

## SUPPORTING INFORMATION

### **Extremely Selective and Fluorescent Detection of Cysteine Or Superoxide: with Aliphatic Ester Hydrolysis**

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## Experimental Section

**General Remarks.** All reagents used herein were used as received from commercial suppliers (Aldrich, Acros, and Junsei companies).  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra were acquired using a Bruker Avance 400 MHz spectrometer. TMS was used as an internal standard.  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectral signals were calibrated internally by the respective protio impurity or carbon resonance of the NMR spectroscopic solvent, *e.g.*, DMSO- $d_6$ . ESI-mass spectrometry was performed on a VG AUTOSPEC ULTIMA by the research support staff at KAIST. This instrument possesses a trisector double focusing magnetic sector analyzer and was operated at a resolution of 80,000. Absorption spectra were measured using a JASCO V-530 UV/Vis spectrophotometer. Fluorescence measurements were carried out with a Shimadzu RF-5301pc spectrofluorophotometer.

**Synthesis of probe 1.** In a 100 mL round bottom flask, Fluorescein (0.50 g, 1.5 mmol) and vinylacetic acid (0.25 mL, 3.0 mmol) were dissolved up in 30 mL of tetrahydrofuran. Then 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (EDC) (0.69 mg, 3.61 mmol) and 4-dimethylaminopyridine (DMAP) (0.367 mg, 3.01 mmol) were added at room temperature. The reaction mixture was allowed to react at room temperature overnight; after this time, the solvent was removed under pressure. The obtained crude material was subjected to column chromatography and purified by using hexane: ethyl acetate (8:2) as an eluent. Probe **1** was obtained as a white solid (0.47 g, 67 % yield).

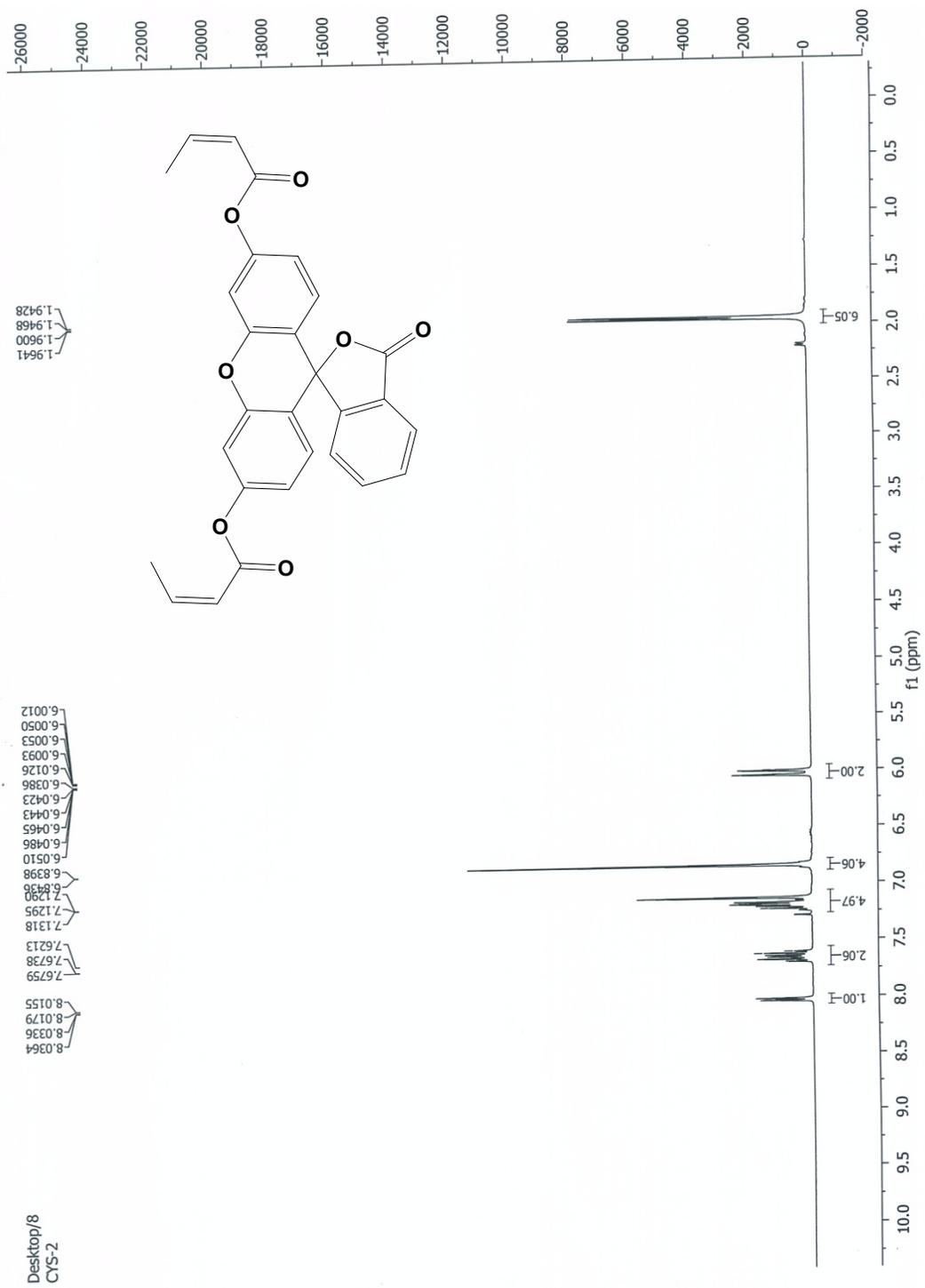
**<sup>1</sup>H NMR spectroscopy:** (400 MHz, CDCl<sub>3</sub>): 8.10 (d, <sup>3</sup>J<sub>(H,H)</sub> = 7.24 Hz, 1H), 7.69-7.60 (2H, m), 7.22-7.12 (5H, m), 6.84 (d, <sup>3</sup>J<sub>(H,H)</sub> = 1.52 Hz, 4H), 6.03 (d, <sup>3</sup>J<sub>(H,H)</sub> = 15.6 Hz, 2H), 1.95 (dd, <sup>4</sup>J<sub>(H,H)</sub>/<sup>3</sup>J<sub>(H,H)</sub> = 1.64, 6.92 Hz, 6H).

**<sup>13</sup>C NMR spectroscopy:** (100 MHz, CDCl<sub>3</sub>): 169.11, 164.10, 152.87, 152.20, 151.57, 147.93, 135.30, 130.06, 128.86, 126.13, 125.15, 124.07, 121.59, 117.83, 116.30, 110.41, 81.76, 18.27.

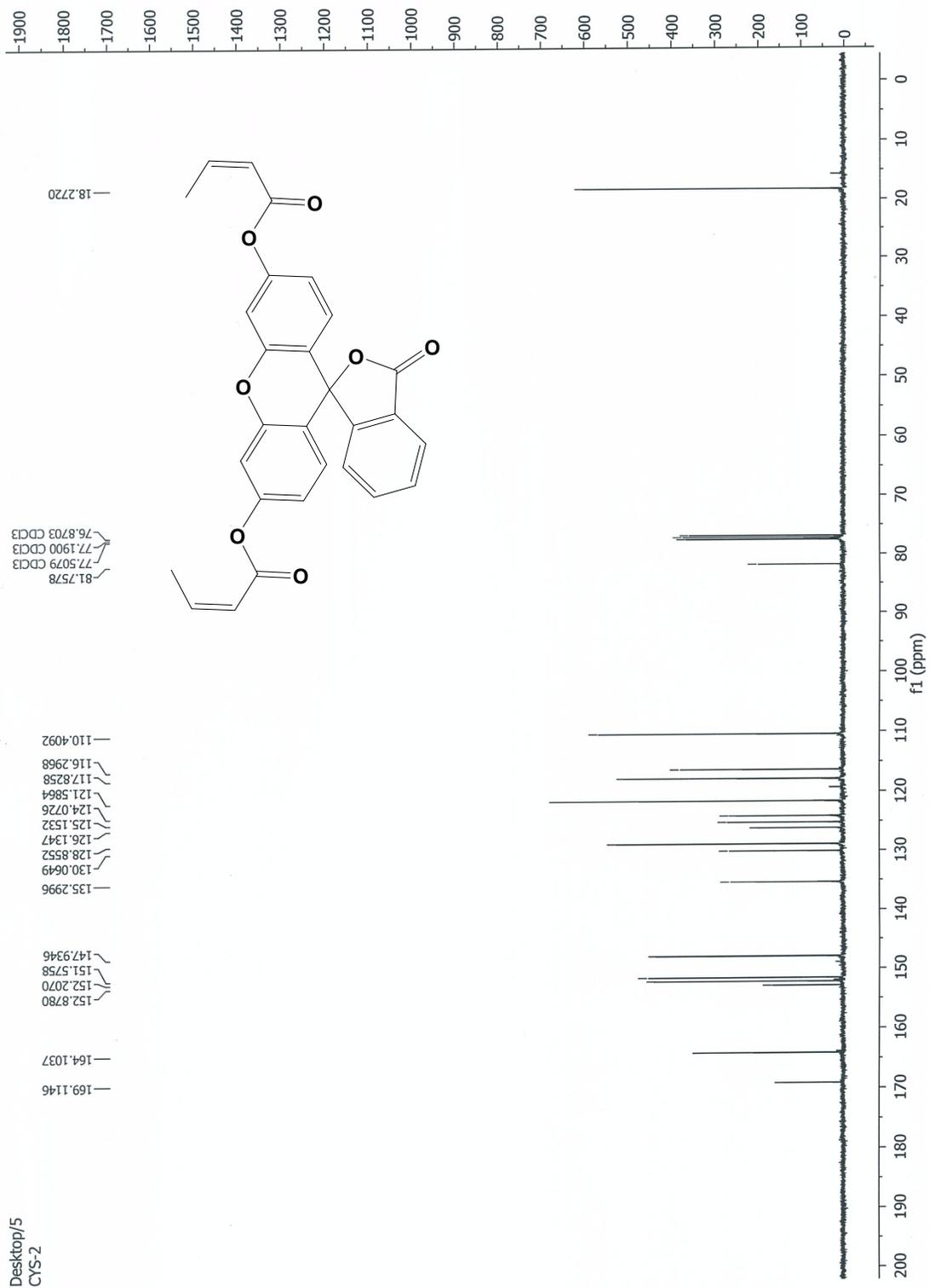
**ESI-MS: Fig. S3** probe-1 [M+Na]<sup>+</sup> = 491.1107 (calc.), 491.1101(exp.)

**ESI-MS: Fig. S4** of probe **1** + **L-cys** [M+H]<sup>+</sup> = 333.068 (calc.), 333.075 (exp.) [**Fluorescein**]

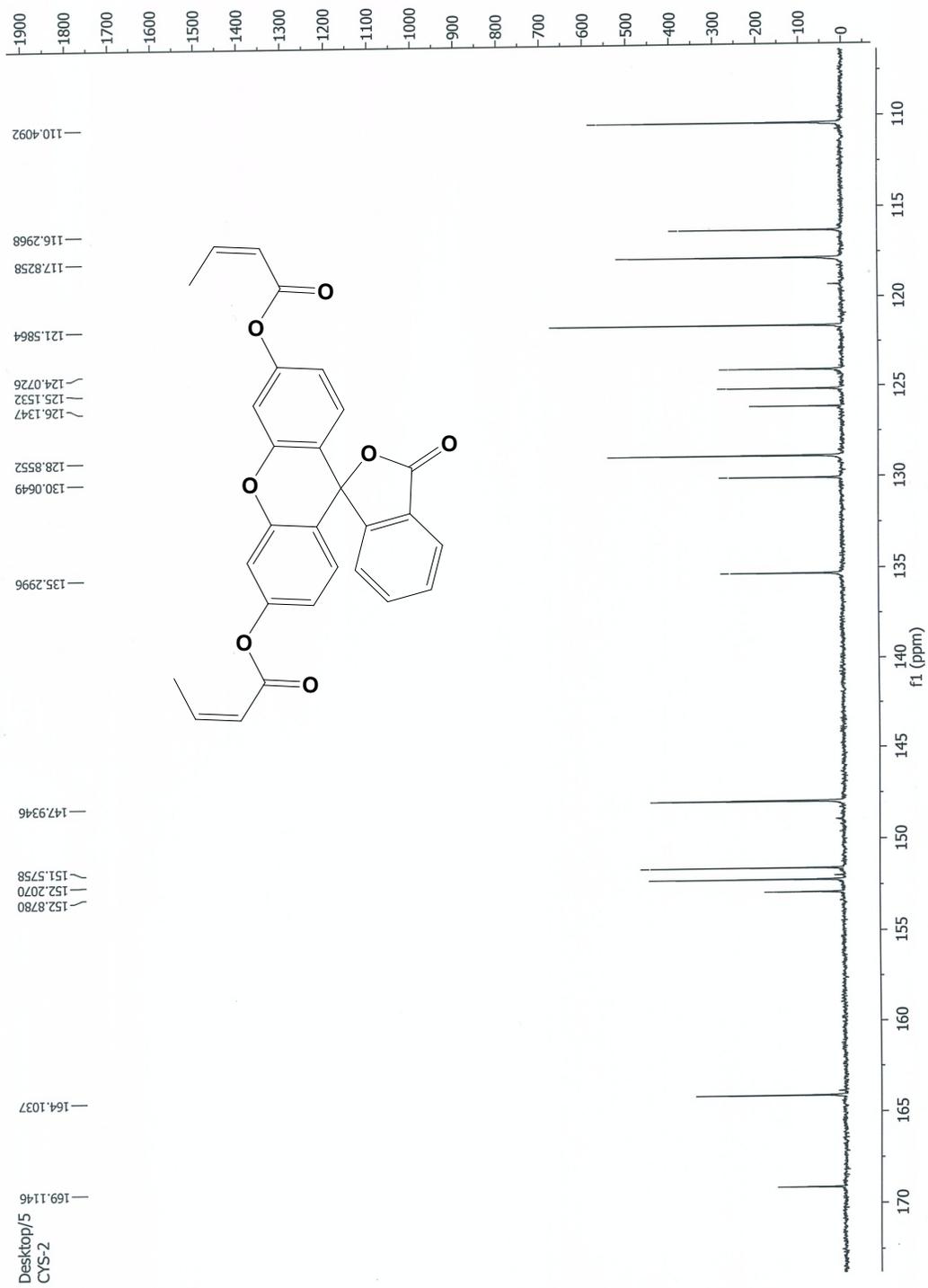
**Analysis of Cys in living cells.** A human neuroblastoma cell line, SH-SY5Y, was grown in a 1:1 mixture of Eagle's Minimum Essential Medium, and Ham's F12 Medium supplemented with 10 % FBS, 100 U/mL penicillin and 100 µg/mL streptomycin and incubated in a humidified 5 % CO<sub>2</sub> incubator at 37° C. Cells were seeded in a 24-well plate in ~80 % confluence for one day before the experiments. Cells were incubated with probe **1** (1~20 µM) or vehicle (DMSO) for 1 h, washed with PBS. For the *N*-ethyl maleimide (NEM) treated samples, cells were pre-incubated with NEM (0.25 mM) for 30 min. Cells were washed with PBS before fluorescence imaging. Fluorescence images of the cells were obtained by an epifluorescence microscope (Olympus, JP IX-71). The excitation source was a 490 nm laser and all images were taken under the same experimental conditions to minimize variations in fluorescence intensity.



**Fig. S1.** <sup>1</sup>H NMR spectrum of Probe 1.



**Fig. S2a.**  $^{13}\text{C}$  NMR spectrum of Probe 1.



**Fig. S2b.**  $^{13}\text{C}$  NMR spectrum of Probe 1 (EXPANDED VIEW).

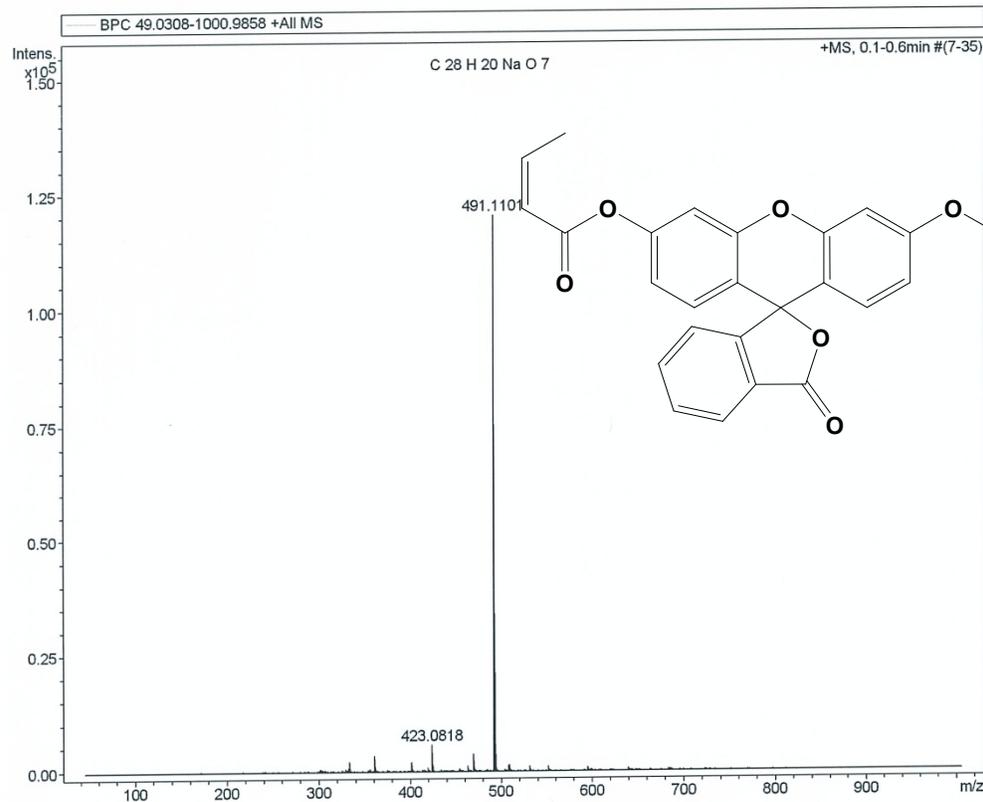
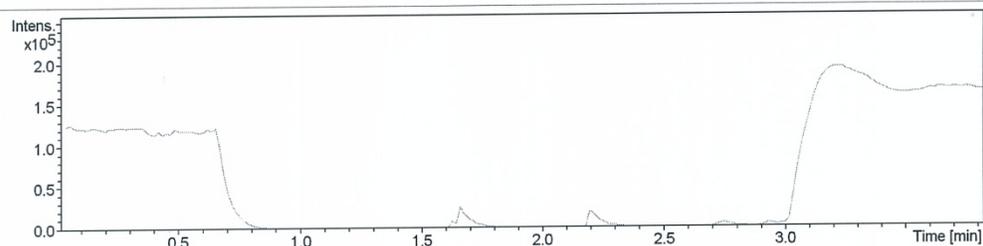
## Generic Display Report

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#Dilution solvent:MeOH

Acquisition Date 7/15/2013 3:51:47 PM

Operator BDAL@KR  
Instrument micrOTOF-Q



**Fig. S3.** ESI-mass spectrum of Probe 1.

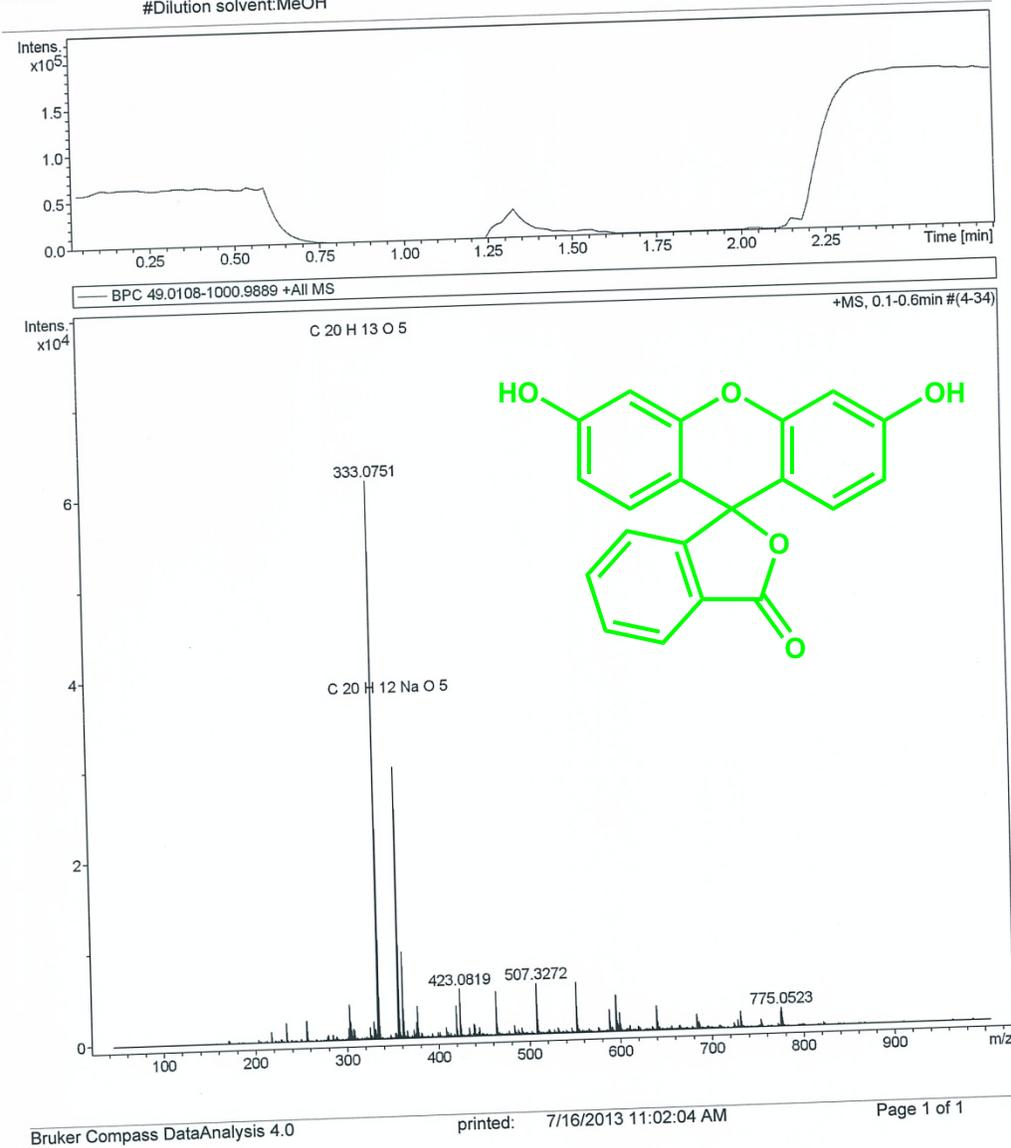
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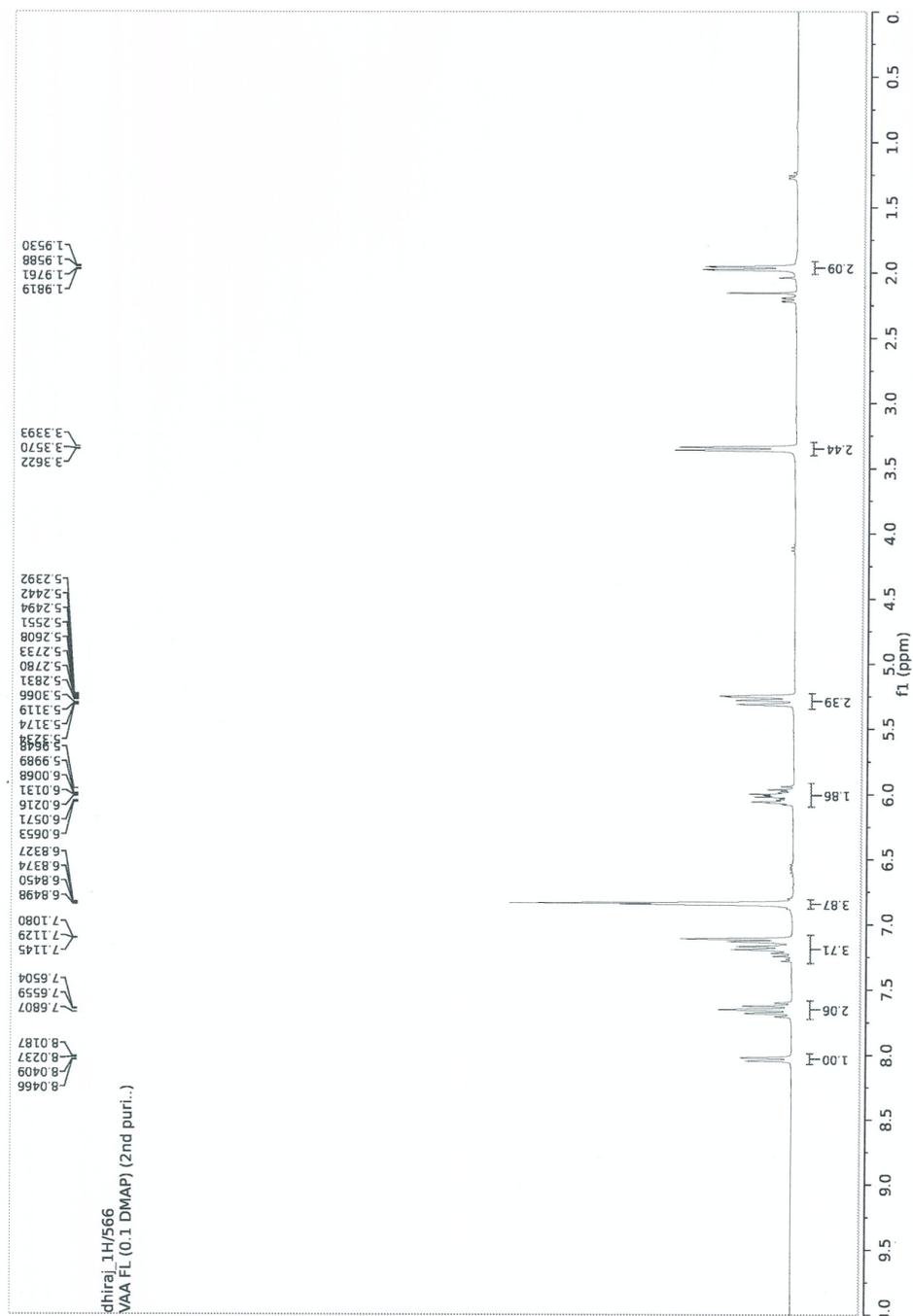
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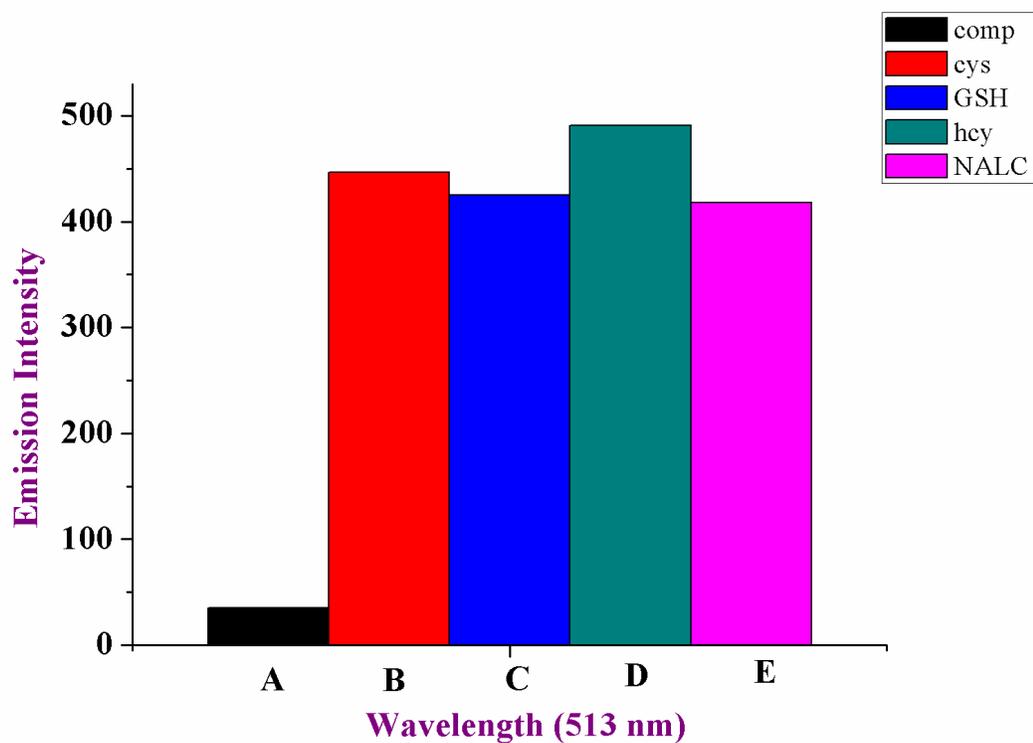
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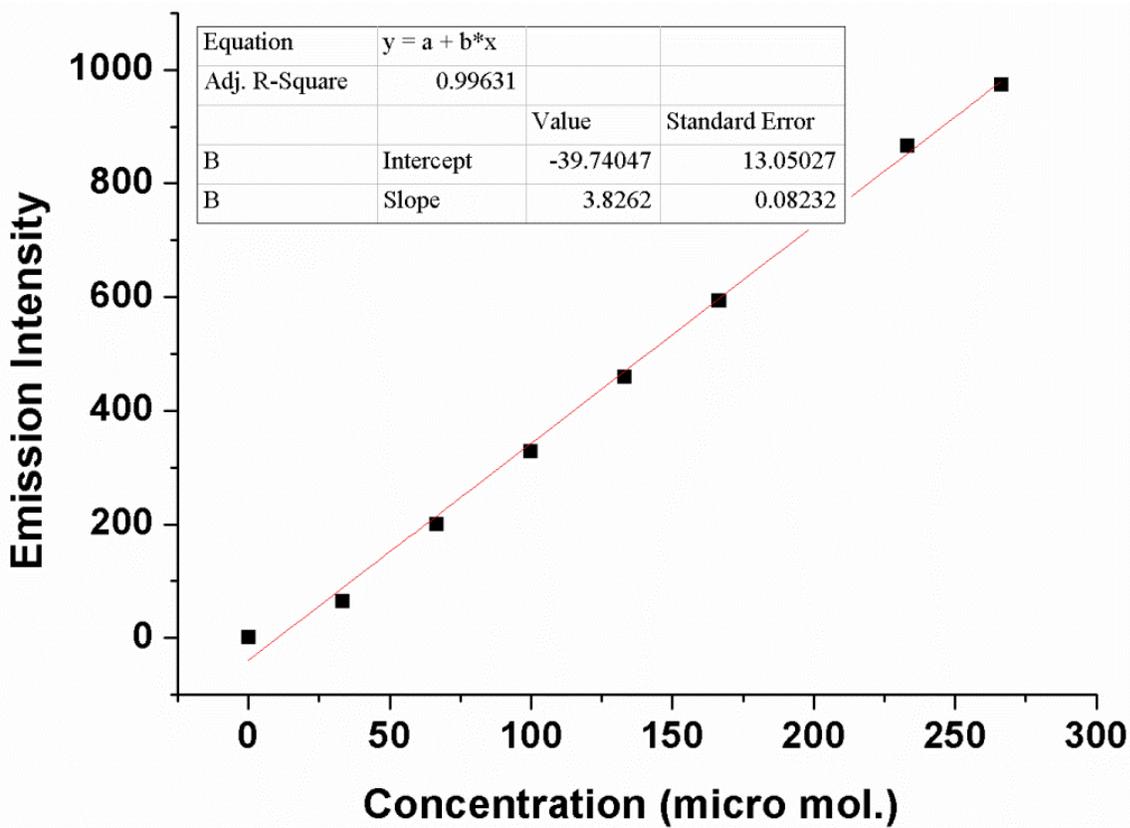
**Fig. S4.** ESI-mass spectrum of Probe 1 + dissolved cysteine. This data aided in identifying fluorescein as a final product.



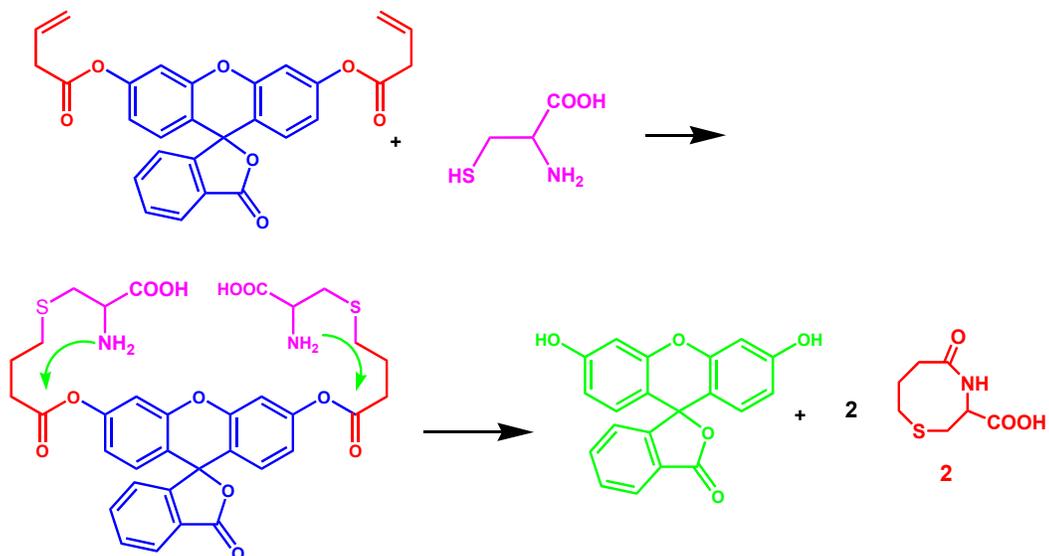
**Fig. S5.**  $^1\text{H}$  NMR spectrum of probe **1** and probe **1a** with 0.2 equiv of DMAP (4-dimethylaminopyridine).



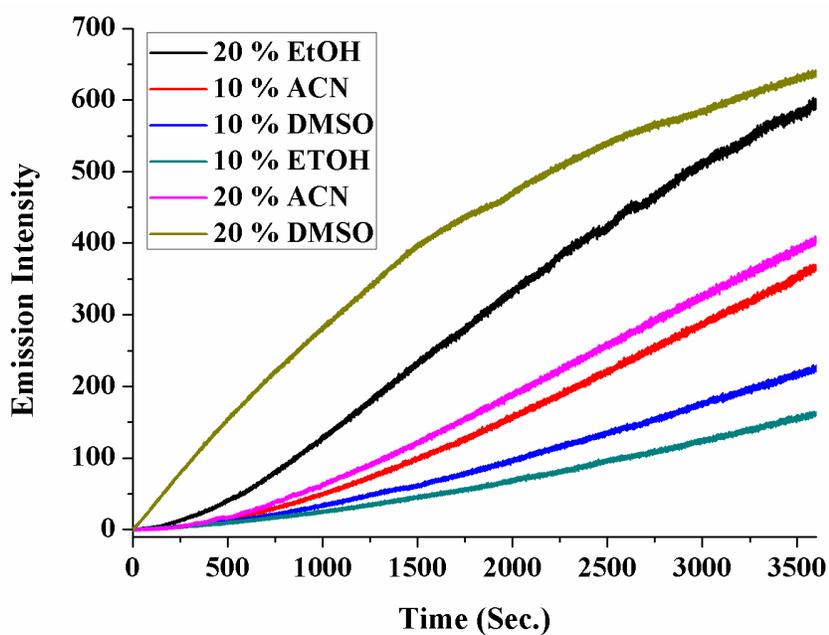
**Fig. S6.** Relative fluorescence intensity of compound **1** ( $4.0 \times 10^{-6}$  M, buffered H<sub>2</sub>O:DMSO 80:20; pH 7.4 PBS) with *Cys* (~10 equiv) with other amino acids (from left to right: A–compound, B–comp + *cys*, C–comp + *cys* + GSH, D–comp + *cys* + GSH, E–comp + *cys* + *N*-acetyl-*L*-*Cys* (~10 equiv).  $\lambda_{exci} = 490$  nm. Slit width = 1.5.



**Fig. S7.** Detection limit determinations for cysteine calculated from the linear fit (33.3 – 266  $\mu$ M in water).

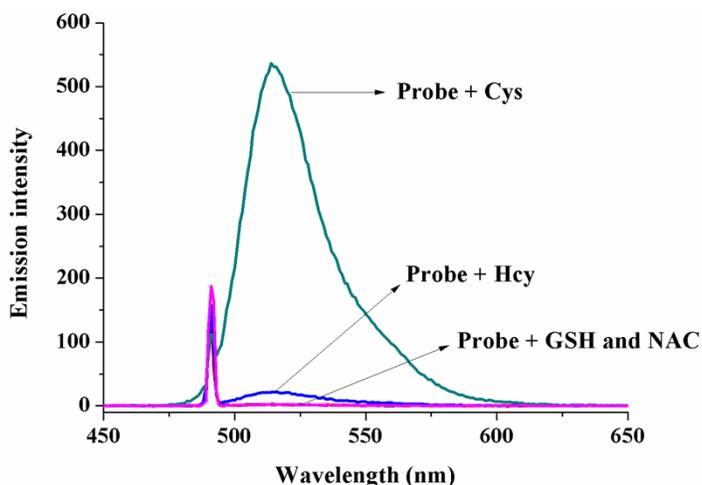


**Fig. S8.** Proposed sensing mechanism with probe 1a.

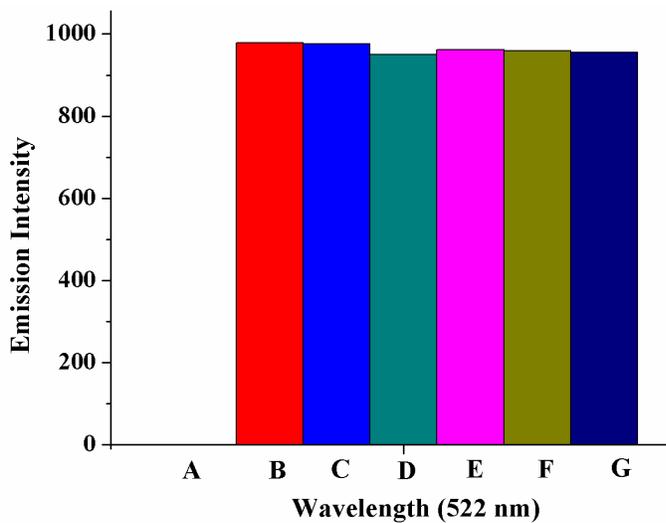


**Fig. S9.** Time-dependent emission spectra of probe ( $2.0 \times 10^{-6}$  M) with Cys (200  $\mu$ M in water)

$\lambda_{exc}$  = 490 nm) slit width Ex. & Em. = 1.5 nm.



**Fig. S10.** Emission spectra of probe ( $4.0 \times 10^{-6}$  M, buffered  $\text{H}_2\text{O}$ : DMSO 80:20; pH 7.4 PBS) with amino acids *L*-Cys, *Hcy*, *N*-acetyl-*L*-Cys and GSH ( $66 \mu\text{M}$  in water) incubated for 1 h at RT. Slit width for Ex. & Em. = 1.5 nm.



**Fig. S11.** Relative fluorescence intensity of probe-1 ( $4 \times 10^{-6}$  M,  $\text{H}_2\text{O}$ : DMSO 80:20;) with  $\text{KO}_2$  and the presence of other interfering ROS (A) comp, (B)  $\text{KO}_2$ , (C)  $\text{KO}_2$  + NaOCl, (D)  $\text{KO}_2$  +  $\text{H}_2\text{O}_2$ . (E)  $\text{KO}_2$  +  $\cdot\text{OH}$ , (F)  $\text{KO}_2$  +  $^t\text{BuOOH}$ , (G)  $\text{KO}_2$  +  $\cdot\text{O}^t\text{Bu}$ , ( $\sim 10$  equiv) incubated for 30 min at RT.  $\lambda_{\text{exci}} = 490$  nm.

**END**