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## **Electronic Supplementary Information (ESI)**

for

# A colorimetric and near-infrared fluorescent probe for biothiols and its application in living cells

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HR-MS spectrum of compound 2





#### 152.2 157.2 157.2 157.2 152.3

-0.0



MS (EI) spectrum of probe 1



HR-MS spectrum of probe 1

#### 2. Additional spectra



Fig. S1 (a) Absorption spectra of probe 1 at different concentrations. (b) Absorbance intensity changes at 380 nm of probe 1 as a function of its concentration from 0-50  $\mu$ M. (c) Absorption

spectra of compound **2** at different concentrations. (d) Absorbance intensity changes at 560 nm of compound **2** as a function of its concentration from 0-50  $\mu$ M. (e) Fluorescence spectra of compound **2** at different concentrations. (f) Fluorescence intensity changes of compound **2** at 706 nm as a function of its concentration from 0-50  $\mu$ M. All spectra were collected in PBS buffer (10 mM, pH 7.4) with 50% DMSO at 37 °C. For fluorescence measurement,  $\lambda_{ex} = 560$  nm with slit width: (10, 10).



**Fig. S2** Time-dependent of fluorescence kinetics spectra of probe **1** (20  $\mu$ M) upon addition of Cys (20  $\mu$ M) in PBS buffer (10 mM, pH 7.4) with 50% DMSO at 37°C. The reaction is monitored at 706 nm.  $\lambda_{ex} = 560$  nm, slit width: (10, 10).



Fig. S3 (a) Absorption spectra changes of probe 1 (20  $\mu$ M) upon addition of different concentrations of Cys in PBS buffer (10 mM, pH 7.4) with 50% DMSO at 37 °C. Final concentration of Cys: 0, 6, 10, 30, 40, 50, 60, 70, 100, 120, 140, 160 and 180  $\mu$ M, respectively. (b) Absorbance changes at 560 nm of probe 1 (20  $\mu$ M) against concentration of Cys. Each spectrum was obtained 30 min after Cys addition.



Fig. S4 (a) Fluorescence spectra changes of probe 1 (20  $\mu$ M) upon addition of different concentrations of Cys in PBS buffer (10 mM, pH 7.4) with 50% DMSO at 37 °C. Final concentration of Cys: 0, 2, 6, 10, 20, 40, 60, 80, 100, 120, 140, 160 and 180  $\mu$ M, respectively. (b) Fluorescence intensity changes at 706 nm of probe 1 (20  $\mu$ M) against concentration of Cys. Each spectrum was obtained 30 min after Cys addition.  $\lambda_{ex} = 560$  nm, slit width: (10, 10).



**Fig. S5** (a) Absorbance responses of probe **1** (20  $\mu$ M) at 560 nm in the presence and absence of Cys (5 eq) under different pHs. (b) Fluorescence intensity responses of probe **1** (20  $\mu$ M) at 706 nm in the presence and absence of Cys (5 eq) under different pHs. All experiment was performed in PBS buffer (10 mM) with 50% DMSO at 37 °C and each data was obtained 30 min after addition of Cys. For fluorescence measurements,  $\lambda_{ex} = 560$  nm, slit width: (10, 10).

#### Data for investigation of the sensing mechanism



This experiment was performed according to a reported procedure (see Ref: X. Yang, Y. Guo and R. Strongin, *Angew. Chem., Int. Ed.*, 2011, **50**, 10690–10693). To a 100 mL flask, probe **1** (50 mg, 0.14 mmol) and Cys (21 mg, 1.25 eq) were combined in 30 mL of MeOH:  $H_2O$  (90: 10, v/v) solution, and the mixture stirred at room temperature for 1 h. Then, Et<sub>3</sub>N (30 µL) was added and the solution stirred overnight. The solvents was removed under reduced pressure and the crude

product was subjected to column chromatography (eluted with  $CH_2Cl_2$ : MeOH, 5:1, v/v) to afford a red solid (23 mg), which was proved to be compound **2** (see below).



Fig. S6 (a) The thin layer chromatography (TLC) plate under different light to compare probe 1, the reference sample of 2 and the isolated product from the reaction mixture of probe 1 with Cys. Spots on the TLC plate are: a. probe 1; b. the isolated reaction product of probe 1 and Cys; c. mixture of b and 2; d. the reference sample of 2. The eluent for TLC: hexane:ethyl acetate = 3:1 (v/v). (b) Fluorescent spectra of probe 1, the isolated product and the reference sample of 2 in PBS buffer (10 mM, pH 7.4) with 50% DMSO at 37 °C.  $\lambda_{ex} = 560$  nm, slit width: (10,10).



Fig. S7 <sup>1</sup>H NMR spectrum of the isolated product from the reaction of probe 1 and Cys, which is identical to that of the reference sample of 2 (see above).



Fig. S8 MS spectrum of the isolated product from the reaction of probe 1 and Cys, which showed the expected mass of 2.



**Fig. S9** (a) Absorption spectral responses of probe **1** (20  $\mu$ M) upon addition of 100  $\mu$ M various anions and Cys. Anions are F<sup>-</sup>, Cl<sup>-</sup>, Br<sup>-</sup>,  $\Gamma$ , NO<sub>3</sub><sup>-</sup>, NO<sub>2</sub><sup>-</sup>, AcO<sup>-</sup>, SCN<sup>-</sup>, CO<sub>3</sub><sup>2-</sup>, SO<sub>4</sub><sup>2-</sup>, SO<sub>3</sub><sup>2-</sup>, S<sub>2</sub>O<sub>3</sub><sup>2-</sup>, HS<sup>-</sup>, S<sup>2-</sup>, CN<sup>-</sup>, O<sub>2</sub>•<sup>-</sup>, ClO<sup>-</sup>, H<sub>2</sub>O<sub>2</sub> and OH•. (b) A comparison of absorbance intensity changes of probe **1** (20  $\mu$ M) at 560 nm upon addition of 100  $\mu$ M various anions and Cys. All experiment was performed in PBS buffer (10 mM, pH 7.4) with 50% DMSO at 37 °C and each spectrum was obtained 30 min after addition of various anions.



Fig. S10 (a) Fluorescent spectral responses of probe 1 (20  $\mu$ M) upon addition of 100  $\mu$ M various anions and Cys. Anions are F<sup>-</sup>, Cl<sup>-</sup>, Br<sup>-</sup>, I<sup>-</sup>, NO<sub>3</sub><sup>-</sup>, NO<sub>2</sub><sup>-</sup>, AcO<sup>-</sup>, SCN<sup>-</sup>, CO<sub>3</sub><sup>2-</sup>, SO<sub>4</sub><sup>2-</sup>, SO<sub>3</sub><sup>2-</sup>,

 $S_2O_3^{2-}$ , HS<sup>-</sup>, S<sup>2-</sup>, CN<sup>-</sup>,  $O_2^{\bullet-}$ , ClO<sup>-</sup>, H<sub>2</sub>O<sub>2</sub> and OH•. (b) A comparison of fluorescence intensity changes of probe 1 (20  $\mu$ M) at 706 nm upon addition of 100  $\mu$ M various anions and Cys. All experiment was performed in PBS buffer (10 mM, pH 7.4) with 50% DMSO at 37 °C and each spectrum was obtained 30 min after addition of various anions.  $\lambda_{ex} = 560$  nm , slit width: (10, 10).



**Fig. S11** (a)-(c): Scanning kinetics of the UV-vis spectra changes of probe **1** (20  $\mu$ M) against time in the presence of 5 equiv of Cys (a), Hcy (b) and GSH (c), respectively. All spectra were measured in PBS buffer (10 mM, pH 7.4) with 50% DMSO at 37 °C. (d) The corresponding absorbance changes of probe **1** at 560 nm against time for Cys, Hcy and GSH.



**Fig. S12** Fluorescence kinetics of probe 1 (20  $\mu$ M) at 706 nm in the absence and presence of biothiols (Cys, Hcy and GSH, 100  $\mu$ M each). All spectra were measured in PBS buffer (10 mM, pH 7.4) with 50% DMSO at 37 °C.  $\lambda_{ex}$ = 560 nm, slit width (10,10).

#### Generation of reactive oxygen species (ROS)

#### ClO<sup>-</sup>

The NaOCl solution was added to the probe 1 testing solution at 25 °C.

#### $H_2O_2 \\$

 $H_2O_2$  was added and the mixtures were stirred for 1 hour at 25 °C.

#### OH•

Hydroxyl radicals were generated by the addition of  $Fe^{2+}$  and  $H_2O_2$  at room temperature in PBS buffer (pH 7.4) and the mixture was then stirred for 30 min. (Ref. Y. Zhou, J.-Y. Li, K.-H. Chu, K. Liu, C. Yao, J.-Y. Li, *Chem Commun.* 2012, **48**, 4677–4679.)

#### $O_2 \bullet^-$

O2<sup>•</sup> was generated by an improved pyrogallol autoxidation method reported recently (Ref. X. Li, *J. Agric. Food Chem.*, 2012, **60**, 6418–6424).