4. Attempted synthesis of *homo*-catenane (catenane appeared to be formed but could not be separated from macrocycle by-products).

General remarks

1,8-Diazidooctane was prepared from 1,8-dibromooctane using NaN₃ in DMSO; 3,5diethynylpyridine was prepared by deprotecting 3,5bis(trimethylsilyl)ethynylpyridine^{S1} using KOH in methanol; 5-*f*butyl-1,3-diethynyl benzene was prepared as previously described.^{S2,S3}

Pyridyl macrocycle S1

The bis-azide **1** (0.155 g, 0.300 mmol) and 3,5-diethynyl pyridine (0.038 g, 0.30 mmol) were dissolved in CH_2Cl_2 (300 mL); DIPEA (0.17 mL, 0.13 g, 1.0 mmol), TBTA (0.032 g, 0.060 mmol) and $[Cu(CH_3CN)_4](PF_6)$ (0.022 g, 0.060 mmol) were added and the reaction stirred at room temperature under a nitrogen atmosphere for 3 days. It was taken to dryness under reduced pressure and purified by preparative TLC (4% CH₃OH in CHCl₃) to give **S1** as a white powder. Yield: 0.082 g (43%).

¹H NMR (CDCl₃): 9.12 (d, ${}^{4}J$ = 2.0 Hz, 2H, py-*H*), 8.32 (t, ${}^{4}J$ = 2.0 Hz, 1H, py-*H*), 8.10 (s, 2H, trz-*H*), 6.72–6.81 (m, 8H, hydroquinone-*H*), 4.83 (t, ${}^{3}J$ = 4.7 Hz, 4H, CH₂-trz), 4.33 (t, ${}^{3}J$ = 4.7 Hz, 4H, hydroquinone-CH₂), 4.02 (t, ${}^{3}J$ = 4.8 Hz, 4H, hydroquinone-CH₂), 3.78 (t, ${}^{3}J$ = 4.8 Hz, 4H, CH₂), 3.63–3.69 (m, 8H, CH₂). ¹³C NMR (d₆-DMSO): 152.9, 152.0, 145.2, 143.5, 129.4, 128.8, 123.3, 115.8, 115.2, 70.0, 69.9, 68.9, 67.5, 67.0, 49.8. HRESI-MS (pos.): 666.2647, calc. for [C₃₃H₃₇N₇O₇·Na]⁺ = 666.2647.

5-^{*t*}Butylphenyl macrocycle S2

The bis-azide **1** (0.155 g, 0.300 mmol) and 5-'butyl-1,3-diethynyl benzene (0.055 g, 0.30 mmol) were dissolved in CH_2Cl_2 (300 mL); DIPEA (0.17 mL, 0.13 g, 1.0 mmol), TBTA (0.032 g, 0.060 mmol) and $[Cu(CH_3CN)_4](PF_6)$ (0.022 g, 0.060 mmol) were added and the reaction stirred at room temperature under a nitrogen atmosphere for 3 days. It was taken to dryness under reduced pressure and purified by preparative TLC (3% CH₃OH in CH₂Cl₂) to give **S2** as a white powder. Yield: 0.075 g (36%).

¹H NMR (CDCl₃): 8.04 (s, 2H, trz-*H*), 8.02 (d, ${}^{4}J$ = 1.5 Hz, 2H, ph-*H*), 7.75 (t, ${}^{4}J$ = 1.5 Hz, 1H, ph-*H*), 6.74–6.81 (m, 8H, hydroquinone-*H*), 4.79 (t, *J* = 4.7 Hz, 4H, CH₂-trz), 4.31 (t, *J* = 4.7 Hz, 4H, hydroquinone-CH₂), 4.02 (t, *J* = 4.8 Hz, 4H, hydroquinone-CH₂), 3.80 (t, *J* = 4.8 Hz, 4H, CH₂), 3.64–3.73 (m, 8H, CH₂), 1.42 (s, 9H, CH₃). ¹³C NMR (CDCl₃): 153.8, 152.9, 152.2, 148.1, 130.9, 122.9, 121.3, 120.7,

115.8, 110.1, 70.9, 70.9, 69.8, 68.2, 67.6, 50.3, 35.2, 31.5. HRESI-MS (pos.): 721.3314, calc. for $[C_{38}H_{46}N_6O_7 \cdot Na]^+ = 721.3320$.

Bis(triazole octyl azide) S3

1,8-Diazidooctane (1.37 g, 7.00 mmol) and 1,3-diethynylbenzene (0.093 mL, 0.088 g, 0.70 mmol) were dissolved in CH_2Cl_2 (50 mL). DIPEA (0.25 mL, 0.18 g, 1.4 mmol), TBTA (0.074 g, 0.14 mmol) and $[Cu(CH_3CN)_4](PF_6)$ (0.052 g, 0.14 mmol) were added and the resulting yellow solution stirred at room temperature under a nitrogen atmosphere for 16 hours. It was carefully concentratedunder reduced pressure and then purified by column chromatography (2% CH₃OH in CHCl₃) to give **S3** (0.24 g, 66%) as a slightly off-white waxy solid.

¹H NMR (CDCl₃): 8.30 (t, ${}^{4}J$ = 1.6 Hz, 1H, ph-*H*), 7.85 (s, 2H, trz-*H*), 7.83 (dd, ${}^{3}J$ = 7.7 Hz, ${}^{4}J$ j,k = 1.6 Hz, 2H, ph-*H*), 7.49 (t, ${}^{3}J$ = 7.7 Hz, 1H, ph-*H*), 4.42 (t, ${}^{3}J$ = 7.1 Hz, 4H, trz-CH₂), 3.25 (t, ${}^{3}J$ = 6.9 Hz, 4H, CH₂-N₃), 1.92–2.01 (m, 4H, trz-CH₂-CH₂), 1.54–1.63 (m, 4H, CH₂-CH₂-N₃), 1.30–1.41 (m, 16H, CH₂). ¹³C NMR (CDCl₃): 147.4, 131.3, 129.5, 125.3, 122.8, 119.9, 51.4, 50.5, 30.3, 28.9, 28.9, 28.8, 26.6, 26.4. HRESI-MS (pos.): 541.3235, calc. for [C₂₆H₃₈N₁₂·Na]⁺ = 541.3235.

Bis(triazolium octyl azide) S4·(BF₄)₂

The bis(triazole) thread **S3** (0.156 g, 0.300 mmol) was dissolved in dry CH_2Cl_2 (25 mL). (Me₃O)(BF₄) (0.097 g, 0.66 mmol) was added and the mixture stirred at room temperature under a nitrogen atmosphere over the weekend. The reaction was quenched by the addition of CH_3OH (1 mL), and then taken to dryness under reduced pressure. Purification by preparative TLC (10% CH_3OH in CH_2Cl_2) gave **S4**·(**BF**₄)₂ as a very pale yellow oil. Yield: 0.163 g (75%).

¹H NMR (d₆-acetone): 9.11 (s, 2H, trz⁺-*H*), 8.30 (s, 1H, ph-*H*), 8.13 (d, ${}^{3}J = 8.3$ Hz, 2H, ph-*H*), 7.98 (t, ${}^{3}J = 8.3$ Hz, 1H, ph-*H*), 4.83 (t, ${}^{3}J = 7.3$ Hz, 4H, trz⁺-*H*), 4.50 (s, 6H, trz⁺-CH₃), 3.32 (t, ${}^{3}J = 6.9$ Hz, 4H, CH₂-N₃), 2.10–2.19 (m, 4H, trz⁺-CH₂-CH₂), 1.55–1.63 (m, 4H, CH₂-CH₂-N₃), 1.34–1.51 (m, 16H, CH₂). ¹³C NMR (CDCl₃): 141.8, 132.3, 130.8, 130.5,129.0, 123.7, 54.2, 51.3, 38.7, 28.9, 28.7, 28.6, 26.5, 25.9 (one resonance not observed). ¹⁹F NMR (d₆-acetone): –151.4 (m). HRESI-MS (pos.): 274.1903, calc. for $[C_{28}H_{44}N_{12}]^{2+} = 274.1900$.



Figure S37. ¹H NMR spectrum of S1 (d₆-DMSO, 293 K, 300 MHz).



Figure S38. ¹³C NMR spectrum of S1 (d₆-DMSO, 293 K, 76 MHz).

NMR Spectra of S2



Figure S39. 1 H NMR spectrum of S2 (d₆-acetone, 293 K, 500 MHz).



Figure S40. ¹³C NMR spectrum of S2 (CDCl₃, 293 K, 76 MHz).

NMR Spectra of bis(triazole octyl azide) S3



Figure S41. ¹H NMR spectrum of S3 (CDCl₃, 293 K, 300 MHz).



Figure S42. ¹³C NMR spectrum of S3 (CDCl₃, 293 K, 76 MHz).

NMR Spectra of bis(triazolium octyl azide) S4·(BF₄)₂



Figure S43. ¹H NMR spectrum of S4·(BF₄)₂ (CDCl₃, 293 K, 300 MHz).



Figure S44. ¹³C NMR spectrum of S4·(BF₄)₂ (CD₃CN, 293 K, 76 MHz).

Solid state structure of S1



Figure S45. Solid state structure of S1 (hydrogen atoms and solvent molecules omitted for clarity).

ROESY NMR Spectra of 8·Cl·10, 5·SO₄·8 and 22·(PF₆)₂



Figure S46. Truncated ROESY NMR spectrum of a 1:1:1 mixture of $8 \cdot (BF_4)_2$, 10 and TBA ·Cl showing selected inter-component couplings (d₆-acetone, 293 K, 500 MHz).



Figure S47. Truncated ROESY NMR spectrum of a 1:1:1 mixture of $5 \cdot (BF_4)_2$, $8 \cdot (BF_4)$ and TBA \cdot SO₄ showing selected inter-component couplings (d₆-DMSO, 293 K, 500 MHz).



Figure S48. Truncated ROESY NMR spectrum of $22 \cdot (PF_6)_2$ showing selected inter-component couplings (1:1 CDCl₃:CD₃OD, 293 K, 500 MHz).

Titration protocols and additional binding isotherms

Spectra for ¹H NMR titrations were recorded at 293 K on a Varian Unity Plus 500 spectrometer with ¹H operating at 500 MHz. Initial sample volumes were 0.50 mL and concentrations were 2.0 mM of host. Solutions (100 mM) of anions as their tetrabutylammonium salts were added in aliquots, the samples thoroughly shaken and spectra recorded. Spectra were recorded at 0, 0.2, 0.4, 0.6, 0.8, 1.0, 1.2, 1.4, 1.6, 1.8, 2.0, 2.5, 3.0, 4.0, 5.0, 7.0 and 10 equivalents. Stability constants were obtained by analysis of the resulting data using the WINEQNMR2^{S4} computer program, monitoring the triazole C–H proton resonance in all cases.

Estimates for the association constant and the limiting chemical shifts were added to the program's input file. The parameters were refined by non-linear leastsquares analysis using WINEQNMR2^{S4} to achieve the best fit between observed and calculated chemical shifts. The input parameters for the final chemical shift and association constant were adjusted based on the program output until convergence was reached. Comparison of the calculated and experimental binding isotherms demonstrated that an appropriate model with an appropriate 1:1 binding stoichiometry was being used. The 1:1 stoichiometry was also confirmed using approximations of Job plots. A graph of $\Delta\delta \cdot \chi_H$ against χ_H was plotted, with a 1:1 binding stoichiometry corresponding to a maximum $\delta \cdot \chi_H$ of approximately 0.5 (χ_H = mole fraction of host, $\Delta\delta$ = change in chemical shift relative to free host).



Figure S49. Change in chemical shift of triazole proton of **3** and **4** upon addition of chloride anion. Circles represent data points, lines represent binding isotherms calculated using WINEQNMR2^{S4} (d_6 -acetone, 293 K).



Figure S50. Change in chemical shift of triazolium proton of 5^{2+} upon addition of anions. Circles represent data points, lines represent binding isotherms calculated using WINEQNMR2^{S4} (CD₃CN, 293 K).



Figure S51. Change in chemical shift of triazolium proton of 8^{2+} and 9^{2+} upon addition of anions. Circles represent data points, lines represent binding isotherms calculated using WINEQNMR2^{S4} (d₆-acetone, 293 K).



Figure S52. Change in chemical shift of hydroquinone resonance of **10** upon addition of either **8**·(**BF**₄)₂ and one equivalent of TBA·anion or **9**·(**BF**₄)₂ and one equivalent of TBA·anion. Circles represent data points, lines represent binding isotherms calculated using WINEQNMR2^{S4} (anion = Cl⁻ or Br, d₆-acetone, 293 K).

Comments on X-ray crystallography

For several of the structures presented, it was only possible to grow small and/or thin single crystals of the macrocycles, and these crystals often diffracted weakly. In these cases, synchrotron radiation or Cu radiation and long exposure times were used, but data were often weak. Nevertheless, the overall structure of the macrocycles and macrocycle–anion complexes is clear.

In some cases poorly-resolved areas of electron density, presumably arising from illdefined solvent molecules, was present. Where this could not be modelled satisfactorily, PLATON-SQUEEZE^{S5,S6} was used to include the electron density in the refinement.

Specific details for each structure are given in the CIF file.

References

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