## Cu(II) and Co(II) ternary complexes of quinolone antimicrobial drug enoxacin and levofloxacin: Structure and biological evaluation

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1				
Cu(1)-O(1)	1.930(6)	N(6)-Cu(1)-N(5)	81.1(3)	
Cu(1)-O(2)	1.939(6)	O(1)-Cu(1)-N(1) <sup>A</sup>	97.3(3)	
Cu(1)-N(6)	1.982(7)	O(2)-Cu(1)-N(1) <sup>A</sup>	98.8(3)	
Cu(1)-N(5)	2.015(7)	N(6)-Cu(1)-N(1) <sup>A</sup>	94.5(3)	
$Cu(1)-N(1)^{A}$	2.278(8)	N(5)-Cu(1)-N(1) <sup>A</sup>	99.5(3)	
Cu(2)-O(5)	1.917(6)	O(5)-Cu(2)-O(4)	93.4(3)	
Cu(2)-O(4)	1.943(6)	O(5)-Cu(2)-N(11)	162.5(3)	
Cu(2)-N(11)	2.000(7)	O(4)-Cu(2)-N(11)	90.9(3)	
Cu(2)-N(12)	2.002(8)	O(5)-Cu(2)-N(12)	91.2(3)	
$Cu(2)-N(7)^{B}$	2.291(7)	O(4)-Cu(2)-N(12)	165.0(3)	
O(1)-Cu(1)-O(2)	93.9(3)	N(11)-Cu(2)-N(12)	80.7(3)	
O(1)-Cu(1)-N(6)	166.1(3)	O(5)-Cu(2)-N(7) <sup>B</sup>	95.9(3)	
O(2)-Cu(1)-N(6)	91.6(3)	O(4)-Cu(2)-N(7) <sup>B</sup>	96.4(3)	
O(1)-Cu(1)-N(5)	89.6(3)	N(11)-Cu(2)-N(7) <sup>B</sup>	100.5(3)	
O(2)-Cu(1)-N(5)	160.7(3)	N(12)-Cu(2)-N(7) <sup>B</sup>	97.4(3)	
2				
Co(1)-O(5)	2.046(3)	O(4)-Co(1)-N(7)	97.24(12)	
Co(1)-O(1)	2.073(2)	O(5)-Co(1)-O(2)	175.81(10)	
Co(1)-O(4)	2.072(3)	O(1)-Co(1)-O(2)	84.48(10)	
Co(1)-N(7)	2.096(3)	O(4)-Co(1)-O(2)	89.34(10)	
Co(1)-O(2)	2.097(3)	N(7)-Co(1)-O(2)	89.72(11)	
Co(1)-N(8)	2.106(3)	O(5)-Co(1)-N(8)	92.32(12)	

Table S1 Selected bond lengths (Å) and angles (deg) for complex 1–4.

O(5)-Co(1)-O(1)	93.43(11)	O(1)-Co(1)-N(8)	96.12(12)
O(5)-Co(1)-O(4)	87.00(11)	O(4)-Co(1)-N(8)	174.47(12)
O(1)-Co(1)-O(4)	89.40(10)	N(7)-Co(1)-N(8)	77.31(13)
O(5)-Co(1)-N(7)	92.77(12)	O(2)-Co(1)-N(8)	91.51(12)
O(1)-Co(1)-N(7)	171.14(12)		
3			
Cu(1)-O(2)	1.912(3)	O(2)-Cu(1)-N(5)	168.07(12)
Cu(1)-O(1)	1.937(2)	O(1)-Cu(1)-N(5)	92.79(11)
Cu(1)-N(4)	2.001(3)	N(4)-Cu(1)-N(5)	81.03(12)
Cu(1)-N(5)	2.005(3)	O(2)-Cu(1)-O(9)	97.88(12)
Cu(1)-O(9)	2.280(3)	O(1)-Cu(1)-O(9)	95.85(11)
O(2)-Cu(1)-O(1)	92.84(11)	N(4)-Cu(1)-O(9)	91.61(12)
O(2)-Cu(1)-N(4)	91.97(12)	N(5)-Cu(1)-O(9)	91.99(12)
O(1)-Cu(1)-N(4)	170.49(11)		
4			
Co(1)-O(2)	2.069(4)	$O(3)^{A}$ -Co(1)-O(3)	96.5(2)
Co(1)-O(2) <sup>A</sup>	2.069(4)	O(2)-Co(1)-N(4)	97.75(18)
Co(1)-O(3) <sup>A</sup>	2.095(4)	$O(2)^{A}$ -Co(1)-N(4)	89.68(18)
Co(1)-O(3)	2.095(4)	$O(3)^{A}$ -Co(1)-N(4)	169.60(19)
Co(1)-N(4)	2.124(5)	O(3)-Co(1)-N(4)	92.63(19)
Co(1)-N(4) <sup>A</sup>	2.124(5)	O(2)-Co(1)-N(4) <sup>A</sup>	89.68(18)
O(2)-Co(1)-O(2) <sup>A</sup>	170.4(2)	$O(2)^{A}$ -Co(1)-N(4) <sup>A</sup>	97.75(18)
O(2)-Co(1)-O(3) <sup>A</sup>	87.85(17)	O(3) <sup>A</sup> -Co(1)-N(4) <sup>A</sup>	92.63(18)
$O(2)^{A} - Co(1) - O(3)^{A}$	85.77(17)	O(3)-Co(1)-N(4) <sup>A</sup>	169.60(19)
O(2)-Co(1)-O(3)	85.77(17)	N(4)-Co(1)-N(4) <sup>A</sup>	78.7(3)
O(2) <sup>A</sup> -Co(1)-O(3)	87.85(17)		
Symmetry transformati	ons used to generate e	equivalent atoms:	

For **1**, A) –*x* + 1, –*y* + 2, –*z* + 1; B) –*x* + 1, –*y* + 1, –*z*. For **4**, A) –*x* + 1, *y*, –*z* + 3/2.

Table S2 Hydrogen	bonds (Å and °	) for comple	xes 1–4.
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D–H…A	$d(\mathbf{H}\cdots\mathbf{A})$	$d(\mathbf{D}\cdots\mathbf{A})$	< DHA
1			
N(7)-H(7)O(16) <sup>C</sup>	2.26	3.157(11)	167.4
N(1)-H(1)O(13) <sup>D</sup>	2.33	3.129(12)	145.8
O(16)-H(16B)O(9) <sup>A</sup>	2.20(8)	2.924(19)	143(12)
O(16)-H(16A)O(3) <sup>A</sup>	2.17(9)	2.876(11)	140(11)
O(15)-H(15A)O(3) <sup>E</sup>	1.99(10)	2.83(2)	166(34)
Symmetry code: A) $-x + 1, -y$	+2, -z+1; C) -x+1, -y -	+ 1, - <i>z</i> + 1; D) <i>x</i> , <i>y</i> + 1, <i>z</i> ; E	(x) - x + 2, -y + 2, -z + 2
1			
2			
O(11)-H(11A)O(3) <sup>A</sup>	2.01(2)	2.843(5)	165(7)
O(14)-H(14A)O(13) <sup>B</sup>	2.26(4)	2.965(17)	141(6)
O(11)-H(11B) N(6) <sup>C</sup>	2 14(4)	2803(8)	149(7)

O(12)-H(12C)O(5)	1.953(16)	2.802(5)	172(7)
O(12)-H(12D)O(11)	2.12(5)	2.826(7)	140(6)
N(3)-H(3A)O(9) <sup>D</sup>	2.12	3.008(10)	166.7
N(3)-H(3B)O(3) <sup>D</sup>	1.82	2.712(4)	168.6
Symmetry code: A) $x + 1, y, z$ ;	B) $-x + 1$ , $-y + 1$ , $-z + 1$ ;	C) $-x + 1$ , $-y + 1$ , $-z$ ; D) $x$ ,	y + 1, z
3			
O(9)-H(9A)O(10) <sup>A</sup>	1.92(3)	2.708(5)	154(5)
O(9)-H(9B)O(11)	1.905(13)	2.751(6)	172(5)
O(10)-H(10A)O(5) <sup>A</sup>	2.35(4)	3.021(7)	136(5)
O(11)-H(11F)O(3) <sup>B</sup>	2.17(4)	2.820(6)	133(5)
Symmetry code: A) $-x + 1, -y$	+1, -z + 1; B) -x, -y + 1, -x	-z + 1.	



Fig. S1. Time-dependent stability studies on complexes **1-4** in TBS buffer solution monitored by UV-vis absorption spectra.



Fig. S2.  $\pi$ - $\pi$  stacking (purple dashed lines) between the aromatic planes of the HEn ligand and bipyridine of complex 1.



Fig. S3. 2D supramolecular network of **3** constructed by hydrogen bonding (green dashed lines) and  $\pi$ - $\pi$  stacking interactions (purple dashed lines) in the *ac* plane.











Fig. S4. UV absorption spectra of **2** (A), **3** (B), HEn (C) and HLevo (D) in the absence (---) and presence (---) of CT DNA with increasing [DNA]/[compound] ratios. The arrows show the changes upon increasing amounts of CT DNA. Inset: plot of [DNA]/( $\epsilon_a$ - $\epsilon_f$ ) versus [DNA].





Fig. S5. Circular dichromism spectra of CT DNA bound with HEn, Hlevo and complexes 2-4 with [DNA] =  $1 \times 10^{-4}$  M.











Fig. S6. Fluorescence emission spectra of EB-DNA in the absence (dashed line) and presence (colored solid lines) of HEn, HLevo and complexes **2-4** as competitive agents with increasing [compound]/[EB] ratios from 1:1 to 6:1.



Fig. S7. Stern-Volmer quenching plot of EB bound to CT-DNA by complexes 1-4 and HEn, HLevo.





Fig. S8. Emission spectra ( $\lambda_{exit}$  = 295 nm) for HSA ([HSA] = 3 µM) in buffer solution in the absence (dashed line) and presence (colored solid lines) of increasing amounts of compound (r = [compound] / [HSA] = 0-10).





Fig. S9. Emission spectra ( $\lambda_{exit} = 295$  nm) for BSA ([BSA] = 3  $\mu$ M) in buffer solution in the absence (dashed line) and presence (colored solid lines) of increasing amounts of compound (r = [compound] / [BSA] = 0-10, except r = 0-7 for **3** and r = 0-6 for **4**).



Fig. S10. Stern-Volmer quenching plot of HSA for complexes 1-4 and HEn, HLevo ligand, respectively.



Fig. S11. Stern-Volmer quenching plot of BSA for complexes 1-4 and HEn, HLevo ligand, respectively.

## **Equations:**

$$\frac{[DNA]}{(\varepsilon_a - \varepsilon_f)} = \frac{[DNA]}{(\varepsilon_b - \varepsilon_f)} + \frac{1}{K_b(\varepsilon_b - \varepsilon_f)} \qquad \text{Eq. (S1)}$$

where [DNA] is the concentration of DNA in base pairs,  $\varepsilon_f$  is the extinction coefficient for the free compound at the corresponding  $\lambda_{max}$ ,  $\varepsilon_a = A_{obsd}$ /[compound] and  $\varepsilon_b$  is the extinction

coefficient for the compound in the fully bound form.

$$\frac{I_0}{I} = 1 + K_{sv}[Q]$$
 Eq. (S2)

where  $I_0$  and I are the emission intensities in the absence and the presence of the complex, respectively.

$$\frac{I_0}{I} = 1 + k_q \tau_0[Q] = 1 + K_{sv}[Q] \qquad \text{Eq. (S3)}$$

where Io = the initial tryptophan fluorescence intensity of SA, I = the tryptophan fluorescence intensity of SA after the addition of the quencher,  $k_q$  = the quenching rate constants of SA,  $K_{sv}$ = the dynamic quenching constant,  $\tau_o$  = the average lifetime of SA without the quencher, [Q] = the concentration of the quencher respectively,  $K_{sv} = k_q \tau_o$  (taking as fluorescence lifetime ( $\tau_o$ ) of tryptophan in SA at around 10<sup>-8</sup> s).

$$\frac{\Delta I / I_0}{[Q]} = nK - K \frac{\Delta I}{I_0} \qquad \text{Eq. (S4)}$$

Where n is the number of binding sites per albumin and K (M<sup>-1</sup>) is the association binding constant, K (M<sup>-1</sup>) may be calculated from the slope in plots  $\Delta I/(I_0[Q])$  versus  $\Delta I/I_0$  and n is given by the ratio of y intercept to the slope.