

Electronic Supplementary Information

A handheld stamping process to fabricate microfluidic paper-based analytical devices with chemically modified surface for clinical assays

Paulo de Tarso Garcia^a, Thiago Miguel Garcia Cardoso^a, Carlos Diego Garcia^b, Emanuel Carrilho^{c,d}, and Wendell Karlos Tomazelli Coltro^{a,d}*

^aInstituto de Química, Universidade Federal de Goiás, 74001-970, Goiânia, GO, Brazil.

^bDepartment of Chemistry, The University of Texas at San Antonio, One UTSA Circle, San Antonio, TX, USA

^cInstituto de Química de São Carlos, Universidade de São Paulo, 13566-970, São Carlos, SP, Brazil

^dInstituto Nacional de Ciência e Tecnologia de Bioanalítica, 13084-971, Campinas, SP, Brazil

***Corresponding Author:**

Professor Wendell K. T. Coltro

Instituto de Química, Universidade Federal de Goiás

Campus Samambaia, 74001-970, P.O. Box 131

Goiânia, GO, Brazil

Fax: +55 62 3521 1097

E-mail: wendell@ufg.br

Paper surface nature with and without paraffin

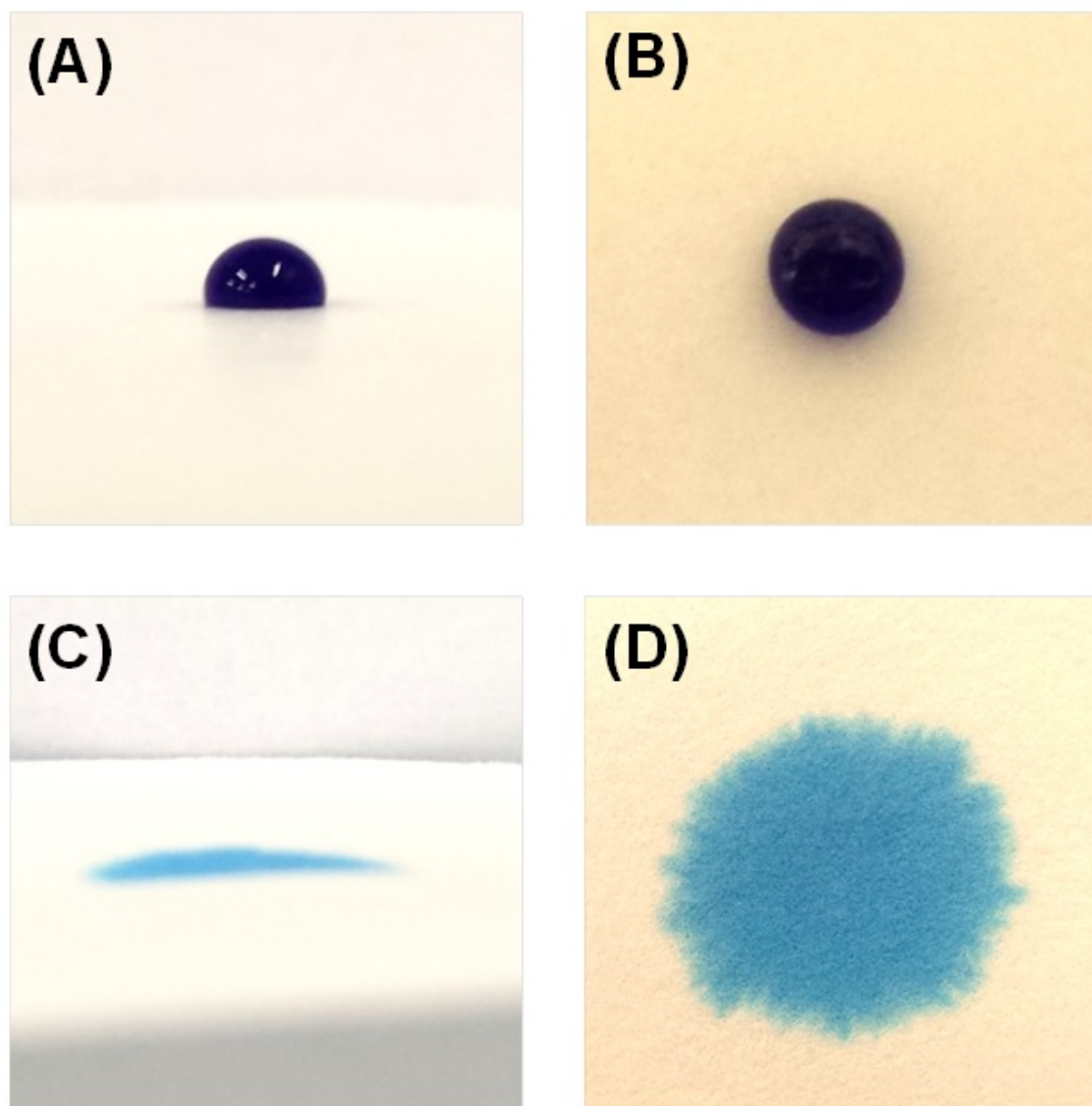


Fig. S-1 Optical images showing the (A, B) hydrophobic and (C, D) hydrophilic properties after and before the impregnation of paraffin on paper surface by the stamping process. Images were captured after the adding of 10 μL of a dye solution.

Lateral flow on μ PADs fabricated with different stamping times.

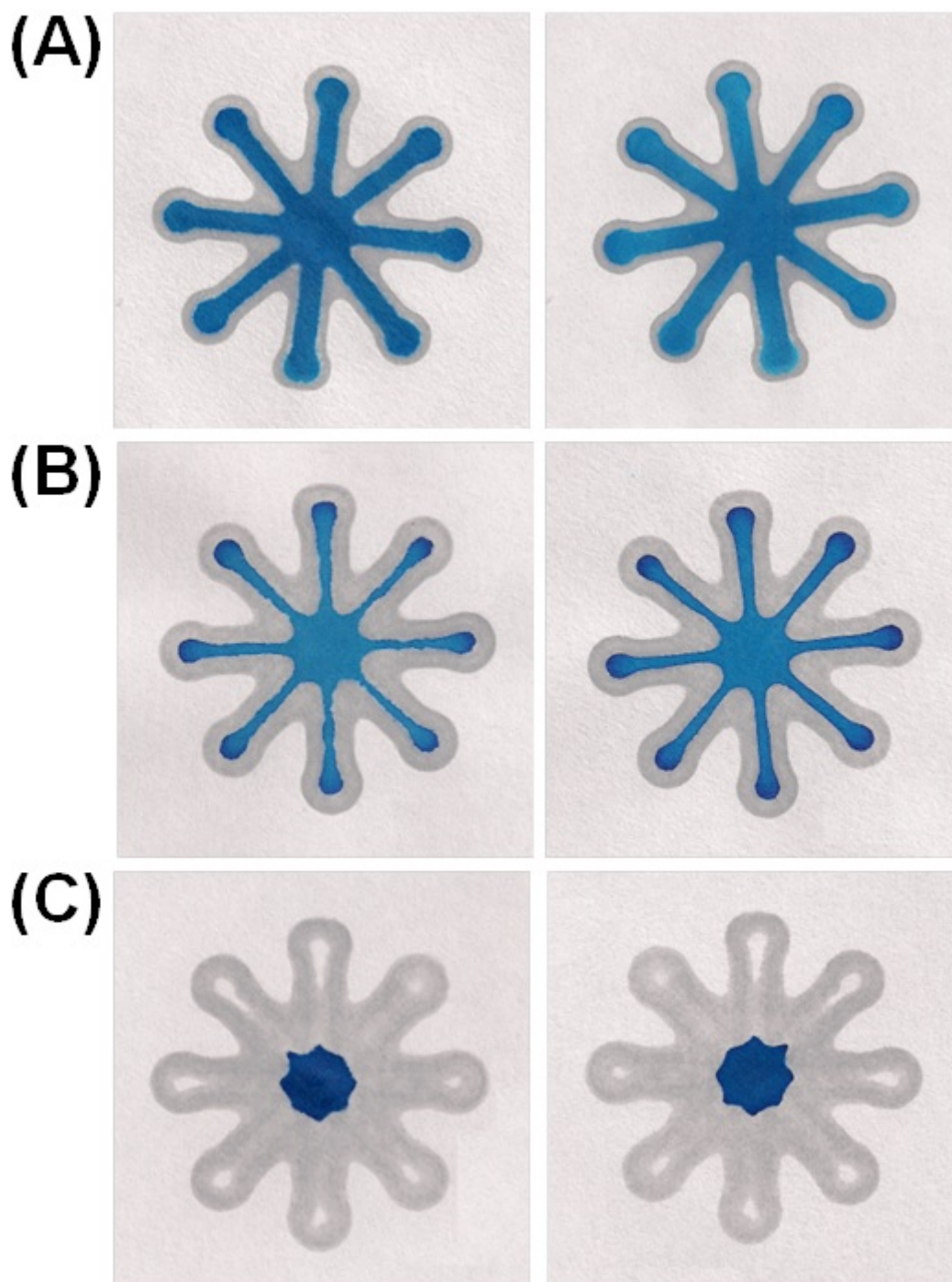


Fig. S-2 Optical images showing the filling of paper channels by lateral flow. Images (A), (B) and (C) depict the distribution of the sample on μ PADs stamped during 2, 5 and 10 s, respectively. Left and right images mean the back and the front views, respectively.

Effect of the stamping time on the amount of paraffin transferred on paper and device-to-device reproducibility

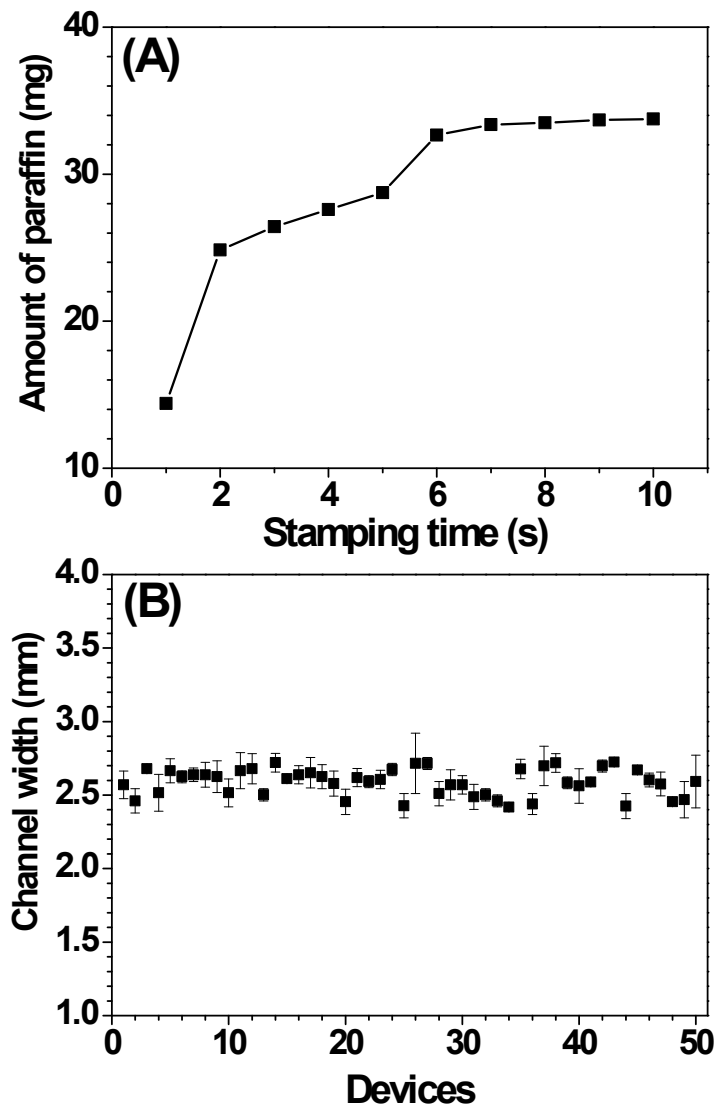


Fig. S-3 Presentation of (A) the amount of paraffin transferred on n-paper as function of the stamping time, and (B) channel width reproducibility for multiple stamped μ PADs. In (B), the stamping time was fixed in 2 s.

Spectroscopic Characterization

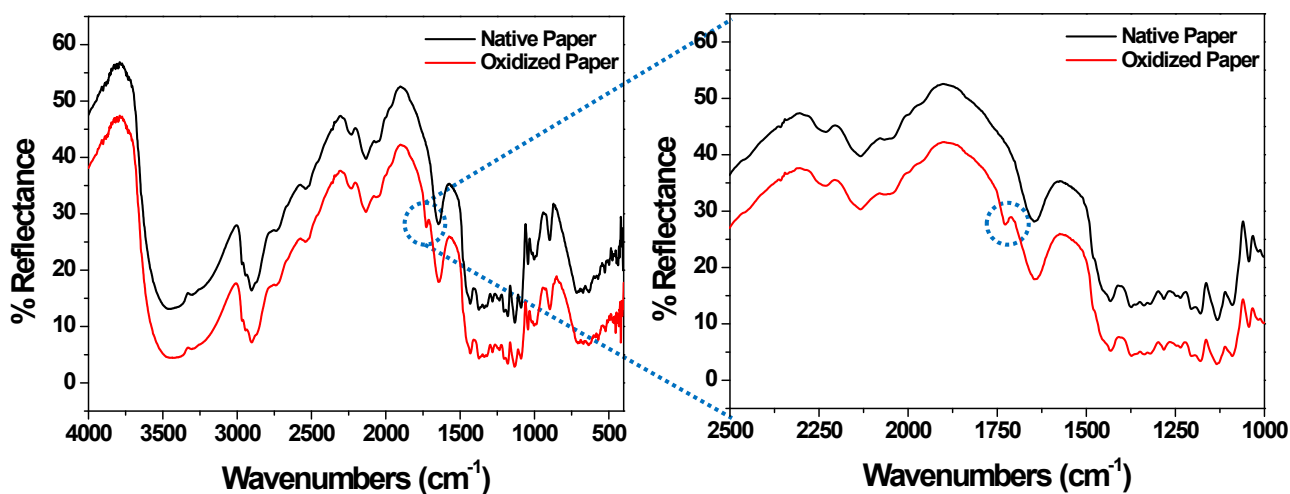


Fig. S-4 Diffuse reflectance infrared fouriertransform spectra for native and oxidized paper surfaces.

Simultaneous clinical assays

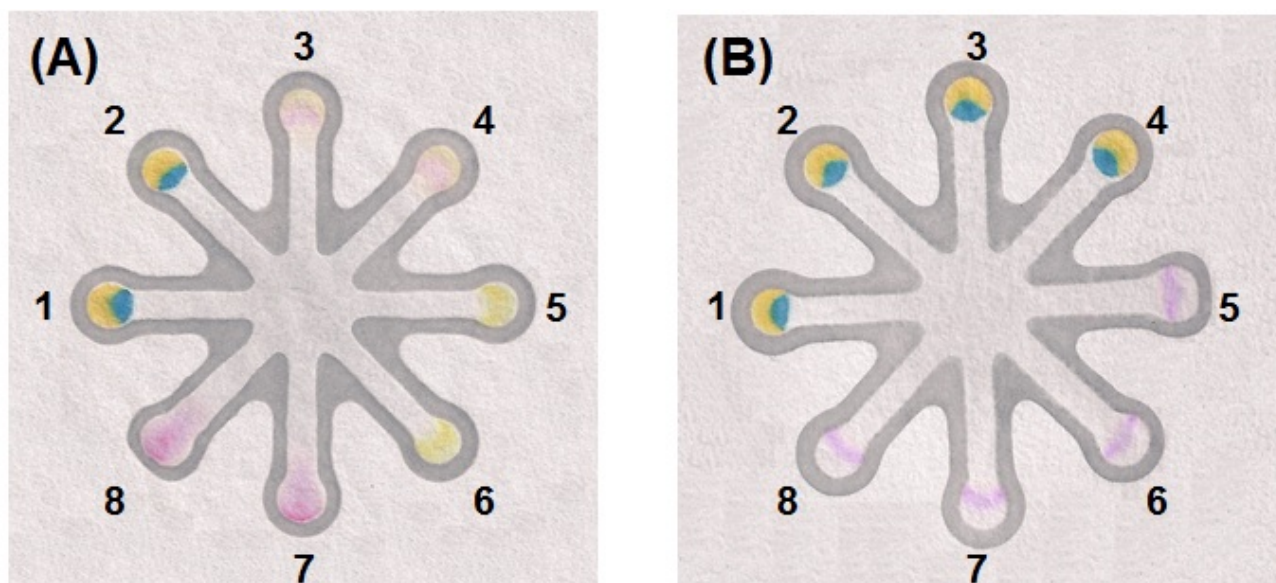


Fig. S-5 Scanned images of typical clinical assays performed on (A) oxidized and (B) native μ PADs. In (A), zones 1-2, 3-4, 5-6 and 7-8 display images for BSA, nitrite, glucose and uric acid, respectively. In (B), zones 1-4 and 5-8 indicate the sensing areas for BSA and nitrite, respectively.

Analytical performance

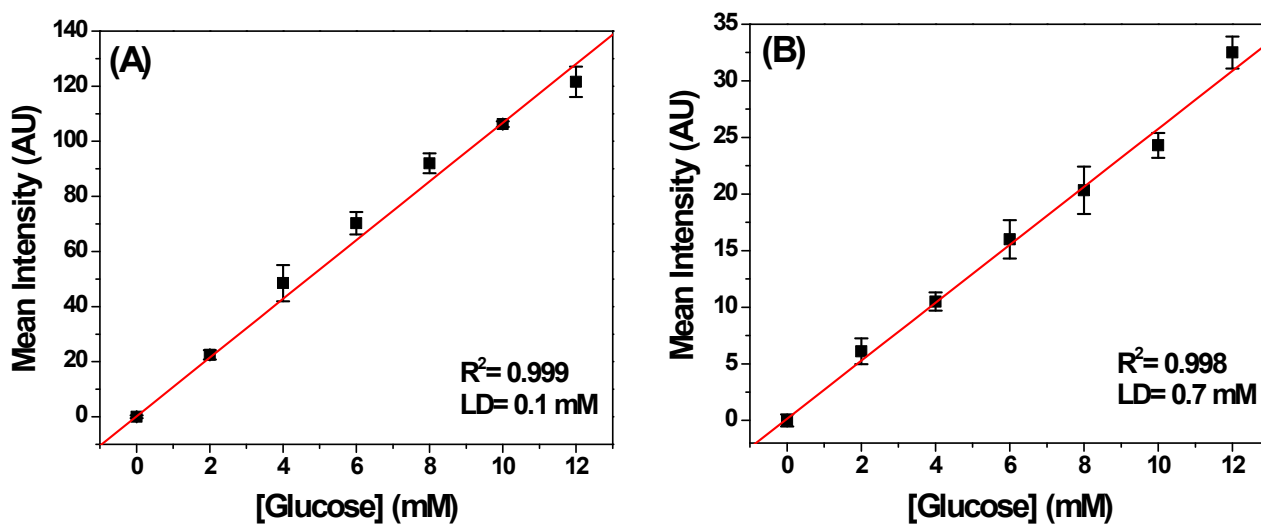


Fig. S-6 Analytical curves for glucose on (A) native and (B) oxidized μ PADs.