

Nanoassembly of an Amphiphilic Cyclodextrin and Zn(II)-Phthalocyanine with Potential for Photodynamic Therapy of Cancer

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S.1 Time-resolved fluorescence measurements

The excitation source was a synchronously mode-locked rhodamine 6G dye laser (Spectra Physics 375B) which provided excitation pulses of about 2 ps full width at half-maximum at a repetition rate of 82 MHz. An excitation wavelength of 570 nm was used. The fluorescence pulses were detected with a microchannel-plate photomultiplier (Hamamatsu R1645U-01, about 200 ps rising-time) and the decay profiles at 675 nm were collected with a computer-controlled multi-channel analyzer card (EG&G Ortec Trump-8k/2k). The

collected data were then analyzed using the non linear least-squares iterative reconvolution procedures based on the Marquardt algorithm. In the case of total fluorescence decay curves, the fitting was performed on the basis of the multiexponential decay law, ¹

$$I(t) = I_0 \sum_i \alpha_i \exp(-t / \tau_i) \quad (1)$$

where $I(t)$ is the total fluorescence decay curve, I_0 is the intensity at time zero, and α_i and τ_i are, respectively, the relative amplitudes and lifetimes of the i^{th} component (the normalization condition is $\sum_i \alpha_i = 1$). In the case of time-resolved anisotropy measurements, the reconvolution fitting procedure was

based on two steps. Fluorescence anisotropy $r(t)$ is defined using the following expression:

$$r(t) = \frac{I_{VV}(t) - I_{VH}(t)}{I_{VV}(t) + 2I_{VH}(t)} = \frac{D(t)}{S(t)} \quad (2)$$

where the sum data, $S(t)$, must be equal to the total intensity $I(t)$. In some cases, an additive constant r_∞ to $r(t)$ was introduced to take into account a long decay contribution due to static interaction with large clusters. In the simple case of spherical molecules, each rotational correlation time, τ_{Rj} , is related to the volume (V_j) of the rotating unit (or of the equivalent sphere) by the following equation:²

$$\tau_{Rj} = \frac{\eta V_j}{k_B T} \quad (3)$$

where η is the microviscosity of the medium, T is the temperature in Kelvin, and k_B is the Boltzmann constant.

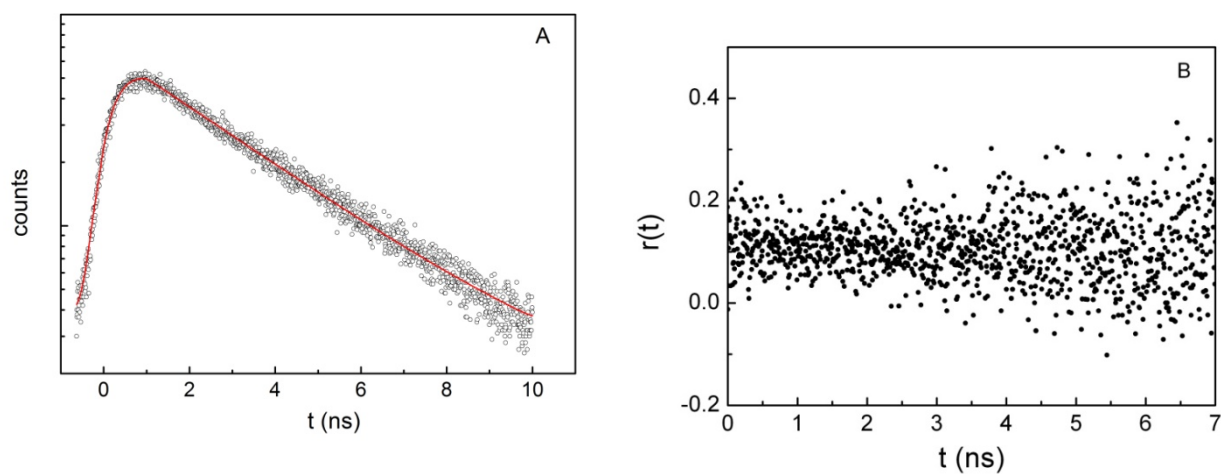


Figure S1. (A) Fluorescence time decay of ZnPc/SC16OH nanoassemblies dispersed in water ($\lambda_{\text{exc}}=570$ nm; $\lambda_{\text{em}}=675$ nm). The continuous red curve is the fit resulting from the convolution of the fluorescence decay and the excitation. (B) Time-resolved fluorescence anisotropy of ZnPc/SC16OH nanoassemblies dispersed in water (ZnPc concentration was fixed at $0.25 \mu\text{g/mL}$).

S.2 Depolarized fluorescence spectra

Depolarized fluorescence spectra were measured by using Equation (4):³

$$r = \frac{I_{VV}I_{HH} - I_{VH}I_{HV}}{I_{VV}I_{HH} + 2I_{VH}I_{HV}} \quad (4)$$

where r is the anisotropy and I_{VV} , I_{HH} , I_{VH} , and I_{HV} are the fluorescence intensities registered with different polarizer orientations (V=vertical, H=horizontal).

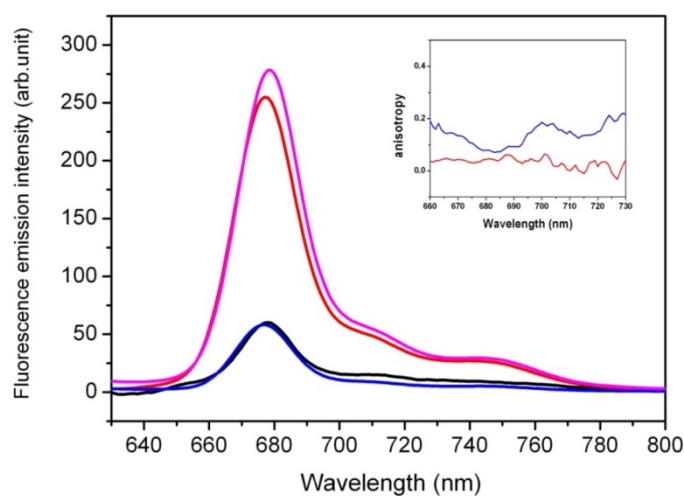


Figure S2: Steady-state fluorescence emission spectra of free ZnPc in DMSO (red trace), ZnPc/SC16OH nanoassemblies in DMSO (violet trace), ZnPc/SC16OH nanoassemblies in CHCl₃ (black trace) and ZnPc/SC16OH nanoassemblies in water (blue trace). ZnPc concentration was fixed at 0.25 $\mu\text{g/mL}$ in all the dispersions, $\lambda_{\text{exc}} = 600 \text{ nm}$. In the inset the anisotropy of ZnPc in DMSO (red trace) and ZnPc/SC16OH nanoassemblies in CHCl₃ (blue trace) is reported.

S3. Overall properties of Nile Red/SC16OH nanoassembly (Nr/SC16OH). SD was calculated on three different batches.

	Nr/SC16OH
Nile Red (% w/w)	0.2
Yield (%)	75
Mean D _H (nm ± SD)	195±8
P.I.	0.198
Zeta Potential (mV ± SD)	-28.3±3.5
Nile Red Actual loading ^a	0.199 ± 0.03 (71.2± 4.2)
(Ent. Efficiency ^b)	

^aActual loading is expressed as the amount of Nile Red (mg) encapsulated per 100 mg of nanoassembly; ^b ratio between actual and theoretical loading x 100.

References

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